

GGN.³⁵⁹ Optimal evaluation of subsolid nodules requires thin-section CT scans (≤ 3 mm thickness) to assess the solid versus ground-glass components.^{359,360}

Radiologic Spectrum According to Histologic Subtype

AAH is the earliest preinvasive lesion for lung adenocarcinoma detectable by thin-section CT. It appears as a small (usually ≤ 5 mm), GGN (Figure 11).^{19,23,129,361–365} AAH characteristically appears as a very faint pure GGN usually measuring ≤ 5 mm.^{130,366} The pure GGN of AAH can be single or multiple.^{129,365,367}

AIS is best demonstrated at CT (preferably thin section) and sometimes can be seen on chest radiography. It is a noninvasive lesion and nonmucinous AIS presents typically as a pure GGN (Figure 12) but sometimes as a part solid or occasionally a solid nodule.^{19,23,128,131,362,367–370} AIS can be bubble like.^{131,365,370,371} Mucinous AIS can appear as a solid nodule or consolidation (Figure 13). The pure GGN of AIS usually appears at thin-section CT as slightly higher attenuation than the very faint GGN of AAH.^{130,366,367} AIS also can be single or multiple.^{19,128,131,365,370}

MIA is variable in its imaging presentation and is, as yet, not fully described, but a provisional description of the nonmucinous type at thin-section CT is a part-solid nodule consisting of a predominant ground-glass component and a small central solid component measuring 5 mm or less (Figure 14).^{47,58} Mucinous MIA (Figure 14) is less common than nonmucinous MIA and appears as a solid or part-solid nodule.^{52,93,126} There is an overlap among imaging features of AAH, AIS, and MIA.

Radiology Recommendation 1

When an opacity in the lung adenocarcinoma spectrum is either a pure GGN or part-solid nodule with a predominant ground-glass component, we recommend that the term BAC no longer be used. These tumors should be classified by the new terms: AIS, MIA, and LPA (strong recommendation, low-quality evidence).

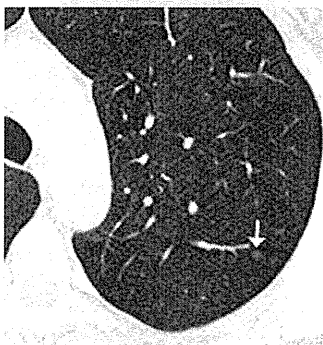


FIGURE 11. CT of preinvasive lesion (AAH or AIS). Axial 2-mm image through the left upper lobe shows a 5 mm pure ground-glass nodule (GGN), which has remained stable for 8 years (arrow). AAH and AIS can be single or multiple. AIS, adenocarcinoma in situ; CT, computed tomography.

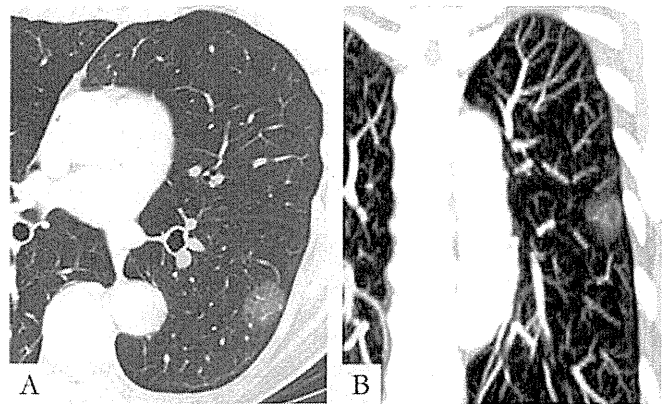


FIGURE 12. CT of a peripheral 2 cm nonmucinous AIS. A, Axial CT section. B, Coronal maximal intensity projection (MIP) image shows a pure GGN in the left lower lobe. Vessels and lung architecture are seen through the nodule. AIS, adenocarcinoma in situ; CT, computed tomography; GGN, ground-glass nodule.

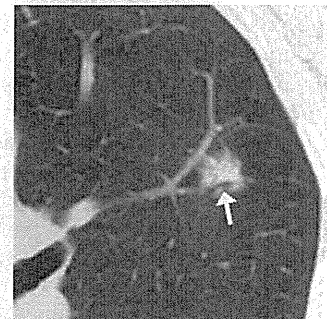


FIGURE 13. CT of mucinous adenocarcinoma in situ; 2 cm predominantly solid nodule with air bronchogram (arrow) is noted in the left upper lobe. CT, computed tomography.

Invasive adenocarcinoma is usually a solid nodule (Figure 15) but may also be part solid (Figure 16) and occasionally a GGN.^{23,58,103,125,129–134,367,370–372} A lobar pattern of ground-glass opacity (GGO) may occur (Figure 17). Bubble-like or cystic lucencies in stage IA adenocarcinoma have been described as correlating with well-differentiated tumors^{131,132,370,371,373,374} and slow growth.^{132,374} Thick (≥ 2 mm) coarse spiculation has been associated with lymph node metastasis, vascular invasion, and decreased survival post resection.^{23,375} For stage IA adenocarcinoma presenting as a part-solid nodule, an extensive ground-glass component suggests a favorable prognosis.^{18,20,23,103,105,376–388} Histologically, the ground-glass component typically corresponds to a lepidic pattern and the solid component to invasive patterns. An intratumoral air bronchogram usually indicates a well-differentiated tumor.^{132,370,375,387} Absence of pleural retraction for stage IA adenocarcinoma is also a favorable prognostic sign.^{375,389} In solid adenocarcinomas, the presence of notches, or concave cuts on thin section CT, has been associated with poor differentiation on histology and adverse outcome.³⁹⁰

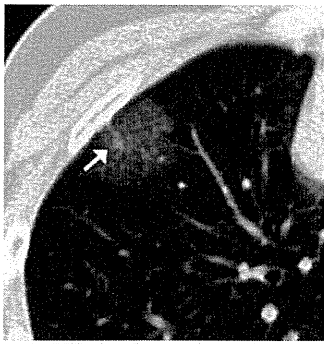


FIGURE 14. CT of nonmucinous minimally invasive adenocarcinoma. Axial 2-mm CT section shows a peripheral, predominantly ground-glass, part-solid nodule in the right upper lobe that includes a 4 × 3 mm solid component (arrow), which corresponded to invasion by pathology. CT, computed tomography.

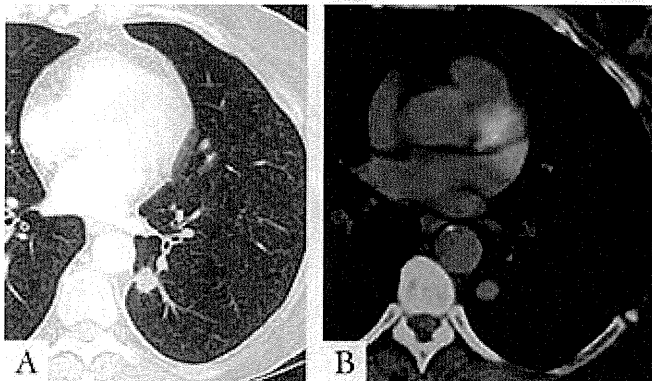


FIGURE 15. CT and FDG-PET of invasive adenocarcinoma. A, Axial CT image and (B) FDG-PET images show a 2-cm spiculated hypermetabolic solid nodule in the left lower lobe. CT, computed tomography; FDG-PET, fluorodeoxyglucose positron emission tomography.

Invasive mucinous adenocarcinoma, formerly called mucinous BAC, characteristically presents in imaging studies as a range of nodules to lobar replacement by a spectrum of patterns including GGO, mixed GGO/solid foci, or consolidation,^{126,128} but intraalveolar mucus may make the CT appearance solid or nearly solid (Figs. 18 and 19).^{125,391} The mucoid component may appear as homogeneous consolidation with soft-tissue attenuation that is lower than that of muscle. After administration of an intravenous iodinated contrast agent, vessels are well shown traversing these regions (CT angiogram sign).^{126,125,133,391} Overlap does occur between imaging features of mucinous and nonmucinous invasive adenocarcinomas.

Radiology Recommendation 2

For overtly invasive adenocarcinomas previously classified as mucinous BAC, we recommend they be separated from nonmucinous adenocarcinomas and be classified as invasive mucinous adenocarcinomas (strong recommendation, moderate quality evidence).

Remark: At CT, this entity is usually solid or mostly solid, has frequent air bronchograms, shows a lobar or multilobar distribution, and frequently consists of multiple nodular or consolidative opacities (former term multicentric BAC).

Size and Growth Rate of Lesions

AAH characteristically is ≤5 mm but in a minority of cases may be up to 12 mm.^{19,276,129,363,364,368} Growth is very slow. Although it has been suggested that a pure GGN less than 5 mm is so unlikely to become a cancer that it needs no follow-up,³⁵⁹ optimal frequency and duration of CT follow-up of a GGN of any size are as yet unclear.

AIS is variable in size, but most are 2 cm or less. Growth is very slow. Suspicious GGNs, i.e., ≥5 mm diameter, are usually followed by at least annual follow-up CT examination, and an increase in size or attenuation is regarded as a sign of probable progression to invasive disease.³⁵⁹ For sizes more than 10 mm, closer follow-up is indicated with CT every 6 months to 1 year. Nevertheless, all recommendations for following suspicious GGNs to date have been based on data from small observational studies and need further evaluation.^{131,361,368,372,387,392–394}

MIA has not yet been well defined in terms of imaging findings, in part, because the histopathologic definition is difficult, and little is known about size and growth rates, but most MIA are less than 2 cm.⁵⁸ Invasive adenocarcinomas of the lung are variable in size and growth rates. For adenocarcinoma less than 2 cm, the smaller the tumor, the less likely there is to be vascular invasion.³⁷³ Size of an adenocarcinoma does predict metastatic disease to the central nervous system: for a node-negative adenocarcinoma of 2 to 6 cm, the probability of metastatic disease to the central nervous system has been reported as 0.14 for a 2 cm tumor, increasing linearly to 0.72 for a 6 cm tumor.^{395,396}

For small solid nodules suspicious for lung cancer at CT, the recommendations for follow-up per Fleischner Society guidelines are currently widely recognized.^{397–399} Nevertheless, these guidelines do not specifically address GGNs and part-solid nodules, as discussed by Godoy and Naidich.³⁵⁹

Because the sizes of many of the clinically problematic nodular lesions at CT are small, how size is measured is especially important. Differences in CT scanners, window settings, and inter- and intraobserver performance are common and may impact critically on assessments of size, especially in the CT follow-up of nodular lesions.^{400–405}

Multiple Primary Lung Cancers

Multifocal lung adenocarcinomas are not uncommon, being found in up to 8 to 22% in surgically resected adenocarcinomas^{406,407} and 18% of adenocarcinomas detected in screening programs.⁶⁴ Multiple lung adenocarcinomas can occur in the setting of multiple AAH, AIS, and invasive adenocarcinoma (Figure 20).³⁶⁵ Similarities or differences in attenuation may provide clues regarding the relative percentage of lepidic versus solid histologic components.³⁵⁹ Subsolid nodules are very rarely metastatic.⁴⁰⁸

Positron Emission Tomography (Scanning)

Elevated standard uptake values (SUVs) on fluorodeoxyglucose positron emission tomography (PET) correlate

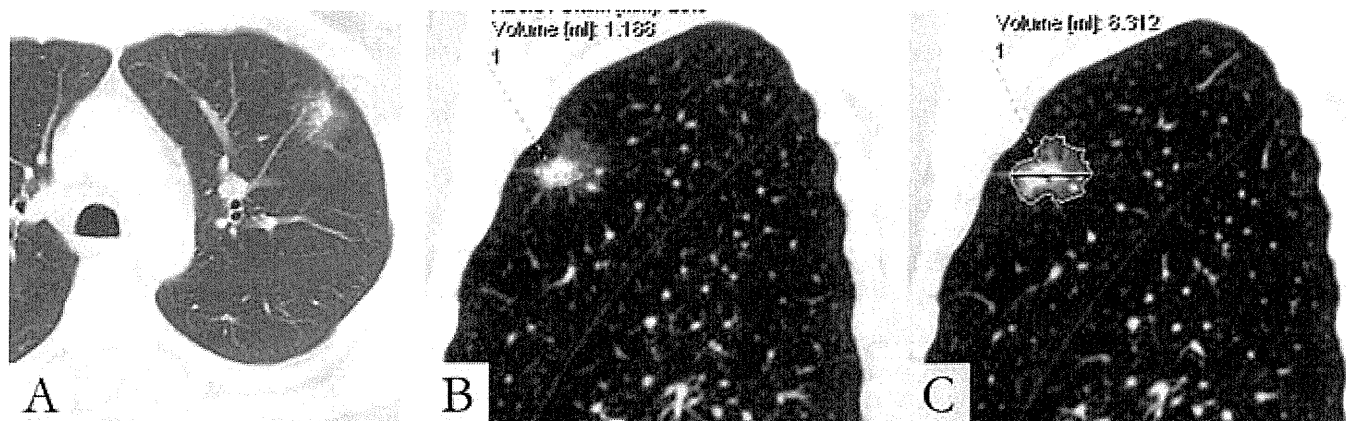


FIGURE 16. Invasive adenocarcinoma. *A*, Axial CT image shows a part-solid nodule in the left upper lobe. *B*, Corresponding sagittal CT images show automated estimation of the volume of (*B*) the solid component (1.188 cm³) and (*C*) the entire lesion (8.312 cm³). In this case, if tumor size were measured only by the invasive component, the size T factor would change from T2a (3.2 cm) to T1a (1.8 cm). Recording of total and invasive sizes are suggested until it is known whether invasive size predicts prognosis better than total size. CT, computed tomography.

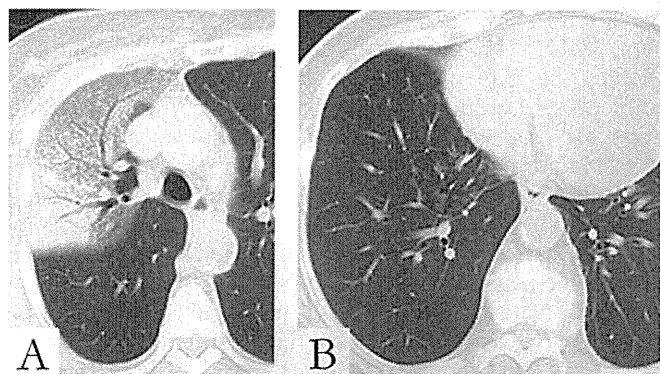


FIGURE 17. CT of nonmucinous lepidic predominant adenocarcinoma. CT images show (*A*) predominantly GGO in the right upper lobe and (*B*) multiple GGN in the right lower lobe. CT, computed tomography; GGN, ground-glass nodule.

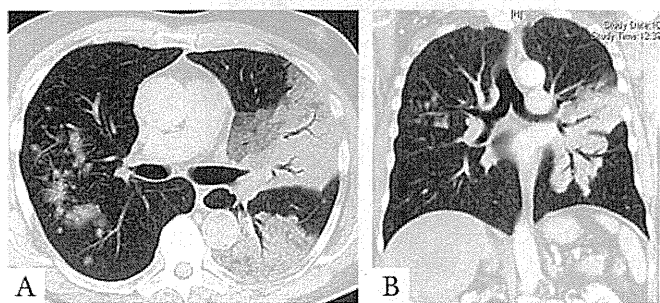


FIGURE 18. CT of invasive mucinous adenocarcinoma. *A*, Axial and (*B*) coronal CT images show multilobar consolidation and nodules mixed with GGO. Air bronchograms are present. CT, computed tomography; GGO, ground-glass opacity.

with cellular proliferation and aggressiveness of the primary cancer (Figures 15 and 19).^{369,409–417} Sensitivity of PET for AIS is usually very low.^{410,414} PET is commonly used for staging and follow-up of invasive adenocarcinoma, and for

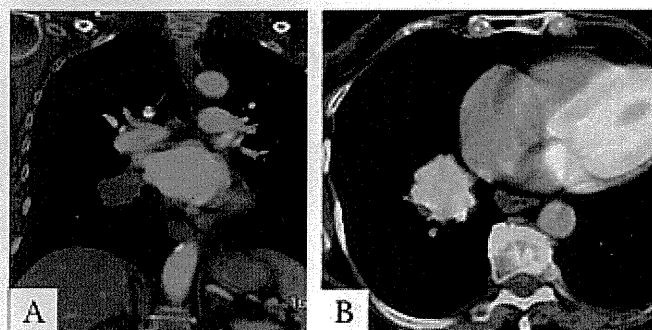


FIGURE 19. CT and FDG PET of invasive mucinous adenocarcinoma. *A*, Coronal CT and (*B*) FDG-PET images show a hypermetabolic hypodense solid 4 cm mass in the right lower lobe. CT, computed tomography; FDG-PET, fluorodeoxyglucose positron emission tomography.

lesions of 7 mm or larger, SUV for adenocarcinoma of the lung tends to be lower than for other histologic types of lung cancer and correlate inversely with survival.^{416,418,419} PET may be more accurate than CT for assessing response to chemotherapy, although more studies are needed.^{420,421} For mucinous versus nonmucinous adenocarcinoma, after adjusting for size of the lesion, no significant difference in SUV has been found.¹²⁵ For a small, well-differentiated adenocarcinoma of low fluorodeoxyglucose avidity (e.g., maximum SUV <2.5), follow-up PET to assess change in SUV as a diagnostic tool unfortunately seems to be of only limited value.⁴²²

Magnetic Resonance

Magnetic resonance has been investigated as a method for differentiating among small AIS, mixed invasive adenocarcinoma/AIS, and invasive adenocarcinoma.^{285,423} In the studies by Ohno et al. and Tanaka et al.,^{285,423} for the distinction of AIS/lepidic predominant (former BAC) from

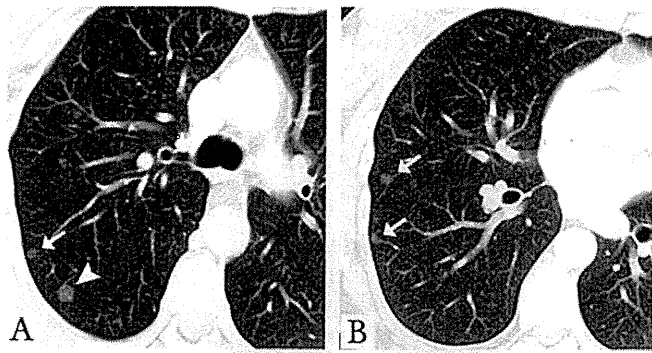


FIGURE 20. CT of multicentric GGNs of AIS/AAH. *A* and *B*, Multiple subsolid nodules (arrows) on axial 3-mm CT images show differing sizes and attenuation. These were presumed to represent preinvasive lesions (AAH and AIS). Because the dominant nodule in the right upper lobe posteriorly near the fissure in part *A* (large arrowhead) appears somewhat dense, it was excised surgically and found to be nonmucinous AIS. AAH, atypical adenomatous hyperplasia; AIS, adenocarcinoma in situ; CT, computed tomography; GGN, ground-glass nodule.

invasive adenocarcinoma, sensitivity was 86% and 97%, and specificity was 100% and 77%, respectively.

Imaging-Guided Percutaneous Needle Biopsy for Molecular and Immunohistochemical Correlations

Percutaneous imaging-guided needle biopsy, whether obtained by aspiration or as a core, allows molecular characterization from even minimal samples.^{200,201,203}

Radiology Recommendations

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Remark: At CT, this entity is usually solid or mostly solid, has frequent air bronchograms, shows a lobar or multilobar distribution, and frequently consists of multiple nodular or consolidative opacities (former term multicentric BAC).

Radiology Considerations for Good Practice

1. Radiologists performing biopsies should obtain sufficient tissue not only for traditional microscopic analysis but also for immunohistochemical and molecular analysis.
2. Thin-section CT technique should be used for part-solid lesions, to record the size of (a) the solid component and

(b) total tumor size, including both solid and ground-glass components (Figure 16).

3. Changes in shape, size, and attenuation help determine follow-up and when intervention is appropriate.

Radiology Research Recommendations

1. What is the natural history of single and multiple GGNs? The frequency of invasive transformation of these lesions is unknown.
2. How should tumor size be measured: single largest diameter, bidimensional, volume? For part-solid lesions, size of both the entire lesion and solid component should be mentioned, because prognosis as a function of size is not yet well established in terms of the dimensions of the solid component. Correlation of the measurement of the solid component of part-solid lesions and size of the invasive component at histopathologic assessment is also needed. Careful attention to thin-section CT technique to assess changes over time in sizes of small adenocarcinomas of the lung is warranted. Inter- and intraobserver differences among radiologists for measurements of the size of a nodule remain an important arena for inquiry.^{400,402} Volumetric measurements also offer promise for assessing changes in size of indeterminate nodules, but error—both human and computer—remains an issue for small GGN, including identifying a possible solid component (Figures 17*B*, *C*).^{405,424,425}
3. What is the CT attenuation according to the newly proposed lung adenocarcinoma histologic subtypes? CT histogram analysis suggests that attenuation characteristics may differ among AAH, AIS, and MIA.³⁶⁶ Further investigations of these lesions using quantitative analysis are in order.
4. In the setting of multiple adenocarcinomas, can careful description of the attenuation (e.g., relative extent of ground glass versus solid components) for each nodule assist in the determination whether the nodules are metastases versus synchronous or metachronous primary carcinomas, similar to the way comprehensive histologic subtyping is helpful pathologically?¹⁰²
5. How can this new classification impact CT screening? Screening may reveal small cancers early in their natural history,^{64,372,426–434} and cost/benefit issues, both medical and economic, remain an arena of active current research.^{424,435–439}
6. What molecular correlations can be made with the spectrum of radiologic patterns of lung adenocarcinoma? Not many studies have attempted correlation of imaging and molecular findings. *EGFR* mutations have been described as correlating with more than 50% GGO,^{271,440,441} with size less than 3.5 cm^{440,442} and with a high SUV level at PET of advanced-stage disease.^{440,443} Ki-67 has been described as associated with high SUV levels at PET^{444,445} and with dedifferentiation of the tumor.⁴⁴⁵

SURGICAL FEATURES

The newly proposed adenocarcinoma classification, particularly introduction of the concepts of AIS and MIA, raises surgical issues to which no definite answer is available yet. This relates to sublobar resection for early-stage lung cancer, role of chest CT in selecting patients for sublobar resection, specific surgical approach for these lesions, extent of lymph node dissection, the role of frozen section analysis, and the treatment of multiple small lung cancers.

Is Sublobar (Limited) Resection Adequate Oncologic Treatment for Some Early Adenocarcinomas?

One of the main reasons for defining the concepts of AIS and MIA in this classification is to raise the question whether these diagnoses can be anticipated by a GGO appearance on CT when presenting as a small, solitary lesion and whether limited resection may be effective therapy for such lesions. Lobectomy is still considered standard surgical treatment for tumors 2 cm or less in size, which have a solid appearance on CT, because such tumors are invasive carcinomas. Whether there can be any change in this standard care for lesions that present with a GGO appearance on CT awaits the results of two randomized trials (Japan Clinical Oncology Group, JCOG 0802 in Japan and CALGB 140503 in North America) that randomize such patients to either lobectomy or sublobar resection. Recently, there have been numerous retrospective studies that have suggested that sublobar (limited) resection for early lung cancers may be adequate surgical treatment; however, these are not randomized trials.^{24–26} Most reports showed no difference in survival or in locoregional recurrence between lobectomy and sublobar resection for tumors 2 cm or less in size. Tumors with a GGN (GGO) appearance on CT are reported to have 100% disease-free survival at 5 years after complete resection.^{18–21}

Can CT be Used to Select Patients for Sublobar Resection?

In performing sublobar resections, several important factors affect the appropriateness of this intervention. These include the location (peripheral versus central), appearance (ground glass versus solid), and size (T1a versus T1b versus T2) of the tumor. CT images, especially obtained by high-resolution CT scan with thin slices, are indispensable to evaluate these factors, and recent studies show rather good image-pathological correlations.³⁵⁹ In recent studies correlating CT findings of GGOs with histopathology, many of these lesions, though not all, correspond to preinvasive, noninvasive, or early forms of neoplastic growth, especially those of adenocarcinoma lineage.^{18–23,359,424}

Is There a Difference in Outcome between Video-Assisted Thoracoscopic surgery versus Thoracotomy in the Treatment of Early-Stage Lung Adenocarcinoma?

Several series suggest that there is no difference in overall survival between patients who have lobectomies performed by video-assisted thoracoscopic surgery (VATS) versus those performed by thoracotomy for clinical stage I non-small cell lung

cancer.^{446,447} Morbidity seems to be lower with the VATS approach. VATS is a standard approach for peripheral wedge resections; VATS segmentectomy is much less widely performed and requires further evaluation.⁴⁴⁸

What can be Expected of Pathologists at Frozen Section?

For a limited resection to be adequate oncologically, a precise pre- and intraoperative diagnosis is critical. The accuracy of intraoperative frozen section analysis in determining whether small lung adenocarcinomas have an invasive component still needs to be defined. The predictive value of frozen section ranges from 93 to 100% but not all articles clearly report the accuracy of frozen section analysis.^{65–67,449}

Evaluation of margins by frozen section may be problematic, especially when stapler cartridges have been used on both sides. Scraping or washing of staple lines with subsequent cytological analysis has been attempted.^{450,451} When a sublobar resection is performed, frozen section analysis of an interlobar, hilar, or any suspicious lymph node is a useful staging evaluation, and when positive nodes are found, a lobectomy is indicated when there is no functional cardiopulmonary limitation.

Should a Systematic Lymph Node Dissection be Performed in Every Early-Stage Adenocarcinoma?

The necessity of systematic hilar and mediastinal lymph node dissection is based on the fact that approximately 20% of pulmonary adenocarcinomas ≤ 20 mm and 5% of cases ≤ 10 mm in size are reported to have nodal metastases.^{452–454} Lobe-specific nodal dissection, which limits dissection to the primary nodal regions draining the involved lobe, has been shown to be a potentially adequate alternative to complete systematic nodal dissection.^{26,455,456} A recently reported multicenter prospective clinical trial randomizing patients with intraoperatively staged T1–2N0 nonhilar N1 NSCLC to lymph node sampling versus systematic nodal dissection showed that systematic nodal dissection identified occult disease in 3.8% of patients but was not associated with a benefit in overall survival.⁴⁵⁷ These results should not be generalized to higher stage tumors. Recent studies also show that in some specific subsets of very early-stage adenocarcinoma, especially GGO lesions, systematic lymph node dissection is not always required.⁴⁵⁸

Multiple Lesions

In the setting of multifocal lung adenocarcinomas, when there is no evidence of mediastinal lymph node invasion, multiple nodules are not a contraindication for surgical exploration.^{64,459} A standard treatment algorithm for multiple lesions has not yet been established. Several factors have to be taken into consideration: number and size of the different nodules, synchronous versus metachronous lesions, ipsilateral versus contralateral, primary versus metastatic lesions, and specific nature (AAH, AIS, and MIA).

Surgery Research Recommendations

1. The precise role of limited resection has not been determined yet because of a lack of randomized prospective trials.
2. The extent of lymph node dissection remains controversial.
3. The accuracy of frozen section in assessing the presence of invasive adenocarcinoma and the accuracy of frozen section or cytology of resection margins in sublobar resections need to be investigated further, and specific guidelines for frozen section analysis should be developed to guide intraoperative decisions.
4. Treatment of multiple lesions has not been standardized.

CLASSIFICATION IN A LOW-RESOURCE SETTING

Although this lung adenocarcinoma classification is written to incorporate special stains and molecular techniques, it is understood that some patients will need to be managed without immunohistochemical or molecular data. This may occur in parts of the world where resources are limited, or it may happen in academic centers where the additional tissue required for special studies is not available. This section briefly outlines how this classification can be applied in such situations.

Pathologic Classification

In the absence of molecular, immunohistochemical, or histochemical testing, the diagnosis and subclassification of lung adenocarcinoma are based purely on light microscopic evaluation of pathologic material.

Resection Specimens

For resection specimens, the two situations where special stains may be useful include solid adenocarcinoma, for which mucin stains can help in the distinction from large cell carcinoma, and for which NE markers can help diagnose LCNEC. In the former situation, if an adenocarcinoma shows a pure solid pattern without acinar, papillary, or lepidic patterns, sometimes intracytoplasmic mucin can be seen on H&E stains. If this cannot be detected, the tumor should be classified as large cell carcinoma, mentioning that it was not possible to perform special stains. If a non-small cell carcinoma shows NE morphology and NE immunohistochemical markers cannot be performed, the tumor should be classified as large cell carcinoma with NE morphology and a specific comment should be made that the tumor could be LCNEC but that material was not available to confirm this immunohistochemically.

Small Biopsies and Cytology

For small biopsies, if clear glandular or squamous differentiation is seen morphologically, the tumor can be classified as adenocarcinoma or squamous cell carcinoma, respectively. If there is some level of uncertainty, this can be reflected by the phrase: poorly differentiated non-small cell carcinoma, favor adenocarcinoma (or squamous cell carcinoma),

mentioning in a comment that special stains were not available, and this diagnosis is based purely on light microscopic morphology. If no morphologic features of glandular or squamous differentiation are seen, the tumor should be classified as poorly differentiated NSCLC-NOS.

Clinical, Radiologic, and Surgical Approach to Aid Management of Patients in the Absence of Molecular or Immunohistochemical Data

Evaluation of patients with lung adenocarcinoma should be no different if the diagnosis is established in the absence of special techniques.

Whenever possible, a chest CT extending to adrenals and liver should be used for radiologic evaluation of such patients. In a low resource setting, chest radiography may reveal the primary lung cancer, pleural effusions, and involvement of lymph nodes or bones; however, given the much lower resolution with radiographs compared with CT, an attempt to obtain a chest CT examination should be made for accurate diagnosis and staging of tumor when possible.

If patients diagnosed in low resource settings may subsequently have tissue tested with molecular or immunohistochemical studies, tissue should be managed appropriately to make this possible.

Clinical management of lung adenocarcinoma patients without information about molecular status such as *EGFR* or *KRAS* mutations consists of standard surgical and chemotherapeutic approaches based on tumor, node, and metastasis (TNM) staging.

IMPLICATIONS OF THIS CLASSIFICATION FOR TNM STAGING

There are several important implications of this new adenocarcinoma classification for staging that need to be considered for the next revision of the TNM classification. The changes relating to the concepts of AIS, MIA, and LPA parallel classification criteria and terminology currently used in breast cancer,⁴⁶⁰ but they would not be applicable to other histologic types of lung cancer. In addition, the comprehensive histologic subtyping approach to assessing invasive adenocarcinomas in this classification provides a useful approach to staging multiple adenocarcinomas.

1. AIS would be classified as Tis. Nevertheless, because carcinoma in situ (CIS) can occur with both lung squamous cell carcinoma and adenocarcinoma, these should be specified as Tis (squamous) or Tis (adenocarcinoma), similar to breast cancer where there is Tis for ductal CIS and Tis for lobular CIS.
2. MIA would be classified as T1mi, similar to microinvasive breast cancer, which defined as an invasive carcinoma with no focus measuring greater than 1 mm; however, the size for MIA is not greater than 5 mm.
3. Also, similar to breast cancer, the size T factor for adenocarcinomas with an in situ or lepidic component may best predict prognosis according only to the size of the invasive component rather than the way it is currently done including total tumor size including both the invasive and the lepidic or in situ components. In

early-stage tumors, the tumor size T factor may need to be adjusted from total tumor size to only the size of the invasive component. This needs to be tested radiologically and pathologically by comparing survival according to total tumor size (GGO plus solid components by CT versus invasive versus in situ/lepidic components pathologically) compared with analysis only by the size of the solid or invasive component by CT and pathology examinations, respectively.

- For multiple lung adenocarcinomas, comprehensive histologic subtyping can help in distinguishing intrapulmonary metastasis versus synchronous or metachronous primaries.¹⁰² The role of molecular testing in this setting is promising but needs further study.³³¹

Many of these concepts need to be tested vigorously in the next 5 years in both early- and advanced-stage lung adenocarcinoma to determine whether they are robust enough to warrant changes in the 8th Edition TNM classification.

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Writing Committee: William D. Travis, Elisabeth Brambilla, Masayuki Noguchi, Andrew G. Nicholson, Kim R. Geisinger, Yasushi Yatabe, David G. Beer, Charles A. Powell, Gregory J. Riely, Paul E. Van Schil, Kavita Garg, John H. M. Austin, Hisao Asamura, Valerie W. Rusch, Fred R. Hirsch, Giorgio Scagliotti, Tetsuya Mitsudomi, Rudolf M. Huber, Yuichi Ishikawa, James Jett, Montserrat Sanchez-Cespedes, Jean-Paul Sculier, Takashi Takahashi, Masahiro Tsuboi, Johan Vansteenkiste, Ignacio Wistuba, and Pan-Chyr Yang.

REFERENCES

- Boyle P, Levin B. World Cancer Report 2008. Lyon: International Agency for Research on Cancer, 2008.
- Parkin DM, Bray F, Ferlay J, et al. Global cancer statistics, 2002. *CA Cancer J Clin* 2005;55:74–108.
- Curado MP, Edwards B, Shin HR, et al. Cancer Incidence in Five Continents, Vol. IX. Lyon: IARC Scientific Publications, 2007.
- Travis WD, Brambilla E, Muller-Hermelink HK, et al. Pathology and Genetics. Tumours of the Lung, Pleura, Thymus and Heart. Lyon, France: IARC Press, 2004.
- Travis WD, Colby TV, Corrin B, et al. Histological Typing of Lung and Pleural Tumors. Berlin: Springer, 1999.
- WHO. Histological Typing of Lung Tumours. Geneva: World Health Organization (WHO), 1967.
- WHO. Histological Typing of Lung Tumors. Geneva: World Health Organization (WHO), 1981.
- Mok TS, Wu YL, Thongprasert S, et al. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. *N Engl J Med* 2009;361:947–957.
- Mitsudomi T, Morita S, Yatabe Y, et al. Gefitinib versus cisplatin plus docetaxel in patients with non-small-cell lung cancer harbouring mutations of the epidermal growth factor receptor (WJTOG3405): an open label, randomised phase 3 trial. *Lancet Oncol* 2010;11:121–128.
- Maemondo M, Inoue A, Kobayashi K, et al. Gefitinib or chemotherapy for non-small-cell lung cancer with mutated *EGFR*. *N Engl J Med* 2010;362:2380–2388.
- Zhou C, Wu Y-L, Chen G, et al. Efficacy results from the randomized phase III OPTIMAL (CTONG 0802) study comparing first-line erlotinib versus carboplatin (CBDCA) plus gemcitabine (GEM) in Chinese advanced non-small cell lung cancer (NSCLC) patients (PTS) with *EGFR* activating mutations. *Ann Oncol* 2010;21 (Suppl. 8):viii1–viii12.
- Scagliotti GV, Park K, Patil S, et al. Survival without toxicity for cisplatin plus pemetrexed versus cisplatin plus gemcitabine in chemonaive patients with advanced non-small cell lung cancer: a risk-benefit analysis of a large phase III study. *Eur J Cancer* 2009;45:2298–2303.
- Scagliotti GV, Parikh P, von Pawel J, et al. Phase III study comparing cisplatin plus gemcitabine with cisplatin plus pemetrexed in chemotherapy-naive patients with advanced-stage non-small-cell lung cancer. *J Clin Oncol* 2008;26:3543–3551.
- Ciuleanu T, Brodowicz T, Zielinski C, et al. Maintenance pemetrexed plus best supportive care versus placebo plus best supportive care for non-small-cell lung cancer: a randomised, double-blind, phase 3 study. *Lancet* 2009;374:1432–1440.
- Scagliotti G, Hanna N, Fossella F, et al. The differential efficacy of pemetrexed according to NSCLC histology: a review of two phase III studies. *Oncologist* 2009;14:253–263.
- Johnson DH, Fehrenbacher L, Novotny WF, et al. Randomized phase II trial comparing bevacizumab plus carboplatin and paclitaxel with carboplatin and paclitaxel alone in previously untreated locally advanced or metastatic non-small-cell lung cancer. *J Clin Oncol* 2004;22:2184–2191.
- Cohen MH, Gootenberg J, Keegan P, et al. FDA drug approval summary: bevacizumab (Avastin) plus Carboplatin and Paclitaxel as first-line treatment of advanced/metastatic recurrent nonsquamous non-small cell lung cancer. *Oncologist* 2007;12:713–718.
- Kodama K, Higashiyama M, Yokouchi H, et al. Prognostic value of ground-glass opacity found in small lung adenocarcinoma on high-resolution CT scanning. *Lung Cancer* 2001;33:17–25.
- Suzuki K, Asamura H, Kusumoto M, et al. “Early” peripheral lung cancer: prognostic significance of ground glass opacity on thin-section computed tomographic scan. *Ann Thorac Surg* 2002;74:1635–1639.
- Takamochi K, Nagai K, Yoshida J, et al. Pathologic N0 status in pulmonary adenocarcinoma is predictable by combining serum carcinoembryonic antigen level and computed tomographic findings. *J Thorac Cardiovasc Surg* 2001;122:325–330.
- Sakurai H, Maeshima A, Watanabe S, et al. Grade of stromal invasion in small adenocarcinoma of the lung: histopathological minimal invasion and prognosis. *Am J Surg Pathol* 2004;28:198–206.
- Adler B, Padley S, Miller RR, et al. High-resolution CT of bronchioalveolar carcinoma. *AJR Am J Roentgenol* 1992;159:275–277.
- Aoki T, Tomoda Y, Watanabe H, et al. Peripheral lung adenocarcinoma: correlation of thin-section CT findings with histologic prognostic factors and survival. *Radiology* 2001;220:803–809.
- El-Sherif A, Gooding WE, Santos R, et al. Outcomes of sublobar resection versus lobectomy for stage I non-small cell lung cancer: a 13-year analysis. *Ann Thorac Surg* 2006;82:408–415.
- Nakamura H, Kawasaki N, Taguchi M, et al. Survival following lobectomy vs limited resection for stage I lung cancer: a meta-analysis. *Br J Cancer* 2005;92:1033–1037.
- Okada M, Koike T, Higashiyama M, et al. Radical sublobar resection for small-sized non-small cell lung cancer: a multicenter study. *J Thorac Cardiovasc Surg* 2006;132:769–775.
- Shah PL, Singh S, Bower M, et al. The role of transbronchial fine needle aspiration in an integrated care pathway for the assessment of patients with suspected lung cancer. *J Thorac Oncol* 2006;1:324–327.
- Edwards SL, Roberts C, McKean ME, et al. Preoperative histological classification of primary lung cancer: accuracy of diagnosis and use of the non-small cell category. *J Clin Pathol* 2000;53:537–540.
- Cataluna JJ, Perpina M, Greses JV, et al. Cell type accuracy of bronchial biopsy specimens in primary lung cancer. *Chest* 1996;109:1199–1203.
- Travis WD, Rekhtman N, Riley GJ, et al. Pathologic diagnosis of advanced lung cancer based on small biopsies and cytology: a paradigm shift. *J Thorac Oncol* 2010;5:411–414.

31. Survey Monkey. Available at: www.surveymonkey.com. 2010.
32. Schunemann HJ, Jaeschke R, Cook DJ, et al. An official ATS statement: grading the quality of evidence and strength of recommendations in ATS guidelines and recommendations. *Am J Respir Crit Care Med* 2006;174:605–614.
33. Schunemann HJ, Oxman AD, Brozek J, et al. GRADE: assessing the quality of evidence for diagnostic recommendations. *ACP J Club* 2008;149:2–3.
34. Schunemann HJ, Oxman AD, Brozek J, et al. Grading quality of evidence and strength of recommendations for diagnostic tests and strategies. *BMJ* 2008;336:1106–1110.
35. Guyatt GH, Oxman AD, Kunz R, et al. Going from evidence to recommendations. *BMJ* 2008;336:1049–1051.
36. Guyatt GH, Oxman AD, Kunz R, et al. What is “quality of evidence” and why is it important to clinicians? *BMJ* 2008;336:995–998.
37. Guyatt GH, Oxman AD, Vist GE, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ* 2008;336:924–926.
38. Guyatt G, Oxman AD, Kunz R, et al. GRADE guidelines: 1. Introduction—GRADE evidence profiles and summary of findings tables. *J Clin Epidemiol*. December 30, 2010 (Epub ahead of print).
39. Loo PS, Thomas SC, Nicolson MC, et al. Subtyping of undifferentiated non-small cell carcinomas in bronchial biopsy specimens. *J Thorac Oncol* 2010;5:442–447.
40. Nicholson AG, Gonzalez D, Shah P, et al. Refining the diagnosis and *EGFR* status of non-small cell lung carcinoma in biopsy and cytologic material, using a panel of mucin staining, TTF-1, cytokeratin 5/6, and P63, and *EGFR* mutation analysis. *J Thorac Oncol* 2010;5:436–441.
41. Barletta JA, Perner S, Iafrate AJ, et al. Clinical significance of TTF-1 protein expression and TTF-1 gene amplification in lung adenocarcinoma. *J Cell Mol Med* 2009;13:1977–1986.
42. Deshpande CG, Geisinger K, Petersen I, et al. Grading of lung adenocarcinoma: architectural versus nuclear approach. *Mod Pathol* 2009;22(Suppl 1):1596.
43. Sica G, Yoshizawa A, Sima CS, et al. A grading system of lung adenocarcinomas based on histologic pattern is predictive of disease recurrence in stage I tumors. *Am J Surg Pathol* 2010;34:1155–1162.
44. Yoshizawa A, Motoi N, Riely GJ, et al. Prognostic significance of the proposed IASLC/ATS/ERS revised classification of lung adenocarcinoma in 514 stage I lung adenocarcinomas. *Mod Pathol*. 2011:24. In press.
45. Thunnissen FB, Beasley MB, Borczuk A, et al. Reproducibility of histopathological subtypes in pulmonary adenocarcinoma. *Mod Pathol* 2010;23:415A.
46. Noguchi M, Morikawa A, Kawasaki M, et al. Small adenocarcinoma of the lung. Histologic characteristics and prognosis. *Cancer* 1995;75:2844–2852.
47. Borczuk AC, Qian F, Kazeros A, et al. Invasive size is an independent predictor of survival in pulmonary adenocarcinoma. *Am J Surg Pathol* 2009;33:462–469.
48. Yim J, Zhu LC, Chiriboga L, et al. Histologic features are important prognostic indicators in early stages lung adenocarcinomas. *Mod Pathol* 2007;20:233–241.
49. Goldstein NS, Mani A, Chmielewski G, et al. Prognostic factors in T1 NO MO adenocarcinomas and bronchioloalveolar carcinomas of the lung. *Am J Clin Pathol* 1999;112:391–402.
50. Clayton F. Bronchioloalveolar carcinomas. Cell types, patterns of growth, and prognostic correlates. *Cancer* 1986;57:1555–1564.
51. Daly RC, Trastek VF, Pairolero PC, et al. Bronchoalveolar carcinoma: factors affecting survival. *Ann Thorac Surg* 1991;51:368–376; discussion 76–77.
52. Manning JT Jr, Spjut HJ, Tschen JA. Bronchioloalveolar carcinoma: the significance of two histopathologic types. *Cancer* 1984;54:525–534.
53. Riquet M, Foucault C, Berna P, et al. Prognostic value of histology in resected lung cancer with emphasis on the relevance of the adenocarcinoma subtyping. *Ann Thorac Surg* 2006;81:1988–1995.
54. Goldstein NS, Thomas M. Mucinous and nonmucinous bronchioloalveolar adenocarcinomas have distinct staining patterns with thyroid transcription factor and cytokeratin 20 antibodies. *Am J Clin Pathol* 2001;116:319–325.
55. Garfield DH, Cadranel J, West HL. Bronchioloalveolar carcinoma: the case for two diseases. *Clin Lung Cancer* 2008;9:24–29.
56. Garfield DH, Cadranel J. The importance of distinguishing mucinous and nonmucinous bronchioloalveolar carcinomas. *Lung* 2009;187:207–208.
57. Garfield DH, Franklin WA. A comparison of survival and disease-specific survival in surgically resected, lymph node-positive bronchioloalveolar carcinoma versus nonsmall cell lung cancer: implications for adjuvant therapy. *Cancer* 2008;113:1107–1108.
58. Travis WD, Garg K, Franklin WA, et al. Evolving concepts in the pathology and computed tomography imaging of lung adenocarcinoma and bronchioloalveolar carcinoma. *J Clin Oncol* 2005;23:3279–3287.
59. West HL, Garfield DH. Bronchioloalveolar carcinoma: not as easy as “BAC.” *J Thorac Oncol* 2009;4:1047–1048.
60. Raz DJ, He B, Rosell R, et al. Bronchioloalveolar carcinoma: a review. *Clin Lung Cancer* 2006;7:313–322.
61. Gandara DR. Bronchioloalveolar carcinoma: the “changing face of lung cancer.” *Clin Lung Cancer* 2006;7:299.
62. Watanabe S, Watanabe T, Arai K, et al. Results of wedge resection for focal bronchioloalveolar carcinoma showing pure ground-glass attenuation on computed tomography. *Ann Thorac Surg* 2002;73:1071–1075.
63. Sakurai H, Dobashi Y, Mizutani E, et al. Bronchioloalveolar carcinoma of the lung 3 centimeters or less in diameter: a prognostic assessment. *Ann Thorac Surg* 2004;78:1728–1733.
64. Vazquez M, Carter D, Brambilla E, et al. Solitary and multiple resected adenocarcinomas after CT screening for lung cancer: histopathologic features and their prognostic implications. *Lung Cancer* 2009;64:148–154.
65. Yamato Y, Tsuchida M, Watanabe T, et al. Early results of a prospective study of limited resection for bronchioloalveolar adenocarcinoma of the lung. *Ann Thorac Surg* 2001;71:971–974.
66. Yamada S, Kohno T. Video-assisted thoracic surgery for pure ground-glass opacities 2 cm or less in diameter. *Ann Thorac Surg* 2004;77:1911–1915.
67. Yoshida J, Nagai K, Yokose T, et al. Limited resection trial for pulmonary ground-glass opacity nodules: fifty-case experience. *J Thorac Cardiovasc Surg* 2005;129:991–996.
68. Koike T, Togashi K, Shirato T, et al. Limited resection for noninvasive bronchioloalveolar carcinoma diagnosed by intraoperative pathologic examination. *Ann Thorac Surg* 2009;88:1106–1111.
69. Motoi N, Szoke J, Riely GJ, et al. Lung adenocarcinoma: modification of the 2004 WHO mixed subtype to include the major histologic subtype suggests correlations between papillary and micropapillary adenocarcinoma subtypes, *EGFR* mutations and gene expression analysis. *Am J Surg Pathol* 2008;32:810–827.
70. Carey FA, Wallace WA, Fergusson RJ, et al. Alveolar atypical hyperplasia in association with primary pulmonary adenocarcinoma: a clinicopathological study of 10 cases. *Thorax* 1992;47:1041–1043.
71. Weng S, Tsuchiya E, Satoh Y, et al. Multiple atypical adenomatous hyperplasia of type II pneumonocytes and bronchio-alveolar carcinoma. *Histopathology* 1990;16:101–103.
72. Nakanishi K. Alveolar epithelial hyperplasia and adenocarcinoma of the lung. *Arch Pathol Lab Med* 1990;114:363–368.
73. Nakahara R, Yokose T, Nagai K, et al. Atypical adenomatous hyperplasia of the lung: a clinicopathological study of 118 cases including cases with multiple atypical adenomatous hyperplasia. *Thorax* 2001;56:302–305.
74. Miller RR. Bronchioloalveolar cell adenomas. *Am J Surg Pathol* 1990;14:904–912.
75. Nakayama H, Noguchi M, Tsuchiya R, et al. Clonal growth of atypical adenomatous hyperplasia of the lung: cytofluorometric analysis of nuclear DNA content. *Mod Pathol* 1990;3:314–320.
76. Niho S, Yokose T, Suzuki K, et al. Monoclonality of atypical adenomatous hyperplasia of the lung. *Am J Pathol* 1999;154:249–254.
77. Sakamoto H, Shimizu J, Horio Y, et al. Disproportionate representation of *KRAS* gene mutation in atypical adenomatous hyperplasia, but even distribution of *EGFR* gene mutation from preinvasive to invasive adenocarcinomas. *J Pathol* 2007;212:287–294.
78. Westra WH, Baas IO, Hruban RH, et al. K-ras oncogene activation in atypical alveolar hyperplasias of the human lung. *Cancer Res* 1996;56:2224–2228.
79. Kohno T, Kunitoh H, Suzuki K, et al. Association of *KRAS* polymor-

- phisms with risk for lung adenocarcinoma accompanied by atypical adenomatous hyperplasias. *Carcinogenesis* 2008;29:957–963.
80. Yoshida Y, Shibata T, Kokubu A, et al. Mutations of the epidermal growth factor receptor gene in atypical adenomatous hyperplasia and bronchioloalveolar carcinoma of the lung. *Lung Cancer* 2005;50:1–8.
 81. Kitamura H, Kameda Y, Nakamura N, et al. Atypical adenomatous hyperplasia and bronchoalveolar lung carcinoma. Analysis by morphometry and the expressions of p53 and carcinoembryonic antigen. *Am J Surg Pathol* 1996;20:553–562.
 82. Takamochi K, Ogura T, Suzuki K, et al. Loss of heterozygosity on chromosomes 9q and 16p in atypical adenomatous hyperplasia concomitant with adenocarcinoma of the lung. *Am J Pathol* 2001;159:1941–1948.
 83. Licchesi JD, Westra WH, Hooker CM, et al. Promoter hypermethylation of hallmark cancer genes in atypical adenomatous hyperplasia of the lung. *Clin Cancer Res* 2008;14:2570–2578.
 84. Nakanishi K, Kawai T, Kumaki F, et al. Expression of human telomerase RNA component and telomerase reverse transcriptase mRNA in atypical adenomatous hyperplasia of the lung. *Hum Pathol* 2002;33:697–702.
 85. Seki N, Takasu T, Mandai K, et al. Expression of eukaryotic initiation factor 4E in atypical adenomatous hyperplasia and adenocarcinoma of the human peripheral lung. *Clin Cancer Res* 2002;8:3046–3053.
 86. Licchesi JD, Westra WH, Hooker CM, et al. Epigenetic alteration of Wnt pathway antagonists in progressive glandular neoplasia of the lung. *Carcinogenesis* 2008;29:895–904.
 87. Kerr KM, MacKenzie SJ, Ramasami S, et al. Expression of Fhit, cell adhesion molecules and matrix metalloproteinases in atypical adenomatous hyperplasia and pulmonary adenocarcinoma. *J Pathol* 2004;203:638–644.
 88. Maeshima AM, Tochigi N, Yoshida A, et al. Clinicopathologic analysis of multiple (five or more) atypical adenomatous hyperplasias (AAHs) of the lung: evidence for the AAH-adenocarcinoma sequence. *J Thorac Oncol* 2010;5:466–471.
 89. Mori M, Rao SK, Popper HH, et al. Atypical adenomatous hyperplasia of the lung: a probable forerunner in the development of adenocarcinoma of the lung. *Mod Pathol* 2001;14:72–84.
 90. Kitamura H, Kameda Y, Ito T, et al. Atypical adenomatous hyperplasia of the lung. Implications for the pathogenesis of peripheral lung adenocarcinoma. *Am J Clin Pathol* 1999;111:610–622.
 91. Koga T, Hashimoto S, Sugio K, et al. Lung adenocarcinoma with bronchioloalveolar carcinoma component is frequently associated with foci of high-grade atypical adenomatous hyperplasia. *Am J Clin Pathol* 2002;117:464–470.
 92. Maeshima AM, Tochigi N, Yoshida A, et al. Histological scoring for small lung adenocarcinomas 2 cm or less in diameter: a reliable prognostic indicator. *J Thorac Oncol* 2010;5:333–339.
 93. Sawada E, Nambu A, Motosugi U, et al. Localized mucinous bronchioloalveolar carcinoma of the lung: thin-section computed tomography and fluorodeoxyglucose positron emission tomography findings. *Jpn J Radiol* 2010;28:251–258.
 94. Oka S, Hanagiri T, Uramoto H, et al. Surgical resection for patients with mucinous bronchioloalveolar carcinoma. *Asian J Surg* 2010;33:89–93.
 95. De Oliveira Duarte Achcar R, Nikiforova MN, Yousem SA. Micropapillary lung adenocarcinoma: *EGFR*, *K-ras*, and *BRAF* mutational profile. *Am J Clin Pathol* 2009;131:694–700.
 96. Nakamura Y, Niki T, Goto A, et al. c-Met activation in lung adenocarcinoma tissues: an immunohistochemical analysis. *Cancer Sci* 2007;98:1006–1013.
 97. Kim YH, Ishii G, Goto K, et al. Dominant papillary subtype is a significant predictor of the response to gefitinib in adenocarcinoma of the lung. *Clin Cancer Res* 2004;10:7311–7317.
 98. Ding L, Getz G, Wheeler DA, et al. Somatic mutations affect key pathways in lung adenocarcinoma. *Nature* 2008;455:1069–1075.
 99. Shedden K, Taylor JM, Enkemann SA, et al. Gene expression-based survival prediction in lung adenocarcinoma: a multi-site, blinded validation study. *Nat Med* 2008;14:822–827.
 100. Sholl LM, Yeap BY, Iafraite AJ, et al. Lung adenocarcinoma with *EGFR* amplification has distinct clinicopathologic and molecular features in never-smokers. *Cancer Res* 2009;69:8341–8348.
 101. Dacic S, Shuai Y, Yousem S, et al. Clinicopathological predictors of *EGFR/KRAS* mutational status in primary lung adenocarcinomas. *Mod Pathol* 2010;23:159–168.
 102. Girard N, Deshpande C, Lau C, et al. Comprehensive histologic assessment helps to differentiate multiple lung primary nonsmall cell carcinomas from metastases. *Am J Surg Pathol* 2009;33:1752–1764.
 103. Lee HY, Han J, Lee KS, et al. Lung adenocarcinoma as a solitary pulmonary nodule: prognostic determinants of CT, PET, and histopathologic findings. *Lung Cancer* 2009;66:379–385.
 104. Yokose T, Suzuki K, Nagai K, et al. Favorable and unfavorable morphological prognostic factors in peripheral adenocarcinoma of the lung 3 cm or less in diameter. *Lung Cancer* 2000;29:179–188.
 105. Lin DM, Ma Y, Zheng S, et al. Prognostic value of bronchioloalveolar carcinoma component in lung adenocarcinoma. *Histol Histopathol* 2006;21:627–632.
 106. Okudela K, Woo T, Mitsui H, et al. Proposal of an improved histological sub-typing system for lung adenocarcinoma—significant prognostic values for stage I disease. *Int J Clin Exp Pathol* 2010;3:348–366.
 107. Silver SA, Askin FB. True papillary carcinoma of the lung: a distinct clinicopathologic entity. *Am J Surg Pathol* 1997;21:43–51.
 108. Amin MB, Tamboli P, Merchant SH, et al. Micropapillary component in lung adenocarcinoma: a distinctive histologic feature with possible prognostic significance. *Am J Surg Pathol* 2002;26:358–364.
 109. Miyoshi T, Satoh Y, Okumura S, et al. Early-stage lung adenocarcinomas with a micropapillary pattern, a distinct pathologic marker for a significantly poor prognosis. *Am J Surg Pathol* 2003;27:101–109.
 110. Kamiya K, Hayashi Y, Douguchi J, et al. Histopathological features and prognostic significance of the micropapillary pattern in lung adenocarcinoma. *Mod Pathol* 2008;21:992–1001.
 111. Kawakami T, Nabeshima K, Makimoto Y, et al. Micropapillary pattern and grade of stromal invasion in pT1 adenocarcinoma of the lung: usefulness as prognostic factors. *Mod Pathol* 2007;20:514–521.
 112. Kuroda N, Hamaguchi N, Takeuchi E, et al. Lung adenocarcinoma with a micropapillary pattern: a clinicopathological study of 25 cases. *APMIS* 2006;114:381–385.
 113. Kuroda N, Hamauzu T, Toi M, et al. Pulmonary adenocarcinoma with micropapillary component: an immunohistochemical study. Case report. *APMIS* 2005;113:550–554.
 114. Maeda R, Isowa N, Onuma H, et al. Lung adenocarcinomas with micropapillary components. *Gen Thorac Cardiovasc Surg* 2009;57:534–539.
 115. Makimoto Y, Nabeshima K, Iwasaki H, et al. Micropapillary pattern: a distinct pathological marker to subclassify tumours with a significantly poor prognosis within small peripheral lung adenocarcinoma ($\leq 20\text{ mm}$) with mixed bronchioloalveolar and invasive subtypes (Noguchi's type C tumours). *Histopathology* 2005;46:677–684.
 116. Miyoshi T, Shirakusa T, Ishikawa Y, et al. Possible mechanism of metastasis in lung adenocarcinomas with a micropapillary pattern. *Pathol Int* 2005;55:419–424.
 117. Roh MS, Lee JI, Choi PJ, et al. Relationship between micropapillary component and micrometastasis in the regional lymph nodes of patients with stage I lung adenocarcinoma. *Histopathology* 2004;45:580–586.
 118. Sanchez-Mora N, Presmanes MC, Monroy V, et al. Micropapillary lung adenocarcinoma: a distinctive histologic subtype with prognostic significance. Case series. *Hum Pathol* 2008;39:324–330.
 119. Tsutsumida H, Nomoto M, Goto M, et al. A micropapillary pattern is predictive of a poor prognosis in lung adenocarcinoma, and reduced surfactant apoprotein A expression in the micropapillary pattern is an excellent indicator of a poor prognosis. *Mod Pathol* 2007;20:638–647.
 120. Wislez M, Antoine M, Baudrin L, et al. Non-mucinous and mucinous subtypes of adenocarcinoma with bronchioloalveolar carcinoma features differ by biomarker expression and in the response to gefitinib. *Lung Cancer* 2010;68:185–191.
 121. Hata A, Katakami N, Fujita S, et al. Frequency of *EGFR* and *KRAS* mutations in Japanese patients with lung adenocarcinoma with features of the mucinous subtype of bronchioloalveolar carcinoma. *J Thorac Oncol* 2010;5:1197–1200.
 122. Yatabe Y, Mitsudomi T. Epidermal growth factor receptor mutations in lung cancers. *Pathol Int* 2007;57:233–244.
 123. Holst VA, Finkelstein S, Yousem SA. Bronchioloalveolar adenocarcinoma of lung: monoclonal origin for multifocal disease. *Am J Surg Pathol* 1998;22:1343–1350.
 124. Furak J, Trojan I, Szoke T, et al. Bronchioloalveolar lung cancer:

- occurrence, surgical treatment and survival. *Eur J Cardiothorac Surg* 2003;23:818–823.
125. Lee HY, Lee KS, Han J, et al. Mucinous versus nonmucinous solitary pulmonary nodular bronchioloalveolar carcinoma: CT and FDG PET findings and pathologic comparisons. *Lung Cancer* 2009;65:170–175.
 126. Miyake H, Matsumoto A, Terada A, et al. Mucin-producing tumor of the lung: CT findings. *J Thorac Imaging* 1995;10:96–98.
 127. Casali C, Rossi G, Marchioni A, et al. A single institution-based retrospective study of surgically treated bronchioloalveolar adenocarcinoma of the lung: clinicopathologic analysis, molecular features, and possible pitfalls in routine practice. *J Thorac Oncol* 2010;5:830–836.
 128. Akira M, Atagi S, Kawahara M, et al. High-resolution CT findings of diffuse bronchioloalveolar carcinoma in 38 patients. *AJR Am J Roentgenol* 1999;173:1623–1629.
 129. Kodama K, Higashiyama M, Yokouchi H, et al. Natural history of pure ground-glass opacity after long-term follow-up of more than 2 years. *Ann Thorac Surg* 2002;73:386–392.
 130. Nagao M, Murase K, Yasuhara Y, et al. Measurement of localized ground-glass attenuation on thin-section computed tomography images: correlation with the progression of bronchioloalveolar carcinoma of the lung. *Invest Radiol* 2002;37:692–697.
 131. Saito H, Yamada K, Hamanaka N, et al. Initial findings and progression of lung adenocarcinoma on serial computed tomography scans. *J Comput Assist Tomogr* 2009;33:42–48.
 132. Yabuuchi H, Murayama S, Murakami J, et al. High-resolution CT characteristics of poorly differentiated adenocarcinoma of the peripheral lung: comparison with well differentiated adenocarcinoma. *Radiat Med* 2000;18:343–347.
 133. Im JG, Han MC, Yu EJ, et al. Lobar bronchioloalveolar carcinoma: 'Angiogram sign' on CT scans. *Radiology* 1990;176:749–753.
 134. Tateishi U, Muller NL, Johkoh T, et al. Mucin-producing adenocarcinoma of the lung: thin-section computed tomography findings in 48 patients and their effect on prognosis. *J Comput Assist Tomogr* 2005;29:361–368.
 135. Clayton F. The spectrum and significance of bronchioloalveolar carcinomas. *Pathol Annu* 1988;23:361–394.
 136. Shah RN, Badve S, Papreddy K, et al. Expression of cytokeratin 20 in mucinous bronchioloalveolar carcinoma. *Hum Pathol* 2002;33:915–920.
 137. Lau SK, Desrochers MJ, Luthringer DJ. Expression of thyroid transcription factor-1, cytokeratin 7, and cytokeratin 20 in bronchioloalveolar carcinomas: an immunohistochemical evaluation of 67 cases. *Mod Pathol* 2002;15:538–542.
 138. Sarantopoulos GP, Gui D, Shintaku P, et al. Immunohistochemical analysis of lung carcinomas with pure or partial bronchioloalveolar differentiation. *Arch Pathol Lab Med* 2004;128:406–414.
 139. Simsir A, Wei XJ, Yee H, et al. Differential expression of cytokeratins 7 and 20 and thyroid transcription factor-1 in bronchioloalveolar carcinoma: an immunohistochemical study in fine-needle aspiration biopsy specimens. *Am J Clin Pathol* 2004;121:350–357.
 140. Finberg KE, Sequist LV, Joshi VA, et al. Mucinous differentiation correlates with absence of *EGFR* mutation and presence of *KRAS* mutation in lung adenocarcinomas with bronchioloalveolar features. *J Mol Diagn* 2007;9:320–326.
 141. Sakuma Y, Matsukuma S, Yoshihara M, et al. Distinctive evaluation of nonmucinous and mucinous subtypes of bronchioloalveolar carcinomas in *EGFR* and *K-ras* gene-mutation analyses for Japanese lung adenocarcinomas: confirmation of the correlations with histologic subtypes and gene mutations. *Am J Clin Pathol* 2007;128:100–108.
 142. Marchetti A, Buttitta F, Pellegrini S, et al. Bronchioloalveolar lung carcinomas: *K-ras* mutations are constant events in the mucinous subtype. *J Pathol* 1996;179:254–259.
 143. Maeshima A, Sakamoto M, Hirohashi S. Mixed mucinous-type and non-mucinous-type adenocarcinoma of the lung: immunohistochemical examination and *K-ras* gene mutation. *Virchows Arch* 2002;440:598–603.
 144. Tam IY, Chung LP, Suen WS, et al. Distinct epidermal growth factor receptor and *KRAS* mutation patterns in non-small cell lung cancer patients with different tobacco exposure and clinicopathologic features. *Clin Cancer Res* 2006;12:1647–1653.
 145. Copin MC, Buisson MP, Leteurte E, et al. Mucinous bronchioloalveolar carcinomas display a specific pattern of mucin gene expression among primary lung adenocarcinomas. *Hum Pathol* 2001;32:274–281.
 146. Awaya H, Takeshima Y, Yamasaki M, et al. Expression of MUC1, MUC2, MUC5AC, and MUC6 in atypical adenomatous hyperplasia, bronchioloalveolar carcinoma, adenocarcinoma with mixed subtypes, and mucinous bronchioloalveolar carcinoma of the lung. *Am J Clin Pathol* 2004;121:644–653.
 147. Sato K, Ueda Y, Shikata H, et al. Bronchioloalveolar carcinoma of mixed mucinous and nonmucinous type: immunohistochemical studies and mutation analysis of the p53 gene. *Pathol Res Pract* 2006;202:751–756.
 148. Tsuta K, Ishii G, Nitadori J, et al. Comparison of the immunophenotypes of signet-ring cell carcinoma, solid adenocarcinoma with mucin production, and mucinous bronchioloalveolar carcinoma of the lung characterized by the presence of cytoplasmic mucin. *J Pathol* 2006;209:78–87.
 149. Sica GL, Yoshizawa AK, Downey RJ, et al. Reassessment of the histologic spectrum of mucinous bronchioloalveolar carcinoma (mBAC). *Mod Pathol* 2008;21:351A.
 150. Gaeta M, Blandino A, Scribano E, et al. Mucinous cystadenocarcinoma of the lung: CT-pathologic correlation in three cases. *J Comput Assist Tomogr* 1999;23:641–643.
 151. Deshpande CG, Yoshizawa A, Motoi N, et al. Clear cell change in lung adenocarcinoma: a cytologic change rather than a histologic variant. *Mod Pathol* 2009;22(Suppl 1):1595.
 152. Cohen PR, Yoshizawa A, Motoi N, et al. Signet ring cell features (SRCF) in lung adenocarcinoma: a cytologic feature or a histologic subtype? *Mod Pathol* 2010;23:404A.
 153. Rodig SJ, Mino-Kenudson M, Dacic S, et al. Unique clinicopathologic features characterize ALK-rearranged lung adenocarcinoma in the western population. *Clin Cancer Res* 2009;15:5216–5223.
 154. Inamura K, Satoh Y, Okumura S, et al. Pulmonary adenocarcinomas with enteric differentiation: histologic and immunohistochemical characteristics compared with metastatic colorectal cancers and usual pulmonary adenocarcinomas. *Am J Surg Pathol* 2005;29:660–665.
 155. Moran CA, Hochholzer L, Fishback N, et al. Mucinous (so-called colloid) carcinomas of lung. *Mod Pathol* 1992;5:634–638.
 156. Rossi G, Murer B, Cavazza A, et al. Primary mucinous (so-called colloid) carcinomas of the lung: a clinicopathologic and immunohistochemical study with special reference to CDX-2 homeobox gene and MUC2 expression. *Am J Surg Pathol* 2004;28:442–452.
 157. Gao ZH, Urbanski SJ. The spectrum of pulmonary mucinous cystic neoplasia: a clinicopathologic and immunohistochemical study of ten cases and review of literature. *Am J Clin Pathol* 2005;124:62–70.
 158. Nakatani Y, Kitamura H, Inayama Y, et al. Pulmonary adenocarcinomas of the fetal lung type: a clinicopathologic study indicating differences in histology, epidemiology, and natural history of low-grade and high-grade forms. *Am J Surg Pathol* 1998;22:399–411.
 159. Nakatani Y, Masudo K, Miyagi Y, et al. Aberrant nuclear localization and gene mutation of beta-catenin in low-grade adenocarcinoma of fetal lung type: up-regulation of the Wnt signaling pathway may be a common denominator for the development of tumors that form morules. *Mod Pathol* 2002;15:617–624.
 160. Sekine S, Shibata T, Matsuno Y, et al. Beta-catenin mutations in pulmonary blastomas: association with morule formation. *J Pathol* 2003;200:214–221.
 161. Li HC, Schmidt L, Greenson JK, et al. Primary pulmonary adenocarcinoma with intestinal differentiation mimicking metastatic colorectal carcinoma: case report and review of literature. *Am J Clin Pathol* 2009;131:129–133.
 162. Hatanaka K, Tsuta K, Watanabe K, et al. Primary pulmonary adenocarcinoma with enteric differentiation resembling metastatic colorectal carcinoma: a report of the second case negative for cytokeratin 7. *Pathol Res Pract*. In press.
 163. Yousem SA. Pulmonary intestinal-type adenocarcinoma does not show enteric differentiation by immunohistochemical study. *Mod Pathol* 2005;18:816–821.
 164. Rossi G, Pelosi G, Graziano P, et al. A reevaluation of the clinical significance of histological subtyping of non-small-cell lung carcinoma: diagnostic algorithms in the era of personalized treatments. *Int J Surg Pathol* 2009;17:206–218.
 165. Rossi G, Papotti M, Barbareschi M, et al. Morphology and a limited

- number of immunohistochemical markers may efficiently subtype non-small-cell lung cancer. *J Clin Oncol* 2009;27:e141–e142; author reply e3–e4.
166. Suh J, Rekhman N, Ladanyi M, et al. Testing of new IASLC/ATS/ERS criteria for diagnosis of lung adenocarcinoma (AD) in small biopsies: minimize immunohistochemistry (IHC) to maximize tissue for molecular studies. *Mod Pathol*. 2011;24 (Supplement 1). In press.
 167. Hanna N, Shepherd FA, Fossella FV, et al. Randomized phase III trial of pemetrexed versus docetaxel in patients with non-small-cell lung cancer previously treated with chemotherapy. *J Clin Oncol* 2004;22:1589–1597.
 168. Yatabe Y, Mitsudomi T, Takahashi T. TTF-1 expression in pulmonary adenocarcinomas. *Am J Surg Pathol* 2002;26:767–773.
 169. Lau SK, Luthringer DJ, Eisen RN. Thyroid transcription factor-1: a review. *Appl Immunohistochem Mol Morphol* 2002;10:97–102.
 170. Camilo R, Capelozzi VL, Siqueira SA, et al. Expression of p63, keratin 5/6, keratin 7, and surfactant-A in non-small cell lung carcinomas. *Hum Pathol* 2006;37:542–546.
 171. Wu M, Wang B, Gil J, et al. p63 and TTF-1 immunostaining. A useful marker panel for distinguishing small cell carcinoma of lung from poorly differentiated squamous cell carcinoma of lung. *Am J Clin Pathol* 2003;119:696–702.
 172. Chu PG, Weiss LM. Expression of cytokeratin 5/6 in epithelial neoplasms: an immunohistochemical study of 509 cases. *Mod Pathol* 2002;15:6–10.
 173. Ordonez NG. Value of thyroid transcription factor-1, E-cadherin, BG8, WT1, and CD44S immunostaining in distinguishing epithelial pleural mesothelioma from pulmonary and nonpulmonary adenocarcinoma. *Am J Surg Pathol* 2000;24:598–606.
 174. Kaufmann O, Dietel M. Thyroid transcription factor-1 is the superior immunohistochemical marker for pulmonary adenocarcinomas and large cell carcinomas compared to surfactant proteins A and B. *Histopathology* 2000;36:8–16.
 175. Kargi A, Gurel D, Tuna B. The diagnostic value of TTF-1, CK 5/6, and p63 immunostaining in classification of lung carcinomas. *Appl Immunohistochem Mol Morphol* 2007;15:415–420.
 176. Khayyata S, Yun S, Pasha T, et al. Value of P63 and CK5/6 in distinguishing squamous cell carcinoma from adenocarcinoma in lung fine-needle aspiration specimens. *Diagn Cytopathol* 2009;37:178–183.
 177. Chu P, Wu E, Weiss LM. Cytokeratin 7 and cytokeratin 20 expression in epithelial neoplasms: a survey of 435 cases. *Mod Pathol* 2000;13:962–972.
 178. Rivera MP, Mehta AC. Initial diagnosis of lung cancer: ACCP evidence-based clinical practice guidelines (2nd edition). *Chest* 2007;132:131S–148S.
 179. Rekhman N, Brandt SM, Sigel CS, et al. Suitability of thoracic cytology for new therapeutic paradigms in non-small cell lung carcinoma: high accuracy of tumor subtyping and feasibility of *EGFR* and *KRAS* molecular testing. *J Thorac Oncol*. In press.
 180. Sigel CS, Friedlander MA, Zakowski MF, et al. Subtyping of non-small cell lung carcinoma (NSCLC): comparison of cytology and small biopsy specimens. *Mod Pathol* 2010;23:414A.
 181. Zhang X, Zhao Y, Wang M, et al. Detection and comparison of epidermal growth factor receptor mutations in cells and fluid of malignant pleural effusion in non-small cell lung cancer. *Lung Cancer* 2008;60:175–182.
 182. Zakowski MF, Hussain S, Pao W, et al. Morphologic features of adenocarcinoma of the lung predictive of response to the epidermal growth factor receptor kinase inhibitors erlotinib and gefitinib. *Arch Pathol Lab Med* 2009;133:470–477.
 183. Wu SG, Gow CH, Yu CJ, et al. Frequent *EGFR* mutations in malignant pleural effusion of lung adenocarcinoma. *Eur Respir J* 2008;32:924–930.
 184. Au NH, Gown AM, Cheang M, et al. P63 expression in lung carcinoma: a tissue microarray study of 408 cases. *Appl Immunohistochem Mol Morphol* 2004;12:240–247.
 185. Ang DC, Ghaffar H, Zakowski MF, et al. Expression of squamous markers in lung adenocarcinoma (AD): clinicopathologic and molecular correlates, and implications for differentiation from squamous cell carcinoma (SqCC). *Mod Pathol* 2010;23:397A.
 186. Ionescu DN, Treaba D, Gilks CB, et al. Non-small cell lung carcinoma with neuroendocrine differentiation—an entity of no clinical or prognostic significance. *Am J Surg Pathol* 2007;31:26–32.
 187. Sterlacci W, Fiegl M, Hilbe W, et al. Clinical relevance of neuroendocrine differentiation in non-small cell lung cancer assessed by immunohistochemistry: a retrospective study on 405 surgically resected cases. *Virchows Arch* 2009;455:125–132.
 188. Chung CK, Zaino R, Stryker JA, et al. Carcinoma of the lung: evaluation of histological grade and factors influencing prognosis. *Ann Thorac Surg* 1982;33:599–604.
 189. Kobayashi N, Toyooka S, Soh J, et al. Risk factors for recurrence and unfavorable prognosis in patients with stage I non-small cell lung cancer and a tumor diameter of 20 mm or less. *J Thorac Oncol* 2007;2:808–812.
 190. Nakazato Y, Minami Y, Kobayashi H, et al. Nuclear grading of primary pulmonary adenocarcinomas—correlation between nuclear size and prognosis. *J Thorac Oncol* 2009;4:S495.
 191. Petersen I, Koth WF, Friedrich KH, et al. Core classification of lung cancer: correlating nuclear size and mitoses with ploidy and clinicopathological parameters. *Lung Cancer* 2009;65:312–318.
 192. Aida S, Shimazaki H, Sato K, et al. Prognostic analysis of pulmonary adenocarcinoma subclassification with special consideration of papillary and bronchioloalveolar types. *Histopathology* 2004;45:468–476.
 193. Gleason DF. Histologic grading of prostate cancer: a perspective. *Hum Pathol* 1992;23:273–279.
 194. Kadota K, Suzuki K, Rusch VW, et al. Nuclear grading system predicts recurrence in stage I lung adenocarcinoma patients. *Mod Pathol*. 2011;24 (Supplement 1). In press.
 195. Li AR, Chitale D, Riely GJ, et al. *EGFR* mutations in lung adenocarcinomas: clinical testing experience and relationship to *EGFR* gene copy number and immunohistochemical expression. *J Mol Diagn* 2008;10:242–248.
 196. Lim EH, Zhang SL, Li JL, et al. Using whole genome amplification (WGA) of low-volume biopsies to assess the prognostic role of *EGFR*, *KRAS*, *p53*, and *CMET* mutations in advanced-stage non-small cell lung cancer (NSCLC). *J Thorac Oncol* 2009;4:12–21.
 197. Savic S, Tapia C, Grilli B, et al. Comprehensive epidermal growth factor receptor gene analysis from cytological specimens of non-small-cell lung cancers. *Br J Cancer* 2008;98:154–160.
 198. Miller VA, Riely GJ, Zakowski MF, et al. Molecular characteristics of bronchioloalveolar carcinoma and adenocarcinoma, bronchioloalveolar carcinoma subtype, predict response to erlotinib. *J Clin Oncol* 2008;26:1472–1478.
 199. Kimura H, Fujiwara Y, Sone T, et al. *EGFR* mutation status in tumour-derived DNA from pleural effusion fluid is a practical basis for predicting the response to gefitinib. *Br J Cancer* 2006;95:1390–1395.
 200. Borczuk AC, Shah L, Pearson GD, et al. Molecular signatures in biopsy specimens of lung cancer. *Am J Respir Crit Care Med* 2004;170:167–174.
 201. Zudaire I, Lozano MD, Vazquez MF, et al. Molecular characterization of small peripheral lung tumors based on the analysis of fine needle aspirates. *Histol Histopathol* 2008;23:33–40.
 202. Gordon GJ, Richards WG, Sugarbaker DJ, et al. A prognostic test for adenocarcinoma of the lung from gene expression profiling data. *Cancer Epidemiol Biomarkers Prev* 2003;12:905–910.
 203. Solomon SB, Zakowski MF, Pao W, et al. Core needle lung biopsy specimens: adequacy for *EGFR* and *KRAS* mutational analysis. *AJR Am J Roentgenol* 2010;194:266–269.
 204. Asano H, Toyooka S, Tokumo M, et al. Detection of *EGFR* gene mutation in lung cancer by mutant-enriched polymerase chain reaction assay. *Clin Cancer Res* 2006;12:43–48.
 205. Otani H, Toyooka S, Soh J, et al. Detection of *EGFR* gene mutations using the wash fluid of CT-guided biopsy needle in NSCLC patients. *J Thorac Oncol* 2008;3:472–476.
 206. Bepler G, Kusmartseva I, Sharma S, et al. RRM1 modulated in vitro and in vivo efficacy of gemcitabine and platinum in non-small-cell lung cancer. *J Clin Oncol* 2006;24:4731–4737.
 207. Olausson KA, Dunant A, Fouret P, et al. DNA repair by ERCC1 in non-small-cell lung cancer and cisplatin-based adjuvant chemotherapy. *N Engl J Med* 2006;355:983–991.
 208. Chang MH, Ahn JS, Lee J, et al. The efficacy of pemetrexed as a third- or fourth-line therapy and the significance of thymidylate synthase

- expression in patients with advanced non-small cell lung cancer. *Lung Cancer* 2010;69:323–329.
209. Monica V, Scagliotti GV, Ceppi P, et al. Differential thymidylate synthase expression in different variants of large-cell carcinoma of the lung. *Clin Cancer Res* 2009;15:7547–7552.
 210. Kang CH, Jang BG, Kim DW, et al. The prognostic significance of ERCC1, BRCA1, XRCC1, and betaII-tubulin expression in patients with non-small cell lung cancer treated by platinum- and taxane-based neoadjuvant chemotherapy and surgical resection. *Lung Cancer* 2010; 68:478–483.
 211. Rosell R, Perez-Roca L, Sanchez JJ, et al. Customized treatment in non-small-cell lung cancer based on *EGFR* mutations and *BRCA1* mRNA expression. *PLoS One* 2009;4:e5133.
 212. Savci-Heijink CD, Kosari F, Aubry MC, et al. The role of desmoglein-3 in the diagnosis of squamous cell carcinoma of the lung. *Am J Pathol* 2009;174:1629–1637.
 213. Monica V, Ceppi P, Righi L, et al. Desmocollin-3: a new marker of squamous differentiation in undifferentiated large-cell carcinoma of the lung. *Mod Pathol* 2009;22:709–717.
 214. Bishop JA, Sharma R, Illei PB. Napsin A and thyroid transcription factor-1 expression in carcinomas of the lung, breast, pancreas, colon, kidney, thyroid, and malignant mesothelioma. *Hum Pathol* 2010;41: 20–25.
 215. Paez JG, Janne PA, Lee JC, et al. *EGFR* mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* 2004; 304:1497–1500.
 216. Lynch TJ, Bell DW, Sordella R, et al. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med* 2004;350:2129–2139.
 217. Pao W, Miller V, Zakowski M, et al. *EGF* receptor gene mutations are common in lung cancers from “never smokers” and are associated with sensitivity of tumors to gefitinib and erlotinib. *Proc Natl Acad Sci USA* 2004;101:13306–13311.
 218. Azzoli CG, Baker S Jr, Temin S, et al. American Society of Clinical Oncology Clinical Practice Guideline update on chemotherapy for stage IV non-small-cell lung cancer. *J Clin Oncol* 2009;27:6251–6266.
 219. Pao W, Kris MG, Iafrate AJ, et al. Integration of molecular profiling into the lung cancer clinic. *Clin Cancer Res* 2009;15:5317–5322.
 220. Shaw AT, Yeap BY, Mino-Kenudson M, et al. Clinical features and outcome of patients with non-small-cell lung cancer who harbor *EML4-ALK*. *J Clin Oncol* 2009;27:4247–4253.
 221. Deterbeck FC, Jantz MA, Wallace M, et al. Invasive mediastinal staging of lung cancer: ACCP evidence-based clinical practice guidelines (2nd edition). *Chest* 2007;132:202S–220S.
 222. Schrupp DS, Giaccone G, Kelsey CR, et al. Non-small cell lung cancer. In VT DeVita, TS Lawrence, SA Rosenberg (Eds.). *Cancer, Principles and Practice of Oncology*, 7th Ed. Philadelphia: Wolters Kluwer; Lippincott, Williams & Wilkins, 2008. Pp. 887–895.
 223. Sculier JP, Chansky K, Crowley JJ, et al. The impact of additional prognostic factors on survival and their relationship with the anatomical extent of disease expressed by the 6th Edition of the TNM Classification of Malignant Tumors and the proposals for the 7th Edition. *J Thorac Oncol* 2008;3:457–466.
 224. Chansky K, Sculier JP, Crowley JJ, et al. The International Association for the Study of Lung Cancer Staging Project: prognostic factors and pathologic TNM stage in surgically managed non-small cell lung cancer. *J Thorac Oncol* 2009;4:792–801.
 225. Janjigian YY, McDonnell K, Kris MG, et al. Pack-years of cigarette smoking as a prognostic factor in patients with stage IIIB/IV Non-small cell lung cancer. *Cancer* 2010;116:670–675.
 226. Miller VA, Kris MG, Shah N, et al. Bronchioloalveolar pathologic subtype and smoking history predict sensitivity to gefitinib in advanced non-small-cell lung cancer. *J Clin Oncol* 2004;22:1103–1109.
 227. West HL, Franklin WA, McCoy J, et al. Gefitinib therapy in advanced bronchioloalveolar carcinoma: Southwest Oncology Group Study S0126. *J Clin Oncol* 2006;24:1807–1813.
 228. Sequist LV, Bell DW, Lynch TJ, et al. Molecular predictors of response to epidermal growth factor receptor antagonists in non-small-cell lung cancer. *J Clin Oncol* 2007;25:587–595.
 229. Sutani A, Nagai Y, Udagawa K, et al. Gefitinib for non-small-cell lung cancer patients with epidermal growth factor receptor gene mutations screened by peptide nucleic acid-locked nucleic acid PCR clamp. *Br J Cancer* 2006;95:1483–1489.
 230. Inoue A, Suzuki T, Fukuhara T, et al. Prospective phase II study of gefitinib for chemotherapy-naïve patients with advanced non-small-cell lung cancer with epidermal growth factor receptor gene mutations. *J Clin Oncol* 2006;24:3340–3346.
 231. Inoue A, Kobayashi K, Usui K, et al. First-line gefitinib for patients with advanced non-small-cell lung cancer harboring epidermal growth factor receptor mutations without indication for chemotherapy. *J Clin Oncol* 2009;27:1394–1400.
 232. Tamura K, Okamoto I, Kashii T, et al. Multicentre prospective phase II trial of gefitinib for advanced non-small cell lung cancer with epidermal growth factor receptor mutations: results of the West Japan Thoracic Oncology Group trial (WJTOG0403). *Br J Cancer* 2008;98: 907–914.
 233. Yoshida K, Yatabe Y, Park JY, et al. Prospective validation for prediction of gefitinib sensitivity by *epidermal growth factor receptor* gene mutation in patients with non-small cell lung cancer. *J Thorac Oncol* 2007;2:22–28.
 234. Douillard JY, Shepherd FA, Hirsh V, et al. Molecular predictors of outcome with gefitinib and docetaxel in previously treated non-small-cell lung cancer: data from the randomized phase III INTEREST trial. *J Clin Oncol* 2010;28:744–752.
 235. Hirsch FR, Varella-Garcia M, Bunn PA Jr, et al. Molecular predictors of outcome with gefitinib in a phase III placebo-controlled study in advanced non-small-cell lung cancer. *J Clin Oncol* 2006;24:5034–5042.
 236. Zhu CQ, da Cunha Santos G, Ding K, et al. Role of *KRAS* and *EGFR* as biomarkers of response to erlotinib in National Cancer Institute of Canada Clinical Trials Group Study BR. 21. *J Clin Oncol* 2008;26: 4268–4275.
 237. Sholl LM, Xiao Y, Joshi V, et al. *EGFR* mutation is a better predictor of response to tyrosine kinase inhibitors in non-small cell lung carcinoma than FISH, CISH, and immunohistochemistry. *Am J Clin Pathol* 2010;133:922–934.
 238. Kawahara A, Yamamoto C, Nakashima K, et al. Molecular diagnosis of activating *EGFR* mutations in non-small cell lung cancer using mutation specific antibodies for immunohistochemical analysis. *Clin Cancer Res* 2010;16:3163–3170.
 239. Brevet M, Arcila M, Ladanyi M. Assessment of *EGFR* mutation status in lung adenocarcinoma by immunohistochemistry using antibodies specific to the two major forms of mutant *EGFR*. *J Mol Diagn* 2010;12:169–176.
 240. Yu J, Kane S, Wu J, et al. Mutation-specific antibodies for the detection of *EGFR* mutations in non-small-cell lung cancer. *Clin Cancer Res* 2009;15:3023–3028.
 241. Fukuoka M, Wu Y, Thongprasert S, et al. Biomarker analyses from a phase III, randomized, open-label, first-line study of gefitinib (G) versus carboplatin/paclitaxel (C/P) in clinically selected patients (pts) with advanced non-small-cell lung cancer (NSCLC) in Asia (IPASS). *J Clin Oncol* 2009;27:521S.
 242. Kubota K, Niho S, Enatsu S, et al. Efficacy differences of pemetrexed by histology in pretreated patients with stage IIIB/IV non-small cell lung cancer: review of results from an open-label randomized phase II study. *J Thorac Oncol* 2009;4:1530–1536.
 243. Zinner RG, Novello S, Peng G, et al. Comparison of patient outcomes according to histology among pemetrexed-treated patients with stage IIIB/IV non-small-cell lung cancer in two phase II trials. *Clin Lung Cancer* 2010;11:126–131.
 244. Gronberg BH, Bremnes RM, Flotten O, et al. Phase III study by the Norwegian lung cancer study group: pemetrexed plus carboplatin compared with gemcitabine plus carboplatin as first-line chemotherapy in advanced non-small-cell lung cancer. *J Clin Oncol* 2009;27:3217–3224.
 245. Sandler A, Gray R, Perry MC, et al. Paclitaxel-carboplatin alone or with bevacizumab for non-small-cell lung cancer. *N Engl J Med* 2006;355:2542–2550.
 246. Soda M, Choi YL, Enomoto M, et al. Identification of the transforming *EML4-ALK* fusion gene in non-small-cell lung cancer. *Nature* 2007; 448:561–566.
 247. Wong DW, Leung EL, So KK, et al. The *EML4-ALK* fusion gene is

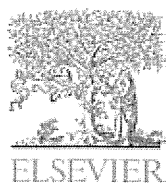
- involved in various histologic types of lung cancers from nonsmokers with wild-type *EGFR* and *KRAS*. *Cancer* 2009;115:1723–1733.
248. Kwak EL, Bang YJ, Camidge DR, et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 2010;363:1693–1703.
 249. Choi YL, Soda M, Yamashita Y, et al. *EML4-ALK* mutations in lung cancer that confer resistance to ALK inhibitors. *N Engl J Med* 2010;363:1734–1739.
 250. Mao C, Qiu LX, Liao RY, et al. *KRAS* mutations and resistance to EGFR-TKIs treatment in patients with non-small cell lung cancer: a meta-analysis of 22 studies. *Lung Cancer* 2010;69:272–278.
 251. Linardou H, Dahabreh IJ, Kanaloupiti D, et al. Assessment of somatic *k-RAS* mutations as a mechanism associated with resistance to EGFR-targeted agents: a systematic review and meta-analysis of studies in advanced non-small-cell lung cancer and metastatic colorectal cancer. *Lancet Oncol* 2008;9:962–972.
 252. Goldstraw P. IASLC Staging Manual in Thoracic Oncology. Orange Park, FL: International Association for the Study of Lung Cancer, Editorial Rx Press, 2009.
 253. Goldstraw P, Crowley J, Chansky K, et al. The IASLC Lung Cancer Staging Project: proposals for the revision of the TNM stage groupings in the forthcoming (seventh) edition of the TNM Classification of malignant tumours. *J Thorac Oncol* 2007;2:706–714.
 254. Pignon JP, Tribodet H, Scagliotti GV, et al. Lung adjuvant cisplatin evaluation: a pooled analysis by the LACE Collaborative Group. *J Clin Oncol* 2008;26:3552–3559.
 255. Pao W, Wang TY, Riely GJ, et al. *KRAS* mutations and primary resistance of lung adenocarcinomas to gefitinib or erlotinib. *PLoS Med* 2005;2:e17.
 256. Kim CF, Jackson EL, Woolfenden AE, et al. Identification of bronchioalveolar stem cells in normal lung and lung cancer. *Cell* 2005;121:823–835.
 257. Weir BA, Woo MS, Getz G, et al. Characterizing the cancer genome in lung adenocarcinoma. *Nature* 2007;450:893–898.
 258. Tanaka H, Yanagisawa K, Shinjo K, et al. Lineage-specific dependency of lung adenocarcinomas on the lung development regulator TTF-1. *Cancer Res* 2007;67:6007–6011.
 259. Colby TV, Leslie KO, Yousem SA. Lungs. In SE Mills (Ed.). *Histology for Pathologists*. 3rd ed. Philadelphia: Lippincott Williams & Wilkins, 2007, pp 473–504.
 260. Wang BY, Gil J, Kaufman D, et al. P63 in pulmonary epithelium, pulmonary squamous neoplasms, and other pulmonary tumors. *Hum Pathol* 2002;33:921–926.
 261. Takeuchi T, Tomida S, Yatabe Y, et al. Expression profile-defined classification of lung adenocarcinoma shows close relationship with underlying major genetic changes and clinicopathologic behaviors. *J Clin Oncol* 2006;24:1679–1688.
 262. Colby TV, Wistuba II, Gazdar A. Precursors to pulmonary neoplasia. *Adv Anat Pathol* 1998;5:205–215.
 263. Westra WH. Early glandular neoplasia of the lung. *Respir Res* 2000;1:163–169.
 264. Tang X, Varella-Garcia M, Xavier AC, et al. Epidermal growth factor receptor abnormalities in the pathogenesis and progression of lung adenocarcinomas. *Cancer Prev Res (Phila Pa)* 2008;1:192–200.
 265. Tang X, Shigematsu H, Bekele BN, et al. EGFR tyrosine kinase domain mutations are detected in histologically normal respiratory epithelium in lung cancer patients. *Cancer Res* 2005;65:7568–7572.
 266. Soh J, Toyooka S, Ichihara S, et al. Sequential molecular changes during multistage pathogenesis of small peripheral adenocarcinomas of the lung. *J Thorac Oncol* 2008;3:340–347.
 267. Yatabe Y, Takahashi T, Mitsudomi T. *Epidermal growth factor receptor* gene amplification is acquired in association with tumor progression of *EGFR*-mutated lung cancer. *Cancer Res* 2008;68:2106–2111.
 268. Tang ZQ, Han LY, Lin HH, et al. Derivation of stable microarray cancer-differentiating signatures using consensus scoring of multiple random sampling and gene-ranking consistency evaluation. *Cancer Res* 2007;67:9996–10003.
 269. Koga T, Hashimoto S, Sugio K, et al. Clinicopathological and molecular evidence indicating the independence of bronchioalveolar components from other subtypes of human peripheral lung adenocarcinoma. *Clin Cancer Res* 2001;7:1730–1738.
 270. Marchetti A, Pellegrini S, Bertacca G, et al. *FHIT* and *p53* gene abnormalities in bronchioalveolar carcinomas. Correlations with clinicopathological data and *K-ras* mutations. *J Pathol* 1998;184:240–246.
 271. Yoshida Y, Kokubu A, Suzuki K, et al. Molecular markers and changes of computed tomography appearance in lung adenocarcinoma with ground-glass opacity. *Jpn J Clin Oncol* 2007;37:907–912.
 272. Terasaki H, Niki T, Matsuno Y, et al. Lung adenocarcinoma with mixed bronchioalveolar and invasive components: clinicopathological features, subclassification by extent of invasive foci, and immunohistochemical characterization. *Am J Surg Pathol* 2003;27:937–951.
 273. Huang CL, Taki T, Adachi M, et al. Mutations of *p53* and *K-ras* genes as prognostic factors for non-small cell lung cancer. *Int J Oncol* 1998;12:553–563.
 274. Shigematsu H, Lin L, Takahashi T, et al. Clinical and biological features associated with *epidermal growth factor receptor* gene mutations in lung cancers. *J Natl Cancer Inst* 2005;97:339–346.
 275. Marchetti A, Martella C, Felicioni L, et al. *EGFR* mutations in non-small-cell lung cancer: analysis of a large series of cases and development of a rapid and sensitive method for diagnostic screening with potential implications on pharmacologic treatment. *J Clin Oncol* 2005;23:857–865.
 276. Ohtsuka T, Watanabe K, Kaji M, et al. A clinicopathological study of resected pulmonary nodules with focal pure ground-glass opacity. *Eur J Cardiothorac Surg* 2006;30:160–163.
 277. Sonobe M, Manabe T, Wada H, et al. Mutations in the *epidermal growth factor receptor* gene are linked to smoking-independent, lung adenocarcinoma. *Br J Cancer* 2005;93:355–363.
 278. Bryant CM, Albertus DL, Kim S, et al. Clinically relevant characterization of lung adenocarcinoma subtypes based on cellular pathways: an international validation study. *PLoS One* 2010;5:e11712.
 279. Yousem SA, Nikiforova M, Nikiforov Y. The histopathology of *BRAF-V600E*-mutated lung adenocarcinoma. *Am J Surg Pathol* 2008;32:1317–1321.
 280. Conde E, Angulo B, Tang M, et al. Molecular context of the *EGFR* mutations: evidence for the activation of *mTOR/S6K* signaling. *Clin Cancer Res* 2006;12:710–717.
 281. Ohtsuka K, Ohnishi H, Furuyashiki G, et al. Clinico-pathological and biological significance of tyrosine kinase domain gene mutations and overexpression of epidermal growth factor receptor for lung adenocarcinoma. *J Thorac Oncol* 2006;1:787–795.
 282. Sonobe M, Manabe T, Wada H, et al. Lung adenocarcinoma harboring mutations in the ERBB2 kinase domain. *J Mol Diagn* 2006;8:351–356.
 283. Ninomiya H, Hiramatsu M, Inamura K, et al. Correlation between morphology and *EGFR* mutations in lung adenocarcinomas. Significance of the micropapillary pattern and the hobnail cell type. *Lung Cancer* 2009;63:235–240.
 284. Hirsch FR, Varella-Garcia M, McCoy J, et al. Increased epidermal growth factor receptor gene copy number detected by fluorescence in situ hybridization associates with increased sensitivity to gefitinib in patients with bronchioalveolar carcinoma subtypes: a southwest oncology group study. *J Clin Oncol* 2005;23:6838–6845.
 285. Tanaka R, Horikoshi H, Nakazato Y, et al. Magnetic resonance imaging in peripheral lung adenocarcinoma: correlation with histopathologic features. *J Thorac Imaging* 2009;24:4–9.
 286. Stenhouse G, Fyfe N, King G, et al. Thyroid transcription factor 1 in pulmonary adenocarcinoma. *J Clin Pathol* 2004;57:383–387.
 287. Kim YT, Kim TY, Lee DS, et al. Molecular changes of *epidermal growth factor receptor (EGFR)* and *KRAS* and their impact on the clinical outcomes in surgically resected adenocarcinoma of the lung. *Lung Cancer* 2008;59:111–118.
 288. Ang DC, Zakowski MF, Ladanyi M, et al. Characteristic morphology and immunoprofile of lung adenocarcinoma with *KRAS* mutations: propensity for solid growth pattern and correlation with TTF-1 expression. *Mod Pathol* 2010;23(Suppl):396A.
 289. Saad RS, Cho P, Silverman JF, et al. Usefulness of Cdx2 in separating mucinous bronchioalveolar adenocarcinoma of the lung from metastatic mucinous colorectal adenocarcinoma. *Am J Clin Pathol* 2004;122:421–427.
 290. Shrestha B, Ebihara Y, Osakabe Y, et al. Immunohistochemical, ultrastructural and molecular study of well differentiated adenocarcinomas of the lung predominantly composed of goblet cells. *Lung Cancer* 1998;22:103–117.
 291. Yatabe Y, Koga T, Mitsudomi T, et al. CK20 expression, CDX2

- expression, K-ras mutation, and goblet cell morphology in a subset of lung adenocarcinomas. *J Pathol* 2004;203:645–652.
292. Sasaki H, Kawano O, Endo K, et al. Uncommon V599E *BRAF* mutations in Japanese patients with lung cancer. *J Surg Res* 2006;133:203–206.
 293. Naoki K, Chen TH, Richards WG, et al. Missense mutations of the *BRAF* gene in human lung adenocarcinoma. *Cancer Res* 2002;62:7001–7003.
 294. Tang Z, Du R, Jiang S, et al. Dual *MET-EGFR* combinatorial inhibition against T790M-EGFR-mediated erlotinib-resistant lung cancer. *Br J Cancer* 2008;99:911–922.
 295. Cappuzzo F, Janne PA, Skokan M, et al. *MET* increased gene copy number and primary resistance to gefitinib therapy in non-small-cell lung cancer patients. *Ann Oncol* 2009;20:298–304.
 296. Engelman JA, Zejnullahu K, Mitsudomi T, et al. *MET* amplification leads to gefitinib resistance in lung cancer by activating *ERBB3* signaling. *Science* 2007;316:1039–1043.
 297. Pao W, Miller VA, Politi KA, et al. Acquired resistance of lung adenocarcinomas to gefitinib or erlotinib is associated with a second mutation in the EGFR kinase domain. *PLoS Med* 2005;2:e73.
 298. Sequist LV, Martins RG, Spigel D, et al. First-line gefitinib in patients with advanced non-small-cell lung cancer harboring somatic *EGFR* mutations. *J Clin Oncol* 2008;26:2442–2449.
 299. Nguyen KS, Kobayashi S, Costa DB. Acquired resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-small-cell lung cancers dependent on the epidermal growth factor receptor pathway. *Clin Lung Cancer* 2009;10:281–289.
 300. Takahashi T, Sonobe M, Kobayashi M, et al. Clinicopathologic features of non-small-cell lung cancer with *EML4-ALK* fusion gene. *Ann Surg Oncol* 2010;17:889–897.
 301. Inamura K, Takeuchi K, Togashi Y, et al. *EML4-ALK* lung cancers are characterized by rare other mutations, a TTF-1 cell lineage, an acinar histology, and young onset. *Mod Pathol* 2009;22:508–515.
 302. Takeuchi K, Choi YL, Togashi Y, et al. *KIF5B-ALK*, a novel fusion oncokinas identified by an immunohistochemistry-based diagnostic system for *ALK*-positive lung cancer. *Clin Cancer Res* 2009;15:3143–3149.
 303. Yoshida A, Tsuta K, Watanabe SI, et al. Frequent *ALK* rearrangement and TTF-1/p63 co-expression in lung adenocarcinoma with signet-ring cell component. *Lung Cancer*. In press.
 304. Joki R, Yamasaki T, Minami S, et al. Combination of morphological feature analysis and immunohistochemistry is useful for screening of *EML4-ALK*-positive lung adenocarcinoma. *J Clin Pathol* 2010;63:1066–1070.
 305. Inamura K, Takeuchi K, Togashi Y, et al. *EML4-ALK* fusion is linked to histological characteristics in a subset of lung cancers. *J Thorac Oncol* 2008;3:13–17.
 306. Mino-Kenudson M, Chirieac LR, Law K, et al. A novel, highly sensitive antibody allows for the routine detection of *ALK*-rearranged lung adenocarcinomas by standard immunohistochemistry. *Clin Cancer Res* 2010;16:1561–1571.
 307. Sakairi Y, Nakajima T, Yasufuku K, et al. *EML4-ALK* fusion gene assessment using metastatic lymph node samples obtained by endobronchial ultrasound-guided transbronchial needle aspiration. *Clin Cancer Res* 2010;16:4938–4945.
 308. Boland JM, Erdogan S, Vasmataz G, et al. Anaplastic lymphoma kinase immunoreactivity correlates with *ALK* gene rearrangement and transcriptional up-regulation in non-small cell lung carcinomas. *Hum Pathol* 2009;40:1152–1158.
 309. Beer DG, Kardias SL, Huang CC, et al. Gene-expression profiles predict survival of patients with lung adenocarcinoma. *Nat Med* 2002;8:816–824.
 310. Borczuk AC, Kim HK, Yegen HA, et al. Lung adenocarcinoma global profiling identifies type II transforming growth factor-(beta) receptor as a repressor of invasiveness. *Am J Respir Crit Care Med* 2005;172:729–737.
 311. Shibata T, Hanada S, Kokubu A, et al. Gene expression profiling of epidermal growth factor receptor/*KRAS* pathway activation in lung adenocarcinoma. *Cancer Sci* 2007;98:985–991.
 312. Berrar D, Sturgeon B, Bradbury I, et al. Survival trees for analyzing clinical outcome in lung adenocarcinomas based on gene expression profiles: identification of neogenin and diacylglycerol kinase alpha expression as critical factors. *J Comput Biol* 2005;12:534–544.
 313. Bianchi F, Nuciforo P, Vecchi M, et al. Survival prediction of stage I lung adenocarcinomas by expression of 10 genes. *J Clin Invest* 2007;117:3436–3444.
 314. Endoh H, Tomida S, Yatabe Y, et al. Prognostic model of pulmonary adenocarcinoma by expression profiling of eight genes as determined by quantitative real-time reverse transcriptase polymerase chain reaction. *J Clin Oncol* 2004;22:811–819.
 315. Guo L, Ma Y, Ward R, et al. Constructing molecular classifiers for the accurate prognosis of lung adenocarcinoma. *Clin Cancer Res* 2006;12:3344–3354.
 316. Hayes DN, Monti S, Parmigiani G, et al. Gene expression profiling reveals reproducible human lung adenocarcinoma subtypes in multiple independent patient cohorts. *J Clin Oncol* 2006;24:5079–5090.
 317. Inamura K, Shimoji T, Ninomiya H, et al. A metastatic signature in entire lung adenocarcinomas irrespective of morphological heterogeneity. *Hum Pathol* 2007;38:702–709.
 318. Larsen JE, Pavey SJ, Passmore LH, et al. Gene expression signature predicts recurrence in lung adenocarcinoma. *Clin Cancer Res* 2007;13:2946–2954.
 319. Liu H, Kho AT, Kohane IS, et al. Predicting survival within the lung cancer histopathological hierarchy using a multi-scale genomic model of development. *PLoS Med* 2006;3:e232.
 320. Sun Z, Wigle DA, Yang P. Non-overlapping and non-cell-type-specific gene expression signatures predict lung cancer survival. *J Clin Oncol* 2008;26:877–883.
 321. Xi L, Lyons-Weiler J, Coello MC, et al. Prediction of lymph node metastasis by analysis of gene expression profiles in primary lung adenocarcinomas. *Clin Cancer Res* 2005;11:4128–4135.
 322. Potti A, Mukherjee S, Petersen R, et al. A genomic strategy to refine prognosis in early-stage non-small-cell lung cancer. *N Engl J Med* 2006;355:570–580.
 323. Chen HY, Yu SL, Chen CH, et al. A five-gene signature and clinical outcome in non-small-cell lung cancer. *N Engl J Med* 2007;356:11–20.
 324. Chitale D, Gong Y, Taylor BS, et al. An integrated genomic analysis of lung cancer reveals loss of *DUSP4* in *EGFR*-mutant tumors. *Oncogene* 2009;28:2773–2783.
 325. Tontonoz G, Brennan C, Prottopopov A, et al. Common and contrasting genomic profiles among the major human lung cancer subtypes. *Cold Spring Harb Symp Quant Biol* 2005;70:11–24.
 326. Aviel-Ronen S, Coe BP, Lau SK, et al. Genomic markers for malignant progression in pulmonary adenocarcinoma with bronchioloalveolar features. *Proc Natl Acad Sci USA* 2008;105:10155–10160.
 327. Chang JW, Liu HP, Hsieh MH, et al. Increased epidermal growth factor receptor (*EGFR*) gene copy number is strongly associated with *EGFR* mutations and adenocarcinoma in non-small cell lung cancers: a chromogenic in situ hybridization study of 182 patients. *Lung Cancer* 2008;61:328–339.
 328. Cappuzzo F, Hirsch FR, Rossi E, et al. Epidermal growth factor receptor gene and protein and gefitinib sensitivity in non-small-cell lung cancer. *J Natl Cancer Inst* 2005;97:643–655.
 329. Dacic S, Ionescu DN, Finkelstein S, et al. Patterns of allelic loss of synchronous adenocarcinomas of the lung. *Am J Surg Pathol* 2005;29:897–902.
 330. Wang X, Wang M, MacLennan GT, et al. Evidence for common clonal origin of multifocal lung cancers. *J Natl Cancer Inst* 2009;101:560–570.
 331. Girard N, Ostrovskaya I, Lau C, et al. Genomic and mutational profiling to assess clonal relationships between multiple non-small cell lung cancers. *Clin Cancer Res* 2009;15:5184–5190.
 332. van Rens MT, Eijken EJ, Elbers JR, et al. p53 mutation analysis for definite diagnosis of multiple primary lung carcinoma. *Cancer* 2002;94:188–196.
 333. Matsuzoe D, Hideshima T, Ohshima K, et al. Discrimination of double primary lung cancer from intrapulmonary metastasis by p53 gene mutation. *Br J Cancer* 1999;79:1549–1552.
 334. Wang X, Christiani DC, Mark EJ, et al. Carcinogen exposure, p53 alteration, and K-ras mutation in synchronous multiple primary lung carcinoma. *Cancer* 1999;85:1734–1739.
 335. Lau DH, Yang B, Hu R, et al. Clonal origin of multiple lung cancers: K-ras and p53 mutations determined by nonradioisotopic single-strand conformation polymorphism analysis. *Diagn Mol Pathol* 1997;6:179–184.
 336. Girard N, Deshpande C, Azzoli CG, et al. Use of epidermal growth factor

- receptor/Kirsten rat sarcoma 2 viral oncogene homolog mutation testing to define clonal relationships among multiple lung adenocarcinomas: comparison with clinical guidelines. *Chest* 2010;137:46–52.
337. Nonami Y, Ohtuki Y, Sasaguri S. Study of the diagnostic difference between the clinical diagnostic criteria and results of immunohistochemical staining of multiple primary lung cancers. *J Cardiovasc Surg (Torino)* 2003;44:661–665.
 338. Vansteenkiste JF, De Belie B, Deneffe GJ, et al. Practical approach to patients presenting with multiple synchronous suspect lung lesions: a reflection on the current TNM classification based on 54 cases with complete follow-up. *Lung Cancer* 2001;34:169–175.
 339. Yoshino I, Nakanishi R, Osaki T, et al. Postoperative prognosis in patients with non-small cell lung cancer with synchronous ipsilateral intrapulmonary metastasis. *Ann Thorac Surg* 1997;64:809–813.
 340. Chung JH, Choe G, Jheon S, et al. Epidermal growth factor receptor mutation and pathologic-radiologic correlation between multiple lung nodules with ground-glass opacity differentiates multicentric origin from intrapulmonary spread. *J Thorac Oncol* 2009;4:1490–1495.
 341. Balak MN, Gong Y, Riely GJ, et al. Novel D761Y and common secondary T790M mutations in epidermal growth factor receptor-mutant lung adenocarcinomas with acquired resistance to kinase inhibitors. *Clin Cancer Res* 2006;12:6494–6501.
 342. Jackman DM, Holmes AJ, Lindeman N, et al. Response and resistance in a non-small-cell lung cancer patient with an epidermal growth factor receptor mutation and leptomenigeal metastases treated with high-dose gefitinib. *J Clin Oncol* 2006;24:4517–4520.
 343. Schmid K, Oehl N, Wrba F, et al. EGFR/KRAS/BRAF mutations in primary lung adenocarcinomas and corresponding locoregional lymph node metastases. *Clin Cancer Res* 2009;15:4554–4560.
 344. Monaco SE, Nikiforova MN, Ciepły K, et al. A comparison of EGFR and KRAS status in primary lung carcinoma and matched metastases. *Hum Pathol* 2010;41:94–102.
 345. Meert AP, Martin B, Delmotte P, et al. The role of EGF-R expression on patient survival in lung cancer: a systematic review with meta-analysis. *Eur Respir J* 2002;20:975–981.
 346. Berghmans T, Paesmans M, Mascaux C, et al. Thyroid transcription factor 1—a new prognostic factor in lung cancer: a meta-analysis. *Ann Oncol* 2006;17:1673–1676.
 347. Mascaux C, Iannino N, Martin B, et al. The role of RAS oncogene in survival of patients with lung cancer: a systematic review of the literature with meta-analysis. *Br J Cancer* 2005;92:131–139.
 348. Nakamura H, Kawasaki N, Taguchi M, et al. Association of HER-2 overexpression with prognosis in nonsmall cell lung carcinoma: a metaanalysis. *Cancer* 2005;103:1865–1873.
 349. Mitsudomi T, Hamajima N, Ogawa M, et al. Prognostic significance of p53 alterations in patients with non-small cell lung cancer: a meta-analysis. *Clin Cancer Res* 2000;6:4055–4063.
 350. Steels E, Paesmans M, Berghmans T, et al. Role of p53 as a prognostic factor for survival in lung cancer: a systematic review of the literature with a meta-analysis. *Eur Respir J* 2001;18:705–719.
 351. Martin B, Paesmans M, Mascaux C, et al. Ki-67 expression and patients survival in lung cancer: systematic review of the literature with meta-analysis. *Br J Cancer* 2004;91:2018–2025.
 352. Martin B, Paesmans M, Berghmans T, et al. Role of Bcl-2 as a prognostic factor for survival in lung cancer: a systematic review of the literature with meta-analysis. *Br J Cancer* 2003;89:55–64.
 353. Mascaux C, Martin B, Paesmans M, et al. Has Cox-2 a prognostic role in non-small-cell lung cancer? A systematic review of the literature with meta-analysis of the survival results. *Br J Cancer* 2006;95:139–145.
 354. Marks JL, Broderick S, Zhou Q, et al. Prognostic and therapeutic implications of EGFR and KRAS mutations in resected lung adenocarcinoma. *J Thorac Oncol* 2008;3:111–116.
 355. Kosaka T, Yatabe Y, Onozato R, et al. Prognostic implication of EGFR, KRAS, and TP53 gene mutations in a large cohort of Japanese patients with surgically treated lung adenocarcinoma. *J Thorac Oncol* 2009;4:22–29.
 356. Yanaihara N, Caplen N, Bowman E, et al. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 2006;9:189–198.
 357. Raponi M, Dossey L, Jatko T, et al. MicroRNA classifiers for predicting prognosis of squamous cell lung cancer. *Cancer Res* 2009;69:5776–5783.
 358. Hansell DM, Bankier AA, MacMahon H, et al. Fleischner Society: glossary of terms for thoracic imaging. *Radiology* 2008;246:697–722.
 359. Godoy MC, Naidich DP. Subsolid pulmonary nodules and the spectrum of peripheral adenocarcinomas of the lung: recommended interim guidelines for assessment and management. *Radiology* 2009;253:606–622.
 360. Lee HY, Goo JM, Lee HJ, et al. Usefulness of concurrent reading using thin-section and thick-section CT images in subcentimetre solitary pulmonary nodules. *Clin Radiol* 2009;64:127–132.
 361. Takashima S, Sone S, Li F, et al. Small solitary pulmonary nodules (< or =1 cm) detected at population-based CT screening for lung cancer: reliable high-resolution CT features of benign lesions. *AJR Am J Roentgenol* 2003;180:955–964.
 362. Ishikawa H, Koizumi N, Morita T, et al. Ultras-small pulmonary opacities on multidetector-row high-resolution computed tomography: a prospective radiologic-pathologic examination. *J Comput Assist Tomogr* 2005;29:621–625.
 363. Kishi K, Homma S, Kurosaki A, et al. Small lung tumors with the size of 1cm or less in diameter: clinical, radiological, and histopathological characteristics. *Lung Cancer* 2004;44:43–51.
 364. Kim HY, Shim YM, Lee KS, et al. Persistent pulmonary nodular ground-glass opacity at thin-section CT: histopathologic comparisons. *Radiology* 2007;245:267–275.
 365. Kim TJ, Goo JM, Lee KW, et al. Clinical, pathological and thin-section CT features of persistent multiple ground-glass opacity nodules: comparison with solitary ground-glass opacity nodule. *Lung Cancer* 2009;64:171–178.
 366. Ikeda K, Awai K, Mori T, et al. Differential diagnosis of ground-glass opacity nodules: CT number analysis by three-dimensional computerized quantification. *Chest* 2007;132:984–990.
 367. Choi JA, Kim JH, Hong KT, et al. CT bronchus sign in malignant solitary pulmonary lesions: value in the prediction of cell type. *Eur Radiol* 2000;10:1304–1309.
 368. Takashima S, Maruyama Y, Hasegawa M, et al. CT findings and progression of small peripheral lung neoplasms having a replacement growth pattern. *AJR Am J Roentgenol* 2003;180:817–826.
 369. Gould MK, Fletcher J, Iannettoni MD, et al. Evaluation of patients with pulmonary nodules: when is it lung cancer?: ACCP evidence-based clinical practice guidelines (2nd edition). *Chest* 2007;132:108S–130S.
 370. Nakazono T, Sakao Y, Yamaguchi K, et al. Subtypes of peripheral adenocarcinoma of the lung: differentiation by thin-section CT. *Eur Radiol* 2005;15:1563–1568.
 371. Zwirwich CV, Vedal S, Miller RR, et al. Solitary pulmonary nodule: high-resolution CT and radiologic-pathologic correlation. *Radiology* 1991;179:469–476.
 372. Yang ZG, Sone S, Takashima S, et al. High-resolution CT analysis of small peripheral lung adenocarcinomas revealed on screening helical CT. *AJR Am J Roentgenol* 2001;176:1399–1407.
 373. Tateishi U, Uno H, Yonemori K, et al. Prediction of lung adenocarcinoma without vessel invasion: a CT scan volumetric analysis. *Chest* 2005;128:3276–3283.
 374. Kojima Y, Saito H, Sakuma Y, et al. Correlations of thin-section computed tomographic, histopathological, and clinical findings of adenocarcinoma with a bubblelike appearance. *J Comput Assist Tomogr* 2010;34:413–417.
 375. Yoshino I, Nakanishi R, Kodate M, et al. Pleural retraction and intra-tumoral air-bronchogram as prognostic factors for stage I pulmonary adenocarcinoma following complete resection. *Int Surg* 2000;85:105–112.
 376. Kondo T, Yamada K, Noda K, et al. Radiologic-prognostic correlation in patients with small pulmonary adenocarcinomas. *Lung Cancer* 2002;36:49–57.
 377. Sakao Y, Nakazono T, Sakuragi T, et al. Predictive factors for survival in surgically resected clinical IA peripheral adenocarcinoma of the lung. *Ann Thorac Surg* 2004;77:1157–1161.
 378. Kuriyama K, Seto M, Kasugai T, et al. Ground-glass opacity on thin-section CT: value in differentiating subtypes of adenocarcinoma of the lung. *AJR Am J Roentgenol* 1999;173:465–469.
 379. Castro CY, Coffey DM, Medeiros LJ, et al. Prognostic significance of

- percentage of bronchioloalveolar pattern in adenocarcinomas of the lung. *Ann Diagn Pathol* 2001;5:274–284.
380. Hashizume T, Yamada K, Okamoto N, et al. Prognostic significance of thin-section CT scan findings in small-sized lung adenocarcinoma. *Chest* 2008;133:441–447.
 381. Dong B, Sato M, Sagawa M, et al. Computed tomographic image comparison between mediastinal and lung windows provides possible prognostic information in patients with small peripheral lung adenocarcinoma. *J Thorac Cardiovasc Surg* 2002;124:1014–1020.
 382. Matsuguma H, Yokoi K, Anraku M, et al. Proportion of ground-glass opacity on high-resolution computed tomography in clinical T1 N0 M0 adenocarcinoma of the lung: a predictor of lymph node metastasis. *J Thorac Cardiovasc Surg* 2002;124:278–284.
 383. Ohde Y, Nagai K, Yoshida J, et al. The proportion of consolidation to ground-glass opacity on high resolution CT is a good predictor for distinguishing the population of non-invasive peripheral adenocarcinoma. *Lung Cancer* 2003;42:303–310.
 384. Okada M, Nishio W, Sakamoto T, et al. Correlation between computed tomographic findings, bronchioloalveolar carcinoma component, and biologic behavior of small-sized lung adenocarcinomas. *J Thorac Cardiovasc Surg* 2004;127:857–861.
 385. Sakao Y, Nakazono T, Tomimitsu S, et al. Lung adenocarcinoma can be subtyped according to tumor dimension by computed tomography mediastinal-window setting. Additional size criteria for clinical T1 adenocarcinoma. *Eur J Cardiothorac Surg* 2004;26:1211–1215.
 386. Seki N, Sawada S, Nakata M, et al. Lung cancer with localized ground-glass attenuation represents early-stage adenocarcinoma in nonsmokers. *J Thorac Oncol* 2008;3:483–490.
 387. Takashima S, Maruyama Y, Hasegawa M, et al. High-resolution CT features: prognostic significance in peripheral lung adenocarcinoma with bronchioloalveolar carcinoma components. *Respiration* 2003;70:36–42.
 388. Nishio R, Akata S, Saito K, et al. The ratio of the maximum high attenuation area dimension to the maximum tumor dimension may be an index of the presence of lymph node metastasis in lung adenocarcinomas 3 cm or smaller on high-resolution computed tomography. *J Thorac Oncol* 2007;2:29–33.
 389. Shim HS, Park IK, Lee CY, et al. Prognostic significance of visceral pleural invasion in the forthcoming (seventh) edition of TNM classification for lung cancer. *Lung Cancer* 2009;65:161–165.
 390. Ikehara M, Saito H, Kondo T, et al. Comparison of thin-section CT and pathological findings in small solid-density type pulmonary adenocarcinoma: prognostic factors from CT findings. *Eur J Radiol*. In press.
 391. Gaeta M, Vinci S, Minutoli F, et al. CT and MRI findings of mucin-containing tumors and pseudotumors of the thorax: pictorial review. *Eur Radiol* 2002;12:181–189.
 392. Nakata M, Sawada S, Saeki H, et al. Prospective study of thoracoscopic limited resection for ground-glass opacity selected by computed tomography. *Ann Thorac Surg* 2003;75:1601–1605; discussion 5–6.
 393. Takashima S, Maruyama Y, Hasegawa M, et al. Prognostic significance of high-resolution CT findings in small peripheral adenocarcinoma of the lung: a retrospective study on 64 patients. *Lung Cancer* 2002;36:289–295.
 394. Hiramatsu M, Inagaki T, Matsui Y, et al. Pulmonary ground-glass opacity (GGO) lesions—large size and a history of lung cancer are risk factors for growth. *J Thorac Oncol* 2008;3:1245–1250.
 395. Austin JHM, Mujoomdar A, Powell CA, et al. Carcinoma of the lung and metastatic disease of the central nervous system. *Am J Respir Crit Care Med* 2008;178:1090.
 396. Mujoomdar A, Austin JHM, Malhotra R, et al. Clinical predictors of metastatic disease to the brain from non-small cell lung carcinoma: primary tumor size, cell type, and lymph node metastases. *Radiology* 2007;242:882–888.
 397. MacMahon H, Austin JHM, Gamsu G, et al. Guidelines for management of small pulmonary nodules detected on CT scans: a statement from the Fleischner Society. *Radiology* 2005;237:395–400.
 398. Eisenberg RL, Bankier AA, Boiselle PM. Compliance with Fleischner Society guidelines for management of small lung nodules: a survey of 834 radiologists. *Radiology* 2010;255:218–224.
 399. MacMahon H. Compliance with Fleischner Society guidelines for management of lung nodules: lessons and opportunities. *Radiology* 2010;255:14–15.
 400. Zhao B, James LP, Moskowitz CS, et al. Evaluating variability in tumor measurements from same-day repeat CT scans of patients with non-small cell lung cancer. *Radiology* 2009;252:263–272.
 401. Ravenel JG, Leue WM, Nietert PJ, et al. Pulmonary nodule volume: effects of reconstruction parameters on automated measurements—a phantom study. *Radiology* 2008;247:400–408.
 402. Jennings SG, Winer-Muram HT, Tarver RD, et al. Lung tumor growth: assessment with CT—comparison of diameter and cross-sectional area with volume measurements. *Radiology* 2004;231:866–871.
 403. Winer-Muram HT, Jennings SG, Meyer CA, et al. Effect of varying CT section width on volumetric measurement of lung tumors and application of compensatory equations. *Radiology* 2003;229:184–194.
 404. Yankelevitz DF, Reeves AP, Kostis WJ, et al. Small pulmonary nodules: volumetrically determined growth rates based on CT evaluation. *Radiology* 2000;217:251–256.
 405. de Hoop B, Gietema H, van de Vorst S, et al. Pulmonary ground-glass nodules: increase in mass as an early indicator of growth. *Radiology* 2010;255:199–206.
 406. Nakata M, Sawada S, Yamashita M, et al. Surgical treatments for multiple primary adenocarcinoma of the lung. *Ann Thorac Surg* 2004;78:1194–1199.
 407. Zwirewich CV, Miller RR, Muller NL. Multicentric adenocarcinoma of the lung: CT-pathologic correlation. *Radiology* 1990;176:185–190.
 408. Park CM, Goo JM, Kim TJ, et al. Pulmonary nodular ground-glass opacities in patients with extrapulmonary cancers: what is their clinical significance and how can we determine whether they are malignant or benign lesions? *Chest* 2008;133:1402–1409.
 409. Okada M, Tauchi S, Iwanaga K, et al. Associations among bronchioloalveolar carcinoma components, positron emission tomographic and computed tomographic findings, and malignant behavior in small lung adenocarcinomas. *J Thorac Cardiovasc Surg* 2007;133:1448–1454.
 410. Higashi K, Ueda Y, Seki H, et al. Fluorine-18-FDG PET imaging is negative in bronchioloalveolar lung carcinoma. *J Nucl Med* 1998;39:1016–1020.
 411. Higashi K, Ueda Y, Yagishita M, et al. FDG PET measurement of the proliferative potential of non-small cell lung cancer. *J Nucl Med* 2000;41:85–92.
 412. Higashi K, Ueda Y, Ayabe K, et al. FDG PET in the evaluation of the aggressiveness of pulmonary adenocarcinoma: correlation with histopathological features. *Nucl Med Commun* 2000;21:707–714.
 413. Ohtsuka T, Nomori H, Watanabe K, et al. Prognostic significance of [(18)F]fluorodeoxyglucose uptake on positron emission tomography in patients with pathologic stage I lung adenocarcinoma. *Cancer* 2006;107:2468–2473.
 414. Raz DJ, Odisho AY, Franc BL, et al. Tumor fluoro-2-deoxy-D-glucose avidity on positron emission tomographic scan predicts mortality in patients with early-stage pure and mixed bronchioloalveolar carcinoma. *J Thorac Cardiovasc Surg* 2006;132:1189–1195.
 415. Sagawa M, Higashi K, Sugita M, et al. Fluorodeoxyglucose uptake correlates with the growth pattern of small peripheral pulmonary adenocarcinoma. *Surg Today* 2006;36:230–234.
 416. Pastorino U, Landoni C, Marchiano A, et al. Fluorodeoxyglucose uptake measured by positron emission tomography and standardized uptake value predicts long-term survival of CT screening detected lung cancer in heavy smokers. *J Thorac Oncol* 2009;4:1352–1356.
 417. Nakayama H, Okumura S, Daisaki H, et al. Value of integrated positron emission tomography revised using a phantom study to evaluate malignancy grade of lung adenocarcinoma: a multicenter study. *Cancer* 2010;116:3170–3177.
 418. Um SW, Kim H, Koh WJ, et al. Prognostic value of 18F-FDG uptake on positron emission tomography in patients with pathologic stage I non-small cell lung cancer. *J Thorac Oncol* 2009;4:1331–1336.
 419. Berghmans T, Dusart M, Paesmans M, et al. Primary tumor standardized uptake value (SUVmax) measured on fluorodeoxyglucose positron emission tomography (FDG-PET) is of prognostic value for survival in non-small cell lung cancer (NSCLC): a systematic review and meta-analysis (MA) by the European Lung Cancer Working Party for the IASLC Lung Cancer Staging Project. *J Thorac Oncol* 2008;3:6–12.
 420. Birchard KR, Hoang JK, Herndon JE Jr, et al. Early changes in tumor size in patients treated for advanced stage nonsmall cell lung cancer do not correlate with survival. *Cancer* 2009;115:581–586.
 421. Sohn HJ, Yang YJ, Ryu JS, et al. [(18)F]Fluorothymidine positron emission tomography before and 7 days after gefitinib treatment pre-

- dicts response in patients with advanced adenocarcinoma of the lung. *Clin Cancer Res* 2008;14:7423–7429.
422. Cloran FJ, Banks KP, Song WS, et al. Limitations of dual time point PET in the assessment of lung nodules with low FDG avidity. *Lung Cancer* 2010;68:66–71.
 423. Ohno Y, Hatabu H, Takenaka D, et al. Dynamic MR imaging: value of differentiating subtypes of peripheral small adenocarcinoma of the lung. *Eur J Radiol* 2004;52:144–150.
 424. van Klaveren RJ, Oudkerk M, Prokop M, et al. Management of lung nodules detected by volume CT scanning. *N Engl J Med* 2009;361:2221–2229.
 425. Oda S, Awai K, Muraio K, et al. Computer-aided volumetry of pulmonary nodules exhibiting ground-glass opacity at MDCT. *AJR Am J Roentgenol* 2010;194:398–406.
 426. Henschke CI, McCauley DI, Yankelevitz DF, et al. Early Lung Cancer Action Project: overall design and findings from baseline screening. *Lancet* 1999;354:99–105.
 427. Henschke CI, Naidich DP, Yankelevitz DF, et al. Early lung cancer action project: initial findings on repeat screenings. *Cancer* 2001;92:153–159.
 428. Lindell RM, Hartman TE, Swensen SJ, et al. Five-year lung cancer screening experience: CT appearance, growth rate, location, and histologic features of 61 lung cancers. *Radiology* 2007;242:555–562.
 429. Hasegawa M, Sone S, Takashima S, et al. Growth rate of small lung cancers detected on mass CT screening. *Br J Radiol* 2000;73:1252–1259.
 430. Henschke CI, Yankelevitz DF, Libby DM, et al. Survival of patients with stage I lung cancer detected on CT screening. *N Engl J Med* 2006;355:1763–1771.
 431. Kakinuma R, Ohmatsu H, Kaneko M, et al. Progression of focal pure ground-glass opacity detected by low-dose helical computed tomography screening for lung cancer. *J Comput Assist Tomogr* 2004;28:17–23.
 432. Sone S, Nakayama T, Honda T, et al. Long-term follow-up study of a population-based 1996–1998 mass screening programme for lung cancer using mobile low-dose spiral computed tomography. *Lung Cancer* 2007;58:329–341.
 433. Pelosi G, Sonzogni A, Veronesi G, et al. Pathologic and molecular features of screening low-dose computed tomography (LDCT)-detected lung cancer: a baseline and 2-year repeat study. *Lung Cancer* 2008;62:202–214.
 434. Wang JC, Sone S, Feng L, et al. Rapidly growing small peripheral lung cancers detected by screening CT: correlation between radiological appearance and pathological features. *Br J Radiol* 2000;73:930–937.
 435. Infante M, Cavuto S, Lutman FR, et al. A randomized study of lung cancer screening with spiral computed tomography: three-year results from the DANTE trial. *Am J Respir Crit Care Med* 2009;180:445–453.
 436. Bepler G. Are we coming full circle for lung cancer screening a second time? *Am J Respir Crit Care Med* 2009;180:384–385.
 437. McMahon PM, Kong CY, Johnson BE, et al. Estimating long-term effectiveness of lung cancer screening in the Mayo CT screening study. *Radiology* 2008;248:278–287.
 438. McMahon PM, Kong CY, Weinstein MC, et al. Adopting helical CT screening for lung cancer: potential health consequences during a 15-year period. *Cancer* 2008;113:3440–3449.
 439. Gatsonis CA. The National Lung Screening Trial: overview and study design. *Radiology*. 2011;258:243–253.
 440. Park EA, Lee HJ, Kim YT, et al. *EGFR* gene copy number in adenocarcinoma of the lung by FISH analysis: investigation of significantly related factors on CT, FDG-PET, and histopathology. *Lung Cancer* 2009;64:179–186.
 441. Yano M, Sasaki H, Kobayashi Y, et al. *Epidermal growth factor receptor* gene mutation and computed tomographic findings in peripheral pulmonary adenocarcinoma. *J Thorac Oncol* 2006;1:413–416.
 442. Chantranuwat C, Sriuranpong V, Huapai N, et al. Histopathologic characteristics of pulmonary adenocarcinomas with and without *EGFR* mutation. *J Med Assoc Thai* 2005;88(Suppl 4):S322–S329.
 443. Huang CT, Yen RF, Cheng MF, et al. Correlation of F-18 fluorodeoxyglucose-positron emission tomography maximal standardized uptake value and *EGFR* mutations in advanced lung adenocarcinoma. *Med Oncol* 2010;27:9–15.
 444. Watanabe K, Nomori H, Ohtsuka T, et al. [F-18]Fluorodeoxyglucose positron emission tomography can predict pathological tumor stage and proliferative activity determined by Ki-67 in clinical stage IA lung adenocarcinomas. *Jpn J Clin Oncol* 2006;36:403–409.
 445. Vesselle H, Salskov A, Turcotte E, et al. Relationship between non-small cell lung cancer FDG uptake at PET, tumor histology, and Ki-67 proliferation index. *J Thorac Oncol* 2008;3:971–978.
 446. Schuchert MJ, Pettiford BL, Keeley S, et al. Anatomic segmentectomy in the treatment of stage I non-small cell lung cancer. *Ann Thorac Surg* 2007;84:926–932.
 447. Shapiro M, Weiser TS, Wisnivesky JP, et al. Thoracoscopic segmentectomy compares favorably with thoracoscopic lobectomy for patients with small stage I lung cancer. *J Thorac Cardiovasc Surg* 2009;137:1388–1393.
 448. Yan TD, Black D, Bannon PG, et al. Systematic review and meta-analysis of randomized and nonrandomized trials on safety and efficacy of video-assisted thoracic surgery lobectomy for early-stage non-small-cell lung cancer. *J Clin Oncol* 2009;27:2553–2562.
 449. Watanabe T, Okada A, Imakiire T, et al. Intentional limited resection for small peripheral lung cancer based on intraoperative pathologic exploration. *Jpn J Thorac Cardiovasc Surg* 2005;53:29–35.
 450. Higashiyama M, Kodama K, Takami K, et al. Intraoperative lavage cytologic analysis of surgical margins in patients undergoing limited surgery for lung cancer. *J Thorac Cardiovasc Surg* 2003;125:101–107.
 451. Utsumi T, Sawabata N, Inoue M, et al. Optimal sampling methods for margin cytology examination following lung excision. *Interact Cardiovasc Thorac Surg* 2010;10:434–436.
 452. Asamura H, Suzuki K, Watanabe S, et al. A clinicopathological study of resected subcentimeter lung cancers: a favorable prognosis for ground glass opacity lesions. *Ann Thorac Surg* 2003;76:1016–1022.
 453. Ginsberg RJ, Rubinstein LV. Randomized trial of lobectomy versus limited resection for T1 N0 non-small cell lung cancer. Lung Cancer Study Group. *Ann Thorac Surg* 1995;60:615–622.
 454. Miller DL, Rowland CM, Deschamps C, et al. Surgical treatment of non-small cell lung cancer 1 cm or less in diameter. *Ann Thorac Surg* 2002;73:1545–1550; discussion 50–51.
 455. Rami-Porta R, Wittekind C, Goldstraw P. Complete resection in lung cancer surgery: proposed definition. *Lung Cancer* 2005;49:25–33.
 456. Ishiguro F, Matsuo K, Fukui T, et al. Effect of selective lymph node dissection based on patterns of lobe-specific lymph node metastases on patient outcome in patients with resectable non-small cell lung cancer: a large-scale retrospective cohort study applying a propensity score. *J Thorac Cardiovasc Surg* 2010;139:1001–1006.
 457. Darling GE, Allen MS, Landreneau RJ, et al. Randomized trial of mediastinal lymph node sampling versus complete lymphadenectomy during pulmonary resection in the patient with N0 or N1 (less than hilar) non-small cell carcinoma: results of the ACOSOG Z0030 Trial. *J Thorac Cardiovasc Surg*. In press.
 458. Nomori H, Iwatani K, Kobayashi H, et al. Omission of mediastinal lymph node dissection in lung cancer: its techniques and diagnostic procedures. *Ann Thorac Cardiovasc Surg* 2006;12:83–88.
 459. Finley DJ, Yoshizawa A, Travis W, et al. Predictors of outcomes after surgical treatment of synchronous primary lung cancers. *J Thorac Oncol* 2010;5:197–205.
 460. Hayes DF, Allred C, Anderson BO, et al. Breast. In: SB Edge, DR Byrd, CC Compton, et al. (Eds.). *AJCC Cancer Staging Manual*, 7th Ed. New York: Springer, 2009. Pp. 347–376.



Clinicopathological findings of non-small-cell lung cancer with high serum progastrin-releasing peptide concentrations

Keita Kudo^a, Fumiyoshi Ohyanagi^a, Atushi Horiike^a, Eisaku Miyauchi^a, Noriko Yanagitani^a, Rira Hoshi^c, Yukitoshi Satoh^d, Noriko Motoi^b, Wakako Hamanaka^b, Yuichi Ishikawa^b, Mingyon Mun^a, Yukinori Sakao^a, Sakae Okumura^a, Ken Nakagawa^a, Takeshi Horai^a, Makoto Nishio^{a,*}

^a Thoracic Oncology Center, Cancer Institute Hospital, Japanese Foundation for Cancer Research, 3-8-31 Ariake, Kouto-ku, Tokyo 135-8550, Japan

^b Department of Pathology, Cancer Institute, Japanese Foundation for Cancer Research, 3-8-31 Ariake, Kouto-ku, Tokyo, Japan

^c Department of Cytology, Cancer Institute, Japanese Foundation for Cancer Research, 3-8-31 Ariake, Kouto-ku, Tokyo, Japan

^d Department of Thoracic Surgery, Kitasato University School of Medicine, 1-15-1 Kitasato, Sagamiharashi, Kanagawa, Japan

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ABSTRACT

Although progastrin-releasing peptide (proGRP) is used as a serum tumor marker for small cell lung cancer (SCLC), high serum pro-GRP concentrations are observed in some non-small-cell lung cancers (NSCLCs). The characteristics of these NSCLCs are not well known. To determine the clinicopathological features of NSCLC in patients with elevated serum proGRP concentrations, serum proGRP values were assessed in 654 advanced lung cancer patients, and positive (>46 pg/mL) NSCLC specimens were subjected to cytological and histopathological reevaluation. Serum proGRP concentrations were positive in 34 of 421 NSCLC patients (8.1%) and 186 of 233 SCLC patients (80%). Histological subtypes of the 34 NSCLC patients at diagnosis were 20 adenocarcinomas, 5 squamous cell carcinomas, 4 large cell carcinomas, and 5 large cell neuroendocrine carcinomas. Six of 27 cytology specimens contained characteristic neuroendocrine morphology. Immunohistochemical analysis showed that 11 of 17 tumors were positive for neuroendocrine markers (64.7%). Twenty of 34 serum proGRP-positive NSCLC patients received platinum-based chemotherapy, and the response rate was 55.0%. These results suggest that serum proGRP-positive NSCLCs may have neuroendocrine differentiation. In addition, serum proGRP-positive NSCLCs may have clinical characteristics that are different from other NSCLCs.

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1. Introduction

Lung cancer is the leading cause of cancer death worldwide. In 2005, the number of deaths due to lung cancer in Japan exceeded 60,000 [1]. Conventionally, lung cancer is classified into small cell lung cancer (SCLC) and non-small-cell carcinoma (NSCLC). Because SCLC has neuroendocrine features, it has a poorer prognosis and shows greater sensitivity to chemotherapy than NSCLC. Although NSCLC is subclassified into adenocarcinoma, squamous cell carcinoma, and large cell carcinoma, some NSCLCs have neuroendocrine differentiation. In 1999, the World Health Organization categorized large cell neuroendocrine carcinoma (LCNEC) as a variant of large cell carcinoma [2]. LCNEC has been reported to have a poor prognosis, even for early-stage disease [3,4]. Different types of NSCLCs differ in their clinical behavior according to the presence or absence of neuroendocrine differentiation. Neuroendocrine differentiation in a tumor is generally determined by

immunohistochemistry and/or electron microscopy, which reveal the characteristic neuroendocrine morphology [2,5]. However, it is difficult to obtain sufficient tissue by biopsy, and limited tumor tissue sampling may make it difficult to diagnose neuroendocrine differentiation in NSCLC. Therefore, the development of a sensitive serum marker for the detection of neuroendocrine differentiation is greatly desired to facilitate the diagnosis of NSCLCs and neuroendocrine tumors.

Progastrin-releasing peptide (proGRP) is a signal peptide that is produced by small cell lung cancer cells (SCLC). Serum proGRP is considered to be a sensitive tumor marker for SCLC. The sensitivity and specificity of serum proGRP as a tumor marker for SCLC is 60–70% and 96%, respectively [6]. Elevated serum proGRP concentrations have been observed in some NSCLC patients, especially LCNEC patients [6,7], suggesting that serum proGRP is a potentially good marker not only for SCLC but also for NSCLC with neuroendocrine features. However, the clinical and pathological characteristics of NSCLCs with elevated serum proGRP concentrations have not been well studied. In the present study, serum proGRP levels were measured in 654 lung cancer patients and the clinical characteristics of serum proGRP-positive NSCLC patients

* Corresponding author. Tel.: +81 03 3520 0111; fax: +81 03 3570 0343.
E-mail address: mnishio@jfcrc.or.jp (M. Nishio).

were analyzed; the histopathology of the surgical or biopsy specimens of the positive patients were also evaluated.

2. Patients and methods

Serum proGRP concentrations were measured in 654 patients who were diagnosed with lung cancer by histology or cytology at the Cancer Institute Hospital of the Japanese Foundation for Cancer Research between April 1998 and April 2006.

An enzyme-linked immunosorbent assay (ELISA) kit (serumlabo ProGRP; Fujirebio Diagnostics Inc., Tokyo, Japan) was used to determine serum proGRP concentrations, and samples were considered positive when their values exceeded 46 pg/mL [8].

The clinical characteristics of serum proGRP-positive NSCLC patients were retrospectively analyzed, including age at diagnosis, gender, smoking history, and TNM stage. Response to platinum-based chemotherapy in serum proGRP-positive NSCLC patients was determined according to RECIST criteria (without confirmation).

In addition, the cytological and histological findings of the surgical or biopsy specimens of these patients were reevaluated. Immunohistochemical (IHC) staining was used to evaluate neuroendocrine differentiation in the tumors. Formalin-fixed paraffin-embedded sections were stained for a panel of epithelial markers, including thyroid transcription factor-1 (TTF-1; Dako EnVision+, Saitama, Japan) and carcinoembryonic antigen (CEA; Nichirei, Tokyo, Japan), and neuroendocrine markers, including chromogranin A (CGA) (Dako EnVision+, Saitama, Japan), synaptophysin (Dako EnVision+, Saitama, Japan), CD56 (neural cell adhesion molecule [NCAM]) (Clone 1B6; Novocastra, and proGRP (Advanced Life Science Institute Inc., Saitama, Japan). IHC staining was performed according to standard protocols with EnVision kits (Dako EnVision+, Saitama, Japan). IHC results were grouped into 3 categories – strongly positive, weakly positive, or negative – by well-trained pathologists (WH and NM).

Statistical calculations were performed using StatView version 5.0 for Windows XP (SAS Institute, Cary, NC). Associations between categorical variables and serum proGRP concentrations were evaluated using Student's *t* test. Survival was measured from the start of chemotherapy to the last follow-up evaluation or death, and survival rates were estimated using the Kaplan–Meier method.

3. Results

3.1. Patient characteristics

Of a total of 654 patients, 421 were diagnosed with NSCLC and 233 with SCLC. Serum proGRP samples were positive in 220 of 654 patients, of which 34 (8.1%) had NSCLC and 186 (80%) had SCLC.

The clinical characteristics of serum proGRP-positive and negative NSCLC patients are shown in Table 1. There were no significant differences in the clinical characteristics between the serum proGRP-positive and -negative NSCLC patients.

In serum ProGRP-positive NSCLC patients, the median age of these patients was 67 years (range, 49–77). There were 22 males and 12 females, and 65% of the patients were heavy smokers (smoking index > 400). Most of the patients (94%) had advanced NSCLC. Serum creatinine concentrations were less than 1.6 mg/dL in all 34 serum proGRP-positive NSCLC patients.

The histological subtypes of the 34 serum proGRP-positive NSCLCs at diagnosis were as follows: 20 adenocarcinomas, 5 squamous cell carcinomas, 4 large cell carcinomas, and 5 LCNECs. The rates of positive serum proGRP in each histological subtype were as follows: 7.7% in 260 adenocarcinomas, 5.9% in 85 squamous

Table 1
Clinical characteristics of NSCLC patients.

Characteristics	ProGRP-positive NSCLC patients	ProGRP-negative NSCLC patients
Total no. of patients	34	387
Age, years		
Median (range)	67 (49–77)	62 (29–87)
Sex		
Male/female	22/12	261/126
Smoking index		
Mean (range)	807.5 (0–1400)	661 (0–3000)
Never/≤400/>400	10/2/22	121/29/237
Histological subtype at diagnosis		
Adenocarcinoma	20	240
Squamous cell carcinoma	5	80
Large cell carcinoma	4	48
LCNEC	5	6
Adenosquamous	0	2
Other	0	11
Stage		
I/II	2	56
III A	7	63
III B	6	106
IV	16	144
Recurrent	3	18

LCNEC: large cell neuroendocrine carcinoma, ProGRP: progastrin-releasing peptide, NSCLC: non-small-cell lung cancer.

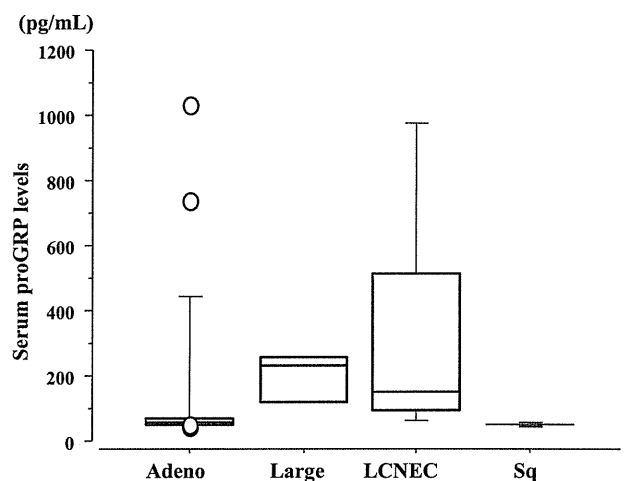


Fig. 1. Serum progastrin-releasing peptide (proGRP) concentrations of 34 non-small-cell lung cancer patients with elevated proGRP. Adeno: adenocarcinoma, Large: large cell carcinoma, LCNEC: large cell neuroendocrine carcinoma, Sq: squamous cell carcinoma.

cell carcinomas, 9.3% in 43 large cell carcinomas, and 44.4% in 11 LCNECs.

The median serum proGRP concentration of the 34 NSCLC patients was 60.7 pg/mL and the range was 46.0–973.0 pg/mL. The serum proGRP concentrations in these 34 NSCLC patients were significantly lower than the serum concentrations in proGRP-positive SCLC patients (median, 469 pg/mL; range, 47.1–344,000 pg/mL) ($P < 0.05$).

Fig. 1 shows the serum proGRP concentrations for each histological subtype of serum proGRP-positive NSCLC. The mean serum proGRP concentration in LCNECs was 147 pg/mL and the range was 78.6–973 pg/mL. These concentrations are relatively high compared to other NSCLCs. On the other hand, serum proGRP concentrations were relatively low, even in serum proGRP-positive squamous cell carcinoma patients (median, 47.4 pg/mL; range, 46–56.7 pg/mL).

Table 2
Immunohistochemistry results of non-small-cell lung cancer with elevated serum progastrin-releasing peptide concentrations.

	Serum proGRP (pg/dL)	Histological subtype at diagnosis	Specimen	Stage	IHC					
					TTF-1	CEA	CGA	Synapto-phisin	NCAM	proGRP
1	973	Large	TBLB	IV	–	++	++	++	+	–
2	363	LCNEC	TBLB	IV	++	–	++	++	++	++
3	269	LCNEC	TBLB	IV	++	++	++	++	++	+
4	229	Large	TBLB	IV	–	–	–	–	–	–
5	78.6	Large	TBLB	IV	–	–	–	–	–	–
6	60.3	Adeno	TBLB	IV	++	–	–	–	–	–
7	56.7	Sq	Surgery	Rec	–	+	–	+	++	–
8	56.0	Adeno	Surgery	Rec	++	–	–	+	–	–
9	55.3	Adeno	TBLB	IIIB	++	–	–	+	+	–
10	54.7	Adeno	TBLB	IV	++	–	–	+	–	–
11	53.1	Adeno	Surgery	IB	++	–	–	+	–	–
12	51.8	Adeno	TBLB	IV	++	+	++	++	++	+
13	51.4	Adeno	Surgery	IB	++	++	++	++	++	++
14	48.3	Sq	TBLB	IB	–	++	–	–	–	–
15	47.4	Sq	Surgery	IIIA	–	++	–	–	–	–
16	47.3	Sq	TBLB	IIIA	–	–	–	+	–	–
17	46	Sq	TBLB	IIIB	–	++	–	–	–	–

IHC: immunohistochemistry, proGRP: progastrin-releasing peptide, TTF-1: thyroid transcription factor 1, CEA: carcinoembryonic antigen, CGA: chromogranin A, NCAM: neural cell adhesion molecule, Adeno: adenocarcinoma, Sq: squamous cell carcinoma, Large: large cell carcinoma, LCNEC: large cell neuroendocrine carcinoma, TBLB: transbronchial lung biopsy, –: negative, +: weakly positive, ++: strongly positive.

3.2. Cytological and histological examination

Cytological specimens corresponding to 27 of the 34 serum proGRP-positive NSCLCs were reevaluated. All 5 cytology specimens diagnosed as LCNEC contained typical neuroendocrine features such as rosette-like and palisading patterns. A rosette-like formation was found in only 1 cytology specimen diagnosed as squamous cell carcinoma, and the other 21 specimens did not contain the typical cytological features of neuroendocrine differentiation.

IHC staining was performed on 17 histological specimens of the 34 serum proGRP-positive NSCLCs (7 adenocarcinomas, 5 squamous cell carcinomas, 3 large cell carcinomas, and 2 LCNECs) to examine neuroendocrine differentiation (Table 2). Four of 17 specimens (24%) showed positive staining (2 weakly and 2 strongly positive) for proGRP, and some neuroendocrine markers were positive in 11 of 17 specimens (64.7%). In particular, 2 of 7 adenocarcinomas, 1 of 3 large cell carcinomas, and 2 of 2 LCNECs showed strongly positive staining for at least 2 out of the 3 neuroendocrine markers CGA, synaptophysin, and NCAM. None of the squamous cell carcinomas showed strongly positive staining for at least 2 out of the 3 neuroendocrine markers CGA, synaptophysin, and NCAM. One of 5 squamous cell carcinomas showed strongly positive staining for NCAM. One of 5 squamous cell carcinomas showed strongly positive staining for NCAM. There was no significant relationship between serum proGRP concentrations and proGRP immunoreactivity.

3.3. Response to chemotherapy

Twenty of 34 serum proGRP-positive NSCLC patients received platinum-based chemotherapy (Table 3). There were 11 partial responses, 4 stable diseases, and no responses observed in the 5 patients. The objective response rate was 55.0%. The median survival of the 20 patients was 11 months, and the 1-year survival rate was 48%.

On the other hand, 232 of 387 serum proGRP-negative NSCLC patients received platinum-based chemotherapy. There were 82 partial responses, 97 stable diseases, and no complete responses observed in the 53 patients. The objective response rate was 35.0%, the median survival was 11.5 months, and the 1-year survival rate was 49.1%.

Table 3

Platinum doublet regimens administered to serum proGRP-positive and -negative patients.

Regimens	ProGRP-positive NSCLC patients (n=20)		ProGRP-negative NSCLC patients (n=232)	
	n	%	n	%
CBDCA/PTX	9	45	141	60.7
CBDCA/GEM	0	0	16	7
CBDCA/VP16	1	5	5	2
CDDP/DOC	3	15	53	23
CDDP/S-1	2	10	4	1.8
CDDP/VNR	2	10	1	0.5
CDDP/VP16	1	5	0	0
CDDP/CPT11	2	10	2	1
CDDP/GEM	0	0	10	4

CBDCA: carboplatin, CDDP: cisplatin, PTX: paclitaxel, GEM: gemcitabine, VP16: etoposide, DOC: docetaxel, VNR: vinorelbine, CPT11: irinotecan, ProGRP: progastrin-releasing peptide.

4. Discussion

In the present study, the positive rate of serum proGRP concentration in NSCLC patients was 8.1% (34/421), and histological neuroendocrine features were detected in 11 of 17 (64.7%) serum proGRP-positive NSCLC specimens. Several studies have examined serum proGRP concentrations in lung cancer, and all of these studies reported that serum proGRP was a specific tumor marker for SCLC [6,9]. The sensitivity and specificity for SCLCs were around 70% and 99%, respectively, and serum proGRP was superior to NSE [10].

Although increases in serum ProGRP concentration have been observed in some NSCLC patients in previous studies, the reported positive rates of serum proGRP in NSCLCs demonstrated a wide range of variability (3–30%) [6,11]. The 8% positive rate found in the present study was relatively higher than in previous studies, with the exception of 2 reports (the Takada study used a lower cutoff for positive, at 34 pg/mL, and the Molina study included a higher proportion of renal failure patients) [11,12]. Although several studies have reported that serum proGRP values were elevated in NSCLC patients, there have been few studies examining the clinicopathological characteristics of serum proGRP-positive NSCLC patients [13]. Only 1 study examined the clinicopathological