insulin signaling, phosphatase and tensin homologue<sup>[69]</sup>, and SH2 domain-containing inositol phosphatase- 2<sup>[20]</sup> occur in HCC. Thus, HCC may be sensitive to insulin stimulation.

## Extrahepatic manifestations

HCV causes extrahepatic manifestations including mixed cryoglobulinemia, Sjögren's syndrome, and non-Hodgkin lymphoma, oral lichen planus, oral squamous cell carcinoma, and malignancies other than HCC<sup>[22-24,70-73]</sup>. In patients with extrahepatic manifestations of HCV, fasting insulin levels and homeostasis model assessment for insulin resistance are significantly higher than for patients without extrahepatic manifestations [22]. Among various extrahepatic manifestations, insulin resistance is associated with oral lichen planus<sup>[23]</sup>, oral squamous cell carcinoma<sup>[24]</sup> and multiple primary cancers including gastric cancer [24]. Although reasons for this association remain unclear, a high prevalence of precancerous lesions and cancers are seen in patients with type 2 diabetes mellitus<sup>[74,75]</sup>, suggesting that insulin resistance or hyperinsulinemia may enhance carcinogenic activities.

# DISTINCTIVE THERAPEUTIC STRATEGY FOR HCV-ASSOCIATED INSULIN RESISTANCE

Despite awareness of the increased risk of insulin resistance, therapeutic guidelines to inhibit distinctive complications of HCV-associated insulin resistance have not yet been established. HCV itself has a significant impact on the development of insulin resistance, and eradication of HCV improves insulin resistance, and eradication of HCV improves insulin resistance [44,46,76]. Thus, anti-viral therapy is a fundamental therapeutic strategy for patients with HCV infection. In addition, amelioration of insulin resistance is considered to inhibit complications and improve prognosis. Here, we summarize treatments which could improve HCV-associated insulin resistance as therapeutic strategies (Figure 1).

# Late evening snack

Proper diet and exercise are fundamental for patients with lifestyle-associated insulin resistance as well as patients with HCV-associated insulin resistance [77-80]. As a nutritional treatment for liver cirrhosis, divided energy intake (4 to 6 meals/d) has been recommended [77-80]. As postprandial hyperglycemia is characteristic of HCV-associated insulin resistance [77-80], a decrease in energy intake per meal reduces postprandial hyperglycemia and hyperinsulinemia. In particular, a late evening snack is reported not only to improve glucose intolerance [81-84], but also to suppress hepatocarcinogenesis in cirrhotic patients [85].

# Coffee consumption

Coffee consumption reduces the risk of elevated serum alanine aminotransferase activity<sup>[86]</sup>, hepatic fibrosis<sup>[87]</sup>, and disease progression in chronic hepatitis C<sup>[88]</sup>. Coffee

consumption also reduces the risk of HCC independent of HCC etiology<sup>[89]</sup>. Caffeine is metabolized by hepatic cytochrome P450 1A2 into 3 metabolites, the dimethylxanthines paraxanthine, theobromine, and theophylline. Of these metabolites, theophylline inhibits transforming growth factor-\(\beta\)-stimulated CTGF expression through PPARy and Smad 2/3-dependent pathways. Since CTGF and transforming growth factor-B are important factors associated with progression of hepatic fibrosis and hepatocarcinogenesis, a metabolite of caffeine, theophylline, may have an inhibitory effect on the development of complications associated with HCV infection. In addition, coffee has significant effects on glucose metabolism [90]. In an animal experiment, the insulin-sensitizing effects of coffee have been demonstrated[91]. Similarly, in a human study, coffee consumption reduced fasting glucose and insulin levels<sup>[90,92]</sup>. Although the mechanisms for the coffeeinduced insulin-sensitizing effect remain unclear, some possibilities exist. Chlorogenic acids, a constituent of coffee, inhibits hepatic glucose-6-phosphate translocation [90,93], limits glucose absorption from the gut by inhibiting Na+dependent transport<sup>[94]</sup>, and increases the secretion of glucose regulating hormone, glucagon-like peptide (GLP)-1, from the gut [90,95,96]. These findings suggest that a constituent of coffee, chlorogenic acid, directly ameliorates HCV-associated insulin resistance. Furthermore, coffee modulates lipid metabolism<sup>[97,98]</sup> and lowers body weight<sup>[90]</sup>, indicating that coffee may suppress the lipid-induced increase in oxidative stress and ameliorates HCV-associated insulin resistance.

### **Phlebotomy**

Hepatic iron overload produces oxidative stress and is a factor responsible for the development of HCV-associated insulin resistance<sup>[4,99-101]</sup>. Although the pathogenesis of hepatic iron overload remains unclear, recent studies showed that iron-regulating molecules are modulated by HCV infection. Hepcidin is a negative regulator of duodenal iron absorption and macrophage iron release<sup>[100]</sup> and decreased hepatic expression of hepcidin is seen in both HCV polyprotein transgenic mice<sup>[102]</sup> and patients with HCV infection<sup>[103-105]</sup>. In addition, upregulation of hepatic expression of transferrin receptor 2, a mediator of iron uptake, is responsible for hepatic iron overload<sup>[106]</sup>.

In order to reduce hepatic iron deposition, dietary iron restriction and phlebotomy are effective. Dietary iron restriction (less than 7 mg/d) decreases serum alanine aminotransferase levels in patients with HCV infection<sup>[107]</sup>. Phlebotomy reduces oxidative stress as well as insulin resistance in patients with HCV infection<sup>[101,108,109]</sup>. A long-term combination treatment with phlebotomy and dietary iron restriction reduces the risk of development of HCC in patients with HCV infection<sup>[110]</sup>.

### Supplementation of zinc

Zinc plays a crucial role in the metabolism of protein, carbohydrate, lipid, nucleic acid, and ammonia<sup>[111-113]</sup>. In fact, zinc supplementation improves glucose disposal

in patients with cirrhosis<sup>[114]</sup>. Zinc also inhibits hepatic inflammation<sup>[115]</sup> and hepatic fibrosis<sup>[116]</sup>. More recently, zinc supplementation was shown to lower the cumulative incidence of HCC in patients with HCV infection<sup>[117]</sup>. It is unclear whether these inhibitory effects of zinc on progression of liver disease are mediated by amelioration of insulin resistance. However, zinc participates in the synthesis, storage and secretion of insulin<sup>[118]</sup> and regulates the binding ability of insulin to bind to its receptor<sup>[113]</sup>. As the serum zinc level is decreased in patients with HCV infection<sup>[115,117]</sup>, supplementation of zinc could be a therapeutic option.

# Anti-diabetic agents

Exogenous insulin and sulfonylurea agents: Antidiabetic agents are effective for decreasing plasma glucose and HbA1c levels, leading to prevention of diabetes mellitus-associated complications including cardiovascular diseases[119,120]. However, it has never been determined whether anti-diabetic agents prevent complications or improve prognosis in patients with HCV infection. Use of exogenous insulin or sulfonylurea agents may worsen hyperinsulinemia. In fact, we, along with others, recently reported an association between exogenous insulin or sulphonylurea treatment and the development of HCC in patients with HCV infection<sup>[29,30,121]</sup>. Use of exogenous insulin is also reported to be associated with the development of colon cancer<sup>[122]</sup> and other malignancies<sup>[123]</sup>. Although a causal relationship between exogenous insulin and the development of HCC remains controversial<sup>[124]</sup>, the reduction of serum insulin levels is a first line therapeutic strategy for insulin resistance [125-128].

Insulin-sensitizing agents: Insulin resistance is associated with a poor response to anti-viral treatment in patients with HCV infection<sup>[10,43–46]</sup>. Amelioration of insulin resistance may improve the response to anti-viral treatment. However, the impact of insulin-sensitizing agents, biguanides and thiazolidinediones, on sustained virologic response (SVR) rates has not yet been established. Recently, metformin, a biguanide agent, has been reported to ameliorate HCV-associated insulin resistance, and increase the SVR rate in HCV genotype 1 infected patients with normalization of homeostasis model assessment for insulin resistance at week 24 of therapy<sup>[129]</sup>. Pioglitazone, a thiazolidinedione agent, has also been reported to ameliorate insulin resistance and increase SVR rates in patients with HCV genotype 4 infection [130]. Although the insulin-sensitizing mechanisms of metformin and of pioglitazone are different, both agents are known to upregulate IRS<sup>[131,132]</sup>, which is the molecule responsible for HCV-associated insulin resistance [3,6,50], and to improve HCV-associated insulin resistance. Because both agents have severe adverse effects, neither is recommended for patients with liver cirrhosis. Biguanides predispose cirrhotic patients to lactic acidosis [133]. Thiazolidinediones cause overproduction of hydrogen peroxide leading to severe hepatotoxicity<sup>[134]</sup>. Thus, further validation for safety is required.

Dipeptidyl peptidase IV (DPPIV) inhibitor is a new therapeutic agent<sup>[135]</sup> and its clinical efficacy in type 2 diabetes has been shown<sup>[136]</sup>. Although no study has examined the effect of DPPIV inhibitor on HCV-associated insulin resistance, we found that activation of DPPIV is a factor responsible for HCV-associated insulin resistance<sup>[27]</sup>. Thus, a DPPIV inhibitor may be suited for ameliorating HCV-associated insulin resistance.

# BCAA supplementation, a possible insulin-sensitizing agent

BCAA are constituents of proteins and are required for protein synthesis<sup>[19,78,137,138]</sup>. In addition, BCAA are reported to modulate glucose metabolism. Leucine and isoleucine induce glucose transporter 1 and 4 translocation to the plasma membrane of muscle cells and improve glucose metabolism in a carbon tetrachloride-induced cirrhotic rat model<sup>[139]</sup>. In addition, leucine enhances the insulin-induced activation of the Akt/mammalian target of the rapamycin pathway in adipocytes of db/db mice[140]. Moreover, isoleucine increases hepatic phosphatidylinositol 3-kinase activity and improves insulin resistance in Zucker fa/fa rats, a model of severe insulin resistance<sup>[141]</sup>. Recently, knockout of the mitochondrial BCAA aminotransferase gene in mice, in which results in elevated plasma BCAA levels, was found to ameliorate insulin resistance<sup>[142]</sup>. Thus, BCAA improve insulin signaling in various animal models via several pathways. In good agreement with these results in animals, in human studies, we have recently shown that BCAA-enriched supplementation reduces insulin resistance in patients with HCV infection<sup>[143,144]</sup>. In a multicenter, randomized, controlled trial, BCAA supplementation led to a reduction in the risk of HCC in cirrhotic patients<sup>[145]</sup>. This suppressive effect on hepatocarcinogenesis was more evident in obese patients with HCV infection<sup>[145]</sup>. Both obesity and HCV induce the development of insulin resistance. Thus, BCAA may improve insulin resistance and subsequently inhibit insulin resistance-induced hepatocarcinogenesis [19,145].

# CONCLUSION

In this review, we summarize the distinctive complications of, and therapeutic strategies for, HCV-associated insulin resistance. Although cardiovascular diseases, renal failure, and infections are well-known complications of lifestyle-associated insulin resistance, these complications are not major causes of death in cirrhotic patients with insulin resistance. HCV-associated insulin resistance rather causes (1) hepatic steatosis, (2) resistance to antiviral treatment, (3) hepatic fibrosis and esophageal varices, (4) hepatocarcinogenesis and proliferation of HCC, and (5) extrahepatic manifestations. These complications are life-threatening, and therapeutic strategies for HCV-associated insulin resistance have to be considered on the basis of its pathogenic mechanisms.

Pathogenic mechanisms for HCV-associated insulin resistance differ from those for lifestyle-associated insulin resistance. Postprandial hyperglycemia, lipid-induced oxidative stress, hepatic iron overload, and depletion of zinc are responsible for the development of HCV-associated insulin resistance. Therefore, a late evening snack, coffee consumption, dietary iron restriction, phlebotomy, and supplementation of zinc are recommended therapeutic strategies. No clinical guidelines for the use of anti-diabetic agents are available for patients with HCV-associated insulin resistance. However, use of exogenous insulin or sulphonylurea may increase the risk for HCC. On the other hand, insulin-sensitizing agents may improve the SVR rate of anti-viral treatment. In addition, BCAA supplementation has an insulin-sensitizing effect as well as a suppressive effect on hepatocarcinogenesis. Thus, in order to ameliorate HCV-associated insulin resistance, various therapeutic approaches are required.

# **REFERENCES**

- Allison ME, Wreghitt T, Palmer CR, Alexander GJ. Evidence for a link between hepatitis C virus infection and diabetes mellitus in a cirrhotic population. J Hepatol 1994; 21: 1135-1139
- 2 Caronia S, Taylor K, Pagliaro L, Carr C, Palazzo U, Petrik J, O'Rahilly S, Shore S, Tom BD, Alexander GJ. Further evidence for an association between non-insulin-dependent diabetes mellitus and chronic hepatitis C virus infection. *Hepatology* 1999; 30: 1059-1063
- 3 Aytug S, Reich D, Sapiro LE, Bernstein D, Begum N. Impaired IRS-1/PI3-kinase signaling in patients with HCV: a mechanism for increased prevalence of type 2 diabetes. *Hepatology* 2003; **38**: 1384-1392
- 4 Furutani M, Nakashima T, Sumida Y, Hirohama A, Yoh T, Kakisaka Y, Mitsuyoshi H, Senmaru H, Okanoue T. Insulin resistance/beta-cell function and serum ferritin level in nondiabetic patients with hepatitis C virus infection. *Liver Int* 2003; 23: 294-299
- Maeno T, Okumura A, Ishikawa T, Kato K, Sakakibara F, Sato K, Ayada M, Hotta N, Tagaya T, Fukuzawa Y, Kakumu S. Mechanisms of increased insulin resistance in non-cirrhotic patients with chronic hepatitis C virus infection. J Gastroenterol Hepatol 2003; 18: 1358-1363
- 6 Kawaguchi T, Yoshida T, Harada M, Hisamoto T, Nagao Y, Ide T, Taniguchi E, Kumemura H, Hanada S, Maeyama M, Baba S, Koga H, Kumashiro R, Ueno T, Ogata H, Yoshimura A, Sata M. Hepatitis C virus down-regulates insulin receptor substrates 1 and 2 through up-regulation of suppressor of cytokine signaling 3. Am J Pathol 2004; 165: 1499-1508
- 7 Narita R, Abe S, Kihara Y, Akiyama T, Tabaru A, Otsuki M. Insulin resistance and insulin secretion in chronic hepatitis C virus infection. J Hepatol 2004; 41: 132-138
- 8 Shintani Y, Fujie H, Miyoshi H, Tsutsumi T, Tsukamoto K, Kimura S, Moriya K, Koike K. Hepatitis C virus infection and diabetes: direct involvement of the virus in the development of insulin resistance. *Gastroenterology* 2004; 126: 840-848
- 9 Negro F. Mechanisms and significance of liver steatosis in hepatitis C virus infection. World J Gastroenterol 2006; 12: 6756-6765
- 10 Romero-Gómez M. Insulin resistance and hepatitis C. World J Gastroenterol 2006; 12: 7075-7080
- Taura N, Ichikawa T, Hamasaki K, Nakao K, Nishimura D, Goto T, Fukuta M, Kawashimo H, Fujimoto M, Kusumoto K, Motoyoshi Y, Shibata H, Abiru N, Yamasaki H, Eguchi K. Association between liver fibrosis and insulin sensitivity in chronic hepatitis C patients. Am J Gastroenterol 2006; 101: 2752-2759
- 12 Tuma P, Vispo E, Barreiro P, Soriano V. [Role of tenofovir in HIV and hepatitis C virus coinfection] Enferm Infecc Microbiol Clin 2008; 26 Suppl 8: 31-37

- Akuta N, Suzuki F, Hirakawa M, Kawamura Y, Yatsuji H, Sezaki H, Suzuki Y, Hosaka T, Kobayashi M, Kobayashi M, Saitoh S, Arase Y, Ikeda K, Kumada H. Amino acid substitutions in the hepatitis C virus core region of genotype 1b are the important predictor of severe insulin resistance in patients without cirrhosis and diabetes mellitus. J Med Virol 2009; 81: 1032-1039
- 14 Eguchi Y, Mizuta T, Ishibashi E, Kitajima Y, Oza N, Nakashita S, Hara M, Iwane S, Takahashi H, Akiyama T, Ario K, Kawaguchi Y, Yasutake T, Iwakiri R, Ozaki I, Hisatomi A, Eguchi T, Ono N, Fujimoto K. Hepatitis C virus infection enhances insulin resistance induced by visceral fat accumulation. *Liver Int* 2009; 29: 213-220
- 15 Kawaguchi T, Nagao Y, Tanaka K, Ide T, Harada M, Kumashiro R, Sata M. Causal relationship between hepatitis C virus core and the development of type 2 diabetes mellitus in a hepatitis C virus hyperendemic area: a pilot study. *Int J Mol Med* 2005; 16: 109-114
- Bianchi G, Marchesini G, Zoli M, Bugianesi E, Fabbri A, Pisi E. Prognostic significance of diabetes in patients with cirrhosis. *Hepatology* 1994; 20: 119-125
- 17 Davila JA, Morgan RO, Shaib Y, McGlynn KA, El-Serag HB. Diabetes increases the risk of hepatocellular carcinoma in the United States: a population based case control study. *Gut* 2005; **54**: 533-539
- 18 El-Serag HB. Hepatocellular carcinoma: recent trends in the United States. Gastroenterology 2004; 127: S27-S34
- 19 Kawaguchi T, Yamagishi S, Sata M. Branched-chain amino acids and pigment epithelium-derived factor: novel therapeutic agents for hepatitis c virus-associated insulin resistance. Curr Med Chem 2009; 16: 4843-4857
- 20 Sumie S, Kawaguchi T, Komuta M, Kuromatsu R, Itano S, Okuda K, Taniguchi E, Ando E, Takata A, Fukushima N, Koga H, Torimura T, Kojiro M, Sata M. Significance of glucose intolerance and SHIP2 expression in hepatocellular carcinoma patients with HCV infection. Oncol Rep 2007; 18: 545-552
- 21 Tazawa J, Maeda M, Nakagawa M, Ohbayashi H, Kusano F, Yamane M, Sakai Y, Suzuki K. Diabetes mellitus may be associated with hepatocarcinogenesis in patients with chronic hepatitis C. *Dig Dis Sci* 2002; 47: 710-715
- 22 Nagao Y, Kawaguchi T, Tanaka K, Kumashiro R, Sata M. Extrahepatic manifestations and insulin resistance in an HCV hyperendemic area. Int J Mol Med 2005; 16: 291-296
- Nagao Y, Kawasaki K, Sata M. Insulin resistance and lichen planus in patients with HCV-infectious liver diseases. J Gastroenterol Hepatol 2008; 23: 580-585
- 24 Nagao Y, Sata M. High incidence of multiple primary carcinomas in HCV-infected patients with oral squamous cell carcinoma. Med Sci Monit 2009; 15: CR453-CR459
- 25 Franz MJ, Bantle JP, Beebe CA, Brunzell JD, Chiasson JL, Garg A, Holzmeister LA, Hoogwerf B, Mayer-Davis E, Mooradian AD, Purnell JQ, Wheeler M. Evidence-based nutrition principles and recommendations for the treatment and prevention of diabetes and related complications. Diabetes Care 2002; 25: 148-198
- 26 Global Guideline for Type 2 Diabetes: recommendations for standard, comprehensive, and minimal care. *Diabet Med* 2006; 23: 579-593
- 27 Itou M, Kawaguchi T, Taniguchi E, Sumie S, Oriishi T, Mitsuyama K, Tsuruta O, Ueno T, Sata M. Altered expression of glucagon-like peptide-1 and dipeptidyl peptidase IV in patients with HCV-related glucose intolerance. J Gastroenterol Hepatol 2008; 23: 244-251
- Qamar AA, Grace ND, Groszmann RJ, Garcia-Tsao G, Bosch J, Burroughs AK, Ripoll C, Maurer R, Planas R, Escorsell A, Garcia-Pagan JC, Patch D, Matloff DS, Makuch R, Rendon G. Incidence, prevalence, and clinical significance of abnormal hematologic indices in compensated cirrhosis. Clin Gastroenterol Hepatol 2009; 7: 689-695
- 29 Donadon V, Balbi M, Ghersetti M, Grazioli S, Perciaccante A, Della Valentina G, Gardenal R, Dal Mas M, Casarin P,

- Zanette G, Miranda C. Antidiabetic therapy and increased risk of hepatocellular carcinoma in chronic liver disease. *World J Gastroenterol* 2009; **15**: 2506-2511
- 30 Kawaguchi T, Taniguchi E, Morita Y, Shirachi M, Tateishi I, Nagata E, Sata M. Association of exogenous insulin or sulphonylurea treatment with an increased incidence of hepatoma in patients with hepatitis C virus infection. Liver Int 2009; Epub ahead of print
- 31 Jansson SP, Andersson DK, Svärdsudd K. Mortality trends in subjects with and without diabetes during 33 years of follow-up. Diabetes Care 2010; 33: 551-556
- 32 Negro F, Sanyal AJ. Hepatitis C virus, steatosis and lipid abnormalities: clinical and pathogenic data. Liver Int 2009; 29 Suppl 2: 26-37
- 33 Ong JP, Younossi ZM, Speer C, Olano A, Gramlich T, Boparai N. Chronic hepatitis C and superimposed nonalcoholic fatty liver disease. *Liver* 2001; 21: 266-271
- 34 Leandro G, Mangia A, Hui J, Fabris P, Rubbia-Brandt L, Colloredo G, Adinolfi LE, Asselah T, Jonsson JR, Smedile A, Terrault N, Pazienza V, Giordani MT, Giostra E, Sonzogni A, Ruggiero G, Marcellin P, Powell EE, George J, Negro F. Relationship between steatosis, inflammation, and fibrosis in chronic hepatitis C: a meta-analysis of individual patient data. Gastroenterology 2006; 130: 1636-1642
- 35 Korenaga M, Wang T, Li Y, Showalter LA, Chan T, Sun J, Weinman SA. Hepatitis C virus core protein inhibits mitochondrial electron transport and increases reactive oxygen species (ROS) production. J Biol Chem 2005; 280: 37481-37488
- McPherson S, Jonsson JR, Barrie HD, O'Rourke P, Clouston AD, Powell EE. Investigation of the role of SREBP-1c in the pathogenesis of HCV-related steatosis. J Hepatol 2008; 49: 1046-1054
- 37 Negro F. Peroxisome proliferator-activated receptors and hepatitis C virus-induced insulin resistance. PPAR Res 2009; 2009: 483485
- 38 Perlemuter G, Sabile A, Letteron P, Vona G, Topilco A, Chrétien Y, Koike K, Pessayre D, Chapman J, Barba G, Bréchot C. Hepatitis C virus core protein inhibits microsomal triglyceride transfer protein activity and very low density lipoprotein secretion: a model of viral-related steatosis. FASEB J 2002; 16: 185-194
- 39 Geelen MJ, Harris RA, Beynen AC, McCune SA. Short-term hormonal control of hepatic lipogenesis. *Diabetes* 1980; 29: 1006-1022
- 40 Koike K. Hepatitis C as a metabolic disease: Implication for the pathogenesis of NASH. Hepatol Res 2005; 33: 145-150
- 41 Kawaguchi T, Osatomi K, Yamashita H, Kabashima T, Uyeda K. Mechanism for fatty acid "sparing" effect on glucose-induced transcription: regulation of carbohydrateresponsive element-binding protein by AMP-activated protein kinase. J Biol Chem 2002; 277: 3829-3835
- 42 Kawaguchi T, Takenoshita M, Kabashima T, Uyeda K. Glucose and cAMP regulate the L-type pyruvate kinase gene by phosphorylation/dephosphorylation of the carbohydrate response element binding protein. Proc Natl Acad Sci USA 2001; 98: 13710-13715
- 43 Cammà C, Bruno S, Di Marco V, Di Bona D, Rumi M, Vinci M, Rebucci C, Cividini A, Pizzolanti G, Minola E, Mondelli MU, Colombo M, Pinzello G, Craxì A. Insulin resistance is associated with steatosis in nondiabetic patients with genotype 1 chronic hepatitis C. Hepatology 2006; 43: 64-71
- 44 Kawaguchi T, Ide T, Taniguchi E, Hirano E, Itou M, Sumie S, Nagao Y, Yanagimoto C, Hanada S, Koga H, Sata M. Clearance of HCV improves insulin resistance, beta-cell function, and hepatic expression of insulin receptor substrate 1 and 2. Am | Gastroenterol 2007; 102: 570-576
- 45 Poustchi H, Negro F, Hui J, Cua IH, Brandt LR, Kench JG, George J. Insulin resistance and response to therapy in patients infected with chronic hepatitis C virus genotypes 2 and 3. J Hepatol 2008; 48: 28-34

- 46 Romero-Gómez M, Del Mar Viloria M, Andrade RJ, Salmerón J, Diago M, Fernández-Rodríguez CM, Corpas R, Cruz M, Grande L, Vázquez L, Muñoz-De-Rueda P, López-Serrano P, Gila A, Gutiérrez ML, Pérez C, Ruiz-Extremera A, Suárez E, Castillo J. Insulin resistance impairs sustained response rate to peginterferon plus ribavirin in chronic hepatitis C patients. Gastroenterology 2005; 128: 636-641
- 47 Shimomura I, Matsuda M, Hammer RE, Bashmakov Y, Brown MS, Goldstein JL. Decreased IRS-2 and increased SREBP-1c lead to mixed insulin resistance and sensitivity in livers of lipodystrophic and ob/ob mice. Mol Cell 2000; 6: 77-86
- 48 Miyanari Y, Atsuzawa K, Usuda N, Watashi K, Hishiki T, Zayas M, Bartenschlager R, Wakita T, Hijikata M, Shimotohno K. The lipid droplet is an important organelle for hepatitis C virus production. Nat Cell Biol 2007; 9: 1089-1097
- 49 Vlotides G, Sörensen AS, Kopp F, Zitzmann K, Cengic N, Brand S, Zachoval R, Auernhammer CJ. SOCS-1 and SOCS-3 inhibit IFN-alpha-induced expression of the antiviral proteins 2,5-OAS and MxA. Biochem Biophys Res Commun 2004; 320: 1007-1014
- Pazienza V, Clément S, Pugnale P, Conzelman S, Foti M, Mangia A, Negro F. The hepatitis C virus core protein of genotypes 3a and 1b downregulates insulin receptor substrate 1 through genotype-specific mechanisms. *Hepatology* 2007; 45: 1164-1171
- 51 Persico M, Capasso M, Persico E, Svelto M, Russo R, Spano D, Crocè L, La Mura V, Moschella F, Masutti F, Torella R, Tiribelli C, Iolascon A. Suppressor of cytokine signaling 3 (SOCS3) expression and hepatitis C virus-related chronic hepatitis: Insulin resistance and response to antiviral therapy. Hepatology 2007; 46: 1009-1015
- 52 Del Campo JA, Romero-Gómez M. Steatosis and insulin resistance in hepatitis C: a way out for the virus? World J Gastroenterol 2009; 15: 5014-5019
- 53 Miyaaki H, Ichikawa T, Nakao K, Matsuzaki T, Muraoka T, Honda T, Takeshita S, Shibata H, Ozawa E, Akiyama M, Miuma S, Eguchi K. Predictive value of suppressor of cytokine signal 3 (SOCS3) in the outcome of interferon therapy in chronic hepatitis C. Hepatol Res 2009; 39: 850-855
- 54 Walsh MJ, Jonsson JR, Richardson MM, Lipka GM, Purdie DM, Clouston AD, Powell EE. Non-response to antiviral therapy is associated with obesity and increased hepatic expression of suppressor of cytokine signalling 3 (SOCS-3) in patients with chronic hepatitis C, viral genotype 1. Gut 2006; 55: 529-535
- Muzzi A, Leandro G, Rubbia-Brandt L, James R, Keiser O, Malinverni R, Dufour JF, Helbling B, Hadengue A, Gonvers JJ, Müllhaupt B, Cerny A, Mondelli MU, Negro F. Insulin resistance is associated with liver fibrosis in non-diabetic chronic hepatitis C patients. J Hepatol 2005; 42: 41-46
- 56 Petrides AS, Vogt C, Schulze-Berge D, Matthews D, Strohmeyer G. Pathogenesis of glucose intolerance and diabetes mellitus in cirrhosis. *Hepatology* 1994; 19: 616-627
- 57 Veldt BJ, Poterucha JJ, Watt KD, Wiesner RH, Hay JE, Rosen CB, Heimbach JK, Janssen HL, Charlton MR. Insulin resistance, serum adipokines and risk of fibrosis progression in patients transplanted for hepatitis C. Am J Transplant 2009; 9: 1406-1413
- Paradis V, Perlemuter G, Bonvoust F, Dargere D, Parfait B, Vidaud M, Conti M, Huet S, Ba N, Buffet C, Bedossa P. High glucose and hyperinsulinemia stimulate connective tissue growth factor expression: a potential mechanism involved in progression to fibrosis in nonalcoholic steatohepatitis. Hepatology 2001; 34: 738-744
- Cammà C, Petta S, Di Marco V, Bronte F, Ciminnisi S, Licata G, Peralta S, Simone F, Marchesini G, Craxi A. Insulin resistance is a risk factor for esophageal varices in hepatitis C virus cirrhosis. *Hepatology* 2009; 49: 195-203
- 60 Svegliati-Baroni G, De Minicis S, Marzioni M. Hepatic fibro-

- genesis in response to chronic liver injury: novel insights on the role of cell-to-cell interaction and transition. *Liver Int* 2008; 28: 1052-1064
- 61 Vincent MA, Montagnani M, Quon MJ. Molecular and physiologic actions of insulin related to production of nitric oxide in vascular endothelium. Curr Diab Rep 2003; 3: 279-288
- 62 Iwakiri Y, Groszmann RJ. Vascular endothelial dysfunction in cirrhosis. *J Hepatol* 2007; 46: 927-934
- 63 **El-Serag HB**. Epidemiology of hepatocellular carcinoma in USA. *Hepatol Res* 2007; **37** Suppl 2: S88-S94
- 64 El-Serag HB, Tran T, Everhart JE. Diabetes increases the risk of chronic liver disease and hepatocellular carcinoma. Gastroenterology 2004; 126: 460-468
- 65 Marra F, Bertolani C. Adipokines in liver diseases. *Hepatology* 2009; 50: 957-969
- 66 Saito K, Inoue S, Saito T, Kiso S, Ito N, Tamura S, Watanabe H, Takeda H, Misawa H, Togashi H, Matsuzawa Y, Kawata S. Augmentation effect of postprandial hyperinsulinaemia on growth of human hepatocellular carcinoma. Gut 2002; 51: 100-104
- 67 Nehrbass D, Klimek F, Bannasch P. Overexpression of insulin receptor substrate-1 emerges early in hepatocarcinogenesis and elicits preneoplastic hepatic glycogenosis. Am J Pathol 1998; 152: 341-345
- 68 Boissan M, Beurel E, Wendum D, Rey C, Lécluse Y, Housset C, Lacombe ML, Desbois-Mouthon C. Overexpression of insulin receptor substrate-2 in human and murine hepatocellular carcinoma. Am J Pathol 2005; 167: 869-877
- 69 Yao YJ, Ping XL, Zhang H, Chen FF, Lee PK, Ahsan H, Chen CJ, Lee PH, Peacocke M, Santella RM, Tsou HC. PTEN/MMAC1 mutations in hepatocellular carcinomas. Oncogene 1999; 18: 3181-3185
- 70 Nagao Y, Sata M, Noguchi S, Tajiri N, Ono N, Fukuda T, Kameyama T, Ueno T. Various extrahepatic manifestations caused by hepatitis C virus infection. *Int J Mol Med* 1999; 4: 621-625
- 71 Nagao Y, Sata M, Tanikawa K, Itoh K, Kameyama T. Lichen planus and hepatitis C virus in the northern Kyushu region of Japan. *Eur J Clin Invest* 1995; **25**: 910-914
- 72 Nagao Y, Tanaka J, Nakanishi T, Moriya T, Katayama K, Kumagai J, Komiya Y, Itoh Y, Myoken Y, Fujihara M, Sata M, Yoshizawa H. High incidence of extrahepatic manifestations in an HCV hyperendemic area. Hepatol Res 2002; 22: 27-36
- 73 Ohtsubo K, Sata M, Kawaguchi T, Morishige S, Takata Y, Oku E, Imamura R, Seki R, Hashiguchi M, Osaki K, Yakushiji K, Kanaji T, Yoshimoto K, Ueno T, Okamura T. Characterization of the light chain-restricted clonal B cells in peripheral blood of HCV-positive patients. *Int J Hematol* 2009; 89: 452-459
- 74 Husseini A, Abu-Rmeileh NM, Mikki N, Ramahi TM, Ghosh HA, Barghuthi N, Khalili M, Bjertness E, Holmboe-Ottesen G, Jervell J. Cardiovascular diseases, diabetes mellitus, and cancer in the occupied Palestinian territory. *Lancet* 2009; 373: 1041-1049
- 75 Ship JA. Diabetes and oral health: an overview. J Am Dent Assoc 2003; 134 Spec No: 4S-10S
- 76 Kawaguchi Y, Mizuta T, Oza N, Takahashi H, Ario K, Yoshimura T, Eguchi Y, Ozaki I, Hisatomi A, Fujimoto K. Eradication of hepatitis C virus by interferon improves whole-body insulin resistance and hyperinsulinaemia in patients with chronic hepatitis C. Liver Int 2009; 29: 871-877
- 77 ASPEN Board of Directors and the Clinical Guidelines Task Force. Guidelines for the use of parenteral and enteral nutrition in adult and pediatric patients. *JPEN J Parenter Enteral Nutr* 2002; 26: 1SA-138SA
- 78 Kato A, Suzuki K. How to select BCAA preparations. Hepatol Res 2004; 30S: 30-35
- 79 Plauth M, Cabré E, Riggio O, Assis-Camilo M, Pirlich M, Kondrup J, Ferenci P, Holm E, Vom Dahl S, Müller MJ,

- Nolte W. ESPEN Guidelines on Enteral Nutrition: Liver disease. *Clin Nutr* 2006; **25**: 285-294
- 80 Tajika M, Kato M, Mohri H, Miwa Y, Kato T, Ohnishi H, Moriwaki H. Prognostic value of energy metabolism in patients with viral liver cirrhosis. *Nutrition* 2002; 18: 229-234
- 81 Korenaga K, Korenaga M, Uchida K, Yamasaki T, Sakaida I. Effects of a late evening snack combined with alphaglucosidase inhibitor on liver cirrhosis. Hepatol Res 2008; 38: 1087-1097
- 82 Okamoto M, Sakaida I, Tsuchiya M, Suzuki C, Okita K. Effect of a late evening snack on the blood glucose level and energy metabolism in patients with liver cirrhosis. Hepatol Res 2003; 27: 45-50
- 83 Sakaida I, Tsuchiya M, Okamoto M, Okita K. Late evening snack and the change of blood glucose level in patients with liver cirrhosis. Hepatol Res 2004; 30S: 67-72
- 84 Tsuchiya M, Sakaida I, Okamoto M, Okita K. The effect of a late evening snack in patients with liver cirrhosis. Hepatol Res 2005; 31: 95-103
- 85 Ohfuji S, Fukushima W, Tanaka T, Habu D, Takeda T, Tamori A, Sakaguchi H, Seki S, Kawada N, Nishiguchi S, Shiomi S, Hirota Y. Does a late evening meal reduce the risk of hepatocellular carcinoma among patients with chronic hepatitis C? *Hepatol Res* 2008; 38: 860-868
- Ruhl CE, Everhart JE. Coffee and caffeine consumption reduce the risk of elevated serum alanine aminotransferase activity in the United States. Gastroenterology 2005; 128: 24-32
- 87 Modi AA, Feld JJ, Park Y, Kleiner DE, Everhart JE, Liang TJ, Hoofnagle JH. Increased caffeine consumption is associated with reduced hepatic fibrosis. *Hepatology* 2010; 51: 201-209
- Freedman ND, Everhart JE, Lindsay KL, Ghany MG, Curto TM, Shiffman ML, Lee WM, Lok AS, Di Bisceglie AM, Bonkovsky HL, Hoefs JC, Dienstag JL, Morishima C, Abnet CC, Sinha R. Coffee intake is associated with lower rates of liver disease progression in chronic hepatitis C. Hepatology 2009; 50: 1360-1369
- 89 Gelatti U, Covolo L, Franceschini M, Pirali F, Tagger A, Ribero ML, Trevisi P, Martelli C, Nardi G, Donato F. Coffee consumption reduces the risk of hepatocellular carcinoma independently of its aetiology: a case-control study. J Hepatol 2005; 42: 528-534
- 90 Tunnicliffe JM, Shearer J. Coffee, glucose homeostasis, and insulin resistance: physiological mechanisms and mediators. Appl Physiol Nutr Metab 2008; 33: 1290-1300
- 91 Shearer J, Farah A, de Paulis T, Bracy DP, Pencek RR, Graham TE, Wasserman DH. Quinides of roasted coffee enhance insulin action in conscious rats. J Nutr 2003; 133: 3529-3532
- 92 Naismith DJ, Akinyanju PA, Szanto S, Yudkin J. The effect, in volunteers, of coffee and decaffeinated coffee on blood glucose, insulin, plasma lipids and some factors involved in blood clotting. Nutr Metab 1970; 12: 144-151
- 93 Gerin I, Van Schaftingen E. Evidence for glucose-6-phosphate transport in rat liver microsomes. FEBS Lett 2002; 517: 257-260
- 94 Welsch CA, Lachance PA, Wasserman BP. Dietary phenolic compounds: inhibition of Na+-dependent D-glucose uptake in rat intestinal brush border membrane vesicles. J Nutr 1989; 119: 1698-1704
- 95 Johnston KL, Clifford MN, Morgan LM. Coffee acutely modifies gastrointestinal hormone secretion and glucose tolerance in humans: glycemic effects of chlorogenic acid and caffeine. Am J Clin Nutr 2003; 78: 728-733
- 96 McCarty MF. A chlorogenic acid-induced increase in GLP-1 production may mediate the impact of heavy coffee consumption on diabetes risk. Med Hypotheses 2005; 64: 848-853
- 97 **Heyden S**. Does coffee influence the lipid metabolism? *Z Ernahrungswiss* 1969; 9: 388-396
- 98 Zeller W. [Effect of coffee on fat metabolism] Med Klin 1968; 63: 707-709

- 99 Garrido Serrano A, Guerrero Igea FJ, Lepe Jiménez JA, Palomo Gil S, Grilo Reina A. Hepatitis C virus infection, increased serum ferritin and hyperinsulinemia. Rev Esp Enferm Dig 2001; 93: 639-648
- 100 Lecube A, Hernández C, Simó R. Glucose abnormalities in non-alcoholic fatty liver disease and chronic hepatitis C virus infection: the role of iron overload. *Diabetes Metab Res* Rev 2009; 25: 403-410
- 101 Mitsuyoshi H, Itoh Y, Sumida Y, Minami M, Yasui K, Nakashima T, Okanoue T. Evidence of oxidative stress as a cofactor in the development of insulin resistance in patients with chronic hepatitis C. Hepatol Res 2008; 38: 348-353
- 102 Nishina S, Hino K, Korenaga M, Vecchi C, Pietrangelo A, Mizukami Y, Furutani T, Sakai A, Okuda M, Hidaka I, Okita K, Sakaida I. Hepatitis C virus-induced reactive oxygen species raise hepatic iron level in mice by reducing hepcidin transcription. Gastroenterology 2008; 134: 226-238
- 103 Fujita N, Sugimoto R, Takeo M, Urawa N, Mifuji R, Tanaka H, Kobayashi Y, Iwasa M, Watanabe S, Adachi Y, Kaito M. Hepcidin expression in the liver: relatively low level in patients with chronic hepatitis C. Mol Med 2007; 13: 97-104
- 104 Nagashima M, Kudo M, Chung H, Ishikawa E, Hagiwara S, Nakatani T, Dote K. Regulatory failure of serum prohepcidin levels in patients with hepatitis C. Hepatol Res 2006; 36: 288-293
- 105 Piperno A, Mariani R, Trombini P, Girelli D. Hepcidin modulation in human diseases: from research to clinic. World J Gastroenterol 2009; 15: 538-551
- 106 Takeo M, Kobayashi Y, Fujita N, Urawa N, Iwasa M, Horiike S, Tanaka H, Kaito M, Adachi Y. Upregulation of transferrin receptor 2 and ferroportin 1 mRNA in the liver of patients with chronic hepatitis C. J Gastroenterol Hepatol 2005; 20: 562-569
- 107 Iwasa M, Iwata K, Kaito M, Ikoma J, Yamamoto M, Takeo M, Kuroda M, Fujita N, Kobayashi Y, Adachi Y. Efficacy of long-term dietary restriction of total calories, fat, iron, and protein in patients with chronic hepatitis C virus. Nutrition 2004; 20: 368-371
- 108 Fujita N, Miyachi H, Tanaka H, Takeo M, Nakagawa N, Kobayashi Y, Iwasa M, Watanabe S, Takei Y. Iron overload is associated with hepatic oxidative damage to DNA in nonalcoholic steatohepatitis. Cancer Epidemiol Biomarkers Prev 2009; 18: 424-432
- 109 Kaito M, Iwasa M, Kobayashi Y, Fujita N, Tanaka H, Gabazza EC, Adachi Y, Kojima Y, Nakagawa N, Watanabe S. Iron reduction therapy by phlebotomy reduces lipid peroxidation and oxidative stress in patients with chronic hepatitis C. J Gastroenterol 2006; 41: 921-922
- 110 Kato J, Miyanishi K, Kobune M, Nakamura T, Takada K, Takimoto R, Kawano Y, Takahashi S, Takahashi M, Sato Y, Takayama T, Niitsu Y. Long-term phlebotomy with low-iron diet therapy lowers risk of development of hepatocellular carcinoma from chronic hepatitis C. J Gastroenterol 2007; 42: 830-836
- 111 Hayashi M, Ikezawa K, Ono A, Okabayashi S, Hayashi Y, Shimizu S, Mizuno T, Maeda K, Akasaka T, Naito M, Michida T, Ueshima D, Nada T, Kawaguchi K, Nakamura T, Katayama K. Evaluation of the effects of combination therapy with branched-chain amino acid and zinc supplements on nitrogen metabolism in liver cirrhosis. Hepatol Res 2007; 37: 615-619
- 112 Katayama K. Ammonia metabolism and hepatic encephalopathy. Hepatol Res 2004; 30S: 73-80
- 113 Marreiro DN, Geloneze B, Tambascia MA, Lerário AC, Halpern A, Cozzolino SM. [Role of zinc in insulin resistance] Arg Bras Endocrinol Metabol 2004; 48: 234-239
- 114 Marchesini G, Bugianesi E, Ronchi M, Flamia R, Thomaseth K, Pacini G. Zinc supplementation improves glucose disposal in patients with cirrhosis. Metabolism 1998; 47: 792-798
- 115 Himoto T, Hosomi N, Nakai S, Deguchi A, Kinekawa F,

- Matsuki M, Yachida M, Masaki T, Kurokochi K, Watanabe S, Senda S, Kuriyama S. Efficacy of zinc administration in patients with hepatitis C virus-related chronic liver disease. *Scand J Gastroenterol* 2007; **42**: 1078-1087
- 116 Takahashi M, Saito H, Higashimoto M, Hibi T. Possible inhibitory effect of oral zinc supplementation on hepatic fibrosis through downregulation of TIMP-1: A pilot study. Hepatol Res 2007; 37: 405-409
- 117 Matsuoka S, Matsumura H, Nakamura H, Oshiro S, Arakawa Y, Hayashi J, Sekine N, Nirei K, Yamagami H, Ogawa M, Nakajima N, Amaki S, Tanaka N, Moriyama M. Zinc supplementation improves the outcome of chronic hepatitis C and liver cirrhosis. J Clin Biochem Nutr 2009; 45: 292-303
- 118 Chausmer AB. Zinc, insulin and diabetes. J Am Coll Nutr 1998; 17: 109-115
- 119 Ajjan RA, Grant PJ. Cardiovascular disease prevention in patients with type 2 diabetes: The role of oral anti-diabetic agents. Diab Vasc Dis Res 2006; 3: 147-158
- Patel A, MacMahon S, Chalmers J, Neal B, Billot L, Woodward M, Marre M, Cooper M, Glasziou P, Grobbee D, Hamet P, Harrap S, Heller S, Liu L, Mancia G, Mogensen CE, Pan C, Poulter N, Rodgers A, Williams B, Bompoint S, de Galan BE, Joshi R, Travert F. Intensive blood glucose control and vascular outcomes in patients with type 2 diabetes. N Engl J Med 2008; 358: 2560-2572
- 121 Komura T, Mizukoshi E, Kita Y, Sakurai M, Takata Y, Arai K, Yamashita T, Ohta T, Shimizu K, Nakamoto Y, Honda M, Takamura T, Kaneko S. Impact of diabetes on recurrence of hepatocellular carcinoma after surgical treatment in patients with viral hepatitis. Am J Gastroenterol 2007; 102: 1939-1946
- 122 Yang YX, Hennessy S, Lewis JD. Insulin therapy and colorectal cancer risk among type 2 diabetes mellitus patients. *Gastroenterology* 2004; **127**: 1044-1050
- 123 Colhoun HM. Use of insulin glargine and cancer incidence in Scotland: a study from the Scottish Diabetes Research Network Epidemiology Group. Diabetologia 2009; 52: 1755-1765
- 124 Kath R, Schiel R, Müller UA, Höffken K. Malignancies in patients with insulin-treated diabetes mellitus. J Cancer Res Clin Oncol 2000; 126: 412-417
- 125 Fogli-Cawley JJ, Dwyer JT, Saltzman E, McCullough ML, Troy LM, Meigs JB, Jacques PF. The 2005 Dietary Guidelines for Americans and insulin resistance in the Framingham Offspring Cohort. Diabetes Care 2007; 30: 817-822
- 126 Graham I, Atar D, Borch-Johnsen K, Boysen G, Burell G, Cifkova R, Dallongeville J, De Backer G, Ebrahim S, Gjelsvik B, Herrmann-Lingen C, Hoes A, Humphries S, Knapton M, Perk J, Priori SG, Pyorala K, Reiner Z, Ruilope L, Sans-Menendez S, Op Reimer WS, Weissberg P, Wood D, Yarnell J, Zamorano JL, Walma E, Fitzgerald T, Cooney MT, Dudina A, Vahanian A, Camm J, De Caterina R, Dean V, Dickstein K, Funck-Brentano C, Filippatos G, Hellemans I, Kristensen SD, McGregor K, Sechtem U, Silber S, Tendera M, Widimsky P, Zamorano JL, Altiner A, Bonora E, Durrington PN, Fagard R, Giampaoli S, Hemingway H, Hakansson J, Kjeldsen SE, Larsen ML, Mancia G, Manolis AJ, Orth-Gomer K, Pedersen T, Rayner M, Ryden L, Sammut M, Schneiderman N, Stalenhoef AF, Tokgözoglu L, Wiklund O, Zampelas A. European guidelines on cardiovascular disease prevention in clinical practice: executive summary. Fourth Joint Task Force of the European Society of Cardiology and other societies on cardiovascular disease prevention in clinical practice (constituted by representatives of nine societies and by invited experts). Eur I Cardiovasc Prev Rehabil 2007; 14 Suppl 2: E1-E40
- 127 Haupt A, Häring HU, Matthaei S. [National practice guidelines. New aspects in therapy of type 2 diabetes mellitus] MMW Fortschr Med 2003; 145: 41-46
- 128 Melkersson KI, Dahl ML, Hulting AL. Guidelines for prevention and treatment of adverse effects of antipsychotic drugs on glucose-insulin homeostasis and lipid metabolism.

- Psychopharmacology (Berl) 2004; 175: 1-6
- 129 Romero-Gómez M, Diago M, Andrade RJ, Calleja JL, Salmerón J, Fernández-Rodríguez CM, Solà R, García-Samaniego J, Herrerías JM, De la Mata M, Moreno-Otero R, Nuñez O, Olveira A, Durán S, Planas R. Treatment of insulin resistance with metformin in naïve genotype 1 chronic hepatitis C patients receiving peginterferon alfa-2a plus ribavirin. Hepatology 2009; 50: 1702-1708
- 130 Khattab M, Emad M, Abdelaleem A, Eslam M, Atef R, Shaker Y, Hamdy L. Pioglitazone improves virological response to peginterferon alpha-2b/ribavirin combination therapy in hepatitis C genotype 4 patients with insulin resistance. Liver Int 2009; Epub ahead of print
- 131 Smith U, Gogg S, Johansson A, Olausson T, Rotter V, Svalstedt B. Thiazolidinediones (PPARgamma agonists) but not PPARalpha agonists increase IRS-2 gene expression in 3T3-L1 and human adipocytes. FASEB J 2001; 15: 215-220
- 132 Yuan L, Ziegler R, Hamann A. Metformin modulates insulin post-receptor signaling transduction in chronically insulintreated Hep G2 cells. Acta Pharmacol Sin 2003; 24: 55-60
- 133 Bailey CJ, Turner RC. Metformin. N Engl J Med 1996; 334: 574-579
- 134 Shishido S, Koga H, Harada M, Kumemura H, Hanada S, Taniguchi E, Kumashiro R, Ohira H, Sato Y, Namba M, Ueno T, Sata M. Hydrogen peroxide overproduction in megamitochondria of troglitazone-treated human hepatocytes. *Hepatology* 2003; 37: 136-147
- 135 Deacon CF, Holst JJ. Dipeptidyl peptidase IV inhibitors: a promising new therapeutic approach for the management of type 2 diabetes. Int J Biochem Cell Biol 2006; 38: 831-844
- 136 DeFronzo RA, Fleck PR, Wilson CA, Mekki Q. Efficacy and safety of the dipeptidyl peptidase-4 inhibitor alogliptin in patients with type 2 diabetes and inadequate glycemic control: a randomized, double-blind, placebo-controlled study. Diabetes Care 2008; 31: 2315-2317
- 137 Suzuki K, Suzuki K, Koizumi K, Ichimura H, Oka S, Takada H, Kuwayama H. Measurement of serum branched-chain amino acids to tyrosine ratio level is useful in a prediction of a change of serum albumin level in chronic liver disease.

- Hepatol Res 2008; 38: 267-272
- 138 Moriwaki H, Miwa Y, Tajika M, Kato M, Fukushima H, Shiraki M. Branched-chain amino acids as a protein- and energy-source in liver cirrhosis. Biochem Biophys Res Commun 2004; 313: 405-409
- 139 Nishitani S, Takehana K, Fujitani S, Sonaka I. Branchedchain amino acids improve glucose metabolism in rats with liver cirrhosis. Am J Physiol Gastrointest Liver Physiol 2005; 288: G1292-G1300
- 140 Hinault C, Mothe-Satney I, Gautier N, Lawrence JC Jr, Van Obberghen E. Amino acids and leucine allow insulin activation of the PKB/mTOR pathway in normal adipocytes treated with wortmannin and in adipocytes from db/db mice. FASEB J 2004; 18: 1894-1896
- 141 Broca C, Breil V, Cruciani-Guglielmacci C, Manteghetti M, Rouault C, Derouet M, Rizkalla S, Pau B, Petit P, Ribes G, Ktorza A, Gross R, Reach G, Taouis M. Insulinotropic agent ID-1101 (4-hydroxyisoleucine) activates insulin signaling in rat. Am J Physiol Endocrinol Metab 2004; 287: E463-E471
- 142 She P, Reid TM, Bronson SK, Vary TC, Hajnal A, Lynch CJ, Hutson SM. Disruption of BCATm in mice leads to increased energy expenditure associated with the activation of a futile protein turnover cycle. Cell Metab 2007; 6: 181-194
- 143 Kawaguchi T, Nagao Y, Matsuoka H, Ide T, Sata M. Branchedchain amino acid-enriched supplementation improves insulin resistance in patients with chronic liver disease. *Int J Mol Med* 2008; 22: 105-112
- 144 Kawaguchi T, Taniguchi E, Itou M, Sumie S, Oriishi T, Matsuoka H, Nagao Y, Sata M. Branched-chain amino acids improve insulin resistance in patients with hepatitis C virusrelated liver disease: report of two cases. Liver Int 2007; 27: 1287-1292
- 145 Muto Y, Sato S, Watanabe A, Moriwaki H, Suzuki K, Kato A, Kato M, Nakamura T, Higuchi K, Nishiguchi S, Kumada H, Ohashi Y. Overweight and obesity increase the risk for liver cancer in patients with liver cirrhosis and long-term oral supplementation with branched-chain amino acid granules inhibits liver carcinogenesis in heavier patients with liver cirrhosis. Hepatol Res 2006; 35: 204-214
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# The Pathogenesis, Complications and Therapeutic Strategy for Hepatitis C Virus-associated Insulin Resistance in the Era of Anti-viral Treatment

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Abstract: Recent experimental and clinical studies have shown that chronic hepatitis C virus (HCV) infection causes insulin resistance. Since insulin resistance decreases response to antiviral treatments, promotes inflammatory and fibrogenic reactions and increases a risk of hepatocellular carcinoma (HCC), amelioration of insulin resistance may be a novel therapeutic target, which could improve the prognosis in patients with HCV-related chronic liver disease. Despite the increased awareness of health risk of insulin resistance, there is no common therapeutic strategy for HCV-associated insulin resistance. Indeed, treatments with exogenous insulin or sulfonylureas may be rather harmful because these treatments are associated with the development of HCC in patients with HCV infection. Meanwhile, we, along with others, have found distinctive treatments which improve HCV-associated insulin resistance. Administration of branched-chain amino acids (BCAA), especially as a late evening snack, improves glucose metabolism by improving insulin-signal cascades in insulin resistance patients with HCV infection. In this paper, we discuss the pathogenesis and complications for HCV-associated insulin resistance and further review a recent clinical therapeutic strategy using these agents for the treatment of this devastating disorder. We also discuss therapeutic potentialities of incretin-based therapies, new anti-diabetic agents for HCVassociated insulin resistance and the significance of insulin resistance in the era of new anti-viral treatments.

Keywords: Hepatitis C virus, insulin resistance, hepatocellular carcinoma, branched-chain amino acids, incretin.

### INTRODUCTION

Since hepatitis C virus (HCV) was identified in 1989 [1, 2], underlying pathophysiology of chronic hepatitis C has been disclosed tremendously [3-7]. Individuals infected with HCV frequently develop chronic infection, which is associated with the development of liver cirrhosis and hepatocellular carcinoma (HCC) [8-10]. In addition, epidemiological data from East-West show an association between HCV infection and insulin resistance [11-21]. Recent basic and clinical researches have revealed the mechanisms of HCVassociated insulin resistance and insulin resistance is now recognized as a sequela of chronic HCV infection [14, 22-30].

Generally, insulin resistance is associated with the development of diabetes mellitus (DM), hypertension and cardiovascular diseases [31]. Besides these complications, insulin resistance is also involved in many events in patients with chronic hepatitis C, including antiviral treatment, fibrogenic reaction and the development of HCC [25, 28, 32-35].

for the progression of any stage of chronic hepatitis C [36]. Despite the accumulated evidences for the risk of insulin resistance, therapeutic strategy for HCV-associated insulin resistance has not been established yet [37]. In HCV-infected patients with diabetes, cardio-vascular disease occupies only

It is now clear that insulin resistance is a critical factor RESISTANCE

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less than 5% of cause of death and is not a significant prognostic factor [38]. Prognostic factors are HCC, liver failure, and esophageal varices even in HCV-infected patients with diabetes [38], and the therapeutic strategy should be considered based on mechanisms and life-threatening complications for HCV-associated insulin resistance.

Recently, we have found that an inactivation of incretin is a causative factor for HCV-associated insulin resistance [39]. Incretin mimetics and incretin enhancer are new antidiabetic agents. Theoretically, it seems that replenishment and/or enhancement of incretin is proper therapeutic approach and these new anti-diabetic agents may ameliorate insulin resistance, prevent the development of HCC and improve the prognosis in patients with HCV infection.

In this review, we summarize the pathogenesis for HCVassociated insulin resistance and propose the clinical therapeutic strategy for the treatment of HCV-associated insulin resistance. In addition, therapeutic potentialities of incretinbased therapies and the significance of insulin resistance in the era of new anti-viral treatments are discussed.

# MECHANISMS FOR HCV-ASSOCIATED INSULIN

## **Indirect Effects of HCV**

Various factors are reported to be associated with the development of HCV-associated insulin resistance (Table 1). Similar to the life style-associated insulin resistance, obesity with decreased serum adiponectin levels [40-42], inflammatory cytokines [41, 43, 44], oxidative stress [45-47], hepatic steatosis [48], pancreatic beta-cell function [49], and serum

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Table 1. Factors Associated with Insulin Resistance in Patients with HCV Infection

Factor	References [40-42]	
Obesity and decreased serum adiponectin levels		
Inflammation and inflammatory cytokines	[41, 43, 44]	
Oxidative stress	[45-47]	
Decreased serum PEDF levels	[77, 187]	
Portal-systemic shunt	[50]	
Hepatic iron accumulation	[57, 58]	
HCV core	[14, 26, 29]	
Amino acid substitutions of the HCV core region (Gln70 (His70) and/or Met91)	[22]	

pigment epithelium-derived factor (PEDF) levels are also involved in the development of HCV-associated insulin resistance. In addition to those common factors, liver specific factors are underlying in the development of HCV-associated insulin resistance.

Ferenci *et al.* reported that portal-systemic shunts caused not only hepatic encephalopathy, but also insulin resistance in patients with liver cirrhosis [50]. Reduced hepatic blood flow may lead liver dysfunction and subsequent insulin resistance. In fact, Tanabe *et al.* reported that occlusion of portal-systemic shunt improved glucose metabolism in patients with liver cirrhosis [51].

Hepatic iron accumulation is one of characters for HCV infection. Hepcidin is a key negative regulator of iron metabolism [52] and hepcidin levels are correlate with hepatic iron accumulation in HCV transgenic mice [53] and patients with chronic hepatitis C [54, 55]. Excessive iron induces reactive oxygen species mediated oxidative stress [56]. In patients with HCV infection, hepatic iron accumulation is associated with insulin resistance [57, 58]. Furthermore, iron depletion by phlebotomy reduces serum and hepatic levels of thioredoxin, a marker of oxidative stress, and homeostasis model assessment-insulin resistance, an index for insulin resistance in patients with chronic hepatitis C [46]. Thus, hepatic iron accumulation may cause insulin resistance through induction of inflammatory cytokines and oxidative stress.

# Direct Effects of HCV on the Development of Insulin Resistance

Direct effects of HCV on the development of insulin resistance are still debatable. Tsochatzis E et al. reported that insulin resistance is not associated with viremia [59]. On the other hand, some previous studies reported that serum HCV RNA levels is associated with insulin resistance in a dose-dependent manner, independent of the visceral adipose tissue area [60-62] and HCV suppression by anti-viral treatment correlates with improvement in insulin resistance [63-65]. These findings suggest a possible role of HCV in the development of insulin resistance. Recently, HCV is now known to directly associate with insulin signaling molecules and cause insulin resistance. HCV core protein causes nuclear translocation of signal transducer and activation of transcription 3 and subsequent up-regulation of suppressor of cytokine signaling (SOCS) 3 proteins in various hepatoma cell

lines [26]. SOCS3 is known to block insulin signaling cascade by ubiquitin-mediated degradation of insulin receptor substrate (IRS)1/2 [66]. Down-regulation of IRS1/2 are seen in livers from HCV-core transgenic mice and in livers of patients with HCV infection [26] and therefore, HCV coreinduced SOCS3 up-regulation may promote proteasomal degradation of IRS1 and IRS2 through ubiquitination, leading to insulin resistance in patients with HCV infection [26, 67] (Fig. 1).

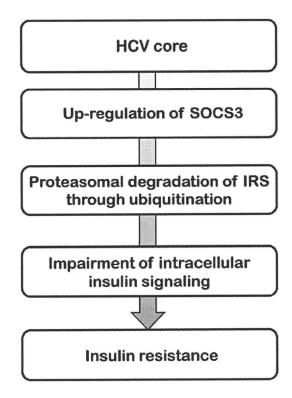


Fig. (1). Direct effects of HCV on the development of insulin resistance.

HCV core protein up-regulates suppressor of cytokine signaling (SOCS) 3 protein. SOCS3 ubiquitinates insulin receptor substrate (IRS) and causes proteasomal degradation of IRS. Since IRS is a central molecule of intarcellular insulin signaling, down-regulation of IRS blocks insulin signaling cascade, leading to insulin resistance in patients with HCV infection.

In addition to down-regulation of IRS1/2, HCV core protein also up-regulates serine phosphorylation of IRS1 or down-regulate tyrosine phosphorylation of IRS1 and subsequently impairs Akt signaling pathway, a downstream signaling of IRS1 in vitro [68] and in HCV-core transgenic mice [29]. Possible mechanism for changes in serine or tyrosine phosphorylation of IRS1 is due to increased activation of mammalian target of rapamycin [69] or Jun-N-terminal kinase [68]. Although changes in serine phosphorylation of IRS1 have not been validated in human liver tissue, downregulation in tyrosine phosphorylation of IRS1 and subsequent Akt signaling pathway has been reported in human liver tissue [24].

## **HCV Genotype and Insulin Resistance**

Although associations between HCV genotypes and the development of insulin resistance are still controversial [26, 59, 70, 71], HCV genotype-specific interactions of insulin resistance are reported in patients with HCV genotype 1b, 3a, and 4 [61, 69].

HCV genotype 1b significantly suppresses IRS1 expression compared to HCV genotype 2 in HepG2 cell [67]. The suppression of IRS1 expression is due to two different pathways. HCV genotype 1b up-regulates SOCS3, which causes proteasomal degradation of IRS1 [26]. Alternatively, HCV genotype 1 activates mammalian target of rapamycin, which suppress IRS1 expression [69]. Recently, Akuta et al. found that amino acid substitutions in the HCV core region of genotype 1b were associated with severe insulin resistance in patients without cirrhosis [22]. Although the precise mechanisms for the development to insulin resistance by amino acid substitutions in the HCV core region are unclear, HCV core seems to impair intracellular insulin signaling through several pathways.

In in vitro experiments, HCV genotype 3a modulates SOCS7 expression and causes down-regulation of IRS1 [69, 72]. In fact, patients infected with HCV genotype 3 frequently associated with insulin resistance [73]. In patients infected with HCV genotype 3, a decrease in total and high molecular weight adiponectin is another causative factor for the development of insulin resistance [74].

High prevalence of insulin resistance is also seen in patients infected with HCV genotype 4 and insulin resistance is a major determinant of both rapid virologic response and sustained virologic response [61, 75]. Normal BMI and no significant fibrosis are characters for patients HCV genotype 4 infected with insulin resistance [61, 76]. Although molecular mechanisms of HCV genotype 4 associated insulin resistance is not unclear, these findings suggest the genotype specific interaction with intracellular insulin signaling.

### COMPLICATIONS OF HCV-ASSOCIATED INSULIN RESISTANCE

Insulin regulates not only glucose metabolism, but also protein synthesis, lipid metabolism and cell proliferation through activation of various intracellular signaling molecules [77]. Therefore, insulin resistance is involved in not only the development of DM, but also non-response to anti-

viral treatment [25, 28] and hepatic fibrosis [32]. In addition, insulin resistance causes esophageal varices [78] and HCC [33-35], life-threatening complications, Recently, the development of lichen planus [79], multiple primary carcinomas [80], and other extrahepatic manifestations [81] are associated with insulin resistance. Thus, insulin resistance could play crucial roles in the development to variety of complications in patients with HCV infection. In this review, we focused on an association between insulin resistance and HCC, a major cause of death for patients with HCV infection.

#### DIABETES MELLITUS AND HCC

DM has been found as a potential risk factor for the development of HCC. Three large population based cohort studies, in Sweden [82], Denmark [83], and the United States [84], reported that the development of HCC was increased 2 to 4 fold in patients with diabetes. Moreover, in some casecontrol studies, the positive association between DM and HCC has been suggested [85-88]. However, in patient HCV infection, an association between DM and HCC remains unclear, because most of these study populations had not been routinely surveyed for serological marker of HCV or contained only small number patients with HCV infection.

Table 2 shows a summary of recent seven cohort studies that investigated an association between DM and the development of HCC. Three of 7 studies reported by Tazawa et al. [89], Chen et al. [90] and Wang et al. [91] found that DM was significantly associated with the development of HCC in patients with HCV infection, with relative risk ranging from 3.1 to 9.4. On the other hand, three studies reported by Ohota et al. [92], Lai et al. [93] and Henderson et al. [94] found that there was no significant association between DM and the development of HCC in patients with HCV infection. Although the reason for this discrepancy is unclear, one possible explanation is that DM is diagnosed based on fasting plasma glucose and hemoglobin A1c levels. Both plasma glucose and hemoglobin A1c levels are not adequate diagnostic markers for DM in patients with HCV infection because of depletion of hepatic glycogen content and increased turnover of erythrocytes [95]. Thus, underdiagnosis of DM is a possible reason for the discrepancy. Recently, measurement of serum insulin levels is reported as a relevant clinical marker for predicting the development of HCC [96].

### INSULIN RESISTANCE/HYPERINSULINEMIA AND HCC

Insulin is known as one of the most important factors not only for a variety of metabolic pathways, but also for cell growth. Insulin stimulates, via phosphorylation of IRS, phosphatidylinositol 3-kinase and Akt cascade [97] as well as Ras and mitogen-activated protein kinase cascade [98]. Since these cascades regulate hepatocyte proliferation and apoptosis, hyperinsulinemia may stimulate growth of HCC. Saito et al. demonstrated that postprandial hyperinsulinemia accelerated tumor doubling time of HCC in patient with cirrhosis [99]. Moreover, several previous studies showed that insulin resistance and subsequent hyperinsulinemia contributed to progression of liver fibrosis in patients with HCV infection, regardless of the degree of steatosis [100, 101]. As

Table 2. A Summary of Seven Cohort Studies for an Association Between Diabetes Mellitus and the Development of HCC in the Population Based on HCV Infection

Year	Country	No. of Cases	No. of Cohort	No. of Cohort with HCV (%)	RR (95% CI)	Adjustment	References
2002	Japan	13	279	279 (100%)	9.4 (ND)*	age, sex, alcohol, blood transfusion, α-fetoprotein, biopsy, IFN, HCV genotype, viral road	[89]
2003	Japan	101	161	161 (100%)	1.58 (0.62-3.99)	age, sex, BMI, alcohol, ALT, triglyc- eride, cholesterol, IFN, HCV geno- type, HCV core protein	[92]
2006	Taiwan	115	54979	2087 (3.8%)	0.62 (0.22-1.76)	age, sex, alcohol, cigarette	[93]
2008	Netherland Canada Germany Switzerland	38	541	541 (100%)	2.07 (0.95-4.47)	age, sex, BMI, bilirubin, albumin, platelet count, HCV genotype, HCV viral road, HBc-Ab	[188]
2008	Taiwan	291	23820	1095 (4.6%)	3.52 (1.29-9.24)*	age, sex, alcohol, cigarette	[90]
2009	Taiwan	111	5929	982 (16.6%)	3.1 (1.7-5.4)*	age, sex, BMI, alcohol, cigarette	[91]
2009	United State	262	32806	32806 (100%)	1.17 (0.90-1.52)	age, sex, race, duration on dialysis	[94]

Abbreviation: RR; relative risk, Cl; confidence interval, ND; no description in the text, IFN; interferon, BMI; body mass index, ALT; alanine aminotransferase. \* P value < 0.05

a pathogenesis of these positive associations, it has been demonstrated that hyperinsulinemia promotes fibrogenesis by stimulating the secretion of connective tissue growth factor from hepatic stellate cells [102]. Therefore, it is suggested that the insulin resistance/hyperinsulimemia causes progression of hepatic fibrosis, leading to development of HCC in patients with HCV infection.

# THERAPEUTIC STRATEGY FOR HCV-ASSOCIATED INSULIN RESISTANCE

In this review, we propose the clinical therapeutic strategy for the treatment of HCV-associated insulin resistance, which can be determined by life-style and stage of the liver disease, but not by definition of metabolic syndrome (Fig. 2).

The International Diabetes Federation definition of metabolic syndrome is central adiposity plus two or more of the following factors; 1) raised concentration of triglycerides, 2) reduced concentration of HDL cholesterol, 3) raised blood pressure, and 4) raised fasting plasma glucose concentration [103]. In general, overeating and less activity induce metabolic syndrome through changes in adipocytokines. Therefore, diet therapy and exercise are recommended to patients with central adiposity [104]. However, none of the adipocytokines is associated with insulin resistance in patients with HCV infection, suggesting life-style may not be a major cause of HCV-associated insulin resistance [105]. Excessive life style modification such as fasting and over exercise could worsen liver function in patients with chronic liver

disease [106-108], diet therapy and exercise are recommended if patients are overeating or less active.

# **Nutritional Therapy**

Treatment for the insulin resistance could be a therapeutic strategy to prevent the development of HCC and to improve the prognosis in patients with HCV infection. Nutritional therapy and exercise are fundamental for patients with metabolic syndrome, which is defined by as well as patients with HCV-associated insulin resistance. However, in patients with liver cirrhosis, glucose metabolism is characterized as postprandial hyperglycemia induced by decreased glucose uptake of the liver [109, 110] and as fasting hypoglycemia induced by decreased glycogen storage in the liver, accompanied with increased ratio of lipid combustion in fasting state [108, 111-114]. Therefore, in order to improve glucose metabolism, divided energy intake into a larger number of meals per day including a late evening snack has been recommended for cirrhotic patients [115].

# Branched-chain Amino Acids (BCAA)

BCAA has been recently demonstrated to play a role in glucose metabolism [116-119], while it improves the complications and prognosis induced by liver cirrhosis, such as hyperammonemia, encephalopathy, or HCC [120-122]. Therefore, nutritional therapy containing BCAA is most essential when considering a nutritional therapy for liver cirrhosis as well as DM. The mechanism of BCAA action on glucose metabolism is suggested as follows. In a rat model

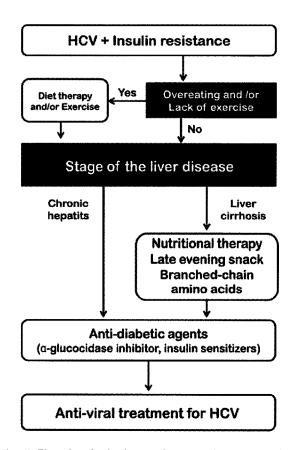


Fig. (2). Flow chart for the therapeutic strategy for HCV-associated insulin resistance. The therapeutic strategy for HCV-associated insulin resistance can be determined not only by life-style, but also by stage of the liver disease.

with liver cirrhosis, leucine and isolocine upregulate glucose uptake on skeletal muscle [123]. Similarly, a dipeptide constituted of leucine and isolocine also stimulates glucose uptake in myotube and skeletal muscle cells [124]. BCAA has potential to improve abnormal glucose metabolism such as insulin resistance through anti-obese effect in mice [125, 126]. In human, BCAA-enriched supplementation is also demonstrated to improve the insulin resistance or glucose tolerance of patients with chronic liver disease or liver cirrhosis [64, 127, 128]. Alternatively, intake of enteral nutrients for liver failure that are conditioned with enriched BCAA and high fisher's ratio as late evening snack would improve glucose metabolism [118].

It has been recently demonstrated that BCAA supplementation is potential to prevent the incidence of HCC [122]. Although the mechanism remains unclear, anti-diabetic and anti-obese effects might reduce the HCC incidence because both DM and obesity are shown to be risk factors for HCC. In addition, BCAA is shown to inhibit in vitro vascularization under the high concentration of glucose and/or insulin or in vivo vascularization of the liver in a diabetic rat model with liver injury [129]. Because HCC is a hypervascular tumor, the inhibitive effect of vascularization through the improvement of glucose metabolism by BCAA might reduce the incidence of HCC.

#### Anti-diabetic Agents

Anti-diabetic agents are generally used if nutritional and exercise therapies are not sufficient to improve hyperglycemia. When using anti-diabetic agents, the risk for adverse effects should be considered carefully because most of them are metabolized in liver, or α-glucocidase inhibitor [130] have been reported to have an adverse effect of severe liver injury that could be life-threatening complications for cirrhotic patients. In addition, use of biguanide in cirrhotic patients would be riskier for lactic acidosis than diabetic patients because lactic acid is also metabolized in liver. Thiazolidinedione is an insulin-sensitizing agent and improves virological response to peginterferon alpha-2b/ribavirin combination therapy in hepatitis C genotype 4 patients with insulin resistance [131]. However, thiazolidinedione causes overproduction of hydrogen peroxide, leading to severe hepatotoxicity [132-134]. Thus, these risks for adverse effects may limit the use of anti-diabetic agents and exogenous insulin tends to be administrated in patients with HCV infection.

### An Association Between Anti-Diabetic Agents and Malignancies

Insulin therapy is generally selected for the treatment of cirrhotic patients with DM, however, insulin therapy has recently raised a question concerning to cancer incidence. In fact, use of anti-diabetic agents has been recently demonstrated to influence the tumor-free survival in diabetic patients. Currie CJ et al. demonstrated that overall tumor-free survival in diabetic patients receiving insulin-based therapy or sulfonylurea, but not metformin, was significantly worse than receiving no diabetes medications [135]. Although insulin or sulfonylurea does not seem to increase any tumor incidence, these agents could influence the incidence of digestive system cancers such as colorectal and pancreatic cancers. Similarly, Yang Y et al. demonstrated that the risk of colorectal cancer was increased by insulin therapy [136]. Li D et al. also demonstrated that the risk of pancreatic cancer was increased by insulin or sulfonylurea and reduced by metformin [137].

Concerning to HCC incidence, Donadon V et al. implied an association between HCC incidence and use of insulin or sulfonylurea [138]. Subsequently, they carried out a largescale survey, demonstrating a direct association of HCC with use of insulin and sulfonylurea and an inverse relationship with metformin [139]. We also demonstrated that insulin or sulfonylurea was an independent risk factor for HCC incidence and hepatocarcinogenic effects of these anti-diabetic agents are evident in patients who were male or non-cirrhotic [140]. In addition, Komura T et al. described that insulin therapy was a significant factor contributing HCC recurrence after surgical treatment [141]. Thus, these results strongly suggest a potential risk factor of insulin for HCC incidence because either insulin administration or sulfonylurea intake increases insulin level in serum.

The mechanism between insulin and cancer incidence is little known. Since insulin has biological activities of cell proliferation, insulin may stimulate cancer cell proliferation and develop the cancer [142, 143]. In addition, it has been shown that the expression of phosphatase and tensin homolog or SH2 domain-containing inositol phosphatase 2, suppressive molecules of insulin signaling in cells, is decreased in HCC tissues, indicating enhanced action of insulin in HCC [35, 144-146]. Thus, insulin therapy might worsen the prognosis of the patients with HCC because suppressors of intracellular insulin signaling are inactivated in HCC and therefore, insulin effects may be more evident in HCC than in hepatocytes.

#### Anti-viral Treatment for HCV

Since HCV itself plays crucial role in the development of insulin resistance, eradication of HCV by anti-viral treatment has a significant impact when patients meet the criteria. We along with others have shown that clearance of HCV improves insulin resistance, beta-cell function, and hepatic IRS1/2 expression [25, 28, 65]. Although clearance of HCV is a fundamental therapeutic strategy for patients with HCV infection, Tsochatzis et al. described that insulin resistance develops early in the course of the disease, and negatively affects treatment response and the development of liver cirrhosis and HCC, irrespective of genotype [147]. Thus, amelioration of insulin sensitivity may inhibit the progression of HCV-associated liver disease.

#### **GUT HORMONES AND GLUCOSE METABOLISM**

The gut has currently been recognized as an endocrine system that regulates glucose metabolism [148, 149]. Among several gut hormones called "incretin", glucagon-like peptide-1 (GLP-1) is well known to be involved in the glucose metabolism. GLP-1 is secreted from endocrine L-cells of the distal intestine and colon in response to enteric nutrient ingestion, such as carbohydrates, fatty acids, essential aminoacids and dietary fiber [150, 151]. GLP-1 exerts a direct insulinotoropic effect on the pancreatic  $\beta$ -cell [151, 152]. In addition, GLP-1 activates adenylate cyclase and subsequently enhances insulin secretion via GLP-1 receptor on the cell-membrane of pancreatic \beta-cell [153], and glucose disposal [154]. GLP-1 also inhibits glucagon secretion via GLP-1 receptor on pancreatic alpha-cells [155]. Thus, GLP-1 exerts carbohydrate assimilation and inhibits gluconeogenesis, consequently, GLP-1 is considered as a therapeutic target for DM as well as insulin resistance [150, 151, 155].

Active type of GLP-1 is rapidly inactivated by dipeptidyl peptidase-IV (DPP-4, enzyme code number 3.4.14.5) [151, 152, 156, 157]. DPP-4 is a membrane-associated peptidase and is widely distributed in numerous tissues, such as intestinal brush-border, endothelial cell and hepatocytes. DPP-4 inactivates GLP-1 within a few minutes. Therefore, DPP-4 inhibitor (incretin enhancer) may be a suitable agent for the treatment of insulin resistance.

# GLP-1 AND GLUCOSE METABOLISM IN LIVER

GLP-1 reduces hepatic glucose production [158]. Although direct effect of GLP-1 on hepatocytes remains unclear, GLP-1 increases glycogen synthesis in hepatocytes by stimulating glycogen synthase alpha via GLP-1 receptor in rat hepatocytes [159, 160]. In addition, GLP-1 receptor agonist improve hepatic glucose homeostasis by promoting he-

patic insulin signaling in diabetic rats [161]. In human study, GLP-1 receptor antagonist promotes hepatocyte proliferation via induced c-AMP [162] and GLP-1 inhibits glucose disposal rather than increasing glucose disposal [163]. These findings indicate that GLP-1 has a direct effect on hepatocytes in the regulation of glucose metabolism.

# HCV-ASSOCIATED INSULIN RESISTANCE AND GLP-1

We have previously demonstrated that the up-regulation of DPP-4 causes a decrease in serum active GLP-1 levels, resulting in a decrease in hepatic glycogen contents and the development of insulin resistance in patients with HCV infection [39]. The mechanism of increased DPP-4 expression is unclear. However, a significant increase in DPP-4 expression is seen in a hepatoma cell line transfected with a HCV non-structural genome region [164]. In addition, eradication of HCV by treatment with interferon-alpha decreases serum DPP-4 activity [165]. These findings may indicate that HCV directly up-regulates DPP-4 expression. Although limited information is available for the effects of DPP-4 inhibitor in HCV-associated insulin resistance, this therapeutic agent could improve the initial step of the development of insulin resistance and is considered as a new therapeutic strategy for HCV-associated insulin resistance.

# THE SIGNIFICANCE OF INSULIN RESISTANCE IN THE ERA OF NEW ANTI-VIRAL TREATMENTS

It is no doubt that these new anti-viral agents will markedly change the treatment for HCV infection in the near future. The most of new antiviral agents for HCV infection are currently in phase I-III [166-172] and the most studied agent is an inhibitor of the HCV non-structural 3 protease, telaprevir or boceprevir [173-183]. The addition of telaprevir or boceprevir to pegylated interferon-α and ribavirin combination therapy significantly enhance sustained virologic response rates even in HCV genotype 1 patients [168, 175, 180, 181, 184, 185]. However, the rates of sustained virologic response of triple therapy with telaprevir, pegylated interferon-α and ribavirin are still up to about 50% in patients who had previously treated by pegylated interferon-α and ribavirin [181]. In addition, the resistance profile of the HCV non-structural 3 protease inhibitor is elucidated. Thus, the triple therapy is not promising to cure all of patients with chronic HCV infection.

Recetnly, Akuta *et al.* examined the impact of substitution of amino acid in the core region of HCV genotype 1b in triple therapy with telaprevir, pegylated interferon- $\alpha$  and ribavirin and identified that substitutions of amino acid 70 and 91 as independent responsible factors associated with early virologic response [186]. Although the significance of insulin resistance in the triple therapy with telaprevir, pegylated interferon- $\alpha$  and ribavirin has never been investigated, insulin resistance may be a crucial factor even in the new era of anti-viral treatments because substitutions of amino acid 70 and 91 in the core region of HCV genotype 1b are closely associated with the development of insulin resistance [22].

#### CONCLUSION

In this review, we summarize the pathogenesis for HCV-associated insulin resistance. Similar to the life style-associated insulin resistance, obesity, inflammatory cytokines, oxidative stress, and serum PEDF levels are involved in the development of HCV-associated insulin resistance. Besides these factors, HCV itself also causes insulin resistance through down-regulation of hepatic IRS1/2. Insulin resistance is responsible for the development of cirrhotic complications including HCC, however, there is no common therapeutic strategy for HCV-associated insulin resistance.

Clearance of HCV by anti-viral treatment is a fundamental therapeutic strategy for patients with HCV infection. In addition, amelioration of insulin sensitivity may inhibit the progression of HCV-associated liver disease, and could improve the survival of these patients. Late evening snack and BCAA are nutritional therapies which could improve insulin resistance. However, use of anti-diabetic agents and exogenous insulin are not always recommended because of adverse effects and possible link to the development of HCC.

HCV also affects insulin resistance through activation of DPP-4 and subsequent inactivation of GLP-1, a key regulator of insulin secretion and hepatic glucose metabolism. Although availability of DPP-4 inhibitor in HCV-associated insulin resistance is yet unclear, this therapeutic agent could improve the early step of the development of insulin resistance and is expected to be a new therapeutic strategy for HCV-associated insulin resistance.

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### LIST OF ABBREVIATIONS

BCAA = branched-chain amino acids

DM = diabetes mellitus

DPP-4 = dipeptidyl peptidase-IV GLP-1 = glucagon-like peptide-1

HCC = hepatocellular carcinoma

HCV = hepatitis C virus

IRS = insulin receptor substrate

PEDF = serum pigment epithelium-derived factor

SOCS = suppressor of cytokine signaling

# REFERENCES

 Choo QL, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M. Isolation of a cDNA clone derived from a blood-borne non-A,

- non-B viral hepatitis genome. Science 1989; 244: 359-62.
- [2] Houghton M. The long and winding road leading to the identification of the hepatitis C virus. J Hepatol 2009; 51: 939-48.
- [3] Asselah T, Bieche I, Sabbagh A, et al. Gene expression and hepatitis C virus infection. Gut 2009; 58: 846-58.
- [4] Maheshwari A, Ray S, Thuluvath P J. Acute hepatitis C. Lancet 2008; 372: 321-32.
- [5] Seeff LB, Hoofnagle JH. Appendix: The national institutes of health consensus development conference management of hepatitis C 2002. Clin Liver Dis 2003; 7: 261-87.
- [6] Thein HH, Yi Q, Dore GJ, Krahn MD. Estimation of stage-specific fibrosis progression rates in chronic hepatitis C virus infection: a meta-analysis and meta-regression. Hepatology 2008; 48: 418-31.
- [7] Thompson ND, Perz JF, Moorman AC, Holmberg SD. Nonhospital health care-associated hepatitis B and C virus transmission: United States, 1998-2008. Ann Intern Med 2009; 150: 33-9.
- [8] Alter MJ, Margolis HS, Krawczynski K, et al. The natural history of community-acquired hepatitis C in the United States. The sentinel counties chronic non-a, non-b hepatitis study team. N Engl J Med 1992; 327: 1899-905.
- [9] Marcellin P. Hepatitis B and hepatitis C in 2009. Liver Int 2009; 29 (Suppl 1): 1-8.
- [10] Poynard T, Bedossa P, Opolon P. Natural history of liver fibrosis progression in patients with chronic hepatitis C. The OBSVIRC, METAVIR, CLINIVIR, and DOSVIRC groups. Lancet 1997; 349: 825-32
- [11] Ali SS, Ali I S, Aamir AH, Jadoon Z, Inayatullah S. Frequency of hepatitis C infection in diabetic patients. J Ayub Med Coll Abbottabad 2007; 19: 46-9.
- [12] Bahtiyar G, Shin JJ, Aytaman A, Sowers JR, Mcfarlane SI. Association of diabetes and hepatitis C infection: epidemiologic evidence and pathophysiologic insights. Curr Diabetes Rep 2004; 4: 194-8
- [13] Gray H, Wreghitt T, Stratton IM, Alexander GJ, Turner RC, O'rahilly S. High prevalence of hepatitis C infection in Afro-Caribbean patients with type 2 diabetes and abnormal liver function tests. Diabet Med 1995; 12: 244-9.
- [14] Kawaguchi T, Nagao Y, Tanaka K, et al. Causal relationship between hepatitis C virus core and the development of type 2 diabetes mellitus in a hepatitis C virus hyperendemic area: a pilot study. Int J Mol Med 2005; 16: 109-14.
- [15] Mehta SH, Brancati FL, Strathdee SA, et al. Hepatitis C virus infection and incident type 2 diabetes. Hepatology 2003; 38: 50-6.
- [16] Mehta SH, Brancati FL, Sulkowski MS, Strathdee SA, Szklo M, Thomas DL. Prevalence of type 2 diabetes mellitus among persons with hepatitis C virus infection in the United States. Ann Intern Med 2000; 133: 592-9.
- [17] Nwokediuko SC, Oli JM. Hepatitis C virus infection in Nigerians with diabetes mellitus. Niger J Clin Pract 2008; 11: 94-9.
- [18] Shaheen M, Echeverry D, Oblad MG, Montoya MI, Teklehaimanot S, Akhtar AJ. Hepatitis C, metabolic syndrome, and inflammatory markers: results from the Third National Health and Nutrition Examination Survey [NHANES III]. Diabetes Res Clin Pract 2007; 75: 320-6.
- [19] Simo R, Hernandez C, Genesca J, Jardi R, Mesa J. High prevalence of hepatitis C virus infection in diabetic patients. Diabetes Care 1996; 19: 998-1000.
- [20] Sporea I, Sirli R, Hogea C, Sink AA, Serban V. Diabetes mellitus and chronic HCV infection. Rom J Intern Med 2009; 47: 141-7.
- [21] Wilson C. Hepatitis C infection and type 2 diabetes in American-Indian women. Diabetes Care 2004; 27: 2116-9.
- [22] Akuta N, Suzuki F, Hirakawa M, et al. Amino acid substitutions in the hepatitis C virus core region of genotype 1b are the important predictor of severe insulin resistance in patients without cirrhosis and diabetes mellitus. J Med Virol 2009; 81: 1032-9.
- [23] Allison ME, Wreghitt T, Palmer CR, Alexander GJ. Evidence for a link between hepatitis C virus infection and diabetes mellitus in a cirrhotic population. J Hepatol 1994; 21: 1135-9.
- [24] Aytug S, Reich D, Sapiro LE, Bernstein D, Begum N. Impaired IRS-1/Pl3-kinase signaling in patients with HCV: a mechanism for increased prevalence of type 2 diabetes. Hepatology 2003; 38: 1384.02
- [25] Kawaguchi T, Ide T, Taniguchi E, et al. Clearance of HCV improves insulin resistance, beta-cell function, and hepatic expression

- of insulin receptor substrate 1 and 2. Am J Gastroenterol 2007; 102: 570-6
- [26] Kawaguchi T, Yoshida T, Harada M, et al. Hepatitis C virus down-regulates insulin receptor substrates 1 and 2 through up-regulation of suppressor of cytokine signaling 3. Am J Pathol 2004; 165: 1499-508
- [27] Negro F. Insulin resistance and HCV: will new knowledge modify clinical management? J Hepatol 2006; 45: 514-9.
- [28] Romero-Gomez M. Insulin resistance and hepatitis C. World J Gastroenterol 2006; 12: 7075-80.
- [29] Shintani Y, Fujie H, Miyoshi H, et al. Hepatitis C virus infection and diabetes: direct involvement of the virus in the development of insulin resistance. Gastroenterology 2004; 126: 840-8.
- [30] Taura N, Ichikawa T, Hamasaki K, et al. Association between liver fibrosis and insulin sensitivity in chronic hepatitis C patients. Am J Gastroenterol 2006; 101: 2752-9.
- [31] Alberti KG, Eckel RH, Grundy SM, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation 2009; 120: 1640-5.
- [32] Balsano C, Alisi A, Nobili V. Liver fibrosis and therapeutic strategies: the goal for improving metabolism. Curr Drug Targets 2009; 10: 505-12.
- [33] El-Serag HB. Hepatocellular carcinoma: recent trends in the United States. Gastroenterology 2004; 127: S27-34.
- [34] Negro F. Correction of insulin resistance in chronic hepatitis C patients not responding to the standard of care: more questions than answers. J Hepatol 2009; 50: 1271-2.
- [35] Sumie S, Kawaguchi T, Komuta M, et al. Significance of glucose intolerance and SHIP2 expression in hepatocellular carcinoma patients with HCV infection. Oncol Rep 2007; 18: 545-52.
- [36] Kawaguchi T, Sata M. Importance of hepatitis C virus-associated insulin resistance: therapeutic strategies for insulin sensitization. World J Gastroenterol 2010, 16: 1943-52.
- [37] Kawaguchi T, Sata M. Importance of hepatitis C virus-associated insulin resistance-therapeutic strategies for insulin sensitization. World J Gastroenterol 2010; 16: 1943-52.
- [38] Bianchi G, Marchesini G, Zoli M, Bugianesi E, Fabbri A, Pisi E. Prognostic significance of diabetes in patients with cirrhosis. Hepatology 1994; 20: 119-25.
- [39] Itou M, Kawaguchi T, Taniguchi E, et al. Altered expression of glucagon-like peptide-1 and dipeptidyl peptidase IV in patients with HCV-related glucose intolerance. J Gastroenterol Hepatol 2008: 23: 244-51.
- [40] Eguchi Y, Mizuta T, Ishibashi E, et al. Hepatitis C virus infection enhances insulin resistance induced by visceral fat accumulation. Liver Int 2009; 29: 213-20.
- [41] Hung CH, Lee CM, Chen CH, et al. Association of inflammatory and anti-inflammatory cytokines with insulin resistance in chronic hepatitis C. Liver Int 2009; 29: 1086-93.
- [42] Lu JY, Chuang LM, Yang WS, et al. Adiponectin levels among patients with chronic hepatitis B and C infections and in response to IFN-alpha therapy. Liver Int 2005; 25: 752-9.
- [43] Knobler H, Zhornicky T, Sandler A, Haran N, Ashur Y, Schattner A. Tumor necrosis factor-alpha-induced insulin resistance may mediate the hepatitis C virus-diabetes association. Am J Gastroenterol 2003; 98: 2751-6.
- [44] Maeno T, Okumura A, Ishikawa T, et al. Mechanisms of increased insulin resistance in non-cirrhotic patients with chronic hepatitis C virus infection. J Gastroenterol Hepatol 2003; 18: 1358-63.
- [45] Koike K, Miyoshi H. Oxidative stress and hepatitis C viral infection. Hepatol Res 2006; 34: 65-73.
- [46] Mitsuyoshi H, Itoh Y, Sumida Y, et al. Evidence of oxidative stress as a cofactor in the development of insulin resistance in patients with chronic hepatitis C. Hepatol Res 2008; 38: 348-53.
- [47] Vidali M, Tripodi MF, Ivaldi A, et al. Interplay between oxidative stress and hepatic steatosis in the progression of chronic hepatitis C. J Hepatol 2008; 48: 399-406.
- [48] Narita R, Abe S, Tabaru A, Otsuki M. Impact of steatosis on insulin secretion in chronic hepatitis C patients. Am J Gastroenterol 2007; 102: 2173-80.

- [49] Narita R, Abe S, Kihara Y, Akiyama T, Tabaru A, Otsuki M. Insulin resistance and insulin secretion in chronic hepatitis C virus infection. J Hepatol 2004; 41: 132-8.
- [50] Ferenci P, Bratusch-Marrain P, Waldhausl WK, Nowotny P, Korn A. Impaired plasma amino-acid clearance in patients with cirrhosis of the liver and portocaval shunt--its relation to insulin resistance. Eur J Clin Invest 1984; 14: 255-61.
- [51] Tanabe N, Ishii M, Sato Y, et al. Effects of collateral vessel occlusion on oral glucose tolerance test in liver cirrhosis. Dig Dis Sci 2000; 45: 581-6.
- [52] Muckenthaler MU. Fine tuning of hepcidin expression by positive and negative regulators. Cell Metab 2008; 8: 1-3.
- [53] Nishina S, Hino K, Korenaga M, et al. Hepatitis C virus-induced reactive oxygen species raise hepatic iron level in mice by reducing hepcidin transcription. Gastroenterology 2008; 134: 226-38.
- [54] Fujita N, Sugimoto R, Takeo M, et al. Hepcidin expression in the liver: relatively low level in patients with chronic hepatitis C. Mol Med 2007; 13: 97-104.
- [55] Nagashima M, Kudo M, Chung H, et al. Regulatory failure of serum prohepcidin levels in patients with hepatitis C. Hepatol Res 2006; 36: 288-93.
- [56] Lecube A, Hernandez C, Simo R. Glucose abnormalities in nonalcoholic fatty liver disease and chronic hepatitis C virus infection: the role of iron overload. Diabetes Metab Res Rev 2009; 25: 403-10.
- [57] Furutani M, Nakashima T, Sumida Y, et al. Insulin resistance/betacell function and serum ferritin level in non-diabetic patients with hepatitis C virus infection. Liver Int 2003; 23: 294-9.
- [58] Sumida Y, Kanemasa K, Fukumoto K, Yoshida N, Sakai K. Hepatic iron accumulation may be associated with insulin resistance in patients with chronic hepatitis C. Hepatol Res 2007; 37: 932-40.
- [59] Tsochatzis E, Manolakopoulos S, Papatheodoridis GV, et al. Serum HCV RNA levels and HCV genotype do not affect insulin resistance in nondiabetic patients with chronic hepatitis C: a multicentre study. Aliment Pharmacol Ther 2009; 30: 947-54.
- [60] Hsu CS, Liu CJ, Liu CH, et al. High hepatitis C viral load is associated with insulin resistance in patients with chronic hepatitis C. Liver Int 2008; 28: 271-7.
- [61] Moucari R, Asselah T, Cazals-Hatem D, et al. Insulin resistance in chronic hepatitis C: association with genotypes 1 and 4, serum HCV RNA level, and liver fibrosis. Gastroenterology 2008; 134: 416-23.
- [62] Yoneda M, Saito S, Ikeda T, et al. Hepatitis C virus directly associates with insulin resistance independent of the visceral fat area in nonobese and nondiabetic patients. J Viral Hepatitis 2007; 14: 600-7.
- [63] Delgado-Borrego A, Jordan SH, Negre B, et al. Reduction of insulin resistance with effective clearance of hepatitis C infection: results from the HALT-C trial. Clin Gastroenterol Hepatol 2010; 8(5): 458-62.
- [64] Kawaguchi T, Taniguchi E, Itou M, et al. Branched-chain amino acids improve insulin resistance in patients with hepatitis C virusrelated liver disease: report of two cases. Liver Int 2007; 27: 1287-92
- [65] Kawaguchi Y, Mizuta T, Oza N, et al. Eradication of hepatitis C virus by interferon improves whole-body insulin resistance and hyperinsulinaemia in patients with chronic hepatitis C. Liver Int 2009; 29: 871-7.
- [66] Rui L, Yuan M, Frantz D, Shoelson S, White MF. SOCS-1 and SOCS-3 block insulin signaling by ubiquitin-mediated degradation of IRS1 and IRS2. J Biol Chem 2002; 277: 42394-8.
- [67] Persico M, Russo R, Persico E, et al. SOCS3 and IRS-1 gene expression differs between genotype 1 and genotype 2 hepatitis C virus-infected HepG2 cells. Clin Chem Lab Med 2009; 47: 1217-25.
- [68] Banerjee S, Saito K, Ait-Goughoulte M, Meyer K, Ray RB, Ray R. Hepatitis C virus core protein upregulates serine phosphorylation of insulin receptor substrate-1 and impairs the downstream akt/protein kinase B signaling pathway for insulin resistance. J Virol 2008; 82: 2606-12
- [69] Pazienza V, Clement S, Pugnale P, et al. The hepatitis C virus core protein of genotypes 3a and 1b downregulates insulin receptor substrate 1 through genotype-specific mechanisms. Hepatology 2007; 45: 1164-71.
- [70] Cua IH, Hui JM, Kench JG, George J. Genotype-specific interac-

- tions of insulin resistance, steatosis, and fibrosis in chronic hepatitis C. Hepatology 2008; 48: 723-31.
- [71] Liu CJ, Jeng YM, Chen PJ, et al. Influence of metabolic syndrome, viral genotype and antiviral therapy on superimposed fatty liver disease in chronic hepatitis C. Antivir Ther 2005; 10: 405-15.
- [72] Pazienza V, Vinciguerra M, Andriulli A, Mangia A. HCV core protein genotype 3a increases SOCS-7 expression through PPAR-{gamma} in Huh-7 cells. J Gen Virol 2010; 91(Pt 7): 1678-86.
- [73] Duseja A, Dhiman RK, Chawla Y, et al. Insulin resistance is common in patients with predominantly genotype 3 chronic hepatitis C. Dig Dis Sci 2009; 54: 1778-82.
- [74] Wang AY, Hickman IJ, Richards AA, Whitehead JP, Prins JB, Macdonald GA. High molecular weight adiponectin correlates with insulin sensitivity in patients with hepatitis C genotype 3, but not genotype 1 infection. Am J Gastroenterol 2005; 100: 2717-23.
- [75] Khattab M, Eslam M, Sharwae MA, Shatat M, Ali A, Hamdy L. Insulin resistance predicts rapid virologic response to peginterferon/ribavirin combination therapy in hepatitis C genotype 4 patients. Am J Gastroenterol 2010; [Epub ahead of print]
- [76] Moucari R, Ripault MP, Martinot-Peignoux M, et al. Insulin resistance and geographical origin: major predictors of liver fibrosis and response to peginterferon and ribavirin in HCV-4. Gut 2009; 58: 1662-9.
- [77] Kawaguchi T, Yamagishi S, Sata M. Branched-chain amino acids and pigment epithelium-derived factor: novel therapeutic agents for hepatitis C Virus-associated insulin resistance. Curr Med Chem 2009: 16(36): 4843-57.
- [78] Camma C, Petta S, Di Marco V, et al. Insulin resistance is a risk factor for esophageal varices in hepatitis C virus cirrhosis. Hepatology 2009; 49: 195-203.
- [79] Nagao Y, Kawasaki K, Sata M. Insulin resistance and lichen planus in patients with HCV-infectious liver diseases. J Gastroenterol Hepatol 2008; 23: 580-5.
- [80] Nagao Y, Sata M. High incidence of multiple primary carcinomas in HCV-infected patients with oral squamous cell carcinoma. Med Sci Monit 2009: 15: CR453-9
- [81] Nagao Y, Kawaguchi T, Tanaka K, Kumashiro R, Sata M. Extrahepatic manifestations and insulin resistance in an HCV hyperendemic area. Int J Mol Med 2005; 16: 291-6.
- [82] Adami HO, Chow WH, Nyren O, et al. Excess risk of primary liver cancer in patients with diabetes mellitus. J Natl Cancer Inst 1996; 88: 1472-7.
- [83] Wideroff L, Gridley G, Mellemkjaer L, et al. Cancer incidence in a population-based cohort of patients hospitalized with diabetes mellitus in Denmark. J Natl Cancer Inst 1997; 89: 1360-5.
- [84] El-Serag HB, Tran T, Everhart JE. Diabetes increases the risk of chronic liver disease and hepatocellular carcinoma. Gastroenterology 2004; 126: 460-8.
- [85] Davila JA, Morgan RO, Shaib Y, Mcglynn KA, El-Serag HB. Diabetes increases the risk of hepatocellular carcinoma in the United States: a population based case control study. Gut 2005; 54: 533-9.
- [86] El-Serag HB, Richardson PA, Everhart JE. The role of diabetes in hepatocellular carcinoma: a case-control study among United States Veterans. Am J Gastroenterol 2001; 96: 2462-7.
- [87] La Vecchia C, Negri E, Decarli A, Franceschi S. Diabetes mellitus and the risk of primary liver cancer. Int J Cancer 1997; 73: 204-7.
- [88] Lagiou P, Kuper H, Stuver SO, Tzonou A, Trichopoulos D, Adami HO. Role of diabetes mellitus in the etiology of hepatocellular carcinoma. J Natl Cancer Inst 2000; 92: 1096-9.
- [89] Tazawa J, Maeda M, Nakagawa M, et al. Diabetes mellitus may be associated with hepatocarcinogenesis in patients with chronic hepatitis C. Dig Dis Sci 2002; 47: 710-5.
- [90] Chen CL, Yang HI, Yang WS, et al. Metabolic factors and risk of hepatocellular carcinoma by chronic hepatitis B/C infection: a follow-up study in Taiwan. Gastroenterology 2008; 135: 111-21.
- [91] Wang CS, Yao WJ, Chang TT, Wang ST, Chou P. The impact of type 2 diabetes on the development of hepatocellular carcinoma in different viral hepatitis statuses. Cancer Epidemiol Biomarkers Prev 2009; 18: 2054-60.
- [92] Ohata K, Hamasaki K, Toriyama K, et al. Hepatic steatosis is a risk factor for hepatocellular carcinoma in patients with chronic hepatitis C virus infection. Cancer 2003; 97: 3036-43.
- [93] Lai MS, Hsieh MS, Chiu YH, Chen TH. Type 2 diabetes and hepa-

- tocellular carcinoma: a cohort study in high prevalence area of hepatitis virus infection. Hepatology 2006; 43: 1295-302.
- [94] Henderson WA, Shankar R, Gill JM, et al. Hepatitis C progressing to hepatocellular carcinoma: the HCV dialysis patient in dilemma. J Viral Hepatitis 2010; 17(1): 59-64.
- [95] Koga M, Kasayama S, Kanehara H, Bando Y. CLD (chronic liver diseases)-HbA1C as a suitable indicator for estimation of mean plasma glucose in patients with chronic liver diseases. Diabetes Res Clin Pract 2008; 81: 258-62.
- [96] Donadon V, Balbi M, Valentia G, et al. Association between type 2 diabetes mellitus and hepatocellular carcinoma: role of insulin resistance and hyperinsulinemia. Hepatology 2009; 50: 1081A.
- [97] Cantley LC. The phosphoinositide 3-kinase pathway. Science 2002: 296: 1655-7.
- [98] Boulton TG, Nye SH, Robbins DJ, et al. ERKs: a family of proteinserine/threonine kinases that are activated and tyrosine phosphorylated in response to insulin and NGF. Cell 1991; 65: 663-75.
- [99] Saito K, Inoue S, Saito T, et al. Augmentation effect of postprandial hyperinsulinaemia on growth of human hepatocellular carcinoma. Gut 2002; 51: 100-4.
- [100] Bugianesi E, Marchesini G, Gentilcore E, et al. Fibrosis in genotype 3 chronic hepatitis C and nonalcoholic fatty liver disease: role of insulin resistance and hepatic steatosis. Hepatology 2006; 44: 1648-55.
- [101] Fartoux L, Poujol-Robert A, Guechot J, Wendum D, Poupon R, Serfaty L. Insulin resistance is a cause of steatosis and fibrosis progression in chronic hepatitis C. Gut 2005; 54: 1003-8.
- [102] Paradis V, Perlemuter G, Bonvoust F, et al. High glucose and hyperinsulinemia stimulate connective tissue growth factor expression: a potential mechanism involved in progression to fibrosis in nonalcoholic steatohepatitis. Hepatology 2001; 34: 738-44.
- [103] Alberti KG, Zimmet P, Shaw J. The metabolic syndrome--a new worldwide definition. Lancet 2005; 366: 1059-62.
- [104] Kelesidis T, Kelesidis I, Chou S, Mantzoros CS. Narrative review: the role of leptin in human physiology: emerging clinical applications. Ann Intern Med 2010; 152: 93-100.
- [105] Cua IH, Hui JM, Bandara P, et al. Insulin resistance and liver injury in hepatitis C is not associated with virus-specific changes in adipocytokines. Hepatology 2007; 46: 66-73.
- [106] Degoricija V, Zjacic-Rotkvic V, Marout J, Sefer S, Troskot B. Clinical and neurohumoral response to posture, physical exercise, and ascites treatment in Child-Pugh C liver cirrhosis: randomized prospective trial. Croat Med J 2003; 44: 178-86.
- [107] Kawaguchi T, Taniguchi E, Itou M, et al. Supplement improves nutrition and stresses caused by examination-associated fasting in patients with liver cirrhosis. Hepatol Res 2008; 38: 1178-85.
- [108] Owen OE, Trapp VE, Reichard GA, Jr., et al. Nature and quantity of fuels consumed in patients with alcoholic cirrhosis. J Clin Invest 1983; 72: 1821-32.
- [109] Kalaitzakis E, Bosaeus I, Ohman L, Bjornsson E. Altered postprandial glucose, insulin, leptin, and ghrelin in liver cirrhosis: correlations with energy intake and resting energy expenditure. Am J Clin Nutr 2007; 85: 808-15.
- [110] Nielsen MF, Caumo A, Aagaard NK, et al. Contribution of defects in glucose uptake to carbohydrate intolerance in liver cirrhosis: assessment during physiological glucose and insulin concentrations. Am J Physiol Gastrointest Liver Physiol 2005; 288: G1135-43.
- [111] Miwa Y, Shiraki M, Kato M, et al. Improvement of fuel metabolism by nocturnal energy supplementation in patients with liver cirrhosis. Hepatol Res 2000; 18: 184-9.
- [112] Muller MJ, Lautz HU, Plogmann B, Burger M, Korber J, Schmidt FW. Energy expenditure and substrate oxidation in patients with cirrhosis: the impact of cause, clinical staging and nutritional state. Hepatology 1992; 15: 782-94.
- [113] Nakaya Y, Harada N, Kakui S, et al. Severe catabolic state after prolonged fasting in cirrhotic patients: effect of oral branched-chain amino-acid-enriched nutrient mixture. J Gastroenterol 2002; 37: 531-6.
- [114] Tajika M, Kato M, Mohri H, et al. Prognostic value of energy metabolism in patients with viral liver cirrhosis. Nutrition 2002; 18: 229-34
- [115] Aspen Board of Directors and the Clinical Guidelines Task Force. Guidelines for the use of parenteral and enteral nutrition in adult and pediatric patients. JPEN J Parenter Enteral Nutr 2002; 26:

- 1SA-138SA.
- [116] Aoyama K, Tsuchiya M, Mori K, et al. Effect of a late evening snack on outpatients with liver cirrhosis. Hepatol Res 2007; 37: 608-14.
- [117] Carty H. Scintigraphy in sports injuries: is it underutilized? Nucl Med Commun 1991: 12: 173-4.
- [118] Tsuchiya M, Sakaida I, Okamoto M, Okita K. The effect of a late evening snack in patients with liver cirrhosis. Hepatol Res 2005; 31: 95-103.
- [119] Utsugi T, Yoshida A, Kanda T, et al. Oral administration of branched chain amino acids improves virus-induced glucose intolerance in mice. Eur J Pharmacol 2000; 398: 409-14.
- [120] Marchesini G, Bianchi G, Merli M, et al. Nutritional supplementation with branched-chain amino acids in advanced cirrhosis: a double-blind, randomized trial. Gastroenterology 2003; 124: 1792-801.
- [121] Muto Y, Sato S, Watanabe A, et al. Effects of oral branched-chain amino acid granules on event-free survival in patients with liver cirrhosis. Clin Gastroenterol Hepatol 2005; 3: 705-13.
- [122] Muto Y, Sato S, Watanabe A, et al. Overweight and obesity increase the risk for liver cancer in patients with liver cirrhosis and long-term oral supplementation with branched-chain amino acid granules inhibits liver carcinogenesis in heavier patients with liver cirrhosis. Hepatol Res 2006; 35: 204-14.
- [123] Nishitani S, Takehana K, Fujitani S, Sonaka I. Branched-chain amino acids improve glucose metabolism in rats with liver cirrhosis. Am J Physiol Gastrointest Liver Physiol 2005; 288: G1292-300.
- [124] Morifuji M, Koga J, Kawanaka K, Higuchi M. Branched-chain amino acid-containing dipeptides, identified from whey protein hydrolysates, stimulate glucose uptake rate in L6 myotubes and isolated skeletal muscles. J Nutr Sci Vitaminol (Tokyo) 2009; 55: 81-6.
- [125] Ikehara O, Kawasaki N, Maezono K, Komatsu M, Konishi A. Acute and chronic treatment of L-isoleucine ameliorates glucose metabolism in glucose-intolerant and diabetic mice. Biol Pharm Bull 2008; 31: 469-72.
- [126] She P, Reid TM, Bronson SK, et al. Disruption of BCATm in mice leads to increased energy expenditure associated with the activation of a futile protein turnover cycle. Cell Metab 2007; 6: 181-94.
- [127] Kawaguchi T, Nagao Y, Matsuoka H, Ide T, Sata M. Branchedchain amino acid-enriched supplementation improves insulin resistance in patients with chronic liver disease. Int J Mol Med 2008; 22: 105-12.
- [128] Urata Y, Okita K, Korenaga K, Uchida K, Yamasaki T, Sakaida I. The effect of supplementation with branched-chain amino acids in patients with liver cirrhosis. Hepatol Res 2007; 37: 510-6.
- [129] Yoshiji H, Noguchi R, Kitade M, et al. Branched-chain amino acids suppress insulin-resistance-based hepatocarcinogenesis in obese diabetic rats. J Gastroenterol 2009; 44: 483-91.
- [130] Carrascosa M, Pascual F, Aresti S. Acarbose-induced acute severe hepatotoxicity. Lancet 1997; 349: 698-9.
- [131] Khattab M, Emad M, Abdelaleem A, et al. Pioglitazone improves virological response to peginterferon alpha-2b/ribavirin combination therapy in hepatitis C genotype 4 patients with insulin resistance. Liver Int 2010; 30(3): 447-54.
- [132] Al-Salman J, Arjomand H, Kemp DG, Mittal M. Hepatocellular injury in a patient receiving rosiglitazone. A case report. Ann Intern Med 2000; 132: 121-4.
- [133] Gouda HE, Khan A, Schwartz J, Cohen RI. Liver failure in a patient treated with long-term rosiglitazone therapy. Am J Med 2001; 111: 584-5.
- [134] Lenhard MJ, Funk WB. Failure to develop hepatic injury from rosiglitazone in a patient with a history of troglitazone-induced hepatitis. Diabetes Care 2001; 24: 168-9.
- [135] Hemkens LG, Grouven U, Bender R, et al. Risk of malignancies in patients with diabetes treated with human insulin or insulin analogues: a cohort study. Diabetologia 2009; 52: 1732-44.
- [136] Yang YX, Hennessy S, Lewis JD. Insulin therapy and colorectal cancer risk among type 2 diabetes mellitus patients. Gastroenterology 2004; 127: 1044-50.
- [137] Li D, Yeung SC, Hassan MM, Konopleva M, Abbruzzese JL. Antidiabetic therapies affect risk of pancreatic cancer. Gastroenterology 2009; 137: 482-8.
- [138] Donadon V, Balbi M, Casarin P, Vario A, Alberti A. Association

- between hepatocellular carcinoma and type 2 diabetes mellitus in Italy: potential role of insulin. World J Gastroenterol 2008; 14: 5695-700.
- [139] Donadon V, Balbi M, Ghersetti M, et al. Antidiabetic therapy and increased risk of hepatocellular carcinoma in chronic liver disease. World J Gastroenterol 2009; 15: 2506-11.
- [140] Kawaguchi T, Taniguchi E, Morita Y, et al. Association of exogenous insulin or sulfonylurea treatment with an increased incidence of hepatoma in patients with HCV infection. Liver Int 2010; 30(3): 479-86
- [141] Komura T, Mizukoshi E, Kita Y, et al. Impact of diabetes on recurrence of hepatocellular carcinoma after surgical treatment in patients with viral hepatitis. Am J Gastroenterol 2007; 102: 1939-46.
- [142] Godsland IF. Insulin resistance and hyperinsulinaemia in the development and progression of cancer. Clin Sci (Lond) 2009; 118: 315-32
- [143] Sandow J. Growth effects of insulin and insulin analogues. Arch Physiol Biochem 2009; 115: 72-85.
- [144] Dong-Dong L, Xi-Ran Z, Xiang-Rong C. Expression and significance of new tumor suppressor gene PTEN in primary liver cancer. J Cell Mol Med 2003; 7: 67-71.
- [145] Wan XW, Jiang M, Cao HF, et al. The alteration of PTEN tumor suppressor expression and its association with the histopathological features of human primary hepatocellular carcinoma. J Cancer Res Clin Oncol 2003; 129: 100-6.
- [146] Wu SK, Wang BJ, Yang Y, Feng XH, Zhao XP, Yang DL. Expression of PTEN, PPM1A and P-Smad2 in hepatocellular carcinomas and adjacent liver tissues. World J Gastroenterol 2007; 13: 4554-9.
- [147] Tsochatzis EA, Manolakopoulos S, Papatheodoridis GV, Archimandritis AJ. Insulin resistance and metabolic syndrome in chronic liver diseases: old entities with new implications. Scand J Gastroenterol 2009; 44: 6-14.
- [148] Creutzfeldt W. The [pre-] history of the incretin concept. Regul Pept 2005; 128: 87-91.
- [149] Seltzer HS, Allen EW, Herron AL, Jr., Brennan MT. Insulin secretion in response to glycemic stimulus: relation of delayed initial release to carbohydrate intolerance in mild diabetes mellitus. J Clin Invest 1967; 46: 323-35.
- [150] Baggio LL, Drucker DJ. Biology of incretins: GLP-1 and GIP. Gastroenterology 2007; 132: 2131-57.
- [151] Drucker DJ. The biology of incretin hormones. Cell Metab 2006; 3: 153-65
- [152] Holst JJ. Glucagon-like peptide-1: from extract to agent. The Claude Bernard Lecture, 2005. Diabetologia 2006; 49: 253-60.
- [153] Thorens B. Expression cloning of the pancreatic beta cell receptor for the gluco-incretin hormone glucagon-like peptide 1. Proc Natl Acad Sci USA 1992; 89: 8641-5.
- [154] Vilsboll T, Krarup T, Madsbad S, Holst JJ. Both GLP-1 and GIP are insulinotropic at basal and postprandial glucose levels and contribute nearly equally to the incretin effect of a meal in healthy subjects. Regul Pept 2003; 114: 115-21.
- [155] Nauck MA, Kleine N, Orskov C, Holst JJ, Willms B, Creutzfeldt W. Normalization of fasting hyperglycaemia by exogenous glucagon-like peptide 1 (7-36 amide) in type 2 (non-insulin-dependent) diabetic patients. Diabetologia 1993; 36: 741-4.
- [156] Deacon CF, Pridal L, Klarskov L, Olesen M, Holst JJ. Glucagonlike peptide 1 undergoes differential tissue-specific metabolism in the anesthetized pig. Am J Physiol 1996; 271: E458-64.
- [157] Hansen L, Deacon CF, Orskov C, Holst JJ. Glucagon-like peptide-1-(7-36)amide is transformed to glucagon-like peptide-1-(9-36)amide by dipeptidyl peptidase IV in the capillaries supplying the L cells of the porcine intestine. Endocrinology 1999; 140: 5356-63.
- [158] Larsson H, Holst JJ, Ahren B. Glucagon-like peptide-1 reduces hepatic glucose production indirectly through insulin and glucagon in humans. Acta Physiol Scand 1997; 160: 413-22.
- [159] Mommsen TP, Andrews PC, Plisetskaya EM. Glucagon-like peptides activate hepatic gluconeogenesis. FEBS Lett 1987; 219: 227-32.
- [160] Redondo A, Trigo MV, Acitores A, Valverde I, Villanueva-Penacarrillo ML. Cell signalling of the GLP-1 action in rat liver. Mol Cell Endocrinol 2003; 204: 43-50.
- [161] Park S, Hong SM, Ahn IS. Exendin-4 and exercise improve hepatic glucose homeostasis by promoting insulin signaling in diabetic rats.

- Metabolism 2010; 59: 123-33.
- [162] Aviv V, Meivar-Levy I, Rachmut IH, Rubinek T, Mor E, Ferber S. Exendin-4 promotes liver cell proliferation and enhances the PDX-1-induced liver to pancreas transdifferentiation process. J Biol Chem 2009; 284: 33509-20.
- [163] Prigeon RL, Quddusi S, Paty B, D'alessio DA. Suppression of glucose production by GLP-1 independent of islet hormones: a novel extrapancreatic effect. Am J Physiol Endocrinol Metab 2003; 285: E701-7.
- [164] Harada T, Kim DW, Sagawa K, et al. Characterization of an established human hepatoma cell line constitutively expressing nonstructural proteins of hepatitis C virus by transfection of viral cDNA. J Gen Virol 1995; 76 (Pt 5): 1215-21.
- [165] Maes M, Bonaccorso S, Marino V, et al. Treatment with interferonalpha (IFN alpha) of hepatitis C patients induces lower serum dipeptidyl peptidase IV activity, which is related to IFN alphainduced depressive and anxiety symptoms and immune activation. Mol Psychiatry 2001; 6: 475-80.
- [166] Asselah T, Benhamou Y, Marcellin P. Protease and polymerase inhibitors for the treatment of hepatitis C. Liver Int 2009; 29 (Suppl 1): 57-67.
- [167] Harrison SA. Small molecule and novel treatments for chronic hepatitis C virus infection. Am J Gastroenterol 2007; 102: 2332-8.
- [168] Lange CM, Sarrazin C, Zeuzem S. Review article: HCV STAT-C era of therapy. Aliment Pharmacol Ther 2010; [Epub ahead of print]
- [169] Moucari R, Marcellin P. Hepatitis C: The role of new interferons in the era of STAT-C. Nat Rev Gastroenterol Hepatol 2009; 6: 509-11.
- [170] Parfieniuk A, Jaroszewicz J, Flisiak R. Specifically targeted antiviral therapy for hepatitis C virus. World J Gastroenterol 2007; 13: 5673-81.
- [171] Pereira AA, Jacobson IM. New and experimental therapies for HCV. Nat Rev Gastroenterol Hepatol 2009; 6: 403-11.
- [172] Schinazi RF, Bassit L, Gavegnano C. HCV drug discovery aimed at viral eradication. J Viral Hepatitis 2010, 17: 77-90.
- [173] Berman K, Kwo PY. Boceprevir, an NS3 protease inhibitor of HCV. Clin Liver Dis 2009; 13: 429-39.
- [174] Flint M, Mullen S, Deatly AM, et al. Selection and characterization of hepatitis C virus replicons dually resistant to the polymerase and protease inhibitors HCV-796 and boceprevir (SCH 503034). Antimicrob Agents Chemother 2009; 53: 401-11.

- [175] Hezode C, Forestier N, Dusheiko G, et al. Telaprevir and peginterferon with or without ribavirin for chronic HCV infection. N Engl J Med 2009; 360: 1839-50.
- [176] Hoofnagle JH. A step forward in therapy for hepatitis C. N Engl J Med 2009; 360: 1899-901.
- [177] Kao JH. Telaprevir for chronic HCV infection. N Engl J Med 2009; 361: 534-5.
- [178] Kronenberger B, Zeuzem S. Current and future treatment options for HCV. Ann Hepatol 2009; 8: 103-12.
- [179] Liu-Young G, Kozal MJ. Hepatitis C protease and polymerase inhibitors in development. AIDS Patient Care STDS 2008; 22: 449-57.
- [180] Mchutchison JG, Everson GT, Gordon SC, et al. Telaprevir with peginterferon and ribavirin for chronic HCV genotype 1 infection. N Engl J Med 2009; 360: 1827-38.
- [181] Mchutchison JG, Manns MP, Muir AJ, et al. Telaprevir for previously treated chronic HCV infection. N Engl J Med 2010; 362: 1292-303.
- [182] Njoroge FG, Chen KX, Shih NY, Piwinski JJ. Challenges in modern drug discovery: a case study of boceprevir, an HCV protease inhibitor for the treatment of hepatitis C virus infection. Acc Chem Res 2008; 41: 50-9.
- [183] Van der Meer AJ, De Knegt RJ. Telaprevir for chronic HCV infection. N Engl J Med 2009; 361: 533-4; author reply 34-5.
- [184] Gentile I, Viola C, Borgia F, Castaldo G, Borgia G. Telaprevir: a promising protease inhibitor for the treatment of hepatitis C virus infection. Curr Med Chem 2009; 16: 1115-21.
- [185] Suzuki F, Akuta N, Suzuki Y, et al. Rapid loss of hepatitis C virus genotype 1b from serum in patients receiving a triple treatment with telaprevir (MP-424), pegylated interferon and ribavirin for 12 weeks. Hepatol Res 2009; 39: 1056-63.
- [186] Akuta N, Suzuki F, Hirakawa M, et al. Amino acid substitutions in the hepatitis C virus core region of genotype 1b affect very early viral dynamics during treatment with telaprevir, peginterferon, and ribavirin. J Med Virol 2010; 82: 575-82.
- [187] Kawaguchi T, Yamagishi SI, Itou M, et al. Pigment epithelium-derived factor inhibits lysosomal degradation of Bcl-xL and apoptosis in HepG2 cells. Am J Pathol 2010; 176(1): 168-76.
- [188] Veldt BJ, Chen W, Heathcote EJ, et al. Increased risk of hepatocellular carcinoma among patients with hepatitis C cirrhosis and diabetes mellitus. Hepatology 2008; 47: 1856-62.

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# Structure-Function Relationships of PEDF

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Abstract: Pigment epithelial-derived factor (PEDF) is a 50-kDa secreted glycoprotein that belongs to the non-inhibitory serpin. It has an  $\alpha/\beta$  core serine-protease inhibitor domain, 3 major  $\beta$ -sheets, and 10  $\alpha$ -helices. Although PEDF does not inhibit either serine or cysteine proteinases, PEDF exerts diverse physiological activities including anti-angiogenesis, anti-vasopermeability, anti-tumor, and neurotrophic activities. Recent studies have shown that a variety of peptides derived from PEDF possess activities similar to those of the parent molecule through interactions with the extracellular matrix, binding to PEDF receptors, nuclear localization and phosphorylation. Thus, peptides derived from PEDF have therapeutic potential for various diseases and therefore, it is important to clarify the structure-function relationship of PEDF. In this review, we summarize structural features of PEDF that could affect various target organs such as blood vessels, tumors, and the central nervous system. In addition, since PEDF is recently identified as a regulator for glucose and lipid metabolism, we also discuss PEDF structures specially related to insulin-sensitizing and triglyceride-reducing properties.

**Keywords:** Pigment epithelial-derived factor, functional domain, anti-angiogenic activity, anti-vasopermeability activity, anti-tumor activity, neurotrophic activity, glucose metabolism, lipid metabolism.

Pigment epithelial-derived factor (PEDF) is widely expressed throughout the human body and has multiple biological activities. A variety of peptides derived from PEDF exerts diverse physiological activities including anti-angiogenesis, anti-vasopermeability, anti-tumor, and neurotrophic activities as shown in Table 1. In this review, we summarize structure-function relationships of PEDF.

## **REGULATION OF SECRETION OF PEDF**

C-terminal amino-acid residues play an important role in the secretion of various proteins [1-5]. The insertion of a reactive center loop (RCL) into the βsheet, which is called "loop-sheet polymerization" is involved in impaired secretion of various types of proteins [6, 7]. PEDF is a secretory protein, and the Cterminal of PEDF contains highly exposed typical RCL [8-10]. Truncation of the C-terminal tail of PEDF (Pro415-Pro418) inhibits the secretion of PEDF by Chinese hamster ovary cells [11]. Since Pro415 is mostly buried and interacts with Phe231 and Lue223, truncation of PEDF at Pro415 causes disruption of the hydrophobic interactions imposed by Pro415 and exposure of Asp414 to the negatively charged Cterminus, resulting in inefficient secretion of PEDF [11]. In addition, not only deletion of Pro373-Ala380, but also alanine substitution at Gly376 and Leu377 inhibits the secretion of PEDF. Gly376 and Leu377 are located within the highly exposed segment of the RCL. Therefore, these two residues are indispensable for (i)

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interactions of PEDF with components of the quality control system in the endoplasmic reticulum and (ii) subsequent efficient secretion of PEDF [11].

# INTERACTIONS OF PEDF WITH THE EXTRA-CELLULAR MATRIX

PEDF accumulates in the extracellular matrix [12]. The extracellular matrix is a complex of proteins, proteoglycans, and glycosaminoglycans, and plays a crucial role in the mechanical strength of cells and the regulation of cell proliferation and differentiation [13, 14]. It has been speculated, therefore, that PEDF exerts its diverse biological activities by interacting with different components of the extracellular matrix [15].

The crystal structure of human PEDF shows an asymmetrical charge distribution, which is one of the structural characters of PEDF [8]. A high density of basic residues exists at the center of  $\beta$  sheet A-strand 2 and 3, and helix F. This region is densely populated with lysines exposed to the surface (aa134, aa137, aa189, aa191, aa212, and aa124), which interact with various glycosaminoglycans [16-18].

The heparin-binding motif is XBBXBX (where B represents basic amino acids: X represents residues excluding acidic amino acids) [19] and is localized at the basic surface of PEDF (aa145-148), which is in the loop region between sheet 2A and helix E [8, 16]. Studies using site-directed mutagenesis showed that three clustered basic amino acid residues, Arg145, Lys146, and Arg148, are necessary for heparin binding [18]. Binding with heparin increases the proteolytic susceptibility of PEDF by trypsin and induces a conformational change in the vicinity of Lys178 of PEDF [20]. Heparin mediates the binding of PEDF to a receptor on the cell surface of Y-79 retinoblastoma

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Table 1. Functional Sites of PEDF

Amino Acids	Function	References
16-26	Anti-angiogenic activity	[28]
24	Protein kinase CK2 phosphorylation site	[59, 137]
	Neutrophic activity	
	Anti-angiogenic activity	
24-57	PEDF receptor binding site	[28]
	Anti-angiogenic activity	
	Apoptotic activity in endothelial cells	
32-380	Neurotrophic activity	[43]
39-57	Chemotaxis activity	[51]
	Apoptotic activity in endothelial cells	
40-64	Anti-tumor activity	[55]
	Osteogenic differentiation activity	
	Collagen I-binding site	
41-44	Collagen I-binding site	[22]
44-77	PEDF receptor (Laminin receptor) binding site	[31]
44-121	Neurotrophic activity	[43]
46-70	PEDF receptor (Laminin receptor) binding site	[31]
	Apoptotic activity in endothelial cells	
	Anti-migration activity in endothelial cells	
	Anti-tube-like formation activity in endothelial cells	
	Anti-angiogenic activity	
58-101	Neuroendocrine differentiation activity	[28]
64	Collagen I-binding site	[22]
78-94	PEDF receptor binding site	[28]
	Neuroendocrine differentiation activity	
	Neurotrophic activity	
78-102	Anti-proliferation activity	[55]
	Collagen I-binding site	
78-121	PEDF receptor (PNPLA2) binding site	[24, 29, 73, 91]
	Neuroendocrine differentiation activity	
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82-121	Neutrophic activity	[93]
90-114	Collagen I-binding site	[55]
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