Student's t-test. In all tests, corrected P values of < 0.05 were considered statistically significant.

Results

Clinical manifestations

After 6 months' treatment, oral prednisolone had been administered to 19 patients in both groups [pulse group: 9 of 11 vs. oral group: 10 of 10; difference not significant (N.S.)], and the median dosage of prednisolone in each group was 10 mg/day (pulse group: 2.5–12.5 vs. oral group: 5–12.5; N.S.). Two patients in the pulse group dropped out of the maintenance therapy. Neither group showed severe or lethal complications.

In both groups, extrapancreatic lesions other than the bile duct lesions were observed in 10 patients (pulse group: 5 of 11 vs. oral group: 5 of 10; N.S.) (Tables 1, 2). No exacerbation of the extrapancreatic lesions was found following either of the treatments (data not shown). Laboratory findings including immunoglobulin, autoantibody, and exocrine function at the treatment start are listed in Tables 1 and 2.

Immunoglobulin

At the beginning of treatment, in both groups, abnormal serum immunoglobulin-G4 (IgG4) values were observed in all patients (Tables 1, 2), and abnormal serum immunoglobulin-G (IgG) values were observed in 13 patients (pulse group: 6 of 11 vs. oral group: 7 of 10; N.S.) (Tables 1, 2). Normalization of the IgG value was shown in all these patients within 6 months (data not shown).

Liver function

At the beginning of treatment, abnormal γ -GTP values were revealed in both groups, in a total of 18 patients (pulse group: 11 of 11 vs. oral group: 7 of 10; N.S.). In the pulse group, the median γ-GTP levels fell, from 222 IU/L (range 65-1,352) at the beginning of treatment to 92 IU/L (22-679) (P < 0.01) after 2 weeks of pulse therapy (Fig. 1a), and to 36 IU/L (19–556) (P < 0.01) after 8 weeks of pulse therapy (Fig. 1a). In the oral group, however, the median γ -GTP fell insignificantly after 2 weeks of prednisolone treatment, from 149 IU/L (range 67-380) at the beginning of treatment to 125 IU/L (63-274) (P = 0.083) (Fig. 1b), although the level fell significantly to 72 IU/L (29–114) (P = 0.027) after 8 weeks of prednisolone treatment (Fig. 1b). When limiting results to the patients who showed diffuse pancreatic swelling, γ -GTP was significantly improved in the pulse

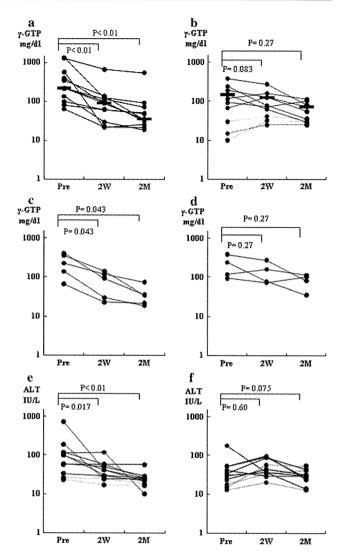


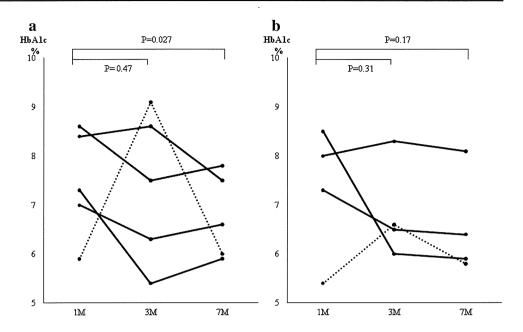
Fig. 1 γ -Guanosine triphosphate (γ -GTP) and alanine aminotransferase (ALT) changes after steroids. The serum levels of γ -GTP after steroid pulse therapy (a), after oral steroid therapy (b), those of ALT after steroid pulse therapy (e) and those after oral steroid therapy (f) were monitored on day 0 (Pre), and 2 weeks (W) and 8 weeks after therapy. To evaluate the therapeutic effect strictly, the patients (dotted lines) who did not show an abnormal value during the clinical course were excluded from this analysis. The serum levels of γ -GTP after 2 weeks on steroids and those of ALT after 2 and 8 weeks on steroids were significantly improved in the pulse group, compared with the oral group. When limiting the patients to those who showed diffuse pancreatic swelling, the serum level of γ -GTP was significantly improved in the pulse group after 2 and 8 weeks of pulse therapy (c), compared with the oral group (d). M Months

group after 2 weeks (P = 0.043) and 8 weeks (P = 0.043) of pulse therapy (Fig. 1c), whereas the improvement in the γ -GTP level was insignificant in the oral group after 2 weeks (P = 0.27) and 8 weeks (P = 0.27) of prednisolone treatment (Fig. 1d).

At the beginning of treatment, abnormal ALT values were revealed in both groups, in a total of 16 patients



Fig. 2 Glycosylated hemoglobin (HbA1C) changes after steroid therapy. The levels of HbA1c after steroid pulse therapy (a) and oral steroid therapy (b) were monitored at months 1, 3, and 7 after therapy, which closely reflect glucose tolerance at months 0, 2, and 6, respectively. To evaluate the therapeutic effect strictly. patients who did not show abnormal values during the clinical course were excluded from this analysis. Dotted lines represent the patients who developed glucose intolerance after 2 months of steroid therapy. The level of HbA1c at month 7 on steroids tended to be improved in the pulse group, compared with the oral group



(pulse group: 10 of 11 vs. oral group: 6 of 10; N.S.). In the pulse group, the median ALT level fell from 100 IU/L (range 33–721) at the beginning of treatment to 41 IU/L (24–117) (P=0.017) after 2 weeks of pulse therapy (Fig. 1e), and to 25 IU/L (10–57) (P<0.01) after 8 weeks of pulse therapy (Fig. 1e). In the oral group, however, the median ALT rose transiently from 46.5 IU/L (range 30–178) at the beginning of treatment to 62 IU/L (29–96) (P=0.60) after 2 weeks of prednisolone (Fig. 1f), but improved to 28 IU/L (14–44) (P=0.075) after 8 weeks of prednisolone treatment (Fig. 1f).

Endocrine function

Before steroid treatment, diabetes mellitus was seen in 7 patients (pulse group: 4 of 11 vs. oral group: 3 of 10; N.S.) (Tables 1, 2). One patient in each group developed glucose intolerance after 2 months of steroid therapy. Including these patients, all the patients with impaired glucose tolerance were treated with dietary measures and received medical therapy while the steroid therapy was maintained. Neither group showed significant improvement in glucose tolerance after 2 months, but at 6 months, the pulse group had improved significantly (P = 0.027), whereas the oral group had not (P = 0.17) (Fig. 2).

Pancreas size

Both groups evidenced pancreatic swelling: diffuse pancreatic swelling was observed in 10 patients (pulse group: 5 of 11 vs. oral group: 5 of 10; N.S.) and focal swelling was observed in 11 patients (pulse group: 6 of 11 vs. oral group: 5 of 10; N.S.) (Tables 1, 2). The change in pancreas size

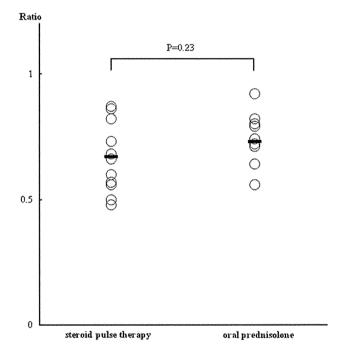


Fig. 3 Pancreas morphology changes after 2 weeks on steroids. Morphological changes of the pancreas after 2 weeks of steroid pulse therapy (*left*) and oral steroid therapy (*right*) were scored. The width of the pancreas along its longest axis was measured on computed tomography (CT) or magnetic resonance imaging (MRI) and compared with the transverse diameter of the vertebral body, referred to in the method of Heuck et al. [29]. The pancreas size on the first image was defined as 100%, and the ratio after 2 weeks with each treatment was measured in the same manner. The *black bars* represent the mean in each group. The two groups showed no significant difference in morphological change of the pancreas after 2 weeks

after 2 weeks showed no significant difference between the groups after treatment (pulse group: 67% vs. oral group: 73.4%, P = 0.23) (Fig. 3).



Fig. 4 Lower bile duct stricture changes after steroids. The changes in lower bile duct stricture in the pulse group (a) and the oral group (b) on day 0 and 2 weeks after therapy were scored as follows: 0 = absent, 1 = <0-25%, 2 = <25-50%, 3 = <50-75%, 4 = <75-100%, referred to in the method of Craig et al. [30]. Bile duct stenosis in the distal third part was significantly improved after 2 weeks in each group

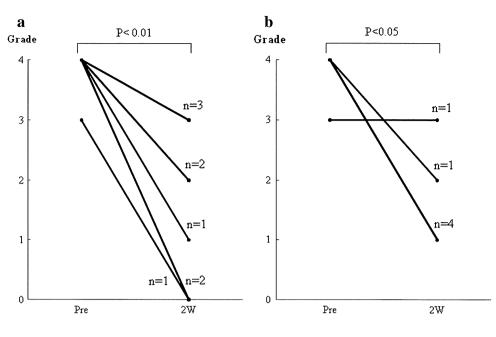
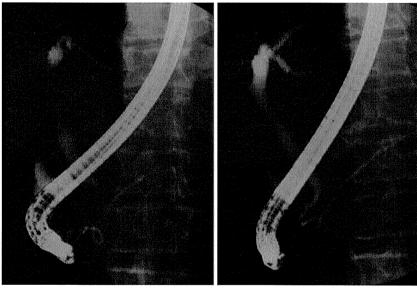


Fig. 5 Endoscopic retrograde cholangiopancreatography (ERCP) images of the impact of steroid pulse therapy on refractory autoimmune pancreatitis (AIP). Although oral prednisolone was commenced, it had had no effect on the biliary stenosis (*left*). Two courses of steroid pulse therapy ameliorated the stenosis dramatically (*right*) (Case 10; reference [31])



Bile duct lesion

After 2 weeks on steroids, significant improvement of lower bile duct stricture was shown in both groups (pulse group: P < 0.01, and oral group: P < 0.05) (Fig. 4). However, there was one patient (case 10) whose lower bile duct stricture did not improve following oral prednisone treatment, but showed definite improvement with steroid pulse therapy [31] (Fig. 5).

Discussion

Several cases have been reported of pancreatic cancer or bile duct cancer concurrent with AIP [16–19]; because the

image findings of AIP often mimic pancreatobiliary malignancies, it is extremely crucial to distinguish AIP from cancers, although this can be difficult [12–15].

In the Asian diagnostic criteria proposed by the Japan-Korea symposium on autoimmune pancreatitis [11], the use of steroids as diagnostic treatment was allowed only when the imaging findings were compatible with AIP and only after there was a negative result for malignancy work-up. Cases of diagnostic treatment using steroids will doubtless increase in future. The usefulness of a 2-week conventional oral steroid diagnostic treatment was also proposed by Moon et al. [20]. However, oral steroid therapy requires a long period for drug tapering, because any patient who has received a glucocorticoid in doses equivalent to at least 20 mg a day of prednisone for >5 days is at risk of



secondary adrenal insufficiency due to hypothalamic—pituitary—adrenal suppression [21–23]. Diagnostic treatment with an oral steroid may cause an undesirable effect when surgical resection is required. Although one article reported that azathioprine was used for refractory AIP [32], it is causative of acute pancreatitis [33]. Therefore, a safe and simple alternative to oral steroid treatment is needed for refractory AIP [34].

Steroid pulse therapy is a well known alternative to oral steroid treatment for autoimmune disorders; it requires no drug tapering [28], and we have already reported cases where steroid pulse therapy was effective for AIP [35], although comparative studies of conventional oral steroid therapy and steroid pulse therapy have not been reported. Here we report that steroid pulse therapy is an effective alternative to oral steroid for the initial treatment of AIP.

The efficacy of oral steroid therapy for AIP is well known, and the improvement of AIP in patients treated with steroids for 2 weeks can be shown in radiographic findings [5, 20, 36]. Our data on pancreas size after 2 weeks did not show a significant difference between oral steroid and pulse therapy. Both therapies were effective for alleviating lower bile duct stricture in the short term and for resolving abnormal IgG values. However, the shortterm change in y-GTP showed significant improvement in the steroid pulse therapy group, but not in the oral steroid group. In one patient, lower bile duct stricture was improved by steroid pulse therapy, although it had not been improved by oral steroids. These findings suggest that steroid pulse therapy may prevent patients from having unnecessary major operations for benign bile duct lesions which do not respond to oral steroid treatment.

Steroid therapy is reported to be effective in approximately half of AIP patients with accompanying diabetes mellitus [37, 38]. The mechanism of steroid action in the recovery of endocrine function in patients with AIP is unclear, but Tanaka et al. [39] have suggested that steroids can suppress the release of cytokines produced by inflammatory cells, and enable islet regeneration and eventual restoration of insulin secretion. However, we have reported that conventional oral steroids did not improve diabetes mellitus in the long term [40]. Although the accumulation of a larger numbers of cases is needed, our study of glycosylated hemoglobin values showed that steroid pulse therapy tended to be more effective than oral steroids for improving glucose tolerance after 6 months of treatment. In our protocol, there was a decisive difference between the two therapies just for the initial 2 weeks, because 20 mg/day of oral prednisolone was prescribed as maintenance therapy after the 2 weeks of steroid pulse therapy. We presume that the steroid pulse therapy was better than the oral steroid therapy for improving glucose tolerance because of its stronger cytokine suppression.

In summary, steroid pulse therapy is an effective alternative initial treatment for AIP, and surpasses conventional oral steroid therapy in the improvement of bile duct lesions. When a differential diagnosis between AIP and pancreatic cancer is difficult clinically or when bile duct lesions do not respond to oral steroid treatment, we recommend steroid pulse therapy as an alternative to oral steroid therapy. In future, the accumulation of a larger number of patients receiving steroid pulse therapy is needed, and prospective studies will be required.

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References

- Sarles H, Sarles JC, Muratore R, Guien C. Chronic inflammatory sclerosis of the pancreas: an autonomous pancreatic disease? Am J Dig Dis. 1961;6:688–98.
- Yoshida K, Toki F, Takeuchi T, Watanabe S, Shiratori K, Hayashi N. Chronic pancreatitis caused by an autoimmune abnormality. Proposal of the concept of autoimmune pancreatitis. Dig Dis Sci. 1995;40(7):1561-8.
- Okazaki K, Chiba T. Autoimmune related pancreatitis. Gut. 2002;51(1):1–4.
- Hamano H, Kawa S, Horiuchi A, Unno H, Furuya N, Akamatsu T, et al. High serum IgG4 concentrations in patients with sclerosing pancreatitis. N Engl J Med. 2001;344(10):732–8.
- Kamisawa T, Egawa N, Nakajima H, Tsuruta K, Okamoto A. Morphological changes after steroid therapy in autoimmune pancreatitis. Scand J Gastroenterol. 2004;39(11):1154–8.
- Uchida K, Okazaki K, Konishi Y, Ohana M, Takakuwa H, Hajiro K, et al. Clinical analysis of autoimmune-related pancreatitis. Am J Gastroenterol. 2000;95(10):2788–94.
- 7. Kamisawa T, Shimosegawa T, Okazaki K, Nishino T, Watanabe H, Kanno A, et al. Standard steroid treatment for autoimmune pancreatitis. Gut. 2009;58(11):1504–7 (epub 26 April 2009).
- Okazaki K, Kawa S, Kamisawa T, Ito T, Inui K, Irie H, et al. Japanese clinical guidelines for autoimmune pancreatitis. Pancreas. 2009;38(8):849–66.
- Kim KP, Kim MH, Kim JC, Lee SS, Seo DW, Lee SK. Diagnostic criteria for autoimmune chronic pancreatitis revisited. World J Gastroenterol. 2006;12(16):2487–96.
- Chari ST, Smyrk TC, Levy MJ, Topazian MD, Takahashi N, Zhang L, et al. Diagnosis of autoimmune pancreatitis: the Mayo Clinic experience. Clin Gastroenterol Hepatol. 2006;4(8):1010–6.
- 11. Otsuki M, Chung JB, Okazaki K, Kim MH, Kamisawa T, Kawa S, et al. Asian diagnostic criteria for autoimmune pancreatitis: consensus of the Japan-Korea symposium on autoimmune pancreatitis. J Gastroenterol. 2008;43(6):403–8.
- Kamisawa T, Egawa N, Nakajima H, Tsuruta K, Okamoto A, Kamata N. Clinical difficulties in the differentiation of autoimmune pancreatitis and pancreatic carcinoma. Am J Gastroenterol. 2003;98(12):2694–9.
- 13. Nakazawa T, Ohara H, Sano H, Ando T, Imai H, Takada H, et al. Difficulty in diagnosing autoimmune pancreatitis by imaging findings. Gastrointest Endosc. 2007;65(1):99–108.



- Okazaki K, Uchida K, Fukui T. Recent advances in autoimmune pancreatitis: concept, diagnosis, and pathogenesis. J Gastroenterol. 2008;43(6):409–18.
- Frulloni L, Scattolini C, Falconi M, Zamboni G, Capelli P, Manfredi R, et al. Autoimmune pancreatitis: differences between the focal and diffuse forms in 87 patients. Am J Gastroenterol. 2009;104(9):2288–94.
- Fukui T, Mitsuyama T, Takaoka M, Uchida K, Matsushita M, Okazaki K. Pancreatic cancer associated with autoimmune pancreatitis in remission. Intern Med. 2008;47(3):151–5.
- Witkiewicz AK, Kennedy EP, Kennyon L, Yeo CJ, Hruban RH. Synchronous autoimmune pancreatitis and infiltrating pancreatic ductal adenocarcinoma: case report and review of the literature. Hum Pathol. 2008;39(10):1548–51.
- Inoue H, Miyatani H, Sawada Y, Yoshida Y. A case of pancreas cancer with autoimmune pancreatitis. Pancreas. 2006;33(2): 208-9
- Oh HC, Kim JG, Kim JW, Lee KS, Kim MK, Chi KC, et al. Early bile duct cancer in a background of sclerosing cholangitis and autoimmune pancreatitis. Intern Med. 2008;47(23):2025–8.
- 20. Moon SH, Kim MH, Park DH, Hwang CY, Park SJ, Lee SS, et al. Is a 2-week steroid trial after initial negative investigation for malignancy useful in differentiating autoimmune pancreatitis from pancreatic cancer? A prospective outcome study. Gut. 2008;57(12):1704–12.
- Fraser CG, Preuss FS, Bigford WD. Adrenal atrophy and irreversible shock associated with cortisone therapy. JAMA. 1952;149(17):1542–3.
- Axelrod L. Perioperative management of patients treated with glucocorticoids. Endocrinol Metab Clin North Am. 2003;32(2): 367–83.
- Alves C, Robazzi TC, Mendonça M. Withdrawal from glucocorticosteroid therapy: clinical practice recommendations. J Pediatr (Rio J). 2008;84(3):192–202.
- Kee TY, Chapman JR, O'Connell PJ, Fung CL, Allen RD, Kable K, et al. Treatment of subclinical rejection diagnosed by protocol biopsy of kidney transplants. Transplantation. 2006;82(1):36–42.
- Wiersinga WM. Management of Graves' ophthalmopathy. Nat Clin Pract Endocrinol Metab. 2007;3(5):396–404.
- Saha M, Powell AM, Bhogal B, Black MM, Groves RW. Pulsed intravenous cyclophosphamide and methylprednisolone therapy in refractory pemphigus. Br J Dermatol. 2010;162(4):790–7.
- Parker BJ, Bruce IN. High dose methylprednisolone therapy for the treatment of severe systemic lupus erythematosus. Lupus. 2007;16(6):387–93.
- Boumpas DT, Chrousos GP, Wilder RL, Cupps TR, Balow JE. Glucocorticoid therapy for immune-mediated diseases: basic and clinical correlates. Ann Intern Med. 1993;119(12):1198–208.

- Heuck A, Maubach PA, Reiser M, Feuerbach S, Allgayer B, Lukas P, et al. Age-related morphology of the normal pancreas on computed tomography. Gastrointest Radiol. 1987;12(1):18–22.
- Craig DA, MacCarty RL, Wiesner RH, Grambsch PM, LaRusso NF. Primary sclerosing cholangitis: value of cholangiography in determining the prognosis. AJR Am J Roentgenol. 1991;157(5): 959-64
- 31. Nakayama S, Matsushita M, Yamashina M, Shimatani M, Uchida K, Okazaki K. Effective steroid mini-pulse therapy for refractory biliary stenosis due to autoimmune pancreatitis (in Japanese with English abstract). Suizou. 2006;21(5):446–52.
- 32. Church NI, Pereira SP, Deheragoda MG, Sandanayake N, Amin Z, Lees WR, et al. Autoimmune pancreatitis: clinical and radiological features and objective response to steroid therapy in a UK series. Am J Gastroenterol. 2007;102(11):2417–25.
- Floyd A, Pedersen L, Nielsen GL, Thorlacius-Ussing O, Sorensen HT. Risk of acute pancreatitis in uses of azathioprine: a population-based case-control study. Am J Gastroenterol. 2003;98(6): 1305–8.
- Matsushita M, Ikeura T, Fukui T, Uchida K, Okazaki K. Refractory autoimmune pancreatitis: azathioprine or steroid pulse therapy? Am J Gastroenterol. 2008;103(7):1834; author reply 1834–5.
- 35. Matsushita M, Yamashina M, Ikeura T, Shimatani M, Uchida K, Takaoka M, et al. Effective steroid pulse therapy for the biliary stenosis caused by autoimmune pancreatitis. Am J Gastroenterol. 2007;102(1):220–1.
- 36. Shigekawa M, Yamao K, Sawaki A, Hara K, Takagi T, Bhatia V, et al. Is (18)F-fluorodeoxyglucose positron emission tomography meaningful for estimating the efficacy of corticosteroid therapy in patients with autoimmune pancreatitis? J Hepatobiliary Pancreat Sci. 2010;17(3):269–74.
- Nishino T, Toki F, Oyama H, Shimizu K, Shiratori K. Long-term outcome of autoimmune pancreatitis after oral prednisolone therapy. Intern Med. 2006;45(8):497–501.
- 38. Nishimori I, Tamakoshi A, Kawa S, Tanaka S, Takeuchi K, Kamisawa T, et al. Influence of steroid therapy on the course of diabetes mellitus in patients with autoimmune pancreatitis: findings from a nationwide survey in Japan. Pancreas. 2006;32(3): 244–8.
- Tanaka S, Kobayashi T, Nakanishi K, Okubo M, Murase T, Hashimoto M, et al. Corticosteroid-responsive diabetes mellitus associated with autoimmune pancreatitis. Lancet. 2000;356 (9233):910-1.
- 40. Uchida K, Yazumi S, Nishio A, Kusuda T, Koyabu M, Fukata M, et al. Long-term outcome of autoimmune pancreatitis. J Gastroenterol. 2009;44(7):726–32.



REVIEW

Recent advances in the concept and diagnosis of autoimmune pancreatitis and IgG4-related disease

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Abstract Recent studies have suggested the existence of two subtypes of autoimmune pancreatitis (AIP): type 1 AIP, related to IgG4 (lymphoplasmacytic sclerosing pancreatitis); and type 2 AIP, related to a granulocytic epithelial lesion (idiopathic duct-centric chronic pancreatitis). Compared with type 2 AIP, the clinicopathological features of type 1 AIP, with increased serum IgG4/IgE levels, abundant infiltration of IgG4 + plasmacytes and lymphocytes, autoantibodies, and steroid responsiveness, are more suggestive of abnormal immunity such as allergy or autoimmunity. Moreover, patients with type 1 AIP often have extrapancreatic lesions, such as sclerosing cholangitis, sclerosing sialadenitis, or retroperitoneal fibrosis, showing pathological features similar to those of the pancreatic lesions. Based on these findings, an international concept of and diagnostic criteria for AIP have been proposed recently. Of interest, many synonyms have been proposed for the conditions of AIP and extrapancreatic lesions associated with IgG4, such as "multifocal idiopathic "IgG4-related autoimmune disease," fibrosclerosis," "IgG4-related sclerosing disease," "systemic IgG4-related plasmacytic syndrome (SIPS)," and "IgG4-related multiorgan lymphoproliferative syndrome," all of which may refer to the same conditions. Therefore, the Japanese Research Committee for "Systemic IgG4-Related Sclerosing Disease" proposed a disease concept and clinical diagnostic criteria based on the concept of multifocal fibrosclerosing disease, in 2009, in which the term "IgG4-related disease" was agreed upon as a minimal consensus to cover these conditions. Although the significance of IgG4 in the development of "IgG4-related disease" remains unclear, we have proposed a hypothesis for the development of type 1 AIP, one of the IgG4-related diseases. The concept and diagnostic criteria of "IgG4-related disease" will be changed in accordance with future studies.

Keywords IgG4 · IgG4-related disease · Autoimmune pancreatitis · Mikulicz disease · Regulatory T cell (Treg)

Abbreviations

11001011	
AIP	Autoimmune pancreatitis
ANA	Anti-nuclear antibody
CA-II	Carbonic anhydrase-II
CTLA-4	Cytotoxic T lymphocyte antigen-4
ERCP	Endoscopic retrograde cholangio-
	pancreatography
FCRL	Fc-receptor-like
IFN-γ	Interferon-γ
IL-4	Interleukin-4
LF	Lactoferrin
LPSP	Lymphoplasmacytic sclerosing pancreatitis
MD	Mikulicz disease
MHC	Major histocompatibility complex
MOLPS	Multiorgan lymphoproliferative disease
PBP	Plasminogen-binding protein
SjS	Sjögren's syndrome
PSC	Primary sclerosing cholangitis
RF	Rheumatoid factor
SIPS	Systemic IgG4 plasmacytic syndrome
SLE	Systemic lupus erythematosus
Treg	Regulatory T cell
UBR2	Ubiquitin-protein ligase E3 component
	n-recognin 2

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Introduction

In 1961, Sarles et al. [1] first observed a case of particular pancreatitis with hypergammaglobulinemia. Yoshida et al. [2] first proposed the concept of autoimmune pancreatitis (AIP). Hamano et al. [3] reported increased serum levels of IgG4 in Japanese patients with AIP. Thereafter, many studies of AIP were reported, mainly by Japanese investigators. The histopathological findings of AIP are characterized by the periductal localization of predominantly CD4-positive T cells, IgG4-positive plasma cells, storiform fibrosis with acinar cell atrophy frequently resulting in stenosis of the main pancreatic duct, and obliterative fibrosis [4-6], which is also called lymphoplasmacytic sclerosing pancreatitis (LPSP) [7]. In 2003, Kamisawa et al. [8] suggested that AIP is a systemic sclerosing disease, based on the findings that the pancreas and other involved organs have fibrosis with abundant infiltration of IgG4-positive plasma cells, which is similar to the concept of multifocal fibrosclerosis proposed by Comings et al. [9]. Further histological and clinical profiling of patients with "AIP" reveals two distinct subtypes, type 1 and type 2 AIP [10, 11]. Type 1 AIP is classified as a pancreatic manifestation of IgG4-related disease, probably a systemic disease with an autoimmune process, whereas type 2 AIP is regarded as a specific pancreatic disease with a granulocytic epithelial lesion (GEL) [12, 13] and occasional coexistence with ulcerative colitis [10, 11].

Of note, patients with Mikulicz's disease (MD) -originally classified as an atypical type of Sjögren's syndromewho usually have bilateral, painless, and symmetrical swelling of the lachrymal, parotid, and submandibular glands [14], show elevated serum levels of IgG4, infiltration of IgG4-positive plasma cells into the glands, and recovery of secretion with steroid treatment. Similar to patients with AIP, these patients often show other organ involvement (OOI) such as AIP, sclerosing cholangitis, retroperitoneal fibrosis, enlarged celiac and hilar lymph nodes, chronic thyroiditis, and interstitial nephritis [4–6, 14–16]. Recently, however, MD has been considered to be completely different from Sjögren's syndrome because of the lack of anti-SS-A/ Ro or anti-SS-B/La antibodies, and the showing of steroid responsiveness [2-6]. The steroid responses and the prognoses of AIP patients with sclerosing cholangitis differ from these features in patients with primary sclerosing cholangitis (PSC), which suggests different pathological conditions. These findings led us to the concept of "IgG4-related disease" such as IgG4-related systemic sclerosing disease [8, 17], systemic IgG4-related plasmacytic syndrome (SIPS) [18], and IgG4-positive multiorgan lymphoproliferative syndrome (IgG4-MOLPS) [19]. Although the pathogenesis and pathophysiology of AIP remain unclear, we will discuss the most recent advances in the concept of AIP, especially IgG4-related type 1 AIP, and the advances in the novel concept of "IgG4-related disease."

Subtypes of autoimmune pancreatitis: type 1 and type 2 AIP

Recent studies have suggested that "AIP" manifests as two distinct subtypes, type 1 and type 2 AIP (Table 1) [10, 11]. Hypergammaglobulinemia, the deposition of immunoglobulins, the presence of autoantibodies, and steroid efficacy in Type 1 AIP confirm the definition of autoimmune disease proposed by Mackay [20]. Different from type 1 AIP, patients with type 2 AIP have no serological markers of autoimmunity. Therefore, it is still in debate as to whether or not type 2 AIP should be classified as a clinical entity of AIP, but the deposition of C3c and IgG in the basement membrane of pancreatic ducts and acini suggests an immune complex-mediated destruction of ducts and acini in type 2 AIP as well as type 1 AIP [21]. The nomenclature of the two subtypes of AIP and international consensus diagnostic criteria were proposed at the meeting of the International Association of Pancreatology held at Fukuoka in 2010 (Tables 2, 3) [22].

Type 1 AIP

In type 1 AIP, whose histological description is called LPSP, the pancreatic histopathology shows the following characteristic features: (1) abundant infiltration of plasma cells (IgG4⁺ cells; high-power field [hpf], IgG4/IgG cells; >40%) and lymphocytes, (2) peculiar storiform or swirling fibrosis, and (3) perivenular infiltration with lymphocytes and plasma cells often leading to obliterative phlebitis. Clinically, type 1 AIP seems to be the pancreatic manifestation of the recently proposed IgG4-related disease, characterized by swelling of the pancreas, elevated serum IgG4 levels, and extrapancreatic lesions (e.g., sclerosing cholangitis, sclerosing sialadenitis, and retroperitoneal fibrosis) associated with the infiltration of abundant IgG4 + plasma cells. Patients with type 1 AIP are often elderly males who have obstructive jaundice, and the pancreatic and extrapancreatic manifestations respond to steroid therapy.

Extrapancreatic lesions in type 1 AIP

A variety of extrapancreatic lesions have been noted in patients with AIP, including lachrymal and salivary gland lesions [23], pulmonary lesions including hilar lymphadenopathy [24], sclerosing cholangitis [25, 26], retroperitoneal fibrosis [27], and tubulointerstitial nephritis (TIN) [15, 28, 29]. Associations were also reported with hypophysitis



Table 1 Subtypes of autoimmune pancreatitis (AIP)

Subtype of AIP	Type 1	Type 2
Other nomenclatures	AIP without GEL	AIP with GEL
	IgG4-related LPSP	IgG4-unrelated IDCP
Prevalence	Asia > USA, Europe	Europe > USA > Asia
Age	High age	Younger
Gender	Male ≫ female	Male = female (NS)
Symptoms	Often obstructive jaundice rare abdominal pain	Often obstructive jaundice abdominal pain like acute pancreatitis
Pancreas images	Swelling (diffuse/segmental/focal)/ mass-forming	Swelling (diffuse/segmental/focal)/mass-forming
Serology	High serum IgG, IgG4, autoAbs (+)	Normal IgG, normal IgG4, autoAbs (-)
Other organ involvement (OOI)	Sclerosing cholangitis	Unrelated to OOI
	Sclerosing sialadenitis	
	Retroperitoneal fibrosis	
	Others	
Ulcerative colitis	Rare	Often
Steroid	Responsive	Responsive
Relapse	High rate	Rare

GEL granulocytic epithelial lesion, LPSP lymphoplasmacytic sclerosing pancreatitis, IDCP idiopathic duct-centric chronic pancreatitis, Abs antibodies, NS not significant

[30], chronic thyroiditis [16, 31], and prostatitis [32]. Other extrapancreatic involvements have been reported in a few cases [33-36]. Though it is not certain that all of these involvements have a relation with AIP, extrapancreatic lesions are prevalent systemically in various organs (Table 4) [29-41], suggesting that type 1 AIP, but not type 2 AIP, may be a pancreatic manifestation of IgG4-related disease. The extrapancreatic lesions appear synchronously or metachronously with the pancreatic lesion(s), share the same pathological conditions, and show favorable responses to steroid therapy; these characteristics suggest a common pathophysiological background. The lesions are usually detected by imaging and blood tests (computed tomography [CT], magnetic resonance imaging [MRI], gallium scintigraphy, fluorodeoxyglucose positron emission tomography [FDG-PET], and IgG4); however, such findings should be confirmed by histological findings. Extrapancreatic lesions sometimes mimic, or are misdiagnosed as, primary lesions of the corresponding organs: lachrymal and salivary gland lesions for Sjögren's syndrome, respiratory lesions for sarcoidosis, and sclerosing cholangitis for PSC. Therefore, it is necessary to differentiate between IgG4-related diseases and inherent diseases of the corresponding organs. Patients with IgG4related sialodacryoadenitis, synonymous with IgG4-related MD [13, 41], usually have symmetrical enlargement of the salivary and lacrimal glands. The IgG4-related central nervous system lesions include infundibulohypophysitis, hypertrophic pachymeningitis, intracranial inflammatory pseudotumor, and orbital pseudotumor [21-41].

Type 2 AIP

Type 2 AIP was proposed from histological examination of pancreases resected from patients with chronic non-alcoholic pancreatitis by American and European pathologists, who reported another histopathological pattern, named idiopathic duct-centric pancreatitis (IDCP) or AIP with GEL [11–13]. The most characteristic feature of type 2 AIP is the GEL, often with destruction and obliteration of the pancreatic duct. Type 2 AIP has swelling of the pancreas, but none or very few IgG4-positive plasma cells, and clinical features show a distinctly different profile from that of type 1 AIP, with no associated serum IgG4, IgG elevation, presence of autoantibodies, or other organ involvement, except for inflammatory bowel disease (approximately 30%).

The concept of IgG4-related disease and proposal of the clinical diagnostic criteria

Patients with IgG4-related disease show diffuse/focal organ enlargement, with mass-forming or nodular/thickened lesions in various organs, occurring synchronously or metachronously, due to the prominent infiltration of lymphocytes and plasmacytes with fibrosis [21–41]; however, the causes of the disease are still not clear. The organs known to be affected include the pancreas, biliary duct, lacrimal/salivary glands, retroperitoneum, central nervous system, thyroid gland, lungs, liver, gastrointestinal tract, kidneys, prostate gland, and lymph nodes [21–41]. Clinical



Table 2 Diagnosis of definitive and probable type 1 AIP using international consensus diagnostic criteria (ICDC) [22]

Diagnosis		imary basis for diagnosis	Imaging evidence	Collateral evidence
Definitive type 1 AIP Histolog Imaging			Typical/indeterminate Typical Indeterminate	Histologically confirmed LPSP (level 1 H) Any non-D level 1/level 2 Two or more from level 1 (+level 2 D*)
	Re	esponse to steroid	Indeterminate	Level 1 S/OOI + Rt or level 1 D + level 2 S/OOI/H + R
Probable typ			Indeterminate	Level 2 S/OOI/H + Rt
	Criterion	Level 1		Level 2
P	Parenchymal imaging	Typical		Indeterminate (including atypical*):
		=	nent with delayed enhancement ciated with rim-like enhancement)	Segmental/focal enlargement with delayed enhancement
D	Ductal imaging (ERP)	-	th of the MPD) or multiple strictures upstream dilatation	Segmental/focal narrowing without marked upstream dilatation (duct size <5 mm)
S	Serology	$IgG4 > 2 \times uppe$	er limit of normal value	IgG4 1-2× upper limit of normal value
	Other organ involveme	ent a or b		a or b
		a. Histology of <i>Any three of the</i>	extrapancreatic organs following	a. Histology of extrapancreatic organs including endoscopic biopsies of bile duct**
		·	plasmacytic infiltration with fibrosis and	Both of the following
		without granul	ocytic infiltration	Marked lymphoplasmacytic infiltration without granulocytic infiltration
		Storiform fibrosi		Abundant (>10 cells/hpf) IgG4-positive cells
		Obliterative phle		b. Physical or radiological evidence
			cells/hpf) IgG4 positive cells	At least one
			ological evidence	Symmetrically enlarged salivary/lacrimal glands
			ple proximal (hilar/intra hepatic) or listal bile duct stricture	Radiological evidence of renal involvement described in association with AIP
		Retroperitoneal t	îbrosis	
н н	Histology of the pancre	eas LPSP (core biop	sy/resection)	LPSP (core biopsy)
		At least 3 of the	following	Any 2 of the following
		Periductal lymph granulocytic in	oplasmacytic infiltrate without filtration	Periductal lymphoplasmacytic infiltrate without granulocytic infiltration
		Obliterative phle	bitis	Obliterative phlebitis
		Storiform fibrosi	s	Storiform fibrosis
		Abundant (>10	cells/hpf) IgG4-positive cells	Abundant (>10 cells/hpf) IgG4positive cells
Diagnostic s	teroid trial			
Response t	to steroid (Rt)#		s) radiologically demonstrable resolution or rement in pancreatic/extrapancreatic manifes	

Table 3 Diagnosis of definitive and probable type 2 AIP using international consensus diagnostic criteria (ICDC) [22]

Diagnosis		Imaging evidence Collateral ev		lence	
Definitive type 2 AIP Probable type 2 AIP				onfirmed IDCP (level 1 H) 0 + level 2 H + Rt	
		Typical/ indeterminate	Level 2 H/clinica	Level 2 H/clinical IBD + Rt	
	Criterion	Level 1		Level 2	
P	Parenchymal imaging	Typical		Indeterminate (including atypical*)	
		Diffuse enlargement with delayed enhancement (sometimes associated with rim-like enhancement)		Segmental/focal enlargement with delayed enhancement	
D	Ductal imaging (ERP)	Long (>1/3 length of the MPD) or strictures without marked upstream	*	Segmental/focal narrowing without marked upstream dilatation (duct size <5 mm)	
OOI	Other organ involvement			Clinically diagnosed inflammatory bowel disease	
Н	Histology, of the pancreas (core biopsy/resection)	IDCP:			
		Both of the following		Both of the following	
		Granulocytic infiltration of duct wal without granulocytic acinar inflam	, ,	Granulocytic and lymphoplasmacytic acinar infiltrate Absent or scant (0–10 cells/hpf) IgG4-positive cells	
		Absent or scant (0-10 cells/hpf) IgG4-positive cells		,	
Diagnostic st	teroid trial				
Response to steroid (Rt)#		Rapid (<2 weeks) resolution or mar	Rapid (<2 weeks) resolution or marked improvement in manifestations		

IBD inflammatory bowel disease, D ductal imaging, Rt response to steroid treatment, H histology, ERP endoscopic retrograde pancreatography

symptoms vary depending on the organ in which the lesions are located, but many cases are treated effectively by steroid therapy [21–41]. The prognosis is not clear; however, some patients develop serious complications such as obstructive jaundice due to hepatic, gallbladder, or pancreatic lesions; hydronephrosis due to retroperitoneal fibrosis; or respiratory symptoms due to "pulmonary lesions" [17–19, 26–29]. Although the infiltration of IgG4-positive cells and increased serum levels of IgG4 are characteristic of IgG4-related disease, the severity of fibrosis seems to be different among the individual involved organs. These conditions are quite similar to multifocal idiopathic fibrosclerosis (MIF) [9].

In addition to MIF, there are many synonyms, such as IgG4-related autoimmune disease [8], "IgG4-related

Table 4 Extrapancreatic lesions complicated with autoimmune pancreatitis. (from Ref. [38])

Close association

Lachrymal gland inflammation

Sialoadenitis

Hilar lymphadenopathy

Interstitial pneumonitis

Sclerosing cholangitis

Retroperitoneal fibrosis

Tubulointerstitial nephritis

Possible association

Hypophysit is

Autoimmune neurosensory hearing loss

Uveitis

Chronic thyroiditis

Pseudotumor (breast, lung, liver)

Gastric ulcer

Swelling of papilla of Vater

IgG4 hepatopathy

Aortitis

Prostatitis

Schonlein-Henoch purpura

Autoimmune thrombocytopenia

sclerosing disease" [17], SIPS [18], and "IgG4 + MOL-PS" [19], all of which may refer to the same conditions. It has been debated which term is the most appropriate. Storiform fibrosis and obliterative phlebitis are characteristic in the pancreatic and biliary tract lesions, but the degree varies depending on the individual organs, e.g., these features are very seldom found in lachrymal/salivary gland lesions or lymph node lesions. The term "IgG4-related sclerosing disease" is mainly based on fibrous swollen organs, whereas the terms "IgG4-SIPS" and "IgG4 + MOLPS" are based on lymphoplasmacytic proliferation and swollen lymph nodes without fibrosis.

Although most patients with type 1 AIP have multiorgan lesions that occur synchronously or metachronously, about 10-20% of the patients show a solitary organ involved without confirmation of other organ involvement. Therefore, it is unclear whether or not the pathogenetic mechanism is the same in individual organs. Based on these findings, the members of the Japanese Research Committees for "Systemic IgG4-Related Sclerosing Disease" (chaired by Professor K. Okazaki) [40] and "IgG4-MOLPS" (chaired by Professor H. Umehara) [41], both of which Committees were supported by the "Research for Intractable Disease Program from the Ministry of Health, Labor and Welfare of Japan", have agreed that the term "IgG4-related disease" be regarded as minimally accepting these conditions at present. To study these conditions, the Japanese Research Committee for "Systemic IgG4-Related Sclerosing Disease" (chaired by Professor K. Okazaki) proposed a disease concept and clinical diagnostic criteria of "systemic IgG4-related sclerosing disease" in 2009 (Table 5) [40]. However, the concept and diagnostic criteria should be changed in accordance with the findings of the future studies.

Immunological approaches to the pathophysiology of AIP and IgG4-related disease

The pathogenesis and pathophysiology of AIP have been studied mainly from immunological approaches and

Table 5 Clinical diagnostic criteria 2009 for IgG4-related disease (proposed by the Japanese Research Committee for "Systemic IgG4-related Sclerosing Disease") [40]

- (1) Clinically, diffuse/focal enlargement, or mass-forming, nodular/thickened lesions in one or more organs
- (2) Elevated levels of serum IgG4 (>135 mg/dl)
- (3) Histopathological findings
 - ① Prominent infiltration of lymphocytes and plasmacytes with fibrosis, but no neutrophilic infiltration
- 2 Abundant infiltration of IgG4-positive plasmacytes (>10/hpf) and/or a ratio of IgG4/IgG-positive cells of >40%
- ③ Storiform/swirling fibrosis
- Obliterative phlebitis

Diagnosis of IgG4-related disease: (1) + (2), $(1) + (3) \oplus 2$, $(2) + (3) \oplus 2$, or $(3) \oplus 2 \oplus 4$

The following cases must be excluded from the diagnosis: malignant tumors developed in organs (e.g., cancers, malignant lymphomas) or similar diseases (e.g., Sjögren's syndrome, primary sclerosing cholangitis), bronchial asthma, and Castleman's disease



studies have focused mainly on IgG4-related type 1 AIP, because little evidence of abnormal immunity has been reported in type 2 AIP.

Humoral immunity

IgG4 and its possible role in IgG4-related diseases

In healthy subjects, IgG1 usually accounts for most of the total IgG [42]. Generally, the amount of IgG4 does not vary with sex or age, and the quantity of IgG4 as well as the IgG4/total IgG ratio tends to remain constant [42]. The ratios for each IgG subclass are 65% of IgG1, 25% of IgG2, 6% of IgG3, and 4% of IgG4 [42]. In IgG4-related diseases, total IgG, IgG1, IgG2, IgG4, and IgE ratios are usually increased compared with healthy subjects, while IgM, IgA, and the ratios of IgG to IgM or IgA, are decreased compared with findings in healthy subjects or those with other diseases [3, 15, 19, 43]. Ratios of IgG subclasses other than IgG4 are somewhat different among individual diseases; in AIP, all subclasses (IgG1-G4) of IgG are increased compared with findings in other types of pancreatitis. In contrast, IgG₁ and IgG₃ in MD show significantly lower negative correlations with IgG4 than those shown in typical Sjögren's syndrome.

Although the association of IgE-mediated allergy and IgG4 antibodies is well known [44], IgG4 characteristics are still poorly understood. Basically, IgG4 has non-active characteristics for immune responses involved in a continuous process referred to as 'Fab-arm exchange', which occurs by the swapping of a heavy chain and attached light chain (half-molecule) with a heavy-light chain pair from another molecule [45], which usually results in asymmetric antibodies with two different antigen-combining sites. While these modified antibodies are hetero-bivalent, they behave as monovalent antibodies [45]. Another aspect of IgG4 mimics IgG rheumatoid factor (RF) activity by interacting with IgG on a solid support [46]. In contrast to conventional RF, which binds via its variable domains, the activity of IgG4 is located in its constant domains, but is inefficient in activating potentially dangerous effector systems due to its low affinity for C1q and the classical Fcy-receptors.

IgG4 seems to be associated with a pathogenic effect in a few situations. In pemphigus, recognition of skin auto-antigens (desmogleins) by IgG4 is at the origin of the disease process [47]. IgG4 Fc—Fc binding may have a pathological role within the inflammatory process, or may even induce inflammation through the aggregation of immunoglobulins, as occurs in a mouse lupus model [48]. Although some earlier reports of AIP suggested the presence of autoantibodies against the systemically distributed antigens described above, it remains unclear whether or not

IgG4-type autoantibodies have a direct role in the pathogenesis of IgG4-related diseases. To date, there have been few reports indicating IgG4 deposition in IgG4-related renal diseases [15, 29]. Therefore, in some IgG4-related diseases, the infiltration of IgG4 + plasma cells might have an association with pathological roles, similar to those in pemphigoid diseases, through IgG4 Fc–IgG Fc binding.

On the other hand, although IgG4 is associated with several clinical conditions, it is generally considered to be a benign, non-pathogenic antibody [49]. Some of these associations suggest a protective effect, such as in allergen-specific immunotherapy, tolerance induction after food avoidance [50], and protection from allergic effects during parasitosis [51, 52]. Recent data on the regulation of IgG4 showed that IgG4-related diseases may reflect an excessive production of anti-inflammatory cytokines such as interleukin (IL)-10, triggering an overwhelming expansion of IgG4-producing plasma cells. In AIP, increased peripheral inducible-memory regulatory T cells (Tregs) are positively correlated with serum levels of IgG4 [53]. In addition, prominent infiltration of Tregs upregulated IL-10 in the livers of patients with IgG4-related sclerosing cholangitis [54]. These findings suggest that IgG4 or IgG4-immune complexes do not act as a pathogenetic factor, but act as an anti-inflammatory factor in IgG4-related diseases [46]. Further studies are necessary to clarify the role of IgG4 in IgG4-related diseases.

The complement system

Patients in active stages of AIP occasionally show decreased complement (C3, C4) with elevated circulating immune complex as well as elevated serum levels of IgG4 and the IgG4 subclass of immune complexes [3, 55]. However, a recent study showed that the classical pathway of complement activation through IgG1 may be involved in the development of AIP, rather than mannose-binding lectin or alternative pathways through IgG4 [56]. Moreover, IgG4 is bound to other isotypes such as IgG1, 2, and 3 with an Fc-Fc interaction immune complex in patients with AIP [46], and thus IgG4 may contribute to the clearance of immune complexes or termination of the inflammatory process by preventing the formation of large immune complexes by blocking the Fcmediated effector functions of IgG1. Compared with findings in systemic lupus erythematosus (SLE), TIN is more often observed in the renal lesions of IgG4-related disease. But, in acute TIN associated with AIP, the deposition of immune complex (IgG and C3) was observed in the glomerular basement membrane but not in the tubular basement membrane, which suggests that membranous glomerulonephritis is also associated with severe TIN associated with IgG4-related disease [15, 29]. Recently, the deposition of C3 and IgG in the basement membrane of pancreatic ducts have been identified in both type 1 and 2 AIP [21].



Autoantibodies

Patients with IgG4-related diseases generally show several autoantibodies in addition to increased IgG and IgG4 [4, 5]. Although some patients with IgG4-related disease have non-specific antibodies such as anti-nuclear antibody (ANA), the association of IgG4-related disease and wellknown autoimmune diseases such as Sjögren's syndrome and SLE is rare. From the viewpoint of IgG4 function, the big mystery is whether IgG4-related disease is an autoimmune or an allergic disease. However, the occasional coexistence of other organ involvement leads us to the concept that there may be common target antigens in the involved organs such as the pancreas, salivary glands, biliary tract, lungs, and renal tubules. Although diseasespecific antibodies have not been identified at present, several disease-related antibodies such as anti-lactoferrin (LF) [57, 58], anti-carbonic anhydrase (CA)-II [57–60], anti-CA-IV [61], anti-pancreatic secretory trypsin inhibitor (PSTI) [62], anti-amylase-alpha [63], anti-heat-shock protein (HSP)-10 [64], and anti-plasminogen-binding protein (PBP) peptide autoantibodies [65] have been reported. Although the patients show increased serum levels of IgG4, the major subclass of these autoantibodies is not necessarily IgG4, but is often IgG1 [62]. CA-II [59], CA-IV [61], LF [58], and PSTI [62] are distributed in the ductal cells of several exocrine organs, including the pancreas, salivary glands, biliary duct, lungs, and renal tubules. Although not all peptides have been studied, immunization with CA-II or LF induced systemic lesions such as pancreatitis, sialadenitis, cholangitis, and interstitial nephritis in mouse models similar to human IgG4related diseases [66, 67]. The high prevalence of the above antibodies suggests that they may be candidates for the target antigens in AIP [58].

Molecular mimicry among microbes and target antigens may be a possible mechanism for breaking down immune tolerance. This hypothesis is based on the concept that infectious agents share one or more epitopes with selfcomponents, or infectious agents cause bystander activation of immune cells with autoaggressive potential [68–70]. Guarneri and colleagues showed significant homology between human CA-II and alpha-CA of Helicobacter pylori, a fundamental enzyme for bacterial survival and proliferation in the stomach [70]. Moreover, the homologous segments contained the binding motif of DRB1*0405 [71], which confers a risk for AIP development [70]. The PBP peptide newly identified in European patients with AIP shows homology with an amino acid sequence of PBP of H. pylori and with the ubiquitin-protein ligase E3 component n-recognin 2 (UBR2), an enzyme highly expressed in acinar cells of the pancreas, while European patients with AIP did not necessarily show LPSP as the typical histopathology of type 1 AIP in IgG4-related diseases [65]. These findings suggest that gastric *H. pylori* infection might trigger AIP in genetically predisposed subjects [68–70].

Diabetes mellitus (DM) complications exist in 43–68% of AIP patients, but autoantibodies against glutamic acid decarboxylase, beta-cell, or tyrosine phosphatase-like protein [67] associated with type 1A DM are rarely observed. These findings suggest that islet cells may not be targeted in the development of DM associated with AIP.

No disease-specific autoantibodies have been identified in IgG4-related disease. The rare association of IgG4related disease and well-known autoimmune diseases such as Sjögren's syndrome and SLE must be discussed.

Cellular immunity

Th1 and Th2 immune balance

The effector cells in IgG4-related diseases have been poorly understood. The presence of autoantibodies, the predominant infiltration of CD4+ and CD8+ T cells, and the expression of HLA-DR antigens in the pancreas [57] suggest that, as well as the infiltration of plasmacytes and B cells, an immunological mechanism may be involved in the development of AIP. CD4⁺ T cells differentiate from naïve T cells (Th0) to Th1, Th2, Th17, and Treg cells [72]. IL-12 induces Thl cells, which produce IL-2, tumor necrosis factor (TNF)-alpha, and interferon (IFN)-gamma, and mediate cellular immunity, macrophage activation, and cytotoxicity, as well as helping B-cell production of opsonizing and complement fixing antibodies [4]. IL-4 induces Th2 cells, which produce IL-4, IL-5, IL-6, and IL-10, promoting humoral and allergic responses [4]. Transforming growth factor (TGF)-beta, IL-6, IL-21, and IL-23 induce Th17 cells, which secrete IL-17, and may be involved in inflammation in mice [73].

In some patients with AIP, Th1 cells are predominant over Th2 type cells in the periphery [58, 74]. On the other hand, a Th2-type immune reaction, as well as the Th1 responses, is induced in the livers of patients with IgG4-related sclerosing cholangitis [54]. This discrepancy may be explained by the shift of Th2 cells from the periphery to local tissues, or by different disease stages. Mouse models with depletion of Tregs induced by neonatal thymectomy (nTx) support the hypothesis that Th1 cells act mainly as effectors in the initial early stage [75]. In Sjögren's syndrome [76] and PSC [77], the major infiltrating cells in the tissue are CD4⁺HLA-DR⁺Th1 cells, although CD8⁺ and B cells are also present. Similar to Sjögren's syndrome, Th1 cytokines may be essential in the induction of AIP, while Th2 cytokines may be involved in the progression of the



disease process, especially the maturation and proliferation of local B cells and plasmacytes [4].

Regulatory T cells

From naïve Th0 cells, TGF-beta can induce CD4+CD25+ Tregs, which have a potent inhibitory function, via the transcription factor Foxp3, to show CD4+ T cell-mediated immune responses such as Th1, Th2, and Th17 [73]. Foxp3 is a member of the forkhead/winged-helix family of transcriptional regulators, and functions as the master regulator in the development and function of Tregs. This suppressive function is mediated by TGF-beta and IL-10, and/or cell-tocell contact via ligation of cytotoxic T lymphocyte antigen-4 (CTLA-4). Recent studies have clarified several subtypes of Tregs [78]. Tregs originating in the thymus are naturally occurring CD4⁺CD25⁺ Tregs (nTregs), which are different from adaptive Tregs (aTregs) induced in the periphery by different antigens [78]. As Tregs expressing Foxp3 are critical in the transfer of immune tolerance, Treg deficiency has been shown to induce various autoimmune diseases in animal experimental models [73]. However, in humans, an increased prevalence of circulating CD4⁺CD25⁺ T cells or a similar level of peripheral CD4+CD25+ T cells was observed in patients with rheumatoid arthritis, Sjögren's

syndrome, and inflammatory bowel disease, compared with healthy controls [79]. Therefore, the evidence of decreased circulating Tregs as shown in the animal studies may not be a general finding in human autoimmune diseases. In IgG4related diseases, the role of Tregs remains unclear. In AIP, in addition to increased soluble CTLA4, circulatory naïve (CD45RA+) Tregs are significantly decreased in the peripheral blood of patients with AIP, whereas the major population of memory (CD45RA-)-Tregs is significantly increased [53]. In addition, prominent infiltration of Tregs with upregulation of IL-10 is observed in the livers of patients with IgG4-related sclerosing cholangitis [58]. These findings suggest that increased memory-Tregs in the periphery and local tissues may be inhibitory immune responses against inflammation in patients with AIP, although decreased naïve Tregs may be pathogenetic.

Our hypothesis for the pathogenesis of AIP as IgG4-related disease

In nTx-BALB/c mouse models immunized with CA-II or LF, CD4⁺ T cells, rather than B cells, are the predominant infiltrates in pancreatitis, sialoadenitis, and cholangitis, which is similar to human AIP [75]. These findings suggest

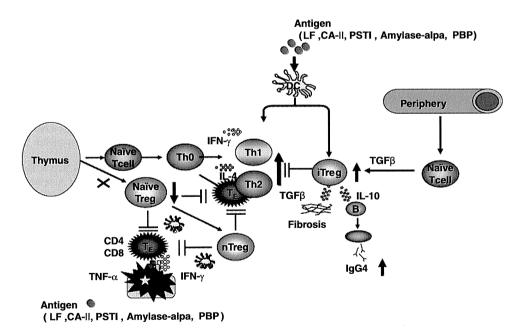


Fig. 1 Hypothesis for the pathogenesis of autoimmune pancreatitis (AIP) in IgG4-related disease. In regard to central tolerance, naïve and natural regulatory T cells (*Tregs*) derived from the thymus suppress autoreactive CD4 or CD8 cells in the normal state. In IgG4-related disease, the basic concept is a biphasic mechanism of "induction" and "progression". Initial response to self-antigens (e.g., lactoferrin [*LF*], carbonic anhydrase II [*CA-II*], CA-IV, pancreatic secretory trypsin inhibitor [*PSTI*], amylase-alpha, and plasminogenbinding protein [*PBP*] peptide of *Helicobacter pylori*) might be

induced by decreased naïve Tregs. Th2 immune responses are followed by a Th1-type immune response with the release of proinflammatory cytokines (interferon- γ [IFN- γ], interleukin [IL]-1beta, IL-2, tumor necrosis factor- α [TNF- α]). Th2-type immune responses, producing IgG, IgG4, and autoantibodies may be involved in the pathophysiology of progression. IgG4 and fibrosis may be regulated by increased IL-10 and transforming growth factor- β (TGF- β), respectively, secreted from inducible memory Tregs. DC ductal cell, iTreg inducible Treg, TE effector T cell, nTreg natural Treg



that the depletion of naïve Tregs in the periphery [80] and major histocompatibility complex (MHC)-class II restricted-autoreactive CD4+ T cells that escape from positive selection in the thymus, may play important roles in the induction of systemic organ lesions. These CD4⁺ T cells probably induce macrophage activation and further proinflammatory reactions during the early stage of AIP as direct cytotoxic effects through Fas ligand expression [81]. On the other hand, CD8⁺ T cells may play roles as effector cells in the MHC class II-deficient mouse [82] and in WBN/Kob rat models [83]. WBN/Kob rats with congenitally decreased peripheral Tregs spontaneously develop sialadenitis, thyroiditis, sclerotic cholangitis, and TIN. Although the target antigens remain unclear, CD8⁺ cells also seem to be effectors. Although rodents lack the IgG4 subclass, deposits of tissue-specific IgG2b, in electrophoretic position similar to human IgG4, were observed in the injured pancreas and lachrymal glands in WBN/Kob rats [83]. These animal models suggest that although CD8⁺ T cells may be partially involved, CD4+ T cells play major roles in the development of experimental systemic lesions, which are similar to the lesions in human IgG4-related diseases [4, 58], although the counterpart of IgG4 in mouse IgG subclasses has not been identified. As TGF-beta is an important regulatory factor in maintaining immune homeostasis [84], TGF-beta-dominant negative mutant mice suggest that the loss of TGF-beta signaling may contribute to AIP [85].

From the above findings, we propose a hypothesis for the pathogenesis of AIP (Fig. 1). The basic concept is a biphasic mechanism of "induction" and "progression." An initial response to self-antigens (e.g., LF, CA-II, CA-IV, PSTI, amylase-alpha, and PBP peptide of *H. pylori*) might be induced by decreased naïve Tregs, followed by a Th1-type immune response with the release of proinflammatory cytokines (IFN-gamma, IL-1-beta, IL-2, TNF-alpha). Then Th2-type immune responses producing IgG, IgG4, and autoantibodies may be involved in the pathophysiology of progression. IgG4 and fibrosis may be regulated by increased IL-10 and TGF-beta, respectively, secreted from inducible memoryTregs. The classical pathway of the complement system may be activated by the IgG1 immune complex.

Conclusion

In conclusion, recent advances support the concept of IgG4-related disease, a unique clinical entity, as a systemic disease. As Tregs seem to play important roles in the progression as well as the induction of the disease, further studies are necessary to clarify the pathogenesis, including studies of genetic backgrounds, disease-specific antigens, and the role of IgG4.

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References

- Sarles H, Sarles JC, Muratore R, Guien C. Chronic inflammatory sclerosis of the pancreas—an autonomous pancreatic disease? Am J Dig Dis. 1961;6:688–98.
- 2. Yoshida K, Toki F, Takeuchi T, Watanabe S, Shiratori K, Hayashi N. Chronic pancreatitis caused by an autoimmune abnormality. Proposal of the concept of autoimmune pancreatitis. Dig Dis Sci. 1995;40:1561–8.
- 3. Hamano H, Kawa S, Horiuchi A, Unno H, Furuya N, Akamatsu T, et al. High serum IgG4 concentrations in patients with sclerosing pancreatitis. New Engl J Med. 2001;344:732–8.
- Okazaki K, Uchida K, Chiba T. Recent concept of autoimmunerelated pancreatitis. J Gastroenterol. 2001;36:293–302.
- Okazaki K, Kawa S, Kamisawa T, Naruse S, Tanaka S, Nishimori I, et al. Clinical diagnostic criteria of autoimmune pancreatitis: revised proposal. J Gastroenterol. 2006;41:626–31.
- Pickartz T, Mayerle J, Lerch M. Autoimmune pancreatitis. Nat Clin Pract Gastroenterol Hepatol. 2007;4:314–23.
- 7. Kawaguchi K, Koike M, Tsuruta K, Okamoto A, Tabata I, Fujita N. Lymphoplasmacytic sclerosing pancreatitis with cholangitis: a variant of primary sclerosing cholangitis extensively involving pancreas. Hum Pathol. 1991;22:387–95.
- Kamisawa T, Funata N, Hayashi Y, Eishi Y, Koike M, Tsuruta K, et al. A new clinicopathological entity of IgG4-related autoimmune disease. J Gastroenterol. 2003;38:982

 –4.
- Comings DE, Skubi KB, Van Eyes J, Motulsky AG. Familial multifocal fibrosclerosis. Ann Intern Med. 1967;66:884

 –92.
- Chari ST, Kloeppel G, Zhang L, Notohara K, Lerch MM, Shimosegawa T, et al. Histopathologic and clinical subtypes of autoimmune pancreatitis: the Honolulu consensus document. Pancreas. 2010;39:549–54.
- Klöppel G, Detlefsen S, Chari ST, Longnecker DS, Zamboni G. Autoimmune pancreatitis: the clinicopathological characteristics of the subtype with granulocytic epithelial lesions. J Gastroenterol. 2010;45:787–93.
- Notohara K, Burgart LJ, Yadav D, Chari S, Smyrk TC. Idiopathic chronic pancreatitis with periductal lymphoplasmacytic infiltration: clinicopathologic features of 35 cases. Am J Surg Pathol. 2003;27:1119–27.
- 13. Zamboni G, Luttges J, Capelli P, Frulloni L, Cavallini G, Pederzoli P, et al. Histopathological features of diagnostic and clinical relevance in autoimmune pancreatitis: a study on 53 resection specimens and 9 biopsy specimens. Virchows Arch. 2004;445:552–63.
- Mikulicz J. Über eine eigenartige symmetrishe Erkrankung der Tränen und Mundspeicheldrüsen. Stuttgart: Beitr z Chir Festschr f Theodor Billroth; 1892. p. 610–30.
- Saeki T, Nishi S, Imai N, Ito T, Yamazaki H, Kawano M, et al. Clinicopathological characteristics of patients with IgG4-related tubulointerstitial nephritis. Kidney Int. 2010;78:1016–23.
- Li Y, Nishihara E, Hirokawa M, Taniguchi E, Miyauchi A, Kakudo K. Distinct clinical, serological, and sonographic characteristics of Hashimoto's thyroiditis based with and without IgG4-positive plasma cells. J Clin Endocrinol Metab. 2010;95: 1309–17.
- Kamisawa T, Okamoto A. Autoimmune pancreatitis: proposal of IgG4-related sclerosing disease. J Gastroenterol. 2006;41: 613–25.



- Yamamoto M, Takahashi H, Ohara M, Suzuki C, Naishiro Y, Yamamoto H, Shinomura Y, Imai K. A new conceptualization for Mikulicz's disease as an IgG4-related plasmacytic disease. Mod Rheumatol. 2006;16:335–40.
- Masaki Y, Dong L, Kurose N, Kitagawa K, Morikawa Y, Yamamoto M, et al. Proposal for a new clinical entity, IgG4positive multi-organ lymphoproliferative syndrome: analysis of 64 cases of IgG4-related disorders. Ann Rheum Dis. 2009;68:1310–5.
- 20. Mackay IR. Autoimmune disease. Med J Aust. 1969;1:696-9.
- Detlefsen S, Bräsen JH, Zamboni G, Capelli P, Klöppel G. The position of complement C3c, immunoglobulin (Ig)G4 and IgG at the basement membrane of pancreatic ducts and acini in autoimmune pancreatitis. Histopathology. 2010;57:825–35.
- Shimosegawa T, Chari ST, Frulloni L, Kamisawa T, Kawa S, Mino-Kenudson M, et al. International consensus diagnostic criteria for autoimmune pancreatitis: guidelines of the International Association Of Pancreatology. Pancreas (in press).
- Kamisawa T, Funata N, Hayashi Y, Tsuruta K, Okamoto A, Amemiya K, et al. Close relationship between autoimmune pancreatitis and multifocal fibrosclerosis. Gut. 2003;52:683–7.
- Saegusa H, Momose M, Kawa S, Hamano H, Ochi Y, Takayama M, et al. Hilar and pancreatic gallium-67 accumulation is characteristic feature of autoimmune pancreatitis. Pancreas. 2003:27:20-5.
- Erkelens GW, Vleggaar FP, Lesterhuis W, van Buuren HR, van der Werf SD. Sclerosing pancreato-cholangitis responsive to steroid therapy. Lancet. 1999;354:43–4.
- Nakazawa T, Ohara H, Yamada T, Ando H, Sano H, Kajino S, et al. Atypical primary sclerosing cholangitis cases associated with unusual pancreatitis. Hepatogastroenterology. 2001;48: 625-30.
- Hamano H, Kawa S, Ochi Y, Unno H, Shiba N, Wajiki M, et al. Hydronephrosis associated with retroperitoneal fibrosis and sclerosing pancreatitis. Lancet. 2002;359:1403

 –4.
- Takeda S, Haratake J, Kasai T, Takaeda C, Takazakura E. IgG4associated idiopathic tubulointerstitial nephritis complicating autoimmune pancreatitis. Nephrol Dial Transplant. 2004;19: 474-6
- Uchiyama-Tanaka Y, Mori Y, Kimura T, Watanabe S, Nozaki Y, Fujita K, et al. Acute tubulointerstitial nephritis associated with autoimmune-related pancreatitis. Am J Kidney Dis. 2004; 43:e18-25.
- Shimatsu A, Oki Y, Fujisawa I, Sano T. Pituitary and stalk lesions (infundibulo-hypophysitis) associated with immunoglobulin G4-related systemic disease: an emerging clinical entity. Endocr J. 2009;56:1033–41.
- Komatsu K, Hamano H, Ochi Y, Takayama M, Muraki T, Yoshizawa K, et al. High prevalence of hypothyroidism in patients with autoimmune pancreatitis. Dig Dis Sci. 2005;50: 1052-7.
- 32. Yoshimura Y, Takeda S, Ieki Y, Takazakura E, Koizumi H, Takagawa K. IgG4-associated prostatitis complicating autoimmune pancreatitis. Intern Med. 2006;45:897–901.
- 33. Okazaki K, Uchida K, Matsushita M, Takaoka M. How to diagnose autoimmune pancreatitis by the revised Japanese clinical criteria. J Gastroenterol. 2007;42(Suppl 18):32–8.
- 34. Ohara H, Nakazawa T, Sano H, Ando T, Okamoto T, Takada H, et al. Systemic extrapancreatic lesions associated with autoimmune pancreatitis. Pancreas. 2005;31:232–7.
- Hamano H, Arakura N, Muraki T, Ozaki Y, Kiyosawa K, Kawa S. Prevalence and distribution of extrapancreatic lesions complicating autoimmune pancreatitis. J Gastroenterol. 2006;41: 1197–205.
- 36. Okazaki K, Kawa S, Kamisawa T, Ito T, Inui K, Irie H, et al. Japanese clinical guidelines for autoimmune pancreatitis. Pancreas. 2009;38:849–66.

- Okazaki K, Kawa S, Kamisawa T, Shimosegawa T, Tanaka M, et al. Japanese consensus guidelines for management of autoimmune pancreatitis: I. Concept and diagnosis of autoimmune pancreatitis. J Gastroenterol. 2010;45:249–65.
- Kawa S, Okazaki K, Kamisawa T, Shimosegawa T, Tanaka M, et al. Japanese consensus guidelines for management of autoimmune pancreatitis: II. Extrapancreatic lesions, differential diagnosis. J Gastroenterol. 2010;45:355-69.
- 39. Kamisawa T, Okazaki K, Kawa S, et al. Japanese consensus guidelines for management of autoimmune pancreatitis: III. Treatment and prognosis of AIP. J Gastroenterol. 2010;45: 471-7
- 40. Okazaki K, Uchida K, Research Committee Members. Proposal of the concept and diagnostic criteria of IgG4-related disease. Annual reports of Research Committee Of Intractable Diseases supported by Ministry of Health, Labour and Welfare of Japan. 2009. p. 25–30 (in Japanese).
- 41. Masaki Y, Sugai S, Umehara H. IgG4-related diseases including Mikulicz's disease and sclerosing pancreatitis: diagnostic insights. J Rheumatol. 2010;37:1380–5.
- 42. Roitt I. Antibodies. In: Roitt I, editor. Roitt's essential immunology. 9th ed. London: Blackwell Science; 1997. p. 43–62.
- Taguchi M, Kihara Y, Nagashio Y, Yamamoto M, Otsuki M, Harada M. Decreased production of immunoglobulin M and A in autoimmune pancreatitis. J Gastroenterol. 2009;44:1133–9.
- Robinson DS, Larche M, Durham SR. Tregs and allergic disease.
 J Clin Invest. 2004;114:1389–97.
- 45. van der Neut Kolfschoten M, Schuurman J, Losen M, Bleeker WK, Martínez-Martínez P, Vermeulen E, et al. Anti-inflammatory activity of human IgG4 antibodies by dynamic Fab arm exchange. Science. 2007;317:1554–7.
- Kawa S, Kitahara K, Hamano H, Ozaki Y, Arakura N, Yoshizawa K, et al. A novel immunoglobulin-immunoglobulin interaction in autoimmunity. PLoS One. 2008;3:e1637.
- Stanley JR, Amagai M. Pemphigus, bullous impetigo, and the staphylococcal scalded-skin syndrome. N Engl J Med. 2006;355: 1800–10.
- 48. Fujii H, Nakatani K, Arita N, Ito MR, Terada M, Miyazaki T, et al. Internalization of antibodies by endothelial cells via fibronectin implicating a novel mechanism in lupus nephritis. Kidney Int. 2003;64:1662–70.
- 49. Aalberse RC, Stapel SO, Schuurman J, Rispens T. Immuno-globulin G4: an odd antibody. Clin Exp Allergy. 2009;39:469–77.
- Ruiter B, Knol EF, van Neerven RJ, Garssen J, Bruijnzeel-Koomen CA, Knulst AC, et al. Maintenance of tolerance to cow's milk in atopic individuals is characterized by high levels of specific immunoglobulin G4. Clin Exp Allergy. 2007;37: 1103–10.
- Hussain R, Poindexter RW, Ottesen EA. Control of allergic reactivity in human filariasis. Predominant localization of blocking antibody to the IgG4 subclass. J Immunol. 1992;148: 2731–7.
- Yazdanbakhsh M, van den Biggelaar A, Maizels RM. Th2 responses without atopy: immunoregulation in chronic helminth infections and reduced allergic disease. Trends Immunol. 2001;22:372-7.
- 53. Miyoshi H, Uchida K, Taniguchi T, Yazumi S, Matsushita M, Takaoka M, et al. Circulating naive and CD4+CD25high regulatory T cells in patients with autoimmune pancreatitis. Pancreas. 2008;36:133–40.
- 54. Zen Y, Fujii T, Harada K, Kawano M, Yamada K, Takahira M, et al. Th2 and regulatory immune reactions are increased in immunoglobin G4-related sclerosing pancreatitis and cholangitis. Hepatology. 2007;45:1538–46.
- 55. Cornell LD, Chicano SL, Deshpande V, Collins AB, Selig MK, Lauwers GY, et al. Pseudotumors due to IgG4 immune-complex



- tubulointerstitial nephritis associated with autoimmune pancreatocentric disease. Am J Surg Pathol. 2007;31:1586–97.
- Muraki T, Hamano H, Ochi Y, Komatsu K, Komiyama Y, Arakura N, et al. Autoimmune pancreatitis and complement activation system. Pancreas. 2006;32:16–21.
- Uchida K, Okazaki K, Konishi Y, Ohana M, Takakuwa H, Hajiro K, et al. Clinical analysis of autoimmune-related pancreatitis. Am J Gastroenterol. 2000;95:2788–94.
- Okazaki K, Uchida K, Ohana M, Nakase H, Uose S, Inai M, et al. Autoimmune-related pancreatitis is associated with autoantibodies and a Th1/Th2-type cellular immune response. Gastroenterology. 2000;118:573–81.
- Nishi H, Tojo A, Onozato ML, Jimbo R, Nangaku M, Uozaki H, et al. Anti-carbonic anhydrase II antibody in autoimmune pancreatitis and tubulointerstitial nephritis. Nephrol Dial Transplant. 2007;22:1273–5.
- 60. Aparisi L, Farre A, Gomez-Cambronero L, Martinez J, De Las Heras G, Corts J, et al. Antibodies to carbonic anhydrase and IgG4 levels in idiopathic chronic pancreatitis: relevance for diagnosis of autoimmune pancreatitis. Gut. 2005;54:703–9.
- Nishimori I, Miyaji E, Morimoto K, Nagao K, Kamada M, Onishi S. Serum antibodies to carbonic anhydrase IV in patients with autoimmune pancreatitis. Gut. 2005;54:274–81.
- 62. Asada M, Nishio A, Uchida K, Kido M, Ueno S, Uza N, et al. Identification of a novel autoantibody against pancreatic secretory trypsin inhibitor in patients with autoimmune pancreatitis. Pancreas. 2006;33:20–6.
- Endo T, Takizawa S, Tanaka S, Takahashi M, Fujii H, Kamisawa T, et al. Amylase alpha-2A autoantibodies: novel marker of autoimmune pancreatitis and fulminant type 1 diabetes. Diabetes. 2009;58:732–7.
- 64. Takizawa S, Endo T, Wanjia X, Tanaka S, Takahashi M, Ko-bayashi T. HSP 10 is a new autoantigen in both autoimmune pancreatitis and fulminant type 1 diabetes. Biochem Biophys Res Commun. 2009;386:192–6.
- Frulloni L, Lunardi C, Simone R, Dolcino M, Scattolini C, Falconi M, et al. Identification of a novel antibody associated with autoimmune pancreatitis. N Engl J Med. 2009;361:2135–42.
- Nishimori I, Bratanova T, Toshkov I, Caffrey T, Mogaki M, Shibata Y, et al. Induction of experimental autoimmune sialoadenitis by immunization of PL/J mice with carbonic anhydrase II. J Immunol. 1995;154:4865–73.
- Ueno Y, Ishii M, Takahashi S, Igarashi T, Toyota T, LaRusso NF. Different susceptibility of mice to immune-mediated cholangitis induced by immunization with carbonic anhydrase II. Lab Invest. 1998;78:629–37.
- Kountouras J, Zavos C, Gavalas E, Tzilves D. Challenge in the pathogenesis of autoimmune pancreatitis: potential role of *Heli-cobacter pylori* infection via molecular mimicry. Gastroenterology. 2007;133:368–9.
- Kountouras J, Zavos C, Chatzopoulos D. A concept on the role of Helicobacter pylori infection in autoimmune pancreatitis. J Cell Mol Med. 2005;9:196–207.
- Guarneri F, Guarneri C, Benvenga S. Helicobacter pylori and autoimmune pancreatitis: role of carbonic anhydrase via molecular mimicry? J Cell Mol Med. 2005;9:741–4.

- Kawa S, Ota M, Yoshizawa K, Horiuchi A, Hamano H, Ochi Y, et al. HLA DRB10405-DQB10401 haplotype is associated with autoimmune pancreatitis in the Japanese population. Gastroenterology. 2002;122:1264–9.
- 72. McGeachy MJ, Cua DJ. The link between IL-23 and Th17 cell-mediated immune pathologies. Semin Immunol. 2007;19:372–6.
- 73. Oukka M. Interplay between pathogenic Th17 and regulatory T cells. Ann Rheum Dis. 2007;66(Suppl 3):iii87–90.
- 74. Yamamoto M, Harada S, Ohara M, Suzuki C, Naishiro Y, Yamamoto H, et al. Clinical and pathological differences between Mikulicz's disease and Sjogren's syndrome. Rheumatology (Oxford). 2005;44:227–34.
- 75. Uchida K, Okazaki K, Nishi T, Uose S, Nakase H, Ohana M, et al. Experimental immune-mediated pancreatitis in neonatally thymectomized mice immunized with carbonic anhydrase II and lactoferrin. Lab Invest. 2002;82:411–24.
- Ajjan RA, McIntosh RS, Waterman EA, Watson PF, Franklin CD, Yeoman CM, et al. Analysis of the T-cell receptor Valpha repertoire and cytokine gene expression in Sjogren's syndrome. Br J Rheumatol. 1998;37:179–85.
- 77. Dienes HP, Lohse AW, Gerken G, Schirmacher P, Gallati H, Löhr HF, et al. Bile duct epithelia as target cells in primary biliary cirrhosis and primary sclerosing cholangitis. Virchows Arch. 1997;431:119–24.
- Valencia X, Lipsky PE. CD4+CD25+FoxP3+ regulatory T cells in autoimmune diseases. Nat Clin Pract Rheumatol. 2007;3:619–26.
- 79. Gottenberg JE, Lavie F, Abbed K, Gasnault J, Le Nevot E, Delfraissy JF, et al. CD4 CD25high regulatory T cells are not impaired in patients with primary Sjogren's syndrome. J Autoimmun. 2005;24:235–42.
- 80. Sakaguchi S, Fukuma K, Kuribayashi K, Masuda T. Organ-specific autoimmune diseases induced in mice by elimination of T cell subset. I. Evidence for the active participation of T cells in natural self-tolerance; deficit of a T cell subset as a possible cause of autoimmune disease. J Exp Med. 1985;161:72–87.
- 81. Demols A, Le Moine O, Desalle F, Quertinmont E, Van Laethem JL, Deviere J. CD4(+)T cells play an important role in acute experimental pancreatitis in mice. Gastroenterology. 2000;118: 582–90.
- Vallance BA, Hewlett BR, Snider DP, Collins SM. T cell-mediated exocrine pancreatic damage in major histocompatibility complex class II-deficient mice. Gastroenterology. 1998;115: 978–87.
- 83. Sakaguchi Y, Inaba M, Tsuda M, Quan GK, Omae M, Ando Y, et al. The Wistar Bonn Kobori rat, a unique animal model for autoimmune pancreatitis with extrapancreatic exocrinopathy. Clin Exp Immunol. 2008;152:1–12.
- 84. Marth T, Strober W, Kelsall BL. High dose oral tolerance in ovalbumin TCR-transgenic mice: systemic neutralization of IL-12 augments TGF-beta secretion and T cell apoptosis. J Immunol. 1996;157:2348–57.
- Hahm KB, Im YH, Lee C, Parks WT, Bang YJ, Green JE, et al. Loss of TGF-beta signaling contributes to autoimmune pancreatitis. J Clin Invest. 2000;105:1057-65.



Involvement of Inducible Costimulator— and Interleukin 10-Positive Regulatory T Cells in the Development of IgG4-Related Autoimmune Pancreatitis

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Objectives: Immunoglobulin G4 (IgG4)-related autoimmune pancreatitis (AIP) is a new clinical entity of pancreatic disorder. There are immunologic and histological abnormalities, including increased serum IgG4 levels and the infiltration of IgG4-positive plasmacytes. However, the role of IgG4 is unclear. Recently, regulatory T cells (Tregs) were reported to contribute to the development of various autoimmune diseases as well as in B-cell shifting to IgG4-producing plasmacytes. We studied Tregs in the pancreas and peripheral blood.

Methods: We recruited 44 patients with IgG4-related AIP. For comparison, we recruited 37 patients with other pancreatic diseases and 27 healthy subjects as controls. We studied infiltrating cells in the pancreas by immunohistochemistry and analyzed inducible costimulator-positive Tregs and interleukin 10-positive Tregs in the peripheral blood by flow

Results: The ratio of Foxp3-positive cells to infiltrated mononuclear cells (Foxp3/Mono) in AIP patients was significantly higher than in patients with alcoholic chronic pancreatitis. In AIP, Foxp3/Mono and IgG4/Mono were positively correlated. Inducible costimulator-positive Tregs were significantly higher in AIP patients than in the patients with other pancreatic diseases and the healthy control group. Interleukin 10-positive Tregs were significantly higher in AIP patients than in the healthy control group. Conclusions: Increased quantities of inducible costimulator-positive

Key Words: IgG4-related disease, autoimmune pancreatitis (AIP), regulatory T cells (Tregs), IgG4, IL-10, inducible costimulator (ICOS)

Tregs may influence IgG4 production in IgG4-related AIP.

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n 1961, Sarles et al observed the first case of idiopathic chronic pancreatitis (CP) with hypergammaglobulinemia, in which an autoimmune mechanism was supposedly involved. In 1991, Kawaguchi et al² reported 2 cases of an unusual lymphoplasmacytic sclerosing inflammatory disease involving the total pancreas, common bile duct, gallbladder, and, in 1 patient, the lip. In addition, 2 patients presented mass-like enlargement of the pancreatic head. Histopathologic characteristics included diffuse lymphoplasmacytic infiltration, marked interstitial fibrosis, acinar atrophy, and obliterative phlebitis of the pancreatic and portal veins, which was termed as lymphoplasmacytic sclerosing pancreatitis (LPSP). In 1995, Yoshida et al³ first proposed the concept of "autoimmune pancreatitis (AIP)," in which patients showed a diffusely enlarged pancreas, a narrowing pancreatogram, increased serum immunoglobulin G (IgG), the presence of autoantibodies, fibrotic changes with lymphocytic infiltration, and steroidal efficacy. In 2001, Hamano et al4 reported that elevated serum IgG4 levels were highly specific and sensitive for the diagnosis of AIP. In 2003, Kamisawa et al⁵ suggested that AIP is a systemic disease, based on the findings that the pancreas and other involved organs have abundant infiltration of IgG4-positive plasma cells. Thereafter, many AIP cases have been reported by Japanese investigators, and AIP has been accepted as a new clinical entity.6-11

On the other hand, reports from Europe¹² and the United States¹³ described unique histological patterns in the resected pancreata of patients with mass-forming, chronic, nonalcoholic pancreatitis with epithelial destruction by granulocytes, which is now supposed to be distinguishable from IgG4-related AIP (or type 1 AIP) and called idiopathic duct centric pancreatitis (IDCP) or AIP with granulocyte epithelial lesions (AIP with GELs) or type 2 AIP. Most of the Japanese AIP cases are LPSP, whereas those concerning IDCP are very few. Although we recently reported the first case of IDCP in Japan with full radiological and histopathologic findings, 15 it still remains unclear whether the clinical manifestations of the Japanese patients with IDCP are similar to those of Western countries. Therefore, Japanese consensus clinical guidelines have focused on IgG4-related AIP (LPSP).⁹⁻¹¹ An overlap in the histological features of the 2 patterns may exist in some patients. Although the pathogenesis is still unclear, the most important issue in managing AIP is to differentiate it from pancreas and biliary malignancy.

Great attention has been focused on the relation between various autoimmune diseases and regulatory T cells (Tregs), which are present in human peripheral blood, ¹⁶⁻²¹ intestinal lamina propria, ²² and the thymus. ^{20,21} Recent reports have shown that Tregs can be classified into 2 groups: (1) naturally occurring Tregs expressing CD4⁺CD25^{high} and (2) naive Tregs expressing the naive T-cell marker CD45RA in addition to CD4 and CD25. Naive Tregs also expressed very high levels of mRNA for Foxp3 and manifested equivalent suppressive activity in vitro. 23,24 We have previously reported that increased quantities of CD4⁺ CD25^{high} Tregs may influence IgG4 production, and naive Tregs may be involved in the development of AIP.25 Patients with IgG4related AIP often show high serum levels of IgG4, but the pathologic significance and mechanism are unknown. Immunoglobulin 4 is well known as a T-helper 2 (T_H2)-dependent isotype. Interleukin (IL) 4 directs naive human B cells to switch to IgG4 and IgE production. Interleukin 10 promotes isotype switching from IgM to IgG1, IgG3, IgG4, and/or IgE. Interleukin 10 also stimulates IgG4 production. The association with the IL-10 response still holds and has strengthened, with the

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TABLE 1. Profiles of Subjects in the Present Study

	n	Sex, n (Female/Male)	Age, Mean (SD), y
Healthy control	27	10/17	61 (13)
Alcoholic CP	20	0/20	59 (11)
Idiopathic CP	17	10/7	55 (15)
AIP	44	14/30	65 (9)
Without steroid	21	9/12	64 (14)
With steroid	23	5/18	67 (8)

development of IL-10-producing Tregs in vivo being found to be inducible costimulator (ICOS) dependent.^{28,29} Of note, a recent report revealed the existence of 2 functional subsets of human Foxp3⁺ Tregs, including ICOS⁺ cells, which produced high amounts of IL-10.³⁰ However, the relationship between Tregs in the peripheral blood and the pancreas remains unclear. In this study, we attempt to clarify that relationship by comparing Tregs in the peripheral blood and the pancreatic tissues.

MATERIALS AND METHODS

Subjects

We examined 44 patients with AIP at Kansai Medical University and its affiliated hospitals (21 untreated patients and 23 patients treated with corticosteroids; 14 women and 30 men; mean age, 65 years; range, 42-81 years). There were 20 patients with alcoholic pancreatitis (0 woman, 20 men; mean age, 59 years; range, 38-75 years), 17 patients with idiopathic pancreatitis (10 women, 7 men; mean age, 55 years; range, 20-72 years), and 27 healthy volunteers (10 women, 17 men; mean age, 61 years; range, 33-82 years), who served as control subjects. Nine AIP cases (6 women and 3 men; mean age, 54 years; range, 56-73 years) and 9 alcoholic pancreatitis cases (9 men; mean age, 53 years; range, 39-75 years) were surgery cases in that. All cases were collected between 1992 and 2009 (Table 1). All the AIP patients were diagnosed according to the clinical diagnostic criteria for AIP proposed by the Research Committee of Intractable Diseases of the Pancreas supported by the Japanese Ministry of Health, Labor, and Welfare, ³¹ and the Asian criteria. ³² Thirty-five AIP patients fulfilled Japanese diagnostic criteria 1 and 2. In brief, they exhibited diffuse enlargement of the pancreas and diffuse narrowing of the main pancreatic duct with an irregular wall. They also showed high levels of serum γ -globulin (>2 g/dL), IgG (>1800 mg/dL), and IgG4 (>135 mg/dL). Because 9 cases showed LPSP with a pancreatectomy specimen, AIP was diagnosed. The diagnosis of CP was made by imaging studies (ultrasound, computed tomography, endoscopic retrograde cholangiopancreatography, and magnetic resonance cholangiopancreatography), a pancreatic function test (secretin stimulation test), or histopathologic examination according to the clinical diagnosis criteria of the Japan Pancreatic Society.³³ Alcoholic CP was considered to be alcoholic in origin if the patient had a daily alcohol intake of more than 80 g/d for more than 10 years. Patients had a diagnosis of idiopathic CP when no apparent causes such as alcoholism, gallstones, or autoimmunity could be identified. This study was approved by the Kansai Medical University's ethics committee, and all patients and healthy volunteers gave informed consents.

Histopathology and Immunohistochemistry

Formalin-fixed and paraffin-embedded specimens were prepared and used for histopathologic and immunohistochemical studies. Sections measuring 4 µm were cut from each paraffin block and stained with hematoxylin and eosin, periodic acid-Schiff after diastase digestion, Azan-Mallory, reticulin, or orcein. The remaining material was used for immunohistochemical analysis. Immunoglobulin G4 immunostaining was performed using a monoclonal antibody for human IgG4 (ZYMED Laboratories, San Francisco, Calif). Immunoglobulin 1 and Foxp3 immunostaining was performed by avidin-biotin complex method with reagents provided by Vector Laboratories (Burlingame, Calif). The antibodies used to identify the inflammatory cells in the pancreas were IgG1 antibodies (Binding Site, Birmingham, UK) and Foxp3 (eBioscience, San Diego, Calif). The deparaffinized sections were pretreated in an EDTA buffer (pH 8.0) in a pressure cooker at 100°C for 5 minutes. After incubation with the first antibody at 4°C overnight, biotinylated rabbit anti-sheep serum IgG (Vector) was used as the secondary antibody (sections for IgG1), and immunoreactive deposits were visualized with 3,3diaminobenzidine tetrahydrochloride. Cells positive for IgG1, IgG4, and Foxp3 were counted under 5 different high-power fields with intense inflammation. The ratios between IgG1, IgG4, Foxp3-positive cells and infiltrated mononuclear cells were calculated in each case by 2 pathologists.³⁴

Flow Cytometric Analysis of Tregs in Peripheral Blood Cells

Flow cytometric analysis was performed according to the method described by Okazaki et al. ³⁵ Briefly, cells were stained with fluorescein isothiocyanate-conjugated anti-CD4 (BD Bioscience, Franklin Lakes, NJ), PE-Cy7-conjugated anti-ICOS (CD278) (eBioscience), and allophycocyanin-conjugated anti-CD25 (BD Bioscience), for Tregs at 4°C for 30 minutes. After staining, the cells were fixed in 1% paraformaldehyde, and 3 color flow cytometric analyses were performed using FACS-LSR (BD Biosciences) and FlowJo (FlowJo LLC, Ashland, Ore).

Flow Cytometric Analysis of Intracellular IL-10

Intracellular staining of IL-10 was performed according to the manufacturer's instruction. In brief, cells magnetically labeled with CD4 and CD25 microbeads (CD4⁺CD25^{high} Treg Isolation Kit; Miltenyi Biotec, Bergisch Gladbach, Germany) were loaded onto auto MACS (Miltenyi Biotec). The cells were then washed with phosphate-buffered saline before analysis. For intracellular staining, CD4⁺CD25^{high} Tregs were stimulated with phorbol 12-myristate 13-acetate, ionomycin for 2 to 4 hours before adding Golgi stop/Golgi plug solution (BD Biosciences). The cells were then stained with fluorescein isothiocyanate-conjugated anti-CD4, allophycocyanin-conjugated anti-CD25, and PE-Cy7-conjugated anti-ICOS (CD278) (eBioscience). After fixation with a fixation buffer (eBioscience) for 20 minutes at 4°C, the cells were permeabilized with a permeabilization buffer (eBioscience) and counterstained with PE-conjugated anti-IL-10 for 30 minutes. Stained cells were analyzed using FACS-LSR (BD Biosciences) and FlowJo (FlowJo LLC).

Statistical Analysis

For all studies, data are expressed as mean (SD); differences were analyzed using the nonparametric Mann-Whitney rank test and Fisher exact test, where P < 0.05 was considered significant.

RESULTS

Immunohistochemical Findings of IgG1 and IgG4

The ratio of IgG4-positive plasma cells to infiltrated mononuclear cells (IgG4/Mono) was significantly higher in AIP (n = 9, 0.122 [SD, 0.049]) than in alcoholic CP (n = 9, 0.013 [SD, 0.007]; P < 0.05; Figs. 1–3). The ratio of IgG1-positive plasma cells to infiltrated mononuclear cells (IgG1/Mono) was significantly lower in AIP (n = 9, 0.041 [SD, 0.024]) than in

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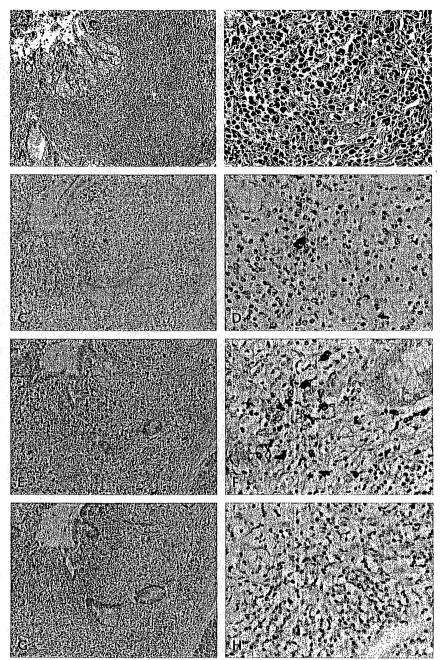


FIGURE 1. Histopathologic and immunohistochemical findings of the pancreas in LPSP. Hematoxylin-eosin staining (original magnification $\times 100$ [A], $\times 400$ [B]). Lymphoplasmacytic sclerosing pancreatitis showed interlobular fibrosis, atrophic pancreatic lobules, and infiltration of lymphoplasmacytes. Immunoglobulin 1–positive cells (original magnification $\times 100$ [C], $\times 400$ [D]) were scattered, whereas lgG4-positive (original magnification $\times 100$ [E], $\times 400$ [F]) and Foxp3-positive cells (original magnification $\times 100$ [G], $\times 400$ [H]) were abundant.

alcoholic CP (n = 9, 0.085 [SD, 0.011]; P < 0.05; Figs. 1–3). The ratio of IgG4/Mono to IgG1/Mono (IgG4/G1) was significantly higher in AIP (n = 9, 2.92 [SD, 1.42]) than in alcoholic CP (n = 9, 0.16 [SD, 0.08]; P < 0.05; Figs. 1–3A–C).

Immunohistochemical Findings of Foxp3

The ratio of Foxp3-positive cells to infiltrated mononuclear cells (Foxp3/Mono) in patients with AIP (n = 9, 0.091)

[SD, 0.023]) was significantly higher than alcoholic CP (n = 9, 0.012 [SD, 0.003]; P < 0.05; Figs. 1–3D).

Correlation Between Foxp3/Mono and IgG4/Mono in Patients With AIP

In the group of patients with AIP, Foxp3/Mono and $\lg G4/M$ Mono were positively correlated (n = 9, P < 0.05; R = 0.91) (Fig. 4).

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