

markedly reduced in the wild-type somites, in concert with formation of skeletal muscle derived from myotomes (Figure 7). Because the myogenic master transcription factor myoD plays essential roles in muscle differentiation [30] and can drive apoptotic pathways as reported by Asakura et al. [37], the perturbed expression of *myod* in the *C4ST-1* morphants may imply an immature developmental state of myotomes and/or disruption of the somitic musculature accompanied by excessive stimulation of the apoptotic cascade, leading to the aberrant morphology in the trunk and tail.

The defects in somites with persistent expression of *myod*, which were evoked by the functional knockdown of *C4ST-1*, have been also reported for zebrafish embryos injected with MOs, which inhibited functions of HS6ST-2, one of the sulfotransferases involved in Hep/HS (heparan sulfate) biosynthesis [38,39]. HS-PGs play roles in various cellular signalling pathways through the high-affinity binding of HS moieties to diverse Hep-binding proteins including growth factors, cytokines and morphogens [40,41]. Recent studies have demonstrated that CS also possesses the capacity to bind various Hep-binding proteins [9,41]. In fact, signalling pathways involving TGF β (transforming growth factor β) and BMP (bone morphogenetic protein) are dramatically affected in growth plates of *C4ST-1*-deficient mice [19]. Hence, the apparently similar muscular defects in both morphants might be indicative of partially overlapping or compensatory functions of CS and HS in the regulation of local signalling pathways via Hep-binding myogenic morphogens such as hedgehog and Wnt [42,43]. Most recently, Nakanaka et al. [44] reported that the decreased Wnt-3a signalling in *sog9* cells lacking *C4ST-1* mRNA is recovered by the introduction of C4ST-1, providing strong support to the notion that the fine structures of CS formed by C4ST-1 are required for efficient signalling inputs mediated by multiple morphogens.

C4ST-1 has been screened as one of the target genes induced by BMP signalling during differentiation of mouse embryonic stem cells [45]. This indicates that C4ST-1 is one of the essential modulators in a sequence of the BMP-dependent cell fate decisions. In zebrafish, BMPs are known to be key regulators of posterior (i.e. trunk and tail) mesoderm patterning, as typified by zebrafish mutants lacking *bmp2b* that do not form a tail [46]. In view of generally mild but definitive phenotypic abnormalities biased to trunk and tail in *C4ST-1* morphants, C4ST-1 might also be one of the downstream targets of the BMP signalling in the posterior half of the body axis and might play important roles in at least several developmental processes induced by BMPs. Therefore elucidation of the transcriptional regulatory mechanism for *C4ST-1* will provide insights into how C4ST-1 participates in body axis formation including muscle development.

Emerging evidence suggests that CS-PGs are crucial environmental modulators in the nervous system of vertebrates. During neuronal development and regeneration, they have apparently contradictory roles as major inhibitors of axonal pathfindings and regeneration and as neuritogenic molecules [2,3]. These functions are exerted mainly through their CS moieties [2,3,9,10]. During zebrafish embryogenesis, CS is abundantly distributed at the interface between the notochord and somites where ventral motor axons located in the middle of each spinal cord hemisegment project into ventral muscle [22]. As reported previously [23], elimination of CS in the trunk by injection with chondroitinase ABC induces axonal projections with abnormal side branches, indicating that CS constrains the outgrowth of the ventral motor nerves through its inhibitory role as a physical barrier or a repulsive cue for axonal growth. In the present study, aberrant axonal outgrowth of ventral motor neurons was also observed in *C4ST-1* morphants (Figure 7).

However, the most common abnormality was characterized by truncated axons rather than abnormally branched and misrouted axons (Table 3). One explanation for this discrepancy is that a low but significant level of CS in the trunk of *C4ST-1* morphants supports fasciculation of the ventral motor nerves, preventing the formation of side branches, whereas a nearly complete loss of CS by treatment with chondroitinase ABC does not. Furthermore, in view of the requirement of myotome-derived cues for the migration of motor axons [24,47], we cannot rule out the possibility that defects in muscle development in *C4ST-1* morphants led to additional indirect effects on the axonal pathfindings, because it has been reported that there was no damage in either the notochord or somites in chondroitinase-treated zebrafish embryos [23].

Although the precise molecular basis of the axonal pathfindings of ventral motor nerves involving 4-O-sulfated CS remains unclear, the high incidence of truncated axons, probably representing axons straying from their pathways, in *C4ST-1* morphants suggests potential bifunctional roles of CS not only as a repulsive cue but also as a permissive/attractive guidance cue for specific axons, as described above. In fact, a substrate uniformly precoated with oversulfated CS variants promoted the outgrowth of neurites in rodent embryonic hippocampal neurons, at least in part, by capturing and presenting several Hep-binding growth factors to neurons [2,3,9]. In addition, habenula nucleus axons derived from the developing rat diencephalon were also able to extend over a substrate precoated with a relatively high concentration of CS-PGs [48]. Interestingly, a CS-PG-coated substrate coexisting with *sema5A* (semaphorin 5A), a bifunctional guidance cue, serves as an inhibitory cue for habenula nucleus axons, resulting from conversion of the attractive property of *sema5A* into an inhibitory one through its specific interaction with the CS moieties of CS-PGs [48]. Therefore further exploration of CS-interacting molecules in the extracellular matrix and unidentified functional CS receptor(s) is required for a better understanding of the apparent contradictory neuroregulatory functions of CS.

In contrast with *C4ST-1* morphants and chondroitinase-treated embryos, functional knockdown of zygotic *ChSy-1* (chondroitin synthase-1), which encodes one of the glycosyltransferases involved in the biosynthesis of the chondroitin backbone, has been reported to have no significant effects on the pathfinding of the motor axons, although CS immunoreactivity was reduced in the morphants [47]. Recently, Izumikawa et al. [49,50] demonstrated that chondroitin polymerization can be achieved by any two combinations of four ChSy family members including ChSy-1. Therefore, if similar biosynthetic machinery is encoded by the zebrafish genome, a single knockdown of ChSy family members may not always lead to a drastic reduction in the amount of CS, which provokes developmental defects. Consequently, our results in conjunction with earlier studies [18,19,23] strongly suggest the critical functions of C4ST-1 in CS biosynthesis and in zebrafish embryogenesis. Thus further analysis focusing on C4ST-1 will facilitate our understanding of the molecular mechanisms underlying the development and pathology of various diseases involving CS.

ACKNOWLEDGEMENTS

We thank Junko Fukuda, Saki Sato and Shunsuke Kataoka for technical assistance.

FUNDING

This work was supported in part by a Scientific Promotion Fund from the Japan Private School Promotion Foundation, Grants-in-Aid for Encouragement of Young Scientists

[numbers 20790080 and 18790073; to T.M.] from the MEXT (Ministry of Education, Culture, Sports, Science and Technology) of Japan, a grant from the Uehara Memorial Foundation (to T.M.), and a Grant-in-aid for Scientific Research-B [number 20390019; to K.S.] and the CREST (Core Research for Evolutional Science and Technology) programme of the JST (Japan Science and Technology Agency; to K.S.).

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Received 11 August 2008/12 December 2008; accepted 6 January 2009

Published as BJ Immediate Publication 6 January 2009, doi:10.1042/BJ20081639

