

receptors in a clinical situation, because these substances should be not only effective but also well-tolerated and safe for long-term treatment. From these standpoints, a strategy using common food materials, such as lemon grass, instead of antibodies or synthetic substances appears to be promising. As lemon grass has been used in Asian foods for many years, its safety for daily intake appears to be guaranteed. Citral itself is used as a spice in many foods and its safety has also been proven by the investigation of National Toxicology Program of USA [15]. Moreover, citral has the advantage of suppressing both $\beta 7$ -integrin and CCR9 expression on lymphocytes in an RALDH2-specific manner. SAMP1/Yit mice we used in this study are very nice model for CD. Chronic sustained inflammation is not reproducible in a chemically induced model. However, for treatment study, treatment of mice for a long time is needed. Citral itself is vulnerable to oxidation, thus blending citral into diet chow will result in a decrease in the concentration of the active form of citral. Dissolving citral in drinking water is difficult due to its lipophilicity. Intraperitoneal injection of citral for a long period has an ethical problem and also may influence disease activity by stress. On the other hand, lemon grass tea drinking is a common habit for Asian people and is good way to ingest citral stably.

One concern we have is that lemon grass might reduce lymphocyte recruitment to levels lower than expected. Actually, vitamin A deficiency has been shown to cause attenuation of T cell recruitment to intestinal mucosa in a physiological condition [9], and vitamin A deficiency can cause many diseases, such as infectious colitis, respiratory infections and night blindness, as well as decrease in growth rate and slow bone development [26,27]. However, treatments for two weeks did not show any significant difference in weight gain or general appearance between mice with lemon grass treatment and control mice. Moreover, in our preliminary study, we also confirmed that even long-term treatment with lemon grass (35 weeks) did not produce any significant changes in body weight, small intestinal weight and length, sizes of Peyer's patches and MLNs in AKR/J mice (data not shown), suggesting that lemon grass did not have any toxic effect at the concentration used.

We speculated that the anti-inflammatory effect of lemon grass on ileitis is due mainly to inhibition of aberrant lymphocyte homing through attenuation

of up-regulation of gut-homing molecules on the surface of lymphocytes, as we did not observe a direct effect of lemon grass on RALDH mRNA expression in DCs or MAdCAM-1 expression in the microvascular endothelium in this study. However, we cannot exclude the possibility that lemon grass is able to exert an inhibitory effect on immunological components of the intestinal immune system other than lymphocytes. It is also well known that lemon grass antimicrobial activity against bacteria or fungi. Therefore, there is also the possibility that microbacterial activity of lemon grass changes the population of commensal bacteria, leading to attenuation of inflammatory activity of ileitis. We previously showed enterobacteria recruitment of lymphocytes by modulating expression of adhesion molecules [28]. In addition, change in bacterial flora by prebiotics treatment altered lymphocyte homing [16]. Thus, it is also possible that lemon grass tea inhibits aberrant lymphocyte migration through modulating bacterial flora. Further studies are needed to determine the mechanism by which this natural herb prevents the development of ileitis and to explore its therapeutic usefulness for human IBD.

ACKNOWLEDGEMENTS

This research was supported by grants from the National Defense Medical College and the Japanese Ministry of Health, Labor and Welfare. We thank Dr. Satoshi Matsumoto, Yakult Central Institute, for providing SAMP1/Yit mice.

REFERENCES

1. Bachmann C, Klivanov AL, Olson TS, Sonnenschein JR, Rivera-Nieves J, Cominelli F, Ley KF, Lindner JR, Pizarro TT. Targeting mucosal addressin cellular adhesion molecule (MAdCAM)-1 to noninvasively image experimental Crohn's disease. *Gastroenterology* 130: 8–16, 2006.
2. Briskin M, Winsor-Hines D, Shyjan A, Cochran N, Bloom S, Wilson J, McEvoy LM, Butcher EC, Kassam N, Mackay CR, Newman W, Ringler DJ. Human mucosal addressin cell adhesion molecule-1 is preferentially expressed in intestinal tract and associated lymphoid tissue. *Am J Pathol* 151: 97–110, 1997.
3. Butcher EC, Williams M, Youngman K, Rott L, Briskin M. Lymphocyte trafficking and regional immunity. *Adv Immunol* 72: 209–253, 1999.
4. Connor MJ, Smit MH. Terminal-group oxidation of retinol by mouse epidermis. Inhibition in vitro and in vivo. *Biochem J* 244: 489–492, 1987.

5. Fujimori H, Miura S, Koseki S, Hokari R, Komoto S, Hara Y, Hachimura S, Kaminogawa S, Ishii H. Intra-vital observation of adhesion of lamina propria lymphocytes to microvessels of small intestine in mice. *Gastroenterology* 122: 734–744, 2002.
6. Ghosh S, Goldin E, Gordon FH, Malchow HA, Rask-Madsen J, Rutgeerts P, Vyhálek P, Zádorová Z, Palmer T, Donoghue S; Natalizumab Pan-European Study Group. Natalizumab for active Crohn's disease. *N Engl J Med* 348: 24–32, 2003.
7. Hosoe N, Miura S, Watanabe C, Tsuzuki Y, Hokari R, Oyama T, Fujiyama Y, Nagata H, Ishii H. Demonstration of functional role of TECK/CCL25 in T lymphocyte-endothelium interaction in inflamed and uninfamed intestinal mucosa. *Am J Physiol Gastrointest Liver Physiol* 286: G458–G466, 2004.
8. Inoue T, Tsuzuki Y, Matsuzaki K, Matsunaga H, Miyazaki J, Hokari R, Okada Y, Kawaguchi A, Nagao S, Itoh K, Matsumoto S, Miura S. Blockade of PSGL-1 attenuates CD14+ monocytic cell recruitment in intestinal mucosa and ameliorates ileitis in SAMP1/Yit mice. *J Leukoc Biol* 77: 287–295, 2005.
9. Iwata M, Hirakiyama A, Eshima Y, Kagechika H, Kato C, Song SY. Retinoic acid imprints gut-homing specificity on T cells. *Immunity* 21: 527–538, 2004.
10. Iwata M, Mukai M, Nakai Y, Iseki R. Retinoic acids inhibit activation-induced apoptosis in T cell hybridomas and thymocytes. *J Immunol* 149: 3302–3308, 1992.
11. Johansson-Lindbom B, Svensson M, Wurbel MA, Malissen B, Márquez G, Agace W. Selective generation of gut tropic T cells in gut-associated lymphoid tissue (GALT): requirement for GALT dendritic cells and adjuvant. *J Exp Med* 198: 963–969, 2003.
12. Matsumoto S, Okabe Y, Setoyama H, Takayama K, Ohtsuka J, Funahashi H, Imaoka A, Okada Y, Umesaki Y. Inflammatory bowel disease-like enteritis and caecitis in a senescence accelerated mouse P1/Yit strain. *Gut* 43: 71–78, 1998.
13. Matsuzaki K, Tsuzuki Y, Matsunaga H, Inoue T, Miyazaki J, Hokari R, Okada Y, Kawaguchi A, Nagao S, Itoh K, Matsumoto S, Miura S. In vivo demonstration of T lymphocyte migration and amelioration of ileitis in intestinal mucosa of SAMP1/Yit mice by the inhibition of MAdCAM-1. *Clin Exp Immunol* 140: 22–31, 2005.
14. Mora JR, Bono MR, Manjunath N, Weninger W, Cavanagh LL, Roseblatt M, Von Andrian UH. Selective imprinting of gut-homing T cells by Peyer's patch dendritic cells. *Nature* 424: 88–93, 2003.
15. Natl Toxicol Program Tech Rep Ser. Toxicology and Carcinogenesis Studies of Citral (Microencapsulated) (CAS No. 5392-40-5) in F344/N rats and B6C3F1 mice (feed studies). [U.S. Department of Health and Human Services.], 2003. http://ntp.niehs.nih.gov/ntp/htdocs/LT_rpts/tr505.pdf
16. Okada Y, Tsuzuki Y, Miyazaki J, Matsuzaki K, Hokari R, Komoto S, Kato S, Kawaguchi A, Nagao S, Itoh K, Watanabe T, Miura S. Propionibacterium freudenreichii component 1,4-dihydroxy-2-naphthoic acid (DHNA) attenuates dextran sodium sulphate induced colitis by modulation of bacterial flora and lymphocyte homing. *Gut* 55: 681–688, 2006.
17. Onawunmi GO, Yisak WA, Ogunlana EO. Antibacterial constituents in the essential oil of *Cymbopogon citratus* (DC.) Stapf. *J Ethnopharmacol* 12: 279–286, 1984.
18. Panes J, Granger DN. Leukocyte-endothelial cell interactions: molecular mechanisms and implications in gastrointestinal disease. *Gastroenterology* 114: 1066–1090, 1998.
19. Papadakis KA, Landers C, Prelin J, Kouroumalis EA, Moreno ST, Gutiérrez-Ramos JC, Hodge MR, Targan SR. CC chemokine receptor 9 expression defines a subset of peripheral blood lymphocytes with mucosal T cell phenotype and Th1 or T-regulatory 1 cytokine profile. *J Immunol* 171: 159–165, 2003.
20. Papadakis KA, Prelin J, Moreno ST, Cheng L, Kouroumalis EA, Deem R, Breaverman T, Ponath PD, Andrew DP, Green PH, Hodge MR, Binder SW, Targan SR. CCR9-positive lymphocytes and thymus-expressed chemokine distinguish small bowel from colonic Crohn's disease. *Gastroenterology* 121: 246–254, 2001.
21. Picarella D, Hurlbut P, Rottman J, Shi X, Butcher E, Ringler DJ. Monoclonal antibodies specific for $\beta 7$ integrin and mucosal addressin cell adhesion molecule-1 (MAdCAM-1) reduce inflammation in the colon of acid mice reconstituted with CD45RBhigh CD4+ T cells. *J Immunol* 158: 2099–2106, 1997.
22. Reif S, Klein I, Lubin F, Shi X, Butcher E, Ringler DJ. Pre-illness dietary factors in inflammatory bowel disease. *Gut* 40: 754–760, 1997.
23. Sandborn WJ, Colombel JF, Enns R, Feagan BG, Hanauer SB, Lawrance IC, Panaccione R, Sanders M, Schreiber S, Targan S, van Deventer S, Goldblum R, Despain D, Hogge GS, Rutgeerts P. Natalizumab induction and maintenance therapy for Crohn's disease. *N Engl J Med* 353: 1912–1925, 2005.
24. Sandborn WJ, Targan SR. Biologic therapy of inflammatory bowel disease. *Gastroenterology* 122: 1592–1608, 2002.
25. Saruta M, Yu QT, Avanesyan A, Fleshner PR, Targan SR, Papadakis KA. (Phenotype and effector function of CC chemokine receptor 9-expressing lymphocytes in small intestinal Crohn's disease. *J Immunol* 178: 3293–3300, 2007.
26. Sommer A, Tarwotjo I, Djunaedi E, West KP Jr, Loeden AA, Tilden R, Mele L. Impact of vitamin A supplementation on childhood mortality.

- A randomised controlled community trial. *Lancet* 1: 1169–1173, 1986.
27. Sommer A. Vitamin A deficiency, child health, and survival. *Nutrition* 13: 484–485, 1997.
 28. Takebayashi K, Hokari R, Kurihara C, Okada Y, Okudaira K, Matsunaga H, Komoto S, Watanabe C, Kawaguchi A, Nagao S, Tsuzuki Y, Miura S. Oral tolerance induced by enterobacteria altered the process of lymphocyte recruitment to intestinal microvessels: roles of endothelial cell adhesion molecules, TGF-beta and negative regulators of TLR signaling. *Microcirculation* 16: 251–264, 2009.
 29. Targan SR, Feagan BG, Fedorak RN, Lashner BA, Panaccione R, Present DH. Natalizumab for the treatment of active Crohn's disease: results of the ENCORE Trial. *Gastroenterology* 132: 1672–1683, 2007.
 30. Yokoyama H, Matsumoto M, Shiraishi H, Ishii H. Simultaneous quantification of various retinoids by high performance liquid chromatography: its relevance to alcohol research. *Alcohol Clin Exp Res* 24: 26S–29S, 2000.
 31. Van Assche G., Van Ranst M, Sciut R, Dubois B, Vermeire S, Noman M, Verbeek J, Geboes K, Robberecht W, Rutgeerts P. Progressive multifocal leukoencephalopathy after natalizumab therapy for Crohn's disease. *N Engl J Med* 353: 362–368, 2005.

VIII. 研究班構成

難治性腸管吸収機能障害Microscopic colitisに関する調査研究班

区 分	氏 名	所 属 等	職 名
研 究 代 表 者	渡 辺 守	東京医科歯科大学大学院医歯学総合研究科消化器病態学	教 授
研 究 分 担 者	平 田 一 郎	藤田保健衛生大学医学部消化管内科学	教 授
	松 井 敏 幸	福岡大学筑紫病院消化器内科	教 授
	三 浦 総 一 郎	防衛医科大学校内科学	教 授
	田 中 正 則	弘前市立病院臨床検査科	医療局長
	緒 方 晴 彦	慶應義塾大学医学部内視鏡センター	教 授
	松 本 主 之	九州大学病院消化管内科	診療准教授
	清 水 誠 治	JR大阪鉄道病院消化器内科	医務部長
	土 屋 輝 一 郎	東京医科歯科大学消化器内科	講 師
	岡 本 隆 一	東京医科歯科大学大学院医歯学総合研究科 消化管先端治療学（消化器病態学）	准教授
事 務 局	岡 本 隆 一	東京医科歯科大学大学院医歯学総合研究科 消化管先端治療学（消化器病態学） 〒113-8519 東京都文京区湯島1-5-45 T E L 03-5803-5877 F A X 03-5803-0268 e-mail rokamoto.gast@tmd.ac.jp	
経 理 事 務 担 当 者	若 山 友 啓	東京医科歯科大学 研究・産学連携推進機構事務部研究推進掛 T E L 03-5803-5872 F A X 03-5803-0179 e-mail t.wakayama.adm@cmn.tmd.ac.jp	

