TABLE 1. Profiles of Subjects in the Present Study

	n	Sex, n (Female/Male)	Age, Mean (SD), y
Healthy control	27	10/17	61 (13)
Alcoholic CP	20	0/20	59 (11)
Idiopathic CP	17	10/7	55 (15)
AIP	44	14/30	65 (9)
Without steroid	21	9/12	64 (14)
With steroid	23	5/18	67 (8)

development of IL-10–producing Tregs in vivo being found to be inducible costimulator (ICOS) dependent.^{28,29} Of note, a recent report revealed the existence of 2 functional subsets of human Foxp3⁺ Tregs, including ICOS⁺ cells, which produced high amounts of IL-10.³⁰ However, the relationship between Tregs in the peripheral blood and the pancreas remains unclear. In this study, we attempt to clarify that relationship by comparing Tregs in the peripheral blood and the pancreatic tissues.

MATERIALS AND METHODS

Subjects

We examined 44 patients with AIP at Kansai Medical University and its affiliated hospitals (21 untreated patients and 23 patients treated with corticosteroids; 14 women and 30 men; mean age, 65 years; range, 42-81 years). There were 20 patients with alcoholic pancreatitis (0 woman, 20 men; mean age, 59 years; range, 38-75 years), 17 patients with idiopathic pancreatitis (10 women, 7 men; mean age, 55 years; range, 20-72 years), and 27 healthy volunteers (10 women, 17 men; mean age, 61 years; range, 33-82 years), who served as control subjects. Nine AIP cases (6 women and 3 men; mean age, 54 years; range, 56-73 years) and 9 alcoholic pancreatitis cases (9 men; mean age, 53 years; range, 39-75 years) were surgery cases in that. All cases were collected between 1992 and 2009 (Table 1). All the AIP patients were diagnosed according to the clinical diagnostic criteria for AIP proposed by the Research Committee of Intractable Diseases of the Pancreas supported by the Japanese Ministry of Health, Labor, and Welfare,³¹ and the Asian criteria.³² Thirty-five AIP patients fulfilled Japanese diagnostic criteria 1 and 2. In brief, they exhibited diffuse enlargement of the pancreas and diffuse narrowing of the main pancreatic duct with an irregular wall. They also showed high levels of serum γ-globulin (>2 g/dL), IgG (>1800 mg/dL), and IgG4 (>135 mg/dL). Because 9 cases showed LPSP with a pancreatectomy specimen, AIP was diagnosed. The diagnosis of CP was made by imaging studies (ultrasound, computed tomography, endoscopic retrograde cholangiopancreatography, and magnetic resonance cholangiopancreatography), a pancreatic function test (secretin stimulation test), or histopathologic examination according to the clinical diagnosis criteria of the Japan Pancreatic Society.³³ Alcoholic CP was considered to be alcoholic in origin if the patient had a daily alcohol intake of more than 80 g/d for more than 10 years. Patients had a diagnosis of idiopathic CP when no apparent causes such as alcoholism, gallstones, or autoimmunity could be identified. This study was approved by the Kansai Medical University's ethics committee, and all patients and healthy volunteers gave informed consents.

Histopathology and Immunohistochemistry

Formalin-fixed and paraffin-embedded specimens were prepared and used for histopathologic and immunohistochemical studies. Sections measuring 4 μm were cut from each paraffin block and stained with hematoxylin and eosin, periodic acid-Schiff after diastase digestion, Azan-Mallory, reticulin, or orcein.

The remaining material was used for immunohistochemical analysis. Immunoglobulin G4 immunostaining was performed using a monoclonal antibody for human IgG4 (ZYMED Laboratories, San Francisco, Calif). Immunoglobulin 1 and Foxp3 immunostaining was performed by avidin-biotin complex method with reagents provided by Vector Laboratories (Burlingame, Calif). The antibodies used to identify the inflammatory cells in the pancreas were IgG1 antibodies (Binding Site, Birmingham, UK) and Foxp3 (eBioscience, San Diego, Calif). The deparaffinized sections were pretreated in an EDTA buffer (pH 8.0) in a pressure cooker at 100°C for 5 minutes. After incubation with the first antibody at 4°C overnight, biotinylated rabbit anti-sheep serum IgG (Vector) was used as the secondary antibody (sections for IgG1), and immunoreactive deposits were visualized with 3,3diaminobenzidine tetrahydrochloride. Cells positive for IgG1, IgG4, and Foxp3 were counted under 5 different high-power fields with intense inflammation. The ratios between IgG1, IgG4, Foxp3-positive cells and infiltrated mononuclear cells were calculated in each case by 2 pathologists.³⁴

Flow Cytometric Analysis of Tregs in Peripheral Blood Cells

Flow cytometric analysis was performed according to the method described by Okazaki et al. ³⁵ Briefly, cells were stained with fluorescein isothiocyanate–conjugated anti-CD4 (BD Bioscience, Franklin Lakes, NJ), PE-Cy7–conjugated anti-ICOS (CD278) (eBioscience), and allophycocyanin-conjugated anti-CD25 (BD Bioscience), for Tregs at 4°C for 30 minutes. After staining, the cells were fixed in 1% paraformaldehyde, and 3 color flow cytometric analyses were performed using FACS-LSR (BD Biosciences) and FlowJo (FlowJo LLC, Ashland, Ore).

Flow Cytometric Analysis of Intracellular IL-10

Intracellular staining of IL-10 was performed according to the manufacturer's instruction. In brief, cells magnetically labeled with CD4 and CD25 microbeads (CD4⁺CD25^{high} Treg Isolation Kit; Miltenyi Biotec, Bergisch Gladbach, Germany) were loaded onto auto MACS (Miltenyi Biotec). The cells were then washed with phosphate-buffered saline before analysis. For intracellular staining, CD4⁺CD25^{high} Tregs were stimulated with phorbol 12myristate 13-acetate, ionomycin for 2 to 4 hours before adding Golgi stop/Golgi plug solution (BD Biosciences). The cells were then stained with fluorescein isothiocyanate-conjugated anti-CD4, allophycocyanin-conjugated anti-CD25, and PE-Cy7-conjugated anti-ICOS (CD278) (eBioscience). After fixation with a fixation buffer (eBioscience) for 20 minutes at 4°C, the cells were permeabilized with a permeabilization buffer (eBioscience) and counterstained with PE-conjugated anti-IL-10 for 30 minutes. Stained cells were analyzed using FACS-LSR (BD Biosciences) and FlowJo (FlowJo LLC).

Statistical Analysis

For all studies, data are expressed as mean (SD); differences were analyzed using the nonparametric Mann-Whitney rank test and Fisher exact test, where P < 0.05 was considered significant.

RESULTS

Immunohistochemical Findings of IgG1 and IgG4

The ratio of IgG4-positive plasma cells to infiltrated mononuclear cells (IgG4/Mono) was significantly higher in AIP (n = 9, 0.122 [SD, 0.049]) than in alcoholic CP (n = 9, 0.013 [SD, 0.007]; P < 0.05; Figs. 1–3). The ratio of IgG1-positive plasma cells to infiltrated mononuclear cells (IgG1/Mono) was significantly lower in AIP (n = 9, 0.041 [SD, 0.024]) than in

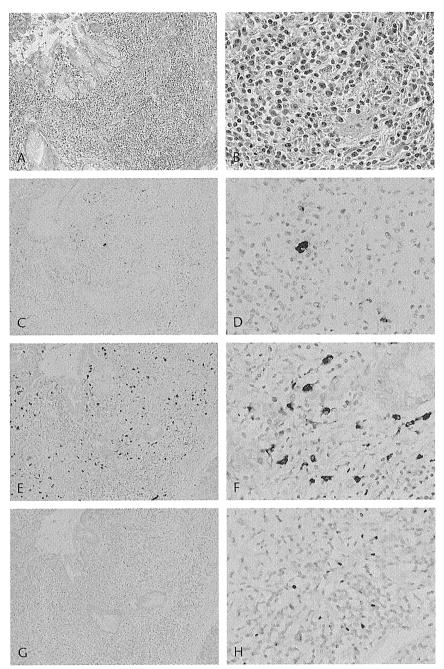


FIGURE 1. Histopathologic and immunohistochemical findings of the pancreas in LPSP. Hematoxylin-eosin staining (original magnification $\times 100$ [A], $\times 400$ [B]). Lymphoplasmacytic sclerosing pancreatitis showed interlobular fibrosis, atrophic pancreatic lobules, and infiltration of lymphoplasmacytes. Immunoglobulin 1–positive cells (original magnification $\times 100$ [C], $\times 400$ [D]) were scattered, whereas IgG4-positive (original magnification $\times 100$ [E], $\times 400$ [F]) and Foxp3-positive cells (original magnification $\times 100$ [G], $\times 400$ [H]) were abundant.

alcoholic CP (n = 9, 0.085 [SD, 0.011]; P < 0.05; Figs. 1–3). The ratio of IgG4/Mono to IgG1/Mono (IgG4/G1) was significantly higher in AIP (n = 9, 2.92 [SD, 1.42]) than in alcoholic CP (n = 9, 0.16 [SD, 0.08]; P < 0.05; Figs. 1–3A–C).

Immunohistochemical Findings of Foxp3

The ratio of Foxp3-positive cells to infiltrated mononuclear cells (Foxp3/Mono) in patients with AIP (n = 9, 0.091

[SD, 0.023]) was significantly higher than alcoholic CP (n = 9, 0.012 [SD, 0.003]; P < 0.05; Figs. 1–3D).

Correlation Between Foxp3/Mono and IgG4/Mono in Patients With AIP

In the group of patients with AIP, Foxp3/Mono and IgG4/Mono were positively correlated (n = 9, P < 0.05; R = 0.91) (Fig. 4).

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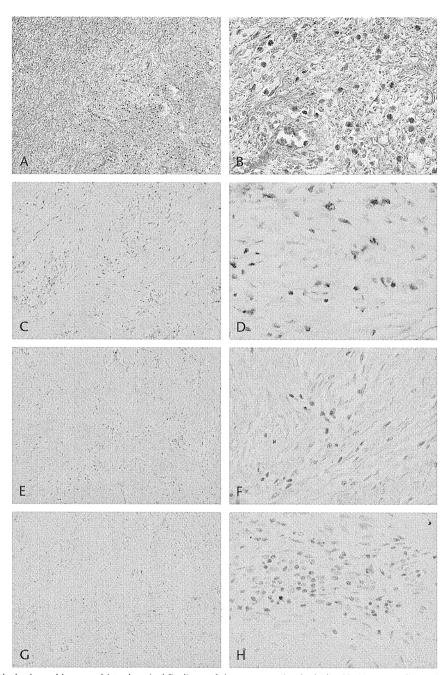


FIGURE 2. Histopathologic and immunohistochemical findings of the pancreas in alcoholic CP. Hematoxylin-eosin staining (original magnification $\times 100$ [A], $\times 400$ [B]). Photomicrograph of alcoholic CP showed dense fibrosis that surrounded atrophic or normal-appearing lobules. Some lobules were divided or replaced by dense fibrous tissue that extended from perilobular fibrosis bundles. Immunoglobulin 4–positive (original magnification $\times 100$ [E], $\times 400$ [F]) and Foxp3-positive cells (original magnification $\times 100$ [G], $\times 400$ [H]) were scattered, whereas IgG1-positive cells (original magnification $\times 100$ [C], $\times 400$ [D]) were abundant.

Circulating CD4⁺CD25^{high}Tregs Were Higher in Patients With AIP

CD4⁺CD25^{high} Tregs (%CD4⁺CD25^{high} Tregs of CD4⁺) were significantly higher in AIP patients (n = 31, 4.99% [SD, 2.70%]) compared with those with alcoholic CP (n = 11, 3.49% [SD, 1.43%]), those with idiopathic CP (n = 17, 2.24% [SD, 1.01%]), and the healthy control group (n = 16, 2.60% [SD, 1.05%], P < 0.05) (Figs. 5 and 6). There was no difference

in these Tregs between AIP patients who underwent steroid therapy (n = 22, 5.12% [SD, 2.84%]) and those who did not (n = 9, 4.52% [SD, 2.40%]).

Circulating ICOS⁺ Tregs Were Higher in Patients With AIP

There was no difference in these ICOS⁺ lymphocytes (% ICOS⁺ cells of lymphocytes) between patients with AIP

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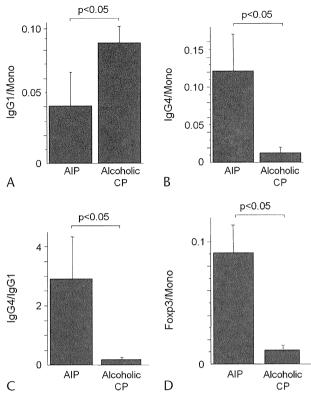


FIGURE 3. The ratio of Foxp3-, IgG1-, and IgG4-positive cells to infiltrated mononuclear cells (Foxp3/Mono, IgG1/Mono, IgG4/Mono) in patients with AIP (n = 9) and alcoholic CP (n = 9). Foxp3/Mono and IgG4/Mono in patients with AIP were significantly increased compared with alcoholic CP. Immunoglobulin 1/Mono in patients with AIP was significantly increased compared with alcoholic CP.

(n = 31, 0.40% [SD, 0.36%]), alcoholic CP (n = 11, 0.43%)[SD, 0.26%]), and idiopathic CP (n = 17, 0.49%) [SD, 0.30%]) and the healthy control group (n = 16, 0.35% [SD, 0.21%]) (Fig. 7A). However, the ratio of ICOS⁺ Tregs was significantly higher in the AIP patients. Inducible costimulator-positive Tregs (% ICOS+ Tregs of lymphocytes) were significantly increased in patients with AIP (n = 31, 0.08% [SD, 0.01%]) compared with those with alcoholic CP (n = 11, 0.03% [SD, 0.02%]), those with idiopathic CP (n = 17, 0.02% [SD, 0.01%]), and the healthy control group (n = 16, 0.02% [SD, 0.01%], P < 0.05) (Fig. 7B). Inducible costimulator-positive Tregs (% ICOS) Tregs of CD4⁺) were significantly increased in patients with AIP (n = 31, 0.18% [SD, 0.15%]) compared with those with alcoholic CP (n = 11, 0.07% [SD, 0.06%]), those with idiopathic CP (n = 17, 0.04% [SD, 0.04%]), and the healthy control (n = 16, 0.04% [SD, 0.03%], P < 0.05) (Fig. 7C). Inducible costimulator-positive Tregs (% ICOS+ Tregs of total Tregs) were significantly increased in patients with AIP (n = 31, 3.45%[SD, 1.58%]) compared with those with alcoholic CP (n = 11, 1.71% [SD, 0.98%]), those with idiopathic CP (n = 17, 1.80%) [SD, 0.86%]), and the healthy control group (n = 16, 1.57%[SD, 0.61%], P < 0.05) (Figs. 5, 7D). There was no difference in these ICOS[‡] Tregs between AIP patients who underwent steroid therapy (n = 22, 3.51% [SD, 1.57%]) and those who did not (n = 9, 3.31% [SD, 1.68%]).

A Comparison of the IL-10-Producing Ability Between AIP Patients and the Healthy Control Group

IL- 10^+ Tregs (% IL- 10^+ Tregs of total Tregs) were significantly higher in AIP patients (n = 16, 3.81% [SD, 1.52%]), compared with the healthy control group (n = 11, 1.38% [SD, 0.64%], P < 0.05) (Figs. 8 and 9). The ratio of IL- 10^+ Tregs and ICOS⁺ Tregs of total Tregs was significantly higher in AIP patients compared with the healthy control group (Figs. 5 and 8). The ratio that ICOS⁺ Tregs express IL-10 (n = 16, 11.92% [SD, 3.73%]) in the AIP group is higher in significance than the ratio that ICOS⁻ Tregs express IL-10 (n = 16, 1.61% [SD, 3.73%]) in the AIP group (Fig. 10).

DISCUSSION

Autoimmune pancreatitis is accepted worldwide as a distinctive type of pancreatitis, which contains 2 forms, IgG4-related AIP (type 1 AIP, LPSP type) and AIP with GEL (type 2 AIP, IDCP type). ¹⁴ Although the precise mechanism remains unclear, autoimmune mechanisms are suspected to be involved in the development of both types of AIP. In addition to pancreatitis, patients with IgG4-related AIP often manifest extrapancreatic lesions such as biliary lesions, sialadenitis, retroperitoneal fibrosis, enlarged celiac and hilar lymph nodes, chronic thyroiditis, and interstitial nephritis, ¹⁰ suggesting that IgG4-related AIP may be the pancreatic manifestation of a systemic disorder, IgG4-related disease. ⁵ On the other hand, ulcerative colitis is sometimes associated with AIP with GEL (IDCP or type 2 AIP). ¹²

Patients with IgG4-related AIP are known to show higher levels of serum IgG4 than patients with other pancreatic diseases or healthy control subjects. 4,5 However, the mechanism of increased IgG4 production is still unclear. Previous reports showed that both activated $\mathrm{CD4}^+$ and $\mathrm{CD8}^+$ T cells increased significantly in the peripheral blood and infiltrated into the pancreas more predominantly than B cells, in addition to abundant infiltration of IgG4-positive plasma cells. $\mathrm{CD4}^+$ T cells differentiate from naive T cells ($\mathrm{T_H0}$) to T-helper 1 ($\mathrm{T_H1}$), $\mathrm{T_H2}$, $\mathrm{T_H17}$, and Tregs. 36 In the peripheral blood of AIP patients, $\mathrm{T_H1}$ cells are predominant over $\mathrm{T_H2}$ cells, 35 whereas $\mathrm{T_H2}$ dominates in the

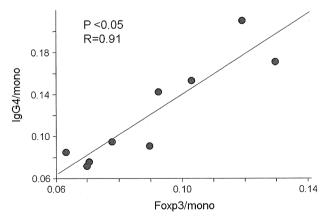


FIGURE 4. Correlation between the ratio of Foxp3-positive cells to infiltrated mononuclear cells (Foxp3/Mono) and the ratio of IgG4-positive cells to infiltrated mononuclear cells (IgG4/Mono) in patients with AIP. Foxp3/Mono and IgG4/Mono are positively correlated (n = 9, P < 0.05; R = 0.91).

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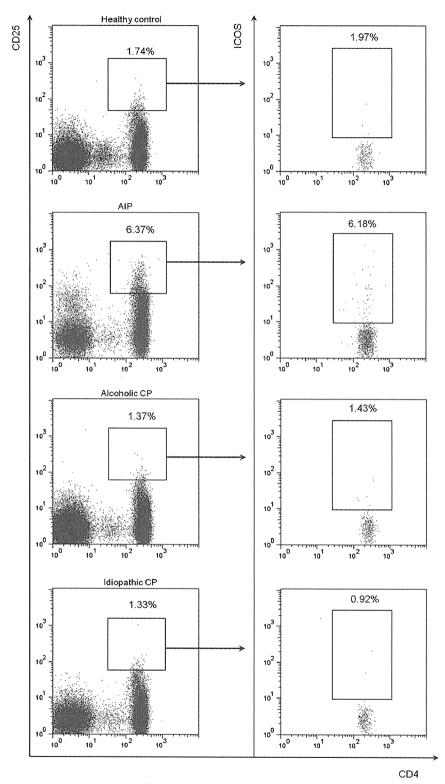


FIGURE 5. Flow cytometric analysis of CD4⁺CD25^{high} Tregs and ICOS⁺ Tregs. Human peripheral blood CD4⁺CD25^{high} Tregs separated into ICOS⁺ subpopulations. In AIP patients (n = 31), the number of CD4⁺CD25^{high} Tregs and ICOS⁺ Tregs increased compared with healthy control (n = 16), alcoholic CP (n = 11), and idiopathic CP (n = 17) group.

involved organs. 37 This discrepancy is possibly caused by the shift of $T_{\rm H2}$ cells from the periphery to local tissues, or different disease stages.

Different from $T_{\rm H}1/T_{\rm H}2$ immune balances, the role of Tregs in AIP is still unclear. Animal models have demonstrated that decreased Tregs induce various autoimmune diseases including

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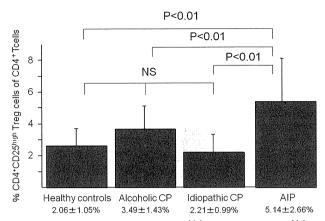


FIGURE 6. Percentage of CD4⁺ CD25^{high} Tregs. CD4⁺CD25^{high} Tregs were significantly increased in patients with AIP (n = 31, 4.99% \pm 2.70%) compared with alcoholic CP (n = 11, 3.49% \pm 1.43%), idiopathic CP (n = 17, 2.24% \pm 1.01%), and healthy control groups (n = 16, 2.60% \pm 1.05%, P < 0.05).

pancreatitis models. ³⁸ Recent studies of immune tolerance and allergy showed that high doses of antigen exposure cause both immune deviation of $T_{\rm H}2$ response in favor of a $T_{\rm H}0/T_{\rm H}1$, and the generation of IL-10– and transforming growth factor β (TGF- β)–producing Tregs. ³⁹ High-dose antigen exposure inhibits the usual $T_{\rm H}2$ T-cell response activation and/or maintenance. In addition, interferon γ and IL-10 induce preferential switching of B-cell response in favor of producing IgG and IgG4 antibodies, respectively, and possibly IgA antibodies under the influence of TGF- β . ⁴⁰ Recent reports showed that CD4 $^+$

CD25^{high}Foxp3⁺ Tregs also produce IL-10.⁴¹⁻⁴³ In general, high amounts of IL-10-producing Tregs are well known as type 1 regulatory (Tr1) cells. These Tregs also produce TGF-β. There is some evidence in adult humans that constitutive CD4+CD25high Tregs and inducible IL-10- and TGF-B-secreting Tr1 cells represent overlapping populations, based on CD25 expression on CD4⁺ Tr1 cells.³⁹ CD4⁺CD25^{high} Tregs also produce IL-10 to educate the antigen-presenting cells.⁴⁴ Akitake et al⁴⁵ reported that peripheral blood mononuclear cells (PBMCs) isolated from a patient with AIP exhibited enhanced production of IgG4 and IL-10 upon stimulation with Toll-like receptor ligands, compared with those from a healthy control. We did not examine immunological relations with Toll-like receptor in AIP patients, but this will need examination in the future. Our previous findings showed increasing inducible memory Tregs but decreasing naive Tregs in the peripheral blood of patients with IgG4-related AIP.^{25,46} In another study of IgG4-related sclerosing cholangitis, prominent Tregs infiltration was observed in the liver biopsy specimens. 25,34,3°

In 1999, Hutloff et al⁴⁷ reported that ICOS is a costimulatory molecule in the CD28 family, whose expression is induced during the activation of CD4 T cells. Most notably, it is expressed on the T-cell subset found in B-cell follicles. It is also associated with the production of IL-10 by T cells. This is also associated with the production of IL-10 by T cells. This is expressed on resting B cells, dendritic cells, and activated monocytes. Inducible costimulator–positive expression is low on naive human and murine cells and is up-regulated within hours after T-cell receptor engagement. At all the costimulator expression appears to be higher on $T_{\rm H2}$ CD4 $^+$ T cells than on $T_{\rm H1}$ -CD4 $^+$ T cells. After activation, ICOS expression persists on recently activated as well as memory $T_{\rm H1}$ and $T_{\rm H2}$ CD4 $^+$ T cells.

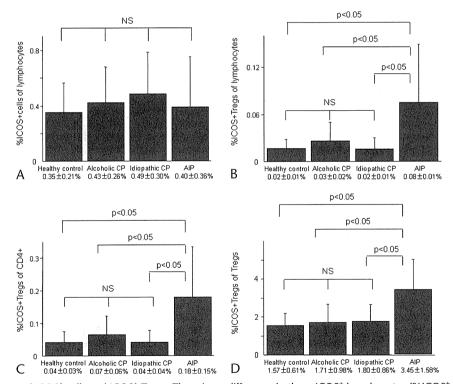


FIGURE 7. Percentage of ICOS⁺ cells and ICOS⁺ Tregs. There is no difference in these ICOS⁺ lymphocytes (%ICOS⁺ cells of lymphocytes) between patients with AIP (n=31), alcoholic CP (n=11), idiopathic CP (n=17), and healthy control (n=16). Inducible costimulator–positive Tregs (%ICOS⁺ Tregs of lymphocytes, CD4⁺ cells, and Tregs) were significantly increased in patients with AIP compared with those with alcoholic CP, those with idiopathic CP, and healthy control (P < 0.05).

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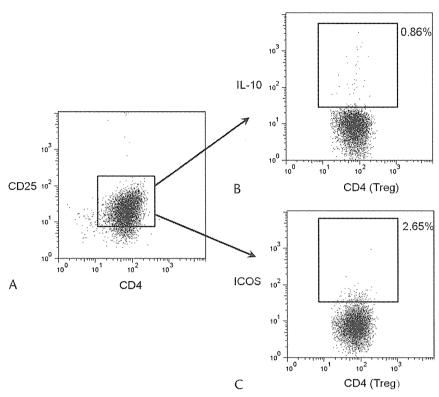


FIGURE 8. Flow cytometric analysis of Tregs (A) that express IL-10 (B) and ICOS (C) in a healthy control. The ratio of IL-10⁺ Tregs in total Tregs (%IL-10⁺ Tregs) was 0.86% (B). The ratio of ICOS⁺ Tregs in total Tregs (%ICOS⁺ Tregs) was 2.65% (C).

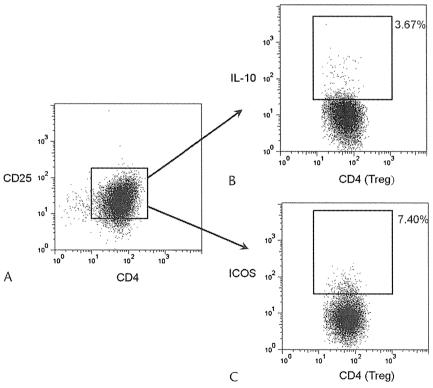


FIGURE 9. Flow cytometric analysis of Tregs (A) that express IL-10 (B) and ICOS (C) in an AIP patient. %IL-10+ Tregs of total Tregs was 3.67% (B). %ICOS+ Tregs was 7.40% (C).

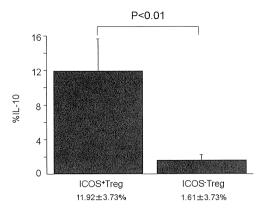


FIGURE 10. The ratio of IL-10⁺ Tregs in ICOS⁺ Tregs and ICOS⁻ Tregs with AIP patients (% IL-10⁺) (n = 16). Interleukin 10–positve cells were significantly increased in ICOS⁺ Tregs compared with ICOS⁻ Tregs (P < 0.05).

Ex vivo, the levels of ICOS expression correlate with a cytokine production pattern; peripheral T cells are either IL-10–producing (high ICOS); IL-4–, IL-5– and IL-13–producing (medium ICOS); or IL-2–, IL-3–, IL-6–, and interferon γ –producing cells (low ICOS). Moreover, ICOS expression is highly expressed on Tregs, and blocking ICOS function on these cells abrogates their regulatory capacity in diabetic lesions and also in an allergic asthma model, possibly through lack of inhibitory IL-10 production. Some studies also suggest that intermediate ICOS expression is associated with high production of $T_{\rm H}2$ cytokines, whereas high levels of ICOS predominantly translate into high IL-10 production. This is in concordance with the recent finding that ICOS expression is high on IL-10–producing

Tregs, which require ICOS presence for optimal regulatory function. 28,56 Recently, Ito et al 30 reported that expression of ICOS by human Foxp3 $^+$ Tregs was shown to distinguish 2 subsets: ICOS $^+$ Foxp3 $^+$ Tregs inhibited dendritic cell function via IL-10, and T cells via TGF- β , whereas ICOS $^-$ Foxp3 $^+$ Tregs used TGF- β only.

In the present study, we identified that circulating ICOS⁺ CD4⁺CD25^{high} Tregs expressing IL-10 were significantly increased in the peripheral blood of the patients with IgG4-related AIP. We also confirmed abundant infiltration of CD4⁺CD25^{high} Tregs in the pancreas tissues of IgG4-related AIP in addition to the liver of IgG4-related sclerosing cholangitis. In general, it has been reported that CD4⁺CD25^{high}Foxp3⁺ Tregs⁴¹⁻⁴³ producing IL-10 are regulated by the ICOS molecule. However, the involvement of the ICOS molecule on Tregs in AIP is still unknown. Therefore, to clarify the role of IgG4 in IgG4-related AIP, in the present study, we focused on the relations among serum levels of IgG4, ICOS molecules, IL-10, and Tregs in the peripheral blood and the pancreas. In addition to increasing numbers of ICOS⁺ Tregs expressing IL-10, the ratio of ICOS⁺ Tregs to ICOS⁻ Tregs in AIP was similar to control subjects, which suggested unpaired function (data not shown).

Taken together, our findings suggested that increasing IL-10 secreted from ICOS⁺ Tregs switched B cells to produce IgG4 in the periphery and abundant infiltration of IgG4-positive cells in the pancreas, which consequently may function to inhibit immune responses against inflammation (Fig. 11). According to this theory, AIP patients may be exposed to high doses of unknown disease-related antigens such as lactoferrin, carbonic anhydrase II, ^{8,38} or pancreatic secretory trypsin inhibitor, ⁵⁷ resulting in activation of both T_H1 type immune cells and Tregs suppressing T_H2-type immune cells. However, the actual mechanism of infiltration of ICOS⁺ Tregs from peripheral blood into

Hypothesis of Pathogenesis In AIP(LPSP)

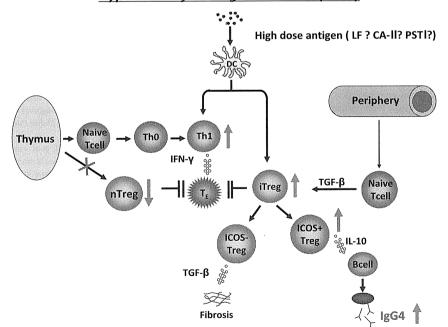


FIGURE 11. Hypothesis of AIP. High-dose antigens (carbonic anhydrase II, lactoferrin, or PST1) induce Tregs from periphery. Increased ICOS⁺ Tregs correlate with the production of IL-10, which may influence the switching of B cells to IgG4-producing plasmacytes and the production of serum IgG4. On the other hand, ICOS⁻ Tregs, which produce TGF-β, may influence fibrosis in AIP. Moreover, decreased naive Tregs may be involved in the pathogenesis of AIP.

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the pancreatic tissues and fibrosis in AIP remains unclear. Future studies should be addressed to clarify this point. In conclusion, increased quantities of ICOS⁺CD4⁺CD25^{high} Tregs may influence IgG4 production in AIP.

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REFERENCES

- Sarles H, Sarles JC, Muratore R, et al. Chronic inflammatory sclerosis
 of the pancreas—an autonomous pancreatic disease? Am J Dig Dis.
 1961:6:688-698.
- Kawaguchi K, Koike M, Tsuruta K, et al. Lymphoplasmacytic sclerosing pancreatitis with cholangitis: a variant of primary sclerosing cholangitis extensively involving pancreas. *Hum Pathol*. 1991:22:387–395.
- Yoshida K, Toki F, Takeuchi T, et al. Chronic pancreatitis caused by an autoimmune abnormality. Proposal of the concept of autoimmune pancreatitis. *Dig Dis Sci.* 1995;40:1561–1568.
- Hamano H, Kawa S, Horiuchi A, et al. High serum IgG4 concentrations in patients with sclerosing pancreatitis. N Engl J Med. 2001;344:732–738.
- Kamisawa T, Funata N, Hayashi Y, et al. A new clinicopathological entity of IgG4-related autoimmune disease. *J Gastroenterol*. 2003;38:982–984.
- Ito T, Nakano I, Koyanagi S, et al. Autoimmune pancreatitis as a new clinical entity. Three cases of autoimmune pancreatitis with effective steroid therapy. *Dig Dis Sci.* 1997;42:1458–1468.
- Horiuchi A, Kawa S, Akamatsu T, et al. Characteristic pancreatic duct appearance in autoimmune chronic pancreatitis: a case report and review of the Japanese literature. Am J Gastroenterol. 1998;93:260–263.
- Uchida K, Okazaki K, Konishi Y, et al. Clinical analysis of autoimmune-related pancreatitis. Am J Gastroenterol. 2000:95:2788–2794.
- Okazaki K, Kawa S, Kamisawa T, et al. Japanese consensus guidelines for management of autoimmune pancreatitis: I. Concept and diagnosis of autoimmune pancreatitis. J Gastroenterol. 2010;45:249–265.
- Kawa S, Okazaki K, Kamisawa T, et al. Japanese consensus guidelines for management of autoimmune pancreatitis: II. Extrapancreatic lesions, differential diagnosis. *J Gastroenterol*. 2010;45:355–369.
- Kamisawa T, Okazaki K, Kawa S, et al. Japanese consensus guidelines for management of autoimmune pancreatitis: III. Treatment and prognosis of AIP. J Gastroenterol. 2010;45:471–477.
- Zamboni G, Luttges J, Capelli P, et al. Histopathological features
 of diagnostic and clinical relevance in autoimmune pancreatitis:
 a study on 53 resection specimens and 9 biopsy specimens.

 Virchows Arch. 2004;445:552–563.
- Notohara K, Burgart LJ, Yadav D, et al. Idiopathic chronic pancreatitis with periductal lymphoplasmacytic infiltration: clinicopathologic features of 35 cases. Am J Surg Pathol. 2003;27:1119–1127.
- Chari ST, Kloeppel G, Zhang L, et al. Histopathologic and clinical subtypes of autoimmune pancreatitis: the Honolulu consensus document. *Pancreas*. 2010;39:549–554.
- Kusuda T, Uchida K, Satoi S, et al. Idiopathic duct-centric pancreatitis (IDCP) with Immunological Studies. *Intern Med.* 2010;49:2569–2575.
- Baecher-Allan C, Brown JA, Freeman GJ, et al. CD4⁺CD25^{high} regulatory cells in human peripheral blood. *J Immunol*. 2001;167:1245–1253.
- Dieckmann D, Plottner H, Berchtold S, et al. Ex vivo isolation and characterization of CD4(+)CD25(+) T cells with regulatory properties from human blood. *J Exp Med*. 2001;193:1303–1310.

- Jonuleit H, Schmitt E, Stassen M, et al. Identification and functional characterization of human CD4(+)CD25(+) T cells with regulatory properties isolated from peripheral blood. *J Exp Med*. 2001;193:1285–1294.
- Levings MK, Sangregorio R, Roncarolo MG. Human CD25(+) CD4(+) t regulatory cells suppress naive and memory T cell proliferation and can be expanded in vitro without loss of function. *J Exp Med.* 2001;193:1295–1302.
- Stephens LA, Mottet C, Mason D, et al. Human CD4(+)CD25(+) thymocytes and peripheral T cells have immune suppressive activity in vitro. Eur J Immunol. 2001;31:1247–1254.
- Wing K, Ekmark A, Karlsson H, et al. Characterization of human CD25⁺ CD4⁺ T cells in thymus, cord and adult blood. *Immunology*. 2002;106:190–199.
- Makita S, Kanai T, Oshima S, et al. CD4+CD25bright T cells in human intestinal lamina propria as regulatory cells. *J Immunol*. 2004;173: 3119–3130.
- Darrasse-Jeze G, Marodon G, Salomon BL, et al. Ontogeny of CD4⁺ CD25⁺ regulatory/suppressor T cells in human fetuses. *Blood*. 2005:105:4715–4721.
- Seddiki N, Santner-Nanan B, Tangye SG, et al. Persistence of naive CD45RA⁺ regulatory T cells in adult life. *Blood*. 2006;107:2830–2838.
- Miyoshi H, Uchida K, Taniguchi T, et al. Circulating naive and CD4⁺CD25^{high} regulatory T cells in patients with autoimmune pancreatitis. *Pancreas*. 2008;36:133–140.
- Punnonen J, Aversa G, Cocks BG, et al. Interleukin 13 induces interleukin 4-independent IgG4 and IgE synthesis and CD23 expression by human B cells. *Proc Natl Acad Sci U S A*. 1993:90:3730-3734.
- Jeannin P, Lecoanet S, Delneste Y, et al. IgE versus IgG4 production can be differentially regulated by IL-10. *J Immunol*. 1998;160:3555–3561.
- Akbari O, Freeman GJ, Meyer EH, et al. Antigen-specific regulatory T cells develop via the ICOS-ICOS-ligand pathway and inhibit allergen-induced airway hyperreactivity. Nat Med. 2002;8:1024–1032.
- Kohyama M, Sugahara D, Sugiyama S, et al. Inducible costimulator–dependent IL-10 production by regulatory T cells specific for self-antigen. *Proc Natl Acad Sci U S A*. 2004;101:4192–4197.
- Ito T, Hanabuchi S, Wang YH, et al. Two functional subsets of FOXP3⁺ regulatory T cells in human thymus and periphery. *Immunity*. 2008;28:870–880.
- Okazaki K, Kawa S, Kamisawa T, et al. Clinical diagnostic criteria of autoimmune pancreatitis: revised proposal. *J Gastroenterol*. 2006;41:626–631.
- Otsuki M, Chung JB, Okazaki K, et al. Asian diagnostic criteria for autoimmune pancreatitis: consensus of the Japan-Korea Symposium on Autoimmune Pancreatitis. *J Gastroenterol*. 2008;43:403–408.
- Japan Pancreas Society. Clinical diagnosis criteria of chronic pancreatitis [in Japanese]. Suizou (J Jpn Pancreas Soc). 2001;16:560–561.
- 34. Koyabu M, Uchida K, Miyoshi H, et al. Analysis of regulatory T cells and IgG4-positive plasma cells among patients of IgG4-related sclerosing cholangitis and autoimmune liver diseases. *J Gastroenterol*. 2010:45:732–741.
- Okazaki K, Uchida K, Ohana M, et al. Autoimmune-related pancreatitis is associated with autoantibodies and a T_H1/T_H2-type cellular immune response. *Gastroenterology*. 2000;118:573–581.
- McGeachy MJ, Cua DJ. The link between IL-23 and T_H17 cell-mediated immune pathologies. Semin Immunol. 2007;19:372–376.
- Zen Y, Fujii T, Harada K, et al. T_H2 and regulatory immune reactions are increased in immunoglobin G4–related sclerosing pancreatitis and cholangitis. *Hepatology*. 2007;45:1538–1546.
- Uchida K, Okazaki K, Nishi T, et al. Experimental immune-mediated pancreatitis in neonatally thymectomized mice immunized with carbonic anhydrase II and lactoferrin. *Lab Invest*. 2002;82:411–424.
- Robinson DS, Larche M, Durham SR. Tregs and allergic disease. J Clin Invest. 2004;114:1389–1397.
- Ling EM, Smith T, Nguyen XD, et al. Relation of CD4⁺CD25⁺ regulatory T-cell suppression of allergen-driven T-cell activation

- to atopic status and expression of allergic disease. *Lancet*. 2004;363:608-615.
- Gerstmayr M, Ilk N, Schabussova I, et al. A novel approach to specific allergy treatment: the recombinant allergen-S-layer fusion protein rSbsC-Bet v 1 matures dendritic cells that prime T_H0/T_H1 and IL-10-producing regulatory T cells. *J Immunol*. 2007;179: 7270-7275.
- Strauss L, Bergmann C, Szczepanski M, et al. A unique subset of CD4⁺ CD25^{high} Foxp3⁺ T cells secreting interleukin-10 and transforming growth factor-beta1 mediates suppression in the tumor microenvironment. *Clin Cancer Res.* 2007;13:4345–4354.
- Uhlig HH, Coombes J, Mottet C, et al. Characterization of Foxp3⁺CD4⁺CD25⁺ and IL-10-secreting CD4⁺CD25⁺ T cells during cure of colitis. *J Immunol.* 2006;177:5852–5860.
- Alpan O, Bachelder E, Isil E, et al. 'Educated' dendritic cells act as messengers from memory to naive T helper cells. *Nat Immunol*. 2004;5:615–622.
- Akitake R, Watanabe T, Zaima C, et al. Possible involvement of T helper type 2 responses to Toll-like receptor ligands in IgG4-related sclerosing disease. Gut. 2010;59:542–545.
- Okazaki K, Uchida K, Fukui T. Recent advances in autoimmune pancreatitis: concept, diagnosis, and pathogenesis. *J Gastroenterol*. 2008:43:409–418.
- Hutloff A, Dittrich AM, Beier KC, et al. ICOS is an inducible T-cell co-stimulator structurally and functionally related to CD28. *Nature*. 1999;397:263–266.
- 48. Yoshinaga SK, Whoriskey JS, Khare SD, et al. T-cell co-stimulation through B7RP-1 and ICOS. *Nature*. 1999;402:827–832.

- Wang S, Zhu G, Chapoval AI, et al. Costimulation of T cells by B7-H2, a B7-like molecule that binds ICOS. *Blood*. 2000;96:2808–2813.
- Brodie D, Collins AV, Iaboni A, et al. LICOS, a primordial costimulatory ligand? *Curr Biol*. 2000;10:333–336.
- Swallow MM, Wallin JJ, Sha WC. B7h, a novel costimulatory homolog of B7.1 and B7.2, is induced by TNFalpha. *Immunity*. 1999;11:423–432.
- McAdam AJ, Chang TT, Lumelsky AE, et al. Mouse inducible costimulatory molecule (ICOS) expression is enhanced by CD28 costimulation and regulates differentiation of CD4⁺ T cells. *J Immunol.* 2000:165:5035–5040.
- Coyle AJ, Lehar S, Lloyd C, et al. The CD28-related molecule ICOS is required for effective T cell-dependent immune responses. *Immunity*. 2000;13:95–105.
- Vieira PL, Wassink L, Smith LM, et al. ICOS-mediated signaling regulates cytokine production by human T cells and provides a unique signal to selectively control the clonal expansion of T_H2 helper cells. *Eur J Immunol.* 2004;34:1282–1290.
- Lohning M, Hutloff A, Kallinich T, et al. Expression of ICOS in vivo defines CD4⁺ effector T cells with high inflammatory potential and a strong bias for secretion of interleukin 10. *J Exp Med*. 2003;197:181–193.
- Herman AE, Freeman GJ, Mathis D, et al. CD4⁺CD25⁺ T regulatory cells dependent on ICOS promote regulation of effector cells in the prediabetic lesion. *J Exp Med*. 2004;199:1479–1489.
- Asada M, Nishio A, Uchida K, et al. Identification of a novel autoantibody against pancreatic secretory trypsin inhibitor in patients with autoimmune pancreatitis. *Pancreas*. 2006;33:20–26.

ORIGINAL ARTICLE—LIVER, PANCREAS, AND BILIARY TRACT

Comparison of steroid pulse therapy and conventional oral steroid therapy as initial treatment for autoimmune pancreatitis

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Abstract

Background The efficacy of oral steroid therapy for autoimmune pancreatitis (AIP) is well known, and oral prednisolone treatment is most usually commenced at 30–40 mg/day, but there have been few reports about comparative studies of oral steroid therapy and steroid pulse therapy as the initial treatment for AIP. We studied the clinical course and image findings to estimate the utility of steroid pulse therapy for AIP, comparing it with oral steroid therapy.

Methods Laboratory and image findings were assessed retrospectively in 11 patients who received steroid pulse therapy, and the findings were compared to those in 10 patients who received conventional oral steroid therapy. Results Change in pancreatic size showed no significant difference between the therapies after 2 weeks of treatment. Significant improvement of lower bile duct strictures after 2 weeks of treatment and that of immunoglobulin values within 6 months were shown with both therapies. However, steroid pulse therapy showed significant improvement of y-guanosine triphosphate (GTP) in 2 weeks and of alanine aminotransferase (ALT) in 2 and 8 weeks, compared with oral steroid therapy. Moreover, there was one patient in whom the lower bile duct stricture was not improved by oral steroid therapy, but it did show improvement with steroid pulse therapy.

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Conclusions Initial steroid pulse therapy is a beneficial alternative to oral steroid therapy for the improvement of bile duct lesions. In future, the accumulation of a larger number of patients receiving steroid pulse therapy is needed, and prospective studies will be required.

Keywords Autoimmune pancreatitis (AIP) · Steroid pulse therapy · Bile duct stricture · Diabetes mellitus · Pancreatic cancer

Introduction

Sarles et al. [1] reported a case of chronic pancreatitis with hypergammaglobulinemia, but the clinical entity was not confirmed thereafter. Autoimmune pancreatitis (AIP), which was first proposed as a clinical entity by Yoshida et al. [2] from Japan, is now generally accepted as a distinctive type of pancreatitis [1]. AIP is characterized by diffuse irregular narrowing of the main pancreatic duct, sausage-like diffuse swelling of the pancreas, high serum levels of IgG or IgG4, and steroid responsiveness [3–6]. Since the fibroinflammatory process of AIP responds well to steroids, autoimmune mechanisms are thought to be involved in the development of AIP. A recent large Japanese study of AIP and guidelines for treatment recommend standard oral steroid therapy with an initial dose of 0.5–0.6 mg/kg/day [7, 8].

While steroid responsiveness as a diagnostic component is not included in the revised Japanese criteria, it is included in the Korean criteria, Mayo Clinic HISORt (Histology, Imaging, Serology, Other organ involvement, and Response to steroids) criteria, and recently proposed Asian criteria [8–11]. The most important issue in AIP management is making the diagnosis of AIP, especially the

mass-forming type, differentiating it from pancreatic or biliary cancers [12–15], although some cases of pancreatic cancer are accompanied by AIP [16–19]. In some tumorforming AIP cases, the efficacy of a steroid trial has been reported as useful in diagnosing AIP by criteria other than the Japanese diagnostic criteria [20]. Moreover, Korean investigators have reported the usefulness of a 2 weeks' trial of oral steroids in differentiating AIP from malignancy, with continuing administration if AIP is diagnosed [20]. However, it has not yet been established whether or not withdrawal of steroids in reconsidering malignancy presents a risk of postoperative adrenal insufficiency [21–23]. Therefore, it is desirable to have an alternative to the discontinuation of steroid administration immediately after surgery.

Steroid pulse therapy is widely used to initiate treatment in patients with rapidly progressive and immunologically mediated disorders such as acute graft rejection, Graves ophthalmopathy, pemphigus, and severe systemic lupus erythematosus [24–27]. Moreover, high doses of systemic steroid can be given with comparative safety within a period of 1 week [28]. We therefore evaluated the efficacy of short-term steroid pulse therapy, in comparison with oral steroid therapy, in patients with AIP.

Methods

Patients and treatment

For this study, we retrospectively examined the records of all 21 AIP patients treated in our hospitals from November 2004 to May 2009. All patients were diagnosed with AIP according to the clinical diagnostic criteria for AIP proposed by the Research Committee of Intractable Diseases of the Pancreas supported by the Japanese Ministry of Health, Labor, and Welfare, and the Japan Pancreas Society. Following diagnosis, 20 patients with AIP were randomly distributed to two treatment groups by their attending physicians. One patient (case 10) was referred to our hospital after the withdrawal of oral steroid for AIP because his bile duct lesion had not responded to the treatment. Eleven patients (cases 1-11; 5 male and 6 female; aged 47-80 years, with a mean age of 66 years, named the "pulse group") (Table 1) received steroid pulse therapy, and ten patients (cases 12-21; 8 male and 2 female, aged 49-72 years, with a mean age of 69 years, named the "oral group") (Table 2) received oral steroid therapy. For the pulse group, the initial dose of methylprednisolone was 500 mg/day for 3 days each week as 1 course, and we treated them with 2 weekly courses. Then oral prednisolone at 20 mg/day was prescribed as maintenance therapy and the dose was tapered off. For the oral group, ten patients commenced oral prednisolone at 30–40 mg/day. Two weeks after the start of the treatment, oral prednisolone at 20 mg/day was prescribed, and the dose was tapered off. This study was approved by the Kansai Medical University ethics committee.

Serological study

We analyzed immunological findings for the following: IgG, IgG4, antinuclear antibodies (ANA), rheumatoid factor (RF), antimitochondrial antibodies (AMA), myeloperoxidase-antineutrophil cytoplasmic antibodies (MPO-ANCA), anti-Sjögren's syndrome A antibodies (SS-A), anti-Sjögren's syndrome B antibodies (SS-B), anti-thyroid peroxidase antibodies (TPOAb), and anti-thyroglobulin antibodies (TgAb). To compare liver and endocrine function in both groups, we evaluated the serum levels of γ-guanosine triphosphate (GTP) and alanine aminotransferase (ALT) on day 0 (data just before the treatment), and at weeks 2 and 8 after therapy, and checked glycosylated hemoglobin values (HbA1c) at months 1, 3, and 7 after therapy, which closely reflected glucose tolerance at months 0, 2, and 6, respectively. In each evaluation, patients who did not show abnormal values during the clinical course were excluded in order to evaluate the therapeutic effect strictly.

Radiological study

All the patients were examined by contrast-enhanced helical computed tomography (CT), magnetic resonance imaging (MRI), and endoscopic retrograde cholangiopancreatography (ERCP) and underwent liver function tests, combined with bile duct drainage and pathological tests as necessary.

For morphological changes, CT, MRI, and ERCP were studied. The width of the pancreas along its longest axis was measured on CT or MRI images and compared with the transverse diameter of the vertebral body according to the method of Heuck et al. [29]. The pancreas size on the first image was defined as 100%.

Cases showing lower bile duct stricture were classified as follows: 0 = absent, 1 = <0-25%, 2 = <25-50%, 3 = <50-75%, and 4 = <75-100%, according to the method of Craig et al. [30]. Using the method described above, pancreas size was evaluated after 2 weeks on steroids, and stricture of the distal third of the common bile duct was measured after 2 weeks and after 8 weeks.

Statistical analysis

Statistical analysis was performed using the Mann–Whitney U-test, Wilcoxon signed-ranks test, paired t-test, and



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Table 1 Background of AIP patients who received steroid pulse therapy

Patient ID	Age (years)/ sex	Symptoms	IgG <1,700 (mg/dl)	IgG 4 <135 (mg/dl)	Amy <130 (IU/L)	T-Bil <0.9 (mg/dl)	ALT <30 (IU/L)	γ-GTP <35 (IU/L)	ANA	PFD <73.4 (%)	DM	Extrapancreatic lesion	Stenosis on ERCP	Morphological change of the pancreas
1	80/F	Jaundice	2,604	1,230	52	0.9	43	260	_	52.1		Sialoadenitis	Head, CBD	FS in head
2	63/M	Epigastralgia	1,714	354	66	0.9	56	222	_	29.7	_	Warthin tumor	Head, CBD	DS
3	54/F	Epigastralgia	1,828	324	561	1.0	100	403	_	NT		Hypothyroidism	Body, CBD	DS
4	71/F	None	1,916	295	78	0.6	60	101	_	97.9		Sialoadenitis, mediastinum LNS	Head	FS in head
5	66/F	Nausea	1,535	235	77	2.2	118	1,311	_	NT	_	Hypothyroidism, retroperitoneal fibrosis	Head to tail, CBD	FS in head
6	66/M	Epigastralgia, jaundice	2,695	1,790	164	12.5	98	137	-	58.1	+	None	Body, CBD	DS
7	47/F	Jaundice	2,453	629	15	1.0	190	65	_	NT	_	None	Head, body to tail, CBD	DS
8	72/M	Epigastralgia	1,692	452	66	1.3	721	1,352	+	30.9	+	None	Head to tail CBD	FS in head
9	72/M	Epigastralgia, jaundice	1,513	411	32	14.1	114	352	-	NT	+	None	Head, CBD	DS
10	63/M	Malaise	1,514	394	76	0.5	20	82		NT		None	Head to tail, CBD	FS in head
11	73/F	Epigastralgia	1,598	373	55	0.6	33	588		59.7	+	None	Head to tail, CBD	FS in head

AIP autoimmune pancreatitis, T Bil total bilirubin, ALT alanine aminotransferase, ID identification, γ-GTP γ-guanosine triphosphate, Amy amylase, ANA antinuclear antibody, PFD pancreatic functional diagnostic test, DM diabetes mellitus, ERCP endoscopic retrograde cholangiopancreatography, CBD common bile duct, FS focal swelling, DS diffuse swelling, NT not tested, LNS lymph node swelling

PFD IgG ANA DM Extrapancreatic Stenosis Morphological Patient ID Age Symptoms IgG4 Amy T-Bil ALT γ-GTP on ERCP <73.4 change of the <1,700 <135 <130 < 0.9 < 30 <35 lesion (years)/sex (mg/dl) (mg/dl) (IU/L) (mg/dl) (IU/L) (IU/L) (%) pancreas 76 32.7 Mediastinum Head to tail, CBD DS 58 1.0 24 12 71/M Jaundice 3,274 1,870 LNS Sialoadenitis, Tail, CBD DS 65.8 13 66/M Thirst 4,060 1,170 64 0.4 18 116 mediastinum LNS 58/F 2,754 95 0.9 13 10 NT Thyroiditis, Body to tail FS in head 14 Epigastralgia 1,110 mediastinum LNS FS in tail Interstitial Tail 15 52/F None 2,190 661 435 0.6 15 15 69.7 pneumonia, Mikulicz tumor Thyroiditis Head, body, CBD DS 73.4 16 68/M Fever 1,622 407 34 0.6 30 30 NT Head FS in head 17 72/M Vomiting 2,010 773 51 0.3 34 67 None

Amy amylase, ANA antinuclear antibody, PFD pancreatic functional diagnostic test, DM diabetes mellitus, ERCP endoscopic retrograde cholangiopancreatography, LNS lymph node swelling, CBD common bile duct, DS diffuse swelling, NT not tested, FS focal swelling

240

149

503

189

17.9

NT

35.7

77.9

+

+

+

None

None

None

None

178

52

52

41

18

19

20

21

55/M

63/M

49/M

66/M

Table 2 Background of AIP patients treated with oral prednisolone

Epigastralgia

Jaundice

Jaundice

Malaise

1,461

2,073

2,065

1,607

659

487

479

200

53

41

62

47

0.6

1.1

0.9

1.0

Head to tail, CBD

Body, tail, CBD

Head, CBD

Head, CBD

DS

DS

FS in head

FS in head

Student's *t*-test. In all tests, corrected P values of < 0.05 were considered statistically significant.

Results

Clinical manifestations

After 6 months' treatment, oral prednisolone had been administered to 19 patients in both groups [pulse group: 9 of 11 vs. oral group: 10 of 10; difference not significant (N.S.)], and the median dosage of prednisolone in each group was 10 mg/day (pulse group: 2.5–12.5 vs. oral group: 5–12.5; N.S.). Two patients in the pulse group dropped out of the maintenance therapy. Neither group showed severe or lethal complications.

In both groups, extrapancreatic lesions other than the bile duct lesions were observed in 10 patients (pulse group: 5 of 11 vs. oral group: 5 of 10; N.S.) (Tables 1, 2). No exacerbation of the extrapancreatic lesions was found following either of the treatments (data not shown). Laboratory findings including immunoglobulin, autoantibody, and exocrine function at the treatment start are listed in Tables 1 and 2.

Immunoglobulin

At the beginning of treatment, in both groups, abnormal serum immunoglobulin-G4 (IgG4) values were observed in all patients (Tables 1, 2), and abnormal serum immunoglobulin-G (IgG) values were observed in 13 patients (pulse group: 6 of 11 vs. oral group: 7 of 10; N.S.) (Tables 1, 2). Normalization of the IgG value was shown in all these patients within 6 months (data not shown).

Liver function

At the beginning of treatment, abnormal γ-GTP values were revealed in both groups, in a total of 18 patients (pulse group: 11 of 11 vs. oral group: 7 of 10; N.S.). In the pulse group, the median γ-GTP levels fell, from 222 IU/L (range 65-1,352) at the beginning of treatment to 92 IU/L (22-679) (P < 0.01) after 2 weeks of pulse therapy (Fig. 1a), and to 36 IU/L (19–556) (P < 0.01) after 8 weeks of pulse therapy (Fig. 1a). In the oral group, however, the median γ-GTP fell insignificantly after 2 weeks of prednisolone treatment, from 149 IU/L (range 67-380) at the beginning of treatment to 125 IU/L (63-274) (P = 0.083) (Fig. 1b), although the level fell significantly to 72 IU/L (29–114) (P = 0.027) after 8 weeks of prednisolone treatment (Fig. 1b). When limiting results to the patients who showed diffuse pancreatic swelling, γ-GTP was significantly improved in the pulse

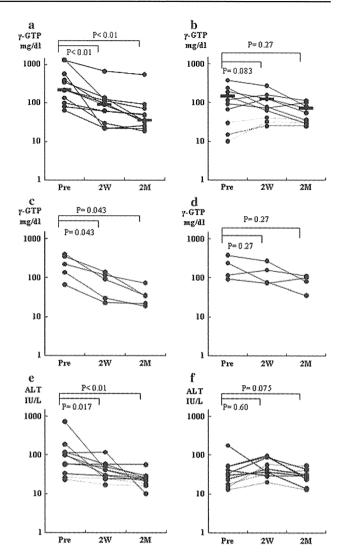


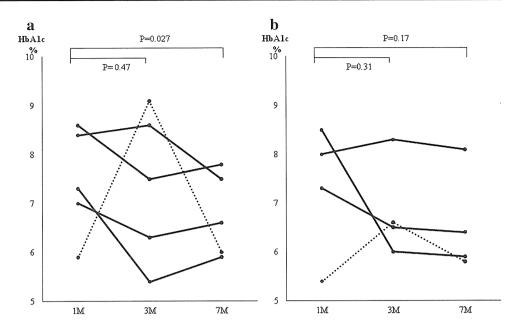
Fig. 1 γ -Guanosine triphosphate (γ -GTP) and alanine aminotransferase (ALT) changes after steroids. The serum levels of γ -GTP after steroid pulse therapy (a), after oral steroid therapy (b), those of ALT after steroid pulse therapy (e) and those after oral steroid therapy (f) were monitored on day 0 (Pre), and 2 weeks (W) and 8 weeks after therapy. To evaluate the therapeutic effect strictly, the patients (dotted lines) who did not show an abnormal value during the clinical course were excluded from this analysis. The serum levels of γ -GTP after 2 weeks on steroids and those of ALT after 2 and 8 weeks on steroids were significantly improved in the pulse group, compared with the oral group. When limiting the patients to those who showed diffuse pancreatic swelling, the serum level of γ -GTP was significantly improved in the pulse group after 2 and 8 weeks of pulse therapy (c), compared with the oral group (d). M Months

group after 2 weeks (P = 0.043) and 8 weeks (P = 0.043) of pulse therapy (Fig. 1c), whereas the improvement in the γ -GTP level was insignificant in the oral group after 2 weeks (P = 0.27) and 8 weeks (P = 0.27) of prednisolone treatment (Fig. 1d).

At the beginning of treatment, abnormal ALT values were revealed in both groups, in a total of 16 patients



Fig. 2 Glycosylated hemoglobin (HbA1C) changes after steroid therapy. The levels of HbA1c after steroid pulse therapy (a) and oral steroid therapy (b) were monitored at months 1, 3, and 7 after therapy, which closely reflect glucose tolerance at months 0, 2, and 6, respectively. To evaluate the therapeutic effect strictly, patients who did not show abnormal values during the clinical course were excluded from this analysis. Dotted lines represent the patients who developed glucose intolerance after 2 months of steroid therapy. The level of HbA1c at month 7 on steroids tended to be improved in the pulse group, compared with the oral group



(pulse group: 10 of 11 vs. oral group: 6 of 10; N.S.). In the pulse group, the median ALT level fell from 100 IU/L (range 33–721) at the beginning of treatment to 41 IU/L (24–117) (P=0.017) after 2 weeks of pulse therapy (Fig. 1e), and to 25 IU/L (10–57) (P<0.01) after 8 weeks of pulse therapy (Fig. 1e). In the oral group, however, the median ALT rose transiently from 46.5 IU/L (range 30–178) at the beginning of treatment to 62 IU/L (29–96) (P=0.60) after 2 weeks of prednisolone (Fig. 1f), but improved to 28 IU/L (14–44) (P=0.075) after 8 weeks of prednisolone treatment (Fig. 1f).

Endocrine function

Before steroid treatment, diabetes mellitus was seen in 7 patients (pulse group: 4 of 11 vs. oral group: 3 of 10; N.S.) (Tables 1, 2). One patient in each group developed glucose intolerance after 2 months of steroid therapy. Including these patients, all the patients with impaired glucose tolerance were treated with dietary measures and received medical therapy while the steroid therapy was maintained. Neither group showed significant improvement in glucose tolerance after 2 months, but at 6 months, the pulse group had improved significantly (P = 0.027), whereas the oral group had not (P = 0.17) (Fig. 2).

Pancreas size

Both groups evidenced pancreatic swelling: diffuse pancreatic swelling was observed in 10 patients (pulse group: 5 of 11 vs. oral group: 5 of 10; N.S.) and focal swelling was observed in 11 patients (pulse group: 6 of 11 vs. oral group: 5 of 10; N.S.) (Tables 1, 2). The change in pancreas size

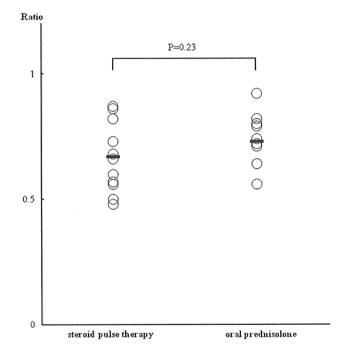


Fig. 3 Pancreas morphology changes after 2 weeks on steroids. Morphological changes of the pancreas after 2 weeks of steroid pulse therapy (*left*) and oral steroid therapy (*right*) were scored. The width of the pancreas along its longest axis was measured on computed tomography (CT) or magnetic resonance imaging (MRI) and compared with the transverse diameter of the vertebral body, referred to in the method of Heuck et al. [29]. The pancreas size on the first image was defined as 100%, and the ratio after 2 weeks with each treatment was measured in the same manner. The *black bars* represent the mean in each group. The two groups showed no significant difference in morphological change of the pancreas after 2 weeks

after 2 weeks showed no significant difference between the groups after treatment (pulse group: 67% vs. oral group: 73.4%, P = 0.23) (Fig. 3).



Fig. 4 Lower bile duct stricture changes after steroids. The changes in lower bile duct stricture in the pulse group (a) and the oral group (b) on day 0 and 2 weeks after therapy were scored as follows: 0 = absent, 1 = <0-25%, 2 = <25-50%, 3 = <50-75%, 4 = <75-100%, referred to in the method of Craig et al. [30]. Bile duct stenosis in the distal third part was significantly improved after 2 weeks in each group

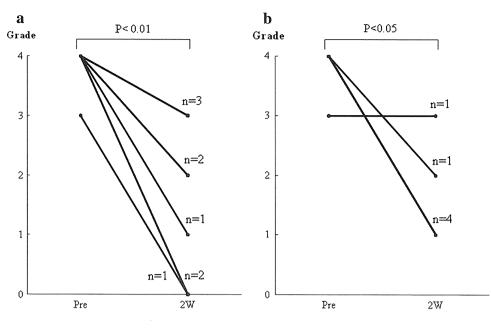
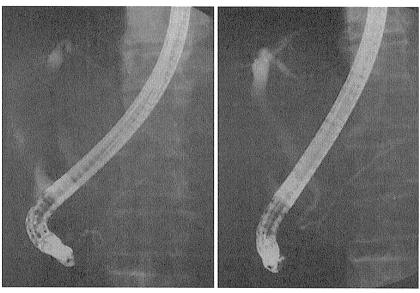


Fig. 5 Endoscopic retrograde cholangiopancreatography (ERCP) images of the impact of steroid pulse therapy on refractory autoimmune pancreatitis (AIP). Although oral prednisolone was commenced, it had had no effect on the biliary stenosis (*left*). Two courses of steroid pulse therapy ameliorated the stenosis dramatically (*right*) (Case 10; reference [31])



Bile duct lesion

After 2 weeks on steroids, significant improvement of lower bile duct stricture was shown in both groups (pulse group: P < 0.01, and oral group: P < 0.05) (Fig. 4). However, there was one patient (case 10) whose lower bile duct stricture did not improve following oral prednisone treatment, but showed definite improvement with steroid pulse therapy [31] (Fig. 5).

Discussion

Several cases have been reported of pancreatic cancer or bile duct cancer concurrent with AIP [16–19]; because the

image findings of AIP often mimic pancreatobiliary malignancies, it is extremely crucial to distinguish AIP from cancers, although this can be difficult [12–15].

In the Asian diagnostic criteria proposed by the Japan-Korea symposium on autoimmune pancreatitis [11], the use of steroids as diagnostic treatment was allowed only when the imaging findings were compatible with AIP and only after there was a negative result for malignancy work-up. Cases of diagnostic treatment using steroids will doubtless increase in future. The usefulness of a 2-week conventional oral steroid diagnostic treatment was also proposed by Moon et al. [20]. However, oral steroid therapy requires a long period for drug tapering, because any patient who has received a glucocorticoid in doses equivalent to at least 20 mg a day of prednisone for >5 days is at risk of



secondary adrenal insufficiency due to hypothalamic-pituitary-adrenal suppression [21–23]. Diagnostic treatment with an oral steroid may cause an undesirable effect when surgical resection is required. Although one article reported that azathioprine was used for refractory AIP [32], it is causative of acute pancreatitis [33]. Therefore, a safe and simple alternative to oral steroid treatment is needed for refractory AIP [34].

Steroid pulse therapy is a well known alternative to oral steroid treatment for autoimmune disorders; it requires no drug tapering [28], and we have already reported cases where steroid pulse therapy was effective for AIP [35], although comparative studies of conventional oral steroid therapy and steroid pulse therapy have not been reported. Here we report that steroid pulse therapy is an effective alternative to oral steroid for the initial treatment of AIP.

The efficacy of oral steroid therapy for AIP is well known, and the improvement of AIP in patients treated with steroids for 2 weeks can be shown in radiographic findings [5, 20, 36]. Our data on pancreas size after 2 weeks did not show a significant difference between oral steroid and pulse therapy. Both therapies were effective for alleviating lower bile duct stricture in the short term and for resolving abnormal IgG values. However, the shortterm change in y-GTP showed significant improvement in the steroid pulse therapy group, but not in the oral steroid group. In one patient, lower bile duct stricture was improved by steroid pulse therapy, although it had not been improved by oral steroids. These findings suggest that steroid pulse therapy may prevent patients from having unnecessary major operations for benign bile duct lesions which do not respond to oral steroid treatment.

Steroid therapy is reported to be effective in approximately half of AIP patients with accompanying diabetes mellitus [37, 38]. The mechanism of steroid action in the recovery of endocrine function in patients with AIP is unclear, but Tanaka et al. [39] have suggested that steroids can suppress the release of cytokines produced by inflammatory cells, and enable islet regeneration and eventual restoration of insulin secretion. However, we have reported that conventional oral steroids did not improve diabetes mellitus in the long term [40]. Although the accumulation of a larger numbers of cases is needed, our study of glycosylated hemoglobin values showed that steroid pulse therapy tended to be more effective than oral steroids for improving glucose tolerance after 6 months of treatment. In our protocol, there was a decisive difference between the two therapies just for the initial 2 weeks, because 20 mg/day of oral prednisolone was prescribed as maintenance therapy after the 2 weeks of steroid pulse therapy. We presume that the steroid pulse therapy was better than the oral steroid therapy for improving glucose tolerance because of its stronger cytokine suppression.

In summary, steroid pulse therapy is an effective alternative initial treatment for AIP, and surpasses conventional oral steroid therapy in the improvement of bile duct lesions. When a differential diagnosis between AIP and pancreatic cancer is difficult clinically or when bile duct lesions do not respond to oral steroid treatment, we recommend steroid pulse therapy as an alternative to oral steroid therapy. In future, the accumulation of a larger number of patients receiving steroid pulse therapy is needed, and prospective studies will be required.

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References

- Sarles H, Sarles JC, Muratore R, Guien C. Chronic inflammatory sclerosis of the pancreas: an autonomous pancreatic disease? Am J Dig Dis. 1961;6:688–98.
- 2. Yoshida K, Toki F, Takeuchi T, Watanabe S, Shiratori K, Hayashi N. Chronic pancreatitis caused by an autoimmune abnormality. Proposal of the concept of autoimmune pancreatitis. Dig Dis Sci. 1995;40(7):1561–8.
- Okazaki K, Chiba T. Autoimmune related pancreatitis. Gut. 2002;51(1):1–4.
- Hamano H, Kawa S, Horiuchi A, Unno H, Furuya N, Akamatsu T, et al. High serum IgG4 concentrations in patients with sclerosing pancreatitis. N Engl J Med. 2001;344(10):732–8.
- Kamisawa T, Egawa N, Nakajima H, Tsuruta K, Okamoto A. Morphological changes after steroid therapy in autoimmune pancreatitis. Scand J Gastroenterol. 2004;39(11):1154–8.
- Uchida K, Okazaki K, Konishi Y, Ohana M, Takakuwa H, Hajiro K, et al. Clinical analysis of autoimmune-related pancreatitis. Am J Gastroenterol. 2000;95(10):2788–94.
- 7. Kamisawa T, Shimosegawa T, Okazaki K, Nishino T, Watanabe H, Kanno A, et al. Standard steroid treatment for autoimmune pancreatitis. Gut. 2009;58(11):1504–7 (epub 26 April 2009).
- 8. Okazaki K, Kawa S, Kamisawa T, Ito T, Inui K, Irie H, et al. Japanese clinical guidelines for autoimmune pancreatitis. Pancreas. 2009;38(8):849–66.
- Kim KP, Kim MH, Kim JC, Lee SS, Seo DW, Lee SK. Diagnostic criteria for autoimmune chronic pancreatitis revisited. World J Gastroenterol. 2006;12(16):2487–96.
- Chari ST, Smyrk TC, Levy MJ, Topazian MD, Takahashi N, Zhang L, et al. Diagnosis of autoimmune pancreatitis: the Mayo Clinic experience. Clin Gastroenterol Hepatol. 2006;4(8):1010–6.
- 11. Otsuki M, Chung JB, Okazaki K, Kim MH, Kamisawa T, Kawa S, et al. Asian diagnostic criteria for autoimmune pancreatitis: consensus of the Japan-Korea symposium on autoimmune pancreatitis. J Gastroenterol. 2008;43(6):403–8.
- Kamisawa T, Egawa N, Nakajima H, Tsuruta K, Okamoto A, Kamata N. Clinical difficulties in the differentiation of autoimmune pancreatitis and pancreatic carcinoma. Am J Gastroenterol. 2003;98(12):2694–9.
- 13. Nakazawa T, Ohara H, Sano H, Ando T, Imai H, Takada H, et al. Difficulty in diagnosing autoimmune pancreatitis by imaging findings. Gastrointest Endosc. 2007;65(1):99–108.



- Okazaki K, Uchida K, Fukui T. Recent advances in autoimmune pancreatitis: concept, diagnosis, and pathogenesis. J Gastroenterol. 2008;43(6):409–18.
- Frulloni L, Scattolini C, Falconi M, Zamboni G, Capelli P, Manfredi R, et al. Autoimmune pancreatitis: differences between the focal and diffuse forms in 87 patients. Am J Gastroenterol. 2009;104(9):2288–94.
- Fukui T, Mitsuyama T, Takaoka M, Uchida K, Matsushita M, Okazaki K. Pancreatic cancer associated with autoimmune pancreatitis in remission. Intern Med. 2008;47(3):151-5.
- Witkiewicz AK, Kennedy EP, Kennyon L, Yeo CJ, Hruban RH. Synchronous autoimmune pancreatitis and infiltrating pancreatic ductal adenocarcinoma: case report and review of the literature. Hum Pathol. 2008;39(10):1548–51.
- Inoue H, Miyatani H, Sawada Y, Yoshida Y. A case of pancreas cancer with autoimmune pancreatitis. Pancreas. 2006;33(2): 208-9
- Oh HC, Kim JG, Kim JW, Lee KS, Kim MK, Chi KC, et al. Early bile duct cancer in a background of sclerosing cholangitis and autoimmune pancreatitis. Intern Med. 2008;47(23):2025–8.
- 20. Moon SH, Kim MH, Park DH, Hwang CY, Park SJ, Lee SS, et al. Is a 2-week steroid trial after initial negative investigation for malignancy useful in differentiating autoimmune pancreatitis from pancreatic cancer? A prospective outcome study. Gut. 2008;57(12):1704–12.
- Fraser CG, Preuss FS, Bigford WD. Adrenal atrophy and irreversible shock associated with cortisone therapy. JAMA. 1952;149(17):1542–3.
- Axelrod L. Perioperative management of patients treated with glucocorticoids. Endocrinol Metab Clin North Am. 2003;32(2): 367–83.
- Alves C, Robazzi TC, Mendonça M. Withdrawal from glucocorticosteroid therapy: clinical practice recommendations. J Pediatr (Rio J). 2008;84(3):192–202.
- 24. Kee TY, Chapman JR, O'Connell PJ, Fung CL, Allen RD, Kable K, et al. Treatment of subclinical rejection diagnosed by protocol biopsy of kidney transplants. Transplantation. 2006;82(1):36–42.
- 25. Wiersinga WM. Management of Graves' ophthalmopathy. Nat Clin Pract Endocrinol Metab. 2007;3(5):396–404.
- Saha M, Powell AM, Bhogal B, Black MM, Groves RW. Pulsed intravenous cyclophosphamide and methylprednisolone therapy in refractory pemphigus. Br J Dermatol. 2010;162(4):790–7.
- Parker BJ, Bruce IN. High dose methylprednisolone therapy for the treatment of severe systemic lupus erythematosus. Lupus. 2007;16(6):387–93.
- Boumpas DT, Chrousos GP, Wilder RL, Cupps TR, Balow JE. Glucocorticoid therapy for immune-mediated diseases: basic and clinical correlates. Ann Intern Med. 1993;119(12):1198–208.

- 29. Heuck A, Maubach PA, Reiser M, Feuerbach S, Allgayer B, Lukas P, et al. Age-related morphology of the normal pancreas on computed tomography. Gastrointest Radiol. 1987;12(1):18–22.
- Craig DA, MacCarty RL, Wiesner RH, Grambsch PM, LaRusso NF. Primary sclerosing cholangitis: value of cholangiography in determining the prognosis. AJR Am J Roentgenol. 1991;157(5): 959–64.
- 31. Nakayama S, Matsushita M, Yamashina M, Shimatani M, Uchida K, Okazaki K. Effective steroid mini-pulse therapy for refractory biliary stenosis due to autoimmune pancreatitis (in Japanese with English abstract). Suizou. 2006;21(5):446–52.
- 32. Church NI, Pereira SP, Deheragoda MG, Sandanayake N, Amin Z, Lees WR, et al. Autoimmune pancreatitis: clinical and radiological features and objective response to steroid therapy in a UK series. Am J Gastroenterol. 2007;102(11):2417–25.
- Floyd A, Pedersen L, Nielsen GL, Thorlacius-Ussing O, Sorensen HT. Risk of acute pancreatitis in uses of azathioprine: a population-based case-control study. Am J Gastroenterol. 2003;98(6): 1305–8.
- 34. Matsushita M, Ikeura T, Fukui T, Uchida K, Okazaki K. Refractory autoimmune pancreatitis: azathioprine or steroid pulse therapy? Am J Gastroenterol. 2008;103(7):1834; author reply 1834–5.
- 35. Matsushita M, Yamashina M, Ikeura T, Shimatani M, Uchida K, Takaoka M, et al. Effective steroid pulse therapy for the biliary stenosis caused by autoimmune pancreatitis. Am J Gastroenterol. 2007;102(1):220–1.
- 36. Shigekawa M, Yamao K, Sawaki A, Hara K, Takagi T, Bhatia V, et al. Is (18)F-fluorodeoxyglucose positron emission tomography meaningful for estimating the efficacy of corticosteroid therapy in patients with autoimmune pancreatitis? J Hepatobiliary Pancreat Sci. 2010;17(3):269–74.
- 37. Nishino T, Toki F, Oyama H, Shimizu K, Shiratori K. Long-term outcome of autoimmune pancreatitis after oral prednisolone therapy. Intern Med. 2006;45(8):497–501.
- 38. Nishimori I, Tamakoshi A, Kawa S, Tanaka S, Takeuchi K, Kamisawa T, et al. Influence of steroid therapy on the course of diabetes mellitus in patients with autoimmune pancreatitis: findings from a nationwide survey in Japan. Pancreas. 2006;32(3): 244–8.
- Tanaka S, Kobayashi T, Nakanishi K, Okubo M, Murase T, Hashimoto M, et al. Corticosteroid-responsive diabetes mellitus associated with autoimmune pancreatitis. Lancet. 2000;356 (9233):910-1.
- Uchida K, Yazumi S, Nishio A, Kusuda T, Koyabu M, Fukata M, et al. Long-term outcome of autoimmune pancreatitis. J Gastroenterol. 2009;44(7):726–32.



Idiopathic Duct-Centric Pancreatitis (IDCP) with Immunological Studies

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Abstract

A 65-year-old woman with elevated serum levels of pancreatic enzymes was referred to our hospital for further examinations. Abdominal US and contrast-enhanced CT demonstrated swelling of the pancreas body and tail. MRCP and ERCP revealed abrupt ending of the MPD in the pancreas body. Under the suspicion of malignancy, distal pancreatectomy and splenectomy were performed. The histopathological findings showed idiopathic duct-centric pancreatitis (IDCP) with granulocytic epithelial lesions (GEL). As most cases of Japanese autoimmune pancreatitis (AIP) are lymphoplasmacytic sclerosing pancreatitis (LPSP), the present case seems to be helpful to clarify the clinical findings of IDCP in Japan.

Key words: autoimmune pancreatitis (AIP), Idiopathic duct-centric pancreatitis (IDCP), lymphoplasmacytic sclerosing pancreatitis (LPSP), granulocytic epithelial lesion (GEL), IgG4

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Introduction

Since Sarles et al reported a case of idiopathic pancreatitis with hypergammaglobulinemia in 1961 (1), several investigators have reported that autoimmune mechanisms may be involved in the etiology of chronic pancreatitis. Yoshida et al first proposed the concept of "autoimmune pancreatitis" (AIP) in 1995 (2). Thereafter, many cases of AIP have been reported mainly from Japan until the disease concept was accepted worldwide. As previously reported, the characteristic features (3, 4) of the Japanese patients with AIP show (i) diffuse enlargement of the pancreas on US, CT and MRI, (ii) irregular narrowing of the pancreatic duct (sclerosing pancreatitis) on endoscopic retrograde cholangiopancreatographic (ERCP) images, (iii) histologically termed lymphoplasmacytic sclerosing pancreatitis (LPSP) with fibrosis, abundant infiltration of lymphocyte and IgG4-positive plasmacytes and obliterative phlebitis, and (iv) it is often associated with extrapancreatic lesions, such as sclerosing cholangitis similar to primary sclerosing cholangitis (PSC), sclerosing cholecystitis, sclerosing sialoadenitis, retroperitoneal fibrosis, interstitial renal tubular disorders, enlarged celiac and hilar lymph nodes, chronic thyroiditis, and pseudotumor of the liver (5-7). On the other hand, in Western countries, another type of AIP different from the AIP commonly observed in Japan has been reported. In a study performed by a group at the Mayo Clinic, it was demonstrated that there may be two histological types of AIP, LPSP and idiopathic duct-centric pancreatitis (IDCP) (6, 8). IDCP was characterized by lobular fibrosis and pancreatic duct damage mainly caused by infiltration of neutrophils without obliterative phlebitis (8). Zamboni et al also recognized a subtype of AIP occurring in a subset of patients who are younger and more commonly have ulcerative colitis and Crohn's disease, which is characterized by the presence of granulocytic epithelial lesions (GEL) (9). There are a number of similarities in the clinical and histopathological findings between AIP

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