

tion of single Gag<sub>206-216</sub> epitope-specific CTL responses, Gag<sub>206-216</sub>-specific CTL responses were induced dominantly but Gag<sub>241-249</sub>-specific CTL responses were undetectable at week 2. In contrast, Gag<sub>241-249</sub>-specific CTL responses were induced dominantly at week 2 in group III. Both groups showed Gag<sub>206-216</sub>-specific and Gag<sub>241-249</sub>-specific CTL responses equivalently at week 6. It may be difficult to compare these results with those in group II animals inducing whole Gag antigen-specific CTL and CD4<sup>+</sup> T-cell responses before challenge; the group II animals elicited Gag<sub>206-216</sub>-specific and Gag<sub>241-249</sub>-specific CTL responses equivalently at week 2. Our results indicate that prophylactic vaccination results in dominant induction of vaccine antigen-specific CTL responses and may delay CTL responses specific for viral antigens other than vaccine antigens (referred to as nonvaccine antigens) after viral exposure.

A significant difference between groups III and IV is the pattern of selection of CTL escape mutation. All group IV animals showed rapid selection of a Gag<sub>206-216</sub>-specific CTL escape mutation, while most group III animals showed no gag mutation at week 5 but selection of the Gag<sub>206-216</sub>-specific CTL escape mutation later, at week 12. Thus, prophylactic vaccination may affect the patterns of viral genome diversification, possibly accelerating selection of CTL escape mutations. Interestingly, Gag<sub>241-249</sub>-specific CTL mutations were not detected even at week 12 in group III animals, although a previous study observed not only the Gag<sub>206-216</sub>-specific CTL escape mutation (GagL216S), but also a Gag<sub>241-249</sub>-specific CTL escape mutation (GagD244E) in the chronic phase of SIV infection in 90-120-Ia-positive macaques (9). These results indicate that delayed, naive-derived Gag<sub>206-216</sub>-specific CTL responses, as well as preceding Gag<sub>241-249</sub>-specific CTL responses, exert strong suppressive pressure on SIV replication in group III animals, implying cooperation between vaccine antigen-specific and non-vaccine antigen-specific CTL responses for virus control.

Rapid selection of the Gag<sub>206-216</sub>-specific CTL escape mutation (GagL216S) in group II and delayed selection of this mutation without a detectable Gag<sub>241-249</sub>-specific CTL escape mutation (GagD244E) in group III suggest that the virus with GagL216S (SIVmac239Gag216S) replicates more efficiently than the virus with GagD244E (SIVmac239Gag244E) under both Gag<sub>206-216</sub>-specific and Gag<sub>241-249</sub>-specific CTL responses. Our previous competition assay did not find a significant difference in viral fitness between these mutant viruses. Possibly, escape of SIVmac239Gag216S from Gag<sub>206-216</sub>-specific CTL pressure may be more efficient than that of SIVmac239Gag244E from Gag<sub>241-249</sub>-specific CTL pressure.

Our analysis revealed that the decline of plasma viral loads from week 3 to week 5 in group II+IV with rapid selection of the GagL216S mutation was significantly less than that in group III without the mutation at week 5, possibly reflecting viral escape from suppressive pressure by Gag<sub>206-216</sub>-specific CTL responses in the former groups around weeks 3 to 5. Even the comparison between groups II and III, both showing dominant Gag<sub>241-249</sub>-specific CTL responses at week 2, revealed a significantly sharper decline in the latter ( $P = 0.0087$ ). Thus, our results suggest three patterns of Gag<sub>206-216</sub>-specific and Gag<sub>241-249</sub>-specific CTL cooperation for virus control after SIVmac239 challenge. First, as observed in group II, dominantly induced Gag<sub>206-216</sub>-specific and Gag<sub>241-249</sub>-specific CTL responses both work against wild-type SIV replication around week 2, but then a mutant virus escaping

from the former CTL responses is selected, and the responses work against this mutant virus replication. Second, as observed in group III, dominantly induced Gag<sub>241-249</sub>-specific CTL responses work against wild-type SIV replication around week 2 and then contribute to virus control, together with delayed, naive-derived Gag<sub>206-216</sub>-specific CTL responses. Third, as observed in group IV, dominantly induced Gag<sub>206-216</sub>-specific CTL responses work against wild-type SIV replication around week 2, but then a mutant virus escaping from Gag<sub>206-216</sub>-specific CTL responses is selected, and delayed, naive-derived Gag<sub>241-249</sub>-specific CTL responses instead work against this mutant virus replication. Viral loads at week 3 in group III looked higher than those in group IV, implying that Gag<sub>206-216</sub>-specific CTL responses may exert a stronger suppressive effect on SIV replication in the acute phase than Gag<sub>241-249</sub>-specific CTL responses. However, viral loads at week 5 in group III looked lower than those in group IV, and the comparison between the two groups showed significantly less decline in the latter ( $P = 0.0303$ ). It is speculated that the third pattern observed in group IV is prone to failure in virus control. Indeed, two of five animals in group IV failed to control SIV replication. Even if vaccines are designed to express multiple antigens, of the vaccine-induced CTLs generated, only several epitope-specific cells may recognize the incoming HIV because of viral diversity and host MHC polymorphisms (18), and cooperation of these vaccine antigen-specific and non-vaccine antigen-specific CTL responses would be required for viral control. Thus, our results may imply a rationale of inducing escape-resistant, epitope-specific CTL memory by prophylactic AIDS vaccines.

In summary, this study showed dominant induction of vaccine antigen-specific CTL responses and delay in non-vaccine antigen-specific CTL responses in the acute phase of SIV infection, clearly describing the impact of prophylactic vaccination on CTL immunodominance and cooperation after virus exposure. Our results indicate that the patterns of cooperation of vaccine antigen-specific and non-vaccine antigen-specific CTL responses affect virus control and selection of CTL escape mutations. These findings provide great insights into antigen design in the development of a CTL-inducing AIDS vaccine.

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