

番号	用途・機能・特徴	研究・開発メーカー	ベクター	プロモーター	ターミネーター	マーカー
762	RNAi	Promega	psiCHECK-1 (Accession No. AY535006)	・T7 ・SV40		Ampicillin
763	RNAi	Promega	psiCHECK-2 (Accession No. AY535007)	・T7 ・SV40 ・HSV-TK		Ampicillin
764	クローニング	Promega	pGEM-T	・T7 ・SP6		Ampicillin
765	クローニング	Promega	pGEM-T Easy	・T7 ・SP6		Ampicillin
766	大腸菌発現	Promega	PinPoint Xa-1 (Accession No. U47626)	・T7 ・SP6 ・Tac		Ampicillin
767	クローニング	STRATGENE	Lambda gt11	lac		Ampicillin
768	クローニング	STRATGENE	Lambda ZAPII	・T3 ・T7 ・lac		Ampicillin
769	クローニング	STRATGENE	pBluescript SK(+/-)	・T3 ・T7 ・lac		Ampicillin
770	・クローニング ・大腸菌発現 ・哺乳類細胞発現	STRATGENE	Lambda ZAP- CMV	・T3 ・T7 ・CMV		・Kanamycin ・Neomycin
771	クローニング	STRATGENE	pBC SK(+/-)	lac		Chloramphenicol
772	クローニング	STRATGENE	pBC KS(+/-)	lac		Chloramphenicol
773	クローニング	STRATGENE	pCMV-Script	CMV		・Kanamycin ・Neomycin
774	・クローニング ・大腸菌発現 ・哺乳類細胞発現	STRATGENE	Zap Express	・T3 ・T7 ・lac ・CMV		・Kanamycin ・Neomycin
775	Bacillus megaterium 発現	Mo Bi Tec	pWH1520	xylA		・Ampicillin ・Tetracycline
776	Bacillus megaterium 発現	Mo Bi Tec	pMM1522	xylA		・Ampicillin ・Tetracycline
777	Bacillus megaterium 発現	Mo Bi Tec	pMM1525	xylA		・Ampicillin ・Tetracycline
778	Bacillus megaterium 発現	Mo Bi Tec	pHIS1522	xylA		・Ampicillin ・Tetracycline
779	Bacillus megaterium 発現	Mo Bi Tec	pHIS1525	xylA		・Ampicillin ・Tetracycline
780	Bacillus megaterium 発現	Mo Bi Tec	pSTREP1525	xylA		・Ampicillin ・Tetracycline
781	Bacillus megaterium 発現	Mo Bi Tec	pSTREPHIS1525	xylA		・Ampicillin ・Tetracycline
782	Bacillus megaterium 発現	Mo Bi Tec	pC-HIS1622	xylA		・Ampicillin ・Tetracycline
783	Bacillus megaterium 発現	Mo Bi Tec	pC-STREP1622	xylA		・Ampicillin ・Tetracycline
784	Bacillus megaterium 発現	Mo Bi Tec	pN-HIS-TEV1622	xylA		・Ampicillin ・Tetracycline
785	Bacillus megaterium 発現	Mo Bi Tec	pN-STREP- TEV1622	xylA		・Ampicillin ・Tetracycline
786	Bacillus megaterium 発現	Mo Bi Tec	pN-STREP- Xa1622	xylA		・Ampicillin ・Tetracycline
787	Bacillus megaterium 発現	Mo Bi Tec	pSTOP1622	xylA		・Ampicillin ・Tetracycline
788	Bacillus megaterium 発現	Mo Bi Tec	pRBBm15	xylA		・Ampicillin ・Tetracycline
789	Bacillus megaterium 発現	Mo Bi Tec	pRBBm13	xylA		・Ampicillin ・Tetracycline
790	Bacillus megaterium 発現	Mo Bi Tec	pRBBm16	xylA		・Ampicillin ・Tetracycline
791	Bacillus subtilis発現	Mo Bi Tec	pHT01	groE		・Ampicillin ・Chloramphenicol
792	Bacillus subtilis発現	Mo Bi Tec	pHT43	groE		・Ampicillin ・Chloramphenicol
793	Bacillus subtilis発現	Mo Bi Tec	pHT08	groE		・Ampicillin ・Chloramphenicol
794	Bacillus subtilis発現	Mo Bi Tec	pHT09	groE		・Ampicillin ・Chloramphenicol
795	Bacillus subtilis発現	Mo Bi Tec	pHT10	groE		・Ampicillin ・Chloramphenicol
796	Lactococcus lactis発 現	Mo Bi Tec	pNZ8148	Nisin A		Chloramphenicol
797	Lactococcus lactis発 現	Mo Bi Tec	pNZ8150	Nisin A		Chloramphenicol

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798	Lactococcus lactis発現	Mo Bi Tec	pNZ8008	Nisin A		Chloramphenicol
799	Bacillus subtilis発現	Mo Bi Tec	pBacTag-FLAG	spac	Tryptophan operon	•Ampicillin •Erythromycin
800	Bacillus subtilis発現	Mo Bi Tec	pBacTag-cMyc	spac	Tryptophan operon	•Ampicillin •Erythromycin
801	Bacillus subtilis発現	Mo Bi Tec	pBacTag-HA	spac	Tryptophan operon	•Ampicillin •Erythromycin
802	Bacillus subtilis発現	Mo Bi Tec	pBacTag-GFP+	spac	Tryptophan operon	•Ampicillin •Erythromycin
803	Bacillus subtilis発現	Mo Bi Tec	pBacTag-CFP	spac	Tryptophan operon	•Ampicillin •Erythromycin
804	Bacillus subtilis発現	Mo Bi Tec	pBacTag-YFP	spac	Tryptophan operon	•Ampicillin •Erythromycin
805	Pichia pastoris発現	Mo Bi Tec	pPICHOLI(1-3)	•T7 •Alcohol dehydrogenase	Alcohol dehydrogenase Terminator	Zeocin
806	Pichia pastoris発現	Mo Bi Tec	pPICHOLI-C	•T7 •CUP-1	Alcohol dehydrogenase Terminator	Zeocin
807	Pichia pastoris発現	Mo Bi Tec	pPICHOLI-HA	•T7 •Alcohol dehydrogenase	Alcohol dehydrogenase Terminator	Zeocin
808	cell label	Mo Bi Tec	pCDNA-3-mito-EosFPwt	•T7 •CMV		•Ampicillin •Neomycin
809	cell label	Mo Bi Tec	pCDNA-3-Flag1-EosFPwt	•T7 •CMV		•Ampicillin •Neomycin
810	cell label	Mo Bi Tec	pCDNA-3-Flag1-td-EosFP	•T7 •CMV		•Ampicillin •Neomycin
811	Broad host range	Mo Bi Tec	pBBR122			•Kanamycin •Chloramphenicol
812	Broad host range	Mo Bi Tec	pBHR1			•Kanamycin •Chloramphenicol
813	大腸菌発現	Mo Bi Tec	pHKperi1	lac	bacteriophage fd	Chloramphenicol
814	大腸菌発現	Mo Bi Tec	pHKperi2	lac	bacteriophage fd	Chloramphenicol
815	大腸菌発現	Mo Bi Tec	pHKcyto1	lac	bacteriophage fd	Chloramphenicol
816	大腸菌発現	Mo Bi Tec	pHKcyto2	lac	bacteriophage fd	Chloramphenicol
817	大腸菌発現	Mo Bi Tec	pAX4(a-c)(+,-)	lac		Ampicillin
818	大腸菌発現	Mo Bi Tec	pAX5(+,-)	lac		Ampicillin
819	大腸菌発現	Mo Bi Tec	pJL3	E. coli lipoprotein		Chloramphenicol
820	大腸菌発現	Mo Bi Tec	pSW1			Tetracyclin
821	大腸菌発現	Mo Bi Tec	pPEPTIDE	T7	T7	Ampicillin
822	promoter cloning	Mo Bi Tec	pBBR RESO			•Kanamycin •Chloramphenicol
823	酵母発現	Mo Bi Tec	pORF-CLONE	CUP1		Ampicillin
824	クローニング	Mo Bi Tec	Exontrap	SV40		Ampicillin
825	クローニング	Mo Bi Tec	p3T	lacI		Ampicillin
826	クローニング	Mo Bi Tec	pCorrect Clone	lambda		Ampicillin
827	クローニング	Mo Bi Tec	pMEX(5-8)	Tac	rrnB	Ampicillin
828	クローニング	Mo Bi Tec	pEG-His1			Ampicillin
829	クローニング	Mo Bi Tec	pMCS5	T7		Ampicillin
830	Y2H	Mo Bi Tec	pBT3-N	CYC1	CYC1	Kanamycin
831	Y2H	Mo Bi Tec	pBT3-C	CYC1	CYC1	Kanamycin
832	Y2H	Mo Bi Tec	pBT3-SUC	CYC1	CYC1	Kanamycin
833	Y2H	Mo Bi Tec	pBT3-STE	CYC1	CYC1	Kanamycin
834	Y2H	Mo Bi Tec	pPR3-N	CYC1	CYC1	Ampicillin
835	Y2H	Mo Bi Tec	pPR3-C	ADH1	CYC1	Ampicillin
836	Y2H	Mo Bi Tec	pPR3-SUC	ADH1	CYC1	Ampicillin
837	Y2H	Mo Bi Tec	pPR3-STE	ADH1	CYC1	Ampicillin
838	Y2H	Mo Bi Tec	pCCw-Alg5	CYC1	CYC1	Kanamycin
839	Y2H	Mo Bi Tec	pAI-Alg5	ADH1	CYC1	Ampicillin
840	Y2H	Mo Bi Tec	pDL2-Alg5	ADH1	CYC1	Ampicillin
841	one-Hybrid	Mo Bi Tec	pGNG2		ADH1	Ampicillin
842	one-Hybrid	Mo Bi Tec	pJG4-5	GAL1	ADH	Ampicillin
843	Y2H	Mo Bi Tec	pGNG1	GAL1,10	ADH	Ampicillin
844	Y2H	Mo Bi Tec	pEG202	ADH1	ADH	Ampicillin

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1	アレルギー治療剤	IgEエポトープ構造を挿したペプチドCε4min, α1,6Galactoseをコードする遺伝子	<i>Lactobacillus johnsonii</i> NCC 2754	菌体内に蓄積または菌体表面に固定。β-glucuronidase融合タンパク質を含む。ラットの鼻腔または胃内投与、自己免疫性アレルギー疾患に対する免疫寛容の誘導	スイス・Institute of Immunology 他	Scheppeler et al. (2005) Intranasal immunisation using recombinant <i>Lactobacillus johnsonii</i> as a new strategy to prevent allergic disease. Vaccine. 23:1126-34.	形質転換	pM112			chloramphenicol resistance (cat)	
	自己免疫疾患治療剤	myelin basic protein (MBP) 遺伝子	<i>Lactobacillus casei</i> ATCC 393	菌体内に蓄積または菌体表面に固定。β-glucuronidase融合タンパク質を含む。ラットの鼻腔または胃内投与、自己免疫性アレルギー疾患に対する免疫寛容の誘導	オランダ・INHO	Maassen et al. (2003) Reduced experimental autoimmune encephalomyelitis after intranasal and oral administration of recombinant lactobacillus expressing myelin antigens. Vaccine. 21:4695-4693	形質転換	pLP402, pLP403	α-amylase, P _{amy} (<i>Lactobacillus amylovorus</i>)	cbh (<i>Lactobacillus plantarum</i> conjugated bile acid hydrolase)	erythromycin resistance	pLP400シリーズの文献 Pounels et al. (2001) Methode in Enzyrol. 336:369-389
	ワクチン	Human Papilloma virus (HPV) 16L-L1抗原遺伝子	<i>Lactobacillus casei</i> CECT 5275	菌体内発現。マウスの皮下へ投与し、HPV 16L-L1特異的IgGの産生	ブラジル・Instituto de Quimica, Universidade de Sao Paulo 他	Aires et al. (2006) Production of human papillomavirus type 16 L1 virus-like particles by recombinant <i>Lactobacillus casei</i> cells. Appl. Env. Microbiol. 72:3444-3450	形質転換	pIA1ac			erythromycin resistance	pIA1acの文献 Gosalbes et al. (2001) List 81:29-35
	ワクチン	重症急性呼吸器感染症 (SARS) ウイルス抗原遺伝子	<i>Lactobacillus casei</i> ELS-S8	Bacillus subtilis由来AgAアンカーペプチドによる菌体表面発現。マウスへ経口および経鼻投与、殺菌SARSウイルスに対する中和抗体の産生	韓国・Bioleaders corporation 他	Lee et al. (2005) Mucosal immunization with surface-displayed severe acute respiratory syndrome coronavirus spike protein on <i>Lactobacillus casei</i> induces neutralizing antibodies in mice. J. Appl. Microbiol. 98:1922-1930	形質転換	pIA1			erythromycin resistance	
	ワクチン	<i>Helicobacter pylori</i> urease B (UrbB) 遺伝子	<i>Lactobacillus plantarum</i> NCIWB 8826	菌体内発現。マウスの胃内へ投与。特異的抗体産生と部分的感染抑制効果	スイス・Centre Hospitalier Universitaire Vaudois 他	Corthesy et al. (2005) Oral immunization of mice with lactose acid bacteria producing <i>Helicobacter pylori</i> urease B subunit partially protects against challenge with <i>Helicobacter felis</i> . J. Infect. Dis. 192:1441-1446	形質転換	pMEC142	lactose dehydrogenase, P _{lac} (<i>Lactobacillus plantarum</i>)		erythromycin resistance	
6	ワクチン	Transmissible gastroenteritis coronavirus (TGEV) glycoprotein SのN-末端配列遺伝子	<i>Lactobacillus casei</i> Shirata	菌体外分泌。マウスへ経口投与。特異的抗体産生と感染抑制効果	シンガポール・National University of Singapore	Ho et al. (2005) Intragastric administration of <i>Lactobacillus casei</i> expressing transmissible gastroenteritis coronavirus spike glycoprotein induced specific antibody production. Vaccine 23:3444-3450	形質転換	pLP500	lactose dehydrogenase, P _{lac} (<i>Lactobacillus casei</i>)	cbh (<i>Lactobacillus plantarum</i> conjugated bile acid hydrolase)	erythromycin resistance	pLP500シリーズの文献 Pounels et al. (2001) Methode in Enzyrol. 336:369-389
	ワクチン	Erysipelothrix rhusiopathiae 由来抗原 (Spa) 遺伝子	<i>Lactococcus lactis</i> subsp. <i>lactis</i> IL1403	菌体外分泌 (一部菌体固定)。マウスへ鼻内、胃内、または腹腔内投与。特異的抗体産生および感染抑制効果	日本・帝産産大 他	Cheun et al. (2004) Protective immunity of Spa-antigen producing <i>Lactococcus lactis</i> against <i>Erysipelothrix rhusiopathiae</i> infection. J. Appl. Microbiol. 96:1347-1353	形質転換	pSECE1			erythromycin resistance	
	ワクチン	<i>Streptococcus pyogenes</i> M proteinの conserved C-repeat region (CRP) 遺伝子	<i>Lactococcus lactis</i> LM2301	マウスへ鼻内および皮下投与。特異的抗体産生と感染抑制効果	アメリカ・Oregon State University 他	Wannan et al. (2004) Mucosal vaccine made from live recombinant <i>Lactococcus lactis</i> protects mice against pharyngeal infection with <i>Streptococcus pyogenes</i> . Infect. Immun. 72:3444-3450	形質転換	pPI6aip			erythromycin resistance	
9	ワクチン	<i>Salmonella enterica</i> serovar Enteritidisの鞭毛抗原 (F1)C 遺伝子	<i>Lactobacillus casei</i> ATCC 393	菌体表面発現。マウスへの胃内投与。抗体非依存的な感染抑制効果	日本・国立医薬品食品衛生研究所 他	Kajikawa et al. (2007) Intragastric immunization with recombinant <i>Lactobacillus casei</i> expressing flagellar antigen confers antibody-independent protective immunity against <i>Salmonella enterica</i> serovar Enteritidis. J. Infect. Immun. 75:2344-2350	形質転換	pLP401	α-amylase, P _{amy} (<i>Lactobacillus amylovorus</i>)	cbh (<i>Lactobacillus plantarum</i> conjugated bile acid hydrolase)	erythromycin resistance	
10	ワクチン	Human immunodeficiency virus (HIV) 抗原遺伝子	<i>Lactococcus lactis</i> subsp. <i>lactis</i> IL1403	菌体表面発現。コレラトキシン使用。マウスへ経口投与。特異的抗体産生、細胞性免疫誘導、および感染抑制効果	日本・横浜国立大学 他	Xin et al. (2003) Immunogenicity and protective efficacy of orally administered recombinant <i>Lactococcus lactis</i> expressing surface-bound HIV Env. Vaccine 21:1423-1428	形質転換	pSGAN332	erythromycin resistance gene promoter		erythromycin resistance	
11	ワクチン	破傷風毒素 Fragment C (TFc) 遺伝子	<i>Lactobacillus plantarum</i> NCIWB 8826および <i>Lactococcus lactis</i> WGI363	菌体内発現。マウスへ胃内投与。破傷風毒素中和抗体の産生	フランス・パスツール研究所 他	Garagetle et al. (2002) Protection against tetanus toxin after intragastric administration of two recombinant lactic acid bacteria: impact of strain viability and in vivo persistence. Vaccine 20:1423-1428	形質転換	pMEC127	lactose dehydrogenase, P _{lac} (<i>Lactobacillus plantarum</i>)		erythromycin resistance	
12	ワクチン	破傷風毒素 ミニトープ遺伝子	<i>Lactobacillus johnsonii</i> NCC 2754	菌体表面発現。マウスへの投与。特異的抗体産生	スイス・Institute of Immunology and Allergology 他	Scheppeler et al. (2002) Recombinant <i>Lactobacillus johnsonii</i> as a mucosal vaccine delivery vehicle. Vaccine 20:1423-1428	形質転換	pM112			chloramphenicol resistance (cat)	
13	ワクチン	ウシ Rotavirus nonstructural protein 4 (NSP4) 抗原遺伝子	<i>Lactococcus lactis</i> NZ9000	菌体内発現または菌体外分泌。菌体ライゼント-アラムアジュバントをウサギの筋肉内へ投与。特異的抗体産生	フランス・INRA	Enouf et al. (2001) Bovine rotavirus nonstructural protein 4 produced by <i>Lactococcus lactis</i> is antigenic and immunogenic. Appl. Env. Microbiol. 67:1423-1428	形質転換	pSEC, pCY1	nisin-inducible promoter, P _{nis}		chloramphenicol resistance (cat)	
14	ワクチン	破傷風毒素 Fragment C (TFc) 遺伝子	<i>Lactobacillus plantarum</i> 256 および <i>Lactobacillus casei</i> ATCC 393	菌体内発現または菌体表面発現。マウスの胃内または鼻腔内投与。特異的抗体および細胞誘導	オランダ・INHO	Shaw et al. (2000) Engineering the microflora to vaccinate the mucosa: serum immunoglobulin G responses and activated draining cervical lymph nodes following mucosal application of tetanus toxin fragment C-expressing <i>Lactobacilli</i> . Vaccine 18:1423-1428	形質転換	pLP503, pLP401	lactose dehydrogenase, P _{lac} (<i>Lactobacillus plantarum</i>)	cbh (<i>Lactobacillus plantarum</i> conjugated bile acid hydrolase)	erythromycin resistance	
15	ワクチン	炭疽菌抗原遺伝子 pag	<i>Lactobacillus casei</i> ATCC 393	菌体内発現。マウスの胃内または鼻腔内投与。特異的抗体産生	オランダ・INHO 他	Zegers et al. (1999) Expression of the protective antigen of <i>Bacillus anthracis</i> by <i>Lactobacillus casei</i> : towards the development of an oral vaccine against anthrax. J. Appl. Microbiol. 87:309-316	形質転換	pLP503	lactose dehydrogenase, P _{lac} (<i>Lactobacillus casei</i>)	cbh (<i>Lactobacillus plantarum</i> conjugated bile acid hydrolase)	erythromycin resistance	
16	ワクチン	コレラ毒素Bサブユニット遺伝子	<i>Lactobacillus paracasei</i> lb1651.4, <i>Lactobacillus plantarum</i> NCIWB 8826	菌体内発現または菌体外分泌	フランス・Transgene S.A. 他	Sios et al. (1998) Production of cholera toxin B subunit in <i>Lactobacillus</i> . FEMS Microbiol. Lett. 169:29-36	形質転換	pIG292	P25	flf2 double vertical hairpins	chloramphenicol resistance (cat)	

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17	ワクチン	マウスインターロイキン-2 (IL-2), IL-6, TIFC遺伝子	<i>Lactococcus lactis</i> W61363	IL-2 (またはIL-6) 分泌およびTIFC遺伝子発現, マウスへ鼻腔内投与, TIFC特異的抗体産生	ベルギー・Ghent大学 他	Steidler et al. (1998) Mucosal delivery of murine interleukin-2 (IL-2) and IL-6 by recombinant strains of <i>Lactococcus lactis</i> coexpressing antigen and cytokine. <i>Infect. Immun.</i> 66:3183-3189	形質転換	pIREX1			erythromycin resistance	pIREX1の文献 Naito et al. <i>NATO ASI Ser. H</i> 98:37-63	
18	抗ウイルス (HIV) 剤	Cyanovirin遺伝子	<i>Lactococcus lactis</i> W61363 および <i>Lactobacillus plantarum</i> NC1MB 8826	分泌または細胞内発現, in vitroで抗ウイルス活性を確認	オーストリア・Vienna医科大学 他	Pusch et al. (2005) Bioengineering lactic acid bacteria to secrete the HIV-1 virucide cyanovirin. <i>J. Acquir. Immune Defic. Syndr.</i>	形質転換	pTSV1, pTSV2	LPS2 (bacteriophage)			erythromycin resistance	
19	抗ウイルス (HIV) 剤	Cyanovirin-H遺伝子	<i>Lactobacillus jensenii</i> 1153	分泌, <i>L. jensenii</i> のマウス腔内定着と cyanovirin発現を確認	アメリカ・Osel, Inc. 他	Liu et al. (2006) Engineered vaginal <i>Lactobacillus</i> strain for mucosal delivery of the human immunodeficiency virus inhibitor cyanovirin-H. <i>Anticancer Agents and Chemotherapy</i> 6:3256	形質転換	pOSEL175	P ₂₁ (<i>Lactococcus lactis</i>), P ₂₃ (self), P ₂₅₀ (self)			erythromycin resistance	
20	抗炎症剤	ヒトインターロイキン10 (IL-10) 遺伝子	<i>Lactococcus lactis</i> W61363	分泌, IL-10遺伝子をホストtthYA遺伝子と置換することで, 生物学的封じ込めに優れる	ベルギー・Ghent大学	Steidler et al. (2003) Biological containment of genetically modified <i>Lactococcus lactis</i> for intestinal delivery of human interleukin 10. <i>Nat. Biotech.</i> 21:785-789	相同組換え (double crossover)	pOR119, pVE6007 (遺伝子組換え中間体のみ保持)	P ₀₅₀₄ (self)			erythromycin resistance (組換え中間体菌株のみ保持)	pOR119, pVE6007の文献 Law et al. (1995) <i>J. Bacteriol.</i> 177:7011-7018
21	抗炎症剤	マウスインターロイキン10 (IL-10) 遺伝子	<i>Lactococcus lactis</i> W61364	分泌, マウスへの腔内投与, 炎症性腸疾患モデルにおいて症状改善	ベルギー・Ghent大学 他	Steidler et al. (2000) Treatment of murine colitis by <i>Lactococcus lactis</i> secreting interleukin-10. <i>Science</i> 289:1352-1355	形質転換	pIREX1				erythromycin resistance	組換え体構築の文献 Schotte et al. (2000) <i>Enz. Microb. Technol.</i> 27:761-765
22	HIV感染阻害剤	ヒトCD4細胞外ドメイン (2D CD4) 遺伝子	<i>Lactobacillus jensenii</i>	分泌, in vitroでHIV-1の感染阻害を確認	アメリカ・Osel, Inc. 他	Chang et al. (2003) Inhibition of HIV infectivity by a natural human isolate of <i>Lactobacillus jensenii</i> engineered to express functional two-domain CD4. <i>Proc. Natl. Acad. Sci. USA</i> 100:11672-11677	形質転換	pOSEL144	P ₂₁ (<i>Lactococcus lactis</i>)			erythromycin resistance	
23	<i>S. mutans</i> 定着阻害剤	<i>S. mutans</i> 定着因子阻害抗体のFv領域遺伝子	<i>Lactobacillus casei</i> ATCC 393	細胞表面発現, ラット口腔内で <i>S. mutans</i> の定着阻害	スウェーデン・Karolinska Institutet at Huddinge Hospital 他	Kruger et al. (2002) <i>In situ</i> delivery of passive immunity by lactobacilli producing single-chain antibodies. <i>Nat. Biotechnol.</i> 20:702-706	形質転換	pLF402	α -amylase, P ₀₅₀₄ (self)	<i>cbh</i> (<i>Lactobacillus plantarum</i> conjugated bile acid hydrolyase)		erythromycin resistance	
24	ワクチン	タイプ16ヒトpapillomaウイルス (HPV-16) E7タンパク質抗原遺伝子	<i>Lactococcus lactis</i> N29000	細胞表面発現, マウスへの鼻腔内投与, 抗原特異的Th1型サイトカイン産生	フランス・INRA 他	Bernandez-Huaman et al. (2004) An inducible surface presentation system improves cellular immunity against human papillomavirus type 16 E7 antigen in mice after nasal administration with recombinant <i>Lactococcus</i> . <i>J. Med.</i>	形質転換	pCWA	nisin-inducible promoter, P ₀₅₀₄			chloramphenicol resistance (cat)	
25	免疫アジュバント	インターロイキン12遺伝子	<i>Lactococcus lactis</i> N29000	分泌, マウスへの鼻腔内投与, 抗原特異的Th1型サイトカイン産生の増強	フランス・INRA 他	Bernandez-Huaman et al. (2003) Intranasal immunization with recombinant <i>Lactococcus lactis</i> secreting murine interleukin-12 enhances antigen-specific Th1 cytokine production. <i>Infect. Immun.</i> 71:1887-1896	形質転換	pSEC	nisin-inducible promoter, P ₀₅₀₄			chloramphenicol resistance (cat)	
26	non-GMOタンパク質発現菌固定システム		<i>Lactococcus lactis</i> N29001	アンカー融合組換えタンパク質の産生, gram-positive enhancer, matrix (GEM) 粒子に結合させる	オランダ・BIONADE Technology 他	Boers et al. (2005) Novel surface display system for proteins on non-genetically modified gram-positive bacteria. <i>Appl. Env. Microbiol.</i>	形質転換	pPA3	nisin-inducible promoter, P ₀₅₀₄			chloramphenicol resistance (cat)	
27		green fluorescent protein (GFP) 遺伝子	<i>Lactobacillus</i>		ブラジル・Universidade Federal de Minas Gerais 他	Wata et al. (2006) Genetic transformation of novel isolates of chicken <i>Lactobacillus</i> bearing probiotic features for expression of heterologous proteins: a tool to develop live oral	相同組換え (single crossover)	pLBS-GFP-Erf					
28		ヒトパピローマウイルス type-16 E7抗原	<i>Lactobacillus plantarum</i> NC1MB 8826, <i>Lactococcus lactis</i> N29000		フランス・INRA 他	Cortes-Perez et al. (2005) Cell-surface display of E7 antigen from human papillomavirus type-16 in <i>Lactococcus lactis</i> and in <i>Lactobacillus plantarum</i> using a new cell-wall anchor from <i>Lactobacilli</i> . <i>J. Drug Target.</i> 13:89-98	形質転換	pSEC, pCWA, pILLCWA	nicA			erythromycin resistance, or chloramphenicol resistance	
29		<i>Streptococcus pneumoniae</i> antigens, PsaA and PspA	<i>Lactobacillus casei</i> CECT 5275		ブラジル・Instituto Butantan 他	Oliveira et al. (2003) Expression of <i>Streptococcus pneumoniae</i> antigens, PsaA (pneumococcal surface antigen A) and PspA (pneumococcal surface protein A) by <i>Lactobacillus casei</i> . <i>FEMS Microbiol. Lett.</i> 227:25-31	形質転換	pIA	lac			erythromycin resistance	
30			<i>Lactobacillus reuteri</i> 105-23C		ドイツ・University of Hohenheim 他	Maier et al. (2003) Identification of <i>Lactobacillus reuteri</i> genes specifically induced in the mouse gastrointestinal tract. <i>Appl. Environ. Microbiol.</i> 69:2044-51	形質転換	pM100, pM200	tdh			chloramphenicol resistance	
31			<i>Lactococcus lactis</i> N29000		フィンランド・University of Helsinki 他	Avall-Jääskeläinen et al. (2003) Surface display of the receptor-binding region of the <i>Lactobacillus brevis</i> S-layer protein in <i>Lactococcus lactis</i> provides nonadhesive <i>laetococci</i> with the ability to adhere to intestinal epithelial	形質転換	pKH 5056	nicA			chloramphenicol resistance	
32		FliC	<i>Lactococcus lactis</i>		韓国・Kyung Hee University	Kim et al. (2003) Display of heterologous proteins on the surface of <i>Lactococcus lactis</i> using the H and W domain of FliB from <i>Lactobacillus delbrueckii</i> subsp. <i>bulgaricus</i> as an anchoring matrix. <i>J. Appl.</i>		pNLI				rythromycin resistance gene (ery)	

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11		classical swine fever virus (CSFV) T cell epitope E290 peptide	Lactobacillus casei		中国・Northeast Agricultural University	Xu et al. (2007) The oral immune efficacy of recombinant lactobacillus casei expressing CSFV E290 peptide and it elicited specific CTL response. Sheng Wu Gong Cheng Xue		pPG-VP2				
14		GFP	Leuconostoc mesenteroides		韓国・Kyung Hee University	Lee et al. (2007) Development of a monitoring vector for Leuconostoc mesenteroides using the green fluorescent protein gene. J Microbiol Biotechnol.		pCW5	p32			
15					インド・National Dairy Research Institute	Sudhanani et al. (2008) Characterisation of pSMA23, a 3.5kbp plasmid of Lactobacillus casei, and application for heterologous expression in Lactobacillus. Plasmid. 59:11-9		pSMA23, pL142, pL157				
16		GFP			中国・Nanjing Agricultural University	Yu et al. (2007) Use of green fluorescent protein to monitor Lactobacillus in the gastro-intestinal tract of chicken. FEMS Microbiol Lett. 275:207-		pLEW415				
17		GFP	Lactobacillus reuteri		台湾・National Chung-Hsing University	Mice protected by oral immunization with Lactobacillus reuteri secreting fusion protein of Escherichia coli enterotoxin subunit protein. FEMS Immunol Med Microbiol. 50:354-		pRIES				
18			Lactobacillus casei		日本・岡山大学	An et al. (2007) Expression of citrate permease gene of plasmid pCMI isolated from Lactococcus lactis subsp. lactis biovar. diacetylactis NIAI N-7 in Lactobacillus casei L-49-4. Appl Microbiol Biotechnol. 74:609-16.		pLECit			rythromycin resistance gene (ery)	
19		L. mesenteroides SY1 (another Kirchl isolate), Leuconostoc sp., and Lactobacillus brevis			韓国・Gyeongsang National University	Characterization of pLME1, a small cryptic plasmid isolated from Leuconostoc mesenteroides SY2. Plasmid. 57:314-23.		pS33E				
20		Foot-and-mouth disease virus (FMDV) VP1 protein			中国・China Agricultural University	Li et al. (2007) Immune responses generated by Lactobacillus as a carrier in DNA immunization against foot-and-mouth disease virus. Vaccine. 25:902-		pRc/CMV2				
21					アメリカ・University of Wisconsin-Madison	Sridhar et al. (2006) Construction and evaluation of food-grade vectors for Lactococcus lactis using aspartate aminotransferase and alpha-galactosidase as selectable markers. J Appl Microbiol. 101:161-71.		pSUN611, pSUN711			lactococcal aspartate aminotransferase gene (aspC), Bifidobacterium longum alpha-galactosidase	
22					台湾・National Chung-Hsing University	Wu et al. (2006) Construction and characterization of nisin-controlled expression vectors for use in Lactobacillus reuteri. Biocatal Biotechnol Biochem.		pNICE				
23					中国・Jilin Agricultural University	Wang et al. (2005) Expression of recombinant Vp6 gene of porcine rotavirus A with non-antibiotic Lactobacillus vector in Escherichia coli. Wei Sheng Wu Xue Bao.		pW425t				
24					ノルウェー・Norwegian Food Research Institute	Sarvig et al. (2005) High-level, inducible gene expression in Lactobacillus casei and Lactobacillus plantarum using versatile expression vectors. Microbiology. 151:2429-		pSIF				
25					韓国・Korea University	Kim et al. (2005) Optimization of technical conditions for the transformation of Lactobacillus acidophilus strains by electroporation. J Appl Microbiol. 99:167-74.		pR123, pKU				
26					ノルウェー・Agricultural University of Norway	Sarvig et al. (2005) Plasmid p256 from Lactobacillus plantarum represents a new type of replicon in lactic acid bacteria, and contains a toxin-antitoxin-like plasmid maintenance system. Microbiology. 151:421-31.		p256				
27		S-layer protein SgsE from Geobacillus stearothermophilus NRS 2004/3a	Lactococcus lactis		オーストリア・University of Natural Resources and Applied Life Sciences	Novotny et al. (2005) Gene cloning, functional expression and secretion of the S-layer protein SgsE from Geobacillus stearothermophilus NRS 2004/3a in Lactococcus lactis. FEMS Microbiol Lett. 242:27-35.		pR124				
28			Lactococcus lactis		マレーシア・Universiti Putra Malaysia	Raha et al. (2005) Cell surface display system for Lactococcus lactis: a novel development for oral vaccine. Appl Microbiol Biotechnol. -		pSVac				
29			Lactobacillus paracasei NFBC 338, Lactococcus lactis		アイルランド・Teagasc, Biotechnology Centre	Desmond et al. (2004) Improved stress tolerance of GroESL-overproducing Lactococcus lactis and probiotic Lactobacillus paracasei NFBC 338. Appl Environ Microbiol.		pR101, pRR02				

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50					中国・Fudan University	Yao et al. (2004) Inoculation of <i>Lactobacillus</i> expressing hCG beta in the vagina induces an anti-hCG beta antibody response in murine vaginal mucosa. <i>J Reprod Immunol</i> . 63:111-117		pIac				
51					フランス・Université de Bourgogne	Sury et al. (2004) Random transposon mutagenesis of <i>Lactobacillus plantarum</i> by using the pG9:IS S1 vector to clone genes involved in the regulation of phenolic acid metabolism. <i>Arch Microbiol</i> . 175:111-117		pG9				
52					アメリカ・Utah State University	Broadbent et al. (2004) Overexpression of <i>Lactobacillus casei</i> D-hydroxyisocaproic acid dehydrogenase in cheddar cheese. <i>Appl Environ Microbiol</i> . 70:4814-20		pIRKH2				
53					アメリカ・North Carolina State University	Bruno-Barona et al. (2004) Expression of a heterologous manganese superoxide dismutase gene in intestinal <i>Lactobacilli</i> provides protection against hydrogen peroxide toxicity. <i>Appl Environ Microbiol</i> . 70:111-117		pIRX563				
54					ロシア・All-Russia State Institute of the Physiology	Tarakanov et al. (2004) Expression vector pLF22 for the lactic acid bacteria. <i>Mikrobiologiya</i> . 73:211-217		pLF22				
55		folate gene cluster, consisting of six genes (folA, folB, folKE, folP, ylgG and folC)	<i>L. gasseri</i> strain ATCC 33323		オランダ・WCFS	Megkarp et al. (2004) Transformation of folate-consuming <i>Lactobacillus gasseri</i> into a folate producer. <i>Appl Environ Microbiol</i> . 70:3146-8	形質転換	pHZ7019	popN		chloramphenicol acetyltransferase (cat)	
56			<i>L. plantarum</i> WCFS1		オランダ・WCFS	Bron et al. (2004) Selection and characterization of conditionally active promoters in <i>Lactobacillus plantarum</i> using alanine racemase as a promoter probe. <i>Appl Environ Microbiol</i> . 70:111-117		pHZ7120, pHZ7121			rythromycin resistance gene (ery)	
57			<i>Lactobacillus sakei</i> , <i>Lactobacillus plantarum</i>		ノルウェー・Norwegian Food Research Institute	Servis et al. (2003) Construction of vectors for inducible gene expression in <i>Lactobacillus sakei</i> and <i>L. plantarum</i> . <i>FEBS Microbiol Lett</i> . 229:119-125						
58					ノルウェー・Norwegian Food Research Institute	Axeleson et al. (2003) Development of an inducible gene expression system for <i>Lactobacillus sakei</i> . <i>Letts Appl Microbiol</i> . 70:111-117		pRV3				
59					スペイン・Instituto de Agroquímica y Tecnología de los Alimentos	Pérez-Arellano et al. (2003) Optimization of the green fluorescent protein (GFP) expression from a lactose-inducible promoter in <i>Lactobacillus casei</i> . <i>FEBS Microbiol Lett</i> . 229:119-125						
60					ドイツ・Universität Kaiserslautern	Neu et al. (2003) New thermosensitive delivery vector and its use to enable nisim-controlled gene expression in <i>Lactobacillus gasseri</i> . <i>Appl Environ Microbiol</i> . 69:1377-82		pINI				
61					フィンランド・University of Helsinki	Takala et al. (2003) Food-grade host/vector expression system for <i>Lactobacillus casei</i> based on complementation of plasmid-associated phospho-beta-galactosidase gene <i>lacZ</i> . <i>Appl Microbiol</i> . 70:111-117		pLEB600				
62					ノルウェー・Agricultural University of Norway	Rawlinson et al. (2002) LasX, a transcriptional regulator of the lactocin S biosynthetic genes in <i>Lactobacillus sakei</i> L5, acts both as an activator and a repressor. <i>Biochimie</i> . 84:559-67. Review		pCIW1				
63					ドイツ・Universität Kaiserslautern	Henrich et al. (2002) Food-grade delivery system for controlled gene expression in <i>Lactococcus lactis</i> . <i>Appl Environ Microbiol</i> . 68:111-117		pLHG1363, pKS500				
64					フィンランド・University of Helsinki	Takala et al. (2002) A food-grade cloning vector for lactic acid bacteria based on the nisim immunity gene <i>nisI</i> . <i>Appl Microbiol Biotechnol</i> . 59:457-71		pLEB 590				
65					アメリカ・University of Illinois at Chicago	Pavlova et al. (2002) Characterization of a cryptic plasmid from <i>Lactobacillus Terreruntum</i> KC59 and its use for constructing a stable <i>Lactobacillus</i> cloning vector. <i>Plasmid</i> . 47:182-92		pSP1				
66					イギリス・Rural Development for Northern Ireland	Thompson et al. (2001) DNA cloning in <i>Lactobacillus helveticus</i> by the exconjugation of recombinant non-containing plasmid constructs from strains of transmissible lactic acid bacteria. <i>Plasmid</i> . 45:111-117						
67		<i>gusA</i> (β -glucuronidase)			オランダ・TNO	Pouwels et al. (2001) <i>Lactobacilli</i> as vehicles for targeting antigens to mucosal tissues by surface exposition of foreign antigens. <i>Methods Enzymol</i> . 260:111-117	形質転換	pLP401-pLP403, pLP501-pLP503, pLP601, pLP602, pLP701-pLP704, pLP803	amy, ldh, xylA, cbh		ery, cat	

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61					フィンランド・Food Research Institute, Agricultural Research Centre of Finland	Luoma et al. (2001) Expression of six peptidases from <i>Lactobacillus helveticus</i> in <i>Lactococcus lactis</i> . <i>Appl Environ Microbiol.</i> 67:1232-8.	形質転換	pKH2172, pKH2171, pKH2150, pKH2170	popN, popX, popC, popD, popR, nisA		ery, cat	
64		GFP			フランス・INRA	Gory et al. (2001) Use of green fluorescent protein to monitor <i>Lactobacillus sakei</i> in fermented meat products. <i>FEBS Microbiol Lett.</i>	相間組換え (double crossover)	pidH::gfp				
71					スペイン・Instituto de Agroquímica y Tecnología de Alimentos	Gosalbes et al. (2000) Integrative food-grade expression system based on the lactose regulon of <i>Lactobacillus casei</i> . <i>Appl Environ Microbiol.</i> 66:4922-8.	形質転換	pLlac				
74					スペイン・Universidad de Oviedo	Martin et al. (2000) Generation of food-grade recombinant lactic acid bacterium strains by site-specific recombination. <i>Appl Environ Microbiol.</i>	部位特異的組換え	pMW6, pEW68				(ery), 最終組換え体には残らない
75					ロシア・Institute of Genetics and Selection of Industrial Microorganisms	Shevelev et al. (2000) Expression of bacillar glutamyl endopeptidase genes in <i>Bacillus subtilis</i> by a new mobilizable single-replicon vector pLF.	形質転換	pLF9, pLF14				
77					韓国・Seoul National University	Cho et al. (2000) Expression of <i>Clostridium thermocellum</i> endoglucanase gene in <i>Lactobacillus gasseri</i> and <i>Lactobacillus johnsonii</i> and characterization of the genetically modified probiotic <i>Lactobacillus</i> .	形質転換	pSD1, pSD2				
78					アメリカ・North Carolina State University	Walker and Klaerner, (2000) An exolative antisense RNA strategy for inhibition of a lactococcal bacteriophage. <i>Appl Environ Microbiol.</i>		pIRK593				
79					アメリカ・Agricultural Research Service, Eastern Regional Research Center	Codre et al. (1999) Cloning and expression of the pediocin operon in <i>Streptococcus thermophilus</i> and other lactic fermentation bacteria. <i>Curr Microbiol.</i> 39:295-301.		pPC418				
76					アメリカ・North Carolina State University	Walker et al. (1999) The proSL chaperone operon of <i>Lactobacillus johnsonii</i> . <i>Appl Environ Microbiol.</i> 65:3033-41.		pIRQ12				
77					イギリス・Department of Agriculture for Northern Ireland	Thompson et al. (1999) Potential of conjugal transfer as a strategy for the introduction of recombinant genetic material into strains of <i>Lactobacillus helveticus</i> . <i>Appl Environ Microbiol.</i> 65:1910-4.		pSA3, pVA197			ery, cat	
78					イタリア・Istituto di Microbiologia	Callegari et al. (1998) The 5-layer gene of <i>Lactobacillus helveticus</i> CNRZ 882: cloning, sequence and heterologous expression. <i>Microbiology.</i> 144:719-726.		pMC01				
79					ベルギー・Université catholique de Louvain	Hois et al. (1997) Efficient secretion of the model antigen M6-p04E in <i>Lactobacillus plantarum</i> NCI 8826. <i>Microbiology.</i> 143:2733-39.		pG2281, pG3237				
80		endo-1,3- β -D-glucanase (bgIM)			ニュージーランド・University of Otago	Heng et al. (1997) Cloning and expression of an endo-1,3- β -D-glucanase gene from <i>Bacillus macerans</i> in <i>Lactobacillus reuteri</i> . <i>Appl Environ Microbiol.</i>	形質転換	pCKQH111	bgI		ery	
81					アメリカ・University of Nebraska	Gold et al. (1996) Cloning and expression of the <i>Zyomonas mobilis</i> "production of ethanol" genes in <i>Lactobacillus casei</i> . <i>Curr Microbiol.</i> 33:256-60.		pW6266, pRS602				
82					アメリカ・University of Wisconsin-Madison	Yoksel et al. (1996) DNA sequence analysis, expression, distribution, and physiological role of the Xaa-prolyl dipeptidyl aminopeptidase gene from <i>Lactobacillus helveticus</i> CNRZ92. <i>Appl Microbiol.</i>		pGK12, pIL253				
83					オランダ・TNO	Pouwels et al. (1996) The potential of <i>Lactobacillus</i> as a carrier for oral immunization: development and preliminary characterization of vector systems for						
84					ドイツ・Institut für Lebensmitteltechnologie	Obert et al. (1995) Two genes encoding the beta-galactosidase of <i>Lactobacillus sakei</i> . <i>Microbiology.</i> 141:3059-65.						
85					スロバキア・Comenius University	Trnava and Jurna. (1995) Transformation of microorganisms with the plasmid vector with the replicon from pACI from <i>Acetobacter pasteurianus</i> . <i>Biochem Biophys Res Commun.</i>		pACKS, pACT72				
86					ドイツ・Institut für Lebensmitteltechnologie	Herne et al. (1994) Expression of <i>Lactobacillus casei</i> ATCC 393 beta-galactosidase encoded by plasmid pLZ15 in <i>Lactococcus lactis</i> CNRZ 1123. <i>Lett Appl Microbiol.</i> 19:345-8.		pLZ15				

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87		chitinase gene from <i>Serratia marcescens</i>	<i>Lactococcus lactis</i> and <i>Lactobacillus plantarum</i>		ノルウェー・Agricultural University of Norway	Brurberg et al. (1994) Expression of a chitinase gene from <i>Serratia marcescens</i> in <i>Lactococcus lactis</i> and <i>Lactobacillus plantarum</i> . <i>Appl. Microbiol.</i>		pM36e, pGV259, pL1253	p32, p59			
88		beta-glucuronidase gene (<i>gusA</i>)			オランダ・Netherlands Institute for Dairy Research (NIZO)	Platteeuw et al. (1994) Use of the <i>Escherichia coli</i> beta-glucuronidase (<i>gusA</i>) gene as a reporter gene for analyzing promoters in lactic acid bacteria. <i>Appl. Environ. Microbiol.</i> 60:587-93. Erratum in: <i>Appl. Environ. Microbiol.</i>		pHZ772	<i>lacA</i> , <i>usp45</i> , a promoter from bacteriophage phi SK11G			
89		manganese superoxide dismutase	<i>Lactococcus lactis</i> and <i>Lactobacillus gasseri</i>		アメリカ・North Carolina State University	Roy et al. (1993) Cloning and expression of the manganese superoxide dismutase gene of <i>Escherichia coli</i> in <i>Lactococcus lactis</i> and <i>Lactobacillus gasseri</i> . <i>Mol. Gen. Genet.</i>		pM36e				
90		conjugated bile acid hydrolase	<i>Lactobacillus plantarum</i> 80		ベルギー・University of Ghent	Christians et al. (1992) Cloning and expression of a conjugated bile acid hydrolase gene from <i>Lactobacillus plantarum</i> by using a direct plate assay. <i>Appl. Environ. Microbiol.</i>		pCBH1				
91		<i>kataA</i>	<i>Lactobacillus casei</i>		ドイツ・Universität Hohenheim	Knauf et al. (1992) Cloning, sequence, and phenotypic expression of <i>kataA</i> , which encodes the catalase of <i>Lactobacillus sake</i> LTH677. <i>Appl. Environ. Microbiol.</i>		pGV210				
92		lysostaphin	<i>Lactobacillus casei</i> 102S		西ドイツ・Universität Hohenheim	Baier et al. (1992) Cloning and expression of the lysostaphin gene in <i>Bacillus subtilis</i> and <i>Lactobacillus casei</i> . <i>Lett. Appl. Microbiol.</i>						
93		beta-galactosidase	<i>L. helveticus</i> SBTZ195		日本・雪印乳業	Hashiba et al. (1992) Establishment of a host-vector system in <i>Lactobacillus helveticus</i> with beta-galactosidase activity as a selection marker. <i>Biosci. Biotechnol. Biochem.</i>		pB610				
94		beta-glucanase	<i>Lactobacillus helveticus</i> strain CNRZ450		イギリス・Department of Agriculture for Northern Ireland	Thompson and Collins (1991) Molecular cloning in <i>Lactobacillus helveticus</i> by plasmid pS2::pHA197 co-integrate formation and conjugal transfer. <i>Appl. Microbiol. Biotechnol.</i>		pSA3, pVA197				
95					アメリカ・North Carolina State University	Muriana and Klaenhammer (1991) Cloning, phenotypic expression, and DNA sequence of the gene for lactacin F, an antimicrobial peptide produced by <i>Lactobacillus</i> spp. <i>J. Bacteriol.</i> 173:1779-88.		pIRK160, pIRK162				
96					フィンランド・Genesis Oy	Sibakov et al. (1991) Secretion of TEM beta-lactamase with signal sequences isolated from the chromosome of <i>Lactococcus lactis</i> subsp. <i>lacticus</i> . <i>Appl. Environ. Microbiol.</i>		pVS2				
97		alpha-amylase			イギリス・Cranfield Institute of Technology	Jones and Garner (1990) Cloning and expression of alpha-amylase from <i>Bacillus amyloliquefaciens</i> in a stable plasmid vector in <i>Lactobacillus plantarum</i> . <i>Lett. Appl. Microbiol.</i>						
98		lipase	<i>Lactobacillus curvatus</i> Lc2-c		西ドイツ・Hohenheim University	Vogel et al. (1990) Expression of the lipase gene from <i>Staphylococcus hyicus</i> in <i>Lactobacillus curvatus</i> Lc2-c. <i>FEMS Microbiol. Lett.</i> 57:289-90.		pLipPS1				
99			<i>Lactobacillus plantarum</i> CCM 1904		フランス・Université Louis-Pasteur	Bringei et al. (1989) Characterization, cloning, curing, and distribution in lactic acid bacteria of pLP1, a plasmid from <i>Lactobacillus plantarum</i> CCM 1904 and its use in shuttle vector construction. <i>Plasmid.</i>		pLP1				
100			<i>Leuconostoc paramesenteroides</i>		オランダ・Netherlands Institute for Dairy Research (NIZO)	David et al. (1989) Plasmid transformation by electroporation of <i>Leuconostoc paramesenteroides</i> and its use in molecular cloning. <i>Appl. Environ. Microbiol.</i> 55:1483-9.		pNZ12				
101						Fons et al. (1997) Isolation and characterization of a plasmid from <i>Lactobacillus fermentum</i> conferring erythromycin resistance. <i>Plasmid.</i>		pLEW415, pLEW3				
102						Wiem et al. (1995) Molecular tools for the genetic modification of dairy lactobacilli. <i>Syst. Appl. Microbiol.</i> 18:493-503.		pK650, pMS58				
103						patent		pK, pEUL1				
104						De Vos et al. (1987) Gene cloning and expression in lactic streptococci. <i>FEMS Microbiol. Lett.</i> 45:1-10.		pHZ12, pSH71				
105						Maguin et al. (1992) New thermoresponsive plasmid for gram-positive bacteria. <i>J. Bacteriol.</i> 174:5633-5638.		pG ⁺ host4, pNV01				
106						Le Chatelier et al. (1993) Biochemical and genetic analysis of the unidirectional theta replication of the <i>S. agalactiae</i> plasmid pLPS01. <i>Plasmid</i> 29:50-56.		pGB305Δ, pLPS01				

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107						Tannock et al. (1994) Molecular characterization of a plasmid-borne (pG1633) erythromycin resistance determinant (ermE) from <i>Lactobacillus reuteri</i> , 100-63. <i>Plasmid</i> 31:60-71		pG1633				
108						Klein et al. (1993) Characterization and sequence analysis of a small cryptic plasmid from <i>Lactobacillus curvatus</i> L11683 and its use for construction of new <i>Lactobacillus</i> cloning vectors. <i>Plasmid</i>		pOU1892, pLC2				
109						Horinouchi et al. (1982) Nucleotide sequence and functional map of pE194, a plasmid that specifies inducible resistance to neomycin, tetracycline, and streptomycin type B antibiotics. <i>J. Bacteriol.</i> 150:804-814		pE194				
110						Horinouchi et al. (1982) Nucleotide sequence and functional map of pC194, a plasmid that specifies inducible chloramphenicol resistance. <i>J.</i>		pC194				
111						Zink et al. (1991) Transformation of <i>Lactobacillus delbrueckii</i> ssp. <i>Lactis</i> by electroporation and cloning of origins of replication by use of a positive selection vector. <i>FEMS Microbiol. Lett.</i> 78:207-212		p.W300, pWS97, pAZ20, pND0151				
117						Perez-Casal et al. (1993) An M protein with a single C repeat prevents phagocytosis of <i>Streptococcus pyogenes</i> : use of a temperature-sensitive shuttle vector to deliver homologous sequences to the chromosome of <i>S. pyogenes</i> . <i>Mol.</i>		p.RS233				
113						Kok et al., 1984;		pWV01				
114						Leenhouts et al., 1991		pSH71				
115						de Vos et al., 1987		pDI25				
116						Xu et al., 1990		pCL2.1				
117						Chang et al., 1995		pWC1				
118						Pillidge et al., 1996		pBM02				
119						Sanchez et al., 2003		pC305				
120						Hayes et al., 1990, 1991		pSL2				
121						Jahns et al., 1991		pVS40				nisin resistance
122						von Wright et al., 1990; von Wright and Raty, 1993		pSK111				
123						Hornig et al., 1991		pWV02				
124						Kiewiet et al., 1993		pCIS28				
125						Lucey et al., 1993		pUCL22				
126						Frere et al., 1993		pCT1138				
127						Pedersen et al., 1994		pWV04				
128						Seegers et al., 1994		pWV05				
129						Seegers et al., 1994		pIL7				
130						Gravsten et al., 1995		pJW563				cadmium resistance
131						Liu et al., 1996, 1997		pND302				
132						Liu et al., 1997		FG2 plasmid				
133						van Kranenburg and deVos, 1998		pNZ4000				
134						Duan et al., 1999		pND324				
135						Seegers et al., 2000		pCIS3				
136						Kearney et al., 2000		pC12000				
137						Jewel and Thompson-Collins, 1989		pcaT				
138						Vagic and Topisirovic, 1993		pA1				
139						Bouis et al., 1989		pLP1				
140						Leer et al., 1992		p8014-2				
141						Skaugen et al., 1989		pC301				
142						Bates and Gilbert, 1989		pLB4				
143						Darming et al., 2003		pLP2000				
144						Darming et al., 2003		pLP9000				
145						Eguchi et al., 2000		pLKL				
146						Eguchi et al., 2000		pLKS				
147						Danielsen, 2002		pMS5057				
148						Iwata et al., 1986		pLY2				
149						Iwata et al., 1986		pLY4				
150						Fons et al., 1997		pLEM3				
151						Aluisin et al., 1999		pLF1311				
152						Paulova et al., 2002		pKC56				
153						Posno et al., 1991		p353-1				
154						Leer et al., 1992		p353-2				
155						Axelsson et al., 1988		pLUL631				
156						Rinckel and Savage, 1990		pLAR33				
157						Tannock et al., 1994		pGT633				
158						Heng et al., 1999		pGT232				
159						Lin et al., 1999		pTE15				
160						Lin et al., 1999		pTE80				
161						Lin et al., 1996, 2001		pTC82				
162						Josson et al., 1989, 1990		pLAB1000				
163						Josson et al., 1989		pLAB2000				
164						Vogel et al., 1991; Klein et al., 1993		pLC2				
165						Zink et al., 1991		pWS97				
166						Chagnaud et al., 1992		pLB10				
167						Azcarate-Peril and Rays, 2002		pLBB1				
168						Boumiquel et al., 2002		pJBL2				
169						Boumiquel et al., 2002		pN42				
170						Damiani et al., 1987		p1				
171						Damiani et al., 1987		p3				
172						Luchansky et al., 1988		pPM4				
173						Kanatani et al., 1992, 1995		pLA103				
174						Kanatani et al., 1995		pLA105				
175						Sano et al., 1997		pLA106				
176						Takiguchi et al., 1989		pLJ1				

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177						Yamamoto and Takano, 1996		pCP53				
178						Fortina et al., 1993; Thompson et al., 1999		pLH1				
179						Fortina et al., 1993		pLH2				
180						Fortina et al., 1993		pLH3				
181						Pridmore et al., 1994		pLH4				
182						Unpublished; GenBank accession no Z50862		pSAK1				
183						Alpert et al., 2003		pRV500				
184						Chassy and Flickinger, 1987		pLZ15				
185						Whitehead et al., 2001		p121BS				
186			<i>B. subtilis</i>			Kok et al., 1984		pGK1			Chloramphenicol resistance	Shuttle No
187			<i>L. lactis, B. subtilis, E. coli, L. acidophilus, L. brevis, L. casei, L. delbrueckii, L. fermentum, L. helveticus, L. pentosus, L. plantarum, L. reuteri</i>			Kok et al., 1984; Posno et al., 1991; Zink et al., 1991; de los Reyes-Gavilan et al., 1990; Badii et al., 1989; Luchansky et al., 1989		pGK12			Chloramphenicol and Erythromycin resistance	Shuttle Yes
188			<i>L. acidophilus</i>			Luchansky et al., 1988		pGKV1			Chloramphenicol and Erythromycin resistance	Shuttle Yes
189			<i>B. subtilis</i>			van der Vossen et al., 1985		pGKV10			Chloramphenicol resistance	Shuttle Yes
190			<i>L. lactis, B. subtilis</i>			van der Vossen et al., 1985		pGKV110			Chloramphenicol and Erythromycin resistance	Shuttle Yes
191			<i>L. lactis, B. subtilis</i>			van der Vossen et al., 1985		pGKV11			Chloramphenicol and Erythromycin resistance	Shuttle Yes
192			<i>L. lactis, B. subtilis, E. coli, L. plantarum</i>			van der Vossen et al., 1985; Josson et al., 1989		pGKV2			Chloramphenicol and Erythromycin resistance	Shuttle Yes
193			<i>L. acidophilus, L. plantarum, L. casei</i>			van der Vossen et al., 1994; Pouwels and Leer, 1993; Gaier et al., 1992		pGKV21			Chloramphenicol and Erythromycin resistance	Shuttle Yes
194			<i>L. lactis, B. subtilis, L. casei, L. johnsonii, L. reuteri</i>			van der Vossen et al., 1987; Gaier et al., 1992; Fremaux et al., 1993; Djordjevic et al., 1994		pGKV210			Chloramphenicol and Erythromycin resistance	Shuttle Yes
195			<i>L. plantarum</i>			Badii et al., 1989		pGKV13			Chloramphenicol and Erythromycin resistance	Shuttle Yes
196			<i>L. lactis, B. subtilis</i>			Kok et al., 1985		pGKV500			Erythromycin resistance	Shuttle No
197			<i>L. lactis</i>			van Belkum et al., 1989		pMG24			Kanamycin resistance	Shuttle No
198			<i>L. lactis</i>			van de Guchte et al., 1989		pMG36			Kanamycin resistance	Shuttle No
199			<i>L. lactis, L. gasserii</i>			van de Guchte et al., 1989; Roy et al., 1993		pMG36a			Erythromycin resistance	Shuttle No
200			<i>L. acidophilus, L. gasserii, L. helveticus</i>			Raya and Knaenhammer, 1992		pTRK170			Chloramphenicol resistance	Shuttle Yes
201			<i>L. plantarum</i>			Hols et al., 1994		pGIP212			Chloramphenicol, Kanamycin, and Spectinomycin	Shuttle Yes
202			<i>L. casei</i>			Perez-Arellano et al., 2001		pLAV1			Erythromycin and Chloramphenicol resistance, lacZ	Shuttle Yes
203			<i>L. casei</i>			Perez-Arellano et al., 2001		pLAV6			Chloramphenicol resistance	Shuttle Yes
204			<i>L. casei</i>			Perez-Arellano et al., 2001		pLAV7			Erythromycin resistance	Shuttle Yes
205			<i>L. casei</i>			Perez-Arellano et al., 2001		pLAV9			Chloramphenicol resistance	Shuttle Yes
206			<i>B. subtilis, L. lactis, E. coli</i>			de Vos et al., 1986		pNZ11			Chloramphenicol and Kanamycin resistance	Shuttle No
207			<i>B. subtilis, L. lactis, E. coli, L. casei, L. curvatus, L. plantarum, L. sakei</i>			de Vos et al., 1986, 1987; Olukoya et al., 1993; Vogel et al., 1992; Gaier et al., 1990; Bringel and Hubert, 1990; Bringel and Hubert, 1990; Gaier et al., 1990		pNZ12			Chloramphenicol and Kanamycin resistance	Shuttle No
208			<i>B. subtilis, S. aureus, E. coli, L. lactis</i>			de Vos et al., 1987		pNZ121			Chloramphenicol and Kanamycin resistance	
209			<i>L. lactis</i>			de Vos et al., 1986		pNZ122			Chloramphenicol and Kanamycin resistance	Shuttle No
210			<i>L. lactis, L. acidophilus, L. casei, L. plantarum</i>			de Vos et al., 1986; Kim et al., 1994; Platteeuw et al., 1994		pNZ123			Chloramphenicol resistance	
211			<i>L. casei, L. plantarum</i>			Platteeuw et al., 1994		pNZ124			Chloramphenicol resistance	Shuttle Yes
212			<i>L. plantarum</i>			de Vos, 1987		pNZ17			Chloramphenicol resistance	Shuttle Yes

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71			<i>L. casei</i>			Solaiman et al., 1992		pNZ18			Chloramphenicol and Kanamycin resistance	Shuttle Yes
711			<i>L. casei</i>			Solaiman et al., 1992; Somkuti et al., 1992		pNZ19			Chloramphenicol and Kanamycin resistance	Shuttle Yes
712			<i>L. lactis, B. subtilis, E. coli</i>			deVos et al., 1989		pNZ220			Kanamycin resistance	Shuttle No
713			<i>L. casei, L. plantarum</i>			Platteeuw et al., 1994		pNZ272			Chloramphenicol resistance	Shuttle Yes
714			<i>B. subtilis, L. lactis, E. coli</i>			Gasson and Anderson, 1985		pCK1			Chloramphenicol and Kanamycin resistance	Shuttle Yes
715			<i>B. subtilis, L. lactis, E. coli</i>			Gasson and Anderson, 1985		pCK17			Chloramphenicol and Kanamycin resistance	Shuttle Yes
716			<i>B. subtilis, L. lactis, E. coli</i>			Gasson and Anderson, 1985		pCK21			Chloramphenicol and Kanamycin resistance	Shuttle Yes
717			<i>L. lactis</i>			Wells et al., 1993		pMIG1			Chloramphenicol and Kanamycin resistance	Shuttle Yes
718			<i>L. lactis</i>			Wells et al., 1993		pMIG2			Chloramphenicol resistance	Shuttle Yes
719			<i>L. lactis</i>			Wells et al., 1993		pMIG2H			Chloramphenicol resistance	Shuttle Yes
720			<i>L. lactis</i>			Wells et al., 1993		pMIG3			Chloramphenicol resistance	Shuttle Yes
721			<i>L. lactis, L. plantarum, L. routeri, L. sakei</i>			von Wright et al., 1987; Aukrust and Bloom, 1992; Ahrne et al., 1992; Axelsson et al., 1992		pVS2			Chloramphenicol and Erythromycin resistance	Shuttle No
722			<i>L. sakei</i>			Axelsson and Holck, 1995		pVSB1			Chloramphenicol resistance, lac	Shuttle Yes
723			<i>L. lactis</i>			von Wright and Raty, 1993		pRW1			Erythromycin, Tetracyclin, and Ampicillin resistance	Shuttle Yes
724			<i>L. casei</i>			Solaiman et al., 1992		pBN183			Ampicillin, Chloramphenicol, and Kanamycin resistance	Shuttle Yes
725			<i>L. casei</i>			Solaiman et al., 1992		pBN183A			Ampicillin, Chloramphenicol, and Kanamycin resistance	Shuttle Yes
726			<i>L. casei</i>			Solaiman et al., 1992		pDBN183			Ampicillin, Chloramphenicol, and Kanamycin resistance	Shuttle Yes
727			<i>L. plantarum</i>			Hols et al., 1994		pGIP331			Chloramphenicol, Kanamycin, and Spectinomycin	Shuttle Yes
728			<i>L. lactis</i>			Xu et al., 1990, 1991		pFX1			Chloramphenicol resistance	Shuttle Yes
729			<i>L. lactis</i>			Xu et al., 1990, 1991		pFX3			Chloramphenicol resistance	Shuttle Yes
730			<i>L. lactis, L. garviae, S. thermophilus, E. faecalis, S. aureus</i>			Pillidge et al., 1996		pCP12			Chloramphenicol resistance	Shuttle No
731			<i>L. lactis, L. casei, L. plantarum, B. subtilis</i>			Sanchez and Mayo, 2003		p21-22			Ampicillin and Erythromycin	Shuttle Yes
732			<i>L. lactis, L. casei, L. plantarum, B. subtilis</i>			Sanchez and Mayo, 2003		p22-25			Ampicillin and Erythromycin	Shuttle Yes
733			<i>L. lactis, L. casei, L. plantarum, B. subtilis</i>			Sanchez and Mayo, 2003		p22-26			Ampicillin and Erythromycin	Shuttle Yes
734			<i>L. lactis</i>			Hayes et al., 1990		pCI3340			Chloramphenicol resistance	Shuttle Yes
735			<i>L. lactis</i>			Hayes et al., 1990		pCI374			Chloramphenicol resistance	Shuttle Yes
736			<i>L. lactis</i>			von Wright et al., 1990		pVS40			nisin resistance	Shuttle No
737			<i>L. lactis</i>			Horng et al., 1991		pKMP1			Chloramphenicol and Erythromycin resistance	Shuttle No
738			<i>L. lactis</i>			Lucey et al., 1993		pCI534			Chloramphenicol resistance	Shuttle No
739			<i>L. lactis, P. acidilactici</i>			Kiewiet et al., 1993		pLR300			Erythromycin resistance	Shuttle No
740			<i>L. lactis</i>			Liu et al., 1996		pND302			cadmium resistance	Shuttle No
741			<i>L. lactis</i>			Liu et al., 1996		pND304			Tetracyclin and Ampicillin resistance	Shuttle Yes
742			<i>L. lactis</i>			Liu et al., 1996		pND624			Ampicillin, nisin, and cadmium	Shuttle No
743			<i>L. lactis</i>			Liu et al., 1996		pND625			nisin and cadmium resistance	Shuttle No
744			<i>L. lactis</i>			Duan et al., 1999		pND421			Ampicillin, Erythromycin, and nisin resistance	Shuttle No

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718			<i>C. divergens</i> , <i>C. piscicola</i> , <i>L. casei</i> , <i>L. plantarum</i>			Ahn et al., 1992		pcaT			Chloramphenicol resistance	
719			<i>E. coli</i> , <i>L. delbrueckii</i> , <i>L. plantarum</i>			Vujcic and Topisirovic, 1993		pA1			Chloramphenicol resistance	Shuttle No
720			<i>B. subtilis</i> , <i>L. plantarum</i>			Bringel et al., 1989		pULP8			Ampicillin and Erythromycin resistance	Shuttle Yes
721			<i>B. subtilis</i> , <i>L. plantarum</i>			Bringel et al., 1989		pULP9			Ampicillin and Erythromycin resistance	Shuttle Yes
727			<i>L. acidophilus</i> , <i>L. brevis</i> , <i>L. casei</i> , <i>L. fermentum</i> , <i>L. helveticus</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Posno et al., 1991; Leer et al., 1992; de los Reyes-Gavilan et al., 1990; Badii et al., 1989; Duckworth et al., 1993		pLP825			Ampicillin and Chloramphenicol resistance	Shuttle Yes
723			<i>L. casei</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Leer et al., 1992; Duckworth et al., 1993		pLP82H			Ampicillin and Chloramphenicol resistance	Shuttle Yes
724			<i>L. casei</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Leer et al., 1992		pLPC37			Chloramphenicol resistance	Shuttle No
725			<i>B. subtilis</i> , <i>L. acidophilus</i> , <i>L. fermentum</i> , <i>L. helveticus</i> , <i>L. plantarum</i> , <i>L. reuteri</i> , <i>L. lactis</i>			Cocconcelli et al., 1991		pPSC1			Chloramphenicol resistance	Shuttle Yes
726			<i>B. subtilis</i> , <i>L. acidophilus</i> , <i>L. fermentum</i> , <i>L. helveticus</i> , <i>L. reuteri</i> , <i>L. lactis</i>			Cocconcelli et al., 1991		pPSC10			Erythromycin resistance	Shuttle Yes
727			<i>B. subtilis</i> , <i>L. acidophilus</i> , <i>L. fermentum</i> , <i>L. helveticus</i> , <i>L. reuteri</i> , <i>L. lactis</i>			Cocconcelli et al., 1991		pPSC11			Chloramphenicol resistance	Shuttle Yes
728			<i>B. subtilis</i> , <i>L. acidophilus</i> , <i>L. fermentum</i> , <i>L. helveticus</i> , <i>L. reuteri</i> , <i>L. lactis</i>			Cocconcelli et al., 1991; Vescovo et al., 1991		pPSC20			Chloramphenicol and Erythromycin resistance	Shuttle Yes
729			<i>B. subtilis</i> , <i>L. acidophilus</i> , <i>L. fermentum</i> , <i>L. helveticus</i> , <i>L. reuteri</i> , <i>L. lactis</i>			Cocconcelli et al., 1991		pPSC22			Chloramphenicol and Erythromycin resistance	Shuttle Yes
720			<i>L. plantarum</i> , <i>L. sake</i>			Holck et al., 1992		pLPV106			Ampicillin and Erythromycin resistance	Shuttle Yes
721			<i>L. sake</i>			Arelsson and Holck, 1995		pLPVII			Erythromycin resistance, lac	Shuttle Yes
722			<i>L. fermentum</i>			Iwata et al., 1986		pLY2			Tetracycline resistance	
723			<i>L. fermentum</i>			Iwata et al., 1986		pLY4			Erythromycin resistance	
724			<i>L. fermentum</i>			Fons et al., 1997		pLEM5			Erythromycin resistance	Shuttle No
725			<i>L. fermentum</i>			Fons et al., 1997		pLEM7			Erythromycin resistance	Shuttle No
726			<i>L. brevis</i> , <i>L. buchneri</i> , <i>L. lactis</i> , <i>E. faecalis</i> , <i>E. faecium</i> , <i>B. subtilis</i> , <i>B. thuringiensis</i> subsp. <i>galleriae</i> , subsp. <i>kurstaki</i> , subsp. <i>finitimus</i> , <i>B. amyloquelaciens</i> , <i>B. flavum</i> , <i>E. coli</i>			Aleshin et al., 1999		pLFVM2			Chloramphenicol resistance	Shuttle Yes
727			<i>L. fermentum</i> , <i>L. jansoni</i> , <i>L. spp.</i> , <i>L. gassori</i> , <i>L. crispatus</i> , <i>L. johnsonii</i> , <i>L. salivarius</i> , <i>S. mutans</i> , <i>S. gordonii</i> , <i>S. sanguis</i>			Pavlova et al., 2002		pSP1			Erythromycin resistance	Shuttle Yes
728			<i>L. plantarum</i>			Pouwels and Leer, 1993		pLPE23M			Erythromycin resistance	Shuttle No
729			<i>L. plantarum</i>			Pouwels and Leer, 1993		pLPE24M			Erythromycin resistance	Shuttle No

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700			<i>L. casei</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Posno et al., 1991		pLPE317			Erythromycin resistance	Shuttle No
701			<i>L. casei</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Posno et al., 1991		pLPE323			Erythromycin resistance	Shuttle No
702			<i>L. casei</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Leer et al., 1992		pLPE350			Chloramphenicol resistance	Shuttle No
703			<i>L. acidophilus</i> , <i>L. casei</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Posno et al., 1991		pLP3537			Ampicillin and Erythromycin resistance	Shuttle Yes
704			<i>L. casei</i> , <i>L. pentosus</i> , <i>L. plantarum</i>			Posno et al., 1991		pLP3537- <i>xyI</i>			Ampicillin and Erythromycin resistance, <i>xyI</i>	Shuttle Yes
705			<i>B. subtilis</i> , <i>E. faecalis</i> , <i>L. delbrueckii</i> , <i>L. fermentum</i> , <i>L. reuteri</i> , <i>L. gasseri</i> , <i>L. salivarius</i> , <i>S. aureus</i> , <i>S. sanguis</i>			Tannock et al., 1994		pGT633			Erythromycin resistance	
706			<i>L. reuteri</i>			Ahn et al., 1992		pLUL631			Erythromycin resistance	
707			<i>L. reuteri</i>			Ahn et al., 1992		pLUL200			Chloramphenicol resistance	Shuttle Yes
708			<i>L. reuteri</i>			Ahn et al., 1992		pLUL201			Chloramphenicol and Erythromycin resistance	Shuttle Yes
709			<i>L. reuteri</i>			Ahn et al., 1992		pLUL202			Chloramphenicol resistance	Shuttle No
710			<i>L. reuteri</i>			Ahn et al., 1992		pLUL634			Erythromycin resistance	Shuttle No
711			<i>L. reuteri</i>			Heng et al., 1999		pNCKH104			Erythromycin resistance	Shuttle Yes
712			<i>L. reuteri</i>			Heng et al., 1999		pNCHK103			Erythromycin resistance	Shuttle Yes
713			<i>L. reuteri</i> , <i>L. fermentum</i>			Lin et al., 1999		pTE15-RO			Ampicillin and Erythromycin resistance	Shuttle Yes
714			<i>L. reuteri</i>			Lin et al., 1999		pTE80-RO			Ampicillin and Erythromycin resistance	Shuttle Yes
715			<i>L. reuteri</i>			Lin et al., 2001		pTC82-RO			Erythromycin resistance	Shuttle No
716			<i>L. plantarum</i>			Scheirfinck et al., 1989		pERM3.2			Ampicillin and Erythromycin resistance	Shuttle Yes
717			<i>B. subtilis</i> , <i>E. faecalis</i> , <i>L. casei</i> , <i>L. plantarum</i>			Josson et al., 1989		pLAB1102			Ampicillin and Chloramphenicol resistance	Shuttle Yes
718			<i>B. subtilis</i> , <i>E. faecalis</i> , <i>L. casei</i> , <i>L. plantarum</i>			Josson et al., 1989		pLAB1301			Ampicillin and Erythromycin resistance	Shuttle Yes
719			<i>B. subtilis</i> , <i>E. faecalis</i> , <i>L. casei</i> , <i>L. plantarum</i>			Josson et al., 1990		pLAB1304			Ampicillin and Erythromycin resistance	Shuttle Yes
720			<i>B. subtilis</i> , <i>E. faecalis</i> , <i>L. casei</i> , <i>L. plantarum</i>			Josson et al., 1990		pLAB1321			Ampicillin and Erythromycin resistance	Shuttle Yes
721			<i>B. subtilis</i> , <i>L. casei</i> , <i>L. lactis</i>			Klein et al., 1993		pJK352			Ampicillin and Chloramphenicol resistance	Shuttle Yes
722			<i>B. subtilis</i> , <i>L. casei</i> , <i>L. lactis</i>			Klein et al., 1993		pJK352d			Chloramphenicol resistance	Shuttle No
723			<i>B. subtilis</i> , <i>L. casei</i> , <i>L. lactis</i>			Klein et al., 1993		pJK353			Ampicillin and Chloramphenicol resistance	Shuttle Yes
724			<i>B. subtilis</i> , <i>L. casei</i> , <i>L. lactis</i>			Klein et al., 1993		pJK354			Ampicillin and Chloramphenicol resistance	Shuttle Yes
725			<i>B. subtilis</i> , <i>L. casei</i> , <i>L. lactis</i>			Klein et al., 1993		pJK355			Chloramphenicol resistance	Shuttle No
726			<i>B. subtilis</i> , <i>L. casei</i> , <i>L. lactis</i>			Klein et al., 1993		pJK356			Chloramphenicol resistance	Shuttle No
727			<i>L. delbrueckii</i>			Zink et al., 1991		pJK300			Ampicillin and Chloramphenicol resistance	Shuttle Yes
728			<i>Lactobacillus</i> spp. 89			Chagnaud et al., 1992		pLE16			Chloramphenicol and Tetracycline resistance	Shuttle Yes
729			<i>L. lactis</i> , <i>L. johnsonii</i>			Azcarate-Peri and Raya, 2002		pSS1			Chloramphenicol, Erythromycin, and Tetracycline resistance	Shuttle No
730			<i>L. lactis</i>			Boumiquel et al., 2002		pN42+pJDC9			Erythromycin resistance	Shuttle Yes
731			<i>L. helveticus</i>			Hashiba et al., 1990		pLHR			Ampicillin and Erythromycin resistance	Shuttle Yes
732			<i>L. helveticus</i>			Hashiba et al., 1992		pBG10			B-gal	Shuttle Yes
733			<i>L. helveticus</i> , <i>L. casei</i>			Yamamoto and Takano, 1996		pCP53d			Tetracycline resistance	Shuttle No
734			<i>L. acidophilus</i>			Damiani et al., 1987		pPV751			Tetracycline resistance	Shuttle Yes
735			<i>L. acidophilus</i>			Damiani et al., 1987		pPV754			Tetracycline resistance	Shuttle Yes
736			<i>L. acidophilus</i>			Luchansky et al., 1988		pTRK13			Chloramphenicol resistance	Shuttle Yes

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327			<i>L. acidophilus</i>			Muriana and Klaenhammer, 1991		pTRK159			Chloramphenicol, Erythromycin, and Tetracycline resistance	Shuttle Yes
328			<i>L. acidophilus</i> , <i>L. casei</i>			Kanatani et al., 1992		pULA105E			Ampicillin and Erythromycin resistance	Shuttle Yes
329			<i>L. acidophilus</i> , <i>L. casei</i> , <i>E. coli</i>			Sano et al., 1997		pLA106PVem			Erythromycin resistance	Shuttle No
330			<i>L. sakei</i> , <i>L. plantarum</i> , <i>L. curvatus</i> , <i>L. casei</i>			Alpert et al., 2003		pRV566			Ampicillin and Erythromycin resistance	Shuttle Yes
341			<i>L. casei</i> , <i>L. lactis</i>			Chassy and Flickinger, 1987; Heme et al., 1994		pLZ15			lac	Shuttle Yes
342			<i>L. delbrueckii</i>			Zink et al., 1991		pAZ20			Ampicillin and Chloramphenicol resistance	Shuttle Yes
343						Mercenier et al., 1989		pTQ219			Erythromycin resistance	Shuttle Yes
344			<i>S. thermophilus</i>			Solaiman and Somkuti, 1993		pMEU5a			Ampicillin and Erythromycin resistance	Shuttle Yes
345			<i>S. thermophilus</i>			Solaiman and Somkuti, 1993		pMEU5b			Ampicillin and Erythromycin resistance	Shuttle Yes
346			<i>S. thermophilus</i>			Solaiman and Somkuti, 1993		pMEU6a			Ampicillin and Erythromycin resistance	Shuttle Yes
347			<i>S. thermophilus</i>			Solaiman and Somkuti, 1993		pMEU6b			Ampicillin and Erythromycin resistance	Shuttle Yes
348			<i>S. thermophilus</i>			Solaiman and Somkuti, 1993		pMEU9			Ampicillin, Chloramphenicol, and Erythromycin resistance	Shuttle Yes
349			<i>S. thermophilus</i>			Solaiman and Somkuti, 1993		pMEU10			Ampicillin, Chloramphenicol, and Erythromycin resistance	Shuttle Yes
350			<i>S. thermophilus</i>			Solaiman and Somkuti, 1995		pEU5xML2201 a/b			Ampicillin and Erythromycin resistance	Shuttle Yes
351			<i>S. thermophilus</i>			Solaiman and Somkuti, 1995		pEU5aML2201 a			Ampicillin and Erythromycin resistance	Shuttle Yes
352			<i>S. thermophilus</i>			Solaiman and Somkuti, 1995		pEU5aCH2201 a/b			Ampicillin and Erythromycin resistance	Shuttle Yes
353			<i>S. thermophilus</i>			Somkuti et al., 1995		pER82			Chloramphenicol and Erythromycin resistance	Shuttle No
354			<i>S. thermophilus</i>			Somkuti et al., 1995		pER82Pb			Chloramphenicol and Erythromycin resistance	Shuttle No
355			<i>S. thermophilus</i>			Solaiman and Somkuti, 1996		pER82pPbID			Ampicillin, Chloramphenicol, and Erythromycin resistance	Shuttle No
356			<i>S. thermophilus</i>			Solaiman and Somkuti, 1996		pEU5aID2201			Ampicillin and Erythromycin resistance	Shuttle No
357			<i>S. thermophilus</i>			Somkuti and Steinberg, 1999		pG341Pa			Ampicillin and Erythromycin resistance	Shuttle Yes
358			<i>S. thermophilus</i>			Somkuti and Steinberg, 1999		pG341Pb			Ampicillin and Erythromycin resistance	Shuttle Yes
359			<i>E. coli</i> , <i>S. thermophilus</i> , <i>L. lactis</i> , <i>E. faecalis</i>			Coderre and Somkuti, 1999		pPC418			Ampicillin and Erythromycin resistance	Shuttle Yes
360			<i>S. thermophilus</i> , <i>S. salivarius</i> , <i>L. lactis</i> , <i>E. coli</i>			Turgeon and Moineau, 2001		pSMQ172cat			Chloramphenicol resistance	Shuttle No
361			<i>S. thermophilus</i> , <i>L. lactis</i>			Su et al., 2002		pND913			Ampicillin and Erythromycin resistance	Shuttle Yes
362			<i>S. thermophilus</i>			Su et al., 2002		pND914			Ampicillin and Erythromycin resistance	Shuttle Yes
363			<i>S. thermophilus</i>			Su et al., 2002		pND915			Ampicillin and Erythromycin resistance	Shuttle Yes
364			<i>S. thermophilus</i>			El Demerdash et al., 2003		pHRM1			shsp	Shuttle No
365			<i>L. cremoris</i> , <i>L. mesenteroides</i> subsp. <i>mesenteroides</i> subsp. <i>dextranicum</i> , <i>L. sakei</i>			Biet et al., 1999		pFBYC018E			Ampicillin and Erythromycin resistance	Shuttle Yes
366			<i>L. cremoris</i> , <i>L. mesenteroides</i> subsp. <i>mesenteroides</i> subsp. <i>dextranicum</i> , <i>L. sakei</i>			Biet et al., 1999		pFBYC18E			Erythromycin resistance	Shuttle No

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100			<i>L. crumoris</i> , <i>L. mesenteroides</i> subsp. <i>mesenteroides</i> subsp. <i>dextranicum</i> , <i>P. acidilactici</i> , <i>L. sakei</i>			Biat et al., 2002		pFBYC050E			Ampicillin and Erythromycin resistance	Shuttle Yes
101			<i>L. crumoris</i> , <i>L. mesenteroides</i> subsp. <i>mesenteroides</i> subsp. <i>dextranicum</i> , <i>P. acidilactici</i> , <i>L. sakei</i>			Biat et al., 2002		pFBYC50E			Erythromycin resistance	Shuttle No
102			<i>L. mesenteroides</i> subsp. <i>mesenteroides</i> . <i>L. paramesenteroides</i> , <i>L. lactis</i> subsp. <i>lactis</i> , <i>S. thermophilus</i> , <i>B. subtilis</i> , <i>L. casei</i>			Coffey et al., 1994		pCI431			Chloramphenicol resistance	Shuttle No
103			<i>P. acidilactici</i> , <i>L. mesenteroides</i> subsp. <i>mesenteroides</i> . <i>L. plantarum</i> , <i>E. faecalis</i>			Benachour et al., 1995		pUCB813			Erythromycin resistance	Shuttle No
104			<i>P. acidilactici</i> , <i>L. mesenteroides</i> subsp. <i>mesenteroides</i> . <i>L. plantarum</i> , <i>E. faecalis</i>			Benachour et al., 1995		pUCB825			Erythromycin resistance	Shuttle No
105			<i>E. coli</i>			Matteuzzi et al., 1990		pMR3			Ampicillin and Chloramphenicol resistance	Shuttle Yes
106			<i>E. coli</i> , <i>B. animalis</i>			Argnani et al., 1996		pDG7			Ampicillin and Chloramphenicol resistance	Shuttle Yes
107			<i>B. longum</i> , <i>E. coli</i>			Missich et al., 1994		pRM2			Spectinomycin and Ampicillin resistance	Shuttle Yes
108			<i>E. coli</i>			Rossi et al., 1996		pDH7			Ampicillin and Chloramphenicol resistance	Shuttle Yes
109			<i>B. animalis</i> , <i>E. coli</i>			Rossi et al., 1996		pKG7			Ampicillin and Chloramphenicol resistance	Shuttle Yes
110			<i>B. animalis</i>			Rossi et al., 1996		pNC7			Ampicillin and Chloramphenicol resistance	Shuttle No
111			<i>B. animalis</i> , <i>E. coli</i>			Rossi et al., 1996		pDGE7			Ampicillin, Chloramphenicol, and Erythromycin resistance	Shuttle Yes
112			<i>B. animalis</i> , <i>B. bifidum</i> , <i>B. infantis</i> , <i>B. longum</i> , <i>B. magnum</i>			Rossi et al., 1998		pLF5			Ampicillin and Chloramphenicol resistance	Shuttle Yes
113			<i>B. animalis</i> , <i>B. bifidum</i> , <i>B. infantis</i> , <i>B. longum</i> , <i>B. magnum</i>			Rossi et al., 1998		pCLJ15			Ampicillin and Erythromycin resistance	Shuttle Yes
114			<i>B. animalis</i> , <i>B. bifidum</i> , <i>B. infantis</i> , <i>B. longum</i> , <i>B. magnum</i>			Rossi et al., 1998		pSPEC1			Spectinomycin and Ampicillin resistance	Shuttle Yes
115			<i>B. animalis</i> , <i>B. bifidum</i> , <i>B. infantis</i> , <i>B. longum</i> , <i>B. magnum</i>			Rossi et al., 1998		pTRE3			Chloramphenicol resistance	Shuttle No
116			<i>B. longum</i>			Matsumura et al., 1997		pBLES100			Spectinomycin resistance	Shuttle Yes
117			<i>B. animalis</i>			Park et al., 1999		pBKJ50F/R			Ampicillin and Chloramphenicol resistance	Shuttle Yes
118			<i>B. animalis</i> , <i>B. infantis</i>			Park et al., 2000		pBKJ36F/R			Ampicillin and Chloramphenicol resistance	Shuttle Yes
119			<i>B. longum</i>			Park et al., 2003		pBES2			Ampicillin and Chloramphenicol resistance	Shuttle Yes
120			<i>B. animalis</i>			Gonzalez Vara et al., 2003		pLAV			Chloramphenicol resistance	Shuttle Yes

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1		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pMH1520	xyIA		-Ampicillin -Tetracycline				
2		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pMM1522	xyIA		-Ampicillin -Tetracycline				pMM1522+signal sequence
3		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pMM1525	xyIA		-Ampicillin -Tetracycline				
4		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pHIS1522	xyIA		-Ampicillin -Tetracycline		His		
5		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pHIS1525	xyIA		-Ampicillin -Tetracycline		His		pHIS1522+signal sequence
6		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pSTREP1525	xyIA		-Ampicillin -Tetracycline	STREP			pHIS1522+signal sequence
7		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pSTREPHIS1525	xyIA		-Ampicillin -Tetracycline	STREP	His		
8		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pC-HIS1622	xyIA		-Ampicillin -Tetracycline		His		
9		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pC-STREP1622	xyIA		-Ampicillin -Tetracycline	STREP			
10		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pN-HIS-TEV1622	xyIA		-Ampicillin -Tetracycline	His			
11		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pN-STREP- TEV1622	xyIA		-Ampicillin -Tetracycline	STREP			
12		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pN-STREP-Xa1622	xyIA		-Ampicillin -Tetracycline	STREP			
13		-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pSTOP1622	xyIA		-Ampicillin -Tetracycline				
14	levansucrase (Lactobacillus reuteri)	-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pRBBm15	xyIA		-Ampicillin -Tetracycline		His		
15	levansucrase (Lactobacillus reuteri)	-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pRBBm13	xyIA		-Ampicillin -Tetracycline	STREP			
16	levansucrase (Lactobacillus reuteri)	-E coli -Bacillus megaterium	Bacillus megaterium	Bacillus megaterium発現	Mo Bi Tec			pRBBm16	xyIA		-Ampicillin -Tetracycline	STREP	His		
17		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pHT01	groE		-Ampicillin				
18		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pHT43	groE		Chloramphenicol -Ampicillin				
19		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pHT08	groE		Chloramphenicol -Ampicillin	His			
20		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pHT09	groE		Chloramphenicol -Ampicillin	STREP			
21		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pHT10	groE		Chloramphenicol -Ampicillin		cMyc		
22		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pBacTag-FLAG	spac		Chloramphenicol -Ampicillin -Erythromycin		FLAG		
23		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pBacTag-cMyc	spac		-Ampicillin -Erythromycin		cMyc		
24		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pBacTag-HA	spac		-Ampicillin -Erythromycin		HA		
25		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pBacTag-GFP+	spac		-Ampicillin -Erythromycin		GFP+		
26		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pBacTag-CFP	spac		-Ampicillin -Erythromycin		CFP		
27		-E coli -Bacillus subtilis	Bacillus subtilis	Bacillus subtilis発現	Mo Bi Tec			pBacTag-YFP	spac		-Ampicillin -Erythromycin		YFP		

番号	区	導入遺伝子	宿主	生産物・機能・特徴	研究・開発	文献	遺伝子組換え法	ベクター名	プロモーター	ターミネーター	マーカー	N末端	C末端	タグ	備考		
28	分	white spot syndrome virus (WSSV) envelope protein VP28	Bacillus subtilis OB1130	-B. subtilisはVP28を分泌・膜口投入し、そのWSSVに対する防御効果を誘導	中国・浙江 中江 大学	Fu LL et al. (2008) Oral vaccination with envelope protein VP28 against white spot syndrome virus in <i>Bacillus subtilis</i> as delivery vehicles. Proc Natl Acad Sci U S A 105:1117-1121 Letz Apgl Microbiol	electroporation	pGS-H1	sigma43 promoter from <i>B. subtilis</i> 168	Kanamycin	Kanamycin						
29		α-amylase gene	Bacillus subtilis OB1130	培養液中にα-amylaseを産生	中・山中大 学	荻本清ら(1994) 淀粉酶基因在枯草杆菌中的表达和分泌 Acta Genetica Sinica 21:307-316 Anra DF et al. (2003) High-level expression of Alicyclobacillus acidocaldarius thioredoxin in <i>Pichia pastoris</i> and <i>Bacillus subtilis</i> . Protein Expr Purif. 30:179-184	electroporation	pUSA186	sigma43 promoter from <i>B. subtilis</i> 168	Kanamycin	Kanamycin						
30		thioredoxin from Alicyclobacillus acidocaldarius (AtRx)	Bacillus subtilis DB428	B. subtilisはAtRxを培養液中に分泌し、その分泌したAtRxは生物活性を有する	日本 大学	Shiroza T et al. (2001) Construction of a chimeric shuttle plasmid via a heterodimer system: secretion of an sFv protein from <i>Bacillus brevis</i> cells capable of inhibiting recombination. Biotechnol Biochem 35:2385-2389 Shiroza T et al. (2003) Production of a single-chain variable action capable of inhibiting the Streptococcus mutans glucosyltransferase in <i>Bacillus brevis</i> . shuttle plasmid secreting its gene product Biochim Biophys Acta 1603:103-110 Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Kim HK et al. (2002) Expression and characterization of Ca(2+)-independent lipase from <i>Bacillus pumilus</i> BZ6. Biotechnol Biochem 36:11:1583(2):205-212 Li W et al. (2007) Characterization of two temperature-inducible promoters newly isolated from <i>B. subtilis</i> . Biochem Biophys Res Commun. 358(4):1148-53	electroporation	pYS08	α-amylase promoter from <i>Bacillus subtilis</i> NA64	Erythromycin	Erythromycin						
31		single chain variable fragment (scFv)	Bacillus brevis HPD-31	B. brevisはscFvを分泌	日本 大学	Shiroza T et al. (2001) Construction of a chimeric shuttle plasmid via a heterodimer system: secretion of an sFv protein from <i>Bacillus brevis</i> cells capable of inhibiting recombination. Biotechnol Biochem 35:2385-2389 Shiroza T et al. (2003) Production of a single-chain variable action capable of inhibiting the Streptococcus mutans glucosyltransferase in <i>Bacillus brevis</i> . shuttle plasmid secreting its gene product Biochim Biophys Acta 1603:103-110 Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Kim HK et al. (2002) Expression and characterization of Ca(2+)-independent lipase from <i>Bacillus pumilus</i> BZ6. Biotechnol Biochem 36:11:1583(2):205-212 Li W et al. (2007) Characterization of two temperature-inducible promoters newly isolated from <i>B. subtilis</i> . Biochem Biophys Res Commun. 358(4):1148-53	electroporation	pYS18	α-amylase promoter from <i>Bacillus subtilis</i> NA64	Erythromycin	Erythromycin						
32		Glutamin合成制御タンパク GltC(gfC)	Bacillus subtilis SMY	B. subtilisはGltCを表現	米・台湾 大学	Shiroza T et al. (2001) Construction of a chimeric shuttle plasmid via a heterodimer system: secretion of an sFv protein from <i>Bacillus brevis</i> cells capable of inhibiting recombination. Biotechnol Biochem 35:2385-2389 Shiroza T et al. (2003) Production of a single-chain variable action capable of inhibiting the Streptococcus mutans glucosyltransferase in <i>Bacillus brevis</i> . shuttle plasmid secreting its gene product Biochim Biophys Acta 1603:103-110 Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Kim HK et al. (2002) Expression and characterization of Ca(2+)-independent lipase from <i>Bacillus pumilus</i> BZ6. Biotechnol Biochem 36:11:1583(2):205-212 Li W et al. (2007) Characterization of two temperature-inducible promoters newly isolated from <i>B. subtilis</i> . Biochem Biophys Res Commun. 358(4):1148-53	electroporation	pSP12	gltC promoter	Neomycin	Neomycin		6xHis				相同組織換えにより発原体上へ導入される
33		Glutamin合成制御タンパク GltC(gfC)	Bacillus subtilis SMY	B. subtilisはGltCを表現	米・台湾 大学	Shiroza T et al. (2001) Construction of a chimeric shuttle plasmid via a heterodimer system: secretion of an sFv protein from <i>Bacillus brevis</i> cells capable of inhibiting recombination. Biotechnol Biochem 35:2385-2389 Shiroza T et al. (2003) Production of a single-chain variable action capable of inhibiting the Streptococcus mutans glucosyltransferase in <i>Bacillus brevis</i> . shuttle plasmid secreting its gene product Biochim Biophys Acta 1603:103-110 Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Pioossi S et al. (2007) Molecular mechanism of the regulation of <i>Bacillus subtilis</i> gGAB expression by gHC. Kim HK et al. (2002) Expression and characterization of Ca(2+)-independent lipase from <i>Bacillus pumilus</i> BZ6. Biotechnol Biochem 36:11:1583(2):205-212 Li W et al. (2007) Characterization of two temperature-inducible promoters newly isolated from <i>B. subtilis</i> . Biochem Biophys Res Commun. 358(4):1148-53	electroporation	pSP16	gltC promoter	Neomycin	Neomycin		6xHis				相同組織換えにより発原体上へ導入される
34		Bacillus pumilus BZ6由来 lipaseBZ6 (Lip)	Bacillus subtilis DB104	B. subtilisはlipase BZ6を分泌	韓国・Korea Research Institute of Bioscience and Biotechnol ogy	Kim HK et al. (2002) Expression and characterization of Ca(2+)-independent lipase from <i>Bacillus pumilus</i> BZ6. Biotechnol Biochem 36:11:1583(2):205-212 Li W et al. (2007) Characterization of two temperature-inducible promoters newly isolated from <i>B. subtilis</i> . Biochem Biophys Res Commun. 358(4):1148-53	electroporation	pMB26	lip promoter	Ampicillin Kanamycin	Ampicillin Kanamycin						
35		bgAB	Bacillus subtilis 1A747	B-galactosidase	Northwest A&F University	Li W et al. (2007) Characterization of two temperature-inducible promoters newly isolated from <i>B. subtilis</i> . Biochem Biophys Res Commun. 358(4):1148-53	electroporation	pSI427	temperature inducible promoter P2	Chloramphenicol Spectinomycin	Chloramphenicol Spectinomycin						
36																	

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37		bgxB	Bacillus subtilis 1A747	B-galactosidase	中・ Northwest A&F University	Li W et al. (2007) Characterization of two temperature-inducible promoters newly isolated from <i>B. subtilis</i> . Biochem Biophys Res Commun. 358(4):1148-53.	electroporation	pSI7427	temperature inducible promoter P7		Chloramphenicol -Spectinomycin		
38			Bacillus subtilis Marburg 168	B. subtilisは培養液中に cystatin SあるいはSAを産生	日・ Biological science laboratories of Kao corporation	Comun. 358(4):1148-53. Extracellular production of cystatin S and its application. Protein Expr Purif. 48:203-210.		pHSP-US	alkaline endoglucanase promoter (from <i>Bacillus sp.</i> KSM-S237)		-Ampicillin -Tetracycline		
39			Bacillus subtilis 1012	B. subtilisはいずれのタンパク質も発現	日・ Kim L et al. (1996)	A xylose-inducible <i>Bacillus subtilis</i> integration vector and its application. Gene. 181(1-2):71-6.		pX	xyIA (from <i>B. megaterium</i>)		Chloramphenicol		染色体上の amyE 部位に plasmid は組み込まれる
40			Bacillus subtilis YB886	B. subtilisは温度依存的に GFPを発現し、蛍光を発する	西・ Universidad Autonoma	Serrano-Heras G et al. (2005) A new plasmid vector for regulated gene expression in <i>Bacillus subtilis</i> . Plasmid. 54(3):278-82.	electroporation	pPR54	λ -phage promoter (temperature inducible)	rmB (from <i>E. coli</i>)	Bleomycin		
41			Bacillus subtilis IHA01	B-galactosidase	独・ university of bayreuth	Hard B et al. (2001) Development of a new integration site within the <i>Bacillus subtilis</i> chromosome and construction of compatible expression cassettes. J Bacteriol. 183(8):2696-9.	electroporation	pAX01	xyIA promoter	t0 (lambda phage)	Erythromycin		染色体上の lacA 部位に plasmid は組み込まれる
42			Bacillus subtilis IHA01	B-galactosidase	独・ university of bayreuth	Hard B et al. (2001) Development of a new integration site within the <i>Bacillus subtilis</i> chromosome and construction of compatible expression cassettes. J Bacteriol. 183(8):2696-9.	electroporation	pAX01	lac promoter	t0 (lambda phage)	Erythromycin		染色体上の lacA 部位に plasmid は組み込まれる
43			Bacillus subtilis 1A747	B-galactosidase	中・ Northwest A&F University	中・ Zhejiang Ming-Ming Y et al. (2006) Construction and characterization of a novel mitose inducible expression vector in <i>Bacillus subtilis</i> . Biotechnol Lett.	electroporation	-pLJ-7 -pGJ203 -pDCA	glv (from <i>B. subtilis</i> maltose utilization operon)		Chloramphenicol -Spectinomycin		
44			Bacillus subtilis 1A747	B-galactosidase	中・ Northwest A&F University	Zhang AL et al. (2007) Assay and characterization of a strong promoter element from <i>B. subtilis</i> . Biochem Biophys Res Commun. 354(1):90-5.	electroporation	pYG123	yxIE		Chloramphenicol		
45			Bacillus subtilis WB600	B. subtilisは培養液中に活性を有する penicillin G acylase を産生	中・ Chinese academy of sciences	Yang S et al. (2001) Expression and purification of extracellular penicillin G acylase in <i>Bacillus subtilis</i> . Protein Expr Purif. 21:60-65.	protoplast method	pEES102	P43		Neomycin		
46			Bacillus subtilis 1012	B-galactosidase -Heat shock protein	独・ University of Bayreuth	Neyen HD et al. (2005) Construction of plasmid-based expression vectors for <i>Bacillus subtilis</i> exhibiting full structural stability.		pHCMC02	lepA (weak constitutive promoter)	trpA	-Ampicillin Chloramphenicol		

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					国	遺伝子組換え法					C末端	その他
47		bgxB or lacZ	Bacillus subtilis 1012 Bacillus subtilis IS58	B-galactosidase	独・University of Bayreuth	Nyuen HD et al. (2005) Construction of plasmid-based expression vectors for Bacillus subtilis exhibiting full structural stability.	pHMC03	gsB (xylase inducible promoter)	tpA	-Ampicillin Chloramphenicol		
48		bgxB or htpG	Bacillus subtilis 1012	B-galactosidase Heat shock protein	独・University of Bayreuth	Nyuen HD et al. (2005) Construction of plasmid-based expression vectors for Bacillus subtilis exhibiting full structural stability.	pHMC04	xyIA (xylase inducible promoter)	tpA	-Ampicillin Chloramphenicol		
49		bgxB or htpG	Bacillus subtilis 1012	B-galactosidase Heat shock protein	独・University of Bayreuth	Nyuen HD et al. (2005) Construction of plasmid-based expression vectors for Bacillus subtilis exhibiting full structural stability.	pHMC05	spac (PTG inducible promoter)	tpA	-Ampicillin Chloramphenicol		
50		Glutamyl endopeptidase gene (geBL from Bacillus licheniformis) B-6839	Bacillus subtilis A.J73	B. subtilisは培養液中に GeBLを産生	露・Institute of Genetics and Selection of genes in Bacillus subtilis by Industrial Microorganism	Shevelev AB et al. (2000) Expression of bacillar glutamyl endopeptidase and a new mobilizable single-replicon vector pLF-sms Plasmid. 43(3):19D-9	pLF9	-cryIAa (endotoxin gene from B. thuringiensis ssp kurstaki) -Kanamycin		Chloramphenicol		
51		-Glutamyl endopeptidase gene (geBL from Bacillus intermedius)	Bacillus subtilis A.J73	B. subtilisは培養液中に GeBLを産生	露・Institute of Genetics and Selection of genes in Bacillus subtilis by Industrial Microorganism	Shevelev AB et al. (2000) Expression of bacillar glutamyl endopeptidase and a new mobilizable single-replicon vector pLF-sms Plasmid. 43(3):19D-9	pLF14	Kanamycin		Chloramphenicol		
52		Bacillus thuringiensis subsp. Morrisoni PC-14由来 cry11Aa	Bacillus brevis	B. brevisはCry11Aaを発現し、Aedes aegyptiとCulex pipiens に対し殺虫活性をもつ	韓・Seoul national university	Roh JY et al. (2010) Expression of Bacillus thuringiensis mesoautocidal toxin in an antimicrobial Bacillus brevis strain. Journal of Asia-Pacific Entomology. 13(6):61-64	pPro11A	cry1Ac		Erythromycin		vectorは宿主の染色体上に組み込まれる
53		*xynA (xylanase from Bacillus subtilis 168) *bgl (glucanase from Bacillus subtilis 108)	Bacillus subtilis 1A304	B. subtilisは培養液中に 糖鎖活性を有する xylanase あるいは glucanase を産生	中・University of Hong Kong	Chan AY et al. (2002) A dual protein expression system in Bacillus subtilis. Protein Expr Purif. 26:337-342.	pSGt	φ105 (thermal induction)	α-amylase (from B. licheniformis)	Chloramphenicol Ampicillin		
54		*birA (Bacillus subtilis biotin biosynthetic enzyme) *prsA (Bacillus subtilis extracytoplasmic molecular chaperone)	Bacillus subtilis WB600	B. subtilisはそれぞれ BirA, PrsA を遺伝体内に SAK を培養中に産生する	加・University of Calgary	Wu SC et al. (1999) Development of improved pUB110-based vectors for expression and secretion studies in Bacillus subtilis. J Biotechnol. 72:185-195.	pUB18P43BIRA pUB18P43PRSA pUB18P43SAKT	P43 (B. subtilis constitutive promoter)	pUB18P43SA KTのみ T1T2 (from E. coli rrmB operon)	Kanamycin		
55		*prsA (Bacillus subtilis extracytoplasmic molecular chaperone) sak (Staphylokinase)	Bacillus subtilis WB600	B. subtilisはそれぞれ BirA, PrsA を遺伝体内に SAK を培養中に産生する	加・University of Calgary	Wu SC et al. (1999) Development of improved pUB110-based vectors for expression and secretion studies in Bacillus subtilis. J Biotechnol. 72:185-195.	pE18P43BIRA pE18P43PRSA pE18P43SAK	P43 (B. subtilis constitutive promoter)	P43 (B. subtilis constitutive promoter)	•Erythromycin •Lincomycin		
56			Bacillus subtilis WB600	B. subtilisはSAKを培養液中に産生する	加・University of Calgary	Development of improved pUB110-based vectors for expression and secretion studies in Bacillus subtilis. J Biotechnol. 72:185-195.	pUB19P43	P43 (B. subtilis constitutive promoter)		Kanamycin		

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57		cryIC (<i>C</i> -endotoxin from <i>Bacillus thuringiensis</i> subsp. <i>Aizawai</i> 7-29) - <i>Bacillus thuringiensis</i> (kto)	- <i>Bacillus thuringiensis</i> 407-OA - <i>Bacillus thuringiensis</i> (kto)	CryICは1つだけの宿主でも発現するが、 <i>Spodoptera littoralis</i> への殺虫活性は407-OA株が高い	国 Sanchis V et al. (1996) Insecticidal <i>Bacillus thuringiensis</i> recombinant strains by using the sporulation non-dependent expression system of cryIIIA and site specific recombination vector J Biotechnol 48:81-96 Kai K et al. (1997) Over-expression and post-translational modification of thermophilic <i>Bacillus</i> cytochrome <i>c</i> -551 in <i>Bacillus subtilis</i> J Ferment Bioeng 84:190-194 Anriov RI et al. (1995) Expression of a <i>cellE</i> gene from <i>Clostridium thermocellum</i> in <i>Bacillus</i> J Ferment Bioeng 79:530-537 Anriov RI et al. (1995) Expression of a <i>cellE</i> gene from <i>Clostridium thermocellum</i> in <i>Bacillus</i> J Ferment Bioeng 79:530-537 Sharipov M et al. (2008) The expression of the <i>cellE</i> proteinase gene of <i>Bacillus intermedius</i> in <i>Bacillus subtilis</i> Microb Drug Deliv 163:39-50 Mull A et al. (1995) Expression and secretion of β -tubulin in <i>Bacillus subtilis</i> Biotechnol 77:255-261 Yue Y et al. (2009) Cloning of amidase gene from <i>Rhodococcus erythropolis</i> and expression by distinct promoters in <i>Bacillus subtilis</i> Mol Catal B Enzym 124:100-105 Takao Y et al. (2005) Hyper expression of <i>Kojibiose phosphorylase</i> gene and trehalase phosphorylase gene from <i>Thermoarabacter brockii</i> ATCC35047 in <i>Bacillus subtilis</i> and selaginose synthesis utilizing two phosphorylases. Plant T et al. (2006) Novel plasmid-based expression vectors for <i>intra-</i> and <i>extracellular</i> production of recombinant proteins in <i>Bacillus subtilis</i> . Protein Expr Purif 46(2):189-95.	electroporation pHTBS-F3-IC cryIIIA	B. <i>stearothermophilus</i> neutral protease gene		-Erythromycin -Ampicillin -Tetracycline							
58		ccsA (<i>Bacillus stearothermophilus</i> PS3 cytochrome <i>c</i> -551)	<i>Bacillus subtilis</i> 1012	B. <i>subtilis</i> は cytochrome を膜タンパク質として発現し、NADH oxidase 活性をもつ	日・Kusvnu Institute of Technology	electroporation pHYc551					-Tetracycline -Ampicillin					
59		<i>cellE</i> (from <i>Clostridium thermocellum</i>)	<i>Bacillus subtilis</i> 168	B. <i>subtilis</i> は培養液中に活性を持つ	日・Nis university	electroporation pUB102		<i>cellE</i> promoter			-Kanamycin -Neomycin					
60		<i>cellE</i> (from <i>Clostridium thermocellum</i>)	<i>Bacillus subtilis</i> 168	B. <i>subtilis</i> は培養液中に活性を持つ - <i>thoxy</i> methylcellulase を発酵表面に産生する	日・Nis university	electroporation pHE9102		<i>cellE</i> promoter			-Erythromycin Chloramphenicol					
61		aprBI (serine proteinase from <i>Bacillus intermedius</i>)	<i>Bacillus subtilis</i> A.J73	B. <i>subtilis</i> は培養液中に活性を持つ - <i>serine</i> protease を産生する	日・Kazan state university	protoplast method	pCS9		aprBI		Erythromycin					
62		bTub1 (β -tubulin from <i>Plasmodium falciparum</i>)	<i>Bacillus brevis</i> 47-50	B. <i>brevis</i> は培養液中に β -tubulin を産生	日・Biozentrum	Tri (s-p) yethylen method	pCB202	MNP (<i>B. brevis</i> midle wall protein promoter)			Erythromycin					
63		ami (Amidase from <i>Rhodococcus erythropolis</i>)	<i>Bacillus subtilis</i>	全ての組織単位が Amidase を産生するが、 <i>ami</i> のプロモーターを用いたものが産生量、活性が共に低い	日・Hayashibara biochemical laboratories	protoplast method	pGI103	- <i>sacB</i> - <i>amyE</i> - <i>p43</i> - <i>degQ</i> - <i>aprE</i>			Chloramphenicol					
64		k6P (<i>kojibiose</i> phosphorylase(KP)) treP (<i>trehalose</i> phosphorylase(TP)) from <i>Thermoarabacter brockii</i> ATCC35047	<i>Bacillus subtilis</i>	B. <i>subtilis</i> は KP あるいは TP を産生するが、 <i>treP</i> を用いてハイブリッドからキナーズの合成に成功	日・University of bayreuth	protoplast method	-pUBAPKP -pUBAFTP	BamP (α -amylase gene from <i>Bacillus amyloliquefaciens</i>)			Kanamycin					
65		- <i>bpaB</i> (β -galactosidase) - <i>trpC</i> (heat shock protein) - <i>pbaE</i> (penicillin binding protein PbaE)	<i>Bacillus subtilis</i> 1012	B. <i>subtilis</i> は IPTG の添加により、菌体内に各タンパク質を産生	日・University of bayreuth	pNDH33	<i>groE</i>				-Ampicillin Chloramphenicol					