

Figure 1 Nucleotide sequence results for the *SYCP3* gene in women with recurrent miscarriage and a control. (**A**) Heterozygous 657T>C mutation in exon 8 of the *SYCP3* gene of one patient with a history of six recurrent miscarriages. (**B**) Normal genotype in one patient with recurrent miscarriage. (**C**) Heterozygous 657T>C mutation in exon 8 of the *SYCP3* gene in one control with a history of one live birth and no miscarriages. From (A–C) sequences are all of the same region, and (A) and (B) sequences are complementary.

production of proteins that were mutated at the C-terminus. However, the effects of the SYCP3 mutations on non-disjunction or the function of the synaptonemal complex have not been clear in mammals so far.

Our patient with 657T>C had repeated miscarriages with euploidy. It may be that the 657T>C mutation is a polymorphism without the specific function ascertained in the Bolor et al. (2009) study in humans. Further studies with larger numbers and a wide range of cases are needed to define whether the SYCP3 mutations can be a cause of recurrent miscarriage.

Sycp3-deficient mice show complete meiotic arrest leading to male infertility (Yuan et al., 2000). Miyamoto et al. (2003) identified in two azoospermia patients a I bp deletion of the SYCP3 gene (643delA) that results in a premature stop codon and truncation of the C-terminal, coiled-coil-forming region of the SYCP3 protein. The mutant protein showed greatly reduced interaction with the wild-type protein in vitro (Miyamoto et al., 2003). Reynolds et al. (2007) suggested that azoospermia associated with a decrease in the DAZ gene function in humans might, in part, be the consequence of failure at synapsis caused by reduced levels of the SYCP protein. However, no female patient with the 643delA mutation of SYCP3 has been reported. The infertile women might have the mutation because embryos with trisomy or monosomy except 45,X are frequently seen by PGD.

Our data showed that among the normal fetal karyotypes, XX and XY were found at a similar frequency, indicating that fetal rather than maternal karyotypes were obtained. In this study, about 90% of patients with abnormal and normal embryonic (fetal) karyotype tended to have repeat miscarriages with abnormal and normal karyotypes, respectively. The results suggest that unexplained recurrent miscarriage should be grouped as two types: one is miscarriage caused by abnormal embryonic karyotype and the other is 'real' unexplained recurrent miscarriage.

Moreover, the prognosis of a successful pregnancy for patients with an abnormal embryonic karyotype was better than for patients with a normal embryonic karyotype (Ogasawara et al., 2000). No therapeutic approach to improve the rate of live birth could be found at this time (Kaandorp et al., 2010). Thus, the gene associated with unexplained recurrent miscarriage with normal embryonic karyotype is more important (Suzumori and Sugiura-Ogasawara, 2010).

The results of our study suggest no clinical significance of routine screening for the presence of the *SYCP3* mutation in women with recurrent miscarriage because we detected only one benign mutation in 101 such patients. Future studies in mammalian animal models are likely to accelerate our understanding of the molecular mechanisms involved in recurrent miscarriage and will provide additional candidate genes to be screened in recurrent miscarriage patients and embryos with genetic factors.

8 Mizutani et al.

Authors' roles

N.S., Y.O., M.N. and M.S.-O. were involved in conception and design; E.M., K.O. and C.Y-N. conducted data analysis; E.M., N.S. and M.S.-O. were involved in drafting the article. All authors agreed final approval of the version to be published.

Acknowledgements

We thank Shintaro Obayashi, M.D., Kenji Asamoto, M.D. and Kinue Katano, M.D. for organizing the collection of samples.

Funding

This study was supported by a Grant-in-Aid for Scientific Research from the Ministry of Health, Labour and Welfare (to M.S.-O.), by the Ministry of Education, Culture, Sports, Science, and Technology of Japan (to N.S.) and by the Japan Medical Association (to M.S.-O.).

References

- Aarabi M, Modarressi MH, Soltanghoraee H, Behjati R, Amirjannati N, Akhondi MM. Testicular expression of synaptonemal complex protein 3 (SYCP3) messenger ribonucleic acid in 110 patients with nonobstructive azoospermia. Fertil Steril 2006;86:325–331.
- Balasch J, Font J, López-Soto A, Cervera R, Jové I, Casals FJ, Vanrell JA. Antiphospholipid antibodies in unselected patients with repeated abortion. *Hum Reprod* 1990;**5**:43–46.
- Bolor H, Mori T, Nishiyama S, Ito Y, Hosoba E, Inagaki H, Kogo H, Ohye T, Tsutsumi M, Kato T et al. Mutations of the SYCP3 gene in women with recurrent pregnancy loss. Am J Hum Genet 2009;84:14–20.
- Carp H, Toder V, Aviram A, Daniely M, Mashiach S, Barkai G. Karyotype of the abortus in recurrent miscarriage. Fertil Steril 2001;75:678–682.
- Coulam CB, Jeyendran RS, Fishel LA, Roussev R. Multiple thrombophilic gene mutations rather than specific gene mutations are risk factors for recurrent miscarriage. *Am J Reprod Immunol* 2006;**55**:360–368.
- Farquharson RG, Pearson JF, John L. Lupus anticoagulant and pregnancy management. *Lancet* 1984;**28**:228–229.
- Goodman CS, Coulam CB, Jeyendran RS, Acosta VA, Roussev R. Which thrombophilic gene mutations are risk factors for recurrent pregnancy loss? *Am J Reprod Immunol* 2006;**56**:230–236.
- Harper J, Coonen E, De Rycke M, Fiorentino F, Geraedts J, Goossens V, Harton G, Moutou C, Pehlivan Budak T, Renwick P et al. What next for

- preimplantation genetic screening (PGS)? A position statement from the ESHRE PGD Consortium Steering Committee. *Hum Reprod* 2010; **25**:821–823
- Kaandorp SP, Goddijin M, van der Post JA, Hutten BA, Verhoeve HR, Hamulyák K, Mol BW, Folkeringa N, Nahuis M, Papatsonis DN et al. Aspirin plus heparin or aspirin alone in women with recurrent miscarriage. N Engl J Med 2010;362:1586–4166.
- Miyamoto T, Hasuike S, Yogev L, Maduro MR, Ishikawa M, Westphal H, Lamb DJ. Azoospermia in patients heterozygous for a mutation in SYCP3. *Lancet* 2003;**362**:1714–1719.
- Nelen W, Steegers E, Eskes T, Blom H. Genetic risk factors for unexplained recurrent pregnancy loss. *Lancet* 1996;**350**:861.
- Ogasawara M, Aoki K, Matsuura E, Sasa H, Yagami Y. Antiβglycoprotein I antibodies and lupus anticoagulant in patients with recurrent pregnancy loss: prevalence and clinical association. *Lupus* 1996; **5**:587–592.
- Ogasawara M, Aoki K, Okada S, Suzumori K. Embryonic karyotype of abortuses in relation to the number of previous miscarriages. *Fertil* Steril 2000;**73**:300–304.
- Platteau P, Staessen C, Michiels A, Van Steirteghem A, Liebaers I, Devroey P. Preimplantation genetic diagnosis for aneuploidy screening in patients with unexplained recurrent miscarriages. *Fertil Steril* 2005; **83**:393–397.
- Rai R, Regan L. Recurrent miscarriage. Lancet 2006;368:601-611.
- Rey E, Kahn SR, David M, Shrier I. Thrombophilic disorders and fetal loss: a meta-analysis. *Lancet* 2003;**361**:901–908.
- Reynolds N, Collier B, Bingham V, Gray NK, Cooke HJ. Translation of the synaptonemal complex component Sycp3 is enhanced *in vivo* by the germ cell specific regulator DAZI. *RNA* 2007;**13**:974–981.
- Sugiura-Ogasawara M, Ozaki Y, Sato T, Suzumori N, Suzumori K. Poor prognosis of recurrent aborters with either maternal or paternal reciprocal translocation. Fertil Steril 2004;81:367–373.
- Sugiura-Ogasawara M, Ozaki Y, Kitaori T, Kumagai K, Suzuki S. Midline uterine defect size correlated with miscarriage of euploid embryos in recurrent cases. Fertil Steril 2010;**93**:1983–1988.
- Suzumori N, Sugiura-Ogasawara M. Genetic factors as a cause of miscarriage. *Curr Med Chem* 2010;17:3431–3437.
- The American College of Obstetrics and Gynecologists. Preimplantation genetic screening for aneuploidy. *Obstet Gynecol* 2009; **I 13**:766–767.
- Yuan L, Liu JG, Zhao J, Brundell E, Daneholt B, Hoog C. The murine SCP3 gene is required for synaptonemal complex assembly, chromosome synapsis, and male fertility. *Mol Cell* 2000;**5**:73–83.
- Yuan L, Liu JG, Hoja MR, Wilbertz J, Nordqvist K, Hoog C. Female germ cell aneuploidy and embryo death in mice lacking the meiosis-specific protein SCP3. *Science* 2002;**296**:1115–1118.



'Non-criteria' aPL tests: report of a task force and preconference workshop at the 13th International Congress on Antiphospholipid Antibodies, Galveston, TX, USA, April 2010

**IL Bertolaccini, O Amengual, T Atsumi, WL Binder, B de Laat, R Forastiero, WH Kutteh, M Lambert, H Matsubayashi, V Murthy, M Petri, JH Rand, M Sanmarco, AE Tebo and SS Pierangeli

Lupus 2011 20: 191

**PSI 10 4137/2020402020402020

DOI: 10.1177/0961203310397082

The online version of this article can be found at: http://lup.sagepub.com/content/20/2/191

> Published by: **S**SAGE

http://www.sagepublications.com

Additional services and information for Lupus can be found at:

Email Alerts: http://lup.sagepub.com/cgi/alerts

Subscriptions: http://lup.sagepub.com/subscriptions

Reprints: http://www.sagepub.com/journalsReprints.nav

Permissions: http://www.sagepub.com/journalsPermissions.nav

>> Version of Record - Feb 8, 2011

What is This?

SPECIAL ARTICLE

'Non-criteria' aPL tests: report of a task force and preconference workshop at the 13th International Congress on Antiphospholipid Antibodies, Galveston, TX, USA, April 2010

ML Bertolaccini¹, O Amengual², T Atsumi², WL Binder³, B de Laat⁴, R Forastiero⁵, WH Kutteh⁶, M Lambert⁷, H Matsubayashi⁸, V Murthy⁹, M Petri¹⁰, JH Rand¹¹, M Sanmarco¹², AE Tebo¹³ and SS Pierangeli¹⁴

¹Lupus Research Unit, The Rayne Institute, King's College London School of Medicine, London, UK; ²Department of Internal Medicine II, Hokkaido University School of Medicine, Sapporo, Japan; ³INOVA Diagnostics Inc., San Diego, California, USA; ⁴Sanquin Research, Sanquin Blood Supply Foundation, Amsterdam, The Netherlands; Department of Clinical Chemistry and Hematology, University Medical Center Utrecht, Utrecht, The Netherlands; ⁵Department of Physiology, Favaloro University, Division of Hematology, Thrombosis, and Haemostasis, University Hospital, Favaloro Foundation, Buenos Aires, Argentina; ⁶Division of Reproductive Endocrinology, Department of Obstetrics and Gynecology, University of Tennessee Health Science Center, Memphis, Tennessee, USA; ⁷Service de Médecine Interne, Hôpital Claude-Huriez, Centre Hospitalier Régional et Universitaire de Lille, Lille, France; ⁸Osaka New ART Clinic, Tokai University School of Medicine, Osaka, Japan; ⁹Division of Rheumatology, Department of Internal Medicine, University of Texas Medical Branch, Galveston, Texas, USA; ¹⁰Department of Medicine, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA; ¹¹Department of Pathology, Montefiore Medical Center, Albert Einstein College of Medicine, Bronx, New York, USA; ¹²Laboratoire d'Immunologie, Hôpital de La Conception, Marseille, France; ¹³Associated Regional and University Pathologists (ARUP) Institute for Clinical and Experimental Pathology, University of Utah School of Medicine, Salt Lake City, Utah, USA; and ¹⁴Antiphospholipid Standardization Laboratory, Division of Rheumatology, Department of Internal Medicine, University of Texas Medical Branch, Galveston, Texas, USA

Abstract: Current classification criteria for definite APS recommend the use of one or more of three positive standardized laboratory assays, including anticardiolipin antibodies (aCL), lupus anticoagulant (LA), and antibodies directed to β_2 glycoprotein I (anti- β_2 GPI) to detect antiphospholipid antibodies (aPL) in the presence of at least one of the two major clinical manifestations (i.e., thrombosis or pregnancy morbidity) of the syndrome. Several other autoantibodies shown to be directed to phospholipids and/or their complexes with phospholipids and/or to proteins of the coagulation cascade, as well as a mechanistic test for resistance to annexin A5 anticoagulant activity, have been proposed to be relevant to APS. A task force of worldwide scientists in the field discussed and analyzed critical questions related to 'non-criteria' aPL tests in an evidence-based manner during the 13th International Congress on Antiphospholipid Antibodies (APLA 2010, 13–16 April 2010, Galveston, Texas, USA). This report summarizes the findings, conclusions, and recommendations of this task force. Lupus (2011) **20**, 191–205.

Key words: autoantibodies; prothrombin; phosphatidylethanolamine; IgA

Introduction

Current classification criteria for definite antiphospholipid syndrome (APS) recommend the use of one or more of three positive standardized laboratory assays to detect antiphospholipid antibodies (aPL) in the presence of at least one of the two major clinical manifestations

(i.e., thrombosis or pregnancy morbidity) of the syndrome. Anticardiolipin antibodies (aCL), anti- β_2 glycoprotein I (β_2 GPI) antibodies, and lupus anticoagulant (LA) are the laboratory tests included in the revised criteria for the classification of APS.

A number of issues regarding the definition of 'aPL positive' are under discussion. For example, there are in daily practice many in vitro 'false positives' for aPL, due to the lack of specificity of the tests, particularly the aCL ELISA. APL antibodies are found in patients with a variety of diseases, such as infectious, malignant, or autoimmune diseases (clinical false positive), but in those cases they are

Correspondence to: Maria Laura Bertolaccini, Lupus Research Unit, The Rayne Institute, King's College London School of Medicine, 4th Floor Lambeth Wing, St Thomas' Hospital, London SE1 7EH, UK Email: maria.bertolaccini@kcl.ac.uk

10.1177/0961203310397082

not associated with clinical manifestations of APS. Furthermore, increasing evidence demonstrates that aPL antibodies are heterogeneous in function and specificity, and individual tests may recognize various subtypes of antibodies, some of which may be 'pathogenic'. In addition, there are patients strongly suspected of having APS by their clinical phenotype, but persistently negative for any currently tested aPL (laboratory and/or clinical false negative). These findings have nurtured the concept of 'seronegative APS' (SNAPS), a much contended setting that is based on a clinical picture highly suggestive of the syndrome in the absence of conventional aPL antibodies, leading investigators to maintain their efforts to identify 'true aPL' in an attempt to better recognize APS patients.

Several autoantibodies shown to be directed to negatively charged phospholipids other than cardiolipin, to other proteins of the coagulation cascade (i.e., prothrombin and/or phosphatidylserine-prothrombin complexes), to some domains of β₂GPI, or to interfere with the anticoagulant activity of annexin A5 (A5), have been proposed to be relevant to APS.2 In some cases, these assays appear to detect specific subsets of pathogenic antibodies, or a particular mechanism in APS. However, the clinical utility of these newly developed assays and their diagnostic value remains elusive. The issue of the value of IgA aPL antibodies and whether this test should be part of the routine diagnostic algorithm has also been a subject of debate. A worldwide task force of scientists in the field – divided into subgroups – discussed and analyzed critical questions related to 'non-criteria' aPL tests in an evidence-based manner during the 13th International Congress on Antiphospholipid Antibodies (APLA 2010, 13–16 April 2010, Galveston, TX, USA). This report summarizes the findings, conclusions, and recommendations of this task force.

Antibodies to phosphatidylethanolamine

(Presented by Drs Sanmarco, Lambert, and Matsubayashi)

Introduction and questions addressed by the task force

Antibodies directed toward phosphatidylethanolamine (anti-PE) deserve particular attention, since they have been described in some instances as the sole aPL in patients that have manifestations of APS. Thus, the goal of this session was to highlight the clinical interest of anti-PE investigation through a brief review of the literature of their clinical associations and clinical experience. Another point opened to the debate was the methodological problems of the anti-PE assays.

Regarding obstetrical complications, anti-PE have been reported to be significantly more frequent in women with unexplained early fetal loss (UFL) than in either those with explained early fetal loss or healthy mothers. Two different studies have shown that anti-PE are a higher independent risk factor for early UFL than either aCL or anti- β_2 GPI antibodies.^{3,4} Interestingly, anti-PE have also been described as the only aPL found in the majority of cases (73%). Likewise, anti-PE have been reported as significantly the most frequent aPL in infertile women (67.5% of aPL-positive sera), where they were found to be the sole aPL in 85% of cases.

Recently, a murine model has reinforced the interest in anti-PE investigation in obstetric complications. Indeed, as reported by Dr Matsubayashi in this session, passive immunization of anti-PE or anti-LDC27 (antigen site in the third domain of kininogen) in pregnant mice causes increased fetal resorption, which correlated with significant increases in apoptosis in the placenta (study in progress). He claimed that this study supports the pathogenic role of anti-PE in pregnancy complications and also suggests the importance of LDC27, the target antigen site for kininogen-dependent anti-PE.

The relationship between anti-PE and thrombosis, the other clinical feature of APS, has also been reported in several studies. In particular, in a multicenter study set up within the framework of the European Forum on aPL, the prevalence of anti-PE was 15% in patients with unexplained venous thromboses and mainly found as the sole aPL.⁵ In this retrospective study, IgG-anti-PE were found to be an independent risk factor for venous thrombosis, with an odds ratio of 6:1. Interestingly, Dr Lambert reported that in a selected population 243 outpatients consulting for idiopathic arterial and/or venous thrombosis, negative for conventional aPL antibodies, 58 were positive for anti-PE (IgM mainly and IgG rarely). Other thrombophilic disorders were not frequently found. During a median follow-up of 34 months, thrombotic recurrence was found in 25% of patients.

Importantly, the task force recognized that no consensual standardized method exists for the measurement of anti-PE and that the heterogeneity of these antibodies increases the difficulties in attempting such a goal. This problem significantly limits

the clinical utility of this assay. The impact of the various ELISA components on the interlaboratory variability of results was analyzed, the conclusion being that the buffer supplement represents the critical factor in anti-PE measurement. To that regard, the results from a recent study showing that buffer supplements with a high lipid content decrease anti-PE reactivity in a dose-dependent manner were presented at this meeting.⁶

Recommendations of the task force

Based on published evidence and the additional studies presented during this session, the detection of anti-PE antibodies may be useful in 'seronegative' APS, in spite of the absence of a consensual method for their detection. The task force recognized that further steps must be made in order to ascertain the place of these antibodies in the diagnostic algorithm of APS, including standardization and proper validation of an anti-PE ELISA test and a prospective study on a broad population with well-documented clinical and biological features of APS (Table 1a).

Antibodies to domains of \(\beta \) glycoprotein I

(Presented by Dr Bas de Laat)

Introduction and questions addressed by the task force

APL antibodies form a heterogeneous population of antibodies recognizing different antigens. 7 β_2 GPI is recognized as the most important antigen in APS, but anti- β_2 GPI antibodies are also regarded as a heterogeneous population of antibodies with reactivity towards different epitopes on β_2 GPI. During the last decade evidence has accumulated for a central role for domain I of β_2 GPI as a primary epitope for aPL antibodies. Iverson et al. were the first to show that a specific population of aPL antibodies showed reactivity towards domain I, with glycine40-arginine43 as the major epitope. Recently Ioannou et al. reported that the epitope possibly comprises a larger region on domains I and II. 11

Two studies have been conducted to investigate the clinical significance of the detection of antidomain I antibodies. The first of these showed that the presence of anti-domain I antibodies was associated more with (predominantly venous) thrombosis compared with anti- $\beta_2 GPI$ antibodies with reactivity towards other domains. ¹²

This observation was recently confirmed in a double-blinded multicenter study including 442 patients, all positive for anti- β₂GPI antibodies.¹³ Anti-domain I antibodies were shown to be present in the plasma of 243/442 patients (55%). From these patients with anti-domain I antibodies in their plasma, 83% had a history of thrombosis resulting in an odds ratio of 3.5:1 (2.3-5.4, 95% CI) for thrombosis. confidence interval, Interestingly, it was also found that anti-domain I antibodies were associated with pregnancy morbidity. Furthermore, recently in vivo data have been generated with respect to domain I. Ioannou et al. conducted a study in which mice were injected with IgG purified from patients diagnosed with APS.¹⁴ After standardized vessel injury, mice injected with antiphospholipid-related IgG displayed increased thrombus size that could be inhibited by domain I of β_2 GPI.

This task force subgroup was charged with investigating whether there is sufficient scientific evidence to recommend the incorporation of the assay to measure anti-domain I antibodies for implementation in the official guidelines for diagnosis of patients with APS.

Recommendations of the task force

The general opinion of the task force was that detection of anti-domain I antibodies is of major importance. This was predominantly based on a double-blinded multicenter study in which it was shown that anti-domain I antibodies were associated more with thrombosis and pregnancy morbidity compared with antibodies with reactivity towards other domains of β_2 GPI. One of the problems that can also be applied to (some of) the other assays that are already included in the official guidelines is lack of prospective data (a) and causality (b):

- (a) Several prospective studies have been performed with regard to the clinical significance of the presence of aPL antibodies regardless of specificity, but there is no consensus as to whether the presence of aPL antibodies is a risk factor for thrombosis (either first or second event). 15-21
- (b) The causality of anti-domain I has been demonstrated only by the use of animal models, and additional clinical studies are needed. 14

Therefore, this task force recommended that the anti-domain I assay may be used in a research-based setting and that more prospective and in vivo data are needed before the anti-domain I

Table 1 Questions and recommendations of the non-criteria aPL task force

1a. Anti-PE antibodies and antibodies to negatively charged phospholipids other than cardiolipin

Test	Questions addressed by task force	Recommendations
Anti-PE antibodies	Is the anti-PE ELISA standardized? What are the challenges with the assay?	Standardization of anti-PE ELISA needed
	Are anti-PE antibodies clinically relevant?	Well-designed clinical studies needed to confirm the diagnostic value of anti-PE antibodies
Antibodies to negatively charged phospholipids other than cardiolipin		
a) Perspectives and experiences from a large reference laboratory in the USA	Are antibodies to negatively charged phospholipids other than cardiolipin important in the diagnosis of APS?	Important to establish whether these antibodies recognize additional APS patients, currently missed with tradition assays
		Address existing technical problems and inconsistencies with the tests
		Anti-PS may be best candidate with respect to relevance are association with recurrent pregnancy loss
Antibodies to negatively charged phospholipids other than cardiolipin		
b) In the obstetric population	Do non-criteria aPL exist and are they found in women with RPL?	Based on clinical studies = yes
	Are there sufficient clinical data to warrant a change in the 2006 Classification criteria	Not at the moment; more conclusive clinical studies are needed
	Do women with RPL who have early pregnancy losses and no thrombosis constitute a unique subgroup of APS with different diagnostic criteria of APS?	Obstetric populations should be stratified (with or withou prior thrombosis and third-trimester losses from first trimester)

Test	Questions addressed by task force	Recommendations
Anti-domain I antibodies	Does the anti-domain I antibodies test recognize 'pathogenic' anti- β_2 GPI antibodies?	0 0
	Is there convincing evidence to include this test in the diagnostic algorithm of APS?	In vivo data with anti-domain I antibodies needed. Standardized consensus protocol for this assay needed Additional clinical studies needed
IgA aCL and IgA anti-β ₂ GPI antibodies	Are IgA aPL (particularly IgA anti- β_2 GPI) clinically significant in patients with clinical manifestations of APS?	IgA anti- β_2 GPI antibodies should be tested in the presence of clinical signs and symptoms of SLE and/or APS, particularly when other aPL tests are negative Evaluation and comparison of multiple, commercially available IgA aPL assays in A larger and well-characterized population of patients needed to confirm the diagnostic value of isolated anti- β_2 GPI positivity Studies needed to determine the role of IgA anti- β_2 GPI antibodies in the pathogenesis of APS IgA anti- β_2 GPI antibodies that bind to domains IV/V of β_2 GPI might represent an important subgroup of clinically relevant aPL antibodies
Anti-prothrombin and anti-prothrombin-PS antibodies; antibodies to negatively charged phospholipids other than cardiolipin	What is the role of anti-prothrombin and anti-prothrombin/phosphatidylserine antibodies in APS? Are antibodies to negatively charged phospholipids other than cardiolipin important in the diagnosis of APS?	aPT-A test in conjunction with other tests may be a good risk marker for thrombosis aPT-A and particularly the anti-PS/PT are good specific tests to confirm APS aPT-A and anti-PS/PT not ready to be included in the diagnostic criteria (standardization of the tests needed) Collaborative studies needed to confirm clinical association with these tests
Annexin A5 (A5R) resistance test; anti- prothrombin and anti-prothrombin-PS antibodies	What is the role of the Annexin A5 resistance test in the diagnosis of APS? What is the role of anti-prothrombin and anti-prothrombin/phosphatidylserine antibodies in APS?	Data on the utility of AnxA5 resistance assay as a mechanistic diagnostic marker for APS are highly promising Developing mechanistic clinical assays that measure APS disease mechanisms is an important and appropriate avenue to pursue Additional data are needed before recommending A5R as standard component of aPL testing panels

assay can be added to the official diagnostic guidelines. This assay needs to be made available to other centers for testing before any recommendation can be made (Table 1b).

Antibodies to negatively charged phospholipids other than cardiolipin: perspectives and experiences from a large reference laboratory in the USA (*Presented by Dr Tebo*)

Introduction and questions addressed by the task force

Antibodies directed against negatively charged phospholipids such as phosphatidic acid (PA), phosphatidylinositol (PI), and phosphatidylserine (PS) have been reported in patients with APS. However, the use of these antibodies in addition to the currently recommended laboratory markers for the diagnosis of APS remains controversial. Some investigators have suggested that testing for these aPL antibodies may help to identify women with recurrent pregnancy loss (RPL) with clinical features of APS who may benefit from treatment, a topic discussed in detail in the next section.² In other such studies, as well as in the context of thrombosis associated with systemic lupus erythematosus, no improvement in the diagnosis performance was observed when these were measured simultaneously with aCL and LA, 23-25,27 Therefore, these assays were not included in the 2006 revised criteria for the classification of APS.¹ In a review of the literature since the laboratory criteria for APS were revised, very few studies have been carried out to examine the relevance for these antibody markers. As such, most of the discussion and recommendations in this article will focus on the few recent investigations on this topic, with reference to some earlier key findings.

Early investigations by Gharavi and colleagues showed that aCL antibodies broadly cross-react to both antiphosphatidylserine (anti-PS) and antiphosphatidylinositol (anti-PI) antibodies.²⁸ Of the three major negatively charged aPL antibodies (anti-PA, anti-PI, and anti-PS), anti-PS has been most extensively investigated in thrombosis- and pregnancy-related morbidity APS. 22-27,29,30 These antibodies, particularly anti-PS, have been shown to be more specific for APS when compared with aCL, since aCL is often found to be positive in disorders.31,32 diseases and other infectious However, the conditions necessary to achieve optimal clinical and analytical performance in these assays are yet to be determined. 1,29 Using aPS assays from two different manufacturers, Tebo et al. could not document a consistent diagnostic utility for this marker for both the IgG and IgM isotypes. 19 In addition, the combined use of these 'non-criteria' aPL antibodies differed significantly between manufacturers, especially for IgM specificities, and their overall combined diagnostic performance was not significantly higher than that of aCL and anti- β_2 GPI assays. 19,30 Of clinical importance, no difference in the magnitude and prevalence of these antibodies was documented between healthy controls and women with recurrent pregnancy loss. 10

Recommendations of the task force

In the evaluation of additional diagnostic markers for APS:

- (a) It is important to determine critically whether, indeed, these antibodies contribute to the identification of additional patients who would otherwise be missed by the current assays or, alternatively, they would be better predictors of disease due to improved analytical and clinical performance. Anti-PA, anti-PI, and anti-PS antibodies in their current format pose significant diagnostic and analytical challenges. First, when they occur, they do so in high association with aCL antibodies and in isolation, and their clinical relevance is questionable and has not been fully investigated.
- (b) In the case of anti-PS antibodies, the conditions required to detect this antibody remain controversial. Even for assays using the same reagents, the results are discordant as there are no formal calibrators or agreed methods of detection. Thus, in addition to not being cost-effective, to choose assays with the best medical benefit rather than a collection of tests with overlapping properties and equivalent or questionable clinical value may be the best practice.
- (c) Based on the current evidence, it would appear that testing for anti-PA, anti-PI, and anti-PS antibodies in the initial diagnostic work-up for APS is not clinically useful, as these antibodies may have overlapping properties with the markers considered diagnostic for this disease.
- (d) It would appear that the anti-PS marker may be the best candidate for further investigation of its relevance and significance, especially in the area of recurrent pregnancy loss, provided an accepted and standardized method is in place. In this case, more prospective studies using an agreed-upon protocol for patient recruitment,

follow-up, and testing for the presence of these antibodies are critical (Table 1a).

Antiphospholipid antibodies other than anticardiolipin antibodies in obstetric APS

(Presented by Dr Kutteh)

Introduction

Several investigators worldwide have advocated the use of a panel of aPL antibodies (aPL) to screen for APS. 33,34 This panel of tests includes not only cardiolipin (CL, diphosphatidyl glycerol) but also phosphatidyl inositol, phosphatidyl glycerol, phosphatidyl serine, and other negatively charged phospholipids. These phospholipids are found in various proportions on virtually every cell in the body, on the inner and outer surface membranes. Controversy has arisen as to the significance of these antibodies and whether treatment should be based solely on positive results of aCL or on positive results of any other aPL.

This ongoing debate of the clinical significance of aCL and other aPL has prompted some clinicians to screen recurrent pregnancy loss (RPL) patients and indentify those that might be missed if only aCL were considered significant. For example, Branch et al. analyzed the 95th and the 99th percentiles of the positive and negative cut-off for a panel of phospholipids among 147 women with RPL, APS, and fertile controls.²³ By using the 99th percentile, they found that 26/147 (17.7%) of women with RPL had positive antibodies to CL and 13/147 (8.8%) with RPL demonstrated binding against phospholipids other than CL or lupus anticoagulant (LA). The cut-off value in phospholipid units was determined by using the 99th percentile of the normal population, approximately threefold the median value. Based on comparison with controls, they concluded that this difference was not clinically significant.

In a much larger, earlier study, Yetman and Kutteh determined the prevalence of aPL among 866 women with RPL 5. In this population, 150 of 866 (17.3%) women with RPL were positive for IgG and/or IgM aCL while only 12 of 288 (4%) of control women without a history of poor obstetrical outcome were positive for the same antibodies (p < 0.001). The same study identified 87 of 866 women with RPL who were negative for aCL but positive for one of the other aPL, considering patients with more than one positive

aPL only once.³⁵ Although this study was retrospective, it suggests that a significant number of women with RPL would not have been identified if they had been tested solely for aCL. The same group recently reported on another group of 872 women with RPL. 36 Positive aCL were detected in 132 of 872 women with RPL (15.1%), LA was detected in 31 of 872 (3.6%), and aPS was identified in 49 of 872 (5.6%) of women with RPL who were negative for aCL and LA.³⁶ Anti-PS antibodies were found in the absence of aCL and LA in women with RPL and two consecutive losses (18/391 or 4.6%), women with three consecutive losses (16/288 or 5.6%), and women with four or more consecutive losses (15/193 or 7.8%). In control women without a history of poor obstetric outcome, positive aCL were detected in 4.9%, positive LA in 1.0%, and positive aPS in 2.8%. Differences in aCL and anti-PS when comparing women with RPL to controls were significant using the two-tailed Fisher exact test.

The lack of standardization among different laboratories has made it difficult for physicians to identify patients with APS and those at risk for a miscarriage. 37–39 This has been used as a reason for not using other aPL as APS criteria, but in fact a great deal of variation exists between laboratories even when assaying aCL. For example, IgG aCL, considered by almost all clinicians and laboratory professionals as the 'gold standard', is still not standardized to the level of uniform agreement in all labs and all assays. In 2009, the College of American Pathologists survey results for sample ACL-06 showed that only 78% of labs could even agree that the sample was positive, while 5.5% of the labs determined the sample was negative, and the remaining 16.5% of the labs indicated that the result was indeterminate! Thus, an international group of investigators has established both clinical and laboratory criteria for the diagnosis of APS.¹ Yet, problems still exist when pregnancy loss patients are referred to fertility clinics that may have had testing performed at different laboratories using different control values cut-off values to determine positive results. Also, standard testing may exclude a population of aPL patients who have had significant obstetric problems but test positive for other aPL and negative for the most commonly assayed aCL and LA.

Basic science supports the significance of aPL other than aCL. Anti-PS antibodies have been shown to inhibit trophoblast development and invasion using an in vitro model system.⁴⁰ Anti-PS retard syncytiotrophoblast formation and

decrease the synthesis of hCG. Both low-molecular weight and unfractionated heparin have been shown to reduce the in vitro binding of anti-PS as well as aCL. I Furthermore, some clinical data have been published suggesting that some women with a diagnosis of RPL and aPL positivity may benefit from treatments that have assisted women with RPL and aCL to deliver healthy offspring. In the synthesis of the synthesi

Questions and answers from the task force

1. Do non-criteria aPL exist and are they found in women with RPL?

The task force generally felt that enough studies had been performed on large populations of patients to demonstrate that these 'non-criteria' aPL do indeed exist. ^{23,33–35}

2. Are there sufficient clinical data to warrant a change to the 2006 criteria for the diagnosis of APS?

The task force acknowledged that several studies have suggested that 'non-criteria' aPL may have clinical significance, but that the current level of evidence did not warrant any changes to the current criteria. Obviously, the task force would like to see more prospective, randomized trials, but acknowledged that a number of obstacles exist to make these types of studies difficult. These challenges include both clinical and laboratory inclusion criteria and the need to use an experienced laboratory in a multicenter study.

3. Do women with recurrent pregnancy loss who have predominantly early pregnancy losses (prior to ten gestational weeks) and no history of thrombosis constitute a unique population that warrants different diagnostic criteria to APS?

Considerable discussion on this topic was generated. It was felt that obstetric populations should be stratified to distinguish women based on their history of prior thrombotic events from those without this history. It was also felt that women with predominantly later-trimester losses (beyond 13 gestational weeks) should be distinguished from those women who had losses that were predominantly in the first trimester. This population of women with early pregnancy losses may be affected differently by the non-criteria aPL through mechanisms other than thrombosis. The task force felt that this should receive strong consideration at the next consensus conference.

Recommendations of the task force

The 'non-criteria' aPL task force agreed that studies from several different investigators clearly demonstrate that there are women with RPL who are negative when tested for aCL and LA but who are positive for other 'non-criteria' aPL. In fact, some of the task force members reiterated previous suggestions that women with RPL without a history of thrombosis should be placed in a separate classification when considering the diagnosis of APS, and that a treatment algorithm be constructed to address this group. However, the task force is uncertain and unwilling at this time to make any changes in the current criteria for the diagnosis of APS. It was agreed that some clinical studies show promise and need to be repeated by other groups, as those available do not have enough power to be considered significant. The task force felt that the significance of a panel of aPL antibodies to diagnose APS is an ongoing debate, with many complex questions that can only be addressed with larger study groups using an experienced central laboratory and multiple sites (Table 1a)

IgA anticardiolipin (aCL) and IgA anti-β₂GPI antibodies

(Presented by Dr Murthy on behalf of Dr Pierangeli's group and by Dr Petri)

Introduction and questions addressed by the task force

The current laboratory criteria for APS include the presence of positive lupus anticoagulant (LA) and/or IgG or IgM isotypes of aCL and/or anti- β 2GPI antibodies, but omit the IgA isotypes for both tests. ¹

a) IgA aCL antibodies

Studies have shown data on the prevalence and significance of IgA aCL antibodies. In unselected patients with systemic lupus erythematosus (SLE), the prevalence of increased titers of IgA aCL has been reported to vary from 1% to 44%. ^{43–51} The lowest reported frequency was that found by Selva-O'Callaghan et al., who detected IgA aCL in only 2 of their 200 (1%) patients with SLE. ⁵² Alarcon-Segovia et al., in an earlier study that included 500 patients with SLE, found increased titers of IgA aCL in 16.6% of their patients. ⁵³ In another study, Spadaro et al. found that IgA aCL was positive in 13 (20%) of their 65 SLE patients. ⁵⁴

In contrast, Weidmann et al. found IgA aCL to be positive in 44% of 92 SLE patients and also found IgA to be the most frequent aCL isotype. 45 The reported frequency for raised IgA aCL was higher (52.5%) in an earlier study by Wilson et al., where patients were preselected for being IgG or IgM aCL positive and/or having APS-associated clinical complications. 46 A prevalence of 83.3% was reported by Lopez et al. in a group of patients with SLE and thrombocytopenia.⁴⁷ As noted, the ethnic group composition of patients can influence the isotypic distribution of aCL. Molina et al. studied African-American, Afro-Caribbean, and Hispanic patients with SLE and found elevated levels of IgA aCL in 16%, 21%, and 14%, respectively. 48 The most important finding was that IgA aCL was the only aCL isotype present in 82% of aCL-positive Afro-Caribbean patients. In contrast, IgA aCL was found to be positive only in 4.4% of Chinese patients with SLE.⁴⁹ In another study, Cucurull et al. found that, although IgA aCL antibodies were present in 51% to 55% of patients with APS, most were also IgG or IgM positive, suggesting that measurement of IgA aCL would add little to IgG and IgM determination.⁵⁰

There is some experimental evidence that IgA aCL antibodies are pathogenic. In a mouse model designed to study thrombus formation, injected IgA immunoglobulins with aCL activity from patients with APS were shown to cause thrombosis. The mean thrombus size using two different IgA immunoglobulin preparations was found to be significantly larger compared with control IgA. 55

Numerous studies have also investigated possible associations between raised levels of aCL and clinical manifestations of APS attributed to these autoantibodies. Several of these studies reported a significant association for IgA aCL with one or more of the main clinical manifestations of APS. Cucurull et al., studying both aCL and anti-B2GPI antibodies in African-American patients with SLE, found an association between thrombotic events and raised levels of both these autoantibodies.⁵⁰ However, the number of their patients with thrombotic events was very small: only 5% of their 100 patients had documented evidence of thrombosis. 50 An association between raised IgA aCL levels and thrombocytopenia in patients with SLE or other collagen vascular diseases has also been reported.⁵⁶ Finally, an association between IgA aCL and recurrent fetal loss and with unexplained spontaneous abortions has been reported in women with SLE.⁵⁷ In a study that tested over 700 samples from an APS registry (APSCORE),

only five samples were positive for IgA aCL alone and four of those were from patients who had presented with at least one of the two major manifestations of APS, according to the Sapporo revised criteria (unpublished observations). Furthermore, although the number of APS patients with IgA aCL positive results only – in the absence of IgG and/or IgM aCL-positive results – is low, its presence seems to be associated with clinical manifestations for the APS.⁵⁷ At this preconference workshop, Dr Michelle Petri showed data from her own laboratory, indicating that isolated IgA aCL positivity is rare but is associated with venous and arterial thrombosis.

b) IgA anti- $\beta_2 GPI$ antibodies

Previous studies have raised the possibility that IgA anti-β₂GPI might be associated with clinical manifestations of APS; those observations showed that SLE patients with APS are more prone to be positive for the IgA isotypes. 58-61 Furthermore, it seems that IgA anti-β₂GPI antibodies are independent risk factors of acute myocardial infarction and atherosclerotic disease in populations without APS (OR 3.4, CI 1.3–9.1),⁶² and the same positive association was found for acute cerebral ischemia. 63-66 A concise report by Yamada et al. also showed anti-B2GPI positivity in the absence of IgG antiβ₂GPI in a subgroup of women with unexplained recurrent pregnancy loss (particularly in the first trimester). 67 Similar findings were reported by Lee et al., indicating that IgA anti-β₂GPI positivity is more common in women who experience unexplained recurrent spontaneous abortion and unexplained fetal death and whose initial test results for other isotypes and LA were negative.⁶⁸ Further characterization of IgA anti-β₂GPI positivity in the absence of IgG anti-β₂GPI positivity associated with vascular morbidity showed that these antibodies may recognize domain IV of β_2 GPI as their epitope. ^{69,70} In patients with SLE, the IgA anti-β₂GPI that recognizes domains IV and V seems to be positively correlated with thrombosis. ^{69–71}

Recently, Kumar et al. (from Dr Pierangeli's group) reported five isolated cases of individuals who were *exclusively* positive for IgA anti- β_2 GPI and had concomitant clinical manifestations of APS. Subsequently, Sweiss et al. reported that the presence of isolated IgA anti- β_2 GPI positivity is associated with an increase in thromboembolic events, especially among patients with SLE. In that study – which included only a small group of SLE patients – IgA anti- β_2 GPI was associated with an increased prevalence of morbidities involving

organs of mucosal immunity. ⁷³ IgA anti- β_2 GPI-isolated positivity has also been reported in both scleroderma and autoimmune hepatitis, and it was shown to correlate with both disease severity and endothelial damage. ^{74,75}

This task force further addressed the question whether IgA anti-β₂GPI may have diagnostic value for APS. First, the task force asked attendees of the 13th International Congress on APL antibodies to fill in a survey questionnaire on the use of IgA anti-β₂GPI assays. Thirty responses were returned and, of those who responded, 47% indicated that they routinely order or perform IgA anti-β₂GPI tests in their units; 25% indicated that they find an unusual number of patients with isolated IgA anti-β₂GPI tests; and 83% responded that those isolated IgA anti-β₂GPI are associated with manifestations of APS. Sixty-three percent of the responses indicated that a higher incidence of isolated IgA anti β₂GPI is seen in patients with SLE. Finally, approximately 44% of the responses indicated that IgA anti-β₂GPI tests should be used in confirmation of the diagnosis of APS.

Second, a group of investigators from Dr Pierangeli's laboratory presented data from a recent study where they examined the prevalence of isolated IgA anti-β₂GPI in 588 subjects with SLE from a large, multi-ethnic, multicenter cohort, Lupus in Minorities: Nature vs nurture (LUMINA), in 200 sera from SLE samples provided by Drs Akhther and Petri, and also in the of 5098 individuals referred to Pierangeli's reference clinical laboratory (APLS) for APS work-up between January 2008 and March 2010 and correlated with the presence of APS-related clinical manifestations. The data were presented at this preconference workshop by Dr Murthy. aCL antibodies (IgG, IgM, IgA isotypes) and IgA anti-β₂GPI antibodies were evaluated by ELISA. IgA anti-β₂GPI titers were determined in two commercial FDA-cleared ELISA kits (kits 1 and 2). The binding of the IgA anti- β_2 GPI-positive sera to domains IV/V of IgA anti-β₂GPI was also examined by ELISA. A total of 149 patients were found to be positive for IgA anti-β₂GPI isotype – 80 from LUMINA, 34 from Dr. Petri's cohort, and 35 from the APLS cohort. Of these, 35 from the LUMINA study, 15 from the Petri cohort, and 25 from the APLS cohort were found to be exclusively positive for the anti-β₂GPI isotype while being negative for the other aPL antibodies, including IgA aCL. 70 A significant number of subjects in the three groups had at least one APS-related clinical manifestation (70% in LUMINA, 100% in the Petri

cohort, and 80% in the APLS group). These manifestations included: venous and arterial thrombosis (i.e., deep vein thrombosis, strokes, myocardial infarction); transient ischemic attacks; thrombocytopenia; miscarriages; and other symptoms such as livedo reticularis, pulmonary hypertension, cognitive dysfunction, and seizures. In kits 1 and 2, 86% and 85%, respectively, of IgA anti- β_2 GPI were found to be positive. All samples were positive for IgA anti- β_2 GPI in at least one kit. The correlation between the two kits was found to be 0.93.

In addition, 55% of the IgA anti- β_2 GPI-positive sera (LUMINA and APLS cohorts) reacted with domains IV/V of the β_2 GPI, and 77% of those had clinical manifestations of APS that included deep vein thrombosis, strokes, myocardial infarction, pulmonary hypertension, seizures, pregnancy losses, skin ulcers, and livedo reticularis

In summary, Pierangeli and collaborators showed that a significant proportion of subjects in three different cohorts were positive solely for IgA anti- β_2 GPI, and many of these had clinical manifestations of APS. Their data confirm that isolated IgA anti- β_2 GPI antibody titers may identify additional patients who have clinical features of APS but who do not meet current diagnostic criteria. We also concluded that IgA anti- β_2 GPI antibodies that bind to domains IV/V of β_2 GPI might represent an important subgroup of clinically relevant aPL antibodies.

Dr Petri also presented data at this preconference workshop proving that anti- β_2 GPI of the IgA isotype is associated with thrombosis in SLE patients. In her studies, IgA anti- β_2 GPI was found in 10.2% of SLE patients, and as the sole anti- β_2 GPI isotype in 13.1%. The association of IgA anti- β_2 GPI antibodies with APS manifestations is shown in Table 1. The IgA anti- β_2 GPI antibody was more strongly associated with deep venous thrombosis than the IgM isotype. Second, the specificity of the association was also shown in those with IgA anti- β_2 GPI alone: 22.1% had venous thrombosis and 11.9% had arterial thrombosis. 69,77

Interestingly, discrepant results and significant lack of concordance among different IgA aCL and IgA anti- β_2 GPI assays were obtained during a wet workshop at APLA 2010, when 26 APS samples were tested simultaneously in six different commercial IgA aCL and anti- β_2 GPI assays, indicating that there may be substantial differences in the performance of various IgA assays.

Recommendations by the task force

a) IgA aCL

IgA aCL antibodies appear to be similar to IgG aCL in terms of thrombogenicity and cofactor requirement. Controversies regarding their prevalence and clinical associations still exist, perhaps due to the use of various nonstandardized assays and from differences in the design of the studies. Because of the very small prevalence of IgA aCL positivity alone in the absence of IgG and/or IgM aCL positivity, IgA aCL testing should be recommended in cases where IgG and IgM aCL are negative and there is a strong suspicion of APS.

b) IgA anti-β₂GPI

Based on the published evidence available (April 2010) – thoroughly reviewed by this group – and the studies presented by members of the task force at the preconference workshop at the 13th Congress on Antiphospholipid International Antibodies (APLA 2010), IgA anti-β2GPI antibodies should be tested in the presence of clinical signs and symptoms of SLE and/or APS, particularly when other aPL tests are negative. The group also recognized that well-designed studies, which should include evaluation and comparison of multiple commercially available assays in larger and wellcharacterized populations of patients, are needed in order to confirm the diagnostic value of isolated anti-β2GPI positivity before this test can be included in the diagnostic criteria of APS. The group also recommended that investigation should be carried out to determine the role of IgA anti-β2GPI antibodies in the pathogenesis of APS (Table 1b)

Antiprothrombin antibodies: aPT-A and aPS-PT

(Presented by Drs Bertolaccini, Forastiero, Binder, and Atsumi)

Introduction and questions addressed by the task force

The presence of antibodies solely targeting human prothrombin (aPT-A) by enzyme-linked immunosorbent assay (ELISA) has been recognized since 1995. Several ELISA methods have been reported, 79-84 most of which use irradiated plates and buffers containing detergent (Tween 20), but the use of non-gamma-irradiated plates has also been proposed. The presence of Tween in the

washing buffer enhances the binding of antibodies to the antigen, and this effect was found in both irradiated and nonirradiated microtiter plates. There is an ample variety of commercial microtiter plates and diverse blocking solutions used by different researchers. A major problem is that several in-house methods do not evaluate binding to empty or blank wells of each serum sample in order to assess nonspecific binding. The use of an irrelevant protein such as bovine serum albumin instead of only buffer for coating the control wells improves the performance of the aPT-A assay.⁸⁴ Several methodologic variations were assessed in an attempt to optimize the aPT-A assay:85 the combination of gamma-irradiated plates, phosphate-buffered saline buffer, and a coating antigen of 10 µg/ml prothrombin was found the most sensitive. In recent years, a number of commercial kits for the detection of aPT-A have been made available. In a collaborative study assessing different in-house and commercial anti-PT assays, a good interassay concordance was found for IgG aPT-A using in-house and commercial kits, while IgM results were discordant between assays.86

Anti-PT antibodies bind not only to prothrombin coated on gamma-irradiated or -activated polyvinyl chloride ELISA plates (aPT-A), but also recognize prothrombin exposed to immobilized phosphatidylserine (phospatidylserine-dependent antiprothrombin antibodies, anti-PS/PT). Antiprothrombin antibodies have been detected against prothrombin-bound, hexagonal (II)-phase phosphatidylethanolamine, but this finding has not been fully investigated.

Although aPT-A and/or aPS-PT are associated with APS-related clinical features and these antibodies correlate with each other, aPT-A and aPS-PT belong to different populations of autoantibodies, even though they can both be present in the same patient. 88

A number of studies have been published with regard to the relationship between APS-related clinical features and the presence of aPT-A, with conflicting conclusions. High levels of aPT-A were found to confer a high risk of myocardial infarction in dyslipidemic middle-aged men without autoimmune disease. Although no association between aPT-A and the risk of thrombosis was found in a systematic review, there are some data suggesting that aPT-A are likely a risk factor of recurrent venous thromboembolism. The majority of these studies were retrospective, and this fact makes it difficult to draw definite conclusions. Although no association between aPT-A are likely a risk factor of recurrent venous thromboembolism.

prospective studies have shown for the first time that the presence of aPT-A is a predictor of first or recurrent thrombosis in aPL patients. 94,95 The results of a 15-year longitudinal study showed that IgG aPT-A is the most useful predictor of thrombosis in SLE patients. In addition, an important observation reported by several recent studies is that the risk of thrombosis progressively increases with the number of positive aPL tests. The quadruple positivity of lupus anticoagulant, aCL, anti- β_2 GPI antibodies, and aPT-A seems to confer the highest risk for thrombosis. 96

Many reports have also shown the clinical utility of anti-PS/PT assay for the diagnosis of APS.88 Galli et al. 89 showed aPS-PT in 95% of their patients with thrombosis, but no differences in prevalence were found between those patients with thrombosis and those without. Funke et al. 9 reported that aPS-PT conferred an odds ratio of 2.8:1 for venous thrombosis and of 4.1:1 for arterial thrombosis in patients with SLE. Atsumi et al. 93 supported these data by showing that the presence of aPS-PT conferred an odds ratio of 3.6:1 for APS in 265 Japanese patients with systemic autoimmune diseases. Bertolaccini et al. 88 the association between confirmed (IgG and/or IgM isotype) and arterial and/or venous thrombosis. Both sensitivity and specificity of aPS-PT for the diagnosis of APS have been shown to be higher than that of aCL. In addition, aPS-PT strongly correlates with the LA, also suggesting that anti-PS/PT may be one of the 'screening' or 'confirming' assays for APS-associated LA. 93,98

Recommendations of the task force

Based on the evidence published in recent years, it appears that the detection of aPT-A in conjunction with the other aPL tests could be useful in the consideration of risk for thrombosis.

The task force members agreed that anti-PT antibody assay — in particular, anti-PS/PT — would potentially contribute to a better recognition of APS. However, the inclusion of anti-PT antibodies as one of the laboratory criteria of APS cannot be warranted at this time, mainly due to poor standardization of aPT-A and/or anti-PS/PT.

Reproducibility of such strong correlations between anti-PS/PT and APS manifestations, which were presented by some investigators, should be confirmed by the collaboration design. A multicentre study was proposed during the workshop discussion, and is currently being designed by task force members (Table 1b)

The annexin A5 resistance test: a mechanistic test for the detection of pathogenic aPL antibodies

(Presented by Dr Rand)

Introduction and questions addressed by the task force

Dr Jacob Rand from the Montefiore Medical Center, New York presented data on the annexin A5 resistance (A5R) test. Dr. Rand provided the committee with a brief historic background on current aPL tests - the aPL immunoassays and the lupus anticoagulant assays - all of which were derived empirically and do not report on thrombogenic mechanisms. The Rand laboratory has developed a novel functional assay that measures a disease mechanism - aPL antibody-mediated disruption of an anticoagulant shield that is composed of annexin A5 (AnxA5). The assay is based on the concept that AnxA5 has potent anticoagulant properties that result from its forming 2-dimensional crystals over phospholipids, blocking the availability of the phospholipids for critical coagulation enzyme reactions. 100-102 Previous research over the past 17 years has yielded strong evidence that aPL antibodies can disrupt this anticoagulant shield and unmask thrombogenic anionic phospholipids, which may thereby contribute to thrombosis and pregnancy complications in patients with APS. 103-107 The A5R assay is a 2-stage coagulation assay that mimics this mechanism on phospholipid suspensions. ^{108–110} The assay measures the effect of patient plasma on the anticoagulant activity of AnxA5; results are reported as percentage prolongation of the coagulation time by AnxA5; patients with percentages lower than the reference range are considered to have AnxA5 resistance. Remarkably, resistance to AnxA5 anticoagulant activity has been correlated with aPL antibodies that recognize an epitope on domain I of β₂GPI.¹⁰⁹ Dr Rand provided details on the methodology and, with Dr Xiao-Xuan Wu, demonstrated the assay in the meeting's wet laboratory demonstration session. The assay is labour intensive and, as mentioned above, requires a 2-stage procedure in which the first stage exposes the phospholipid suspension to patient plasma, and the suspension is then centrifuged and washed for the second stage in which the phospholipid is used to coagulate a normal pooled plasma.

Dr Rand presented the task force with data collected from five studies on coded samples from

597 patients – all of which were obtained from collaborators at outside institutions. The available evistrongly supports the utility of this mechanistic assay in defining a subgroup of patients in whom this disease mechanism occurs. The pooled data indicated that about half (52%) of patients with symptomatic APS by current consensus criteria have AnxA5 resistance, whereas 2-5% of disease-free controls and patients with non-APS thrombosis have that abnormality. Interestingly, 27% of patients who tested positive for aPL antibodies but did not have a history for thrombosis also tested positive for AnxA5 resistance. Since many of the latter were patients with autoimmune conditions such as SLE, Dr Rand hypothesized that these patients might have an increased risk for future thrombosis – a concept that would need to be validated in prospective longitudinal observational studies.

Recommendations of the task force

The task force committee concluded that data on the utility of AnxA5 resistance assay as a mechanistic diagnostic marker for APS are highly promising. The committee also felt that the concept of developing mechanistic clinical assays that measure APS disease mechanisms was an important and appropriate avenue to pursue. The committee would like to see additional data before recommending A5R as a standard component of aPL testing panels. In addition, the assay needs to be made available for other centers to be tested before any recommendation can be made (Table 1b).

Acknowledgements

All authors participated equally in the preparation of this manuscript

The following collaborators participated in the studies presented by Dr Murthy: R Aguilar-Valenzuela¹, LA MartÍnez-MartÍnez¹, V Murthy¹, S Jatwani¹, AM Seif², GS Alarcón¹, E Papalardo², J Liu⁴, LM Vila¹, S Najam¹, T McNearney¹, EB Gonzalez¹, R Maganti⁶, W Binder⁵, M Teodorescu³, JD Reveille⁶, R Willis⁷, J Tarantino⁸, M Petri⁸, and E Akhter⁸.

¹University of Texas Medical Branch, Galveston, Texas, USA; ²University of Alabama at Birmingham, Birmingham, Alabama, USA; ³University of Texas–Houston Health Sciences Center, Houston, Texas, USA; ⁴University of Puerto Rico Medical Sciences Campus, San Juan, Puerto Rico; ⁵Theratest Laboratories, Lombard, Illinois, USA; ⁶INOVA Diagnostics, San Diego, California, USA; ⁶Department of Microbiology,

University of the West Indies, Kingston, Jamaica; ⁷Comprehensive Bleeding Disorders, Peoria, Illinois, USA; ⁸John Hopkins School of Medicine, Baltimore, Maryland, USA.

Funding

ML Bertolaccini is funded by the Louise Gergel Fellowship; B de Laat is funded by a personal grant from the Netherlands Heart Foundation (grant number NHS2006T053).

JH Rand is funded by grants from the National Institutes of Health, National Heart, Lung and Blood Institute (RC1 HL101031 and R01 HL061331).

Conflict of interest statement

None declared.

References

- 1 Miyakis S, Lockshin MD, Atsumi T, et al. International consensus statement on an update of the classification criteria for definite antiphospholipid syndrome (APS). J Thromb Haemost 2006; 4: 295-306
- 2 Bertolaccini ML, Hughes GR, Khamashta MA. Revisiting antiphospholipid antibodies: from targeting phospholipids to phospholipid binding proteins. Clin Lab 2004; 50: 653–665.
- 3 Gris JC, Quere I, Sanmarco M, et al. Antiphospholipid and antiprotein syndromes in non-thrombotic, non-autoimmune women with unexplained recurrent primary early foetal loss. The Nimes Obstetricians and Haematologists Study – NOHA. Thromb Haemost 2000; 84: 228–236.
- 4 Sugi T, Matsubayashi H, Inomo A, Dan L, Makino T. Antiphosphatidylethanolamine antibodies in recurrent early pregnancy loss and mid-to-late pregnancy loss. *J Obstet Gynaecol Res* 2004; 30: 326–332.
- 5 Sanmarco M, Gayet S, Alessi MC, et al. Antiphosphatidylethanolamine antibodies are associated with an increased odds ratio for thrombosis. A multicenter study with the participation of the European Forum on antiphospholipid antibodies. *Thromb Haemost* 2007; 97: 949–954.
- 6 Sanmarco M. ELISA for antiphosphatidylethanolamine antibody detection: high impact of assay buffer on results. *J Immunol Methods* 2010; 358: 9–16.
- 7 de Laat B, Mertens K, de Groot PG. Mechanisms of disease: antiphospholipid antibodies from clinical association to pathologic mechanism. *Nat Clin Pract Rheumatol* 2008; 4: 192–199.
- 8 de Laat B, Derksen RH, van Lummel M, Pennings MT, de Groot PG. Pathogenic anti-beta2-glycoprotein I antibodies recognize domain I of beta2-glycoprotein I only after a conformational change. *Blood* 2006; 107: 1916–1924.
- 9 Iverson GM, Victoria EJ, Marquis DM. Anti-beta2 glycoprotein I (beta2GPI) autoantibodies recognize an epitope on the first domain of beta2GPI. Proc Natl Acad Sci U S A 1998; 95: 15542–15546.
- 10 Iverson GM, Reddel S, Victoria EJ, et al. Use of single point mutations in domain I of beta 2-glycoprotein I to determine fine antigenic specificity of antiphospholipid autoantibodies. J Immunol 2002: 169: 7097–7103.
- 11 Ioannou Y, Pericleous C, Giles I, Latchman DS, Isenberg DA, Rahman A. Binding of antiphospholipid antibodies to

- discontinuous epitopes on domain I of human beta(2)-glycoprotein I: mutation studies including residues R39 to R43. *Arthritis Rheum* 2007; 56: 280–290.
- 12 de Laat B, Derksen RH, Urbanus RT, de Groot PG. IgG antibodies that recognize epitope Gly40-Arg43 in domain I of beta 2-glycoprotein I cause LAC, and their presence correlates strongly with thrombosis. *Blood* 2005; 105: 1540-1545.
- 13 de Laat B, Pengo V, Pabinger I, et al. The association between circulating antibodies against domain I of beta2-glycoprotein I and thrombosis: an international multicenter study. J Thromb Haemost 2009; 7: 1767–1773.
- 14 Ioannou Y, Romay-Penabad Z, Pericleous C, et al. In vivo inhibition of antiphospholipid antibody-induced pathogenicity utilizing the antigenic target peptide domain I of beta2-glycoprotein I: proof of concept. J Thromb Haemost 2009; 7: 833–842.
- 15 Petri M. The lupus anticoagulant is a risk factor for myocardial infarction (but not atherosclerosis): Hopkins Lupus Cohort. *Thromb Res* 2004; 114: 593–595.
- 16 Finazzi G, Brancaccio V, Moia M, et al. Natural history and risk factors for thrombosis in 360 patients with antiphospholipid antibodies: a four-year prospective study from the Italian Registry. Am J Med 1996; 100: 530-536.
- 17 Abu-Shakra M, Gladman DD, Urowitz MB, Farewell V. Anticardiolipin antibodies in systemic lupus erythematosus: clinical and laboratory correlations. Am J Med 1995; 99: 624-628.
- 18 Ahmed E, Stegmayr B, Trifunovic J, Weinehall L, Hallmans G, Lefvert AK. Anticardiolipin antibodies are not an independent risk factor for stroke: An incident case-referent study nested within the MONICA and Vasterbotten cohort project. Stroke 2000; 31: 1289–1293.
- 19 Brey RL, Abbott RD, Curb JD, et al. Beta(2)-glycoprotein 1-dependent anticardiolipin antibodies and risk of ischemic stroke and myocardial infarction: the Honolulu Heart Program. Stroke 2001; 32: 1701–1706.
- 20 van Goor MP, Alblas CL, Leebeek FW, Koudstaal PJ, Dippel DW. Do antiphospholipid antibodies increase the long-term risk of thrombotic complications in young patients with a recent TIA or ischemic stroke? *Acta Neurol Scand* 2004; 109: 410-415.
- 21 Ginsburg KS, Liang MH, Newcomer L, et al. Anticardiolipin antibodies and the risk for ischemic stroke and venous thrombosis. Ann Intern Med 1992; 117: 997–1002.
- 22 Laroche P, Berard M, Rouquette AM, Desgruelle C, Boffa MC. Advantage of using both anionic and zwitterionic phospholipid antigens for the detection of antiphospholipid antibodies. Am J Clin Pathol 1996; 106: 549-554.
- 23 Branch DW, Silver R, Pierangeli S, van Leeuwen I, Harris EN. Antiphospholipid antibodies other than lupus anticoagulant and anticardiolipin antibodies in women with recurrent pregnancy loss, fertile controls, and antiphospholipid syndrome. *Obstet Gynecol* 1997; 89: 549–555.
- 24 Bertolaccini ML, Roch B, Amengual O, Atsumi T, Khamashta MA, Hughes GRV. Multiple antiphospholipid tests do not increase the diagnostic yield in antiphospholipid syndrome. *Br J Rheumatol* 1998; 37: 1229–1232.
- 25 Fialova L, Mikulikova L, Matous-Malbohan I, Benesova O, Zwinger A. Prevalence of various antiphospholipid antibodies in pregnant women. *Physiol Res* 2000; 49: 299–305.
- 26 Franklin RD, Kutteh WH. Antiphospholipid antibodies (APA) and recurrent pregnancy loss: treating a unique APA positive population. *Hum Reprod* 2002; 17: 2981–2985.
- 27 Amoroso A, Mitterhofer AP, Del Porto F, *et al.* Antibodies to anionic phospholipids and anti-beta2-GPI: association with thrombosis and thrombocytopenia in systemic lupus erythematosus. *Hum Immunol* 2003; 64: 265–273.
- 28 Gharavi AE, Harris EN, Asherson RA, Hughes GR. Anticardiolipin antibodies: isotype distribution and phospholipid specificity. *Ann Rheum Dis* 1987; 46: 1–6.
- 29 Tebo AE, Jaskowski TD, Phansalkar AR, Litwin CM, Branch DW, Hill HR. Diagnostic performance of phospholipid-specific assays for the evaluation of antiphospholipid syndrome. Am J Clin Pathol 2008; 129: 870–875.
- 30 Tebo AE, Jaskowski TD, Hill HR, Branch DW. Clinical relevance of multiple antibody specificity testing in anti-phospholipid

- syndrome and recurrent pregnancy loss. Clin Exp Immunol 2008; 154; 332–338.
- 31 Campbell AL, Pierangeli SS, Wellhausen S, Harris EN. Comparison of the effects of anticardiolipin antibodies from patients with the antiphospholipid syndrome and with syphilis on platelet activation and aggregation. *Thromb Haemost* 1995; 73: 529–534
- 32 Gharavi AE, Pierangeli SS. Origin of antiphospholipid antibodies: Induction of aPL by viral peptides. *Lupus* 1998; Suppl. 2: S52–S54.
- 33 Matzner W, Chong P, Xu G, Ching W. Characterization of antiphospholipid antibodies in women with recurrent spontaneous abortions. J Reprod Med 1994; 39: 27–30.
- 34 Gilman-Sachs A, Lubinski J, Beer AE, Brend S, Beaman KD. Patterns of anti-phospholipid antibody specificities. J Clin Lab Immunol 1991; 35: 83–88.
- 35 Yetman DL, Kutteh WH. Antiphospholipid antibody panels and recurrent pregnancy loss: prevalence of anticardiolipin antibodies compared with other antiphospholipid antibodies. *Fertil Steril* 1996; 66: 540–546.
- 36 Kutteh WH, Corey A, Jaslow CR. Antiphospholipid antibodies and recurrent pregnancy loss: Prevalence of anticardiolipin antibodies, the lupus anticoagulant, and antiphosphatidyl serine antibodies. *Lupus* 2010; 19: C156 (abstract).
- 37 Peaceman AM, Silver RK, MacGregor SN, Socol ML. Interlaboratory variation in antiphospholipid antibody testing. Am J Obstet Gynecol 1992; 166: 1780-1784; discussion 4-7.
- 38 Carreras LO, Forastiero RR, Martinuzzo ME. Which are the best biological markers of the antiphospholipid syndrome? J Autoimmun 2000; 15: 163–172.
- 39 Kutteh WH, Franklin RD. Assessing the variation in antiphospholipid antibody (APA) assays: comparison of results from 10 centers. Am J Obstet Gynecol 2004; 191: 440–448.
- 40 Katsuragawa H, Kanzaki H, Inoue T, Hirano T, Mori T, Rote NS. Monoclonal antibody against phosphatidylserine inhibits in vitro human trophoblastic hormone production and invasion. *Biol Reprod* 1997; 56: 50-58.
- 41 Franklin RD, Kutteh WH. Effects of unfractionated and low molecular weight heparin on antiphospholipid antibody binding in vitro. *Obstet Gynecol* 2003; 101: 455–462.
- 42 Branch DW, Khamashta MA. Antiphospholipid syndrome: obstetric diagnosis, management and controversies. Obstet Gynecol 2003; 101: 1333–1344.
- 43 Greco TP, Amos MD, Conti-Kelly AM, Naranjo JD, Ijdo JW. Testing for the antiphospholipid syndrome: importance of IgA anti-beta 2-glycoprotein I. *Lupus* 2000; 9: 33–41.
- 44 Wilson WA, Faghiri Z, Taheri F, Gharavi AE. Significance of IgA antiphospholipid antibodies. *Lupus* 1998; 7: S110–S113.
- 45 Weidmann CE, Wallace DJ, Peter JB, Knight PJ, Bear MB, Klinenberg JR. Studies of IgG, IgM and IgA antiphospholipid antibody isotypes in systemic lupus erythematosus. *J Rheumatol* 1988; 15: 74-79.
- 46 Tajima C, Suzuki Y, Mizushima Y, Ichikawa Y. Clinical significance of immunoglobulin A antiphospholipid antibodies: possible association with skin manifestations and small vessel vasculitis. *J Rheumatol* 1998; 25: 1730–1736.
- 47 Lopez LR, Santos ME, Espinoza LR, La Rosa FG. Clinical significance of immunoglobulin A versus immunoglobulins G and M anti-cardiolipin antibodies in patients with systemic lupus erythematosus. Correlation with thrombosis, thrombocytopenia, and recurrent abortion. *Am J Clin Pathol* 1992; 98: 449–454.
- 48 Molina JF, Gutierrez-Urena S, Molina J, et al. Variability of anticardiolipin antibody isotype distribution in 3 geographic populations of patients with systemic lupus erythematosus. *J Rheumatol* 1997; 24: 291–296.
- 49 Wong KL, Liu HW, Ho K, Chan K, Wong R. Anticardiolipin antibodies and lupus anticoagulant in Chinese patients with systemic lupus erythematosus. *J Rheumatol* 1991; 18: 1187–1192.
- 50 Cucurull E, Gharavi AE, Diri E, Mendez E, Kapoor D, Espinoza LR. IgA anticardiolipin and anti-beta2-glycoprotein I are the most prevalent isotypes in African American patients with systemic lupus erythematosus. Am J Med Sci 1999; 318: 55-60.

- 51 Kalunian KC, Peter JB, Middlekauff HR, *et al.* Clinical significance of a single test for anti-cardiolipin antibodies in patients with systemic lupus erythematosus. *Am J Med* 1988; 85: 602–608.
- 52 Selva-O'Callaghan A, Ordi-Ros J, Monegal-Ferran F, Martinez N, Cortes-Hernandez F, Vilardell-Tarres M. IgA anticardiolipin antibodies relation with other antiphospholipid antibodies and clinical significance. *Thromb Haemost* 1998; 79: 282–285.
- 53 Alarcon-Segovia D, Delezé M, Oria CV, et al. Antiphospholipid antibodies and the antiphospholipid syndrome in systemic lupus erythematosus: a prospective analysis of 500 consecutive patients. *Medicine (Baltimore)* 1989; 68: 353–365.
- 54 Spadaro A, Riccieri V, Terracina S, Rinaldi T, Taccari E, Zoppini A. Class specific rheumatoid factors and antiphospholipid syndrome in systemic lupus erythematosus. *Lupus* 2000; 9: 56–60.
- 55 Pierangeli SS, Liu XW, Barker JH, Anderson G, Harris EN. Induction of thrombosis in a mouse model by IgG, IgM and IgA immunoglobulins from patients with the antiphospholipid syndrome. Thromb Haemost 1995; 74: 1361–1367.
- 56 Lakos G, Kiss E, Regeczy N, et al. Isotype distribution and clinical relevance of anti-beta2-glycoprotein I (beta2-GPI) antibodies: importance of IgA isotype. Clin Exp Immunol 1999; 117: 574–579.
- 57 Lee RM, Brown MA, Branch DW, Ward K, Silver RM. Anticardiolipin and anti-beta2-glycoprotein-I antibodies in preeclampsia. Obstet Gynecol 2003; 102: 294–300.
- 58 Diri E, Cucurull E, Gharavi AE, *et al.* Antiphospholipid (Hughes') syndrome in African-Americans: IgA aCL and abeta2 glycoprotein-I is the most frequent isotype. *Lupus* 1999; 8: 263–268.
- 59 Bertolaccini ML, Atsumi T, Escudero-Contreras A, Khamashta MA, Hughes GRV. The value of IgA antiphospholipid testing for the diagnosis of antiphospholipid (Hughes) syndrome in systemic lupus erythematosus. *J Rheumatol* 2001; 28: 2637–2643.
- 60 Danowski A, Kickler TS, Petri M. Anti-beta2-glycoprotein I: prevalence, clinical correlations, and importance of persistent positivity in patients with antiphospholipid syndrome and systemic lupus erythematosus. J Rheumatol 2006; 33: 1775–1779.
- 61 Petri M. Update on anti-phospholipid antibodies in SLE: the Hopkins' Lupus Cohort. *Lupus* 2010; 19: 419–423.
- 62 Staub HL, Norman GL, Crowther T, et al. Antibodies to the atherosclerotic plaque components beta2-glycoprotein I and heat-shock proteins as risk factors for acute cerebral ischemia. Arq Neuropsiquiatr 2003; 61: 757–763.
- 63 Staub HL, Franck M, Ranzolin A, Norman GL, Iverson GM, von Muhlen CA. IgA antibodies to beta2-glycoprotein I and atherosclerosis. Autoimmun Rev 2006; 6: 104–106.
- 64 Ranzolin A, Bohn JM, Norman GL, et al. Anti-beta2-glycoprotein I antibodies as risk factors for acute myocardial infarction. Arg Bras Cardiol 2004; 83: 37–40; 141–144.
- 65 Staub HL, von Muhlen CA, Norman GL. Beta2-glycoprotein I IgA antibodies and ischaemic stroke. *Rheumatology (Oxford)* 2006; 45: 645-646. (author reply 6.
- 66 Franck M, Staub HL, Petracco JB, et al. Autoantibodies to the atheroma component beta2-glycoprotein I and risk of symptomatic peripheral artery disease. Angiology 2007; 58: 295–302.
- 67 Yamada H, Tsutsumi A, Ichikawa K, Kato EH, Koike T, Fujimoto S. IgA-class anti-beta2-glycoprotein I in women with unexplained recurrent spontaneous abortion. *Arthritis Rheum* 1999; 42: 2727–2728.
- 68 Lee RM, Branch DW, Silver RM. Immunoglobulin A anti-beta2-glycoprotein antibodies in women who experience unexplained recurrent spontaneous abortion and unexplained fetal death. Am J Obstet Gynecol 2001; 185: 748–753.
- 69 Akhter E, Binder W, Shums Z, Magder L, Petri M. Novel antiphospholipid antibodies and stroke in SLE. *Lupus* 2010; 19: C124 (abstract)
- 70 Martinez-Martinez LA, Aguilar-Valenzuela R, Seif A, Binder W, Alarcon GS, Pierangeli S. Do clinically relevant IgA anti-b2glyco-protein I (anti-b2GPI) antibodies bind to DIV/V of b2GPI? *Lupus* 2010; 19: C130 (abstract).
- 71 Iverson GM, von Muhlen CA, Staub HL, Lassen AJ, Binder W, Norman GL. Patients with atherosclerotic syndrome, negative in anti-cardiolipin assays, make IgA autoantibodies that preferentially target domain 4 of beta2-GPI. J Autoimmun 2006; 27: 266-271.

- 72 Kumar S, Papalardo E, Sunkureddi P, Najam S, Gonzalez EB, Pierangeli SS. Isolated elevation of IgA anti-beta2glycoprotein I antibodies with manifestations of antiphospholipid syndrome: a case series of five patients. *Lupus* 2009; 18: 1011–1014.
- 73 Sweiss NJ, Bo R, Kapadia R, et al. IgA anti-beta2-glycoprotein I autoantibodies are associated with an increased risk of thromboembolic events in patients with systemic lupus erythematosus. *PLoS One* 2010; 5.
- 74 Gabeta S, Norman GL, Gatselis N, et al. IgA anti-b2GPI antibodies in patients with autoimmune liver diseases. J Clin Immunol 2008; 28: 501–511.
- 75 Boin F, Franchini S, Colantuoni E, Rosen A, Wigley FM, Casciola-Rosen L. Independent association of antibeta(2)-glycoprotein I antibodies with macrovascular disease and mortality in scleroderma patients. *Arthritis Rheum* 2009; 60: 2480–2489.
- 76 Seif A, Alarcón GS, Aguilar-Valenzuela R, et al. Are IgA antib2glycoprotein I clinically relevant? Lupus 2010; 19: C133 (abstract).
- 77 Mehrani T and Petri M. Association of IgA Anti-b2 glycoprotein I with clinical and laboratory manifestations of systemic lupus erythematosus. J Rheumatol 2010[Epub ahead of print].
- 78 Arvieux J, Darnige L, Caron C, Reber G, Bensa JC, Colomb MG. Development of an ELISA for autoantibodies to prothrombin showing their prevalence in patients with lupus anticoagulants. *Thromb Haemost* 1995; 74: 1120–1125.
- 79 Pengo V, Biasolo A, Brocco T, Tonetto S, Ruffatti A. Autoantibodies to phospholipid-binding plasma proteins in patients with thrombosis and phospholipid reactive antibodies. *Thromb Haemost* 1996; 75: 721–724.
- 80 Bertolaccini ML, Atsumi T, Khamashta MA, Amengual O, Hughes GR. Autoantibodies to human prothrombin and clinical manifestations in 207 patients with systemic lupus erythematosus. *J Rheumatol* 1998; 25: 1104–1108.
- 81 Vaarala O, Puurunen M, Manttari M, Manninen V, Aho K, Palosuo T. Antibodies to prothrombin imply a risk of myocardial infarction in middle-aged men. *Thromb Haemost* 1996; 75: 456-459.
- 82 Horbach DA, van Oort E, Donders RC, Derksen RH, de Groot PG. Lupus anticoagulant is the strongest risk factor for both venous and arterial thrombosis in patients with systemic lupus erythematosus. Comparison between different assays for the detection of antiphospholipid antibodies. *Thromb Haemost* 1996; 76: 916–924.
- 83 Galli M, Beretta G, Daldossi M, Bevers EM, Barbui T. Different anticoagulant and immunological properties of anti-prothrombin antibodies in patients with antiphospholipid antibodies. *Thromb Haemost* 1997; 77: 486–491.
- 84 Forastiero RR, Martinuzzo ME, Cerrato GS, Kordich LC, Carreras LO. Relationship of anti beta2-glycoprotein I and anti prothrombin antibodies to thrombosis and pregnancy loss in patients with antiphospholipid antibodies. *Thromb Haemost* 1997; 78: 1008–1014.
- 85 Donohoe S, Mackie IJ, Isenberg D, Machin SJ. Anti-prothrombin antibodies: assay conditions and clinical associations in the anti-phospholipid syndrome. *Br J Haematol* 2001; 113: 544–549.
- 86 Tincani A, Morozzi G, Afeltra A, et al. Antiprothrombin antibodies: a comparative analysis of homemade and commercial methods. A collaborative study by the Forum Interdisciplinare per la Ricerca nelle Malattie Autoimmuni (FIRMA). Clin Exp Rheumatol 2007; 25: 268–274.
- 87 Rauch J, Tannenbaum M, Neville C, Fortin PR. Inhibition of lupus anticoagulant activity by hexagonal phase phosphatidyleth-anolamine in the presence of prothrombin. *Thromb Haemost* 1998; 80: 936–941.
- 88 Bertolaccini ML, Atsumi T, Koike T, Hughes GR, Khamashta MA. Antiprothrombin antibodies detected in two different assay systems. Prevalence and clinical significance in systemic lupus erythematosus. *Thromb Haemost* 2005; 93: 289–297.
- 89 Galli M, Luciani D, Bertolini G, Barbui T. Anti-beta 2-glycoprotein I, antiprothrombin antibodies, and the risk of thrombosis in the antiphospholipid syndrome. *Blood* 2003; 102: 2717–2723.

- 90 Zanon E, Saggiorato G, Ramon R, Girolami A, Pagnan A, Prandoni P. Anti-prothrombin antibodies as a potential risk factor of recurrent venous thromboembolism. *Thromb Haemost* 2004; 91: 255–258.
- 91 Palosuo T, Virtamo J, Haukka J, *et al.* High antibody levels to prothrombin imply a risk of deep venous thrombosis and pulmonary embolism in middle-aged men a nested case-control study. *Thromb Haemost* 1997; 78: 1178–1182.
- 92 Tsutsumi A, Hayashi T, Chino Y, et al. Significance of antiprothrombin antibodies in patients with systemic lupus erythematosus: clinical evaluation of the antiprothrombin assay and the antiphosphatidylserine/prothrombin assay, and comparison with other antiphospholipid antibody assays. Mod Rheumatol 2006; 16: 158–164.
- 93 Atsumi T, Ieko M, Bertolaccini ML, et al. Association of autoantibodies against the phosphatidylserine-prothrombin complex with manifestations of the antiphospholipid syndrome and with the presence of lupus anticoagulant. Arthritis Rheum 2000; 43: 1982–1993.
- 94 Forastiero R, Martinuzzo M, Pombo G, et al. A prospective study of antibodies to beta2-glycoprotein I and prothrombin, and risk of thrombosis. J Thromb Haemost 2005; 3: 1231–1238.
- 95 Bizzaro N, Ghirardello A, Zampieri S, et al. Anti-prothrombin antibodies predict thrombosis in patients with systemic lupus erythematosus: a 15-year longitudinal study. J Thromb Haemost 2007; 5: 1158-1164
- 96 Forastiero R, Martinuzzo M, Iglesias Varela ML, Cerrato G. Multiple (triple or quadruple) aPL positivity is much more prevalent in patients with definite antiphospholipid syndrome. *Lupus* 2010; 19: C109 (abstract).
- 97 Funke A, Bertolaccini ML, Atsumi T, Amengual O, Khamashta MA, Hughes GRV. Autoantibodies to prothrombin-phosphatidylserine complex: clinical significance in systemic lupus erythematosus. Arthritis Rheum 1998; 41: S240 (abstract).
- 98 Atsumi T, Koike T. Antiprothrombin antibody: why do we need more assays? *Lupus* 2010; 19: 436–439.
- 99 Binder W, Lewis S, Shums Z. Clinical significance of IgG and IgM autoantibodies that target the complex of phosphatidylserine and prothrombin. *Lupus* 2010; 19: C134 (abstract).

- 100 Gerke V, Moss SE. Annexins: from structure to function. *Physiol Rev* 2002; 82: 331–371.
- 101 Mortimer JC, Laohavisit A, Macpherson N, et al. Annexins: multifunctional components of growth and adaptation. J Exp Bot 2008: 59: 533-544
- 102 Tait JF, Sakata M, McMullen BA, et al. Placental anticoagulant proteins: isolation and comparative characterization of four members of the lipocortin family. *Biochemistry* 1988; 27: 6268–6276.
- 103 Rand JH, Wu XX, Andree HA, et al. Antiphospholipid antibodies accelerate plasma coagulation by inhibiting annexin-V binding to phospholipids: a "lupus procoagulant" phenomenon. Blood 1998; 92: 1652–1660.
- 104 Rand JH, Wu XX, Quinn AS, et al. Human monoclonal antiphospholipid antibodies disrupt the annexin A5 anticoagulant crystal shield on phospholipid bilayers: evidence from atomic force microscopy and functional assay. Am J Pathol 2003; 163: 1193–1200.
- 105 Hanly JG, Smith SA. Anti-beta2-glycoprotein I (GPI) autoantibodies, annexin V binding and the anti-phospholipid syndrome. Clin Exp Immunol 2000; 120: 537–543.
- 106 Gaspersic N, Ambrozic A, Bozic B, Majhenc J, Svetina S, Rozman B. Annexin A5 binding to giant phospholipid vesicles is differentially affected by anti-beta2-glycoprotein I and anti-annexin A5 antibodies. *Rheumatology (Oxford)* 2007; 46: 81–86.
 107 Tomer A, Bar-Lev S, Fleisher S, Shenkman B, Friger M,
- 107 Tomer A, Bar-Lev S, Fleisher S, Shenkman B, Friger M, Abu-Shakra M. Antiphospholipid antibody syndrome: the flow cytometric annexin A5 competition assay as a diagnostic tool. Br J Haematol 2007; 139: 113–120.
- 108 Rand JH, Wu XX, Lapinski R, et al. Detection of antibodymediated reduction of annexin A5 anticoagulant activity in plasmas of patients with the antiphospholipid syndrome. Blood 2004; 104: 2783–2790.
- 109 de Laat B, Wu XX, van Lummel M, Derksen RH, de Groot PG, Rand JH. Correlation between antiphospholipid antibodies that recognize domain I of beta2-glycoprotein I and a reduction in the anticoagulant activity of annexin A5. *Blood* 2007; 109: 1490–1494.
- 110 Rand JH, Arslan AA, Wu XX, et al. Reduction of circulating annexin A5 levels and resistance to annexin A5 anticoagulant activity in women with recurrent spontaneous pregnancy losses. Am J Obstet Gynecol 2006; 194: 182–188.

Novel Assays of Thrombogenic Pathogenicity in the Antiphospholipid Syndrome Based on the Detection of Molecular Oxidative Modification of the Major Autoantigen β_2 -Glycoprotein I

Yiannis Ioannou,¹ Jing-Yun Zhang,² Miao Qi,³ Lu Gao,² Jian Cheng Qi,³ De-Min Yu,⁴ Herman Lau,³ Allan D. Sturgess,³ Panayiotis G. Vlachoyiannopoulos,⁵ Haralampos M. Moutsopoulos,⁵ Anisur Rahman,⁶ Charis Pericleous,⁶ Tatsuya Atsumi,⁷ Takao Koike,⁷ Stephane Heritier,⁸ Bill Giannakopoulos,³ and Steven A. Krilis³

Objective. Beta-2-glycoprotein I (β_2 GPI) constitutes the major autoantigen in the antiphospholipid syndrome (APS), a common acquired cause of arterial and venous thrombosis. We recently described the novel observation that β_2 GPI may exist in healthy individuals in a free thiol (biochemically reduced) form. The present study was undertaken to quantify the levels of total,

Dr. Ioannou's work was supported by Arthritis Research UK (Clinician Scientist Fellowship award, grant 17821). Dr. Zhang's work was supported by Tianjin Medical University, Tianjin, China. Dr. Pericleous' work was supported by Arthritis Research UK (project grant 18491). Dr. Krilis' work was supported by grants from the National Health and Medical Research Council of Australia.

National Health and Medical Research Council of Australia.

¹Yiannis Ioannou, PhD, MRCP: St. George Hospital and University of New South Wales, Sydney, New South Wales, Australia and University College London, London, UK; ²Jing-Yun Zhang, MD, PhD, Lu Gao, MD, PhD: St. George Hospital and University of New South Wales, Sydney, New South Wales, Australia and Metabolic Disease Hospital and Tianjin Medical University, Tianjin, China; ³Miao Qi, MSc, Jian Cheng Qi, MD, PhD, Herman Lau, MBBS, FRACP, Allan D. Sturgess, MBBS, FRACP, PhD, Bill Giannako-poulos, MBBS, PhD, FRACP, Steven A. Krilis, MBBS, PhD, FRACP: St. George Hospital and University of New South Wales, Sydney, New South Wales, Australia; ⁴De-Min Yu, MD, PhD: Metabolic Disease Hospital and Tianjin Medical University, Tianjin, China; ⁵Panayiotis G. Vlachoyiannopoulos, MD, PhD, Haralampos M. Moutsopoulos, MD, FACP, FRCP, PhD: National University of Athens Medical School, Athens, Greece; ⁶Anisur Rahman, PhD, FRCP, Charis Pericleous, PhD: University College London, London, UK; ⁷Tatsuya Atsumi, MD, PhD, Takao Koike, MD, PhD: Hokkaido University School of Medicine, Sapporo, Japan; ⁸Stephane Heritier, PhD: The George Institute for Global Health and Sydney University, Sydney, New South Wales, Australia.

Drs. Giannakopoulos and Krilis contributed equally to this work.
Dr. Atsumi has received consulting fees, speaking fees, and/or honoraria from Mitsubishi Tanabe, Takeda, Pfizer, Bristol-Myers Squibb, Chugai, Eisai, Abbott Japan, and MBL (less than \$10,000 each). Dr. Koike has received consulting fees, speaking fees, and/or honoraria from Abbott Immunology, Bristol-Myers Squibb, Chugai, Mitsubishi Tanabe, Takeda, Pfizer, and Eisai (less than \$10,000 each).

reduced, and posttranslationally modified oxidized $\beta_2 \text{GPI}$ in APS patients compared to various control groups.

Methods. In a retrospective multicenter analysis, the proportion of β_2 GPI with free thiols in serum from healthy volunteers was quantified. Assays for measurement of reduced as well as total circulating β_2 GPI were developed and tested in the following groups: APS (with thrombosis) (n = 139), autoimmune disease with or without persistent antiphospholipid antibodies (aPL) but without APS (n = 188), vascular thrombosis without APS or aPL (n = 38), and healthy volunteers (n = 91).

Results. Total β_2 GPI was significantly elevated in patients with APS (median 216.2 μ g/ml [interquartile range 173.3–263.8]) as compared to healthy subjects (median 178.4 μ g/ml [interquartile range 149.4–227.5] [P < 0.0002]) or control patients with autoimmune disease or vascular thrombosis (both P < 0.0001). The proportion of total β_2 GPI in an oxidized form (i.e., lacking free thiols) was significantly greater in the APS group than in each of the 3 control groups (all P < 0.0001).

Conclusion. This large retrospective multicenter

Address correspondence to Bill Giannakopoulos, MBBS, PhD, FRACP, Departments of Immunology, Rheumatology, and Medicine, St. George Hospital, University of New South Wales, Gray Street, Kogarah 2217. Sydney, New South Wales, Australia (e-mail: bill.giannakopoulos@unsw.edu.au); or to Steven A. Krilis, MBBS, PhD, FRACP, Department of Immunology, Allergy and Infectious Diseases, St. George Hospital, University of New South Wales, 2 South Street, Kogarah 2217, Sydney, New South Wales, Australia (e-mail: s.krilis@unsw.edu.au).

Submitted for publication December 16, 2010; accepted in revised form March 29, 2011.

study shows that posttranslational modification of β_2 GPI via thiol-exchange reactions is a highly specific phenomenon in the setting of APS thrombosis. Quantification of posttranslational modifications of β_2 GPI in conjunction with standard laboratory tests for APS may offer the potential to more accurately predict the risk of occurrence of a thrombotic event in the setting of APS.

The antiphospholipid syndrome (APS) is an autoimmune condition characterized by vascular thrombosis of the arterial and/or venous systems as well as recurrent miscarriages (1). Beta-2-glycoprotein I (β_2 GPI) is the major autoantigen in APS (2). A number of studies have provided robust evidence that autoantibodies to β_2 GPI are a significant risk factor for arterial thrombosis in young adults (3,4). In vivo and ex vivo studies by multiple groups have shown anti- β_2 GPI autoantibodies to be directly thrombogenic (5).

At present it is not possible to stratify the risk for development of thrombosis in antiphospholipid antibody (aPL)–positive patients based on clinical features or use of currently available laboratory assays (6). The development of novel assays that could be used to stratify future thrombosis risk in patients with APS would hold immense clinical utility in informing the decision as to whether initiation of prophylactic therapy or intensification of therapy is warranted.

Beta-2-glycoprotein I is an evolutionarily conserved 50-kd protein circulating in the blood in relative abundance ($\sim 4 \mu M$) (7). The physiologic role of β_2 GPI is pleiotropic, with functional studies implicating a role in processes relating to coagulation (8), angiogenesis (9), and clearance of apoptotic cells (10). The crystal structure of β_2 GPI, which has been ascertained based on the purified native protein, reveals that it does not possess free thiols (11,12). We have recently shown, however, that in vivo β_2 GPI circulates in a free thiol form and that this free thiol form of β_2 GPI is involved in the protection of endothelial cells against oxidative stressinduced cell injury (13). Beta-2-glycoprotein I can also participate in redox thiol-exchange reactions by acting as a substrate for oxidoreductase enzymes such as thioredoxin 1 (14). However, the proportion of β_2 GPI circulating in the reduced state is unknown. Also unknown is whether the redox state of this autoantigen differs in patients with pathogenic anti- β_2 GPI antibodies and a history of thrombosis.

In the present study we demonstrated that, in serum/plasma derived from healthy subjects, β_2 GPI exists in a reduced biochemical state as the dominant molecular phenotype. Detailed in vitro quantitative as-

says to assess the levels of total and reduced β_2 GPI were developed and used to screen >450 samples. Levels of both total and oxidized β_2 GPI were found to be elevated in patients with APS as compared to disease and healthy control groups. These findings have implications with respect to understanding the antigenic drive for pathogenic aPL, as well as the potential for development of assays for purposes of thrombosis risk stratification.

PATIENTS AND METHODS

Patient samples. Samples were collected through an international collaborative multicenter effort involving 5 centers (University of New South Wales [Sydney, Australia], University of Athens [Athens, Greece], University College London [London, UK], Tianjin Medical University [Tianjin, China], and Hokkaido University School of Medicine [Sapporo, Japan]). An APS group, 2 disease control groups, and 1 healthy control group were studied. The disease control groups consisted of an autoimmune disease group (with or without aPL, but with no clinical features of APS) and a clinical event control group (clinical features of APS, but no aPL or autoimmune disease).

APS group. A total of 139 samples from patients with APS were collected and analyzed (24 from Sydney, 38 from Athens, 22 from London, and 55 from Sapporo). Every APS patient fulfilled the revised consensus classification criteria for vascular thrombosis-associated APS (1). All serologic tests for aPL were performed using standard commercially available kits and in accordance with the revised classification criteria. A venous thrombotic event was diagnosed based on a combination of clinical assessment and appropriate imaging with either Doppler ultrasonography or venography to confirm deep venous thrombosis, or isotope ventilation/perfusion scanning or computed tomography (CT) (with or without angiography) to confirm pulmonary embolism. An arterial event was diagnosed based on clinical findings along with one or more of the following: electrocardiographic evidence of myocardial ischemia or infarction, confirmation of infarction by brain CT or magnetic resonance imaging, or confirmation of peripheral vascular disease or arterial thrombosis by Doppler ultrasonography or angiography.

Autoimmune disease control group. Of the 189 autoimmune disease controls, samples from 188 were analyzed (42 from Sydney, 43 from Athens, 29 from London, and 74 from Sapporo). One sample (from a patient with systemic lupus erythematosus [SLE] and no aPL) was found to be deficient in β_2 GPI and was withdrawn from the study. Among the autoimmune disease controls, 74 had persistently positive serologic findings for aPL satisfying the serologic component of the APS classification criteria (1), but did not have APS given the lack of a clinical event. All patients with SLE fulfilled the American College of Rheumatology revised classification criteria (15), and those with Sjögren's syndrome fulfilled the revised European classification criteria (16).

Clinical event control group. Thirty-eight samples from aPL-negative patients with a clinical event were collected and analyzed (26 from Sydney and 12 from Tianjin). Clinical events were diagnosed as described above for the APS group.