

Fig. (3). HCV-infected apoptotic cells, but not HCV particles, regulate mDC function to activate NK cells and T cells. The HCV JFH1 strain does not directly stimulate mDCs or pDCs. In this case, pDCs and mDCs do not directly respond to the RNA virus. Our recent finding is that dsRNA in HCV-infected apoptotic vesicles is an immune-stimulator for mDCs that triggers the innate system for the activating T cell (Th1 dominant) and NK cells. Cell/cell contact between mDCs and NK cells is indispensable to increased NK cell cytotoxicity.

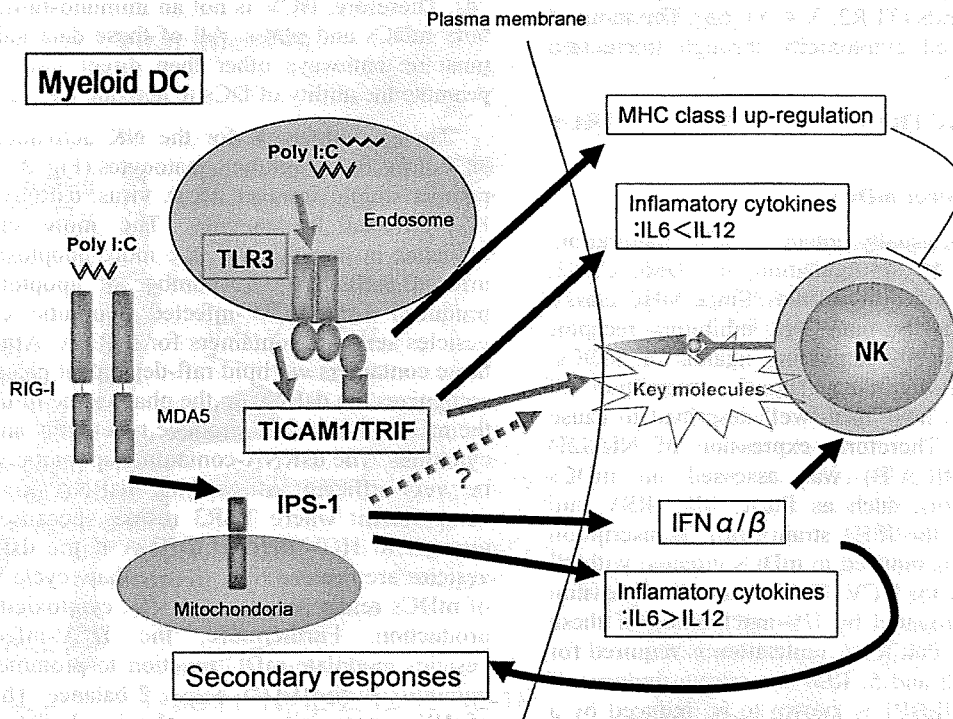


Fig. (4). The TLR3/TICAM1 signal leads to expression of unknown molecules on mDC, which activate NK cell *via* cell/cell contact. Among the three poly I:C receptors (TLR3, RIG-I and MDA5), the TLR3/TICAM1 signal plays an important role in regulating mDC capacity to activate NK cells. This mDC-mediated NK activation is dependent on the direct interaction of mDC and NK cells. Besides IL-15, there must be other key molecules that are induced on mDC by the TLR3/TICAM1 signal to activate NK cells because IL-15 is normally induced by poly I:C in TICAM1^{-/-} mice. NK activity is also regulated by type I IFN mediated by MDA5/IPS-1 pathway. Although IL-12 is produced mainly by the TICAM1 signal, IL-12 is not a functional entity that can enhance NK cytotoxicity by poly I:C-stimulated mDCs.

through poly I:C-stimulated DCs requires cell/cell contact. Although type I IFN is a key molecule regulating NK activation, the IFN α response in TICAM1^{-/-} mice is comparable to that in wild type mice. This is because another poly I:C receptor, MDA5, recognizes poly I:C and leads to IFN α production [19].

Poly I:C-dependent IL-12 production is impaired in TICAM1^{-/-} mice [13,19]. However, neutralization of IL-12 with anti IL-12 antibodies does not affect dsRNA-induced NK activation [13]. All of these data suggest that the TLR3/TICAM1 signal leads to up-regulation of a certain NK-activating ligand on mDCs. Lucas *et al.* showed that type I IFN secretion and trans-presentation of IL-15 by

mDCs are crucial to DC-mediated NK activation in response to poly I:C [14]. However, induction of IL-15 by mDCs is normal after stimulation with poly I:C in TICAM1-/- mice (unpublished data). We are now searching for the unknown molecules that regulate NK activation on mDCs (Fig. 4), other than IL-15. This unidentified factor may contribute to elucidation of the way NK cells are activated by mDCs during HCV infection.

CONCLUSION

NK cells are principal components of innate immunity and play a key role in early immune responses to viruses. The pathway mediating NK activation has been precisely examined *in vivo* by MCMV (DNA virus) infection. However, the receptors that sense virus-derived DNA are still unknown, except for TLR9 and DAI [75]. Many systems (such as TLR3, RIG-I, MDA5, TLR7 and TLR8) have been shown to trigger innate immunity against virus-derived RNA, but the mechanism of NK cell activation is less studied in RNA virus infection than in DNA virus infection. pDCs, mDCs, macrophages, monocytes and other cells invaded by RNA viruses presumably constitute a complicated network that activates NK cells. The tropism of RNA viruses appears to be the determinant of the mechanisms by which NK cells participate in innate immunity.

ACKNOWLEDGEMENTS

We thank all the past and present members of the Seya lab and, in particular, Shingai M. and Akazawa T. We also thank Dr. Wakita T. for the gift of the *in vitro* system to propagate the JFH1 strain.

REFERENCES

- [1] Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S. Functions of natural killer cells. *Nat Immunol* 2008; 9: 503-10.
- [2] Ljunggren HG, Karre K. Host resistance directed selectively against H-2-deficient lymphoma variants: analysis of the mechanism. *J Exp Med* 1985; 162: 1745-59.
- [3] Gonzalez S, Lopez-Soto A, Suarez-Alvarez B, Lopez-Vazquez A, Lopez-Larrea C. NKG2D ligands: key targets of the immune response. *Trends Immunol* 2008; 29: 397-403.
- [4] Mistry AR, O'Callaghan CA. Regulation of ligands for the activating receptor NKG2D. *Immunology* 2007; 121: 439-47.
- [5] Nausch N, Cerwenka A. NKG2D ligands in tumor immunity. *Oncogene* 2008; 27: 5944-58.
- [6] Lanier LL. Evolutionary struggles between NK cells and viruses. *Nat Rev Immunol* 2008; 8: 259-68.
- [7] Salih HR, Holdenrieder S, Steinle A. Soluble NKG2D ligands: prevalence, release, and functional impact. *Front Biosci* 2008; 13: 3448-56.
- [8] Sivori S, Falco M, Della Chiesa M, *et al.* CpG and double-stranded RNA trigger human NK cells by Toll-like receptors: induction of cytokine release and cytotoxicity against tumors and dendritic cells. *Proc Natl Acad Sci USA* 2004; 101: 10116-21.
- [9] Girart MV, Fuertes MB, Domaica CI, Rossi LE, Zwirner NW. Engagement of TLR3, TLR7, and NKG2D regulate IFN-gamma secretion but not NKG2D-mediated cytotoxicity by human NK cells stimulated with suboptimal doses of IL-12. *J Immunol* 2007; 179: 3472-9.
- [10] Hart OM, Athie-Morales V, O'Connor GM, Gardiner CM. TLR7/8-mediated activation of human NK cells results in accessory cell-dependent IFN-gamma production. *J Immunol* 2005; 175: 1636-42.
- [11] Arase H, Mocarski ES, Campbell AE, Hill AB, Lanier LL. Direct recognition of cytomegalovirus by activating and inhibitory NK cell receptors. *Science* 2002; 296: 1323-6.
- [12] Smith HR, Heusel JW, Mehta IK, *et al.* Recognition of a virus-encoded ligand by a natural killer cell activation receptor. *Proc Natl Acad Sci USA* 2002; 99: 8826-31.
- [13] Akazawa T, Ebihara T, Okuno M, *et al.* Antitumor NK activation induced by the Toll-like receptor 3-TICAM-1 (TRIF) pathway in myeloid dendritic cells. *Proc Natl Acad Sci USA* 2007; 104: 252-7.
- [14] Lucas M, Schachterle W, Oberle K, Aichele P, Dieffenbach A. Dendritic cells prime natural killer cells by trans-presenting interleukin 15. *Immunity* 2007; 26: 503-17.
- [15] Newman KC, Riley EM. Whatever turns you on: accessory-cell-dependent activation of NK cells by pathogens. *Nat Rev Immunol* 2007; 7: 279-91.
- [16] Honda K, Taniguchi T. IRFs: master regulators of signalling by Toll-like receptors and cytosolic pattern-recognition receptors. *Nat Rev Immunol* 2006; 6: 644-658.
- [17] Diebold SS, Kaisho T, Hemmi H, Akira S, Reis e Sousa C. Innate antiviral responses by means of TLR7-mediated recognition of single-stranded RNA. *Science* 2004; 303: 1529-31.
- [18] Yoneyama M, Kikuchi M, Natsukawa T, *et al.* The RNA helicase RIG-I has an essential function in double-stranded RNA-induced innate antiviral responses. *Nat Immunol* 2004; 5: 730-737.
- [19] Kato H, Takeuchi O, Sato S, *et al.* Differential roles of MDA5 and RIG-I helicases in the recognition of RNA viruses. *Nature* 2006; 441: 101-5.
- [20] Uematsu S, Akira S. Toll-like receptors and Type I interferons. *J Biol Chem* 2007; 282: 15319-23.
- [21] Matsumoto M, Funami K, Tanabe M, *et al.* Subcellular localization of Toll-like receptor 3 in human dendritic cells. *J Immunol* 2003; 171: 3154-62.
- [22] Kato H, Takeuchi O, Mikamo-Sato E, *et al.* Length-dependent recognition of double-stranded ribonucleic acids by retinoic acid-inducible gene-1 and melanoma differentiation-associated gene 5. *J Exp Med* 2008; 205: 1601-10.
- [23] Hornung V, Ellegast J, Kim S, *et al.* 5'-Triphosphate RNA is the ligand for RIG-I. *Science* 2006; 314: 994-7.
- [24] O'Neill LA, Bowie AG. The family of five: TIR-domain-containing adaptors in Toll-like receptor signalling. *Nat Rev Immunol* 2007; 7: 353-64.
- [25] Moynagh PN. TLR signalling and activation of IRFs: revisiting old friends from the NF-kappaB pathway. *Trends Immunol* 2005; 26: 469-76.
- [26] Oshiumi H, Matsumoto M, Funami K, Akazawa T, Seya T. TICAM-1, an adaptor molecule that participates in Toll-like receptor 3-mediated interferon-beta induction. *Nat Immunol* 2003; 4: 161-7.
- [27] Sasai M, Oshiumi H, Matsumoto M, *et al.* Cutting Edge: NF-kappaB-activating kinase-associated protein 1 participates in TLR3/Toll-IL-1 homology domain-containing adapter molecule-1-mediated IFN regulatory factor 3 activation. *J Immunol* 2005; 174: 27-30.
- [28] Kawai T, Takahashi K, Sato S, *et al.* IPS-1, an adaptor triggering RIG-I- and Mda5-mediated type I interferon induction. *Nat Immunol* 2005; 6: 981-8.
- [29] Seth RB, Sun L, Ea CK, Chen ZJ. Identification and characterization of MAVS, a mitochondrial antiviral signaling protein that activates NF-kappaB and IRF 3. *Cell* 2005; 122: 669-682.
- [30] Xu LG, Wang YY, Han KJ, Li LY, Zhai Z, Shu HB. VISA is an adapter protein required for virus-triggered IFN-beta signaling. *Mol Cell* 2005; 19: 727-40.
- [31] Meylan E, Curran J, Hofmann K, *et al.* Cardif is an adaptor protein in the RIG-I antiviral pathway and is targeted by hepatitis C virus. *Nature* 2005; 437: 1167-72.
- [32] Takeuchi O, Akira S. MDA5/RIG-I and virus recognition. *Curr Opin Immunol* 2008; 20: 17-22.
- [33] Kumagai Y, Takeuchi O, Kato H, *et al.* Alveolar macrophages are the primary interferon-alpha producer in pulmonary infection with RNA viruses. *Immunity* 2007; 27: 240-52.
- [34] Cervantes-Barragan L, Zust R, Weber F, *et al.* Control of coronavirus infection through plasmacytoid dendritic-cell-derived type I interferon. *Blood* 2007; 109: 1131-7.
- [35] Lee HK, Lund JM, Ramanathan B, Mizushima N, Iwasaki A. Autophagy-dependent viral recognition by plasmacytoid dendritic cells. *Science* 2007; 315: 1398-401.
- [36] Melchjorsen J, Jensen SB, Malmgaard L, *et al.* Activation of innate defense against a paramyxovirus is mediated by RIG-I and TLR7 and TLR8 in a cell-type-specific manner. *J Virol* 2005; 79: 12944-51.

- [37] Yount JS, Gitlin L, Moran TM, Lopez CB. MDA5 participates in the detection of paramyxovirus infection and is essential for the early activation of dendritic cells in response to Sendai Virus defective interfering particles. *J Immunol* 2008; 180: 4910-8.
- [38] Bhoj VG, Sun Q, Bhoj EJ, *et al.* MAVS and MyD88 are essential for innate immunity but not cytotoxic T lymphocyte response against respiratory syncytial virus. *Proc Natl Acad Sci USA* 2008; 105: 14046-51.
- [39] Shingai M, Inoue N, Okuno T, *et al.* Wild-type measles virus infection in human CD46/CD150-transgenic mice: CD11c-positive dendritic cells establish systemic viral infection. *J Immunol* 2005; 175: 3252-61.
- [40] de Swart RL, Ludlow M, de Witte L, *et al.* Predominant infection of CD150+ lymphocytes and dendritic cells during measles virus infection of macaques. *PLoS Pathog* 2007; 3: e178.
- [41] Liu P, Jamaluddin M, Li K, Garofalo RP, Casola A, Brasier AR. Retinoic acid-inducible gene I mediates early antiviral response and Toll-like receptor 3 expression in respiratory syncytial virus-infected airway epithelial cells. *J Virol* 2007; 81: 1401-11.
- [42] Hornung V, Schlender J, Guenther-Biller M, *et al.* Replication-dependent potent IFN-alpha induction in human plasmacytoid dendritic cells by a single-stranded RNA virus. *J Immunol* 2004; 173: 5935-43.
- [43] Honda K, Yanai H, Negishi H, *et al.* IRF-7 is the master regulator of type-I interferon-dependent immune responses. *Nature* 2005; 434: 772-7.
- [44] Lund JM, Alexopoulou L, Sato A, *et al.* Recognition of single-stranded RNA viruses by Toll-like receptor 7. *Proc Natl Acad Sci USA* 2004; 101: 5598-603.
- [45] Diebold SS, Kaisho T, Hemmi H, Akira S, Reis e Sousa C. Innate antiviral responses by means of TLR7-mediated recognition of single-stranded RNA. *Science* 2004; 303: 1529-31.
- [46] Waibler Z, Detje CN, Bell JC, Kalinke U. Matrix protein mediated shutdown of host cell metabolism limits vesicular stomatitis virus-induced interferon-alpha responses to plasmacytoid dendritic cells. *Immunobiology* 2007; 212: 887-94.
- [47] Yount JS, Gitlin L, Moran TM, Lopez CB. MDA5 participates in the detection of paramyxovirus infection and is essential for the early activation of dendritic cells in response to Sendai Virus defective interfering particles. *J Immunol* 2008; 180: 4910-8.
- [48] Wang JP, Liu P, Latz E, Golenbock DT, Finberg RW, Libraty DH. Flavivirus activation of plasmacytoid dendritic cells delineates key elements of TLR7 signaling beyond endosomal recognition. *J Immunol* 2006; 177: 7114-21.
- [49] Ho LJ, Wang JJ, Shiao MF, *et al.* Infection of human dendritic cells by dengue virus causes cell maturation and cytokine production. *J Immunol* 2001; 166: 1499-506.
- [50] Granelli-Piperno A, Golebiowska A, Trumphyeller C, Siegal FP, Steinman RM. HIV-1-infected monocyte-derived dendritic cells do not undergo maturation but can elicit IL-10 production and T cell regulation. *Proc Natl Acad Sci USA* 2004; 101: 7669-74.
- [51] Beignon AS, McKenna K, Skoberne M, *et al.* Endocytosis of HIV-1 activates plasmacytoid dendritic cells via Toll-like receptor-viral RNA interactions. *J Clin Invest* 2005; 115: 3265-75.
- [52] McCartney SA, Thackray LB, Gitlin L, Gilfillan S, Virgin IV HW, Colonna M. MDA-5 recognition of a murine norovirus. *PLoS Pathog* 2008; 4: e1000108.
- [53] Narvaez CF, Angel J, Franco MA. Interaction of rotavirus with human myeloid dendritic cells. *J Virol* 2005; 79: 14526-35.
- [54] Bauer S, Pigisch S, Hangel D, Kaufmann A, Hamm S. Recognition of nucleic acid and nucleic acid analogs by Toll-like receptors 7, 8 and 9. *Immunobiology* 2008; 213: 315-28.
- [55] Triantafyllou K, Orthopoulos G, Vakakis E, *et al.* Human cardiac inflammatory responses triggered by Coxsackie B viruses are mainly Toll-like receptor (TLR) 8-dependent. *Cell Microbiol* 2005; 7: 1117-26.
- [56] Triantafyllou K, Vakakis E, Orthopoulos G, *et al.* TLR8 and TLR7 are involved in the host's immune response to human parechovirus 1. *Eur J Immunol* 2005; 35: 2416-23.
- [57] Gerosa F, Gobbi A, Zorzi P, *et al.* The reciprocal interaction of NK cells with plasmacytoid or myeloid dendritic cells profoundly affects innate resistance functions. *J Immunol* 2005; 174: 727-34.
- [58] Barr DP, Belz GT, Reading PC, *et al.* A role for plasmacytoid dendritic cells in the rapid IL-18-dependent activation of NK cells following HSV-1 infection. *Eur J Immunol* 2007; 37: 1334-42.
- [59] Hanabuchi S, Watanabe N, Wang YH, *et al.* Human plasmacytoid predendritic cells activate NK cells through glucocorticoid-induced tumor necrosis factor receptor-ligand (GITRL). *Blood* 2006; 107: 3617-23.
- [60] Poggi A, Carosio R, Spaggiari GM, *et al.* NK cell activation by dendritic cells is dependent on LFA-1-mediated induction of calcium-calmodulin kinase II: inhibition by HIV-1 Tat C-terminal domain. *J Immunol* 2002; 168: 95-101.
- [61] Andoniou CE, van Dommelen SL, Voigt V, *et al.* Interaction between conventional dendritic cells and natural killer cells is integral to the activation of effective antiviral immunity. *Nat Immunol* 2005; 6: 1011-9.
- [62] Yang T, Flint MS, Webb KM, Chambers WH. CD161B: C1rB interactions mediate activation of enhanced lysis of tumor target cells following NK cell: DC co-culture. *Immunol Res* 2006; 36: 43-50.
- [63] Kijima M, Yamaguchi T, Ishifune C, *et al.* Dendritic cell-mediated NK cell activation is controlled by Jagged2-Notch interaction. *Proc Natl Acad Sci USA* 2008; 105: 7010-5.
- [64] Pan PY, Gu P, Li Q, Xu D, Weber K, Chen SH. Regulation of dendritic cell function by NK cells: mechanisms underlying the synergism in the combination therapy of IL-12 and 4-1BB activation. *J Immunol* 2004; 172: 4779-89.
- [65] Welte S, Kuttruff S, Waldhauer I, Steinle A. Mutual activation of natural killer cells and monocytes mediated by NKp80-AICL interaction. *Nat Immunol* 2006; 7: 1334-42.
- [66] Ebihara T, Masuda H, Akazawa T, *et al.* Induction of NKG2D ligands on human dendritic cells by TLR ligand stimulation and RNA virus infection. *Int Immunol* 2007; 19: 1145-55.
- [67] Draghi M, Pashine A, Sanjanwala B, *et al.* NKp46 and NKG2D recognition of infected dendritic cells is necessary for NK cell activation in the human response to influenza infection. *J Immunol* 2007; 178: 2688-98.
- [68] Wakita T, Pietschmann T, Kato T, *et al.* Production of infectious hepatitis C virus in tissue culture from a cloned viral genome. *Nat Med* 2005; 11: 791-6.
- [69] Auffermann-Gretzinger S, Keeffe EB, Levy S. Impaired dendritic cell maturation in patients with chronic, but not resolved, hepatitis C virus infection. *Blood* 2001; 97: 3171-6.
- [70] Bain C, Fatmi A, Zoulim F, Zarski JP, Trepo C, Inchauspe G. Impaired allostimulatory function of dendritic cells in chronic hepatitis C infection. *Gastroenterology* 2001; 120: 512-4.
- [71] Longman RS, Talal AH, Jacobson IM, Albert ML, Rice CM. Presence of functional dendritic cells in patients chronically infected with hepatitis C virus. *Blood* 2004; 103: 1026-9.
- [72] Longman RS, Talal AH, Jacobson IM, Rice CM, Albert ML. Normal functional capacity in circulating myeloid and plasmacytoid dendritic cells in patients with chronic hepatitis C. *J Infect Dis* 2005; 192: 497-503.
- [73] Ebihara T, Shingai M, Matsumoto M, Wakita T, Seya T. Hepatitis C virus-infected hepatocytes extrinsically modulate dendritic cell maturation to activate T cells and natural killer cells. *Hepatology* 2008; 48: 48-58.
- [74] Shiina M, Rehmann B. Cell culture-produced hepatitis C virus impairs plasmacytoid dendritic cell function. *Hepatology* 2008; 47: 385-95.
- [75] Takaoka A, Wang Z, Choi MK, *et al.* DAI (DLM-1/ZBP1) is a cytosolic DNA sensor and an activator of innate immune response. *Nature* 2007; 448: 501-5.

