This statement was agreed on by 89% of the participants at the ILCA consensus meeting, but 11% disagreed with this statement. At the JSH consensus meeting, 84% of the participants agreed with this statement, and 16% disagreed. The outcome should be evaluated by both overall survival and incidence of recurrence.

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Disclosure Statement

The author declares that he has no financial conflict of interest.

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Original Article

Prolonged treatment with pegylated interferon α 2b plus ribavirin improves sustained virological response in chronic hepatitis C genotype 1 patients with late response in a clinical real-life setting in Japan

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Aim: This study was conducted to clarify the factors related to sustained virological response (SVR) to pegylated interferon α 2b (PEG-IFN) plus ribavirin (RBV) combination therapy administered for 48 weeks in patients with chronic hepatitis C virus (CHCV) and to evaluate the usefulness of prolonged treatment in patients with late virological response (LVR).

Methods: Of 2257 patients registered at 68 institutions, those with genotype 1 and high viral load were selected to participate in two studies. Study 1 (standard 48-week group, $n = 1480$) investigated SVR-determining factors in patients who received the treatment for ≤52 weeks, whereas study 2 compared SVR rates between patients with LVR who received treatment for either 36–52 weeks (48-week group, $n = 223$) or 60–76 weeks (72-week group, $n = 73$).

Results: In study 1, SVR rate was 44.9%; that in male subjects (50.4%) was significantly ($P < 0.0001$) higher than in female subjects (36.4%). SVR rate significantly ($P < 0.0001$) decreased with 10-year age increments in both sexes. Multivariate logistic regression analysis revealed that age, F score, platelet count, and HCV load were SVR-related factors. In study 2, SVR rate in the 72-week group (67.1%) was significantly ($P = 0.0020$) higher than in the 48-week group (46.2%).

Conclusions: Patients with CHCV genotype 1 infection should be treated with PEG-IFN plus ribavirin combination therapy as early as possible, and 72 weeks' treatment is recommended in patients with LVR regardless of age.

Key words: chronic hepatitis C virus, elderly patients, pegylated interferon, prolonged treatment, ribavirin

INTRODUCTION

THE TOTAL NUMBER of patients infected with the hepatitis C virus (HCV) is estimated at 170 million worldwide, of whom 1.5–1.7 million are Japanese. Treatment of HCV infection began with interferon (IFN) monotherapy before the discovery of HCV in 1989. At that time, responders to treatment were mostly limited to patients with HCV genotypes 2 or 3 infection, which is highly sensitive to IFN. The sustained virological response (SVR: HCV-RNA negative at 24 weeks after end of treatment) to IFN monotherapy in genotype 1 patients known from that time to be difficult to treat was only about 5%. SVR rate has since increased thanks to concomitant administration of the antiviral drug ribavirin (RBV), and with the development of the long-acting

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IFN product pegylated interferon (PEG-IFN) it has increased to 50%.[1–4] Today, PEG-IFN plus ribavirin regimen is internationally recognized as a standard therapy for chronic hepatitis C virus (CHCV) infection.[5,6] Early clinical trials of this regimen focused on specific patient populations. Subsequently, several multinational studies such as WIN-R,[7] HALT-C,[8] EPIC3,[9] and REPEAT Study[10] have been conducted in the general clinical setting. The results of the IDEAL Study[11] directly comparing PEG-IFN α 2a versus PEG-IFN α 2b have also been published. From these studies, variables predictive of SVR have been identified, including ethnicity, sex, age, and weight as demographic parameters, staging and hepatic steatosis as histological parameters, viral load, genotype, NS5A, and core mutation as virologic parameters, alanine aminotransferase (ALT) and γ-glutamyl transpeptidase (GGT) as biochemical parameters, and even the timing of viral negativity as a treatment variable.[12–15] More recently, the SVR rate was reported to increase in association with decrease in the relapse rate with 72-week treatment in patients with delayed HCV-RNA negativity.[15,16] However, the majority of patients participating in previous studies in western countries were aged in their 40s on average, and the influence of aging of the patient population has not been studied adequately.

We therefore examined SVR-determining factors with 48-week PEG-IFN α 2b plus RBV combination therapy in the prevailing Japanese clinical setting characterized by increasing numbers of elderly patients. We also compared SVR rate between 48-week and 72-week treatment in patients with late virological response (LVR) defined as achieving HCV-RNA negativity in the period from weeks 13 to 24 after the start of treatment so as to examine the significance of prolonged treatment.

METHODS

Patients

A MULTICENTER STUDY was conducted at 68 institutions in Tokyo and Yamanashi prefectures (PERFECT Study Group; see Appendix I) to survey the actual state of combination therapy with PEG-IFN α 2b (PegIntron; Schering Plough, Kenilworth, NJ) and RBV (Rebetol, Schering Plough) in 2008. A total of 2257 chronic hepatitis C virus (CHCV) patients seen from December 2004 who completed combination treatment by September 2007 were registered regardless of genotype, history of IFN treatment, and ALT levels. The presence of HCV in serum had to be confirmed by Cobas Amplicor HCV Monitor, version 2.0 (Roche Diagnostic, Tokyo) for registration.

Excluded from this study were pregnant or possibly pregnant and lactating women, and patients with severe heart disease, chronic kidney failure or creatinine clearance of ≤50 mL/min, current or history of severe psychiatric disorder, and autoimmune hepatitis.

Demographic characteristics examined included age, sex, height and weight, the presence or absence of diabetes mellitus, hypertension, heavy drinking, and history of IFN therapy and hepatic cancer. Hepatic histological data recorded were stage (F0–F4) and grade (A0–A3). Laboratory tests recorded were ALT, platelet count, albumin, and α-fetoprotein (AFP) before the start of PEG-IFN α 2b plus RBV combination therapy.

As indicated in Figure 1, of the total 2257 patients registered, patients with genotype 1 and high viral load (>100 KIU/mL: Amplicor PCR quantitation) who satisfied the following conditions were included in this study: patients who received treatment for ≤52 weeks (standard 48-week treatment group, $n = 1480$) in study 1, and patients with LVR who received treatment for either 36–52 weeks (48-week treatment group, $n = 223$) or 60–76 weeks (72-week treatment group, $n = 73$) in study 2.

This multicenter study was approved by IRB at each participating institution. The study protocol was carried out according to the ethical guidelines of the 1975 Declaration of Helsinki. Informed consent was obtained from each patient.

Treatment

PEG-IFN α 2b was administered subcutaneously once weekly at a dose of 1.5 μg/kg. Dose reduction and treatment discontinuation followed the instructions given in the package insert, i.e., the dose was reduced by half if WBC decreased to <1500/mm^3, neutrophils to <750/mm^3 or platelet count to <80000/mm^3, and treatment was discontinued if WBC decreased to <1000/mm^3, neutrophils to <500/mm^3 or platelet count to <50000/mm^3. RBV was administered in two divided doses of 600, 800, or 1000 mg/day in patients weighing <60, 60–<80, and ≥80 kg, respectively. Dose reduction and treatment discontinuation followed the package insert, i.e., dose was reduced from 600 mg/day to 400 mg/day, from 800 mg/day to 600 mg/day, or from 1000 mg/day to 600 mg/day if hemoglobin (Hb) concentration decreased to <10 g/dL, and administration was discontinued if Hb decreased to 8.5 g/dL. Duration of treatment was 48 weeks as a rule. In LVR patients who did

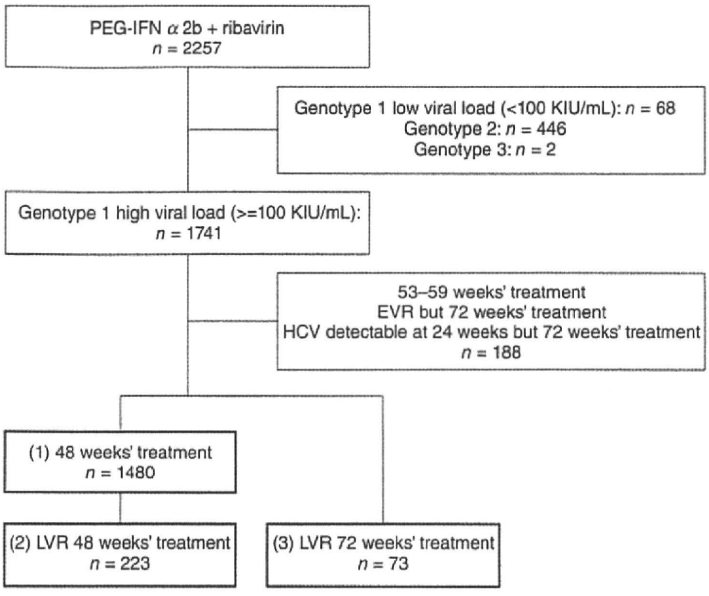

Figure 1 Flow-chart of study subjects. (1) 48 weeks' treatment (48-week standard therapy group): patients with genotype 1 and high viral load who received pegylated interferon α 2b (PEG-IFN α 2b) + ribavirin (RBV) for 52 weeks. Multiple logistic regression analysis was used to evaluate the response to PEG-IFN α 2b + RBV in this group (2) Late virological response (LVR) 48 weeks' treatment: patients with genotype 1 and high viral load who received PEG-IFN α 2b + RBV for 36–52 weeks (3) LVR 72 weeks' treatment: patients with genotype 1 and high viral load who received PEG-IFN α 2b + RBV for 60–76 weeks. SVR rate was compared between LVR 48 weeks' treatment group (2) and LVR 72 weeks' treatment group (3). EVR, early virologic response; HCV, hepatitis C virus.

not achieve HCV-RNA negativity by week 12, treatment could be extended for 48 weeks or longer based on individual patients' desire and investigators' judgment.

Evaluation of response to treatment

Determination of genotype and measurement of HCV-RNA levels were performed at each center. Pre-treatment HCV-RNA levels were determined by Amplicor PCR quantitation. Viral negativity was defined as HCV below detection limit (<50 IU/mL) by Amplicor qualitative analysis (Roche Molecular Systems, NJ).

SVR was defined as HCV below detection limit at 24 weeks after the end of PEG-IFN α 2b plus RBV combination therapy by Amplicor HCV qualitative analysis.

Statistical analysis

All statistical analyses were performed using SAS, version 9.13 (SAS Institute, Cary, NC). Intergroup comparison of SVR rate was performed by Fisher's exact test; that of background variables by Fisher's exact test and Mann–Whitney *U*-test. Trend of SVR rate by age was assessed by Cochran–Armitage test, and intergroup comparison after adjustment of stratification factors was conducted by Mantel–Haenstzel method. Determination of factors associated with SVR was conducted by a stepwise procedure using the results of logistic univariate analysis ($P < 0.2$) into logistic multivariate analysis. All tests were two-sided, with significance level set at $P < 0.05$.

RESULTS

Study 1: SVR-related factors in patients receiving standard 48-week treatment

AS INDICATED IN Table 1 and Figure 1, 1480 subjects (male, $n = 898$ [60.7%]; median age, 57 [range, 13–79] years) were eligible for analysis. SVR rate based on ITT was 44.9%. SVR rate in subjects who completed and who discontinued treatment was 56.5% ($n = 1110$) and 10.3% ($n = 370$), respectively, a statistically significant difference ($P < 0.0001$). SVR rate in male subjects (50.4%; 453/898) was significantly ($P < 0.0001$) higher than in female subjects (36.4%; 212/582). SVR rate significantly ($P < 0.0001$) decreased as age increased by 10 years in both male and female subjects (Fig. 2); the odds ratio for SVR decreasing with 10-year increase in age was 0.688 (95% CI, 0.604–0.784; $P < 0.0001$) in male subjects and 0.546 (0.449–0.663; <0.0001) in female subjects, indicating that the influence of aging was greater in female than in male subjects. There was no bias of older versus younger age among patients who had and had not previously

Table 1 Pretreatment characteristics of chronic hepatitis C virus (CHCV) patients with HCV-1b RNA who received pegylated interferon α 2b + ribavirin standard therapy for 48 weeks

Characteristic	Value (n = 1480)
Sex (male/female)	898/582
Age (years)	57 (13–79)
History of HCC (yes/no/unknown)	8/1405/67
Previous IFN treatment (yes/no/unknown)	459/688/333
Diabetes (yes/no/unknown)	44/480/956
Hypertension (yes/no/unknown)	105/417/958
Ongoing alcohol use (yes/no/unknown)	157/456/867
Grade (A0/A1/A2/A3/unknown)	14/499/478/55/434
Stage (F0/F1/F2/F3/F4/unknown)	36/469/316/176/48/435
ALT (IU/L)	63 (8.4–910)
Platelets (×10⁴/μL)	16.6 (4.3–47.7)
Viral load (KIU/mL)	1900 (100–5100)

Data expressed as median (range). HCC, hepatocellular carcinoma; ALT, alanine aminotransferase; IFN, interferon.

received IFN. Whereas, multivariate logistic regression analysis revealed that older age (<55/≥55 years), degree of progression of hepatic fibrosis (F0–1/2–4), low platelet count (≥16/<16×10⁴/μL), and high viral load (<1900/≥1900 KIU/mL) are resistance factors to SVR (Table 2). In multivariate logistic regression analysis, sex was not selected.

Study 2: usefulness of prolonged treatment in LVR patients

Of the patients who completed standard 48-week treatment, 223 patients (20.0%) showed LVR (Fig. 1), and median duration of treatment was 48 weeks. Compared with patients who exhibited early virologic response (EVR) defined as HCV-RNA negative within 12 weeks after the start of treatment, those with LVR were older (median age, 58 vs 55 years; $P = 0.0043$) and had higher viral load (median, 2700 vs 1620 KIU/mL; $P < 0.0001$) and lower platelet count (median, 16.5 vs 17.3 × 10⁴/μL; $P = 0.0162$). SVR rate based on treatment analysis was 56.5 in all, 79.2% in EVR and 46.2% in LVR, respectively. In multivariate logistic regression analysis of SVR-related factors in LVR patients who completed standard 48-week treatment, age (10-year groups) was selected as a significant factor.

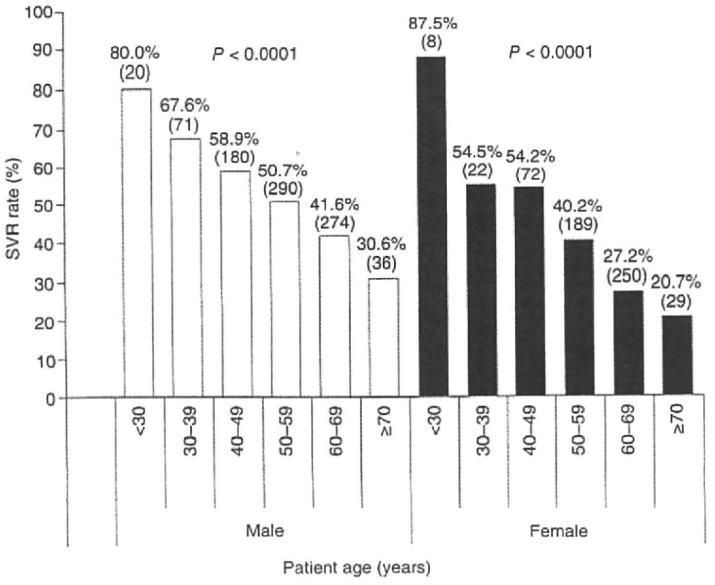

Figure 2 Sustained virological response (SVR) rate to 48 weeks' standard treatment with pegylated interferon α 2b (PEG-IFN α 2b) + ribavirin in male and female patients stratified by age. Cochran–Armitage test was used to study the underlying trend.

Table 2 Independent factors associated with sustained virological response in genotype 1 chronic hepatitis C virus patients who received pegylated interferon α 2b + ribavirin standard therapy for 48 weeks

	Odds ratio	95% confidence interval	*P*-value†
Age <55/≥55 years	0.414	0.293–0.585	<0.0001
Stage 0–1/2–4	0.633	0.442–0.906	0.0124
Platelets <16/≥16 × 10^4/μL	1.876	1.305–2.696	0.0007
Viral load </≥1900 KIU/mL	0.663	0.471–0.935	0.0192

†Multiple logistic regression analysis.

Prolonged treatment was conducted in 73 LVR patients (Fig. 1), with mean duration of 72 weeks. As shown in Table 3, whereas among LVR patients there were significantly ($P = 0.0061$) more female subjects in 72-week group than 48-week group, no intergroup difference of other factors was observed. Overall, SVR rate based on treatment analysis was significantly ($P = 0.0020$) higher in 72-week treatment group than in 48-week treatment group (67.1% [49/73] vs 46.2% [103/223]; Fig. 3A).

When stratified by sex, SVR rate with 48-week and 72-week treatment was 51.4% and 68.6% ($P = 0.0809$) in male subjects and 37.3% and 65.9% ($P = 0.0039$) in female subjects, with SVR in 72-week treatment being significantly higher in female subjects and indicating that, in LVR patients, efficacy comparable to male subjects is achieved in female subjects with 72-week treatment.

In patients aged <55 years SVR rate in the 48- and 72-week treatment groups was 57.6% and 78.9% ($P = 0.1100$) in male subjects and 40.0% and 76.9% ($P = 0.0724$) in female subjects, respectively, with higher SVR rates for the 72-week treatment group (Fig. 3B). In patients aged ≥55 years this parameter was 44.6% and 53.8% ($P = 0.5619$) in male subjects and 37.1% and 60.7% ($P = 0.0425$) in female subjects, respectively, with higher SVR rates for the 72-week treatment group than for the 48-week treatment group as in the case of the younger age group (Fig. 3C).

DISCUSSION

Study 1: SVR-related factors in patients receiving standard 48-week treatment

SVR RATE WITH standard 48-week treatment in this study was 44.9%, roughly equal to the 45% reported in previous clinical trials in Japan.[4,17–19] The present results are also similar to those of clinical trials conducted in patients aged in their mid-40s in western countries and in the general clinical setting.[1–4] Age was

Table 3 Comparison of clinical and virological characteristics between groups receiving pegylated interferon α 2b + ribavirin therapy for 48 and 72 weeks among patients showing late virological response

	48 weeks' group ($n = 223$)	72 weeks' group ($n = 73$)
Sex (male/female)	140/83*	32/41*
Age (years)	58 (21–75)	56 (22–71)
History of HCC (yes/no/unknown)	1/221/11	0/73/0
Previous IFN treatment (yes/no/unknown)	68/113/42	29/32/12
Diabetes (yes/no/unknown)	11/71/141	1/34/38
Hypertension (yes/no/unknown)	18/62/143	6/29/38
Ongoing alcohol use (yes/no/unknown)	17/75/131	6/27/40
Grade (A0/A1/A2/A3/unknown)	2/66/82/6/67	0/21/26/4/22
Stage (F0/F1/F2/F3/F4/unknown)	7/68/45/32/5/66	2/16/20/12/2/21
ALT (IU/L)	61.5 (14–550)	52 (17–254)
Platelets (×10^4/μL)	16.5 (8.5–43.2)	16.6 (4.3–40.2)
Viral load (KIU/mL)	2700 (160–5100)	2100 (130–5000)

Data expressed as median (range). *$P = 0.006$. ALT, alanine aminotransferase; HCC, hepatocellular carcinoma; IFN, interferon.

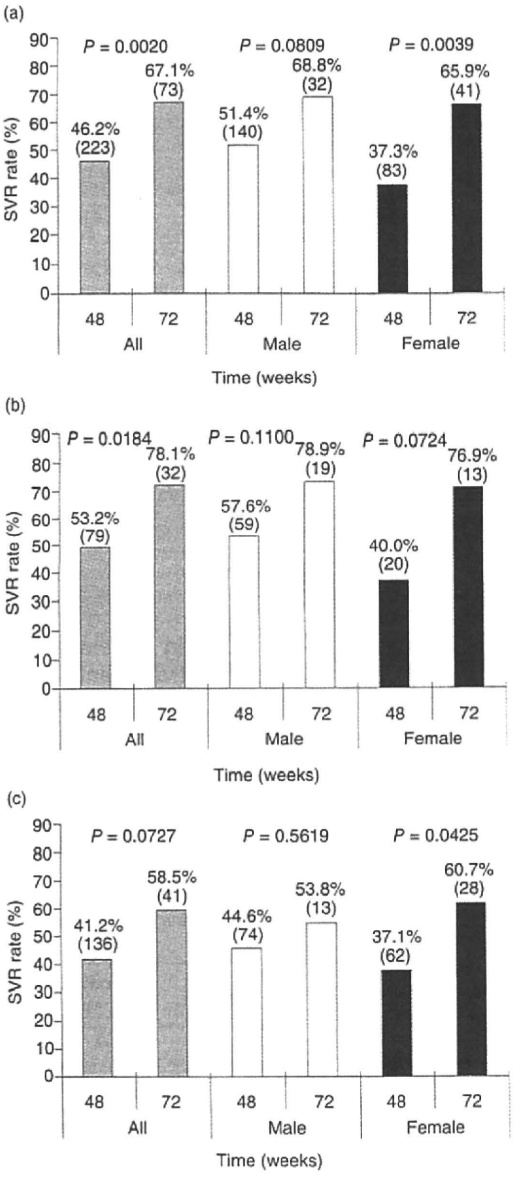

Figure 3 Sustained virological response (SVR) rate based on treatment analysis between groups receiving pegylated interferon α 2b (PEG-IFN α 2b) + ribavirin therapy for 48 and 72 weeks who exhibited late virological response (LVR). (A) Overall; (b) patients aged <55 years; (c) patients aged ≥55 years. Data on age not available for 7 male patients and 1 female patient.

selected among factors for SVR with PEG-IFN plus RBV combination therapy in an aging patient population, the examination of which was the objective of this study, and SVR rate decreased stepwise with 10-year age increase. Of particular note was the greater impact of aging observed in female than male subjects.

Lower efficacy in elderly female patients infected with HCV genotype 1 has already been reported in Japan.[20] A low SVR rate was also observed in elderly female subjects in this study. Although female sex was considered a favorable prognostic factor in some Western studies, there is no established opinion on sex difference. Change associated with aging of the patient population in Japan is considered to account for this phenomenon observed in the present study. This may be due to decrease in compliance among elderly women; on the other hand, however, there was no difference between male and female subjects aged ≥55 years in the rate of completion of treatment. Although the rate of dose reduction of RBV tended to be slightly higher in female subjects (data not shown), the difference was not significant. These findings suggest the influence of factors other than adherence to treatment for the low SVR rate among elderly women. One possible factor for reduced SVR rate among these individuals may be the effect of menopause. In women, insulin resistance begins to worsen after the age of 50 years,[21,22] and this is reported more closely associated with the effect of menopause than age itself.[23]

The presence of insulin resistance has been reported to lower efficacy of PEG-IFN and RBV combination therapy.[24–27] Insulin resistance is also a cause of advanced fibrosis and fatty change of the liver.[28–31] It is possible that such changes combined with other factors associated with metabolic syndrome interact in a complex way to reduce the efficacy of this therapy.[32–35] In fact, the incidence of non-alcoholic fatty liver disease (NAFLD) among elderly Asians was reported higher in women as compared with that in men.[36–38] However, while older age, advanced fibrosis, low platelet count and high HCV load were selected as factors for reduction of SVR rate in our multivariate logistic regression analysis, sex was not selected. It is therefore necessary to examine further the confounding of these selected factors with sex. It also should be taken into consideration that, due to limitations imposed by the retrospective nature of this study, data on factors affecting the efficacy of PEG-IFN plus RBV therapy such as insulin resistance, steatosis, and core mutation are lacking. A large-scale prospective study is

required to examine the lower efficacy observed in elderly women.

Study 2: usefulness of prolonged treatment in LVR patients

EVR (viral load reduced by 2 log or undetected in week 12) has been used for determining continuation or discontinuation of treatment in western countries. Recently, however, EVR was divided into complete EVR (HCV RNA <50 IU/mL at week 12) and partial EVR (>2 log drop in HCV RNA but still detectable [>50 IU/mL]). Fried *et al.*[15] and Berg *et al.*[16] reported that the SVR rate was a high 68–84% in patients showing complete EVR but only 17–29% in those with partial EVR with treatment for 48 weeks. They also reported that treatment for 72 weeks was effective in patients with partial EVR. In the clinical study for health registration in Japan, the SVR rate by timing of HCV-RNA negativity at 4, 12, and 24 weeks was 100%, 71.1%, and 36.4%, respectively, and no patient with HCV-RNA negativity after 25 weeks achieved SVR.[4] With these studies as reference, patients with LVR were defined as those who were positive (>50 IU/mL) at week 12 and became negative (<50 IU/mL) by week 24. To minimize the influence of treatment discontinuation, only patients who completed the standard duration of treatment were selected as subjects in this study. In the comparison of patient background, there was no significant intergroup difference except for a significantly greater number of female subjects in the 72-week treatment group. This finding might be related to the observation that it was already widely believed that efficacy in elderly women in Japan is low and that duration of treatment was at the discretion of individual physicians. Nevertheless, it is noteworthy that the SVR rate was significantly higher in the 72-week treatment group than in the 48-week treatment group and that a high 60% SVR rate was achieved with 72-week treatment in elderly female patients, a population in whom a relatively low SVR was observed with standard 48-week treatment.

This retrospective study had the limitation that duration of treatment was at the sole discretion of each participating physician. A prospective study is necessary to demonstrate whether 72-week treatment in elderly women with LVR is more efficaous than 48-week treatment in male patients. Although the number of younger subjects examined was rather low, it is noteworthy that an SVR rate of >75% was observed with 72-week treatment in both male and female patients. This also should be confirmed by prospective study.

CONCLUSIONS

PATIENTS WITH CHCV genotype 1 infection should be treated with PEG-IFN and ribavirin combination therapy as early as possible. Seventy-two weeks' treatment is recommended in patients with LVR, regardless of age.

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APPENDIX I

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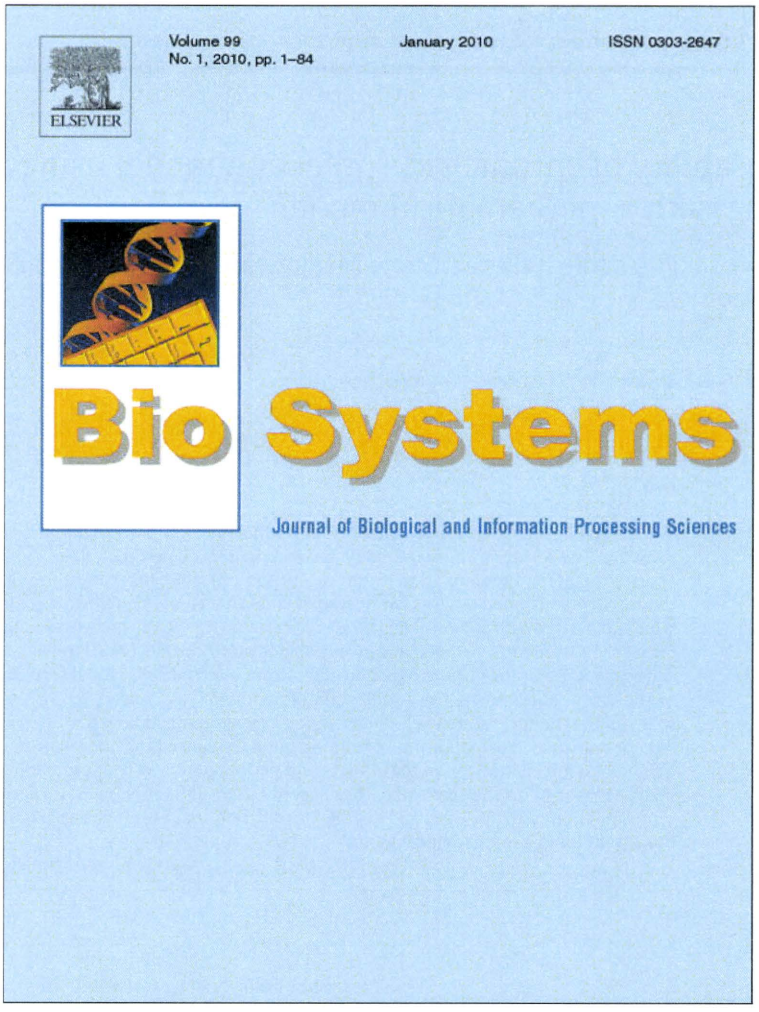

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Reproducibility and usability of chronic virus infection model using agent-based simulation; comparing with a mathematical model

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ABSTRACT

We created agent-based models that visually simulate conditions of chronic viral infections using two software. The results from two models were consistent, when they have same parameters during the actual simulation. The simulation results comprise a transient phase and an equilibrium phase, and unlike the mathematical model, virus count transit smoothly to the equilibrium phase without overshooting which correlates with actual biology in vivo of certain viruses. We investigated the effects caused by varying all the parameters included in concept; increasing virus lifespan, uninfected cell lifespan, uninfected cell regeneration rate, virus production count from infected cells, and infection rate had positive effects to the virus count during the equilibrium period, whereas increasing the latent period, the lifespan-shortening ratio for infected cells, and the cell cycle speed had negative effects. Virus count at the start did not influence the equilibrium conditions, but it influenced the infection development rate. The space size had no intrinsic effect on the equilibrium period, but virus count maximized when the virus moving speed was twice the space size. These agent-based simulation models reproducibly provide a visual representation of the disease, and enable a simulation that encompasses parameters those are difficult to account for in a mathematical model.

1. Introduction

All viruses need hosts as a basis for their life. When a virus enters the host body, it invades cells and uses both its own enzymes and those of the host cells to replicate. Host cells infected by viruses launch a self-defense system known as the innate immune system (See and Wark, 2008; Naniche, 2009), which inhibits viral replication and uses the human leukocyte antigen system and cytokines to elicit an immune response. Immune cells that have received signals from host cells activate other immune cells, neutralize viruses in the serum by means of antibodies, and prevent the virus from replicating and proliferating by destroying or curing host cells. Viral infection is a disorder based on the interactions between viruses and cells.

The power relationship between these agents changes along with the progression of the disease. In the very early stages of infection, as the host defense mechanisms are immature, the virus has the ability to overwhelm the host cells, actively replicate, and proliferate. Subsequently, as the capacity of the immune system improves, the speed of viral proliferation drops and the virus count reaches a peak. Infected host cells begin to be disrupted by the immune system or virus particles, and symptoms appear as a result. If the immune system is stronger than the virus, then the viral counts decline, and, in transient viral disorders, the virus is finally eliminated and the host recovers. In chronic viral disorders, however, the power relationship between the virus and host cells reaches equilibrium, and a long-term power balance is maintained with the virus count reaching a plateau.

Mathematical models have been proposed to study the dynamics of such viral disorders, and are regarded as being of value in understanding this phenomenon (Ho et al., 1995; Nowak et al., 1996; Neumann et al., 1998). However, these models are difficult to understand for clinicians, and their applicability is somewhat limited in everyday practice. In clinical research, measurements of viral dynamics in patients for short duration have been made for human

Abbreviations: HIV, human immunodeficiency virus; HBV, hepatitis B virus; HCV, hepatitis C virus.

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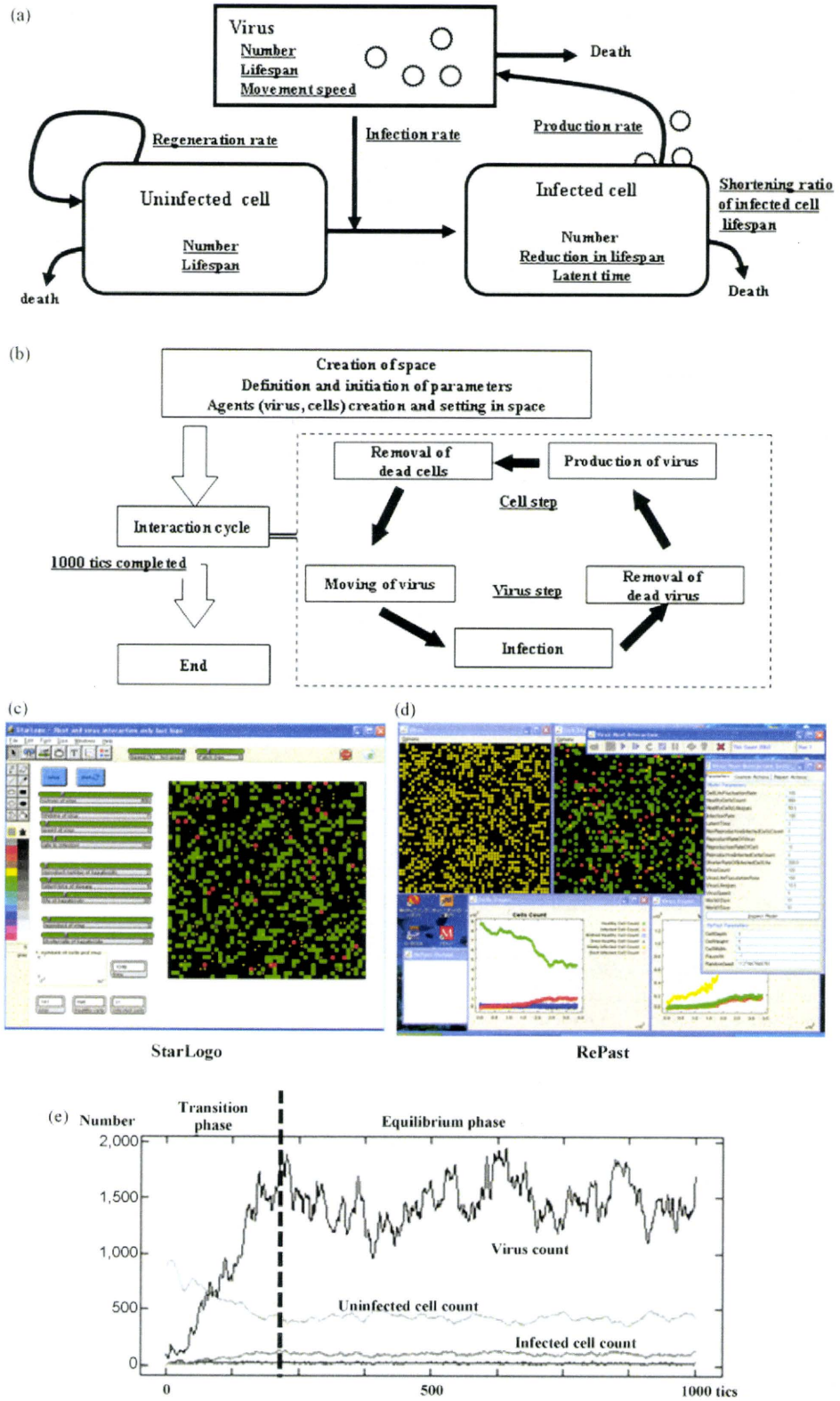

Fig. 1. Simulation design and an example of simulation results. (a) Model concept. Viruses, uninfected cells, and infected cells were treated as agents, and parameters were set for each of these and for interactions between agents (underlined). (b) Flowchart of the program. After preparing the simulation, we entered the interaction cycle, in which virus steps (such as movement) and cell steps were repeated. One cycle was counted as 1 tic, and the simulation concluded after 1000 tics. (c and d) Simulation screen using (c) StarLogo and (d) RePast. Yellow circles are viruses, green squares are uninfected cells, and orange and red indicate infected cells, with orange indicating the latent period. In StarLogo, all the agents are shown on the same screen, but in RePast, viruses and cells are shown in separate windows. (e) Example of a simulation chart in StarLogo. After the start of simulation the virus count and infected cell count increase while the uninfected cell count decreases, with equilibrium state reached after a certain number of tics.

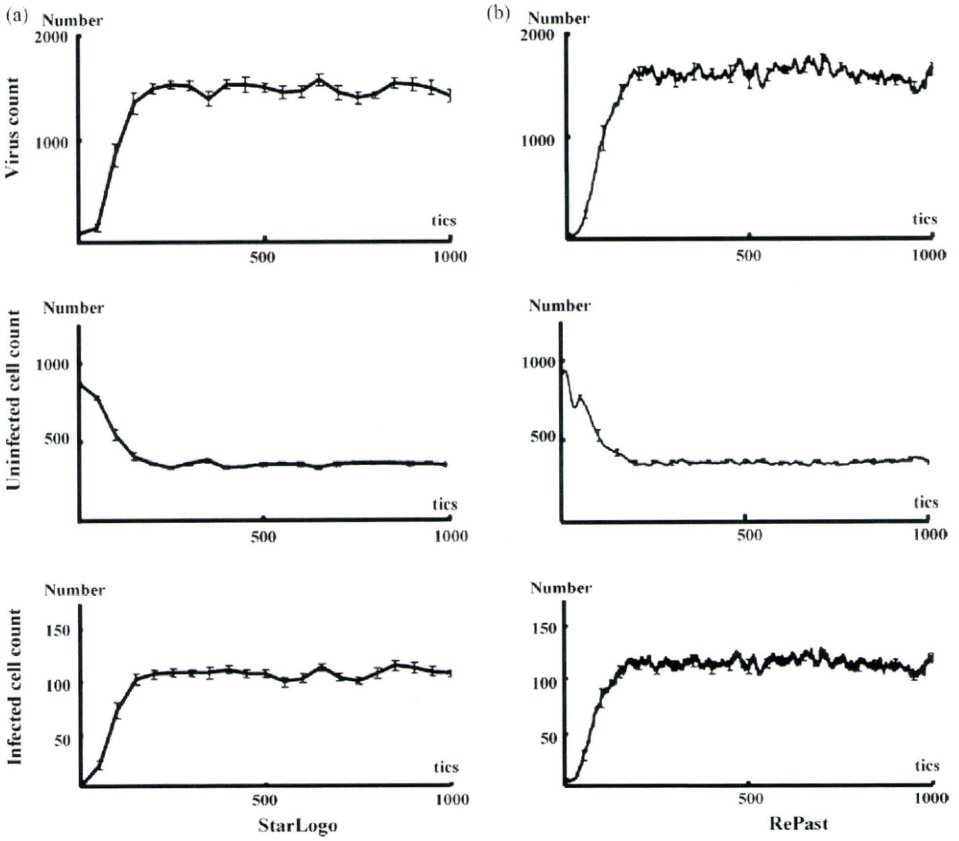

Fig. 2. Comparison of simulation results in (a) StarLogo and (b) RePast. The results were consistent when the parameters were made consistent. (Virus count [average ± SD]: StarLogo 1458.03 ± 173.1, RePast 1462.71 ± 178.8, $p = 0.94$. Uninfected cell count: 364.24 ± 30.4, 368.11 ± 33.4, $p = 0.83$. Infected cell count: 105.73 ± 13.0, 107.74 ± 13.0, $p = 0.24$. Unpaired Student's t-test.) Parameter values were set as follows: initial virus count, 100; uninfected cell count, 880; infected cell count, 0; virus speed of movement, 5 grids/tic; infection rate, 10%; uninfected cell regeneration rate, 1%; latent period, 3 tics; and virus reproduction rate, 5/cells/tic. The following parameter settings were taken from actual measurements: virus lifespan, 4.5 tics; uninfected cell lifespan, 49.8 tics; and infected cell lifespan, 6.7 tics.

immunodeficiency virus (HIV) (Ho et al., 1995), hepatitis B virus (HBV) (Nowak et al., 1996) and hepatitis C virus (HCV) (Neumann et al., 1998), and research is also underway on a range of models based on animal experiments and cell culture systems. As chronic viral disorders persist over long periods of time complete follow-up of viral dynamics is difficult. Furthermore, limitations of items that can be measured, such as the difficulty of measuring whole numbers of host cells, make it extremely difficult to investigate their consistency in mathematical models.

The recent ascend of dynamic-models owes much to advances in computers. Computer performance has improved markedly in recent years, not only in terms of their calculating capacity but also with regard to image displays, and models that offer a visual representation of viral disorders are now being reported (Gilbert and Bankes, 2002; Duca et al., 2007; Shapiro et al., 2008; Castiglione et al., 2007). One advantage of such visual models is that by providing a visual representation, they make understanding the disease status easy. Another benefit is that they enable parameters to be identified that are hidden as background noise in mathematical models. However, these models have some problems; it is difficult to prove the reproducibility of the simulation results derived from different languages or libraries, difficult to prove the validity of the model's concepts, and difficult to prove that the simulation results accurately reflect the reality. In this study, we created agent-based computer models that visually simulate the conditions of chronic viral infections using two software. The reproducibility of two agent-based computer models and the differences between agent-based models and the mathematical model were analyzed. This agent-based model enabled us to investigate how each parameter included in the concept affects the conditions of chronic viral infections.

2. Methods

2.1. Selection of Software

In this study, we used two different types of softwares: StarLogo version 2.0 (http://education.mit.edu/starlogo/) supplied by MIT Media Laboratory and Recursive Porous Agent Simulation Toolkit (RePast-3.0, http://repast.sourceforge.net/) supplied by the Argonne National Laboratory. StarLogo uses Logo, one of the simplest programming languages, and has a fixed graphical user interface. RePast is a library that uses Java, another programming language, which also has a fixed graphical user interface.

Logo is an assembly language, and StarLogo carries out processing sequentially. Java is an object-oriented language, and RePast has a faster processing speed than StarLogo. In addition, StarLogo has a number of stipulations to simplify simulations, such as parameters can only be set up to five decimal places and the simulation space is also fixed as 51 × 51 square grids. RePast, on the other hand, has fewer such restrictions. Thus, it offers a higher degree of freedom in program settings than StarLogo. Taking simulation space as an example, in spite of the restrictions imposed by the underlying operating system's image display system, any number of grids can be set and a hexagonal grid could also be chosen rather than a square one. However, users must stipulate and set all parameters themselves. This means that they must first declare the shape of the grid and the number of grids they will use to fill the simulation space. Java is also more difficult to learn than Logo, and debugging and correcting the program is also more difficult. Thus, it is difficult to judge whether or not the results agree with the planned simulation.

In effect, these two different types of softwares are polar opposites. It is simple to start a simulation in StarLogo, but producing results takes time and it is difficult to carry out more complex simulations. In RePast it is difficult to compose the program and judge whether or not the planned study has actually been achieved, but the

simulation itself takes only a short time to complete and there are lesser restrictions in the construction of a simulation model.

2.2. Concept for Modeling

We applied the basic virus–host interaction mathematical model to the agent-based simulation system with slight modifications. The mathematical model was used to describe the dynamics of HIV (Ho et al., 1995), HBV (Nowak et al., 1996), and HCV (Neumann et al., 1998) and the only agents involved were host cells and viruses, without the inclusion of immune cells. The effects of the immune system are expressed by varying parameters such as lifespan of host cells and viruses.

Fig. 1a illustrates the study concept. Viruses have the ability to infect healthy host cells (uninfected cells) and the infected cells produce new viruses. Both cells and viruses have definite lifespans, and the lifespan of infected cells is usually shorter than that of uninfected cells. Uninfected cells automatically regenerate within the space, whereas infected cells only arise due to infection of uninfected cells. Viruses also lack the ability to regenerate themselves and are only produced from infected cells.

2.3. Parameter Settings

In the present study, as the StarLogo settings are circumscribed, we limited the simulation space to 51 × 51 square grids. However, we made an exception here while investigating the effects of size of space on the simulation results. The numbers of viruses, uninfected cells, and infected cells could only be set before the start of the simulation. As described in the later, our simulation ran in cycles, with 1 cycle defined as 1 tic.

In mathematical simulation models, the death rate is required as a parameter. However, in our program we set lifespans for viruses and uninfected cells. These lifespans were not uniform, but were set to have a deviation of about 10%. The lifespan of cells was shortened by infection with ratio decided beforehand.

The infection ratio was meaningful only when an infected cell and a virus coincidentally occupied the same grid, and this was used to calculate the probability of the infection occurring in that situation. The virus production rate was set as the number of viruses produced by an infected cell during 1 tic. Infected cells could be set as a parameter indicating the latent period between the time of virus infection and the time of virus replication.

In order to emulate the tissue repair capacity, we set uninfected cell regeneration rate such that grids without any cells had a specified probability of producing uninfected cells on top of themselves. As a result, the more the cell count declined within a space the more regenerated uninfected cells were produced, whereas the number of regenerated cells declined as cell count increased.

The number of grids through which a virus could move in 1 tic was set as the speed of movement, and the direction of movement was set within a range of 90° toward the top of the simulation space. The program used a circulatory method of movement; when a virus arrived at the top of the space, it was translocated to the bottom, and moved again toward the top. Cells were fixed on the grid.

2.4. Simulation Flowchart

Fig. 1b shows a flowchart of the program. First, the simulation space was produced, after which each parameter was defined and the initial settings were made. Next the agents – viruses and uninfected and infected cells – were produced. The simulation cycle was as follows. Viruses moved to a new grid, and if an uninfected cell was present, this was infected with a probability based on the infection rate. The lifespan of the virus decreased, and viruses that had completed their lifespan and those that had caused an infection were removed from the space. Infected cells produced new viruses, the lifespans of both uninfected and infected cells decreased. Then, cells that had completed their lifespan were eliminated and a new cycle began. The program was set such that the simulation ended after this cycle had repeated 1000 times. This meant that one simulation was complete after 1000 tics.

2.5. Data Collection

The RePast model was programmed such that data for each tic was saved automatically as a text file at the end of the simulation. This text file could be opened by a database software. The StarLogo model was programmed to stop the simulation and collect data after every 50 tics.

2.6. Mathematical Model

In order to compare the results of this agent-based simulation, we used a viral infection mathematical model, which we improved as follows.

$$\frac{dT}{dt} = s[2601 - (T + I)] - dT - bVT \quad (1)$$

$$\frac{dI}{dt} = bVT - dI \quad (2)$$

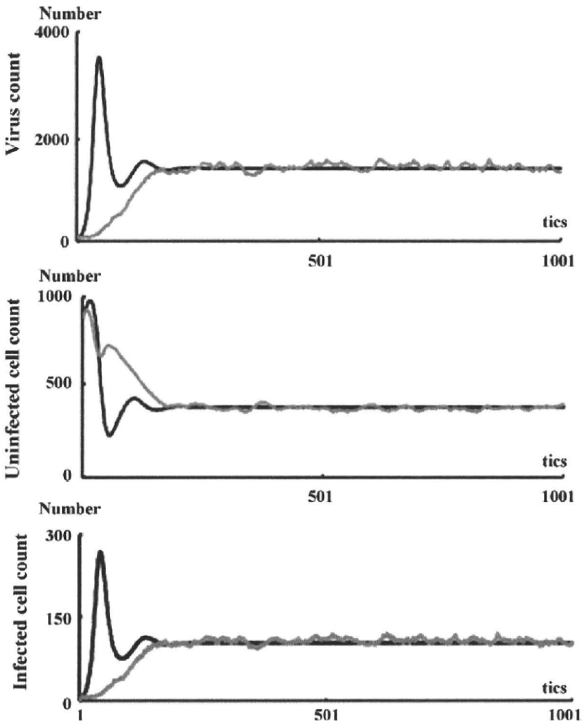

Fig. 3. Comparison of results of agent-based simulation and mathematical simulation. Both sets of results were consistent for the equilibrium phase, but differed in the shift in transition phase. Black line: mathematical model; grey line: results of simulation in RePast. Parameter values were set as follows: initial virus count, 100; uninfected cell count, 880; infected cell count, 0; virus speed of movement, 5 grids/tic; infection rate, 10%; uninfected cell regeneration rate, 1%; latent period, 3 tics; virus reproduction rate, 5/cells/tic; virus lifespan, 10 tics; uninfected cell lifespan, 50 tics; and cell lifespan-shortening ratio as a result of infection, 69%.

$$\frac{dV}{dt} = pI - cV \quad (3)$$

where, T is the uninfected cell count, I is the infected cell count, and V is the virus count. Uninfected cells are supplied to the space with a probability $s[2601 - (T+I)]$, as the number of grids in this agent-based simulation model was 2601 (51 × 51). The death rate of uninfected cells is d, the death rate of infected cells is δ, and the death rate of viruses is c. The infection rate is indicated by β. Viruses are released from infected cells at a probability p.

2.7. Statistical Analysis

Statistical analyses were performed by statistical tests using the program StatView 5.0 (SAS Institute Inc.). All tests of significance were two-tailed, with p values of <0.05 considered to be significant.

3. Results

3.1. Reproducibility of Chronic Viral Infection Disease Models Using Agent-based Simulation Methods

We constructed the chronic viral infection model with StarLogo library. Fig. 1c shows the simulation screen, and Fig. 1e shows one sample result. Immediately after the start of the simulation, the virus count temporarily dropped in accordance with the onset of an infection. Subsequently, the virus count started to increase with an increase in the infected cells and a decrease in the uninfected cells. After a certain number of tics (around 300 in this example), although the virus count, infected cell count, and uninfected cell count had some fluctuation, an equilibrium state was reached. We use the following descriptive terms in this paper: the transient phase is the period during which virus growth peaks, and the equilibrium phase is the period during which an equilibrium state is

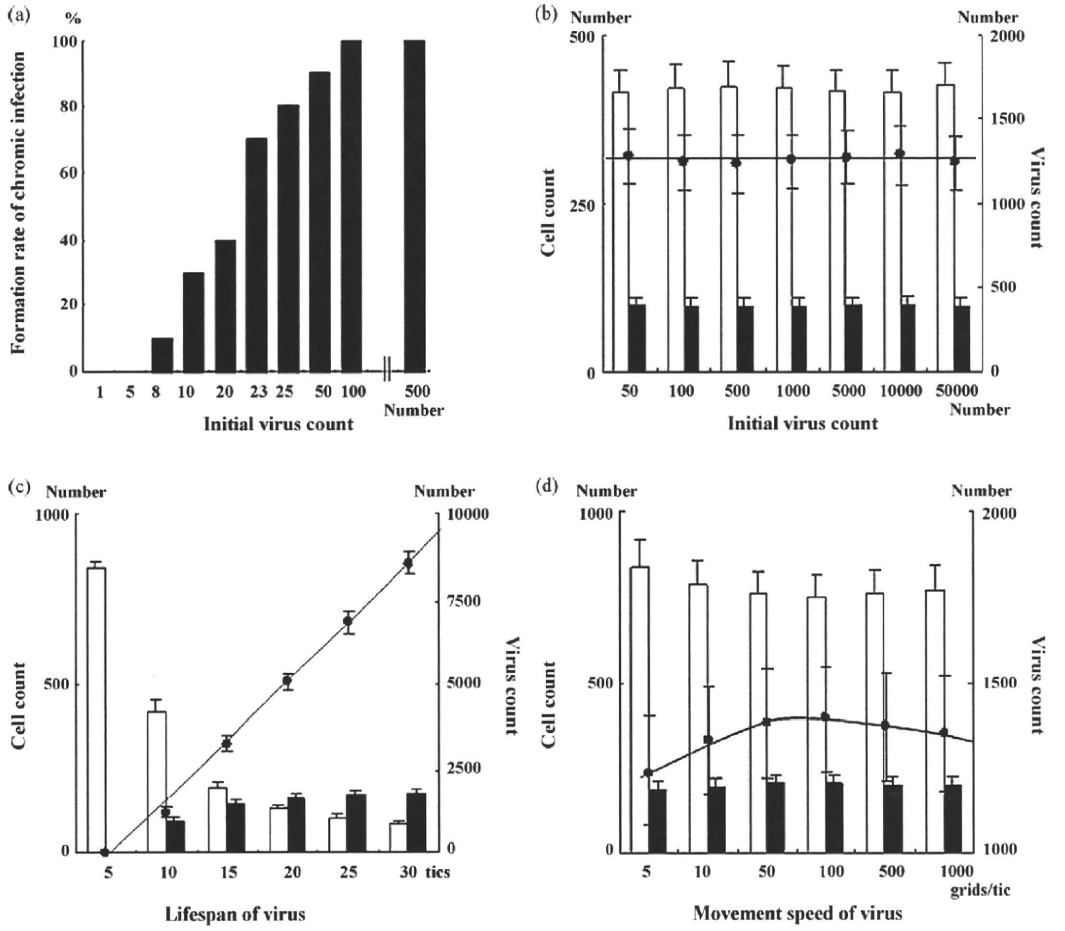

Fig. 4. Effects of changes in viral parameters. (a) The higher the initial virus count, the greater is the increase in the rate of formation of chronic infection, but (b) there was no effect on the conditions in the equilibrium phase. (c) Extending the virus lifespan increased the virus count. (d) Increasing the speed of virus movement to 100 grids/tic increased the virus count, but increasing it to 500 grids/tic had the opposite effect, with a slight declining trend. (a) Black bars: number of infections produced; (b–d) black circles: virus count; line: virus count approximation curve; white bars: uninfected cell count; black bars: infected cell count.

established. When the simulation was performed multiple times, the features described above were maintained, and the average values for virus, infected cell, and uninfected cell counts during the equilibrium state were all consistent.

Fig. 1d shows the simulation screen of the RePast. When we attempted setting all the initial parameters to the same values as those in the StarLogo, the results were not consistent. When we recalculated the parameters from the simulation results, in RePast, the parameters were largely maintained at the levels of the settings, but in StarLogo, the lifespans of both cell types became shorter than the settings while the simulation was in progress. We made the results of both simulations consistent by using the same parameters during the actual simulation (Fig. 2a and b).

3.2. Comparison Between Agent-based Simulation Models and Mathematical Simulation Model

We investigated whether the results of a chronic viral infection disease model produced by RePast would be consistent with the results of a mathematical model. For the mathematical model, we carried out an approximate integration using a four-dimensional Runge–Kutta method to ensure that the uninfected cell count and infected cell count would be in the same class. Parameters were always fixed as constant between simulations. The simulation results were consistent for the equilibrium phase, but transitions in virus count during the transient phase varied, with a shift to equilibrium state following two overshoots in the mathematical model, but a monotonic increase following a logistic curve in the agent-based model (Fig. 3). In the mathematical model, when the equilibrium condition was calculated with $dT/dt = dI/dt = dV/dt = 0$, the equilibrium-phase virus count, uninfected cell count, and infected cell count were very similar to those of the agent-based model (virus count: mathematical model 371.8/space, agent-based model 371.1 ± 32.4/space [average ± SD]; uninfected cell count: mathematical model 1605/space, agent-based model 1454 ± 194/space; infected cell count: mathematical model 115.9/space, agent-based model 108.3 ± 14.2/space).

3.3. Usability of the Models; Effect of Changing Parameters

We investigated the changes in the equilibrium phase brought about by changing each parameter. All the investigations below were carried out by using RePast, and we used the average values from ten simulations.

3.4. Viral Parameters

The lower the virus counts at the beginning of the simulation, the lower the probability of a chronic infection (Fig. 4a). However, the initial virus count did not have any effect on the equilibrium

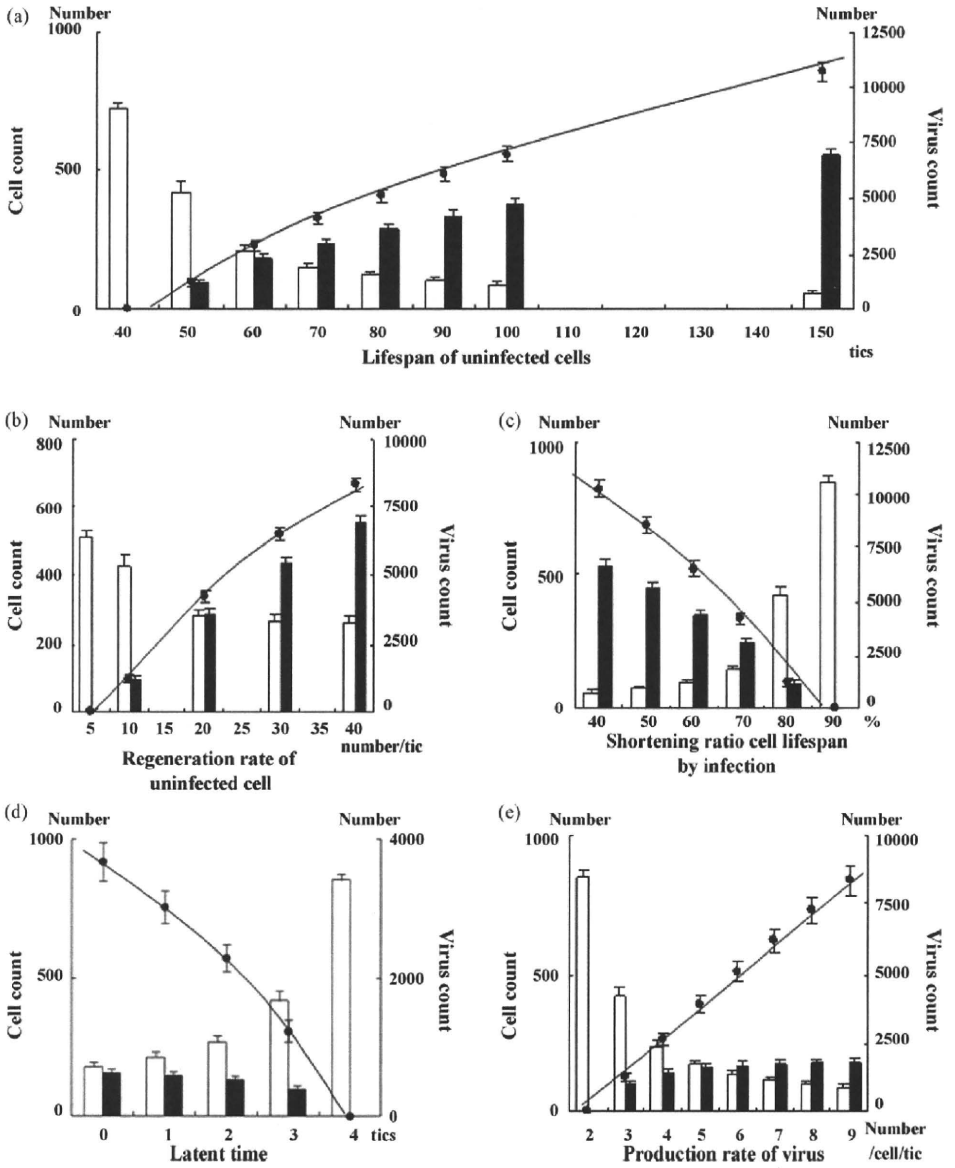

Fig. 5. Effects of changes in cell parameters. (a) Extending the uninfected cell lifespan and (b) increasing the uninfected cell regeneration rate increased the virus count. (c) Raising the lifespan-shortening ratio as a result of infection shortened the lifespan of infected cells, thereby decreasing the virus count. (d) Extending the latent period shortened the period of virus production from infected cells, thereby decreasing the virus count. (e) Increasing the virus production count resulted in a linear increase in equilibrium-phase virus count. Black circles: virus count; line: virus count approximation curve; white bars: uninfected cell count; black bars: infected cell count.

phase itself (Fig. 4b). Extending the lifespan of viruses resulted in a linear increase in equilibrium-phase virus count (Fig. 4c). Although the infected cell count increased, the rate of increase gradually declined. Changing the speed of viral movement resulted in the equilibrium-phase virus count to eventually decline after 100 grids/tic was reached, allowing movement over an area twice the size of the simulation space (Fig. 4d).

3.5. Uninfected Cell Parameters

Extending the lifespan of uninfected cells led to an increased virus count during the equilibrium phase (Fig. 5a). Increasing the uninfected cell regeneration rate also contributed to increased equilibrium-phase virus count (Fig. 5b). In both the cases, the increases in virus count and infected cell count were not linear, but showed a tendency for the rate of increase to decline gradually.

3.6. Infected Cell Parameters

We carried out an investigation of the effects of variation in the lifespan-shortening ratio on the virus count on the assumption that cell lifespan is shortened by infection. When this ratio was increased, the virus count decreased (Fig. 5c). An extended latent period was also related to a decreased virus count (Fig. 5d). However, the virus production from infected cells led to a linear increase in the virus count (Fig. 5e).