

■ Table 42-1
Causes of Fanconi syndrome

Hereditary
• Dent disease
• Lowe syndrome
• Mitochondriopathies
• Cystinosis
• Galactosemia
• Hereditary fructose intolerance
• Glycogen storage disease type I (von Gierke disease)
• Fanconi-Bickel syndrome
• Tyrosinemia
• Wilson disease
• Idiopathic Fanconi syndrome
Acquired
• Nephrotic syndrome
• Myeloma
• Sjögren syndrome
• Renal transplantation
• Acute tubulointerstitial nephritis with uveitis (TINU) syndrome
• Autoimmune interstitial nephritis and membranous nephropathy
• Anorexia nervosa
• Untreated condition of distal renal tubular acidosis
Exogenous substances
• Drugs
• Chemical compounds
• Heavy metals

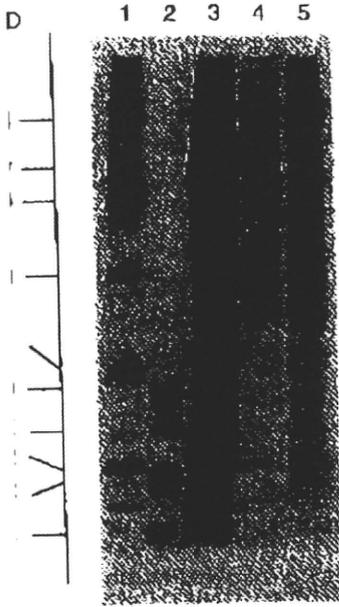
Dent disease is associated with inactivating mutations in *CLCN5* gene, which encodes 746 amino acids renal specific chloride channel-5 (ClC-5) (8, 47–49). ClC-5 belongs to the family of voltage-dependent chloride channels, which function as homodimeric proteins. ClC-5 is co-expressed with the vacuolar H⁺-ATPase and plays a key role in endosomal acidification that is a crucial function in the receptor-mediated endocytic pathway (50). More than 80 distinct *CLCN5* mutations are reported in patients with Dent disease. They are nonsense, missense, frameshift, splice-site, insertional, and deletional mutations, which result in total or partial loss of function. There are no genotype-phenotype correlations as various mutations are associated with different clinical phenotypes, even within the same family.

Numerous filtered proteins are bound to megalin and cubilin in the luminal membrane of proximal tubules, and the protein-receptor complex is incorporated into the endosome. The ligand and receptor are disassociated in the endosome; the receptor is recycled back to the luminal membrane and the reabsorbed proteins go into lysosome for further processing. This disassociation is dependent on acidification of the lumen of endosome by increased concentration of H⁺ and Cl⁻ due to the function of H⁺-ATPase and ClC-5 chloride channel. An abnormal endocytosis pathway due to ClC-5 dysfunction disturbs the recycling of megalin and cubilin, the back to the luminal membrane, and the expression of megalin and cubilin in the luminal membrane of proximal tubules, leading to LMW proteinuria, hypercalciuria, hyperphosphaturia, and nephrolithiasis. Proper acidification is also important for protein degradation in the endosome. Immunohistochemical analysis of proximal tubule cells in patients with Dent disease revealed an inverted polarity of the H⁺-ATPase, with redistribution to basolateral regions, suggesting that the loss of ClC-5 channel alters the function of components that co-distribute and physically interact with it (51).

Total urine protein ranges from 0.5–2.5 g a day, but may reach 4 g or higher in patients with Dent disease (45, 52). More than 60% of the filtered proteins are LMW proteins with molecular weight less than 45 KD (● Fig. 42-3). Nephrotic syndrome does not occur. LMW proteinuria is the most consistent and one of the earliest presenting abnormalities. Urinary beta 2-microglobulin, a LMW protein (MW = 11.6 KD), is excreted in amounts 100–300 times the upper limit of the normal. Albumin is also excreted in the urine. The pattern of proteins representing the increased excretion of several LMW proteins as well as albumin (MW = 65 KD) is termed as *tubular proteinuria* (52, 53). The terms of LMW proteinuria and tubular proteinuria have usually been used interchangeably (53).

Patients with Dent disease manifest hypercalciuria in the range of 4–10 mg kg⁻¹ of body weight a day in children and 4–6 mg kg⁻¹ of body weight a day in adults (54). Nephrocalcinosis is also found in children (● Fig. 42-4). Defective endocytosis of parathyroid hormone (PTH) in patients with Dent disease resulting in its persistence in the lumen of the proximal tubule stimulates 25-hydroxyvitamin D3 1-hydroxylase to produce more 1,25-dihydroxyvitamin D3 resulting in the increased serum levels of this vitamin. 25-hydroxyvitamin D3 is presented to 25-hydroxyvitamin D3 1-hydroxylase in the form of a complex with the vitamin D3-binding protein. As this complex is lost in the urine as a result of defective endocytosis leading to LMW proteinuria, the precursor

of urine from the family members of Dent polyacrylamide gel and stained by silver (MW proteinuria. Lanes 1 and 2, molecular , 12-year-old boy with Dent disease; Lane 4, e 5, his father.



nin D3 could be in short supply. The e of increased 1, 25-dihydroxyvitamin depend on the delicate balance between The slightly elevated serum levels of 1, amin D3 in patients with Dent disease eased intestinal Ca^{2+} reabsorption result- iuria (absorptive hypercalciuria). In fact, tients with *CLCN5* mutations manifest even though some of these do exhibit s (55). This may be explained by the out mice model; *Clc-5* disruption pro- rystal agglomeration, as well as a redistri- rystal-binding molecule annexin A2, in pithelial cells (56). f renal insufficiency and progression to failure are quite variable. Significant de- lar filtration rate is seen in children with d the patients fall into end-stage renal d of the age of 40s. Renal biopsy demon- or focal global glomerulosclerosis with tubular dilatation, and interstitial infil- cytes. Medullary nephrocalcinosis is a re in patients with Dent disease. Patients of age manifest medullary nephrocalci- e mechanism of progressive renal failure

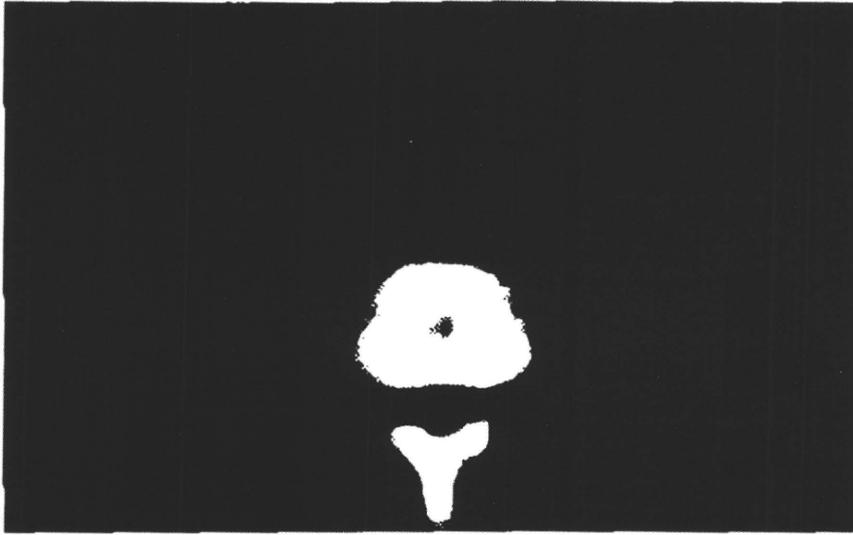
is not known in patients with Dent disease. Nephrocalci- nosis can be a candidate to disturb the glomerular filtra- tion rate. High urinary concentrations of potentially bioactive proteins including insulin, insulin like growth factor-1 (IGF-1), and the chemokine monocyte chemoat- tractant protein-1 (MCP-1) may contribute to interstitial fibrosis that will lead to progressive renal failure in patients with Dent disease (57). Generalized proximal tubule dysfunction is associated with increased cell prolif- eration, dedifferentiation, and oxidative stress resulting in interstitial fibrosis and eventual renal failure (58). Patients and carrier females often manifest nephrolithiasis, and renal stone is calcium phosphate stone that is also seen in patients with distal renal tubular acidosis.

Dent disease is genetically heterogeneous. Mutations in the *OCRL1* gene are identified in a subset of patients with the Dent disease phenotype (59). Unlike patients with typical Lowe syndrome, typical facial features, men- tal retardation, metabolic acidosis, and ocular abnormal- ities are usually absent in patients with Dent disease who have *OCRL1* mutation. The phosphoinositol 4,5-bispho- sphate phosphatase (PIP₂ 5-phosphatase) activity is markedly reduced in skin fibroblasts cultured from patients with Dent disease due to *OCRL1* mutations, and protein expression, measured by Western blotting, is reduced or absent. PIP₂ 5-phosphatase participates the trafficking and recycling of endosome in the proximal tubules. Defective PIP₂ 5-phosphatase activity can lead to endosomal dysfunction leading to LMW proteinuria. Unlike the patients with typical Lowe syndrome, none of patients have metabolic acidosis. These observations and findings suggest that *OCRL1* mutations can cause the isolated renal phenotype of Dent disease and affected individuals lack the cataracts, typical facial features, renal tubular acidosis, and neurologic abnormalities that are characteristic to Lowe syndrome. It is difficult to explain that *OCRL1* mutations can cause the isolated renal phenotype of Dent disease. However, it is possible that another phosphatase, *INPP5B*, which shares amino acid homology with *OCRL1*, can compensate phosphatase activity in patients with Dent disease due to *OCRL1* mutations.

There are no specific interventions at present that will change the natural course of renal manifestations and progressive renal failure in patients with Dent disease. Hypercalciuria is corrected by thiazide diuretics therapy in doses similar to effective doses for idiopathic hypercal- ciuria, presumably by stimulating the reabsorption of calcium in the distal convoluted tubule, where *Clc-5* channel is not expressed (60). However, this is not a long-term study which provides the evidence that it is

■ Figure 42-4

Abdominal CT demonstrating bilateral medullary nephrocalcinosis in a 12-year-old boy with Dent disease.



effective to prevent or delay the progression of end stage renal failure. In animal experiment using *clcn5* (mouse chloride channel 5 gene) knock-out mice, high citrate diets can delay the progression of nephrocalcinosis and end stage renal failure (61, 62). Treatment with an angiotensin-converting enzyme (ACE) inhibitor or angiotensin receptor blocker (ARB) may delay progression of end stage renal failure (63).

Low Syndrome

The oculocerebrorenal syndrome of Lowe (OCRL) is a rare X-linked disorder that is characterized by a complex phenotype that involves major abnormalities of the eyes (particularly congenital cataracts), central nervous system abnormality and FS (64). Cataracts are recognized in all the male patients. Lens opacities are present in all carriers of females by slit-lamp examination and this is a reliable screen for carriers (65). Other ocular abnormalities include glaucoma, microphthalmos, and corneal keloid formation. Visual acuity is frequently disturbed. Central nervous system abnormalities include infantile hypotonia (floppy infant), areflexia, and mental retardation. The patients manifest mild ventriculomegaly and cysts in the periventricular regions in the brain. Status epilepticus is sometimes recognized. FS is a major clinical feature and occurs in the 1st year of life, but the severity and age of onset vary. The patients manifest LMW proteinuria, glucosuria, aminoaciduria, hyperphosphaturia, hypercalciuria, hypophosphatemic rickets, hyperchloremic

metabolic acidosis, and progressive renal insufficiency (66). Renal insufficiency progresses at variable rate among patients, and it progresses to end-stage renal failure by 30s or 40s.

Like in patients with Dent disease, megalin is nearly absent from the urine in patients with Lowe syndrome suggesting of the decreased expression of megalin in the luminal membrane of the proximal tubule cells. Thus, urinary excretion of retinol-binding protein and the lysosomal enzyme N-acetyl-glucosaminidase are significantly increased in young boys with OCRL (67).

The gene (*OCRL1*) that is responsible for OCRL encodes a 105 KD Golgi protein with phosphoinositol 4,5-bisphosphate phosphatase (PIP₂ 5-phosphatase) activity (68). PIP₂ 5-phosphatase is mainly a lipid phosphatase that may control cellular levels of a critical metabolite, phosphatidylinositol 4,5-bisphosphate, and is involved in the inositol phosphatase signaling pathway (69). PIP₂ 5-phosphatase is present in cultured skin fibroblast and it is not present in peripheral blood cells. PIP₂ 5-phosphatase activity is markedly reduced in fibroblasts from patients with Lowe syndrome (70). However, this biochemical test for carrier diagnosis is not reliable; lyonization produces a highly variable pattern of tissue expression in females.

Deficiency of PIP₂ 5-phosphatase leads to cellular accumulation of its substrate PIP₂. PIP₂ accumulates in lysosomal membrane (71). PIP₂ is involved in signal transduction, vesicle trafficking and actin polymerization. Absence of PIP₂ 5-phosphatase activity leads to a reduction in the number and length of actin stress fibers, a tendency of actin fibers to depolymerize when provoked,

nal distribution of two actin-binding proteins, α -actinin. This disruption of actin has significant effects on epithelial function affecting cell-cell contacts such as tight junctions or by altering membrane transport proteins (72). Trans-Golgi network altered actin polymerization can explain the phenotype in Lowe syndrome. PIP₂ 5-phosphatase is localized to endosome and Golgi membranes along with clathrin, the mannose 6-phosphate receptor, and the early endosomal antigen 1 marker. OCRL1 phosphatase interacts with clathrin terminal domain and the clathrin adaptor protein AP-2. This interaction is important for PIP₂ 5-phosphatase in endosomal targeting and sorting (73, 74). OCRL1 is present in the early endocytic pathway, including in clathrin-coated pits, and demonstrate a close association between OCRL1 and adaptor molecules in the endocytic trafficking of receptor in the

cytoplasm; a C-terminal RhoGAP domain. OCRL1 has PIP₂ 5-phosphatase activity that binds to Rac1. Activated Rac GTPase stably associates with the AP-2 domain. In this sense, the protein OCRL1 can play a bifunctional role. Loss of the PIP domain and the resulting alteration in function may contribute to mental retardation in Lowe syndrome, as observed in other forms of X-linked mental retardation (76).

OCRL1 mutations can cause the isolated renal phenotype and affected individuals lack the characteristic facial features, renal tubular acidosis, and other abnormalities that are characteristic to Lowe syndrome. It is difficult to explain that OCRL1 mutations cause the isolated renal phenotype of Lowe syndrome; however, another phosphatase, INPP5B, has amino acid homology with OCRL1, can have phosphatase activity in patients with Denton-Drobny syndrome and OCRL1 mutations.

Over 100 OCRL1 mutations have been described in Lowe syndrome; nearly all are clustered in the region of exon 15, and almost none are in exons 1-9 (70).

Management is supportive and includes taking care of electrolyte imbalances, anticonvulsants, speech therapy, and ophthalmologic complications. The eye abnormalities usually appear early in life. Bicarbonate therapy is usually given in a dose of 2-3 mmol kg⁻¹ of body weight a day. Sodium or potassium phosphate can be given in a dose of 4 g a day for phosphate depletion and if vitamin D can be given.

Mitochondriopathies

The mitochondria (mt) have a major role in fatty acid oxidation, tricarboxylic acid cycle, urea cycle, and ATP production through the process of oxidative phosphorylation. Oxidative phosphorylation occurs at the level of the respiratory chain in the inner membrane of the mt (77). The respiratory chain comprises five complexes (Fig. 42-5). Complex I (NADH-coenzyme Q reductase) carries reducing equivalents from NADH to coenzyme Q and consists of different polypeptides, seven of which are encoded by mitochondrial DNA (mtDNA). Complex II (succinate-coenzyme Q reductase) carries reducing equivalents from FADH₂ to coenzyme Q and contains five polypeptides that are all encoded only by mtDNA. Complex III (reduced coenzyme Q-cytochrome c reductase) carries reducing equivalents from coenzyme Q to cytochrome c and contains 11 subunits, one of which is encoded by mtDNA. Complex IV (cytochrome c oxidase) transfers reducing equivalents from cytochrome c to oxygen. This complex is composed of cytochromes a and a₃, and 13 protein subunits, three of which are encoded by mtDNA. The fifth complex is ATP synthetase.

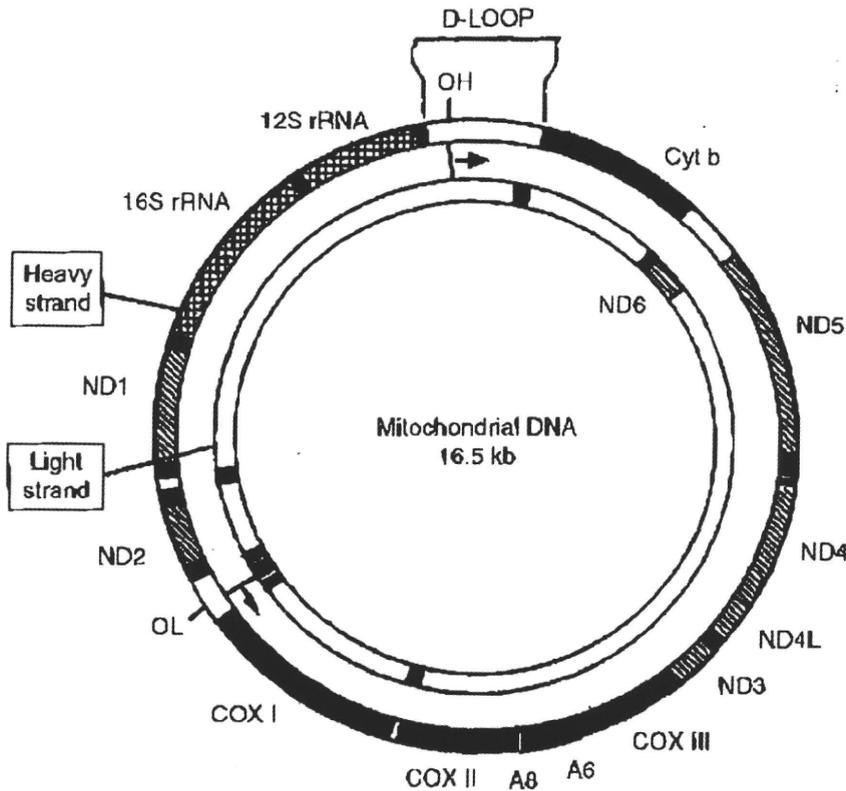
Oxidative phosphorylation consists of oxidative reactions that lead to oxygen consumption and phosphorylation of ADP to ATP. Each mt has its own 2-10 mtDNA. MtDNA genome is a 16.5 kb circular double stranded DNA with an asymmetrical base composition. The heavy strand contains more guanine residues while the light strand contains more cytosine residues. The mtDNA contains 37 genes; 13 encode for polypeptides of the respiratory chain, 2 for the ribosomal RNAs and 22 for transfer RNAs (78). Mitochondrial injury is due to congenital insults or may be the result of secondary events. Genetic defects of one or several polypeptide enzyme complexes of the oxidative phosphorylation system in the mt DNA or nuclear DNA which encodes structural or functional mitochondrial proteins can give rise to mitochondriopathies (mitochondrial cytopathies, mitochondrial diseases) (79).

Mitochondriopathies are multi-systemic disease and may begin at any age. Affected organs are diverse including central nervous system, muscle, liver, heart, kidney, gut, endocrine system, bone marrow, ear, eye, and skin (Table 42-2). They display extreme heterogeneity, and make unpredictable the extent and manifestations of disease presentation (80). With the course of the disease, the numbers of organs involved are increased.

The screening for mitochondriopathies includes the determination of plasma lactate, pyruvate, keton bodies and their ratios in fasted and fed patients, polarographic, and spectrometric studies to evaluate the different

■ Figure 42-5

Map of mitochondria genome. Regions encoding cytochrome b (ctb b), various subunits of NADH-coenzyme Q reductase (ND), cytochrome c oxidase (COX), ATPase, and ribosomal RNAs (rRNA) are indicated (Niaudet P, Rötig A. The kidney in mitochondrial cytopathies. *Kidney Int* 1997;51:1000-1007).



enzymatic complexes of the respiratory chain, muscle histologic studies, and genetic analysis (4).

Renal disease may be the first sign of mitochondriopathies, or it may appear simultaneously with neurological and neuromuscular signs (81). FS is particularly frequent in newborns, infants, and young children (82-84), whereas tubulointerstitial nephropathy is more frequent in children and young adults (85, 86), and can be associated with hereditary focal segmental glomerulosclerosis due to the mitochondrial transfer RNA gene mutation (87-89) and collapsing glomerulopathy due to the mutations in the gene *COQ2* encoding the para-hydroxybenzoate-polyprenyl-transferase enzyme of the CoQ10 synthesis pathway (CoQ2 nephropathy) and in the gene *PDSS2* encoding for decaprenyl diphosphate synthase (90, 91).

Most patients with FS due to mitochondriopathies manifest moderate FS including failure to thrive, dehydration, aminoaciduria, glucosuria, proteinuria, LMW proteinuria, phosphaturia, uricosuria, hypercalciuria, and bicarbonaturia. Many patients manifest FS by the age of 2 years. Extra-renal manifestations including neurological

symptoms, myopathy, hepatic dysfunction, clinical features of Pearson syndrome, partial adrenal insufficiency, cardiac involvement, diabetes mellitus, deafness, and ophthalmoplegia often manifest in the patients (12, 92, 93). Patients who manifested proximal tubular acidosis with hypercalciuria or Bartter syndrome were described (94, 95). Some patients manifest progressive renal failure (96). Histological analysis reveals tubular dilatations, tubular atrophy and cytoplasmic vacuolization of the tubules. Bizarre giant mitochondria are frequently observed (97).

No satisfactory treatment is presently available to alter the course of mitochondriopathies. The treatment is mainly symptomatic: supplements of sodium bicarbonate, potassium, vitamin D3, phosphate, and water are necessary. Carnitine is given in case of secondary carnitine deficiency. It includes avoidance of drugs that interfere with the respiratory chain such as valproate and barbiturates, or that inhibit mitochondrial protein synthesis such as tetracyclines and chloramphenicol. Dietary recommendations include a high lipid and low carbohydrate diet in patients with complex I deficiency. Hypercaloric diet and parenteral nutrition should be avoided in these patients.

Symptoms in patients with mitochondrialopathies

System	Symptoms
Respiratory system	Apnea, lethargy, hypotonia, coma, psychomotor retardation, cerebellar ataxia, stroke-like episodes, myoclonus, seizures, dementia, spasticity, headache, hemiparesis
Musculoskeletal	Myopathy, poor head control, limb weakness, myalgia, exercise intolerance
Liver	Hepatomegaly, liver dysfunction
Cardiac	Cardiomyopathy, arrhythmia
Kidney	Fanconi syndrome, tubulointerstitial nephropathy, nephrotic syndrome (focal segmental glomerulosclerosis, collapsing nephropathy), renal failure
Gastrointestinal	Vomiting, diarrhea, villous atrophy, cholemic pseudoobstruction, pancreatic dysfunction
Endocrine	Diabetes mellitus, growth hormone deficiency, hypoparathyroidism, hypothyroidism, adrenal insufficiency
Hematology	Sideroblastic anemia, neutropenia, thrombocytopenia
Ear, nose, and throat	Hearing loss
Eyes	Progressive extrarenal ophthalmoplegia, pigmentary retinal degeneration, ptosis, diplopia, cataract
Skin	Mottled pigmentation, trichothiodystrophy

reviewed in detail in Chapter 41 of this book. Here, therefore, only a short description is included. Cystinosis is an autosomal recessive lysosomal storage disease characterized by an accumulation of cystine in various organs, notably kidney, cornea, bone marrow, liver, and spleen (98). Renal dysfunction is the most characteristic feature of the disease and determines the clinical presentation and course of the disease. Cystinosis is the most common form of the FS in Western countries. Many patients, particularly in North America, have blonde or red hair. Other organs frequently affected include the thyroid, causing painful photophthalmopathy and hypothyroidism respectively.

Several clinical forms of the disease exist and are distinguished by the age at onset and severity of the symptoms. The most severe form, infantile cystinosis, is characterized by failure to thrive, polyuria, polydipsia, fluid and electrolyte loss, aminoaciduria, phosphaturia, renal tubular acidosis, and rickets within 12 months of age. Some of the patients with infantile cystinosis manifest severe growth failure. Renal function is usually normal at presentation. However, progressive glomerular impairment leads to focal segmental glomerulosclerosis and eventually to end stage renal disease by 10 years of age without treatment (99).

Patients with infantile cystinosis manifest FS, including hyperchloremic metabolic acidosis, aminoaciduria, hypokalemia, hypophosphatemia, glucosuria, and phosphaturia. There have been several patients of nephropathic cystinosis presenting with features of secondary Bartter syndrome (hypokalemia, hyperchloremic metabolic alkalosis, hyperreninemia, and hyperaldosteronism), suggesting abnormalities of Na^+ and Cl^- reabsorption (100, 101). Patients with cystinosis often manifest medullary nephrocalcinosis (102).

Renal histopathologic changes in infantile cystinosis include severe lesions of proximal tubules; typical alterations to the glomerular podocytes, which become multinucleated giant cells; and the presence of cystine crystals, mostly in interstitial cells and podocytes (103). The proximal tubule is the first clinical target of the disease, but cystine crystals are rarely found in the tubular cells of patients with cystinosis. Cystine crystal deposition in the cornea leads to photophobia. Continuous widespread cystine accumulation eventually leads to rickets and retinal, endocrinologic (hypothyroidism and impaired glucose tolerance), hepatic, gastrointestinal, muscular, and neurological abnormalities.

Two less severe and less common forms of cystinosis are juvenile (or late-onset) and ocular cystinosis. Patients with juvenile cystinosis manifest glomerular impairment between 12 and 15 years of age but do not suffer from severe tubulopathy or growth failure. Progression to end stage renal failure is slow and occurs at

Table 42-3

Clinical manifestations of infantile cystinosis

At presentation
Common
Failure to thrive
Polyuria and polydipsia
Fanconi syndrome
Vitamin D resistant rickets
Progressive renal failure
Photophobia
Hypothyroidism
Uncommon ^a
Bartter syndrome
Nephrotic syndrome
Diabetes insipidus
Pot-renal transplantation
Dysphagia
Myopathy
Exocrine pancreatic insufficiency
Diabetes mellitus
Central nervous system deterioration
Primary hypogonadism

Adapted from Gahl et al. (114)

^amay be transient and coexist with common manifestations

variable ages (104). Patients with ocular cystinosis do not involve kidney.

Infantile cystinosis is caused by mutations of the *CNTS* gene encoding cystinosin, a lysosomal transport protein, leading to complete abolition of cystine transport (105). Cystinosin has 367 amino acids and seven transmembrane domains. Cystine transport is dependent on the pH gradient, and the transport of cystine out of the lysosome is driven by the high H^+ content within the lysosomal lumen that is produced by the activity of the H^+ -ATPase. A range of mutations in *CNTS* gene has been described, but a single mutation, a 57-kb intragenic deletion, accounts for as many as three quarters of all European cases (105). The adolescent and ocular forms have one severe and one mild *CTNS* mutation, leading to reduced transport activity. The sparing of the kidney in patients with ocular cystinosis reflects tissue-specific expression of splicing factors, or the increased endogenous level of *CTNS* mRNA normally seen in the kidney (106). Individuals who are heterozygous for severe *CTNS* mutations reveal elevated levels of leukocyte cystine but are completely asymptomatic.

The pathophysiology of tubular cystine transport defects in patients with cystinosis is poorly understood, reflecting of an animal model for the disease. Knock-out mice model lacking cystinosin gene do not manifest signs of FS, despite accumulation of lysosomal cystine in the proximal tubules (107). Cystine-loaded proximal tubular cells demonstrate loss of free phosphate and defective ATP production and inhibition of Na^+ -dependent transporters (108). ATP depletion can reduce proximal tubular Na^+ , K^+ -ATPase activity leading to increased Na^+ delivery into the distal tubules and Bartter syndrome (101). A cell culture demonstrated that cells accumulated with intracellular cystine undergo apoptosis at a rate two- to four-fold higher than controls (109). Another works suggests that increased oxidative stress and altered redox status in proximal tubule cells cultured from the urine of patients with cystinosis are associated with proximal tubule dysfunction (110).

The diagnosis of cystinosis is confirmed by demonstrating elevated cystine levels in peripheral leukocytes (97). Corneal crystals detected by slit-lamp examination are diagnostic in childhood cystinosis because these crystals are not seen in patients with other hereditary FS. However, this finding is not sensitive for early diagnosis. The renal pathologic findings in infantile cystinosis consist of a chronic tubulointerstitial nephropathy, with characteristic multinucleated podocytes and intracellular crystalline inclusions in interstitial histiocytes (111). Although numerous multinucleated podocytes are the most characteristic pathologic findings, they are not found in the sclerotic glomeruli and detected only in low frequency (<4%). The cystine crystals are birefringent under polarized light in only alcohol-fixed tissue or in unfixed frozen tissue, because they are water-soluble and not retained in the tissue after routine histologic preparation with aqueous solutions (112).

The management and treatment for infantile cystinosis involve supportive therapy to maintain fluid balance and replace electrolyte losses at initial presentation. Early diagnosis and oral cysteamine, a cystine-depleting agent, can delay the progression of end stage renal failure and other organ involvement. Oral cysteamine therapy given at doses of 60–90 mg kg^{-1} of body weight (or between 1.3 and 1.95 g m^{-2}) a day divided every 6 h generally achieves approximately 90% depletion of cellular cystine, as measured in circulating leukocytes (<1.0 nmol half-cystine/mg protein) (113). The dosage recommended for adults is 500 mg every 6 h, but higher dosages are often required to achieve satisfactory cystine depletion. On the basis of its beneficial effects in maintaining thyroid function and depleting muscle of cystine, oral cysteamine

d continue in patients after renal transplan-
 to preserve other organs. Administration of
 nine eye-drops, given 6–12 times a day, can
 eal cystine crystals and lessen visual symp-
)ther therapies to supply potassium, alkali-
 including citrate or bicarbonate, phosphate,
 D3 are required. When the growth velocity
 yved and the patient remains below the 3rd
 height after one year of therapy, growth
 apy may be considered.

nia

is an autosomal recessive disease of galactose
 Nursing infants must move large amounts of
 ough Leloir pathway in order to utilize the
 ons for energy (● Fig. 42-6). Galactose is the
 on source in mammalian neonates, since it
 ed into glycogen more efficiently than is

frequent form is classic galactosemia that is
 deficient activity of galactose-1-phosphate
 rase (GALT) encoded by *GALT1* (116).
 es the reaction of galactose-1-phosphate
 s uridine diphosphate glucose to uridine
 galactose plus glucose-phosphate. Uri-
 ate galactose can be further metabolized
 ose or CO₂ and H₂O via glycolysis. Milk is
 e of galactose. Accumulated gal-1-p due to
 T and exposure to galactose lead to acute
 of multiple organ systems, including liver,
 , brain, and eye. Affected infant patients
 iting, diarrhea, failure to thrive, develop-
 liver dysfunction, coagulopathy, renal tubu-
 on, cerebral edema, vitreous hemorrhage,
Salmonella sepsis. They sometimes manifest jaun-
 -conjugated hyperbilirubinemia and may have
 sis. Liver damage leads to hepatomegaly and
 is potentially lethal. Neonatal screening
 udes galactosemia, anticipating that early
 l intervention would prevent long-term
 such as mental retardation, premature
 e, and speech delay. Although a galactose-
 it prevents the neonatal death, many
 patients continue to develop debilitating
 (117, 118). Clinically evident speech delay
 signs are more frequent than other findings.
 ovarian failure is nearly universal in females
 mia. The predominant manifestation due to

kidney damage is FS including hyperaminoaciduria,
 LMW proteinuria, hyperphosphaturia, and bicarbona-
 turia (118). Patients placed on a galactose-restricted diet
 are never truly free of galactose insult, as a significant
 amount of galactose is found in non-dairy foodstuffs
 such as vegetables and fruits (119, 120). More important-
 ly, galactose moieties can be produced endogenously from
 UDP-glucose via the UDP-4-galactose epimerase reac-
 tion, and natural turnover of glycoproteins/glycolipid;
 the rate of endogenous galactose synthesis ranges from
 0.53–1.05 mg kg⁻¹ of body weight a day (121, 122).
 Once the lactose is formed intracellularly, it will be con-
 verted to gal-1-p by GALT. The less common form of
 galactosemia is a deficiency of galactose kinase (GALK),
 which forms gal-1-p from galactose. These patients do not
 manifest either the acute toxicity syndrome or chronic
 complications seen in patients with classic galactosemia.
 They manifest cataracts. Since GALK-deficient patients
 do not accumulate gal-1-p in their tissues, gal-1-p is con-
 sidered to play a significant role in the pathogenesis of
 classic galactosemia (123, 124). GALT deficiency results in
 accumulation of toxic galactose leading to the unfolded
 proteins, altered calcium homeostasis and subsequently
 endoplasmic reticulum (ER) stress (125). ER stress caused
 by GALT-deficiency might contribute to accelerated apo-
 ptosis seen in the granulosa cells maturing follicles in
 galactosemic females, leading to premature ovarian failure
 (126). Formation of galactitol from galactose by aldose
 reductase has been proposed as a pathogenetic mecha-
 nism and is at least responsible for cataract formation.

The diagnosis is suggested by galactose or galactose
 1-phosphate in serum, or in the urine. The diagnosis is
 confirmed by demonstrating deficient GALT activity in
 red blood cells, fibroblasts, leukocytes, or hepatocytes.

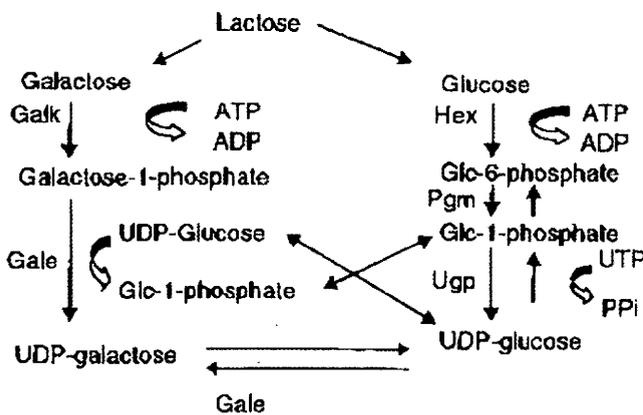
Treatment of this disorder is elimination of galactose
 from the diet. Acute symptoms and signs resolve within a
 few days after starting the diet therapy. However, devel-
 opmental delay, speech disturbance, ovarian dysfunction,
 and growth retardation are common outcomes in this
 disorder (127).

Hereditary Fructose Intolerance

Hereditary fructose intolerance (HFI) is an autosomal
 recessive disorder caused by a deficiency of aldolase B,
 an enzyme of liver, intestine, and renal cortex catalyzing
 the metabolism of fructose of exogenous origin (128).
 Frequency of HFI is estimated at 1 in 20,000 live births.
 Aldolase B catalyses the specific and reversible cleavage of

Figure 42-6

Composite diagram of the Leloir pathway and uridine diphosphate (UDP)-glucose pyrophosphorylase pathway. *Galk*, galactokinase; *galt*, galactose-1-phosphate uridylyltransferase; *Gale*, UDP-galactose 4-epimerase; *Hex*, hexokinase; *Pgm*, phosphoglucomutase; *Ugp*, UDP-glucose pyrophosphorylase (Leslie ND. *Insights into pathogenesis of galatosemia. Annu Rev Nutr* 2003;23:59–80).



fructose-1,6-bisphosphate (FBP) and fructose-1-phosphate (F1P) into dihydroxyacetone phosphate and D-glyceraldehyde-3-phosphate, or D-glyceraldehyde, respectively. Aldolase B is equally active with FBP and F1P, whereas aldolase A and aldolase C, the other two vertebrate isozymes, are more active with FBP than with F1P. Aldolase B is encoded in Aldolase B gene (*ALDOB*) mapped to chromosome 9q21.3–q22.2 (129, 130). Missense and nonsense mutations, large and small gene deletions and mutations in the splicing region have been identified in *ALDOB* of HFI patients (131).

Affected individuals manifest symptomatic hypoglycemia, vomiting and life-threatening episodes shortly after the intake of fructose or related sugars including sucrose and sorbitol (132). Prolonged ingestion leads to failure to thrive, hepatomegaly, jaundice, hepatic cirrhosis, and nephrocalcinosis, and may lead to convulsions, coma, and death from severe liver and kidney failure. Symptoms of HFI appear during infancy when infants with HFI are fed a formula or foods including fruits, vegetables, and sweetened cereals that contain sucrose. Patients with HFI may develop a protective aversion to sweets and fruits, which is a reason that diagnosis is frequently missed, and which also explains that reliable prevalence numbers for different populations do not exist.

HFI is associated with proximal tubule dysfunction leading to aminoaciduria, bicarbonaturia, phosphaturia, and lactic acidosis. These manifestations appear rapidly after the ingestion of fructose (133, 134).

The development of lactic acidosis adds significantly to the metabolic acidosis (135). Chronic fructose ingestion leads to nephrocalcinosis and impaired distal tubular function. In contrast, resolution of proximal tubule dysfunction can take days or weeks with strict restriction of fructose and sucrose (136).

Aldolase B coexists abundantly in endocytosis zones of the proximal tubule cells with H^+ -ATPase (137). Nonfunctional aldolase B impairs the coupling of H^+ -ATPase to glycolysis and endosomal acidification that will lead to FS.

Diagnosis includes the metabolic response to an intravenous fructose load or an enzymatic assay of liver or intestinal biopsy samples. However, both of them are bothering and invasive (138). Fructose breath hydrogen test is one of the standard procedures for the diagnosis. However, it can develop life-threatening adverse effects during the test (139). Molecular analysis is available for the diagnosis.

Strict avoidance of foods or drugs containing fructose, sucrose, and sorbitol is the predominant treatment.

Glycogen Storage Disease Type I (von Gierke Disease)

Glycogen storage disease type I (GSD-I) is a group of autosomal recessive disorders with an incidence of 1 in 100,000. There are two major subtypes. Glycogen storage disease type Ia (GSD-Ia, von Gierke disease) is common and is caused by a deficiency in glucose-6-phosphatase-alpha (G6Pase-alpha), a key enzyme in glucose homeostasis that catalyzes the hydrolysis of glucose-6-phosphate (G6P) to glucose and phosphate in the terminal step of gluconeogenesis and glycogenolysis (140). G6Pase-alpha is a hydrophobic endoplasmic reticulum-associated transmembrane protein. Glycogen storage disease type Ib (GSD-Ib) is caused by a deficiency of microsomal glucose-6-phosphatase transporter (G6PT). G6PT translocates G6P from cytoplasm to the lumen of the endoplasmic reticulum. Therefore, G6PT and G6Pase-alpha work in concert to maintain glucose homeostasis. Whereas G6Pase is exclusively expressed in gluconeogenic cells, G6PT is ubiquitously expressed and its deficiency generally causes a more severe phenotype.

Patients with GSD-Ia manifest a phenotype of disturbed glucose homeostasis characterized by fast life-threatening hypoglycemia, hepatomegaly, nephromegaly, hypercholesterolemia, hypertriglyceridemia, hyperuricemia, lactic acidemia, neutrophilia, and growth retardation (141, 142). Infants with GSD-Ia typically present with

epatomegaly at 6–8 months of age. Approx- of adolescent and adult patients develop adenoma (HCA), which can lead to con- idity and mortality (143). The incidence of tocellular carcinoma is recently increasing tients can live longer than before (144). The SD-Ia and GSD-Ib are associated with re- of life, independent functioning, and ele- f internalizing distress, and parental stress lthy peers.

lications include renal enlargement, gout renal stones, nephrocalcinosis, Fanconi-like chronic renal disease leading to renal insuf- . Hepatomegaly is a common finding in uricemia and uric acid stone in GSD-Ia by a combination of increased synthesis of competitive inhibition of renal tubular ex- c acid (urate) by lactate (146). Proximal iction has been observed in patients with its manifest proximal renal tubular acidosis bicarbonate in the urine, hyperphosphat- ed aminoaciduria and increased excretion oglobulin which are ameliorated by inten- py (147, 148). This finding suggests that c control can prevent proximal tubular dys- nic renal disease is a long-term complication. reveal interstitial fibrosis, tubular atrophy, renal glomerulosclerosis with marked glo- ment membrane (GBM) thickening and patients with GSD-Ia (149–151). Glycogen resent in the areas of abnormal GBM. The nt in the mesangium and in the epithelial, endothelial cells is increased. Recent treat- ficantly alleviated the metabolic abnormal- yed the clinical manifestation of chronic and renal insufficiency in patients with ver, glomerular hyperfiltration, hypercal- raturia that worsens with age, and urinary tion still occur in metabolically compen- with GSD-Ia (152, 153). Although the hanism responsible for chronic renal dis- rly understood, activation of the angioten- ggested to have an important role for the sion (154). The expression of TGF-beta 1 e is increased in a patient with GSD-Ia oteinuria, interstitial fibrosis, and tubular

ve of treatment is to maintain normogly- id metabolic complications and lactic oglycemia is accomplished at night with ding of glucose or with orally administered

uncooked cornstarch (156). A single dose (1.75–2.5 g kg⁻¹ of body weight) of uncooked cornstarch will maintain serum glucose concentration higher than 3.9 mmol L⁻¹ for 7 h in most young adults (157, 158).

Liver transplantation is indicated in the patients when medical treatment fails to control the metabolic problems or when HCA or hepatocellular carcinoma develops. Living-donor liver transplantation is a viable option to re- store normal metabolic balance and keeping normal renal function (159). Hepatocyte transplantation can be a po- tential therapeutic intervention to prevent hypoglycemia despite the discontinuation of cornstarch meals (160).

Fanconi-Bickel Syndrome

Fanconi-Bickel syndrome (FBS) is an autosomal recessive disorder characterized by failure to thrive, “doll-like” face, hepatomegaly, nephromegaly, and severe rickets. Patients with FBS manifest glycogen accumulation in hepatocytes and proximal tubular cells, fasting hypoglycemia, galac- tose intolerance, and FS including glucosuria, aminoacid- uria, hyperuricosuria, hyperphosphaturia, proteinuria, and sodium and potassium wastage (161, 162). Some patients manifest cataracts in neonatal period (163). Overall prognosis of FGS is considered as favorable (164). However, some patients manifest neonatal diabetes mellitus and galactosemia and die of hepatic failure dur- ing infancy (165).

FBS is caused by the mutations in facilitative glucose transporter gene (*SLC2A2*, also referred to as *GLUT2*) expressed in liver, kidney, intestine, and pancreatic islet cells (166). Over 60 mutations in *SLC2A2* were reported (167). This facilitative glucose transporter is expressed in hepatocytes, pancreatic beta cells, and renal and intestinal epithelial cells and is important for the exchange of glu- cose between these cell types and the bloodstream (168). Renal histology reveals an increase in mesangial cellularity, glomerulosclerosis, and patchy swelling of epithelial foot process and irregularly thickened lamina rara interna in the glomeruli, and vacuolization of epithelial nuclei in the proximal tubule cells suggesting the presence of glyco- gen in a 7 year-old patient with FBS (169).

The therapy for FBS is directed at the renal solute losses including sodium bicarbonate and potassium- sodium phosphate; treatment of rickets including active vitamin D3; and frequent feeding including night-time supplementation to prevent ketosis. Uncooked cornstarch has been shown to lessen hypoglycemia and improved growth (170). Galactose-free milk is also used for infant patients (165, 171).

Tyrosinemia I

Hereditary tyrosinemia type I (TI) is an autosomal recessive disorder of an amino acid metabolism. TI is due to the defect in the fumarylacetoacetate hydrolase (FAH) gene (172, 173). FAH is the last enzyme in the tyrosine catabolic pathway.

Patients with TI display a variety of clinical symptoms, such as liver damage from infancy that advances to cirrhosis, reduced coagulation factors, hypoglycemia, high plasma concentrations of methionine, phenylalanine, and aminolevulinic acid, high risk of hepatocellular carcinoma, and tubular and glomerular renal dysfunction (174).

Progressive renal damage begins from early infancy in severe form. Chronic liver damage with a high incidence of hepatoma (hepatocellular carcinoma) is characteristic in milder form (175). Even a patient without clinical manifestations of TI can manifest hepatoma during childhood (176). Accumulated fumarylacetoacetate in the patients with TI is pathogenic for hepatoma. Patients with milder form of TI are at risk for acute exacerbation of liver dysfunction. A common presentation mode is the "acute hepatic crisis" in which ascites, jaundice, and gastrointestinal bleeding are precipitated by an acute event such as an infection. Acute hepatic crises usually resolve spontaneously but on occasion progress to complete liver failure and encephalopathy. Acute, painful peripheral neuropathy may appear and can lead to transient paralysis. Autonomic dysfunction with hypertension and tachycardia can be associated with this acute neuropathy (177). Plasma tyrosine and methionine levels usually are elevated in untreated patients. The presence of succinylacetone in plasma and urine is diagnostic of TI. A rapid ultra performance liquid chromatography tandem mass spectrometric method is used for mass screening of tyrosinemia (178).

FS and developmental hypophosphatemic rickets are features of the kidney involvement. Generalized aminoaciduria, renal tubular acidosis, and mild proteinuria are also often seen, whereas glucosuria is less common because plasma glucose levels are low. Kidney enlargement is common, and nephrocalcinosis can be seen (179). FS leads to carnitine deficiency (180). Glomerulosclerosis and impaired GFR may be seen with time.

Disturbances in tyrosine metabolism lead to increased levels of succinylacetone and succinylacetoacetate. However, the mechanisms causing liver failure, cirrhosis, renal tubular dysfunction, and hepatocarcinoma are still unknown. Apoptosis of hepatocytes and renal tubular epithelial cells are characteristic features of this disease and the apoptotic signal in this disease seems to be initiated by

fumarylacetoacetate (181, 182). Accumulated maleylacetoacetate and fumarylacetoacetate in affected tissues can react with free sulfhydryl groups and reduce intracellular levels of glutathione. They may be capable of acting as alkylating agents. Maleylacetoacetate and fumarylacetoacetate are not detectable in plasma or urine but are converted to succinylacetoacetate. Succinylacetone, a metabolite of succinylacetoacetate, is structurally similar to maleic acid, which is known to induce FS and may be the cause of tubular dysfunction of HI. Experimentally, succinylacetone administration to rats leads to FS (183, 184).

Treatment with a low-phenylalanine and low-tyrosine diet dramatically improves the renal tubular dysfunction (185). However, this treatment cannot necessarily improve the hepatic involvement. Moreover, there is a risk of inducing deficiencies of phenylalanine or tyrosine. The formation of pathogenic fumarylacetoacetate is prevented by 2- (2-nitro-4-trifluoromethylbenzoyl)-1,3-cyclohexanedione (NTBC). NTBC is used for the patients with TI during the first 6 months of life in addition to a diet low in tyrosine and phenylalanine. NTPC clearly improves the vital prognosis and quality of life in the patients (186). However, some patients with NTBC treatment develop hepatoma. A rise of alpha-fetoprotein (AFP), a slow AFP decrease, and never normalizing levels of AFP are important predictors of hepatoma development (187). Liver transplantation has been used for patients with liver failure and to prevent the development of hepatoma (174). Liver transplantation leads to rapid correction of FS (188).

Wilson Disease

Wilson disease (WD, progressive hepatolenticular degeneration) is an autosomal recessive inborn error of copper (Cu) metabolism that affects numerous organ systems (189). Biliary excretion of Cu and incorporation into ceruloplasmin is impaired, leading to liver damage, neuronal degeneration, and impairment of other organs from accumulation of Cu in patients with WD.

The majority of patients with WD presents with either predominantly hepatic or neuropsychiatric symptoms, and with either clinically asymptomatic or symptomatic liver involvement. Approximately 40% of patients presents with liver disease, 40% with extrapyramidal symptoms, and 20% with psychiatric or behavioral abnormalities. Symptoms rarely occur before 6 years of age. Hepatic involvement includes acute hepatitis, fulminant hepatic failure, or progressive chronic liver disease in the form of either chronic

is or cirrhosis of the macronodular type. Neu-involvements are variable. The most common tation is bulbar symptoms characterized by th speech and swallowing, and drooling. They anifest dysarthria and coordination defects

movements accompanied by involuntary One third of the patients with WD manifest listurbances. The remaining patients with ent with symptoms including hemolytic ane- acture, arrhythmias, FS, hyperpigmentation, er ring, cataract, and gynecological problems utable to the involvement of the organs.

used by a mutation in the gene *ATP7B* that type Cu transporting ATPase beta polypep- *ATP7B*) (190). This ATPase is targeted to the 1, suggesting that its role in Cu dependent kes place in this organelle. The disease estimated to be between 1 in 5,000 and 1 in the carrier frequency is approximately

orbed by the intestinal cells and stored with ein in a non-toxic form. The Cu is later y the circulation by a Cu transporter 15,000 protein, Cu-transporting P-type ATPase 1 ich is located on the membrane of entero- 42-7) (192). It is then transported to the liver dbumin, from where it is accepted by hepa- ATOX1 chaperone protein directs Cu to its ts in the hepatocytes. Some of Cu bounds to ain for storage, and the remainders are ex- *ATP7B*-regulated biliary canaliculi. *ATP7B* : the transfer of Cu to apoceruloplasmin to Cu binding protein, ceruloplasmin (193). n is released into the blood, carries 90% of nt in the plasma, and acts as a source for gans. Mutations of the *ATP7A* gene result in f Cu in enterocytes, preventing entry of Cu ulation and thus causing a complete Cu enkes kinky hair disease) (192).

s in *ATP7B* gene lead to a reduction in the of apoceruloplasmin into ceruloplasmin, uly present at low levels in the patients of e to excrete Cu into the biliary canaliculi : effect to hepatocytes. Excess Cu damages , which produce oxidative damage to the ws spillage of Cu into the blood, thereby other tissues including the brain, kidney, l cells, initiating toxic effects.

accumulation of Cu in the kidney leads to dysfunction in patients with WD. Patients atures of FS before the onset of hepatic

failure and is characterized by intermittent glucosuria, aminoaciduria, hyperphosphaturia, hyperuricosuria, and proteinuria (194-196). Patients can manifest rickets or osteomalacia, hypercalciuria, urolithiasis, nephrocalcinosis, decreased urine concentrating ability and distal renal tubular acidosis are reported (197). Glomerular function decreases as the disease progresses, but death from extra-renal causes occurs before the onset of renal failure.

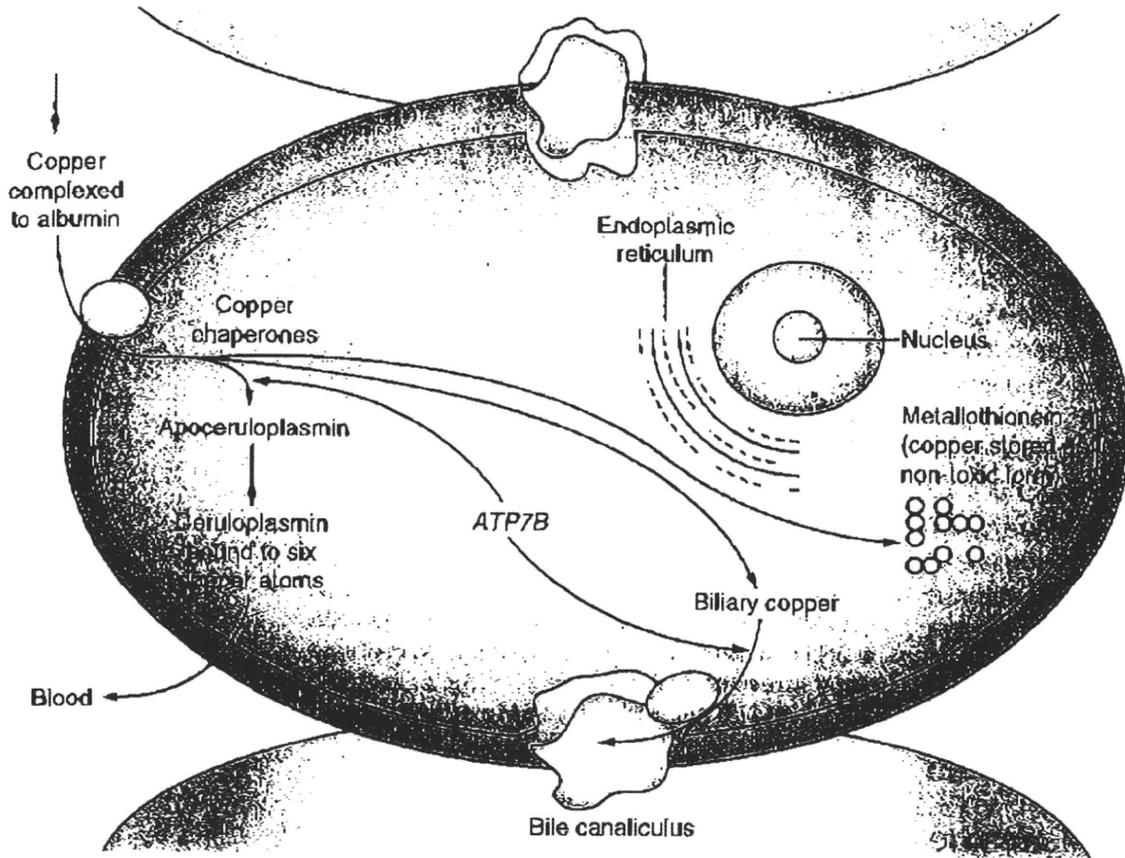
Histological analysis of the patients with WD revealed no alteration on light microscopy or only flattened proximal tubule cells without recognizable brush border (198). Electron microscopy disclosed loss of the brush borders, electron dense deposit in the subapical region of tubular cytoplasm probably representing metalloproteins, and cavitation of the mitochondria with disruption of the normal cristae pattern (199).

All subjects presenting with symptomatic or asymptomatic liver disease with no apparent causes or with extrapyramidal features along with a past or family history of similar hepatic or neurological illnesses in other siblings should be screened for WD (200). Measurement of the serum ceruloplasmin is valuable for diagnosis. Any value below 200 mg L^{-1} is abnormal, and reduced levels are seen in up to 95% of the patients with WD. An estimation of 24 h urinary Cu excretion is another reliable test for the diagnosis of WD. Normal excretion is between 20 and $50 \text{ } \mu\text{g}$ a day, and Cu excretion is increased to in excess of $100 \text{ } \mu\text{g}$ a day in patients with WD. Serum free Cu is a measure of nonceruloplasmin toxic Cu in the blood, and normal value range from 1.3 to $1.9 \text{ } \mu\text{mol L}^{-1}$ (8 - $12 \text{ } \mu\text{g dL}^{-1}$) in parallel with the increased urinary Cu excretion, because of saturation of the hepatic storage of Cu. A hepatic biopsy and a measurement of its Cu content are helpful for diagnosis. 80% of the patients manifest increased hepatic Cu ($>250 \text{ } \mu\text{g/g}$ of dry tissue weight). Genetic analysis of *ATP7B* gene is a helpful confirmation of WD. Brain MRI is a very sensitive method for revealing abnormalities in patients with WD. Generalized brain atrophy and hyper-intensity in the basal ganglia, white matter, thalamus, or brainstem are common findings in the patients. Patients exhibit characteristic features on MRI; "face of the giant panda" is seen in the midbrain and "face of the miniature panda" is seen in the tegmentum region of the pons in T2-weighted images (201, 202).

WD has a fatal outcome if not treated appropriately and in a timely manner. The aim of treatment for WD is to remove the toxic deposit of Cu from the body to produce a negative Cu balance, and to prevent its re-accumulation (190, 203). Successful therapy is measured in terms of a restoration of normal levels of free serum Cu and its excretion in the urine. The average daily diet

Figure 42-7

Schematic representation of copper metabolism within a liver cell. *ATP7B*, causative gene for Wilson disease. (Das SK, Ray K. Wilson's disease: an update. *Nat Clin Pract Neurol* 2006;2:482-493).



contains 2–4 mg of Cu, and 0.8 mg is normally lost into the feces. Patients should avoid Cu-rich foods including chocolate, nuts, shellfish, mushrooms, and liver. D-penicillamine therapy (20 mg kg⁻¹ of body weight a day) has been the most commonly used chelating agent that may reverse multiple tissue dysfunction including FS. However, use of D-penicillamine has been questioned, because of reported side effects. The side effects from D-penicillamine can occur both early and late in the treatment period. Early side effects include a hypersensitivity reaction characterized by fever, skin rash, and lymphadenopathy. Delayed side effects including Goodpasture's syndrome, polymyositis, systemic lupus erythematosus, and bone marrow suppression are caused by immunological reactions. Trientine is another effective chelating agent (750–2,000 mg a day for adults). Ammonium tetrathiomolybdate (2–3 mg kg⁻¹ of body weight a day in six doses along with meals and in the interval between meals) is a potent agent in removing Cu from the body and may be the drug of choice for patients with neurologic disease to prevent the immediate worsening of

symptoms that can occur with D-penicillamine therapy. Zinc acetate or sulfate induces intestinal metallothionein and helps in the prevention of Cu absorption from the gut (3–5 mg kg⁻¹ of body weight a day in three divided doses before meals). D-penicillamine should be taken 2 h after meals to avoid any interaction with the zinc. Zinc acetate or zinc sulfate has been used successfully in asymptomatic or presymptomatic affected family with WD, and is equally as effective as D-penicillamine in a group of patients predominantly with neurological disease (204, 205). The use of trientine, tetrathiomolybdate, and zinc has been advocated, although results of long-term trials are awaited. The best therapeutic approach remains controversial and there is no universally accepted regimen. Liver transplantation is effective for the patients with progressive liver failure or acute liver failure. Liver transplantation is also indicated for patients with WD in whom medical therapy is ineffective. Symptomatic patients with WD require lifelong treatment, because an interruption to therapy or inadequate treatment can lead to fatalities within 9 months to 3 years (189, 203).

Fanconi Syndrome

significant numbers of genetic causes for the leading to FS are identified, there exist patients with idiopathic FS. Idiopathic FS should be diagnosed when all other known causes have been excluded. Idiopathic FS can be inherited as an autosomal dominant, autosomal recessive (212, 213), and X-linked (214). However, most of the familial forms are autosomal dominant inheritance.

Patients with idiopathic FS manifest failure to thrive, recurrent episodes of dehydration, and rickets. They often have clinical features of FS, including polyuria, polydipsia, hypophosphatemia, proximal renal tubular dysfunction, nocturia, aciduria, glucosuria, and proteinuria. Renal filtration rate is usually normal during childhood, but some develop chronic renal insufficiency and renal failure 10–30 years after the onset of disease (208, 209, 213). Nephrocalcinosis and genu valgum (knee) are seen in some patients (213). Osteoporosis, bone fracture, and scoliosis due to osteomalacia are serious complications in adult patients with idiopathic FS (Fig. 42-8).

Renal biopsy reveals chronic tubulointerstitial nephropathy. The interstitium demonstrates patchy fibrosis, tubular atrophy and focal collections of

mononuclear inflammatory cells. Occasional cystically dilated tubules are seen containing eosinophilic proteinaceous material that stains positive for PAS (213).

Treatment for idiopathic FS remains symptomatic. Careful follow-up of these patients is necessary to prevent recurrent bouts of dehydration, electrolyte imbalance, and metabolic bone diseases. Glomerular function and nephrocalcinosis must be checked regularly. Renal transplantation has been done in a few patients who had end-stage renal failure (209).

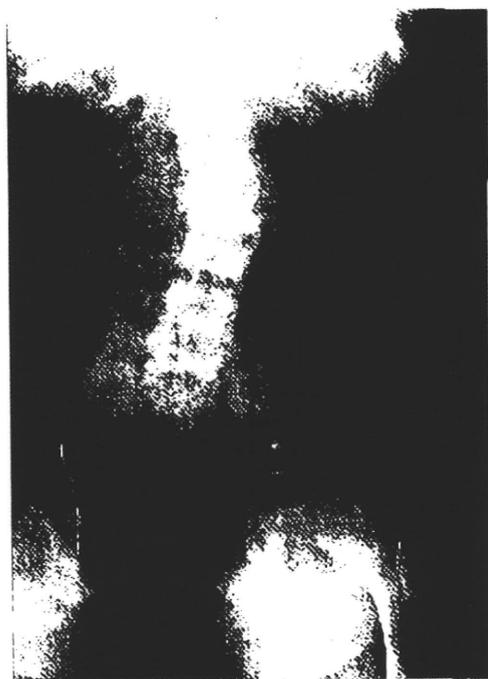
Acquired Fanconi Syndrome

Nephrotic syndrome is associated with FS (215). The renal pathology is focal segmental glomerulosclerosis. Although the true pathogenesis is not clarified, mitochondrialopathies can manifest FS, focal segmental glomerulosclerosis leading to nephrotic syndrome, and both (79–84).

Immunological or hematological disorders are associated with dysproteinuria leading to FS. They are multiple myeloma, Sjögren syndrome, and amyloidosis. Almost all of the patients with these diseases are adults. In early stages of myeloma, light chain nephrotoxicity often presents with proximal tubular functional abnormalities leading to FS. Proximal tubule dysfunction is the most common mode of renal involvement and it can manifest in a variety of ways. Endocytosis in the proximal tubules is overloaded and cell stress responses that include phosphorylation of MAPKs, prominently, p38 MAPK, and nuclear transcription factors NF-kappa B, AP-1 are activated resulting in production of inflammatory and proinflammatory cytokines, TNF-alpha, interleukin-6, 8 and monocyte chemo-attractant protein-1 (216). These proximal tubule alterations often progress to a severe tubulointerstitial nephritis and end stage renal failure.

Sjögren syndrome is an autoimmune connective tissue disorder that affects exocrine glands. Renal involvement of Sjögren syndrome is mainly manifested as tubular disorders; 70% of the patients manifest distal renal tubular acidosis (217). Urinary concentration defect, proteinuria and LMW proteinuria are often seen in the patients. Only 4% of the patients manifest FS (217). Patients with FS and Sjögren syndrome manifest osteomalacia including bony deformities of rib cage, bilateral humeral shaft fractures, and marked cortical bone thinning (218). Characteristic histological feature of Sjögren syndrome is chronic interstitial nephritis, with diffuse or focal plasmacytoid lymphocytic infiltration. In the late stage of the disease, tubulointerstitial fibrosis is severe. Corticosteroid or/and immunosuppressant therapy can improve the prognosis.

Figure 42-8. Lateral view of spine showing scoliosis and osteopenia in a 35-year-old patient with idiopathic Fanconi syndrome.



FS has appeared rarely after renal transplantation (219). Acute tubular necrosis, chronic rejection reaction, and nephrotoxic drugs can induce the progression of FS in the patients.

Acute tubulointerstitial nephritis with uveitis (TINU) syndrome is an immunological disease that leads to tubulointerstitial nephritis and anterior uveitis (220). Patients with TINU syndrome manifest asthenia, malaise, weight loss, nocturia, and thirst. Patients also manifest incomplete or complete FS including proteinuria, LMW proteinuria, glucosuria, aminoaciduria, bicarbonaturia, phosphaturia, and uricosuria due to proximal tubule dysfunction and acute renal failure (221). Urine concentration is decreased in the patients. Corticosteroid therapy can improve renal and eye manifestations.

Autoimmune interstitial nephritis and membranous nephropathy is a distinct disorder. The patients manifest failure to thrive, multiple renal tubular disorders including FS and proteinuria (222, 223). Renal biopsy revealed interstitial nephritis with lymphocytic infiltration and fibrosis, and membranous nephropathy. In advanced stage, focal segmental glomerulosclerosis and tubular atrophy develop. Immunofluorescence analysis shows linear staining of IgG along the glomerular capillaries and the tubular basement membrane. These renal lesions result from an autoimmune response to the 58-kD tubular basement membrane autoantibody (224). This disorder is genetically related to HLA B7 serotype (224).

A patient with anorexia nervosa is described to manifest reversible FS like condition including glucosuria, phosphaturia, and uricosuria, although the precise pathogenesis is not known (225). These manifestations subside with nutritional recovery.

Untreated patients with distal renal tubular acidosis manifest LMW proteinuria, generalized aminoaciduria, phosphaturia, uricosuria, and hypercalciuria (226, 227). These proximal tubular abnormalities are transient and disappear by the alkali and potassium therapy. Although the precise pathogenic mechanisms underlying the development of proximal tubular dysfunction remains unclear, decreased pH in the cytoplasm of the proximal tubule cells resulting from the intracytoplasmic accumulation of H^+ due to luminal membrane H^+ -ATPase dysfunction can disturb trafficking of endosome.

Exogenous Factors

Drugs

Numerous drugs and herbs are implicated in the pathogenesis of FS. Drugs and herbs are usually filtered from

the glomerulus and reabsorbed in the proximal tubules. They include outdated tetracycline (228), aminoglycosides (229, 230), salicylate (231), valproic acid (232, 233), and Chinese herbs (234, 235). Aminoglycoside antibiotics reduce glucose reabsorption in kidney tissue by reducing mRNA, protein expression, and function of the Na^+ -dependent glucose transporter, which is located in the luminal membrane of the proximal tubule (236). Covalent binding of salicylate or its metabolites to mitochondria in proximal tubule cells alters the function of mitochondria (231). Valproic acid produces the defects of mitochondrial respiratory chain and lysosomal enzyme activity in the proximal tubule cells leading to multiple renal transport abnormalities (13, 237). Chinese herbs containing aristolochic acids cause proximal tubular injury, and this is called as aristolochic acid-related nephropathy.

A number of cancer chemotherapy agents are associated with renal glomerular and tubular dysfunctions including FS. The nephrotoxicity of cancer chemotherapy agents is dose dependent and often irreversible. Ifosfamide is an alkylating agent widely used in the treatment of various solid tumors. Chloroacetaldehyde (CAA), one of the main metabolites of ifosfamide, contributes to inhibit endocytosis in the proximal tubule cells (238). CAA decreases total glutathione and ATP levels in the proximal tubule cells. CAA also inhibits endosomal H^+ -ATPase activity, which disturbs intracellular vesicle trafficking (239). Patients receiving ifosfamide who have received prior cisplatin are at significantly higher risk of developing FS than are those who have received no prior nephrotoxic therapy (240). When the patients manifest FS, renal sonography reveals hyperechogenicity of the parenchyma with good corticomedullary differentiation (241). Taurine can protect against ifosfamide-induced renal dysfunction without compromising its anti-tumor activity (242). Cisplatin also reduces glucose reabsorption in kidney tissue by reducing mRNA, protein expression, and function of the Na^+ -dependent glucose transporter (243). Cisplatin inhibits various types of amino acid transporters in the proximal tubule cells leading to a generalized aminoaciduria (243).

Imanitinib mesylate is a specific tyrosine kinase inhibitor that is the first line therapy for patients with chronic myeloid leukemia. This agent induces partial FS including phosphaturia and uricosuria with mild renal failure. Combined blockade of both platelet-derived growth factor receptor and c-Kit receptor tyrosine kinase in proximal tubules causes partial FS (244).

Nucleotide reverse transcriptase inhibitors that are used as anti-human immunodeficiency virus (HIV) agents including adefovir, cidofovir, and tenofovir induce

nic diabetes insipidus, and acute renal failure. Adefovir and cidofovir interact with organic anion transporters (OAT); these drugs enter into proximal tubules by activated OAT located in the basolateral membrane. However, their efflux into the tubular lumen is reabsorbed by inactivated multidrug-resistance-1 (MDR1) located in the luminal membrane. Drugs are accumulated in the proximal tubule due to mitochondrial damage and tubular toxicity of adefovir and cidofovir is proportional to mitochondrial ATP expression (246). Histologic and ultrastructural examination reveals tubular degenerative changes in proximal tubules with swollen and dysmorphic mitochondria. In tubular cells, respiratory chain components and mitochondrial DNA (cytochrome oxidase I) are selectively deficient in renal tubular cells. Mitochondrial DNA is quantitatively reduced. In contrast to adefovir and cidofovir, renal toxicity of tenofovir is much less frequent. Tenofovir has little tubular toxicity and it does not interact with OAT; therefore, the precise mechanisms of nephrotoxicity of tenofovir remain unknown.

Compounds

Non-selective herbicide, and colloidal bismuth subcitrate cause FS (251, 252). Large amount of these compounds are usually ingested in a suicide attempt. They cause FS and acute renal failure. Treatment with the chelating agent sodium 2,3-dimercaptosuccinate in combination with hemodialysis is effective in reducing the serum bismuth level. Methylenediamine (diachrome) (253), 6-mercaptopurine and thiothamizone also lead to FS (255).

Metals

Heavy metals such as lead, cadmium, mercury, chromium are a major environmental and occupational hazard. They are very toxic at very low doses. The first target organ of heavy metal toxicity is the kidney. renal damage by heavy metals depends on the metal, dose, route, and duration of exposure. Both acute and chronic intoxication have been demonstrated to cause Fanconi's syndrome, with various levels of severity ranging from mild renal dysfunctions like acquired FS to severe renal failure (256). Lead poisoning leads to FS, predominantly in children (257). As lead is non-biodegradable and has a long biological half-life, aminoaciduria and

glycosuria persist up to 13 years after childhood severe lead poisoning (258).

Cadmium intoxication leads to FS after a long exposure (259). The industrial waste contaminating cadmium in the Jinzu River basin in Toyama prefecture in Japan produced a lot of patients with *Itai-Itai* (ouch-ouch) disease that is compatible to FS with severe osteomalacia. Patients complained severe bone pains that are derived from advanced non-traumatic multiple bone fractures. Cadmium produces free radicals that alter mitochondrial activity or induce mitochondrial gene deletion in the proximal tubules (260, 261). Cadmium inhibits H⁺-ATPase, which results in a Fanconi-like syndrome (6).

Therapy

Identification of the underlying cause for FS is a first step and is critical to direct specific therapy. Avoidance of offending nutrients in galactosemia, HFI, and tyrosinemia and avoidance of Cu-rich foods in WD are therapeutically critical. Specific treatments with Cu-chelating agents including D-penicillamine, trientine, and ammonium tetrathiomolybdate, and zinc are effective for WD. Immunosuppressive drugs are used for immunologically induced disorders including Sjögren syndrome, TINU syndrome and autoimmune interstitial nephritis and membranous nephropathy. These treatments can completely resolve FS.

When specific therapy does not exist, therapy is directed at the biochemical abnormalities secondary to renal solute and fluid losses and the metabolic bone diseases. Proximal renal tubular acidosis usually requires large amount of alkali (2–15 mEq kg⁻¹ of body weight a day) divided into four to six daily doses. High dose of alkali can produce volume expansion, further bicarbonate wasting and potassium loss in the patients with FS. 1–3 mg kg⁻¹ of body weight a day of hydrochlorothiazide can reduce the dose of alkali by preventing the volume expansion. Administration of potassium salt of citrate, bicarbonate, or acetate fulfills the dual purpose of treating acidosis and preventing hypokalemia. Sodium wasting and dehydration are treated with combination of sodium bicarbonate, citrate, and chloride, depending on the degree of acidosis. Ensuring adequate fluid and electrolyte intake is essential, especially in the case of infants or gastrointestinal diseases. Early intervention with intravenous replacement therapy is required for the patients with FS who manifest vomiting and diarrhea.

Hypophosphatemia and impaired renal vitamin D3 metabolism in patients with FS lead to rickets and other metabolic bone diseases. 1–3 g of phosphate supplementation

is necessary as neutral phosphate (the mixture of sodium phosphate dibasic 1.94 g and potassium phosphate monobasic 0.34 g contains 0.5 g of phosphate) divided into four to six daily doses. Supplementation of 1,25-dihydroxyvitamin D₃ or dihydrotachysterol is effective to treat or prevent rickets and osteomalacia. Vitamin D₃ therapy improves the hypophosphatemia and lessens the risk of hyperparathyroidism. Hypercalcemia and hypercalciuria are toxic side effects of vitamin D₃ therapy. An adequate amount of physical activity, as well as appropriate diet with calcium, phosphate, and vitamin D₃, is necessary to prevent bone deformations, non-traumatic fractures leading to bone pain, deterioration of motor development and disability (262).

Aminoaciduria, glucosuria, proteinuria, LMW proteinuria, and uricosuria usually do not induce clinical symptoms and do not require specific treatments.

Growth failure is a major complication in FS. Despite correction of electrolyte abnormalities, some patients manifest severe growth retardation, especially those with cystinosis and Fanconi-Bickel syndrome. A patient with FS was reported to have growth hormone deficiency (263). Supplemental growth hormone has been used successfully in a few patients with FS.

References

- De Toni G. Remarks on the relations between renal and rickets (renal dwarfism) and renal diabetes. *Acta Paediatr* 1933;16:479-484.
- Debré R, Marie J, Cléret F et al. Rachitisme tradit coexistent avec une nephrite chronique et une glycosurie. *Arch Med Enf* 1934;37:597-606.
- Fanconi G. Die nicht diabetischen glykosurien und hyperglykämien des altern kinds. *Jahrb Kinderheilkd* 193;133:257-300.
- Marshansky V, Bourgoïn S, Londino I et al. Receptor-mediated endocytosis in kidney proximal tubules; recent advances and hypothesis. *Electrophoresis* 1997;18:2661-2676.
- Brown D, Stow JL. Protein trafficking and polarity in kidney epithelium; from cell biology to physiology. *Physiol Rev* 1996;76:245-297.
- Herak-Kramberger CM, Stow JL. Protein trafficking and polarity in kidney vacuolar H⁺-ATPase and endocytosis in rat cortex. *Kidney Int* 1998;53:1713-1726.
- Marshansky V, Richard M, Bartle J et al. Regulation of renal albumin reabsorption by endosomal proton transport [Abstract]. *J Am Soc Nephrol* 1996;7:1311.
- Lloyd SE, Pearce SH, Fisher SE et al. A common molecular basis for three inherited kidney stone diseases. *Nature* 1996;379:3445-3449.
- Norden AGW, Lapsley M, Igarashi Tet al. Urinary megalin deficiency implicates abnormal tubular endocytotic function in Fanconi syndrome. *J Am Soc Nephrol* 2002;13:123-133.
- Sakarcan A. The Fanconi syndrome of cystinosis: insights into the pathophysiology. *Tur J Paediatr* 2002;44:279-282.
- Rech VC, Athaydes GA, Feksa LR et al. Inhibition of creatine kinase activity by cystine in the kidney of young rats. *Pediatr Res* 2006;60:190-195.
- Niaudet P, Rötig A. The kidney in mitochondrial cytopathies. *Kidney Int* 1997;51:1000-1007.
- Hawkins E, Brewer E. Renal toxicity induced by valproic acid (Depaken). *Pediatr Pathol* 1993;13:863-868.
- Magen D, Sprecher E, Zelikovic I et al. A novel missense mutation in SLC5A2 encoding SGLT2 underlies autosomal-recessive renal glucosuria and aminoaciduria. *Kidney Int* 2005;67:34-41.
- Bingham C, Ellard S, Cheret C et al. The generalized aminoaciduria seen in patients with hepatocyte nuclear factor-1 alpha mutation is a feature of all patients with diabetes and is associated with glucosuria. *Diabetes* 2001;50:2047-2052.
- Tokaymat A, Sakarcan A, Neiberger R. Idiopathic Fanconi syndrome in a family. I. Clinical aspects. *J Am Soc Nephrol* 1992;2:1310-1317.
- Haffner D, Weinfurth A, Seidel C et al. Body growth in primary de Toni-Debré-Fanconi syndrome. *Pediatr Nephrol* 1997;11:40-45.
- Flyvbjerg A, Dørup I, Everis ME et al. Evidence that potassium deficiency induces growth retardation through reduced circulating levels of growth hormone and insulin-like growth factor I. *Metabolism* 1991;40:769-775.
- Tsao T, Fawcett J, Pervenzas FC et al. Expression of insulin-like growth factor-I and transforming growth factor-beta in hypokalemic nephropathy in the rat. *Kidney Int* 2001;59:96-105.
- Brünger M, Hütler HN, Krapf R. Effect of chronic metabolic acidosis on the growth hormone/IGF-I endocrine axis: new cause of growth hormone insensitivity in humans. *Kidney Int* 1997;51:216-221.
- Hsu SY, Tsai HJ, Tsau YK. Comparison of growth in primary Fanconi syndrome and proximal renal tubular acidosis. *Pediatr Nephrol* 2005;20:460-464.
- Tsilchorazidou T, Yovos JG. Hypophosphataemic osteomalacia due to de Toni-Debré-Fanconi syndrome in a 42-year old girl. *Hormones (Athens)* 2005;4:171-176.
- Urabe Y, Tagami T, Suwabe T et al. A patient with symptomatic osteomalacia associated with Fanconi syndrome. *Mod Rheumatol* 2005;15:207-212.
- Morisaki I, Abe K, Sobue S. Orofacial manifestations in a child with Fanconi's syndrome. *Oral Surg Oral Med Oral Pathol* 1989;68:171-174.
- Armando N. Proximal tubule endocytic apparatus as the specific renal uptake mechanism for vitamin D binding protein/25-(OH) D₃ complex. *Nephrology* 2006;11:510-515.
- Gahl WA. Cystinosis coming of age. *Adv Pediatr* 1986;33:95-126.
- Deshpande P, Ali U. Primary Fanconi syndrome. *Ind Pediatr* 1997;34:547-549.
- Brewer BD, Tsai HC, Norris RC. Evidence for impairment of metabolism of 25-hydroxyvitamin D₃ in children with Fanconi syndrome. *Clin Res* 1976;24:154A.
- Scheinman SJ. X-linked hypercalciuric nephrolithiasis: clinical syndromes and chloride channel mutation. *Kidney Int* 1998;53:2-17.
- Kaunisto K, Parkkila S, Rajaniemi H et al. Carbonic anhydrase XIV: Luminal expression suggests key role in renal acidification. *Kidney Int* 2002;61:2111-2118.
- Levinson DJ, Sorensen LB. Renal handling of uric acid in normal and gouty subject: Evidence for a 4-component system. *Ann Rheum Dis* 1980;39:173-179.
- Meisel AD, Diamond HS. Hyperuricosuria in the Fanconi syndrome. *Am J Med Sci* 1977;273:109-115.

- Guisan B, Diezi J. Effects of uricosuric and antiuric on urate transport in human brush-border membrane. *J Pharmacol Exp Ther* 1997;280:839–845.
- Mura H, Chairoungdua A et al. Molecular identification of a urate anion exchanger that regulates blood urate concentration. *Nature* 2002;417:447–452.
- T, Igarashi T et al. Exercise-induced acute renal failure and renal hypouricemia: Results of a questionnaire-based study. *Nephrol Dial Transplant* 2004;19:1447–1453.
- L, Hayward C et al. SLC2A9 is a newly identified urate transporter influencing serum urate concentration, urate excretion and renal function. *Nat Genet* 2008;40:437–442.
- wards the physiological function of uric acid. *Free Rad Biol Med* 2005;48:615–631.
- Sharratt P, Cutillas PR et al. Quantitative amino acid analysis: Very low excretion of polypeptides >750 Da in Fanconi syndrome. *Kidney Int* 2004;66:1994–2003.
- handling of proteins and polypeptides. In *Handbook of Renal Physiology*, Windhager EE (ed.). New York, Raven Press, 1992, pp. 2039–2082.
- C, Jacobsen C et al. Cubilin is an albumin binding protein important for renal tubular albumin reabsorption. *J Clin Invest* 2003;113:1353–1361.
- nsen BI. Renal albumin absorption in physiology and pathology. *Physiol Rev* 2006;86:440–449.
- man M. Hypercalcaemic rickets associated with renal tubular dysfunction. *Arch Dis Child* 1964;39:240–249.
- orden AG, Freest TG et al. Dent's disease; a familial syndrome with low-molecular weight proteinuria, hypercalcaemia, metabolic bone disease, progressive renal tubular dysfunction and male predominance. *QJM* 1994;87:473–493.
- rey HB, Kaplan BS et al. Dent disease presenting as Fanconi syndrome and hypercalcaemia. *Kidney Int* 2008;74:1033–1037.
- la T, Higuchi A et al. The low molecular weight of proteins in children urine. *Acta Paediatr Jpn* 1980;22:1–5.
- ikawa H, Shiraga H et al. Hypercalcaemia and nephrocalcinosis with idiopathic low-molecular-weight proteinuria: Is the disease identical to Dent's disease in United States? *Am J Kidney Dis* 1995;69:242–247.
- ce SHS, Gunter H et al. Idiopathic low molecular weight proteinuria associated with hypercalcaemia, nephrocalcinosis and renal tubular dysfunction in children is due to mutations of the renal chloride channel. *J Clin Invest* 1997;99:967–974.
- SE, Igarashi T et al. Mutations of CLCN5 in Japanese patients with idiopathic low molecular weight proteinuria, hypercalcaemia and nephrocalcinosis. *Kidney Int* 1997;52:911–916.
- omi J, Ohara T et al. Clinical and genetic studies of Dent's disease in Japanese families with Dent's disease. *Kidney Int* 2000;57:527–531.
- t M, Fuhmann JK et al. Physiological functions of the renal chloride channel are gleaned from human genetic disease and mouse models. *Physiol Rev* 2005;85:779–807.
- shi T, van der Smitten P et al. Altered polarity and function of Na⁺-ATPase without ultrastructural changes in kidneys of Fanconi syndrome patients. *Kidney Int* 2003;63:1285–1295.
- cheinman SJ, Dunham PB et al. X-linked recessive Fanconi syndrome with renal failure. *N Engl J Med* 1991;325:681–686.
- cheinman SJ, Deschodt-Lanckman MM et al. Tubular dysfunction defined by a study of Dent's (CLCN5 mutation) and Fanconi syndrome. *Kidney Int* 2000;57:240–249.
54. Scheinman SJ. X-linked hypercalcaemic nephrolithiasis: Clinical syndromes and chloride channel mutations. *Kidney Int* 1998;53:3–17.
55. Ludwig M, Utsch B, Balluch B et al. Hypercalcaemia in patients with CLCN5 mutations. *Pediatr Nephrol* 2006;21:1241–1250.
56. Gailly P, Jouret E, Martin D et al. A novel renal carbonic anhydrase type III plays a role in proximal tubule dysfunction. *Kidney Int* 2008;74:52–61.
57. Carr G, Simmons NL, Sayer JA et al. Disruption of clc-5 leads to redistribution of annexin A2 and promotes calcium crystal agglomeration in collecting duct epithelial cells. *Cell Mol Life Sci* 2006;63:367–377.
58. Norden AGW, Lapsley M, Lee PJ et al. Glomerular protein sieving and implications for renal failure in Fanconi syndrome. *Kidney Int* 2001;60:1885–1892.
59. Hoopes RR Jr, Raja KM, Koich A et al. Evidence for genetic heterogeneity in Dent's disease. *Kidney Int* 2004;65:1615–1620.
60. Raja KA, Schurman S, D'Mello M et al. Responsiveness of hypercalcaemia to thiazide in Dent's disease. *J Am Soc Nephrol* 2002;13:2938–2944.
61. Cebotaru V, Kaul S, Devuyst O et al. High citrate diet delays progression of renal insufficiency in the CLC-5 knockout mouse model of Dent's disease. *Kidney Int* 2005;68:642–652.
62. Guggino SE. Mechanism of disease: What can mouse models tell us about the molecular process underlying Dent disease? *Nat Clin Pract Nephrol* 2007;3:449–455.
63. Copelvitich L, Nash MA, Kaplan BS. Hypothesis: Dent disease is an underrecognized cause of focal glomerulosclerosis. *Clin J Am Soc Nephrol* 2007;2:914–918.
64. Lowe CU, Terrey M, MacLachlan EA. Organic aciduria, decreased renal ammonia production, hydrophthalmos and mental retardation: A clinical entity. *Am J Dis Child* 1952;83:164–184.
65. Lin T, Lewis RA, Nussbaum RL. Molecular confirmation of carriers of Lowe syndrome. *Ophthalmology* 1999;106:119–122.
66. Charnas LR, Bernardini I, Rader D et al. Clinical and laboratory findings in the oculocerebrorenal syndrome of Lowe, with special reference to growth and renal function. *N Engl J Med* 1991;324:1318–1325.
67. Laube G, Russel-Egitt I, van't Hoff W. Early proximal tubular dysfunction in Lowe's syndrome. *Arch Dis Child* 2004;89:479–480.
68. Altree O, Olivio JM, Okabe I et al. The Lowe's oculocerebrorenal syndrome gene encodes a protein highly homologous to inositol polyphosphate 5-phosphatase. *Nature* 1992;358:239–242.
69. Zhang X, Jefferson AB, Auethavekiat V et al. The protein deficient in Lowe syndrome is a phosphatidylinositol 4,5-bisphosphate 5-phosphatase. *Proc Natl Acad Sci USA* 1995;92:4853–4856.
70. Lin T, Orrison BM, Leahey AM et al. Spectrum of mutations in the OCRL1 gene in the Lowe oculocerebrorenal syndrome. *Am J Hum Genet* 1997;60:1384–1388.
71. Zhang X, Hartz PA, Philip E et al. Cell lines from kidney proximal tubules of a patient with Lowe syndrome lack OCRL1 inositol polyphosphate 5-phosphatase and accumulate phosphatidylinositol 4,5-bisphosphate. *J Biol Chem* 1998;273:1574–1582.
72. Suchy SF, Nussbaum RL. The deficiency of PIP₂ 5-phosphatase in Lowe syndrome affects actin polymerization. *Am J Hum Genet* 2002;71:1420–1427.
73. Ungewickell A, Ward M, Ungewickell B et al. The inositol polyphosphate 5-phosphatase Ocrl associates with endosome that are partially coated with clathrin. *Proc Natl Acad Sci USA* 2004;101:13501–13506.
74. Lowe M. Structure and function of Lowe syndrome protein. *Traffic* 2005;6:711–719.

75. Erdmann KS, Mao Y, McCrea HJ et al. A role of Lowe syndrome protein OCRL in early steps of the endocytotic pathway. *Dev Cell* 2007;13:377–390.
76. Fauchere A, Desbois P, Satre V et al. Lowe syndrome protein OCRL interacts with Rac GTPase in the trans-Golgi network. *Hum Mol Genet* 2003;12:2449–2456.
77. Hatefi Y. The mitochondrial electron transport and oxidative phosphorylation system. *Annu Rev Biochem* 1985;54:1015–1069.
78. Clayton DA. Structure and function of the mitochondrial genome. *J Inherib Metab Dis* 1992;15:439–447.
79. DiMauro S, Bonilla E, Lombes A et al. Mitochondrial encephalomyopathies. *Neurol Clin* 1990;8:483–506.
80. Niaudet P. Mitochondrial disorders and the kidney. *Arch Dis Child* 1998;78:387–390.
81. Ueda Y, Ando A, Nagata T et al. A boy with mitochondrial disease: Asymptomatic proteinuria without neuromyopathy. *Pediatr Nephrol* 2004;19:107–110.
82. Morris AA, Taylor RW, Birchi-Marchin MA et al. Neonatal Fanconi syndrome due to deficiency of complex III of the respiratory chain. *Pediatr Nephrol* 1995;9:407–411.
83. Kuwertz-Broking E, Koch HG, Marquardt T et al. Renal Fanconi syndrome: First sign of partial respiratory chain complex IV deficiency. *Pediatr Nephrol* 2000;14:495–498.
84. Au KM, Lau SC, Mak YF et al. Mitochondrial DNA deletion in a girl with Fanconi syndrome. *Pediatr Nephrol* 2007;22:136–140.
85. Tzen CY, Tsai JD, Wu TY et al. Tubulointerstitial nephritis associated with a novel mitochondrial point mutation. *Kidney Int* 2001;59:846–854.
86. Szabolcs MJ, Seigle R, Shanake S et al. Mitochondrial DNA deletion: A cause of chronic tubulointerstitial nephropathy. *Kidney Int* 1994;45:1388–1396.
87. Mochizuki H, Joh K, Kawame H et al. Mitochondrial encephalomyopathies preceded by de Toni-Debré-Fanconi syndrome or focal segmental glomerulosclerosis. *Clin Nephrol* 1996;46:347–352.
88. Gucer S, Talim B, Asan E et al. Focal segmental glomerulosclerosis associated with mitochondrial cytopathy: Report of two cases with special emphasis on podocytes. *Pediatr Dev Pathol* 2005;8:710–717.
89. Hotta O, Inoue CN, Miyabayashi S et al. Clinical and pathologic features of focal segmental glomerulosclerosis with mitochondrial tRNA^{Leu(UUR)} gene mutation. *Kidney Int* 2001;59:1236–1243.
90. Barisoni L, Diomedè-Camassei F, Santorelli FM et al. Collapsing glomerulopathy associated with inherited mitochondrial injury. *Kidney Int* 2008;74:237–243.
91. Lopez LC, Schuelke M, Quinzii CM et al. Leigh syndrome with nephropathy and CoQ10 deficiency due to decaprenyl diphosphate synthase subunit 2 (PDSS2) mutations. *Am J Hum Genet* 2006;79:1125–1129.
92. Niaudet P, Heidet L, Munnich A et al. Deletion of the mitochondrial DNA in a case of de Toni-Debré-Fanconi syndrome and Pearson syndrome. *Pediatr Nephrol* 1994;8:164–168.
93. Zaffanello M, Zamboni G. Therapeutic approach in a case of Pearson's syndrome. *Minerva Pediatr* 2005;57:143–146.
94. Matsutani H, Mizusawa Y, Shimoda M et al. Partial deficiency of cytochrome c oxidase with isolated proximal renal tubular acidosis and hypercalciuria. *Clin Nephrol Urol* 1992;12:221–224.
95. Goto Y, Itami N, Kajii N et al. Renal tubular involvement mimicking Bartter syndrome in a patient with Kearns-Sayre syndrome. *J Pediatr* 1990;116:904–910.
96. Moraes CT, Shanske S, Triachler HJ et al. Mitochondrial DNA depletion with variable tissue expression: A novel genetic abnormality in mitochondrial disease. *Am J Hum Genet* 1991;48:492–501.
97. Gilber RD, Erams M. Pearson's syndrome presenting with Fanconi syndrome. *Ultrastruct Pathol* 1996;20:473–475.
98. Gahl WA, Thoene JG, Schneidell JA. Cystinosis. *N Engl J Med* 2003;347:111–121.
99. van't Hoff WG, Ledermann SE, Waldron M et al. Early-onset chronic renal failure as a presentation of infantile nephropathy cystinosis. *Pediatr Nephrol* 1995;9:483–484.
100. Pennesi M, Marchetti E, Crovella S et al. A new mutation in two siblings with cystinosis presenting with Bartter syndrome. *Pediatr Nephrol* 2005;20:217–219.
101. Yildiz B, Durmus-Aydogdu S, Kural N et al. A patient with cystinosis presenting transient features of Bartter syndrome. *Turk J Pediatr* 2006;48:260–262.
102. Theodoropoulos DS, Shawker TH, Heinrichs C et al. Medullary nephrocalcinosis in nephropathic cystinosis. *Pediatr Nephrol* 1995;9:412–418.
103. Gubler MC, Lacoste M, Sich M et al. The pathology of the Kidney in Cystinosis. Paris, France, Elsevier, 1999.
104. Servais A, Moriniere V, Grünfeld JP et al. Late onset nephropathic cystinosis: Clinical presentation, outcome, and genotyping. *Clin J Am Soc Nephrol* 2008;3:27–35.
105. Town M, Jean G, Cherqui S et al. A novel gene encoding an integral membrane protein is mutated in nephropathic cystinosis. *Nature Genet* 1998;18:319–324.
106. Anikster Y, Lucero C, Guo J et al. Ocular nonnephropathic cystinosis: Clinical, biochemical, and molecular correlations. *Pediatr Res* 2000;47:17–23.
107. Cherqui S, Sevin C, Hamard G et al. Intralysosomal cystine accumulation in mice lacking cystinosis, the protein defective in cystinosis. *Mol Cell Biol* 2002;22:7622–7632.
108. Cervinkys I, Schlatter E, Hirsch JR et al. Inhibition of Na⁺-dependent transporters in cystine-loaded human renal cells: Electrophysiological studies on the Fanconi syndrome. *J Am Soc Nephrol* 2002;13:2085–2093.
109. Park MA, Thoene JG. Potential role of apoptosis in development of the cystinotic phenotype. *Pediatr Nephrol* 2005;20:441–446.
110. Wülmer MJ, de Graaf-Hess A, Blom HJ et al. Elevated oxidative glutathione in cystinotic proximal tubular epithelial cells. *Biochem Biophys Res Commun* 2005;337:610–614.
111. Bonsib SM, Horvth F Jr. Multinucleated podocytes in a child with nephrotic syndrome and Fanconi's syndrome: A unique clue to the diagnosis. *Am J Kidney Dis* 1999;34:966–971.
112. Spear GS, Slusser RJ, Tousimis AJ et al. Cystinosis. An ultrastructural and electron-probe study of the kidney with unusual findings. *Arch Pathol* 1971;91:206–221.
113. Kleit R, Gahl WA. Pharmacological treatment of nephropathic cystinosis with cysteamine. *Expert Opin Pharmacother* 2004;5:2255–2262.
114. Gahl WA, Kuehl EM, Iwata F et al. Corneal crystals in nephropathic cystinosis: natural history and treatment with cysteamine eye drops. *Mol Genet Metab* 2000;71:100–120.
115. Kleigman RM, Sparks JW. Perinatal galactose metabolism. *J Pediatr* 1985;107:831–841.
116. Tyfield L, Reichardt J, Fridovich-Keil J et al. Classical galactosemia and mutation at the galactose-1-uridylyl transferase (GALT) gene. *Human Mutat* 1999;13:417–430.
117. Waggoner DD, Buist NRM, Donnel GN et al. Long-term prognosis in galactosemia: Results in a survey of 350 cases. *J Inherib Metab Dis* 1990;13:802–818.
118. Waggoner DD, Buist NRM. Long-term complications in treated galactosemia-175 U.S. cases. *Int Pediatr* 1993;8:97–199.

- ross KC. Hidden sources of galactose in the environment. *Diabetes* 1995;154:S87-S92.
- nieri M, Gross KC et al. The effect of dietary fruits and urinary galactitol excretion in galactose-1-phosphate uridylyltransferase deficiency. *J Inher Metab Dis* 1993;16:91-100.
- ite PJ, Reynold RA. The rate of de novo galactose synthesis in patients with galactose-1-phosphate uridylyltransferase deficiency. *J Genet Metab* 2004;81:22-30.
- im I, Lin Z et al. Endogenous synthesis of galactose in patients with hereditary galactosemia. *Lancet* 1974;1:1074.
- Wells HJ, Segal S. Galactose metabolism in a patient with galactokinase deficiency. *Eur J Clin Invest* 1974;4:1007-1010.
- Additional findings in galactokinase deficiency. *J Clin Invest* 1974;53:1007-1008.
- ig M, Slepak VZ et al. Involvement of endoplasmic reticulum stress in a novel classic galactosemia model. *Mol Genet Metab* 2004;81:8-17.
- g LY, Choung AL et al. Inhibitor of apoptosis proteins and apoptosis in galactosemic rats. *Cell Tissue Res* 2004;315:417-425.
- Galactosemia in infancy: Diagnosis, management, and prognosis. *Am J Clin Nutr* 1997;23:563-469.
- Fructose intolerance. *J Med Genet* 1963;1:353-365.
- H, Tolan DR, Penhoet EB. Complete amino acid human aldolase B derived from cDNA and genomic DNA. *J Biol Chem* 1984;259:2738-2742.
- uki H, Joh K et al. Human aldolase b gene: Characterization of the genomic aldolase B gene and analysis of mutations. *J Biochem* 1991;110:1051-1055.
- itagliano L, Santamaria R et al. Structural and functional analysis of aldolase B mutants related to hereditary fructose intolerance. *Eur J Clin Invest* 2002;32:152-156.
- Fructose intolerance. *Int J Biochem* 1989;11:89-93.
- An experimental renal acidification defect in patients with fructose intolerance: I. Its resemblance to renal tubular acidosis. *J Clin Invest* 1967;47:1389-1398.
- An experimental renal acidification defect in patients with fructose intolerance: II. Its distinction from classic renal tubular acidosis and its resemblance to the renal acidification defect in the Fanconi syndrome of children with cystinosis. *J Clin Invest* 1968;47:1648-1663.
- Little JA, Patten RL et al. Pathogenesis of acidotic fructose intolerance. *Metabolism* 1979;28:1133-1138.
- grass GLAI, Oberholzer VG et al. Fructosemia. Observations on 10 cases. *Am J Med* 1968;45:826-838.
- ry LS, Zhang L et al. Interaction between aldolase B and H⁺-ATPase: Evidence for direct coupling of glycolytic ATP-hydrolyzing proton pump. *J Biol Chem* 2001;276:30413-30418.
- Gitzelmann R. The diagnosis of hereditary fructose intolerance. *Acta Paediatr Scand* 1981;36:297-316.
- r C, Böhme HJ et al. Fructose breath hydrogen test: A non-invasive diagnostic procedure? *Dig Dis* 2003;21:276-278.
- ern D, Mansfield BC et al. Type I glycogen storage disease: The glucose-6-phosphatase complex. *Curr Mol Biol* 1987;1:143-149.
141. von Gierke E. Hepato-nephro-megalia glycogenica (Glykogenespeicher-krankheit der Leber und Nieren). *Beitr Pathol Anat* 1929;82:497-513.
142. Kim SY, Vhen LY, Yiu WH et al. Neutrophilia and elevated serum cytokines are implicated in glycogen storage disease type Ia. *FEBS Lett* 2007;581:3833-3838.
143. Rocco Di, Calevo MG, Taro M et al. Hepatocellular adenoma and metabolic balance in patients with type Ia glycogen storage disease. *Mol Genet Metab* 2008;93:398-401.
144. Reddy SK, Kishnani PS, Sullivan JA et al. Resection of hepatocellular adenoma in patients with glycogen storage disease type Ia. *J Hepatol* 2007;47:658-663.
145. Reitsma-Bierens WCC. Renal complications in glycogen storage disease type I. *Eur J Pediatr* 1993;152:S60-S62.
146. Hers HG, van Hoof F, de Borsy T. Glycogen storage disease. In: *The Metabolic Basis of Inherited Disease*, 6th edn. Scriver CR, Beaudet AL, Sly WS et al. (eds.) New York, McGraw-Hill Inc, 1989, pp. 425-437.
147. Matsuo N, Tsuchiya M, Cho H et al. Proximal renal tubular acidosis in a child with type I glycogen storage disease. *Acta Paediatr Scand* 1986;75:332-335.
148. Chen YT, Scheinman JJ, Park HK et al. Amelioration of proximal renal tubular dysfunction in type I glycogen storage disease with dietary therapy. *N Engl J Med* 1990;323:590-593.
149. Chen YT, Coleman RA, Scheinman JJ et al. Renal disease in type I glycogen storage disease. *N Engl J Med* 1988;318:7-11.
150. Verani R, Bernstein J. Renal glomerular and tubular abnormalities in glycogen storage disease type I. *Arch Pathol Lab Med* 1988;112:271-274.
151. Baker L, Dahlem S, Goldfarb S et al. Hyperfiltration and renal disease in glycogen storage disease. *Kidney Int* 1989;35:1345-1350.
152. Weinstein DA, Somers MJ, Wolfsdorf JL. Decreased urinary citrate excretion in type Ia glycogen storage disease. *J Pediatr* 2001;138:378-382.
153. Rake JP, Visser G, Labrune P et al. Glycogen storage disease type I: Diagnosis, management, clinical course and outcome. Results of the European study on glycogen storage disease type I (ESGSD I). *Eur J Pediatr* 2002;161:S20-S34.
154. Yiu WH, Pan C-J, Ruel RA et al. Angiotensin mediates renal fibrosis in the nephropathy of glycogen storage disease type I. *Kidney Int* 2008;73:716-723.
155. Urushibara M, Kagami S, Ito M et al. Transforming growth factor-beta in renal disease with glycogen storage disease I. *Pediatr Nephrol* 2004;19:676-678.
156. Greene HL, Slonim AE, O'Neill JA Jr. et al. Continuous nocturnal intragastric feeding for management of type I glycogen storage disease. *N Engl J Med* 1976;294:423-425.
157. Wolfsdorf JL, Crigler JF Jr. Cornstarch regimens for nocturnal treatment of young adults with type I glycogen storage disease. *Am J Clin Nutr* 1997;65:1507-1511.
158. Chen YT, Cornblath M, Sidbury JB et al. Cornstarch therapy in type I glycogen storage disease. *N Engl J Med* 1984;310:171-175.
159. Jyer SG, Chen CL, Wang CC et al. Long-term results of living donor liver transplantation for glycogen storage disorders in children. *Liver Transpl* 2007;13:848-852.
160. Lee KW, Lee JH, Shin SW et al. Hepatocyte transplantation for glycogen storage type Ib. *Cell Transplant* 2007;16:629-637.
161. Fanconi G, Bickel H. Die chronische Aminoacidurie (Aminosäurendiabetes oder nephrotisch-glukosurischer Zwerchwuchs) bei der Glykogenose und der Cystinose. *Helv Paediatr Acta* 1949;4:359-396.
162. Manz F, Bickel H, Brodehl J et al. Fanconi-Bickel syndrome. *Pediatr Nephrol* 1987;1:509-519.