

Figure 5. Effect of BCAA on HCV replication in cells in low-amino-acid medium. (A) Effect of BCAA on H77S.3/GLuc2A replication in Huh-7.5 cells. (B) Mx-1 expression in H77S.3/GLuc2A-transfected Huh-7.5 cells supplemented with BCAA. (C) Viability of Huh-7.5 cells. (D) Effect of BCAA on JFH-1 replication continuously infecting Huh-7 cells. (E) Mx-1 expression in continuously JFH-1-infected Huh-7 cells supplemented with BCAA. (F) Viability of Huh-7 cells.

to these metabolic aspects, recent reports have shown that mTORC1 participates in IFN signaling and antiviral defense responses,^{9,10} although the precise signaling pathway has not yet been clarified. In the present study, we evaluated mTORC1 signaling in CH-C livers using gene expression profiling of 91 patients (Figure 1, Supplementary Table 1). We observed a significant negative correlation between plasma Fischer's ratio and hepatic expression of BCAT1, an important catalytic enzyme of BCAA (Figure 1A). Moreover, BCAT1 expression was correlated positively with PDCD4 expression, which in turn is regulated negatively by pS6K at the transcriptional level (Figure 1D).¹⁶ Thus, the expression of BCAT1 appears to be a negative indicator of mTORC1 signaling in the liver, and the plasma Fischer's ratio is partially reflected by mTORC1 signaling in the liver and muscle.

Interestingly, the expression of c-myc was correlated significantly with BCAT1 (Figure 1C) as reported previously.¹⁵ Several studies observed up-regulated c-myc expression in advanced stages of CH-C¹⁹ but, on the other hand, c-myc recently was shown to be a target of

mTORC1 in hepatic cells.¹⁷ The existence of a feedback mechanism between c-myc and mTORC1 signaling to maintain liver homeostasis (Figure 1E) is plausible, although the precise mechanisms need to be confirmed.

Impaired mTORC1 signaling is suggested to affect the IFN- α -induced signaling pathway. To address this, the relationship between mTORC1 and IFN signaling was assessed using a cell culture system. In low-amino-acid medium ($\times 1/5$, $\times 1/30$, and $\times 1/100$ DMEM), expression of pSTAT1 was decreased substantially, correlating with the impaired mTORC1 signaling represented by decreased p-mTOR and pS6K expression in Huh-7 cells (Figure 2A).

The relationship between mTORC1 and IFN signaling was confirmed further by the knock-down experiment of Raptor, a specific subunit of mTORC1 (Figure 2B), although a more precise analysis should be performed to confirm this relationship. Importantly, when Huh-7 cells were stimulated by IFN- α , pSTAT1 induction was repressed significantly in low-amino-acid medium ($\times 1/5$ DMEM) or in Raptor knocked-down conditions (Figure 2C). It therefore could be speculated that IFN treat-

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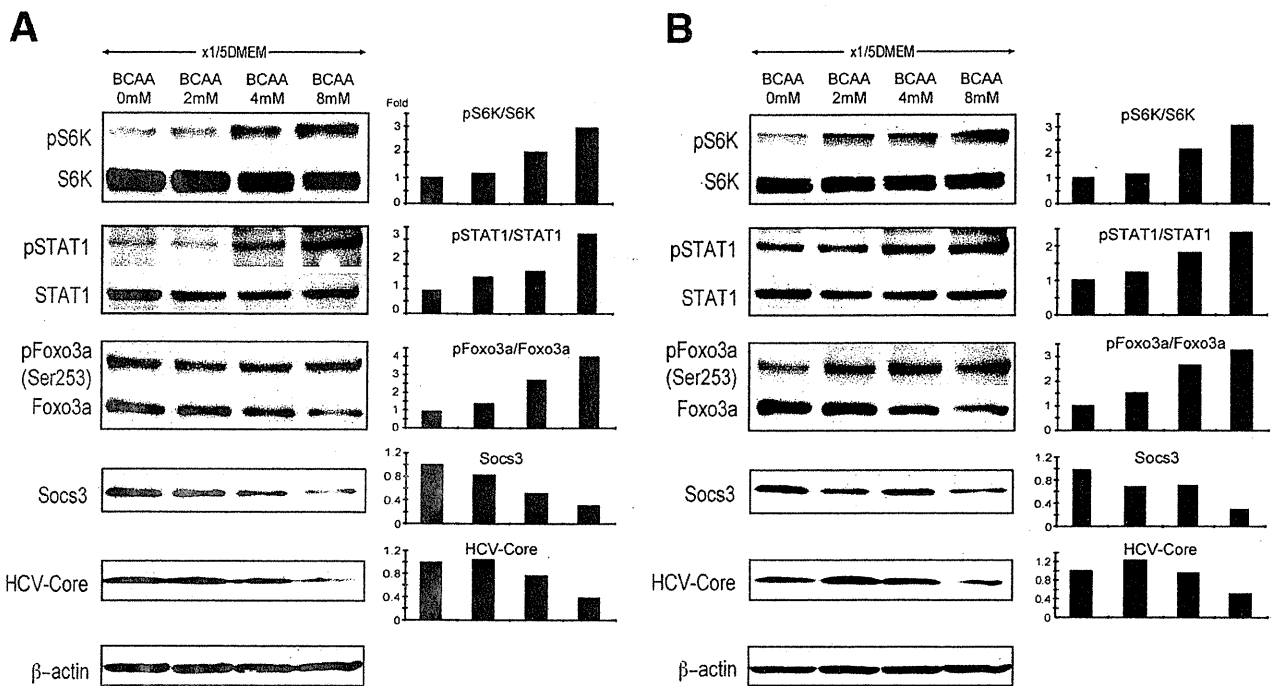


Figure 6. Expression of S6K, STAT1, Foxo3a, Socs3, and HCV core in H77S.3/GLuc2A-transfected Huh-7.5 cells or continuously JFH-1-infected Huh-7 cells supplemented with BCAA.

ment of patients with liver malnutrition and impaired mTORC1 signaling would lead to reduced induction of ISGs. Importantly, BCAA was able to restore impaired IFN signaling through increased binding of ISGF3 γ to its targets (Figure 2D–F).

Besides cross-talk of mTORC1 and IFN signaling, we revealed that Foxo3a also is involved in the IFN inhibitory pathway. In low-amino-acid medium, expression of pFoxo3a (ser253) was decreased substantially whereas that of Socs3 was increased. A decreased pFoxo3a/Foxo3a ratio indicates nuclear accumulation of Foxo3a before activation of its target genes, and this was confirmed by immunofluorescent staining (Figure 3C). The expression of Foxo3a was significantly positively correlated with that of Socs3 in CH-C liver (Figure 3F). These findings prompted us to identify a putative FBE in the Socs3 promoter region (Figure 4A). In fact, Socs3 promoter reporter activity was activated by overexpression of Foxo3a, and mutation of FBE impaired Foxo3a-dependent Socs3 promoter activation. Conversely, induction of Socs3 was not observed when expression of Foxo3a was knocked down by siRNA in low-amino-acid medium. Socs3 induction in low-amino-acid medium was owing to increased binding of Foxo3a to the FBE, which was confirmed by ChIP (Figure 4D). Therefore, in addition to impaired mTORC1 signaling, the Foxo3a-mediated Socs3 IFN inhibitory pathway might be involved in impaired IFN signaling in patients with liver malnutrition (Figure 4E).

Finally, we examined whether BCAA could restore impaired IFN signaling and inhibit HCV replication in cells

under conditions of malnutrition. Importantly, BCAA could repress replication of the recombinant genotype 1a-derived HCV, H77S.3/GLuc2A, in a dose-dependent manner (Figure 5A). H77S.3/GLuc2A RNA produces infectious virus¹⁴ and, therefore, the results indicate that BCAA might act on a naive HCV infection. Moreover, BCAA inhibited JFH-1-infected Huh-7 cells in which JFH-1 continuously was infecting in a dose-dependent manner. These results indicate that BCAA had an inhibitory effect on either naive or persistent HCV infection irrespective of genotypes (1a and 2a). Consistent with these results, BCAA induced the expression of pSTAT1 and Mx protein in a dose-dependent manner, and repressed Socs3 expression through increasing the ratio of pFoxo3a (ser243) to Foxo3a in a dose-dependent manner (Figures 5 and 6). Therefore, BCAA potentially could restore impaired IFN signaling and inhibit HCV replication in a CH-C state of malnutrition.

In conclusion, we addressed the clinical significance of the nutritional state of the liver on the treatment response of Peg-IFN and RBV combination therapy for CH-C. Although further studies are required to fully define the precise mechanisms underlying mTOR and IFN signaling, we showed that plasma values of Fischer's ratio are a useful nutritional parameter associated with treatment response. Fischer's ratio reflects mTORC1 signaling in the liver, which is correlated with IFN signaling and related to Socs3 IFN inhibitory signaling through Foxo3a. The potential usefulness of BCAA for the augmentation of IFN signaling could suggest a new therapeutic application for advanced-stage CH-C.

Supplementary Material

Note: To access the supplementary material accompanying this article, visit the online version of *Gastroenterology* at www.gastrojournal.org, and at doi: 10.1053/j.gastro.2011.03.051.

Appendix A

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Acknowledgments

Participating investigators from the Hokuriku Liver Study Group are listed in Appendix A.

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Conflicts of interest

The authors disclose no conflicts.

Supplementary Materials and Methods

Plasma Amino Acid Analysis

Plasma sample amino acid concentrations were measured by high-performance liquid chromatography-electrospray ionization-mass spectrometry followed by derivatization.¹ An MSQ Plus LC/MS system (Thermo Fischer Scientific, Waltham, MA) equipped with an electrospray ionization source was used in positive ionization mode for selected ion monitoring. Xcalibur version 1.4 SR1 software (Thermo Fischer Scientific, Yokohama, Japan) was used for data collection and processing. The high-performance liquid chromatography separation system consisted of an L-2100 pump, L-2200 autosampler, and L-2300 column oven (Hitachi High-Technologies Corporation, Tokyo, Japan). A Wakosil-II 3C8-100HG column (100, 2.1, 3 mm; Wako Pure Chemical Industries, Osaka, Japan) was used for the separation, and the mobile phase consisted of eluent A (25-mmol/L ammonium formate in water, pH 6.0) and eluent B (water:acetonitrile = 40:60).

Western Blotting

The expression of HCV core protein, Socs3, Foxo3a, phospho-Foxo3a (Ser253) (pFoxo3a), STAT1, pSTAT1 (Tyr701), S6K, pS6K, p-mTOR (Ser2448), Raptor, and β -actin were evaluated with mouse anti-core (Affinity BioReagents, Golden, CO), mouse anti-Socs3 (Santa Cruz Biotechnology, Santa Cruz, CA), rabbit anti-Foxo3a, rabbit anti- β -actin (Sigma-Aldrich, St Louis, MO), rabbit anti-phospho-Foxo3a (Ser253), rabbit anti-STAT1, rabbit anti-p-STAT1 (Tyr701), rabbit anti-p70 S6K, rabbit anti-pS6K, rabbit anti-p-mTOR (Ser2448), and rabbit anti-Raptor (Cell Signaling Technology, Beverly, MA), respectively. Densitometric analysis was conducted directly on the blotted membrane using a charge coupled device camera system (LAS-3000 Mini; FujiFilm, Tokyo, Japan) and Scion Image software (Frederick, MD).

Primer Sequences for PCR and siRNA

Primer sequences for PCR and siRNA were as follows: 2'5'OAS: forward 5'-CTC AGA AAT ACC CCA GCC AAA TC-3', reverse 5'-GTG GTG AGA GGA CTG AGG AA-3'; Socs3: forward 5'-TAC CAC CTG AGT CTC CAG CTT CTC-3', reverse 5'-CCT GGC AGT TCT CAT TAG TTC AGC ATT C-3'; Foxo3a: forward 5'-TGC TGT ATG CAA GAA CTT TCC AGT AGC AG-3', reverse 5'-ACT CTA GCC CCC ATG CTA CTA GTG-3'; glyceraldehyde-3-phosphate dehydrogenase: forward 5'-GAA GGT GAA GGT CGG AGT-3', reverse 5'-GAA GAT GGT GAT GGG ATT TC-3', siFoxo3a (SASI_Hs01_00119127; Sigma) sense: 5'-GAA UGA UGG GCU GAC UGA AdTdT-3', antisense: 5'-UUC AGU CAG CCC AUC AUU CdTdT-3'. Small interfering Raptor was purchased as

part of KIAA1303 siGENOME SMART pool siRNA reagents from Dharmacon, Inc (Lafayette, CO).

Construction of ISRE-Luc Reporter and FBEmut-luc Reporter Plasmids

Oligonucleotides containing the ISRE tandem repeat sequence (sense 5'-TCG AGA ACT GAA ACT GAA ACT GAA ACT GAA ACT GAA ACT GAA ACT GAA ACT GAA A-3', antisense 5'-AGC TTT TCA GTT TCA GTT TCA GTT TCA GTT TCA GTT TCA GTT TCA GTT TCA GTT TCA GTT TCA GTT TCA GTT C-3', consensus 5'-GAA Ann GAA ACT-3') were annealed, and integrated into Xho I and Hind III sites of the pGL4.23 luciferase vector (Promega). The human Socs3 promoter region (-109/+217) was amplified by genomic PCR using specific primers (forward, 5'-TGC TGC GAG TAG TGA CTA AAC ATT ACA AG-3' and reverse, 5'-CCG TGA AGT CCA CAA AGG AGC CTT C-3') and cloned into the EcoR V site of the pGL4.10-luc2 reporter vector (Promega). The Socs3 FBE mutant reporter vector was created by substituting 2 adenines in the putative FBE with guanines (wild-type sequence 5'-CTAAACA-3', mutated sequence 5'-CTGAGCA-3').

ChIP Assay

For the ChIP assay using the anti-ISGF3 γ antibody, 1×10^6 Huh-7 cells were treated with IFN- α (0 or 100 U/mL) and BCAA (2 mmol/L) in low-amino-acid medium for 6 hours. For ChIP using the anti-Foxo3a antibody, 1×10^6 Huh-7 cells were cultured in low-amino-acid medium for 24 hours.

Cells were cross-linked with 1% formaldehyde in PBS for 10 minutes at 37°C, and the reaction was stopped with 250 mmol/L glycine for 10 minutes. Cells were suspended in sodium dodecyl sulfate-lysis buffer (1% sodium dodecyl sulfate, 10 mmol/L ethylenediaminetetraacetic acid [EDTA], 50 mmol/L Tris-HCl [pH 8.1]), complete protease inhibitor cocktail (Roche Applied Science), and incubated for 30 minutes at 10°C. Cell lysate was sonicated with Bioruptor (Cosmo Bio, Tokyo, Japan) to obtain chromatin fragments and diluted 10-fold in ChIP dilution buffer (0.01% sodium dodecyl sulfate, 1.1% Triton-X 100, 1.2 mmol/L EDTA, 16.7 mmol/L Tris-HCl [pH 8.1], 150 mmol/L NaCl, complete protease inhibitor cocktail). Chromatin fragments were incubated with 2 μ g ISGF3 γ antibody (Santa Cruz Biotechnology), 2 μ g Foxo3a antibody (H-144; Santa Cruz Biotechnology), or normal rabbit immunoglobulin G for 18 hours at 4°C. Dynabeads (30 μ L) protein G (Invitrogen) was added and incubated for 1 hour at 4°C. The beads were washed with low-salt-wash buffer (0.1% sodium dodecyl sulfate, 1% Triton-X 100, 2.0 mmol/L EDTA, 20 mmol/L Tris-HCl [pH 8.1], 150 mmol/L NaCl), high-salt-wash buffer (0.1% sodium dodecyl sulfate, 1% Triton-X 100, 2.0 mmol/L EDTA, 20 mmol/L Tris-HCl [pH 8.1], 500 mmol/L NaCl), LiCl wash buffer (250 mmol/L LiCl, 1% NP-40, 1% de-

oxycholate, 1.0 mmol/L EDTA, 1.0 mmol/L Tris-HCl [pH 8.1]) and Tris-EDTA buffer. Immunoprecipitated chromatin fragments were eluted with elution buffer (1% sodium dodecyl sulfate, 100 mmol/L NaHCO₃, 10 mmol/L dithiothreitol), and reverse cross-linked by incubating for 6 hours at 65°C in elution buffer containing 200 mmol/L NaCl. DNA fragments were purified and quantified by real-time detection PCR with primers for putative ISRE in the 2'5'OAS promoter region (forward, 5'-AAA TGC ATT TCC AGA GCA GAG TTC AGA G-3', reverse, 5'-GGG TAT TTC TGA GAT CCA TCA TTG ACA GG-3') or putative FBE in the Socs3 promoter region (forward, 5'-TGC TGC GAG TAG TGA CTA AAC ATT ACA AG -3', reverse, 5'-AGC GGA GCA GGG AGT CCA AGT C -3'). Values were normalized by the measurement of input DNA.

pH77S.3/GLuc2A

pH77S.2 is a modification of pH77S² containing an additional mutation within the E2 protein (N476D in the polyprotein) that promotes infectious virus yields from RNA-transfected cells (Yi et al, unpublished data). To monitor replication, the GLuc sequence, fused at its C terminus to the foot-and-mouth disease virus 2A autoprotease, was inserted between p7 and NS2 of pH77S.2 (Supplementary Figure 4). To insert the GLuc-coding sequence between p7 and NS2 in pH77S.2, followed by the foot-and-mouth disease virus 2A protein-coding sequence, Mlu I, EcoR V, and Spe I restriction sites were created between the p7 and NS2 coding sequences by site-directed mutagenesis. DNA coding for GLuc was subcloned into the Mlu I and EcoR V sites of the modified plasmid after PCR amplification using the primers: 5'-ATA ATA TTA CGC GTA TGG GAG TCA AAG TTC TGT TTG CC-3' (sequence corresponding to the N-terminal GLuc is italicized and that corresponding to Mlu I is underlined) and 5'-ATA AAT AGAT ATC GTC ACC ACC GGC CCC CTT GAT CTT-3' (C terminal GLuc is italicized and EcoR V is underlined). A DNA fragment encoding the 17 amino acids of the foot-and-mouth disease virus 2A protein was generated by annealing the following complementary oligonucleotides: 5'-ATA TGA TAT CAA CTT TGA CCT TCT CAA GTT GGC CGG CGA CGT

CGA GTC CAA CCC AGG GCC CAC TAG CAT AT-3' and 5'-ATA TGC TAG TGG GCC CTG GGT TGG ACT CGA CGT CGC CGG CCA ACT TGA GAA GGT CAA AGT TGA TAT CAT AT-3' (underlined sequences indicate EcoR V and Spe I sites). The annealed oligonucleotides were digested by both restriction enzymes and the product inserted into the corresponding sites of pH77S.2 containing GLuc to generate pH77S.2/GLuc2A. Q41R is a cell-culture adaptive mutation within the NS3 protease domain of pH77S. Because it is not essential for production of infectious virus (Yi et al, unpublished data), pH77S.2 and pH77S.2/GLuc2A constructs underwent this mutation by site-directed mutagenesis of a PCR fragment spanning the Afe I and BsrG I sites to replace Gln₄₁ with wild-type Arg. The resulting plasmids (pH77S.2/R41Q and pH77S.2/GLuc2A/R41Q) were redesignated pH77S.3 and pH77S.3/GLuc2A, respectively.^{3,4} GLuc has several advantages over other luciferase reporter enzymes in that it is smaller and allows more sensitive detection than either firefly or Renilla luciferase.^{3,4} In addition, a signal sequence directs its secretion into cell-culture media, allowing real-time dynamic measurements of GLuc expression without the need for cell lysis. H77S.3/GLuc2A RNA produces infectious virus, although with lower efficiency than H77S.3 RNA (10-fold less).

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Pre-treatment prediction of response to pegylated-interferon plus ribavirin for chronic hepatitis C using genetic polymorphism in *IL28B* and viral factors

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Background & Aims: Pegylated interferon and ribavirin (PEG-IFN/RBV) therapy for chronic hepatitis C virus (HCV) genotype 1 infection is effective in 50% of patients. Recent studies revealed an association between the *IL28B* genotype and treatment response. We aimed to develop a model for the pre-treatment prediction of response using host and viral factors.

Methods: Data were collected from 496 patients with HCV genotype 1 treated with PEG-IFN/RBV at five hospitals and universities in Japan. *IL28B* genotype and mutations in the core and IFN sensitivity determining region (ISDR) of HCV were analyzed to predict response to therapy. The decision model was generated by data mining analysis.

Results: The *IL28B* polymorphism correlated with early virological response and predicted null virological response (NVR) (odds ratio = 20.83, $p < 0.0001$) and sustained virological response (SVR) (odds ratio = 7.41, $p < 0.0001$) independent of other covariates. Mutations in the ISDR predicted relapse and SVR independent of *IL28B*. The decision model revealed that patients with the minor *IL28B* allele and low platelet counts had the highest NVR (84%) and lowest SVR (7%), whereas those with the major *IL28B* allele and mutations in the ISDR or high platelet counts had the lowest NVR (0–17%) and highest SVR (61–90%). The model had high reproducibility and predicted SVR with 78% specificity and 70% sensitivity.

Conclusions: The *IL28B* polymorphism and mutations in the ISDR of HCV were significant pre-treatment predictors of response to PEG-IFN/RBV. The decision model, including these host and viral factors may support selection of optimum treatment strategy for individual patients.

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Introduction

Hepatitis C virus (HCV) infection is the leading cause of cirrhosis and hepatocellular carcinoma worldwide [1]. The successful eradication of HCV, defined as a sustained virological response (SVR), is associated with a reduced risk of developing hepatocellular carcinoma. Currently, pegylated interferon (PEG-IFN) plus ribavirin (RBV) is the most effective standard of care for chronic hepatitis C but the rate of SVR is around 50% in patients with HCV genotype 1 [2,3], the most common genotype in Japan, Europe, the United States, and many other countries. Moreover, 20–30% of patients with HCV genotype 1 have a null virological response (NVR) to PEG-IFN/RBV therapy [4]. The most reliable method for predicting the response is to monitor the early decline of serum HCV-RNA levels during treatment [5] but there is no established method for prediction before treatment. Because PEG-IFN/RBV therapy is costly and often accompanied by adverse effects such as flu-like symptoms, depression and hematological abnormalities, pre-treatment predictions of those patients who are unlikely to benefit from this regimen enables ineffective treatment to be avoided.

Recently, it has been reported through a genome-wide association study (GWAS) of patients with genotype 1 HCV that single nucleotide polymorphisms (SNPs) located near the *IL28B* gene are strongly associated with a response to PEG-IFN/RBV therapy in

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Table 1. Baseline characteristics of all patients, and patients assigned to the model building or validation groups.

	All patients n = 496	Model group n = 331	Validation group n = 165
Gender: male	250 (50%)	170 (51%)	80 (48%)
Age (years)	57.1 ± 9.9	56.8 ± 9.7	57.5 ± 10.2
ALT (IU/L)	78.6 ± 60.8	78.1 ± 61.4	79.7 ± 59.6
GGT (IU/L)	59.3 ± 63.6	58.9 ± 62.0	60.2 ± 66.9
Platelets (10 ⁹ /L)	154 ± 53	153 ± 52	154 ± 56
Fibrosis: F3-4	121 (24%)	80 (24%)	41 (25%)
HCV-RNA: >600,000 IU/ml	409 (82%)	273 (82%)	136 (82%)
ISDR mutation: ≤1	220 (88%)	290 (88%)	145 (88%)
Core 70 (Arg/Gln or His)	293 (59%)/203 (41%)	197 (60%)/134 (40%)	96 (58%)/69 (42%)
Core 91 (Leu/Met)	299 (60%)/197 (40%)	200 (60%)/131 (40%)	99 (60%)/66 (40%)
<i>IL28B</i> : Minor allele	151 (30%)	101 (31%)	50 (30%)
SVR	194 (39%)	129 (39%)	65 (39%)
Relapse	152 (31%)	103 (31%)	49 (30%)
NVR	150 (30%)	99 (30%)	51 (31%)

ALT, alanine aminotransferase; GGT, gamma-glutamyltransferase; ISDR, interferon sensitivity determining region; Arg, arginine; Gln, glutamine; His, histidine; Leu, leucine; Met, methionine; Minor, heterozygote or homozygote of minor allele; SVR, sustained virological response; NVR, null virological response.

Japanese [6], European [7], and a multi-ethnic population [8,9]. The last three studies focused on the association of SNPs in the *IL28B* region with SVR [7–9] but we found a stronger association with NVR [6]. In addition to these host genetic factors, we have reported that mutations within a stretch of 40 amino acids in the NS5A region of HCV, designated as the IFN sensitivity determining region (ISDR), are closely associated with the virological response to IFN therapy: a lower number of mutations is associated with treatment failure [10–13]. Amino acid substitutions at positions 70 and 91 of the HCV core region (Core70, Core91) also have been reported to be associated with response to PEG-IFN/RBV therapy: glutamine (Gln) or histidine (His) at Core70 and methionine (Met) at Core91 are associated with treatment resistance [4,14]. The importance of substitutions in the HCV core and ISDR was confirmed recently by a Japanese multicenter study [15]. How these viral factors contribute to response to therapy is yet to be determined. For general application in clinical practice, host genetic factors and viral factors should be considered together.

Data mining analysis is a family of non-parametric regression methods for predictive modeling. Software is used to automatically explore the data to search for optimal split variables and to build a decision tree structure [16]. The major advantage of decision tree analysis over logistic regression analysis is that the results of the analysis are presented in the form of flow chart, which can be interpreted intuitively and readily made available for use in clinical practice [17]. The decision tree analysis has been utilized to define prognostic factors in various diseases [18–25]. We have reported recently its usefulness for the prediction of an early virological response (undetectable HCV-RNA within 12 weeks of therapy) to PEG-IFN/RBV therapy in chronic hepatitis C [26].

This study aimed to define the pre-treatment prediction of response to PEG-IFN/RBV therapy through the integrated analysis of host factors, such as the *IL28B* genetic polymorphism and various clinical covariates, as well as viral factors, such as mutations in the HCV core and ISDR and serum HCV-RNA load. In addition,

for the general application of these results in clinical practice, decision models for the pre-treatment prediction of response were determined by data mining analysis.

Materials and methods

Patients

This was a multicentre retrospective study supported by the Japanese Ministry of Health, Labor and Welfare. Data were collected from a total of 496 chronic hepatitis C patients who were treated with PEG-IFN alpha and RBV at five hospitals and universities throughout Japan. Of these, 98 patients also were included in the original GWAS analysis [6]. The inclusion criteria in this study were as follows (1) infection by genotype 1b, (2) lack of co-infection with hepatitis B virus or human immunodeficiency virus, (3) lack of other causes of liver disease, such as autoimmune hepatitis, and primary biliary cirrhosis, (4) completion of at least 24 weeks of therapy, (5) adherence of more than 80% to the planned dose of PEG-IFN and RBV for the NVR patients, (6) availability of DNA for the analysis of the genetic polymorphism of *IL28B*, and (7) availability of serum for the determination of mutations in the ISDR and substitutions of Core70 and Core91 of HCV. Patients received PEG-IFN alpha-2a (180 µg) or 2b (1.5 µg/kg) subcutaneously every week and were administered a weight adjusted dose of RBV (600 mg for <60 kg, 800 mg for 60–80 kg, and 1000 mg for >80 kg daily) which is the recommended dosage in Japan. Written informed consent was obtained from each patient and the study protocol conformed to the ethical guidelines of the Declaration of Helsinki and was approved by the institutional ethics review committee. The baseline characteristics are listed in Table 1. For the data mining analysis, 67% of the patients (331 patients) were assigned randomly to the model building group and 33% (165 patients) to the validation group. There were no significant differences in the clinical backgrounds between these two groups.

Laboratory and histological tests

Blood samples were obtained before therapy and were analyzed for hematologic tests and for blood chemistry and HCV-RNA. Sequences of ISDR and the core region of HCV were determined by direct sequencing after amplification by reverse-transcription and polymerase chain reaction as reported previously [4,11]. Genetic polymorphism in one tagging SNP located near the *IL28B* gene (rs8099917) was determined by the GWAS or DigiTag2 assay [27]. Homozygosity (GG) or heterozygosity (TG) of the minor sequence was defined as having the *IL28B* minor allele, whereas homozygosity for the major sequence (TT) was

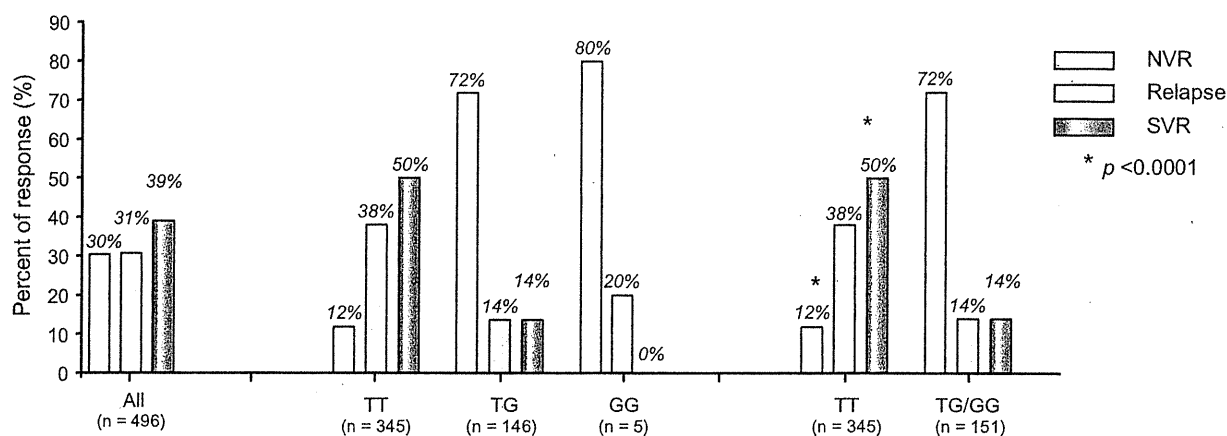


Fig. 1. Association between the IL28B genotype (rs8099917) and treatment response. The rates of response to treatment are shown for each rs8099917 genotype. The rate of null virological response (NVR), relapse, and sustained virological response (SVR) is shown. The *p* values are from Fisher's exact test. The rate of NVR was significantly higher ($p < 0.0001$) and the rate of SVR was significantly lower ($p < 0.0001$) in patients with the IL28B minor allele compared to those with the major allele.

defined as having the IL28B major allele. In this study, NVR was defined as a less than 2 log reduction of HCV-RNA at week 12 and detectable HCV-RNA by qualitative PCR with a lower detection limit of 50 IU/ml (Amplicor, Roche Diagnostic systems, CA) at week 24 during therapy. RVR (rapid virological response) and complete early virological response (cEVR) were defined as undetectable HCV-RNA at 4 weeks and 12 weeks during therapy and SVR was defined as undetectable HCV-RNA 24 weeks after the completion of therapy. Relapse was defined as reappearance of HCV-RNA after the completion of therapy. The stage of liver fibrosis was scored according to the METAVIR scoring system: F0 (no fibrosis), F1 (mild fibrosis: portal fibrosis without septa), F2 (moderate fibrosis: few septa), F3 (severe fibrosis: numerous septa without cirrhosis) and F4 (cirrhosis). Percentage of steatosis was quantified in 111 patients by determining the average proportion of hepatocytes affected by steatosis.

Statistical analysis

Associations between pre-treatment variables and treatment response were analyzed by univariate and multivariate logistic regression analysis. Associations between the IL28B polymorphism and sequences of HCV were analyzed by Fisher's exact test. SPSS software v.15.0 (SPSS Inc, Chicago, IL) was used for these analyses. For the data mining analysis, IBM-SPSS Modeler version 13.0 (IBM-SPSS Inc, Chicago, IL) software was utilized as reported previously [26]. The patients used for model building were divided into two groups at each step of the analysis based on split variables. Each value of each variable was considered as a potential split. The optimum variables and cut-off values were determined by a statistical search algorithm to generate the most significant division into two prognostic subgroups that were as homogeneous as possible for the probability of SVR. Thereafter, each subgroup was evaluated again and divided further into subgroups. This procedure was repeated until no additional significant variable was detected or the sample size was below 15. To avoid over-fitting, 10-fold cross validation was used in the tree building process. The reproducibility of the resulting model was tested with the data from the validation patients.

Results

Association between the IL28B (rs8099917) genotype and the PEG-IFN/RBV response

The rs8099917 allele frequency was 70% for TT ($n = 345$), 29% for TG ($n = 146$), and 1% for GG ($n = 5$). We defined the IL28B major allele as homozygous for the major sequence (TT) and the IL28B minor allele as homozygous (GG) or heterozygous (TG) for the minor sequence. The rate of NVR was significantly higher (72% vs. 12%, $p < 0.0001$) and the rate of SVR was significantly lower (14% vs. 50%, $p < 0.0001$) in patients with the IL28B minor allele compared to those with the major allele (Fig. 1).

Effect of the IL28B polymorphism, substitutions in the ISDR, Core70, and Core91 of HCV on time-dependent clearance of HCV

Patients were stratified according to their IL28B allele type, the number of mutations in the ISDR, the amino acid substitutions in Core70 and Core91, and the rate of undetectable HCV-RNA at 4, 8, 12, 24, and 48 weeks after the start of therapy were analyzed (Fig. 2A-D). The rate of undetectable HCV-RNA was significantly higher in patients with the IL28B major allele than the minor allele, in patients with two or more mutations in the ISDR compared to none or only one mutation, in patients with arginine (Arg) at Core70 rather than Gln/His, and in patients with leucine (Leu) at Core91 rather than Met. The difference was most significant when stratified by the IL28B allele type. The rate of RVR and cEVR was significantly more frequent in patients with the IL28B major allele compared with those with the IL28B minor allele: 9% vs. 3% for RVR ($p < 0.005$) and 57% vs. 11% for cEVR ($p < 0.0001$). These findings suggest that IL28B has the greatest impact on early virological response to therapy.

Association between substitutions in the ISDR and relapse after the completion of therapy

Patients were stratified according to the IL28B allele, number of mutations in the ISDR, and amino acid substitutions of Core70 and Core91, and the rate of relapse was analyzed (Fig. 3A and B). Among patients who achieved cEVR, the rate of relapse was significantly lower in patients with two or more mutations in the ISDR compared to those with only one or no mutations (15% vs. 31%, $p < 0.005$) (Fig. 3 B). On the other hand, the relapse rate was not different between the IL28B major and minor alleles within patients who achieved RVR (3% vs. 0%) or cEVR (28% vs. 29%) (Fig. 3A). Amino acid substitutions of Core70 and Core91 were not associated with the rate of relapse (data not shown).

Factors associated with response by multivariate logistic regression analysis

By univariate analysis, the minor allele of IL28B ($p < 0.0001$), one or no mutations in the ISDR ($p = 0.03$), high serum level of

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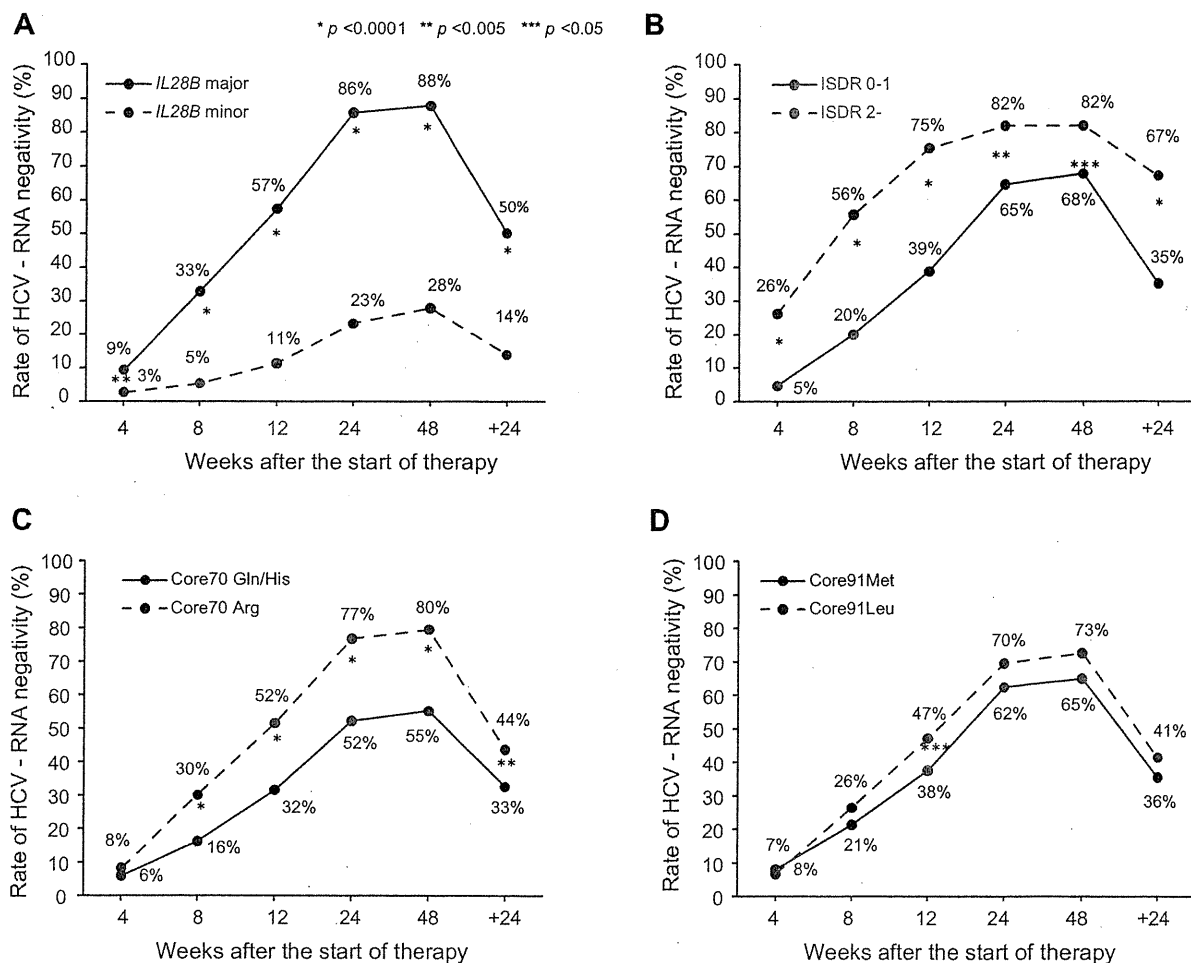


Fig. 2. Effect of *IL28B* mutations in the ISDR, Core70, and Core91 of HCV on time-dependent clearance of HCV. The rate of undetectable HCV-RNA was plotted for serial time points after the start of therapy (4, 8, 12, 24, and 48 weeks) and for 24 weeks after the completion of therapy. Patients were stratified according to (A) the *IL28B* allele (minor allele vs. major allele), (B) the number of mutations in the ISDR (0-1 mutation vs. 2 or more mutations), amino acid substitutions of (C) Core70 (Gln/His vs. Arg), and (D) Core91 (Met vs. Leu). The *p* values are from Fisher's exact test.

HCV-RNA ($p = 0.035$), Gln or His at Core70 ($p < 0.0001$), low platelet counts ($p = 0.009$), and advanced fibrosis ($p = 0.0002$) were associated with NVR. By multivariate analysis, the minor allele of *IL28B* (OR = 20.83, 95%CI = 11.63–37.04, $p < 0.0001$) was associated with NVR independent of other covariates (Table 2). Notably, mutations in the ISDR ($p = 0.707$) and at amino acid Core70 ($p = 0.207$) were not significant in multivariate analysis due to the positive correlation with the *IL28B* polymorphism ($p = 0.004$ for ISDR and $p < 0.0001$ for Core70, Fig. 4).

Genetic polymorphism of *IL28B* also was associated with SVR (OR = 7.41, 95% CI = 4.05–13.57, $p < 0.0001$) independent of other covariates, such as platelet counts, fibrosis, and serum levels of HCV-RNA. Mutation in the ISDR was an independent predictor of SVR (OR = 2.11, 95% CI = 1.06–4.18, $p = 0.033$) but the amino acid at Core70 was not (Table 3).

Factors associated with the *IL28B* polymorphism

Patients with the *IL28B* minor allele had significantly higher serum level of gamma-glutamyltransferase (GGT) and a higher

frequency of hepatic steatosis (Table 4). When the association between the *IL28B* polymorphism and HCV sequences was analyzed, Gln or His at Core70, that is linked to resistance to PEG-IFN and RBV therapy [4,14,15], was significantly more frequent in patients with the minor *IL28B* allele than in those with the major allele (67% vs. 30%, $p < 0.0001$) (Fig. 4). Other HCV sequences with an IFN resistant phenotype also were more prevalent in patients with the minor *IL28B* allele than those with the major allele: Met at Core91 (46% vs. 37%, $p = 0.047$) and one or no mutations in the ISDR (94% vs. 85%, $p = 0.004$) (Fig. 4).

Data mining analysis

Data mining analysis was performed to build a model for the prediction of SVR and the result is shown in Fig. 5. The analysis selected four predictive variables, resulting in six subgroups of patients. Genetic polymorphism of *IL28B* was selected as the best predictor of SVR. Patients with the minor *IL28B* allele had a lower probability of SVR and a higher probability of NVR than those with the major *IL28B* allele (SVR: 14% vs. 50%, NVR: 72% vs.

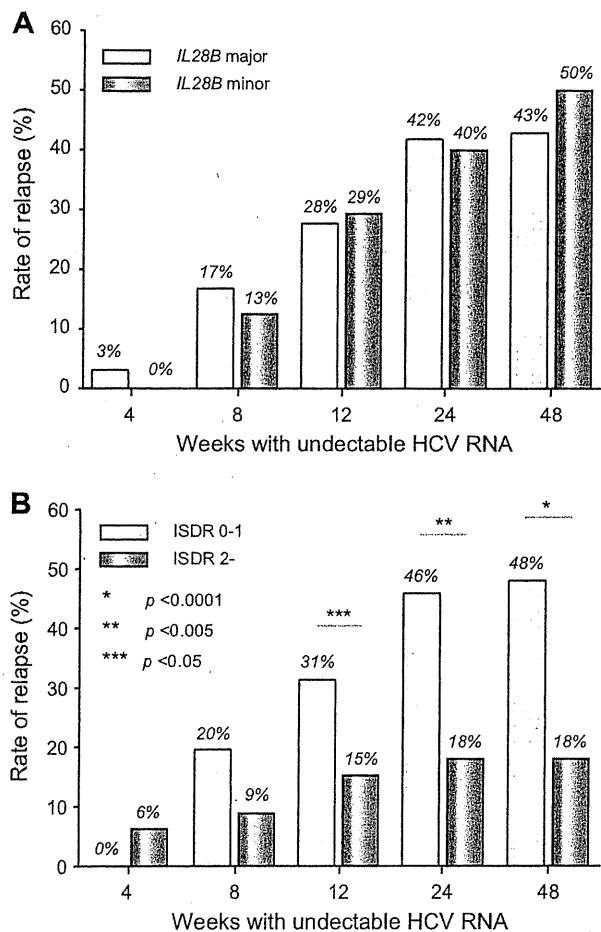


Fig. 3. Association between relapse and the *IL28B* allele or mutations in the ISDR. The rate of relapse was calculated for patients who had undetectable HCV-RNA at serial time points after the start of therapy (4, 8, 12, 24, and 48 weeks). Patients were stratified according to (A) the *IL28B* allele (minor allele vs. major allele) and (B) the number of mutations in the ISDR (0–1 mutation vs. 2 or more mutations). The *p* values are from Fisher's exact test.

12%). After stratification by the *IL28B* allele, patients with low platelet counts ($<140 \times 10^9/L$) had a lower probability of SVR and higher probability of NVR than those with high platelet counts ($\geq 140 \times 10^9/L$): for the minor *IL28B* allele, SVR was 7% vs. 19%, and NVR was 84% vs. 62%, and for the major *IL28B* allele, SVR was 32% vs. 66% and NVR was 16% vs. 8%. Among patients with the major *IL28B* allele and low platelet counts, those with two or more mutations in the ISDR had a higher probability of SVR and lower probability of relapse than those with one or no mutations in the ISDR (SVR: 75% vs. 27%, and relapse: 8% vs. 57%). Among patients with the major *IL28B* allele and high platelet counts, those with a low HCV-RNA titer ($<600,000$ IU/ml) had a higher probability of SVR and lower probability of NVR and relapse than those with a high HCV-RNA titer (SVR: 90% vs. 61%, NVR: 0% vs. 10%, and relapse: 10% vs. 29%). The sensitivity and specificity of the decision tree were 78% and 70%, respectively. The area under the receiver operating characteristic (ROC) curve of the model was 0.782 (data not shown). The pro-

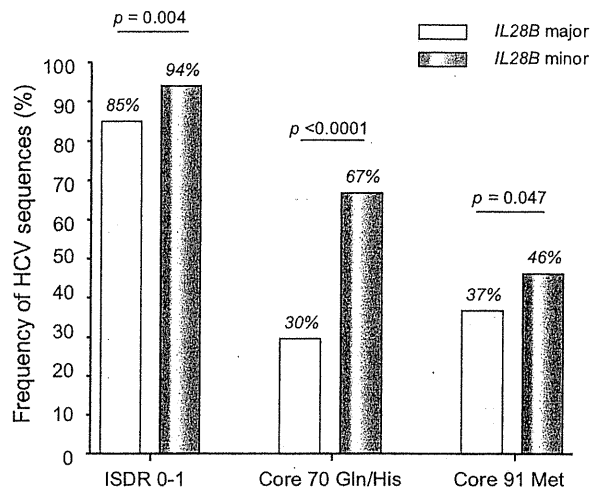


Fig. 4. Associations between the *IL28B* allele and HCV sequences. The prevalence of HCV sequences predicting a resistant phenotype to IFN was higher in patients with the minor *IL28B* allele than those with major allele. (A) 0 or 1 mutation in the ISDR of NS5A, (B) Gln or His at Core70, and (C) Met at Core91. *p* values are from Fisher's exact test.

portion of patients with advanced fibrosis (F3–4) was 39% (84/217) in patients with low platelet counts ($<140 \times 10^9/L$) compared to 13% (37/279) in those with high platelet counts ($\geq 140 \times 10^9/L$).

Validation of the data mining analysis

The results of the data mining analysis were validated with 165 patients who differed from those used for model building. Each patient was allocated to one of the six subgroups for the validation using the flow-chart form of the decision tree. The rate of SVR and NVR in each subgroup was calculated. The rates of SVR and NVR for each subgroup of patients were closely correlated between the model building and the validation patients ($r^2 = 0.99$ and 0.98) (Fig. 6).

Discussion

The rate of NVR after 48 weeks of PEG-IFN/RBV therapy among patients infected with HCV of genotype 1 is around 20–30%. Previously, there have been no reliable baseline predictors of NVR or SVR. Because more potent therapies, such as protease and polymerase inhibitor of HCV [28,29] and nitazoxanide [30], are in clinical trials and may become available in the near future, a pre-treatment prediction of the likelihood of response may be helpful for patients and physicians, to support clinical decisions about whether to begin the current standard of care or whether to wait for emerging therapies. This study revealed that the *IL28B* polymorphism was the overwhelming predictor of NVR and is independent of host factors and viral sequences reported previously. The *IL28B* encodes a protein also known as IFN-lambda 3, which is thought to suppress the replication of various viruses including HCV [31,32]. The results of the current study and the findings of the GWAS studies [6–9] may provide the rationale for developing diagnostic testing or an IFN-lambda based therapy for chronic hepatitis C in the future.

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Table 2. Factors associated with NVR analyzed by univariate and multivariate logistic regression analysis.

	Univariate			Multivariate		
	Odds ratio	95%CI	p value	Odds ratio	95%CI	p value
Gender: female	0.98	0.67-1.45	0.938	1.29	0.75-2.23	0.363
Age	1.01	0.97-1.01	0.223	0.99	0.97-1.02	0.679
ALT	1.00	1.00-1.00	0.867	1.00	0.99-1.00	0.580
GGT	1.004	1.00-1.01	0.029	1.00	1.00-1.00	0.715
Platelets	0.95	0.91-0.99	0.009	0.92	0.87-0.98	0.006
Fibrosis: F3-4	2.23	1.46-3.42	0.0002	1.97	1.09-3.57	0.025
HCV-RNA: $\geq 600,000$ IU/ml	1.83	1.05-3.19	0.035	2.49	1.17-5.29	0.018
ISDR mutation: ≤ 1	2.14	1.08-4.22	0.030	0.96	0.78-1.18	0.707
Core 70 (Gln/His)	3.23	2.16-4.78	<0.0001	1.41	0.83-2.42	0.207
Core 91 (Met)	1.39	0.95-2.06	0.093	1.21	0.72-2.04	0.462
<i>IL28B</i> : Minor allele	19.24	11.87-31.18	<0.0001	20.83	11.63-37.04	<0.0001

ALT, alanine aminotransferase; GGT, gamma-glutamyltransferase; ISDR, interferon sensitivity determining region; Gln, glutamine; His, histidine; Met, methionine; Minor allele, heterozygote or homozygote of minor allele.

Table 3. Factors associated with SVR analyzed by univariate and multivariate logistic regression analysis.

	Univariate			Multivariate		
	Odds ratio	95%CI	p value	Odds ratio	95%CI	p value
Gender: female	0.81	0.56-1.16	0.253	0.86	0.55-1.35	0.508
Age	0.97	0.95-0.99	0.0003	0.99	0.96-1.01	0.199
ALT	1.00	1.00-1.00	0.337	1.00	1.00-1.01	0.108
GGT	1.00	1.00-1.00	0.273	1.00	1.00-1.00	0.797
Platelets	1.12	1.01-1.16	<0.0001	1.13	1.08-1.19	<0.0001
Fibrosis: F0-2	2.64	1.65-4.22	<0.0001	1.87	1.07-3.28	0.029
HCV-RNA: <600,000 IU/ml	2.49	1.55-3.98	0.0001	2.75	1.55-4.90	0.001
ISDR mutation: $2 \leq$	3.78	2.14-6.68	<0.0001	2.11	1.06-4.18	0.033
Core 70 (Arg)	1.61	1.11-2.28	0.012	0.84	0.52-1.35	0.470
Core 91 (Leu)	1.28	0.88-1.85	0.185	1.26	0.81-1.96	0.300
<i>IL28B</i> : Major allele	6.21	3.75-10.31	<0.0001	7.41	4.05-13.57	<0.0001

ALT, alanine aminotransferase; GGT, Gamma-glutamyltransferase; ISDR, interferon sensitivity determining region; Arg, arginine; Leu, leucine; Major allele, homozygote of major allele.

Among baseline factors, *IL28B* was the most significant predictor of NVR and SVR. Moreover, the *IL28B* allele type was also correlated with early virological response: the rate of RVR and cEVR was significantly high for the *IL28B* major allele compared to the *IL28B* minor allele: 9% vs. 3% for RVR and 57% vs. 11% for cEVR (Fig. 2). On the other hand, the relapse rate was not different between the *IL28B* genotypes within patients who achieved RVR or cEVR (Fig. 3). We believe that optimal therapy should be based on baseline features and a response-guided approach. Our findings suggest that the *IL28B* genotype is a useful baseline predictor of virological response which should be used for selecting the treatment regimen: whether to treat patients with PEG-IFN and RBV or to wait for more effective future therapy including direct acting antiviral drugs. On the other hand, baseline *IL28B* genotype might not be suitable for determining the treatment duration in patients who started PEG-IFN/RBV therapy

and whose virological response is determined because the *IL28B* genotype is not useful for the prediction of relapse. The duration of therapy should be personalized based on the virological response. Future studies need to explore whether the combination of baseline *IL28B* genotype and response-guided approach further improves the optimization of treatment duration.

The SVR rate in patients having the *IL28B* minor allele was 14% in the present study while it was 23% in Caucasians and 9% in African Americans in a study by McCarthy et al. [33]. On the other hand, the SVR rate in patients having the *IL28B* minor allele was 28% in genotypes 1/4 compared to 80% in genotypes 2/3 in a study by Rauch et al. [9]. These data imply that the impact of the *IL28B* polymorphism on response to therapy may be different in terms of race, geographical areas, or HCV genotypes, and that our data need to be validated in future studies including different populations and geographical areas before generalization.

Table 4. Factors associated with *IL28B* genotype.

	<i>IL28B</i> major allele n = 345	<i>IL28B</i> minor allele n = 151	p value
Gender: male	166 (48%)	84 (56%)	0.143
Age (years)	57 ± 10	57 ± 10	0.585
ALT (IU/L)	79 ± 60	78 ± 62	0.842
Platelets (10 ⁹ /L)	153 ± 54	155 ± 52	0.761
GGT (IU/L)	51 ± 45	78 ± 91	0.001
Fibrosis: F3-4	76 (22%)	45 (30%)	0.063
Steatosis:			
>10%	16/88 (18%)	13/23 (57%)	0.024
>30%	6/88 (7%)	6/23 (26%)	0.017
HCV-RNA: >600,000 IU/ml	284 (82%)	125 (83%)	1.000

ALT, alanine aminotransferase; GGT, gamma-glutamyltransferase.

Four GWAS studies have shown the association between a genetic polymorphism near the *IL28B* gene and response to PEG-IFN plus RBV therapy. The SNPs that showed significant association with response were rs12979860 [8] and rs8099917 [6,7,9]. There is a strong linkage-disequilibrium (LD) between these two SNPs as well as several other SNPs near the *IL28B* gene in Japanese patients [34] but the degree of LD was weaker in Caucasians and Hispanics [8]. Thus, the combination of SNPs is not useful for predicting response in Japanese patients but may improve the predictive value in patients other than Japanese who have weaker LD between SNPs.

Other significant predictors of response independent of *IL28B* genotype were platelet counts, stage of fibrosis, and HCV RVA load. A previous study reported that platelet count is a predictor of response to therapy [35], and the lower platelet count was related with advanced liver fibrosis in the present study. The association between response to therapy and advanced fibrosis independent of the *IL28B* polymorphism is consistent with a recent study by Rauch et al. [9].

There is agreement that the viral genotype is significantly associated with the treatment outcome. Moreover, viral factors such as substitutions in the ISDR of the NS5A region [10] or in the amino acid sequence of the HCV core [4] have been studied in relation to the response to IFN treatment. The amino acid Gln or His at Core70 and Met at Core91 are repeatedly reported to be associated with resistance to therapy [4,14,15] in Japanese patients but these data wait to be validated in different populations or other geographical areas. In this study, we confirmed that patients with two or more mutations in the ISDR had a higher rate of undetectable HCV-RNA at each time point during therapy. In addition, the rate of relapse among patients who achieved cEVR was significantly lower in patients with two or more mutations in ISDR compared to those with only one or no mutations (15% vs. 31%, $p < 0.05$). Thus, the ISDR sequence may be used to predict a relapse among patients who achieved virological response during therapy, while the *IL28B* polymorphism may be used to predict the virological response before therapy. A higher number of mutations in the ISDR are reported to have close association with SVR in Japanese [11–13,15,36] or Asian [37,38] populations but data from Western countries have been controversial [39–42]. A meta-analysis of 1230 patients including 525 patients from Europe has shown that there was a positive correlation

between the SVR and the number of mutations in the ISDR in Japanese as well as in European patients [43] but this correlation was more pronounced in Japanese patients. Thus, geographical factors may account for the different impact of ISDR on treatment response, which may be a potential limitation of our study.

To our surprise, these HCV sequences were associated with the *IL28B* genotype: HCV sequences with an IFN resistant phenotype were more prevalent in patients with the minor *IL28B* allele than those with the major allele. This was an unexpected finding, as we initially thought that host genetics and viral sequences were completely independent. A recent study reported that the *IL28B* polymorphism (rs12979860) was significantly associated with HCV genotype: the *IL28B* minor allele was more frequent in HCV genotype 1-infected patients compared to patients infected with HCV genotype 2 or 3 [33]. Again, patients with the *IL28B* minor allele (IFN resistant genotype) were infected with HCV sequences that are linked to an IFN resistant phenotype. The mechanism for this association is unclear, but may be related to an interaction between the *IL28B* genotype and HCV sequences in the development of chronic HCV infection as discussed by McCarthy et al., since the *IL28B* polymorphism was associated with the natural clearance of HCV [44]. Alternatively, the HCV sequence within the patient may be selected during the course of chronic infection [45,46]. These hypotheses should be explored through prospective studies of spontaneous HCV clearance or by testing the time-dependent changes in the HCV sequence during the course of chronic infection.

How these host and viral factors can be integrated to predict the response to therapy in future clinical practice is an important question. Because various host and viral factors interact in the same patient, predictive analysis should consider these factors in combination. Using the data mining analysis, we constructed a simple decision tree model for the pre-treatment prediction of SVR and NVR to PEG-IFN/RBV therapy. The classification of patients based on the genetic polymorphism of *IL28B*, mutation in the ISDR, serum levels of HCV-RNA, and platelet counts, identified subgroups of patients who have the lowest probabilities of NVR (0%) with the highest probabilities of SVR (90%) as well as those who have the highest probabilities of NVR (84%) with the lowest probability of SVR (7%). The reproducibility of the model was confirmed by the independent validation based on a second group of patients. Using this model, we can rapidly develop an

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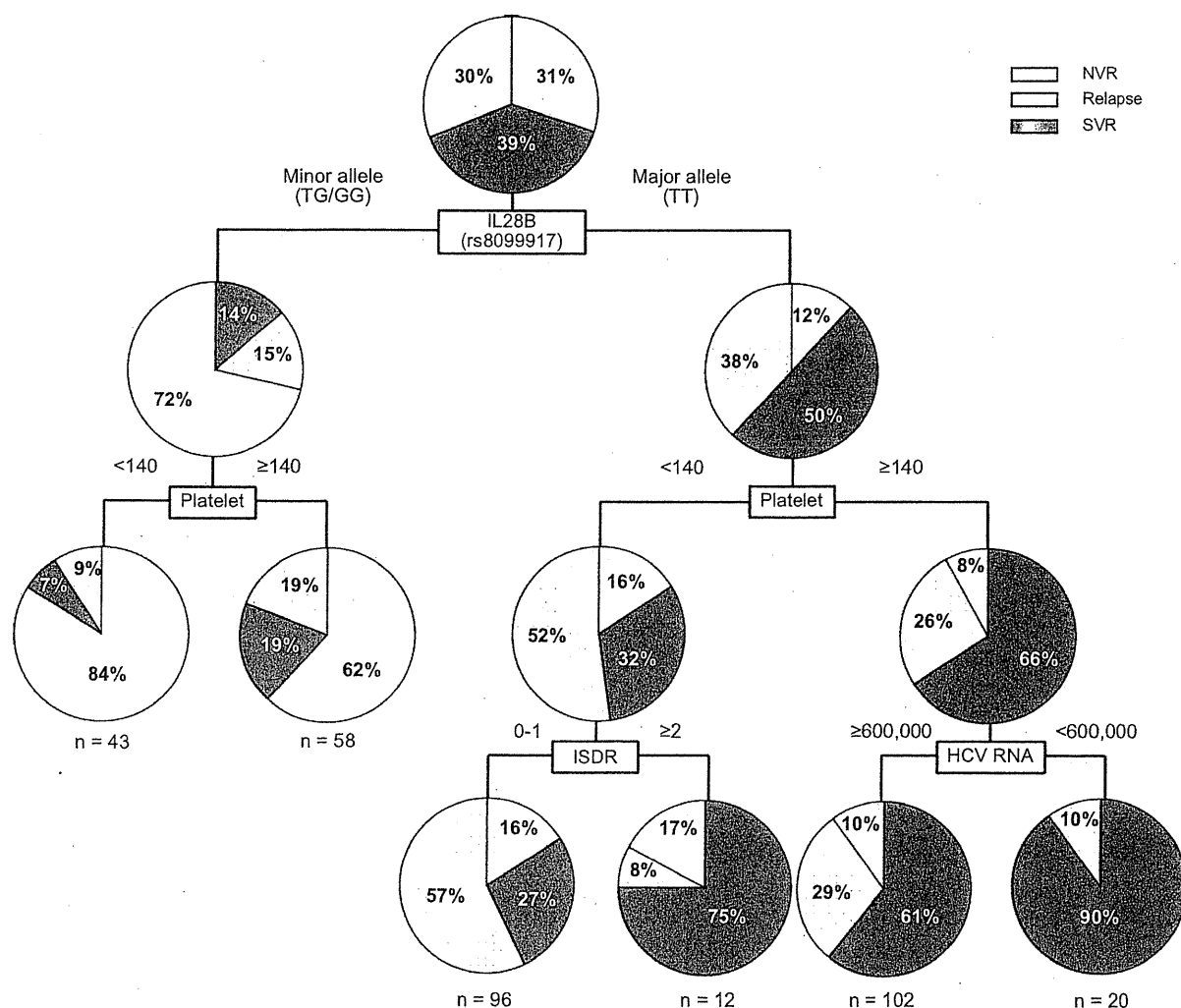


Fig. 5. Decision tree for the prediction of response to therapy. The boxes indicate the factors used for splitting. Pie charts indicate the rate of response for each group of patients after splitting. The rate of null virological response, relapse, and sustained virological response is shown.

estimate of the response before treatment, by simply allocating patients to subgroups by following the flow-chart form, which may facilitate clinical decision making. This is in contrast to the calculating formula, which was constructed by the traditional logistic regression model. This was not widely used in clinical practice as it is abstruse and inconvenient. These results support the evidence based approach of selecting the optimum treatment strategy for individual patients, such as treating patients with a low probability of NVR with current PEG-IFN/RBV combination therapy or advising those with a high probability of NVR to wait for more effective future therapies. Patients with a high probability of relapse may be treated for a longer duration to avoid a relapse. Decisions may be based on the possibility of a response against a potential risk of adverse events and the cost of the therapy, or disease progression while waiting for future therapy.

We have previously reported the predictive model of early virological response to PEG-IFN and RBV in chronic hepatitis C

[26]. The top factor selected as significant was the grade of steatosis, followed by serum level of LDL cholesterol, age, GGT, and blood sugar. The mechanism of association between these factors and treatment response was not clear at that time. To our interest, a recent study by Li et al. [47] has shown that high serum level of LDL cholesterol was linked to the *IL28B* major allele (CC in rs12979860). High serum level of LDL cholesterol was associated with SVR but it was no longer significant when analyzed together with the *IL28B* genotype in multivariate analysis. Thus, the association between treatment response and LDL cholesterol levels may reflect the underlining link of LDL cholesterol levels to *IL28B* genotype. Steatosis is reported to be correlated with low lipid levels [48] which suggest that *IL28B* genotypes may be also associated with steatosis. In fact, there were significant correlations between the *IL28B* genotype and the presence of steatosis in the present study (Table 4). In addition, the serum level of GGT, another predictive factor in our previous study, was signif-

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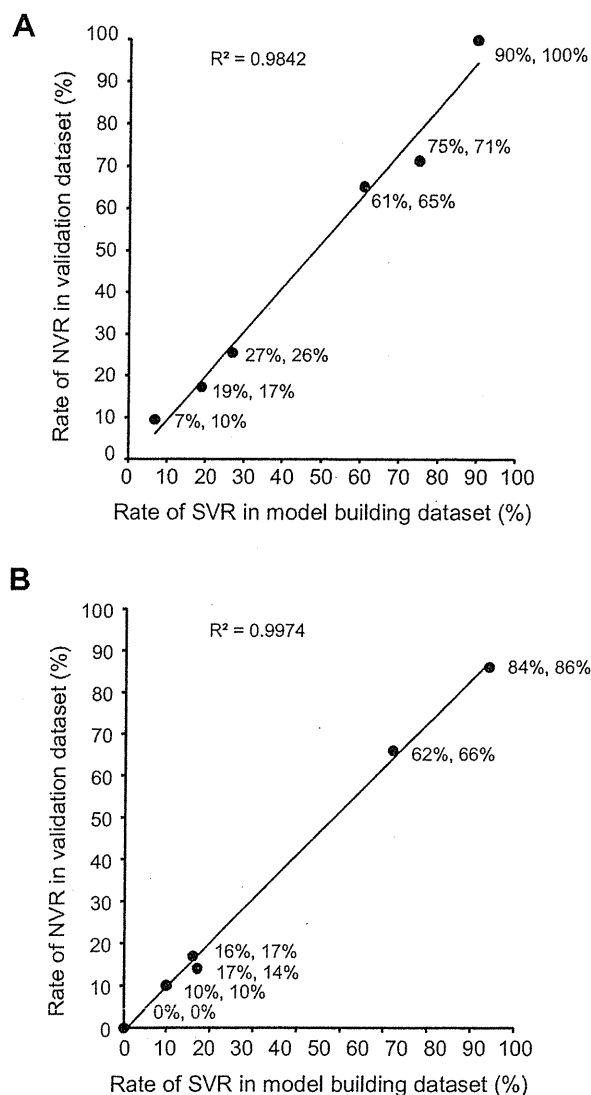


Fig. 6. Validation of the CART analysis. Each patient in the validation group was allocated to one of the six subgroups by following the flow-chart form of the decision tree. The rate of (A) sustained virological response (SVR) and (B) null virological response (NVR) in each subgroup was calculated and plotted. The X-axis represents the rate of SVR or NVR in the model building patients and the Y-axis represents those in the validation patients. The rate of SVR and NVR in each subgroup of patients is closely correlated between the model building and the validation patients (correlation coefficient: $r^2 = 0.98-0.99$).

icantly associated with *IL28B* genotype in the present study (Table 4). The serum level of GGT was significantly associated with NVR when examined independently but was no longer significant when analyzed together with the *IL28B* genotype. These observations indicate that some of the factors that we have previously identified may be associated with virological response to therapy through the underlining link to the *IL28B* genotype.

In conclusion, the present study highlighted the impact of the *IL28B* polymorphism and mutation in the ISDR on the pre-treatment prediction of response to PEG-IFN/RBV therapy. A decision model including these host and viral factors has the potential to

support selection of the optimum treatment strategy for individual patients, which may enable personalized treatment.

Conflict of interest

The authors who have taken part in this study declare that they do not have anything to disclose regarding funding or conflict of interest with respect to this manuscript.

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Differential interferon signaling in liver lobule and portal area cells under treatment for chronic hepatitis C

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Background & Aims: The mechanisms of treatment resistance to interferon (IFN) and ribavirin (Rib) combination therapy for hepatitis C virus (HCV) infection are not known. This study aims to gain insight into these mechanisms by exploring hepatic gene expression before and during treatment.

Methods: Liver biopsy was performed in 50 patients before therapy and repeated in 30 of them 1 week after initiating combination therapy. The cells in liver lobules (CLL) and the cells in portal areas (CPA) were obtained from 12 patients using laser capture microdissection (LCM).

Results: Forty-three patients were infected with genotype 1 HCV, 20 of who were viral responders (genotype 1-Rsp) with treatment outcome of SVR or TR, while 23 were non-responders (genotype 1-nonRsp) with NR. Only seven patients were infected with genotype 2. Before treatment, the expression of *IFN* and *Rib-stimulated genes* (IRSGs), apoptosis-associated genes, and immune reaction gene pathways was greater in genotype 1-nonRsp than in Rsp. During treatment, IRSGs were induced in genotype 1-Rsp, but not in nonRsp. IRSG induction was irrelevant in genotype 2-Rsp and was mainly impaired in CLL but not in CPA. Pathway analysis revealed that many immune regulatory pathways were induced in CLL from genotype 1-Rsp, while growth factors related to angiogenesis and fibrogenesis were more induced in CPA from genotype 1-nonRsp.

Conclusions: Impaired IRSGs induction in CLL reduces the sensitivity to treatment for genotype 1 HCV infection. CLL and CPA in the liver might be differentially involved in treatment resistance. These findings could be useful for the improvement of therapy for HCV infection.

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Introduction

A human liver infected with hepatitis C virus (HCV) develops chronic hepatitis, cirrhosis, and in some instances, hepatocellular carcinoma (HCC). Although interferon (IFN) and ribavirin (Rib) combination therapy has become a popular modality for treating patients with chronic hepatitis C (CH-C), about 50% of patients relapse, particularly those with genotype 1b and high viral load [8]. The reasons for treatment failure are poorly understood. Many studies of IFN and Rib combination therapy for CH-C suggested that patients who cleared HCV viremia early during therapy tended to show favorable outcomes. On the other hand, patients who needed a longer period to clear HCV had poorer outcomes [4,7,17], and those who showed no response (no or minimal decrease in HCV-RNA) to IFN and Rib combination therapy hardly ever achieved a sustained viral response (SVR).

To elucidate the underlying mechanism of treatment resistance, expression profiles in the liver [3,6,20] and peripheral mononuclear cells (PBMC) [10,21] during IFN treatment for CH-C patients have been examined. In chronic viral hepatitis, increased numbers of immune regulatory cells infiltrate the liver. These liver-infiltrating lymphocytes (LILs) might play important roles for virus eradication and are potentially linked to treatment outcome. Previously, we selectively isolated cells in liver lobules (CLL) and cells in the portal area (CPA) from biopsy specimens using laser capture microdissection (LCM) and analyzed their gene expression profiles [11,19]. From these profile analyses, it could be inferred that the majority of CLL were hepatocytes and the majority of CPA were lymphocytes, although other cellular components such as Kupffer cells, endothelial cells, myofibroblasts, and bile duct cells co-existed as well.

To gain further insight into the mechanisms of therapy resistance, we analyzed expression profiles in CLL and CPA in addition to whole liver tissues during IFN therapy for CH-C.

Keywords: HCV; IFN; LCM; Gene expression.

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Abbreviations: HCV, hepatitis C virus; HBV, hepatitis B virus; miRNA, micro RNA; CH-B, chronic hepatitis B; CH-C, chronic hepatitis C; HCC-B, hepatitis B-related hepatocellular carcinoma; HCC-C, hepatitis C-related hepatocellular carcinoma; OCT, optimum cutting temperature.



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Materials and methods

Patients

Patients with CH-C were enrolled in this study at the Graduate School of Medicine, Kanazawa University Hospital, Japan, between 2001 and 2007 (Tables 1 and 2). Prior to the study, we obtained the required approvals, namely: informed consent from all participating patients and ethics approval from the ethics committee for human genome/gene analysis research at Kanazawa University Graduate School of Medical Science. Thirty patients were administered IFN- α 2b (6 MU: every day for 2 weeks, then three times a week for 22 weeks) (Schering-Plough K.K., Tokyo, Japan) and Rib (10–13 mg/kg/day) combination therapy for 24 weeks (Table 1). Twenty patients were administered Peg-IFN- α 2b and Rib combination therapy for 48 weeks (Table 2). The final outcome of the treatment was assessed at 24 weeks after cessation of the combination therapy. In addition, 10 samples of normal liver tissues obtained during surgery for metastatic liver cancer were used as controls.

We defined treatment outcomes according to the decrease in viremia as follows: sustained viral response (SVR), clearance of HCV viremia at 24 weeks after cessation of therapy; transient response (TR), no detectable HCV viremia at 24 weeks but relapse during the follow-up period; and nonresponse (NR), HCV viremia detected at the cessation of therapy. We defined a patient who achieved SVR or TR as a viral responder (Rsp) and a patient who exhibited an NR as a non-responder (nonRsp). As patient 10 stopped treatment at 5 weeks due to an adverse side effect, we grouped this patient as Rsp based on the observed viral decline within 2 weeks (Table 1).

HCV genotype was classified by the methods described by Okamoto et al. [16]. Twenty-three patients were infected with genotype 1b and seven patients were infected with genotype 2 (2a; 6, 2b; 1) (Tables 1 and 2).

Patient serum was aliquoted and stored at -20 °C until use. HCV-RNA was serially monitored by quantitative real-time detection (RTD)-PCR (COBAS® AmpliPrep/COBAS® TaqMan® System®) [9] before treatment, at 48 h, 2 weeks and 24 weeks after initiation of therapy and at 24 weeks after cessation of therapy.

The grading and staging of chronic hepatitis were histologically assessed according to the method described by Desmet et al. (Table 1) [5].

Table 1. Characteristics of study patients who received IFN and ribavirin combination therapy.

Pt.No.	Sex	Age (yr)	Genotype	ALT (IU/ml)		Liver histology			LCM	HCV-RNA (Log IU/ml)			Viral kinetics		Viral response	Outcome		
				Before therapy	During therapy	Before therapy	During therapy	F		A	F	A	Before therapy	48 h			2 wk	24 wk
1	M	48	1b	83	45	1	1	1	1	+	6.6	4.5	3.5	-	1.1	0.5	Rsp	SVR
2	M	32	1b	192	95	1	1	1	1	-	6.4	3.9	3.2	-	1.3	0.4	Rsp	SVR
3	F	50	1b	57	37	1	1	1	1	-	5.8	2.5	1.5	-	1.7	0.5	Rsp	TR
4	M	36	1b	119	117	1	1	1	1	+	6.1	4.4	4.2	+	0.9	0.1	nonRsp	NR
5	M	54	1b	82	69	1	1	1	1	-	6.6	5.1	3.9	+	0.8	0.6	nonRsp	NR
6	M	43	1b	143	116	1	1	1	1	-	6.3	4.4	4.1	+	1.0	0.2	nonRsp	NR
7	M	48	1b	33	30	1	1	1	1	+	1.5	0.0	0.0	-	>0.8	-	Rsp	SVR
8	M	52	1b	316	374	1	2	1	1	-	4.7	5.1	3.9	+	-0.2	0.6	nonRsp	NR
9	M	45	1b	112	39	1	0	2	0	-	6.2	5.1	5.7	+	0.6	-0.3	nonRsp	NR
10	M	48	1b	48	30	2	2	2	1	+	6.4	4.0	2.6	NA	1.2	0.8	Rsp	NA
11	M	52	1b	114	80	2	2	2	1	-	6.1	3.7	3.0	-	1.2	0.4	Rsp	TR
12	F	63	1b	38	30	2	1	2	1	-	5.2	4.2	4.5	+	0.5	-0.2	nonRsp	NR
13	M	58	1b	90	83	2	2	2	2	+	6.9	4.9	5.6	+	1.0	-0.4	nonRsp	NR
14	F	61	1b	87	43	2	1	2	1	+	6.5	3.9	3.7	+	1.3	0.1	nonRsp	NR
15	F	64	1b	133	111	2	1	3	2	-	6.0	4.4	3.6	+	0.8	0.4	nonRsp	NR
16	F	62	1b	251	159	3	2	3	2	-	4.8	2.7	1.5	-	1.1	0.6	Rsp	SVR
17	M	54	1b	211	205	3	2	3	2	+	6.7	0.0	0.0	-	>3.4	-	Rsp	SVR
18	F	68	1b	153	145	3	2	3	2	+	4.9	4.3	3.5	+	0.3	0.4	nonRsp	NR
19	F	69	1b	64	43	3	2	3	2	-	4.4	1.5	0.0	-	1.5	0.8	Rsp	SVR
20	M	49	1b	91	83	3	2	3	2	+	6.6	4.2	3.8	+	1.2	0.2	nonRsp	NR
21	M	55	1b	187	196	4	1	4	2	-	5.8	5.1	5.6	+	0.4	-0.3	nonRsp	NR
22	F	45	1b	113	75	4	2	3	3	-	5.7	4.2	2.7	-	0.8	0.8	Rsp	TR
23	M	60	1b	86	49	4	2	3	1	-	6.3	3.5	3.5	+	1.4	0.0	nonRsp	NR
24	F	51	2b	98	90	1	1	1	1	-	2.7	1.5	0.0	-	0.6	0.8	Rsp	SVR
25	M	37	2a	241	211	1	0	1	0	-	4.0	1.5	0.0	-	1.3	0.8	Rsp	SVR
26	F	45	2a	91	33	2	1	2	1	-	5.4	2.2	1.5	-	1.6	0.4	Rsp	TR
27	M	46	2a	101	45	2	1	2	1	+	3.6	0.0	0.0	-	>1.8	-	Rsp	SVR
28	M	54	2a	196	177	3	2	2	1	+	4.2	0.0	0.0	-	>2.1	-	Rsp	SVR
29	F	68	2a	234	135	3	1	3	2	+	4.6	3.1	0.0	-	0.8	1.7	Rsp	SVR
30	M	67	2a	155	163	4	2	4	2	-	3.9	1.5	0.0	-	1.2	0.8	Rsp	SVR

First phase decline was determined by subtracting HCV-RNA at 48 h from before therapy.

Second phase decline was determined by subtracting HCV-RNA at 2 wk from 48 h.

NA, not applicable; LCM, laser capture microdissection; ALT, alanine aminotransferase; SVR, sustained viral response; A, activity; NR, nonresponse; F, fibrosis; TR, transient response; Rsp, viral responder, patients with SVR or TR; nonRsp, non-viral responder; patients with NR; HCV-RNA was assayed by COBAS® AmpliPrep/COBAS® TaqMan® System® (Log IU/mL).

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Table 2. Characteristics of patients who received Peg-IFN and ribavirin combination therapy and normal control.

Pt.No.	Sex	Age (yr)	Genotype	ALT (IU/ml)	Liver histology		HCV-RNA (Log IU/ml)			Viral response	Outcome		
					Before therapy	Before therapy		Before therapy	2 wk			4 wk	24 wk
						F	A						
1	M	57	1b	68	1	1	6.5	-	-	-	Rsp	SVR	
2	F	56	1b	31	1	1	6.5	4.4	-	-	Rsp	SVR	
3	M	63	1b	50	1	1	6.1	-	-	-	Rsp	SVR	
4	M	44	1b	45	1	1	6.5	3.7	-	-	Rsp	SVR	
5	F	51	1b	27	2	1	6.5	4.1	-	-	Rsp	SVR	
6	M	58	1b	72	2	1	6.2	-	-	-	Rsp	SVR	
7	M	60	1b	71	2	2	6.2	3.9	-	-	Rsp	SVR	
8	F	52	1b	58	2	2	6.5	4.1	-	-	Rsp	SVR	
9	F	62	1b	60	3	2	5.9	3.8	-	-	Rsp	SVR	
10	M	55	1b	106	3	2	6.4	-	-	-	Rsp	SVR	
11	M	30	1b	31	1	1	6.4	6.1	5.9	+	nonRsp	NR	
12	F	55	1b	23	1	2	6.5	6.1	5.9	+	nonRsp	NR	
13	M	58	1b	129	1	2	6.3	6.0	5.8	+	nonRsp	NR	
14	M	42	1b	326	2	1	6.6	6.2	5.8	+	nonRsp	NR	
15	F	61	1b	77	2	1	6.1	5.9	5.7	+	nonRsp	NR	
16	F	44	1b	31	2	2	5.5	5.3	4.7	+	nonRsp	NR	
17	M	51	1b	38	2	2	6.5	6.2	5.9	+	nonRsp	NR	
18	F	55	1b	97	2	2	6.7	6.3	6.1	+	nonRsp	NR	
19	M	59	1b	31	3	2	6.7	5.9	5.7	+	nonRsp	NR	
20	F	53	1b	71	3	2	5.9	5.8	5.8	+	nonRsp	NR	
21	F	51	-	18	0	0	-	-	-	-	-	-	
22	F	78	-	13	0	0	-	-	-	-	-	-	
23	M	75	-	20	0	0	-	-	-	-	-	-	
24	M	34	-	12	0	0	-	-	-	-	-	-	
25	M	64	-	30	0	0	-	-	-	-	-	-	
26	M	78	-	9	0	0	-	-	-	-	-	-	
27	M	53	-	19	0	0	-	-	-	-	-	-	
28	F	64	-	12	0	0	-	-	-	-	-	-	
29	F	60	-	20	0	0	-	-	-	-	-	-	
30	M	66	-	26	0	0	-	-	-	-	-	-	

SVR, sustained viral response; NR, nonresponse; Rsp, viral responder, patients with SVR or TR; nonRsp, non-viral responder; patients with NR.

Preparation of liver tissue samples

Liver biopsy samples were taken from all the patients at around 1 week before treatment and at 1 week after starting therapy (Fig. 1A). The biopsy samples were divided into three parts: the first part was immersed in formalin for histological assessment, the second was immediately frozen in liquid nitrogen tank for future RNA isolation, and the final part was frozen in OCT compound for LCM analysis and stored at -80°C until use. As a control, a liver tissue sample was surgically obtained from a patient who showed no clinical signs of hepatitis and was analyzed as described previously [11].

CLL and CPA were isolated by LCM using a CRI-337 (Cell Robotics, Albuquerque, NM, USA) (Supplementary Fig. 1) from the liver biopsy specimens frozen in OCT compound. The detailed procedure for LCM is described in the Supplementary materials and methods and was performed as previously described [11,19].

RNA isolation and Affymetrix gene chip analysis

Total RNA in each liver biopsy specimen was isolated using the RNAqueous[®] kit (Ambion, Austin, TX, USA). Total RNA in the specimens frozen for LCM was isolated with a carrier nucleic acid (20 ng poly C) using RNAqueous[®]-Micro (Ambion). The quality of the isolated RNA was estimated after electrophoresis using an

Agilent 2001 Bioanalyzer (Palo Alto, CA, USA). Aliquots of total RNA (50 ng) isolated from the liver biopsy specimens were subjected to amplification with the WT-Ovation[™] Pico RNA Amplification System (NuGen, San Carlos, CA, USA) as recommended by the manufacturer. About 10 μg of cDNA was amplified from 50 ng total RNA, and 5 μg of cDNA was used for fragmentation and biotin labeling using the FL-Ovation[™] cDNA Biotin Module V2 (NuGen) as recommended by the manufacturer. The biotin-labeled cDNA was suspended in 220 μl of hybridization cocktail (NuGen), and 200 μl was used for the hybridization. Half of the total RNA isolated from the LCM specimens was amplified twice with the TargetAmp[™] 2-Round Aminoallyl-aRNA Amplification Kit 1.0 (EPICENTRE, Madison, WI, USA). Twenty-five micrograms of amplified antisense RNA were used for biotin labeling according to the manufacturer's protocol *Biotin-X-X-NHS* (provided by EPICENTRE). The biotin-labeled aRNA was suspended in 300 μl of hybridization cocktail (Affymetrix Inc., Santa Clara, CA, USA), and 200 μl was used for the hybridization with the Affymetrix Human 133 Plus 2.0 microarray chip containing 54,675 probes. After stringent washing, the microarray chips were stained with streptavidin-phycoerythrin, and probe hybridization was determined using a GeneChip[®] Scanner 3000 (Affymetrix). Data files (CEL) were obtained with the GeneChip[®] Operating Software 1.4 (GCOS) (Affymetrix). All the expression data were deposited in Gene Expression Omnibus (GEO; <http://www.ncbi.nlm.nih.gov/geo/>) (NCBI) and the accession ID is GSM 425,995. The experimental procedure is described in detail in the Supplementary materials and methods.

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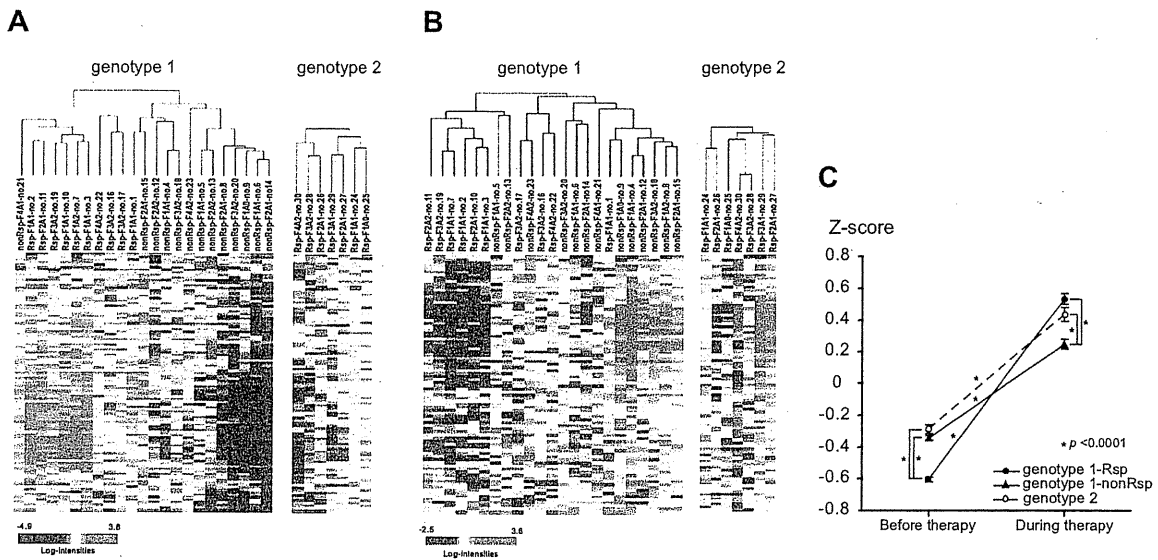


Fig. 1. (A) Hierarchical clustering of expression in genotype 1 and genotype 2 patients during treatment according to fold induction of IRSGs. (B) Hierarchical clustering of expression in genotype 1 and genotype 2 patients before treatment. (C) Serial changes in standardized expression values (Z-score) of IRSGs from genotype 1-Rsp, genotype 1-nonRsp, and genotype 2 patients before and during treatment.

Statistical and pathway analysis of gene chip data

Statistical analysis and hierarchical clustering were performed by BRB-ArrayTools (<http://linus.nci.nih.gov/BRB-ArrayTools.htm>). A class comparison tool based on univariate or paired *t*-tests was used to find differentially expressed genes ($p < 0.005$). To confirm statistical significance, 2000 random permutations were performed, and all of the *t*-tests were re-computed for each gene. The gene set comparison was analyzed using the BioCarta and the KEGG pathway data bases. The Fisher and Kolmogorov-Smirnov tests were performed for statistical evaluation ($p < 0.005$) (BRB-ArrayTools). Functional ontology enrichment analysis was performed to compare the Gene Ontology (GO) process distribution of differentially expressed genes ($p < 0.05$) using MetaCore™ (GeneGo, St. Joseph, MI, USA).

For the comparison of standardized expression values among different pathway groups, standard units (Z-score) of each gene expression value were calculated as:

$$Z_i = \frac{X_i - X_m}{S}$$

where X_i is the raw expression value, X_m is the mean of the expression values in the pathway, and S is the standard deviation of the expression values.

The standard units in each pathway were expressed as mean \pm SEM. A *P*-value of less than 0.05 was considered significant. Multivariate analysis was performed using a logistic regression model with a stepwise method using JMP7 for Windows (SAS Institute, Cary, NC, USA).

Quantitative real-time detection (RTD)-PCR

We performed quantitative real-time detection PCR (RTD)-PCR using TaqMan Universal Master Mix (PE Applied Biosystems, CA). Primer pairs and probes for Mx1, IFI44 and IFTM1, and GAPDH were obtained from TaqMan assay reagents library (Applied Biosystems, CA).

Results

Serial changes in HCV-RNA after initiation of IFN- α 2b and Rib combination therapy

Serial changes in HCV-RNA were monitored at 48 h, 2 weeks, and 24 weeks after the initiation of therapy (Table 1). The biphasic

viral decline after the initiation of IFN therapy has been characterized [14,15,18]. We calculated the first phase decline by comparing viral load before therapy and after 48 h, and the second phase decline by comparing viral load after 48 h and 2 weeks (Table 1) [14,15,18]. Both the first and the second phase declines could be associated with treatment outcome and interestingly, viral responders (Rsp) who achieved SVR or TR showed more than a 1-log drop of first phase decline (Log/24 h) and more than a 0.3-log drop of second phase decline (Log/w) (Table 1). In contrast, non-responders (nonRsp) who exhibited NR failed to meet the criteria. The first phase decline of Rsp and nonRsp were 1.38 ± 0.65 log/24 h and 0.77 ± 0.44 log/24 h ($p = 0.005$), respectively. The second phase decline of Rsp and nonRsp were 0.71 ± 0.34 log/w and 0.11 ± 0.34 log/w ($p = 0.0001$), respectively. Therefore, the classification of Rsp or nonRsp according to the treatment outcome might be feasible based on the viral kinetic responses to IFN. All but one patient infected with genotype 2 HCV eliminated the virus within 2 weeks. There were no significant differences in the degree of histological activity or staging, nor in the sex, age, or alanine aminotransferase (ALT) level among these patients (Table 1). The amount of HCV-RNA was significantly lower in genotype 2 patients (4.06 ± 0.32 log IU/ml) than in genotype 1 patients (5.70 ± 1.10 log IU/ml) (Table 1).

Identification of IFN- α 2b plus Rib-induced genes in the livers of patients with chronic hepatitis C infection

To identify the genes induced in the liver by combination treatment with IFN- α 2b plus Rib, the gene expression profiles from samples taken around 1 week before and 1 week after initiation of therapy were compared. The pairwise *t*-test comparison showed that 798 genes were up-regulated and 220 genes were down-regulated significantly ($p < 0.005$). The 100 most up-regulated genes according to *p* values were selected; these are listed in Supplementary Table 1. Many of the interferon-stimulated