

Table 1. Demographics and Clinical Data

Carrier	Mutation	Age, y	Family Age at Onset, Mean	Years From Onset	Sex	Mini-Mental State Examination Score	Clinical Dementia Rating	Neocortex SUVR ^a
<i>PSEN1</i> _a	L219P	36 ^b	54	-18	F	30	0.0	0.54
<i>PSEN1</i> _b	dE9	38 ^b	44	-6	F	29	0.0	0.80
<i>APP</i>	V717I	48 ^b	54	-6	M	28	0.5 ^b	1.24 ^b
<i>PSEN1</i> _c	L219P	62	65	-3	M	29	0.5 ^b	0.96 ^b
<i>PSEN1</i> _d	Y115G	52 ^b	49	3	F	22 ^b	1.0 ^b	1.39 ^b
<i>PSEN1</i> _e	C236T	55	58	-3	M	...	3.0 ^b	0.76
<i>PSEN1</i> _f	L85P	31 ^b	26	5	M	...	3.0 ^b	0.72
<i>PSEN1</i> _g	L173S	45 ^b	37	8	F	...	3.0 ^b	0.93 ^b

Cohort	Mean (SD)	Female-Male Ratio	Mean (SD)	Mean (SD)	Mean (SD)
HC (n=30)	69.8 (6.6)	18/12	29.3 (0.9)	0.0 (0.0)	0.60 (0.08)
Sporadic AD (n=30)	73.6 (9.4)	14/16	22.1 (5.0) ^b	1.2 (0.7) ^b	1.21 (0.18) ^b

Abbreviations: AD, Alzheimer disease; Ellipsis, not applicable; HCs, healthy control subjects; SUVR, standardized uptake value ratio.

^aThe mean of the SUVR_{pons} for frontal, cingulate, parietal, lateral temporal, and occipital cortex.

^bSignificantly different from HCs (z score, >2).

risk factors for AD.^{6,7} The main feature of *APP*, *PSEN1*, and *PSEN2* mutations (involved in different steps of the APP processing pathway) is increased production and deposition of Aβ, especially Aβ₄₂.^{8,9} Despite some clinical heterogeneity associated with *PSEN1* mutations,^{10,11} these various genetic mutations lead to increased levels of Aβ in the brain before symptoms arise.¹¹

Amyloid β-peptide imaging with positron emission tomography (PET) allows early and accurate diagnosis of AD.^{12,13} Pittsburgh Compound B (PiB), the most widely used amyloid tracer, provides quantitative information on Aβ burden in vivo, which has led to new insights on Aβ deposition in the brain. The use of this technique has shown a robust difference in PiB retention between healthy control subjects (HCs) and subjects with AD^{12,13} and has demonstrated inverse correlations of Aβ burden with glucose hypometabolism in some brain regions,¹⁴ cerebrospinal fluid Aβ₄₂,¹⁵ and rate of cerebral atrophy.¹⁶ About 25% to 35% of asymptomatic age-matched HCs present with cortical PiB retention that correlates with subtle memory impairment and greater risk of cognitive decline, likely representing preclinical AD.¹⁷⁻¹⁹ Two previous studies^{20,21} reported high striatal PiB retention in *PSEN1* mutation carriers, while a third study²² reported a novel *APP* mutation in which Aβ remains in an oligomeric form showing mild cortical PiB retention. From these few studies, it is difficult to infer the significance of PiB retention in asymptomatic mutation carriers. Therefore, it is crucial to examine more patients with familial AD (FAD) having early-onset or variable mutations to better define the role of Aβ in this population and its association with cognitive status.

The objectives of the study were as follows: (1) to evaluate the pattern of PiB retention in subjects with distinct but different autosomal dominant mutations associated with FAD vs that in age-matched HCs and subjects with probable sporadic AD (SAD), and (2) to correlate Aβ burden as measured by PiB with available clinical and cog-

nitive data, and (3) to compare the regional brain patterns of PiB retention and fluorodeoxyglucose F 18 (FDG) uptake.

METHODS

PARTICIPANTS

Written informed consent for participation in this study was obtained from all subjects or caregivers before imaging. The study was approved by the Austin Health (Melbourne, Australia) human research ethics committee and by the Osaka City University Medical School (Osaka, Japan) institutional ethics committee.

Eight subjects who were carriers of *APP* or *PSEN1* mutations were studied using PiB and FDG PET imaging. All subjects were aware that they were carrying a mutation linked to AD. Specific mutations are listed in **Table 1**.

PiB and FDG PET studies of mutation carriers were compared with those of a well-characterized cohort of 30 HCs and 30 subjects with probable SAD. The latter subjects met the criteria for probable AD as outlined by the National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer Disease and Related Disorders Association.²³

Subjects underwent a neurologic examination. In addition to the Clinical Dementia Rating (CDR) and the Mini-Mental State Examination (MMSE), subjects without complete impairment underwent various neuropsychological tasks designed to assess a broad range of cognitive domains, although no specific test to assess striatal function was administered.

IMAGING PROCEDURES

All subjects underwent T1-weighted magnetic resonance (MR) imaging for screening and subsequent coregistration with PET images. Each subject received approximately 370 megabecquerels (MBq) of PiB by intravenous injection over 1 minute. Imaging was performed in Melbourne for the *PSEN1*_{a-c} (as listed in Table 1) and *APP* mutation carriers (Allegro PET camera; Phillips, Amsterdam, the Netherlands) and in Osaka for the *PSEN1*_{d-g} mutation carriers (Eminence-B PET imaging system;

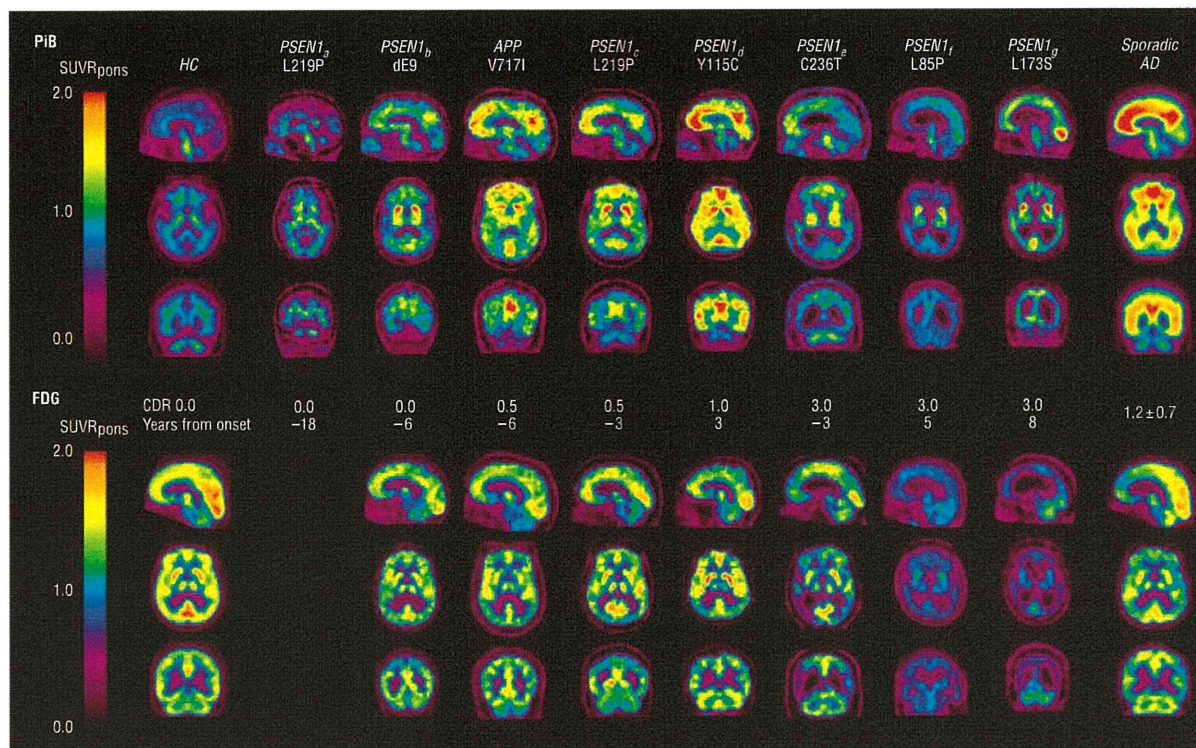


Figure 1. Positron emission tomography (PET) images. Representative parametric sagittal, transaxial, and coronal Pittsburgh Compound B (PiB) PET (top) and fluorodeoxyglucose F 18 (FDG) PET (bottom) images of 8 mutation carriers, as well as representative images of the healthy control (HC) and probable sporadic Alzheimer disease (AD) cohorts. Subjects are arranged according to their Clinical Dementia Rating (CDR) and years from onset for their respective pedigrees. All mutation carriers show high PiB retention in the striata, as well as in the ventrofrontal and posterior cingulate or precuneus areas in most of them. There was no clear pattern of FDG hypometabolism across the subjects studied. SUVR indicates standardized uptake value ratio.

Shimadzu, Osaka, Japan). A 20- to 30-minute emission acquisition was then performed in 3-dimensional mode starting 40 minutes after injection of PiB. In 7 of 8 subjects, static FDG images were obtained 45 minutes after injection of approximately 250 MBq of FDG.

IMAGE ANALYSIS

Coregistration of PET images was performed using a computer program (SPM5; [Statistical Parametric Mapping 5]; Medical Research Council Cognition and Brain Sciences Unit, Cambridge, England),²⁴ and an MR imaging-defined region-of-interest template was then applied to PET images. The mean standardized uptake value (SUV) was obtained from the region of interest for cortical, subcortical, and cerebellar regions.

Because of the reported presence of plaques in the cerebellar cortex of patients with FAD,²⁵ PiB retention and FDG uptake were normalized to pons radioactivity to generate standardized uptake value ratio pons (SUVR_{pons}). Regional PiB and FDG PET imaging SUVR_{pons} of mutation carriers were then compared with those of a well-characterized (although significantly older) cohort of 30 HCs without evidence of A β deposition in the brain (PiB negative) and 30 subjects with probable SAD.

STATISTICAL ANALYSIS

z Scores exceeding 2 were considered significantly different from HCs or subjects with probable SAD. Correlations were assessed using Pearson product moment correlation analyses. Group data are expressed as the mean (SD).

RESULTS

Demographic data for mutation carriers and for the HC and probable SAD cohorts are given in Table 1. All symptomatic mutation carriers had onset of cognitive decline within the expected range of their respective pedigrees. At the time of PET imaging, 3 subjects had severe impairment, and it was impossible to obtain MMSE scores.

The pattern of PiB retention in all mutation carriers, independent of mutation type, differed from that in subjects with probable SAD, with higher PiB retention in the striatum among the mutation carriers (**Figure 1**). Older mutation carriers had higher striatal PiB retention. **Table 2** gives the regional SUVR_{pons} of mutation carriers vs HCs and subjects with probable SAD. Another common feature was high PiB retention in the prefrontal, orbitofrontal, and gyrus rectus regions in most (6 of 8) mutation carriers (Figure 1). Despite their sharing the same *PSEN1* mutation (L219P) (Table 1), the degree of PiB retention was higher and more widespread in the 62-year-old subject closer to the age at onset for his pedigree, while PiB retention was restricted to the caudate nuclei in the 36-year-old subject (18 years away from the age at onset for her pedigree). A cortical PiB retention pattern similar to that usually seen in SAD was observed in 2 women with dementia (the 45-year-old *PSEN1*_g mutation carrier and the 52-year-old *PSEN1*_d mutation carrier) and in 2 men without dementia (the 48-year-old *APP*

Table 2. Individual Cerebral Regional Pittsburgh Compound B Retention SUVR_{pons} in Familial AD, 30 HCs and 30 Subjects With Probable Sporadic AD

Location	<i>PSEN1</i> _a L219P	<i>PSEN1</i> _b dE9	<i>APP</i> V717L	<i>PSEN1</i> _c L219P	<i>PSEN1</i> _d Y115C	<i>PSEN1</i> _e C236T	<i>PSEN1</i> _f L85P	<i>PSEN1</i> _g L173S	Mean (SD)	
									HCs	Sporadic AD
Frontal	0.53	0.91 ^a	1.32 ^a	1.04 ^a	1.50 ^a	0.84 ^a	0.63	0.87 ^a	0.59 (0.10)	1.30 (0.23)
Orbitofrontal	0.57	0.82 ^a	1.34 ^a	1.01 ^a	1.40 ^a	0.88 ^a	0.68	0.89 ^a	0.61 (0.09)	1.26 (0.22)
Gyrus rectus	0.70	0.92 ^a	1.31 ^a	1.17 ^a	1.65 ^a	0.93 ^a	0.57	1.01 ^a	0.62 (0.09)	1.34 (0.27)
Anterior cingulate	0.62	0.91 ^a	1.31 ^a	1.17 ^a	1.53 ^a	0.77	0.76	1.02 ^a	0.63 (0.11)	1.30 (0.21)
Posterior cingulate	0.58	0.81	1.65 ^a	1.17 ^a	1.65 ^a	0.73	0.82 ^a	0.95 ^a	0.62 (0.09)	1.33 (0.19)
Parietal	0.54	0.81 ^a	1.00 ^a	0.81 ^a	1.34 ^a	0.68	0.73	1.00 ^a	0.55 (0.09)	1.14 (0.20)
Occipital	0.47	0.72	0.91 ^a	0.68	1.14 ^a	0.74	0.74	0.96 ^a	0.64 (0.07)	0.96 (0.17)
Lateral temporal	0.51	0.68	1.24 ^a	0.94 ^a	1.34 ^a	0.66	0.66	0.95 ^a	0.59 (0.08)	1.17 (0.20)
Mesial temporal	0.56	0.60	0.80 ^a	0.69	0.83 ^a	0.50	0.55	0.42 ^a	0.61 (0.07)	0.82 (0.11)
Caudate nuclei	1.04 ^a	1.37 ^a	1.76 ^a	1.46 ^a	1.97 ^a	0.70 ^b	1.19 ^a	1.59 ^a	0.64 (0.10)	1.32 (0.27)
Putamen	0.76	1.15 ^a	1.25 ^a	1.34 ^a	1.57 ^a	0.96 ^a	1.15 ^a	1.38 ^a	0.63 (0.08)	1.16 (0.22)
Thalamus	0.63	0.82	1.19 ^a	1.06 ^a	1.28 ^a	0.82	0.72	1.04 ^a	0.72 (0.07)	1.03 (0.16)
Midbrain	0.88	0.95	0.96	0.88	0.98	0.90	0.80	0.92	0.87 (0.10)	0.96 (0.09)
Cerebellum	0.47	0.47	0.63 ^a	0.65 ^a	0.59	0.62 ^a	0.65 ^a	0.51	0.48 (0.06)	0.52 (0.09)
Striatum	0.90 ^a	1.26 ^a	1.50 ^a	1.40 ^a	1.77 ^a	0.83 ^a	1.17 ^a	1.49 ^a	0.63 (0.09)	1.24 (0.26)
Neocortex ^c	0.54	0.80 ^a	1.24 ^a	0.96 ^a	1.39 ^a	0.76	0.72	0.93 ^a	0.60 (0.08)	1.21 (0.18)

Abbreviations: AD, Alzheimer disease; HCs, healthy control subjects; SUVR, standardized uptake value ratio.

^aSignificantly different from HCs (z score, >2).

^bSevere caudate nuclei atrophy.

^cThe mean of the SUVR_{pons} for frontal, cingulate, parietal, lateral temporal, and occipital cortex.

mutation carrier and the 62-year-old *PSEN1*_c mutation carrier). Half of the mutation carriers demonstrated significantly higher cerebellar PiB retention than that in HCs (Table 2).

Although global and regional FDG uptake was lower in most symptomatic mutation carriers (Table 3), there was no common pattern of FDG uptake among the subjects studied (Figure 1). Three subjects (the *PSEN1*_e, *PSEN1*_f, and *PSEN1*_g mutation carriers) showed marked global glucose hypometabolism in which reduced FDG uptake was associated with severe brain atrophy, while the *PSEN1*_c mutation carrier demonstrated asymmetric FDG uptake but less atrophy on MR imaging than that in the other 2 subjects. Three other subjects (the *PSEN1*_b, *APP*, and *PSEN1*_i mutation carriers) showed an almost normal pattern of FDG uptake. The *PSEN1*_d mutation carrier had lower FDG uptake than that among HCs in the parietal cortex, as is usually observed in SAD, and had high FDG uptake in the anterior cingulate, lateral temporal, and striatum (Table 3).

Striatal and cortical PiB retention was not associated with mutation type, disease severity, or cognitive impairment (Figure 2). In contrast to PiB findings, the FDG posterior cortical index correlated with MMSE score ($r=0.85$, $P=.02$), CDR ($r=-0.84$, $P=.02$), and years from onset for the respective families ($r=-0.78$, $P=.04$). There was no regional or global correlation between PiB retention and FDG uptake. Given the dichotomy of the CDR and the MMSE score, mutation carriers were separated into 2 subgroups according to their disease severity (MMSE score >20 or ≤20 and CDR >2 or ≤2) for further comparison. There was no significant difference between the subgroups in striatal or neocortical PiB retention, while the most cognitively impaired subgroup (MMSE score ≤20 and CDR >2) had significantly lower

striatal FDG uptake ($P=.03$) and FDG posterior cortical index ($P=.01$).

COMMENT

In vivo amyloid PET imaging has allowed new insights on Aβ deposition in the brain, facilitating research into the causes, diagnosis, and future treatment of dementias in which Aβ may have a role.^{12,13,26} We examined the pattern and degree of PiB retention in familial cases with *PSEN1* and *APP* mutations. All mutation carriers showed some degree of increased PiB retention. Although the degree of cortical retention was generally lower than that usually observed in SAD, the striatal retention was remarkably high. Early onset and rapid progression of the disease indicate that other factors besides Aβ deposition have a role in the cognitive impairment process in FAD, in which Aβ upregulation and deposition represent an early and necessary but not sufficient immediate cause of cognitive decline.

The pattern of PiB retention was similar to that reported in 2 previous studies^{20,21} of *PSEN1* mutation carriers. Postmortem studies^{21,27} of patients with FAD had shown Aβ deposits in the striatum. The high PiB retention in the striatum is difficult to reconcile with the clinical phenotype given the similar symptoms in SAD, in which this pattern of retention is not observed. However, this pattern is constant across different mutation types, and it has been proposed that Aβ deposition in FAD starts in the striata.²⁰ High striatal PiB retention was accompanied by significant retention in the frontal regions, although the retention was not as high as that observed in SAD. As in a high percentage of HCs, PiB retention is observed in the frontal and posterior cingu-

Table 3. Individual Cerebral Regional Fluorodeoxyglucose F 18 Uptake SUVR_{pons}, in Familial AD, 30 HCs, and 30 Subjects With Probable Sporadic AD

Location	<i>PSEN1</i> _{L219P}	<i>PSEN1</i> _{dE9}	APP V717L	<i>PSEN1</i> _{L219P}	<i>PSEN1</i> _{Y115C}	<i>PSEN1</i> _{C236T}	<i>PSEN1</i> _{L85P}	<i>PSEN1</i> _{L173S}	Mean (SD)	
									HCs	Sporadic AD
Frontal	NA	1.32	1.40	1.30	1.43	1.23	0.80 ^a	0.89 ^a	1.43 (0.23)	1.20 (0.16)
Orbitofrontal	NA	1.32	1.45	1.35	1.36	1.26	0.81 ^a	0.88 ^a	1.42 (0.18)	1.21 (0.13)
Gyrus rectus	NA	1.37	1.42	1.48	1.58	1.29	0.82 ^a	0.95 ^a	1.39 (0.15)	1.24 (0.12)
Anterior cingulate	NA	1.44	1.38	1.37	1.55	1.40	0.91 ^a	1.03 ^a	1.37 (0.17)	1.17 (0.14)
Posterior cingulate	NA	1.46	1.55	1.45	1.48	1.36	0.82 ^a	1.01 ^a	1.62 (0.19)	1.24 (0.18)
Parietal	NA	1.25	1.32	1.01	1.05	1.02	0.79 ^a	0.87	1.37 (0.26)	1.06 (0.24)
Occipital	NA	1.45	1.40	1.15	1.37	1.37	0.80 ^a	1.00	1.45 (0.22)	1.22 (0.21)
Lateral temporal	NA	1.32	1.41	1.26	1.47	1.10 ^a	0.81 ^a	0.86 ^a	1.39 (0.14)	1.07 (0.16)
Mesial temporal	NA	1.02	0.99	0.96	1.17	0.82	0.79 ^a	0.54 ^a	1.05 (0.13)	0.95 (0.12)
Caudate nuclei	NA	1.61	1.39	1.51	1.86	1.27 ^b	1.06 ^a	1.04 ^a	1.61 (0.20)	1.47 (0.20)
Putamen	NA	1.63	1.60	1.62	1.93	1.56	1.08 ^a	1.13 ^a	1.68 (0.17)	1.57 (0.15)
Thalamus	NA	1.30 ^a	1.47	1.72	1.73	1.66	0.88 ^a	1.15 ^a	1.70 (0.13)	1.49 (0.14)
Midbrain	NA	0.90 ^a	0.92 ^a	1.25	1.22	1.26	0.93 ^a	0.96 ^a	1.25 (0.12)	1.16 (0.14)
Cerebellum	NA	1.33	1.20	1.22	1.47	1.37	1.01 ^a	1.17	1.33 (0.15)	1.26 (0.15)
Striatal	NA	1.62	1.50	1.57	1.90 ^a	1.47	1.07 ^a	1.09 ^a	1.64 (0.12)	1.51 (0.17)
Posterior cortex ^c	NA	1.34	1.42	1.24	1.33	1.16	0.80 ^a	0.92 ^a	1.46 (0.17)	1.15 (0.16)

Abbreviations: AD, Alzheimer disease; HCs, healthy control subjects; NA, not applicable; SUVR, standardized uptake value ratio.

^aSignificantly different from HCs (z score, >2).

^bSevere caudate nuclei atrophy.

^cThe mean of the SUVR_{pons} for posterior cingulate, parietal, and lateral temporal cortex.

late regions of a significant proportion of cognitively impaired or minimally impaired mutation carriers. These findings in mutation carriers are in agreement with postmortem findings showing that a high percentage of nondemented older individuals have amyloid plaques (with deposits occurring well before the onset of dementia)^{28,29} and with evidence indicating that neuropathologic changes precede the clinical phenotype by many years.^{30,31}

At least 3 of 8 mutation carriers in our study demonstrated marked atrophy on MR imaging. Most extrastriatal PiB retention was confined to the ventrofrontal regions, with 4 mutation carriers showing a pattern of cortical PiB retention similar to the pattern observed in SAD.

Semiquantitative measures of Aβ burden are usually generated by normalizing the regional SUV to the cerebellar cortex, a region unaffected by senile plaque deposition in SAD.³² In contrast, mutation carriers in our study showed a pattern of Aβ deposition different from that of subjects with probable SAD, with higher PiB retention in the cerebellum of mutation carriers reflecting cerebellar Aβ deposition (Table 2).

Postmortem measurements of the distribution and density of diffuse and neuritic Aβ plaques have not consistently correlated with the degree of cognitive impairment in AD.^{33,34} The best correlation has been observed with neurofibrillary tangles and soluble levels of Aβ.^{35,36} While the exact mechanism by which Aβ might produce synaptic loss and neuronal death is controversial,^{1,37} it is likely that PiB retention in nondemented individuals^{13,17-19,38} and in cognitively unimpaired or minimally impaired mutation carriers reflects preclinical AD in a classic neuropathologic view.³¹ This “delay” in the manifestation of the phenotype may be attributed to different idiosyncratic or cellular susceptibility or vul-

nerability to Aβ, variations in Aβ conformation affecting toxicity or PiB binding, or both.³⁹⁻⁴² There is also the issue of mutation type, whereby some *PSEN1* mutations are more aggressive and evolve faster than others, while others are associated with movement disorders such as spastic paraparesis or extrapyramidal signs. Larner and Doran¹¹ have reviewed the phenotypic manifestations of some of the *PSEN1* mutations discussed herein. These hypotheses would justify early involvement of the striatum and would help explain why some older individuals with significant Aβ burden are cognitively unimpaired, while others with genetic predisposing factors (despite lower Aβ burden) have already developed the full clinical AD phenotype.

Regarding PiB binding to different Aβ species with or without posttranslational modification or in a fibrillary oligomer or monomer form, it has been reported that PiB binds with higher affinity to one kind of N-terminal-truncated Aβ₁₋₄₂₍₄₃₎ species in senile plaques, specifically that truncated at position 3 (Aβ₃[pE]), displaying a 5-fold higher affinity for Aβ₃(pE)₁₋₄₂₍₄₃₎ than for Aβ₁₋₄₂₍₄₃₎.⁴³ This is relevant to PiB binding because, besides the usual senile and diffuse plaques observed in SAD, cotton wool plaques are observed in the brains of *PSEN1* mutation carriers.^{27,44} Cotton wool plaques are generally large with a clear rim and are composed mainly of neuropil elements and Aβ₂ species, ubiquitously located in the cortex and basal ganglia.^{27,44} Cotton wool plaques are particularly important not only because of their distribution in the striatum and cortex but also because they are mildly stained with thioflavin S,⁴⁵ in contrast to conventional Aβ plaques seen in SAD or normal aging. Cotton wool plaques should be studied more extensively to explain their etiologic mechanism and how they might contribute to the different patterns of PiB retention among FAD,

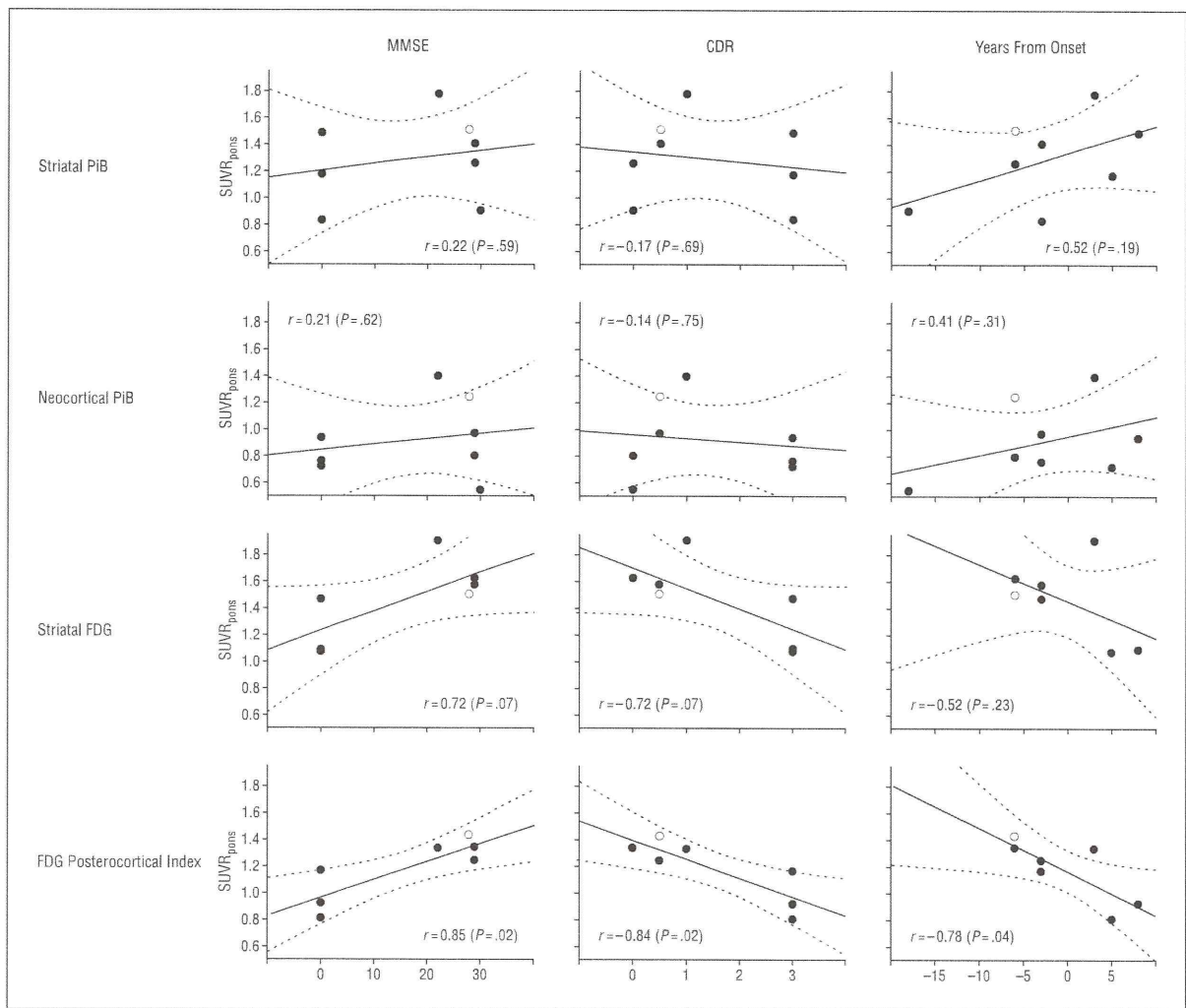


Figure 2. Correlational analysis. Pearson product moment correlation linear correlation analysis shows lack of association between striatal or neocortical Pittsburgh Compound B (PiB) retention and disease severity data or years from onset for the respective pedigrees. Conversely, there are strong correlations between the same variables and fluorodeoxyglucose F 18 (FDG) postero-cortical uptake. CDR indicates Clinical Dementia Rating; MMSE, Mini-Mental State Examination; and SUVR, standardized uptake value ratio.

SAD, and normal aging. Longitudinal studies combined with postmortem assessment of A β are needed to elucidate this point.

Although PiB investigations (in accord with previous studies^{20,21}) showed higher A β burden in the striata of all of the mutation carriers studied herein, there was no clear pattern of FDG hypometabolism, neither the typical temporoparietal hypometabolism observed in SAD⁴⁶ nor the more restricted temporoparietal hypometabolism reported in asymptomatic at-risk subjects with known APP or chromosome 14-linked mutations^{47,48} and in subjects with a strong family history of AD.⁴⁹ Despite their having generally lower FDG uptake than HCs, most mutation carriers did not show significant regional differences vs HCs or subjects with probable SAD. The reason may be the variance of FDG uptake among HCs and subjects with probable SAD, a variance that precluded achieving higher z scores among the mutation carriers. The 2 subjects with extremely low FDG uptake (the PSEN1_I and PSEN1_G mutation carriers) also have the most

severe brain atrophy and the most severe cognitive impairment. The third subject with atrophy and marked cognitive impairment (the PSEN1_I mutation carrier) showed an asymmetric pattern of FDG uptake. In FAD as in SAD, PiB seems to be a more sensitive and accurate biomarker than FDG for early detection of disease.⁵⁰ This might be because A β deposition starts approximately 10 years before any cognitive or memory decline is noted²⁹ and much earlier than the synaptic and neuronal loss that is reflected in regional glucose hypometabolism. Conversely, FDG uptake correlates with MMSE score and (as in SAD) might prove to be a better marker of disease progression than PiB.⁵¹ Evaluation of more familial mutation cases with PiB and longitudinal follow-up are warranted to establish the usefulness of PiB as an early and reliable method of detecting A β deposition, while also providing insights on the progression of A β deposition and assessing its prognostic accuracy. An international consortium, the Dominantly Inherited Alzheimer Network (<http://www.dian-info.org/>), has been organized to

comprehensively assess the origin of AD through FAD and to evaluate the usefulness of different biomarkers such as amyloid imaging. This kind of multidisciplinary approach should define the role of amyloid imaging in the evaluation of asymptomatic mutation carriers at risk of developing FAD.

In conclusion, although A β deposition (as in SAD) seems to precede the clinical manifestation of dementia, the pattern of A β deposition in FAD is not related to disease severity and (irrespective of mutation type) differs from that observed in SAD. When disease-specific therapies aimed at preventing or slowing AD progression become available, amyloid imaging studies will have an important role in the identification of A β deposits in at-risk mutation carriers before the development of symptoms.

Accepted for Publication: April 21, 2009.

Author Affiliations: Department of Nuclear Medicine, Centre for Positron Emission Tomography, Austin Health, Heidelberg (Drs Villemagne, Mulligan, O'Keefe, and Rowe and Mr Jones), The Mental Health Research Institute, The University of Melbourne, Melbourne (Drs Villemagne and Masters), and Prince of Wales Medical Research Institute, Randwick, and The University of New South Wales, Sydney (Dr Brooks), Australia; and Neuroscience, Gerontology, and Neurology, Osaka City University Medical School, Osaka (Drs Ataka, Miki, Shimada, and Mori), Molecular Neurology, Graduate School of Medical Science, Kyoto Prefectural University of Medicine, Kyoto (Drs Mizuno, Kondo, and Nakagawa), and Molecular Imaging Research Program, RIKEN, Kobe (Dr Wada and Watanabe), Japan.

Correspondence: Victor L. Villemagne, MD, Department of Nuclear Medicine, Centre for Positron Emission Tomography, Austin Health, 145 Studley Rd, Heidelberg, Victoria, Australia 3084 (villemagne@petnm.unimelb.edu.au).

Author Contributions: Drs Villemagne, Mori, and Rowe had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. *Study concept and design:* Villemagne, Rowe, and Mori. *Acquisition of data:* Villemagne, Ataka, Mizuno, Brooks, Wada, Kondo, Jones, Watanabe, Mulligan, Nakagawa, Miki, Shimada, O'Keefe, Mori, and Rowe. *Analysis and interpretation of data:* Villemagne, Ataka, Masters, Mori, and Rowe. *Drafting of the manuscript:* Villemagne, Mori, and Rowe. *Critical revision of the manuscript for important intellectual content:* Villemagne, Masters, Mori, and Rowe. *Statistical analysis:* Villemagne. *Study supervision:* Villemagne, Mori, and Rowe.

Financial Disclosure: None reported.

Funding Support: This study was supported in part by the Austin Hospital Medical Research Foundation, Neurosciences Victoria, The University of Melbourne.

REFERENCES

- Masters CL, Cappai R, Barnham KJ, Villemagne VL. Molecular mechanisms for Alzheimer's disease: implications for neuroimaging and therapeutics. *J Neurochem*. 2006;97(6):1700-1725.
- Villemagne VL, Cappai R, Barnham KJ, et al. The A β centric pathway of Alzheimer's disease. In: Barrow CJ, Small BJ, eds. *A β Peptide and Alzheimer's Disease*. London, England: Springer-Verlag; 2006:5-32.
- Cappai R, White AR. Amyloid β . *Int J Biochem Cell Biol*. 1999;31(9):885-889.
- St George-Hyslop PH. Molecular genetics of Alzheimer's disease. *Biol Psychiatry*. 2000;47(3):183-199.
- Bird TD. Genetic aspects of Alzheimer disease. *Genet Med*. 2008;10(4):231-239.
- Tanzi RE, Bertram L. New frontiers in Alzheimer's disease genetics. *Neuron*. 2001;32(2):181-184.
- Sherrington R, Rogaev EI, Liang Y, et al. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature*. 1995;375(6534):754-760.
- Younkin SG. The AAP and PS1/2 mutations linked to early onset familial Alzheimer's disease increase the extracellular concentration and A β 1-42 (43). *Rinsho Shinkeigaku*. 1997;37(12):1099.
- Neve RL, McPhie DL, Chen Y. Alzheimer's disease: a dysfunction of the amyloid precursor protein(1). *Brain Res*. 2000;886(1-2):54-66.
- Menéndez M. Pathological and clinical heterogeneity of presenilin 1 gene mutations. *J Alzheimers Dis*. 2004;6(5):475-482.
- Larner AJ, Doran M. Clinical phenotypic heterogeneity of Alzheimer's disease associated with mutations of the presenilin-1 gene. *J Neurol*. 2006;253(2):139-158.
- Klunk WE, Engler H, Nordberg A, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. *Ann Neurol*. 2004;55(3):306-319.
- Rowe CC, Ng S, Ackermann U, et al. Imaging β -amyloid burden in aging and dementia. *Neurology*. 2007;68(20):1718-1725.
- Edison P, Archer HA, Hinz R, et al. Amyloid, hypometabolism, and cognition in Alzheimer disease: an [¹¹C]PIB and [¹⁸F]FDG PET study. *Neurology*. 2007;68(7):501-508.
- Fagan AM, Mintun MA, Mach RH, et al. Inverse relation between in vivo amyloid imaging load and cerebrospinal fluid A β_{42} in humans. *Ann Neurol*. 2006;59(3):512-519.
- Archer HA, Edison P, Brooks DJ, et al. Amyloid load and cerebral atrophy in Alzheimer's disease: an [¹¹C]-PIB positron emission tomography study. *Ann Neurol*. 2006;60(1):145-147.
- Mintun MA, Larossa GN, Sheline YI, et al. [¹¹C]PIB in a nondemented population: potential antecedent marker of Alzheimer disease. *Neurology*. 2006;67(3):446-452.
- Pike KE, Savage G, Villemagne VL, et al. β -Amyloid imaging and memory in nondemented individuals: evidence for preclinical Alzheimer's disease. *Brain*. 2007;130(pt 11):2837-2844.
- Villemagne VL, Pike KE, Darby D, et al. A β deposits in older non-demented individuals with cognitive decline are indicative of preclinical Alzheimer's disease. *Neuropsychologia*. 2008;46(6):1688-1697.
- Klunk WE, Price JC, Mathis CA, et al. Amyloid deposition begins in the striatum of presenilin-1 mutation carriers from two unrelated pedigrees. *J Neurosci*. 2007;27(23):6174-6184.
- Koivunen J, Verkkoniemi A, Aalto S, et al. PET amyloid ligand [¹¹C]PIB uptake shows predominantly striatal increase in variant Alzheimer's disease. *Brain*. 2008;131(pt 7):1845-1853.
- Tomiya T, Nagata T, Shimada H, et al. A new amyloid β variant favoring oligomerization in Alzheimer's-type dementia. *Ann Neurol*. 2008;63(3):377-387.
- McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology*. 1984;34(7):939-944.
- Friston KJ, Frith CD, Liddle PF, Frackowiak RS. Comparing functional (PET) images: the assessment of significant change. *J Cereb Blood Flow Metab*. 1991;11(4):690-699.
- Lippa CF, Saunders AM, Smith TW, et al. Familial and sporadic Alzheimer's disease: neuropathology cannot exclude a final common pathway. *Neurology*. 1996;96(2):406-412.
- Rowe CC, Ackerman U, Browne W, et al. Imaging of amyloid β in Alzheimer's disease with 18F-BAY94-9172, a novel PET tracer: proof of mechanism. *Lancet Neurol*. 2008;7(2):129-135.
- Tabira T, Chui DH, Nakayama H, Kuroda S, Shibuya M. Alzheimer's disease with spastic paresis and cotton wool type plaques. *J Neurosci Res*. 2002;70(3):367-372.
- Forman MS, Mufson EJ, Leurgans S, et al. Cortical biochemistry in MCI and Alzheimer disease: lack of correlation with clinical diagnosis. *Neurology*. 2007;68(10):757-763.
- Morris JC, Price AL. Pathologic correlates of nondemented aging, mild cognitive impairment, and early-stage Alzheimer's disease. *J Mol Neurosci*. 2001;17(2):101-118.
- Bennett DA, Schneider JA, Arvanitakis Z, et al. Neuropathology of older persons

- without cognitive impairment from two community-based studies. *Neurology*. 2006;66(12):1837-1844.
31. Price JL, Morris JC. Tangles and plaques in nondemented aging and "preclinical" Alzheimer's disease. *Ann Neurol*. 1999;45(3):358-368.
 32. Ikonomic MD, Klunk WE, Abrahamson EE, et al. Post-mortem correlates of in vivo PIB-PET amyloid imaging in a typical case of Alzheimer's disease. *Brain*. 2008;131(pt 6):1630-1645.
 33. Prohovnik I, Perl DP, Davis KL, Libow L, Lesser G, Haroutunian V. Dissociation of neuropathology from severity of dementia in late-onset Alzheimer disease. *Neurology*. 2006;66(1):49-55.
 34. Giannakopoulos P, Herrmann FR, Bussiere T, et al. Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease. *Neurology*. 2003;60(9):1495-1500.
 35. McLean CA, Cherny RA, Fraser FW, et al. Soluble pool of A β amyloid as a determinant of severity of neurodegeneration in Alzheimer's disease. *Ann Neurol*. 1999;46(6):860-866.
 36. Näslund J, Haroutunian V, Mohs R, et al. Correlation between elevated levels of amyloid β -peptide in the brain and cognitive decline. *JAMA*. 2000;283(12):1571-1577.
 37. Walsh DM, Klyubin I, Fadeeva JV, et al. Naturally secreted oligomers of amyloid β protein potently inhibit hippocampal long-term potentiation in vivo. *Nature*. 2002;416(6880):535-539.
 38. Aizenstein HJ, Nebes RD, Saxton JA, et al. Frequent amyloid deposition without significant cognitive impairment among the elderly. *Arch Neurol*. 2008;65(11):1509-1517.
 39. Deshpande A, Mina E, Glabe C, Busciglio J. Different conformations of amyloid β induce neurotoxicity by distinct mechanisms in human cortical neurons. *J Neurosci*. 2006;26(22):6011-6018.
 40. Lockhart A, Ye L, Judd DB, et al. Evidence for the presence of three distinct binding sites for the thioflavin T class of Alzheimer's disease PET imaging agents on β -amyloid peptide fibrils. *J Biol Chem*. 2005;280(9):7677-7684.
 41. Walker LC, Rosen RF, Levine H III. Diversity of A β deposits in the aged brain: a window on molecular heterogeneity? *Rom J Morphol Embryol*. 2008;49(1):5-11.
 42. Rosen RF, Walker LC, Levine H III. PIB binding in aged primate brain: enrichment of high-affinity sites in humans with Alzheimer's disease [published online ahead of print March 27, 2009]. *Neurobiol Aging*. 2009. doi:10.1016/j.neurobiolaging.2009.02.011.
 43. Maeda J, Ji B, Irie T, et al. Longitudinal, quantitative assessment of amyloid, neuroinflammation, and anti-amyloid treatment in a living mouse model of Alzheimer's disease enabled by positron emission tomography. *J Neurosci*. 2007;27(41):10957-10968.
 44. Karlstrom H, Brooks WS, Kwok JB, et al. Variable phenotype of Alzheimer's disease with spastic paraparesis. *J Neurochem*. 2008;104(3):573-583.
 45. Takao M, Ghetti B, Hayakawa I, et al. A novel mutation (G217D) in the Presenilin 1 gene (*PSEN1*) in a Japanese family: presenile dementia and parkinsonism are associated with cotton wool plaques in the cortex and striatum. *Acta Neuropathol*. 2002;104(2):155-170.
 46. Silverman DH, Small GW, Chang CY, et al. Positron emission tomography in evaluation of dementia: regional brain metabolism and long-term outcome. *JAMA*. 2001;286(17):2120-2127.
 47. Kennedy AM, Frackowiak RS, Newman SK, et al. Deficits in cerebral glucose metabolism demonstrated by positron emission tomography in individuals at risk of familial Alzheimer's disease. *Neurosci Lett*. 1995;186(1):17-20.
 48. Rossor MN, Kennedy AM, Frackowiak RS. Clinical and neuroimaging features of familial Alzheimer's disease. *Ann N Y Acad Sci*. 1996;777:49-56.
 49. Mosconi L, Sorbi S, de Leon MJ, et al. Hypometabolism exceeds atrophy in presymptomatic early-onset familial Alzheimer's disease. *J Nucl Med*. 2006;47(11):1778-1786.
 50. Ng S, Villemagne VL, Berlangieri S, et al. Visual assessment versus quantitative assessment of ¹¹C-PIB PET and ¹⁸F-FDG PET for detection of Alzheimer's disease. *J Nucl Med*. 2007;48(4):547-552.
 51. Engler H, Forsberg A, Almkvist O, et al. Two-year follow-up of amyloid deposition in patients with Alzheimer's disease. *Brain*. 2006;129(pt 11):2856-2866.

Announcement

Visit www.archneurol.com. You can send an e-mail to a friend that includes a link to an article and a note if you wish. Links will go to short versions of articles whenever possible.

認知症

アルツハイマー病の分子病態入門

—実地医家に必要な知識—

山川義宏・安宅鈴香・森 啓*

大阪市立大学医学部老年科神経内科・*同 脳神経科学/やまかわ・よしひろ あたか・すすか もり・ひろし

はじめに●

認知症の原因疾患として主にアルツハイマー病 Alzheimer disease (AD), 前頭側頭型認知症, 脳血管性認知症があげられ, 日本ではかつて脳血管性認知症が最も多かったが, 1990年代に入ってからADが多くなった。2000年のわが国の推定認知症有病率は7.2%であり, ADはその約40%を占める。一方でAD病因論としてアミロイドβ蛋白 amyloid beta protein (Aβ)による老人斑の形成およびリン酸化タウによる神経原線維変化が認められ, 不溶性Aβの沈着が神経細胞死につながるとするアミロイドカスケード仮説が提唱されてきたが, 最近ではAβのオリゴマーの段階での神経毒性が判明し, オリゴマーこそ病因の主体であるとするオリゴマー仮説が有力視されつつある。

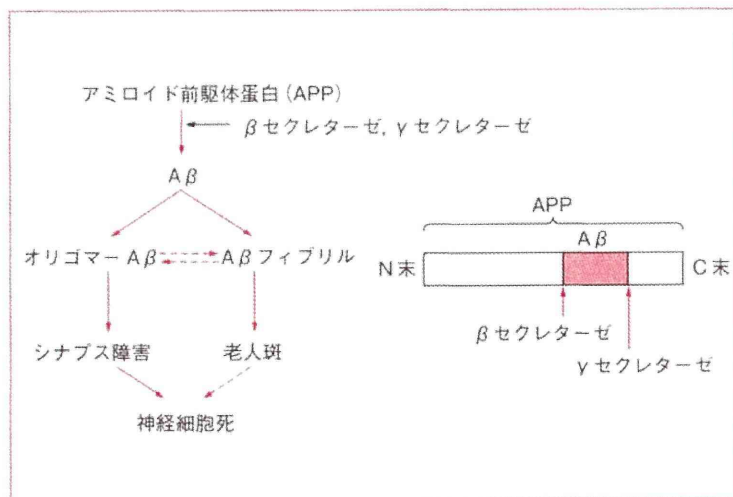
ADの病理●

アミロイド前駆体蛋白 amyloid precursor protein (APP)がβおよびγセクレターゼによっ

て蛋白加水分解を受け, Aβが切り出される(図1)。灰白質のうち神経細胞やグリア細胞の細胞体以外の部分であるニューロピルと呼ばれる細胞外組織周辺にAβが沈着, 異常凝集し, これに伴い神経突起やグリア細胞の変性が起こり, 老人斑 senile plaque (SP)が形成される。SPは最初, 新皮質, 特に前頭・側頭葉の下面に, 次いで一次運動感覚野を除く新皮質に広がってゆく。記憶と関係の深い海馬には初期にはSPは認められない。一方でSPの形成からしばらく遅れて嗅内野, 海馬, 視床を中心に, 神経細胞内における神経原線維変化 neurofibrillary tangle (NFT)が認められるようになり, 前頭・側頭・後頭連合野を含む皮質, 脳幹へ広がってゆく。NFTは2本の線維が捩れ合わさったような構造 paired helical filaments (PHF)の集積物であり, 高度にリン酸化されたタウからなる。Aβの凝集とNFTとの関係はまだ明確ではないが, SP, 神経原線維変化に伴い, 神経細胞数の減少が認められるようになり, グリオーシス, 大脳白質病変などの二次的

図1 アミロイドカスケード仮説およびオリゴマー仮説

図で老人斑が神経細胞死につながるという流れがアミロイドカスケード仮説, オリゴマーAβによるシナプス障害が神経細胞死につながるという流れがオリゴマー仮説, オリゴマーAβのすべてがAβフィブリルを形成するわけではない。矢印は仮想反応を含む。



- AD では SP の形成から遅れて NFT が現れ、初期には海馬には SP が認められない。
- AD ではコリン系、ドパミン系、セロトニン系と広く神経細胞の減少が認められる。
- オリゴマー A β の神経毒性に注目したオリゴマー仮説が有力となってきている。

変化を伴いながら認知機能が低下してゆく。SP および NFT の脳での広がりかたに関し、ブランクのステージ分類がある。ステージ I, II は認知症レベルではなく、ステージ III, IV は初期の AD、あるいは AD の可能性のある段階、ステージ V, VI は確実な AD とされる。SP, NFT 形成あるいは単純萎縮を介し、最終的に起こる神経細胞脱落の分布を 6 層構造でみると II, III 層で最も強い。領域別でみると海馬、下側頭回、内・外側後頭側頭回で最も強い。一方で皮質下でも神経細胞脱落を起こしやすい神経核があり、特にアセチルコリン作動性のマイネルト核では約 30% にまで神経細胞が減少することに注目したコリン仮説は学説としては今や下火となっている。他、ドパミン作動性の青斑核(19~33% への減少)、セロトニン作動性の縫線核(64% への減少)などがみられる¹⁾。

オリゴマー仮説●

NFT と神経細胞脱落の分布領域はほぼ一致しており、NFT と神経細胞死の関連性は密接とされる。一方で A β 単体(モノマー)は可溶性であり、細胞外液中に溶解している状態では神経細胞に対して特別な毒性を発揮しない。SP にみられるのは A β 単体が重合・凝集し、不溶化してフィブリル(線維)となって沈着したものであり、これが神経毒性をもつものと考えられてきた。しかしオリゴマー A β を形成した段階で、まだ可溶性を保ったオリゴマー A β がシナプス後膜に結合すると細胞死を引き起こし、特に海馬シナプスにおける長期増強 long-term potentiation (LTP) を強く抑制することが判明し、また家族性の AD のある一家系にみられた APP の変異体からはフィブリルや SP が形成されず、オリゴマー A β のみ形成されたことが強く示唆されたことからオリゴ

マー A β の神経毒性に注目したオリゴマー仮説が有力となってきている²⁾。

画像診断●

AD では CT, MRI にて海馬を含めた側頭葉前半部と内側部に萎縮が目立ち、頭頂後頭葉にも萎縮が及ぶ。SPECT では頭頂・側頭葉の連合野皮質で血流低下がまずみられ、進行に伴い前頭葉の連合野皮質でも血流低下がみられるようになる。FDG-PET でもまず頭頂・側頭葉の連合野皮質で糖代謝の低下がみられ、前頭葉の連合野皮質での糖代謝の低下とつづく。病理学的変化が起こりにくい一次運動感覚野は SPECT や FDG-PET 所見でも最後まで血流、糖代謝が保たれる。AD では mild cognitive impairment (MCI) と呼ばれる軽度認知機能低下を呈する段階ですでに後部帯状回や楔前部での血流低下・糖代謝の低下が認められる。後部帯状回や楔前部は NFT が初期より出現する嗅内野、海馬と密接な連絡線維をもつ。一方で海馬を含め、側頭葉内側部の血流低下や糖代謝の低下は初期には認められない。

図 2 に PIB-PET の典型的画像を示した。若年健常者では皮質に PIB 集積を認めず、白質の非特異的集積が主体。健常高齢者では加齢に伴い、わずかに皮質に PIB 集積を認める。健常高齢者の PIB シグナルがびまん性老人斑に起因すると結論するには、なお慎重な検証が必要である。AD では明らかに皮質とくに楔前部、後頭帯状回、前頭葉、頭頂葉、外側側頭葉優位に PIB の集積がみられる。MCI は AD ほど著明ではないが、AD と同様の分布で PIB 集積の上昇がみられる症例群がある。逆に PIB 集積の低い MCI 群は AD への移行率が低いことが示されつつある。PIB の意義についての結論は今後の詳細な分析を待たなければならないが、現時点で、その実際的な有用

- AD では MCI の段階ですでに後部帯状回や楔前部で血流低下・糖代謝の低下が認められる。
- 進行に伴い、前頭葉の連合野皮質でも血流低下・糖代謝の低下がみられるようになる。
- MCI には AD 同様に高い PIB 集積を示す症例群と、PIB 集積の低い症例群とが存在する。

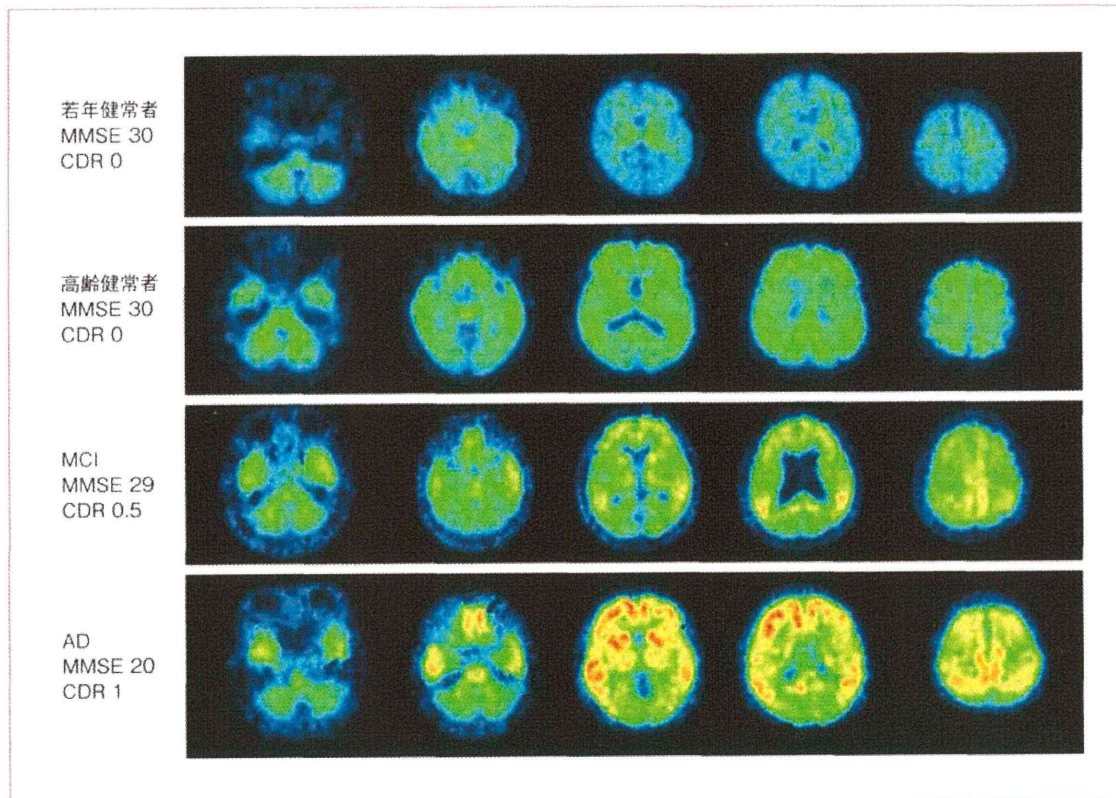


図2 PIB-PET 画像

最上段は若年健常者、2段目が高齢健常者、3段目がMCI、4段目が Alzheimer 病の患者の PIB-PET 画像。各患者の Mini-Mental State Examination (MMSE) と Clinical Dementia Rating (CDR) の点数を付記した。

性は評価に堪えるレベルであろうと考えている。

予防・治療●

本項では AD の病理・病態の正しい理解に裏打ちされた内容が重要である。病因として原因分子であるアミロイドを生成する基質 (APP) および酵素 (presenilin-1, presenilin-2) の阻害薬および消去法が検討されている。現在 β -site APP cleaving enzyme 1 (BACE1) 阻害薬をはじめとす

る β セクレターゼ阻害薬³⁾、 γ セクレターゼ阻害薬により $A\beta$ の産生を抑制する方法や $A\beta$ ワクチン療法などが現在研究開発されている。 $A\beta$ ワクチン療法の作用機序は大きく受動免疫、能動免疫に分けられ、受動免疫によるものに $A\beta$ 抗体、能動免疫によるものに粘膜免疫ワクチン (AAV/ $A\beta$ 経口ワクチン、SeV/ $A\beta$ 経鼻ワクチン)、 $A\beta$ cyclic DNA ワクチン、ペプチド経鼻ワクチンなどがあり、現在も研究がなされている⁴⁾。その他

Medical Practice vol.26 no.12 2009 2035

Presented by Medical*Online

- ADの治療ではセクレターゼ阻害薬, A β ワクチン療法などが現在研究開発されている.
- ADの予防法として生活習慣病への対策, 運動療法などが最近注目されつつある.
- ADでは一般には一次運動野が最後まで保たれるが, 運動障害を顕著に呈する例もある.

ADの危険因子として近年注目を集めるようになった生活習慣病(高脂血症, 高血圧, 糖尿病)および運動療法が新しい視点としてのAD予防法となる可能性が検討されはじめていることを付記しておきたい.

ADにおける運動障害に関する議論は多くない. ただ, presenilin-1変異にみる家族性アルツハイマー病に代表されるように痙性対麻痺, 歩行障害が顕著な臨床像を呈する報告もあり, 該当症例を感覚系と運動系の単なる個別合併症と考えるより, 病態の進行に連鎖した病因論としての視点に立った検討が必要である可能性があり, 今後の重要な研究課題として捉えたい.


おわりに●

オリゴマー仮説のさらなる検証, PET study

を含めた画像診断におけるさらなる知見, 新しい治療法の研究開発に今後の期待がかかる. ADの分子生物学的側面に対する正しい理解こそが正しい診断と治療に直結するものと思われ, 本項がその一助となれば幸いである.

文 献

- 1) 中野今治ほか: よくわかるアルツハイマー病—実際にかかわる人のために—. 永井書店, 2004
- 2) 山本圭一ほか: 可溶性アミロイド β オリゴマーの毒性とアルツハイマー病. 実験医学 26(16): 2560-2565, 2008
- 3) 特集アルツハイマー病研究の最前線—基礎と臨床—. 神経研究の進歩 49(3), 2005
- 4) 田平 武: 治療戦略—免疫療法を中心として—特集アルツハイマー病研究の最前線. 最新医学 61(12): 2428-2433, 2006



Medical Practice 2008年臨時増刊号 (25巻)

新・図解救急・応急処置ガイド

救急・応急時に必ず役立つ基本手技と処置のすべて

編集●Medical Practice編集委員会

◆第一線で活躍する実地医家が, 救急患者の救急・応急処置を的確に施行するために必要なすべてを盛り込んだ実践ガイド, 好評『図解救急・応急処置ガイド 縮刷版』の大改訂版.

●B5判・1,148頁・2色刷/定価**9,450円**(本体9,000円+税5%) 雑誌12078-6

Ⓐ 文光堂 <http://www.bunkodo.co.jp> 〒113-0033 東京都文京区本郷7-2-7 tel.03-3813-5478/fax.03-3813-7241

