24. Harigai M, Koike R, Miyasaka N, Group PPuA-TNFTPS. Pneumocystis pneumonia associated with infliximab in Japan. N Engl J Med 2007;357:1874–76.
25. Yoshizawa H, Kudo H, Iwano K, Yamada A, Aikawa T, Miyamoto Y, et al. Causes of death in patients with rheumatoid arthritis: analysis of 117 cases for 13 years. Ryumachi 1990;30:255–63.
26. Toyoshima H, Kusaba T, Yamaguchi M. Cause of death in autopsied RA patients. Ryumachi 1993;33:209–14.
27. Suzuki A, Ohosone Y, Obana M, Mita S, Matsuoka Y, Irimajiri S. Cause of death in 81 autopsied patients with rheumatoid arthritis. J Rheumatol 1994;21:33–6.
28. Takeuchi T, Tatsuki Y, Nagami Y, Ishiguro N, Tanaka Y, Yamanaka H, et al. Postmarketing surveillance of the safety profile of infliximab in 5000 Japanese patients with rheumatoid arthritis. Ann Rheum Dis 2008;67:189–94.
29. Hakoda M, Oiwa H, Kasagi F, Masunari N, Yamada M, Suzuki G, et al. Mortality of rheumatoid arthritis in Japan: a longitudinal cohort study. Ann Rheum Dis 2005;64:1451–5.
30. Yamanaka H, Inoue E, Tanaka E, Nakajima A, Taniguchi A, Terai C, et al. Influence of methotrexate dose on its efficacy and safety in rheumatoid arthritis patients: evidence based on the variety of prescribing approaches among practicing Japanese rheumatologists in a single institute-based large observational cohort (IORRA). Mod Rheumatol 2007;17:98–105.
31. Nakajima A, Kamitsuji S, Saito A, Tanaka E, Nishimura K, Horikawa N, et al. Disability and patient's appraisal of general health contribute to depressed mood in rheumatoid arthritis in a large clinical study in Japan. Mod Rheumatol 2006;16:151–7.
32. Yamada T, Nakajima A, Inoue E, Tanaka E, Hara M, Tomatsu T, et al. Increased risk of tuberculosis in patients with rheumatoid arthritis in Japan. Ann Rheum Dis 2006;65:1661–3.
33. Yamanaka H, Inoue E, Singh G, Tanaka E, Nakajima A, Taniguchi A, et al. Improvement of disease activity of rheumatoid arthritis patients from 2000 to 2006 in a large observational cohort study IORRA in Japan. Mod Rheumatol 2007;17:283–9.
34. Tanaka E, Kamitsuji S, Inoue E, Yamada T, Nakajima A, Takeuchi E, et al. Nonsteroidal anti-inflammatory drug use dose not affect short-term endoscopic and histologic outcomes after *Helicobacter pylori* eradication in patients with rheumatoid arthritis. Mod Rheumatol 2007;17:228–34.
35. Furuya T, Kotake S, Inoue E, Nanke Y, Yago T, Kobashigawa T, et al. Risk factors associated with incident clinical vertebral and nonvertebral fractures in Japanese women with rheumatoid arthritis: a prospective 54-month observational study. J Rheumatol 2007;34:303–10.
36. Momohara S, Inoue E, Ikari K, Kawamura K, Tsukahara S, Mochizuki T, et al. Risk factors for total knee arthroplasty in rheumatoid arthritis. Mod Rheumatol 2007;17:476–80.
37. Matsuda Y, Singh G, Yamanaka H, Tanaka E, Urano W, Taniguchi A, et al. Validation of a Japanese version of the Stanford health assessment questionnaire in 3,763 patients with rheumatoid arthritis. Arthritis Rheum 2003;49:784–8.
38. Tsuji F, Seki I, Aono H, Odani N, Mizutani K, Okamoto M, et al. Bucillamine mechanism inhibiting IL-1beta-induced VEGF production from fibroblast-like synoviocytes. Int Immunopharmacol 2007;7:1569–76.
39. Iwatani M, Inoue E, Nakamura T, Nakajima A, Hara M, Tomatsu T, et al. Efficacy profile of bucillamine in rheumatoid arthritis patients in a large observational cohort study, IORRA. Mod Rheumatol 2006;16:376–80.
40. Kauppi M, Sokka T, Hannonen P. Survey nonresponse is associated with increased mortality in patients with rheumatoid arthritis and in a community population. J Rheumatol 2005;32:807–10.
41. Sokka T. National databases and rheumatology research I: Longitudinal databases in Scandinavia. Rheum Dis Clin North Am 2004;30:851–67.
42. Barchielli A, Baizi D. Nine-year follow-up of a survey on smoking habits in Florence (Italy): higher mortality among non-responders. Int J Epidemiol 2002;31:1038–42.
43. Solomon DH, Goodson NJ, Katz JN, Weinblatt ME, Avorn J, Setoguchi S, et al. Patterns of cardiovascular risk in rheumatoid arthritis. Ann Rheum Dis 2006;65:1608–12.
44. Libby P. Inflammation and cardiovascular disease mechanisms. Am J Clin Nutr 2006;83:456S–60S.
45. Sidiropoulos PI, Siakka P, Pagonidis K, Raptopoulou A, Kritikos H, Tsetis D, et al. Sustained improvement of vascular endothelial function during anti-TNFalpha treatment in rheumatoid arthritis patients. Scand J Rheumatol 2009;38:6–10.
46. Maradit-Kremers H, Crowson CS, Nicola PJ, Ballman KV, Roger VL, Jacobsen SJ, et al. Increased unrecognized coronary heart disease and sudden deaths in rheumatoid arthritis: a population-based cohort study. Arthritis Rheum 2005;52:402–11.
47. Moriguchi M, Terai C, Kaneko H, Koseki Y, Kajiyama H, Uesato M, et al. A novel single-nucleotide polymorphism at the 5′-flanking region of SAA1 associated with risk of type AA amyloidosis secondary to rheumatoid arthritis. Arthritis Rheum 2001;44:1266–72.
48. Calguneri M, Ureten K, Akif Ozturk M, Onat AM, Ertenli I, Kiraz S, et al. Extra-articular manifestations of rheumatoid arthritis: results of a university hospital of 526 patients in Turkey. Clin Exp Rheumatol 2006;24:305–8.

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The Association of a Nonsynonymous Single-Nucleotide Polymorphism in *TNFAIP3* With Systemic Lupus Erythematosus and Rheumatoid Arthritis in the Japanese Population

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Objective. **Genome-wide association (GWA) studies in systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA) in Caucasian populations have independently identified risk variants in and near the tumor necrosis factor α (TNFα)–induced protein 3 gene (*TNFAIP3*), which is crucial for the regulation of TNF-mediated signaling and Toll-like receptor signaling. The aim of this study was to assess the role of *TNFAIP3* in the development of SLE and RA in Japanese subjects.**

Methods. **We selected 2 single-nucleotide polymorphisms (SNPs) from previous GWA studies. Rs2230926 is a nonsynonymous SNP in *TNFAIP3* and is associated with SLE, while rs10499194 is an intergenic SNP associated with RA. We then performed 2 independent sets of SLE case–control comparisons (717 patients and 1,362 control subjects) and 3 sets of RA case–control comparisons (3,446 patients and 2,344 control subjects) using Japanese subjects. We genotyped SNPs using TaqMan assays.**

Results. **We observed a significant association between rs2230926 and an increased risk of SLE and RA in the Japanese population (for SLE, odds ratio [OR] 1.92, 95% confidence interval [95% CI] 1.53–2.41, $P = 1.9 \times 10^{-8}$; for RA, OR 1.35, 95% CI 1.18–1.56, $P = 2.6 \times 10^{-5}$). The intergenic SNP rs10499194 was also associated with SLE and RA, while the risk allele for RA in Caucasians was protective against the diseases in our population.**

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Conclusion. **We demonstrated a significant association between the nonsynonymous variant in *TNFAIP3* and the risk for SLE and RA in the Japanese population. *TNFAIP3*, similar to *STAT4* and *IRF5*, may be a common genetic risk factor for SLE and RA that is shared between the Caucasian and Japanese populations.**

Systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA) represent multigenic diseases and are considered to be caused by interactions between susceptibility genes and environmental factors that result in an abnormal immune response. In fact, familial and linkage studies have provided strong evidence for the role of multiple genetic factors in the development of SLE and RA (1). In addition, association-based approaches in candidate loci using single-nucleotide polymorphisms (SNPs) have also identified several genes that contribute to these diseases. More recently, genome-wide association (GWA) studies in SLE and RA have revealed many susceptibility genes and pathways that contribute to disease development (2).

Familial and linkage studies have also shown familial aggregation of RA, SLE, and other immune-mediated diseases (1). In fact, several gene polymorphisms, including *PTPN22*, *STAT4*, and *IRF5* variants, have been shown to predispose to SLE and RA. Recent GWA studies in Caucasian populations have also identified the tumor necrosis factor α (TNFα)–induced protein 3 gene (*TNFAIP3*) as another common genetic risk factor for SLE and RA (3–6). *TNFAIP3*, also known as the A20 protein, is a negative regulator of the NF-κB signaling pathway that is essential in the pathogenesis of both SLE and RA (7). The association of *TNFAIP3* with diseases has been independently reported in SLE and RA, and it is of great interest that the peaks in association in the GWA studies are different between SLE and RA. In Caucasian populations, the significantly associated SNP markers for SLE, including the nonsynonymous SNP termed rs2230926, are located in the *TNFAIP3* region, while those for RA are located in the intergenic region between *TNFAIP3* and the oligodendrocyte transcription factor 3 gene (*OLIG3*). In addition to the difference in the diseases themselves, the association between *TNFAIP3* polymorphisms and these diseases in the Asian populations remains unclear (8).

In order to elucidate a genetic role for *TNFAIP3* in the development of SLE and RA in the Japanese population, we investigated 2 independent case–control cohorts of patients with SLE and 3 independent cohorts of patients with RA.

PATIENTS AND METHODS

Subjects. The subjects in the SLE study group comprised 2 cohorts of Japanese patients with SLE and unrelated control subjects. An SLE case–control cohort from the RIKEN (SLE cohort 1) consisted of 376 patients (mean age 43.2 years, 90.3% women) and 934 unrelated control subjects (mean age 52.6 years, 25.0% women). An SLE case–control cohort at Hokkaido University (SLE cohort 2) consisted of 341 patients (mean age 46.2 years, 88.3% women) and 428 unrelated control subjects (mean age 47.7 years, 28.7% female). All patients with SLE fulfilled the 1997 American College of Rheumatology (ACR) revised criteria for SLE (9).

The subjects in the RA component of the study comprised 3 cohorts of Japanese patients with RA and unrelated control subjects. The first cohort of patients with RA from BioBank Japan (RA cohort 1) consisted of 1,112 patients (mean age 60.5 years, 89.7% female, 69.7% positive for rheumatoid factor [RF]), and 934 unrelated control subjects. The second cohort from RIKEN (RA cohort 2) consisted of 830 patients (mean age 64.3 years, 83.7% women, 75.0% RF positive), and 658 unrelated control subjects (mean age 48.6 years, 57.4% women). The 934 unrelated control subjects in the first cohort of RA patients were the same as those used in SLE cohort 1. An RA case–control cohort from the Institute of Rheumatology Rheumatoid Arthritis (IORRA) cohort (RA cohort 3), which is a prospective observational cohort of patients with RA studied at Tokyo Women's Medical University, comprised 1,504 patients (mean age 59.3 years, 84% women, 88% RF positive), and 752 control subjects (mean age 38.4 years, 50% women). All patients with RA met the 1987 ACR (formerly, the American Rheumatism Association) revised criteria for a diagnosis of RA (10).

All subjects entered into this study were self-identified as Japanese and were recruited through several medical institutions located in Japan. DNA samples from the patients in the first cohort of RA patients in BioBank Japan were provided by the Leading Project for Personalized Medicine from the Ministry of Education, Culture, Sports, Science and Technology, Japan (11). All subjects provided informed consent prior to their participation in this study, and the study was preapproved by the ethics committee of each institution.

SNPs. For the selection of SNPs required to genotype in and near *TNFAIP3*, we reviewed previous GWA studies of SLE and RA (3–6). We then selected 2 SNPs, rs2230926 and rs10499194. SNP rs2230926 is a nonsynonymous variant in exon 3 of *TNFAIP3* and was strongly associated with SLE in the GWA study by Musone et al (5). Although the GWA study of SLE by Graham et al indicated that rs5029939, located in intron 2 of the gene, is most significantly associated with a predisposition to SLE (6), there is strong linkage disequilibrium (LD) (r^2 = 0.86) between these SNPs according to HapMap phase II data for Japanese and evidence that rs5029939 may be substituted by rs2230926 (Figure 1). Two previous GWA studies in RA revealed that rs10499194 and rs6920220, which are located between *TNFAIP3* and *OLIG3*, were significantly associated risk variants for RA (3,4). The HapMap data for Japanese individuals indicate that the minor allele frequency (MAF) of rs6920220 is 0.011, and that the MAF for control subjects in RA cohort 3 (IORRA) was <0.01. Results of a recent study in Korean populations also indicated

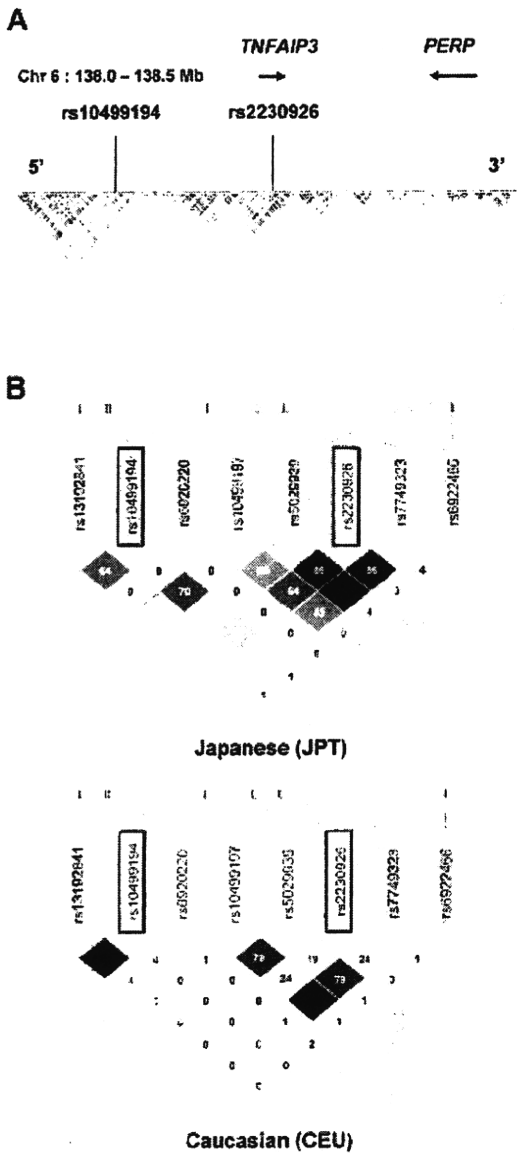

Figure 1. Pairwise linkage disequilibrium (LD) patterns for polymorphisms in the *TNFAIP3* region, according to HapMap phase II data. **A,** Pairwise LD pattern in the expanded *TNFAIP3* region derived from the HapMap data for Japanese patients, with r^2 values. *OLIG3* is located ~370 kbp away from *TNFAIP3* in the 5′ region and is not shown. **B,** Pairwise LD patterns for single-nucleotide polymorphisms (SNPs) in the *TNFAIP3* region that were significantly associated with systemic lupus erythematosus and rheumatoid arthritis in previous genome-wide association studies. The upper and lower panels were constructed using HapMap data for Japanese and Caucasian patients, respectively. The diagram shows pairwise LD values as quantified using the r^2 value. A stronger LD is depicted graphically by the densely shaded boxes. The boxed areas show the 2 SNPs genotyped.

that the variant was too rare (MAF <0.01) to be evaluated for associations (8).

Based on HapMap data for Japanese individuals, pairwise LD patterns for the SNPs in and near *TNFAIP3*, which were significantly associated with SLE and RA in the previous GWA studies, are presented in Figure 1 (for SLE, rs13192841, rs10499197, rs5029939, rs2230926, rs7749323, and rs6922466; for RA, rs10499194 and rs6920220).

Genotyping. We genotyped SNPs using TaqMan assays. For the selected SNPs, predesigned TaqMan SNP genotyping assays were used (probe ID: rs2230926, C.770116_10; rs10499194, C.1575581_10; Applied Biosystems, Foster City, CA). Fluorescence was detected using an ABI Prism 7900HT Sequence Detection System (Applied Biosystems). Genotyping assessment was performed on >98% of the samples, for all of the polymorphisms genotyped. All of the SNPs were in Hardy-Weinberg equilibrium in control subjects, according to chi-square statistics ($P > 0.01$).

Case–control association tests. We first performed allele frequency comparisons of rs2230926 and rs10499194 in SLE cohort 1 and RA cohort 1. Then, further case–control association studies were conducted using SLE cohort 2 and RA cohorts 2 and 3, to validate the associations in the first cohorts. In the replication studies, we genotyped the SNPs with a P value less than 0.05 in either SLE cohort 1 or RA cohort 1 (the P value was determined after correction for conditional logistic analysis, as described below).

Measurement of autoantibodies. Sera from 1,104 patients in RA cohort 1 were available for the measurement of anti–cyclic citrullinated peptide (anti-CCP) antibodies and RF. Anti-CCP antibodies were measured using the Mesacup CCP test (Medical and Biological Laboratories, Woburn, MA), and RF was measured by enzyme-linked immunosorbent assay.

Statistical analysis. The case–control association of each SNP was tested with the Cochran-Armitage trend test. The genotype and allele frequencies for patients and control subjects were used to calculate the odds ratios (ORs) and 95% confidence interval (95% CIs) using Woolf's method. For the combined analysis, we used the Mantel-Haenszel test. We performed conditional logistic regression analysis to evaluate the effect of each polymorphism conditional on the remaining polymorphisms, using Statistica software (StatSoft, Tulsa, OK). We calculated pairwise LD indices between pairs of SNPs (the r^2 value), using HaploView software, version 4.0 (http://www.broad.mit.edu/haploview/ haploview). We calculated the population attributable risk (PAR) using the following formula: $PAR = f(OR - 1)/(1 + f[OR - 1])$, where f is the allele frequency in the control subjects. PAR is defined as the reduction in incidence that would be achieved if the population had been entirely unexposed. We calculated the statistical power of association using the R software program (http:// www.r-project.org).

RESULTS

Our results revealed a significant association between rs2230926 and both SLE and RA when comparing allele frequency in the patients and control subjects in the first cohort (for SLE, OR 1.92, 95% CI

Table 1. Association study of rs2230926 and rs10499194 with SLE in Japanese subjects*

dbSNP number, major/minor allele	No. of patients	No. of controls	Minor allele frequency		OR (95% CI)	*P*
			Patients	Controls		
rs2230926, G/T						
SLE 1	376	934	0.113	0.062	1.92 (1.43–2.58)	1.2×10^{-5}
SLE 2	341	428	0.116	0.064	1.91 (1.33–2.73)	3.0×10^{-4}
Combined analysis†	717	1,362	0.114	0.063	1.92 (1.53–2.41)	1.9×10^{-8}
rs10499194, T/C						
SLE 1	376	933	0.084	0.061	1.42 (1.03–1.95)	0.030

* SLE = systemic lupus erythematosus; dbSNP = Database of Single-Nucleotide Polymorphisms; OR = odds ratio; 95% CI = 95% confidence interval.
† By the Mantel-Haenszel method.

1.43–2.58, $P = 1.2 \times 10^{-5}$; for RA, OR 1.52, 95% CI 1.20–1.92, $P = 5.6 \times 10^{-4}$) (Tables 1 and 2). We also observed an association between rs10499194 and SLE patients in cohort 1 (OR 1.42, 95% CI 1.03–1.95, $P = 0.030$) (Table 1). However, the T allele appeared to represent a susceptibility allele in the SLE and RA patients in cohort 1, whereas the C allele appeared to be a risk allele for RA in Caucasians (3). We speculated that this association could be secondary to the moderate LD between rs2230926 and rs10499194 ($r^2 = 0.14$) according to data on control subjects in SLE cohort 1, and we subsequently performed a conditional logistic regression analysis to evaluate the effects of each polymorphism conditional on the remaining polymorphisms. The results of this analysis indicated that rs10499194 did not retain the statistically significant association when conditionally evaluated on rs2230926 ($P = 0.73$), while rs2230926 retained the significant association when conditionally evaluated on rs10499194 ($P = 3.4 \times 10^{-4}$). We concluded that rs2230926 was primarily associated with SLE located at this locus, and therefore genotyped only rs2230926 for replication studies in SLE (3–6).

The results of a case–control association study in SLE cohort 2 confirmed the significant association between rs2230926 and the risk of SLE (OR 1.91, 95% CI 1.33–2.73, $P = 3.0 \times 10^{-4}$). A combined analysis also confirmed a significant association (OR 1.92, 95% CI 1.53–2.41, $P = 1.9 \times 10^{-8}$, PAR = 0.055). In RA cohort 2 a statistically significant association between rs2230926 and a predisposition for RA was also replicated; however, this was not replicated in RA cohort 3 (for cohort 2, OR 1.39, 95% CI 1.07–1.81, $P = 0.013$; for cohort 3, OR 1.19, 95% CI 0.94–1.50, $P = 0.15$) (Table 2). In RA cohort 3, the statistical power required to detect an association at rs2230926 was 0.54 at a significance level of $\alpha = 0.05$ when we presumed that the OR for RA was 1.4 (the combined OR for RA cohorts 1 and 2 was 1.46). It was possible that the statistical power for RA cohort 3 may have been insufficient. A combined analysis on these data suggested a significant association (OR 1.35, 95% CI 1.18–1.56, $P = 2.6 \times 10^{-5}$, PAR = 0.024).

We observed no significant association of rs10499194 in RA cohort 1, but the statistical power to detect the association in this study was insufficient (1 –

Table 2. Association study of rs2230926 and rs10499194 with RA in Japanese subjects*

dbSNP number, minor/major allele	No. of Patients	No. of controls	Minor allele frequency		OR (95% CI)	*P*
			Patients	Controls		
rs2230926, G/T						
RA cohort 1	1,112	934	0.091	0.062	1.52 (1.20–1.92)	5.6×10^{-4}
RA cohort 2	825	655	0.100	0.074	1.39 (1.07–1.81)	0.013
RA cohort 3	1,478	747	0.087	0.075	1.19 (0.94–1.50)	0.15
Combined analysis†	3,415	2,326	0.092	0.069	1.35 (1.18–1.56)	2.6×10^{-5}
rs10499194, T/C						0.26
RA cohort 1	1,112	933	0.069	0.061	1.15 (0.90–1.48)	0.26
RA cohort 2	827	650	0.072	0.048	1.52 (1.11–2.08)	0.0090
RA cohort 3	1,472	716	0.073	0.059	1.32 (1.02–1.73)	0.038
Combined analysis†	3,411	2,299	0.071	0.056	1.30 (1.11–1.53)	8.4×10^{-4}

* RA = rheumatoid arthritis; dsSNP = Database of Single-Nucleotide Polymorphisms; OR = odds ratio; 95% CI = 95% confidence interval.
† By the Mantel-Haenszel method.

$\beta = 0.31$) considering the previously reported OR of 0.75 and a significance level of $\alpha = 0.05$ (3). Therefore, we genotyped rs10499194 in RA cohorts 2 and 3 for confirmation. Unlike in RA cohort 1, a significant association of rs10499194 was observed in RA cohorts 2 and 3 (for cohort 2, OR 1.52, 95% CI 1.11–2.08, $P = 0.0090$; for cohort 3, OR 1.32, 95% CI 1.02–1.73, $P = 0.038$) (Table 2). However, the risk allele for Caucasian patients with RA was protective against RA in our population, just as was observed in SLE cohort 1. The combined analysis showed a significant association of rs10499194 with RA (OR 1.30, 95% CI 1.11–1.53, $P = 8.4 \times 10^{-4}$).

We stratified patients in RA cohorts 1 and 3 according to the presence of anti-CCP antibodies and RF and examined for the association between *TNFAIP3* polymorphisms (rs2230926 and rs10499194) and RA susceptibility (see Supplementary Table 1, available in the online version of this article at http://www3.interscience.wiley.com/journal/76509746/home). When the patients were stratified according to anti-CCP antibody status, the G allele of rs2230926 was found to confer increased risk for RA in anti-CCP antibody–positive patients relative to anti-CCP antibody–negative patients (for anti-CCP antibody–positive patients, OR 1.36, 95% CI 1.15–1.62, $P = 4.0 \times 10^{-4}$; for anti-CCP–negative patients, OR 1.16, 95% CI 0.83–1.61, $P = 0.39$ in the combined analysis). A similar trend was observed when patients were stratified according to RF status. A stratified analysis on rs10499194 also showed that the disease susceptibility allele in Japanese patients with RA (the T allele) conferred higher risk in autoantibody-positive patients than in autoantibody-negative patients.

DISCUSSION

In the current study, rs2230926, located in exon 3 of *TNFAIP3*, was shown to be significantly associated with a predisposition to both SLE and RA in 2 and 3 independent cohorts of subjects, respectively. Our results confirmed that *TNFAIP3* is one of the common genetic risk factors for both SLE and RA, similar to *STAT4* and *IRF5*, in the Japanese and Caucasian populations (2). In addition, recent studies in Caucasian patients with RA have demonstrated that the *TNFAIP3* variant conferred an increased risk of RA in anti-CCP antibody– and RF-positive patients compared with anti-CCP antibody– and RF-negative patients (12,13). Our analysis stratified according to the autoantibodies confirmed this observation in Japanese patients with RA.

TNFAIP3 encodes a cytoplasmic zinc finger protein that is also known as the A20 protein. The A20 protein is required for negative regulation of the NF-κB signaling pathway, which is mediated by innate immune receptors such as TNF receptors and Toll-like receptors, and it prevents overstimulation of the innate immune response (7,14). The disease-associated variant, rs2230926 (T/G), is a nonsynonymous variant that results in a phenylalanine-to-cysteine change at residue 127 of the A20 protein (5). The risk allele is known to be the G allele that encodes Cys. Musone et al have reported that Cys^{127} A20 protein was only modestly, but consistently, less effective at inhibiting TNF-induced NF-κB activity than the Phe^{127} protein (5). This result suggests that reduced negative regulatory activity of A20 protein may allow excessive immune activity, leading to enhanced autoreactivity.

GWA studies of SLE patients in Caucasian populations have suggested that several polymorphisms in the *TNFAIP3* region, including the nonsynonymous SNP rs2230926, are associated with a predisposition to the disease. The genetic significance of rs2230926 was evident in the Japanese patients with SLE or RA entered into our study, although its precise role in Caucasian patients with RA remains unclear. The intergenic SNP rs10499194 is one of the landmark polymorphisms identified in Caucasian patients with RA (3,15), although the significant association with RA could not be replicated in several Caucasian populations (3,12). Because rs10499194 is also associated with RA susceptibility and autoantibody status in our population, rs10499194 could be a landmark for disease causal variants in Japanese patients with RA. However, considering the inverted susceptibility allele of rs10499194 between Japanese patients (T allele) and Caucasian patients (C allele), this association of rs10499194 would appear to be secondary, as a result of LD between rs10499194 and the disease causal variants. This finding is further supported by the lack of independent association at rs10499194 in SLE when conditioned with the rs2230926 genotype, suggesting that the association observed in rs10499194 may be partially influenced by rs2230926.

Taking into account the biologic impact of rs2230926 demonstrated by Musone et al (5), rs2230926 seems likely to be an important candidate for a causal variant in *TNFAIP3* (5). However, additional polymorphisms that are located in the intergenic region of *OLIG3* and *TNFAIP3* as well as that of *TNFAIP3* and *PERP* may also independently exercise an effect on disease susceptibility, a hypothesis that was previously raised by Musone et al (5) and Graham et al (6). Further mapping of the *TNFAIP3* region in Asian and Caucasian

populations is required for the precise determination of the additional causal polymorphisms present in patients with RA or SLE.

In conclusion, we confirm that *TNFAIP3* is a genetic risk factor for the development of both SLE and RA in the Japanese population. Although the nonsynonymous SNP rs2230926 is a strong causal variant candidate in this region, a search for additional causal variants in *TNFAIP3* is required.

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AUTHOR CONTRIBUTIONS

All authors were involved in drafting the article or revising it critically for important intellectual content, and all authors approved the final version to be published. Dr. Kochi had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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REFERENCES

1. Alarcon-Segovia D, Alarcon-Riquelme ME, Cardiel MH, Caeiro F, Massardo L, Villa AR, et al. Familial aggregation of systemic lupus erythematosus, rheumatoid arthritis, and other autoimmune diseases in 1,177 lupus patients from the GLADEL cohort. Arthritis Rheum 2005;52:1138–47.
2. Gregersen PK, Olsson LM. Recent advances in the genetics of autoimmune disease. Annu Rev Immunol 2009;27:363–91.
3. Plenge RM, Cotsapas C, Davies L, Price AL, de Bakker PI, Maller J, et al. Two independent alleles at 6q23 associated with risk of rheumatoid arthritis. Nat Genet 2007;39:1477–82.
4. Thomson W, Barton A, Ke X, Eyre S, Hinks A, Bowes J, et al. Rheumatoid arthritis association at 6q23. Nat Genet 2007;39: 1431–3.
5. Musone SL, Taylor KE, Lu TT, Nititham J, Ferreira RC, Ortmann W, et al. Multiple polymorphisms in the TNFAIP3 region are independently associated with systemic lupus erythematosus. Nat Genet 2008;40:1062–4.
6. Graham RR, Cotsapas C, Davies L, Hackett R, Lessard CJ, Leon JM, et al. Genetic variants near TNFAIP3 on 6q23 are associated with systemic lupus erythematosus. Nat Genet 2008;40:1059–61.
7. Lee EG, Boone DL, Chai S, Libby SL, Chien M, Lodolce JP, et al. Failure to regulate TNF-induced NF-κB and cell death responses in A20-deficient mice. Science 2000;289:2350–4.
8. Lee HS, Korman BD, Le JM, Kastner DL, Remmers EF, Gregersen PK, et al. Genetic risk factors for rheumatoid arthritis differ in caucasian and Korean populations. Arthritis Rheum 2009;60: 364–71.
9. Hochberg MC, for the Diagnostic and Therapeutic Criteria Committee of the American College of Rheumatology. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus [letter]. Arthritis Rheum 1997;40:1725.
10. Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum 1988;31:315–24.
11. Nakamura Y. The BioBank Japan Project. Clin Adv Hematol Oncol 2007;5:696–7.
12. Perdigones N, Lamas JR, Vigo AG, de la Concha EG, Jover JA, Urcelay E, et al. 6q23 polymorphisms in rheumatoid arthritis Spanish patients. Rheumatology (Oxford) 2009;48:618–21.
13. Patsopoulos NA, Ioannidis JP. Susceptibility variants for rheumatoid arthritis in the TRAF1-C5 and 6q23 loci: a meta-analysis. Ann Rheum Dis 2009. E-pub ahead of print.
14. Liu YC, Penninger J, Karin M. Immunity by ubiquitylation: a reversible process of modification. Nat Rev Immunol 2005;5: 941–52.
15. Orozco G, Hinks A, Eyre S, Ke X, Gibbons LJ, Bowes J, et al. Combined effects of three independent SNPs greatly increase the risk estimate for RA at 6q23. Hum Mol Genet 2009;18:2693–9.