

**Fig. 2** A representative example of breath-by-breath time courses of  $P_{ETCO_2}$ ,  $\dot{V}_E$ ,  $V_T$  and RR at rest and during exercise in an untrained subject during open-loop respiratory chemoreflex control of  $\dot{V}_E$  and  $P_{ETCO_2}$  in hypercapnia test (a) and hyper/hypoventilation test (b).

Accompanying increases in  $F_{ICO_2}$  from 0 to 0.06,  $P_{ETCO_2}$ ,  $\dot{V}_E$ ,  $V_T$  and RR increase both at rest and during exercise.  $P_{ETCO_2}$  and  $\dot{V}_E$  at rest and during exercise reach steady-states in 8–10 min

alcohol or caffeine for 3 h preceding the tests. The subjects remained in a fasting state throughout the two tests that lasted approximately 4 h. Before testing, an intravenous catheter (0.47 mm ID, 24 gauge) for blood collection was placed in the forearm of the subject seated in a comfortable chair. Venous blood samples (2.5 ml) were collected at 11 min of each 12-min trial described below.

**Hypercapnia test** Each hypercapnia test consisted of four trials each under resting and exercising conditions

(Figs. 2a, 3a). Hypercapnia was induced by changing the level of inspired  $CO_2$  concentrations ( $F_{ICO_2} = 0.0, 0.035, 0.05$  or  $0.06$ ,  $F_{IO_2} = 0.80$  with  $N_2$  balance). The subject breathed each gas mixture for 12 min and then breathed room air during the interposing interval of approximately 10–15 min (Miyamoto et al. 2004). The 12-min duration was long enough to permit carbon dioxide tension to reach its new steady-state value at the central chemoreceptors (Lloyd and Cunningham 1963; Honda et al. 1983; Poon and Greene 1985; Cummin and Saunders 1987). The order

of the hypercapnia trials was randomized in each subject. All trials were performed under hyperoxic condition to suppress O<sub>2</sub>-sensitive chemoreflexes (Lloyd and Cunningham 1963; Ohyabu et al. 1982; Robbins 1988; Mohan and Duffin 1997).

**Hyper/hypoventilation test** Each hyper/hypoventilation test consisted of four trials each under resting and exercising conditions (Figs. 2b, 3b). Subjects were instructed to breathe at different respiratory rates and tidal volumes to match a visual display of ventilation curves on a screen monitor (Miyamoto et al. 2004). For three levels of hyperventilation, the subjects breathed according to ventilation curves at respiratory rates and tidal volumes mimicking those recorded during the hypercapnia trials ( $F_{\text{ICO}_2} = 0.035, 0.05$  or  $0.06$ ). For one level of hypoventilation, the subjects were asked to breathe following a breathing curve at 92% respiratory rate and 88% tidal volume measured when  $F_{\text{ICO}_2}$  was 0.0%. Each trial lasted 12 min with interposing intervals. All trials were started after  $\dot{V}_E$  and  $P_{\text{ETCO}_2}$  values recovered to the resting levels. All trials were performed under hyperoxic condition ( $F_{\text{ICO}_2} = 0.80, F_{\text{IO}_2} = 0.0$  with N<sub>2</sub> balance).

#### Data analysis

Because preliminary measurements indicated that both  $\dot{V}_E$  and  $P_{\text{ETCO}_2}$  reached steady states in the last 2 min of each trial, the steady-state  $\dot{V}_E$  and  $P_{\text{ETCO}_2}$  were obtained by averaging the respective data for the last 2 min. To characterize the controller property, we performed linear regression of  $\dot{V}_E$  against  $P_{\text{ETCO}_2}$  [ $\dot{V}_E = S(P_{\text{ETCO}_2} - B)$ ]; where  $S$  is the slope, and  $B$  is the  $P_{\text{ETCO}_2}$ -intercept (Lloyd and Cunningham 1963; Cummin and Saunders 1987). To characterize the plant property, we fitted a metabolic hyperbola ( $P_{\text{ETCO}_2} = A/\dot{V}_E + C$ ) modified from the original metabolic hyperbola (Cunningham et al. 1986; Whipp and Pardy 1986) to the measured data (see Appendix). Hereafter in this paper, we refer to our metabolic hyperbola as the modified metabolic hyperbola. The measured operating point for each subject was defined to be the steady-state values of  $\dot{V}_E$  and  $P_{\text{ETCO}_2}$  obtained during the  $F_{\text{ICO}_2} = 0.00$  trial without visual feedback (i.e., during spontaneous breathing).

#### Statistical analysis

All values are presented as mean (SD). Statistical significance was accepted at  $P < 0.05$ . Two way analysis of variance (ANOVA) was performed with exercise stimulus as one factor (i.e., the difference between rest and exercise conditions) and exercise training as the other factor (i.e.,

the difference between untrained and trained subjects). When the interaction effect was statistically significant ( $P < 0.05$ ), post-hoc analysis using Tukey test was performed for pair-wise comparisons (Glantz and Slinker 2001).

#### Results

The trained group had higher maximal oxygen uptake ( $\dot{V}_{\text{O}_{2\text{max}}}$ ) [3.6 (0.3) vs. 2.7 (0.1) l/min,  $P < 0.01$ ], higher  $\dot{V}_{\text{O}_{2\text{max}}}$  per kg body weight [59.1(6.7) vs. 44.6 (6.9) ml/min/kg,  $P < 0.01$ ] and higher ventilatory threshold [2.5 (0.5) vs. 1.4 (0.3) l/min,  $P < 0.01$ ] compared to the untrained group. The trained group had the slightly higher value of  $P_{\text{ETCO}_2}$  at  $\dot{V}_{\text{O}_{2\text{max}}}$  [39.3 (2.4) vs. 36.3 (4.8) mmHg], but was not significantly different between two groups. The maximal minute ventilation ( $\dot{V}_{\text{Emax}}$ ) was also higher in the trained than in the untrained group [148.9 (12.9) vs. 121.5 (22.0) l/min,  $P < 0.01$ ]. The higher  $\dot{V}_E$  was due to larger  $V_T$ , because RR was not significantly different between two groups.

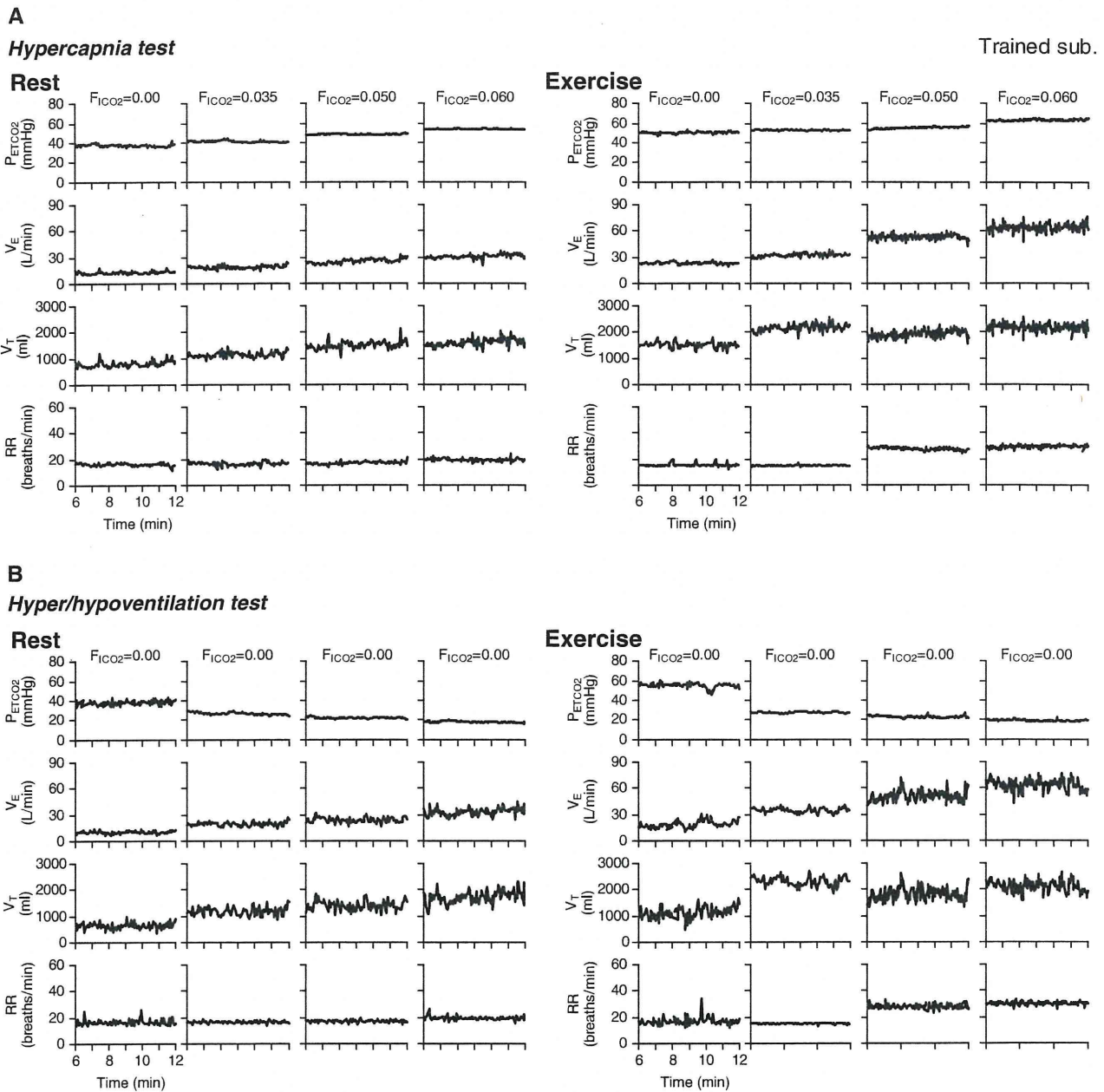
Table 1 summarizes the gas-exchange variables at rest and during exercise under spontaneous breathing ( $F_{\text{ICO}_2} = 0.0$ ). Exercise stimulus significantly increased all the gas-exchange variables in both untrained and trained groups. The significant interaction effect observed for  $\dot{V}_E$  indicated that the exercise-induced  $\dot{V}_E$  increase was smaller in the trained than in the untrained group. Although the exercise stimulus significantly increased plasma potassium level, it did not significantly affect lactate level. There were no significant differences in plasma potassium and lactate levels between the untrained and trained groups.

Panel A of Figs. 2 and 3 show representative examples of breath-by-breath time courses of  $P_{\text{ETCO}_2}$ ,  $\dot{V}_E$ ,  $V_T$  and RR under various  $F_{\text{ICO}_2}$  at rest and during exercise in an untrained and a trained subject, respectively. When  $F_{\text{ICO}_2}$  increased from 0 to 0.06,  $P_{\text{ETCO}_2}$ ,  $\dot{V}_E$ ,  $V_T$  and RR increased in both trained and untrained subjects. Both  $P_{\text{ETCO}_2}$  and  $\dot{V}_E$  at rest and during exercise reached steady states in 8–10 min.

Panel B of Figs. 2 and 3 illustrate the effects of changes in  $\dot{V}_E$  on  $P_{\text{ETCO}_2}$  at rest and during exercise in an untrained and a trained subject, respectively. Hypoventilation increased  $P_{\text{ETCO}_2}$  while hyperventilation decreased  $P_{\text{ETCO}_2}$ .  $P_{\text{ETCO}_2}$  reached steady states in 8–10 min at rest and during exercise in both untrained and trained subjects.

Figure 4 shows the characteristics of controller (A) and plant (C) (left panels) at rest and during exercise obtained from the same untrained subject shown in Fig. 2, and the controller (B) and plant (D) characteristics (right panels)





**Fig. 3** A representative example of breath-by-breath time courses of  $P_{ETCO_2}$ ,  $\dot{V}_E$ ,  $V_T$  and RR at rest and during exercise in a trained subject during open-loop respiratory chemoreflex control of  $\dot{V}_E$  and  $P_{ETCO_2}$  in

hypercapnia test (**a**) and hyper/hypoventilation test (**b**). Hypoventilation increases  $P_{ETCO_2}$  while hyperventilation decreases  $P_{ETCO_2}$ .  $P_{ETCO_2}$  reaches steady states in 8–10 min at rest and during exercise

obtained from the same trained subject shown in Fig. 3.  $\dot{V}_E$  increased linearly with increase in  $P_{ETCO_2}$  at rest and during exercise, both in representative trained and untrained subjects (Fig. 4a, b) and in pooled data ( $r^2 = 0.808$ – $0.995$  in all subjects; Fig. 5). The slope of the regression line, or the controller gain, was significantly increased by the exercise stimulus but was not different between the untrained and trained groups (Table 2). Significant interaction and main effects were observed for the  $P_{ETCO_2}$ -intercept ( $B$ ). A post-

hoc analysis revealed that the  $P_{ETCO_2}$ -intercept ( $B$ ) during exercise was greater in the trained than in the untrained group (Table 2).

The effects of voluntary  $\dot{V}_E$  changes on  $P_{ETCO_2}$  at rest and during exercise are shown in Figs. 4c and d, 6. The plant property approximated the modified metabolic hyperbola reasonably in both untrained and trained groups ( $r^2 = 0.962$ – $0.996$  in all subjects). The horizontal lines indicate the asymptotes of the modified hyperbolas. The

**Table 1** Effects of regular exercise training on the response of gas-exchange variables from rest to exercise during the spontaneous breathing (0%  $F_{ICO_2}$  trial)

	Untrained (UT) ( $n = 7$ )		Trained (Tr) ( $n = 9$ )		ANOVA ( $P$ value)		
	Rest (R)	Exercise (Ex)	Rest (R)	Exercise (Ex)	Main effect		Interaction effect
					UT vs. Tr	R vs. E	UT vs. Tr $\times$ R vs. Ex
$\dot{V}_E$ (L/min)	10.6 $\pm$ 1.5	31.2 $\pm$ 4.0	12.1 $\pm$ 2.0	24.6 $\pm$ 2.7**	0.015	<0.001	<0.001
$P_{ETCO_2}$ (mmHg)	38.7 $\pm$ 2.1	45.0 $\pm$ 4.9	39.3 $\pm$ 3.6	49.1 $\pm$ 3.3	ns	<0.001	ns
VT (mL)	702 $\pm$ 126	1351 $\pm$ 404	770 $\pm$ 155	1300 $\pm$ 230	ns	<0.001	ns
RR (breaths/min)	16.2 $\pm$ 4.6	25.0 $\pm$ 5.8	16.6 $\pm$ 4.7	20.2 $\pm$ 3.7	ns	0.004	ns
$\dot{V}_{O_2}$ (mL/min)	249 $\pm$ 79	836 $\pm$ 43	258 $\pm$ 23	842 $\pm$ 60	ns	<0.001	ns
$\dot{V}_{CO_2}$ (mL/min)	181 $\pm$ 18	762 $\pm$ 80	219 $\pm$ 21	734 $\pm$ 63	ns	<0.001	ns
$K^+$ (mmol/L)	4.1 $\pm$ 0.2	4.3 $\pm$ 0.2	4.0 $\pm$ 0.2	4.3 $\pm$ 0.1	ns	<0.001	ns
$LA^-$ (mmol/L)	1.3 $\pm$ 0.4	1.3 $\pm$ 0.4	1.2 $\pm$ 0.6	1.0 $\pm$ 0.4	ns	ns	ns

Values are mean  $\pm$  SD

$\dot{V}_E$ , minute ventilation;  $P_{ETCO_2}$ , end-tidal pressures for  $CO_2$ ;  $V_T$ , tidal volume; RR, respiratory rate;  $\dot{V}_{O_2}$ , oxygen uptake;  $\dot{V}_{CO_2}$ , carbon dioxide output;  $LA^-$ , blood lactic acid concentration;  $K^+$ , plasma potassium concentration

\*\*  $P < 0.01$  vs. UT during exercise

modified metabolic hyperbola shifted rightward and upward during exercise as predicted by an increased metabolism. The exercise stimulus increased the numerator A of the modified metabolic hyperbola but decreased the asymptote parameter C in both groups (Table 2).

Exercise stimulus increased the controller gain (S) by more than 50% in both groups. In contrast, exercise stimulus decreased the plant gain ( $G_P$ ) calculated by the reciprocal of the slope of the modified metabolic hyperbola at the operating point, in both groups (Table 2). The significant interaction effect suggests that the absolute value in  $G_P$  during exercise was smaller in the untrained than in the trained group. During exercise, the estimated total loop gain at the operating point, i.e., product of the controller gain and plant gain, was significantly higher in the trained than in the untrained group.

The respiratory equilibrium diagram was constructed by plotting the controller and plant properties together on the same graph (Figs. 4e and f, 7). The intersection point between the controller and plant curves predicts the closed-loop operating point of respiration. Exercise stimulus moved the operating point rightward and upward in both groups, indicating that the exercise stimulus increased both  $P_{ETCO_2}$  and  $\dot{V}_E$ . Furthermore, the increase in operating  $\dot{V}_E$  during exercise was smaller and the increase in  $P_{ETCO_2}$  was greater in the trained group than that in the untrained group.

As shown in Fig. 8a and b,  $P_{ETCO_2}$  and  $\dot{V}_E$  predicted from the intersection point on the respiratory equilibrium diagram conformed reasonably well to the values actually measured. The regression line for  $P_{ETCO_2}$  was

$y = 0.98x + 2.3$  ( $r^2 = 0.921$ , SEE = 1.1,  $P < 0.001$ ). The regression line for  $\dot{V}_E$  was  $y = 0.97x + 0.80$  ( $r^2 = 0.985$ , SEE = 2.0,  $P < 0.001$ ).

## Discussion

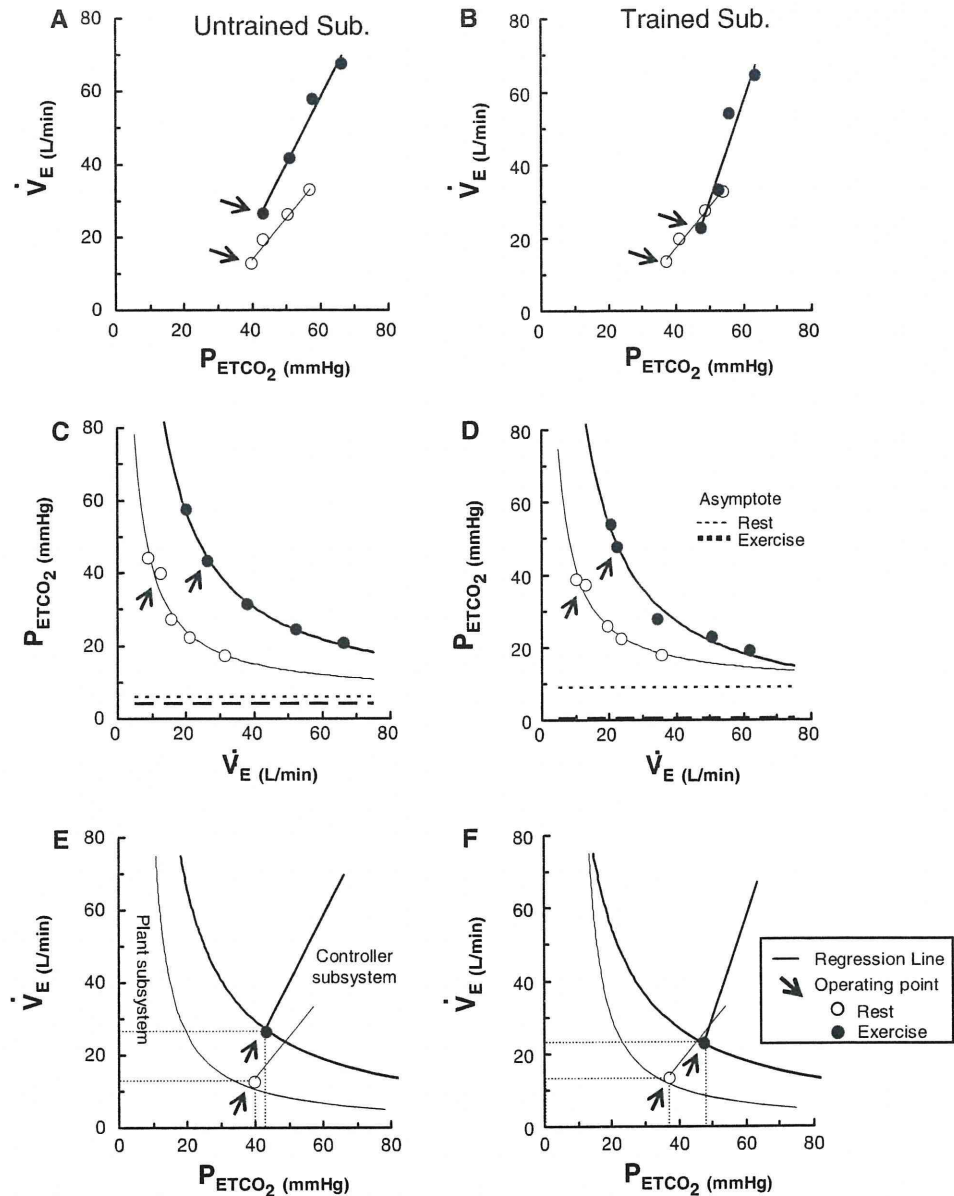
To the best of our knowledge, this is the first report that quantitatively describes the mechanism of attenuated exercise hyperpnea in endurance-trained athletes by integrating the controller and plant properties in the respiratory equilibrium diagram. The present results show that the adaptation mechanism of central controller, but not that of peripheral plant, contributes to the attenuation of exercise hyperpnea at an iso-metabolic rate in trained subjects. In addition, the exercise-induced upward shift of the controller property is less in endurance-trained than in untrained subjects, indicating that the additive exercise drive to breathe is less in trained subjects without necessarily a change in central chemoreflex threshold.

### Interpretation of the operating point of respiration using the respiratory equilibrium diagram

Our group has shown that the operating point of respiration at rest (Miyamoto et al. 2004) and during exercise (Ogoh et al. 2008) can be described by the point of intersection of the controller and plant curves in the respiratory equilibrium diagram. The concept of using the respiratory equilibrium diagram has been proposed by Mahamed et al. (2001). Indeed, operating  $P_{ETCO_2}$  and  $\dot{V}_E$  predicted from the



**Fig. 4** Characteristics of central controller (a, b), peripheral plant (c, d) and equilibrium diagram (e, f) at rest and during exercise in representative untrained and trained subjects. **a** and **b** The central controller is characterized by a linear  $P_{ETCO_2}$ – $\dot{V}_E$  relation.  $\dot{V}_E$  increases linearly with increase in  $P_{ETCO_2}$  during resting and exercising conditions in untrained (a) and trained (b) subjects. **c** and **d** The peripheral plant is characterized by a modified metabolic hyperbola. There is a good fit between measured data and the modified hyperbola in the two representative subjects. **e** and **f** The operating points estimated from the equilibrium diagram (intersection of central controller and peripheral plant plots) are very close to the measured values at rest (open circles) and during exercise (closed circles) in both representative cases

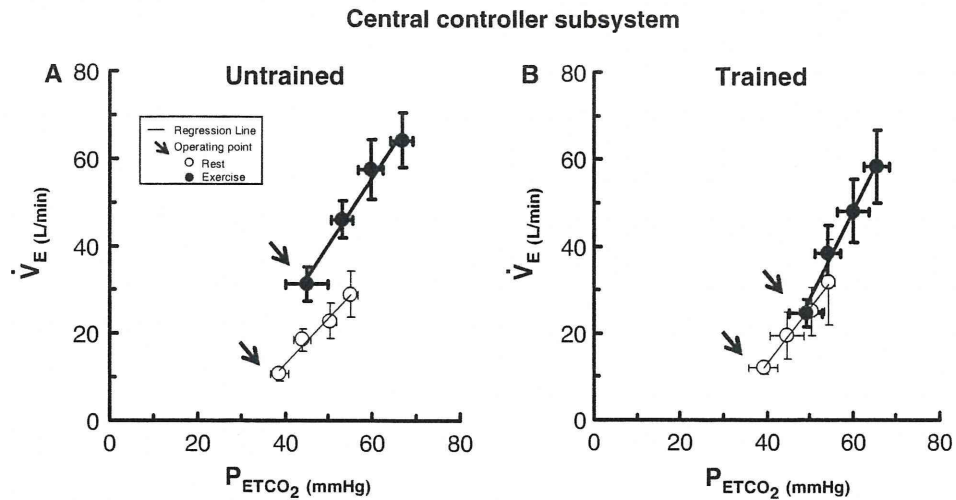


intersection point on the respiratory equilibrium diagram conformed reasonably well to the values actually measured regardless of rest, exercise, and/or physical conditions (Fig. 8). The present investigation extends previous studies by demonstrating the shifts of the operating points in the respiratory equilibrium diagram during exercise in untrained and trained subjects (Fig. 7).

Exercise stimulus moved the plant property right and upward (Fig. 6), reflecting increased metabolism. In the untrained group, if the exercise stimulus had not affected the controller property, the operating point during exercise would have been the intersecting point between the fine line and the bold hyperbola in Fig. 7a. In that case,  $P_{ETCO_2}$  would have increased to approximately 50 mmHg.

However, the exercise stimulus moved the controller property toward higher  $\dot{V}_E$  (Figs. 5, 7), which effectively stabilized  $P_{ETCO_2}$  within the normal range, at the expense of exercise hyperpnea.

In the trained group, because the intersection point between the fine line and bold hyperbola and that between the bold line and bold hyperbola is very close (Fig. 7b), the exercise-induced upward shift of the controller property did not contribute much to stabilize  $P_{ETCO_2}$ , thus attenuating exercise hyperpnea resulting in increased  $P_{ETCO_2}$  (Table 2). McConnell and Semple (1996) and Caillaud et al. (1993) reported that the endurance athletes showed the greatest rise in  $P_{aCO_2}$  or  $P_{ETCO_2}$  from rest to exercise. Taylor and Jones (1979) and Casaburi et al. (1987b) also



**Fig. 5** Characteristics of central controller subsystem at rest and during exercise obtained from pooled data of all untrained and trained subjects.  $\dot{V}_E$  increases linearly with  $P_{ETCO_2}$  during resting and exercising conditions in both groups ( $r^2 = 0.808\text{--}0.995$  in all subjects). The slope of the regression line for pooled data, which represents the gain of the controller, is increased by exercise in both groups. On the other hand, exercise decreases the intercept ( $B$ ) in

untrained, but increases  $B$  in trained subjects. **a** (untrained): The averaged regression line is  $\dot{V}_E = 1.1(P_{ETCO_2} - 27.3)$  at rest and  $\dot{V}_E = 1.5(P_{ETCO_2} - 21.3)$  during exercise. **b** (trained): The averaged regression line is  $\dot{V}_E = 1.2(P_{ETCO_2} - 27.3)$  at rest and  $\dot{V}_E = 2.0 \times (P_{ETCO_2} - 34.2)$  during exercise. Arrows denote operating points. Horizontal and vertical bars indicate  $\pm SD$

**Table 2** Effects of regular exercise training on changes in central controller and peripheral plant properties, and respiratory total loop gain from rest to exercise

	Untrained (UT)		Trained (Tr)		ANOVA ( $P$ value)		
	$(n = 7)$		$(n = 9)$		Main effect	Interaction effect	
	Rest (R)	Exercise (Ex)	Rest (R)	Exercise (Ex)	UT vs. Tr	R vs. E	UT vs. Tr $\times$ R vs. Ex
<b>Central controller</b>							
$S$ ( $\text{mL min}^{-1} \text{mmHg}^{-1}$ )	$1.1 \pm 0.3$	$1.5 \pm 0.5$	$1.2 \pm 0.4$	$2.0 \pm 0.5$	ns	<0.001	ns
$B$ (mmHg)	$26.6 \pm 5.5$	$21.3 \pm 11.1$	$27.3 \pm 7.3$	$34.2 \pm 5.3^*$	0.039	ns	0.018
<b>Peripheral plant</b>							
$A$ ( $\text{mL min mmHg}$ )	$305 \pm 76$	$1170 \pm 201$	$344 \pm 84$	$1154 \pm 193$	ns	<0.001	ns
$C$ ( $\text{mL min}^{-1}$ )	$8.5 \pm 4.2$	$5.3 \pm 4.4$	$7.6 \pm 2.2$	$1.5 \pm 3.4$	ns	0.004	ns
$G_p$ ( $\text{mL min}^{-1} \text{mmHg}$ )	$-2.7 \pm 0.7$	$-1.2 \pm 0.3$	$-2.4 \pm 0.6$	$-1.9 \pm 0.3^*$	ns	<0.001	0.018
Total loop gain	$2.8 \pm 0.7$	$2.0 \pm 1.0$	$3.0 \pm 1.4$	$3.7 \pm 1.1^{**}$	0.041	ns	0.047

Values are mean  $\pm$  SD

Central controller,  $\dot{V}_E = S(P_{ETCO_2} - B)$ ;  $S$ , central controller gain;  $B$ ,  $P_{ETCO_2}$ -intercept; Peripheral plant,  $P_{ETCO_2} = A/\dot{V}_E + C$ ;  $G_p$ , peripheral plant gain at operating point

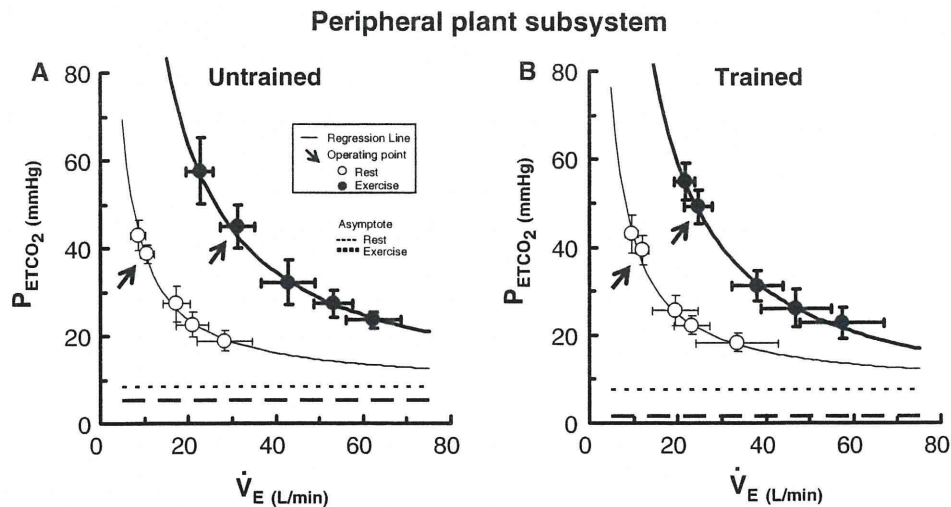
\*\*  $P < 0.01$  and \*  $P < 0.05$  vs. UT during exercise

showed that exercise training increased  $P_{aCO_2}$  and reduced  $\dot{V}_E$  for any given level of work or  $\dot{V}_{O_2}$ . These observations are consistent with our findings. The significance of training-induced change in controller property during exercise, resulting in decreased  $\dot{V}_E$  may be in augmenting the total loop gain of the respiratory chemoreflex (Table 2), rather than in stabilizing  $\dot{V}_{O_2}$ , as discussed in the next paragraphs.

Interpretation of the total loop gain of respiratory control using the respiratory equilibrium diagram

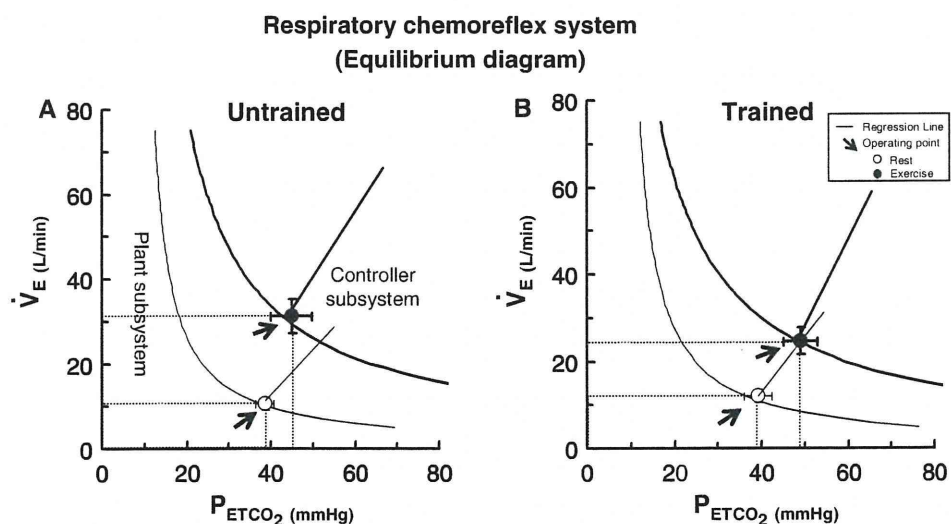
Respiratory homeostasis is maintained by a powerful feedback control system mediated by  $P_{aCO_2}$  (Defares 1964; Milhorn 1966; Cunningham et al. 1986; Duffin et al. 2000). The magnitude of this control capability can be expressed as the “total loop gain” (Berger et al. 1977;





**Fig. 6** Characteristics of peripheral plant subsystem at rest and during exercise obtained from pooled data of all untrained and trained subjects. The  $\dot{V}_E - P_{ETCO_2}$  relationship approximates the modified metabolic hyperbola reasonably well both at rest and during exercise in both groups ( $r^2 = 0.962-0.996$  in all subjects). The hyperbolic plant property shifts rightward and upward during exercise as predicted by increased metabolism. The mean value of the numerator A of the parabola increases from rest to exercise, while the asymptote constant C decreases in both groups. There is little difference between

two groups in the exercise-induced shift, although the asymptote constant C tends to be lower in trained subjects than in untrained subjects. **a** (untrained): The averaged fitted hyperbola is  $P_{ETCO_2} = 305/\dot{V}_E + 8.5$  at rest and  $P_{ETCO_2} = 1.170/\dot{V}_E + 5.3$  during exercise. **b** (trained): The averaged fitted hyperbola is  $P_{ETCO_2} = 344/\dot{V}_E + 7.6$  at rest and  $P_{ETCO_2} = 1.154/\dot{V}_E + 1.5$  during exercise. Arrows denote operating points. Horizontal and vertical bars indicate  $\pm SD$

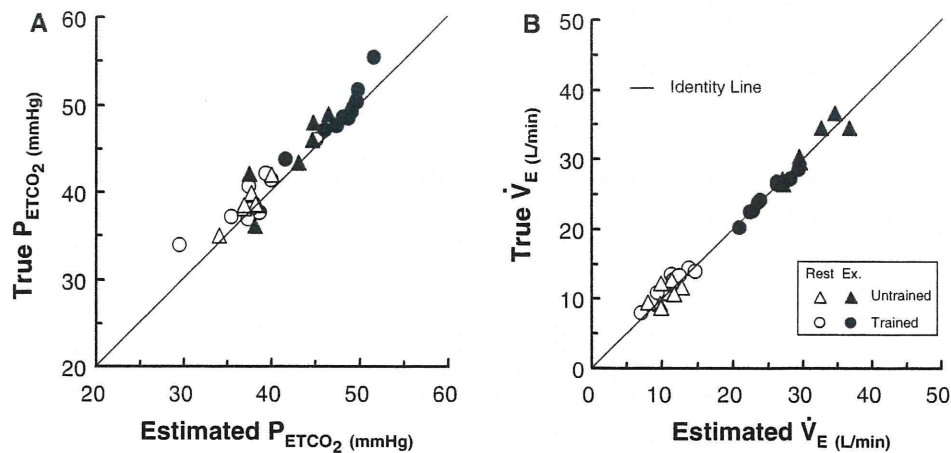


**Fig. 7** Equilibrium diagrams at rest and during exercise in untrained and trained subjects. The operating points of chemoreflex system estimated as the intersection between the controller and plant curves are very close to those measured during closed-loop spontaneous breathing at rest (open circles) and during exercise (closed circles) in untrained (a) and trained (b) groups. In untrained group (a), exercise shifts the operating point by shifting the controller curve to the

direction of decreased  $P_{ETCO_2}$ , which compensates for the shift of the plant curve accompanying increased metabolism. Compared with untrained group, strenuous regular exercise training almost abolishes the exercise-induced upward shift of the controller, but not the plant curve, thus attenuates exercise hyperpnea

Honda et al. 1983; Khoo 2000). In the respiratory equilibrium diagram, the total loop gain of respiratory control is calculated from the product of controller gain and plant gain at the intersection point. In the present study, the

total loop gain was not different between the untrained and trained subjects at rest, but was significantly higher in the trained than in the untrained subjects during exercise (Table 2).



**Fig. 8** Correlation between operating points estimated by the equilibrium diagram and those measured. **a** The estimated  $P_{ETCO_2}$  correlate closely with measured  $P_{ETCO_2}$  ( $y = 0.98x + 2.3$ ,  $r^2 = 0.927$ ,  $SEE = 1.1$ ,  $P < 0.001$ ). The solid line indicates the line of identity. **b** The estimated  $\dot{V}_E$  also correlate closely with measured

$\dot{V}_E$  ( $y = 0.97x + 0.80$ ,  $r^2 = 0.985$ ,  $SEE = 2.0$ ,  $P < 0.001$ ). The solid line indicates the line of identity. Open and closed symbols indicate rest and exercise conditions, respectively. Triangles and circles indicate untrained and trained subjects, respectively

Based on the modified metabolic hyperbola, plant gain decreases markedly as  $\dot{V}_E$  increases (Fig. 6). Because  $\dot{V}_E$  increases significantly during exercise, the plant gain would have been much smaller, if the exercise stimulus had not changed the plant property. The right and upward shift of the plant property during exercise contributed to increase the plant gain at higher  $\dot{V}_E$  range. Despite the right and upward shift in the plant property, however, the plant gain at the operating point was significantly decreased during exercise in both the untrained and trained groups. The exercise-induced increase in controller gain may therefore be important to compensate for the decreased plant gain during exercise.

Although the total loop gain did not differ between the untrained and trained groups at rest, it was significantly higher in the trained than in the untrained group during exercise (Table 2). The increased total loop gain in the trained group is the result of an increase in plant gain because the equilibrium point is at a higher  $P_{CO_2}$ ; and therefore is not related to adaptation in the chemoreflex control of  $CO_2$ .

#### Exercise-induced shift in the controller property

A large number of the physiologists accept the exercise-induced upward shift of the controller property, although the effect of exercise stimulus on the controller gain ( $CO_2$  sensitivity) varies among studies. Casey et al. (1987) demonstrated that the central chemoreceptor threshold is unchanged by exercise, and supported the neuro-humoral theory of exercise ventilation. The concept of an exercise drive to breathe that is additive to the chemoreflex drive to

breathe is now considered a common understanding. In the present study, the exercise stimulus increased the controller gain (S) both in untrained and trained subjects. However, the  $P_{ETCO_2}$ -intercept (B) during exercise was greater in the trained than in the untrained group, implying that exercise-induced upward shift of the controller property was less in the trained than in the untrained subjects (Table 2; Fig. 5). On the other hand, the  $P_{ETCO_2}$ -intercept (B) hardly shifted indicating that the chemoreflex threshold did not change. These findings thus suggest that the additive exercise drive to breathe is less in trained subjects, but not necessarily due to a change in central chemoreflex threshold.

A variety of mechanisms have been postulated to explain the exercise-induced upward shift of the controller property during exercise, leading to exercise hyperpnea (Eldridge and Waldrop 1991; Strange et al. 1993; Eldridge 1994; Mateika and Duffin 1995; Harms and Dempsey 1999), such as “irradiation” of signals from the motor cortex (Wood et al. 2003), stimulation of neural receptors in the exercising muscles (Kao 1963; McCloskey and Mitchell 1972; Smith et al. 2006; Amann et al. 2008), stimulation of chemoreceptors by humoral factors released from the exercising muscles (Casaburi et al. 1987b; Johnson et al. 1998), stimulation of chemoreceptors in the lungs by mixed venous  $P_{CO_2}$  (Wasserman et al. 1986), and thermoregulation (Hayashi et al. 2006). Furthermore, the recent experiments of Bell (2006) and Dempsey (2006) have shown that both peripheral afferent feedback and central command contribute. The fact that the effects of exercise stimulus on the controller property differ between untrained and trained subjects may contribute, in part, to the diverse results reported on the exercise-induced change in the controller property. The training-induced change in



controller property during exercise is probably independent of humoral mechanisms, because the exercise was performed at an intensity below the ventilatory threshold (Table 1). Sporer et al. (2007) showed that the entrainment of breathing rhythm to exercise rhythm may also be a factor affecting the exercise drive to breathe. However, it is unlikely that this mechanism is involved in the difference in controller property between two groups, because there were no intergroup differences in the breathing patterns at rest and during exercise. Consequently, other neural drives originating from the central nervous system, afferents from the working limbs or afferents from the heart, which is additive to the chemoreflex drive to breathe, may be involved in exercise-induced upward shift of the controller property, leading to exercise hyperpnea.

According to the results of Duffin (2005), a decrease in the central-chemoreflex threshold [ $P_{\text{ETCO}_2}$ -intercept ( $B$ )] can be explained by the effects of acute and/or chronic acid–base adjustments (e.g., reduced [strong ion difference]). In our study, it is unlikely that a difference in acid–base response to exercise between two groups contributed to the observed intergroup difference in  $P_{\text{ETCO}_2}$ -intercept ( $B$ ), since exercise was performed at a relatively low intensity (below the ventilatory threshold, with no intergroup difference in plasma lactate) (Table 1).

Another important factor that could contribute to a decrease in the  $P_{\text{ETCO}_2}$ -intercept ( $B$ ) during exercise is a change in the arterial-to-central difference in  $P_{\text{CO}_2}$ , which is primarily determined by changes in cerebral blood flow or cerebrovascular  $\text{CO}_2$  reactivity (Peebles et al. 2007; Ainslie and Duffin 2009). In the previous study, we showed that an increase in cerebrovascular  $\text{CO}_2$  reactivity during exercise compensated for an attenuated respiratory chemoreflex system controllability during exercise, especially under hypercapnic condition (Ogoh et al. 2008). Although the interaction between systemic and cerebral  $\text{CO}_2$  controlling mechanisms during exercise was not examined in the present study, we speculate that intergroup differences in the control of cerebral blood flow and cerebrovascular  $\text{CO}_2$  reactivity during exercise might have contributed, at least in part, to the observed differences.

To better understand the integrated characterization of the human chemoreflex system controlling ventilation using an equilibrium diagram, the interpretation and estimation of chemoreflex responsiveness using steady-state method should be addressed. Berkenbosch et al. (1989) and Mohan et al. (1999) demonstrated that the differences between rebreathing and steady-state affect the interpretation of results: the steady-state estimates are artefactually lower. Both Ainslie et al. (2008) and the review by Ogoh and Ainslie (2009) state that cerebral blood flow is increased in moderate exercise and this change is also

affected by fitness; the gradient is reduced during exercise. This change with respect to testing at rest will produce an increase in slope during exercise compared to at rest. Whether or not this increase accounts for the whole of the change is unknown, but if it does then the increase in the controller gain ( $S$ ) during exercise could well be artefactual. Indeed in Fig. 4a, taking only the two highest  $P_{\text{CO}_2}$  points where the gradient can be assumed to be minimized by the increased cerebral blood flow (Mohan et al. 1999), the two lines at rest and during exercise appear to be parallel.

#### Exercise-induced shift in the plant property

In the past, many researchers have explained the exercise-induced changes in plant property using the conventional metabolic hyperbola (Wasserman et al. 1986; Whipp and Pardy 1986). The equation that expresses the relation between  $\dot{V}_E$  and  $P_{\text{aCO}_2}$  during exercise disregards the scaling factors representing ventilatory work-related  $\text{CO}_2$  production. However, because ventilatory work-related  $\text{CO}_2$  production occurs and  $V_{\text{D}}/V_{\text{T}}$  changes with variation in  $\dot{V}_E$  in the “actual life” physiological system, we modified the conventional metabolic hyperbola to explain the exercise-induced changes in plant property (see Appendix). In the modified metabolic hyperbola [ $P_{\text{ETCO}_2} = A/\dot{V}_E + C$ ], exercise-induced change in the plant property is characterized by an increase in  $A$  and a decrease in  $C$ . The increase in  $A$  may result from increases in basal metabolic demand ( $\alpha$  value) and/or  $V_{\text{D}}/V_{\text{T}}$  (Appendix, Eq. 3). The decrease in  $C$  may result from decreases in  $V_{\text{D}}/V_{\text{T}}$  and/or metabolic cost of breathing ( $\beta$  value) (Appendix Eq. 3).  $V_{\text{D}}/V_{\text{T}}$  is unlikely to increase during exercise, because the exercise stimulus increases  $V_{\text{T}}$  but decreases  $V_{\text{D}}$  due to improved  $\dot{V}_A/Q$  mismatch.  $\beta$  value probably decreases with reduced airway resistance, consequently reduced oxygen cost of breathing. Therefore, the increase in  $A$  may be attributed to the increase in basal metabolic demand, and the decrease in  $C$  to decreases in both  $V_{\text{D}}/V_{\text{T}}$  and  $\beta$  value.

Numerous reports have consistently shown that regular training induces a substantial reduction in  $\dot{V}_E$  during exercise (Byrne-Quinn et al. 1971; Taylor and Jones 1979; Martin et al. 1979; Yerg et al. 1985; Casaburi et al. 1987b; Caillaud et al. 1993). The ventilatory requirement seems to be more reduced at higher exercise intensity level. Casaburi et al. (1987a, b) suggests that the reduced ventilatory response during exercise may be related to peripheral factors such as decreased blood lactate concentration, reduced  $\text{CO}_2$  production caused by increased fatty acid metabolism, and other metabolite factors. In this study, however, exercise training does not affect the

exercise-induced shift in the plant property, probably due to the low exercise intensity. If the exercise task had been performed at a higher intensity level, we might have detected a difference in the plant property between trained and untrained subjects.

**Limitations**

$P_{ETCO_2}$  measurement has been used as an estimate of  $P_{aCO_2}$ . Jones et al. (1979) reported that the difference between  $P_{aCO_2}$  and  $P_{ETCO_2}$  was influenced by  $\dot{V}_{CO_2}$  and  $V_T$ , but not by breathing frequency and exercise. Furthermore,  $P_{ETCO_2}$  is higher than  $P_{aCO_2}$  when  $\dot{V}_{CO_2}$ ,  $F_{ICO_2}$  and  $\dot{V}_E$  are increased, whereas it is lower under normal conditions. Therefore, controller gain at rest estimated using  $P_{ETCO_2}$  may be underestimated compared with using  $P_{aCO_2}$ . Furthermore, the estimation of chemoreflex responsiveness using steady-state methods has limitations that affect the interpretation of the results, as discussed in detail above. In addition, to characterize the plant subsystem, subjects voluntarily generated non-physiological respiration, which might have affected the  $P_{ETCO_2}$  response to  $\dot{V}_E$ . However, because we compared the effects of training on the respiratory chemoreflex system under the same conditions and there were no intergroup differences in  $\dot{V}_{CO_2}$  and  $V_T$  during exercise, the interpretation of the observed differences in the system properties between two groups may be rational.

Becker et al. (1996) reported that 30-min isocapnic hyperoxia leads to hyperventilation; this effect increases as  $F_{IO_2}$  increases and is substantial at high levels of  $O_2$  ( $F_{IO_2} = 0.75$ ). In our experiment, 15-min hyperoxia ( $F_{IO_2} = 0.80$ ) in each trial was used throughout all tests. Although the effect of hyperoxia may have affected our experimental results in both groups, it could not fully explain the observed differences between groups.

Arguably the major limitation of this study is the cross-sectional design. A possibility remains that the observed intergroup differences in ventilatory response to exercise and its physiological determinants do not reflect the effect of regular strenuous exercise training per se. In this regard, other factors (e.g., genetic differences, etc.) poorly controlled in the cross-sectional design of this study may have contributed, at least in part, to the observed differences.

Notwithstanding the limitations, previous numerous reports have consistently shown that exercise training attenuates exercise hyperpnea with increased  $P_{aCO_2}$  for any given level of work or  $\dot{V}_{O_2}$  (Byrne-Quinn et al. 1971; Taylor and Jones 1977; Martin et al. 1979; Yerg et al. 1985; Casaburi et al. 1987b). Based on our respiratory equilibrium model, it is reasonable to speculate that the reduced ventilatory requirement at an iso-metabolic rate during exercise in trained subjects or induced by exercise

training should arise from the exercise-induced adaptive change in the controller property. In the future, a longitudinal study would provide more valuable information on the physiological determinants of the blunted ventilatory response to exercise in aerobically trained versus untrained subjects.

**Conclusion**

Adaptation of the respiratory controller, but not that of plant, contributes to the attenuation of exercise hyperpnea at an iso-metabolic rate in trained subjects. Our experimental findings demonstrated that the exercise-induced upward shift of the controller property is less in endurance-trained than in untrained subjects, and that this effect is not due to a change in chemoreflex threshold. Whether training induces changes in neural drive originating from the central nervous system, afferents from the working limbs, or afferents from the heart, which is additive to the chemoreflex drive to breathe, cannot be determined from these results.

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**Conflict of interest** None.

**Appendix**

The metabolic hyperbola has been described conventionally by

$$P_{aCO_2} = 863 \times \dot{V}_{CO_2} / \dot{V}_A \tag{1}$$

where  $\dot{V}_A$  is alveolar ventilation (Cunningham et al. 1986; Whipp and Pardy 1986). If we approximate  $\dot{V}_A$  by  $\dot{V}_E \times [1 - V_D/V_T]$  and take the metabolic work of respiratory muscles (Harms and Dempsey 1999) into consideration, Eq. 1 can be rewritten as

$$P_{aCO_2} = 863 \times (\alpha + \beta \times \dot{V}_E) / (\dot{V}_E \times [1 - V_D/V_T]) \tag{2}$$

where  $\alpha$  and  $\beta$  are scaling factors representing  $CO_2$  production unrelated and related to respiratory work, respectively. Rearranging Eq. 2 yields



$$P_{\text{aco}_2} = A/\dot{V}_E + C \quad (3)$$

where  $A = 863 \times \alpha / (1 - V_D/V_T)$  and  $C = 863 \times \beta / (1 - V_D/V_T)$

We fit the modified hyperbola (Eq. 3) to the changes in  $P_{\text{aCO}_2}$  in response to alterations in  $\dot{V}_E$ .

## References

- Ainslie PN, Duffin J (2009) Integration of cerebrovascular CO<sub>2</sub> reactivity and chemoreflex control of breathing: mechanisms of regulation, measurement, and interpretation. *Am J Physiol Regul Integr Comp Physiol* 296:R1473–R1495
- Ainslie P, Cotter JD, George KP, Lucas S, Murrell C, Shave R, Thomas K, Williams MJA, Atkinson G (2008) Elevation in cerebral blood flow velocity with aerobic fitness throughout healthy human ageing. *J Physiol* 586:4005–4010
- Amann M, Proctor LT, Sebranek JJ, Eldridge MW, Pegelow DF, Dempsey JA (2008) Somatosensory feedback from the limbs exerts inhibitory influences on central neural drive during whole body endurance exercise. *J Appl Physiol* 105:1714–1724
- Beaver WL, Wasserman K, Whipp BJ (1986) A new method for detecting anaerobic threshold by gas exchange. *J Appl Physiol* 60:2020–2027
- Becker HF, Polo O, Mcnamara SG, Berthon-Jones M, Sullivan CE (1996) Effect of different levels of hyperoxia on breathing in healthy subjects. *J Appl Physiol* 81:1683–1690
- Bell HJ (2006) Respiratory control at exercise onset: an integrated systems perspective. *Respir Physiol Neurobiol* 152:1–15
- Berger AJ, Mitchell RA, Severinghaus JW (1977) Regulation of respiration (third of three parts). *N Engl J Med* 297:194–201
- Berkenbosch A, Bovill JG, Dahan A, DeGoede J, Olivier IC (1989) The ventilatory CO<sub>2</sub> sensitivities from Read's rebreathing method and the steady-state method are not equal in man. *J Physiol* 411:367–377
- Byrne-Quinn E, Weil JV, Sodal IE, Filley GF, Grover RF (1971) Ventilatory control in the athlete. *J Appl Physiol* 30:91–98
- Caillaud C, Anselme F, Mercier J, Prefaut C (1993) Pulmonary gas exchange and breathing pattern during and after exercise in highly trained athletes. *Eur J Appl Physiol* 67:431–437
- Casaburi R, Storer TW, Ben-Dov I, Wasserman K (1987a) Effect of endurance training on possible determinants of  $\dot{V}_{O_2}$  during heavy exercise. *J Appl Physiol* 62:199–207
- Casaburi R, Storer TW, Wasserman K (1987b) Mediation of reduced ventilatory response to exercise after endurance training. *J Appl Physiol* 63:1533–1538
- Casey K, Duffin J, McAvoy GV (1987) The effect of exercise on the central-chemoreceptor threshold in man. *J Physiol* 383:9–18
- Cummin RC, Saunders KB (1987) The ventilatory response to inhaled CO<sub>2</sub>. In: *The control of breathing in man*. Manchester University Press, Manchester, pp 45–67
- Cunningham DJC, Robbins PA, Wolff CB (1986) Integration of respiratory responses to changes in alveolar partial pressures of CO<sub>2</sub> and O<sub>2</sub> and in arterial pH. In: *Handbook of Physiology*. American Physiological Society, Bethesda, pp 476–528
- Defares JG (1964) Principles of feedback control and their application to the respiratory control system. In: *Handbook of physiology*. American Physiological Society, Bethesda, pp 649–680
- Dempsey JA (2006) Challenges for future research in exercise physiology as applied to the respiratory system. *Exerc Sport Sci Rev* 34:92–98
- Duffin J (1994) Neural drives to breathing during exercise. *Can J Appl Physiol* 19:289–304
- Duffin J (2005) Role of acid-base balance in the chemoreflex control of breathing. *J Appl Physiol* 99:2255–2265
- Duffin J, Mcavoy GV (1988) The peripheral-chemoreceptor threshold to carbon dioxide in man. *J Physiol* 406:15–26
- Duffin J, Mohan RM, Vasiliou P, Stephenson R, Mahamed S (2000) A model of the chemoreflex control of breathing in humans: model parameters measurement. *Respir Physiol* 120:13–26
- Eldridge FL (1994) Central integration of mechanisms in exercise hyperpnea. *Med Sci Sports Exerc* 26:319–327
- Eldridge FL, Waldrop TG. (1991) Neural control of breathing during exercise. In: *Exercise, pulmonary physiology and pathophysiology*. Dekker, New York, pp 309–370
- Glantz SA, Slinker BK (2001) *Primer of Applied regression & analysis of variance*, 2nd edn. McGraw Hill, New York
- Harms C, Dempsey JA (1999) Cardiovascular consequences of exercise hyperpnea. *Exerc Sport Sci Rev* 27:37–62
- Hayashi K, Honda Y, Ogawa T, Kondo N, Nishiyasu T (2006) Relationship between ventilatory response and body temperature during prolonged submaximal exercise. *J Appl Physiol* 100:414–420
- Honda Y, Hayashi F, Yoshida A, Ohyabu Y, Nishibayashi Y, Kimura H (1983) Overall “gain” of the respiratory control system in normoxic humans awake and asleep. *J Appl Physiol* 55:1530–1535
- Johnson RA, Johnson SM, Mitchell GS (1998) Catecholaminergic modulation of respiratory rhythm in an in vitro turtle brain stem preparation. *J Appl Physiol* 85:105–114
- Jones NL, Robertson DG, Kane JW (1979) Difference between end-tidal and arterial  $P_{\text{CO}_2}$  in exercise. *J Appl Physiol* 47:954–960
- Kao FF (1963) An experimental study of the pathways involved in exercise hyperventilation employing cross-circulation techniques. In: *The regulation of human respiration*. Blackwell Scientific Publications, Oxford, pp 461–502
- Khoo MCK (2000) Determinants of ventilatory instability and variability. *Respir Physiol* 122:167–182
- Lloyd BB, Cunningham DJC (1963) Quantitative approach to the regulation of human respiration. In: *The regulation of human respiration*. Blackwell Scientific Publications, Oxford, pp 331–349
- Mahamed S, Ali AF, Ho D, Wang B, Duffin J (2001) The contribution of chemoreflex drives to resting breathing in man. *Exp Physiol* 86:109–116
- Martin BJ, Sparks KE, Zwillich CW, Weil JV (1979) Low exercise ventilation in endurance athletes. *Med Sci Sports* 11:181–185
- Mateika JH, Duffin J (1995) A review of the control of breathing during exercise. *Eur J Applied Physiol* 71:1–27
- McCloskey DI, Mitchell JH (1972) Reflex cardiovascular and respiratory responses originating in exercising muscle. *J Physiol* 224:173–186
- McConnell AK, Semple ES (1996) Ventilatory sensitivity to carbon dioxide: the influence of exercise and athleticism. *Med Sci Sports Exerc* 28:685–691
- Milhorn HT Jr (1966) *The application of control theory to physiological systems*. Saunders, Philadelphia, pp 148–157
- Miyamoto T, Inagaki M, Takaki H, Kawada T, Yanagiya Y, Sugimachi M, Sunagawa K (2004) Integrated characterization of the human chemoreflex system controlling ventilation, using an equilibrium diagram. *Eur J Appl Physiol* 93:340–346
- Miyamura M, Yamashina T, Honda Y (1976) Ventilatory responses to CO<sub>2</sub> rebreathing at rest and during exercise in untrained subjects and athletes. *Jpn J Physiol* 26:245–254
- Mohan R, Duffin J (1997) The effect of hypoxia on the ventilatory response to carbon dioxide in man. *Respir Physiol* 108:101–115
- Mohan RM, Amara CE, Cunningham DA, Duffin J (1999) Measuring the central-chemoreflex sensitivity in man: rebreathing and steady-state methods compared. *Respir Physiol* 115:23–33

- Ogoh S, Ainslie PN (2009) Cerebral blood flow during exercise: mechanisms of regulation. *J Appl Physiol* 107:1370–1380
- Ogoh S, Hayashi N, Inagaki M, Ainslie PN, Miyamoto T (2008) Interaction between the ventilatory and cerebrovascular responses to hypo- and hypercapnia at rest and during exercise. *J Physiol* 586:4327–4338
- Ohyabu Y, Yoshida A, Hayashi F, Honda Y (1982) Ventilatory response to CO<sub>2</sub> after brief stimulations of the peripheral chemoreceptors in man. *Jpn J Physiol* 32:627–636
- Peebles K, Celi L, McGrattan K, Murrell C, Thomas K, Ainslie PN (2007) Human cerebrovascular and ventilatory CO<sub>2</sub> reactivity to end-tidal, arterial and internal jugular vein PCO<sub>2</sub>. *J Physiol* 584:347–357
- Poon CS, Greene JG (1985) Control of exercise hyperpnea during hypercapnia in humans. *J Appl Physiol* 59:792–797
- Robbins PA (1988) Evidence for interaction between the contributions to ventilation from the central and peripheral chemoreceptors in man. *J Physiology* 401:503–518
- Smith SA, Mitchell JH, Garry MG (2006) The mammalian exercise pressor reflex in health and disease. *Exp Physiol* 91:89–102
- Sporer BC, Foster GE, Sheel AW, McKenzie DC (2007) Entrainment of breathing in cyclists and non-cyclists during arm and leg exercise. *Respir Physiol Neurobiol* 155:64–70
- Strange S, Secher NH, Pawelczyk JA, Karpakka J, Christensen NJ, Mitchell JH, Saltin B (1993) Neural control of cardiovascular responses and of ventilation during dynamic exercise in man. *J Physiol* 470:693–704
- Taylor R, Jones NL (1979) The reduction by training of CO<sub>2</sub> output during exercise. *Eur J Cardiol* 9:53–62
- Wasserman K, Whipp BJ, Casaburi R (1986) The respiratory system. control of breathing. In: *Handbook of physiology*. American Physiological Society, Bethesda, pp 595–620
- Whipp BJ, Pardy RL (1986) Breathing during exercise. In: *Handbook of physiology*. American Physiological Society, Bethesda, pp 605–629
- Wood HE, Fatemian M, Robbins PA (2003) A learned component of the ventilatory response to exercise in man. *J Physiol* 553:967–974
- Yerg JE 2nd, Seals DR, Hagberg JM, Holloszy JO (1985) Effect of endurance exercise training on ventilatory function in older individuals. *J Appl Physiol* 58:791–794



