

- novel M cell-specific carbohydrate-targeted mucosal vaccine effectively induces antigen-specific immune responses.** *J Exp Med* 2007, **204**:2789-2796.
- This paper describes a new procedure that made it possible to efficiently deliver protein antigens into M cells for mucosal IgA response.
7. Coombes JL, Powrie F: **Dendritic cells in intestinal immune regulation.** *Nat Rev Immunol* 2008, **8**:435-446.
 8. Mora JR, Bono MR, Manjunath N, Weninger W, Cavanagh LL, Roseblatt M, von Andrian UH: **Selective imprinting of gut-homing T cells by Peyer's patch dendritic cells.** *Nature* 2003, **424**:88-93.
 9. Johansson-Lindbom B, Svensson M, Wurbel MA, Malissen B, Marquez G, Agace W: **Selective generation of gut tropic T cells in gut-associated lymphoid tissue (GALT): requirement for GALT dendritic cells and adjuvant.** *J Exp Med* 2003, **198**:963-969.
 10. Mora JR, Iwata M, Eksteen B, Song S-Y, Junt T, Senman B, Otipoby KL, Yokota A, Takeuchi H, Ricciardi-Castagnoli P *et al.*: **Generation of gut-homing IgA-secreting B cells by intestinal dendritic cells.** *Science* 2006, **314**:1157-1160.

This paper describes a mechanistic link between gut-trafficking of B cells and their IgA production by PP DC-derived retinoic acid.

 11. Iwata M, Hirakiyama A, Eshima Y, Kagechika H, Kato C, Song S-Y: **Retinoic acid imprints gut-homing specificity on T cells.** *Immunity* 2004, **21**:527-538.

This paper reports for the first time that retinoic acid is necessary and sufficient to induce gut-homing receptors on T cells, and gut DCs can metabolize dietary vitamin A into retinoic acid.

 12. Dullaers M, Li D, Xue Y, Ni L, Gayet I, Morita R, Ueno H, Palucka KA, Banchereau J, Oh S: **A T cell-dependent mechanism for the induction of human mucosal homing immunoglobulin A-secreting plasmablasts.** *Immunity* 2009, **30**:120-129.
 13. Cerutti A, Rescigno M: **The biology of intestinal immunoglobulin A responses.** *Immunity* 2008, **28**:740-750.
 14. Sato A, Hashiguchi M, Toda E, Iwasaki A, Hachimura S, Kaminogawa S: **CD11b⁺ Peyer's Patch dendritic cells secrete IL-6 and induce IgA secretion from naïve B cells.** *J Immunol* 2003, **171**:3684-3690.
 15. Tezuka H, Abe Y, Iwata M, Takeuchi H, Ishikawa H, Matsushita M, Shiohara T, Akira S, Ohteki T: **Regulation of IgA production by naturally occurring TNF/INOS-producing dendritic cells.** *Nature* 2007, **448**:929-993.

This paper describes gut DCs expressing iNOS that is important for IgA production, including IgA class switching, in a TLR-dependent manner.

 16. Uematsu S, Fujimoto K, Jang M, Yang B, Jung Y, Nishiyama M, Sato S, Tsujimoto T, Yamamoto M, Yokota Y *et al.*: **Regulation of humoral and cellular gut immunity by lamina propria dendritic cells expressing Toll-like receptor 5.** *Nat Immunol* 2008, **9**:769-776.
 17. Balazs M, Martin F, Zhou T, Kearney J: **Blood dendritic cells interact with splenic marginal zone B cells to initiate T-independent-immune responses.** *Immunity* 2002, **17**:341-352.
 18. Litinsky MB, Nardelli B, Hilbert DM, He B, Schaffer A, Casali P, Cerutti A: **DCs induce CD40-independent immunoglobulin class switching through BlyS and APRIL.** *Nat Immunol* 2002, **3**:822-829.
 19. He B, Xu W, Santini PA, Polydorides AD, Chiu A, Estrella J, Shan M, Chadburn A, Villanacci V, Plebani A *et al.*: **Intestinal bacteria trigger T cell-independent immunoglobulin A2 class switching by inducing epithelial-cell secretion of the cytokine APRIL.** *Immunity* 2007, **26**:812-826.
 20. Sanchez J, Holmgren J: **Cholera toxin structure, gene regulation and pathophysiological and immunological aspects.** *Cell Mol Life Sci* 2008, **65**:1347-1360.
 21. World Health Organization, Initiative for Vaccine Research Team. **State of the art of vaccine research and development, 2005.**
 22. Hill DR, Ford L, Lalloo DG: **Oral cholera vaccines; use in clinical practice.** *Lancet Infect Dis* 2006, **6**:361-373.

Same as reference [24*].

 23. Lopez AL, Clemens JD, Deen J, Jadar L: **Cholera vaccines for the developing world.** *Human Vaccines* 2008, **4**:165-169.

Same as reference [24*].

 24. Sanchez J, Holmgren J: **Virulence factors, pathogenesis and vaccine protection in cholera and ETEC diarrhea.** *Curr Opin Immunol* 2005, **17**:388-398.

References [22*,23*,24*] are critical comprehensive review articles dealing with currently available cholera vaccines.

 25. Black RE, Levin MM, Clemens ML, Young CR, Svennerholm AM, Holmgren J: **Protective efficacy in humans of killed whole-vibrio oral cholera vaccine with and without the B subunit of cholera toxin.** *Infect Immun* 1987, **55**:1116-1120.
 26. Clemens JD, Sack DA, Harris JR, Chakraborty J, Khan MR, Stanton BF, Kay BA, Khan MU, Yunus M, Atkinson W *et al.*: **Field trial of oral cholera vaccines in Bangladesh.** *Lancet* 1986, **328**:124-127.
 27. Levine MM, Kaper JB, Herrington D, Ketley J, Losonsky G, Tacket CO, Tall B, Cryz S: **Safety, immunogenicity, and efficacy of recombinant live cholera vaccines, CVD 103 and CVD 103 HgR.** *Lancet* 1988, **2**:467-470.
 28. Tacket CO, Cohen MB, Wasserman SS, Losonsky G, Livio S, Kotloff K, Edelman R, Kaper JB, Cryz SJ, Giannella RA *et al.*: **Randomized, double blind, placebo-controlled, multicentered trial of the efficacy of a single dose of live cholera vaccine CVD103-HgR in preventing cholera following challenge with *Vibrio cholerae* O1 El tor inaba three months after vaccination.** *Infect Immun* 1999, **67**:6341-6345.
 29. Richie EE, Punjabi NH, Sidharta Y, Peetosutan K, Sukandar M, Wasserman SS, Lesmana M, Wangsasaputra F, Pandam S, Levine MM *et al.*: **Efficacy trial of single-dose live oral cholera vaccine CVD 103-HgR in North Jakarta, Indonesia, a cholera-endemic area.** *Vaccine* 2000, **18**:2399-2410.
 30. Haq TA, Mason HS, Clements JD, Arntzen CJ: **Oral immunization with a recombinant bacterial antigen produced in transgenic plants.** *Science* 1995, **268**:714-716.
 31. Arakawa T, Chong DKX, Langridge HR: **Efficacy of a food plant-based oral cholera toxin B subunit vaccine.** *Nat Biotech* 1998, **16**:292-297.
 32. Tacket CO, Mason HS, Losonsky G, Clements JD, Levine MM, Arntzen CJ: **Immunogenicity in humans of a recombinant bacterial antigen delivered in transgenic potato.** *Nat Med* 1998, **4**:607-609.
 33. Tacket CO: **Plant-derived vaccines against diarrheal diseases.** *Vaccine* 2005, **23**:1866-1869.
 34. Tacket CO: **Plant-based vaccines against diarrheal diseases.** *Trans Am Clin Climatol Assoc* 2007, **118**:79-87.
 35. Nochi T, Takagi H, Yuki Y, Yang L, Masumura T, Mejima M, Nakanishi U, Matsumura A, Uozumi A, Hiroi T *et al.*: **Rice-based mucosal vaccine as a global strategy for cold-chain- and needle-free vaccination.** *Proc Natl Acad Sci U S A* 2007, **104**:10986-10991.

This paper describes novel strategies of foreign antigen expression in rice seeds for the development of needle/syringe-free and cold-chain-free mucosal vaccine.

 36. Ma S, Jevnikar AM: **Transgenic rice for allergy immunotherapy.** *Proc Natl Acad Sci U S A* 2005, **102**:17255-17256.
 37. Mason HS, Warzecha H, Mor T, Arntzen CJ: **Edible plant vaccines: applications for prophylactic and therapeutic molecular medicine.** *Trends Mol Med* 2002, **8**:324-329.
 38. Pascual DW: **Vaccines are for dinner.** *Proc Natl Acad Sci U S A* 2007, **104**:10757-10758.

Pattern Recognition Receptors and Inflammation

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Infection of cells by microorganisms activates the inflammatory response. The initial sensing of infection is mediated by innate pattern recognition receptors (PRRs), which include Toll-like receptors, RIG-I-like receptors, NOD-like receptors, and C-type lectin receptors. The intracellular signaling cascades triggered by these PRRs lead to transcriptional expression of inflammatory mediators that coordinate the elimination of pathogens and infected cells. However, aberrant activation of this system leads to immunodeficiency, septic shock, or induction of autoimmunity. In this Review, we discuss the role of PRRs, their signaling pathways, and how they control inflammatory responses.

Introduction

Inflammation is a protective response by the body to ensure removal of detrimental stimuli, as well as a healing process for repairing damaged tissue (Medzhitov, 2008). Inflammation is caused by various factors such as microbial infection, tissue injury, and cardiac infarction. Classically, inflammation is characterized by five symptoms: redness, swelling, heat, pain, and loss of tissue function. These macroscopic symptoms reflect increased permeability of the vascular endothelium allowing leakage of serum components and extravasation of immune cells. The inflammatory response is then rapidly terminated and damaged tissues are repaired. However, overproduction of cytokines (a cytokine storm) by immune cells to overwhelm pathogens can be fatal. A cytokine storm can also be caused by noninfectious diseases such as graft-versus-host disease (GVHD). Inflammatory responses are also critical for the pathogenesis of autoimmune diseases.

The innate immune system is the major contributor to acute inflammation induced by microbial infection or tissue damage (Akira et al., 2006; Beutler et al., 2006). Furthermore, innate immunity is also important for the activation of acquired immunity. Although innate immune cells including macrophages and dendritic cells (DCs) play important roles, nonprofessional cells such as epithelial cells, endothelial cells, and fibroblasts also contribute to innate immunity. Germline-encoded pattern recognition receptors (PRRs) are responsible for sensing the presence of microorganisms. They do this by recognizing structures conserved among microbial species, which are called pathogen-associated molecular patterns (PAMPs). Recent evidence indicates that PRRs are also responsible for recognizing endogenous molecules released from damaged cells, termed damage-associated molecular patterns (DAMPs). Currently, four different classes of PRR families have been identified. These families include transmembrane proteins such as the Toll-like receptors (TLRs) and C-type lectin receptors (CLRs), as well as cytoplasmic proteins such as the Retinoic acid-inducible gene (RIG)-I-like receptors (RLRs) and NOD-like receptors (NLRs).

These PRRs are expressed not only in macrophages and DCs but also in various nonprofessional immune cells. With the exception of some NLRs, the sensing of PAMPs or DAMPs by PRRs upregulates the transcription of genes involved in inflammatory responses. These genes encode proinflammatory cytokines, type I interferons (IFNs), chemokines and antimicrobial proteins, proteins involved in the modulation of PRR signaling, and many uncharacterized proteins. The expression patterns of the inducible genes differ among activated PRRs.

The inflammatory response is orchestrated by proinflammatory cytokines such as tumor necrosis factor (TNF), interleukin (IL)-1, and IL-6. These cytokines are pleiotropic proteins that regulate the cell death of inflammatory tissues, modify vascular endothelial permeability, recruit blood cells to inflamed tissues, and induce the production of acute-phase proteins. Although TNF and IL-6 are mainly regulated at the transcriptional and translational levels, the production of IL-1 β is regulated by a two-step mechanism. The first step is the expression of an IL-1 β zymogen, pro-IL-1 β , which is regulated by the synthesis of its mRNA in a TLR signal-dependent manner. However, IL-1 β maturation requires cleavage of pro-IL-1 β by a protease, caspase-1, which is activated independently of TLR signaling. The complex that activates caspase-1, called the inflammasome, is composed of NLRs, ASC, and caspase-1 (see Review by K. Schroder and J. Tschopp on page 821 of this issue). Type I IFNs, including multiple forms of IFN- α and single forms of IFN- β , IFN- ω , etc., are also involved in the modulation of inflammation (Honda et al., 2006). Type I IFNs play central roles in antiviral responses by inducing apoptotic cell death in virally infected cells, rendering cells resistant to virus infection, activating acquired immunity, and stimulating hematopoietic stem cell turnover and proliferation. Secreted type I IFNs alert the surrounding cells via type I IFN receptors by triggering a signaling cascade that leads to the phosphorylation and nuclear translocation of IFN-stimulated gene factor 3 (ISGF3), a complex composed of Signal Transducers and Activators of Transcription 1 (STAT1), STAT2, and IFN-regulatory factor (IRF) 9. ISGF3

Table 1. PRRs and Their Ligands

PRRs	Localization	Ligand	Origin of the Ligand
TLR			
TLR1	Plasma membrane	Triacyl lipoprotein	Bacteria
TLR2	Plasma membrane	Lipoprotein	Bacteria, viruses, parasites, self
TLR3	Endolysosome	dsRNA	Virus
TLR4	Plasma membrane	LPS	Bacteria, viruses, self
TLR5	Plasma membrane	Flagellin	Bacteria
TLR6	Plasma membrane	Diacyl lipoprotein	Bacteria, viruses
TLR7 (human TLR8)	Endolysosome	ssRNA	Virus, bacteria, self
TLR9	Endolysosome	CpG-DNA	Virus, bacteria, protozoa, self
TLR10	Endolysosome	Unknown	Unknown
TLR11	Plasma membrane	Profilin-like molecule	Protozoa
RLR			
RIG-I	Cytoplasm	Short dsRNA, 5'triphosphate dsRNA	RNA viruses, DNA virus
MDA5	Cytoplasm	Long dsRNA	RNA viruses (Picornaviridae)
LGP2	Cytoplasm	Unknown	RNA viruses
NLR			
NOD1	Cytoplasm	lE-DAP	Bacteria
NOD2	Cytoplasm	MDP	Bacteria
CLR			
Dectin-1	Plasma membrane	β -Glucan	Fungi
Dectin-2	Plasma membrane	β -Glucan	Fungi
MINCLE	Plasma membrane	SAP130	Self, fungi

induces expression of IFN-inducible antiviral genes such as protein kinase R (PKR) and 2'5'-oligoadenylate synthase (OAS) among others. PKR suppresses the proliferation of virus-infected cells and 2'5'-OAS activates RNase L, which cleaves viral nucleotides in order to inhibit virus replication.

In this Review, we discuss how distinct PRRs (with particular emphasis on TLRs and RLRs) sense the presence of pathogens and cellular insults and the mechanisms by which these PRR signals elicit inflammation.

TLRs and Their Ligands

The TLR family is one of the best-characterized PRR families and is responsible for sensing invading pathogens outside of the cell and in intracellular endosomes and lysosomes (Akira et al., 2006). TLRs are characterized by N-terminal leucine-rich repeats (LRRs) and a transmembrane region followed by a cytoplasmic Toll/IL-1R homology (TIR) domain. Ten TLRs have been identified in humans and 12 in mice. Different TLRs recognize the different molecular patterns of microorganisms and self-components (Table 1).

TLR2 senses various components from bacteria, mycoplasma, fungi, and viruses. These components include the lipoproteins of bacteria and mycoplasma. TLR2 recognizes its ligands by forming a heterodimer with either TLR1 or TLR6 (Figure 1). The resulting TLR1/TLR2 and TLR6/TLR2 complexes recognize distinct ligands (triacyl and diacyl lipoproteins, respectively). The crystal structures of the extracellular domains of TLR2, TLR1, and TLR6 revealed that they form M-shaped

structures and that their cognate ligands interact with internal pockets formed by the TLR1/TLR2 or TLR6/TLR2 heterodimers (Jin et al., 2007). Stimulation with TLR2 ligands, such as triacyl and diacyl lipoproteins, induces the production of various proinflammatory cytokines (but not type I IFNs) in macrophages and DCs. However, another report showed that TLR2 in inflammatory monocytes induced type I IFNs in response to viral infection, suggesting that the cellular responses to TLR2 ligands differ depending on the cell types involved (Barbalat et al., 2009). TLR10 is related to TLR1 and TLR6 based on sequence similarity. TLR10 seems to be functional in humans, although mouse TLR10 is disrupted by insertion of an endogenous retrovirus. The ligand for TLR10 has not been identified.

TLR4 recognizes lipopolysaccharide (LPS) together with myeloid differentiation factor 2 (MD2) on the cell surface. LPS is a component derived from the outer membrane of Gram-negative bacteria and is known to be a cause of septic shock. The crystal structure of a complex comprising TLR4, MD2, and LPS revealed that two complexes of TLR4-MD2-LPS interact symmetrically to form a TLR4 homodimer (Park et al., 2009). TLR4 is also involved in the recognition of viruses by binding to viral envelope proteins. In addition, TLR4 modulates the pathogenesis of H5N1 avian influenza virus infection by recognizing a DAMP rather than the virus itself (Imai et al., 2008). Acute lung injury caused by avian influenza virus infection produces endogenous oxidized phospholipids, which stimulate TLR4. Mice lacking TLR4 were found to be resistant to avian flu-induced lethality.

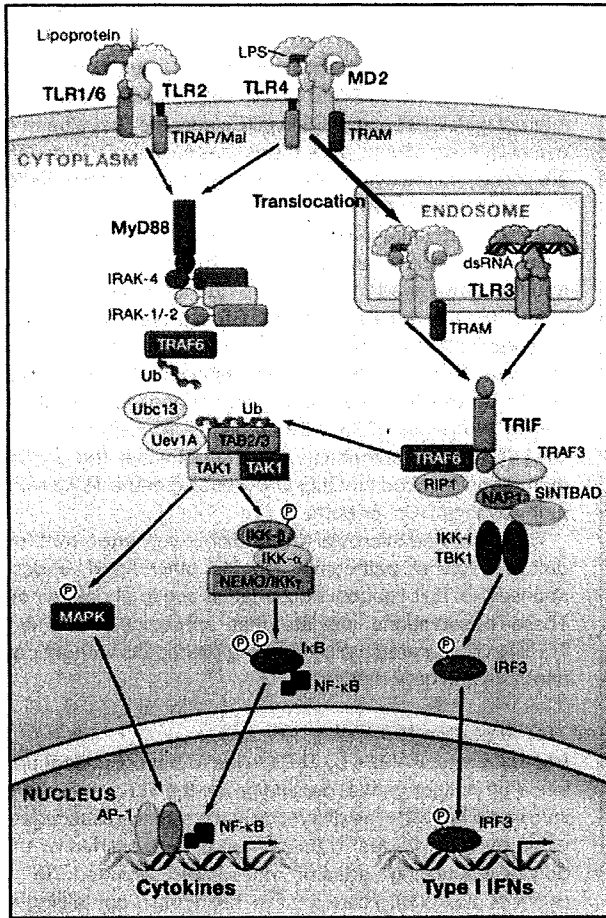


Figure 1. TLR2, TLR3, and TLR4 Signaling Pathways

Lipoproteins and LPS are recognized on the cell surface by a heterodimer of TLR1/6 and TLR2, and by 2 sets of TLR4/MD2 complexes, respectively. Ligand stimulation recruits MyD88 and TIRAP to the TLR, and a complex of IRAKs and TRAF6 is subsequently formed. TRAF6 acts as an E3 ubiquitin ligase and catalyzes formation of a K63-linked polyubiquitin chain on TRAF6 itself and generation of an unconjugated polyubiquitin chain with an E2 ubiquitin ligase complex of Ubc13 and Uev1A. Ubiquitination activates a complex of TAK1, TAB1, and TAB2/3 resulting in the phosphorylation of NEMO and the activation of an IKK complex. Phosphorylated I κ B is degraded, and the freed NF- κ B translocates to the nucleus where it drives expression of cytokine genes. Simultaneously, TAK1 activates MAP kinase cascades leading to the activation of AP-1, which is also critical for the induction of cytokine genes. LPS induces translocation of TLR4 to the endosome together with TRAM. TLR3 is present in the endosome and recognizes dsRNA. TLR3 and TLR4 activate TRIF-dependent signaling, which activates NF- κ B and IRF3 resulting in the induction of proinflammatory cytokine genes and type I IFNs. TRAF6 and RIP1 activate NF- κ B, whereas TRAF3 is responsible for phosphorylation of IRF3 by TBK1/IKK- ϵ . NAP1 and SINTBAD are required for the activation of TBK1/IKK- ϵ . Phosphorylated IRF3 translocates into the nucleus to induce expression of type I IFN genes.

TLR5 is highly expressed by DCs of the lamina propria (LPDCs) in the small intestine, where it recognizes flagellin from flagellated bacteria. In response to flagellin, LPDCs induce B cells to differentiate into IgA-producing plasma cells and trigger the

differentiation of naive T cells into antigen-specific Th17 and Th1 cells (Uematsu et al., 2008). TLR11, which is present in mice but not in humans, shows close homology to TLR5. TLR11 recognizes uropathogenic bacteria and a profilin-like molecule derived from the intracellular protozoan *Toxoplasma gondii* (Yarovinsky et al., 2005).

A set of TLRs, comprising TLR3, TLR7, TLR8, and TLR9, recognize nucleic acids derived from viruses and bacteria, as well as endogenous nucleic acids in pathogenic contexts (Akira et al., 2006). Activation of these TLRs leads to the production of type I IFNs in addition to proinflammatory cytokines. TLR3 detects viral double-stranded (ds) RNA in the endolysosome. TLR3 is involved in the recognition of polyinosinic polycytidylic acid (poly I:C), a synthetic dsRNA analog. Although inoculation of mice with poly I:C induces the production of cytokines as well as type I IFNs in mice, TLR3 is essential for the production of cytokines such as IL-12p40, but not type I IFNs in sera (Kato et al., 2006). The crystal structure of TLR3 bound to dsRNA revealed that dsRNA binds to the N-terminal and C-terminal portions of TLR3 LRRs, and this ligand binding dimerizes two TLR3 molecules (Choe et al., 2005; Liu et al., 2008). Mouse TLR7 and human TLR7/8 recognize single-stranded (ss) RNAs from RNA viruses, as well as small purine analog compounds (imidazoquinolines). TLR7 also detects RNAs from bacteria such as Group B Streptococcus in endolysosomes in conventional DCs (cDCs) (Mancuso et al., 2009). TLR9 senses unmethylated DNA with CpG motifs derived from bacteria and viruses. Although the CpG motif was thought to be essential for TLR9 stimulation, the DNA sugar backbone of 2' deoxyribose also mediates TLR9 recognition (Haas et al., 2008). In addition to DNA, TLR9 also recognizes hemozoin, a crystalline metabolite of hemoglobin produced by the malaria parasite (Coban et al., 2005). TLR9 directly binds to hemozoin, and a crude extract of the malaria parasite elicits parasite-antigen-specific immune responses via TLR9 (Coban et al., 2010). However, another report shows that TLR9 recognizes malaria parasite DNA contained in purified hemozoin, and that hemozoin only transports malaria parasite DNA to the endosome, where TLR9 is present (Parroche et al., 2007). Further studies will clarify the role of TLR9 in the recognition of malaria parasite components. TLR7 and TLR9, but not TLR3, are highly expressed in plasmacytoid DCs (pDCs), a cell type that produces large amounts of type I IFNs in response to virus infection.

Accumulating evidence underscores the importance of the localization of TLRs in the cell for their recognition by ligand (Barton and Kagan, 2009). Given that self-nucleotides are potent TLR ligands and may facilitate autoimmunity, TLRs that recognize self-nucleotides are compartmentalized to avoid unwanted activation. Although TLR1, TLR2, TLR4, TLR5, and TLR6 are present on the plasma membrane, TLR3, TLR7, and TLR9 are mainly present on the endoplasmic reticulum (ER) membrane. It has been proposed that self-nucleic acids are degraded by extracellular or endosomal DNases prior to recognition by TLRs. Nucleic acid-sensing TLRs are recruited from the ER to endolysosomes following stimulation by their ligands (Figure 2). The mechanism by which nucleotide-recognizing TLRs are recruited from the ER to the endolysosome compartment remains to be clarified. However, a forward genetics screen in mice

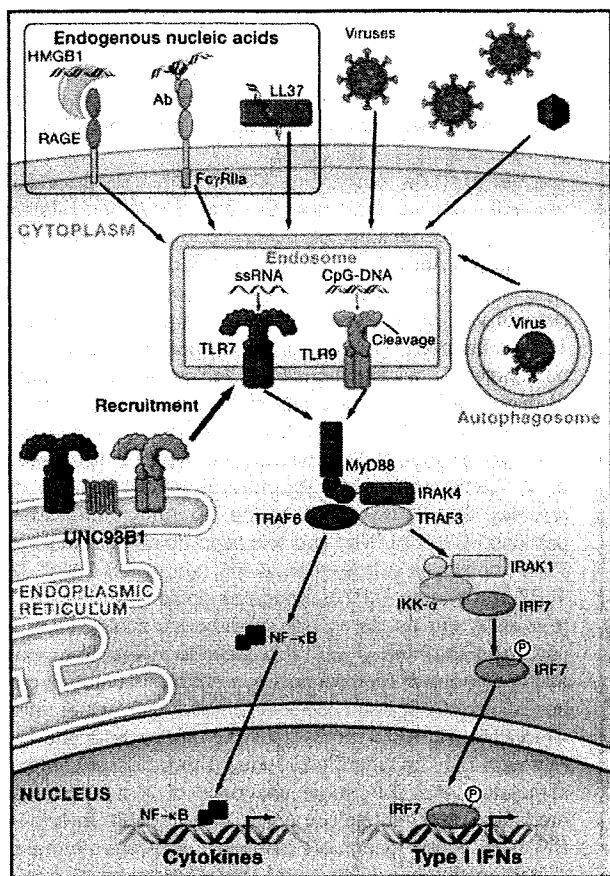


Figure 2. Nucleic Acid Sensing by TLR7 and TLR9

TLR7 and TLR9 recognize viral ssRNA and CpG DNA, respectively. Stimulation with ligands or infection by viruses induces trafficking of TLR7 and TLR9 from the ER to the endolysosome via UNC93B1. TLR9 undergoes cleavage by proteases present in the endolysosome. A complex of MyD88, IRAK-4, TRAF6, TRAF3, IRAK-1, IKK- α , and IRF7 is recruited to the TLR. Phosphorylated IRF7 translocates into the nucleus and upregulates the expression of type I IFN genes. Viruses that have entered the cytoplasm are engulfed by autophagosomes and deliver viral nucleic acids to the endolysosome. An HMGB1-DNA complex released from damaged cells is captured by RAGE. Autoantibodies recognizing self-DNA or -RNA bind to Fc γ RIIIa. LL37, an antimicrobial peptide, associates with endogenous DNA. These proteins are responsible for the delivery of endogenous nucleic acids to endolysosomes where they are recognized by TLR7 or TLR9.

revealed that UNC93B1 (an ER protein with 12 membrane-spanning domains) is responsible for TLR3, TLR7, and TLR9 signaling by governing the translocation of these TLRs from the ER to the endolysosome (Kim et al., 2008; Tabeta et al., 2006). When TLR9 is recruited from the ER to the endolysosome, it undergoes processing by proteases, such as cathepsins, in the endolysosome (Ewald et al., 2008; Park et al., 2008). The processed form of TLR9 is responsible for CpG-DNA recognition. It has been shown that cathepsins B, K, and L and asparagine endopeptidase are required for TLR9 responses (Asagiri et al., 2008; Matsumoto et al., 2008; Sepulveda et al., 2009). Currently, it remains unclear

whether TLR7 is also cleaved in the endolysosome, although endosomal acidification is required for the sensing of TLR7 ligands.

TLR7 and TLR9 are essential for virus-induced type I IFN production by pDCs (Kato et al., 2005). Viral nucleotides can interact with TLR7 and TLR9 in pDCs after they have been endocytosed. Alternatively, once the viruses invade pDCs and virions are present in the cytoplasm, they can be delivered to the endolysosome where TLR7 and TLR9 are recruited for viral sensing. pDCs take advantage of a cellular process called autophagy in which self-proteins and damaged organelles are degraded in double-membraned vesicles called autophagosomes (Lee et al., 2007). In the absence of ATG5, a protein essential for autophagosome formation, pDCs fail to produce type I IFNs in response to virus infection, suggesting that the cytoplasmic virions are engulfed by autophagosomes and then fuse with lysosomes. However, ATG5 is also required for responses to CpG-DNA. Therefore, autophagy may control either the endosomal maturation required for CpG-DNA sensing or the TLR9 signaling pathways in pDCs, or both.

TLR-mediated microbial recognition is very important for host defense against pathogens. On the other hand, excess responses to TLR ligands induce lethal septic shock syndrome. These observations indicate that appropriate activation of TLRs is vital for eradicating invading pathogens without causing harmful damage to the host.

TLR Signaling Pathways

Recognition of PAMPs by TLRs leads to transcriptional upregulation of distinct genes, depending on the TLRs and cell types involved (Figure 1). The difference in the signaling cascades activated by the individual TLRs can be partly explained by the TIR domain-containing adaptor molecules recruited to TLRs (Akira et al., 2006). There are five TIR domain-containing adaptors including MyD88, TIR domain-containing adaptor inducing IFN- β (TRIF; also known as TICAM-1), TIRAP/Mal, TRIF-related adaptor molecule (TRAM), and Sterile- α and Armadillo motif-containing protein (SARM). TLR signaling is roughly divided into two distinct pathways depending on the usage of the distinct adaptor molecules, MyD88 and TRIF.

The MyD88-Dependent Signaling Pathway

MyD88 is composed of a death domain (DD) in addition to a TIR domain. MyD88 is essential for the downstream signaling of various TLRs, with the exception of TLR3. Children with MyD88 deficiency suffer from recurrent pyogenic bacterial infections. TLR2 and TLR4 signaling requires TIRAP/Mal for bridging between TLR and MyD88. MyD88 interacts with IL-1R-associated kinase (IRAK)-4, a serine/threonine kinase with an N-terminal death domain. IRAK-4 activates other IRAK family members, IRAK-1 and IRAK-2 (Kawagoe et al., 2008). The IRAKs then dissociate from MyD88 and interact with TNFR-associated factor 6 (TRAF6), which acts as an E3 ubiquitin protein ligase. Together with an E2 ubiquitin-conjugating enzyme complex comprising Ubc13 and Uev1A, TRAF6 catalyzes the formation of a lysine 63 (K63)-linked polyubiquitin chain on TRAF6 itself as well as the generation of an unconjugated free polyubiquitin chain (Xia et al., 2009). A complex of TGF- β -activated kinase 1 (TAK1), TAK1-binding protein 1 (TAB1), TAB2, and TAB3 is activated by the unconjugated free K63 polyubiquitin chain and

phosphorylates I κ B kinase (IKK)- β and MAP kinase kinase 6. Subsequently, the IKK complex, composed of IKK- α , IKK- β , and NF- κ B essential modulator (NEMO), phosphorylates I κ B α , an NF- κ B inhibitory protein. Phosphorylated I κ B undergoes degradation by the ubiquitin-proteasome system, thereby freeing NF- κ B to translocate into the nucleus and activate expression of proinflammatory cytokine genes. Activation of the MAP kinase cascade is responsible for the formation of another transcription factor complex, AP-1, that targets cytokine genes.

TLR7 and TLR9 signaling induces the production of type I IFNs in addition to other NF- κ B-dependent cytokines in a MyD88-dependent manner. In pDCs, MyD88 forms a complex with IRAK-1, TRAF6, TRAF3, IKK- α , and IRF7, and phosphorylated IRF7 translocates to the nucleus to activate the expression of genes encoding type I IFNs (Figure 2). In cDCs, IRF1, but not IRF7, is activated downstream of TLR7 and TLR9, resulting in the activation of IFN- β gene expression (Negishi et al., 2006; Schmitz et al., 2007).

The TRIF-Dependent Signaling Pathway

In response to stimulation with dsRNA, TLR3 recruits another adaptor protein, TRIF. TLR4 triggers both MyD88-dependent and TRIF-dependent signaling. TLR4, but not TLR3, requires another adaptor, TRAM, for activating TRIF. A splice variant of TRAM called the TRAM adaptor with GOLD domain (TAG) acts as the negative regulator of TRIF-dependent signaling (Palsson-McDermott et al., 2009). TRIF associates with TRAF3 and TRAF6 through TRAF-binding motifs present in its N-terminal portion. TRIF also contains a C-terminal receptor-interacting protein (RIP) homotypic interaction motif (RHIM) and interacts with RIP1 and RIP3 via this motif. In humans, SARM functions as an inhibitor of TRIF-dependent signaling (Carty et al., 2006). The TNFR-associated death domain protein (TRADD), an essential adaptor for TNFR signaling, is involved in the TRIF-dependent signaling pathway (Ermolaeva et al., 2008; Pobeziinskaya et al., 2008). TRADD forms a complex with FAS-associated death domain-containing protein (FADD) and RIP1, and TRADD mediates ubiquitination of RIP1, an event required for NF- κ B activation. FADD activates caspase-8 or caspase-10 in response to poly I:C, and the cleaved form of caspases activates NF- κ B (Takahashi et al., 2006).

TRAF3 is important for activating two IKK-related kinases, TANK-binding kinase 1 (TBK1) and IKK- i (also known as IKK- ϵ) (Hacker et al., 2006; Oganessian et al., 2006). TRAF3 undergoes K63-linked auto-ubiquitination in response to TLR3 and acts as an E3 ubiquitin ligase. TRAF3 activation is negatively regulated by a deubiquitination enzyme DUBA (Kayagaki et al., 2007), and MyD88-dependent signaling triggers K48-linked ubiquitination of TRAF3. Proteasome-mediated degradation of TRAF3 is important for the activation of MAP kinases and the production of proinflammatory cytokines (Tseng et al., 2009). A recent study identified an E2 ubiquitin ligase, Ubc5, as a molecule required for IRF3 activation by catalyzing K63-type polyubiquitin chain formation (Zeng et al., 2009). TBK1 and IKK- i phosphorylate IRF3 and IRF7; IRF3 and IRF7 dimers translocate to the nucleus, resulting in induction of type I IFNs and expression of IFN-inducible genes. IKK- i also phosphorylates STAT1 to facilitate the induction of a set of IFN-inducible genes including *Adar1*, *Irf3*, and *Irf7* (Tenover et al., 2007).

The activation of TBK1 and IKK- i is modulated by various proteins. TBK1 and IKK- i interact with TRAF family member-associated NF- κ B activator (TANK) (also known as I-TRAF), NAK-associated protein 1 (NAP1), and the TBK1 adaptor (SINTBAD), which is similar to NAP1 (Guo and Cheng, 2007; Ryzhakov and Randow, 2007; Sasai et al., 2006). These molecules contain a TBK1-binding motif and show similarities in their coiled-coil domains. However, *TANK*^{-/-} cells do not show impaired type I IFN production in response to dsRNA stimulation (Kawagoe et al., 2009). Although knockdown of either NAP1 or SINTBAD impairs TRIF signaling, the relationship between these molecules in TRIF signaling is not yet fully understood.

RLRs and Virus Recognition

The RIG-I-like receptor (RLR) family is composed of RIG-I, melanoma differentiation-associated gene 5 (MDA5), and LGP2 (Takeuchi and Akira, 2009; Yoneyama and Fujita, 2008). RLRs are composed of two N-terminal caspase recruitment domains (CARDs), a central DEAD box helicase/ATPase domain, and a C-terminal regulatory domain. They are localized in the cytoplasm and recognize the genomic RNA of dsRNA viruses and dsRNA generated as the replication intermediate of ssRNA viruses. The expression of RLRs is greatly enhanced in response to type I IFN stimulation or virus infection.

Mouse fibroblasts and cDCs lacking RIG-I are defective in the production of type I IFNs and inflammatory cytokines in response to various RNA viruses (Kato et al., 2006). These include the Paramyxoviridae such as Newcastle disease virus (NDV) and Sendai virus (SeV), vesicular stomatitis virus (VSV), influenza virus, and Japanese encephalitis virus (JEV). In contrast, cells lacking MDA5 respond normally to these viruses. Meanwhile, the IFN responses to several Picornaviridae, including encephalomyocarditis virus (EMCV), Mengo virus, and Theiler's virus, are abrogated in *MDA5*^{-/-} cells, but not in cells lacking RIG-I. In addition to JEV, another flavivirus, hepatitis C virus (HCV), is also recognized by RIG-I, whereas both RIG-I and MDA5 redundantly recognize Dengue virus and West Nile virus (Loo et al., 2008; Sumpter et al., 2005). A double-stranded segmented RNA virus, reovirus, induces IFN production mainly through MDA5. However, the absence of both RIG-I and MDA5 completely abrogates this IFN production, suggesting that both RIG-I and MDA5 are involved in the recognition of reovirus (Kato et al., 2008; Loo et al., 2008). Mouse embryonic fibroblasts (MEFs) derived from *RIG-I*^{-/-}*MDA5*^{-/-} mice failed to produce type I IFNs to any of the RNA viruses tested, indicating that RIG-I and MDA5 are essential and sufficient for evoking type I IFN production in response to RNA viruses.

RIG-I recognizes relatively short dsRNA (up to 1 kb), and the presence of a 5' triphosphate end greatly enhances its type I IFN-inducing activity (Figure 3). It has been postulated that 5' triphosphate ssRNA synthesized by in vitro transcription is a RIG-I ligand (Hornung et al., 2006; Pichlmair et al., 2006). However, recent studies have shown that T7 polymerase commonly produces extended byproducts generating dsRNA (Schlee et al., 2009; Schmidt et al., 2009). Chemically synthesized 5' triphosphate ssRNA failed to stimulate RIG-I, indicating that RNA needs to be double-stranded for activation of RIG-I. Regarding a minimum length, the 19-mer or 21-mer dsRNA

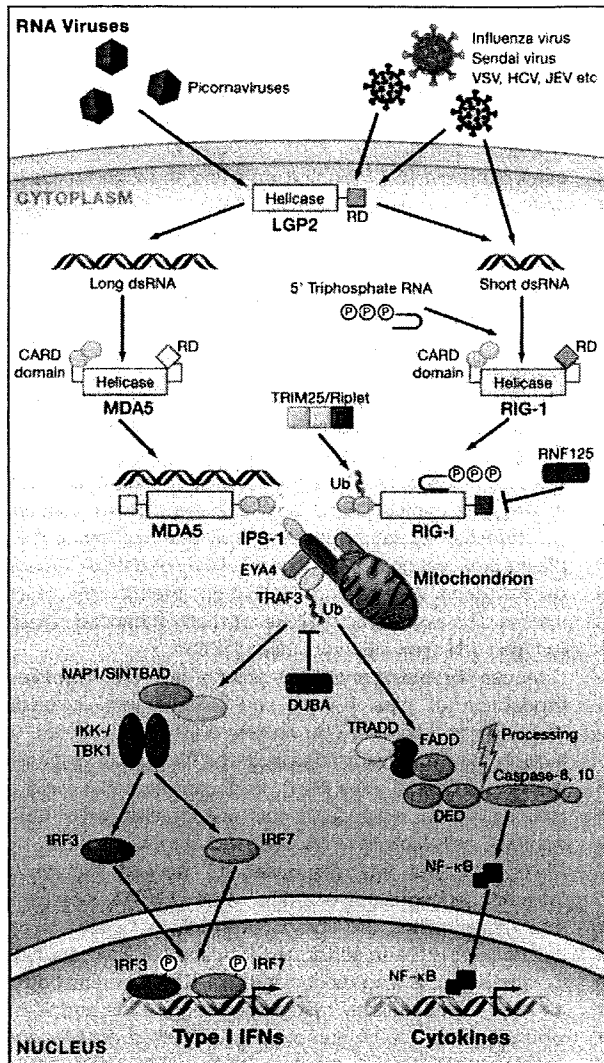


Figure 3. Recognition of RNA Viruses by RLRs

RIG-I and MDA5 recognize different RNA viruses by detecting short dsRNAs with 5' triphosphate ends and long dsRNAs, respectively. LGP2 functions as a positive regulator in RIG-I-mediated and MDA5-mediated virus recognition. Activation of RIG-I is positively and negatively regulated by ubiquitin ligases TRIM25 and RNF125, respectively. RIG-I and MDA5 interact with IPS-1 through homophilic interactions between CARD domains. IPS-1 then activates signaling cascades leading to the expression of type I IFN genes via EYA4, TRAF3, NAP1/SINTBAD, TBK1/IKK-1, and IRF3/7. RLR signaling mediates polyubiquitination of TRAF3, which is removed by a deubiquitinase, DUBA. Simultaneously, IPS-1 signaling induces nuclear translocation of NF-κB via TRADD and FADD and caspase-8/-10. A cleaved fragment of caspase-8/-10 is responsible for the activation of NF-κB.

with a 5' triphosphate end is able to potently induce type I IFNs. In contrast, a 5' triphosphate end is not always necessary, as chemically synthesized dsRNAs with a 5' monophosphate end or those without a 5' phosphate can potently activate RIG-I (Takahashi et al., 2008; Kato et al., 2008). The amount of IFNs produced by these RNAs is low compared to dsRNA with a 5'

triphosphate end. HCV genomic RNA has been screened for RNA sequences that activate RIG-I. Poly(U)- or poly(A)-rich sequences from the HCV RNA 3' untranslated region (UTR) are responsible for RIG-I-mediated IFN production, based on RNAs generated by in vitro transcription using T7 polymerase (Saito et al., 2008). However, in vitro-transcribed RNAs with low U/A content also potently activate RIG-I. Therefore, further studies are required to clarify whether particular RNA sequences are important for RIG-I-mediated recognition.

VSV produces dsRNA in infected cells (Kato et al., 2008). Disruption of dsRNA among RNAs from VSV-infected cells reduces the IFN-β-inducing activity, suggesting that the presence of dsRNA in VSV-infected cells is important for recognition by RIG-I. Interestingly, the dsRNA fragments produced by VSV infection are about 2.0–2.5 kb, and much shorter than the size of the VSV genomic RNA. It has been reported that defective interfering (DI) particles are generated in VSV-infected cells, and that the sizes of DI particle dsRNAs are about 2.2 kb (Pattnaik et al., 1995). Thus, the dsRNA generated during the course of VSV replication may be derived from DI particles, although further studies are needed to clarify the source of this dsRNA. As DI particles are known to strongly induce type I IFNs, RIG-I may play a role in detecting the presence of dsRNA in DI particles. In contrast, it is difficult to detect dsRNA in cells infected with influenza virus. Genomic RNA from influenza virus lost its IFN-inducing activity after a phosphatase treatment that removed the 5' triphosphate end. The sequences of the 5' and 3' ends of the viral RNA are partially complementary to each other, and it has been suggested that a panhandle structure can form (Hsu et al., 1987). Thus, it is tempting to speculate that short panhandle dsRNA with a 5' triphosphate is responsible for recognition by RIG-I.

In contrast to RIG-I, MDA5 detects long dsRNA (more than 2 kb) such as poly I:C. *MDA5*^{-/-} mice show severely reduced production of type I IFNs in response to poly I:C inoculation in vivo, whereas their production of IL-12p40 is not impaired (Kato et al., 2006). Shortening the length of the poly I:C by treatment with a dsRNA-specific nuclease converts poly I:C from an MDA5 ligand to a RIG-I ligand, indicating that long, but not short, dsRNA is recognized by MDA5. EMCV produces high levels of dsRNA in infected cells, and the 5' end of its genomic RNA is covalently linked to a small protein, VPg. Investigation of RNA from EMCV-stimulated cells has revealed that higher-order structure RNA containing both dsRNA and ssRNA, but not simple dsRNA as the replication intermediate, has MDA5-stimulating activity (Pichlmair et al., 2009).

LGP2, the third member of the RLR family, lacks a CARD, and in vitro studies have suggested that LGP2 functions as a negative regulator of RIG-I and MDA5 responses by sequestering dsRNA or inhibiting RIG-I conformational changes (Rothenfusser et al., 2005; Saito et al., 2007; Yoneyama et al., 2005). However, the generation of *LGP2*^{-/-} mice and mice with a point mutation, D30A, that disrupts the ATPase activity of LGP2 revealed that LGP2 positively regulates production of type I IFNs in response to RNA viruses recognized by both RIG-I and MDA5 (Sato et al., 2010). Nevertheless, LGP2 is dispensable for type I IFN production following transfection by synthetic RNAs. These results suggest that LPS2 may modify viral RNA by removing

proteins from viral ribonucleoprotein (RNP) complexes or unwinding complex RNA structures to facilitate MDA5-mediated and RIG-I-mediated recognition of dsRNA.

RLRs contain a C-terminal regulatory domain, which is responsible for the binding to dsRNAs. The recent solution of RLR C-terminal regulatory domain structures revealed that the C-terminal domains of LGP2 and RIG-I have a large basic surface forming an RNA-binding loop (Cui et al., 2008; Takahashi et al., 2008, 2009). The RIG-I and LGP2 C-terminal domains bind to the termini of dsRNA. Although the MDA5 C-terminal domain also has a large basic surface, it is extensively flat because of the open conformation of the RNA-binding loop. Therefore, the RNA-binding activity of MDA5 is much weaker than that of RIG-I and LGP2. RLRs catalyze ATP via the DExD/H helicase domain, and the ATPase activity is essential for RLRs to induce type I IFN production. Although RIG-I has helicase activity, it is not clear if the unwinding of dsRNA by RIG-I is required for triggering the signaling pathway. RIG-I might change its conformation to expose the CARDs and dimerize by catalyzing ATP, or alternatively, RIG-I may act as a translocase for dsRNA.

The RLR Signaling Pathways

The RIG-I conformation is known to be modulated by ubiquitination. TRIM25 and Riplet (also known as RNF135) act as E3 ubiquitin ligases that mediate the K63-linked polyubiquitination of RIG-I (Gack et al., 2007; Pichlmair et al., 2009). This modification is required for the activation of RLR signaling. On the other hand, K48-type polyubiquitination of RIG-I by RNF125 leads to RIG-I degradation by the proteasome and inhibition of RIG-I signaling (Arimoto et al., 2007).

The CARDs of RLRs are responsible for triggering signaling cascades by interacting with the N-terminal CARD-containing adaptor IFN- β -promoter stimulator 1 (IPS-1) (also known as MAVS, CARDIF, or VISA) (Kawai and Akira, 2006) (Figure 3). IPS-1 is localized on the mitochondrial membrane, and cleavage of IPS-1 by an HCV NS3/4A protease, which dislodges it from the mitochondrial membrane, results in abrogation of RLR signaling. NLRX1 (also known as NOD9), an NLR family member, is localized on the mitochondrial membrane and acts as an inhibitor of IPS-1 signaling (Moore et al., 2008). However, another report showed that NLRX1 is a mitochondrial matrix protein responsible for the generation of reactive oxygen species. Therefore, further studies are required to clarify the role of NLRX1 in RLR signaling (Arnoult et al., 2009). IPS-1 activates TRAF3 and TRADD (Michallet et al., 2008). The downstream signaling molecules for the expression of IFN-inducible genes are shared between the IPS-1 and TRIF signaling pathways. The recently identified protein Eyes absent 4 (EYA4) associates with IPS-1 and NLRX1 and stimulates the expression of IFN-inducible genes in response to DNA stimulation (Okabe et al., 2009). EYA4 is a phosphatase for both phosphotyrosine and phosphothreonine, and the threonine phosphatase activity is essential for enhancing IFN- β gene activation. Identification of molecules that are dephosphorylated by EYA4 will be important for understanding the mechanism for how EYA4 regulates IFN responses.

In contrast to pDCs, autophagy negatively regulates IFN responses in fibroblasts and cDCs by suppressing RLR signaling (Jounai et al., 2007; Tal et al., 2009). In the absence of

autophagy-related proteins, such as ATG5 or ATG16L1, damaged mitochondria accumulate together with IPS-1, and reactive oxygen species associated with dysfunctional mitochondria are responsible for the overproduction of type I IFNs.

Recognition of Cytoplasmic DNA

Although DNA with a CpG motif is sensed by TLR9, introduction of dsDNA into cells evokes type I IFN responses in a TLR9-independent manner. Right-handed spiral B-DNA, but not left-handed Z-DNA, activates these responses (Ishii et al., 2006). Although the sequence specificity has not been clearly observed, synthetic poly (dA:dT) is more potent for induction of IFN responses than poly (dI:dC) or poly (dG:dC). Transfection of dsDNA leads to the activation of IRF3 via TBK1 and IKK- α , but activation of NF- κ B is barely detected in response to DNA. It is believed that cytoplasmic DNA recognition is important for the production of type I IFNs to infection with DNA viruses. Infection with intracellular bacteria such as *Listeria monocytogenes* and *Legionella pneumophila* induces type I IFN production in response to bacterial DNA in the cytoplasm (Stetson and Medzhitov, 2006). In addition, the adjuvant effect of DNA vaccines can be explained by TLR9-independent cytoplasmic recognition of DNAs that activates the TBK1-IRF3-dependent pathway (Ishii et al., 2008).

Two studies have shown that poly (dA:dT) can be transcribed into dsRNA by polymerase III and that dsRNA is recognized in a RIG-I-IPS-1-dependent manner in human cells or transformed mouse MEFs (Figure 4) (Ablasser et al., 2009; Chiu et al., 2009; Choi et al., 2009). However, primary DCs prepared from RIG-I-deficient mice do not show a defect in IFN production, suggesting that cytoplasmic DNAs are sensed in both a polymerase III-RIG-I-dependent and -independent manner depending on the species or cell types involved. A cytoplasmic DNA-binding protein, DNA-dependent activator of IRF (DAI, also known as Zbp1 or DLM1), interacts with TBK1 and activates type I IFN responses (Takaoka et al., 2007). Nevertheless, *Zbp1*^{-/-} MEFs are still capable of producing type I IFNs in response to cytoplasmic DNA, suggesting that cytoplasmic DNA is redundantly recognized by as-yet unidentified receptors (Ishii et al., 2008).

Expression cloning of a gene inducing IFN- β promoter activation identified STING (also known as MITA, ERIS, or TMEM173), a transmembrane protein expressed in the ER (Ishikawa and Barber, 2008; Sun et al., 2009; Zhong et al., 2008). Cells and mice lacking *STING* show impaired IFN production in response to both RNA and DNA stimulation. STING associates with a member of the translocon-associated protein (TRAP) complex that is required for protein translocation across the ER in resting cells. In response to transfection of dsDNA, STING translocates from the ER to the Golgi apparatus and subsequently to cytoplasmic punctate structures where it colocalizes with TBK1 (Ishikawa et al., 2009). The exocyst complex is involved in secretory vesicle sorting and is especially required for post-Golgi transport to the plasma membrane. Sec5, a component of the exocyst complex, colocalizes with STING. Activation of the RaiB GTPase promotes the assembly of TBK1 and Sec5, leading to phosphorylation of Sec5 by TBK1. *STING*^{-/-} mice are highly susceptible to HSV-1 or VSV infection, indicating a role for STING in host defense against virus infection. Interestingly,

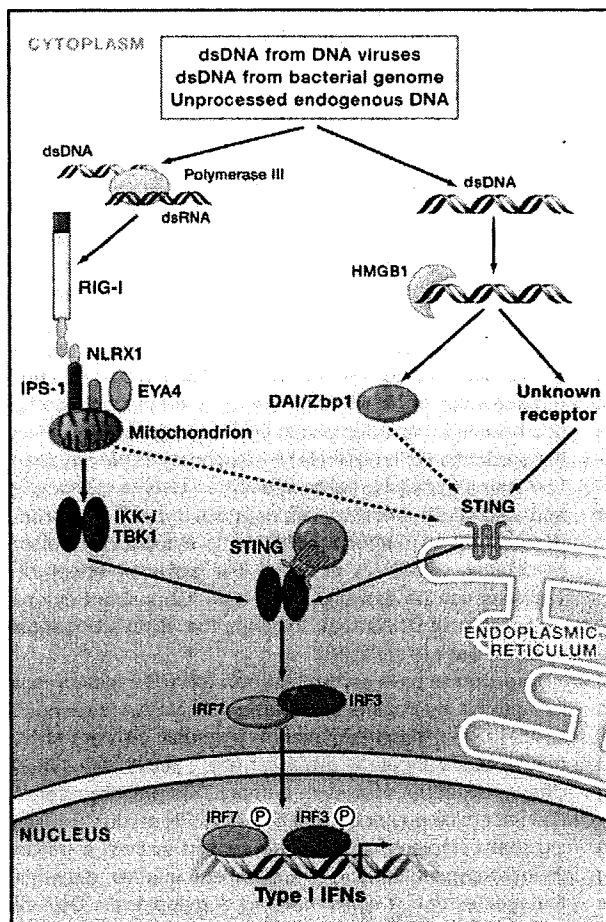


Figure 4. Recognition of Cytoplasmic DNA and IFN Production
 Double-stranded DNA (dsDNA) accumulates in the cytoplasm after infection by viruses or bacteria or through failure to process endogenous DNA. The intracellular DNA is captured by HMGB1 and recognized by an unidentified cytoplasmic receptor or DAI. Alternatively, dsDNA is transcribed into dsRNA by polymerase III in a species- and cell type-specific manner. Generated dsRNA is recognized by RIG-I and production of type I IFNs is induced. Following DNA stimulation, an ER protein STING translocates from the ER to the cytoplasmic punctate structure, where STING colocalizes with TBK1.

autophagy-related gene 9a (ATG9a) colocalizes with STING (Saitoh et al., 2009). Loss of ATG9a, but not another autophagy-related gene (ATG7), greatly enhances the assembly of STING with TBK1 in response to dsDNA, followed by increased production of type I IFNs. Notably, the introduction of dsRNA did not induce the translocation of STING; it will be interesting to explore how STING regulates the IFN response to dsRNA. It has been shown that TBK1 activation by RaIb is involved in cell transformation and tumor progression (Chien et al., 2006). Furthermore, a recent study revealed that TBK1 is required for KRAS-driven cancer formation, probably by activating the NF- κ B-induced antiapoptotic gene *BCL-xL* (Barbie et al., 2009). Thus, TBK1 is a pleiotropic kinase involved in both innate immunity and tumorigenesis.

High-mobility group box (HMGB) proteins were originally identified as nuclear proteins with DNA-binding capacity. HMGB1 is also known to be secreted in response to cell damage and evokes inflammatory responses. Recently, HMGB proteins present in the cytoplasm were found to act as the initial sensors for cytoplasmic nucleic acids that lead to the activation of downstream receptors such as RLRs, TLRs, and unknown DNA receptors (Yanai et al., 2009).

Although stimulation with TLR ligands alone does not lead to activation of the inflammasome, the introduction of dsDNA induces not only the production of pro-IL- β but also its cleavage via caspase-1. Sensing of cytoplasmic DNA by absent-in-melanoma 2 (AIM2) activates the inflammasome via ASC and caspase-1, leading to the production of IL-1 β (see Review by K. Schroder and J. Tschopp on page 821).

NLR- and CLR-Mediated Pathogen Recognition

The NLR family consists of cytoplasmic pathogen sensors that are composed of a central nucleotide-binding domain and C-terminal leucine-rich repeats (Inohara et al., 2005). The N-terminal portions of most NLRs harbor protein-binding motifs, such as CARDs, a pyrin domain, and a baculovirus inhibitor of apoptosis protein repeat (BIR) domain. NLRs harboring a pyrin domain or a BIR domain in their N terminus are not involved in the transcriptional activation of inflammatory mediators and are components of the inflammasome that regulates caspase-1 activation. NOD1 and NOD2, which harbor CARDs in addition to NOD and LRR domains, activate NF- κ B via an adaptor, RIP2/RICK. NOD1 and NOD2 induce transcriptional upregulation of proinflammatory cytokine genes. NOD1 and NOD2 recognize the structures of bacterial peptidoglycans, g-D-glutamyl-meso-diaminopimelic acid (iE-DAP) and muramyl dipeptide (MDP), respectively. As TLRs also recognize bacterial peptidoglycan components, TLRs and NODs synergistically activate proinflammatory cytokine production. In addition to bacteria, a recent study found that the expression of NOD2 is involved in 5'-triphosphate RNA-induced type I IFN production and host defense against respiratory syncytial virus infection (Sabbah et al., 2009).

CLRs comprise a transmembrane receptor family characterized by the presence of a carbohydrate-binding domain. CLRs recognize carbohydrates on microorganisms such as viruses, bacteria, and fungi. CLRs either stimulate the production of proinflammatory cytokines or inhibit TLR-mediated immune complexes. The functions of CLRs are described in detail elsewhere (Geijtenbeek and Gringhuis, 2009), and here we give just a few examples of CLR-mediated microbial recognition. Dectin-1 and dectin-2 are immunoreceptor tyrosine-based activation motif (ITAM)-coupled CLRs responsible for sensing β -glucans from fungi. DCs activated by dectin-1 or dectin-2 are able to instruct T cells to confer protective immunity against *Candida albicans* (Robinson et al., 2009). The macrophage C-type lectin MINCLE (also known as Clec4e and Clec5f9) senses infection by fungi such as *Malassezia* and *Candida* (Yamasaki et al., 2009). In addition, MINCLE is responsible for the detection of an endogenous protein, spliceosome-associated protein 130 (SAP130), which is a component of U2 snRNP from necrotic host cells (Yamasaki et al., 2008). CLEC9A is

expressed on CD8⁺ DCs in the spleen and also recognizes necrotic cells for cross-priming. CLRs activate intracellular signaling either via the ITAM domain of CLRs or via adaptors harboring an ITAM domain, such as FcR γ , DAP10, or DAP12. The Syk tyrosine kinase is activated by ITAM-containing proteins either directly or indirectly via ITAM-containing adaptors. Engagement of Syk with CLRs activates MAP kinases, the transcription factor NF-AT, and NF- κ B through CARD9. As a result, proinflammatory cytokines are produced. However, the signaling mechanism for the production of proinflammatory cytokines is not well understood.

Transcriptional Regulation of Inflammatory Mediators

Activation of PRR signaling pathways leads to the nuclear translocation of a set of transcription factors, including NF- κ B, AP-1, IRFs, and C/EBP β . These factors cooperatively regulate the transcription of their target genes. Furthermore, remodeling of chromatin is important for controlling the transcriptional regulation of a set of TLR-inducible genes (Ramirez-Carrozzi et al., 2006). In addition, epigenetic controls of a set of TLR-inducible genes are critical for induction of tolerance to LPS, a well-known phenomenon rendering cells refractory to a second exposure to LPS (Foster et al., 2007) (see Review by S.T. Smale on page 833 of this issue). Here, we focus on the TLR-inducible proteins that modulate inflammatory responses. Besides cytokines, chemokines, and IFNs, TLR stimulation upregulates the expression of hundreds of genes in macrophages. These genes are potentially involved in antimicrobial defense (e.g., defensins, lipocaline), metabolic changes, and tissue repair (e.g., secretory leukocyte peptidase inhibitor). Nevertheless, the functions of various TLR-inducible genes remain largely unexplored. Some gene products are associated with the positive and negative regulation of inflammatory responses by controlling TLR-signaling pathways. The genes immediately induced by TLR ligands potentially regulate TLR signaling or responses after their expression.

It is well known that the components of NF- κ B are TLR-induced genes that undergo positive feedback regulation by NF- κ B (Figure 5). In addition, I κ B ζ and activating transcription factor 3 (ATF3) are nuclear factors that are rapidly induced by the TLRs (Gilchrist et al., 2006; Yamamoto et al., 2004). I κ B ζ harbors C-terminal ankyrin repeat motifs that bind to the NF- κ Bp50 subunit, and expressed I κ B ζ positively regulates the induction of a set of genes including IL-6 and IL-12p40, which are transcribed later. In contrast, ATF3 negatively regulates the transcription of genes encoding IL-6 and IL-12p40 by increasing histone deacetylase activity. Epigenetic changes can be regulated by TLR-inducible proteins. Trimethyl-histone 3 lysine 27 demethylase, Jmjd3, is expressed in response to TLR stimulation via NF- κ B and is recruited to the transcription start sites of LPS-inducible genes (De Santa et al., 2007). Given that Jmjd3 is recruited to the transcription start site of various LPS-inducible genes, it may be responsible for the fine-tuning of LPS-induced gene expression in macrophages (De Santa et al., 2009).

In response to TLR ligand stimulation, genes encoding RNA-binding proteins with a CCCH-type zinc finger motif (Zf) are rapidly induced. These proteins include Zc3h12a, Zc3h12c, Zc3hav1, and tristetraprolin (TTP; also known as Zfp36)

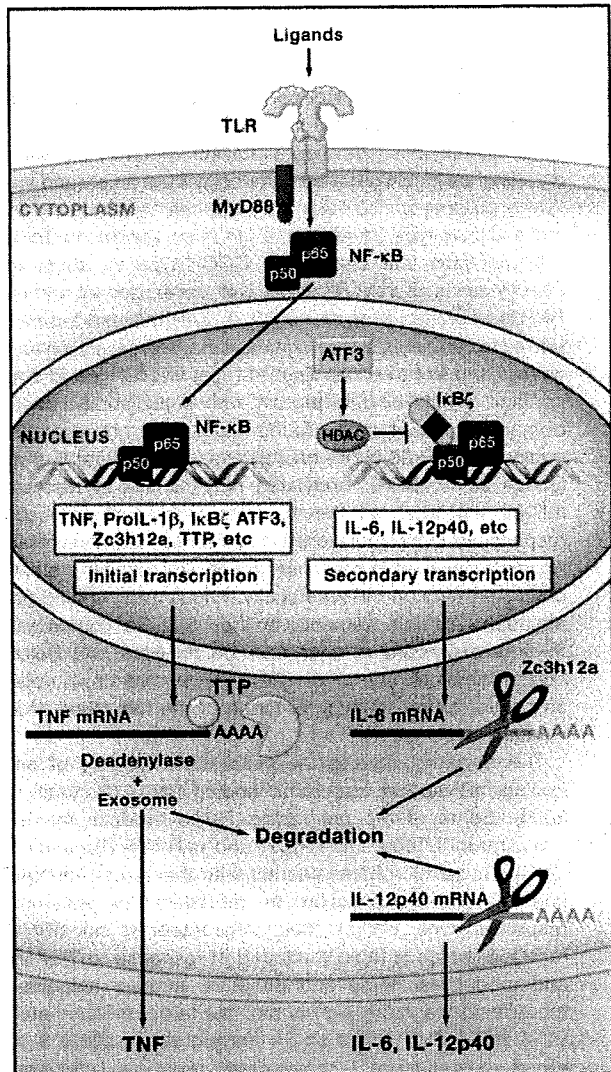


Figure 5. TLR-Inducible Proteins Regulate Inflammation

TLR signaling initially induces genes encoding TNF, Pro-IL-1 β , I κ B ζ , ATF3, Zc3h12a, TTP, and so on. Proteins generated from these genes modulate inflammatory protein expression by various mechanisms. I κ B ζ positively regulates transcription of genes including IL-6 and IL-12p40 by associating with NF- κ Bp50. In contrast, ATF3 negatively regulates transcription of these genes by activating HDAC, which suppresses gene expression by epigenetic regulation. Zc3h12a acts as an RNase degrading IL-6 and IL-12p40 mRNA, whereas TTP binds to TNF mRNA and recruits a deadenylase, which induces its degradation through an exosome.

(Matsushita et al., 2009). TTP harbors two CCCH-type Zf domains and associates with mRNA via AU-rich elements present in the 3'UTR, leading to removal of the poly(A) tail by recruitment of a deadenylase (Carrick et al., 2004). The deadenylated mRNAs traffic to exosomes where the RNA is degraded. TTP is required to prevent the generation of autoimmune arthritis in mice through controlling mRNAs encoding TNF. Zc3h12a and Zc3h12c are composed of a CCCH-type Zf domain and an

RNase domain (Matsushita et al., 2009). Zc3h12a is responsible for degrading mRNAs for TLR-inducible proteins such as IL-6 and IL-12p40 in macrophages. Zc3h12a controls its target mRNAs via the 3'UTR independently of AU-rich elements; the RNase activity is essential for mRNA degradation. *Zc3h12a*^{-/-} mice spontaneously develop a fatal inflammatory disease characterized by highly elevated immunoglobulin levels and autoantibody production. RLR signaling induces another gene encoding a CCCH-type Zf protein, ZAP (also known as Zc3hav1). ZAP contains four clusters of CCCH-type Zf domains and directly binds to viral RNAs as well as exosomes where viral RNA is degraded (Gao et al., 2002). ZAP overexpression is reported to confer resistance against various viruses such as retroviruses and alphaviruses. Zcchc11 is a CCHC-type Zf domain-containing RNA-binding protein with a nucleotidyltransferase domain. A recent study found that Zcchc11 adds terminal uridines to miR-26 family microRNAs, which target IL-6 mRNA (Jones et al., 2009). Uridylated miR-26 fails to repress IL-6 mRNA, and Zcchc11 thereby potentiates IL-6 production in response to TNF stimulation. It was reported that a set of mRNAs rapidly expressed in response to TNF stimulation are critically controlled through mRNA stability. These mRNAs tend to have abundant AU-rich elements in their 3'UTRs compared with mRNAs expressed at later time points (Hao and Baltimore, 2009). Therefore, control of mRNA decay may be as important as control of transcription in terms of the regulation of innate immune responses.

TLR signaling induces the expression not only of protein-coding mRNAs but also of noncoding RNAs (Guttman et al., 2009). Some of the noncoding RNAs produce microRNAs including miR-146a/b, miR-147, and miR-155 (Taganov et al., 2006). These microRNAs interact with their target mRNAs and fine-tune their expression to modulate the inflammatory response. One of the best characterized microRNAs is miR-155, which is rapidly induced in response to TLR ligands and modulates innate and adaptive immune responses by modulating the expression of multiple target mRNAs encoding PU.1, IKK- β , SHIP1, and so on (Faraoni et al., 2009). A recent report shows that miR-21 negatively regulates TLR4 signaling by the tumor suppressor PDCD4, a protein required for NF- κ B activation (Sheedy et al., 2009).

PRR-Dependent Recognition of Self-Nucleotides in Autoimmunity

Under physiological conditions, PRRs strictly discriminate self and microbial components to prevent the development of autoimmune disease. A hallmark of autoimmune disease is the production of antibodies recognizing self-antigens. In human autoimmune diseases such as systemic lupus erythematosus (SLE), immune complexes are formed by autoantibodies and deposited in the kidneys, causing glomerular nephritis. It has been shown that serum type I IFN levels in SLE patients positively correlate with disease severity.

TLRs, especially TLR7 and TLR9, have been implicated in the production of autoantibodies in various mouse models. Self-nucleic acids released from damaged cells are rapidly degraded by serum nucleases and do not encounter TLR7 or TLR9. However, when self-nucleic acids interact with cellular proteins

such as HMGB1, RNPs, antimicrobial peptides, and autoantibodies, endocytosis of the resulting nucleic acid-protein complexes is facilitated and induces TLR7- and TLR9-mediated type I IFN production (Figure 2). For instance, HMGB1 interacts with the receptor for advanced glycation end-products (RAGE) on the cell surface, and HMGB1-DNA-containing immune complexes released from damaged cells stimulate TLR9 and RAGE, which cooperate to stimulate pDCs and B cells (Tian et al., 2007). Endogenous RNA-protein complexes, such as U1 snRNP, can activate autoreactive B cells and DCs via TLR7 (Vollmer et al., 2005). Furthermore, nucleic acids bound to autoantibodies are endocytosed through B cell receptors and Fc γ RIIA in B cells and DCs (Means et al., 2005; Viglianti et al., 2003). Recognition of immune complexes by these receptors leads to the activation of autoreactive B cells as well as the production of type I IFNs, thus exacerbating the causes of autoimmune diseases. Similarly, the cationic antimicrobial peptide LL37 forms a complex with self-DNA or self-RNA, thereby facilitating endocytosis by pDCs and stimulating TLR-mediated IFN responses (Ganguly et al., 2009; Lande et al., 2007). LL37 is highly expressed in psoriasis skin lesions, and the production of IFNs contributes to the pathogenesis of this disease. Taken together these data show that the responses of nucleic acid-sensing TLRs are positively regulated by endogenous proteins that facilitate endocytosis of nucleic acids. Type I IFNs and cytokines produced by the TLRs cause inflammation, and this positive feedback mechanism is involved in the exacerbation of autoimmune diseases.

The contributions of nucleic acid-sensing TLRs to autoimmunity have been examined using mouse models that develop autoimmune disease spontaneously. Duplication of the *Tlr7* gene accounts for the autoimmune phenotypes associated with Y chromosome-linked autoimmune accelerator (Yaa) mice (Pisitkun et al., 2006; Subramanian et al., 2006). Reciprocally, TLR7 deficiency in lupus-prone MRL *lpr/lpr* mice results in reduced levels of autoantibodies recognizing RNA-containing antigens and abrogation of disease (Christensen et al., 2006). But the relationship between TLR9 and autoimmune disease mouse models is complicated. Although CpG-DNA has been used as an adjuvant to generate organ-specific autoimmune disease in mouse models, the lack of TLR9 exacerbated the severity of disease in the MRL *lpr/lpr* murine lupus model. Although the TLR7 and TLR9 signaling pathways are indistinguishable, there is a clear difference in the roles of TLR7 and TLR9 in the establishment of systemic autoimmune disease. A mutation of amino acid D34 in UNC93B1 that changes its ability to bind to TLR7 and TLR9 has been identified (Fukui et al., 2009). The mutant UNC93B1 increases TLR7 responses but causes hyporesponsiveness to TLR9. Given that TLR7 is important for the generation of autoimmune disease models in mice, it is possible that UNC93B1 suppresses RNA sensing by skewing the response toward DNA.

Appropriate clearance of nucleic acids is essential for preventing autoimmune disease. DNase II is located in the lysosomes of macrophages and is required for the degradation of DNA from apoptotic cells. In the absence of DNase II, undigested DNA accumulates in macrophages, and the resulting production of IFN- β and TNF causes embryonic lethality (Kawane et al.,

2003). The type I IFNs are produced independently of TLR9, suggesting that recognition of DNA by intracellular receptors is responsible for the embryonic lethality of DNase II-deficient mice (Okabe et al., 2005). Similarly, deficiency in DNase I, a major DNase present in serum, results in the generation of SLE-like autoimmune disease in mice; a mutation in DNase I has been identified in human SLE patients (Lee-Kirsch et al., 2007). Aicardi-Goutières syndrome (AGS) is a human disorder characterized by fatal encephalopathy owing to overproduction of IFN- α . Recent studies have revealed that AGS is caused by a mutation in 3' repair exonuclease 1 (Trex1) (Crow et al., 2006). It has been shown that DNA derived from endogenous retroelements accumulates in *Trex1*^{-/-} cells and stimulates the induction of IFN-inducible genes (Stetson et al., 2008). In addition, mutations in components of RNase H2 are also involved in AGS (Rice et al., 2009). Nucleic acid accumulation because of a deficiency in these nucleases probably leads to recognition of the nucleic acids by cytoplasmic PRRs resulting in the production of type I IFNs. Interestingly, mutations in MDA5 that disrupt its signaling ability correlate with resistance to type I diabetes, although the mechanism for the contribution of MDA5 to this disease remains unclear (Nejentsev et al., 2009). Future studies will clarify how cytoplasmic PRRs contribute to the generation of autoimmune disease.

PRR Activation in Acute and Chronic Inflammation

PRRs recognizing non-nucleic acid ligands are also involved in various inflammatory and autoimmune diseases. It is well known that activation of TLR signaling by bacterial components leads to acute inflammation, culminating in septic shock in some cases. TLR ligands also are commonly used as adjuvants to generate organ-specific autoimmune disease models in mice for arthritis or encephalitis, for example.

A mutation in NOD2 that generates a truncated form of the protein is frequently found in patients with Crohn's disease, an inflammatory bowel disorder (Cho, 2008). This disease may arise through impairment of proper responses to intestinal bacteria and aberrant bacterial growth causing increased inflammatory responses in the intestine. Mutations in the autophagy-related gene ATG16L1 are also implicated in susceptibility to Crohn's disease. In the absence of ATG16L1 in mice, overactivation of the inflammasome occurs, and increased production of IL-1 β and IL-18 contributes to inflammation in the intestine (Saitoh et al., 2008). Furthermore, ATG16L1 is critical for the normal function of intestinal Paneth cells (Cadwell et al., 2008).

Lack of negative regulators for TLRs can cause autoimmune disease based on various mouse models. Loss of the protein tyrosine phosphatase SHP1 in mice induced by N-ethyl-N-nitrosourea (ENU) mutagenesis leads to the generation of inflammatory lesions together with overactivation of macrophages in response to TLR stimulation (Croker et al., 2008). The inflammation is suppressed by loss of MyD88, suggesting that microbe-induced hyperactivation of TLR signaling is responsible for the generation of this inflammatory disease. A20 is a protein with dual enzyme domains, comprising an E3 ubiquitin ligase and a deubiquitinase that removes K63-linked polyubiquitin chains. A20 negatively regulates NF- κ B activation downstream of TLR2 and NOD1/NOD2 (Boone et al., 2004; Hitotsumatsu

et al., 2008). In the absence of A20, mice develop multiorgan inflammatory disorders that lead to premature death. Another example of a TLR negative regulator is TANK. Generation of mice lacking TANK revealed that TANK is critical for controlling TLR signaling in macrophages as well as antigen receptor signaling in B cells, by inhibiting the activation of TRAF6 (Kawagoe et al., 2009). *TANK*^{-/-} mice spontaneously develop autoimmune glomerular nephritis depending on the presence of MyD88 or IL-6. These results suggest that aberrant activation of innate immune cells and B cells is responsible for the generation of autoimmune diseases resembling human SLE. These studies indicate that negative regulation of PRRs, especially TLR signaling, is important for coordinated innate immune responses.

It is well known that TLR-dependent inflammation contributes significantly to the pathogenesis of ischemia-reperfusion myocardial injury. Mice lacking TLR2, TLR4, or MyD88 show reduced infarcted regions in response to cardiac as well as cerebral ischemic-reperfusion injury (Arumugam et al., 2009). In contrast, pretreatment of mice with TLR ligands such as LPS induces tolerance and reduces the infarct size after ischemic-reperfusion injury. The effects of TLR agonists may be explained by tolerance induction through initial stimulation of TLRs. TLRs are similarly involved in the pathogenesis of cerebral ischemia-reperfusion injury models. Other extensive studies have shown that TLR2 and TLR4 are required for the production of atherosclerotic lesions in mice with hyperlipidemia.

Given that TLR2 and TLR4 are clearly responsible for the severity of inflammation induced by nonmicrobial agents, it has been postulated that endogenous molecules directly activate these TLRs. It has been shown that HMGB1 released from necrotic cells stimulates TLR2 and TLR4 to induce the production of proinflammatory cytokines (Lotze and Tracey, 2005). Furthermore, heat shock proteins (HSPs) such as HSP60, HSP70, gp96, and HSP22 are also reported to be recognized by TLR2 and TLR4 (Asea et al., 2002; Ohashi et al., 2000; Roelofs et al., 2006; Warger et al., 2006). However, most studies describing endogenous protein ligands for TLR2 and TLR4 used recombinant proteins generated in an *Escherichia coli* expression system. We need to interpret the data carefully as it is very difficult to completely eliminate the possibility of contamination by *E. coli* components that stimulate TLRs. Nevertheless, it is apparent that these TLRs contribute to inflammatory diseases caused by nonmicrobial agents. Further studies will uncover how these agents activate TLR-induced inflammation.

Future Perspectives

In this Review, we have discussed how the sensing of pathogens and cellular components by PRRs triggers inflammation and its consequences. The cellular mechanisms for individual PRRs to induce pleiotropic outcomes are complex, and we are far from predicting the entire immune response, which is mediated by crosstalk among the various PRRs. Furthermore, we do not know much about the dynamic regulation of immune cell activation or behavior. Investigations into how inflammation is physiologically controlled to cause appropriate outcomes are in their infancy. We believe that one direction for future research is to define the dynamics of immune cell activation and behavior

in vivo by using imaging techniques, as immune cell activation is differentially regulated depending on the cell type involved. A second important approach is to integrate accumulating knowledge using a systems biology strategy. Indeed, there have been several attempts to incorporate systems biology into immunology research. Using a systems biology approach, Aderem and colleagues identified ATF3 and C/EBP γ as a suppressor and amplifier of TLR-mediated gene expression, respectively (Gilchrist et al., 2006; Litvak et al., 2009). There have also been attempts to comprehensively understand the transcription networks activated in response to PRR stimulation in DCs. Recently, Regev and coauthors selected 125 transcription regulators involved in TLR-mediated gene expression based on gene expression profiles and constructed a network model by knocking down each of them in turn (Amit et al., 2009). However, multiple PRRs are activated simultaneously in the course of microbial infection. Thus, the dynamic changes in transcriptional networks activated in response to inflammatory stimuli are likely to be highly complex. In the future, the merging of imaging, systems biology, and immunology will uncover the dynamics of PRR-mediated inflammatory responses and their role in autoimmune disease.

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REFERENCES

- Ablasser, A., Bauernfeind, F., Hartmann, G., Latz, E., Fitzgerald, K.A., and Hornung, V. (2009). RIG-I-dependent sensing of poly(dA:dT) through the induction of an RNA polymerase III-transcribed RNA intermediate. *Nat. Immunol.* *10*, 1065–1072.
- Akira, S., Uematsu, S., and Takeuchi, O. (2006). Pathogen recognition and innate immunity. *Cell* *124*, 783–801.
- Amit, I., Garber, M., Chevrier, N., Leite, A.P., Donner, Y., Eisenhaure, T., Guttman, M., Grenier, J.K., Li, W., Zuk, O., et al. (2009). Unbiased reconstruction of a mammalian transcriptional network mediating pathogen responses. *Science* *326*, 257–263.
- Arimoto, K., Takahashi, H., Hishiki, T., Konishi, H., Fujita, T., and Shimotohno, K. (2007). Negative regulation of the RIG-I signaling by the ubiquitin ligase RNF125. *Proc. Natl. Acad. Sci. USA* *104*, 7500–7505.
- Arnoult, D., Soares, F., Tattoli, I., Castanier, C., Philpott, D.J., and Girardin, S.E. (2009). An N-terminal addressing sequence targets NLRX1 to the mitochondrial matrix. *J. Cell Sci.* *122*, 3161–3168.
- Arumugam, T.V., Okun, E., Tang, S.C., Thundiyil, J., Taylor, S.M., and Woodruff, T.M. (2009). Toll-like receptors in ischemia-reperfusion injury. *Shock* *32*, 4–16.
- Asagiri, M., Hirai, T., Kunigami, T., Kamano, S., Gober, H.J., Okamoto, K., Nishikawa, K., Latz, E., Golenbock, D.T., Aoki, K., et al. (2008). Cathepsin K-dependent toll-like receptor 9 signaling revealed in experimental arthritis. *Science* *319*, 624–627.
- Asea, A., Rehli, M., Kabingu, E., Boch, J.A., Bare, O., Auron, P.E., Stevenson, M.A., and Calderwood, S.K. (2002). Novel signal transduction pathway utilized by extracellular HSP70: role of toll-like receptor (TLR) 2 and TLR4. *J. Biol. Chem.* *277*, 15028–15034.
- Barbalat, R., Lau, L., Locksley, R.M., and Barton, G.M. (2009). Toll-like receptor 2 on inflammatory monocytes induces type I interferon in response to viral but not bacterial ligands. *Nat. Immunol.* *10*, 1200–1207.
- Barbie, D.A., Tamayo, P., Boehm, J.S., Kim, S.Y., Moody, S.E., Dunn, I.F., Schinzel, A.C., Sandy, P., Meylan, E., Scholl, C., et al. (2009). Systematic RNA interference reveals that oncogenic KRAS-driven cancers require TBK1. *Nature* *462*, 108–112.
- Barton, G.M., and Kagan, J.C. (2009). A cell biological view of Toll-like receptor function: regulation through compartmentalization. *Nat. Rev. Immunol.* *9*, 535–542.
- Beutler, B., Jiang, Z., Georgel, P., Crozat, K., Croker, B., Rutschmann, S., Du, X., and Hoebe, K. (2006). Genetic analysis of host resistance: Toll-like receptor signaling and immunity at large. *Annu. Rev. Immunol.* *24*, 353–389.
- Boone, D.L., Turer, E.E., Lee, E.G., Ahmad, R.C., Wheeler, M.T., Tsui, C., Hurley, P., Chien, M., Chai, S., Hitotsumatsu, O., et al. (2004). The ubiquitin-modifying enzyme A20 is required for termination of Toll-like receptor responses. *Nat. Immunol.* *5*, 1052–1060.
- Cadwell, K., Liu, J.Y., Brown, S.L., Miyoshi, H., Loh, J., Lennerz, J.K., Kishi, C., Kc, W., Carrero, J.A., Hunt, S., et al. (2008). A key role for autophagy and the autophagy gene Atg16l1 in mouse and human intestinal Paneth cells. *Nature* *456*, 259–263.
- Carrick, D.M., Lai, W.S., and Blakeshear, P.J. (2004). The tandem CCH zinc finger protein tristetraprolin and its relevance to cytokine mRNA turnover and arthritis. *Arthritis Res. Ther.* *6*, 248–264.
- Carty, M., Goodbody, R., Schroder, M., Stack, J., Moynagh, P.N., and Bowie, A.G. (2006). The human adaptor SARM negatively regulates adaptor protein TRIF-dependent Toll-like receptor signaling. *Nat. Immunol.* *7*, 1074–1081.
- Chien, Y., Kim, S., Bumeister, R., Loo, Y.M., Kwon, S.W., Johnson, C.L., Balakireva, M.G., Romeo, Y., Kopelovich, L., Gale, M., Jr., et al. (2006). RalB GTPase-mediated activation of the I κ B family kinase TBK1 couples innate immune signaling to tumor cell survival. *Cell* *127*, 157–170.
- Chiu, Y.H., Macmillan, J.B., and Chen, Z.J. (2009). RNA polymerase III detects cytosolic DNA and induces type I interferons through the RIG-I pathway. *Cell* *138*, 576–591.
- Cho, J.H. (2008). The genetics and immunopathogenesis of inflammatory bowel disease. *Nat. Rev. Immunol.* *8*, 458–466.
- Choe, J., Kelker, M.S., and Wilson, I.A. (2005). Crystal structure of human toll-like receptor 3 (TLR3) ectodomain. *Science* *309*, 581–585.
- Choi, M.K., Wang, Z., Ban, T., Yanai, H., Lu, Y., Koshiba, R., Nakaima, Y., Hangai, S., Savitsky, D., Nakasato, M., et al. (2009). A selective contribution of the RIG-I-like receptor pathway to type I interferon responses activated by cytosolic DNA. *Proc. Natl. Acad. Sci. USA* *106*, 17870–17875.
- Christensen, S.R., Shupe, J., Nickerson, K., Kashgarian, M., Flavell, R.A., and Shlomchik, M.J. (2006). Toll-like receptor 7 and TLR9 dictate autoantibody specificity and have opposing inflammatory and regulatory roles in a murine model of lupus. *Immunity* *25*, 417–428.
- Coban, C., Ishii, K.J., Kawai, T., Hemmi, H., Sato, S., Uematsu, S., Yamamoto, M., Takeuchi, O., Itagaki, S., Kumar, N., et al. (2005). Toll-like receptor 9 mediates innate immune activation by the malaria pigment hemozoin. *J. Exp. Med.* *201*, 19–25.
- Coban, C., Igari, Y., Yagi, M., Reimer, T., Koyama, S., Aoshi, T., Ohata, K., Tsukui, T., Takeshita, F., Sakurai, K., et al. (2010). Immunogenicity of whole parasite vaccines against plasmodium falciparum involves malarial hemozoin and host TLR9. *Cell Host Microbe* *7*, 50–61.
- Croker, B.A., Lawson, B.R., Rutschmann, S., Berger, M., Eidenschenk, C., Blasius, A.L., Moresco, E.M., Sovath, S., Cengia, L., Shultz, L.D., et al. (2008). Inflammation and autoimmunity caused by a SHP1 mutation depend on IL-1, MyD88, and a microbial trigger. *Proc. Natl. Acad. Sci. USA* *105*, 15028–15033.
- Crow, Y.J., Hayward, B.E., Parmar, R., Robins, P., Leitch, A., Ali, M., Black, D.N., van Bokhoven, H., Brunner, H.G., Hamel, B.C., et al. (2006). Mutations in the gene encoding the 3'-5' DNA exonuclease TREX1 cause Aicardi-Goutieres syndrome at the AGS1 locus. *Nat. Genet.* *38*, 917–920.

- Cui, S., Eisenacher, K., Kirchhofer, A., Brzozka, K., Lammens, A., Lammens, K., Fujita, T., Conzelmann, K.K., Krug, A., and Hopfner, K.P. (2008). The C-terminal regulatory domain is the RNA 5'-triphosphate sensor of RIG-I. *Mol. Cell* 29, 169–179.
- De Santa, F., Totaro, M.G., Prosperini, E., Notarbartolo, S., Testa, G., and Natoli, G. (2007). The histone H3 lysine-27 demethylase Jmjd3 links inflammation to inhibition of polycomb-mediated gene silencing. *Cell* 130, 1083–1094.
- De Santa, F., Narang, V., Yap, Z.H., Tusi, B.K., Burgold, T., Austenaa, L., Bucci, G., Caganova, M., Notarbartolo, S., Casola, S., et al. (2009). Jmjd3 contributes to the control of gene expression in LPS-activated macrophages. *EMBO J.* 28, 3341–3352.
- Ermolaeva, M.A., Michallet, M.C., Papadopoulou, N., Utermohlen, O., Kranioti, K., Kollias, G., Tschopp, J., and Pasparakis, M. (2008). Function of TRADD in tumor necrosis factor receptor 1 signaling and in TRIF-dependent inflammatory responses. *Nat. Immunol.* 9, 1037–1046.
- Ewald, S.E., Lee, B.L., Lau, L., Wickliffe, K.E., Shi, G.P., Chapman, H.A., and Barton, G.M. (2008). The ectodomain of Toll-like receptor 9 is cleaved to generate a functional receptor. *Nature* 456, 658–662.
- Faraoni, I., Antonetti, F.R., Cardone, J., and Bonmassar, E. (2009). miR-155 gene: a typical multifunctional microRNA. *Biochim. Biophys. Acta* 1792, 497–505.
- Foster, S.L., Hargreaves, D.C., and Medzhitov, R. (2007). Gene-specific control of inflammation by TLR-induced chromatin modifications. *Nature* 447, 972–978.
- Fukui, R., Saitoh, S., Matsumoto, F., Kozuka-Hata, H., Oyama, M., Tabeta, K., Beutler, B., and Miyake, K. (2009). Unc93B1 biases Toll-like receptor responses to nucleic acid in dendritic cells toward DNA- but against RNA-sensing. *J. Exp. Med.* 206, 1339–1350.
- Gack, M.U., Shin, Y.C., Joo, C.H., Urano, T., Liang, C., Sun, L., Takeuchi, O., Akira, S., Chen, Z., Inoue, S., and Jung, J.U. (2007). TRIM25 RING-finger E3 ubiquitin ligase is essential for RIG-I-mediated antiviral activity. *Nature* 446, 916–920.
- Ganguly, D., Chamilos, G., Lande, R., Gregorio, J., Meller, S., Facchinetti, V., Homey, B., Barrat, F.J., Zal, T., and Gilliet, M. (2009). Self-RNA-antimicrobial peptide complexes activate human dendritic cells through TLR7 and TLR8. *J. Exp. Med.* 206, 1983–1994.
- Gao, G., Guo, X., and Goff, S.P. (2002). Inhibition of retroviral RNA production by ZAP, a CCCH-type zinc finger protein. *Science* 297, 1703–1706.
- Geijtenbeek, T.B., and Gringhuis, S.I. (2009). Signalling through C-type lectin receptors: shaping immune responses. *Nat. Rev. Immunol.* 9, 465–479.
- Gilchrist, M., Thorsson, V., Li, B., Rust, A.G., Korb, M., Roach, J.C., Kennedy, K., Hai, T., Bolouri, H., and Aderem, A. (2006). Systems biology approaches identify ATF3 as a negative regulator of Toll-like receptor 4. *Nature* 441, 173–178.
- Guo, B., and Cheng, G. (2007). Modulation of the interferon antiviral response by the TBK1/IKKi adaptor protein TANK. *J. Biol. Chem.* 282, 11817–11826.
- Guttman, M., Amit, I., Garber, M., French, C., Lijn, M.F., Feldser, D., Huarte, M., Zuk, O., Carey, B.W., Cassady, J.P., et al. (2009). Chromatin signature reveals over a thousand highly conserved large non-coding RNAs in mammals. *Nature* 458, 223–227.
- Haas, T., Metzger, J., Schmitz, F., Heit, A., Muller, T., Latz, E., and Wagner, H. (2008). The DNA sugar backbone 2' deoxyribose determines toll-like receptor 9 activation. *Immunity* 28, 315–323.
- Hacker, H., Redecke, V., Blagojev, B., Kratchmarova, I., Hsu, L.C., Wang, G.G., Kamps, M.P., Raz, E., Wagner, H., Hacker, G., et al. (2006). Specificity in Toll-like receptor signalling through distinct effector functions of TRAF3 and TRAF6. *Nature* 439, 204–207.
- Hao, S., and Baltimore, D. (2009). The stability of mRNA influences the temporal order of the induction of genes encoding inflammatory molecules. *Nat. Immunol.* 10, 281–288.
- Hitotsumatsu, O., Ahmad, R.C., Tavares, R., Wang, M., Philpott, D., Turer, E.E., Lee, B.L., Shiffin, N., Advincula, R., Malynn, B.A., et al. (2008). The ubiquitin-editing enzyme A20 restricts nucleotide-binding oligomerization domain containing 2-triggered signals. *Immunity* 28, 381–390.
- Honda, K., Takaoka, A., and Taniguchi, T. (2006). Type I interferon [corrected] gene induction by the interferon regulatory factor family of transcription factors. *Immunity* 25, 349–360.
- Hornung, V., Ellegast, J., Kim, S., Brzozka, K., Jung, A., Kato, H., Poeck, H., Akira, S., Conzelmann, K.K., Schlee, M., et al. (2006). 5'-Triphosphate RNA is the ligand for RIG-I. *Science* 314, 994–997.
- Hsu, M.T., Parvin, J.D., Gupta, S., Krystal, M., and Palese, P. (1987). Genomic RNAs of influenza viruses are held in a circular conformation in virions and in infected cells by a terminal panhandle. *Proc. Natl. Acad. Sci. USA* 84, 8140–8144.
- Imai, Y., Kuba, K., Neely, G.G., Yaghubian-Malhami, R., Perkmann, T., van Loo, G., Ermolaeva, M., Veldhuizen, R., Leung, Y.H., Wang, H., et al. (2008). Identification of oxidative stress and Toll-like receptor 4 signaling as a key pathway of acute lung injury. *Cell* 133, 235–249.
- Inohara, Chamailard, McDonald, C., and Nunez, G. (2005). NOD-LRR proteins: role in host-microbial interactions and inflammatory disease. *Annu. Rev. Biochem.* 74, 355–383.
- Ishii, K.J., Coban, C., Kato, H., Takahashi, K., Torii, Y., Takeshita, F., Ludwig, H., Sutter, G., Suzuki, K., Hemmi, H., et al. (2006). A Toll-like receptor-independent antiviral response induced by double-stranded B-form DNA. *Nat. Immunol.* 7, 40–48.
- Ishii, K.J., Kawagoe, T., Koyama, S., Matsui, K., Kumar, H., Kawai, T., Uematsu, S., Takeuchi, O., Takeshita, F., Coban, C., and Akira, S. (2008). TANK-binding kinase-1 delineates innate and adaptive immune responses to DNA vaccines. *Nature* 451, 725–729.
- Ishikawa, H., and Barber, G.N. (2008). STING is an endoplasmic reticulum adaptor that facilitates innate immune signalling. *Nature* 455, 674–678.
- Ishikawa, H., Ma, Z., and Barber, G.N. (2009). STING regulates intracellular DNA-mediated, type I interferon-dependent innate immunity. *Nature* 461, 788–792.
- Jin, M.S., Kim, S.E., Heo, J.Y., Lee, M.E., Kim, H.M., Paik, S.G., Lee, H., and Lee, J.O. (2007). Crystal structure of the TLR1-TLR2 heterodimer induced by binding of a tri-acylated lipopeptide. *Cell* 130, 1071–1082.
- Jones, M.R., Quinton, L.J., Blahna, M.T., Neilson, J.R., Fu, S., Ivanov, A.R., Wolf, D.A., and Mizgerd, J.P. (2009). Zcchc11-dependent uridylation of micro-RNA directs cytokine expression. *Nat. Cell Biol.* 11, 1157–1163.
- Jounai, N., Takeshita, F., Kobiyama, K., Sawano, A., Miyawaki, A., Xin, K.Q., Ishii, K.J., Kawai, T., Akira, S., Suzuki, K., and Okuda, K. (2007). The Atg5 Atg12 conjugate associates with innate antiviral immune responses. *Proc. Natl. Acad. Sci. USA* 104, 14050–14055.
- Kato, H., Sato, S., Yoneyama, M., Yamamoto, M., Uematsu, S., Matsui, K., Tsujimura, T., Takeda, K., Fujita, T., Takeuchi, O., and Akira, S. (2005). Cell type-specific involvement of RIG-I in antiviral response. *Immunity* 23, 19–28.
- Kato, H., Takeuchi, O., Sato, S., Yoneyama, M., Yamamoto, M., Matsui, K., Uematsu, S., Jung, A., Kawai, T., Ishii, K.J., et al. (2006). Differential roles of MDA5 and RIG-I helicases in the recognition of RNA viruses. *Nature* 441, 101–105.
- Kato, H., Takeuchi, O., Mikamo-Sato, E., Hirai, R., Kawai, T., Matsushita, K., Hiiragi, A., Dermody, T.S., Fujita, T., and Akira, S. (2008). Length-dependent recognition of double-stranded ribonucleic acids by retinoic acid-inducible gene-1 and melanoma differentiation-associated gene 5. *J. Exp. Med.* 205, 1601–1610.
- Kawagoe, T., Sato, S., Matsushita, K., Kato, H., Matsui, K., Kumagai, Y., Saitoh, T., Kawai, T., Takeuchi, O., and Akira, S. (2008). Sequential control of Toll-like receptor-dependent responses by IRAK1 and IRAK2. *Nat. Immunol.* 9, 684–691.
- Kawagoe, T., Takeuchi, O., Takabatake, Y., Kato, H., Isaka, Y., Tsujimura, T., and Akira, S. (2009). TANK is a negative regulator of Toll-like receptor signaling and is critical for the prevention of autoimmune nephritis. *Nat. Immunol.* 10, 965–972.

- Kawai, T., and Akira, S. (2006). Innate immune recognition of viral infection. *Nat. Immunol.* 7, 131–137.
- Kawane, K., Fukuyama, H., Yoshida, H., Nagase, H., Ohsawa, Y., Uchiyama, Y., Okada, K., Iida, T., and Nagata, S. (2003). Impaired thymic development in mouse embryos deficient in apoptotic DNA degradation. *Nat. Immunol.* 4, 138–144.
- Kayagaki, N., Phung, Q., Chan, S., Chaudhari, R., Quan, C., O'Rourke, K.M., Eby, M., Pietras, E., Cheng, G., Bazan, J.F., et al. (2007). DUBA: a deubiquitinase that regulates type I interferon production. *Science* 318, 1628–1632.
- Kim, Y.M., Brinkmann, M.M., Paquet, M.E., and Ploegh, H.L. (2008). UNC93B1 delivers nucleotide-sensing toll-like receptors to endolysosomes. *Nature* 452, 234–238.
- Lande, R., Gregorio, J., Facchinetti, V., Chatterjee, B., Wang, Y.H., Homey, B., Cao, W., Wang, Y.H., Su, B., Nestle, F.O., et al. (2007). Plasmacytoid dendritic cells sense self-DNA coupled with antimicrobial peptide. *Nature* 449, 564–569.
- Lee, H.K., Lund, J.M., Ramanathan, B., Mizushima, N., and Iwasaki, A. (2007). Autophagy-dependent viral recognition by plasmacytoid dendritic cells. *Science* 315, 1398–1401.
- Lee-Kirsch, M.A., Gong, M., Chowdhury, D., Senenko, L., Engel, K., Lee, Y.A., de Silva, U., Bailey, S.L., Witte, T., Vyse, T.J., et al. (2007). Mutations in the gene encoding the 3'-5' DNA exonuclease TREX1 are associated with systemic lupus erythematosus. *Nat. Genet.* 39, 1065–1067.
- Litvak, V., Ramsey, S.A., Rust, A.G., Zak, D.E., Kennedy, K.A., Lampano, A.E., Nykter, M., Shmulevich, I., and Aderem, A. (2009). Function of C/EBPdelta in a regulatory circuit that discriminates between transient and persistent TLR4-induced signals. *Nat. Immunol.* 10, 437–443.
- Liu, L., Botos, I., Wang, Y., Leonard, J.N., Shiloach, J., Segal, D.M., and Davies, D.R. (2008). Structural basis of toll-like receptor 3 signaling with double-stranded RNA. *Science* 320, 379–381.
- Loo, Y.M., Fornek, J., Crochet, N., Bajwa, G., Perwitasari, O., Martinez-Sobrido, L., Akira, S., Gill, M.A., Garcia-Sastre, A., Katze, M.G., and Gale, M., Jr. (2008). Distinct RIG-I and MDA5 signaling by RNA viruses in innate immunity. *J. Virol.* 82, 335–345.
- Lotze, M.T., and Tracey, K.J. (2005). High-mobility group box 1 protein (HMGB1): nuclear weapon in the immune arsenal. *Nat. Rev. Immunol.* 5, 331–342.
- Mancuso, G., Gambuzza, M., Midiri, A., Biondo, C., Papiserigi, S., Akira, S., Teli, G., and Beninati, C. (2009). Bacterial recognition by TLR7 in the lysosomes of conventional dendritic cells. *Nat. Immunol.* 10, 587–594.
- Matsumoto, F., Saitoh, S., Fukui, R., Kobayashi, T., Tanimura, N., Konno, K., Kusumoto, Y., Akashi-Takamura, S., and Miyake, K. (2008). Cathepsins are required for Toll-like receptor 9 responses. *Biochem. Biophys. Res. Commun.* 367, 693–699.
- Matsushita, K., Takeuchi, O., Standley, D.M., Kumagai, Y., Kawagoe, T., Miyake, T., Satoh, T., Kato, H., Tsujimura, T., Nakamura, H., and Akira, S. (2009). Zc3h12a is an RNase essential for controlling immune responses by regulating mRNA decay. *Nature* 458, 1185–1190.
- Means, T.K., Latz, E., Hayashi, F., Murali, M.R., Golenbock, D.T., and Luster, A.D. (2005). Human lupus autoantibody-DNA complexes activate DCs through cooperation of CD32 and TLR9. *J. Clin. Invest.* 115, 407–417.
- Medzhitov, R. (2008). Origin and physiological roles of inflammation. *Nature* 454, 428–435.
- Michallet, M.C., Meylan, E., Ermolaeva, M.A., Vazquez, J., Rebsamen, M., Curran, J., Poeck, H., Bscheider, M., Hartmann, G., Konig, M., et al. (2008). TRADD protein is an essential component of the RIG-like helicase antiviral pathway. *Immunity* 28, 651–661.
- Moore, C.B., Bergstralh, D.T., Duncan, J.A., Lei, Y., Morrison, T.E., Zimmermann, A.G., Accavitti-Loper, M.A., Madden, V.J., Sun, L., Ye, Z., et al. (2008). NLRX1 is a regulator of mitochondrial antiviral immunity. *Nature* 457, 573–577.
- Negishi, H., Fujita, Y., Yanai, H., Sakaguchi, S., Ouyang, X., Shinohara, M., Takayanagi, H., Ohba, Y., Taniguchi, T., and Honda, K. (2006). Evidence for licensing of IFN-gamma-induced IFN regulatory factor 1 transcription factor by MyD88 in Toll-like receptor-dependent gene induction program. *Proc. Natl. Acad. Sci. USA* 103, 15136–15141.
- Nejentsev, S., Walker, N., Riches, D., Egholm, M., and Todd, J.A. (2009). Rare variants of IFIH1, a gene implicated in antiviral responses, protect against type 1 diabetes. *Science* 324, 387–389.
- Oganesyan, G., Saha, S.K., Guo, B., He, J.Q., Shahangian, A., Zarnegar, B., Pery, A., and Cheng, G. (2006). Critical role of TRAF3 in the Toll-like receptor-dependent and -independent antiviral response. *Nature* 439, 208–211.
- Ohashi, K., Burkart, V., Flohe, S., and Kolb, H. (2000). Cutting edge: heat shock protein 60 is a putative endogenous ligand of the toll-like receptor-4 complex. *J. Immunol.* 164, 558–561.
- Okabe, Y., Kawane, K., Akira, S., Taniguchi, T., and Nagata, S. (2005). Toll-like receptor-independent gene induction program activated by mammalian DNA escaped from apoptotic DNA degradation. *J. Exp. Med.* 202, 1333–1339.
- Okabe, Y., Sano, T., and Nagata, S. (2009). Regulation of the innate immune response by threonine-phosphatase of Eyes absent. *Nature* 460, 520–524.
- Palsson-McDermott, E.M., Doyle, S.L., McGettrick, A.F., Hardy, M., Husebye, H., Banahan, K., Gong, M., Golenbock, D., Espevik, T., and O'Neill, L.A. (2009). TAG, a splice variant of the adaptor TRAM, negatively regulates the adaptor MyD88-independent TLR4 pathway. *Nat. Immunol.* 10, 579–586.
- Park, B., Brinkmann, M.M., Spooner, E., Lee, C.C., Kim, Y.M., and Ploegh, H.L. (2008). Proteolytic cleavage in an endolysosomal compartment is required for activation of Toll-like receptor 9. *Nat. Immunol.* 9, 1407–1414.
- Park, B.S., Song, D.H., Kim, H.M., Choi, B.S., Lee, H., and Lee, J.O. (2009). The structural basis of lipopolysaccharide recognition by the TLR4-MD-2 complex. *Nature* 458, 1191–1195.
- Parroche, P., Lauw, F.N., Goutagny, N., Latz, E., Monks, B.G., Visintin, A., Halmen, K.A., Lamphier, M., Olivier, M., Bartholomeu, D.C., et al. (2007). Malaria hemozoin is immunologically inert but radically enhances innate responses by presenting malaria DNA to Toll-like receptor 9. *Proc. Natl. Acad. Sci. USA* 104, 1919–1924.
- Pattnaik, A.K., Ball, L.A., LeGrone, A., and Wertz, G.W. (1995). The termini of VSV DI particle RNAs are sufficient to signal RNA encapsidation, replication, and budding to generate infectious particles. *Virology* 206, 760–764.
- Pichlmair, A., Schulz, O., Tan, C.P., Naslund, T.I., Liljestrom, P., Weber, F., and Reis e Sousa, C. (2006). RIG-I-mediated antiviral responses to single-stranded RNA bearing 5'-phosphates. *Science* 314, 997–1001.
- Pichlmair, A., Schulz, O., Tan, C.P., Rehwinkel, J., Kato, H., Takeuchi, O., Akira, S., Way, M., Schiavo, G., and Reis e Sousa, C. (2009). Activation of MDA5 requires higher-order RNA structures generated during virus infection. *J. Virol.* 83, 10761–10769.
- Pisitkun, P., Deane, J.A., Difilippantonio, M.J., Tarasenko, T., Satterthwaite, A.B., and Bolland, S. (2006). Autoreactive B cell responses to RNA-related antigens due to TLR7 gene duplication. *Science* 312, 1669–1672.
- Pobezinskaya, Y.L., Kim, Y.S., Choksi, S., Morgan, M.J., Li, T., Liu, C., and Liu, Z. (2008). The function of TRADD in signaling through tumor necrosis factor receptor 1 and TRIF-dependent Toll-like receptors. *Nat. Immunol.* 9, 1047–1054.
- Ramirez-Carrozzi, V.R., Nazarian, A.A., Li, C.C., Gore, S.L., Sridharan, R., Imbalzano, A.N., and Smale, S.T. (2006). Selective and antagonistic functions of SWI/SNF and Mi-2beta nucleosome remodeling complexes during an inflammatory response. *Genes Dev.* 20, 282–296.
- Rice, G.I., Bond, J., Asipu, A., Brunette, R.L., Manfield, I.W., Carr, I.M., Fuller, J.C., Jackson, R.M., Lamb, T., Briggs, T.A., et al. (2009). Mutations involved in Aicardi-Goutieres syndrome implicate SAMHD1 as regulator of the innate immune response. *Nat. Genet.* 41, 829–832.
- Robinson, M.J., Osorio, F., Rosas, M., Freitas, R.P., Schweighoffer, E., Gross, O., Verbeek, J.S., Ruland, J., Tybulewicz, V., Brown, G.D., et al. (2009). Dectin-2 is a Syk-coupled pattern recognition receptor crucial for Th17 responses to fungal infection. *J. Exp. Med.* 206, 2037–2051.
- Roelofs, M.F., Boelens, W.C., Joosten, L.A., Abdollahi-Roodsaz, S., Geurts, J., Wunderink, L.U., Schreurs, B.W., van den Berg, W.B., and Radstake, T.R.

- (2006). Identification of small heat shock protein B8 (HSP22) as a novel TLR4 ligand and potential involvement in the pathogenesis of rheumatoid arthritis. *J. Immunol.* **176**, 7021–7027.
- Rothenfusser, S., Goutagny, N., DiPerna, G., Gong, M., Monks, B.G., Schoenemeyer, A., Yamamoto, M., Akira, S., and Fitzgerald, K.A. (2005). The RNA helicase Lgp2 inhibits TLR-independent sensing of viral replication by retinoic acid-inducible gene-1. *J. Immunol.* **175**, 5260–5268.
- Ryzhakov, G., and Randow, F. (2007). SINTBAD, a novel component of innate antiviral immunity, shares a TBK1-binding domain with NAP1 and TANK. *EMBO J.* **26**, 3180–3190.
- Sabbah, A., Chang, T.H., Harnack, R., Frohlich, V., Tominaga, K., Dube, P.H., Xiang, Y., and Bose, S. (2009). Activation of innate immune antiviral responses by Nod2. *Nat. Immunol.* **10**, 1073–1080.
- Saito, T., Hirai, R., Loo, Y.M., Owen, D., Johnson, C.L., Sinha, S.C., Akira, S., Fujita, T., and Gale, M., Jr. (2007). Regulation of innate antiviral defenses through a shared repressor domain in RIG-I and LGP2. *Proc. Natl. Acad. Sci. USA* **104**, 582–587.
- Saito, T., Owen, D.M., Jiang, F., Marcotrigiano, J., and Gale, M., Jr. (2008). Innate immunity induced by composition-dependent RIG-I recognition of hepatitis C virus RNA. *Nature* **454**, 523–527.
- Saitoh, T., Fujita, N., Jang, M.H., Uematsu, S., Yang, B.G., Satoh, T., Omori, H., Noda, T., Yamamoto, N., Komatsu, M., et al. (2008). Loss of the autophagy protein Atg16L1 enhances endotoxin-induced IL-1 β production. *Nature* **456**, 264–268.
- Saitoh, T., Fujita, N., Hayashi, T., Takahara, K., Satoh, T., Lee, H., Matsunaga, K., Kageyama, S., Omori, H., Noda, T., et al. (2009). Atg9a controls dsDNA-driven dynamic translocation of STING and the innate immune response. *Proc. Natl. Acad. Sci. USA*. Published online November 19, 2009. 10.1073/pnas.0911267106.
- Sasai, M., Shingai, M., Funami, K., Yoneyama, M., Fujita, T., Matsumoto, M., and Seya, T. (2006). NAK-associated protein 1 participates in both the TLR3 and the cytoplasmic pathways in type I IFN induction. *J. Immunol.* **177**, 8676–8683.
- Satoh, S., Kato, H., Kumagai, Y., Yoneyama, M., Sato, S., Matsushita, K., Tsujimura, T., Fujita, T., Akira, S., and Takeuchi, O. (2010). LGP2 is a positive regulator of RIG-I- and MDA5-mediated antiviral responses. *Proc. Natl. Acad. Sci. USA* **107**, 1512–1517.
- Schlee, M., Roth, A., Hornung, V., Hagmann, C.A., Wimmenauer, V., Barchet, W., Coch, C., Janke, M., Mihailovic, A., Wardle, G., et al. (2009). Recognition of 5' triphosphate by RIG-I helicase requires short blunt double-stranded RNA as contained in panhandle of negative-strand virus. *Immunity* **31**, 25–34.
- Schmidt, A., Schwerdt, T., Hamm, W., Hellmuth, J.C., Cui, S., Wenzel, M., Hoffmann, F.S., Michallet, M.C., Besch, R., Hopfner, K.P., et al. (2009). 5'-triphosphate RNA requires base-paired structures to activate antiviral signaling via RIG-I. *Proc. Natl. Acad. Sci. USA* **106**, 12067–12072.
- Schmitz, F., Heit, A., Guggemoos, S., Krug, A., Mages, J., Schiemann, M., Adler, H., Drexler, I., Haas, T., Lang, R., and Wagner, H. (2007). Interferon-regulatory-factor 1 controls Toll-like receptor 9-mediated IFN- β production in myeloid dendritic cells. *Eur. J. Immunol.* **37**, 315–327.
- Sepulveda, F.E., Maschalidi, S., Colisson, R., Heslop, L., Ghirelli, C., Sakka, E., Lennon-Dumenil, A.M., Amigorena, S., Cabanie, L., and Manoury, B. (2009). Critical role for asparagine endopeptidase in endocytic Toll-like receptor signaling in dendritic cells. *Immunity* **31**, 737–748.
- Sheedy, F.J., Palsson-McDermott, E., Hennessy, E.J., Martin, C., O'Leary, J., Ruan, Q., Johnson, D.P., Chen, Y., and O'Neill, L.A. (2009). Negative regulation of TLR4 via targeting of the proinflammatory tumor suppressor PDCD4 by the microRNA miR-21. *Nat. Immunol.* **11**, 141–147.
- Stetson, D.B., and Medzhitov, R. (2006). Recognition of cytosolic DNA activates an IRF3-dependent innate immune response. *Immunity* **24**, 93–103.
- Stetson, D.B., Ko, J.S., Heidmann, T., and Medzhitov, R. (2008). Trex1 prevents cell-intrinsic initiation of autoimmunity. *Cell* **134**, 587–598.
- Subramanian, S., Tus, K., Li, Q.Z., Wang, A., Tian, X.H., Zhou, J., Liang, C., Bartov, G., McDaniel, L.D., Zhou, X.J., et al. (2006). A Tlr7 translocation accelerates systemic autoimmunity in murine lupus. *Proc. Natl. Acad. Sci. USA* **103**, 9970–9975.
- Sumpter, R., Jr., Loo, Y.M., Foy, E., Li, K., Yoneyama, M., Fujita, T., Lemon, S.M., and Gale, M., Jr. (2005). Regulating intracellular antiviral defense and permissiveness to hepatitis C virus RNA replication through a cellular RNA helicase, RIG-I. *J. Virol.* **79**, 2689–2699.
- Sun, W., Li, Y., Chen, L., Chen, H., You, F., Zhou, X., Zhou, Y., Zhai, Z., Chen, D., and Jiang, Z. (2009). ERIS, an endoplasmic reticulum IFN stimulator, activates innate immune signaling through dimerization. *Proc. Natl. Acad. Sci. USA* **106**, 8653–8658.
- Tabeta, K., Hoebe, K., Janssen, E.M., Du, X., Georgel, P., Crozat, K., Mudd, S., Mann, N., Sovath, S., Goode, J., et al. (2006). The Unc93b1 mutation 3d disrupts exogenous antigen presentation and signaling via Toll-like receptors 3, 7 and 9. *Nat. Immunol.* **7**, 156–164.
- Taganov, K.D., Boldin, M.P., Chang, K.J., and Baltimore, D. (2006). NF- κ B-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. *Proc. Natl. Acad. Sci. USA* **103**, 12481–12486.
- Takahashi, K., Kawai, T., Kumar, H., Sato, S., Yonehara, S., and Akira, S. (2006). Roles of caspase-8 and caspase-10 in innate immune responses to double-stranded RNA. *J. Immunol.* **176**, 4520–4524.
- Takahashi, K., Yoneyama, M., Nishihori, T., Hirai, R., Kumeta, H., Narita, R., Gale, M., Jr., Inagaki, F., and Fujita, T. (2008). Nonself RNA-sensing mechanism of RIG-I helicase and activation of antiviral immune responses. *Mol. Cell* **29**, 428–440.
- Takahashi, K., Kumeta, H., Tsuduki, N., Narita, R., Shigemoto, T., Hirai, R., Yoneyama, M., Horiuchi, M., Ogura, K., Fujita, T., and Inagaki, F. (2009). Solution structures of cytosolic RNA sensor MDA5 and LGP2 C-terminal domains: identification of the RNA recognition loop in RIG-I-like receptors. *J. Biol. Chem.* **284**, 17465–17474.
- Takaoka, A., Wang, Z., Choi, M.K., Yanai, H., Negishi, H., Ban, T., Lu, Y., Miyagishi, M., Kodama, T., Honda, K., et al. (2007). DAI (DLM-1/ZBP1) is a cytosolic DNA sensor and an activator of innate immune response. *Nature* **448**, 501–505.
- Takeuchi, O., and Akira, S. (2009). Innate immunity to virus infection. *Immunol. Rev.* **227**, 75–86.
- Tal, M.C., Sasai, M., Lee, H.K., Yordy, B., Shadel, G.S., and Iwasaki, A. (2009). Absence of autophagy results in reactive oxygen species-dependent amplification of RLR signaling. *Proc. Natl. Acad. Sci. USA* **106**, 2770–2775.
- Tenover, B.R., Ng, S.L., Chua, M.A., McWhirter, S.M., Garcia-Sastre, A., and Maniatis, T. (2007). Multiple functions of the IKK-related kinase IKKepsilon in interferon-mediated antiviral immunity. *Science* **315**, 1274–1278.
- Tian, J., Avalos, A.M., Mao, S.Y., Chen, B., Senthil, K., Wu, H., Parroche, P., Drabic, S., Golenbock, D., Sirois, C., et al. (2007). Toll-like receptor 9-dependent activation by DNA-containing immune complexes is mediated by HMGB1 and RAGE. *Nat. Immunol.* **8**, 487–496.
- Tseng, P.H., Matsuzawa, A., Zhang, W., Mino, T., Vignali, D.A., and Karin, M. (2009). Different modes of ubiquitination of the adaptor TRAF3 selectively activate the expression of type I interferons and proinflammatory cytokines. *Nat. Immunol.* **11**, 70–75.
- Uematsu, S., Fujimoto, K., Jang, M.H., Yang, B.G., Jung, Y.J., Nishiyama, M., Sato, S., Tsujimura, T., Yamamoto, M., Yokota, Y., et al. (2008). Regulation of humoral and cellular gut immunity by lamina propria dendritic cells expressing Toll-like receptor 5. *Nat. Immunol.* **9**, 769–776.
- Vigilanti, G.A., Lau, C.M., Hanley, T.M., Miko, B.A., Shlomchik, M.J., and Marshak-Rothstein, A. (2003). Activation of autoreactive B cells by CpG dsDNA. *Immunity* **19**, 837–847.
- Vollmer, J., Tluk, S., Schmitz, C., Hamm, S., Jurk, M., Forsbach, A., Akira, S., Kelly, K.M., Reeves, W.H., Bauer, S., and Krieg, A.M. (2005). Immune stimulation mediated by autoantigen binding sites within small nuclear RNAs involves Toll-like receptors 7 and 8. *J. Exp. Med.* **202**, 1575–1585.
- Warger, T., Hilf, N., Rechtsteiner, G., Haselmayer, P., Carrick, D.M., Jonuleit, H., von Landenberg, P., Rammensee, H.G., Nicchitta, C.V., Radsak, M.P.,

- and Schild, H. (2006). Interaction of TLR2 and TLR4 ligands with the N-terminal domain of Gp96 amplifies innate and adaptive immune responses. *J. Biol. Chem.* *281*, 22545–22553.
- Xia, Z.P., Sun, L., Chen, X., Pineda, G., Jiang, X., Adhikari, A., Zeng, W., and Chen, Z.J. (2009). Direct activation of protein kinases by unanchored polyubiquitin chains. *Nature* *461*, 114–119.
- Yamamoto, M., Yamazaki, S., Uematsu, S., Sato, S., Hemmi, H., Hoshino, K., Kaisho, T., Kuwata, H., Takeuchi, O., Takeshige, K., et al. (2004). Regulation of Toll/IL-1-receptor-mediated gene expression by the inducible nuclear protein I κ B ζ . *Nature* *430*, 218–222.
- Yamasaki, S., Ishikawa, E., Sakuma, M., Hara, H., Ogata, K., and Saito, T. (2008). Mincle is an ITAM-coupled activating receptor that senses damaged cells. *Nat. Immunol.* *9*, 1179–1188.
- Yamasaki, S., Matsumoto, M., Takeuchi, O., Matsuzawa, T., Ishikawa, E., Sakuma, M., Tateno, H., Uno, J., Hirabayashi, J., Mikami, Y., et al. (2009). C-type lectin Mincle is an activating receptor for pathogenic fungus, *Malassezia*. *Proc. Natl. Acad. Sci. USA* *106*, 1897–1902.
- Yanai, H., Ban, T., Wang, Z., Choi, M.K., Kawamura, T., Negishi, H., Nakasato, M., Lu, Y., Hangai, S., Koshiba, R., et al. (2009). HMGB proteins function as universal sentinels for nucleic-acid-mediated innate immune responses. *Nature* *462*, 99–103.
- Yarovinsky, F., Zhang, D., Andersen, J.F., Bannenberg, G.L., Serhan, C.N., Hayden, M.S., Hieny, S., Sutterwala, F.S., Flavell, R.A., Ghosh, S., and Sher, A. (2005). TLR11 activation of dendritic cells by a protozoan profilin-like protein. *Science* *308*, 1626–1629.
- Yoneyama, M., and Fujita, T. (2008). Structural mechanism of RNA recognition by the RIG-I-like receptors. *Immunity* *29*, 178–181.
- Yoneyama, M., Kikuchi, M., Matsumoto, K., Imaizumi, T., Miyagishi, M., Taira, K., Foy, E., Loo, Y.M., Gale, M., Jr., Akira, S., et al. (2005). Shared and unique functions of the DExD/H-box helicases RIG-I, MDA5, and LGP2 in antiviral innate immunity. *J. Immunol.* *175*, 2851–2858.
- Zeng, W., Xu, M., Liu, S., Sun, L., and Chen, Z.J. (2009). Key role of Ubc5 and lysine-63 polyubiquitination in viral activation of IRF3. *Mol. Cell* *36*, 315–325.
- Zhong, B., Yang, Y., Li, S., Wang, Y.Y., Li, Y., Diao, F., Lei, C., He, X., Zhang, L., Tien, P., and Shu, H.B. (2008). The adaptor protein MITA links virus-sensing receptors to IRF3 transcription factor activation. *Immunity* *29*, 538–550.

TANK is a negative regulator of Toll-like receptor signaling and is critical for the prevention of autoimmune nephritis

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The intensity and duration of immune responses are controlled by many proteins that modulate Toll-like receptor (TLR) signaling. TANK has been linked to positive regulation of the transcription factors IRF3 and NF- κ B. Here we demonstrate that TANK is not involved in interferon responses and is a negative regulator of proinflammatory cytokine production induced by TLR signaling. TLR-induced polyubiquitination of the ubiquitin ligase TRAF6 was upregulated in *Tank*^{-/-} macrophages. Notably, *Tank*^{-/-} mice spontaneously developed fatal glomerulonephritis owing to deposition of immune complexes. Autoantibody production in *Tank*^{-/-} mice was abrogated by antibiotic treatment or the absence of interleukin 6 (IL-6) or the adaptor MyD88. Our results demonstrate that constitutive TLR signaling by intestinal commensal microflora is suppressed by TANK.

Toll-like receptors (TLRs) recognize microbial components and elicit innate as well as adaptive immune responses. Stimulation with TLR ligands induces the production of proinflammatory cytokines and type I interferons in cells of the innate immune system through intracellular signaling cascades¹⁻³. After stimulation, TLRs trigger the recruitment of adaptor molecules containing Toll-interleukin 1 receptor (IL-1R) homology domains. One adaptor, MyD88, is essential for the 'downstream' signaling of various TLRs, except for TLR3 (refs. 4-6). MyD88 interacts with the kinase IRAK4, which activates IRAK1 and IRAK2 (ref. 7). In turn, the IRAKs dissociate from MyD88 and interact with TRAF6 (A002312), which acts as a ubiquitin protein ligase. Together with an E2 ubiquitin-conjugating enzyme complex composed of Ubc13 and Uev1A, TRAF6 catalyzes formation of a lysine 63-linked polyubiquitin chain on TRAF6 itself and on the transcription factor NF- κ B modulator NEMO (also called IKK γ). The kinase TAK1 is also recruited to TRAF6 and phosphorylates the kinases IKK β and MEKK6 (ref. 8). Subsequently, the inhibitor of NF- κ B (I κ B) kinase (IKK) complex, composed of IKK α , IKK β and NEMO, is formed. NF- κ B binds to I κ B α in resting cells and is sequestered in the cytoplasm. Phosphorylation of I κ B by the IKK complex leads to its degradation by the ubiquitin-proteasome system, thereby freeing NF- κ B to translocate to the nucleus and activate the expression of genes encoding proinflammatory cytokines. Activation of the mitogen-activated protein kinase cascade is responsible for gene expression induced by the transcription activator AP-1. In plasmacytoid dendritic cells (DCs), MyD88-dependent

signaling activates the production of type I interferons through the transcription factor IRF7 (refs. 1,9).

Many proteins control TLR signaling to ensure that the strength and duration of TLR signals is appropriate for any given immune response. TLRs have been linked to the development of autoimmune diseases, and aberrant activation of innate immunity may contribute to rheumatoid arthritis, inflammatory bowel disease and systemic lupus erythematosus^{10,11}. Endogenous RNA molecules such as U1snRNP can activate autoreactive B cells and DCs through TLR7 (ref. 12). Furthermore, duplication of the *Th7* gene accounts for the autoimmune phenotypes associated with Yaa mice (Y chromosome-linked autoimmune accelerator)¹³. TLR9 is also involved in the recognition of immune complexes of DNA and anti-double-stranded DNA (anti-dsDNA) antibodies together with the B cell antigen receptor (BCR)¹⁴. Signaling proteins that inhibit TLR signaling include IRAKM, ST2, SIGIRR, SOCS1, the tumor suppressor CYLD and A20 (refs. 15-21). Cells lacking any one of those proteins produce more proinflammatory cytokines in response to TLR stimulation. Furthermore, mice lacking SOCS1 or A20 have immune disorders that lead to premature death. In addition, the immunosuppressive cytokine IL-10 suppresses colitis development by inhibiting TLR responses²²⁻²⁴. Such studies indicate that negative regulation of TLR signaling is important for coordinated innate immune responses.

TANK (also known as I-TRAF) has been identified as a TRAF-binding protein^{25,26}. Among the six reported TRAF family members, TRAF1, TRAF2, TRAF3, TRAF5 and TRAF6 interact with TANK²⁵⁻²⁸.

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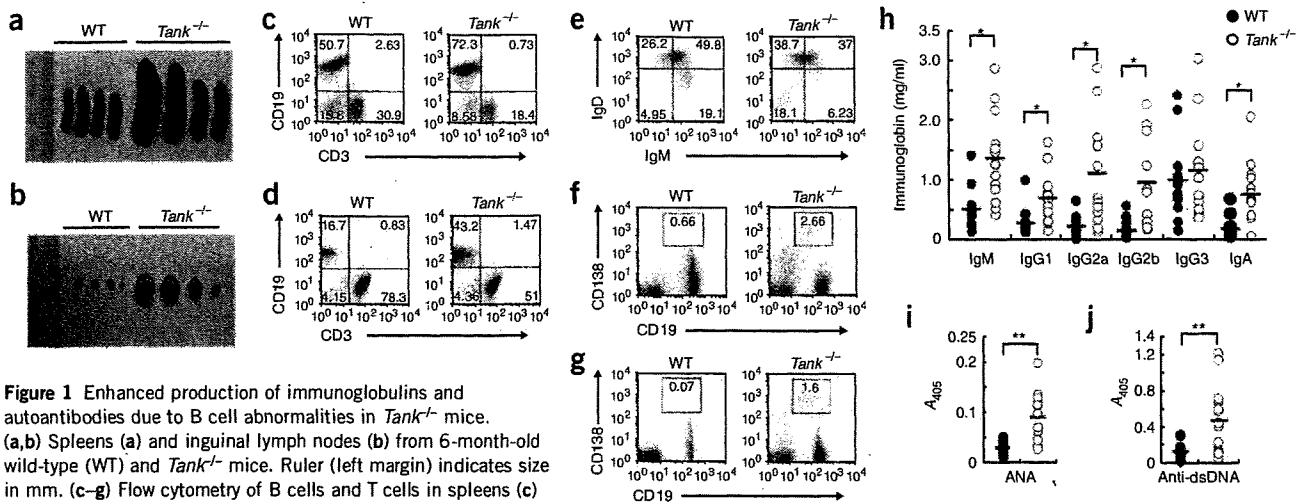


Figure 1 Enhanced production of immunoglobulins and autoantibodies due to B cell abnormalities in *Tank*^{-/-} mice. (a,b) Splenocytes (a) and inguinal lymph nodes (b) from 6-month-old wild-type (WT) and *Tank*^{-/-} mice. Ruler (left margin) indicates size in mm. (c–g) Flow cytometry of B cells and T cells in spleens (c) and lymph nodes (d), expression of IgM and IgD on splenic B cells (e), and expression of CD138 and CD19 on cells in spleens (f) and lymph nodes (g) from wild-type and *Tank*^{-/-} mice. Numbers in quadrants and outlined areas indicate percent cells in each. (h) Enzyme-linked immunosorbent assay (ELISA) of basal titers of immunoglobulin isotypes in serum from unimmunized 3-month-old wild-type and *Tank*^{-/-} mice. (i,j) ELISA of antinuclear antibodies (ANA; i) and anti-dsDNA antibodies (j) in serum from 12-month-old wild-type and *Tank*^{-/-} mice. A₄₀₅, absorbance at 405 nm. Each symbol represents an individual mouse; small horizontal lines indicate the mean (h–j). **P* < 0.005 and ***P* < 0.001, versus *Tank*^{-/-} cells (h–j;). Data are representative of at least five experiments (a,b); three independent experiments (c–g); or single experiments with a total of 13 (h) or 12 (i,j) mice per genotype (h–j).

TANK has been linked to the positive regulation of NF- κ B activation. In addition to TRAF family members, Ikki and TBK1 are TANK-binding partners^{29,30}. These proteins phosphorylate IRF3 and IRF7, which are transcription factors essential for the expression of type I interferon and interferon-inducible genes^{31,32}. TBK1 and Ikki are activated in response to recognition of viruses through TLRs and RNA helicase RIG-I-like receptors (RLRs)^{33,34}. TRAF3 is required for the activation of TBK1 and Ikki downstream of TLRs and RLRs^{35,36}. It has been reported that TANK functions as an adaptor that bridges TRAF3 and TBK1-Ikki and that TANK is required for the production of type I interferon in response to viral infection or TLR stimulation³⁷. However, the functions of TANK *in vivo* have not yet been clarified.

Here we used *Tank*-deficient (*Tank*^{-/-}) mice to demonstrate that TANK is not involved in interferon responses but is a negative regulator of TLR and BCR signaling. Macrophages and B cells from *Tank*^{-/-} mice had more canonical NF- κ B activation in response to stimulation of TLRs and the BCR. TLR-induced polyubiquitination of TRAF6 was upregulated in *Tank*^{-/-} macrophages, which indicates that TANK suppresses TLR signaling by controlling TRAF ubiquitination. *Tank*^{-/-} mice spontaneously developed lupus-like autoimmune nephritis. Autoantibody production in *Tank*^{-/-} mice was abolished in the absence of IL-6 or MyD88 but not in the absence of tumor necrosis factor (TNF). Furthermore, treatment of *Tank*^{-/-} mice with antibiotics resulted in lower autoantibody production, which indicates that IL-6 produced by constitutive TLR stimulation resulting from intestinal commensal microflora is important for the development of disease.

RESULTS

Tank^{-/-} mice develop lupus-like nephritis

To investigate the physiological functions of TANK *in vivo*, we generated *Tank*^{-/-} mice by homologous recombination in embryonic stem cells. We targeted exons 3 and 4 of mouse *Tank* with a neomycin-resistance cassette in embryonic stem cells and established *Tank*^{-/-} mice (Supplementary Fig. 1a). We confirmed homologous recombination of the *Tank* locus by Southern blot analysis (Supplementary Fig. 1b). Expression of *Tank* mRNA and TANK protein was abrogated

in *Tank*^{-/-} macrophages (Supplementary Fig. 1c,d). *Tank*^{-/-} mice were born from interbred *Tank*^{+/-} mice at the expected mendelian ratios and grew normally.

Tank^{-/-} mice had splenomegaly and lymphadenopathy (Fig. 1a,b). Flow cytometry showed a higher percentage of CD19⁺ B cells in the spleen and lymph nodes of *Tank*^{-/-} mice (Fig. 1c,d). Immunoglobulin M-low (IgM^{lo}) IgD^{hi} mature B cells accumulated in the spleen of *Tank*^{-/-} mice (Fig. 1e). The percentage of CD19⁺CD138⁺ plasma cells was also much higher in the spleen and lymph nodes of *Tank*^{-/-} mice (Fig. 1f,g). In contrast, the percentage of Foxp3⁺CD4⁺ regulatory T cells did not differ between wild-type and *Tank*^{-/-} mice (Supplementary Fig. 2a). Consistent with the larger B cell populations, basal serum concentrations of IgM, IgG1, IgG2a, IgG2b and IgA were significantly higher, by 1.2-fold to 6.2-fold, in *Tank*^{-/-} mice than in wild-type mice (Fig. 1h). Notably, we detected antinuclear antibodies and anti-dsDNA antibodies in the serum of *Tank*^{-/-} mice (Fig. 1i,j).

Tank^{-/-} mice began to spontaneously die at 3 months after birth, and about 50% had died by 12 months after birth (Fig. 2a). Histological studies showed that 24-week-old *Tank*^{-/-} mice had glomerulonephritis with mesangial cell proliferation and expansion of the mesangial matrix (Fig. 2b,c). The glomerular structure was devastated in terminally ill *Tank*^{-/-} mice (data not shown), which suggested that renal failure was the cause of death. Although we found infiltration of lymphocytes in the liver and lungs of *Tank*^{-/-} mice, we detected no histological changes in their intestine, heart or joints (data not shown). In addition, we found deposition of IgG, IgM and complement components C3 and C1q in the glomeruli of *Tank*^{-/-} mice (Fig. 2d). Such depositions are characteristic of lupus-like nephritis and suggest that deposition of immune complexes of autoantibodies was the cause of the glomerulonephritis in *Tank*^{-/-} mice.

TANK is a negative regulator of TLR responses

Next we examined the type I interferon responses of *Tank*^{-/-} cells to virus infection. In contrast to the results obtained by *in vitro* studies, interferon- β (IFN- β) production in response to infection with

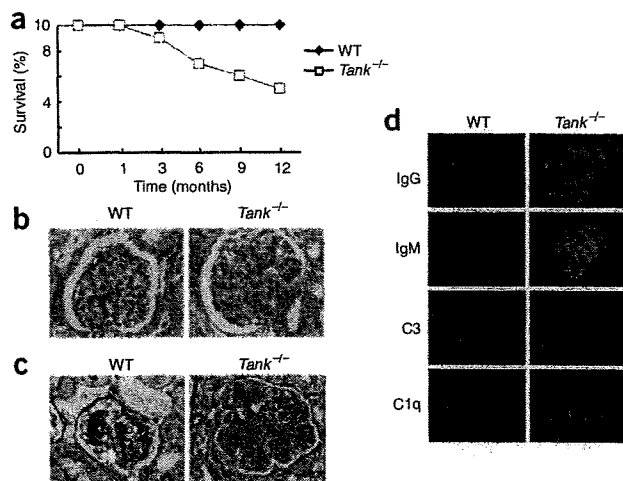


Figure 2 Development of lethal glomerulonephritis in *Tank*^{-/-} mice. (a) Survival of wild-type and *Tank*^{-/-} mice monitored for 1 year. (b,c) Kidney sections from 6-month-old wild-type and *Tank*^{-/-} mice, stained with hematoxylin and eosin (b) or periodic acid-Schiff (c). (d) Kidney sections from 6-month-old wild-type and *Tank*^{-/-} mice, stained with fluorescein isothiocyanate (FITC)-labeled anti-mouse IgG, IgM, C3 and C1q. Original magnification, $\times 100$ (b-d). Data are representative of single experiments with a total of ten mice per genotype (a), or five (b,c) or two (d) experiments.

Newcastle disease virus did not differ between wild-type and *Tank*^{-/-} conventional DCs derived from bone marrow cells (Fig. 3a). Wild-type and *Tank*^{-/-} conventional DCs also produced similar amounts of IL-6 (Fig. 3b). Newcastle disease virus is recognized by the RNA helicase RIG-I in conventional DCs, which indicates that TANK is not essential for the activation of signaling pathways by RLRs. TRAF3 has been shown to be activated downstream of TLR7 and TLR9 in plasmacytoid DCs. However, bone marrow plasmacytoid DCs induced by the cytokine Flt3L from *Tank*^{-/-} mice produced more rather than less IFN- α and IL-6 in response to A- or D-type CpG DNA (Fig. 3c,d). Collectively these results indicate that TANK is not essential for type I interferon responses.

Next we examined the production of proinflammatory cytokines in macrophages in response to a set of TLR ligands, including MALP-2 (TLR6-TLR2), polyinosinic-polycytidylic acid (poly(I:C); TLR3), lipopolysaccharide (LPS; TLR4), the synthetic imidazoquinoline resiquimod (R-848; TLR7) and CpG DNA (TLR9). The production of IL-6 and TNF in response to these TLR ligands, except poly(I:C), was much higher in *Tank*^{-/-} peritoneal macrophages than in wild-type cells (Fig. 3e,f). Of note, the enhanced cytokine production in response

to LPS stimulation in *Tank*^{-/-} macrophages was less substantial than that induced by other TLR ligands. Conventional DCs from *Tank*^{-/-} mice also showed excessive cytokine production in response to these TLR ligands (data not shown).

We subsequently assessed the function of TANK in cytokine responses to stimulation with TLR ligands *in vivo*. We chose R-848, because the enhancement in cytokine production in *Tank*^{-/-} macrophages was most pronounced after stimulation with R-848. We injected R-848 into the peritoneum of 8-week-old mice and measured serum concentrations of IL-6 and IFN- α 1 and 3 h later. *Tank*^{-/-} mice had significantly more of these serum cytokines at both time points than did wild-type mice (Fig. 3g,h). Together these results indicate that TANK is a negative regulator of TLR-mediated responses but is not an essential positive regulator of type I interferon responses *in vivo*.

TANK controls TRAF6 ubiquitination

We examined whether the enhanced cytokine production in *Tank*^{-/-} macrophages was evident at the level of transcription. In response to R-848 stimulation, wild-type macrophages showed induction of the expression of *Il6*, *Tnf*, *Il12b*, *Ptgs2* (encoding cyclooxygenase 2), *Nfkbiz* (encoding I κ B ζ) and *Nos2* (encoding nitric oxide synthase 2). The expression of these genes was enhanced in *Tank*^{-/-} macrophages in response to R-848 stimulation (Fig. 4a), which indicated that initial TLR-induced gene expression was enhanced in *Tank*^{-/-} macrophages. Next we analyzed activation of the transcription factors NF- κ B and AP-1 by electrophoretic mobility-shift assay (EMSA). In response to R-848 stimulation, activation of NF- κ B and AP-1 was enhanced in *Tank*^{-/-} macrophages compared with that in wild-type macrophages (Fig. 4b,c).

Figure 3 Enhanced proinflammatory cytokine production in response to TLR stimulation in *Tank*^{-/-} mice. (a,b) ELISA of IFN- α (a) and IL-6 (b) in culture supernatants of wild-type and *Tank*^{-/-} bone marrow-derived DCs infected for 24 h with Newcastle disease virus (NDV). (c,d) ELISA of IFN- α (c) and IL-6 (d) in culture supernatants of wild-type and *Tank*^{-/-} DCs stimulated for 24 h with 0.1 or 1 μ M CpG DNA. (e,f) ELISA of IL-6 (e) and TNF (f) in culture supernatants of wild-type and *Tank*^{-/-} peritoneal macrophages stimulated for 24 h with MALP-2 (10 ng/ml), poly(I:C) (100 μ g/ml), LPS (100 ng/ml), R-848 (10 nM) or CpG DNA (1 μ M). ND, not detectable. Data are representative of three (a-d) or five (e,f) experiments (error bars, s.d.). (g,h) ELISA of IL-6 (g) and IFN- α (h) in serum from wild-type mice ($n = 5$) and *Tank*^{-/-} mice ($n = 5$) injected intraperitoneally with 30 nmol R-848. Each symbol represents an individual mouse; small horizontal lines indicate the mean. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.005$, versus *Tank*^{-/-} mice (two-tailed Student's *t*-test). Data are representative of a single experiment.

