Figure 2 Crude hepatocellular carcinogenesis (HCC) curves in patients of groups A, B, C and D. There was no significant difference in the HCC appearance rate among the four groups (log-rank test, $P = 0.42$).

≥ 12 months was 0.022 after adjustments for significant covariates, but was not significantly different ($P = 0.43$).

We also assessed the effects of splenectomy and long-term IFN therapy on hepatocarcinogenesis by comparing patients of group D (splenectomy + IFN administration for ≥ 12 months) with those of group A. The combination therapy reduced the hazard ratio to 0.03 (multivariate analysis with adjustments for significant covariates), though it was significant ($P = 0.83$). We also assessed compared patients of groups C and B (splenectomy alone). Administration of IFN for ≥ 12 months reduced the hazard ratio to 0.03 (multivariate analysis after adjustments for significant covariates), but was not significant ($P = 0.83$). These results suggest that the combination of splenectomy plus long-term IFN decreased the likelihood of hepatocarcinogenesis.

Effect of splenectomy and IFN combination therapy on survival

During the observation period, one of the 16 patients of group D (Case 3) died (Fig. 1). The survival rates for groups A, B, C and D were 84.2, 90.9, 87.5 and 100% at the end of the third year, 72.0, 90.9, 87.5 and 100% at the fifth year, 41.4, 36.4, 83.3 and 83.3% at the tenth year, respectively (Fig. 3). The survival rate for patients of group D was the highest compared with the other groups (log-rank test, $P = 0.002$). We also compared the effect of combination therapy on the survival rate of

patients of group A and group D. The survival rate of group D was significantly higher than of group A (log-rank test, $P = 0.004$). We also compared the effect of combination therapy on the survival rate of patients of group C and group D. The survival rate of group D was not significantly different compared with group C (log-rank test, $P = 0.29$). The combination therapy significantly improved the hazard ratio of survival to 9.69 ($P = 0.028$, multivariate analysis with adjustments for significant covariates, Table 2). These results suggest that the splenectomy simply increased the ability for patients to undergo IFN and may not directly improve patient survival.

DISCUSSION

CHRONIC HEPATITIS C virus (HCV) will continue to cause significant morbidity and mortality through to at least 2015.[10] HCV infection remains a common cause of chronic liver disease and is an increasing indication for liver transplantation. Thrombocytopenia (platelet counts $< 150 \times 10^3/\mu L$) is a common complication in patients with chronic liver disease (CLD), and is reported in as many as 76% of cirrhotic patients.[11]The ability to increase platelet levels could significantly reduce the need for platelet transfusions and facilitate the use of IFN-based antiviral therapy and other medically indicated treatments in patients with liver disease. Current treatment options for severe
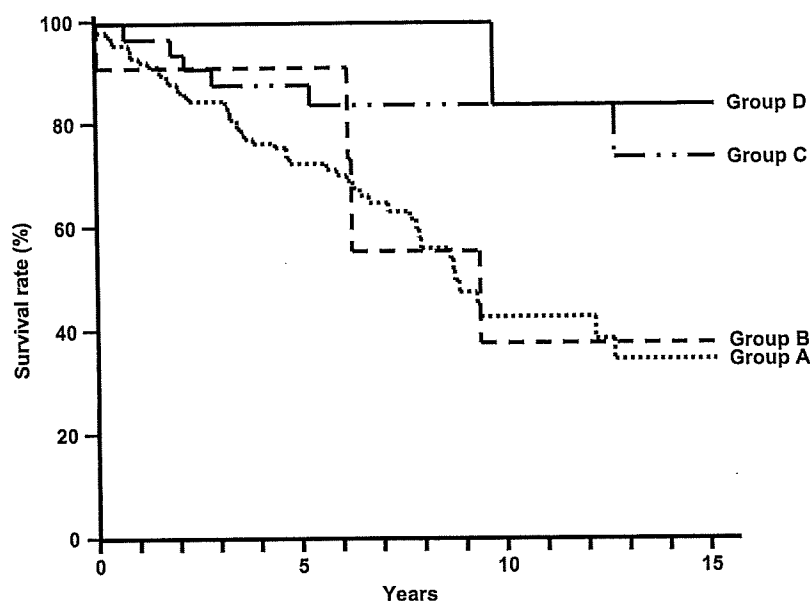
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Figure 3 Survival rates for patients of groups A, B, C and D. The survival rate was significantly different for group A, B, C and D (log-rank test, $P = 0.002$). The survival rate of patients of group D was significantly higher than that of group A (log-rank test, $P = 0.004$).

thrombocytopenia include platelet transfusion, splenic artery embolization and splenectomy. We studied the usefulness of the combination therapy of splenectomy and long-term IFN in patients with advanced HCV-related cirrhosis and thrombocytopenia.

With regard to the usefulness of splenectomy, some studies reported that splenectomy improved PLT counts in cirrhotic patients with thrombocytopenia.[2,3] Furthermore, Shimada *et al.*[12] reported that splenectomy resulted in significant falls in ammonia levels and rises in serum albumin. Thus, there is evidence that splenectomy is beneficial and results in recovery of liver function by improving of blood supply to the liver.[6,13] In the present study, at 6 months after splenectomy, leukocyte count increased 1.6 times, PLT count increased 2.3 times, and total bilirubin decreased nearly 0.6 times,

Table 2 Significance of combined therapy of survival rate in patients of advanced hepatitis C virus-related cirrhosis with low platelet count (time-dependent proportional hazard model)

Factors	Category	Hazard ratio (95% CI)	P
Combined therapy (splenectomy + IFN)	1: no 2: yes	1 9.69 (1.28–76.9)	0.028

IFN, interferon therapy.

relative to prior the procedure. Furthermore, liver function test results also improved in most patients with splenectomy.

With regard to the value of IFN therapy after splenectomy, Hayashi *et al.*[6] reported that splenectomy in patients with HCV cirrhosis can be done safely to allow application of antiviral treatment and potentially avoid transplantation.[6] In this study, only three of 16 (18.8%) patients discontinued IFN therapy after splenectomy. Among the three patients, IFN therapy was discontinued because of thrombocytopenia in only one (6.3%) patient. On the other hand, 13 (81.3%) of the 16 patients on combination therapy were able to complete the full course of IFN therapy, continue IFN therapy or stopped therapy due to NR. Thus, it may be said that IFN therapy is safe in most patients with advanced HCV-related cirrhosis and thrombocytopenia. Furthermore, the present results indicate that splenectomy is an effective method in patients with chronic HCV infection and hypersplenism to increase peripheral leukocyte and platelet counts so that subsequent IFN therapy can be better tolerated. In this study, regarding the reduction of IFN dosages during treatment when comparing group C and D, group D did not have any cases who a reduction in IFN dosages was necessitated by thrombocytopenia ($P = 0.03$). Hayashi *et al.*[6] reported that five of their seven patients underwent splenectomy and then completed a full course of pegylated IFN and ribavirin

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treatment or stopped therapy due to NR, and that none of their patients required dose reductions or treatment discontinuation due to thrombocytopenia. In the present study, the viral response to IFN therapy was SVR in four (36.4%) patients, BR in one (9.1%) and NR in six (54.5%). SVR was not significantly different between group C and D ($P = 0.43$). This result might be a reason that group D had more patients with HCV genotype 1 and higher HCV-RNA than group C. All patients with SVR of group D had genotype 2, suggesting that SVR seems to be achievable by combination therapy in patients with HCV-related cirrhosis with genotype 2 and thrombocytopenia.

We also analyzed the effect of the combination therapy on hepatocarcinogenesis in patients with advanced HCV-related cirrhosis and low PLT count. Chen *et al.*[14] reported that the 5-year tumor-free survival rate was significantly higher after hepatectomy and splenectomy than after hepatectomy alone (37 vs. 27.3%, respectively, $P = 0.003$). In contrast, Yao *et al.*[15] reported that splenectomy in early stage of tumor inoculation stimulated tumor growth and metastasis in their rat model of HCC.[15] In this study, the HCC appearance rate in patients who underwent splenectomy alone (group B) was not significantly different from that of the control (log-rank test, $P = 0.52$). In addition, the HCC appearance rate in patients who received the combination therapy was also not significantly different from the control (log-rank test, $P = 0.50$). We previously reported that long-term IFN therapy for 12 months or longer reduced the rate of hepatocarcinogenesis in patients with liver cirrhosis caused by HCV.[5] Multivariate analysis of long-term follow-up showed that the combination therapy, including IFN administration for ≥ 12 months, decreased the hazard ratio of hepatocarcinogenesis to 0.03, though this was not significant ($P = 0.83$). The reason for the lack of significance might be the small population sample of this study. Yoshida *et al.*[16] reported that IFN therapy significantly reduced the risk for HCC, especially among virologic and biochemical responders. That the combination therapy decreased the hazard ratio of hepatocarcinogenesis to 0.03 suggests the ability of long-term IFN to inhibit HCC, especially among non-responders.

We also examined the effects of the combination therapy on survival. In this study, multivariate analysis using time-dependent variables showed significant improvement of survival in patients who received the combination therapy (group D) compared with the control group (group A) (hazard ratio 3.40, $P = 0.017$; 95% CI 1.24–9.35). This may be considered the crucial

finding of this study. In splenectomy, Morimasa *et al.*[17] reported no difference in survival rate between splenectomy and endoscopic injection sclerotherapy (EIS) for esophageal varices. Similarly, the survival rate in the splenectomy group in this study (group B) was not significantly different from the control ($P = 0.88$). Furthermore, the survival rate of group D was not significantly different compared with group C (log-rank test, $P = 0.29$). These results suggest that the splenectomy increased the ability for patients to undergo IFN and that the combination therapy of splenectomy and long-term IFN significantly improved survival rate in patients with advanced HCV-related cirrhosis and thrombocytopenia. The likely mechanism of action of the combination therapy is first improvement of leucopenia and thrombocytopenia following splenectomy, which allowed administration of IFN, and then IFN produced remission of liver fibrosis, control of necroinflammatory process, and induced suppression of the HCC growth process, consequently leading to improvement of survival rate. Moreno and Muriel[18] reported that IFN resulted in remission of liver fibrosis, and that control of the necroinflammatory process can therefore induce suppression of the HCC growth process. Our results also suggested that patients with NR may need to continue the combination therapy with long-term IFN therapy.

"Pegylated IFN plus ribavirin" and "eltrombopag" are promising drugs and can be potentially used in combination therapy. Recent multicenter trials have demonstrated the superiority of pegylated IFN plus ribavirin compared to pegylated IFN alone or non-pegylated combination therapy.[19,20] In addition, several promising novel agents that stimulate TPO and increase PLT count, such as the oral platelet growth factor eltrombopag, are currently in development for the prevention and/or treatment of thrombocytopenia.[21] Eltrombopag may be a substitute for splenectomy or PSE. Thus, combination therapy of pegylated IFN plus ribavirin after splenectomy or eltrombopag may improve survival rate and reduce the rate of hepatocarcinogenesis.

Our study had certain limitations. In particular, in this study, four (25%) of the patients who underwent combination therapy had a history of splenectomy. A randomized control study with a larger number of cases should be conducted to confirm the effectiveness of this therapy.

In conclusion, the combination therapy of splenectomy and long-term IFN decreased the rate of hepatocarcinogenesis and significantly improved the survival rate in patients with advanced HCV-related cirrhosis and low PLT count.

ACKNOWLEDGMENTS

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GASTROENTEROLOGY

Efficacy of entecavir treatment for lamivudine-resistant hepatitis B over 3 years: Histological improvement or entecavir resistance?

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Abstract

Background and Aims: Long-term lamivudine therapy is required for patients with chronic hepatitis B, because hepatitis reappears frequently after it has withdrawn. However, hepatitis B virus (HBV) mutants resistant to lamivudine emerge frequently accompanied by breakthrough hepatitis.
Methods: Effects of entecavir were evaluated in 19 patients who had developed breakthrough hepatitis during lamivudine therapy for longer than 5 years. This study is a subgroup analysis of a previously reported study. Entecavir, in either 0.5 or 1.0 mg/day doses, was given to 10 and nine patients for 52 weeks, respectively, and then all received 1.0 mg/day entecavir for an additional 68–92 weeks.
Results: There were no differences in biochemical and virological responses in the two groups of patients with respect to the two different initial doses of entecavir. Serum levels of alanine aminotransferase were normalized in 17 (90%) patients, and hepatitis B e antigen (HBeAg) disappeared from the serum in two (14%) of the 14 patients who were HBeAg-positive before. Furthermore, a decrease in histological activity index score greater than 2 points was achieved in nine of the 11 (82%) patients in whom annual liver biopsies were performed during 3 years while they received entecavir. HBV mutants resistant to entecavir emerged in five of the 19 (26%) patients, and hepatitis flare occurred in two of them (40%).
Conclusion: Entecavir in the long term would be useful for histological improvement of breakthrough hepatitis induced by lamivudine-resistant HBV mutants in patients with chronic hepatitis B. However, the relatively high rate of entecavir resistance is a concern, and other strategies need to be considered when available.

Introduction

Worldwide, an estimated 400 million people are infected with hepatitis B virus (HBV) persistently, and some of them develop fatal liver disease, such as decompensated cirrhosis and hepatocellular carcinoma.[1] In 1995, lamivudine was introduced to the treatment of chronic hepatitis B for which interferon (IFN) had previously been the only option.[2,3] Although lamivudine is efficient for treatment of chronic hepatitis B, drug-resistant HBV variants with mutations in the tyrosine–methionine–aspartate–aspartate (YMDD) motif occur increasingly more frequently with treatment duration, to higher than 60% within 5 years.[4–7] Furthermore, these YMDD mutants are often accompanied by breakthrough hepatitis, and it is difficult to obtain disease control with lamivudine.

Subsequently, adefovir dipivoxil has been approved for treatment of chronic hepatitis B,[8,9] and more recently entecavir.[10–12] Entecavir is superior to lamivudine as the first-line treatment, and both adefovir add-on lamivudine and entecavir as switch therapy have also been employed for treatment of breakthrough.[13,14]

The present study represent a subgroup analysis of our previously reported multicenter randomized controlled trial.[12] From a single center, biological and virological responses to entecavir were examined among 19 patients who had developed hepatitis breakthrough during long-term lamivudine therapy, with particular focus on histological responses to entecavir over 3 years and the rate of development of entecavir resistance. Because patients had been randomized to both the low (0.5 mg) and higher (1.0 mg) doses of entecavir, we were also able to compare results between these two different doses.

Methods

Patients

During 10 years from November 1995 to December 2004, 704 patients with chronic hepatitis B received 100 mg lamivudine/day and were followed for more than 5 years in the Department of Hepatology of Toranomon Hospital in metropolitan Tokyo. Lamivudine-resistant YMDD mutants developed in 274 (39%) of the patients, accompanied by breakthrough hepatitis in 176 (64% of those with mutants). Medication was changed so they received the other antivirals. The present study is a subgroup analysis of our previously reported multicenter randomized controlled trial.[12] After entecavir became available, 19 of them were switched to it and the treatment was continued for up to 3 years. None of them were infected with hepatitis C virus (HCV) or HIV type 1, or had autoimmune hepatitis. They were followed for liver function tests and serum markers of HBV infection monthly. At the start of entecavir therapy, chronic hepatitis was diagnosed in them all by liver biopsies performed under laparoscopy and/or ultrasonic imaging; cirrhosis was detected in no patients. Liver biopsies were performed annually for 3 years on 12 of the 19 (63%) patients, for evaluating the efficacy of long-term entecavir in improving histology of the liver. The study design conformed to the 1975 Declaration of Helsinki, and was approved by the ethics committee of the institution. All patients gave their informed consent to participate in this study.

Markers of HBV infection

Hepatitis B surface antigen (HBsAg) and the corresponding antibody (anti-HBs) were determined by hemagglutination (MyCell; Institute of Immunology, Tokyo, Japan), and hepatitis e antigen (HBeAg) by enzyme-linked immunosorbent assay (ELISA) (F-HBe; Sysmex, Kobe, Japan). HBV-DNA was determined by reverse transcription polymerase chain reaction (RT-PCR) with commercial kits (Amplicor, Tokyo, Japan; Roche, Tokyo, Japan), and the result was expressed in log genome equivalents (LGE)/mm with the cut-off value of 2.6 LGE/mL over a dynamic range of 2.6–7.6 LGE/mL. The six major genotypes (A–F) were determined serologically by ELISA (HBV Fenotype EIA; Institute of Immunology). The method employs the combination of epitopes on preS2-region products that is specific for each genotype.[15,16]

Analyses for viral resistance

YMDD mutants were determined by PCR followed by restriction fragment length polymorphism after the method of Chayama *et al*.[4] HBV mutants resistant to entecavir were examined at the baseline and sequentially while patients received entecavir. HBV-DNA was extracted from the serum and amplified by PCR, and nucleotides corresponding to amino acids 1–344 of the reverse transcriptase were sequenced directly by the dideoxy-chain method of Sanger *et al*.[17]

Treatment with entecavir

The 19 patients were randomized to receive two different regimens of entecavir in a double-blind study. Thus, 0.5 and 1.0 mg ente-cavir was given daily to 10 and nine patients, respectively, for the first 52 weeks. Thereafter, patients in both groups received 1.0 mg/day entecavir, and the treatment was continued for an additional 68–92 weeks (120–144 weeks in total).

Response to entecavir

Biochemical response was defined by the normalization of serum alanine aminotransferase (ALT; < 50 IU/L in our laboratory), virological response by the disappearance of HBV-DNA from serum detectable by Amplicor (sensitivity, < 2.6 LGE/mL), and histological response by a decrease in histology activity index (HAI) score of 2 points or more. Necroinflammatory activity and fibrosis were evaluated by the METAVIR score as well.

Statistical analysis

Frequencies were compared between groups by the Mann–Whitney U-test and Fisher's exact test, and medians by the Wilcoxon signed rank test. Normalization in ALT levels and loss of HBV-DNA from the serum, as well as the development of entecavir-resistant HBV mutants, were compared by the method of Kaplan–Meier, and differences were evaluated by the log–rank test with use of the production limit method. $P < 0.05$ was considered significant. Analysis of data was performed with SPSS software (SPSS, Chicago, IL, USA).

Results

Comparison of baseline characteristics between patients given 0.5 and 1.0 mg entecavir daily for 52 weeks and then 1.0 mg for an additional 68–92 weeks

Table 1 compares demographic, biochemical, hematological and virological characteristics between 10 and nine patients with chronic hepatitis B who were randomized to receive 0.5 and 1.0 mg entecavir, respectively, daily for the initial 52 weeks. Thereafter, they all received 1.0 mg entecavir daily for an additional 68–92 weeks (120–144 weeks in total). There were no differences in age, sex, pretreatment ALT levels, platelet counts, frequency of HBeAg, distribution of HBV genotypes, HBV-DNA levels and types of YMDD mutants between the two groups of patients.

Normalization of ALT and loss of HBV-DNA from the serum in patients who received long-term entecavir treatment

Figure 1 a depicts ALT levels in 10 and nine patients who received 0.5 and 1.0 mg entecavir daily, respectively, during the initial 52 weeks; thereafter, they all received 1.0 mg entecavir daily for an additional 68–92 weeks (120–144 weeks in total). In both groups, ALT levels increased slightly during 2–4 weeks after the start of entecavir therapy, and then decreased sharply. ALT levels were lowered within the upper limit of normal (≤ 50 IU/L) 12 and 8 weeks after the start of 0.5 and 1.0 mg entecavir daily, respectively. After then, ALT levels decreased and stayed within the

Table 1 Patients with breakthrough hepatitis induced by lamivudine-resistant hepatitis B virus (HBV) mutants who were treated with two doses of entecavir during the initial 52 weeks

	Initial daily dose of entecavir		
	Total ($n = 19$)	0.5 mg ($n = 10$)	1.0 mg ($n = 9$)
Duration of entecavir (weeks)	120–144	120–144	124–140
Age (years)	38 (29–65)	37 (29–65)	39 (30–49)
Men	17 (89%)	9 (90%)	8 (89%)
ALT (IU/L)	119 (46–1708)	111 (46–1708)	275 (49–442)
Platelets ($\times 10^3/mm^3$)	190 (93–265)	180 (93–235)	190 (108–265)
HBeAg	14 (74%)	7 (70%)	7 (78%)
Genotypes (A : B : C)	1:0:18	1:0:9	0:0:9
HBV-DNA (LGE/mL)	7.2 (5.2–8.6)	7.2 (5.2–8.6)	6.6 (5.7–8.2)
YMDD mutants (I : V : I/V)	11:3:5	6:2:2	5:1:3

Median values are shown with the range in parentheses, and the ratio of HBV genotypes, as well as YIDD, YVDD and both YMDD mutants, is indicated. ALT, alanine aminotransferase; HBeAg, hepatitis e antigen; LGE, log geometric equivalents.
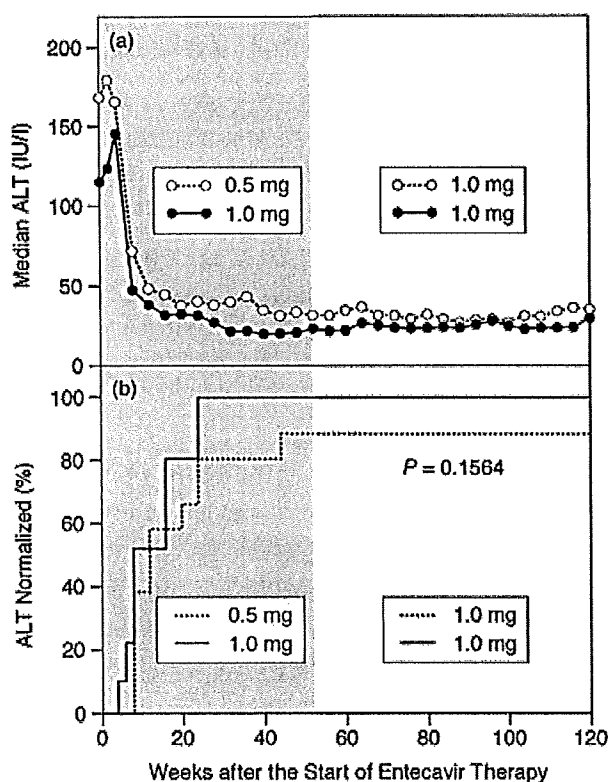

Figure 1 Alanine aminotransferase (ALT) levels in the 19 patients with breakthrough hepatitis induced by lamivudine-resistant hepatitis B virus mutants who received entecavir for 120 weeks. Of them, 10 patients received 0.5 mg and the remaining nine 1.0 mg entecavir daily during the initial 52 weeks (shaded), and thereafter both groups received 1.0 mg entecavir daily. The mean ALT levels (a) and the normalization of serum ALT ($\leq$ 50 IU/L) (b) are illustrated.

normal limit among patients in both groups. Collectively in the 19 patients, the ALT level was normalized in 47% at week 12 and in 83% at week 24. Figure 1(b) compares the normalization of ALT levels between patients who received 0.5 and 1.0 mg entecavir daily during the initial 52 weeks. There were no statistical differences in the normalization of ALT levels between patients given 0.5 and 1.0 mg entecavir. Of the 14 patients positive for HBeAg at the start of entecavir, two (14%) lost HBeAg and seroconverted to anti-HBe, while HBsAg was not cleared from the serum in any of the 19 patients.

The loss of HBV-DNA from serum was compared between patients given 0.5 and 1.0 mg entecavir daily during the initial 52 weeks. A sharp decrease in HBV-DNA by more than 2 logs was achieved at 4 weeks in patients given the initial 0.5 mg entecavir daily, and at 8 weeks in those receiving the initial 1.0 mg entecavir daily. Twenty-four weeks after the start, HBV-DNA levels stabilized and stayed approximately 1 log lower in the patients with the initial 0.5 than 1.0 mg entecavir daily. The loss of HBV-DNA detectable by the quantitative method varied in patients with two different initial entecavir doses. At 24 weeks after the start of entecavir therapy, HBV-DNA became undetectable in 20% and 11%, respectively, of the patients with the initial 0.5 and 1.0 mg entecavir daily; the loss increased to 50% and 33% at 120 weeks, respectively. However, there were no significant differences in the loss of HBV-DNA between the patients receiving 0.5 and 1.0 mg entecavir daily during the initial 25 weeks.

Improvement of liver histology in the patients who were switched to entecavir after the development of breakthrough hepatitis during long-term lamivudine treatment

Of the 19 patients switched to receive entecavir, 12 (63%) underwent serial liver biopsies at the baseline and annually for 3 years while they were treated with entecavir. METAVIR scores for fibrosis stages at the start of entecavir were: F1 in six (50%) patients; F2 in three (25%); and F3 in three (25%). Activity grades were: A1 in six (50%) patients and A2 in six (50%). After they had received entecavir for 1 year, the fibrosis stage improved in two (17%), was

Table 2 Improvement in histology activity scores after entecavir during 3 years in the 12 patients who had developed breakthrough hepatitis induced by lamivudine-resistant HBV mutants

Features		Before	After	Decrement	Differences (*P*-value)
Periportal and/or bridging necrosis	Median (range)	1 (0–3)	0 (0–1)	1 (0–3)	0.003
	Mean ± SD	1.2 ± 0.9	0.1 ± 0.3	1.1 ± 0.8	
Lobular degeneration and focal necrosis	Median (range)	2 (0–3)	1 (0–1)	1 (0–2)	0.014
	Mean ± SD	2.0 ± 1.0	0.9 ± 0.3	1.0 ± 1.0	
Portal inflammation	Median (range)	1 (0–3)	1 (0–1)	1 (0–2)	0.015
	Mean ± SD	1.8 ± 1.0	0.8 ± 0.4	0.9 ± 0.9	
Fibrosis	Median (range)	2 (1–3)	1 (1–3)	0 (0–2)	0.059
	Mean ± SD	2.0 ± 1.0	1.4 ± 0.8	0.5 ± 1.1	
Total HAI score	Median (range)	6 (3–12)	3 (2–5)	3 (1–8)	0.002
	Mean ± SD	7.0 ± 2.7	3.2 ± 0.9	3.5 ± 2.4	

HAI, histology activity index; SD, standard deviation.

unchanged in nine (75%), and worsened in the remaining one (8%). The activity grade improved in nine (75%) patients and was unchanged in the remaining three (25%); it did not worsen in any patient.

One of the 12 patients could not receive liver biopsy 3 years after the start of therapy, because entecavir-resistant HBV mutants developed. Table 2 summarizes changes in HAI scores in the 11 patients who had received long-term entecavir treatment. After 3 years on entecavir therapy, improvement in HAI scores by 2 points or greater was achieved in nine (82%) of the 11 patients. Significant improvement was gained in the total HAI score, as well as scores for periportal and/or bridging necrosis, lobular degeneration/focal necrosis, and portal inflammation. Fibrosis score did not improve significantly ($P = 0.059$); it increased in two patients.

Clinical and virological courses of the representative patient are illustrated in Figure 2 and histological findings in yearly biopsies in Figure 3. The patient developed resistance to lamivudine and was switched to IFN. Hepatitis was exacerbated in him, however, and he was started on lamivudine again. IFN was given intermittently to him when ALT levels were elevated. Because he did not respond to IFN, entecavir was given to him. At that time, he had a HBV-DNA level of more than 7.6 LGE/mL and an HAI score of 8 in the liver biopsy. Soon after entecavir was started, HBV-DNA levels decreased sharply along with the normalization of ALT levels. He seroconverted from HBeAg to anti-HBe 1 year after the start of entecavir treatment. Histological improvement, increasing in parallel with the duration of entecavir treatment, was demonstrated by yearly liver biopsies in comparison with the baseline findings (Fig. 3). Necroinflammatory signs decreased remarkably along with narrowed portal areas, although the stage of fibrosis did not improve appreciably.

HBV mutants resistant to entecavir

Figure 4 illustrates the development of entecavir-resistant HBV mutants that increased in parallel with the duration of treatment. Entecavir-resistant HBV mutants developed in three of the 10 (30%) patients by 18, 84 and 120 weeks; and two of the nine (22%) patients by 132 and 148 weeks, respectively, who received 0.5 and 1.0 mg entecavir daily during the first year; thereafter, they all were given 1.0 mg entecavir daily for the next 68–92 weeks.

During the initial 130 weeks (~2.5 years), therefore, entecavir-resistant HBV mutants developed in three of the 10 (30%) patients with the initial entecavir dose of 0.5 mg daily, in remarkable contrast to no emergence of such mutants in any of the nine patients that received 1.0 mg daily.

Alanine aminotransferase levels were elevated in only two of the five (40%) patients infected with entecavir-resistant HBV mutants, however. These two patients were switched to receive adefovir in combination with lamivudine, and breakthrough hepatitis resolved in them both. All the five patients who developed entecavir-resistant HBV mutants had been infected with lamivudine-resistant YMDD mutants with M204V in the presence or absence of M204I. In outstanding contrast, entecavir-resistant HBV mutants did not develop in any of the 11 patients who had been infected with YMDD mutants with M204I alone.

No adverse effects developed in any of the 19 patients. Breakthrough hepatitis occurred in only one of the five (20%) patients in whom entecavir-resistant mutants emerged.

Discussion

We have previously reported in the Journal that entecavir suppresses serum HBV-DNA to undetectable levels and normalizes ALT levels in more than 30% , respectively, in lamivudine-resistance patients with chronic hepatitis B at 52 weeks.[12] In the present report, we have followed 19 patients from one of the participating centers for 3 years so as to establish longer-term histological efficacy and rates of viral resistance with entecavir treatment of lamivudine-resistant chronic hepatitis B.

As in the earlier report,[12] among the 19 patients described here, ALT levels were normalized in more than 90% of them 8–12 weeks after the start of entecavir until the end of treatment. Although the median HBV-DNA level dropped by 3 logs and remained low during the entecavir therapy, they became undetectable in only 20–40% of the 19 patients. In a previous report, also, the loss of detectable HBV-DNA from the serum was achieved in only 27 of the 141 (19%) patients with lamivudine-resistant HBV mutants after they had received 1.0 mg entecavir daily for 52 weeks.[14] In a remarkable contrast, entecavir is much more efficient in treatment-naive patients who had received it for 1–2 years; HBV-DNA disappeared from the serum in 67–90% of them.[10,11,18] These differences could be attributed to some lamivudine-resistant
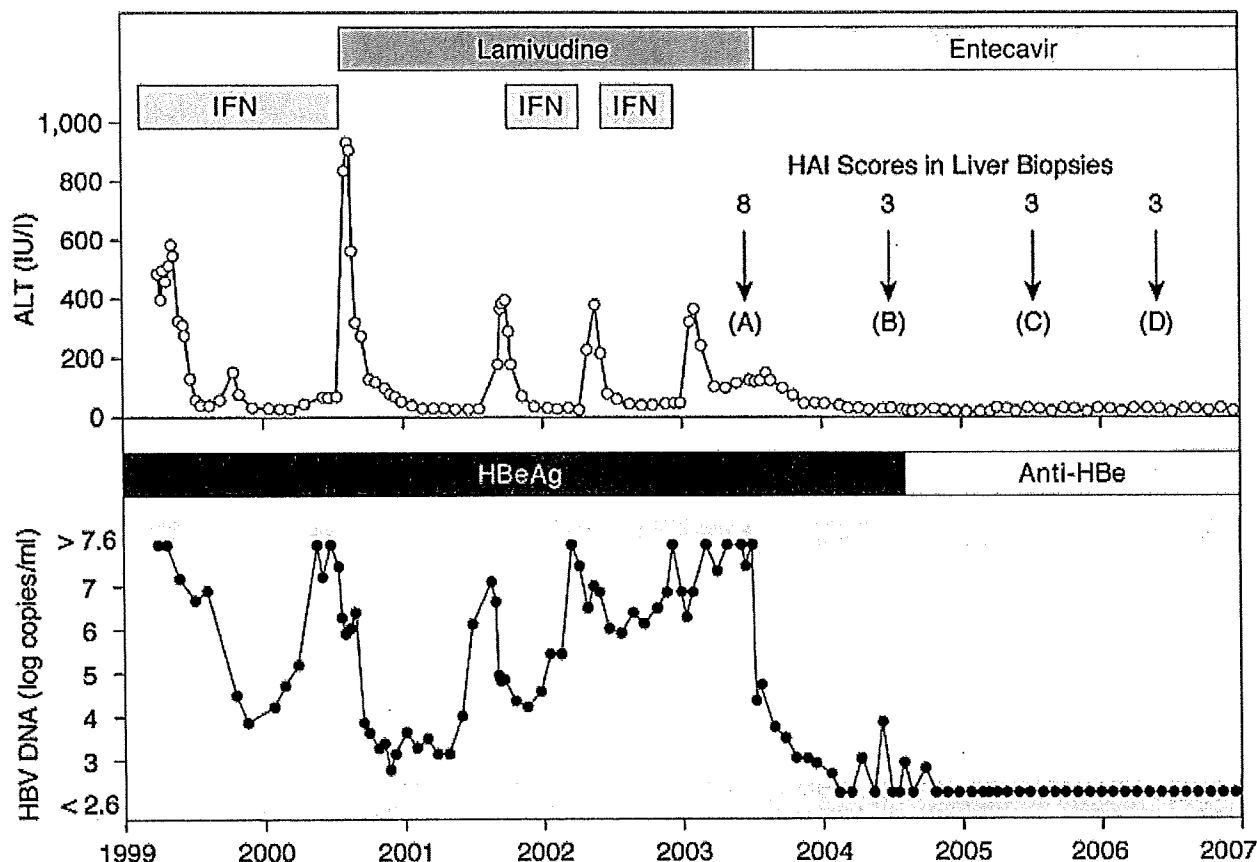
Figure 2 Clinical course of the representative patient. Fluctuating levels of alanine aminotransferase (ALT) and hepatitis B virus (HBV)-DNA are illustrated. Antiviral treatments as well as duration of hepatitis B e antigen (HBeAg) and anti-HBe are indicated by horizontal bars. Also given are time points when four liver biopsies were undertaken, along with histological activity index scores on the top. IFN, interferon.

HBV mutants contributing to the development of entecavir-resistance.[13,18]

Entecavir is a cyclopentyl guanosine analog and can inhibit the polymerase of hepadnaviridae selectively by interfering with priming and reverse transcription, as well as synthesis of minus- and plus-stranded HBV-DNA species.[19] In an *in vitro* expression system with HepG2 cells, entecavir exhibited an antiviral activity with EC_{50} of 0.00375 µM, which is 1500-fold higher than 10 µM of lamivudine.[20] Dose-dependent pharmacological activity of entecavir was evident in a randomized double-blind trial.[21] Although 0.01 mg entecavir daily decreased HBV-DNA by 2.41 logs at 22 weeks, the antiviral activity was significantly lower than 4.31 and 4.72 logs, respectively, of 0.1 and 0.5 mg daily; they were both higher than 3.36 logs by 100 mg lamivudine daily, however. Accordingly, normalization of ALT was more frequent by treatments with 0.1 and 0.5 mg entecavir daily (69% and 83%, respectively) than with 100 mg lamivudine daily (59%).

Significant decrease in HAI scores has been reported in patients with chronic hepatitis B who had received lamivudine for 1–3 years.[22,23] Furthermore, decreases in hepatic inflammation may improve the fibrosis stage. Entecavir therapy for 52 weeks has achieved histological improvement in 55–72% of patients in phase

III clinical trials.[10,11,14] In corroboration of these results, fibrosis stage and inflammation grade improved in the present series of patients who had received entecavir for 3 years, with a significant decrease in the HAI score (Table 2). Histological improvement would have been gained by long-term entecavir therapy, and it may further increase, should entecavir be continued further.

Long-term entecavir treatment, however, may be hampered by the development of drug-resistant mutants. Although entecavir-resistant HBV mutants rarely occur in treatment-naive patients,[18] they can emerge rather frequently in the patients infected with lamivudine-resistant HBV mutants.[14,24] In the present study, entecavir-resistant HBV mutants developed in five of the 19 (26%) lamivudine-resistant patients during 144 weeks of treatment. The incidence was comparable with 32% in the lamivudine-resistant patients who had received entecavir for 3 years.[24] Only two (40%) of them developed hepatitis flares and they were switched to receive adefovir in combination with lamivudine. Entecavir-resistant HBV mutants emerging in patients with lamivudine-resistant mutants are reported to be replication-impaired and rarely induce breakthrough hepatitis.[25] It should be found out how entecavir-resistant HBV mutants can be managed with antiviral nucleos(t)ide analogs other than lamivudine and entecavir, or

433

(a) Before the start of entecavir
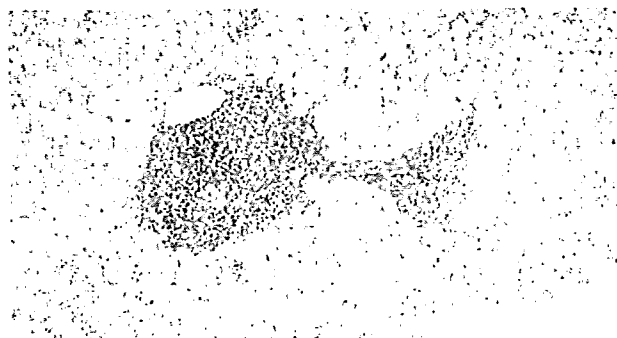
(b) One year after the start of entecavir
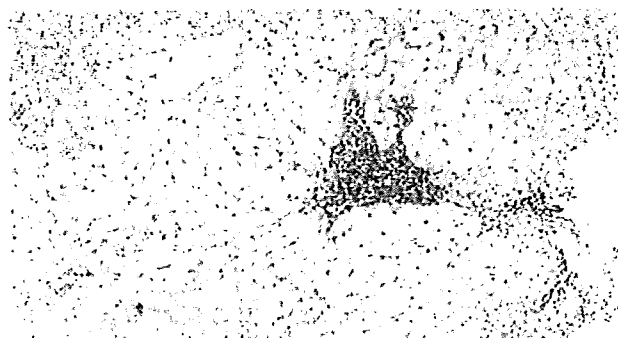
(c) Two years after the start of entecavir
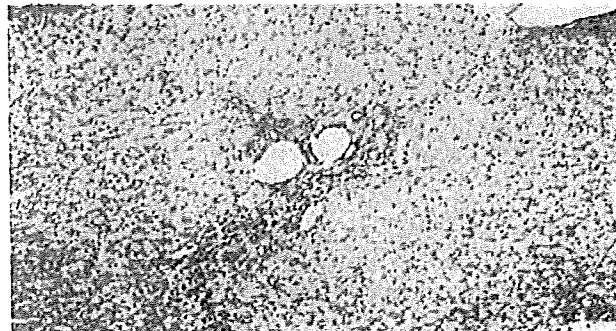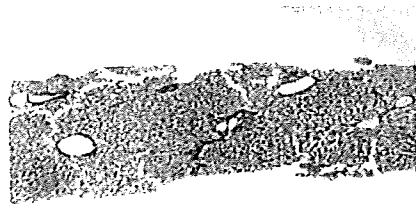
(D) Three years after the start of entecavir
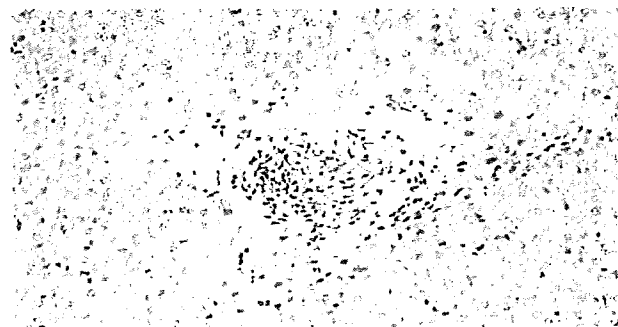
Figure 3 Histological changes in the representative patient during 3-year entecavir treatment (Fig. 2). With hematoxylin–eosin stain on the left, marked enlargement of portal areas is evident along with infiltration of mononuclear cells before the switch from lamivudine to entecavir (a). They decreased increasingly during the 3-year treatment with entecavir (b–d). Stage of fibrosis did not change appreciably by the staining for silver on the right.
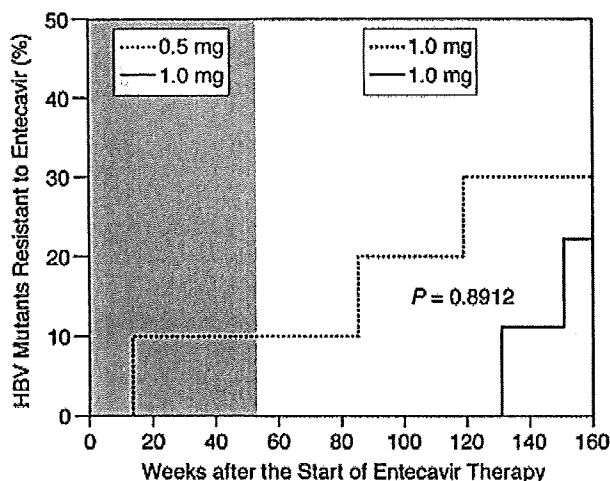
Figure 4 Development of entecavir-resistant hepatitis B virus (HBV) mutants during the 3-year treatment. The 10 patients with the initial entecavir dose of 0.5 mg daily and the nine with that of 1.0 mg daily are compared.

combination thereof. It has been proposed that adefovir add-on lamivudine is efficacious with negligible drug resistance over 3 years.[26,27]

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Original Paper

Intervirology

Association of Amino Acid Substitution Pattern in Core Protein of Hepatitis C Virus Genotype 2a High Viral Load and Virological Response to Interferon-Ribavirin Combination Therapy

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Abstract

Background: Substitution of amino acids (aa) 70 and 91 in the core region of HCV genotype 1b is a useful pretreatment predictor of poor response to interferon + ribavirin combination therapy, but the impacts of aa substitutions in the core region of HCV genotype 2a are still not clear. *Methods:* 154 consecutive Japanese adults with a high viral load (≥100 kIU/ml) of genotype 2a who could complete combination therapy for 24 weeks were evaluated. To examine the differences in virological characteristics between non-sustained virological response (non-SVR) and rapid responder (SVR patients who could achieve a HCV-RNA-negative status within 8 weeks), 86 patients could be analyzed by pretreatment substitution patterns of the core region. *Results:* SVR was achieved in 127 of 154 patients (83%), and rapid response in 113 of 127 (90%). In all 154 patients, multivariate analysis identified younger age, lower level of viremia, and higher level of albumin as significant determinants of SVR. As sig-nificant determinants of rapid response in 86 patients, multivariate analysis identified substitution of aa 4 (non-asparagine) in addition to the significant determinants of SVR. *Conclusions:* Our results suggest that the aa substitution pattern of the core region in patients with a high titer of genotype 2a may partly affect the virological response to combination therapy.

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Introduction

The response to interferon (IFN)-related therapy varies according to hepatitis C virus (HCV) genotype [1, 2]. In Japan, about 70% of patients with chronic hepatitis C are infected with HCV genotype 1b, and about 25% with genotype 2a [3]. Sustained virological response (SVR) to 48-week IFN + ribavirin combination therapy is about 50% in genotype 1b infection, and SVR to 24-week combination therapy is more than 80% in genotype 2 infection [4–9].

IFN + ribavirin combination therapy carries potential serious side effects and is costly, especially when used

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long enough to achieve a high SVR. For these reasons, especially in genotype 2 infection, it is necessary to identify those patients who could achieve SVR with a shorter treatment course (≤16 weeks) to free them of unnecessary side effects and reduce costs, preferably as early as possible [6–8]. Furthermore, we also sometimes encounter treatment-resistant patients infected with genotype 2a [3, 10]. The underlying mechanism(s) of the different virological responses to treatment in patients infected with genotype 2a is still unclear. Hence, the pretreatment predictors of the efficacy of IFN + ribavirin combination therapy were investigated in the present study.

Amino acid (aa) substitutions at position 70 and/or 91 in the HCV core region of infected patients with genotype 1b and a high viral load (≥100 kIU/ml) are predictors of poor virological response to 48- and 72-week pegylated IFN (PEG-IFN) + ribavirin combination therapies [11–15], and also affect clinical outcomes, including insulin resistance and hepatocarcinogenesis [16–18]. However, it is unknown whether the aa substitutions of the core region in patients infected with genotype 2a who have high viral load might also be as useful as the pretreatment virological predictive factors apart from the genotype and viral load.

The present study included 154 Japanese adults with genotype 2a and a high viral load, who received combination therapy for 24 weeks. The aim of the study was to investigate the treatment efficacy and pretreatment predictive factors including virological features.

Materials and Methods

Study Population

A total of 190 HCV genotype 2a-infected Japanese adult patients were consecutively recruited into the study protocol of the combination therapy with IFN (PEG-IFNα-2b or IFNα-2b) + ribavirin for 24 weeks between March 2002 and February 2008 at Toranomon Hospital, Tokyo, Japan. Among these, 154 patients, who could complete a total of 24 weeks of combination therapy, were enrolled in this retrospective study and fulfilled the following criteria: (1) They were negative for hepatitis B surface antigen (radioimmunoassay, Dainabot, Tokyo, Japan), positive for anti-HCV (third-generation enzyme immunoassay, Chiron Corp., Emerville, Calif., USA), and positive for HCV-RNA qualitative analysis with PCR (Amplicor, Roche Diagnostic Systems, Calif., USA). (2) They were naive to ribavirin therapy. (3) They were infected with HCV genotype 2a alone. (4) Each had a high viral load (≥100 kIU/ml) by quantitative analysis of HCV-RNA with PCR (Amplicor GT HCV Monitor v2.0 using the 10-fold dilution method, Roche Molecular Systems, Inc., Pleasanton, Calif., USA) within the preceding 2 months of enrolment. (5) They had no hepatocellular carcinoma. (6) Their body weight was >40 kg. (7) All

were free of coinfection with human immunodeficiency virus. (8) None had been treated with antiviral or immunosuppressive agents within the preceding 3 months of enrolment. (9) None was an alcoholic; lifetime cumulative alcohol intake was <500 kg (mild to moderate alcohol intake). (10) None had other forms of hepatitis, such as hemochromatosis, Wilson's disease, primary biliary cirrhosis, alcoholic liver disease, and autoimmune liver disease. (11) None of the females were pregnant or lactating mothers. (12) All patients had completed a 24-week follow-up program after cessation of treatment, and SVR could be evaluated. (13) Each signed a consent form of the study protocol that had been approved by the Human Ethics Review Committee.

Table 1 summarizes the profiles and data of the 154 patients at the commencement of combination therapy with IFN + ribavirin. The study included 92 men and 62 women, aged 20–70 years (median 53). In all patients, the total duration of treatment was 24 weeks. In 43 of the 154 (27.9%) patients, the dose of ribavirin was reduced during treatment due to a fall in Hb concentration.

With regard to the treatment protocol, 104 (67.5%) patients received PEG-IFNα-2b + ribavirin for 24 weeks, and the remaining 50 (32.5%) patients received IFNα-2b + ribavirin for 24 weeks. They received PEG-IFNα-2b at a median dose of 1.5 μg/kg (range 1.0–1.8) subcutaneously each week, or IFNα-2b at a median dose of 6 million units (range 3–6) intramuscularly each day (7 times per week for the initial 2 weeks, followed by 3 times per week for 22 weeks). They also received oral ribavirin at a median dose of 11.2 mg/kg (range 5.4–14.1) daily.

The treatment efficacy was evaluated by HCV-RNA positivity based on qualitative PCR analysis at the end of treatment (nonvirological response; NVR), and by HCV-RNA negativity based on qualitative PCR analysis at 24 weeks after the completion of therapy (SVR). Furthermore, rapid responders were defined as SVR patients who could achieve a negative status within 8 weeks after the start of treatment, based on qualitative PCR analysis.

Laboratory Tests

Blood samples were obtained at least once every month before, during, and after treatment, and were analyzed for alanine aminotransferase (ALT) and HCV-RNA levels. The serum samples were frozen at −80° within 4 h of collection and thawed at the time of measurement. HCV genotype was determined by PCR using a mixed primer set derived from the nucleotide sequences of the NS5 region [19]. HCV-RNA levels were measured by quantitative PCR (Amplicor GT HCV Monitor v2.0 using the 10-fold dilution method, Roche Molecular Systems, Inc.) at least once every month before, during, and after therapy. The dynamic range of the assay was 5–5,000 kIU/ml. Samples collected during and after therapy that showed undetectable levels of HCV-RNA (<5 kIU/ml) were checked by qualitative PCR (Amplicor HCV v2.0, Roche Molecular Systems, Inc.), which has a higher sensitivity than quantitative analysis, and the results were expressed as positive or negative. The lower limit of the assay was 50 IU/ml.

Histopathological Examination of Liver Biopsies

Liver biopsy specimens were obtained percutaneously or at peritoneoscopy using a modified Vim Silverman needle with an internal diameter of 2 mm (Tohoku University style, Kakinuma Factory, Tokyo, Japan), fixed in 10% formalin, and stained with hematoxylin and eosin, Masson's trichrome, silver impregnation, and periodic acid-Schiff after diastase digestion. All specimens

Table 1. Profile and laboratory data of 154 patients infected with HCV genotype 2a

Demographic data	
Number of patients	154
Sex, M/F	92/62
Age, years*	53 (20–70)
History of blood transfusion	43 (27.9%)
Family history of liver disease	26 (16.9%)
Body mass index*	22.7 (17.9–31.8)
Laboratory data*	
Serum aspartate aminotransferase, IU/l	43 (7–404)
Serum alanine aminotransferase, IU/l	54 (8–651)
Serum albumin, g/dl	3.9 (3.2–4.7)
γ-Glutamyl transpeptidase, IU/l	38 (9–406)
Leukocytes, /mm^3	4,800 (2,200–9,000)
Hemoglobin, g/dl	14.4 (9.9–17.8)
Platelet count, ×10⁴/mm³	17.9 (6.1–32.9)
Indocyanine green retention rate at 15 min, %	13 (4–35)
Serum iron, μg/dl	136 (22–336)
Serum ferritin, μg/l	124 (<10–820)
Level of viremia, kIU/ml	720 (5->5,000)
α-Fetoprotein, μg/l	4 (2–103)
Total cholesterol, mg/dl	174 (107–275)
High-density lipoprotein cholesterol, mg/dl	47 (15–109)
Low-density lipoprotein cholesterol, mg/dl	105 (48–201)
Triglycerides, mg/dl	98 (36–449)
Uric acid, mg/dl	5.6 (2.5–9.4)
Fasting plasma glucose, mg/dl	93 (75–187)
Histological findings	
Stage of fibrosis (F1/F2/F3/ND)	58/23/16/57
Grade of activity (A1/A2/ND)	57/40/57
Hepatocyte steatosis (<5% (absent)/≥5% (present)/ND)	35/52/67
Treatment	
PEG-IFNα-2b/IFNα-2b	104/50
Ribavirin dose, mg/kg*	11.2 (5.4–14.1)
Total duration of treatment, weeks	24
Past history of IFN monotherapy	56 (36.4%)

Data are number and percentages of patients, except those denoted by asterisk (*), which represent the median (range) values. ND = Not determined.

for examinations contained ≥6 portal areas. Histopathological diagnosis was confirmed by an experienced liver pathologist (H. K.) who was blinded to the clinical data. Chronic hepatitis was diagnosed based on histological assessment according to the scoring system of Desmet et al. [20].

Nucleotide Sequencing of the Core Region

We determined the sequences of aa 1–191 in the core region by the direct sequencing method using pretreatment sera of patients who could be analyzed due to adequate serum samples obtained at the start of combination treatment. These sequences were compared with the consensus sequences of genotype 2a, which were determined by comparing the sequences obtained in this study and prototype sequence (HCV J6) [21]. HCV-RNA was extracted from serum samples at the start of treatment and reverse tran-

scribed with random primer and MMLV reverse transcriptase (Takara Syuzo, Tokyo, Japan). Nucleic acids were amplified by nested PCR using the following primers. Nucleotide sequences of the core region: the first-round PCR was performed with 2ACF5 (sense, 5'-GCA AGA CTG CTA GCC GAG TA-3') and 2ACR6 (antisense, 5'-ATC TGA GCT GCG AGC ATC AC-3') primers, and the second-round PCR with 2ACF3N (sense, 5'-CCT TGT GGT ACT GCC TGA TA-3') and 2ACR8 (antisense, 5'-CCA GGT GAT GCT GTC ATT AG-3') primers. All samples were initially denatured at 95° for 2 min. The 35 cycles of amplification were set as follows: denaturation for 30 s at 95°, annealing of primers for 30 s at 55°, and extension for 1 min at 72° with an additional 7 min for extension. Then 1 μl of the first PCR product was transferred to the second PCR reaction. Other conditions for the second PCR were the same as the first PCR, except that the second PCR prim-

Table 2. Factors associated with sustained virological response to combination therapy with IFN + ribavirin for 24 weeks in 154 patients infected with HCV genotype 2a, identified by multivariate analysis

Factor	Category	Odds ratio (95% CI)	p
Age, years	1: ≥50	1	
	2: <50	6.37 (1.76–23.3)	0.005
Serum albumin, g/dl	1: <3.9	1	
	2: ≥3.9	3.19 (1.17–8.73)	0.024
Level of viremia, kIU/ml	1: ≥1,000	1	
	2: <1,000	2.86 (1.11–7.41)	0.030

Only variables that achieved statistical significance (p < 0.05) on multivariate logistic regression are shown.

Table 3. Amino acid substitutions in the core region in non-SVR and rapid response to combination therapy with IFN + ribavirin for 24 weeks in 86 patients infected with HCV genotype 2a

	Non-SVR (n = 25)	Rapid response (n = 61)	p*
Presence of substitution site			
aa 4	1 (4.0%)	15 (24.6%)	0.032
aa 23	2 (8.0%)	0 (0%)	0.082
aa 70	1 (4.0%)	0 (0%)	NS
aa 91	0 (0%)	4 (6.6%)	NS
aa 110	11 (44.0%)	34 (55.7%)	NS

* Non-SVR vs. rapid response (Fisher's exact probability test; statistical significance (p < 0.05), marginal significance (p < 0.10)). aa = Amino acid; SVR = sustained virological response; NS = not significant.

ers were used instead of the first PCR primers. The amplified PCR products were purified by the QIA quick PCR purification kit (Qiagen, Tokyo, Japan) after agarose gel electrophoresis and then used for direct sequencing. Dideoxynucleotide termination sequencing was performed with the Big Dye Deoxy Terminator Cycle Sequencing kit (PerkinElmer, Tokyo, Japan).

To avoid false-positive results, the procedures recommended by Kwok and Higuchi [22] to prevent contamination were strictly applied to these PCR assays. No false-positive results were observed in this study.

Statistical Analysis

Non-parametric tests were used to analyze the aa substitutions in HCV core between the groups, including the Mann-Whitney U test, χ^2 test and Fisher's exact probability test. Uni- and multivariate logistic regression analyses were used to determine the factors that significantly contributed to SVR and rapid response. We also calculated the odds ratios and 95% confidence intervals (CI). All p values <0.05 calculated by the two-tailed test were considered significant. Variables that achieved statistical significance (p < 0.05) or marginal significance (p < 0.10) on univariate analysis were entered into multiple logistic regression analysis to identify significant independent factors. Potential predictive factors associated with SVR included the following variables: sex, age, history of blood transfusion, familial history of liver disease, body mass index, aspartate aminotransferase (AST), ALT, albumin, γ-glutamyl transpeptidase (γGTP), leukocyte count, hemoglobin, platelets, indocyanine green retention rate at 15 min (ICG R15), iron, ferritin, level of viremia, α-fetoprotein, total cholesterol, high-density lipoprotein cholesterol, low-density lipoprotein cholesterol, triglycerides, uric acid, fasting blood glucose, type of IFN, ribavirin dose/body weight, and past history of IFN monotherapy. Furthermore, in addition to potential predictive factors associated with SVR, potential predictive factors associated with a rapid response also included aa substitution in the core region. Statistical analyses were performed using the SPSS software (SPSS, Inc., Chicago, Ill., USA).

Results

Virological Response Rates to Combination Therapy

The virological response could be evaluated in all 154 patients. SVR was achieved in 127 of 154 (82.5%) patients, and rapid response in 113 of 127 (90.0%). Only 5 of 154 (3.2%) patients were considered NVR.

Predictive Factors Associated with SVR in Multivariate Analysis

We then analyzed the data of all 154 patients to determine those factors that could predict SVR. Univariate analysis identified 5 parameters associated with SVR that achieved statistical significance or marginal significance. These included age (<50 years; p < 0.001), serum albumin (≥3.9 g/dl; p = 0.003), level of viremia (<1,000 kIU/ml; p = 0.049), history of blood transfusion (absent; p = 0.064), and ALT (≥30 IU/l; p = 0.088).

Multivariate analysis identified 3 parameters that independently influenced SVR, including age (<50 years; p = 0.005), serum albumin (≥3.9 g/dl; p = 0.024), and level of viremia (<1,000 kIU/ml; p = 0.030) (table 2).

Fig. 1. Sequences of aa 1–30 and aa 61–110 in the core region at the commencement of combination therapy in 86 patients infected with high HCV viral load genotype 2a. Dashes indicate aa identical to the consensus sequence of genotype 2a, and substituted aa are shown by standard single-letter codes. The aa patterns at positions that are probably associated with sensitivity to therapy are shown in boldface characters. NSR = Non-SVR; RR = rapid response.

```
Consensus      10         20         30        70        80        90        100       110
         MSTNPKPQRK TKRNTNRRPQ DVKFPGGGQI//RRQPIPKDRR STGKSWGKPG YPWPLYGNEG LGWAGWLLSP RGSRPSWGPT Efficacy
HCJ6     ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N
Case 1   ---------- ---------- ----------//---------- ------R--  ---------- ---------- --------N  NSR
     2   ---------- ---------- ----------//---------- ------R--  ---------- ---------- --------N  NSR
     3   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   NSR
     4   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  NSR
     5   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  NSR
     6   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   NSR
     7   ---T------ ---------- ----------//------R--H  ---------- ---------- ---------- --------   NSR
     8   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   NSR
     9   ---------- ---------- --R-------//---------- ---------- ---------- ---------- --------   NSR
    10   ---------- ---------- ----------//-------H--  ---------- ---------- ---------- ---H------ NSR
    11   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  NSR
    12   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  NSR
    13   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  NSR
    14   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  NSR
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    19   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   NSR
    20   ---------- SN-------- ----------//---------- ---------- ---------- ---------- --------   NSR
    21   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  NSR
    22   --------Q  ----I----- ----------//---H------ P--------- ---------- Q--------- --------N  NSR
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    25   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  NSR
    26   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  RR
    27   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    28   ---IS----- -H-------- ----------//---------- ---------- ---------- ---------- --------N  RR
    29   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    30   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    31   --------Q  -Q-------- ----------//---------- ---R------ ---------- ---------- --------   RR
    32   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    33   ---V------ ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    34   ---I------ ---------- ----------//---------- T--------- ---------- ---------- --------N  RR
    35   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    36   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    37   ---------- ---------- ----------//-------SQ-  -S-R----- ---------- ---------- --------   RR
    38   ---------- ---------- ----------//---------- ---------- ---------- ---------- ---C------ RR
    39   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    40   ---D------ ---------- ----------//-------H--  ---------- ---------- ---------- -----C---- RR
    41   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    42   ---I------ ----I----- ----------//---------- ---------- ---------- C--------- --------N  RR
    43   ---------- -----I---- ----------//---------- ---------- ---------- ---------- --------N  RR
    44   ---------- -----I---- ----------//---------- ---------- ---------- ---------- --------N  RR
    45   ---------- ---------- ----------//---H------ -P-------- ---------- ---------- --------   RR
    46   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    47   ---------- ---------- ----------//---------- ---R--R--  ---------- ---------- --------   RR
    48   ---Y------ -Q-------- -----------//---------- P-----R-- ---------- ---------- --------N  RR
    49   ---I------ ---------- ----------//------RV--  ---------- ---F----- ---------- G--------- RR
    50   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  RR
    51   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    52   ---------- ---------- --------R-//---------- ---------- ---------- ---------- --------   RR
    53   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  RR
    54   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    55   ---D------ ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    56   ---------I ---------- ----------//---------- ---------- ---------- C---Y---- ----T----  RR
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    58   ---------- -H-------- ----------//---------- ---------- ---------- ---------- --------N  RR
    59   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
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    61   ---------I ---------- ----------//------R--- ---------- ---------- ---------- --------N  RR
    62   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  RR
    63   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------   RR
    64   ---T-----I -----Y---- N---------//---------- P--------- ---------- ---------- --------N  RR
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    68   ---------- ---------- ----------//---------- ------R-- ---------- ---------- --------N  RR
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    72   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------N  RR
    73   ---I------ --------Q  ----------//---------- ---------- ---------- ---------- --------N  RR
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    77   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    78   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    79   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    80   ---I------ ---------- ----------//---------- ---R--R-- ---------- C--------- --------N  RR
    81   ---------- ---------- ----------//---------- ---------- ---------- C--------- --------   RR
    82   ---I------ ---------- ----------//---------- P--------- ---------- ---------- --------S  RR
    83   ---------- -H-------- ----------//-------Q- ---------- ---------- ---------- --------   RR
    84   ---------- ---------- ----------//---------- ---------- ---------- ---------- --------S  RR
    85   ---------- ---------- ----------//---------- -A-------- ---------- ---------- --------S  RR
    86   ---T------ ---------- ----------//---------- ---------- ---------- C--------- --------N  RR
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Table 4. Patient profile and laboratory data of non-SVR and rapid response to combination therapy with IFN + ribavirin for 24 weeks in 86 patients infected with HCV genotype 2a, who could be analyzed by the nucleotide sequences of the core region

	Non-SVR	Rapid response	p^a
Demographic data			
Number of patients	25	61	
Sex, M/F	12/13	39/22	NS
Age, years*	58 (34–64)	51 (20–66)	0.006
History of blood transfusion	10 (40.0%)	15 (24.6%)	NS
Family history of liver disease	4 (16.0%)	10 (16.4%)	NS
Body mass index*	23.1 (19.5–30.0)	22.7 (17.9–31.1)	NS
Laboratory data*			
Serum aspartate aminotransferase, IU/l	31 (19–200)	42 (7–125)	NS
Serum alanine aminotransferase, IU/l	44 (14–357)	53 (8–280)	NS
Serum albumin, g/dl	3.8 (3.2–4.2)	3.9 (3.2–4.5)	0.005
γ-Glutamyl transpeptidase, IU/l	38 (14–141)	39 (10–406)	NS
Leukocytes, /mm³	4,600 (3,100–8,000)	4,800 (2,400–9,000)	NS
Hemoglobin, g/dl	14.2 (12.5–17.8)	14.7 (11.1–17.2)	NS
Platelet count, $\times 10^4$/mm³	16.3 (8.0–32.9)	18.0 (10.6–30.6)	NS
Indocyanine green retention rate at 15 min, %	16 (5–26)	12 (6–35)	NS
Serum iron, μg/dl	152 (30–284)	144 (26–304)	NS
Serum ferritin, μg/l	133 (16–756)	123 (10–820)	NS
Level of viremia, kIU/ml	1,200 (93–>5,000)	680 (5–4,600)	0.053
α-Fetoprotein, μg/l	5 (2–103)	5 (2–48)	NS
Total cholesterol, mg/dl	172 (117–236)	184 (137–264)	NS
High-density lipoprotein cholesterol, mg/dl	49 (27–82)	49 (15–101)	NS
Low-density lipoprotein cholesterol, mg/dl	102 (48–150)	109 (73–198)	NS
Triglycerides, mg/dl	108 (55–276)	97 (39–418)	NS
Uric acid, mg/dl	5.2 (3.2–8.7)	5.7 (2.5–8.7)	NS
Fasting blood glucose, mg/dl	92 (80–126)	93 (77–109)	NS
Treatment			
Ribavirin dose, mg/kg*	11.1 (8.0–12.9)	11.2 (7.3–14.0)	NS
Past history of IFN monotherapy	9 (36.0%)	19 (31.1%)	NS

Data are number and percentages of patients, except those denoted by asterisk (*), which represent the median (range) values.

[a] Non-SVR vs. rapid responder (Mann-Whitney U test or χ^2 test; statistical significance ($p < 0.05$), marginal significance ($p < 0.10$)). SVR = Sustained virological response; NS = not significant.

Treatment Efficacy according to Substitution Patterns in Amino Acids of the HCV Core Region

To examine the differences in virological characteristics between non-SVR and rapid response, 86 patients (25 of 27 non-SVR patients and 61 of 113 rapid responders) could be analyzed by the nucleotide sequences of HCV core region due to adequate serum samples obtained at the start of combination treatment. Figure 1 shows the sequences of aa 1–30 and aa 61–110 of the core region in 86 patients at the commencement of combination therapy. Substitutions at aa 4 (non-asparagine) of HCV core were significantly more frequent in rapid response (n = 15, 24.6%) than non-SVR (n = 1, 4.0%) patients (p = 0.032).

Inversely, substitutions at aa 23 (non-lysine) were more frequent in non-SVR (n = 2, 8.0%) than rapid response (n = 0, 0%) patients (p = 0.082). There were no significant differences in the other substitution sites, including aa 70, aa 91, and aa 110 of the previous report [11], concerning the treatment efficacy of rapid response and non-SVR (table 3).

Predictive Factors Associated with Rapid Response in Multivariate Analysis

We then evaluated the data of all 86 patients who could be analyzed by the nucleotide sequences of core region to determine those factors that could predict rapid re-

Table 5. Factors associated with rapid response to combination therapy with IFN + ribavirin for 24 weeks in 86 patients infected with HCV genotype 2a, identified by multivariate analysis

Factor	Category	Odds ratio (95% CI)	p
Age, years	1: ≥50	1	
	2: <50	7.46 (1.79–31.3)	0.006
Level of viremia kIU/ml	1: ≥1,000	1	
	2: <1,000	4.33 (1.36–13.9)	0.013
Substitution of aa 4	1: asparagine	1	
	2: non-asparagine	9.97 (1.12–89.0)	0.039

Only variables that achieved statistical significance (p < 0.05) on multivariate logistic regression are shown.

sponse. Univariate analysis identified 5 parameters associated with rapid response that achieved statistical significance or marginal significance. As potential predictors of rapid response, table 4 indicates age (<50 years; p = 0.006), serum albumin (≥3.9 g/dl; p = 0.005), and level of viremia (<1,000 kIU/ml; p = 0.053). Furthermore, table 3 shows aa substitution of the core region in the pretreatment sample (substitution of aa 4 [non-asparagine], p = 0.032, and aa 23 [lysine], p = 0.082).

Multivariate analysis identified 3 parameters that independently influenced rapid response, including age (<50 years; p = 0.006), level of viremia (<1,000 kIU/ml; p = 0.013), and substitution of aa 4 (non-asparagine; p = 0.039) (table 5).

Discussion

Previous reports indicated that viral factors (e.g., viral load, aa substitutions in the NS5A region, early viral kinetics, and periods from the start of treatment to initial point of undetectable HCV-RNA) and host factors (e.g., body mass index, fibrosis stage, and level of soluble interleukin-2 receptor) might be important predictors of treatment response to 24-week IFN + ribavirin combination therapy in patients infected with HCV genotype 2a, in addition to treatment-related factors (e.g., treatment duration and ribavirin dose) [6–9, 23–28]. Using multivariate analysis, the present study identified viral- (viral load and substitution of aa 4) and host-related factors (age and serum albumin levels as surrogate markers of liver fibrosis [3, 11]) that influenced the virological response to 24-week combination therapy in patients with genotype 2a

infection and a high viral load. IFN + ribavirin combination therapy carries potential serious side effects and is costly, especially when used long enough to achieve a high SVR. For these reasons, especially in genotype 2 infection, it is necessary to identify those patients who could achieve SVR with a shorter treatment course (≤16 weeks) to free them of unnecessary side effects and reduce costs, preferably as early as possible [6–8]. Identification of these viral and host factors before the start of combination therapy should help design better therapeutic regimens.

Amino acid substitutions at position 70 and/or 91 in the core region of patients with genotype 1b infection and a high viral load are predictors of poor virological response to 48- and 72-week PEG-IFN + ribavirin combination therapy [11–15] and also affect clinical outcomes, including insulin resistance and hepatocarcinogenesis [16–18]. This study, based on 24-week combination therapy in patients with genotype 2a infection and a high viral load, identified substitution of aa 4 in the core region as the significant determinant of treatment efficacy, but did not identify substitutions of aa 70, aa 91, and aa 110. These discrepant findings might be due to the difference of genotype and treatment duration. Other mechanisms could be also explained by the small number of NVR patients with genotype 2a (only 3%), compared with about 25% of patients infected with genotype 1b [11]. Previous studies reported that the core region might be associated with resistance to IFN monotherapy involving the Jak-STAT signaling cascade [29–32]. The present result could also be interpreted to mean that aa substitutions in the core region are associated with those proteins involved in resistance to IFN monotherapy, such as SOCS proteins, which are known to inhibit IFN-α-induced activation of the Jak-STAT pathway and expression of the antiviral proteins 2′,5′-OAS and MxA [33]. Furthermore, this result also indicates that aa substitutions in the core region might serve as surrogate markers for other proteins associated with resistance to the antiviral actions of IFN. One limitation of this study based on the small number of patients was that only the nucleotide sequences of rapid response within SVR patients were analyzed, although all of SVR patients should have been investigated (e.g., possible type II error). Further large-scale studies that examine the structural and functional impacts of aa substitutions during combination therapy should be conducted to confirm the above findings.

In conclusion, our results suggest that the aa substitution pattern in the core region in patients with a high viral load of HCV genotype 2a may partly affect the viro-

logical response to combination therapy. The limitations of this study were that it did not investigate other viral factors, such as the number of substitutions in aa 2193–2228 (the region corresponding to the IFN sensitivity-determining region [ISDR] of genotype 1b [34–36]) or aa 2163–2228 of NS5A in genotype 2a [10, 26, 37], the geographic diversities of the genotype 2a core region (distribution of consensus sequence), and other races apart from Asians in Japan. Further prospective studies, matched for aa substitutions of the core region and large groups of patients of different races, are required to determine the virological response to 24-week IFN + ribavirin combination therapy in patients with a high viral load of HCV genotype 2a.

Acknowledgment

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