

Consequently, exposure of cultured human cells to heavy metals dramatically altered the gene expression of oxidative-responsive genes. However, in human tissues of the GSE7967 study, the p53 signaling pathway differed from that of heavy metals in the GDS2780 study. Overall, the gene expression signals were weaker than those examined in the GDS2780 study. The GSE7967 study examined cord blood collected at birth from infants whose mothers were exposed or unexposed to arsenic (0.1–68.63 mg/g), showing activation of inflammation and NFκB signaling in infants born to mothers exposed to arsenic at high concentration. Therefore, after downloading the datasets, we selected four subjects according to blood concentrations of 0.1, 1.76, 9.66, and 68.63 mg/g; then gene expression of the arsenic (As) exposure-induced responses were visualized in the p53 signaling pathway map. The highest concentration subject showed Gadd45, p53-inducible ribonucleotide reductase small subunit 2 (p53R2), spermatogenic leucine zipper 1 (TSP1), cyclinB, Cdc2, Fas, Noxa and ATR that were higher than those of the subject with the low concentration. However, p53 was opposite: high in the low-exposure subject and low in the high-exposure subject, suggesting that the down-regulation of p53 facilitates apoptosis and promotes cell proliferation.

Previous works described in our study showed that GSS (glutathione synthetase) and PRDX2 (peroxiredoxin 2) regulated TRADD (TNFRSF1A-associated via death domain), NUDT1 (nucleoside diphosphate linked moiety X-type motif 1), SOD1 (superoxide dismutase 1, soluble), and INSIG1 (Insulin induced gene 1) in the low-exposure group (mean blood concentration 0.142 μg/g), and that NUDT1 regulated TRADD, TXNRD2 (thioredoxin reductase 2), and PRDX2 in the high-exposure group

(21.41 $\mu\text{g/g}$) using the theoretical algorithm for identifying optimal gene expression networks (TAO-Gen), which is a Bayesian network algorithm used to describe gene interaction networks [18, 132-134] (Figure. 13.3). In fact, NUDT1 is a DNA repair and recombination protein. The H_2O_2 treatment significantly increased this gene and other oxidative-stress genes involved in cell cycle arrest [135]. Results of our analyses suggest that anti-oxidative stress-related genes play key roles in protection against cellular damage in the low-exposure group, but a DNA damage-related gene was dominant in the high-exposure group, in which cell damage would progress. Datasets used in this chapter are fundamental exposure to environmental stressors in normal tissues and cell lines, Therefore, this discrepancy indicates that gene expression signatures in human clinical tissues or epidemiological studies apparently reflect more inflammation than those of experimental materials, which show acute toxicity in animals after short exposure to oxidants in cell cultures.

13.3 CONCLUSION

In this chapter, we have reviewed gene expression signatures of oxidative stress-mediated signaling pathways by environmental stressors and proposed categorical pathways and canonical pathways of oxidative stress in rat and human systems. Analyses of gene expression signatures in environmental related disease such as neuronal disorders, cancer and diabetes is an important approach in etiology and risk assessment for human health to elucidate the underlying mechanisms of induced health effects. It will take

many more genetic and reverse genetic analyses, combined with functional analysis studies. Furthermore, we have shown that oxidative stress have been associated with many signaling pathways and different environmental stressors impact different molecules, but they connects with the others to the same goals like apoptosis or cell cycle. From a therapeutic point of view, researchers must consider that the best biomarker and/or therapy for oxidative stress-related disease may relies on a combination of several different agents, each targeting specifically one aspect of the oxidative stress machinery.

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13.5 REFERENCES

- [1] Sone H, Akanuma H, Fukuda T. Oxygenomics in environmental stress. *Redox Rep.* 2010;15:98-114.
- [2] Gibb S. Toxicity testing in the 21st century: a vision and a strategy. *Reprod Toxicol.* 2008;25:136-138.
- [3] Woods CG, Heuvel JP, Rusyn I. Genomic profiling in nuclear receptor-mediated toxicity. *Toxicol Pathol.* 2007;35:474-494.
- [4] Franco R, Panayiotidis MI. Environmental toxicity, oxidative stress, human disease and the "black box" of their synergism: how much have we revealed? *Mutat Res.* 2009;674:1-2.
- [5] Hansen JM, Zhang H, Jones DP. Differential oxidation of thioredoxin-1, thioredoxin-2, and glutathione by metal ions. *Free Radic Biol Med.* 2006;40:138-145.
- [6] Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem Biol Interact.* 2006;160:1-40.
- [7] Bau DT, Wang TS, Chung CH, Wang AS, Wang AS, Jan KY. Oxidative DNA adducts and DNA-protein cross-links are the major DNA lesions induced by arsenite. *Environ Health Perspect.* 2002;110 Suppl 5:753-756.

- [8] Kawai Y, Furuhata A, Toyokuni S, Aratani Y, Uchida K. Formation of acrolein-derived 2'-deoxyadenosine adduct in an iron-induced carcinogenesis model. *J Biol Chem.* 2003;278:50346-50354.
- [9] Chiu HJ, Fischman DA, Hammerling U. Vitamin A depletion causes oxidative stress, mitochondrial dysfunction, and PARP-1-dependent energy deprivation. *Faseb J.* 2008;22:3878-3887.
- [10] Knott L, Hartridge T, Brown NL, Mansell JP, Sandy JR. Homocysteine oxidation and apoptosis: a potential cause of cleft palate. *In Vitro Cell Dev Biol Anim.* 2003;39:98-105.
- [11] Nebert DW, Petersen DD, Fornace AJ, Jr. Cellular responses to oxidative stress: the [Ah] gene battery as a paradigm. *Environ Health Perspect.* 1990;88:13-25.
- [12] Cheng Y, Chang LW, Cheng LC, Tsai MH, Lin P. 4-Methoxyestradiol-induced oxidative injuries in human lung epithelial cells. *Toxicol Appl Pharmacol.* 2007;220:271-277.
- [13] Mendrick DL. Genomic and genetic biomarkers of toxicity. *Toxicology.* 2008;245:175-181.
- [14] Luhe A, Suter L, Ruepp S, Singer T, Weiser T, Albertini S. Toxicogenomics in the pharmaceutical industry: hollow promises or real benefit? *Mutat Res.* 2005;575:102-115.
- [15] Wall ME, Dyck PA, Brettin TS. SVDMAN--singular value decomposition analysis of microarray data. *Bioinformatics.* 2001;17:566-568.

- [16] Yeung KY, Ruzzo WL. Principal component analysis for clustering gene expression data. *Bioinformatics*. 2001;17:763-774.
- [17] Portier CJ, Toyoshiba H, Sone H, Parham F, Irwin RD, Boorman GA. Comparative analysis of gene networks at multiple doses and time points in livers of rats exposed to acetaminophen. *Altex*. 2006;23 Suppl:380-384.
- [18] Toyoshiba H, Yamanaka T, Sone H, et al. Gene interaction network suggests dioxin induces a significant linkage between aryl hydrocarbon receptor and retinoic acid receptor beta. *Environ Health Perspect*. 2004;112:1217-1224.
- [19] Bumgarner RE, Yeung KY. Methods for the inference of biological pathways and networks. *Methods Mol Biol*. 2009;541:225-245.
- [20] Huang JC, Babak T, Corson TW, et al. Using expression profiling data to identify human microRNA targets. *Nat Methods*. 2007;4:1045-1049.
- [21] Li H, Lu L, Manly KF, et al. Inferring gene transcriptional modulatory relations: a genetical genomics approach. *Hum Mol Genet*. 2005;14:1119-1125.
- [22] Kultz D. Molecular and evolutionary basis of the cellular stress response. *Annu Rev Physiol*. 2005;67:225-257.
- [23] Hamilton ML, Van Remmen H, Drake JA, et al. Does oxidative damage to DNA increase with age? *Proc Natl Acad Sci U S A*. 2001;98:10469-10474.
- [24] von Zglinicki T, Saretzki G, Ladhoff J, d'Adda di Fagagna F, Jackson SP. Human cell senescence as a DNA damage response. *Mech Ageing Dev*. 2005;126:111-117.

- [25] Lambeth JD. Nox enzymes, ROS, and chronic disease: an example of antagonistic pleiotropy. *Free Radic Biol Med.* 2007;43:332-347.
- [26] Collado M, Blasco MA, Serrano M. Cellular senescence in cancer and aging. *Cell.* 2007;130:223-233.
- [27] Trachootham D, Alexandre J, Huang P. Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach? *Nat Rev Drug Discov.* 2009;8:579-591.
- [28] Kimbro KS, Simons JW. Hypoxia-inducible factor-1 in human breast and prostate cancer. *Endocr Relat Cancer.* 2006;13:739-749.
- [29] Kruse JP, Gu W. Modes of p53 regulation. *Cell.* 2009;137:609-622.
- [30] Benz CC, Yau C. Ageing, oxidative stress and cancer: paradigms in parallax. *Nat Rev Cancer.* 2008;8:875-879.
- [31] Vousden KH, Ryan KM. p53 and metabolism. *Nat Rev Cancer.* 2009;9:691-700.
- [32] Capri M, Salvioli S, Sevini F, et al. The genetics of human longevity. *Ann N Y Acad Sci.* 2006;1067:252-263.
- [33] Sanchez-Capelo A. Dual role for TGF-beta1 in apoptosis. *Cytokine Growth Factor Rev.* 2005;16:15-34.
- [34] Chalmers L, Kaskel FJ, Bamgbola O. The role of obesity and its bioclinical correlates in the progression of chronic kidney disease. *Adv Chronic Kidney Dis.* 2006;13:352-364.
- [35] Eleuteri E, Magno F, Gnemmi I, et al. Role of oxidative and nitrosative stress biomarkers in chronic heart failure. *Front Biosci.* 2009;14:2230-2237.

- [36] Albano E. Oxidative mechanisms in the pathogenesis of alcoholic liver disease. *Mol Aspects Med.* 2008;29:9-16.
- [37] Vogelstein B, Lane D, Levine AJ. Surfing the p53 network. *Nature.* 2000;408:307-310.
- [38] Vousden KH, Lane DP. p53 in health and disease. *Nat Rev Mol Cell Biol.* 2007;8:275-283.
- [39] Marchenko ND, Moll UM. The role of ubiquitination in the direct mitochondrial death program of p53. *Cell Cycle.* 2007;6:1718-1723.
- [40] Chao C, Wu Z, Mazur SJ, et al. Acetylation of mouse p53 at lysine 317 negatively regulates p53 apoptotic activities after DNA damage. *Mol Cell Biol.* 2006;26:6859-6869.
- [41] Chao C, Herr D, Chun J, Xu Y. Ser18 and 23 phosphorylation is required for p53-dependent apoptosis and tumor suppression. *EMBO J.* 2006;25:2615-2622.
- [42] Blattner C, Tobiasch E, Litfen M, Rahmsdorf HJ, Herrlich P. DNA damage induced p53 stabilization: no indication for an involvement of p53 phosphorylation. *Oncogene.* 1999;18:1723-1732.
- [43] Chao C, Hergenahn M, Kaeser MD, et al. Cell type- and promoter-specific roles of Ser18 phosphorylation in regulating p53 responses. *J Biol Chem.* 2003;278:41028-41033.
- [44] Brooks CL, Gu W. Ubiquitination, phosphorylation and acetylation: the molecular basis for p53 regulation. *Curr Opin Cell Biol.* 2003;15:164-171.

- [45] Poyton RO, Ball KA, Castello PR. Mitochondrial generation of free radicals and hypoxic signaling. *Trends Endocrinol Metab.* 2009;20:332-340.
- [46] van Faassen EE, Bahrami S, Feelisch M, et al. Nitrite as regulator of hypoxic signaling in mammalian physiology. *Med Res Rev.* 2009;29:683-741.
- [47] Taylor CT, Cummins EP. The role of NF-kappaB in hypoxia-induced gene expression. *Ann N Y Acad Sci.* 2009;1177:178-184.
- [48] Maziere C, Maziere JC. Activation of transcription factors and gene expression by oxidized low-density lipoprotein. *Free Radic Biol Med.* 2009;46:127-137.
- [49] Bertout JA, Majmundar AJ, Gordan JD, et al. HIF2alpha inhibition promotes p53 pathway activity, tumor cell death, and radiation responses. *Proc Natl Acad Sci U S A.* 2009;106:14391-14396.
- [50] Moriya J, Minamino T, Tateno K, et al. Inhibition of Semaphorin As a Novel Strategy for Therapeutic Angiogenesis. *Circ Res.* 2009.
- [51] Fosslien E. Cancer morphogenesis: role of mitochondrial failure. *Ann Clin Lab Sci.* 2008;38:307-329.
- [52] Koli K, Myllarniemi M, Keski-Oja J, Kinnula VL. Transforming growth factor-beta activation in the lung: focus on fibrosis and reactive oxygen species. *Antioxid Redox Signal.* 2008;10:333-342.
- [53] Zhang H, Jiang Z, Chang J, et al. Role of NAD(P)H oxidase in transforming growth factor-beta1-induced monocyte chemoattractant protein-1 and interleukin-6 expression in rat renal tubular epithelial cells. *Nephrology (Carlton).* 2009;14:302-310.

- [54] Chan DW, Liu VW, To RM, et al. Overexpression of FOXG1 contributes to TGF-beta resistance through inhibition of p21WAF1/CIP1 expression in ovarian cancer. *Br J Cancer*. 2009;101:1433-1443.
- [55] Yu X, Riley T, Levine AJ. The regulation of the endosomal compartment by p53 the tumor suppressor gene. *FEBS J*. 2009;276:2201-2212.
- [56] Feng Z, Zhang H, Levine AJ, Jin S. The coordinate regulation of the p53 and mTOR pathways in cells. *Proc Natl Acad Sci U S A*. 2005;102:8204-8209.
- [57] Levine AJ, Feng Z, Mak TW, You H, Jin S. Coordination and communication between the p53 and IGF-1-AKT-TOR signal transduction pathways. *Genes Dev*. 2006;20:267-275.
- [58] Budanov AV, Karin M. p53 target genes *sestrin1* and *sestrin2* connect genotoxic stress and mTOR signaling. *Cell*. 2008;134:451-460.
- [59] Levine AJ, Hu W, Feng Z. The P53 pathway: what questions remain to be explored? *Cell Death Differ*. 2006;13:1027-1036.
- [60] Crichton D, Wilkinson S, O'Prey J, et al. DRAM, a p53-induced modulator of autophagy, is critical for apoptosis. *Cell*. 2006;126:121-134.
- [61] Morselli E, Tasmir E, Maiuri MC, et al. Mutant p53 protein localized in the cytoplasm inhibits autophagy. *Cell Cycle*. 2008;7:3056-3061.
- [62] Tasmir E, Chiara Maiuri M, Morselli E, et al. A dual role of p53 in the control of autophagy. *Autophagy*. 2008;4:810-814.
- [63] Tasmir E, Maiuri MC, Galluzzi L, et al. Regulation of autophagy by cytoplasmic p53. *Nat Cell Biol*. 2008;10:676-687.

- [64] Lespagnol A, Duflaut D, Beekman C, et al. Exosome secretion, including the DNA damage-induced p53-dependent secretory pathway, is severely compromised in TSAP6/Steap3-null mice. *Cell Death Differ.* 2008;15:1723-1733.
- [65] Schorey JS, Bhatnagar S. Exosome function: from tumor immunology to pathogen biology. *Traffic.* 2008;9:871-881.
- [66] Tezel G. TNF-alpha signaling in glaucomatous neurodegeneration. *Prog Brain Res.* 2008;173:409-421.
- [67] Aguilera-Aguirre L, Bacsi A, Saavedra-Molina A, Kurosky A, Sur S, Boldogh I. Mitochondrial dysfunction increases allergic airway inflammation. *J Immunol.* 2009;183:5379-5387.
- [68] Bertram KM, Baglole CJ, Phipps RP, Libby RT. Molecular regulation of cigarette smoke induced-oxidative stress in human retinal pigment epithelial cells: implications for age-related macular degeneration. *Am J Physiol Cell Physiol.* 2009;297:C1200-1210.
- [69] Lal N, Kumar J, Erdahl WE, et al. Differential effects of non-steroidal anti-inflammatory drugs on mitochondrial dysfunction during oxidative stress. *Arch Biochem Biophys.* 2009;490:1-8.
- [70] Nelson GM, Ahlborn GJ, Allen JW, et al. Transcriptional changes associated with reduced spontaneous liver tumor incidence in mice chronically exposed to high dose arsenic. *Toxicology.* 2009;266:6-15.
- [71] Bailey SM. A review of the role of reactive oxygen and nitrogen species in alcohol-induced mitochondrial dysfunction. *Free Radic Res.* 2003;37:585-596.

- [72] Kim GJ, Chandrasekaran K, Morgan WF. Mitochondrial dysfunction, persistently elevated levels of reactive oxygen species and radiation-induced genomic instability: a review. *Mutagenesis*. 2006;21:361-367.
- [73] Protti A, Singer M. Bench-to-bedside review: potential strategies to protect or reverse mitochondrial dysfunction in sepsis-induced organ failure. *Crit Care*. 2006;10:228.
- [74] Shults CW. Mitochondrial dysfunction and possible treatments in Parkinson's disease--a review. *Mitochondrion*. 2004;4:641-648.
- [75] Lee HK, Cho YM, Kwak SH, Lim S, Park KS, Shim EB. Mitochondrial dysfunction and metabolic syndrome-looking for environmental factors. *Biochim Biophys Acta*. 2009.
- [76] Williamson G, Manach C. Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. *Am J Clin Nutr*. 2005;81:243S-255S.
- [77] Manach C, Williamson G, Morand C, Scalbert A, Remesy C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am J Clin Nutr*. 2005;81:230S-242S.
- [78] Zhou C, Huang Y, Przedborski S. Oxidative stress in Parkinson's disease: a mechanism of pathogenic and therapeutic significance. *Ann N Y Acad Sci*. 2008;1147:93-104.

- [79] Drechsel DA, Patel M. Role of reactive oxygen species in the neurotoxicity of environmental agents implicated in Parkinson's disease. *Free Radic Biol Med.* 2008;44:1873-1886.
- [80] Almon RR, Lai W, DuBois DC, Jusko WJ. Corticosteroid-regulated genes in rat kidney: mining time series array data. *Am J Physiol Endocrinol Metab.* 2005;289:E870-882.
- [81] Jin JY, Almon RR, DuBois DC, Jusko WJ. Modeling of corticosteroid pharmacogenomics in rat liver using gene microarrays. *J Pharmacol Exp Ther.* 2003;307:93-109.
- [82] Sullivan CJ, Teal TH, Luttrell IP, Tran KB, Peters MA, Wessells H. Microarray analysis reveals novel gene expression changes associated with erectile dysfunction in diabetic rats. *Physiol Genomics.* 2005;23:192-205.
- [83] Lattanzi W, Bernardini C, Gangitano C, Michetti F. Hypoxia-like transcriptional activation in TMT-induced degeneration: microarray expression analysis on PC12 cells. *J Neurochem.* 2007;100:1688-1702.
- [84] Erlandsen SE, Fykse V, Waldum HL, Sandvik AK. Octreotide induces apoptosis in the oxyntic mucosa. *Mol Cell Endocrinol.* 2007;264:188-196.
- [85] Chen H, Huang XN, Stewart AF, Sepulveda JL. Gene expression changes associated with fibronectin-induced cardiac myocyte hypertrophy. *Physiol Genomics.* 2004;18:273-283.
- [86] Guan H, Arany E, van Beek JP, et al. Adipose tissue gene expression profiling reveals distinct molecular pathways that define visceral adiposity in offspring of

- maternal protein-restricted rats. *Am J Physiol Endocrinol Metab.* 2005;288:E663-673.
- [87] Sakurai H, Bush KT, Nigam SK. Heregulin induces glial cell line-derived neurotrophic growth factor-independent, non-branching growth and differentiation of ureteric bud epithelia. *J Biol Chem.* 2005;280:42181-42187.
- [88] Koh S, Chung H, Xia H, Mahadevia A, Song Y. Environmental enrichment reverses the impaired exploratory behavior and altered gene expression induced by early-life seizures. *J Child Neurol.* 2005;20:796-802.
- [89] Kubisch CH, Gukovsky I, Lugea A, et al. Long-term ethanol consumption alters pancreatic gene expression in rats: a possible connection to pancreatic injury. *Pancreas.* 2006;33:68-76.
- [90] Kodavanti UP, Schladweiler MC, Ledbetter AD, et al. The spontaneously hypertensive rat: an experimental model of sulfur dioxide-induced airways disease. *Toxicol Sci.* 2006;94:193-205.
- [91] Bruder ED, Lee JJ, Widmaier EP, Raff H. Microarray and real-time PCR analysis of adrenal gland gene expression in the 7-day-old rat: effects of hypoxia from birth. *Physiol Genomics.* 2007;29:193-200.
- [92] Almon RR, DuBois DC, Yao Z, Hoffman EP, Ghimbovschi S, Jusko WJ. Microarray analysis of the temporal response of skeletal muscle to methylprednisolone: comparative analysis of two dosing regimens. *Physiol Genomics.* 2007;30:282-299.

- [93] Chan MM, Lu X, Merchant FM, Iglehart JD, Miron PL. Gene expression profiling of NMU-induced rat mammary tumors: cross species comparison with human breast cancer. *Carcinogenesis*. 2005;26:1343-1353.
- [94] Kendzioriski C, Irizarry RA, Chen KS, Haag JD, Gould MN. On the utility of pooling biological samples in microarray experiments. *Proc Natl Acad Sci U S A*. 2005;102:4252-4257.
- [95] Aplin AC, Gelati M, Fogel E, Carnevale E, Nicosia RF. Angiopoietin-1 and vascular endothelial growth factor induce expression of inflammatory cytokines before angiogenesis. *Physiol Genomics*. 2006;27:20-28.
- [96] Rampil IJ, Moller DH, Bell AH. Isoflurane modulates genomic expression in rat amygdala. *Anesth Analg*. 2006;102:1431-1438.
- [97] Collins JF. Gene chip analyses reveal differential genetic responses to iron deficiency in rat duodenum and jejunum. *Biol Res*. 2006;39:25-37.
- [98] Guzelian J, Barwick JL, Hunter L, Phang TL, Quattrochi LC, Guzelian PS. Identification of genes controlled by the pregnane X receptor by microarray analysis of mRNAs from pregnenolone 16alpha-carbonitrile-treated rats. *Toxicol Sci*. 2006;94:379-387.
- [99] Gebel S, Gerstmayer B, Kuhl P, Borlak J, Meurrens K, Muller T. The kinetics of transcriptomic changes induced by cigarette smoke in rat lungs reveals a specific program of defense, inflammation, and circadian clock gene expression. *Toxicol Sci*. 2006;93:422-431.

- [100] Su Y, Simmen FA, Xiao R, Simmen RC. Expression profiling of rat mammary epithelial cells reveals candidate signaling pathways in dietary protection from mammary tumors. *Physiol Genomics*. 2007;30:8-16.
- [101] Rowe WB, Blalock EM, Chen KC, et al. Hippocampal expression analyses reveal selective association of immediate-early, neuroenergetic, and myelinogenic pathways with cognitive impairment in aged rats. *J Neurosci*. 2007;27:3098-3110.
- [102] Volpicelli F, Caiazzo M, Greco D, et al. Bdnf gene is a downstream target of Nurr1 transcription factor in rat midbrain neurons in vitro. *J Neurochem*. 2007;102:441-453.
- [103] Stemmer K, Ellinger-Ziegelbauer H, Ahr HJ, Dietrich DR. Carcinogen-specific gene expression profiles in short-term treated Eker and wild-type rats indicative of pathways involved in renal tumorigenesis. *Cancer Res*. 2007;67:4052-4068.
- [104] Impey S, McCorkle SR, Cha-Molstad H, et al. Defining the CREB regulon: a genome-wide analysis of transcription factor regulatory regions. *Cell*. 2004;119:1041-1054.
- [105] Bush EW, Hood DB, Papst PJ, et al. Canonical transient receptor potential channels promote cardiomyocyte hypertrophy through activation of calcineurin signaling. *J Biol Chem*. 2006;281:33487-33496.
- [106] Zhou Z, Cornelius CP, Eichner M, Bornemann A. Reinnervation-induced alterations in rat skeletal muscle. *Neurobiol Dis*. 2006;23:595-602.

- [107] Bursztyn M, Gross ML, Goltser-Dubner T, et al. Adult hypertension in intrauterine growth-restricted offspring of hyperinsulinemic rats: evidence of subtle renal damage. *Hypertension*. 2006;48:717-723.
- [108] Thomas H, Senkel S, Erdmann S, et al. Pattern of genes influenced by conditional expression of the transcription factors HNF6, HNF4alpha and HNF1beta in a pancreatic beta-cell line. *Nucleic Acids Res*. 2004;32:e150.
- [109] Schumann A, Nutten S, Donnicola D, et al. Neonatal antibiotic treatment alters gastrointestinal tract developmental gene expression and intestinal barrier transcriptome. *Physiol Genomics*. 2005;23:235-245.
- [110] Roy S, Khanna S, Kuhn DE, et al. Transcriptome analysis of the ischemia-reperfused remodeling myocardium: temporal changes in inflammation and extracellular matrix. *Physiol Genomics*. 2006;25:364-374.
- [111] Tugues S, Morales-Ruiz M, Fernandez-Varo G, et al. Microarray analysis of endothelial differentially expressed genes in liver of cirrhotic rats. *Gastroenterology*. 2005;129:1686-1695.
- [112] Akavia UD, Shur I, Rechavi G, Benayahu D. Transcriptional profiling of mesenchymal stromal cells from young and old rats in response to Dexamethasone. *BMC Genomics*. 2006;7:95.
- [113] Zhou M, Roma A, Magi-Galluzzi C. The usefulness of immunohistochemical markers in the differential diagnosis of renal neoplasms. *Clin Lab Med*. 2005;25:247-257.

- [114] Schiffer D, Giordana MT, Mauro A, Migheli A, Germano I, Giaccone G. Immunohistochemical demonstration of vimentin in human cerebral tumors. *Acta Neuropathol.* 1986;70:209-219.
- [115] Niehans GA, Manivel JC, Copland GT, Scheithauer BW, Wick MR. Immunohistochemistry of germ cell and trophoblastic neoplasms. *Cancer.* 1988;62:1113-1123.
- [116] Iwakuma T, Lozano G. Crippling p53 activities via knock-in mutations in mouse models. *Oncogene.* 2007;26:2177-2184.
- [117] Marine JC, Jochemsen AG. Mdmx as an essential regulator of p53 activity. *Biochem Biophys Res Commun.* 2005;331:750-760.
- [118] Tang Y, Zhao W, Chen Y, Zhao Y, Gu W. Acetylation is indispensable for p53 activation. *Cell.* 2008;133:612-626.
- [119] Gong X, Kole L, Iskander K, Jaiswal AK. NRH:quinone oxidoreductase 2 and NAD(P)H:quinone oxidoreductase 1 protect tumor suppressor p53 against 20s proteasomal degradation leading to stabilization and activation of p53. *Cancer Res.* 2007;67:5380-5388.
- [120] Lai Z, Yang T, Kim YB, et al. Differentiation of Hdm2-mediated p53 ubiquitination and Hdm2 autoubiquitination activity by small molecular weight inhibitors. *Proc Natl Acad Sci U S A.* 2002;99:14734-14739.
- [121] Wang W, Ho WC, Dicker DT, et al. Acridine derivatives activate p53 and induce tumor cell death through Bax. *Cancer Biol Ther.* 2005;4:893-898.

- [122] Kawata K, Yokoo H, Shimazaki R, Okabe S. Classification of heavy-metal toxicity by human DNA microarray analysis. *Environ Sci Technol.* 2007;41:3769-3774.
- [123] Fry RC, Navasumrit P, Valiathan C, et al. Activation of inflammation/NF-kappaB signaling in infants born to arsenic-exposed mothers. *PLoS Genet.* 2007;3:e207.
- [124] Chang L, Zhou B, Hu S, et al. ATM-mediated serine 72 phosphorylation stabilizes ribonucleotide reductase small subunit p53R2 protein against MDM2 to DNA damage. *Proc Natl Acad Sci U S A.* 2008;105:18519-18524.
- [125] Kollberg G, Darin N, Benan K, et al. A novel homozygous RRM2B missense mutation in association with severe mtDNA depletion. *Neuromuscul Disord.* 2009;19:147-150.
- [126] Liu X, Xue L, Yen Y. Redox property of ribonucleotide reductase small subunit M2 and p53R2. *Methods Mol Biol.* 2008;477:195-206.
- [127] Spinazzola A, Invernizzi F, Carrara F, et al. Clinical and molecular features of mitochondrial DNA depletion syndromes. *J Inherit Metab Dis.* 2009;32:143-158.
- [128] Tynismaa H, Suomalainen A. Mouse models of mitochondrial DNA defects and their relevance for human disease. *EMBO Rep.* 2009;10:137-143.
- [129] Ceryak S, Zingariello C, O'Brien T, Patierno SR. Induction of pro-apoptotic and cell cycle-inhibiting genes in chromium (VI)-treated human lung fibroblasts: lack of effect of ERK. *Mol Cell Biochem.* 2004;255:139-149.

- [130] Fanzo JC, Reaves SK, Cui L, et al. Zinc status affects p53, gadd45, and c-fos expression and caspase-3 activity in human bronchial epithelial cells. *Am J Physiol Cell Physiol.* 2001;281:C751-757.
- [131] Shih RS, Wong SH, Schoene NW, Lei KY. Suppression of Gadd45 alleviates the G2/M blockage and the enhanced phosphorylation of p53 and p38 in zinc supplemented normal human bronchial epithelial cells. *Exp Biol Med (Maywood).* 2008;233:317-327.
- [132] Toyoshiba H, Sone H, Yamanaka T, et al. Gene interaction network analysis suggests differences between high and low doses of acetaminophen. *Toxicol Appl Pharmacol.* 2006;215:306-316.
- [133] Yamanaka T, Toyoshiba H, Sone H, Parham FM, Portier CJ. The TAO-Gen algorithm for identifying gene interaction networks with application to SOS repair in *E. coli*. *Environ Health Perspect.* 2004;112:1614-1621.
- [134] Sone H, Imanishi S, Akanuma H, et al. Gene Expression Signatures of Environmental Chemicals in Cancer and in Developmental Disorders In: Zhao B DM, Cadeans E, ed. *The roles of free radicals in biology and medicine*. Beijing: Medimond; 2009:45-52.
- [135] Chua PJ, Yip GW, Bay BH. Cell cycle arrest induced by hydrogen peroxide is associated with modulation of oxidative stress related genes in breast cancer cells. *Exp Biol Med (Maywood).* 2009;234:1086-1094.