

## Cell Metabolism

### Crosstalk between GR and mTOR in Skeletal Muscle

#### Cell Culture

L6 rat myoblasts, C2C12 mouse myoblasts, and COS-7 cells were obtained from American Type Culture Collection (Manassas, VA) and maintained in DMEM supplemented with 10% fetal bovine serum (Invitrogen, Carlsbad, CA). Culture conditions for myotube formation, drug treatment, and amino acids deprivation are described in the Supplemental Information.

#### In Silico Promoter Analysis

Putative FoxO1- and FoxO3-binding sequences, as well as putative GREs which are conserved between rat and human genomes, were searched for in the genomic regions (−5000 to +2000) of KLF15, REDD1, atrogin-1, and MuRF1 using rVISTA 2.0 as described in the Supplemental Information. KLF15-binding sequences (see the Supplemental Information) were searched for in the promoters of rat atrogin-1 (−4141 to +1191) and MuRF1 (−3223 to +1547) genes.

#### Chromatin Immunoprecipitation Assay

Cells or crushed tissues were treated with 1% formaldehyde in PBS for 10 min at 37°C, incubated in 125 mM glycine for 5 min, resuspended in buffer S (50 mM Tris [pH 8.0], 1% SDS, 10 mM EDTA) supplemented with 1 mM DTT, 100 nM MG132, and protease and phosphatase inhibitor cocktail (Nacalai Tesque, Kyoto, Japan), and incubated at 10°C for 10 min. Samples were sheared to an average size of 500 bp by sonication. Lysates corresponding to  $2 \times 10^6$  cells or 200 mg of crushed tissues were diluted 10-fold in buffer D (0.01% SDS, 1.1% Triton X-100, 1.2 mM EDTA, 16.7 mM Tris [pH 8.1], 167 mM NaCl) supplemented with 100 nM MG132, and protease and phosphatase inhibitor cocktail, and incubated with 5  $\mu$ g of antibodies listed in the Supplemental Information at 4°C for 18 hr. Protein A or G agarose/salmon sperm DNA (Millipore, Billerica, MA) was added and further incubated at 4°C for 1 hr. Precipitated DNA were quantified as described in the Supplemental Information.

#### Indirect Immunofluorescent Staining and Fluorescence Imaging

Muscle cryosections were treated with 0.1% Triton X-100, blocked with 5% goat serum/1% BSA in PBS, and incubated with antibodies listed in the Supplemental Information. After washing with PBS, specimens were incubated with secondary antibodies labeled with Alexa Fluor 488 or Alexa Fluor 568 (Invitrogen, 1:1000) and analyzed as described in the Supplemental Information. For imaging cultured myotubes, GFP was expressed in myotubes by infecting 10 multiplicity of infection of Ax1CAgfp (RIKEN DNA Bank, Tsukuba, Japan).

#### Statistical Analysis

Data were analyzed with Student's *t* test for unpaired data. *P* values below 0.05 were considered statistically significant. Graphs represent means  $\pm$  SD or means  $\pm$  SEM as specified in each figure legend.

#### SUPPLEMENTAL INFORMATION

Supplemental Information include one figure, Supplemental Experimental Procedures, and Supplemental References and can be found with this article at doi:10.1016/j.cmet.2011.01.001.

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#### REFERENCES

- Afring, R.P., Miller, W.J., and Buse, M.G. (1988). Effects of diabetes and starvation on skeletal muscle branched-chain alpha-keto acid dehydrogenase activity. *Am. J. Physiol.* *254*, E292–E300.
- Beesley, A.H., Firth, M.J., Ford, J., Weller, R.E., Freitas, J.R., Perera, K.U., and Kees, U.R. (2009). Glucocorticoid resistance in T-lineage acute lymphoblastic leukaemia is associated with a proliferative metabolism. *Br. J. Cancer* *100*, 1926–1936.
- Bentzinger, C.F., Romanino, K., Cloetta, D., Lin, S., Mascarenhas, J.B., Oliveri, F., Xia, J., Casanova, E., Costa, C.F., Brink, M., et al. (2008). Skeletal muscle-specific ablation of raptor, but not of rictor, causes metabolic changes and results in muscle dystrophy. *Cell Metab.* *8*, 411–424.
- Bernardi, R., Guernah, I., Jin, D., Grisendi, S., Alimonti, A., Teruya-Feldstein, J., Cordon-Cardo, C., Simon, M.C., Raffi, S., and Pandolfi, P.P. (2006). PML inhibits HIF-1 $\alpha$  translation and neoangiogenesis through repression of mTOR. *Nature* *442*, 779–785.
- Csibi, A., Cornille, K., Leibovitch, M.P., Poupon, A., Tintignac, L.A., Sanchez, A.M., and Leibovitch, S.A. (2010). The translation regulatory subunit eIF3f controls the kinase-dependent mTOR signaling required for muscle differentiation and hypertrophy in mouse. *PLoS ONE* *5*, e8994. 10.1371/journal.pone.0008994.
- Cunningham, J.T., Rodgers, J.T., Arlow, D.H., Vazquez, F., Mootha, V.K., and Puigserver, P. (2007). mTOR controls mitochondrial oxidative function through a YY1-PGC-1 $\alpha$  transcriptional complex. *Nature* *450*, 736–740.
- DeYoung, M.P., Horak, P., Sofer, A., Sgroi, D., and Ellisen, L.W. (2008). Hypoxia regulates TSC1/2–mTOR signaling and tumor suppression through REDD1-mediated 14-3-3 shuttling. *Genes Dev.* *22*, 239–251.
- Evans, R.M. (2005). The nuclear receptor superfamily: a rosetta stone for physiology. *Mol. Endocrinol.* *19*, 1429–1438.
- Fisch, S., Gray, S., Heymans, S., Haldar, S.M., Wang, B., Pfister, O., Cui, L., Kumar, A., Lin, Z., Sen-Banerjee, S., et al. (2007). Kruppel-like factor 15 is a regulator of cardiomyocyte hypertrophy. *Proc. Natl. Acad. Sci. USA* *104*, 7074–7079.
- Gilson, H., Schakman, O., Combaret, L., Lause, P., Grobet, L., Attaix, D., Ketelslegers, J.M., and Thissen, J.P. (2007). Myostatin gene deletion prevents glucocorticoid-induced muscle atrophy. *Endocrinology* *148*, 452–460.
- Glass, D.J. (2003). Signalling pathways that mediate skeletal muscle hypertrophy and atrophy. *Nat. Cell Biol.* *5*, 87–90.
- Gray, S., Wang, B., Orihuela, Y., Hong, E.G., Fisch, S., Haldar, S., Cline, G.W., Kim, J.K., Peroni, O.D., Kahn, B.B., and Jain, M.K. (2007). Regulation of gluconeogenesis by Kruppel-like factor 15. *Cell Metab.* *5*, 305–312.
- Gu, L., Gao, J., Li, Q., Zhu, Y.P., Jia, C.S., Fu, R.Y., Chen, Y., Liao, Q.K., and Ma, Z. (2008). Rapamycin reverses NPM-ALK-induced glucocorticoid resistance in lymphoid tumor cells by inhibiting mTOR signaling pathway, enhancing G1 cell cycle arrest and apoptosis. *Leukemia* *22*, 2091–2096.
- Hoffman, E.P., and Nader, G.A. (2004). Balancing muscle hypertrophy and atrophy. *Nat. Med.* *10*, 584–585.
- Hu, Z., Wang, H., Lee, I.H., Du, J., and Mitch, W.E. (2009). Endogenous glucocorticoids and impaired insulin signaling are both required to stimulate muscle wasting under pathophysiological conditions in mice. *J. Clin. Invest.* *119*, 3059–3069.
- Hundal, H.S., Babij, P., Taylor, P.M., Watt, P.W., and Rennie, M.J. (1991). Effects of corticosteroid on the transport and metabolism of glutamine in rat skeletal muscle. *Biochim. Biophys. Acta* *1092*, 376–383.
- Izumiya, Y., Hopkins, T., Morris, C., Sato, K., Zeng, L., Viereck, J., Hamilton, J.A., Ouchi, N., LeBrasseur, N.K., and Walsh, K. (2008). Fast/Glycolytic muscle fiber growth reduces fat mass and improves metabolic parameters in obese mice. *Cell Metab.* *7*, 159–172.
- Ma, K., Mallidis, C., Artaza, J., Taylor, W., Gonzalez-Cadavid, N., and Bhasin, S. (2001). Characterization of 5'-regulatory region of human myostatin gene: regulation by dexamethasone in vitro. *Am. J. Physiol. Endocrinol. Metab.* *281*, E1128–E1136.

- Mammucari, C., Milan, G., Romanello, V., Masiero, E., Rudolf, R., Del Piccolo, P., Burden, S.J., Di Lisi, R., Sandri, C., Zhao, J., et al. (2007). FoxO3 controls autophagy in skeletal muscle in vivo. *Cell Metab.* 6, 458–471.
- Matthews, S.E. (1999). Proteins and amino acids. In *Modern Nutrition and Health and Diseases*, 9th ed., M.E. Shils, J.A. Olson, M. Shike, and A.C. Ross, eds. (Baltimore: Williams & Wilkins), pp. 11–48.
- Meijsing, S.H., Pufall, M.A., So, A.Y., Bates, D.L., Chen, L., and Yamamoto, K.R. (2009). DNA binding site sequence directs glucocorticoid receptor structure and activity. *Science* 324, 407–410.
- Menconi, M., Fareed, M., O'Neal, P., Poylin, V., Wei, W., and Hasselgren, P.O. (2007). Role of glucocorticoids in the molecular regulation of muscle wasting. *Crit. Care Med.* 35, S602–S608.
- Mizushima, N., Levine, B., Cuervo, A.M., and Klionsky, D.J. (2008). Autophagy fights disease through cellular self-digestion. *Nature* 451, 1069–1075.
- Moresi, V., Williams, A.H., Meadows, E., Flynn, J.M., Potthoff, M.J., McAnally, J., Shelton, J.M., Backs, J., Klein, W.H., Richardson, J.A., et al. (2010). Myogenin and class II HDACs control neurogenic muscle atrophy by inducing E3 ubiquitin ligases. *Cell* 143, 35–45.
- Munck, A., Guyre, P.M., and Holbrook, N.J. (1984). Physiological functions of glucocorticoids in stress and their relation to pharmacological actions. *Endocr. Rev.* 5, 25–44.
- Newgard, C.B., An, J., Bain, J.R., Muehlbauer, M.J., Stevens, R.D., Lien, L.F., Haqq, A.M., Shah, S.H., Arlotto, M., Slentz, C.A., et al. (2009). A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. *Cell Metab.* 9, 311–326.
- Ning, Y.M., and Sanchez, E.R. (1993). Potentiation of glucocorticoid receptor-mediated gene expression by the immunophilin ligands FK506 and rapamycin. *J. Biol. Chem.* 268, 6073–6076.
- Risson, V., Mazelin, L., Roceri, M., Sanchez, H., Moncollin, V., Corneloup, C., Richard-Bulteau, H., Vignaud, A., Baas, D., Defour, A., et al. (2009). Muscle inactivation of mTOR causes metabolic and dystrophin defects leading to severe myopathy. *J. Cell Biol.* 187, 859–874.
- Sancak, Y., Bar-Peled, L., Zoncu, R., Markhard, A.L., Nada, S., and Sabatini, D.M. (2010). Regulator-Rag complex targets mTORC1 to the lysosomal surface and is necessary for its activation by amino acids. *Cell* 141, 290–303.
- Sandri, M. (2008). Signaling in muscle atrophy and hypertrophy. *Physiology (Bethesda)* 23, 160–170.
- Sandri, M., Sandri, C., Gilbert, A., Skurk, C., Calabria, E., Picard, A., Walsh, K., Schiaffino, S., Lecker, S.H., and Goldberg, A.L. (2004). Foxo transcription factors induce the atrophy-related ubiquitin ligase atrogin-1 and cause skeletal muscle atrophy. *Cell* 117, 399–412.
- Schakman, O., Gilson, H., and Thissen, J.P. (2008). Mechanisms of glucocorticoid-induced myopathy. *J. Endocrinol.* 197, 1–10.
- Sengupta, S., Peterson, T.R., and Sabatini, D.M. (2010). Regulation of the mTOR complex 1 pathway by nutrients, growth factors, and stress. *Mol. Cell* 40, 310–322.
- She, P., Reid, T.M., Bronson, S.K., Vary, T.C., Hajnal, A., Lynch, C.J., and Hutson, S.M. (2007). Disruption of BCATm in mice leads to increased energy expenditure associated with the activation of a futile protein turnover cycle. *Cell Metab.* 6, 181–194.
- Stitt, T.N., Drujan, D., Clarke, B.A., Panaro, F., Timofeyeva, Y., Kline, W.O., Gonzalez, M., Yancopoulos, G.D., and Glass, D.J. (2004). The IGF-1/PI3K/Akt pathway prevents expression of muscle atrophy-induced ubiquitin ligases by inhibiting FOXO transcription factors. *Mol. Cell* 14, 395–403.
- Suzuki, N., Motohashi, N., Uezumi, A., Fukada, S., Yoshimura, T., Itoyama, Y., Aoki, M., Miyagoe-Suzuki, Y., and Takeda, S. (2007). NO production results in suspension-induced muscle atrophy through dislocation of neuronal NOS. *J. Clin. Invest.* 117, 2468–2476.
- Um, S.H., D'Alessio, D., and Thomas, G. (2006). Nutrient overload, insulin resistance, and ribosomal protein S6 kinase 1, S6K1. *Cell Metab.* 3, 393–402.
- Waddell, D.S., Baehr, L.M., van den Brandt, J., Johnsen, S.A., Reichardt, H.M., Furlow, J.D., and Bodine, S.C. (2008). The glucocorticoid receptor and FOXO1 synergistically activate the skeletal muscle atrophy-associated MuRF1 gene. *Am. J. Physiol. Endocrinol. Metab.* 295, E785–E797.
- Wagenmakers, A.J. (1998). Protein and amino acid metabolism in human muscle. *Adv. Exp. Med. Biol.* 441, 307–319.
- Wang, H., Kubica, N., Ellisen, L.W., Jefferson, L.S., and Kimball, S.R. (2006). Dexamethasone represses signaling through the mammalian target of rapamycin in muscle cells by enhancing expression of REDD1. *J. Biol. Chem.* 281, 39128–39134.
- Yan, H., Frost, P., Shi, Y., Hoang, B., Sharma, S., Fisher, M., Gera, J., and Lichtenstein, A. (2006a). Mechanism by which mammalian target of rapamycin inhibitors sensitize multiple myeloma cells to dexamethasone-induced apoptosis. *Cancer Res.* 66, 2305–2313.
- Yoshikawa, N., Nagasaki, M., Sano, M., Tokudome, S., Ueno, K., Shimizu, N., Imoto, S., Miyano, S., Suematsu, M., Fukuda, K., et al. (2009). Ligand-based gene expression profiling reveals novel roles of glucocorticoid receptor in cardiac metabolism. *Am. J. Physiol. Endocrinol. Metab.* 296, E1363–E1373.
- Yu, L., McPhee, C.K., Zheng, L., Mardones, G.A., Rong, Y., Peng, J., Mi, N., Zhao, Y., Liu, Z., Wan, F., et al. (2010). Termination of autophagy and reformation of lysosomes regulated by mTOR. *Nature* 465, 942–946.
- Zhao, J., Brault, J.J., Schild, A., Cao, P., Sandri, M., Schiaffino, S., Lecker, S.H., and Goldberg, A.L. (2007). FoxO3 coordinately activates protein degradation by the autophagic/lysosomal and proteasomal pathways in atrophying muscle cells. *Cell Metab.* 6, 472–483.



