

200924011A

厚生労働科学研究費補助金

第3次対がん総合戦略研究事業

血管新生とリンパ管新生の同時制御による
制癌法の確立に関する研究

平成21年度 総括・分担研究報告書

研究代表者 佐藤 靖史

平成22(2010)年 4月

目 次

I. 総括研究報告		
血管新生とリンパ管新生の同時制御による制癌法の確立に関する研究	-----	1
佐藤 靖史		
II. 分担研究報告		
1. 骨髄細胞・内皮前駆細胞に関する研究	-----	4
高倉 伸幸		
2. リンパ管内皮細胞の遺伝子発現に関する研究	-----	6
渡部 徹郎		
III. 研究成果の刊行に関する一覧表	-----	8
IV. 研究成果の刊行物・別刷	-----	11

血管新生とリンパ管新生の同時制御による制癌法の確立に関する研究

研究代表者 佐藤靖史 東北大学加齢医学研究所教授

研究要旨

大腸菌を用いて、活性を保持したレコンビナント Vasohibin-1 蛋白の調整法を確立した。腹腔内など VASH1 の局所療法の有用性を提示した。Vasohibin ファミリーのもう1つの分子である Vasohibin-2 について、VASH-1 と拮抗して血管新生を促進すること、しかも Vasohibin-2 はがん細胞に発現し、腫瘍血管新生を促進して腫瘍発育に寄与していることを明らかにし、Vasohibin-2 が、がん治療の新たな分子標的になることを示した。

分担研究者

高倉伸幸・大阪大学微生物病研究所
教授
渡部徹郎・東京大学大学院医学系研究
科 准教授

A. 研究目的

癌の治療標的として腫瘍血管新生と腫瘍リンパ管新生が注目されている。理想としては血管新生とリンパ管新生の双方を同時に制御することが望ましいが、そのような治療法は未だ確立していない。本研究は、癌治療への Vasohibin ファミリー分子の解析を進めることで、本邦発の新しい治療法として国民の癌医療に大きく貢献することを目的としている。

B. 研究方法

- (1) 大腸菌を用いたレコンビナント Vasohibin-1 蛋白の調整法について、精製方法やリフォールディングなどの条件を検討する。
- (2) 精製された Vasohibin-1 の抗腫瘍活性を、マウスモデルを用いて検証する。
- (3) Vasohibin ファミリーのもう1つの分子である Vasohibin-2 のがん組織における局在を検証する。
- (4) がんにおける Vasohibin-2 の意義を、動物モデルを用いて検証する。
(倫理面への配慮)
全ての動物実験は所属施設での審査を受けた後に行う。

C. 研究結果

- (1) 内皮細胞を障害せずに、広いスペクトルで血管新生とリンパ管新生を抑制する Vasohibin-1 について、大腸菌由来組換え蛋白のリフォールディングによる分散

画分（活性画分）調製法の改良を進め、活性を有する組換え蛋白の調整法を確立した。

(2) 動物モデルにおいて、この組換え Vasohibin-1 蛋白を腫瘍局所に注射すると抗腫瘍効果は得られるが、血中での安定化を目的に PEG 化修飾した蛋白の全身投与では、PEG 化によって活性化が低下するため十分な抗腫瘍効果は得られなかった。一方、腹膜播種すると腹水を形成して急速に個体を死に至らしめるヒト卵巣癌由来 SKOV-3 細胞に Vasohibin-1 遺伝子を安定導入すると、mock コントロールではヌードマウスの腹腔内移植で 4 週間以内に全て死亡するのに対し、Vasohibin-1 遺伝子導入群では腹水も生じず、全て 10 週間以上生存するという顕著な効果を観察した。

(3) Vasohibin-1 ホモログの Vasohibin-2 は、血管新生局所に浸潤する骨髄由来単核球に発現し、Vasohibin-1 と拮抗して血管新生を促進することが判明しているが、癌細胞の一部にも発現することが判明した。そこで Vasohibin-2 発現陰性の腫瘍細胞に Vasohibin-2 を導入してマウスに移植すると、腫瘍血管新生と腫瘍発育が増強することが判明した。

D. 考察

レコンビナント Vasohibin-1 蛋白の調整法は確立できたが、血管をルートとして全身投与する治療法への応用は、現時点では未だ困難である。しかし、腹腔内投与など、局所療法への応用は、可能性の高い手法として、今後検討したい。

今年度の最も大きな収穫は、がんにおける Vasohibin-2 の意義が明らかになったことである。Vasohibin-2 はがん細胞に発現し、腫瘍血管新生を促進して腫瘍発育に寄与している。よって、Vasohibin-1 を外因性に投与すると同時に Vasohibin-2 の

作用を阻害することが、がん治療における望ましい治療法として提唱したい。

E. 結論

レコンビナント Vasohibin-1 蛋白の大量調整法を確立すると共に、がんにおける Vasohibin-2 の意義が明らかにし、Vasohibin-2 が、がん治療の新たな分子標的となえう可能性を示した。がん治療においては、外因性の Vasohibin-1 蛋白を投与に Vasohibin-2 の作用阻害を併用することの有用性が示唆された。

F. 健康危険情報

健康に対する危険性を認めない。

G. 研究発表

1. 論文発表

Heishi T, Hosaka T, Suzuki Y, Miyashita H, Oike Y, Takahashi T, Nakamura T, Arioka S, Mitsuda Y, Takakura T, Hojo K, Matsumoto M, Yamauchi C, Ohta H, Sonoda H, Sato Y. Endogenous angiogenesis inhibitor vasohibin1 exhibits a broad-spectrum anti-lymphangiogenic activity and suppresses lymph node metastasis. *Am. J. Pathol.* 176:1950-1958, 2010.

Tamaki K, Sasano H, Maruo Y, Takahashi Y, Miyashita M, Moriya T, Sato Y, Hirakawa H, Tamaki N, Watanabe M, Ishida T, Ohuchi N. Vasohibin-1 as a potential predictor of aggressive behavior of ductal carcinoma in situ of the breast. *Cancer Sci.* 101:1051-1058, 2010.

Yoshida T, Sato Y, Morita I, Abe M. Pigpen, a nuclear coiled body component protein is involved in angiogenesis. *Cancer Sci.* 2010 (Epub ahead of print).

Miura S, Mitsui K, Heishi T, Shukunami C, Sekiguchi K, Kondo J, Sato Y, Hiraki Y. Impairment of VEGF-A-stimulated lamellipodial extensions and motility of vascular endothelial cells by Chondromodulin-I, a cartilage-derived angiogenesis inhibitor. *Exp Cell Res.* 316: 775-788 2010.

Komi Y, Sogabe Y, Ishibashi N, Sato Y, Moriwaki H, Shimokado K, Kojima S. Acyclic retinoid inhibits angiogenesis via suppressing MAPK pathway. *Lab. Invest.* 90: 52-60, 2010.

Suzuki H, Ohkuchi A, Matsubara S, Takei Y, Murakami M, Shibuya M, Suzuki M, Sato Y. The effect of recombinant PlGF-2 on hypertension induced by full-length mouse sFlt-1 adenoviral vector in pregnant mice. *Hypertension* 54:1129-1135, 2009.

Taniguchi K, Sasaki K, Watari K, Yasukawa H, Imaizumi T, Ayada T, Okamoto F, Ishizaki T, Kato R, Kohno R, Kimura H, Sato Y, Ono M, Yonemitsu Y, Yoshimura A. Suppression of Sproutys has a therapeutic effect for a mouse model of ischemia by enhancing angiogenesis. *PLoS ONE* 4:e5467, 2009.

Hosaka T, Kimura H, Heishi T, Suzuki Y, Miyashita H, Ohta H, Sonoda H, Moriya T, Suzuki S, Kondo T, Sato Y. Vasohibin-1 expressed in endothelium of tumor vessels regulates angiogenesis. *Am. J. Pathol.* 175:430-439, 2009.

Kimura H, Miyashita H, Suzuki Y, Kobayashi M, Watanabe K, Sonoda H, Ohta H, Fujiwara T, Shimosegawa T and Sato Y. Distinctive localization and opposed roles of vasohibin-1 and vasohibin-2 in the regulation of angiogenesis. *Blood* 113:4810-4818, 2009.

Naito H, Kidoya H, Sato Y, and Takakura N. Induction and expression of anti-angiogenic vasohibins in the hematopoietic stem/progenitor cell population. *J. Biochem.* 145: 653-659, 2009.

2. 学会発表

Sato Y. The significance of vasohibin, an angiogenesis inhibitor expressed in endothelium, in cancers and other pathological conditions. NHRI Conference Series 2009 International Conference on Vascular Biology Zhunan, Taiwan 2009.

Sato Y. Vasohibin expressed in endothelium of tumor vessels regulates angiogenesis. 7th International Symposium on the biology of endothelial cells. Viena, Austria 2009.

宮下浩輝, 佐藤靖史. Vasohibin-1 Prevents Cellular Senescence and Maintains Vascular Endothelial Cells. 10th International Symposium on Mechanisms of Vasodilatation, 松島, 2009.

佐藤靖史. 血管を維持する内因性調節機構. 第17回日本血管細胞生物医学学会, 東京, 2009.

木村洋, 宮下浩輝, 鈴木康弘, 小林美穂, 渡辺和秀, 園田光, 大田英樹, 藤原隆, 下瀬川徹, 佐藤靖史. Distinctive Localization and Opposed Roles of Vasohibin-1 and Vasohibin-2 in the Reculation of Angiogenesis. 10th International Symposium on Mechanisms of Vasodilatation, 松島, 2009.

小林美穂, 鈴木康弘, 佐藤靖史. Vasohibin-1-induced detyronation of α -tubulin triggers an inhibition of endothelial cell migration. 第 68 回日本癌学会学術総会. 横浜, 2009.

吉永浩介, 伊藤潔, 八重樫伸生, 佐藤靖史, 笹野公伸, 太田英樹, 園田光. The roles of vasohibin, an intrinsic antiangiogenesis inhibitor, in cervical carcinomas. 第 68 回日本癌学会学術総会. 横浜, 2009.

小林美穂, 鈴木康弘, 佐藤靖史. 血管新生抑制因子 Vasohibin-1 はチューブリンの脱チロシン化を誘導することで内皮細胞の遊走阻害を引き起こす. 第 17 回日本血管細胞生物医学会, 東京, 2009.

高橋詳史, 中村崇宣, 小林美穂, 鈴木康弘, 宮下浩輝, 佐藤靖史. Vasohibin-2 Expressed in Tumors Accelerates Tumor Growth by Promoting Angiogenesis. The 1st International Global COE Symposium, 仙台, 2009.

H. 知的財産権の出願・登録状況

1. 特許

佐藤靖史 固形癌の存在の検出法 特許出願 2009-190830

佐藤靖史, 近藤丘 抗バゾヒビンモノクローナル抗体のセット 特許出願 PCT/JP2009/64862

佐藤靖史 血管新生促進因子 特許出願 PCT/JP2009/051359

2. 実用新案登録

なし

3. その他

なし

骨髓細胞・内皮前駆細胞に関する研究に関する研究

研究分担者 高倉 伸幸 大阪大学微生物病研究所教授

研究要旨: vasohibin-1 は血管新生抑制因子として、抗腫瘍血管新生にむけた臨床応用に期待されている。腫瘍内の血管に特異的に vasohibin-1 をデリバリーする方法論を得るため、腫瘍内皮細胞に特異的に発現する分子の単離を行い、TSR(仮名)を同定した。

A. 研究目的

血管新生抑制因子として単離された vasohibin-1(以下 vash1)を腫瘍内の内皮細胞に特異的かつ大量に運搬するためには腫瘍の内皮細胞に特異的に発現する分子を単離し、これら分子をターゲットとして薬剤を導入する必要がある。本研究では、腫瘍内の内皮細胞に特異的に発現する分子の単離を試みた。

B. 研究方法

低酸素あるいはVEGFで発現が亢進する分子をリストアップし、腫瘍内の内皮細胞特異的発現性を解析した。

(倫理面への配慮)

本研究はマウスを用いた解析であり、実験動物の使用に関しては、大阪大学微生物病研究所の定める倫理規定に則って研究を行った。

C. 研究結果

本研究においてほぼ腫瘍領域全般に内皮細胞全体に発現するTSRを単離した。本TSRは内皮細胞膜に発現する受容体であり、その結合因子TSRLも同じく腫瘍内の内皮細胞に同時に発現が観察された。TSRは従来より、血管内皮細胞成長因子(VEGF)によって発現が誘導され、血管新生時に血管拡大に機能することを報告してきたが、血管透過性の抑制効果があることも解明された。

D. 考察

TSRに対する抗体、あるいはTSRLを利用して腫瘍内の血管内皮細胞にvash1の導入が可能であると考えられる。PEG化リポソームの表面にTSRLをコーティングしたDDS法を構築しており、本法によるvash1のデリバリーに向け検討を継続する。

E. 結論

TSRは腫瘍内の血管内皮細胞特異性が高く腫瘍特異的DDSに応用可能と考えられた。

F. 健康危険情報

G. 研究発表

1. 論文発表

Han Y, Ueno M, Nagahama Y, Takakura N. Identification and characterization of stem cell-specific transcription of *PSF1* in spermatogenesis. *Biochem Biophys Res Commun*. 380:609-613, 2009.

Ueno M, Itoh M, Sugihara K, Asano M, and Takakura N. Both alleles of *PSF1* are required for maintenance of pool size of immature hematopoietic cells and acute bone marrow regeneration. *Blood* 113:555-562, 2009.

Takakura N and Kidoya H. Maturation of blood vessels by haematopoietic stem cells and progenitor cells : involvement of apelin/APJ and Angiopoietin/Tie2 interactions in vessel caliber size regulation. *Thrombosis and Haemostasis* 101: 999-1005, 2009.

Naito H, Kidoya H, Sato Y, Takakura N. Induction and expression of anti-angiogenic vasohibins in the hematopoietic stem/progenitor cell population. *J Biochem*. 145: 653-659, 2009.

Katoh SY, Kamimoto T, Yamakawa D, Takakura N. Lipid rafts serve as signaling platforms for Tie2 receptor tyrosine kinase in vascular endothelial cells. *Exp Cell Res*. 315: 2818-2823, 2009.

Nagahama Y, Ueno M, Miyamoto S, Morii E, Minami T, Mochizuki N, Saya H, Takakura N. PSF1, a DNA replication factor expressed widely in stem and progenitor cells, drives tumorigenic and metastatic properties. *Cancer Res*. 70: 1215-1224. 2010.

Murakami A, Takasugi H, Ohnuma S, Koide Y, Sakurai A, Takeda S, Hasegawa T, Sasamori J, Konno T, Hayashi K, Watanabe Y, Mori K, Sato Y, Takahashi A, Mochizuki N, Takakura N. Sphingosine 1-Phosphate Regulates Vascular Contraction via S1P3 Receptor: Investigation Based on a New S1P3 Receptor Antagonist. Mol Pharmacol. in press.

Nagahama Y, Ueno M, Haraguchi N, Mori M, Takakura N. PSF3 marks malignant colon cancer and has a role in cancer cell proliferation. Biochem Biophys Res Commun. 392: 150-154, 2010.

Kido H, Naito H, and Takakura N. Apelin induces enlarged and non-leaky blood vessels for functional recovery from ischemia. Blood, in press.

2. 学会発表

Takakura N : Endogenous negative feedback regulation of angiogenesis by micro-RNA, 第 68 回日本癌学会学術総会、 2009 年 10 月 1-3 日、横浜、ほか 7 件

H. 知的財産権の出願・登録状況
(予定を含む。)

1. 特許取得
なし
2. 実用新案登録
なし
3. その他
なし

リンパ管内皮細胞の遺伝子発現に関する研究

分担研究者 渡部 徹郎 東京大学大学院医学系研究科・准教授

研究要旨 癌の悪性化を抑制するためには血管新生とリンパ管新生の両方を同時に制御できる治療法を開発することが急務であり、内因性の血管・リンパ管新生抑制因子である Vasohibin-1 に注目が集まっている。本研究では Vasohibin ならびにさまざまな調節因子による血管・リンパ管新生抑制作用の分子機構の解明を試みた。本年度は主にリンパ管新生を調節する Ets-2 ならびに HoxD8 転写因子の機能についての研究を行い、Ets-2 の機能を阻害することによってリンパ管新生が阻害されることを見出した。

A. 研究目的

リンパ管は末梢組織で血管から漏出した間質液、タンパク質、細胞などを血管系へと環流することにより血液の量や組成を一定に保ち、成体の恒常性を維持するとともに、癌組織から細胞を運んでリンパ節転移を引き起こしたりもする。このためリンパ管形成機構の解明は医学的に必要性が高いが、その研究の歴史は比較的新しく、未解明な点が多く残されている。本研究においては Vasohibin ならびに他の調節因子によるリンパ管新生抑制作用の分子機構の解明を試みるために、本年度は血管新生において重要な役割を果たす Ets-2 と HoxD8 転写因子に注目し、リンパ管新生における作用について検討した。

B. 研究方法

我々はヒト臍帯静脈内皮細胞 (HUVEC) ならびにヒト皮膚由来リンパ管内皮細胞 (HDLEC) を用いて Ets-1 の作用を検討した。HUVEC ならびに HDLEC において Ets-2 ならびに HoxD8 の発現を増減させるためにそれぞれアデノウイルスと siRNA を用いた。さらに個体レベルで両者のリンパ管新生に対する作用を検討するためにマウス横隔膜上の新生リンパ管において腹腔内に注射したアデノウイルスによりそれぞれの転写因子を発現させた。

(倫理面への配慮)

ヒトの遺伝子解析ならびに相手方の同意を得る研究は本研究計画には含まれていない。

C. 研究結果

我々は昨年度 Ets-1 がリンパ管新生を誘導することを報告したが、Ets-2 の発現を低下させてもリンパ管新生は抑制されなかった。しかし Ets-2 の発現を低下させるとリンパ管新生に重要な役割を果たす PDGF 受容体 (PDGFRβ) の発現が低下し、リンパ管新生が抑制された。さらにマウス個体レベルで Ets-2 の機能を阻害する Tm-Ets 変異体を発現させることによりリンパ管新生が抑制されることが確認された (Yoshimatsu, Watabe et al., 投稿準備中)。

さらに Prox1 の標的遺伝子として HoxD8 転写因子を同定し、HoxD8 が Prox1 の発現を亢進するポジティブフィードバック因子であることを明らかにした。さらに HoxD8 をマウス個体のリンパ管において発現するとその管径が増大することを見出した (Harada, Watabe et al., 2009, Journal of Cell Science)。

D. 考察

以上の結果から Ets-2 ならびに HoxD8 がリンパ管新生を抑制するための新たな標的となりうることが示唆された。

E. 結論

Ets-2 と HoxD8 はリンパ管内皮細胞においてそれぞれ Prox1 の機能と発現を調節することにより VEGFR3 などのリンパ管新生シグナル伝達因子の発現調節を行ない、リンパ管新生を誘導していることが明らかになった。

F. 健康危険情報

該当なし

G. 研究発表

1. 論文発表
1. Suzuki Y, Ohga N, Morishita Y, Hida K, Miyazono K, Watabe T: BMP-9 induces proliferation of multiple types of endothelial cells in vitro and in vivo. *Journal of Cell Science*. 2010, in press.
2. Harada K, Yamazaki T, Iwata C, Yoshimatsu Y, Sase H, Mishima K, Morishita Y, Hirashima M, Oike Y, Suda T, Miura N, **Watabe T**, Miyazono K. Identification of targets of Prox1 during in vitro vascular differentiation from embryonic stem cells: functional roles of HoxD8 in lymphangiogenesis. *Journal of Cell Science*. 2009, 122: 3923-3930. 2009
3. Yamamizu K, Kawasaki K, Katayama S, **Watabe T**, Yamashita JK: Enhancement of vascular progenitor potential by protein kinase A through dual induction of Flk-1 and Neuropilin-1. *Blood* 114: 3707-3716. 2009
4. Sase H, **Watabe T**, Kawasaki K, Miyazono K, Miyazawa K: VEGFR2-PLCγ axis is essential for endothelial specification of VEGFR2+ vascular progenitor cells. *Journal of Cell Science*. 122: 3303-3311. 2009
5. Saito RA, **Watabe T**, Horiguchi K, Kohyama T, Saitoh M, Nagase T, Miyazono K: TTF-1 inhibits TGF-β-mediated epithelial-to-mesenchymal transition in lung adenocarcinoma cells. *Cancer Research* 69:2783-2791. 2009
6. Yamazaki T, Yoshimatsu Y, Morishita Y, Miyazono K, **Watabe T**: COUP-TFII regulates the functions of Prox1 in lymphatic endothelial cells through direct interaction. *Genes to Cells* 14:425-434.2009

2. 学会発表

国際学会および集会

The 36th International Congress on Physiological Sciences (Kyoto) 2009年7月28~31日

Watabe T. Roles of signal and transcriptional networks during formation of lymphatic vessels. (Symposium)

The 7th Japan-Korean Joint

Symposium on Vascular Biology (Seoul, Korea) 2009年8月20~21日

Watabe T, Yoshimatsu Y, Yamazaki T, Morikawa M, Miyazono K. Roles of Ets family members in the regulation of Prox1-mediated lymphatic differentiation of endothelial cells. (Oral)

● 国内学会および集会

第68回日本癌学会学術総会(横浜)2009年10月1~3日

Watabe T, Yoshimatsu Y, Yamazaki T, Morikawa M, Miyazono K. Roles of transcriptional networks during the formation of lymphatic vessels. (International session)

第17回日本血管生物医学会(東京)2009年10月8~9日

Watabe T, Kokudo T, Suzuki Y, Yoshimatsu Y, Yamazaki T, Miyazono K. Snail is required for TGF-β-induced endothelial-mesenchymal transition of embryonic stem cell-derived endothelial cells (Symposium)

第32回日本分子生物学会年会(横浜)2009年12月9~12日

Watabe T, Kokudo T, Suzuki Y, Yoshimatsu Y, Yamazaki T, Miyazono K. Snail is required for TGF-β-induced endothelial-mesenchymal transition of embryonic stem cell-derived endothelial cells (Workshop)

H. 知的財産権の出願・登録状況

1. 特許取得
特になし
2. 実用新案登録
特になし
3. その他
特になし

研究成果の刊行に関する一覧表レイアウト

書籍

著者氏名	論文タイトル名	書籍全体の編集者名	書籍名	出版社名	出版地	出版年	ページ

雑誌

発表者氏名	論文タイトル名	発表誌名	巻号	ページ	出版年
Suzuki H, Ohkuchi A, Matsubara S, Takei Y, Murakami M, Shibuya M, Suzuki M, Sato Y.	The effect of recombinant PlGF-2 on hypertension induced by full-length mouse sFlt-1 adenoviral vector in pregnant mice.	Hypertension	54	1129-1135	2009
Taniguchi K, Sasaki K, Watari K, Yasukawa H, Imaizumi T, Ayada T, Okamoto F, Ishizaki T, Kato R, Kohno R, Kimura H, Sato Y, Ono M, Yonemitsu Y, Yoshimura A.	Suppression of Sproutys has a therapeutic effect for a mouse model of ischemia by enhancing angiogenesis.	PLoS ONE	4	e5467	2009
Hosaka T, Kimura H, Heishi T, Suzuki Y, Miyashita H, Ohta H, Sonoda H, Moriya T, Suzuki S, Kondo T, Sato Y.	Vasohibin-1 expressed in endothelium of tumor vessels regulates angiogenesis.	Am. J. Pathol.	175	430-439	2009
Kimura H, Miyashita H, Suzuki Y, Kobayashi M, Watanabe K, Sonoda H, Ohta H, Fujiwara T, Shimosegawa T and Sato Y.	Distinctive localization and opposed roles of vasohibin-1 and vasohibin-2 in the regulation of angiogenesis.	Blood	113	4810-4818	2009
Naito H, Kidoya H, Sato Y, and Takakura N.	Induction and expression of anti-angiogenic vasohibins in the hematopoietic stem/progenitor cell population.	J. Biochem.	145	653-659	2009
Miura S, Mitsui K, Heishi T, Shukunami C, Sekiguchi K, Kondo J, Sato Y, Hiraki Y.	Impairment of VEGF-A-stimulated lamellipodial extensions and motility of vascular endothelial cells by Chondromodulin-I, a cartilage-derived angiogenesis inhibitor.	Exp Cell Res.	316	775-788	2010

Komi Y, Sogabe Y, Ishibashi N, Sato Y, Moriwaki H, Shimokado K, Kojima S.	Acyclic retinoid inhibits angiogenesis via suppressing MAPK pathway.	Lab. Invest.	90	52-60	2010
Heishi T, Hosaka T, Suzuki Y, Miyashita H, Oike Y, Takahashi T, Nakamura T, Arioka S, Mitsuda Y, Takakura T, Hojo K, Matsumoto M, Yamauchi C, Ohta H, Sonoda H, Sato Y.	Endogenous angiogenesis inhibitor vasohibin1 exhibits a broad-spectrum anti-lymphangiogenic activity and suppresses lymph node metastasis.	Am. J. Pathol.	(Epub ahead of print)		2010
Tamaki K, Sasano H, Maruo Y, Takahashi Y, Miyashita M, Moriya T, Sato Y, Hirakawa H, Tamaki N, Watanabe M, Ishida T, Ohuchi N.	Vasohibin-1 as a potential predictor of aggressive behavior of ductal carcinoma in situ of the breast.	Cancer Sci.	101	1051-1058	2010
Yoshida T, Sato Y, Morita I, Abe M.	Pigpen, a nuclear coiled body component protein is involved in angiogenesis.	Cancer Sci.	(Epub ahead of print)		2010
Han Y, Ueno M, Nagahama Y, Takakura N.	Identification and characterization of stem cell-specific transcription of PSF1 in spermatogenesis.	Biochem Biophys Res Commun.	380	609-613	2009
Ueno M, Itoh M, Sugihara K, Asano M, and Takakura N.	Both alleles of <i>PSF1</i> are required for maintenance of pool size of immature hematopoietic cells and acute bone marrow regeneration.	Blood	113	555-562	2009
Takakura N and Kidoya H	Maturation of blood vessels by haematopoietic stem cells and progenitor cells : involvement of apelin/APJ and Angiopoietin/Tie2 interactions in vessel caliber size regulation.	Thrombosis and Haemostasis	101	999-1005	2009
Katoh SY, Kamimoto T, Yamakawa D, Takakura N.	Lipid rafts serve as signaling platforms for Tie2 receptor tyrosine kinase in vascular endothelial cells.	Exp Cell Res.	315	2818-2823	2009
Nagahama Y, Ueno M, Miyamoto S, Morii E, Minami T, Mochizuki N, Saya H, Takakura N.	PSF1, a DNA replication factor expressed widely in stem and progenitor cells, drives tumorigenic and metastatic properties.	Cancer Res.	70	1215-1224	2010
Nagahama Y, Ueno M, Haraguchi N, Mori M, Takakura N.	PSF3 marks malignant colon cancer and has a role in cancer cell proliferation.	Biochem Biophys Res Commun.	392	150-154	2010

Murakami A, Takasugi H, Ohnuma S, Koide Y, Sakurai A, Takeda S, Hasegawa T, Sasamori J, Konno T, Hayashi K, Watanabe Y, Mori K, Sato Y, Takahashi A, Mochizuki N, Takakura N.	Sphingosine 1-Phosphate Regulates Vascular Contraction via S1P3 Receptor: Investigation Based on a New S1P3 Receptor Antagonist.	Mol Pharmacol.	In press		
Kidoya H, Naito H, and Takakura N.	Apelin induces enlarged and non-leaky blood vessels for functional recovery from ischemia.	Blood	In press		
Harada K, Yamazaki T, Iwata C, Yoshimatsu Y, Sase H, Mishima K, Morishita Y, Hirashima M, Oike Y, Suda T, Miura N, Watabe T, Miyazono K	Identification of targets of Prox1 during in vitro vascular differentiation from embryonic stem cells: functional roles of HoxD8 in lymphangiogenesis.	Journal of Cell Science	122	3923-3930	2009
Yamamizu K, Kawasaki K, Katayama S, Watabe T, Yamashita JK	Enhancement of vascular progenitor potential by protein kinase A through dual induction of Flk-1 and Neuropilin-1.	Blood	114	3707-3716	2009
Sase H, Watabe T, Kawasaki K, Miyazono K, Miyazawa K	VEGFR2-PLCgamma1 axis is essential for endothelial specification of VEGFR2+ vascular progenitor cells.	Journal of Cell Science	122	3303-3311	2009
Saito RA, Watabe T, Horiguchi K, Kohyama T, Saitoh M, Nagase T, Miyazono K	TTF-1 inhibits TGF- β -mediated epithelial-to-mesenchymal transition in lung adenocarcinoma cells.	Cancer Research	69	2783-2791	2009
Yamazaki T, Yoshimatsu Y, Morishita Y, Miyazono K, Watabe T	COUP-TFII regulates the functions of Prox1 in lymphatic endothelial cells through direct interaction.	Genes to Cells	14	425-434	2009
Suzuki Y, Ohga N, Morishita Y, Hida K, Miyazono K, Watabe T	BMP-9 induces proliferation of multiple types of endothelial cells in vitro and in vivo.	Journal of Cell Science	in press		2010

Hypertension

JOURNAL OF THE AMERICAN HEART ASSOCIATION

American Heart
Association® 

Learn and Live SM

Effect of Recombinant Placental Growth Factor 2 on Hypertension Induced by Full-Length Mouse Soluble fms-Like Tyrosine Kinase 1 Adenoviral Vector in Pregnant Mice

Hirotsada Suzuki, Akihito Ohkuchi, Shigeki Matsubara, Yuji Takei, Masato Murakami, Masabumi Shibuya, Mitsuaki Suzuki and Yasufumi Sato

Hypertension 2009;54;1129-1135; originally published online Sep 28, 2009;

DOI: 10.1161/HYPERTENSIONAHA.109.134668

Hypertension is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75214

Copyright © 2009 American Heart Association. All rights reserved. Print ISSN: 0194-911X. Online ISSN: 1524-4563

The online version of this article, along with updated information and services, is located on the World Wide Web at:

<http://hyper.ahajournals.org/cgi/content/full/54/5/1129>

Data Supplement (unedited) at:

<http://hyper.ahajournals.org/cgi/content/full/HYPERTENSIONAHA.109.134668/DC1>

Subscriptions: Information about subscribing to Hypertension is online at
<http://hyper.ahajournals.org/subscriptions/>

Permissions: Permissions & Rights Desk, Lippincott Williams & Wilkins, a division of Wolters Kluwer Health, 351 West Camden Street, Baltimore, MD 21202-2436. Phone: 410-528-4050. Fax: 410-528-8550. E-mail:
journalpermissions@lww.com

Reprints: Information about reprints can be found online at
<http://www.lww.com/reprints>

Pregnancy Hypertension/Preeclampsia

Effect of Recombinant Placental Growth Factor 2 on Hypertension Induced by Full-Length Mouse Soluble fms-Like Tyrosine Kinase 1 Adenoviral Vector in Pregnant Mice

Hirotada Suzuki, Akihide Ohkuchi, Shigeki Matsubara, Yuji Takei, Masato Murakami, Masabumi Shibuya, Mitsuaki Suzuki, Yasufumi Sato

Abstract—The first aim of our study was to develop a pregnant mouse model for preeclampsia using adenoviral vector containing mouse full-length soluble fms-like tyrosine kinase 1 (sFlt-1) but not truncated sFlt-1. The second aim was to evaluate effects of recombinant mouse (rm) vascular endothelial growth factor (VEGF) and rm placental growth factor (PIGF) on a preeclampsia model induced by adenoviral vector containing mouse full-length sFlt-1. We injected adenoviral vector containing mouse full-length sFlt-1 on day 8.5 or 9.5 of gestation into pregnant Institute of Cancer Research mice, resulting in hypertension, proteinuria, and similar glomerular histological changes as those seen in human preeclamptic women with glomerular endotheliosis on day 16.5 or 17.5 of gestation. The preeclampsia models were treated with 100 $\mu\text{g}/\text{kg}$ of rmVEGF164 (n=5), 100 $\mu\text{g}/\text{kg}$ of rmPIGF-2 (n=5), or vehicle (n=7) twice a day for 2 days IP. The rmVEGF164 treatment significantly decreased the mean blood pressure on day 16.5 or 17.5 of gestation compared with the vehicle treatment (85 ± 4 versus 97 ± 2 mm Hg; $P=0.018$). The rmPIGF-2 treatment also significantly decreased the mean blood pressure on day 16.5 or 17.5 of gestation compared with the vehicle treatment (86 ± 3 versus 97 ± 2 mm Hg; $P=0.018$). However, proteinuria was not affected by either rmVEGF164 or rmPIGF-2. In conclusion, we, for the first time, created a mouse preeclampsia model using mouse full-length sFlt-1. VEGF and PIGF may be promising for ameliorating hypertension in women with preeclampsia. Additional study of PIGF as a potential drug for preeclampsia is warranted. (*Hypertension*. 2009;54:1129-1135.)

Key Words: adenoviral vector ■ soluble fms-like tyrosine kinase 1 ■ vascular endothelial growth factor ■ placental growth factor ■ preeclampsia ■ animal models ■ therapy

Preeclampsia is associated with maternal and infantile morbidity and mortality.^{1,2} It has been shown that the concentration of soluble fms-like tyrosine kinase 1 (sFlt-1), a circulating antiangiogenic protein, is increased in women with preeclampsia,^{3,4} and increased levels of sFlt-1 and reduced levels of free placental growth factor (PIGF) are potentially useful for predicting the subsequent development of preeclampsia.^{4,5} sFlt-1 acts by adhering to the receptor-binding domains of vascular endothelial growth factor (VEGF)-A and PIGF, preventing their interaction with endothelial receptors on the cell surface. Recent studies have indicated that patients with cancer receiving anti-VEGF antibody therapy may have an increased incidence of proteinuria and hypertension because of a decrease in their circulating VEGF levels.⁶ Nonpregnant and pregnant rodents administered anti-VEGF antibodies or sFlt-1 manifested proteinuria and hypertension.^{3,7,8} These results strongly indicate that increases in sFlt-1 and decreases in VEGF/PIGF in the maternal circulation may cause the occurrence of preeclampsia.

sFlt-1, a human natural soluble form of the VEGF receptor (VEGFR) 1, is produced in conditioned culture medium of human umbilical vein endothelial cells⁹ and in the trophoblasts.^{10,11} sFlt-1 is encoded by the flt-1 gene and is truncated between N-terminal immunoglobulin-like domains 6 and 7.¹² Because the N-terminal first and second domains of Flt-1 are necessary and sufficient for the binding of VEGF-A with near-native affinity,^{13,14} truncated sFlt-1¹⁻³ containing the first to third domains, but not full-length sFlt-1, has been used for the studies evaluating the effect of sFlt-1 on hypertension and proteinuria in mouse or rat models.^{3,7,15-19} However, there are 2 differences between the full-length sFlt-1 and truncated sFlt-1¹⁻³: first, the full-length sFlt-1 has a 31-amino acid carboxyl lesion derived from an intron, which is significantly homologous to that in mammals,¹¹ and, second, the truncated sFlt-1¹⁻³ lacks the immunoglobulin-like loop 4, which is essential to stabilize receptor dimerization of the extracellular domains of Flt-1, in addition to VEGF.^{13,14} Therefore, the effect of sFlt-1 on the occurrence of hyperten-

Received April 27, 2009; first decision May 17, 2009; revision accepted September 1, 2009.

From the Department of Obstetrics and Gynecology (H.S., A.O., S.M., Y.T., M.Su.), Jichi Medical University School of Medicine, Tochigi, Japan; Division of Genetics (M.M., M.Sh.), Institute of Medical Science, University of Tokyo, Tokyo, Japan; Institute of Development, Aging, and Cancer (Y.S.), Department of Vascular Biology, University of Tohoku, Sendai, Japan.

Correspondence to Akihide Ohkuchi, Department of Obstetrics and Gynecology, Jichi Medical University School of Medicine, 3311-1 Yakushiji, Shimotsuke-shi, Tochigi 329-0498, Japan. E-mail okuchi@jichi.ac.jp

© 2009 American Heart Association, Inc.

Hypertension is available at <http://hyper.ahajournals.org>

DOI: 10.1161/HYPERTENSIONAHA.109.134668

Downloaded from hyper.ahajournals.org at TOHOKU UNIVERSITY on April 6, 2010

sion or proteinuria may be different between truncated sFlt-1 and natural full-length sFlt-1.

It has been reported that the effect of excess circulating sFlt-1 can be ameliorated by the administration of recombinant VEGF-A.^{7,19} However, in humans, a decrease in PIGF is related to the later occurrence of preeclampsia.^{20–22} Therefore, we also hypothesized that not only VEGF-A, but also PIGF, may play a pivotal role in the occurrence of hypertension and proteinuria in both humans and rodents. To our knowledge, the effect of the administration of PIGF into a rodent model of preeclampsia via a mouse (m)-sFlt-1 adenoviral vector has not been examined.

First, we evaluated an adenovirus encoding the full-length mouse-sFlt-1 gene (Ad m-sFlt-1) for the induction of hypertension and proteinuria in pregnant mice. Second, we evaluated the effects of recombinant mouse (rm) VEGF164 (rm-VEGF164) and rmPIGF-2 on hypertension and proteinuria in a mouse preeclampsia model induced by Ad m-sFlt-1.

Methods

An expanded Materials and Methods section is available in the online Data Supplement (available at <http://hyper.ahajournals.org>). Briefly, in the first experiment, an Ad m-sFlt-1, which was created in our previous study,²³ and an adenovirus encoding β -galactosidase gene (Ad LacZ) were propagated in HEK293 cells. The viral lysates were purified and concentrated through 2 cycles of CsCl step gradients.²⁴ Nine- to 12-week-old CD1 (Institute of Cancer Research) mice were injected in the tail vein with 3×10^8 plaque-forming unit (PFUs; low dose), 1×10^9 PFUs (medium dose), or 2×10^9 PFUs (high dose) of Ad m-sFlt-1 ($n=9$, $n=6$, and $n=6$, respectively) and with 3×10^8 PFUs (low dose), 1×10^9 PFUs (medium dose), or 2×10^9 PFUs (high dose) of Ad LacZ ($n=7$, $n=6$, and $n=7$, respectively) on day 8.5 or 9.5 of gestation. The control pregnancy mice were not injected with anything ($n=7$). The mean blood pressures (MBPs) were measured by the tail-cuff method (Softron Ltd) on 4 different days: (1) before mating, (2) just before the injection of adenovirus, (3) on day 13.5 or 14.5 of gestation, and (4) on day 16.5 or 17.5 of gestation. The urine albumin:creatinine (Alb/Cr) ratios on day 16.5 or 17.5 of gestation were measured. In the second experiment, pregnant mice were injected in their tail vein with 2×10^9 PFUs (high dose) of Ad m-sFlt-1 on day 8.5 or 9.5 of gestation. rmVEGF164 (100 μ g/kg diluted in 500 μ L of PBS; $n=5$) was administered IP twice a day for 2 days from the evening on day 14.5 or 15.5 of gestation. In other mice, rmPIGF-2 (100 μ g/kg diluted in 500 μ L of PBS; $n=5$) and the vehicle (500 μ L of PBS; $n=7$) were administered IP twice a day for 2 days. The MBP and Alb/Cr ratio were also measured. All of the animal housing and experiments were approved by the institutional animal care and research advisory committee of both the University of Tohoku and Jichi Medical University. The pharmacokinetics of rmVEGF164 and rmPIGF-2 in nonpregnant mice and pregnant mice are shown in the online Data Supplement (available at <http://hyper.ahajournals.org>, Figure S1A through S1D).

Results

Expression of Proteins by the Adenoviral Vector

In mice administered Ad LacZ, β -galactosidase activity was observed in the liver but not in the placenta by 5-bromo-4-chloro-3-indolyl β -D-galactoside staining, suggesting that Ad m-sFlt-1 was expressed in the liver. The levels of mouse sFlt-1 (nanograms per milliliter) on day 16.5 or 17.5 of gestation increased significantly in the medium and high doses of Ad m-sFlt-1 compared with the medium and high doses of Ad LacZ, respectively (85 [58 to 95] versus, 12 [12 to 29], $P=0.002$; 93 [82 to 130] versus 25 [21 to 33], $P=0.001$, respectively; Figure 1A). The levels of mouse sFlt-1 on day 16.5 or 17.5 of gestation

were not different among the control and the mice administered the low, medium, and high dose of Ad LacZ ($P=0.76$ by Kruskal-Wallis test).

Plasma Levels of Angiogenic Factors in Mice Administered Ad m-sFlt-1

The plasma levels of mouse VEGF-A (picograms per milliliter) on day 16.5 or 17.5 of gestation in the mice administered high-dose Ad m-sFlt-1 were significantly lower than in the mice administered high dose Ad LacZ (47 [43 to 52] versus 95 [93 to 113]; $P=0.001$; Figure 1B). On the contrary, the levels of mouse PIGF-2 (picograms per milliliter) on day 16.5 or 17.5 of gestation were almost the same among the control mice, the high-dose Ad LacZ group, and the high-dose Ad m-sFlt-1 group (21 [14 to 26], 25 [21 to 35], and 30 [23 to 38]; Figure 1C).

Blood Pressure and Proteinuria in Pregnant Mice Administered Ad m-sFlt-1

In the control mice and the mice administered low, medium, and high doses of Ad LacZ, MBP (millimeters of mercury) was almost the same during the prepregnancy period and during pregnancy (Figure 1D). In the mice administered high-dose Ad m-sFlt-1, MBP was significantly increased between day 8.5 or 9.5 and day 13.5 or 14.5 (76 ± 2 versus 91 ± 4 ; $P=0.028$) and between day 13.5 or 14.5 and day 16.5 or 17.5 (91 ± 4 versus 101 ± 3 ; $P=0.028$), although such increases were not seen in the mice administered low or medium doses of Ad m-sFlt-1 (Figure 1E). The MBP on day 16.5 or 17.5 of gestation in the high-dose Ad m-sFlt-1 group was significantly higher than that in the high-dose Ad LacZ group (101 ± 3 versus 81 ± 11 ; $P=0.010$) and the control group (83 ± 4 ; $P=0.005$). The urine Alb/Cr ratios (milligrams per gram) on day 16.5 or 17.5 of gestation in the mice administered low, medium, and high doses of Ad m-sFlt-1 were significantly increased compared with those in the mice administered low, medium, and high doses of Ad LacZ, respectively (5.8 [4.5 to 14] versus 2.7 [2.4 to 2.9], $P=0.030$; 92 [43 to 148] versus 10 [4.9 to 22], $P=0.015$; 58 [30 to 161] versus 5.2 [4.0 to 13], $P=0.020$, respectively), although the urine Alb/Cr ratio in the low-dose Ad m-sFlt-1 group was not significantly different from that in the control mice (8.3 [3.6 to 11]; Figure 1F).

Histopathology in Mice Administered Ad m-sFlt-1

Glomerular histologies viewed by light microscopy in mice administered high-dose Ad m-sFlt-1 (Figure S2D and S2E) and those in the mice administered high-dose Ad LacZ (Figure S2A and S2B), scanning electron microscopy of glomerulus in mice administered high-dose Ad m-sFlt-1 and Ad Lac Z (Figure S2F and S2C), and the mean fetal and placental weights among the control mice, the high-dose Ad LacZ group, and the high-dose Ad m-sFlt-1 group (Figure S2G and S2H) were shown in the online Data Supplement. These results were written in the online Data Supplement.

Relationship Among the Plasma Levels of Mouse sFlt-1, Mouse VEGF-A, Blood Pressure, and Proteinuria

Among all of the data in control mice, mice administered Ad LacZ, and mice administered Ad m-sFlt-1, there was an

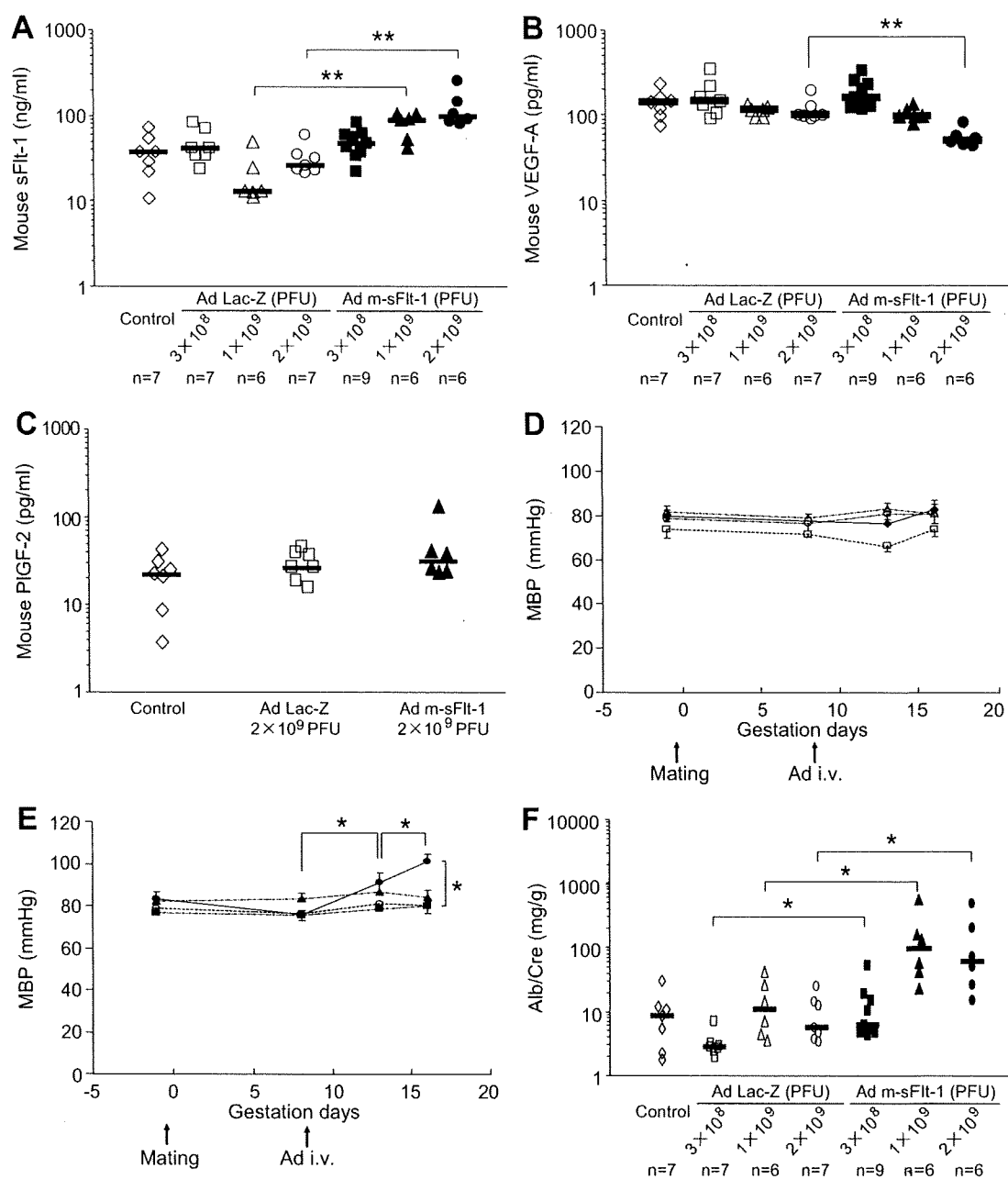


Figure 1. Data from pregnant mice administered nothing, Ad LacZ, and Ad m-sFlt-1. A, Plasma levels of m-sFlt-1 on day 16.5 or 17.5 of gestation. B, Plasma levels of m-VEGF-A (picograms per milliliter) on day 16.5 or 17.5 of gestation. C, Plasma levels of m-PlGF-2 (picograms per milliliter) on day 16.5 or 17.5 of gestation. D, MBP (millimeters of mercury) during the pre-pregnancy period, on day 8.5 or 9.5 of gestation, on day 13.5 or 14.5 of gestation, and on day 16.5 or 17.5 of gestation. \diamond , \square , \triangle , and \circ represent the values in the control pregnant mice and pregnant mice administered low-dose (3×10^8 PFU), medium-dose (1×10^9 PFU), and high-dose (2×10^9 PFU) Ad LacZ, respectively. E, MBP (millimeters of mercury) during the pre-pregnancy period, on day 8.5 or 9.5 of gestation, on day 13.5 or 14.5 of gestation, and on day 16.5 or 17.5 of gestation. \circ , \blacksquare , \blacktriangle , and \bullet represent the values in pregnant mice administered high-dose Ad LacZ and mice administered low-dose, medium-dose, and high-dose Ad m-sFlt-1, respectively. F, Urine Alb/Cre ratios (milligrams per gram) on day 16.5 or 17.5 of gestation. * $P < 0.05$; ** $P < 0.01$.

inverse relationship between the plasma levels of \log_{10} sFlt-1 and \log_{10} VEGF-A ($r = -0.29$; $P = 0.042$; Figure 2A); positive relationships between the plasma levels of \log_{10} sFlt-1 and MBP ($r = 0.24$; $P = 0.098$; Figure 2B) and between the plasma levels of \log_{10} sFlt-1 and urine \log_{10} (Alb/Cre; $r = 0.44$; $P = 0.002$; Figure 2C); and inverse relationships between the plasma levels of \log_{10} VEGF-A and MBP ($r = -0.33$; $P = 0.023$; Figure 2D) and between the plasma

levels of \log_{10} VEGF-A and urine \log_{10} (Alb/Cre; $r = -0.44$; $P = 0.002$; Figure 2E). Thus, the circulating levels of sFlt-1 are significantly positively related to the degree of proteinuria, whereas the circulating levels of VEGF-A are significantly inversely related to both blood pressure and proteinuria. Our results indicate that decreased levels of circulating VEGF-A cause the increases in blood pressure seen in the mice administered Ad m-sFlt-1.

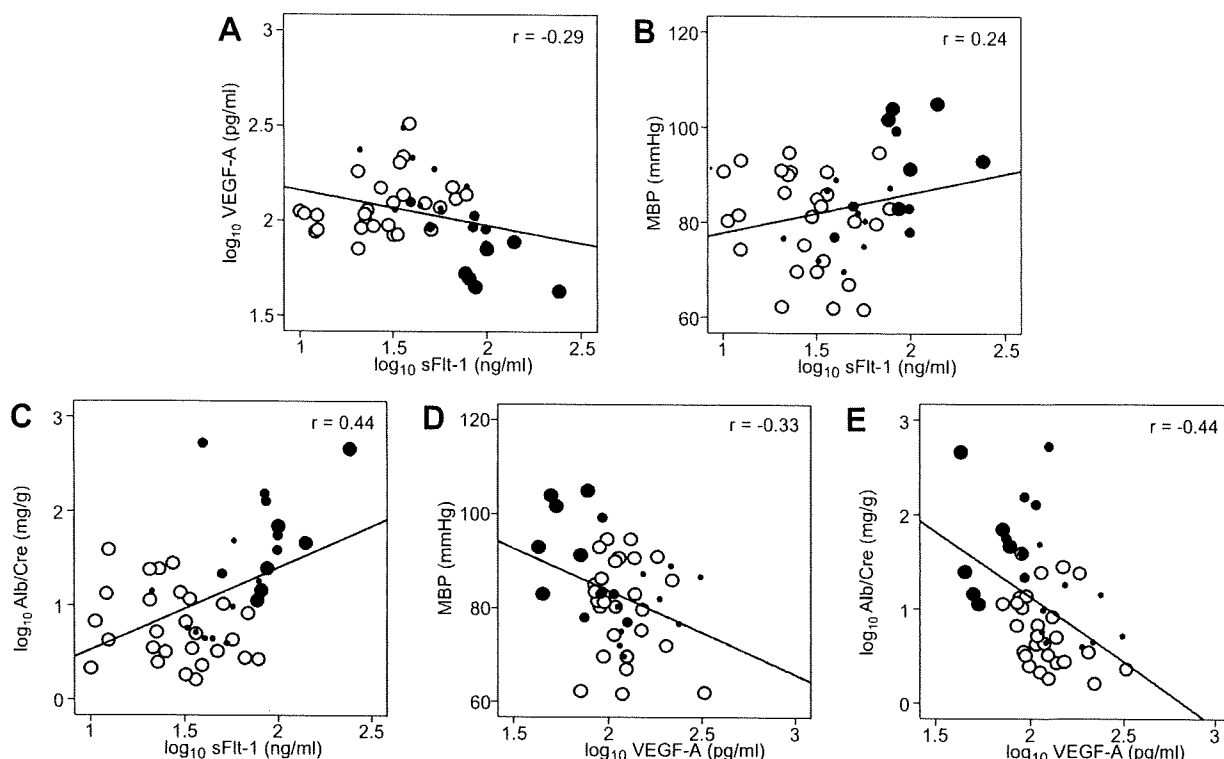


Figure 2. Scattergrams showing the relationship between the plasma levels of \log_{10} sFlt-1 and \log_{10} VEGF-A (A; $r = -0.29$; $P = 0.042$), between the plasma levels of \log_{10} sFlt-1 and MBP (B; $r = 0.24$; $P = 0.098$), between the plasma levels of \log_{10} sFlt-1 and urine \log_{10} (Alb/Cre) (C; $r = 0.44$; $P = 0.002$), between the plasma levels of \log_{10} VEGF-A and MBP (D; $r = -0.33$; $P = 0.023$), and between the plasma levels of \log_{10} VEGF-A and urine \log_{10} (Alb/Cre) (E; $r = -0.44$; $P = 0.002$). The large open circles represent combined data from the control mice and mice administered low, medium, and high doses of Ad LacZ. The small, medium, and large closed circles represent data from mice administered low, medium, and high doses of Ad m-sFlt-1, respectively.

Effects of rmVEGF164 and rmPIGF-2 in Mice Administered High-Dose Ad m-sFlt-1

In pregnant mice administered high-dose Ad m-sFlt-1 compared with mice administered vehicle, the levels of mouse VEGF-A (pg/ml) 3 to 4 hours after the last administration of rmVEGF164 were significantly increased (356 [121 to 793] versus 59 [56 to 68]; $P = 0.010$; Figure 3A), and the level of mouse PIGF-2 (pg/ml) 3 to 4 hours after the last administration of rmPIGF-2 was also significantly increased (244 [133 to 244] versus 60 [52 to 74]; $P = 0.010$; Figure 3B).

The rmVEGF164 treatment significantly decreased the MBP (mm Hg) on day 16.5 or 17.5 of gestation compared with the vehicle treatment (85 ± 4 versus 97 ± 2 ; $P = 0.018$; Figure 3C). The rmPIGF-2 treatment also significantly decreased the MBP on day 16.5 or 17.5 of gestation compared with the vehicle treatment (86 ± 3 versus 97 ± 2 ; $P = 0.018$). However, the urine Alb/Cre levels were not affected by treatment with either rmVEGF164 or rmPIGF-2 (Figure 3D). The rmVEGF164 treatment and rmPIGF-2 treatment did not ameliorate glomerular histology viewed by light microscopy in pregnant mice administered high-dose Ad m-sFlt-1 compared with those administered vehicle.

Discussion

In this study, we created a pregnant mouse model of preeclampsia, showing hypertension, proteinuria, and glomerular change, like endotheliosis, by transfecting a high dose

(2×10^9 PFUs) of adenovirus encoding full-length m-sFlt-1. In addition, we revealed that rmVEGF164 and rmPIGF-2 ameliorate the hypertension induced by the administration of a high dose of Ad m-sFlt-1 in pregnant mice.

Development of a Pregnant Mouse Model of Preeclampsia Using Ad m-sFlt-1 and the Relationships Between the Serum Levels of m-sFlt-1/m-VEGF-A/m-PIGF-2 and Hypertension/Proteinuria

We, for the first time, created a mouse preeclampsia model using full-length m-sFlt-1 instead of truncated m-sFlt-1, which has been used in previous mouse preeclampsia models.^{3,7,15-19} In the previous rat models using truncated m-sFlt-1, both hypertension and proteinuria emerged after low-dose (1×10^8 -PFU) administration of Ad m-sFlt-1.³ On the contrary, we needed a higher dose of Ad m-sFlt-1 to generate both hypertension and proteinuria. In our study, increases in the plasma levels of sFlt-1 were related to the occurrence of proteinuria, and decreases in the plasma levels of VEGF-A were related to the occurrence of both hypertension and proteinuria. Therefore, the circulating levels of sFlt-1 and VEGF-A may be important for the occurrence of hypertension and proteinuria. It is possible that the in vivo expression of sFlt-1 in the liver per administered dose of adenovirus or the circulating levels of sFlt-1 or VEGF-A were different between the 2 studies.

It is well known that the administration of sFlt-1 into rodents results in the occurrence of proteinuria.^{3,7,16,25} Kamba

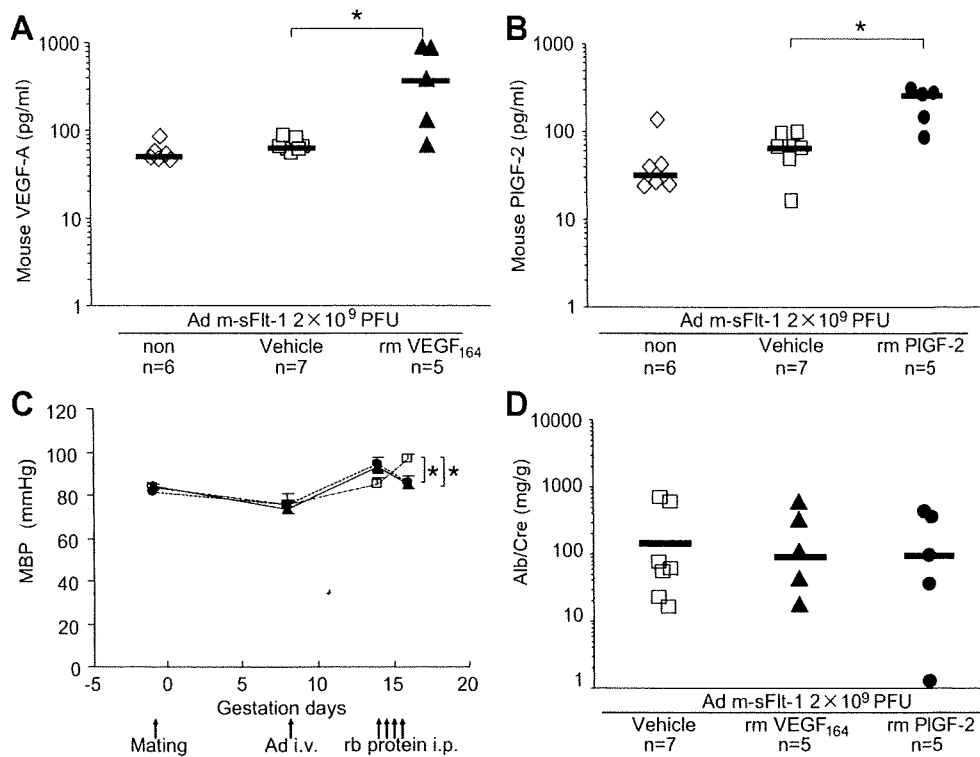


Figure 3. Data in pregnant mice administered high-dose Ad m-sFlt-1 after treatment with vehicle, rmVEGF164, or rmPIGF-2. A, The levels of mouse VEGF-A 3 to 4 hours after the last administration of nothing, vehicle (twice a day, 4 times), or rmVEGF164 (100 μg/kg, twice a day, 4 times). B, The levels of mouse PlGF-2 3 to 4 hours after the last administration of nothing, vehicle (twice a day, 4 times), or rmPIGF-2 (100 μg/kg, twice a day, 4 times). C, MBP (millimeters of mercury) during the prepregnancy period, just before the injection of Ad m-sFlt-1, just before the administration of recombinant proteins or vehicle, and at 1 to 2 hours after the last administration of recombinant proteins or vehicle. □, ▲, and ● represent the values in pregnant mice administered a high dose of Ad m-sFlt-1 with vehicle (n=7), rmVEGF164 (n=5), or rmPIGF-2 (n=5), respectively. D, Urine Alb/Cre ratios on day 16.5 or 17.5 of gestation in pregnant mice administered a high dose of Ad m-sFlt-1 with vehicle, rmVEGF164, or rmPIGF-2. *P<0.05.

et al²⁵ reported that proteinuria, but not hypertension, emerged in nonpregnant mice administered adenovirus-truncated m-sFlt-1. Sugimoto et al⁷ reported that IV administration of an sFlt-1/Fc chimera protein into nonpregnant mice resulted in proteinuria 3 hours after the administration. Maynard et al³ reported that IV administration of adenovirus-truncated m-sFlt-1 into nonpregnant and pregnant rats resulted in severe proteinuria. In addition, the effect of adenovirus-truncated m-sFlt-1 in pregnant rats on the occurrence of proteinuria was more severe than that of adenovirus-soluble endoglin, both of which can cause both hypertension and proteinuria in pregnant mice.¹⁶ However, the proteinuria presented in this mouse model is fairly modest compared with what is presented in the rats where the Alb/Cre ratio is frequently >1000 mg/g.³ One possibility is that the dose using the full-length sFlt1 is not enough, because it tends to have a poor bioavailability in contrast to truncated sFlt1. Another possibility is that proteinuria may depend on the background of the mouse strains used.

In our experiment, the decrease in the circulating levels of VEGF-A, but not PlGF-2, was related to the increase in MBP. The effect of VEGF-A on blood pressure has been reported in humans.^{26–29} The blocking of signal transduction of VEGF-A by anti-VEGF monoclonal antibody^{26,27} and tyrosine kinase inhibitors, such as sorafenib²⁸ and sumatinib,²⁹ induces hyper-

tension. On the contrary, the administration of rmVEGF-A results in a decrease of MBP in rats.^{30,31} Thus, an appropriate circulating VEGF-A level appears to be important for the maintenance of normal blood pressure. Although the detailed mechanism by which a decreased level of VEGF-A affects the occurrence of hypertension has not been elucidated, the modulation of the production of NO by VEGF-A via endothelial NO synthase activity in endothelial cells may be related to the change in blood pressure.^{32,33}

The mouse PlGF-2 was not altered in this mouse model of preeclampsia. The circulating levels of PlGF-1 are low in preeclamptic women,^{3–5,19–21} because the human ELISA assay measured free PlGF but not total PlGF. We could not know whether the assay of the mouse PlGF-2 in our study measured a free or total PlGF-2 assay. We speculate that the assay of the mouse VEGF-A measured a free VEGF-A and that the assay of the mouse PlGF-2 measured a total PlGF-2, because the levels of VEGF-A were decreased but the levels of PlGF-2 were not decreased by the administration of high-dose Ad m-sFlt-1.

Effect of rmVEGF164 and rmPIGF-2 on Blood Pressure and Proteinuria in a Pregnant Mouse Preeclampsia Model

We revealed that rmVEGF164 ameliorates the hypertension induced by the administration of a high dose of Ad m-sFlt-1

in pregnant mice. Li et al¹⁹ reported the therapeutic effect of VEGF-A in preeclamptic rat models induced by the IV administration of truncated Ad m-sFlt-1 on 8 days of gestation; the SC administration of 800 $\mu\text{g}/\text{kg}$ per day of recombinant human VEGF121 for 6 days during day 11 to day 16 of gestation resulted in the amelioration of systolic blood pressure. In our study, the administered doses of rmVEGF164 were lower, and the treatment duration of rmVEGF164 was shorter compared with the study of Li et al¹⁹; however, the MBP after the administration of rmVEGF164 decreased. Therefore, the administration of recombinant VEGF-A to women with preeclampsia may be an effective treatment for this condition, especially for women with early onset preeclampsia, in whom the delay of birth by weeks may contribute to the reduction of neonatal complications and neonatal stay in the newborn intensive care unit.³⁴

We, for the first time, revealed that rmPlGF-2 ameliorates the hypertension induced by high doses of Ad m-sFlt-1 in pregnant mice. To the best of our knowledge, this is the first experiment that showed the antihypertensive effect of rmPlGF on the hypertension induced by Ad m-sFlt-1 in pregnant mice. Hypotension induced by VEGF-A is mainly mediated by VEGFR2.³⁰ Because PlGF binds only to VEGFR1 and has little or no direct mitogenic or permeability-enhancing activity,^{35,36} we supposed that the hypotensive effect of PlGF is very weak. However, surprisingly, the antihypertensive effect of PlGF was as strong as the antihypertensive effect of VEGF-A in our preeclampsia mouse model. Recently, Osol et al³⁷ reported that PlGF had a vasodilatory effect on numerous arteries and veins in rats; pregnancy significantly enhanced sensitivity to PlGF in rat uterine arteries; the vasodilatory effect of PlGF during pregnancy was mainly attributed to the activation of VEGFR1 but not VEGFR2; VEGFR1 was upregulated in the uterine artery wall during gestation; and PlGF dilation was principally mediated by the release of NO in rat uterine arteries. In addition, Osol et al³⁷ also showed that both rat mesenteric and human SC arteries dilated in response to PlGF in an NO-independent manner. These observations clearly suggest that PlGF has the ability to dilate vessels during pregnancy; that is, PlGF has a potentially antihypertensive effect during pregnancy.

Possible Mechanism by Which Hypertension and Proteinuria Emerge in a Pregnant Mouse Administered Recombinant sFlt-1

Recently, Bridges et al⁸ reported that placental and vascular superoxide productions were increased and plasma VEGF-A concentrations were decreased in pregnant rats administered recombinant sFlt-1 chronically during days 13 to 18 of gestation. Vasorelaxations to both acetylcholine and sodium nitroprusside were decreased in pregnant rats administered recombinant sFlt-1, and the decrease of vasorelaxation to acetylcholine was attenuated by the addition of the superoxide scavenger Tiron, indicating elevated maternal sFlt-1, via the decrease of VEGF, results in increased oxidative stress that contributes to vascular dysfunction during pregnancy.⁸ VEGF contributes to the maintenance of an appropriate balance of pro-oxidant and antioxidant factors via manganese superoxide dismutase³⁸ and NADPH oxidase,³⁹ and regulates

NO production.^{32,33} We also observed that the plasma VEGF levels were decreased in pregnant mice administered high doses of Ad m-sFlt-1 and the treatment of VEGF ameliorated hypertension induced by Ad m-sFlt-1. Therefore, increased oxidative stress and vascular dysfunction might be factors in hypertension in the present model, although we did not measure the oxidative stress. Taken together, VEGF antagonism may induce endothelial cell oxidative stress and contribute to renal dysfunction and hypertension.

Conclusions

We, for the first time, created a mouse preeclampsia model using full-length m-sFlt-1 instead of truncated m-sFlt-1, which has been used in previous mouse preeclampsia models.^{3,7,15-19} Not only rmVEGF164, but also PlGF-2, ameliorated hypertension in the mouse preeclampsia model induced by full-length m-sFlt-1. Additional study of PlGF as a potential drug for preeclampsia is warranted.

Perspectives

Our study suggested a possible new therapy using PlGF for preeclamptic women. However, there are several unsolved issues. How many doses of PlGF-2 are sufficient for the amelioration of hypertension and proteinuria in the pregnant mouse model of preeclampsia using Ad m-sFlt-1? When should the administration of PlGF be started to restrict the occurrence of hypertension and proteinuria? What kinds of angiogenic factors show the best therapeutic effects? In addition, if possible, we should make a mouse/rat model using adenoviral human sFlt-1, because the effect of m-sFlt-1 on mouse VEGF-A/PlGF might be different from that of human sFlt-1 on human VEGF-A/PlGF. If the human PlGF-1/PlGF-2 is used for the prevention/therapy of preeclampsia, we should carefully monitor the occurrence of possible adverse effects, such as lung edema, the development of new blood vessels in nontargeted tissues.

Sources of Funding

This work was funded by a Grant-in-Aid for Scientific Research from the Japan Society for the Promotion of Science (No. 18791169).

Disclosures

None.

References

1. Nagaya K, Fetters MD, Ishikawa M, Kubo T, Koyanagi T, Saito Y, Sameshima H, Sugimoto M, Takagi K, Chiba Y, Honda H, Mukubo M, Kawamura M, Satoh S, Neki R. Causes of maternal mortality in Japan. *JAMA*. 2000;283:2661-2667.
2. Hecher K, Campbell S, Doyle P, Harrington K, Nicolaides K. Assessment of fetal compromise by Doppler ultrasound investigation of the fetal circulation: arterial, intracardiac, and venous blood flow velocity studies. *Circulation*. 1995;91:129-138.
3. Maynard SE, Min JY, Merchan J, Lim KH, Li J, Mondal S, Libermann TA, Morgan JP, Sellke FW, Stillman RE, Epstein FH, Sukhatme VP, Karumanchi SA. Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *J Clin Invest*. 2003;111:649-658.
4. Levine RJ, Maynard SE, Qian C, Lim KH, England LJ, Yu KF, Schisterman EF, Thadhani R, Sachs BP, Epstein FH, Sibai BM, Sukhatme VP, Karumanchi SA. Circulating angiogenic factors and the risk of preeclampsia. *N Engl J Med*. 2004;350:672-683.

5. Levine RJ, Lam C, Qian C, Yu KF, Maynard SE, Sachs BP, Sibai BM, Epstein FH, Romero R, Thadhani R, Karumanchi SA; for the CPEP Study Group. Soluble endoglin and other circulating antiangiogenic factors in preeclampsia. *N Engl J Med*. 2006;355:992–1005.
6. Kabbinnar F, Hurwitz HI, Fehrenbacher L, Meropol NJ, Novotny WF, Lieberman G, Griffing S, Bergsland E. Phase II, randomized trial comparing bevacizumab plus fluorouracil (FU)/leucovorin (LV) with FU/LV alone in patients with metastatic colorectal cancer. *J Clin Oncol*. 2003;21:60–65.
7. Sugimoto H, Hamano Y, Charytan D, Cosgrove D, Kieran M, Sudhakar A, Kalluri R. Neutralization of circulating vascular endothelial growth factor (VEGF) by anti-VEGF antibodies and soluble VEGF receptor 1 (sFlt-1) induces proteinuria. *J Biol Chem*. 2003;278:12605–12608.
8. Bridges JP, Gilbert JS, Colson D, Gilbert SA, Dukes MP, Ryan MJ, Granger JP. Oxidative stress contributes to soluble Fms-like tyrosine kinase-1 induced vascular dysfunction in pregnant rats. *Am J Hypertens*. 2009;22:564–568.
9. Kendall RL, Thomas KA. Inhibition of vascular endothelial cell growth factor activity by an endogenously encoded soluble receptor. *Proc Natl Acad Sci U S A*. 1993;90:10705–10709.
10. Clark DE, Smith SK, He Y, Day KA, Licence DR, Corps AN, Lammoglia R, Charnock-Jones DS. A vascular endothelial growth factor antagonist is produced by the human placenta and released into the maternal circulation. *Biol Reprod*. 1998;59:1540–1548.
11. Yamaguchi S, Iwata K, Shibuya M. Soluble Flt-1 (soluble VEGFR-1), a potent natural antiangiogenic molecule in mammals, is phylogenetically conserved in avians. *Biochem Biophys Res Commun*. 2002;291:554–559.
12. Chen H, Ikeda U, Shimpō M, Maeda Y, Shibuya M, Ozawa K, Shimada K. Inhibition of vascular endothelial growth factor activity by transfection with the soluble FLT-1 gene. *J Cardiovasc Pharmacol*. 2000;36:498–502.
13. Barleon B, Totzke F, Herzog C, Blanke S, Kremmer E, Siemeister G, Marmé D, Martiny-Baron G. Mapping of the sites for ligand binding and receptor dimerization at the extracellular domain of the vascular endothelial growth factor FLT-1. *J Biol Chem*. 1997;272:10382–10388.
14. Shinkai A, Ito M, Anazawa H, Yamaguchi S, Shitarai K, Shibuya M. Mapping of the sites involved in ligand association and dissociation at the extracellular domain of the kinase insert domain-containing receptor for vascular endothelial growth factor. *J Biol Chem*. 1998;273:31283–31288.
15. Kuo CJ, Farnebo F, Yu EY, Christofferson R, Swearingen RA, Carter R, von Recum HA, Yuan J, Kamihara J, Flynn E, D'Amato R, Folkman J, Mulligan RC. Comparative evaluation of the antitumor activity of antiangiogenic proteins delivered by gene transfer. *Proc Natl Acad Sci U S A*. 2001;98:4605–4610.
16. Venkatesha S, Toporsian M, Lam C, Hanai J, Mammoto T, Kim YM, Bdoolah Y, Lim KH, Yuan HT, Libermann TA, Stillman RE, Roberts D, D'Amore PA, Epstein FH, Sellke FW, Romero R, Sukhatme VP, Letarte M, Karumanchi SA. Soluble endoglin contributes to the pathogenesis of preeclampsia. *Nat Med*. 2006;12:642–649.
17. Lu F, Bytautiene E, Tamayo E, Gamble P, Anderson GD, Hankins GD, Longo M, Saade GR. Gender-specific effect of overexpression of sFlt-1 in pregnant mice on fetal programming of blood pressure in the offspring later in life. *Am J Obstet Gynecol*. 2007;197:418.e1–e5.
18. Lu F, Longo M, Tamayo E, Maner W, Al-Hendy A, Anderson GD, Hankins GD, Saade GR. The effect of over-expression of sFlt-1 on blood pressure and the occurrence of other manifestations of preeclampsia in unrestrained conscious pregnant mice. *Am J Obstet Gynecol*. 2007;196:396.e1–e7.
19. Li Z, Zhang Y, Ying Ma J, Kapoun AM, Shao Q, Kerr I, Lam A, O'Young G, Sannajust F, Stathis P, Schreiner G, Karumanchi SA, Protter AA, Pollitt NS. Recombinant vascular endothelial growth factor 121 attenuates hypertension and improves kidney damage in a rat model of preeclampsia. *Hypertension*. 2007;50:686–692.
20. Espinoza J, Romero R, Nien JK, Gomez R, Kusanovic JP, Gonçalves LF, Medina L, Edwin S, Hassan S, Carstens M, Gonzalez R. Identification of patients at risk for early onset and/or severe preeclampsia with the use of uterine artery Doppler velocimetry and placental growth factor. *Am J Obstet Gynecol*. 2007;196:326.e1–326.e13.
21. Ohkuchi A, Hirashima C, Matsubara S, Suzuki H, Takahashi K, Arai F, Watanabe T, Kario K, Suzuki M. Alterations in placental growth factor levels before and after the onset of preeclampsia are more pronounced in women with early onset severe preeclampsia. *Hypertens Res*. 2007;30:151–159.
22. Krauss T, Pauer HU, Augustin HG. Prospective analysis of placenta growth factor (PlGF) concentrations in the plasma of women with normal pregnancy and pregnancies complicated by preeclampsia. *Hypertens Pregnancy*. 2004;23:101–111.
23. Kondo K, Hiratsuka S, Subbalakshmi E, Matsushime H, Shibuya M. Genomic organization of the flt-1 gene encoding for vascular endothelial growth factor (VEGF) receptor-1 suggests an intimate evolutionary relationship between the 7-Ig and the 5-Ig tyrosine kinase receptor. *Gene*. 1998;208:297–305.
24. Watanabe K, Hasegawa Y, Yamashita H, Shimizu K, Ding Y, Abe M, Ohta H, Imagawa K, Hojo K, Maki H, Sonoda H, Sato Y. Vasohibin as an endothelium-derived negative feedback regulator of angiogenesis. *J Clin Invest*. 2004;114:898–907.
25. Kamba T, Tam BY, Hashizume H, Haskell A, Sennino B, Mancuso MR, Norberg SM, O'Brien SM, Davis RB, Gowen LC, Anderson KD, Thurston G, Joho S, Springer ML, Kuo CJ, McDonald DM. VEGF-dependent plasticity of fenestrated capillaries in the normal adult microvasculature. *Am J Physiol Heart Circ Physiol*. 2006;290:H560–H576.
26. Yang JC, Haworth L, Sherry RM, Hwu P, Schwartzentruber DJ, Topalian SL, Steinberg SM, Chen HX, Rosenberg SA. A randomized trial of bevacizumab, an anti-vascular endothelial growth factor antibody, for metastatic renal cancer. *N Engl J Med*. 2003;349:427–434.
27. Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, Berlin J, Baron A, Griffing S, Holmgren E, Ferrara N, Fyfe G, Rogers B, Ross R, Kabbinnar F. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N Engl J Med*. 2004;350:2335–2342.
28. Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Siebels M, Negrier S, Chebreau C, Solska E, Desai AA, Rolland F, Demkow T, Hutson TE, Gore M, Freeman S, Schwartz B, Shan M, Simantov R, Bukowski RM; for the TARGET Study Group. Sorafenib in advanced clear-cell renal-cell carcinoma. *N Engl J Med*. 2007;356:125–134.
29. Chu TF, Rupnick MA, Kerkela R, Dallabrida SM, Zurawski D, Nguyen L, Woulfe K, Pravda E, Cassiola F, Desai J, George S, Morgan JA, Harris DM, Ismail NS, Chen JH, Schoen FJ, Van den Abbeele AD, Demetri GD, Force T, Chen MH. Cardiotoxicity associated with tyrosine kinase inhibitor sunitinib. *Lancet*. 2007;370:2011–2019.
30. Li B, Ogasawara AK, Yang R, Wei W, He GW, Zioncheck TF, Bunting S, de Vos AM, Jin H. KDR (VEGF receptor 2) is the major mediator for the hypotensive effect of VEGF. *Hypertension*. 2002;39:1095–1100.
31. Yang R, Ogasawara AK, Zioncheck TF, Ren Z, He GW, DeGuzman GG, Pelletier N, Shen BQ, Bunting S, Jim H. Exaggerated hypotensive effect of vascular endothelial growth factor in spontaneously hypertensive rats. *Hypertension*. 2002;39:815–820.
32. Parsons-Wingeter P, Chandrasekharan UM, McKay TL, Radhakrishnan K, DiCorleto PE, Albarran B, Farr AG. A VEGF165-induced phenotypic switch from increased vessel density to increased vessel diameter and increased endothelial NOS activity. *Microvasc Res*. 2006;72:91–100.
33. Sandrim VC, Palei AC, Metzger IF, Gomes VA, Cavalli RC, Tanus-Santos JE. Nitric oxide formation is inversely related to serum levels of antiangiogenic factors soluble fms-like tyrosine kinase-1 and soluble endogline in preeclampsia. *Hypertension*. 2008;52:402–407.
34. Sibai BM, Mercer BM, Schiff E, Friedman SA. Aggressive versus expectant management of severe preeclampsia at 28 to 32 weeks' gestation: a randomized controlled trial. *Am J Obstet Gynecol*. 1994;171:818–822.
35. Park JE, Chen HH, Winer J, Houck KA, Ferrara N. Placenta growth factor: potentiation of vascular endothelial growth factor bioactivity, in vitro and in vivo, and high affinity binding to Flt-1 but not to Flk-1/KDR. *J Biol Chem*. 1994;269:25646–25654.
36. Sawano A, Takahashi T, Yamaguchi S, Aonuma M, Shibuya M. Flt-1 but not KDR/Flk-1 tyrosine kinase is a receptor for placenta growth factor, which is related to vascular endothelial growth factor. *Cell Growth Differ*. 1996;7:213–221.
37. Osol G, Celia G, Gokina N, Barron C, Chien E, Mandala M, Luksha L, Kublickiene K. Placental growth factor is a potent vasodilator of rat and human resistance arteries. *Am J Physiol Heart Circ Physiol*. 2008;294:H1381–H1387.
38. Abid MR, Schoots IG, Spokes KC, Wu SQ, Mawhinney C, Aird WC. Vascular endothelial growth factor-mediated induction of manganese superoxide dismutase occurs through redox-dependent regulation of forkhead and IκappaB/NF-κappaB. *J Biol Chem*. 2004;279:44030–44038.
39. González-Pacheco FR, Deudero JJ, Castellanos MC, Castilla MA, Alvarez-Arroyo MV, Yagüe S, Caramelo C. Mechanisms of endothelial response to oxidative aggression: protective role of autologous VEGF and induction of VEGFR2 by H2O2. *Am J Physiol Heart Circ Physiol*. 2006;291:H1395–H1401.