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Pegylated interferon alpha-2b (Peg-IFN α-2b) affects early virologic response dose-dependently in patients with chronic hepatitis C genotype 1 during treatment with Peg-IFN α-2b plus ribavirin

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SUMMARY. Chronic hepatitis C (CH-C) genotype 1 patients who achieved early virologic response have a high probability of sustained virologic response (SVR) following pegylated interferon (Peg-IFN) plus ribavirin therapy. This study was conducted to evaluate how reducing drug doses affects complete early virologic response (c-EVR) defined as hepatitis C virus (HCV) RNA negativity at week 12. Nine hundred eighty-four patients with CH-C genotype 1 were enrolled. Drug doses were evaluated independently on a body weight base from doses actually taken. From multivariate analysis, the mean dose of Peg-IFN α-2b during the first 12 weeks was the independent factor for c-EVR ($P = 0.02$), not ribavirin. The c-EVR rate was 55% in patients receiving ≥1.2 μg/kg/week of Peg-IFN, and declined to 38% at 0.9–1.2 μg/kg/week, and 22% in patients given <0.9 μg/kg/week ($P < 0.0001$). Even with stratified analysis according to ribavirin dose, the dose-dependent effect of Peg-IFN on c-EVR was observed, and similar c-EVR rates were obtained if the dose categories of Peg-IFN were the same. Furthermore, the mean dose of Peg-IFN during the first 12 weeks affected HCV RNA negativity at week 24 ($P < 0.0001$) and SVR ($P < 0.0001$) in a dose-dependent manner. Our results suggest that Peg-IFN was dose-dependently correlated with c-EVR, independently of ribavirin dose. Thus, maintaining the Peg-IFN dose as high as possible during the first 12 weeks can yield HCV RNA negativity and higher c-EVR rates, leading to better SVR rates in patients with CH-C genotype 1.

Keywords: chronic hepatitis C, drug dose, early virologic response, HCV RNA negativity, pegylated interferon plus ribavirin, sustained virologic response.

Abbreviations: c-EVR, complete EVR; CH-C, chronic hepatitis C; EVR, early virologic response; G-CSF, granulocyte-macrophage colony stimulating factor; Hb, haemoglobin; HCV, hepatitis C virus; Peg-IFN, pegylated interferon; Plt, platelet; SVR, sustained virologic response; WBC, white blood cell.

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INTRODUCTION

Pegylated interferon (Peg-IFN) plus ribavirin therapy can improve anti-viral efficacy for patients with chronic hepatitis C [1–5], and the prognosis of patients in whom hepatitis C virus (HCV) is successfully eradicated improves markedly [6–10]. However, HCV still persists in approximately half of genotype 1 patients treated with Peg-IFN plus ribavirin [2–4]. Therefore, the treatment method needs to be well managed in order to maximize the virologic response in these patients with HCV genotype 1.

In order to achieve sustained virologic response (SVR), earlier virologic response is very important for patients with chronic hepatitis C (CH-C) genotype 1. A high SVR rate (65–72%) was found in patients who achieved early virologic response (EVR) defined as a 2-log decrease in HCV RNA level at week 12, but only 0–3% SVR was seen in patients without EVR [3,11]. Additionally, complete EVR (c-EVR), which means HCV RNA negativity at week 12, is more strongly related to SVR [3].

The relationship between drug exposure and anti-viral effect has been reported in several papers [2,11–15]. McHutchison *et al.* [12] demonstrated that the SVR rate in patients who received ≥80% of their total planned doses of Peg-IFN and ribavirin for ≥80% of the scheduled duration of therapy was significantly higher than that of patients who received <80% of one or both drugs (51% *vs* 34%) and also suggested that the impact of dose reduction was greatest in patients for whom the dose had to be decreased within the first 12 weeks of treatment. In a subsequent analysis, reducing the dose of Peg-IFN and ribavirin to <80% of the full planned dose within the first 12 weeks was reported to reduce EVR rate from 80 to 33% [11]. Thus, drug adherence during the first 12 weeks has been shown to be very important for attaining EVR and SVR, but it remains obscure whether either drug can be reduced to a certain degree without adversely affecting the treatment efficacy.

In the present study, we examined the correlation between c-EVR and drug doses which are evaluated on a body weight basis from drug doses actually taken, in order to clarify the necessary drug exposure of Peg-IFN and ribavirin for achieving a higher c-EVR rate in patients with CH-C genotype 1.

PATIENTS AND METHODS

Patients

The current study was a retrospective, multicenter trial conducted by Osaka University Hospital and other institutions participating in the Osaka Liver Forum. A total of 984 patients with CH-C treated with a combination of Peg-IFN α-2b plus ribavirin were enrolled in this study between December 2004 and September 2006. The baseline characteristics of the patients are summarized in Table 1. All patients were Japanese, their mean age was 56.3 ± 10.1 years, and 56% were males. The mean serum alanine aminotransferase level was 79 ± 61 IU/L.

Patients eligible for this study were those who were infected with HCV genotype 1 and had a viral load of more than 10^5 IU/mL, but were negative for hepatitis B surface antigen or anti-human immunodeficiency virus. Patients were excluded from this study if they had decompensated cirrhosis or other forms of liver disease (alcohol liver disease, autoimmune hepatitis). Informed consent was obtained from each patient included in this study. This study was conducted according to the ethical guidelines of the 1975 Declaration of Helsinki and informed consent was obtained from each patient.

Table 1 Baseline characteristics of patients

Factor	Mean ± SD or number
n	984
Age (year)	56.3 ± 10.1
Sex: male/female	555/429
Body weight (kg)	61.8 ± 11.5
History of interferon treatment	
Naïve/experienced (relapser/nonresponder)*	575/409(160/182)
White blood cells (per mm^3)	5052 ± 1550
Neutrophils (per mm^3)	2577 ± 1092
Red blood cells ($\times 10^4/mm^3$)	442 ± 47
Haemoglobin (g/dL)	14.1 ± 1.4
Platelets ($\times 10^4/mm^3$)	15.9 ± 5.5
AST (IU/L)	66 ± 45
ALT (IU/L)	79 ± 61
Serum HCV RNA (kIU/mL)†	1600
Histology (METAVIR)‡	
Fibrosis; 0/1/2/3/4	49/314/197/105/18
Activity; 0/1/2/3	23/329/304/27

AST, aspartate aminotransferase; ALT, alanine aminotransferase; HCV, hepatitis C virus.
*Viral response to previous treatment was unknown in 57 patients, and 10 patients had discontinued treatment. †Data shown are median values. ‡301 missing.

Treatment

All patients received Peg-IFN α-2b (PEGINTRON; Schering-Plough, Kenilworth, NJ, USA) plus ribavirin (REBETOL; Schering-Plough) for the duration of the study of 48 weeks. Peg-IFN α-2b was given subcutaneously once weekly at a dosage of 60–150 μg/kg based on body weight (body weight 35–45 kg, 60 μg; 46–60 kg, 80 μg; 61–75 kg, 100 μg; 76–90 kg, 120 μg; 91–120 kg, 150 μg) and ribavirin was given orally twice a day at a total dose of 600–1000 mg/day based on body weight (body weight ≤60 kg, 600 mg; 60–80 kg, 800 mg; >80 kg, 1000 mg), according to a standard treatment protocol for Japanese patients.

Dose reduction

Dose modification followed, as a rule, the manufacturer's drug information according to the intensity of the haematological adverse effects. The dose of Peg-IFN α-2b was reduced to 50% of the assigned dose if the white blood cell (WBC) count declined to <1500/mm^3, the neutrophil count to <750/mm^3 or the platelet (Plt) count to <8 × $10^4/mm^3$, and was discontinued if the WBC count declined to <1000/

mm^3, the neutrophil count to $<500/mm^3$ or the Plt count to $<5 \times 10^4/mm^3$. Ribavirin was also reduced from 1000 to 600 mg, or 800 to 600 mg, or 600 to 400 mg if the haemoglobin (Hb) level decreased to <10 g/dL, and was discontinued if the Hb level decreased to <8.5 g/dL. Both Peg-IFN α-2b and ribavirin had to be discontinued if there was a need to discontinue one of the drugs. During this therapy, ferric medicine or haematopoetic growth factors, such as erythropoietin alpha, or granulocyte-macrophage colony stimulating factor (G-CSF), were not administered.

Virologic assessment and definition of virologic response

Serum HCV RNA level was quantified using the COBAS AMPLICOR HCV MONITOR test, version 2.0 (detection range 6–5000 kIU/mL; Roche Diagnotics, Branchburg, NJ, USA) and qualitatively analysed using the COBAS AMPLICOR HCV test, version 2.0 (lower limit of detection 50 IU/mL). The c-EVR was defined as the absence of detectable serum HCV RNA at treatment week 12, and SVR was defined as the absence of detectable serum HCV RNA at week 72. Patients with less than a 2-log decrease in HCV RNA level at treatment week 12 compared with the baseline had to stop treatment and were regarded as nonresponders. All patients with detectable serum HCV RNA at treatment week 24 were also considered nonresponders and excluded from further treatment.

Assessment of drug exposure

The amounts of Peg-IFN α-2b and ribavirin actually taken by each patient during the first 12 weeks of the treatment were evaluated by reviewing the medical records. The mean doses of both drugs were calculated individually as averages on the basis of body weight at baseline: Peg-IFN α-2b expressed as μg/kg/week, and ribavirin expressed as mg/kg/day.

Evaluation of impact of drug exposure on c-EVR

We evaluated the relationship between the drug exposure of both drugs and c-EVR by univariate and multivariate analysis for c-EVR, using the factors of mean administration doses of both drugs during the first 12 weeks and the factors at baseline. Furthermore, Peg-IFN α-2b dose (average dose per body weight and per week) was classified into five categories (up to 0.6 μg/kg; from 0.6 to <0.9 μg/kg; from 0.9 to <1.2 μg/kg; from 1.2 to <1.5 μg/kg; from 1.5 μg/kg and above). Ribavirin exposure was classified into four categories (up to 8 mg/kg; from 8 to <10 mg/kg; from 10 to <12 mg/kg; from 12 mg/kg and above), in order to examine the impact of Peg-IFN dose exposure on c-EVR. This impact was also evaluated based on the percentage of the total prescribed dose and compared with that based on the mean dose per body weight.

Statistical analysis

Baseline data for various demographic, biochemical and virologic characteristics of the patients are expressed as mean ± SD or median values. To analyse the relationship between baseline data including drug exposure and c-EVR, univariate analysis using the Mann–Whitney *U*-test or chi-squared test and multivariate analysis using logistic regression analysis were performed. The significance of trends in values was determined with the Mantel–Haenszel chi-square test. A two-tailed *P*-value < 0.05 was considered significant. Statistical analysis was conducted with SPSS version 15.0J (SPSS Inc., Chicago, IL, USA).

RESULTS

Progress of patients treated with Peg-IFN α-2b and ribavirin

Of the 984 patients, 81 discontinued treatment because of adverse events (n = 74) or voluntary withdrawal (n = 7) by treatment week 12. The 903 patients who completed 12 weeks of treatment were assessed for c-EVR. During 12–48 weeks of treatment, 331 of the nonresponders and nine of breakthrough discontinued treatment, as did 91 patients (adverse events, n = 71; voluntary withdrawal, n = 20). A total of 472 patients completed 48 weeks of treatment.

Drug reduction and virologic response

Peg-IFN α-2b was reduced without discontinuation in 29% (n = 266) and ribavirin was reduced without discontinuation in 40% (n = 359) of the 903 patients who completed 12 weeks of treatment. The c-EVR rate was 49% (445/903) and HCV RNA was negative at week 24 in 60% (542/903) of patients who completed 12 weeks of treatment. Of the 445 patients with c-EVR, 327 patients achieved SVR (73%). Only 7% of the 458 patients without c-EVR did so.

Impact of dose exposure of Peg-IFN α-2b and ribavirin on c-EVR

The mean dose of Peg-IFN α-2b actually taken during the first 12 weeks by each patient was 1.33 μg/kg/week (range 0.41–2.16 μg/kg/week; median 1.40 μg/kg/week) and that of ribavirin was 10.4 mg/kg/day (range 2.9–16.2 mg/kg/day; median 10.6 mg/kg/day).

The mean doses of both drugs and the factors at baseline correlated with the c-EVR were assessed by univariate and multivariate logistic regression analyses. Univariate analysis showed that factors significantly associated with c-EVR were age, sex, WBC, neutrophils, red blood cells, Hb, Plt, aspartate aminotransferase, the degree of liver fibrosis and the mean doses of Peg-IFN α-2b and ribavirin during the first 12 weeks (Table 2). The factors selected as significant by the univari-

Table 2 Univariate analysis for c-EVR among patients who completed 12 weeks treatment

Factor	c-EVR (+)	c-EVR (−)	*P*-value
n	445	458	
Age (year)	54.4 ± 10.4	57.5 ± 9.6	<0.001
Sex: male/female	267/178	237/221	0.01
Serum HCV RNA (kIU/mL)*	1500	1600	0.28
White blood cells (per mm^3)	5336 ± 1536	4818 ± 1547	<0.001
Neutrophils (per mm^3)	2789 ± 1133	2398 ± 1038	<0.001
Red blood cells ($\times 10^4/mm^3$)	450 ± 46	435 ± 49	<0.001
Haemoglobin (g/dL)	14.3 ± 1.4	13.9 ± 1.4	<0.001
Platelets ($\times 10^4/mm^3$)	17.3 ± 5.2	15.0 ± 5.6	<0.001
AST (IU/L)	62 ± 44	69 ± 44	<0.001
ALT (IU/L)	77 ± 64	80 ± 57	0.07
Histology (METAVIR)†			
Fibrosis: 0–2/3–4	273/37	247/74	<0.001
Activity: 0–1/2–3	171/139	159/162	0.16
Peg-IFN dose (μg/kg/week)‡	1.39 ± 0.22	1.28 ± 0.30	<0.001
Ribavirin dose (mg/kg/day)‡	10.6 ± 1.7	10.1 ± 2.1	0.002

c-EVR, complete early virologic response; HCV, hepatitis C virus; AST, aspartate aminotransferase; ALT, alanine aminotransferase; Peg-IFN, pegylated interferon.
*Data shown are median values. †272 missing. ‡Mean doses during 0–12 weeks.

Table 3 Multivariate analysis for c-EVR among patients who completed 12 weeks treatment

Factor	Category	Odds ratio	95% CI	*P*-value
Age	by 1 year	0.982	0.966–0.999	0.04
Sex	male/female	–	–	NS
Neutrophils	by 100/mm^3	1.017	1.002–1.033	0.03
Red blood cells	by $1 \times 10^4/mm^3$	–	–	NS
Haemoglobin	by 1 g/dL	–	–	NS
Platelets	by $1 \times 10^4/mm^3$	1.051	1.014–1.088	<0.01
AST	by 1 IU/L	–	–	NS
Fibrosis*	0–2/3–4	–	–	NS
Peg-IFN dose†	by 0.1 μg/kg/week	1.079	1.011–1.151	0.02
Ribavirin dose†	by 1 mg/kg/day	–	–	NS

95% CI, 95% confidence interval; Peg-IFN, c-EVR, complete early virologic response; pegylated interferon; N.S., No Significant difference; AST, aspartate aminotransferase.
*METAVIR fibrosis score. †Mean doses during 0–12 weeks.

ate analysis were evaluated by multivariate logistic regression analysis. The mean dose of Peg-IFN α-2b during the first 12 weeks was the independent factor for c-EVR ($P = 0.02$), apart from the neutrophils ($P = 0.03$) and Plt value at baseline ($P < 0.01$) and age ($P = 0.04$) (Table 3). In contrast, the mean dose of ribavirin during the first 12 weeks showed no correlation with c-EVR.

The c-EVR rates were 54% (137/253) and 56% (246/443) for patients who received ≥1.5 and 1.2–1.5 μg/kg/week of Peg-IFN α-2b on average during the first 12 weeks, and declined to an average rate of 38% (40/105) in patients given 0.9–1.2 μg/kg/week of Peg-IFN α-2b, and an average rate of 22% (22/102) in patients given <0.9 μg/kg/week ($P < 0.0001$) (Table 4). The c-EVR rate among the patients with ≥1.2 μg/kg/week of Peg-IFN α-2b was significantly higher than that of the patients with <1.2 μg/kg/week [≥1.2 μg/kg/week, 55% (383/696) *vs* <1.2 μg/kg/week, 30% (62/207), $P < 0.0001$].

Next, we analysed the impact of Peg-IFN α-2b on c-EVR in stratified analysis according to ribavirin dose. Figure 1 shows the relationship of c-EVR and the degree of Peg-IFN α-2b exposure for two groups of ribavirin doses: the group with ≥10.6 mg/kg/day of ribavirin and that with <10.6 mg/kg/day (10.6 mg/kg/day was the median value). In either group, the mean dose of Peg-IFN α-2b was dose-dependently correlated with c-EVR ($P < 0.0001$), and c-EVR rates were very similar in both groups if the dose categories of Peg-IFN α-2b were the same.

Table 4 The c-EVR rate according to Peg-IFN and ribavirin doses during weeks 0–12 for patients who completed 12 weeks treatment

Ribavirin dose (mg/kg/day)**	Peg-IFN α-2b dose (μg/kg/week),*				
	≥1.5	1.2–1.5	0.9–1.2	<0.9	Total
≥12	57% (60/105)	61% (22/36)	38% (6/16)	22% (2/9)	54% (90/166)
10–12	54% (46/85)	58% (154/267)	36% (14/39)	23% (11/47)	51% (225/438)
8–10	50% (25/50)	53% (52/99)	52% (15/29)	18% (4/22)	48% (96/200)
<8	46% (6/13)	44% (18/41)	24% (5/21)	21% (5/24)	34% (34/99)
Total	54% (137/253)	56% (246/443)	38% (40/105)	22% (22/102)	49% (445/903)

c-EVR, complete early virologic response; Peg-IFN, pegylated interferon.
*$P < 0.0001$ for comparison of the four Peg-IFN groups. **$P = 0.05$ for comparison of the four ribavirin groups.

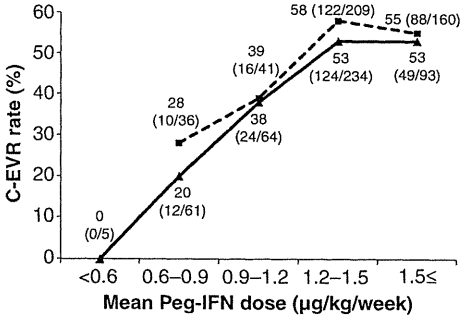

Fig. 1 Complete-EVR rate according to pegylated interferon alpha-2b (Peg-IFN α-2b) and ribavirin doses during weeks 0–12 for patients who completed 12 weeks of treatment. (-▲-) Group with the mean ribavirin dose <10.6 mg/kg/day. (··■··) Group with the mean ribavirin dose ≥10.6 mg/kg/day. The Peg-IFN α-2b dose was dose-dependently correlated with c-EVR in both groups ($P < 0.0001$). There was no significant difference between the two ribavirin-dose groups ($P = 0.19$).

c-EVR rates according to Peg-IFN α-2b drug exposure using a percentage cut off and mean dose cut off

Table 5 shows the c-EVR rates according to the category of Peg-IFN α-2b doses during the first 12 weeks based on the percentage of the total prescribed dose and the mean doses. The whole c-EVR rate was 54% (377/698) for patients who received more than 80% of the prescribed dose, and 43% (47/109) in patients given 60–80% of the prescribed dose, and 21% (21/96) in patients given <60% of the prescribed dose of Peg-IFN α-2b. Among patients given ≥80% of the prescribed dose of Peg-IFN α-2b, the c-EVR rate was significantly lower in patients given <1.2 μg/kg/week of Peg-IFN α-2b than those given ≥1.2 μg/kg/week (32% *vs* 55%, $P < 0.05$). On the other hand, even in patients given 60–80% of the prescribed dose of Peg-IFN α-2b, if they were given ≥1.2 μg/kg/week of Peg-IFN α-2b, a higher c-EVR rate was attained in comparison with those given <1.2 μg/kg/week (71% *vs* 38%, $P = 0.01$); the c-EVR rate in patients given 60–80% of the prescribed dose and ≥1.2 μg/kg/week of Peg-IFN α-2b was not inferior to that in patients given ≥80% of the prescribed dose and ≥1.2 μg/kg/week of Peg-IFN α-2b.

Impact of dose exposure of Peg-IFN α-2b during the first 12 weeks of the treatment on HCV RNA negativity at week 24 and SVR

Patients positive for HCV RNA at week 24 week during Peg-IFN α-2b and ribavirin treatment were regarded as non-responders and stopped treatment [11]. We analysed the

Table 5 The c-EVR rate according to Peg-IFN dose during weeks 0–12 based on the percentage of the planned dose and the mean doses

Peg-IFN α-2b dose (μg/kg/week)	≥80%	60–80%	<60%	Total
≥1.2	55%* (371/679)	71%** (12/17)	–	55% (383/696)
<1.2	32% (6/19)	38% (35/92)	22% (21/96)	30% (62/207)
Total	54% (377/698)	43% (47/109)	21% (21/96)	49% (445/903)

c-EVR, complete early virologic response; Peg-IFN, pegylated interferon.
*$P < 0.05$; patients with ≥1.2 μg/kg/week *vs* <1.2 μg/kg/week among the patients with more than 80% of the total prescribed dose of Peg-IFN α-2b. **$P = 0.01$; patients with ≥1.2 μg/kg/week *vs* <1.2 μg/kg/week among the patients with more than 60–80% of the total prescribed dose of Peg-IFN α-2b.

relationship between the dose exposure to Peg-IFN α-2b during the first 12 weeks and HCV RNA negative rates at week 24 or SVR in 903 patients completing 12 weeks of treatment. As a result, HCV RNA negative rates at week 24 and SVR rates declined according to the decrease in the dose of Peg-IFN α-2b during the 12 weeks of treatment; patients given ≥1.5, 1.2–1.5, 0.9–1.2 and <0.9 μg/kg/week of Peg-IFN α-2b during the first 12 weeks of the treatment showed HCV RNA negativity of 63%, 66%, 48% and 39%, respectively ($P < 0.0001$), and SVR of 46%, 43%, 30% and 20%, respectively ($P < 0.0001$).

DISCUSSION

Adherence to ribavirin was reported to be the important factor for EVR as well as that to Peg-IFN in most previous studies [2,11,12]. However, the drug exposure of Peg-IFN α-2b and ribavirin had not been analysed independently with respect to their individual influence on the anti-viral effect in these studies. Adherence to both drugs may be related factors, i.e. most patients who can tolerate a high dose of Peg-IFN are in good condition and thus can also receive a high dose of ribavirin. In the present study, the impact of the dose of Peg-IFN α-2b and ribavirin on the anti-viral effect was evaluated by multivariate logistic regression analysis, using the mean administration doses of both drugs during the first 12 weeks and baseline factors. As a result, the dose exposure of Peg-IFN α-2b was found to be the significant factor affecting c-EVR as well as baseline factors such as age, neutrophils and Plt values, but not ribavirin. This suggests that the c-EVR rate can be raised by maintaining the dose of Peg-IFN α-2b during the first 12 weeks in patients with disadvantageous factors at baseline. In fact, the c-EVR rate was higher in those who received ≥1.2 μg/kg of Peg-IFN α-2b than in those given <1.2 μg/kg of Peg-IFN α-2b for aged patients over 60 years of age (≥1.2 μg/kg; 46% *vs* <1.2 μg/kg; 28%, $P < 0.01$) or for patients with a low Plt value ($<12 \times 10^4/mm^3$)(≥1.2 μg/kg; 45% *vs* <1.2 μg/kg; 22%, $P < 0.001$). Therefore, a marked dose reduction of Peg-IFN α-2b should not be risked at the start even for aged patients or patients with lower Plt value, which is indicative of advanced fibrosis. The administration of ≥1.2 μg/kg/week of Peg-IFN α-2b is desirable as a starting dose for achieving c-EVR even in these patients: that of <1.2 μg/kg/week can lead to a non-viral response or a late viral response. Independent evaluation of the c-EVR rate according to the degree of the ribavirin dose showed a stepwise decline as the total cumulative dose of Peg-IFN α-2b decreased. Therefore, the dose of Peg-IFN α-2b should be maintained as high as possible even in patients who have to reduce Peg-IFN α-2b to <1.2 μg/kg/week. Using G-CSF for patients who develop severe neutropenia and are forced to decrease Peg-IFN can be beneficial, especially in the first 12 weeks.

The goal of 80% of the planned drug dosage for 80% of the assigned duration was derived from an adherence criterion that had been adopted previously for assessment of the efficacy of other pharmaceutical agents, such as drugs to treat cancer and human immunodeficiency virus [16]. However, in Peg-IFN plus ribavirin therapy for patients with CH-C, the planned administration dose [17,18] differs on a body weight basis by 27% for Peg-IFN α-2b and 40% for ribavirin among patients of 50–100 kg of body weight, which would be equivalent to the same rate differences for 80% of the planned drug dosage. In detail, the target dose of Peg-IFN α-2b scheduled to be administered is 1.5 μg/kg, but the usual dose for the individual patient is from 1.28 to 1.76 μg/kg/week based on body weight among patients weighing 50–100 kg according to the practice guidelines of the American Association for the Study of Liver Diseases and the manufacturer's drug information in the USA and Europe [17,18]. The range of ribavirin dose per kg of body weight is from 12 to 20 mg/kg/day. Therefore, in this study, the drug exposure was assessed from the average dose per kg of body weight.

In the evaluation of c-EVR rates according to Peg-IFN α-2b drug exposure using a percentage cut off and mean dose cut off in this study, the c-EVR rate of patients given <1.2 μg/kg/week of Peg-IFN α-2b was low (32%) even in those who received ≥80% of the total planned doses of Peg-IFN α-2b. If given ≥1.2 μg/kg/week of Peg-IFN α-2b, the c-EVR rate (71%) in patients who received 60–80% of the total doses was not inferior to that in patients given ≥80% of the total dose of Peg-IFN α-2b (54%). This means that patients whose starting dose of Peg-IFN α-2b is <1.5 μg/kg/week should not have their dosage reduced to 80% of the planned dose (<1.2 μg/kg/week) in order to have a higher probability of c-EVR, while those given ≥1.5 μg/kg/week of Peg-IFN α-2b at the start can have their dosage reduced to 80% (≥1.2 μg/kg/week) without lowering the c-EVR rate. Thus, the drug dose on a body weight basis itself should be examined as an index of the drug exposure in order to evaluate the anti-viral effect of both drugs accurately for patients with CH-C.

As for the impact of the drug exposure to ribavirin on c-EVR, the drug dose of ribavirin during the first 12 weeks was shown to have no relationship with the c-EVR rate, although it was precisely evaluated in this study, using doses actually taken on body weight. However, ribavirin can be more effective for decreasing the viral relapse after interferon or Peg-IFN α-2b and ribavirin combination therapy in patients with CH-C genotype 1 [2,3,19–24]. Recently, Shiffman *et al.* [15] have reported that a higher starting dose of ribavirin (1000–1600 mg/day) plus a regular dose of Peg-IFN α-2b with epoetin was associated with a lower relapse rate in treatment with CH-C genotype 1. Considering the viral relapse after treatment, it is thought that the ribavirin dose should not be reduced quickly in patients with mild side effects, even though it does not affect c-EVR. In fact, among the patients who attained c-EVR, a higher rate of viral relapse was found in the patients given <10 mg/kg/day of the mean ribavirin dose during 48 weeks in comparison

with those given ≥10 mg/kg/day of the mean ribavirin dose in this study [26.9% (49/182) *vs* 12.4% (26/209), $P < 0.001$] (data not shown). It seems possible to start ribavirin at a lower dose and increase it by degrees with monitoring of Hb level during treatment of patients with mild anaemia or ischemic heart disease, because the ribavirin dose appears to affect the viral relapse as the total dose over 48 weeks, not during the first 12 weeks.

In conclusion, our results have demonstrated that Peg-IFN α-2b is dose-dependently correlated with c-EVR and maintaining as high a drug dose of Peg-IFN α-2b as possible (≥1.2 μg/kg/week) during the first 12 weeks can yield higher c-EVR rates, leading to better treatment outcomes for patients with CH-C genotype 1.

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Anticancer Chemotherapy Inhibits MHC Class I–Related Chain A Ectodomain Shedding by Downregulating ADAM10 Expression in Hepatocellular Carcinoma

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Abstract

MHC class I–related chain A (MICA) is a ligand for the NKG2D-activating immunoreceptor that mediates activation of natural killer (NK) cells. The ectodomain of MICA is shed from tumor cells, which may be an important means of evading antitumor immunity. We previously reported that patients with hepatocellular carcinoma (HCC) display high levels of soluble MICA in circulation, which could be downregulated by chemotherapy. The present study shows that anti-HCC drugs suppress MICA ectodomain shedding by inhibiting expression of a disintegrin and metalloproteinase 10 (ADAM10). Both ADAM10 and CD44, a typical substrate of the ADAM10 protease, were expressed in human HCC tissues and HCC cells but not in normal liver tissues or cultured hepatocytes. Small interfering RNA–mediated knockdown experiments revealed that ADAM10 is a critical sheddase for both MICA and CD44 in HCC cells. Of interest is the finding that epirubicin clearly downregulated ADAM10 expression and MICA shedding in HCC cells; its suppressive effect on MICA shedding was abolished in ADAM10-depleted cells. Epirubicin treatment also enhanced the NKG2D-mediated NK sensitivity of HCC cells. Patients with HCC had significantly higher levels of serum-soluble CD44, which correlated well with serum-soluble MICA levels, thus suggesting a close link between ADAM10 activity and MICA shedding in these patients. Soluble MICA and CD44 levels were downregulated with a significant correlation in patients treated by transarterial chemoembolization using epirubicin. In conclusion, anticancer drugs can modulate expression of ADAM10, which is critically involved in MICA ectodomain shedding. Epirubicin therapy may have a previously unrecognized effect on antitumor immunity in HCC patients. [Cancer Res 2009;69(20):8050–7]

Introduction

Hepatocellular carcinoma (HCC) is one of the leading causes of cancer deaths worldwide. Chronic liver disease caused by hepatitis virus infection and nonalcoholic steatohepatitis leads to a predisposition for HCC, with liver cirrhosis, in particular, being considered a premalignant condition (1, 2). With regard to treatment, surgical resection or percutaneous techniques such as ethanol injection and radiofrequency ablation are considered to be choices for curable treatment of localized HCC, whereas transcatheter arterial chemoembolization (TACE) is a well-established technique for more advanced HCC (3). The liver contains a large compartment of innate immune cells [natural killer (NK) cells and natural killer T cells] and acquired immune cells (T cells; refs. 4, 5), but the activation of these immune cells after HCC treatments remains unclear. If such treatments can efficiently activate abundant immune cells in the liver, this could lead to the establishment of attractive new strategies for HCC treatment.

MHC class I–related chain A and B (MICA and MICB) are ligands for NKG2D expressed on a variety of immune cells (6). In contrast to classic MHC class I molecules, MICA/B are rarely expressed on normal cells but frequently on tumor cells (7–10). The engagement of MICA/B and NKG2D strongly activates NK cells and costimulates T cells, enhancing their cytolytic activity and cytokine production (11). Thus, the MICA/B-NKG2D pathway is an important mechanism by which the host immune system recognizes and kills transformed cells (12). In addition to those membrane-bound forms, MICA/B molecules are also cleaved proteolytically from tumor cells and appear as soluble forms in sera of patients with malignancy (13–15). Soluble MICA/B in circulation downregulates NKG2D expression and disturbs NKG2D-mediated antitumor immunity (9, 10, 13). We previously reported that soluble MICA could be detected in sera of HCC patients (16) and that TACE treatment reduces the levels of soluble MICA and thereby upregulates the expression of NKG2D (17). Thus, cancer therapy may have a beneficial effect on NKG2D-mediated immune responses.

The release of soluble MICA/B from tumor cells is impaired by metalloproteinase inhibitors, suggesting the involvement of members of the metzincin superfamily, such as ADAM proteins (14, 18). In addition, ERp5, related to protein disulfide isomerase, is required for the MICA shedding as it reduces disulfide bond of the α3 domain of MICA (19). Although it may not be a direct protease for MICA, it may enable proteolytic cleavage through conformational change. Recently, it was reported that MICA shedding of 293T fibroblast cells and HeLa cervical cancer cells was inhibited by silencing of the ADAM10 and ADAM17 proteases (20). This suggests that ADAM family proteins may be a therapeutic target for enhancing antitumor immunity, but how to therapeutically modulate these proteins is still not clear. Furthermore, it remains to be determined whether ADAMs can regulate MICA shedding in a clinical setting.

In the present study, we showed that ADAM10, but not ADAM17, was critically required for MICA shedding in human HCC cells. Of importance is the discovery that epirubicin, a widely used anti-HCC drug, was capable of downregulating ADAM10 expression and

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Note: Supplementary data for this article are available at Cancer Research Online (http://cancerres.aacrjournals.org/).

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activity in HCC cells; it can thus inhibit MICA shedding and enhance NK sensitivity. ADAM10 was immunohistochemically detected in HCC tissues and a correlation was observed between soluble MICA levels and ADAM10 activity determined by soluble CD44 levels in HCC patients. The present study sheds light on previously unrecognized effects of an anticancer drug on modulating ADAM family proteins and MICA shedding and thus suggests a promising aspect for chemoimmunotherapy against human HCC.

Materials and Methods

Liver tissues and immunohistochemistry. Human HCC tissues (n = 8) and normal liver tissues (n = 2) obtained at surgical resection were used. Informed consent, under an institutional review board–approved protocol,

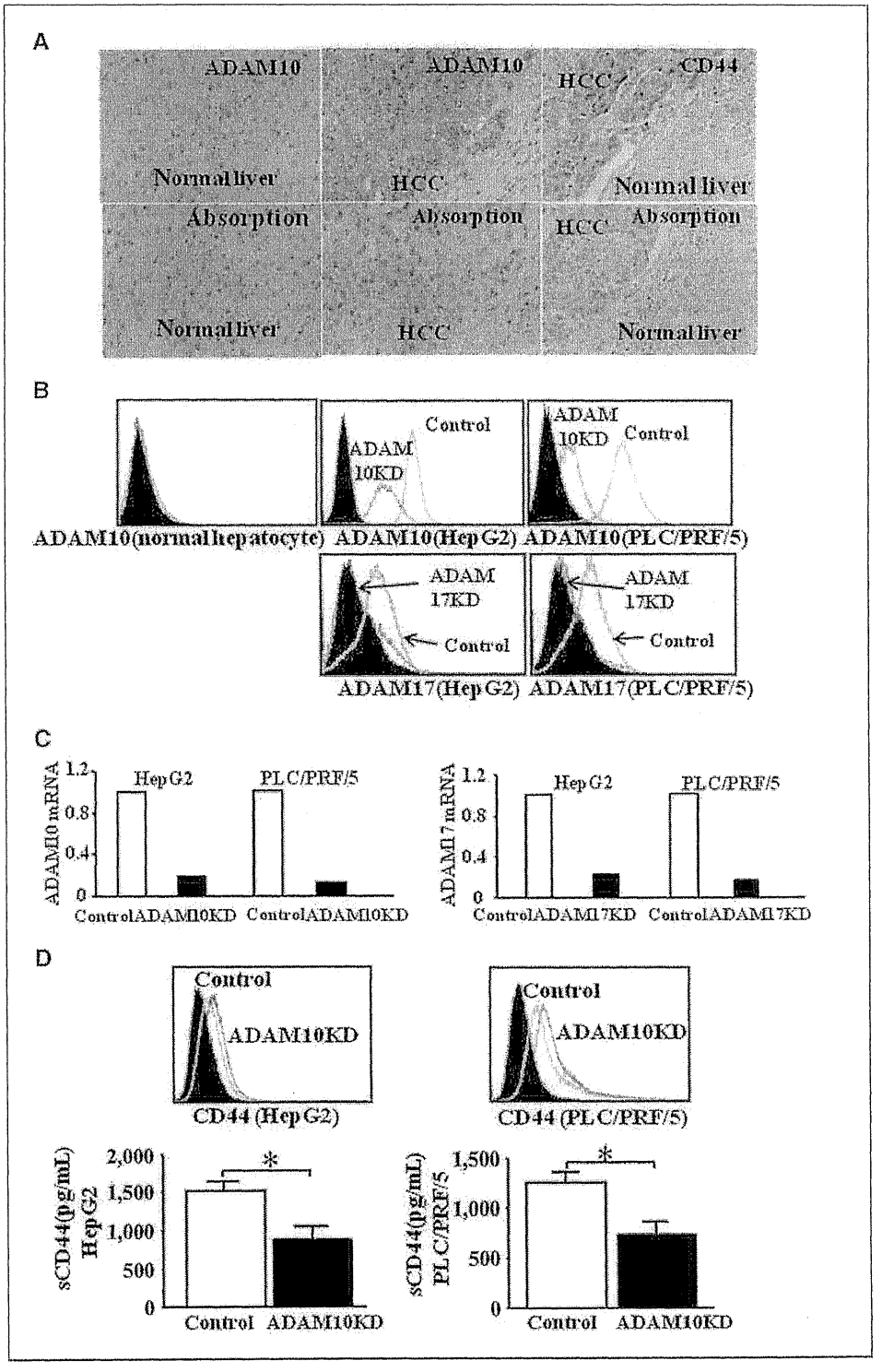

Figure 1. Expression of ADAM10 and CD44 in human HCC tissues and ADAM10 or ADAM17 knockdown in human HCC cells. *A*, immunohistochemical detection of ADAM10 and CD44 in human HCC tissues (n = 8) and normal liver tissues (n = 2). Liver sections were stained with the corresponding antibodies (*top panels*). Both primary antibodies were incubated with recombinant CD44 and ADAM10 proteins and then applied to liver sections in parallel as the absorption test (*bottom panels*). Representative images are shown. *B* and *C*, expression of ADAM10 or ADAM17 in human primary hepatocyte and HCC cell lines (*HepG2* and *PLC/PRF/5*). Cells were treated with ADAM10 siRNA, ADAM17 siRNA, or control siRNA, and subjected to analysis of ADAM10 or ADAM17 expression by flow cytometry (*B*) or real-time RT-PCR (*C*). *Histograms,* anti-ADAM10 or anti-ADAM17 staining of ADAM10 or ADAM17 siRNA-treated cells (*ADAM10KD* or *ADAM17KD, black dotted line*) and control siRNA-treated cells (*Control, gray line*), respectively. *Closed histograms,* control IgG staining. *D,* the expression of membrane-bound CD44 on HCC cells treated with ADAM10 siRNA (ADAM10KD, *black line*) or control siRNA (*Control, gray line*) was evaluated by flow cytometry (*top panels*). *Closed histograms,* control IgG staining. Soluble CD44 (*sCD44*) production from HCC cells treated with ADAM10 siRNA or control siRNA were evaluated by specific ELISA (*bottom panels*). *, $P < 0.05$.

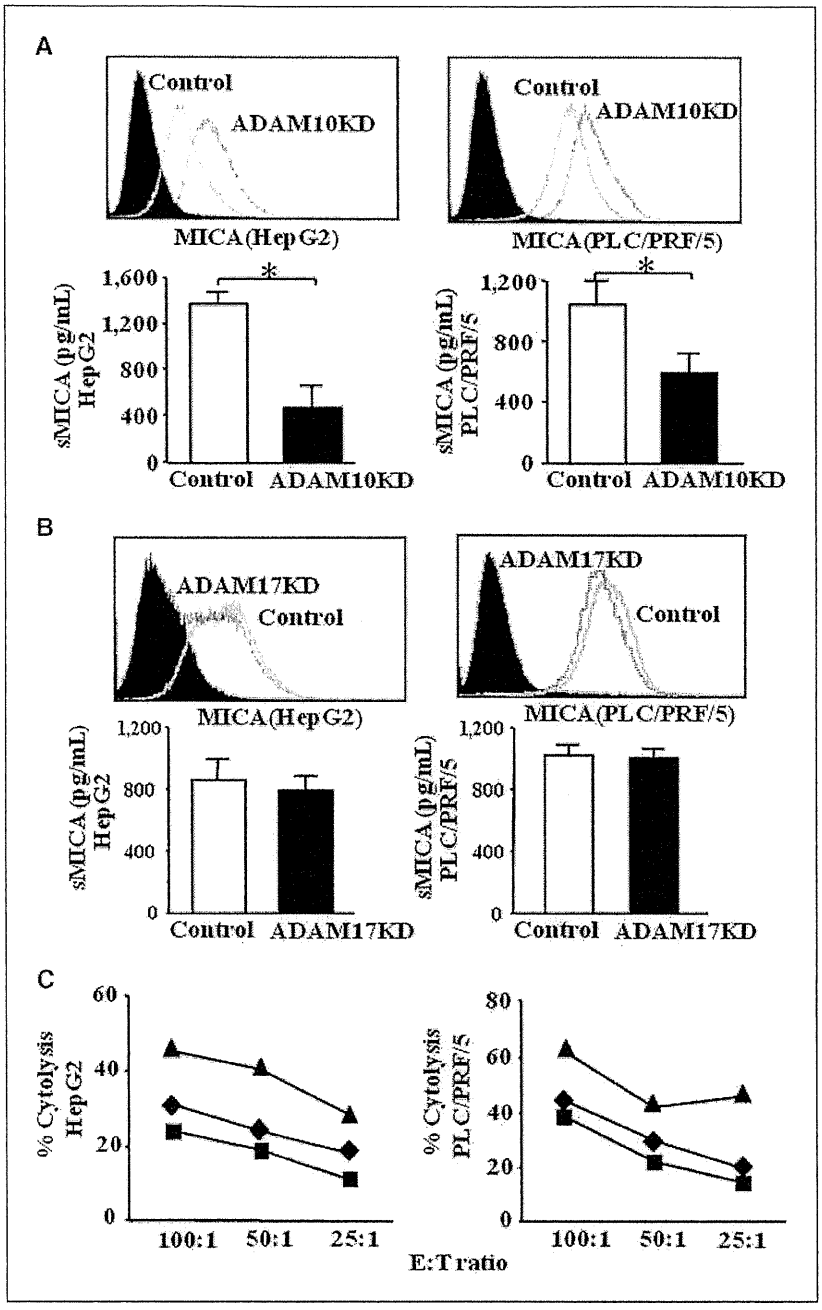

Figure 2. Expression of MICA in ADAM10 or ADAM17 knockdown HCC cells and NK sensitivity in ADAM10 knockdown HCC cells. *A* and *B*, the expression of membrane-bound MICA on HCC cells treated with ADAM10 siRNA (*ADAM10KD, black line*; *A*), ADAM17 siRNA (*ADAM17KD, black line*; *B*), or control siRNA (*Control, gray line*) was evaluated by flow cytometry (*top panels*). *Closed histograms,* control IgG staining. Soluble MICA (*sMICA*) production from HCC cells treated with ADAM10 siRNA (*A*), ADAM17 siRNA (*B*), or control siRNA were evaluated by specific ELISA (*bottom panels*). *, $P < 0.05$. *C*, HCC cells treated with ADAM10 siRNA or control siRNA were subjected to ^{51}Cr-release assay against NK cells. Cytolytic activity of NK cells against control HCC cells (■) or ADAM10 knockdown HCC cells without (▲) or with blocking antibody of MICA/B (6D4; ♦). Representative results are shown. Similar results were obtained from three independent experiments.

was obtained from all patients before sample acquisition. Liver sections were subjected to immunohistochemical staining using the ABC procedure (Vector Laboratories, Burlingame, CA). The primary antibodies used were anti-ADAM10 and anti-CD44 (R&D Systems). To confirm the specificity of the staining, primary antibodies were incubated with recombinant CD44 or ADAM10 protein (R&D Systems, Minneapolis, MN) for 3 h and then applied onto liver sections in parallel with staining of the primary antibodies as the absorption test.

HCC cell lines. Human HCC cell lines HepG2 and PLC/PRF/5 were purchased from the American Type Culture Collection and were cultured with DMEM supplemented with 10% fetal bovine serum (GIBCO/Life Technologies, Grand Island, NY) in a humidified incubator at 5% CO_2 and 37°C.

RNA silencing. The small interfering RNA (siRNA) method was used to knockdown ADAM10 and ADAM17. Stealth RNAi oligonucleotide targeting ADAM10 or ADAM17 and scrambled oligonucleotides as a control were purchased from Invitrogen (Carlsbad, CA). Cells were transfected by RNAi Max transfection reagent (Invitrogen) with 50 nmol/L siRNA. At 24 h posttransfection, the cells were analyzed for specific depletion of the mRNAs of ADAM10 and ADAM17 by real-time reverse transcription-PCR (RT-PCR; Applied Biosystems, Foster City, CA). The following siRNAs were used: ADAM10, 5′-AUAUCUGGGCAAUCACAG-CUUCUCG-3′; scramble control, 5′-AUACUUGGUCAACGCACUUCGAUGG-3′; ADAM17, 5′-UGAACAAGCUCUUCAGGUGGUUCUC-3′; scramble control, 5′-UGAUUAGAACUCUCGACUGGUGCUC-3′.

ELISA. The supernatants of cultured cells were harvested at 24 h after transfection with siRNA as well as sera from HCC patients (n = 97) and age-matched healthy volunteers (n = 32) were subjected to analysis of soluble MICA and soluble CD44 levels. Informed consent, under an institutional review board–approved protocol, was obtained from all patients before sample acquisition. The levels of soluble MICA and soluble CD44 were

determined by DuoSet MICA eELISA kit (R&D Systems) and soluble CD44std ELISA (Abcam, Cambridge, MA), respectively.

Flow cytometry. For the detection of membrane-bound MICA and CD44, cells were incubated with an anti-MICA–specific antibody (2C10, Santa Cruz Biotechnology, Santa Cruz, CA) or anti-CD44 antibody (R&D Systems) and stained with phycoerythrin (PE)-goat anti-mouse immunoglobulin (Beckman Coulter) as a secondary reagent and then subjected to flow cytometric analysis. For the detection of ADAM10 or ADAM17, cells were fixed and permeabilized with Cytofix/Cytoperm (BD Biosciences, San Jose, CA) and stained with PE-conjugated anti-ADAM10 or anti-ADAM17 antibody (R&D Systems). Flow cytometric analysis was performed using a FACScan flow cytometer (Becton Dickinson).

Plasmid construction of pMyc-MICA. MICA full coding cDNA was isolated from Huh7, human HCC cells, using a conventional RT-PCR method (Supplementary Fig. S1, DDBJ/EMBL/Genbank accession number AB506764) and inserted into the *Hin*dIII-*Xba*I site of pcDNA3 (Invitrogen). A C-myc tag was placed between the leader peptide and the α1 domain of MICA by site-specific mutagenesis using a QuikChange site-directed mutagenesis kit (Stratagene, La Jolla, CA) referred to as pMyc-MICA. Cells were transfected with pMyc-MICA using a Lipofectamine LTX reagent (Invitrogen). The green fluorescent protein (GFP)–expressing vector (pEGFP-C1, Clontech, Mountain View, CA) was cotransfected to evaluate the transfection efficiency.

Immunoprecipitation. Cells or tissues were homogenized in lysis buffer containing 1% NP40, 0.5% sodium deoxycholate, 0.1% SDS, 50 μg/mL aprotinin, 100 μg/mL phenylmethylsulfonyl fluoride, 1 mmol/L sodium orthovanadate, 50 mmol/L sodium fluoride, and PBS. To the cell supernatants, 0.5% NP40 and a cocktail of protease inhibitors were added. The protein contents of the samples were determined by BCA protein assay kit (Pierce, Rackford, IL). Immunoprecipitation with anti–c-Myc beads was performed for 1 h at 4°C. Immunocomplexes were eluted by a c-Myc–tagged peptide solution (MBL, Woburn, MA). The samples after immunoprecipitation were treated with 250 mU of N-glycosidase F (Roche, Mannheim, Germany) for 3 h at 37°C.

Western blotting. The total cellular protein was electrophoretically separated using SDS-12% polyacrylamide gels and transferred onto polyvinylidene difluoride membrane. The membrane was blocked in TBS-Tween containing 5% skim milk for 1 h and then probed with anti-Myc mouse monoclonal antibody (Cell Signaling Technology, Danvers, MA) at 4°C overnight. Horseradish peroxidase–conjugated anti-rabbit antibody and SuperSignal West Pico System (Pierce) were used for the detection of blots.

Real-time RT-PCR. Total RNA was isolated using RNeasy Mini Kit (Qiagen K.K., Tokyo, Japan) and was reverse transcribed using SuperScript III First-Strand Synthesis System (Invitrogen). The mRNA levels were evaluated using ABI PRISM 7900 Sequence Detection System (Applied Biosystems). Ready-to-use assays (Applied Biosystems) were used for the quantification of ADAM10 (Hs00153853_m1), ADAM17 (Hs00234221_m1), MICA (Hs00792195_m1), β-actin (Hs99999903_m1), and CD44 (Hs00174139_m1) mRNAs according to the manufacturer's instructions. The thermal cycling conditions for all genes were 2 min at 50°C and 10 min at 95°C, followed by 40 cycles at 95°C for 15 s and 60°C for 1 min. β-Actin mRNA from each sample was quantified as an endogenous control of internal RNA.

WST-8 assay. HepG2 and PLC/PRF/5 cells were treated with different concentrations of epirubicin for 24 h. Cell growth of epirubicin-treated HCC cells was determined by WST-8 assay (Nacalai Tesque, Kyoto, Japan) as previously described (21).

NK cell analysis. NK cells were isolated from human peripheral blood mononuclear cells by magnetic cell sorting using CD56 MicroBeads (Miltenyl Biotech, Auburn, CA) as previously described (16). The cytolytic ability of NK cells was assessed by 4-h ^{51}Cr-releasing assay with or without MICA/B-blocking antibody (6D4; ref. 7), which binds to the α1 and α2 domains of MICA and MICB. 6D4 was a generous gift from Drs. Veronika Groh and Thomas Spies (Fred Hutchinson Cancer Research Center, Seattle, WA).

Statistics. All values were expressed as the mean and SD. The statistical significance of differences between the groups was determined by applying Student's *t* test or two-sample *t* test with Welch correction after each group

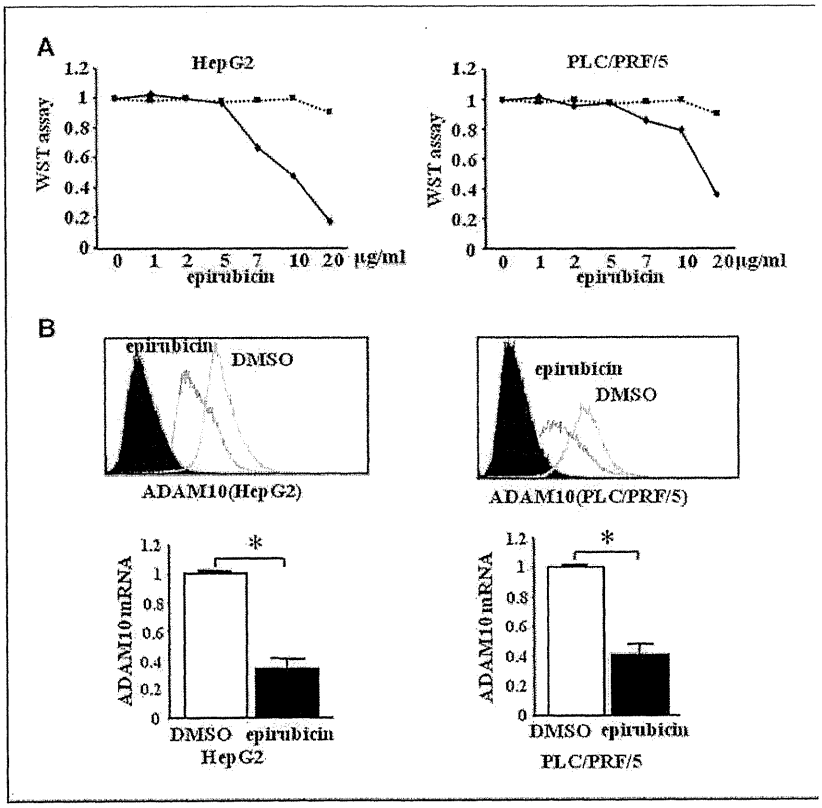

Figure 3. Expression of ADAM10 in epirubicin-treated HCC cells. *A*, the cytotoxicity of epirubicin to human HCC cells was evaluated by WST-8 assay. Cells were treated with different doses of epirubicin (*solid lines*) or vehicle (*DMSO*; *dotted lines*) for 24 h, and the viability of the cells was evaluated by the WST-8 assay. *B*, ADAM10 expression of epirubicin-treated HCC cells. Cells were treated with a nontoxic dose of 1 μg/mL epirubicin (*black lines*) or vehicle (DMSO, *gray lines*) for 24 h and their ADAM10 expression was evaluated by flow cytometry (*top panels*). *Closed histograms*, control IgG staining. Total RNA was extracted at 24 h of epirubicin treatment and mRNA levels of ADAM10 were evaluated by real-time RT-PCR (*bottom panels*). *, $P < 0.05$.

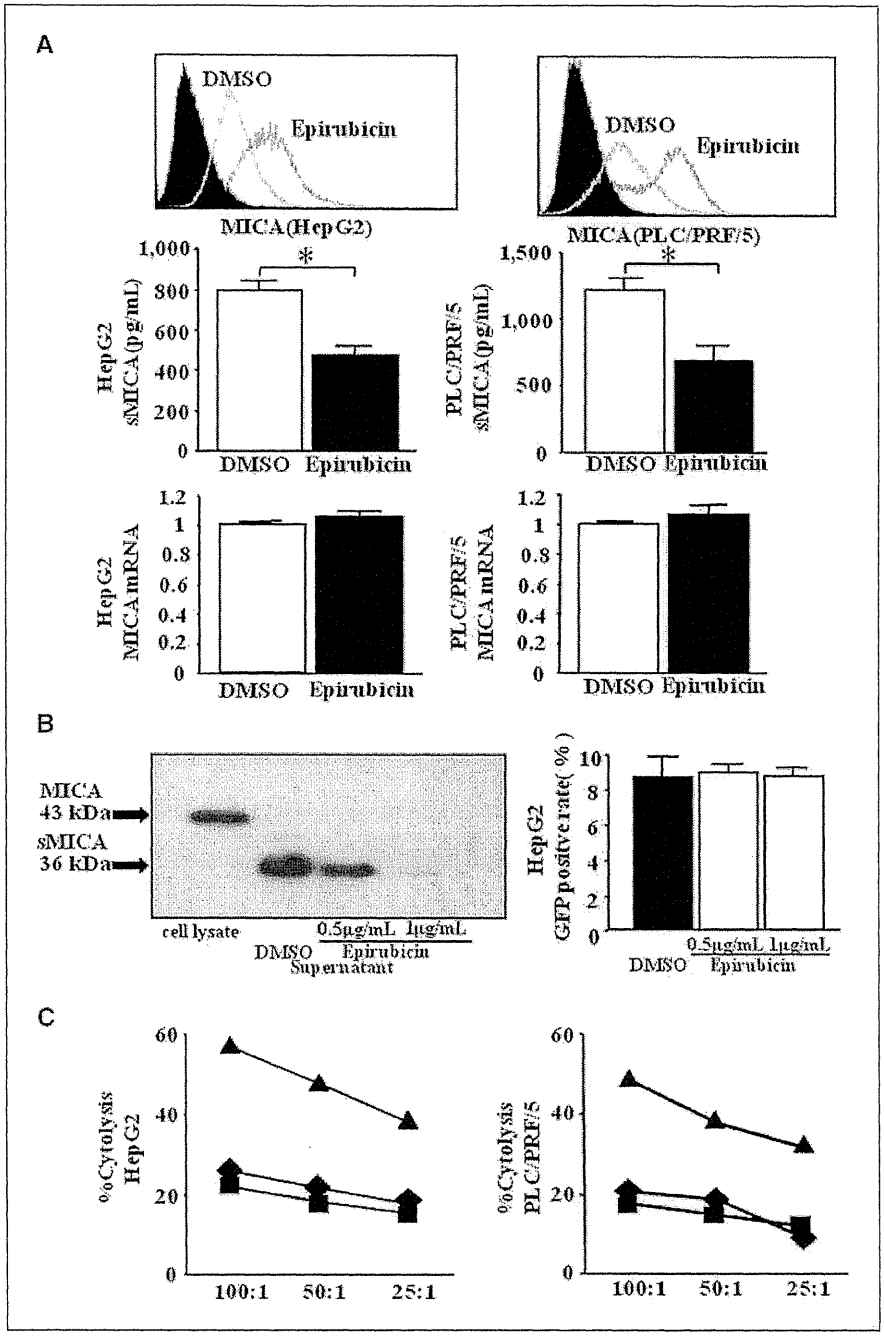

Figure 4. Expression and shedding of MICA in epirubicin-treated HCC cells. *A,* HCC cells were treated with a nontoxic dose of 1 μg/mL epirubicin (*black lines*) or vehicle (DMSO, *gray lines*) for 24 h and their expression of membrane-bound MICA and MICA mRNA was evaluated by flow cytometry (*top panels*) and real-time RT-PCR (*bottom panels*), respectively. *Closed histograms,* control IgG staining in flow cytometry. At the same time, 24-h culture supernatants were subjected to the analysis of soluble MICA (*sMICA*) levels by ELISA (*middle panels*). *, $P < 0.05$. *B,* HepG2 cells were transfected with pMyc-MICA and pEGFP-C1, cultured with 0.5 to 1 μg/mL epirubicin or vehicle (DMSO) for 24 h. Cell lysates from HepG2 cells and 24-h culture supernatants of epirubicin- or vehicle-treated HepG2 cells were immunoprecipitated with anti-Myc. The resulting immunoprecipitates were eluted, treated with N-glycanase, and subjected to Western blot analysis for MICA (*left*). Transfection efficacies were equal in all treatment groups as evidenced by similar GFP-positive cell rates (*right*). *C,* the cytolytic activity of NK cells against HCC cells. Vehicle-treated cells (■) or epirubicin-treated cells without (▲) or with blocking antibody of MICA/B (6D4; ♦) were subjected to ^{51}Cr-release assay. Representative results are shown. Similar results were obtained from three independent experiments.

had been tested with equal variance and Fisher's exact probability test. We defined statistical significance as $P < 0.05$.

Results

ADAM10 and CD44 are overexpressed in human HCC. ADAM10 was detected in all human HCC tissues tested by immunohistochemistry but not in normal liver tissues (Fig. 1*A*). Flow cytometric analysis revealed that ADAM10 was strongly expressed in a variety of HCC cell lines, including HepG2, PLC/PRF/5 (depicted in Fig. 1*B*), and Hep3B (data not shown), but faintly in primary hepatocytes. CD44, a typical substrate of the ADAM10 protease, was also expressed in all human HCC tissues but not in normal liver tissues (Fig. 1*A*). The data suggest that overexpression of ADAM10 and CD44 is a characteristic of human HCC like other malignancies (22).

ADAM10 is involved in MICA shedding of HCC cells but ADAM17 is not. To examine the involvement of ADAM family proteins in MICA ectodomain shedding, ADAM10 or ADAM17 were knocked down in HCC cells using a siRNA-mediated procedure. ADAM10 expression was clearly suppressed in HepG2 cells and PLC/PRF/5 cells at both mRNA and protein levels (Fig. 1*B* and *C*). Both cell lines expressed CD44 on the cellular surface and produced significant levels of soluble CD44 (Fig. 1*D*), indicating that CD44 is expressed and shed from those cell lines. ADAM10 knockdown (KD)

led to an increase in CD44 expression on HCC cells and a decrease in soluble CD44 levels in culture supernatants (Fig. 1*D*). Because ADAM10 has been established as being a sheddase for CD44, siRNA-mediated knockdown of ADAM10 suppressed not only the expression but also the activity of ADAM10 in HCC cells. HepG2 and PLC/PRF/5 cells also expressed ADAM17, which was clearly knocked down by a siRNA-mediated procedure (Fig. 1*B*).

HepG2 cells and PLC/PRF/5 cells expressed membrane-bound MICA and also produced soluble MICA (Fig. 2*A*). Knockdown of ADAM10 for both cell lines clearly upregulated MICA expression on their cellular surface and downregulated soluble MICA levels in their culture supernatant (Fig. 2*A*). In contrast, knockdown of ADAM17 did not affect the expression of membrane-bound MICA or the production of soluble MICA (Fig. 2*B*). We also examined the involvement of ADAM17 in MICA shedding of phorbol 12-myristate 13-acetate (PMA)–stimulated HCC cells because ADAM17 is considered to primarily affect stimulated shedding. The expression of membrane-bound MICA and the soluble MICA production were equal between PMA-stimulated ADAM17KD-HCC cells and control HCC cells (Supplementary Fig. S2). Thus, ADAM10, but not ADAM17, is critically involved in the shedding of MICA in HCC cells.

We next evaluated the cytolytic activity of NK cells against HCC cells. The cytolytic activity of NK cells against ADAM10KD-HepG2 cells was higher than that against control HepG2 cells. This activity was inhibited by blocking of anti-MICA/B antibody, suggesting that the increase of NK sensitivity depended on the increased expression of membrane-bound MICA on ADAM10KD-HepG2 cells, although we could not exclude the possibility of the involvement of MICB in this cytotoxicity (Fig. 2*C*). Similar results were also obtained with ADAM10KD-PLC/PRF/5 cells.

Epirubicin suppresses ADAM10 expression in HCC cells. We examined the biological modification of human HCC cells by adding epirubicin, which is commonly used in anti-HCC chemotherapy. We first examined the cytotoxicity of epirubicin to human HCC cells by WST-8 assay. Adding >5 μg/mL of epirubicin resulted in a significant decrease in cell growth of both HepG2 and PLC/PRF/5 cells (Fig. 3*B*). Based on these findings, we used 1 μg/mL of epirubicin to evaluate the biological effect on human HCC cells without toxicity. Both HepG2 cells and PLC/PRF/5 cells were cultured for 24 h with epirubicin and then subjected to analysis of ADAM10 expression. Epirubicin suppressed ADAM10 expression at the mRNA and protein levels in both cell lines (Fig. 3*C*). Although the data are not shown, doxorubicin also suppressed ADAM10 expression in HCC cells.

Epirubicin inhibits MICA ectodomain shedding and enhances susceptibility to NK cells of HCC cells. The above observations led us to investigate whether epirubicin or doxorubicin treatment would affect MICA ectodomain shedding in HCC cells. Epirubicin treatment led to an increase in membrane-bound MICA expression and a decrease in soluble MICA production in both HepG2 and PLC/PRF/5 cells (Fig. 4*A*). The mRNA levels of MICA did not change after exposure to epirubicin in both HCC cells (Fig. 4*A*). Similar data were obtained with doxorubicin-treated cells (data not shown).

To confirm whether the soluble MICA detected by ELISA was actually reflected in the cleaved form, we transfected Myc-tagged MICA into HepG2 cells and collected culture supernatants as well as cellular lysates. Immunprecipitates from these samples with anti-Myc were subjected to Western blot analysis after treatment with N-glycosidase. MICA in the culture supernatants migrated faster than cellular MICA (Fig. 4*B*), indicating that the MICA detected by ELISA is actually processed and released from full-length MICA. Epirubicin treatment led to a decrease in soluble MICA protein in HepG2 cells (Fig. 4*B*).

We next evaluated whether the epirubicin treatment could also modify the NK sensitivity of human HCC cells. Epirubicin-treated HepG2 cells or PLC/PRF/5 cells were more susceptible to NK cells than nontreated HepG2 or PLC/PRF/5 cells (Fig. 4*C*). The cytolytic activity against epirubicin-treated HCC cells was significantly decreased to the control levels by adding the anti-MICA/B blocking antibody. These results showed that the addition of epirubicin enhanced the NK sensitivity of HCC cell through increased

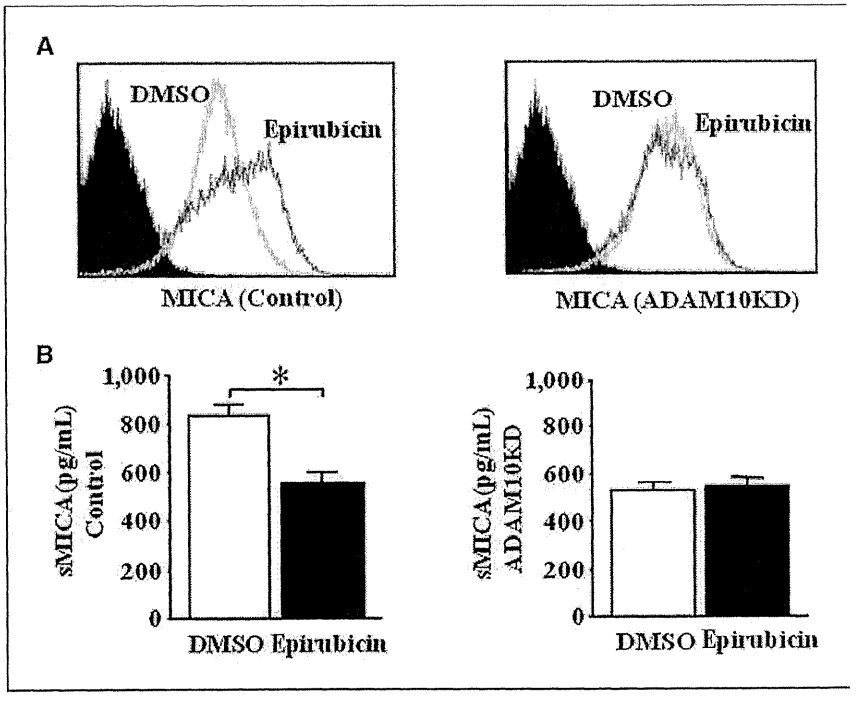

Figure 5. The epirubicin-mediated modification of MICA is ADAM10 dependent. HepG2 cells were transfected with ADAM10 siRNA (*ADAM10KD*) or control siRNA (*Control*) and further cultured with 1 μg/mL of epirubicin (*black lines*) or vehicle (DMSO, *gray line*) for 24 h. The expression of membrane-bound MICA (*MICA*) was evaluated by flow cytometry (*A*), and the soluble MICA (*sMICA*) production in the culture supernatant was evaluated by specific ELISA (*B*). Similar results were obtained from two independent experiments. *, $P < 0.05$.

expression of membrane-bound MICA, although the possibility of MICB involvement could not be excluded. The doxorubicin-treated human HCC cells showed similar results to those obtained from epirubicin-treated HCC cells (data not shown).

Epirubicin inhibits MICA ectodomain shedding through suppression of ADAM10. To examine whether the suppressive effect of epirubicin on MICA shedding occurred through downregulation of ADAM10, HepG2 cells were transfected with ADAM10 siRNA or scramble siRNA as a control and then treated with epirubicin. Consistent with earlier observations, epirubicin upregulated MICA surface expression and downregulated the levels of soluble MICA in control cells (Fig. 5). In contrast, neither upregulation of surface MICA nor downregulation of soluble MICA levels was observed in ADAM10KD-HepG2 cells. These results suggest that the suppressive effect of epirubicin on MICA shedding is mediated by ADAM10 downregulation. We also found similar results with ADAM10KD-PLC/PRF/5 cells (data not shown).

Soluble CD44 and soluble MICA levels in patients with HCC. We have shown that ADAM10 is expressed in human HCC tissues. However, it is not clear whether ADAM10 activity in HCC tissues is actually involved in MICA shedding in patients. Because ADAM10 was reported to be the constitutive functional sheddase of CD44 (23), we examined the soluble CD44 levels in HCC patients, which might be produced from tumor cells through ADAM10 activity. As shown in Fig. 6*A*, the soluble CD44 levels in HCC patients (n = 97) were significantly higher than those in age-matched healthy volunteers (n = 32). More importantly, soluble MICA levels in HCC patients significantly correlated with soluble CD44 levels (Fig. 6*B*), suggesting a close link between MICA shedding and ADAM10 activity.

We further examined soluble CD44 levels before and 2 weeks after TACE in HCC patients. Whereas the levels did not change in nontreated HCC patients during the 2-week interval (n = 9; 306.7 ± 82.5 ng/mL and 309.9 ± 79.9 ng/mL after 2 weeks), they were significantly decreased in epirubicin-based TACE-treated HCC patients (n = 21; 339.7 ± 78.1 ng/mL before TACE and 308.9 ± 81.4 ng/mL after TACE, $P < 0.003$). The changes of soluble CD44 in TACE treatment correlated significantly with those of soluble MICA (P = 0.0002; Fig. 6*C*). These results indicated that ADAM10-mediated CD44 shedding was decreased after TACE in HCC patients, implying that this reduction of ADAM10 activity might be related to the decline in MICA shedding.

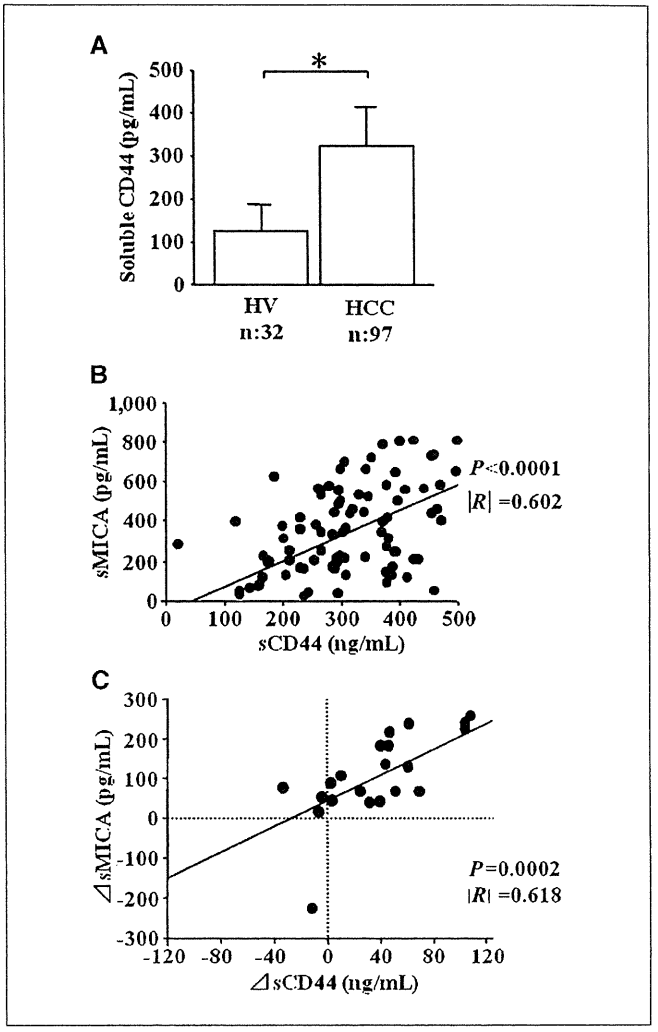

Figure 6. Correlation between soluble CD44 and soluble MICA in human HCC patients. *A* and *B*, soluble CD44 levels and MICA levels in healthy volunteers and HCC patients. Soluble CD44 levels (*sCD44*) and soluble MICA levels (*sMICA*) were determined for sera of HCC patients (n = 97) and age-matched healthy volunteers (*HV*; n = 32). *A*, comparison of sCD44 levels between groups; *B*, correlation between sCD44 levels and sMICA levels in 97 HCC patients. *, $P < 0.05$. *C*, correlation of sCD44 levels and sMICA levels during TACE therapy. HCC patients (n = 21) treated with epirubicin-based TACE therapy were enrolled and examined for sMICA and sCD44 levels before and 2 wk after therapy. Changes in sMICA (ΔsMICA = serum level of sMICA before TACE treatment − serum level of sMICA after TACE treatment) and those in sCD44 levels (ΔsCD44 = serum level of sCD44 before TACE treatment − serum level of sCD44 after TACE treatment) are plotted.

Discussion

MICA shedding is thought to be a principal mechanism by which tumor cells escape from NKG2D-mediated immunosurveillance (13). Thus, inhibition of MICA shedding should be a reasonable strategy for enhancing antitumor immunity. In the present study, we showed that ADAM10 was overexpressed in human HCC tissues and that ADAM10 knockdown resulted in increased expression of membrane-bound MICA, decreased production of soluble MICA, and upregulation of NK sensitivity of human HCC cells. These results point to ADAM10 as a therapeutic target for inhibiting MICA shedding, thereby ameliorating immunity against HCC. Waldhauer and colleagues recently showed that both ADAM10 and ADAM17 proteases are critically involved in the proteolytic release of soluble MICA of human 293T fibroblast cells and HeLa cervix carcinoma cells (20). Interestingly, in the present study, ADAM17 knockdown failed to affect MICA expression in human HepG2 cells or PLC/PRF/5 cells. Thus, ADAM10, not ADAM17, plays an essential role in the shedding of MICA in human HCC cells. Anderegg and colleagues (23) reported that only ADAM10, not ADAM17, contributed to shedding of CD44 molecules in human melanoma cells although both ADAM10 and ADAM17 proteases were significantly expressed in human melanoma tissues, suggesting that ADAM10 and ADAM17 do not always work in a similar manner. A recent report showed that ADAM10, but not ADAM17, could directly bind to calmodulin (24), which may involve the difference of MICA cleavage between ADAM10 and ADAM17 proteases. Recently, Boutet and colleagues reported that ADAM17 regulates proteolytic shedding of the MICB protein, which is another ligand for the NKG2D receptor on immune cells (25). We previously showed that both soluble MICA and MICB significantly increased in the sera of HCC patients and that therapeutic intervention for HCC leads to reduction of soluble

MICA levels, but not of soluble MICB levels (17), suggesting a more important role of soluble MICA in regulating NKG2D expression after HCC therapy. This led us to focus on the mechanism of MICA shedding in the present study.

Our results revealed that anticancer drugs such as epirubicin and doxorubicin downregulated ADAM10 expression and activity, thereby inhibiting MICA ectodomain shedding. The ADAM family proteins, which are highly expressed in some tumors, play a role in secreting growth factors, such as HB-EGF, and migration of cells. Thus, it is speculated that these proteins could be potential targets for tumor treatment (22). The present study is the first to show that clinically available anticancer drugs have an ability to modulate the expression of ADAM family proteins. They seem to suppress ADAM10 expression at a transcriptional level, but the precise mechanism of this suppression is not yet known.

The MICA ELISA system may not equally detect all soluble MICA (MICA molecules have >60 allelic variants). Our finding that soluble MICA could be detected in all HCC patients suggests that this system was applicable for our cohort of HCC patients. However, special caution should be paid for the use of this ELISA system for widely polymorphic MICA. Because CD44 is well known to be released into circulation from tumors by proteolytic cleavage of ADAM10 (23), the activity of ADAM10 in HCC tissues may be correlated with soluble CD44 levels. If so, our data suggest a close link between ADAM10 activity and the shedding of MICA in HCC. Furthermore, the decline in soluble MICA levels correlated well with the decline in soluble CD44 levels as early as 2 weeks after epirubicin-based TACE therapy. Reducing the tumor volume by such therapy may have led to both decreases but it is also possible that epirubicin suppresses ADAM10 activity, thereby inhibiting the shedding of MICA and CD44. Epirubicin may have a previously unrecognized role in cancer therapy; that is, affecting ADAM10 activity and MICA shedding rather than simply serving as a direct toxic agent for tumor cells.

Our data suggest that anti-HCC chemotherapy could remodel HCC cells, enhancing sensitivity to NK cells by upregulating MICA expression on the cellular surface. A concomitant decline in soluble MICA levels ameliorates NK cell ability by upregulating its NKG2D expression. We previously showed that activation of local innate antitumor immunity in liver tissues resulted in eliciting tumor-specific acquired immunity (21). If liver innate immunity is efficiently activated after anti-HCC chemotherapy, an additional antitumor effect against HCC cells could be expected. Immune modulators such as α-galactosylceramide have been shown to efficiently activate liver innate immune cells, including NK cells (21, 26). The combination therapy of anti-HCC chemotherapy and immunotherapy targeting NK cells might improve the antitumor effect of unresectable HCC and the prognosis of HCC patients.

In spite of recent progress in HCC therapies, there remains significant room for improvement, especially with respect to advanced liver cancer. We have shown here that anti-HCC chemotherapy resulted in enhanced NK sensitivity of HCC cells through inhibition of the activity of ADAM10 protease followed by modification of MICA expression. These findings indicate that efficient activation of liver innate immunity after anti-HCC chemotherapy might represent a particularly promising approach to suppress tumor growth and promote regression in liver cancer patients.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

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Factors affecting efficacy in patients with genotype 2 chronic hepatitis C treated by pegylated interferon alpha-2b and ribavirin: reducing drug doses has no impact on rapid and sustained virological responses

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SUMMARY. Reducing the dose of drug affects treatment efficacy in pegylated interferon (Peg-IFN) and ribavirin combination therapy for patients with hepatitis C virus (HCV) genotype 1. The aim of this study was to investigate the impact of drug exposure, as well as the baseline factors and the virological response on the treatment efficacy for genotype 2 patients. Two-hundred and fifty patients with genotype 2 HCV who were to undergo combination therapy for 24 weeks were included in the study, and 213 completed the treatment. Significantly more patients who achieved a rapid virological response (RVR), defined as HCV RNA negativity at week 4, achieved a sustained virological response (SVR) (92%, 122/133) compared with patients who failed to achieve RVR (48%, 38/80) ($P < 0.0001$). Multivariate logistic-regression analysis showed that only platelet counts [odds ratio (OR), 1.68; confidence interval (CI), 1.002–1.139] and RVR (OR, 11.251; CI, 5.184–24.419) were independently associated with SVR, with no correlation being found for the mean dose of Peg-IFN and ribavirin for RVR and SVR. Furthermore, in the stratification analysis of the timing of viral clearance, neither mean dose of Peg-IFN ($P = 0.795$) nor ribavirin ($P = 0.649$) affected SVR in each group. Among the patients with RVR, the lowest dose group of Peg-IFN (0.77 ± 0.10 μg/kg/week) and ribavirin (6.9 ± 0.90 mg/kg/day) showed 100% and 94% of SVR. Hence, RVR served as an important treatment predictor, and drug exposure had no impact on both SVR and RVR in combination therapy for genotype 2 patients.

Keywords: chronic hepatitis C, drug exposure, genotype 2, peginterferon and ribavirin combination therapy.

INTRODUCTION

The current standard of care for chronic hepatitis C (CHC) patients consists of combination therapy using pegylated interferon (Peg-IFN) and ribavirin [1–3]. Large, randomized clinical trials have demonstrated that 42–52% of hepatitis C virus (HCV) genotype 1 'difficult-to-treat' patients achieved sustained virological response (SVR), whereas 76–84% of HCV genotype 2 or 3 infected patients treated with Peg-IFN and ribavirin achieved SVR [4–6]. It also has been shown that in HCV genotype 2 and 3 infected patients, 24-week treatment regimens are just as effective as 48-week regimens [6,7]. Therefore, current guidelines recommend a 24-week treatment for these patients in contrast to 48 weeks for genotype 1 patients [1–3]. However, as side effects are common and treatment is expensive for this therapy, it would be ideal to be able to further reduce the total amount of drug medication

Abbreviations: ALT, alanine aminotransferase; BMI, body mass index; CHC, chronic hepatitis C; c-EVR, complete early virological response; ETR, end of treatment response; γ-GTP, γ-glutamyl transpeptidase; HCV, hepatitis C virus; IFN, interferon; NPV, negative predictive value; Peg-IFN, pegylated interferon; RVR, rapid virological response; SVR, sustained virological response.

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without loss of treatment efficacy for genotype 2 and 3 patients.

In HCV genotype 1 patients, reducing drug doses affects treatment efficacy. In our investigation of HCV genotype 1 patients, the rate of complete early virological response (c-EVR), defined as HCV RNA negativity at week 12, was affected by the mean dose of Peg-IFN during the first 12 weeks dose-dependently ($P < 0.0001$) [8]. Furthermore, we showed that only 4% relapse was found in patients given ≥12 mg/kg/day of ribavirin among those with c-EVR, and the relapse rate showed a decline in relation to the increase in the dose of ribavirin ($P = 0.0002$) [9]. On the contrary, it remains to be determined whether treatment efficacy can be preserved by further reducing both drug doses in genotype 2 and 3 patients. Because lower doses are expected to cause fewer adverse effects, it is important to find whether reduced drug doses can be used while retaining efficacy.

In the present study, we retrospectively evaluated the efficacy of Peg-IFN alpha-2b and ribavirin combination therapy for 24 weeks in patients infected with HCV genotype 2 and analysed the factors that affected the treatment efficacy, with particular interests in the drug impact of Peg-IFN and ribavirin.

PATIENTS AND METHODS

Patient selection and study design

Patients considered to be eligible for this study were those infected with HCV genotype 2 who underwent Peg-IFN alpha-2b (Schering-Plough K.K., Tokyo, Japan) and ribavirin (Schering-Plough K.K.) combination therapy from December 2005 to July 2007 at 29 medical institutions taking part in the Osaka Liver Forum and had completed the 24-week observation after a clinical course of 24 weeks. Patients with the following criteria were excluded: hepatitis B virus or human immunodeficiency virus coinfection, decompensated liver disease, severe cardiac, renal, haematological or chronic pulmonary disease, poorly controlled psychiatric disease, poorly controlled diabetes and immunologically mediated disease. Liver biopsy had been performed within 24 months prior to the treatment, and histological results were classified according to the METAVIR scoring system [10].

Written informed consent was obtained from each patient, and the study protocol was reviewed and approved according to the ethical guidelines of the 1975 Declaration of Helsinki by institutional review boards at the respective sites.

Patients were treated with Peg-IFN alpha-2b plus ribavirin for the duration of the study of 24 weeks. Peg-IFN alpha-2b and ribavirin dosages were based on body weight according to the manufacturer's instructions: Peg-IFN alpha-2b was given subcutaneously weekly (45 kg or less, 60 μg/dose; 46–60 kg, 80 μg/dose; 61–75 kg, 100 μg/dose; 76–90 kg, 120 μg/dose; 91 kg or more, 150 μg/dose), and ribavirin was given orally daily (60 kg or less, 600 mg/day; 61–80 kg, 800 mg/day; 81 kg or more, 1000 mg/day). The drug doses were also modified based on the manufacturer's instructions according to the intensity of the haematologic adverse effects.

Virological tests

Serum HCV RNA level was quantified by PCR assay (COBAS Amplicor HCV Test v2.0, Chugai-Roche Diagnostics, Tokyo, Japan), with a sensitivity limit of 5000 IU/mL and a dynamic range from 5000 to 5 000 000 IU/mL [11].

Serum HCV RNA was assessed by qualitative PCR assay (COBAS Amplicor HCV Monitor Test v2.0, Chugai-Roche Diagnostics), with a detection limit of 50 IU/mL [12].

Assessment of efficacy

Serum HCV RNA (qualitatively or quantitatively) was measured at weeks 4, 8, 12 and 24 during treatment and after 24 weeks of follow-up without treatment. Patients were classified as having a rapid virological response (RVR) if serum HCV RNA was undetectable (<50 IU/mL) at week 4 and at the end of treatment response (ETR) at week 24 of treatment. SVR was defined as undetectable HCV RNA at week 24 after treatment. Patients with an ETR who seroreverted to HCV RNA during follow-up were classified as relapsers.

Drug exposure

The amounts of Peg-IFN alpha-2b and ribavirin actually taken by each patient during the treatment period were evaluated by reviewing the medical records. The mean doses of both drugs were calculated individually as averages on the basis of body weight at baseline; Peg-IFN alpha-2b expressed as μg/kg/week and ribavirin as mg/kg/day.

Data collection

The medical records were retrospectively reviewed and the factors necessary for this examination were extracted: age, sex, body weight, body mass index (BMI), basic laboratory assessments, liver histology, quantitative and qualitative HCV RNA, dose of Peg-IFN alpha-2b and ribavirin received at each administration, and the response to treatment.

Statistical analysis

This study was a retrospective study and, for treatment results and the analysis of related factors, analysis was carried out only for cases in which the treatment had been completed (per-protocol analysis). Continuous variables are reported as the mean with standard deviation (SD) or

median level, while categorical variables are shown as the count and proportion. In univariate analysis, the Mann–Whitney *U*-test was used to analyse continuous variables, while chi-squared and Fisher's exact tests were used for analysis of categorical data. Variables with $P < 0.05$ at univariate analysis were retained for the multivariate logistic-regression analysis. Stepwise and multivariate logistic-regression models were used to explore the independent factors that could be used to predict a virological response. The significance of trends in values was determined with the Mantel–Haenszel chi-square test. For all tests, two-sided *P*-values were calculated and the results were considered statistically significant if $P < 0.05$. Statistical analysis was performed using the SPSS program for Windows, version 15.0J (SPSS, Chicago, IL, USA).

RESULTS

The baseline characteristics for the total cohort are shown in Table 1. Most of the patients were female (56%) with a mean age of 54 years. Seventy per cent of the patients were treatment naïve. Of the 250 patients, liver biopsies were performed for 174 patients, and 18 of them had advanced fibrosis (F 3–4).

Of the total of 250 patients, 37 (15%) were withdrawn from treatment because of adverse events: decreased haemoglobin ($n = 10$), psychiatric problems including depression ($n = 9$), fatigue ($n = 3$), thrombocytopenia, neutropenia, pyrexia, rash, cerebral haemorrhage, bleeding of ocular fundus, dyspnea, dizziness, jaundice, transaminase rise, gastrointestinal symptoms ($n = 1$) and other adverse events ($n = 4$). Eight of these patients who discontinued treatment prematurely had SVR (8/37; 22%).

Drug adherence

Seventy-nine of the 213 patients (37%) required dose reduction of Peg-IFN alpha-2b, 99 (46%) of ribavirin because of adverse events (not including patients who later discontinued treatment because of adverse event). Neutropenia (24/79; 30%) and thrombocytopenia (24/79; 30%) were the most common adverse events for dose reduction of Peg-IFN alpha-2b, and decreased haemoglobin (82/99; 83%) for that of ribavirin.

Virological response

Of the 213 patients who completed 24 weeks of treatment and 24 weeks of follow-up, 160 (75%) patients were clear of HCV RNA at week 4, 191 (90%) at week 8, 196 (92%) at week 12. ETR was observed for 195 (92%), and SVR for 160 (75%). The relapse rate was 18% (35/195).

Virological response according to the timing of viral clearance

Positive and negative prediction of sustained virological response according to the timing of viral clearance

We examined SVR rates according to the timing of viral clearance for the case in which HCV RNA was cleared during the treatment (Fig. 1a). The SVR rate was 92% (122/133) for patients clear of HCV RNA until week 4, 64% (37/58) from week 5 until week 8, 20% (1/5) from week 9 until

Table 1 Baseline demographic and viral characteristics of patients

Number of cases	250	
Age (years)*	54.0 ± 12.4	(22–76)
Sex (male/female)	110/140	
Body weight (kg)*	60.3 ± 11.7	(39–99)
Body mass index (kg/m^2)*	23.1 ± 3.2	(16–35)
Past IFN therapy (naïve/experienced)†	175/70	
HCV RNA (KIU/mL)‡	1700	(4–5000 <)
Fibrosis (0/1/2/3/4)§	18/98/40/14/4	
Activity (0/1/2/3)§	15/81/70/8	
White blood cells ($/mm^3$)*	5210 ± 1,750	(2100–13 870)
Neutrophils ($/mm^3$)*	2700 ± 1,250	(590–9020)
Red blood cells ($\times 10^4/mm^3$)*	436 ± 48	(307–554)
Haemoglobin (g/dL)*	13.9 ± 1.4	(10–18)
Platelets ($\times 10^4/mm^3$)*	18.3 ± 6.4	(4–41)
ALT (IU/L)*	79 ± 77	(13–581)
γ-GTP (U/L)*	56 ± 65	(7–479)
Creatinine(mg/dL)*	0.7 ± 0.1	(0.4–1.1)

IFN, interferon; HCV, hepatitis C virus; ALT, alanine aminotransferase; γ-GTP, γ-glutamyl transpeptidase. *Values expressed as mean ± SD (range), †interferon treatment history was not known for five patients, ‡values expressed as median (range), §data for 76 patients are missing.