TABLE II. Patients Analyzed for Serial Serum Hepatitis A virus

| Patient | Age/sex | Diagnosis | Onset | The last day of HAV (+) | The first day of HAV (-) | Initial viral load (logcopies/ml) |
|---------|---------|---|-------|-------------------------|--------------------------|--------------------------------------|
| 1 | 38/M | Acute hepatitis | 1990 | 27 | 36 | 2.1 |
| 2 | 37/M | Acute hepatitis | 1992 | 21 | , <u> </u> | 4.0 |
| 3 | 33/M | Severe acute hepatitis | 1993 | 11 | 25 | 4.8 |
| 4 | 35/M | Fulminant hepatitis | 1994 | 13 | 33 | 3.3 |
| 5 | 46/M | Severe acute hepatitis | 1994 | 21 | 23 | 2.1 |
| 6 | 29/F | Acute hepatitis | 1995 | 20 | 34 | 2.5 |
| 7 | 41/M | Acute hepatitis | 1995 | 15 | _ | 4.0 |
| 8 | 42/M | Acute hepatitis | 1995 | _ | 62 | _ |
| 9 | 23/M | Acute hepatitis | 1997 | 13 | 19 | 3.2 |
| 10 | 52/F | Fulminant hepatitis | 1999 | 19 | 20 | 4.7 |
| 11 | 43/F | Acute hepatitis | 2000 | 15 | 20 | 5.8 |
| 12 | 60/M | Acute hepatitis | 2000 | 16 | 23 | 3.0 |
| 13 | 54/F | Fulminant hepatitis | 2001 | 7 | 27 | 3.7 |
| 14 | 23/F | Severe acute hepatitis | 2001 | 12 | | 4.6 |
| 15 | 42/M | Acute hepatitis | 2001 | 14 | 19 | 4.3 |
| 16 | 51/M | Acute hepatitis | 2001 | 15 | 21 | 2.9 |
| 17 | 38/M | Acute hepatitis | 2001 | 27 | 34 | 3.1 |
| 18 | 36/M | Acute hepatitis/auto- immune hepatitis | 2001 | 46 | 51 | 2.0 |
| 19 | 60/M | Acute hepatitis | 2001 | 32 | | 5.5 |
| 20 | 61/M | Severe acute hepatitis | 2002 | 15 | | 3.3 |

HAV, hepatitis A virus.

acute hepatitis, and 2.65 ± 1.64 in acute hepatitis. The differences were significant between severe acute hepatitis and acute hepatitis (P < 0.001), and fulminant hepatitis + severe acute hepatitis and acute hepatitis (P < 0.001) (Fig. 1). In fulminant hepatitis and severe acute hepatitis, the HAV RNA levels were 3.21 ± 1.65 in 6 who died and 4.06 ± 0.93 in 15 who recovered (P = 0.28).

Time Course of Viral Loads

The characteristics and time courses of HAV load and ALT levels of patients examined by serial serum samples are shown in Table II and Figures 2 and 3. Viral loads ranged between 2.0 and 5.8 logcopies/ml. The dynamics of viremia were correlated with those of ALT.

In a 52-year-old female patient with fulminant hepatitis (patient 10), viral loads fluctuated and then declined gradually in concordance with the ALT level (Fig. 2A). This might be related to the fact that she was treated with steroid pulse therapy, although the viremia duration was 19 days after onset and it was not prolonged.

A 36-year-old Japanese male (patient 18) presenting with jaundice was admitted to a local hospital in China (Fig. 2B). IgM anti-HAV antibody was positive and he was diagnosed as hepatitis A. After a month of hospitalization, liver function tests improved and he came back to Japan. Two weeks after his return, he was hospitalized with general malaise and itching. Laboratory tests revealed re-elevation of ALT and T-Bil levels (peak T-Bil 8.2 mg/dl), IgG 2,190 mg/dl, ANA < 40 \times , and ASMA < 40 \times . Liver histology showed centrilobular necrosis and marked plasma cell accumulation, leading to a diagnosis of his repeated and prolonged liver injury of acute onset of autoimmune hepatitis triggered by

HAV infection [Fujiwara et al., 2008]. Corticosteroid was introduced and liver function tests normalized within 2 weeks. HAV was detected only on admission, but not at severe exacerbation upon histological examination, suggesting that the severe liver injury was not induced by HAV.

Duration of Viremia

HAV did not disappear during the observation period in 5 (25%; 2 severe acute and 3 acute hepatitis) of 20 patients examined by serial serum samples, with the last time point of HAV detection being 12, 15, 15, 21, and 32 days after onset, respectively. In one of the other 15 patients (patient 8), HAV was not detected at all during follow-up, and HAV disappeared in 14 (70%). Viremia persisted for 18.9 ± 9.6 (7–46) days after clinical onset, similar to the duration of the previous qualitative detection by RT-nested PCR [Fujiwara et al., 1997].

The time courses of the HAV loads of all patients are shown in Figure 3. In the 14 patients in whom HAV disappeared, viremia persisted for 14.2 ± 5.8 days in fulminant hepatitis and severe acute hepatitis, and 21.4 ± 10.6 days in acute hepatitis (P=0.19 by Student's t-test), with the first day of negative HAV being 25.6 ± 4.9 days from onset in fulminant hepatitis and severe acute hepatitis, and 28.6 ± 11.0 in acute hepatitis (P=0.50 by Welch's t-test) (Table II). Therefore, the duration of viremia was not associated with disease severity statistically, although it was relatively longer in cases with non-severe infection than in those with fulminant and severe infections.

DISCUSSION

The reason why the severity of hepatitis A varies among patients is not clear. It has been suggested that

 $J.\ Med.\ Virol.\ DOI\ 10.1002/jmv$

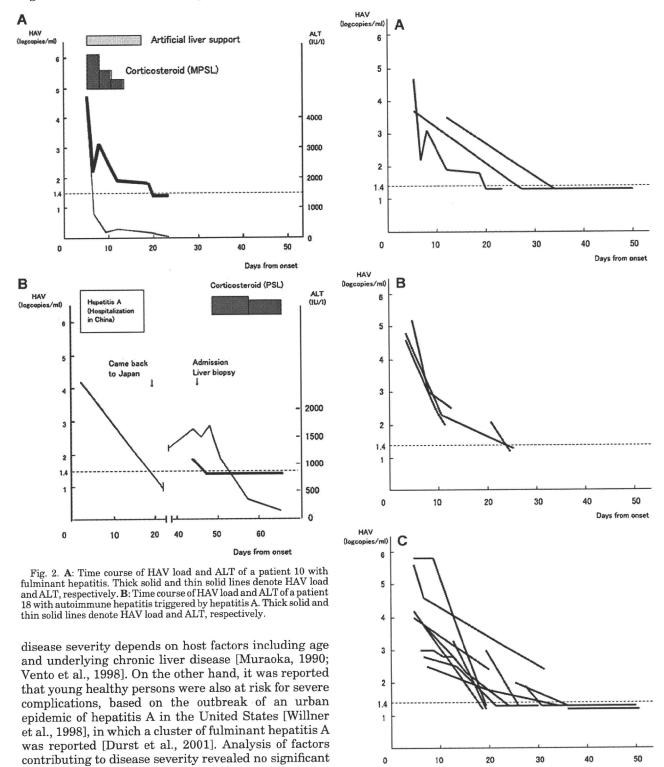


Fig. 3. A: Time courses of HAV loads of three patients with fulminant hepatitis. B: Time courses of HAV loads of four patients with severe acute hepatitis. C: Time courses of HAV loads of 13 patients with acute hepatitis.

Some articles have reported that the pathogenicity of HAV could be related to cooperative mutations within 5'NTR and P2 protein including non-structural protein

differences in patients' factors including age [Fujiwara

et al., 1995, 2000], suggesting that viral factors and the

individual immune responses might determine the

J. Med. Virol. DOI 10.1002/jmv

Days from onset

206 Fujiwara et al.

2B and 2C, based on the study of cultured cells [Zhang et al., 1995] and simians [Raychaudhuri et al., 1998]. However, association between the clinical severity of hepatitis A and HAV genome had not yet been examined. Therefore, to identify differences in HAV for different severities of hepatitis, the HAV genome in the sera from hepatitis A patients with various clinicopathological features was analyzed. Past studies also suggested an association between infection severity and genomic variations in certain parts of the HAV genome including 5'NTR [Fujiwara et al., 2002, 2009], and non-structural protein 2B and 2C [Fujiwara et al., 2007a,b, 2009].

The detection of HAV in clinical samples started after 1990 using the RT-PCR method. Yotsuyanagi et al. [1993] reported that serum HAV RNA was detected only before ALT reached peak levels, as determined by RT-PCR. As ALT levels have already passed their peak in most patients prior to admission, a more sensitive detection method was needed. A technique of nested RT-PCR was previously developed with primers located in 5'NTR for detecting HAV RNA in the majority of patients' sera in the early convalescent phase of hepatitis A [Fujiwara et al., 1997]. In the present study, HAV RNA in serum was quantitatively detectable in almost all patients with hepatitis A in their early convalescent phase by real-time RT-PCR.

It is usually impossible to determine the exact duration of viremia in human HAV infection, as samples can obviously not be readily obtained before clinical onset. Under the special condition of using serum specimens obtained from participants in a clinical trial of hepatitis B vaccine in the United States, Bower et al. [2000] reported that HAV viremia was detected 17 days before ALT peak and persisted for 79 days beyond, as determined by nested RT-PCR. They used three pairs of primers for the VP3-VP1 junction region, the VP1-P2A junction region and 5'NTR, a method more sensitive than ours or those of others. They also demonstrated the presence of high concentrations of HAV during the period before the start of ALT elevation, but the concentrations were low during the convalescent phase: HAV concentrations were $10^4 - 10^5$ plaque-forming units (pfu)/ml before ALT peak, $10^2 - 10^4$ pfu/ml at the peak. 10-100 pfu/ml after the peak, and 1-10 pfu/ml 30 days later.

In a Korean study, using nested RT-PCR with primer sets located at the VP1 region, Kwon et al. [2000] reported a mean duration of HAV viremia of 30 days. In an Italian study, using nested RT-PCR with primers for 5'NTR, Sagnelli et al. [2003] reported that clearance of HAV viremia was observed within 20 days of clinical onset in all patients with a typical self-limiting course. A Dutch study reported that HAV viremia was detected for a median period of 42 days after onset, based on nested RT-PCR with primers at the VP3-VP1 and VP1-P2A regions [Tjon et al., 2006].

Regarding the quantitation of HAV by real-time RT-PCR, several reports have been published. In a French study using real-time RT-PCR quantifying 5'NTR of

HAV, viremia was observed to persist for 60 days after onset [Costa-Mattioli et al., 2002]. Hepatitis A viral loads on admission ranged from 1.8×10^3 to $7.71 \times$ 10^6 copies/ml, with a mean value of 6.38×10^5 copies/ml. Normann et al. [2004] described the time course of HAV viremia and serum viral load in 11 German patients by real-time RT-PCR, reporting long durations of viremia, from 45 days to 490 days after the onset of jaundice, although the viral dynamics of the described patients seemed to be quite different from those in other reports, including ours. Their measured viral loads ranged from 2.0×10^3 to 3.1×10^5 genome equivalents/ml. In an Indian study, six of seven patients with non-severe infection had peak viral loads at admission, whereas three patients with severe infection had peak loads at 15 or 30 days after admission, with the authors suggesting that severe HAV infection may be triggered by diminished cellular immunity [Hussain et al., 2006].

In the current study, initial HAV load was higher in severely infected cases (fulminant hepatitis and severe acute hepatitis) than in cases with non-severe infection, although there was no relation to outcome in the severely infected ones. Interestingly, Rezende et al. [2003] reported that HAV-related liver failure results from an excessive host response associated with a marked reduction in viral load, which represents a distinct discrepancy between their data and ours. As they did not report their schedule of serum sampling, which would provide critical data about viremia in acute hepatitis, a reasonable explanation for this discrepancy cannot be offered. In their study, a higher bilirubin level at admission was associated with fulminant hepatitis A, suggesting an advanced stage of illness of their patients. It is supposed that higher viral replication at onset may induce excessive host responses and severe diseases. and that viral load would be reduced rapidly in the advanced stage of illness as a result of the destruction of large numbers of HAV-infected hepatocytes. In contrast, a longer duration of viremia may be associated with a mild host immune response of non-severe infection, even in the absence of a statistically significant difference.

In summary, HAV RNA was detected quantitatively in the majority of the sera of hepatitis A cases in the early convalescent phase by real-time PCR, and examination of HAV viremia by real-time PCR was useful for analyzing the clinical severity of liver injury during the course of hepatitis A. It might be suggested in part that genetic variations in some regions of HAV including 5'NTR, 2B and 2C might influence cooperatively the replication of the virus and the cellular immune response of the human host, and in large part that vigorous clearance of HAV-infected hepatocytes by the immune response might be the cause of severe disease. Of course, this concept is still at a speculative level and awaits more concrete evidence.

ACKNOWLEDGMENTS

We are indebted to all our colleagues at the liver unit of our hospital who cared for the patients described

J. Med. Virol. DOI 10.1002/jmv

herein. We thank Dr. Makoto Miki (Yokohama-Higashi National Hospital), Professor Kazuyuki Suzuki (Iwate Medical University), Professor Isao Sakaida, Professor Kiwamu Okita (Yamaguchi University), and Professor Eiji Tanaka (Shinshu University) for clinical samples.

REFERENCES

- Bower WA, Nainan OV, Han X, Margolis HS. 2000. Duration of viremia in hepatitis A virus infection. J Infect Dis 182:12–17.
- Costa-Mattioli M, Monpoeho S, Nicand E, Aleman MH, Billaudel S, Ferré V. 2002. Quantification and duration of viraemia during hepatitis A infection as determined by real-time RT-PCR. J Viral Hepat 9:101-106.
- Durst RY, Goldsmidt N, Namestnick J, Safadi R, Ilan Y. 2001. Familial cluster of fulminant hepatitis A infection. J Clin Gastroenterol 32:453-454.
- Fujiwara K, Ehata T, Yokosuka O, Imazeki F, Ohto M, Ohtake Y, Sugai T, Hirasawa H. 1995. The recent increase of severe type A hepatitis in Chiba area. Int Hepatol Commun 3:S37.
- Fujiwara K, Yokosuka O, Ehata T, Imazeki F, Saisho H, Miki M, Omata M. 1997. Frequent detection of hepatitis A viral RNA in serum during the early convalescent phase of acute hepatitis A. Hepatology 26:1634–1639.
- Fujiwara K, Yokosuka O, Ehata T, Imazeki F, Saisho H. 2000. PCR-SSCP analysis of 5' nontranslated region of hepatitis A viral RNA: Comparison with clinicopathological features of hepatitis A. Dig Dis Sci 45:2422–2427.
- Fujiwara K, Yokosuka O, Fukai K, Imazeki F, Saisho H, Omata M. 2001. Analysis of full-length hepatitis A virus genome in sera from patients with fulminant and self-limited acute type A hepatitis. J Hepatol 35:112-119.
- Fujiwara K, Yokosuka O, Ehata T, Saisho H, Saotome N, Suzuki K, Okita K, Kiyosawa K, Omata M. 2002. Association between severity of type A hepatitis and nucleotide variations in the 5' nontranslated region of hepatitis A virus RNA: Strains from fulminant hepatitis have fewer nucleotide substitutions. Gut 51:82–88.
- Fujiwara K, Yokosuka O, Imazeki F, Saisho H, Saotome N, Suzuki K, Okita K, Tanaka E, Omata M. 2003. Analysis of the genotype-determining region of hepatitis A viral RNA in relation to disease severities. Hepatol Res 25:124-134.
- Fujiwara K, Yokosuka O, Imazeki F, Saisho H, Miki M, Omata M. 2005. Do high levels of viral replication contribute to fulminant hepatitis A? Liver Int 25:194–195.
- Fujiwara K, Yokosuka O, Imazeki F, Miki M, Suzuki K, Okita K, Tanaka E, Masao Omata M. 2007a. Genetic analysis of hepatitis A virus protein 2C in sera from patients with fulminant and selflimited hepatitis A. Hepatogastroenterol 54:871–877.
- Fujiwara K, Yokosuka O, Imazeki F, Miki M, Suzuki K, Okita K, Tanaka E, Masao Omata M. 2007b. Analysis of hepatitis A virus protein 2B in sera from various severities of hepatitis A. J Gastroenterol 42:560-566.
- Fujiwara K, Fukuda Y, Yokosuka O. 2008. Precise histological evaluation of liver biopsy specimen is indispensable for diagnosis

- and treatment of acute-onset autoimmune hepatitis. J Gastroenterol 43:951–958.
- Fujiwara K, Kojima H, Yonemistu Y, Yasui S, Imazeki F, Miki M, Suzuki K, Sakaida I, Okita K, Tanaka E, Omata M, Yokosuka O. 2009. Phylogenetic analysis of hepatitis A virus in sera from patients with hepatitis A of various severities. Liver Int 29:838–845.
- Hussain Z, Das BC, Husain SA, Polipalli SK, Ahmed T, Begum N, Medhi S, Verghese A, Raish M, Theamboonlers A, Poovorawan Y, Kar P. 2006. Virological course of hepatitis A virus as determined by real time RT-PCR: Correlation with biochemical, immunological and genotypic profiles. World J Gastroenterol 12:4683-4688.
- Kwon OS, Byun KS, Yeon JE, Park SH, Kim JS, Kim JH, Bak YT, Kim JH, Lee CH. 2000. Detection of hepatitis A viral RNA in sera of patients with acute hepatitis A. J Gastroenterol Hepatol 15:1043–1047.
- Muraoka H. 1990. Clinical and epidemiological study on factors of serious development of viral hepatitis type A. Nippon Shokakibyo Gakkai Zasshi 87:1383–1391.
- Normann A, Jung C, Vallbracht A, Flehmig B. 2004. Time course of hepatitis A viremia and viral load in the blood of human hepatitis A patients. J Med Virol 72:10–16.
- Raychaudhuri G, Govindarajan S, Shapiro M, Purcell RH, Emerson SU. 1998. Utilization of chimeras between human (HM-175) and simian (AGM-27) strains of hepatitis A virus to study the molecular basis of virulence. J Virol 72:7467-7475.
- Rezende G, Roque-Afonso AM, Samuel D, Gigou M, Nicand E, Ferre V, Dussaix E, Bismuth H, Féray C. 2003. Viral and clinical factors associated with the fulminant course of hepatitis A infection. Hepatology 38:613–618.
- Sagnelli E, Coppola N, Marrocco C, Onofrio M, Scarano F, Marotta A, Scolastico C, Catuogno A, Salzillo A, Sagnelli C, Piccinino F, Filippini P. 2003. HAV replication in acute hepatitis with typical and atypical clinical course. J Med Virol 71:1-6.
- Tjon GM, Coutinho RA, van den Hoek A, Esman S, Wijkmans CJ, Hoebe CJ, Wolters B, Swaan C, Geskus RB, Dukers N, Bruisten SM. 2006. High and persistent excretion of hepatitis A virus in immunocompetent patients. J Med Virol 78:1398–1405.
- Vento S, Garofano T, Renzini C, Cainelli F, Casali F, Ghironzi G, Ferraro T, Concia E. 1998. Fulminant hepatitis associated with hepatitis A virus superinfection in patients with chronic hepatitis C. N Engl J Med 338:286-290.
- Willner IR, Uhl MD, Howard SC, Williams EQ, Riely CA, Waters B. 1998. Serious hepatitis A: An analysis of patients hospitalized during an urban epidemic in the United States. Ann Intern Med 128:111-114.
- Yotsuyanagi H, Iino S, Koike K, Yasuda K, Hino K, Kurokawa K. 1993. Duration of viremia in human hepatitis A viral infection as determined by polymerase chain reaction. J Med Virol 40:
- Zhang H, Chao S-F, Ping LH, Grace K, Clarke B, Lemon SM. 1995. An infectious cDNA clone of a cytopathic hepatitis A virus: Genomic regions associated with rapid replication and cytopathic effect. Virology 212:686-697.



doi:10.1111/j.1440-1746.2010.06322.x

HEPATOLOGY

Long-term follow-up of patients with hepatitis B e antigen negative chronic hepatitis B

Dan Bekku,*¹ Makoto Arai,*¹ Fumio Imazeki,* Yutaka Yonemitsu,* Tatsuo Kanda,* Keiichi Fujiwara,* Kenichi Fukai,* Kenichi Sato,[†] Sakae Itoga,[†] Fumio Nomura[†] and Osamu Yokosuka*

Departments of *Medicine and Clinical Oncology and †Department of Molecular Diagnosis, Graduate School of Medicine, Chiba University, Chiba, Japan

Key words

HBe antibody, hepatitis B virus, long-term follow-up.

Accepted for publication 5 March 2010.

Correspondence

Makoto Arai, Department of Medicine and Clinical Oncology (K1), Graduate School of Medicine, Chiba University, Inohana 1-8-1, Chiba 260-8670, Japan. Email: araim-cib@umin.ac.ip

¹These authors contributed equally to this article.

Abstract

Background and Aim: After hepatitis B virus (HBV) e antigen (HBeAg) seroconversion, HBV-DNA continues to replicate, and HBeAg-negative patients still face the risk of liver disease progression. We investigated the predictive factors for alanine aminotransferase (ALT) elevation, antiviral drug use, and hepatocellular carcinoma (HCC) occurrence in HBeAg-negative patients.

Methods: Age, sex, ALT, platelet counts, HBV-DNA levels, genotype, antidiabetic drug use, body mass index, smoking, and alcohol consumption were analyzed for a total of 244 HBV carriers who were HBeAg-negative.

Results: Of 244 HBeAg-negative patients, 158 (64.8%) showed normal ALT levels at baseline. Multivariate Cox hazard regression analysis identified high HBV-DNA levels and high ALT at baseline as independent risk factors for ALT elevation in the patients with normal ALT at baseline. The threshold ALT and HBV-DNA levels were determined to be 31 IU/L and 5.3 logcopies/mL, respectively. Seventeen (7.0%) patients used antiviral drugs. Multivariate Cox hazard regression analysis identified high HBV-DNA levels (threshold, 5.7 log copies/mL), the use of antidiabetic drugs, and daily alcohol consumption at baseline as an independent risk factor for the use of antiviral drugs in HBeAg-negative patients. In 10 patients (4.1%), HCC was detected, and a low platelet count (threshold, 10.0×10^4 /mm³) was associated with the occurrence of HCC.

Conclusion: This study identified predictors of future active liver disease in HBeAgnegative patients, i.e. ALT elevation, unavoidable use of antiviral drugs, and occurrence of HCC.

Introduction

Chronic hepatitis caused by hepatitis B virus (HBV) often follows a fluctuating course characterized by periods of active hepatitis interspersed with quiescence. Therefore, close follow-up is necessary to understand the natural history of HBV patients. On the other hand, patients in which HBV is truly inactive have persistently quiescent disease with an excellent prognosis. Determining an accurate prognosis for HBV carriers based on clinical presentation is important for clinical management of the disease. Various studies have been performed to distinguish the positive and negative prognostic factors among HBV carriers. ¹⁻³

Hepatitis e antigen (HBeAg) seroconversion is an important event in the natural history of HBV infection. HBV-infected patients usually have a very good prognosis after HBeAg seroconversion. Therefore, HBeAg seroconversion has become an important treatment goal during follow-up of HBV carriers. However, it has also been shown that HBV-DNA replication and hepatic inflammation in seroconverted patients continue despite the

persistent loss of HBeAg; thus, HBeAg-negative patients are likely to develop liver cirrhosis or hepatocelluar carcinoma.⁶ In this study, we focused on the natural history of patients with HBeAg-negative chronic hepatitis B, particularly with respect to alanine aminotransferase (ALT) elevation, antiviral drugs, and hepatocellular carcinoma (HCC).

Recently, prognostic factors for HBeAg-negative patients have been investigated in Taiwan and Canada. The We expect to identify a unique constellation of prognostic factors for HBeAg-negative chronic hepatitis B in the Japanese population, due to differences in race and HBV genotype.

Methods

Patients

-199-

Between January 1985 and April 2007, all patients visiting the Chiba University Hospital with HBV infection were approached for participation in the study. This study was carried out only at

one institute, Chiba University Hospital and was approved by ethical the committee of Chiba University. Written informed consent was obtained from all of the patients in accordance with the Declaration of Helsinki. New patients since 1985 and those who were already being followed-up in 1985 were eligible for inclusion in the study. A total of 881 patients were enrolled; of which, 862 were HBsAg positive at enrollment, and 319 were hepatitis B e antibody (HBeAb) positive. Patients who were positive for hepatitis C virus antibody or hepatitis D virus antibody or who had other potential cause of chronic liver diseases (autoimmune hepatitis, primary biliary cirrhosis) were excluded. Patients followed for less than 12 months were also excluded from the analysis. In total, 244 patients were included in the analysis. Serum samples from patients were stored at -20°C and the oldest sample for each patient was used for defining the level of HBV-DNA. The date of evaluation of HBV DNA level by PCR was defined as the baseline. Patient consent was obtained for storage and analysis of serum samples.

Laboratory methods

Serum ALT level was measured using a routine automated method. HBeAg and HBeAb were measured by standard enzyme-linked immunosorbent assays. Patients were screened for hepatitis C virus, hepatitis delta virus, and human immunodeficiency virus antibodies by a third-generation enzyme-linked immunosorbent assay.

HBV-DNA quantitative assay and genotyping

To investigate the level of HBV-DNA in serum, we chose polymerase chain reaction (PCR) assay with an accurate range of 500–200 000 copies/mL (Amplicor HBV monitor test, Roche Diagnostic Systems, Basel, Switzerland). The six major genotypes of HBV (A–F) were determined by enzyme-linked immunosorbent assay (ELISA) (HBV Genotype EIA, Institute of Immunology, Co., Ltd, Tokyo, Japan).

Statistical analysis

ALT elevation was defined as a change from normal ALT (<42 IU/L) to elevated ALT (\geq 42 IU/L), and normalization was defined as a change from elevated ALT to normal from one visit to the next. Baseline data are presented as mean \pm standard deviation (SD). Differences in clinical parameters between groups were analyzed by unpaired *t*-test, Welch *t*-test, and χ^2 tests. The Cox proportional hazards model was used to identify predictive factors for future ALT elevation/normalization, use of antiviral drugs, and HCC occurrence using SPSS version 16.1 software (SPSS Inc., Chicago, IL, USA).

Results

Patient characteristics

To investigate the natural course of HBV carriers with HBeAb, 244 carriers (HBeAg-negative and HBeAb-positive) were enrolled in the study. Follow-up was terminated when the use of

antiviral drugs was started or the occurrence of HCC. The baseline clinical and virological characteristics of the 244 HBeAgnegative carriers are shown in Table 1. Because liver biopsy was performed only in 44 (18.0%) out of 244 patients, liver biopsy results could not be analyzed further. Age, sex, ALT, platelet count (PLT), HBV-DNA level, genotype, antidiabetic drug use, body mass index, smoking, and alcohol consumption were analyzed. The average (± SD) period of follow-up was 103.6 ± 74.8 months. Seventeen (7.0%) patients used antiviral drugs (lamivudine in eight and entecavir in nine) and HCC was detected in 10 (4.1%) patients. Two (0.82%) patients died of HCC. In addition, one died of intrahepatic cholangiocarcinoma, one of liver failure due to gastrointestinal bleeding, and one of tongue cancer during the follow-up period. In Japan, the majority of HBV cases are genotype C and B and these genotypes do not cause HBV carrier by way of horizontal infection in adults; therefore, the HBV infection in our HBV carriers mainly occurred by vertical infection or infection during childhood.9 Thus, the period of HBV infection roughly coincided with the age of HBV carriers in Japan.

ALT and HBV-DNA levels

One hundred and fifty-eight of 244 (64.8%) HBeAg-negative patients had normal ALT levels at baseline. Of these 158 subjects, 85 (53.8%) continued to have normal ALT levels during follow-up, whereas 73 (46.2%) showed fluctuation of ALT levels with intermittently elevated ALT (Fig. 1). A total of 34 (21.5%) patients had ALT \geq 84 IU/L (more than double the normal limit). Of the 86 patients who had elevated ALT levels at baseline, ALT elevation persisted in 10 (11.6%) and 76 (88.4%) showed ALT fluctuations with intermittently elevated ALT. Although HBV-DNA levels were associated with higher ALT levels in general, correlation was weak ($r^2 = 0.13$).

Platelet count

Patients were sub classified based on PLT as follows: (I) < $100\,000$ (II) $100\,000-149\,000$ (III) $150\,000-199\,000$ (IV) $200\,000-249\,000$ (V) > $250\,000$ and more (/mm³). The numbers of patients in groups I, II, III, IV, and V were 17, 28, 73, 68, and 58, respectively. A total of 84 (34.4%) patients reached a lower platelet count at the end of follow-up.

Risk factors for future ALT elevation in patients with normal ALT levels

Although 158 (64.8%) out of 244 HBeAb-positive patients had normal ALT levels at baseline, 73 patients showed fluctuation of ALT levels with intermittently elevated ALT. We investigated the risk factors for future ALT elevation in these patients. The predictive factors of ALT elevation (ALT > 42 IU/L) in patients with normal ALT levels were HBV-DNA and ALT levels at baseline (Table 2). We carried out an additional univariate analysis changing the threshold of HBV DNA from 3.5 to 7.0 log copies/mL in 0.1 log increments and that of ALT from 15 to 41 IU/L in 1.0 increments. We determined the threshold when the value of probability was smallest; the thresholds for ALT and HBV-DNA levels were 31 IU/L and 5.3 logcopies/mL, respectively. The time

Table 1 Baseline characteristics of hepatitis B virus (HBV) e antigen (HBeAg)-negative patients

| | Total | Normal ALT | Elevated ALT | P |
|---|------------------|------------------|------------------|----------|
| Number | 244 | 158 | 86 | |
| Age(years) : (mean ± SD) | 44.1 ± 12.5 | 44.1 ± 13.1 | 44.0 ± 11.4 | NS* |
| <30 | 35 (14.3%) | 24 (15.2%) | 11 (12.8%) | |
| 30–39 | 52 (21.3%) | 32 (20.3%) | 20 (23.2%) | |
| 40–49 | 66 (27.0%) | 44 (27.8%) | 22 (25.6%) | |
| 50- | 91 (37.3%) | 58 (36.7%) | 33 (38.4%) | |
| Sex | | | | <0.001** |
| Male | 141 (57.8%) | 76 (48.1%) | 66 (75.9%) | |
| Female | 103 (42.2%) | 82 (51.9%) | 21 (24.1%) | |
| Alanine aminotransferase (ALT) (IU/L) (mean ± SD) | 58.9 ± 108.1 | 20.9 ± 8.7 | 127.9 ± 160 | <0.001* |
| <20 | 84 | | | |
| 21–30 | 47 | | | |
| 31–40 | 27 | | | |
| 42–84 | 47 | | | |
| 85– | 39 | | | |
| Platelet count (x104/mm³) (mean ± SD) | 205.5 ± 69.6 | 211.4 ± 60 | 193.3 ± 81.8 | NS* |
| HBV-DNA (log copies/mL) (mean ± SD) | 4.3 ± 1.5 | 3.8 ± 1.1 | 5.1 ± 1.7 | <0.001* |
| <4.0 | 116 (47.5%) | 91 (57.6%) | 25 (29.1%) | |
| 4.0-4.9 | 54 (22.1%) | 38 (24.1%) | 16 (18.6%) | |
| 5.0-5.9 | 27 (11.1%) | 18 (11.4%) | 9 (10.5%) | |
| 6.0–6.9 | 26 (10.7%) | 5 (3.2%) | 21 (24.4%) | |
| 7.0– | 16 (6.6%) | 3 (1.9%) | 13 (15.1%) | |
| Genotype | | | | NS** |
| A | 3 (1.2%) | 2 (1.3%) | 1 (1.2%) | |
| В | 30 (12.3%) | 16 (10.1%) | 14 (16.3%) | |
| C | 87 (35.7%) | 49 (31%) | 38 (44.2%) | |
| Not detected | 124 (50.8%) | 91 (57.6%) | 33 (38.4%) | |
| Liver Histology ($n = 44$) | | | | |
| Fibrosis 4/3/2/1 | 7/8/9/20 | 0/1/4/13 | 7/7/5/7 | NS** |
| Activity 3/2/1 | 7/16/21 | 1/4/13 | 6/12/8 | NS** |
| Use of anti-Diabetes drug | 20 (8.2%) | 3 (1.9%) | 6 (7.0%) | NS** |
| Body mass index (kg/m 2) (mean \pm SD) | 23.3 ± 3.3 | 23.1 ± 3.2 | 24.0 ± 3.5 | NS** |
| Smoker/ ever smoker/ non-smoker | 32/15/89 | 16/5/56 | 16/10/33 | NS** |
| Daily alcohol consumption | 46 (27.1%) | 24 (23.1%) | 22 (33.3%) | NS** |
| Follow-up (months) (mean ± SD) | 103.6 ± 74.8 | 109.5 ± 76.1 | 101.8 ± 74.6 | NS* |

^{*}Unpaired t-test and ** χ^2 test. NS, not significant difference.

interval from a visit with a normal ALT to a visit with an elevated ALT was used for Kaplan-Meier and Cox regression analysis. Kaplan-Meier curves were constructed for ALT and HBV-DNA levels (Fig. 2).

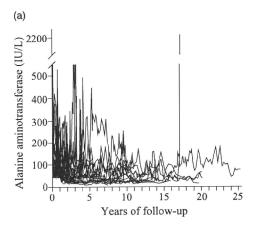
Risk factors for future use of antiviral drugs for HBV in HBeAg-negative patients

Seventeen (7.0%) patients used an antiviral drug (lamivudine in 8 and entecavir in 9). We investigated the risk factors for future use of antiviral drugs for HBV. The time interval from baseline to the use of an antiviral drug for HBV was used for Cox regression analysis. HBV-DNA levels, use of antidiabetic drugs, and daily alcohol consumption were predictive of future antiviral drug use for HBV, according to the results of multivariate Cox hazard regression analysis. Hazard ratios for HBV-DNA levels, antidiabetic drug use, and daily alcohol consumption were 1.519 (1.130–2.042, 95% confidence interval [CI]), 3.769 (1.203–11.81), and 3.011 (1.086–8.348), respectively. We repeated the univariate

analysis, changing the threshold for HBV DNA from 3.5 to 7.0 log copies/mL in 0.1 log increments. We determined the threshold when the probability value was lowest; the HBV-DNA threshold level was 5.7 log copies/mL. Kaplan–Meier curves were constructed for HBV-DNA levels, antidiabetic drug use, and daily alcohol consumption (Fig. 3).

Risk factors for hepatocellular carcinoma in HBeAg-negative patients

In 10 patients (4.1%), HCC was detected. We investigated the risk factors for HCC in HBeAg-negative patients. The time interval from baseline to occurrence of HCC was used for Cox regression analysis. According to the results of multivariate Cox regression analysis, PLT was predictive of the development of HCC. The hazard ratio for PLT was 0.807 (0.724–0.899, 95% CI). We performed univariate analyses, changing the PLT threshold from 8.0 to 30.0×10^4 /mm³ in 1.0×10^4 /mm³ increments. We determined the threshold when the value of probability was smallest; the



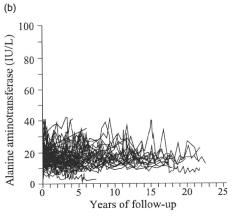
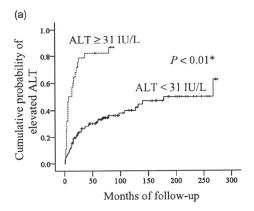


Figure 1 Level of alanine aminotransferase (ALT) in (a) patients with normal ALT at baseline and intermittently elevated ALT during follow-up (n = 73) and (b) patients with normal ALT at baseline and during follow-up (n = 85).



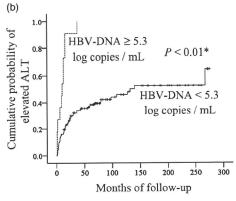


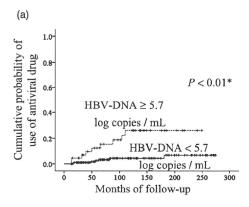
Figure 2 Cumulative occurrence of abnormal alanine aminotransferase (ALT) in HBeAg-negative patients with normal ALT based on (a) ALT and (b) HBV-DNA levels. We determined the threshold for ALT and HBV-DNA levels when the probability value was lowest in the univariate analysis. Kaplan–Meier curves show the time to ALT elevation. Solid lines indicated the control group. *A significant difference was determined by log-rank test.

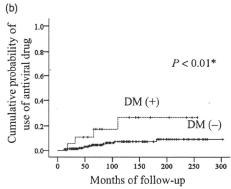
 Table 2
 Univariate and multivariate analysis of factors associated with alanine aminotransferase (ALT) elevation in hepatitis B virus (HBV) e antigen (HBeAg)-negative patients with normal ALT levels

| | | Univ | variate analy | /sis | | Mult | ivariate ana | lysis |
|---------------------------|-------------------|-------------------|-----------------|--|-------------------|-------------------|--------------|--|
| | Standard error | Wald statistic | <i>P</i> -value | Hazard ratio (95% confidence interval) | Standard error | Wald statistic | P-value | Hazard ratio (95% confidence interval) |
| Sex (Male) | 0.263 | 0.203 | 0.652 | 1.126 (0.673–1.885) | | | | |
| Age (years) | 0.011 | 5.704 | 0.017 | 1.027 (1.005-1.049) | 0.252 | 0.068 | 0.794 | 1.015 (0.572-1.534) |
| HBV-DNA | 0.109 | 17.773 | < 0.001 | 1.587 (1.280-1.966) | 0.111 | 10.602 | 0.001 | 1.437 (1.155–1.788) |
| Genotype | | | | | | | | |
| В | 0.459 | 0.22 | 0.639 | 0.806 (0.328-1.982) | | | | |
| С | 0.435 | 0.055 | 0.815 | 1.107 (0.472-2.600) | | | | |
| Alanine aminotransferase | 0.014 | 42.440 | < 0.001 | 1.097 (1.067-1.128) | 0.015 | 29.496 | < 0.001 | 1.086 (1.054–1.119) |
| Platelet count | 0.019 | 5.928 | 0.015 | 0.955 (0.920-0.991) | 0.021 | 0.754 | 0.385 | 0.982 (0.942-1.023) |
| Use of anti-diabetes drug | 0.427 | 0.470 | 0.493 | 1.340 (0.581-3.091) | | | | |
| Body mass index (kg/m²) | 0.042 | 0.033 | 0.855 | 0.992 (0.913-1.078) | | | | |
| Smoker and ever smoker | 0.374 | 0.111 | 0.739 | 1.133 (0.544-2.359) | | | | |
| Daily alcohol consumption | 0.333 | 0.512 | 0.474 | 1.269 (0.661–2.435) | | | | |

Journal of Gastroenterology and Hepatology 26 (2011) 122-128

© 2010 Journal of Gastroenterology and Hepatology Foundation and Blackwell Publishing Asia Pty Ltd





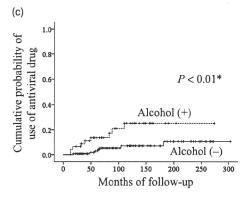


Figure 3 Cumulative occurrence of antiviral drug use for hepatitis B virus (HBV) in HBeAg-negative patients based on (a) HBV-DNA levels (b) use of antidiabetic drug, and (c) daily alcohol consumption. We determined the threshold for HBV-DNA levels when the probability value was lowest in the univariate analysis. Kaplan–Meier curves show the time to use of antiviral drugs for HBV. Solid lines indicated the control group. *A significant difference was determined by log-rank test.

PLT threshold was 10.0×10^4 /mm³. Kaplan–Meier curves were constructed for PLT (Fig. 4).

Stratification analyses of risk factors for clinical outcomes in HBeAg-negative patients by age, sex, and HBV genotype

The stratification analyses by age, sex, and HBV-genotype were performed to evaluate the risk factors for future ALT elevation in

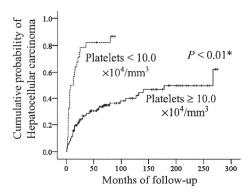


Figure 4 Cumulative occurrence of hepatocellular carcinoma (HCC) based on the platelet counts. We determined the threshold for HBV-DNA levels when the probability value was lowest in the univariate analysis. Kaplan–Meier curves show the time to HCC. Solid lines indicated the control group. *A significant difference was observed by log-rank test.

patients with normal ALT levels, future use of antiviral drugs for HBV, and HCC in HBeAg-negative patients (Table 3). The age threshold was 45 years, which was the average age of all the patients. We did not perform stratification analysis for patients infected with HBV genotype B because the number of such cases was very small.

Discussion

Most patients who have undergone HBeAg seroconversion have normal serum ALT levels, which is indicative of a good clinical outcome. 10 Therefore, various therapies for early seroconversion have been used.⁵ Recently, HBeAg-negative viral mutants have been shown to be responsible for continuous HBV-DNA replication.⁷ That is, there exists the possibility that liver disease will get worse after HBeAg seroconversion. In fact, previous reports revealed that HBeAg status is not a predictive factor for HCC, 11,12 and fulminant hepatitis can occur by the infection of HBV with HBeAg-negative. 13 HBeAg-negative patients should be monitored closely, even though most of these patients show normal ALT levels and no progressive liver disease.14 Therefore, predictive factors for active liver disease in HBeAg-negative patients need to be identified in order to facilitate optimal disease management. This study provides data regarding the prediction of future active liver disease, i.e. ALT elevation, unavoidable use of antiviral drugs, and occurrence of HCC.

Many previous reports have attempted to define a threshold HBV-DNA level that corresponds to the presence of active liver disease. ¹⁵ A National Institute of Health workshop demonstrated that an HBV-DNA level of 10⁵ copies/mL could be used to distinguish active HBV infection from inactive HBV infection. ¹⁶ Other studies also suggested that the threshold HBV-DNA level lies somewhere between 10⁴ and 10⁶ copies/mL. ⁸ In this study, in order to clarify the natural course of HBeAg-negative patients with normal ALT levels, we used a HBV-DNA threshold of 10^{5.3} copies/mL. By log rank analysis, the ALT levels in patients with >10^{5.3} copies/mL HBV-DNA level were significantly higher than in patients with HBV-DNA below this level. In HCV patients, ALT is

Table 3 Stratification analysis multivariate analysis of factors associated with alanine aminotransferase (ALT) elevation in hepatitis B virus (HBV) e antigen (HBeAg)-negative patients with normal ALT levels, future use of antiviral drugs for HBV, and occurrence of hepatocellular carcinoma

| ≥4 n Factors Ha: | | 11 080 | Age (years) | | | | | ñ | Sex | | | | denotype | |
|---|------------------------|-----------------|---------------|--------------------------|-----------------|--------------------------------|--------------------------|-----------------|-----------------|---|-----------------|---------|--------------------------|---------|
| | ≥45 years | | | <45 years | | | Male | | | Female | | | U | |
| | n=126 | | | n=118 | | | n = 141 | | | n = 103 | | | n = 87 | |
| | Hazard ratio (95% CI) | P-value Factors | | Hazard ratio (95% CI) | P-value Factors | Factors | Hazard ratio (95% CI) | P-value Factors | Factors | Hazard ratio (95% CI) | P-value Factors | Factors | Hazard ratio (95% CI) | P-value |
| Future ALT elevation HBV-DNA 1.535 | 1.535 | 0.004 ALT | ALT | 1.106 | <0.001 | ALT | 1.194 | <0.001 | ALT | 1.060 (1.015–1.108) | 0.008 | ALT | 1.149 (1.075–1.228) | <0.001 |
| ALT | | <0.001 | | | | | * | | HBV-DNA | 1.739 (1.213–2.492) | 0.003 | HBV-DNA | 1.902 (1.223–2.956) | 0.004 |
| Future use of Alcohol 4.7 anti-viral drugs (1.3 for HBV | 4.744 (1.362–16.52) | 0.014 | 0.014 HBV-DNA | 2.238 (1.107–4.526) | 0.025 | 0.025 HBV-DNA 1.486 (1.053- | 1.486 (1.053–2.098) | 0.024 | DM ⁺ | 86.14 (1.842–4027.3) 0.408 (0.187–0.890) | 0.023 | Alcohol | 5.617 (1.431–22.05) | 0.013 |
| Occurrence of PLT 0.7 hepatocellular (0.1 carcinoma | 0.772 (0.659–0.905) | 0.001 | <u></u> | | | PLT | 0.832 (0.731–0.948) | 0.006 | PLT⁺ | 0.775 (0.635–0.947) | 0.013 | PLT | 0.833 | 900.00 |

use of antidiabetic medication; PLT, platelet count confidence interval; DM, antiviral drugs and HCC occurrence Alcohol, Daily alcohol consumption; BMI, Body mass index; Cl, patients were used in subgroup for HBV

a poor surrogate marker for inflammation and fibrosis.¹⁷ Therefore, even if the patient's ALT level was within normal limits, they should still be monitored closely, and HCV eradication therapy is recommended under certain circumstances. Similarly, even if the ALT levels are within normal limits in HBV-infected patients who are HBeAg-negative, the higher their ALT levels were, the more frequently their ALT levels would be high in the future, which might cause progressive liver disease.18

Some of the patients with progressive liver disease caused by HBV infection were treated with the antiviral drugs lamivudine and entecavir. The use of lamivudine or entecavir might result in mutant HBV resistance to antiviral drugs19,20 and the associated costs are not trivial. The baseline levels of HBeAg, ALT, and HBV-DNA, and the presence of either chronic hepatitis or cirrhosis have been established as determinants for eligibility for antiviral treatment.21 According to treatment guidelines in the Unites States (National Guideline Clearinghouse, http://www.guideline. gov), patients with HBeAg-negative chronic hepatitis B should be considered for antiviral treatment based on their HBV-DNA and ALT levels (serum HBV-DNA >20 000 IU/mL and elevated ALT >2 times normal). In this study, only four out of 17 patients treated with an antiviral drug showed normal ALT levels at baseline, and all four patients showed elevated ALT levels in 8-57 months later. Therefore, this study revealed that patients with high HBV-DNA levels tended to have high ALT levels at baseline or in the future; as a result, such patients have a tendency for future treatment with and antiviral drug.

Hepatocellular carcinoma occurrence was noted in only 10 cases (4.1%). The only predictive factor for HCC occurrence was PLT, which meant that patients with advanced liver disease tended to develop HCC later, because the decrease in PLT corresponded to the extent of liver fibrosis. Four patients (1.6%) died of liverrelated diseases and one (0.4%) died of cancer in another organ. The number of deaths was too small to determine the predictive factors for death of HBeAg-negative HBV carriers. Further analysis is needed to properly address this factor.

Stratification analyses of risk factors for clinical outcomes by age, sex, and HBV-genotype were performed. Because the numbers of female patients with future use of antiviral drugs for HBV (n = 3), HCC occurrence (n = 3), or who were under 45 years old with HCC occurrence (n = 3) were very small, it was not possible to properly evaluate these subgroups. The risk factors among subgroups for future ALT elevation in patients with normal ALT levels, and for HCC were almost equal to those of the entire patient population. However, daily alcohol consumption, not HBV-DNA level, was predictive of future use of antiviral drugs for HBV in patients ≥45 years old or in patients infected with HBV genotype C. In these subgroups, alcohol consumption was an important factor for predicting the clinical course of HBV carriers; i.e. advising patients to abstain from drinking might reduce the need for antiviral drugs in the future.

Coffee or caffeine consumption is reported to be strongly related to ALT levels and HCC occurrence. 22-24 In our study, we did not survey caffeine consumption; therefore, further analysis is needed to determine the importance of coffee or caffeine consumption as a predictive factor of the clinical course in HBeAgnegative HBV carriers.

In conclusion, we established that low HBV-DNA levels and ALT levels at baseline were good predictors for future ALT elevation in HBeAg-negative HBV carriers with normal ALT levels. In addition, this study provides data on the prediction of unavoidable antiviral drug use and HCC occurrence.

References

- 1 Fattovich G, Bortolotti F, Donato F. Natural history of chronic hepatitis B: special emphasis on disease progression and prognostic factors. J. Hepatol. 2008; 48: 335-52.
- 2 Di Marco V, Lo Iacono O, Camma C et al. The long-term course of chronic hepatitis B. Hepatology 1999; 30: 257-64.
- 3 Fattovich G, Olivari N, Pasino M et al. Long-term outcome of chronic hepatitis B in Caucasian patients: mortality after 25 years. Gut 2008: 57: 84-90.
- 4 Hoofnagle JH, Dusheiko GM, Seeff LB et al. Seroconversion from hepatitis B e antigen to antibody in chronic type B hepatitis. Ann. Intern. Med. 1981; 94: 744–8.
- 5 Korenman J, Baker B, Waggoner J et al. Long-term remission of chronic hepatitis B after alpha-interferon therapy. Ann. Intern. Med. 1991; 114: 629–34.
- 6 Sumi H, Yokosuka O, Seki N et al. Influence of hepatitis B virus genotypes on the progression of chronic type B liver disease. Hepatology 2003; 37: 19–26.
- 7 Lin CL, Liao LY, Liu CJ et al. Hepatitis B viral factors in HBeAg-negative carriers with persistently normal serum alanine aminotransferase levels. Hepatology 2007; 45: 1193–8.
- 8 Feld JJ, Ayers M, El-Ashry D et al. Hepatitis B virus DNA prediction rules for hepatitis B e antigen-negative chronic hepatitis B. Hepatology 2007; 46: 1057–70.
- 9 Orito E, Ichida T, Sakugawa H et al. Geographic distribution of hepatitis B virus (HBV) genotype in patients with chronic HBV infection in Japan. Hepatology 2001; 34: 590–4.
- 10 Lai CL, Yuen MF. The natural history of chronic hepatitis B. *J. Viral. Hepat.* 2007; **14** (Suppl. 1): 6–10.
- 11 Murata K, Sugimoto K, Shiraki K et al. Relative predictive factors for hepatocellular carcinoma after HBeAg seroconversion in HBV infection. World J. Gastroenterol. 2005; 11: 6848–52.
- 12 Pokorski RJ, Ohlmer U. Long-term morbidity and mortality in Chinese insurance applicants infected with the hepatitis B virus. J. Insur. Med. 2001; 33: 143–64.

- 13 Fujiwara K, Yokosuka O, Ehata T et al. The two different states of hepatitis B virus DNA in asymptomatic carriers: HBe-antigenpositive versus anti-HBe-positive asymptomatic carriers. Dig. Dis. Sci. 1998; 43: 368–76.
- 14 Kumar M, Sarin SK, Hissar S et al. Virologic and histologic features of chronic hepatitis B virus-infected asymptomatic patients with persistently normal ALT. Gastroenterology 2008; 134: 1376–84.
- 15 Mels GC, Bellati G, Leandro G et al. Fluctuations in viremia, aminotransferases and IgM antibody to hepatitis B core antigen in chronic hepatitis B patients with disease exacerbations. Liver 1994; 14: 175-81.
- 16 Hoofnagle JH, Doo E, Liang TJ et al. Management of hepatitis B: summary of a clinical research workshop. Hepatology 2007; 45: 1056–75.
- 17 Sanai FM, Benmousa A, Al-Hussaini H et al. Is serum alanine transaminase level a reliable marker of histological disease in chronic hepatitis C infection? Liver Int. 2008; 28: 1011–18.
- 18 Lai M, Hyatt BJ, Nasser I et al. The clinical significance of persistently normal ALT in chronic hepatitis B infection. J. Hepatol. 2007; 47: 760–7.
- 19 Villet S, Ollivet A, Pichoud C et al. Stepwise process for the development of entecavir resistance in a chronic hepatitis B virus infected patient. J. Hepatol. 2007; 46: 531–8.
- 20 Sa-nguanmoo P, Tangkijvanich P, Payungporn S et al. Dynamics of HBV DNA levels, HBV mutations and biochemical parameters during antiviral therapy in a patient with HBeAg-negative chronic hepatitis B. Asian Pac. J. Allergy Immunol. 2007; 25: 183–8.
- 21 Buster EH, van Erpecum KJ, Schalm SW et al. Treatment of chronic hepatitis B virus infection—Dutch national guidelines. Neth. J. Med. 2008; 66: 292–306.
- 22 Ruhl CE, Everhart JE. Coffee and caffeine consumption reduce the risk of elevated serum alanine aminotransferase activity in the United States. *Gastroenterology* 2005; 128: 24–32.
- 23 Ruhl CE, Everhart JE. Coffee and tea consumption are associated with a lower incidence of chronic liver disease in the United States. Gastroenterology 2005; 129: 1928–36.
- 24 Larsson SC, Wolk A. Coffee consumption and risk of liver cancer: a metaanalysis. *Gastroenterology* 2007; 132: 1740–5.

PMID: 20357731



2009.06.03 Received: 2009.12.11 Accepted: Published: 2010.04.01

Aminofeel® improves the sensitivity to taste in patients with HCV-infected liver disease

Authors' Contribution:

- A Study Design
- **B** Data Collection
- C Statistical Analysis
- **D** Data Interpretation
- E Manuscript Preparation
- F Literature Search
- **G** Funds Collection

Yumiko Nagao¹⁰⁰⁰⁰⁰¹, Hisako Matsuoka¹⁸, Takumi Kawaguchi¹⁸¹, Michio Sata^{1,2MEDEG}

¹ Department of Digestive Disease Information & Research, Kurume University School of Medicine, Kurume, Japan

² Division of Gastroenterology, Department of Medicine, Kurume University School of Medicine, Kurume, Japan

Source of support: This study was supported in part by Health and Labour Sciences Research Grants for Research on Hepatitis from the Ministry of Health, Labour and Welfare of Japan

Summary

Background:

Patients with chronic liver diseases have a taste disorder and altered zinc metabolism. We investigated the effects of a supplement enriched with branched-chain amino acids (BCAA) (Aminofeel®) on sensitivity to different tastes in patients with hepatitis C virus (HCV) infected liver disease.

Material/Methods:

Nine patients (mean age 63.3±9.1 years) with HCV-related liver diseases were identified and examined for sensitivity to different tastes. Eight patients had no awareness of taste disorders, and 3 patients had oral lichen planus. We examined 4 tastes (sweet, salty, sour, and bitter) using a Taste Disk® and sensitivity to different tastes was rated on a 6-point scale (I, II, III, IV, V, and VI). Each patient was given one sachet of Aminofeel® after breakfast and another at bedtime for 90 days.

Results:

Only one patient was aware of a taste disorder before administration of Aminofeel®, but 4 patients had decreased gustatory sensitivity in the sour taste test, and 2 had it in the bitter taste test. Sensitivity to sour tastes significantly increased after the administration of Aminofeel® (P=0.03). Sensitivity to sweet tastes increased after the administration of Aminofeel® (P=0.06). Zinc value significantly increased after the administration of Aminofeel® (P=0.02).

Conclusions:

Patients with HCV-infected liver disease have decreased sensitivity to different tastes and decreased zinc levels. Some patients were unaware that they had a taste disorder. Aminofeel® improved sensitivity to different tastes and increased zinc values. Thus, Aminofeel® is a useful therapeutic agent for taste disorders.

key words:

taste disorder • zinc • hepatitis C virus (HCV) • lichen planus • branched-chain amino acids (BCAA) • Aminofeel®

Abbreviations:

BCAA - branched-chain amino acids; Anti-HCV - anti-bodies to HCV; CLEIA - chemiluminescent enzyme immunoassay; HBsAg - hepatitis B surface antigen; HCC - hepatocellular carcinoma; HCV - hepatitis C virus; IFN - interferon

Full-text PDF:

http://www.medscimonit.com/fulltxt.php?ICID=878481

Word count:

1517 **Tables:** 4 Figures: 5

21

References:

Author's address:

Yumiko Nagao, Department of Digestive Disease Information & Research, Kurume University School of Medicine, 67 Asahi-machi, Kurume 830-0011, Japan, e-mail: nagao@med.kurume-u.ac.jp

Product Investigation Med Sci Monit, 2010; 16(4); PI7-12

BACKGROUND

In Japan, the number of patients seeking treatment from otolaryngologists for taste disorders is approximately 240,000/year; the number has almost doubled in the last 13 years [1]. The main treatment for taste disorders is zinc administration. And many studies have found zinc deficiency in patients with liver diseases [2–4]. Therefore, patients with chronic liver disease may have taste disorders and altered zinc metabolism [5,6]. Several factors, such as poor dietary intake, impaired intestinal absorption, or excessive urinary losses may be responsible for the reduced whole-body zinc content [7].

Decreases in levels of serum branched-chain amino acids (BCAA) are often seen in patients with chronic liver diseases and these decreases lead to a decline in production of detoxified ammonia and albumin. Therefore, BCAAs are used for the treatment of hepatic encephalopathy and hypoalbuminemia [8,9]. The Department of Digestive Disease Information & Research, Kurume University School of Medicine and Seikatsu Bunkasya Co. Inc. (Tokyo, Japan) developed the BCAA-enriched supplement (Aminofeel®) and facilitated the commercialization of the product. On March 1, 2007, Seikatsu Bunkasya Co. Inc. released Aminofeel®. A dose of Aminofeel® contains 5.0 mg zinc as well as 3200.0 mg BCAA [10]. We previously reported that Aminofeel® is a useful therapeutic agent for decreasing insulin resistance in male patients with chronic viral diseases of the liver [10,11]. The administration of Aminofeel® in men for 90 days increases serum albumin levels significantly and also increases serum zinc levels [10].

There are few reports that have used objective outcomes to show that the sense of taste is disordered in patients with liver disease. Accordingly, in this study, we conducted objective, gustatory tests and objectively studied sensitivity to different tastes before and after administration of Aminofeel® in patients with hepatitis C virus (HCV)-infected liver disease.

MATERIAL AND METHODS

Subjects

A prospective, consecutive-patient entry study was conducted. Eligibility criteria were chronic viral liver disease, sufficient food intake, and serum albumin concentrations >3.5 g/dl and <4.0 g/dl. Patients with hepatic encephalopathy, ascites, hepatocellular carcinoma (HCC) or renal failure were excluded.

This study included 9 Japanese patients (3 males and 6 females) with HCV-infected liver disease. Eight patients, who had visited our clinic at the Kurume University Hospital in Japan between September 2006 and December 2006, had no awareness of taste disorders associated with their HCV-infected liver disease. The other patient visited our clinic with the main complaint of a taste disorder on December 2008. Patients ranged in age from 51 to 78 years, with an average age of 63.3±9.1 years.

Informed consent for participation in the study was obtained from each patient. The study protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in prior approval by the Ethics Committee of the

Kurume University School of Medicine. None of the subjects were institutionalized.

Methods

Taste test

Taste functions were analyzed using a Taste Disk® (Sanwa Kagaku Kenkyusho Co. Inc, Nagoya, Japan).

The region of taste buds are located is on the front twothirds of the tongue, which is innervated by chorda tympani; the area located on the rear one-third of the tongue is innervated by the glossopharyngeal nerve, and the region located on the soft palate is innervated by the greater petrosal nerve. We used a taste kit to check the different dominant nerves involved in taste in each area. The type and concentration of test fluid and the are as shown in Table 1. To stimulate tastes we put a paper 5 millimeters in diameter and moistened with a reagent in the measurement site (Figure 1). Gustatory criteria for 4 tastes (sweet, salty, sour, and bitter) were examined using a 6-point scale (I, II, III, IV, V, and VI). Numbers I, II, and III were standard values (I - minimum standard value, II - the median of the standard value, III – the upper limit of the standard value). Numbers IV, V, and VI were abnormal values (IV - a slight taste disorder, V - a medium taste disorder, VI - a severe taste disorder). Taste disorders were evaluated in the area of the right chorda tympani.

Liver function tests

Sera from all 9 patients were used for the following liver function tests: serum aspartate aminotransferase (AST), alanine aminotransferase (ALT), albumin (Alb), and zinc. Sera were also examined for the presence or absence of HCV or HBV infection. Anti-bodies to HCV (anti-HCV) antibodies and hepatitis B virus surface antigen (HBsAg) were measured by a chemiluminescent enzyme immunoassay (CLEIA) kit and a chemiluminescent immunoassay (CLIA), respectively. Ultrasonography for all subjects was done to examine the shape of the liver and lesions in the liver.

Design of the administration of Aminofeel®

Each patient was given one sachet of Aminofeel®, a BCAA-enriched supplement including zinc, after breakfast and another at bedtime for 90 days.

Statistical analysis

All data are expressed as mean ± standard error. Differences between two groups were analyzed using the Mann-Whitney U test. Statistical comparisons before administration of the Aminofeel® and after 90 days were done using Wilcoxon's test. All statistical analyses were conducted using JMP Version 6 (SAS Institute, Cary, NC, USA). The level of statistical significance was defined as 0.05.

RESULTS

The characteristics of the 9 patients studied are shown in Table 2. The diagnosis of liver disease included: chronic hepatitis C (n=5), liver cirrhosis (n=3), and post interferon

Table 1. A gustatory criterion by the kind of test fluid and the concentration.

| Taste | | I | II | III | IV | V | VI |
|--------------|---------------------|--------|-------|--------|--------|--------|----------------|
| Sweet taste | (Sucrose) | 0.30% | 2.50% | 10.00% | 20.00% | 80.0% | Unobservable V |
| Salty taste | (Sodium chloride) | 0.30% | 1.25% | 5.00% | 10.00% | 20.00% | Unobservable V |
| Sour taste | (Acidum tartaricum) | 0.02% | 0.20% | 2.00% | 4.00% | 8.00% | Unobservable V |
| Bitter taste | (Quinine) | 0.001% | 0.02% | 0.10% | 0.50% | 4.00% | Unobservable V |

Table 2. Characteristics of 9 patients with HCV-related liver diseases before administration of Aminofeel®.

| | | | | | | Oral liche | en planus | ! | Score o | f tast | e | | La | boratory d | ata | |
|----|-----|-----|------------------------|---------------------|----------------------------------|------------|-------------------------------|---------|---------|--------|--------|------------------------|-----------------------|--------------------------|----------------------------|-------------------------|
| | | | | Subjective | | | | | | | | AST | ALT | Alb | PLT | Zinc |
| No | Sex | Age | Liver disease | symptom of taste | Systemic disease | _ | | _ | | _ | | (IU/L) | (IU/L) | (g/dL) | (/mm³) | (µg/dL) |
| | | | uisease | disorder | uisease | Occurrence | Site | Sweet | Salty | Sour | Bitter | St. value; 13–33 | St. value; 8–42 | St. value; 4.0–5.0 | St. value; 13.0–36.0 | St, value; 80–130 |
| 1 | F | 66 | LC-C | Negative | Hypertenstion | Positive | Buccal mucosa | III | Ш | ٧ | M | 92 | 40 | 3.54 | 13.7 | 71 |
| 2 | F | 78 | CH-C | Negative | Negative | Negative | | 11 | 1 | III | II | 45 | 44 | 3.88 | 8.7 | 73 |
| 3 | F | 56 | CH-C | Negative | Hypertenstion | Negative | | - 11 | Эll | Ш | II. | 24 | 18 | 3.94 | 25.0 | 84 |
| 4 | М | 58 | CH-C | Negative | Negative | Negative | | П | II | II | II | 46 | 70 | 3.92 | 15.8 | 85 |
| 5 | F | 69 | CH-C | Negative | Negative | Negative | | 1 | 11 | IV | II | 46 | 45 | 3.99 | 18.1 | 71 |
| 6 | F | 54 | LC-C | Negative | Negative | Negative | | II. | II | 11 | - 11 | 33 | 24 | 3.88 | 18.0 | 126 |
| 7 | F | 72 | LC-C | Negative | Hypertenstion, Hyperlipidemia | Negative | | II | II | | II | 34 | 29 | 3.87 | 20.5 | 93 |
| 8 | М | 51 | CH-C | Negative | Diabetes mellitus | Positive | Buccal mucosa and tongu | ll e | VI | ٧ | V | 65 | 104 | 3.68 | 13.8 | 87 |
| 9 | M | 66 | CH-C post IFN (SVR) | Positive | Hypothyroidism | Positive | Buccal mucosa | VI | II | VI | ٧ | 21 | 15 | 4.3 | 8.6 | 67 |

CH-C — chronic hepatitis C; LC-C — HCV-related liver cirrhosis; SVR — Sustained virological response; AST — serum aspartate aminotransferase; ALT — alanine aminotransferase; Alb — albumin; PLT — platelets.

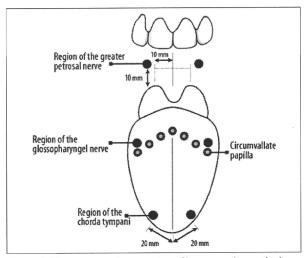


Figure 1. Measurement of tastes using a filter-paper disc method.

(IFN) treatment for chronic hepatitis C (n=1). After we succeeded in eliminating HCV by IFN treatment, one patient, a 66 year old man, developed a taste disorder. Of the 9 patients, 3 had oral lichen planus. There was one patient with oral lichen planus of the tongue. The serum zinc value of the 66 year old man with a complaint of a taste disorder was decreased (67 μ g/dL).

The distributions of gustatory sensitivity before and after administration of Aminofeel® are as shown in Figures 2–5. There was only one patient who was aware of a taste disorder before administration of Aminofeel®, but 4 patients had decreased gustatory sensitivity in the sour taste test, and 2 had it in the bitter taste test. Sensitivity to the sour taste was significantly increased 90 days after the administration of Aminofeel® (P=0.03, Figure 4). Sensitivity to the sweet taste was increased 90 days after the administration of Aminofeel® (P=0.06, Figure 2).

Product Investigation Med Sci Monit, 2010; 16(4): PI7-12

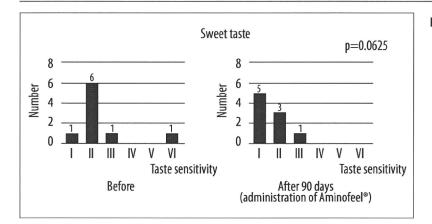


Figure 2. The effects of sweet tastes after administration of Aminofeel®. Sensitivity to sweet tastes was increased 90 days after the administration of Aminofeel® (P=0.06).

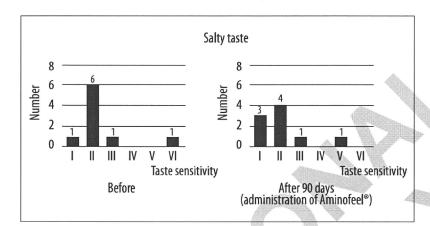


Figure 3. The effects of salty tastes after administration of Aminofeel®.

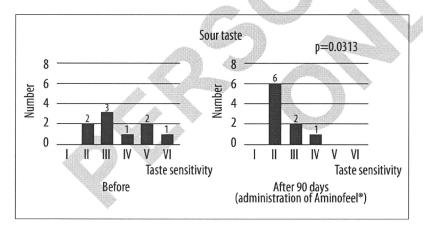


Figure 4. The effects of sour tastes after administration of Aminofeel®. Sensitivity to sour tastes was significantly increased 90 days after the administration of Aminofeel® (P=0.03).

We analyzed for differences before and after the administration of Aminofeel® in AST, ALT, albumin, platelet count and zinc values. These laboratory data are shown in Table 3. The serum zinc value was significantly increased (p=0.02).

DISCUSSION

The number of patients with taste disorders is increasing [1,12]. It is presumed that approximately 240,000/year receive medical treatment for taste disorders from otolaryngologists in Japan [1]. Ikeda et al reported, after administering questionnaires to 1,559 members of the Japan Society of Stomato-pharyngology, that the main treatment used was administration of zinc preparations such as polaprezinc [1]. One reason for the increased number of patients with taste disorders is that the elderly population has increased

year after year [13] and it is believed that taste disorders increase with age.

Taste disorders are symptoms of neurological derangement for which there are many reasons such as use of numerous drugs, idiopathic factors, zinc deficiency, psychogenic factors, systemic diseases, etc [5,6,14]. Zinc is essential for many metabolic and enzymatic functions [15]. A zinc deficiency in man has been found to occur not only as a result of nutritional factors, but also in various disease states, including malabsorption syndromes, acrodermatitis enteropathica, Crohn's disease, alcoholism and liver cirrhosis [15].

We found that sensitivity to tastes and zinc levels are decreased in patients with HCV-infected liver disease. Some patients had decreased sensitivity of taste despite the fact that they were

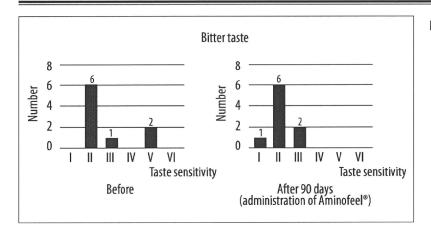


Figure 5. The effects of bitter tastes after administration of Aminofeel®.

Table 3. Laboratory data.

| | Before | Administration of Aminofeel® | p value |
|----------------------------|-----------|------------------------------|---------|
| AST (IU/L) (mean ±SD) | 45.1±22.1 | 44.8±18.3 | NS |
| ALT (IU/L) (mean ±SD) | 43.2±28.3 | 44.0±25.9 | NS |
| Alb (g/dL) (mean ±SD) | 3.9±0.2 | 4.0±0.2 | NS |
| PLT (/mm³) (mean ±SD) | 15.8±5.3 | 15.8±5.8 | NS |
| Zinc (μg/dL) (mean ±SD) | 84.1±18.0 | 108.4±23.5 | 0.0209 |

SD — standard deviation; NS — no significance; AST — serum aspartate aminotransferase; ALT — alanine aminotransferase; Alb — albumin; PLT — platelets.

unaware of their taste disorder. In addition, Aminofeel®, a BCAA-enriched supplement, improved sensitivity to tastes and increased zinc levels. Thus, because Aminofeel® contains zinc, it is a useful therapeutic agent for taste disorders. Hayashi et al reported that combination treatment with BCAA and zinc supplements in cirrhotic liver patients with hypoalbuminemia or hypozincemia showed significantly higher efficacy in correcting amino acid imbalances and significantly greater ability to metabolize ammonia than when BCAA was given alone during the 6 months of the study period [16]. There is a report that zinc only treatment did not improve taste disorder in liver cirrhosis [17]. Combination treatment with BCAA and zinc may be useful for improvement of gustatory sensibility, although it is not clear that whether the combination treatment is more effective for sensitivity to tastes in patients with liver diseases than zinc only or not.

Several studies and our previous reports suggest that HCV infection antedates insulin resistance [18,19], and that insulin resistance is associated with extrahepatic manifestations such as lichen planus [20,21]. We have already reported that Aminofeel® improved insulin resistance and β cell function in male patients with chronic liver disease [10]. A post-marketing surveillance study of Aminofeel® confirmed the usefulness of this supplement (data not shown).

CONCLUSIONS

Aminofeel® is a supplement that improves the sensitivity to tastes by increasing zinc levels. It also improves insulin resistance in patients with chronic liver disease. It is hoped that this supplement improves the prognosis of liver disease and the quality of life of these patients.

REFERENCES:

- Ikeda M, Aiba T, Ikui A et al: Taste disorders: a survey of the examination methods and treatments used in Japan. Acta Otolaryngol, 2005; 125: 1203–10
- Marchesini G, Fabbri A, Bianchi G et al: Zinc supplementation and amino acid-nitrogen metabolism in patients with advanced cirrhosis. Hepatology, 1996; 23: 1084–92
- Rodríguez-Moreno F, González-Reimers E, Santolaria-Fernández F et al: Zinc, copper, manganese, and iron in chronic alcoholic liver disease. Alcohol, 1997; 14: 39–44
- Hatano R, Ebara M, Fukuda H et al: Accumulation of copper in the liver and hepatic injury in chronic hepatitis C. J Gastroenterol Hepatol, 2000; 15: 786–91
- Smith FR, Henkin RI, Dell RB: Disordered gustatory acuity in liver disease. Gastroenterology, 1976; 70: 568–71
- Burch RE, Sackin DA, Ursick JA et al: Decreased taste and smell acuity in cirrhosis. Arch Intern Med, 1978; 138: 743

 –46
- McClain CJ, Marsano L, Burk RF, Bacon B: Trace metals in liver disease. Semin Liver Dis, 1991; 11: 321–39
- Muto Y, Sato S, Watanabe A et al: Effects of oral branchedchain amino acid granules on event-free survival in patients with liver cirrhosis. Clin Gastroenterol Hepatol, 2005; 3: 705–13
- Marchesini G, Zoli M, Dondi C et al: Anticatabolic effect of branchedchain amino acid-enriched solutions in patients with liver cirrhosis. Hepatology, 1982; 2: 420–25
- Kawaguchi T, Nagao Y, Matsuoka H et al: Branched-chain amino acidenriched supplementation improves insulin resistance in patients with chronic liver disease. Int J Mol Med, 2008; 22: 105–12
- Kawaguchi T, Taniguchi E, Itou M et al: Branched-chain amino acids improve insulin resistance in patients with hepatitis C virus-related liver disease: report of two cases. Liver Int, 2007; 27: 1287–92
- Hoffman HJ, Ishii EK, MacTurk RH: Age-related changes in the prevalence of smell/taste problems among the United States adult population. Results of the 1994 disability supplement to the National Health Interview Survey (NHIS). Ann NY Acad Sci, 1998; 855: 716–22
- Ikeda M, Ikui A, Komiyama A et al: Causative factors of taste disorders in the elderly, and therapeutic effects of zinc. J Laryngol Otol, 2008; 199: 155-60
- Hamada N, Endo S, Tomita H: Characteristics of 2278 patients visiting the Nihon University Hospital Taste Clinic over a 10-year period with special reference to age and sex distributions. Acta Otolaryngol Suppl, 2002; 546: 7–15
- 15. Prasad AS: The role of zinc in gastrointestinal and liver disease. Clin Gastroenterol, $1983;\,12:713-41$

- Hayashi M, Ikezawa K, Ono A et al: Evaluation of the effects of combination therapy with branched-chain amino acid and zinc supplements on nitrogen metabolism in liver cirrhosis. Hepatol Res, 2007; 37: 615–19
- 17. Sturniolo GC, D'Incà R, Parisi G et al: Taste alterations in liver cirrhosis: are they related to zinc deficiency? J Trace Elem Electrolytes Health Dis, 1992; 6: 15–19
- Hui JM, Sud A, Farrell GC et al: Insulin resistance is associated with chronic hepatitis C and virus infection fibrosis progression. Gastroenterology, 2003; 125: 1695–704
- Kawaguchi T, Yoshida T, Harada M et al: Hepatitis C virus down-regulates insulin receptor substrates 1 and 2 through up-regulation of suppressor of cytokine signaling 3. Am J Pathol, 2004; 165: 1499–508
- Nagao Y, Kawaguchi T, Tanaka K et al: Extrahepatic manifestations and insulin resistance in an HCV hyperendemic area. Int J Mol Med, 2005; 16: 201–26
- Nagao Y, Kawasaki K, Sata M: Insulin resistance and lichen planus in patients with HCV-infectious liver diseases. J Gastroenterol Hepatol, 2008; 23: 580–85





RESEARCH Open Access

Dental problems delaying the initiation of interferon therapy for HCV-infected patients

Yumiko Nagao^{1*}, Michio Sata^{1,2†}

Abstract

Background: There has been little discussion about the importance of oral management and interferon (IFN) therapy, although management of the side effects of therapy for chronic hepatitis C has been documented. This study determined whether dental problems delayed the initiation of IFN therapy for hepatitis C virus (HCV)-infected patients.

Results: We analyzed 570 HCV-infected patients who were admitted to our hospital from December 2003 to June 2010 for treatment consisting of pegylated IFN (Peg-IFN) monotherapy or Peg-IFN/ribavirin combination therapy. The group comprised 274 men and 296 women with a mean age 57.2 years. Of the 570 patients, six could not commence Peg-IFN therapy, despite their admission, because of dental problems such as periodontitis, pupitis, and pericoronitis. The ages of six whose dental problems delayed the initiation of Peg-IFN ranged from 25 to 67 years, with a mean age of 47.3 ± 15.2 years. IFN therapy was deferred for 61.3 ± 47.7 days. Among the six subjects for whom IFN treatment was delayed, only one had a salivary flow that was lower than the normal value.

Conclusions: Treatment of dental infections is required before IFN therapy for HCV infection can be started. To increase the depth of understanding of oral health care, it is hoped that dentists and medical specialists in all areas will hold discussions to generate cooperation.

Background

In Japan, hepatocellular carcinoma (HCC) is the fourth leading cause of death in males and the sixth in females according to a recent survey. The incidence of HCC has increased in Japan throughout the past several decades [1]. Hepatitis C virus (HCV) is the major cause of HCC in Japan, with 70% of cases being HCV-related. It is assumed that between one and two million Japanese people are chronically infected with HCV [1].

Interferon (IFN) therapy for chronic hepatitis C is the only treatment for completely eliminating the virus. Combination therapy with pegylated IFN (Peg-IFN) and ribavirin has been recommended widely as the first choice for chronic hepatitis C patients with high viral loads. The sustained virological response (SVR) rate after 48 weeks of treatment at a standard dose is approximately 40 to 50% [2-5]. It has been shown that

IFN therapy decreases the rate of development of HCC and improves the long-term prognosis [6-9].

Although IFN therapy has therapeutic benefits, the treatment produces a number of well-described side effects that are dominated by fatigue, influenza-like syndrome and neuropsychiatric symptoms [2-5,10-12] and management of such side effects is required during therapy. Among the side effects in a Japanese Phase III trial of Peg-IFN alfa-2a/alfa-2b and ribavirin, dental problems have been documented in patients with chronic hepatitis C. Meanwhile, it has been reported that hepatitis C infected patients have significant oral health needs [13-16] and that experience of dental caries is significantly worse for HCV-infected patients than patients in general [13].

Therefore, in the present study, we determined whether dental problems delayed the initiation of IFN therapy for HCV-infected patients.

Methods

Patients

A total of 570 HCV-infected patients who admitted to the Kurume University Hospital from December 2003

¹Department of Digestive Disease Information & Research, Kurume University School of Medicine, Kurume, Fukuoka, 830-0011, Japan Full list of author information is available at the end of the article



© 2010 Nagao and Sata; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

^{*} Correspondence: nagao@med.kurume-u.ac.jp

[†] Contributed equally

to June 2010 for treatment with Peg-IFN monotherapy or Peg-IFN/ribavirin combination therapy were studied (Table 1). The 570 patients were 274 men and 296 women with a mean age of 57.2 ± 11.6 years. They were consulted by one oral surgeon for each patient about presence of oral infection before commencing IFN treatment. All HCV-infected patients treated with IFN therapy at our hospital were required to undergo hospitalization for two weeks for therapeutic management and education about liver diseases.

We determined whether dental problems delayed the initiation of IFN therapy for these patients. Patients who underwent Peg-IFN therapy during dental treatment were excluded. Informed consent was obtained from all patients after the purpose and methods of the study were explained.

Salivary flow

We used a simple and low-cost test for xerostomia detection, which requires chewing on a piece of gauze for 2 min. The results from 531 of 570 patients were quantified using the Saxon test. A salivary flow rate ≤ 2 g/2 min was judged as decreased salivary secretion.

Serological assays

Serum samples were examined for the presence or absence of markers of HCV and HBV infection. The HCV RNA level before IFN therapy was analyzed by quantitative PCR assay (COBAS AMPLICOR HCV MONITOR v 2.0 Test, COBAS AmpliPrep/COBAS Taq-Man HCV Test, Roche Molecular Systems, New Jersey, US) [17,18]. HCV genotype was determined by polymerase chain reaction assay, using a mixture of primers for the subtype, as reported previously [19].

Table 1 Characteristics of 570 patients

| Men/Women | v | 274/29 | 5 |
|---------------------|---|----------|---------|
| Age (mean ± SD) yea | rs | 57.2 ± 1 | 1.6 |
| Liver disease | AH-C | 1 | (0.2% |
| | CH-C | 471 | (82.6% |
| | CH-(B+C) | 3 | (0.5% |
| | CH-C and post HCC treatment | 20 | (3.5% |
| | LC-C | 45 | (7.9%) |
| | LC-C and post HCC treatment | 30 | (5.3%) |
| Peg-IFN therapy | Peg-IFN alfa-2a monotherapy | 104 | (18.2%) |
| | Peg-IFN alfa-2a monotherapy and trial | 1 | (0.2%) |
| | Peg-IFN alfa-2a/RBA | 14 | (2.5% |
| | Peg-IFN alfa-2a/RBA and trial | 5 | (0.9%) |
| | Peg-IFN alfa-2b/RBA | 438 | (76.8%) |
| | Peg-IFN alfa-2b/RBA→Peg-IFN alfa-2a monotherapy | 4 | (0.7%) |
| | Peg-IFN alfa-2b/RBA→Peg-IFN alfa-2a monotherapy→Peg-IFN alfa-2a/RBA | 1 | (0.2%) |
| | Peg-IFN alfa-2b/RBA→Peg-IFN alfa-2a monotherapy→Peg-IFN alfa-2b/RBA | 1 | (0.2%) |
| | Peg-IFN alfa-2b/RBA→Peg-IFN alfa-2a/RBA | 2 | (0.4%) |
| HCV genotype | 1a | 2 | (0.4%) |
| | 1a or 1b | 1 | (0.2%) |
| | 1b | 401 | (70.4%) |
| | 2a | 121 | (21.2%) |
| | 2b | 24 | (4.2%) |
| | 3a | 1 | (0.2%) |
| | combination (1a and 1b) | 1 | (0.2%) |
| | combination (1b and 2b) | 1 | (0.2%) |
| | combination (1b and 3a) | 1 | (0.2%) |
| | combination (2a and 2b) | 2 | (0.4%) |
| | indeterminable | 3 | (0.5%) |
| | untested | 12 | (2.1%) |

CH-C: chronic hepatitis C, CH-(B+C): chronic hepatitis B and C, LC-C: liver cirrhosis, HCC: hepatocelular carcinoma, Peg-IFN: pegylated interferon, RBV: ribavirin

Therapeutic response was judged after IFN therapy as: SVR - normalization of alanine aminotranferase (ALT) levels and HCV RNA negative for six months or more after treatment; transient response (TR) - normalization of ALT levels and undetectable HCV RNA during IFN treatment but HCV RNA-positive after IFN treatment; non-responder (NR) - neither normal nor negative results for six months or more.

As shown in Table 1, chronic hepatitis C with HCV genotype 1b was the most common. Patients with genotypes 2a/2b underwent Peg-IFN monotherapy and those with genotypes 1a/1b, a combination of Peg-IFN and ribayirn.

Results

Dental problems delayed the initiation of IFN therapy Of 570 patients with HCV-related liver diseases, we documented six whose dental problems delayed the initiation of Peg-IFN therapy. Their ages ranged from

25 to 67 years, with a mean age of 47.3 ± 15.2 years. There were two men and four women (Table 2). These six patients could not commence IFN therapy, despite their admission for this treatment, and their therapy was deferred for 61.3 ± 47.7 days. Patient no. 1 had an acute odontogenic periostitis, resulting from periapical inflammation of endodontic origin. This was treated successfully by nonsurgical endodontics and administration of antibiotics. Patient no. 2 had an acute alveolar abscess, resulting from periodontal disease. His four molars were extracted after local anti-inflammation treatment. Patient no. 3 had a periapical periodontitis of the right mandibular second molar. The molar was extracted. Patient no. 4 had multiple dental problems with pain. After extirpation of dental pulps and extraction of teeth, she received IFN treatment. Patient no. 5 had apical periodontitis with gingival abscess, consequently her teeth were endodontically treated. Patient no. 6 had trismus and painful swallowing caused by pericoronitis of her

Table 2 Characteristics of six patients whose dental problems delayed the initiation of IFN therapy

| No. | Age | Sex | Liver Disease | HCV RNA | HCV genotype | Dental problems that delayed the initiation of Peg-IFN therapy | Period to onset of IFN treatment after dental therapy (days) | Underlying disease | IFN therapy | Effect of IFN treatment |
|-----|-----|-----|------------------|--------------------|-----------------|--|--|--|----------------------------|-------------------------------|
| 1 | 50 | F | CH-C | 980 kIU/ml | 1b | #1. Acute periostitis of the right maxilla, #2. Periapical periodontitis of the right maxillary first molar | 49 | Gallbladder polyp | Peg-IFN alfa-2b/ RBA | TR |
| 2 | 67 | М | CH-C | 3,940 kIU/ mI | 1b | #1. Acute alveolar abscess of bilateral mandibular molars, #2. Periodontal diseases of the right mandibular first and second molars, the left mandibular first molar, and the left maxilla first and second molars | 105 | Gastric ulcer | Peg-IFN alfa-2b/ RBA | NR |
| 3 | 36 | М | CH-C | over 500 kIU/ml | 1b | Periapical periodontitis of the right mandibular second molar | 4 | None | Peg-IFN alfa-2b/ RBA | SVR |
| 4 | 47 | F | CH-C | 43 kIU/ml | 2a | #1. Pulpitis of the right maxillary first premolar, the left maxillary second premolar, and the right mandibular second premolar, #2. Tooth stumps of the left maxillary canine and second premolar, and the right mandibular first premolar, #3. Dental caries of the right maxillary lateral incisor | 97 | Hypertension, Adjustment disorder, Gallstone | Peg-IFN alfa-2a | SVR |
| 5 | 59 | F | LC-C | 471 kIU/ml | 2a | #1. Periapical periodontitis and gingival abscess of the right mandibular lateral incisor, #2. Dental caries of bilateral mandibular central incisors | 105 | Depression, Hypertension, Osteoarthritis of the spine, Esophageal varices | Peg-IFN alfa-2b/ RBA | SVR |
| 6 | 25 | F | CH-C | 6.2 logIU/ mL | 1b | #1. Pericoronitis of the right mandibular wisdom tooth, #2. Horizontal impacted wisdom teeth of bilateral mandibles | 8 | None | Peg-IFN alfa-2b/ RBA | SVR |

CH-C: chronic hepatitis C, LC-C: liver cirrhosis, Peg-IFN: pegylated interferon, RBV: ribavirin, TR: transient biochemical responders, NR: nonresponder, SVR: sustained virological response