

**Table 4**  
**Characteristics and typical appearance of cirrhotic nodules at MR imaging**

	Signal Intensity		Dynamic MR Imaging <sup>a</sup>	Kupffer Cell Density	SPIO <sup>b</sup>	Hepatocellular Function	Gd-EOB-DTPA <sup>c</sup>
	T1WI	T2WI					
RNs	Iso—Hyper (siderotic nodules; T1/T2WI, hypo)	Iso—Hypo	Iso—Hyper	Similar <sup>d</sup>	Iso <sup>d</sup>	Similar <sup>d</sup>	Iso <sup>d</sup>
DNs	Low grade Hypo—Hyper	Hypo Hypo—Slightly hyper	Iso—Hyper	Various <sup>d</sup>	Hypo—Hyper <sup>d</sup>	Various <sup>d</sup>	Hypo—Hyper <sup>d</sup>

Abbreviations: DNs, dysplastic nodules; Gd-EOB-DTPA, gadoxetic acid; Hypo, hypointense; Hyper, hyperintense; Iso, isointense; RNs, regenerative nodules; SPIO, superparamagnetic iron oxide.

<sup>a</sup> T1-weighted GRE image on delayed phase after administration of gadolinium-based contrast agents.

<sup>b</sup> T2-weighted GRE image after administration of SPIO.

<sup>c</sup> T1-weighted GRE image on hepatocyte-selective phase after administration of Gd-EOB-DTPA.

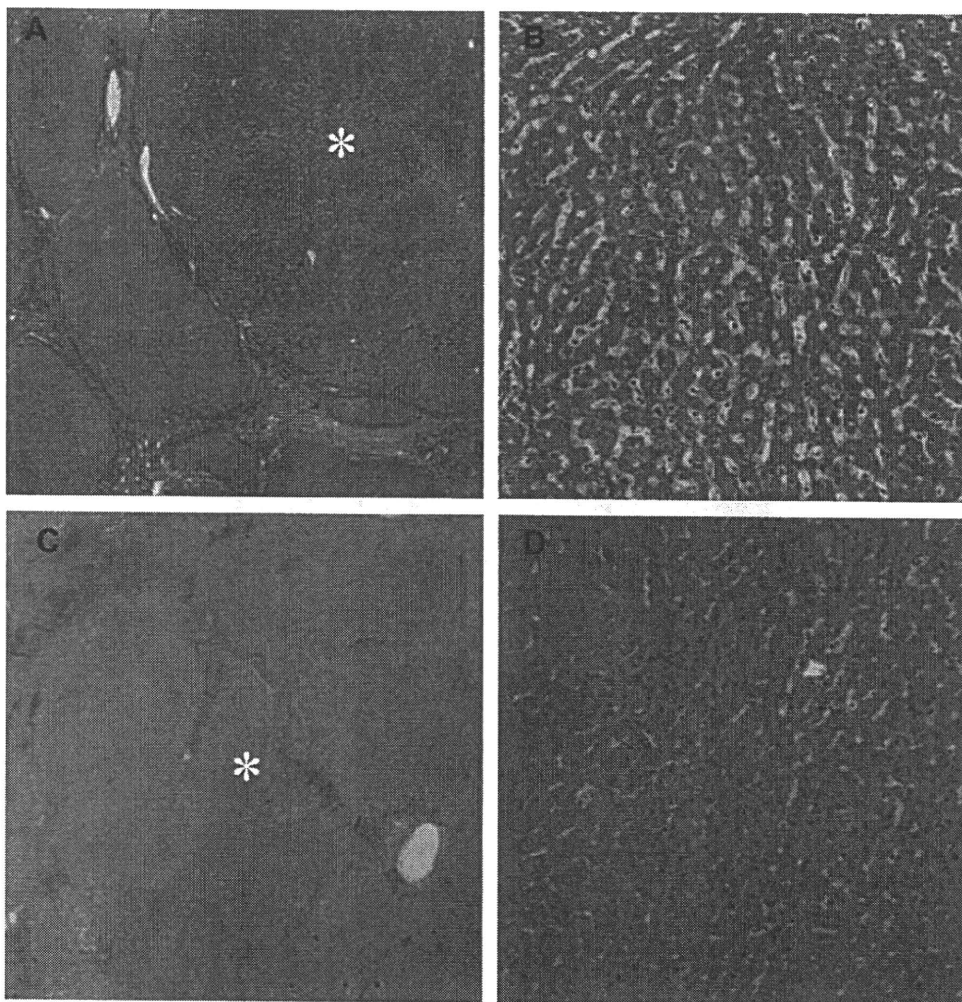
<sup>d</sup> Appearance is described in comparison with the surrounding hepatic parenchyma.

Data from Refs.<sup>47–55,59–61</sup>

elastography in an independent population of 35 healthy individuals and 48 patients with varying degrees of chronic liver disease showed a sensitivity of 86% and specificity of 85% for the detection of stages 2 to 4 fibrosis compared with liver histology from biopsy. A high negative predictive value (97%) for excluding the presence of fibrosis was also noted, suggesting that MR elastography might have a role in improving the ability to risk-stratify patients for liver biopsy to exclude occult advanced fibrosis.<sup>39</sup> MR elastography therefore appears to show promise for the noninvasive staging of liver fibrosis, particularly in patients with advanced fibrosis.

Diffusion-weighted magnetic resonance imaging is a technique that assesses the freedom of diffusion of water protons within tissue by applying motion-sensitizing gradients that cause diffusing protons to lose signal. Recent advances

in MR imaging technology have facilitated the performance of diffusion-weighted MR imaging of the liver, and it has also been used to detect liver fibrosis. Prior studies have reported that apparent diffusion coefficient (ADC) values acquired from b values of 500 (seconds/mm<sup>2</sup>) and greater correlated significantly with liver fibrosis stage, and that ADC values with a combination of b value of 0 and 1000 (seconds/mm<sup>2</sup>) showed the highest correlation ( $r = -0.654$ ,  $P < .001$ ).<sup>40</sup> On the other hand, several studies noted that there was no significant correlation between fibrosis stage and the ADC value using low b values (b values, 50 to 400 seconds/mm<sup>2</sup>), because diffusion-weighted imaging with a low b value was influenced by perfusion contamination.<sup>40,41</sup> Luciani and colleagues,<sup>42</sup> reported that ADC calculated from low b values was significantly reduced in cirrhosis. Thus, the fast component diffusion-weighted MR imaging



**Fig. 16.** Photomicrographs (Azan-Mallory stain, original magnification  $\times 20$  and  $\times 100$ ) in a patient with HCV shows low-grade dysplastic nodule (asterisk) (A, B) and high-grade dysplastic nodule (asterisk) (C, D). (Courtesy of Osamu Nakashima, MD, Department of Pathology, Kurume University School of Medicine, Japan.)

obtained with low  $b$  values may provide information related to microperfusion changes in diffuse liver disease whereas the slow component diffusion-weighted MR imaging obtained with high  $b$  values has been suggested to reflect a decrease in water proton diffusion.<sup>43</sup> The principles of diffusion-weighted MR imaging is discussed further in this presentation on functional MR imaging techniques.

In vivo MR spectroscopy (MRS) is most commonly used to assess signals from hydrogen ( $^1\text{H}$ ) and phosphorus ( $^{31}\text{P}$ ). Although  $^1\text{H}$ -based MRS allows for the quantification of certain metabolites and lipids,  $^{31}\text{P}$ -based MRS provides insights on processes, including cell turnover and energy state, based on the substantial  $^{31}\text{P}$  concentrations within hepatocytes.<sup>44</sup> Previous studies have suggested MRS may be useful in detecting hepatic fibrosis.<sup>45,46</sup> An increased levels of hepatic phosphomonoesters (PME) have been reported in patients with established cirrhosis,<sup>45,46</sup> and an increasing PME to phosphodiester (PDE) ratio has been reported to correlate with worsening

necroinflammatory and fibrosis scores on liver histology.<sup>47</sup> It has also been suggested that a PME and PDE ratio 0.2 or less is correlated with mild hepatitis and 0.3 or greater is correlated with cirrhosis in a study involving patients with chronic hepatitis C.<sup>48</sup> Despite some preliminary promising data,  $^{31}\text{P}$ -based MRS is not widely used due to specific technical requirements. The role of MRS in the detection of liver inflammation and fibrosis requires further investigation.

#### CIRRHOSIS-ASSOCIATED HEPATOCELLULAR NODULES

##### *Regenerative Nodules*

Regenerative nodules form in response to necrosis, altered circulation, or other stimuli,<sup>49</sup> and may progress along a well-described carcinogenic pathway to become dysplastic nodules or hepatocellular carcinomas.<sup>50</sup> These nodules are present in all cirrhotic livers and are surrounded by fibrous septa (see Fig. 10).<sup>16</sup> The nodules may be monoacinar or multiacinar, depending on

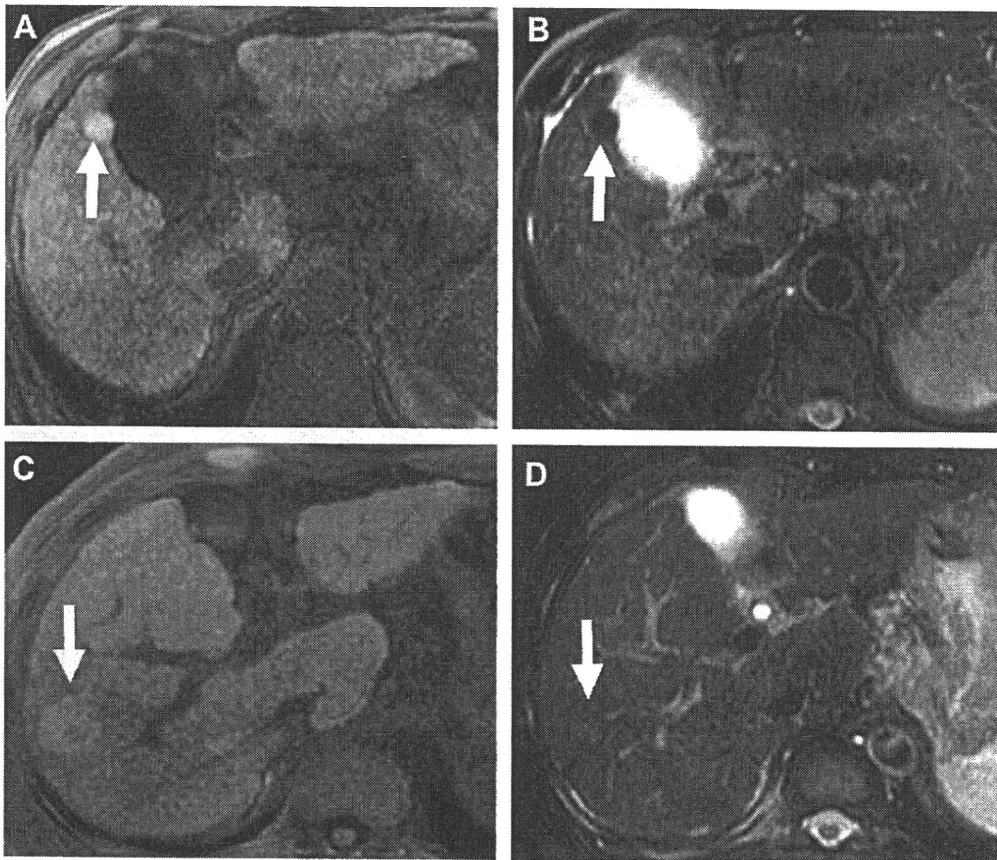
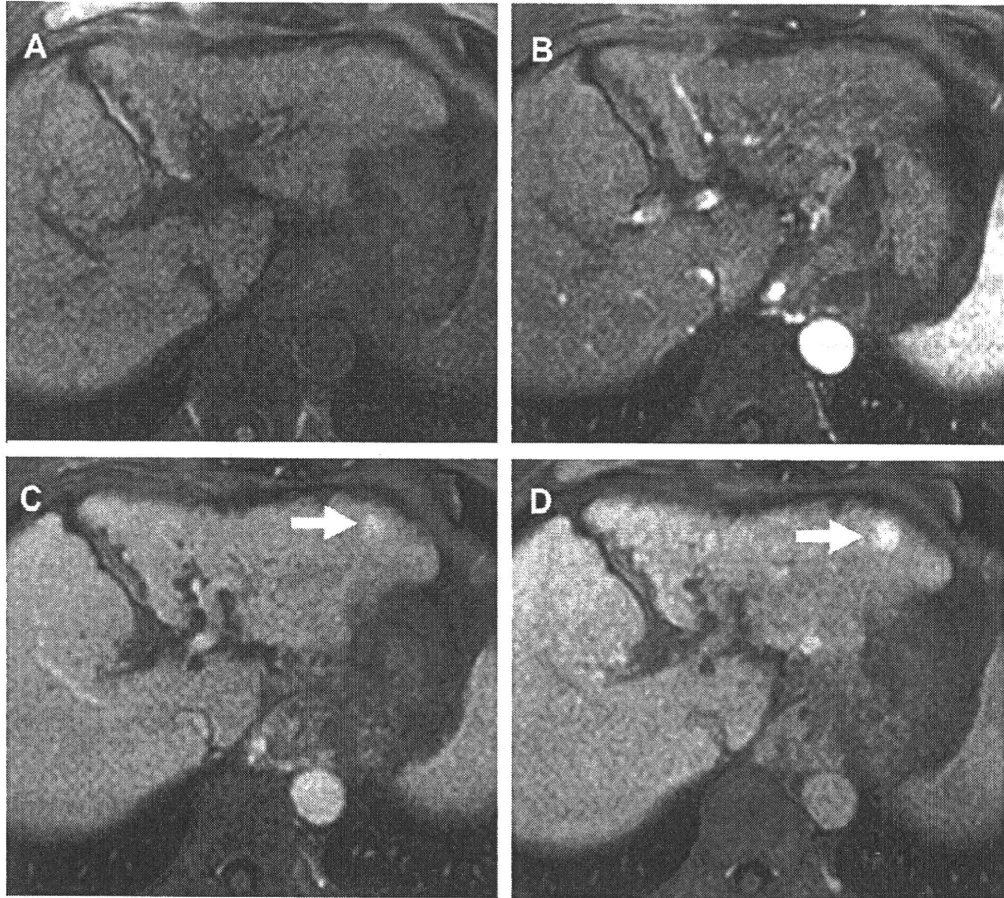


Fig. 17. Low-grade (A, B) and high-grade dysplastic nodules (C, D). All nodules were hyperintense on T1-weighted fat-saturated 3D GRE images (TR/TE = 3.6/1.7 ms, flip angle = 15°) (A, C) (arrows). In T2-weighted fat-saturated TSE images, in contrast, a low-grade dysplastic nodule is observed as hypointense (B) (arrow), whereas a high-grade nodule is observed as slightly hyperintense (D) (arrow).

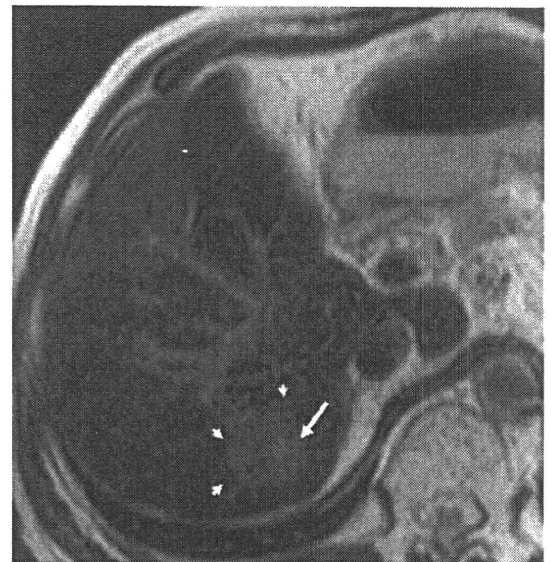




**Fig. 18.** Dynamic enhancement patterns of a high-grade dysplastic nodule in axial T1-weighted fat-saturated 3D GRE images (TR/TE = 3.6/1.7 ms, flip angle = 15°), presenting before (A) and in the arterial phase (30 s) (B), portal phase (90 s), and (C) equilibrium phase (4 min), and (D) after intravenous injection of Gd-EOB-DTPA. Portal and equilibrium phases (arrow) show increased enhancement of high-grade dysplastic nodule.

whether they contain one or more terminal portal tracts, and can also be classified by size as of the micronodular ( $\leq 3$  mm) (Fig. 14), macronodular ( $>3$  mm) (see Fig. 14), or mixed type (features of both micro- and macronodular types).<sup>51</sup>

MR imaging demonstrates regenerative nodules with greater sensitivity than any other imaging modality. These nodules usually appear isointense to hypointense (Fig. 15) on T2-weighted images relative to the surrounding inflammatory fibrous septa, and isointense to hyperintense (see Fig. 15) relative to background liver parenchyma on T1-weighted images.<sup>52</sup> The accumulation of iron within regenerative nodules (siderotic nodules) may cause hypointensity on both T1- and T2-weighted images (see Fig. 15) owing to susceptibility effects.<sup>53</sup> With regard to blood supply on dynamic imaging, regenerative nodules are usually enhanced to the same or greater degree than the background liver in the portal venous phase,<sup>54</sup> owing to the large contribution from the portal vein, with minimal contribution from the hepatic artery (Table 4).<sup>55</sup>



**Fig. 19.** T2-weighted TSE image shows an iso- to slightly high-signal-intensity nodule (arrowheads) with a focus of higher signal intensity (arrow) within the nodule. This higher signal intensity focus within the nodule shows the presence of HCC.



### Dysplastic Nodules

Dysplastic nodules are considered an intermediate, premalignant step along the hepatocarcinogenesis process, and can also be classified by the degree of dysplasia as low- or high-grade.<sup>56</sup>

Low-grade dysplastic nodules are sometimes vaguely nodular but are often distinct from the surrounding cirrhotic liver because of the presence of peripheral fibrous scar.<sup>56</sup> This nodule is not a true capsule, but rather condensation of scarring as is seen around all cirrhotic nodules. Low-grade dysplastic nodules show mild increase in cell density with a uniform pattern, and without cytologic atypia.<sup>56</sup> Architectural changes beyond clearly regenerative features are not present; these

lesions do not contain pseudoglands or markedly thickened trabeculae (Fig. 16).<sup>56</sup> High-grade dysplastic nodules may also be distinctly or vaguely nodular in the background of cirrhosis, although they also lack a true capsule, similar to low-grade dysplastic nodules; however, they are more likely to show a vaguely nodular pattern than low-grade dysplastic nodules.<sup>56</sup> A high-grade dysplastic nodule is defined as having architectural and/or cytologic atypia, but the atypia is insufficient for a diagnosis of HCC.<sup>56</sup> These lesions most often show increased cell density, sometimes more than 2 times higher than the surrounding nontumoral liver, often with an irregular trabecular pattern (see Fig. 16).<sup>56</sup> On MR imaging, dysplastic nodules have variable

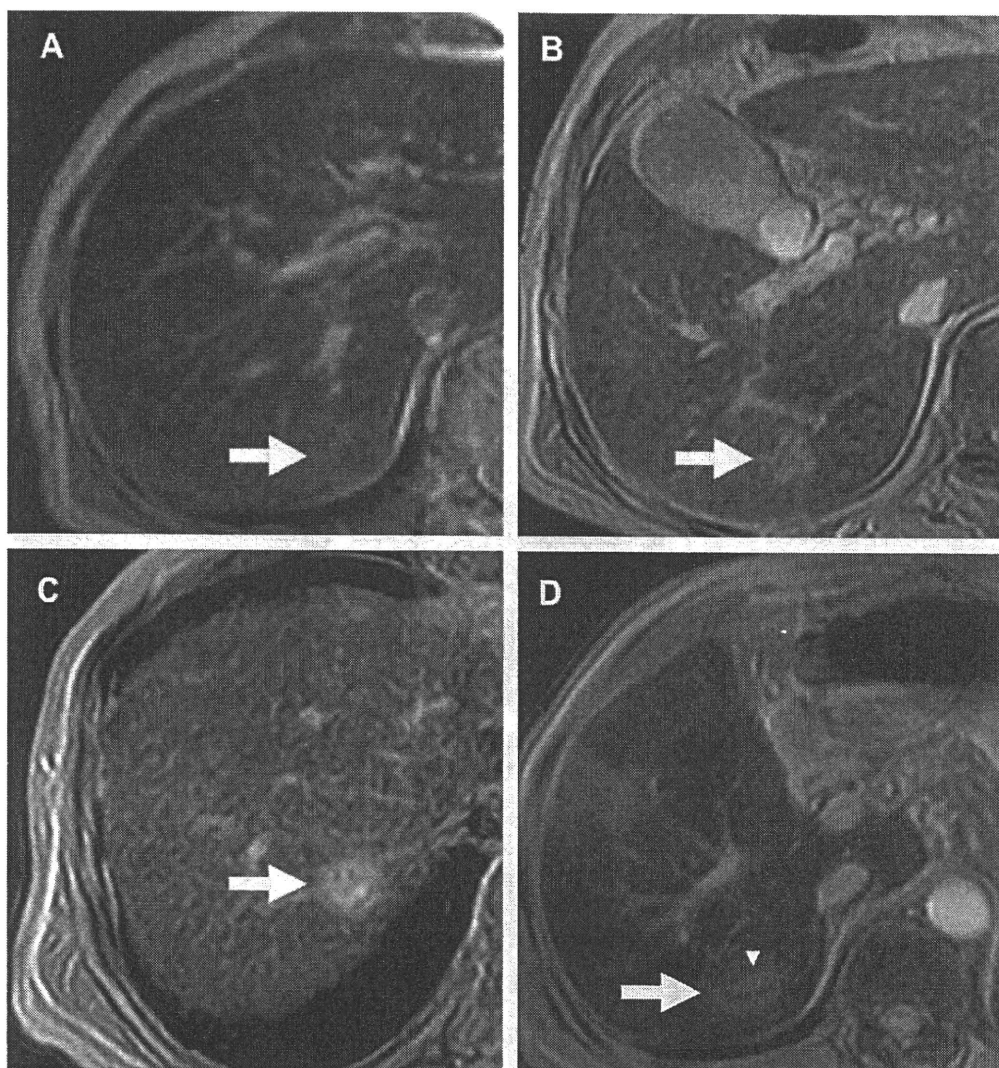


Fig. 20. T2-weighted GRE images (TE, 9.5 ms) after administration of SPIO in well-differentiated hepatocellular carcinoma (HCC) shows various signal intensities depending on Kupffer cell function within the nodule (A–D) (arrow). A dysplastic nodule with a central focus of HCC is observed as “a nodule within a nodule” (D) (arrowhead).

appearances, and their signal intensity characteristics overlap with those of regenerative nodules and well-differentiated HCC. On T2-weighted images, most dysplastic nodules are usually hypointense, and only rarely hyperintense (**Fig. 17**). It has been suggested that high-grade dysplastic nodules tend to have slightly higher signal intensity on T2-weighted images (see **Fig. 17**)<sup>57</sup>; however, the distinction from HCC and a high-grade dysplastic nodule may be difficult even on pathology. On T1-weighted images dysplastic nodules characteristically demonstrate high signal intensity, which may be related to deposition of copper,  $\text{Fe}^{3+}$ , or glycogen, or a high protein or lipid content (see **Fig. 17**).<sup>58,59</sup> However, the appearance on T1-weighted images cannot be used to distinguish low- and high-grade dysplastic nodules because both display variable (low, iso-, or high) signal intensity.<sup>57</sup>

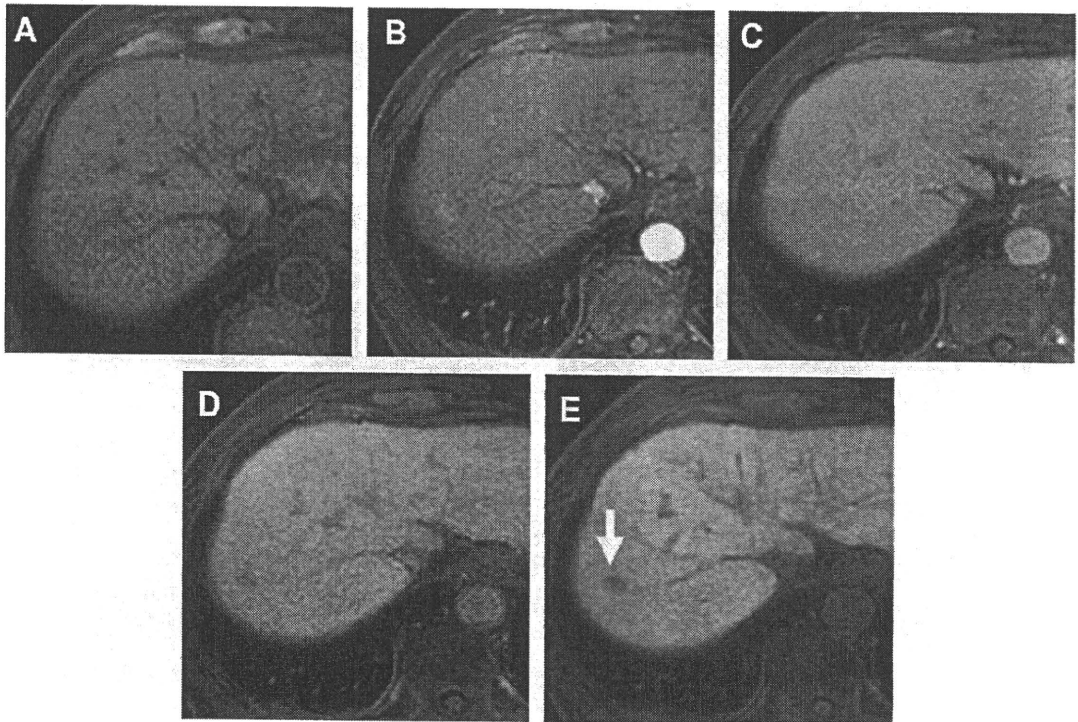
With regard to blood supply, dysplastic nodules are typically hypovascular lesions with predominantly portal venous blood supply, although increased arterial flow is seen in a small minority of cases (**Fig. 18**) (see **Table 4**).<sup>60</sup> The signal intensity characteristics of some high-grade dysplastic nodules that receive increasing supply from the hepatic artery may overlap with those of HCC

nodules during the process of hepatocarcinogenesis.<sup>61</sup> The hepatocarcinogenesis theory has been supported by the description of a dysplastic nodule with a central focus of HCC on T2-weighted images as “a nodule within a nodule.”<sup>62</sup>

The classic MR appearance is a focus of high signal intensity within a low-signal-intensity nodule on T2-weighted images (**Fig. 19**). This focus of HCC may also be enhanced in the arterial phase.<sup>63</sup> Despite the possibility of HCC developing within dysplastic nodules, the development of this tumor may not be a linear process because HCC is recognized to occur in patients with chronic HBV but without cirrhosis.

#### **Liver-Specific MR Contrast Agents (SPIO, Gd-EOB-DTPA) for Liver Nodules**

Because the density of Kupffer cells within regenerative nodules is similar to that in the surrounding nonneoplastic hepatic parenchyma, these nodules take up SPIO through Kupffer cell phagocytosis. On T2-weighted GRE and T2-weighted spin-echo sequences after administration of SPIO, regenerative nodules show the same signal intensity as that of surrounding hepatic parenchyma. In contrast, because Kupffer cell density within



**Fig. 21.** Dynamic enhancement patterns of well-differentiated HCC in axial 3D fat-saturated T1-weighted GRE images (TR/TE = 3.6/1.7 ms, flip angle = 15°), presenting before (A) and in the arterial phase (B), portal phase (C), equilibrium phase (D), and hepatocyte-selective phase (E) after intravenous injection of Gd-EOB-DTPA. Well-differentiated HCC is commonly observed as hypointense in the hepatocyte-selective phase (arrow).

dysplastic nodules and well-differentiated HCC is variable, the signal intensity of these nodules may also vary after administration of SPIO.<sup>57,64</sup> It has been suggested that the extent of SPIO uptake may reflect the degree of Kupffer cell function (Fig. 20).<sup>65</sup> Signal intensity characteristics of dysplastic nodules after administration of SPIO also overlap with those of regenerative nodules and well-differentiated HCC, and uptake of SPIO into these nodules may cause a decrease in detection.

Regenerative nodules generally have normal hepatocellular function and therefore demonstrate uptake of hepatocellular contrast agents such as Gd-EOB-DTPA. As dedifferentiation proceeds, the number of expressed organic anion transporters decreases, with a resulting progressive decrease in the uptake of hepatocellular agents.<sup>66</sup> It is considered that the appearance of HCC at hepatocyte-selective phases with hepatocellular agents is dependent on the degree of tumor

differentiation. However, hepatocytes in well-differentiated HCC may retain enough hepatocellular function to take up hepatocellular agents, and hence may be overlooked at this phase of imaging, or appear similar to a regenerative or dysplastic nodule (see Table 4).

In the authors' experience, most well-differentiated HCCs diagnosed by needle biopsy are clearly observed as hypointense to liver at hepatocyte-selective phases on Gd-EOB-DTPA-enhanced MR imaging (Fig. 21). Nevertheless, some well-differentiated HCCs are observed as isointense or hyperintense. Conclusive differentiation of dysplastic nodules from well-differentiated HCCs appears difficult (Fig. 22). Moreover, the diagnostic differentiation of dysplastic nodules from other cirrhosis-associated hepatocellular nodules may be difficult even on histopathologic analysis, and the use of molecular genetics-based techniques may be necessary in future.<sup>61</sup>

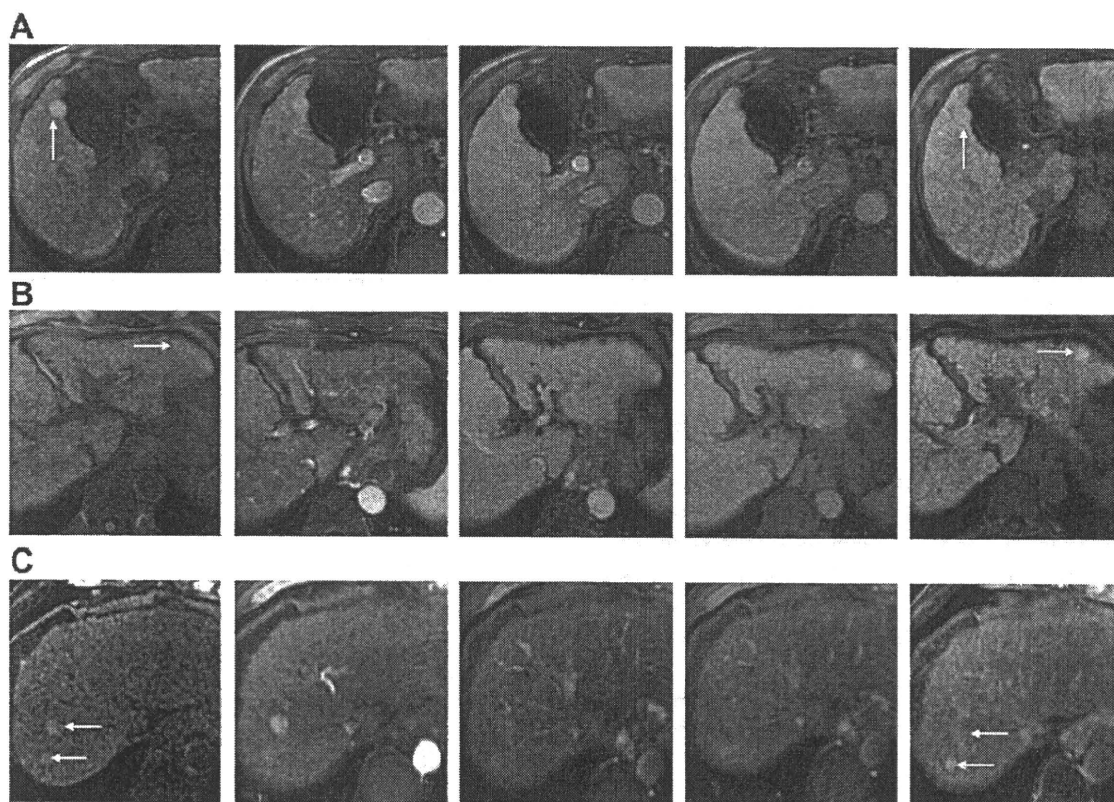


Fig. 22. Dynamic enhancement patterns of a low-grade dysplastic nodule (A), high-grade dysplastic nodule (B), and well-differentiated HCCs (C) in axial T1-weighted fat-saturated 3D GRE images (TR/TE = 3.6/1.7 ms, flip angle = 15°), presenting before and in the arterial phase, portal phase, equilibrium phase, and hepatocyte-selective phase after intravenous injection of Gd-EOB-DTPA. In the hepatocyte-selective phase, each nodule is observed as isointense or hyperintense owing to the uptake of hepatocellular agents (arrows). In series (C), both nodules were well-differentiated HCCs. In the hepatocyte-selective phase, both HCCs are observed as isointense and hyperintense, respectively. All these HCCs were diagnosed by needle biopsy.



## REFERENCES

- Mortele KJ, Ros PR. MR imaging in chronic hepatitis and cirrhosis. *Semin Ultrasound CT MR* 2002;23(1):79–100.
- Popper H. Pathologic aspects of cirrhosis. A review. *Am J Pathol* 1978;87:228–58.
- Garcia-Tsao G, Lim JK. Members of Veterans Affairs Hepatitis C Resource Center Program. Management and treatment of patients with cirrhosis and portal hypertension: recommendations from the Department of Veterans Affairs Hepatitis C Resource Center Program and the National Hepatitis C Program. *Am J Gastroenterol* 2009;104(7):1802–29.
- Sorrell MF, Belongia EA, Costa J, et al. National Institutes of Health Consensus Development Conference Statement: management of hepatitis B. *Ann Intern Med* 2009;150:104–10.
- Global surveillance and control of hepatitis C. Report of a who consultation organized in collaboration with the viral hepatitis prevention board, Antwerp, Belgium. *J Viral Hepat* 1999;6:35–47.
- Davis GL, Albright JE, Cook SF, et al. Projecting future complications of chronic hepatitis C in the United States. *Liver Transpl* 2003;9:331–8.
- Afdhal NH. The natural history of hepatitis C. *Semin Liver Dis* 2004;24(Suppl 2):3–8.
- Maher JJ. Alcoholic liver disease. In: Feldman M, Friedman LS, Sleisenger MH, editors. *Gastrointestinal and liver disease*, vol. II. Philadelphia: Saunders; 2002. p. 1375–91.
- Breitkopf K, Nagy LE, Beier JI, et al. Current experimental perspectives on the clinical progression of alcoholic liver disease. *Alcohol Clin Exp Res* 2009;33:1647–55.
- Greenfield V, Cheung O, Sanyal AJ. Recent advances in nonalcoholic fatty liver disease. *Curr Opin Gastroenterol* 2008;24(3):320–7.
- Itoh H, Sakai T, Takahashi N, et al. Periportal high intensity on T2-weighted MR images in acute viral hepatitis. *J Comput Assist Tomogr* 1992;16:564–7.
- Matsui O, Kadoya M, Takashima T, et al. Intrahepatic periportal abnormal intensity on MR images: an indication of various hepatobiliary diseases. *Radiology* 1989;171:335–8.
- Fisher MR, Gore RM. Computed tomography in the evaluation of cirrhosis and portal hypertension. *J Clin Gastroenterol* 1985;7:173–81.
- Ito K, Mitchell DG, Gabata T. Enlargement of hilar periportal space: a sign of early cirrhosis at MR imaging. *J Magn Reson Imaging* 2000;11:136–40.
- Ito K, Mitchell DG. Imaging diagnosis of cirrhosis and chronic hepatitis. *Intervirol* 2004;47:134–43.
- Ito K, Mitchell DG, Siegelman ES. Cirrhosis: MR imaging features. *Magn Reson Imaging Clin N Am* 2002;10:75–92.
- Lafortune M, Matricardi L, Denys A, et al. Segment 4 (the quadrate lobe): a barometer of cirrhotic liver disease at US. *Radiology* 1998;206:157–60.
- Ito K, Mitchell DG, Gabata T, et al. Expanded gallbladder fossa: simple MR imaging sign of cirrhosis. *Radiology* 1999;211:723–6.
- Ito K, Mitchell DG, Kim MJ, et al. Right posterior hepatic notch sign: a simple diagnostic MR finding of cirrhosis. *J Magn Reson Imaging* 2003;18:561–6.
- Okazaki H, Ito K, Fujita T, et al. Discrimination of alcoholic from virus-cirrhosis on MR imaging. *AJR Am J Roentgenol* 2000;175:1677–81.
- Ito K, Mitchell DG, Outwater EK, et al. Primary sclerosing cholangitis: MR imaging features. *AJR Am J Roentgenol* 1999;172(6):1527–33.
- Brancatelli G, Federle MP, Ambrosini R. Cirrhosis: CT and MR imaging evaluation. *Eur J Radiol* 2007;61(1):57–69.
- Wenzel JS, Donohoe A, Ford KL 3rd, et al. Primary biliary cirrhosis: MR imaging findings and description of MR imaging periportal halo sign. *AJR Am J Roentgenol* 2001;176:885–9.
- Kobayashi S, Matsui O, Gabata T, et al. MRI findings of primary biliary cirrhosis: correlation with Scheuer histologic staging. *Abdom Imaging* 2005;30(1):71–6.
- Martí-Bonmatí L. MR contrast agents in hepatic cirrhosis and chronic hepatitis. *Semin Ultrasound CT MR* 2002;23(1):101–13.
- Vilgrain V. Ultrasound of diffuse liver disease and portal hypertension. *Eur Radiol* 2001;11:1563–77.
- Van Beers BE, Leconte I, Materne R, et al. Hepatic perfusion parameters in chronic liver disease: dynamic CT measurements correlated with disease severity. *AJR Am J Roentgenol* 2001;176:667–73.
- Semelka RC, Chung JJ, Hussain SM, et al. Chronic hepatitis: correlation of early patchy and late linear enhancement patterns on gadolinium-enhanced MR images with histopathology: initial experience. *J Magn Reson Imaging* 2001;13:385–91.
- Faria SC, Ganesan K, Mwangi I, et al. MR imaging of liver fibrosis: current state of the art. *Radiographics* 2009;29(6):1615–35.
- Yeh MM, Brunt EM. Pathology of nonalcoholic fatty liver disease. *Am J Clin Pathol* 2007;128(5):837–47.
- Bedossa P, Dargere D, Paradis V. Sampling variability of liver fibrosis in chronic hepatitis C. *Hepatology* 2003;38:1449–57.
- Blici NC, Semelka RC. Contrast agents for MR imaging of the liver. *Radiol Clin North Am* 2005;43(5):887–98.
- Lucidarme O, Baleston F, Cadi M, et al. Non-invasive detection of liver fibrosis: Is superparamagnetic iron oxide particle-enhanced MR imaging a contributive technique? *Eur Radiol* 2003;13(3):467–74.
- Aguirre DA, Behling CA, Alpert E, et al. Liver fibrosis: noninvasive diagnosis with double contrast

- material-enhanced MR imaging. *Radiology* 2006; 239:425–37.
35. Martí-Bonmatí L, Lonjedo E, Poyatos C, et al. MnDPDP enhancement characteristics and differentiation between cirrhotic and noncirrhotic livers. *Invest Radiol* 1998;33(10):717–22.
  36. Murakami T, Baron RL, Federle MP, et al. Cirrhosis of the liver: MR imaging with mangafodipir trisodium (Mn-DPDP). *Radiology* 1996;198(2):567–72.
  37. Muthupillai R, Lomas DJ, Rossman PJ, et al. Magnetic resonance elastography by direct visualization of propagating acoustic strain waves. *Science* 1995;269:1854–7.
  38. Muthupillai R, Ehman RL. Magnetic resonance elastography. *Nat Med* 1996;2:601–3.
  39. Yin M, Talwalkar JA, Glaser KJ, et al. Assessment of hepatic fibrosis with magnetic resonance elastography. *Clin Gastroenterol Hepatol* 2007;5(10):1207–13.
  40. Taouli B, Tolia AJ, Losada M, et al. Diffusion-weighted MRI for quantification of liver fibrosis: preliminary experience. *AJR Am J Roentgenol* 2007;189:799–806.
  41. Boulanger Y, Amara M, Lepanto L, et al. Diffusion-weighted MR imaging of the liver of hepatitis C patients. *NMR Biomed* 2003;16:132–6.
  42. Luciani A, Vignaud A, Cavet M, et al. Liver cirrhosis: intravoxel incoherent motion MR imaging—pilot study. *Radiology* 2008;249:891–9.
  43. Koinuma M, Ohashi I, Hanafusa K, et al. Apparent diffusion coefficient measurements with diffusion-weighted magnetic resonance imaging for evaluation of hepatic fibrosis. *J Magn Reson Imaging* 2005;22:80–8.
  44. Jalan R, Taylor-Robinson SD, Hodgson HJF. In vivo hepatic magnetic resonance spectroscopy: clinical or research tool? *J Hepatol* 1999;25:414–24.
  45. Khan SA, Cox IJ, Hamilton G, et al. In vivo and in vitro nuclear magnetic resonance spectroscopy as a tool for investigating hepatobiliary disease: a review of H and P MRS applications. *Liver Int* 2005;25:273–81.
  46. Munakata T, Griffiths RD, Martin PA, et al. An in vivo <sup>31</sup>P MRS study of patients with liver cirrhosis: progress towards a non-invasive assessment of disease severity. *NMR Biomed* 1993;6:168–72.
  47. Van Wassenaeer-van Hall HN, van der Grond J, van Hattum J, et al. <sup>31</sup>P magnetic resonance spectroscopy of the liver: correlation with standardized serum, clinical, and histological changes in diffuse liver disease. *Hepatology* 1995;21:443–9.
  48. Menon DK, Sargentoni J, Taylor-Robinson SD, et al. Effect of functional grade and etiology on in vivo hepatic phosphorus-31 magnetic resonance spectroscopy in cirrhosis: biochemical basis of spectral appearances. *Hepatology* 1995; 21:417–27.
  49. Cho SG, Kim MY, Kim HJ, et al. Chronic hepatitis: in vivo proton MR spectroscopic evaluation of the liver and correlation with histopathologic findings. *Radiology* 2001;221:740–6.
  50. Coleman WB. Mechanisms of human hepatocarcinogenesis. *Curr Mol Med* 2003;3(6):573–88.
  51. Lee RG. Fibrosis and cirrhosis. In: Lee RG, editor. *Diagnostic liver pathology*. St Louis (MO): Mosby-Year Book, Inc; 1994. p. 281–308.
  52. Krinsky GA, Lee VS. MR imaging of cirrhotic nodules. *Abdom Imaging* 2000;25:471–82.
  53. Zhang J, Krinsky GA. Iron-containing nodules of cirrhosis. *NMR Biomed* 2004;17(7):459–64.
  54. Seale MK, Catalano OA, Saini S, et al. Hepatobiliary-specific MR contrast agents: role in imaging the liver and biliary tree. *Radiographics* 2009;29(6):1725–48.
  55. Lim JH, Kim EY, Lee WJ, et al. Regenerative nodules in liver cirrhosis: findings at CT during arterial portography and CT hepatic arteriography with histopathologic correlation. *Radiology* 1999;210(2):451–8.
  56. International Consensus Group for Hepatocellular Neoplasia. Pathologic diagnosis of early hepatocellular carcinoma: a report of the international consensus group for hepatocellular neoplasia. *Hepatology* 2009;49(2):658–64.
  57. Hanna RF, Aguirre DA, Kased N, et al. Cirrhosis-associated hepatocellular nodules: correlation of histopathologic and MR imaging features. *Radiographics* 2008;28(3):747–69.
  58. Amano S, Ebara M, Yajima T, et al. Assessment of cancer cell differentiation in small hepatocellular carcinoma by computed tomography and magnetic resonance imaging. *J Gastroenterol Hepatol* 2003; 18:273–9.
  59. Ebara M, Fukuda H, Kojima Y, et al. Small hepatocellular carcinoma: relationship of signal intensity to histopathologic findings and metal content of the tumor and surrounding hepatic parenchyma. *Radiology* 1999;210:81–8.
  60. Matsui O, Kadota M, Kameyama T, et al. Benign and malignant nodules in cirrhotic livers: distinction based on blood supply. *Radiology* 1991;178:493–7.
  61. Willatt JM, Hussain HK, Adusumilli S, et al. MR imaging of hepatocellular carcinoma in the cirrhotic liver: challenges and controversies. *Radiology* 2008; 247(2):311–30.
  62. Mitchell DG, Rubin R, Siegelman ES, et al. Hepatocellular carcinoma within siderotic regenerative nodules: appearance as a nodule within a nodule on MR images. *Radiology* 1991;178(1):101–3.
  63. Goshima S, Kanematsu M, Matsuo M, et al. Nodule-in-nodule appearance of hepatocellular carcinomas: comparison of gadolinium-enhanced and ferumoxides-enhanced magnetic resonance imaging. *J Magn Reson Imaging* 2004;20(2):250–5.

64. Lim JH, Choi D, Cho SK, et al. Conspicuity of hepatocellular nodular lesions in cirrhotic livers at ferumoxides-enhanced MR imaging: importance of Kupffer cell number. *Radiology* 2001;220(3):669–76.
65. Tonan T, Fujimoto K, Azuma S, et al. Evaluation of small (< or = 2 cm) dysplastic nodules and well-differentiated hepatocellular carcinomas with ferucarbotran-enhanced MRI in a 1.0-T MRI unit: utility of T2\*-weighted gradient echo sequences with an intermediate-echo time. *Eur J Radiol* 2007; 64(1):133–9.
66. Gandhi SN, Brown MA, Wong JG, et al. MR contrast agents for liver imaging: what, when, how. *Radiographics* 2006;26:1621–36.



