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Electrostatically constrained α-helical peptide inhibits replication of HIV-1 resistant to enfuvirtide

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ABSTRACT

α-Helical peptides, such as T-20 (enfuvirtide) and C34, derived from the gp41 carboxyl-terminal heptad repeat (C-HR) of HIV-1, inhibit membrane fusion of HIV-1 and the target cells. Although T-20 effectively suppresses the replication of multi-drug resistant HIV variants both in vitro and in vivo, prolonged therapy with T-20 induces emergence of T-20 resistant variants. In order to suppress the emergence of such resistant variants, we introduced charged and hydrophilic amino acids, glutamic acid (E) and lysine (K), at the solvent accessible site of C34. In particular, the modified peptide, SC34EK, demonstrates remarkably potent inhibition of membrane fusion by the resistant HIV-1 variants as well as wild-type viruses. The activity was specific to HIV-1 and little influenced by serum components. We found a strong correlation between the anti-HIV-1 activities of these peptides and the thermostabilities of the 6-helix bundles that are formed with these peptides. We also obtained the crystal structure of SC34EK in complex with a 36 amino acid sequence (N36) comprising the amino-terminal heptad repeat of HIV-1. The EK substitutions in the sequence of SC34EK were directed toward the solvent and generated an electrostatic potential, which may result in enhanced α-helicity of the peptide inhibitor. The 6-helix bundle complex of SC34EK with N36 appears to be structurally similar to that of C34 and N36. Our approach to enhancing α-helicity of the peptide inhibitor may enable future design of highly effective and specific HIV-1 inhibitors.

1. Introduction

Enfuvirtide (T-20), which has been clinically approved as the first fusion inhibitor of HIV-1, is derived from a 36 amino acid region of the carboxyl-terminal heptad repeat (C-HR) of gp41, an HIV-1 transmembrane envelope glycoprotein, which plays central role in the fusion of HIV-1 with host cells. T-20 prevents the formation of a 6-helix bundle, which is comprised of a trimer of dimers formed from the amino-terminal heptad repeat (N-HR) and the carboxyl-terminal heptad repeat (C-HR) in an antiparallel orientation. Six-helix formation by physiological gp41 enables host cell and virus membranes to contact and fuse, enabling the virus entry into the cells. Therefore, inhibition of the formation of this 6-helix bundle prevents fusion of HIV-1 and targeted host cell membranes (Derdeyn et al., 2000; Wild et al., 1992). Notably, T-20 effectively suppresses the replication of HIV-1 variants, which are resistant to multiple reverse transcriptase and protease inhibitors, and has been used in the optimized regimens for HIV-1-infected patients harboring multi-drug resistant HIV-1 variants (Lalezari et al., 2003; Lazzarin et al., 2003).

Emergence of T-20-resistant HIV-1 was reported not only in patients receiving T-20 monotherapy in a phase I clinical trial (Wei et al., 2002), but also in patients treated with a combination of T-20

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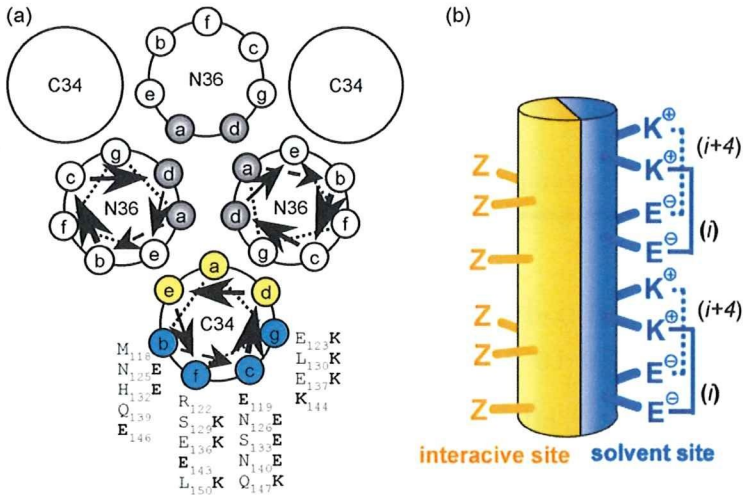

Fig. 1. Helical wheel representation of the 6-helix bundle structure and the design of SC34EK. (a) Amino acid residues at positions a, d, e of C34 are interactive sites that form the 6-helix complex with N36, while the remaining amino acid residues at positions b, c, f, g are solvent accessible sites, which are substituted with Glu (E) or Lys (K) in SC34EK. (b) The design concept of introducing the EK motif to the solvent accessible site. The α-helical C-HR peptide could be divided into interactive (red) and solvent (blue) sites. Z indicates the original amino acids of C34. Only amino acid residues at solvent sites were replaced by E at the *i* position and K at the *i* + 4 position.

and other inhibitors in subsequent phases II and III trials (Matthews et al., 2004; Poveda et al., 2002). These resistant variants frequently acquired mutations in gp41, especially in amino acids 36–45 of the N-HR region (Aquaro et al., 2006; Cabrera et al., 2006; Mink et al., 2005; Poveda et al., 2002; Rimsky et al., 1998; Wei et al., 2002) (Fig. 1). Additionally, complementary mutations in the C-HR region, such as S138A mutation, were found in some T-20 resistant variants (Cabrera et al., 2006; Poveda et al., 2004; Xu et al., 2005). Introduction of these complementary mutations compensates for impaired HIV-1 replication stemming from the primary mutations that give rise to resistance. The N43D mutation in the N-HR region that confers resistance to T-20 is a well documented example (Xu et al., 2005).

Although T-20 inhibits gp41-mediated fusion (Derdeyn et al., 2000; Wild et al., 1992), it has additional effects on HIV-1 replication. For instance, baseline sensitivity of HIV-1 to T-20 is influenced not only by the amino acid sequence of gp41, but also by the co-receptor specificity (CCR5/CXCR4) defined by the structure of the V3 loop of gp120, a glycoprotein capping gp41, which binds to the CD4 cells (Derdeyn et al., 2000; Derdeyn et al., 2001). Moreover, substitutions within the CD4 binding domain of gp120 also contribute to the resistance of the virus to T-20 (Baldwin and Berkhout, 2006). Thus, the mode of action and the mechanism of resistance to T-20 seem to be complicated. In contrast, another fusion inhibitor known as C34, has been clearly shown to bind to the N-HR in vitro and act as a decoy of gp41 C-HR and prevent the formation of the 6-helix bundle (Chan et al., 1997; Liu et al., 2005; Xu et al., 2007). Its inhibitory effect is over 10-fold greater than that of T-20 (Armand-Ugon et al., 2003; Nameki et al., 2005). Thus, C34 appears to be a suitable peptide to employ in the rational design of an improved HIV fusion inhibitor, based on the interaction between the peptide and the target.

It has been reported that α-helicity of the C-HR and N-HR peptide complexes correlates with the anti-HIV-1 activity of the peptide inhibitor (Chan et al., 1998), suggesting that enhancement of α-helicity of C34 may provide higher affinity to the N-HR region, thus resulting in more potent anti-HIV-1 activity. To design potent fusion inhibitors using the enhancement of α-helicity approach, we divided the α-helical peptide C34 into two characteristic interactive (a, d, e) and solvent accessible (b, c, f, g) sites according to the reported N36/C34 structure (Fig. 1) (Chan et al., 1997). When HIV-1 gp41 is folded, a tryptophan-rich domain (WRD) in the N-terminus of C-HR plays an important role in tight and specific binding, through the interaction of the hydrophobic aromatic ring with a deep groove formed by the N-HR coiled coil (Chan and Kim, 1998; Salzwedel et al., 1999). In fact, C34 contains the N-terminal WRD, which binds to a hydrophobic pocket formed by the amino acid residues L57, W60 and K63 on the N-HR trimer surface (Chan et al., 1998; Ferrer et al., 1999), resulting in higher anti-HIV-1 activity of C34 compared to T-20, which lacks the N-terminal WRD. On the other hand, the solvent accessible site appears to contribute little to the formation of the 6-helix bundles, as demonstrated by the crystal structure of C34 bound to N36 (Chan et al., 1997). Therefore, amino acids in the interactive site are indispensable for binding, whereas those in the solvent accessible site may be replaceable (Fig. 1). To enhance the α-helicity of C34, we introduced a series of systematic replacements of amino acid residues in the solvent accessible site, where the original amino acid residues were substituted with charged and hydrophilic glutamic acid (E) or lysine (K) with the intention of forming possible intrahelical salt-bridges (Marqusee and Baldwin, 1987) (Fig. 1b). We obtained two peptides, SC34 and SC34EK (Fig. 2a), both of which gratifyingly demonstrated increased anti-HIV-1 activity (Otaka et al., 2002).

In this study, we demonstrate that SC34EK maintains highly potent activity against T-20 resistant clones of HIV-1, as well as several clinical isolates, and we reveal that the enhanced α-helicity of SC34EK is indeed involved in the improvement of activity. The activities are specific to HIV-1 and are not influenced by serum components. Structural analysis indicates that electrostatic interactions introduced by EK substitutions enhance the conformational stability of the 6-helix bundle, thus preventing HIV-1 fusion with the host cell. The information from our investigations involving the enhanced α-helicity of SC34EK should enable further design of highly effective and specific HIV-1 inhibitors.

2. Materials and methods

2.1. Cells and viruses

MT-2 and 293T cells were grown in RPMI1640- and Dulbecco's modified Eagle medium (DMEM)-based culture medium, respec-

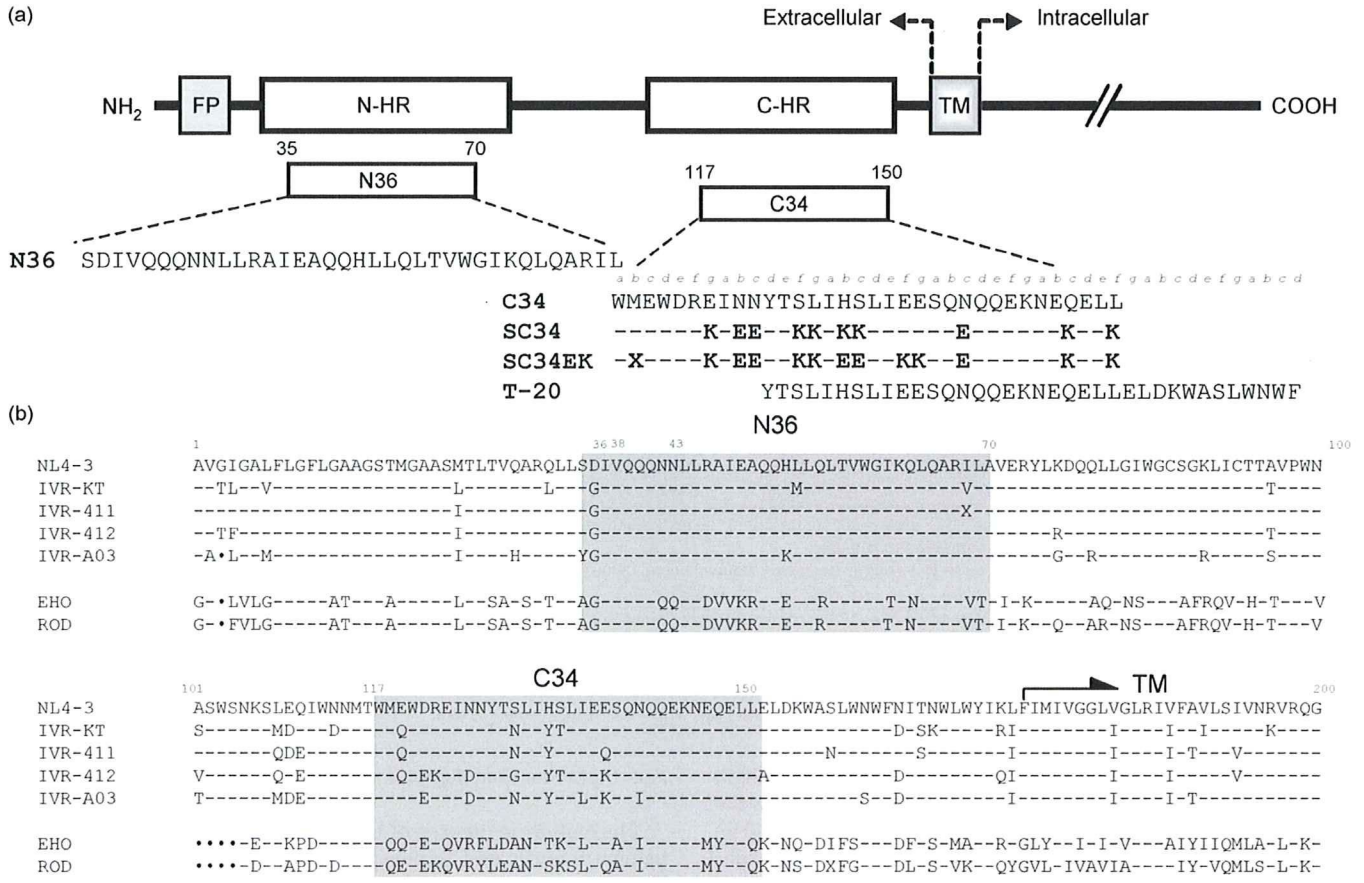

Fig. 2. Schematic view of gp41 and C34 derivatives and amino acid alignment of gp41. (a) The locations of the fusion peptide (FP), the amino-terminal heptad repeat region (N-HR), the carboxyl-terminal heptad repeat region (C-HR), and the transmembrane domain (TM) and the amino acid sequences of N36, T-20, C34 and its derivatives are shown. The residue numbers of each peptide correspond to their positions in gp41 of the NL4-3 strain. The X in SC34EK indicates norleucine, introduced to avoid oxidation of the methionine residues. No differences between the original methionine- and norleucine-containing peptide were observed (Otaka et al., 2002). (b) Alignment of amino acid sequence of clinical isolates (KT, IVR411, IVR412 and IVR-A03; GenBank accession number; AB222704, AB222705, AB222706 and AB222703, respectively) and HIV-2 strains (EHO and ROD) are shown. Corresponding regions of N36 and C34 are indicated in gray. Identical or deleted amino acids from the sequence of NL4-3 are indicated with a bar or a dot, respectively. The X in amino acid sequences of IVR411 and ROD indicates the mixture of I and V for IVR411, and mixture of I and M for ROD.

tively. HeLa-CD4-LTR-β-gal cells were kindly provided by Dr. M. Emerman through the AIDS Research and Reference Reagent Program, Division of AIDS, National Institute of Allergy and Infectious Disease (NIAID) (Bethesda, MD) and were used for the drug susceptibility assay (MAGI assay) as described previously (Kimpton and Emerman, 1992; Kodama et al., 2001; Maeda et al., 1998). The activity of test compounds was determined as the concentration that blocked HIV-1 replication by 50% (EC_{50}).

Laboratory HIV-1 (III_B) and HIV-2 (EHO and ROD) strains were used. An HIV-1 infectious clone pNL4-3 was used for constructions and for the production of HIV-1 variants as described (Nameki et al., 2005). A wild-type HIV-1, HIV-1_{WT}, was generated by transfection of pNL4-3 into 293T cells. Clinical isolates obtained from drug-naïve and heavily drug-experienced patients, were kindly provided by Dr. S. Oka (AIDS Clinical Center, International Medical Center of Japan, Tokyo, Japan). Their co-receptor tropisms were determined using NCK45 cells as described previously (Kajiwara et al., 2006).

2.2. Antiviral agent

The peptide-based fusion inhibitors used in this study were synthesized as described previously (Otaka et al., 2002), and the sequences can be identified in Fig. 2a. 3′-Azido-3′-deoxythymidine (AZT) and 2′,3′-dideoxycytidine (ddC) were purchased from Sigma (St. Louis, MO, USA). MKC-442 was provided by Dr. S. Shigeta (Fukushima Medical University, Fukushima, Japan).

2.3. Determination of drug susceptibility of HIV-1

The peptide sensitivity of infectious clones was determined using the MAGI assay with as described previously (Kodama et al., 2001; Maeda et al., 1998). The activity of test compounds was determined as the concentration that blocks HIV-1 replication by 50% (EC_{50}). For clinical isolates, PHA-stimulated peripheral blood mononuclear cells (PBMCs) were used as described previously (Kodama et al., 2001). PBMCs (10^6 cells/ml) were exposed to test compounds and HIV-1, and were cultured in the presence of interleukin 2 for 7 days. Amounts of p24 protein in the supernatants of the cultures were then determined using the commercially available p24 antigen enzyme linked solvent assay kit.

2.4. Construction of recombinant HIV-1 clone

Recombinant infectious HIV-1 clones with substituted V3 regions, pNL-$V3_{ADA}$ and pNL-$V3_{SF162}$ were generated using pNL4-3. The V3 region, corresponding to n.t. 7029–7249 of pNL4-3, was amplified using primers containing appropriate BglII and NheI restriction enzyme cleavage sites for directional cloning into pBS-

Table 1
Antiviral activity of gp41-derived peptides against gp41 and gp120 V3 recombinant virus[a]

Clone	Tropism[b]	EC_{50} (nM)					
		ddC	N36	T-20	C34	SC34	SC34EK
gp41 recombinant virus							
WT[c]		404 ± 196	180 ± 70	35 ± 17	3.2 ± 0.9	1.4 ± 0.7	0.7 ± 0.3
L33S		289 ± 24	39 ± 11	**>1000**	2.9 ± 0.9	1.3 ± 0.1	0.9 ± 0.3
V38A		714 ± 109	407 ± 76	**402 ± 68**	**96 ± 29**	2.0 ± 0.5	1.1 ± 0.6
V38E		291 ± 57	41 ± 14	**>1000**	**492 ± 85**	**37 ± 12**	**4.3 ± 1.3**
N43K		321 ± 8.5	234 ± 63	114 ± 19	**50 ± 9.5**	2.5 ± 0.3	2.7 ± 0.3
N43D		430 ± 42	461 ± 266	**>1000**	**>100**	**9.0 ± 6.6**	1.0 ± 0.8
D36S/V38M		296 ± 88	178 ± 31	42 ± 6.4	7.2 ± 4.0	1.9 ± 0.1	0.8 ± 0.3
V38E/N42S		273 ± 105	227 ± 20	**>1000**	**322 ± 7.5**	**32 ± 3.1**	3.2 ± 1.0
ΔFNSTW/L33S/N43K[d]		276 ± 39	152 ± 31	**>1000**	**248 ± 56**	2.7 ± 0.3	**4.4 ± 0.5**
ΔFNSTW/D36G/I37K/N126K/L204I[d]		246 ± 67	547 ± 7.8	**754 ± 174**	**67 ± 21**	4.6 ± 0.9	2.9 ± 0.8
gp120 V3 recombinant virus							
V3-ADA	R5	362 ± 102	360 ± 91	**289 ± 19**	6.8 ± 3.3	0.7 ± 0.4	2.0 ± 0.2
V3-SF162[e]	R5	995 ± 219	383 ± 9.9	19 ± 2.8	7.8 ± 3.5	0.5 ± 0.2	0.5 ± 0.2
V3-CH1[f]	R5X4	649 ± 4.5	**2207 ± 42**	16 ± 1	5.6 ± 0.1	1.3 ± 0.1	0.7 ± 0.1
V3-CH2[g]	R5	1515 ± 177	192 ± 13	35 ± 32	3.8 ± 0.1	0.4 ± 0	0.9 ± 0.8

[a] Anti-HIV-1 activity was determined using the MAGI assay. All data represent means ± standard deviation obtained from the results of three independent experiments. Bold indicates over 5-fold increase in EC_{50} value compared to HIV-1_{WT}.

[b] The co-receptor tropism was determined using NCK45 cells as described (Kajiwara et al., 2006).

[c] HIV-1_{NL4-3} served as a wild-type virus.

[d] ΔFNSTW is the deletion of five amino acids at position 364–368 in the gp120 V4 region of HIV-1_{NL4-3} (Nameki et al., 2005). Fusion inhibitor resistant variants used have been previously reported (Armand-Ugon et al., 2003; Nameki et al., 2005).

[e] The V3 region of NL4-3 gp120 was replaced with the corresponding region of HIV-1_{SF162}.

[f] HIV-$1_{V3\text{-}CH1}$ has mutations in the gp120 V3 region of primary isolate HIV-1_{KMT}, where GKI is substituted by GEI.

[g] HIV-$1_{V3\text{-}CH2}$ has mutation in the gp120 V3 region of the primary isolate HIV-1_{KMT}, where GKI is substituted by GQI.

$gp120_{WT}$. The resulting amplified V3 region was subjected to BglII and NheI digestion, subcloned into pBS-$gp120_{WT}$ containing the corresponding region in the DNA fragment of EcoRI–NheI (1510 bp containing gp120 V1, V2 and V3, n.t. 5740–7249 of pNL4-3) and subsequently ligated into pNL4-3. pNL-$V3_{CH1}$ and $V3_{CH2}$, CCR5 and dual (CXCR4 and CCR5) tropic molecular clones, were kindly donated by Dr. Y. Maeda, Kumamoto University (Kumamoto, Japan) (Foda et al., 2001; Maeda et al., 2000).

Recombinant infectious HIV-1 clones carrying various mutations in gp120 and/or gp41 were also generated using pNL4-3. Briefly, the desired mutations were introduced using site directed mutagenesis into the region of pSL-$gp41_{WT}$ flanked by the NheI–BamHI restriction enzyme sites (1220 bp containing gp120 V4, V5 and gp41 ectodomain n.t. 7250–8469 of pNL4-3) (Weiner et al., 1994). After restriction enzyme digestion and purification the NheI–BamHI fragments were ligated into pNL4-3, generating a series of molecular clones with the desired mutations.

Each molecular clone was transfected into 293T cells (10^5 cells/6-well culture plate). After 48 h, MT-2 cells (10^6 cells/well) were added and co-cultured with the 293T cells for an additional 24 h. When an extensive cytopathic effect was observed, the supernatants were harvested and stored at −80 °C for further use.

Table 2
Antiviral activity of gp41-derived peptides against clinical isolates[a]

Strain	EC_{50} (nM)				
	AZT	T-20	C34	SC34	SC34EK
NL4-3 (WT)[b]	2.0	36	3.2	0.36	0.4
KT (WT)[b]	2.0	11	0.2	0.1	0.03
IVR411	**7600**	4.1	0.2	3.1	0.04
IVR412	**9060**	23	7.2	**4.8**	0.1
IVR-A03	**1200**	7.0	**17**	**4.1**	0.7

[a] Anti-HIV-1 activity was determined using the amounts of p24 protein in the supernatants of the PHA-stimulated PBMC cultures using commercially available ELISA kit (Kodama et al., 2001). Bold indicates over 5-fold increase in EC_{50} value compared to HIV-1_{WT}.

[b] HIV-1_{NL4-3} and HIV-1_{KT} served as controls.

2.5. Determination of gp41 amino acid sequence

Nucleotide sequences of the clinical isolates were determined using an automated sequencer. Briefly, DNA was extracted from PBMCs infected with the clinical isolates, subjected to nested PCR for the gp41 coding region, and then directly sequenced as described previously (Nameki et al., 2005).

2.6. Measurement of circular dichroism (CD) spectra

N-HR peptides (N36, $N36_{V38A}$ or $N36_{N43D}$) and C-HR peptides (C34 or SC34EK) were incubated at 37 °C for 30 min (the final concentration of both the N-HR peptide and the C-HR peptide were 10 μM in pH 7.4, 12 mM phosphate-buffered solution containing 50 mM NaCl). The wavelength-dependence of molar ellipticity [θ] was monitored at 25 °C as the average of eight scans, and the thermal stability was estimated by monitoring the change in the CD signal at 222 nm in a spectropolarimeter (Model J-710; Jasco, Tokyo, Japan) equipped with a thermoelectric temperature controller. The midpoint of thermal unfolding transition (melting temperature [T_m]) of each complex was determined as described previously (Otaka et al., 2002). The percentages of a-helicity in 6-helix complexes were calculated by comparing the CD signal at 222 nm of N36/C34 or N36/SC34EK complexes in a spectropolarimeter.

2.7. Crystallization, data collection and refinement

Samples for crystallization were prepared by mixing solutions of N36 and SC34EK dissolved in 10 mM sodium acetate buffer at a concentration of 10 mg/mL. The mixture was incubated for 30 min at 37 °C, then was passed through a 22 μm filter. Crystallization was performed by the hanging drop vapor diffusion method at 4 °C. Droplets were prepared of equal amounts (2 μL) of reservoir solution and the peptide solution. Hexagonal prism crystals were obtained under the following conditions: 100 mM sodium acetate buffer (pH 4.0), 200 mM ammonium sulphate, 14% polyethylene glycol monomethyl ether 2000. After screening of

Table 3
Antiviral activity of HIV-1 gp41-derived peptides against HIV-2[a]

HIV-2 strain	EC_{50} (nM)				
	ddC	T-20	C34	SC34	SC34EK
WT[b]	404 ± 196	35 ± 17	3.2 ± 0.9	1.4 ± 0.7	0.7 ± 0.3
HIV-2_{EHO}[c]	925 ± 188	14 ± 3.0 (×0.4)	**639 ± 87 (×200)**	**68 ± 10 (×49)**	**17 ± 1.2** (×24)
HIV-2_{ROD}[d]	**1808 ± 927**	**176 ± 68 (×5)**	**>1000 (>×313)**	**251 ± 29 (×179)**	**115 ± 33** (×164)

[a] Anti-HIV-2 activity was determined using the MAGI assay. All data represent mean ± standard deviation obtained from the results of three independent experiments. Bold indicates over 5-fold increase in EC_{50} value compared to HIV-1_{WT}.
[b] HIV-1_{NL4-3} served as a wild-type virus.
[c] HIV-2_{EHO} was dual-tropic HIV-2.
[d] HIV-2_{ROD} was T-tropic HIV-2.

various cryo-conditions, the suitable condition was found to be the addition of 35% xylitol to the peptide solution and a slight increase in the amount of the precipitant (*ca* 14.5%). The obtained crystals were easily broken by direct transfer from the crystallization condition to the cryo-condition, but the transfer of the fragile crystals could be accomplished by gradual change in conditions using stepwise increase in the amount (0–35% in five steps) of the cryoprotectant.

Data were collected at a beamline BL38B1 of SPring-8. Collected data were processed using DENZO and SCALEPACK from the HKL2000 package (Otwinowski and Minor, 1997). A molecular replacement solution was found using AMoRe (Navaza, 2001), with a molecular model of the HIV-1 gp41 core structure (PDB code: 1AIK). Model refinements and reconstruction were performed using REFMAC5 (Murshudov et al., 1999) and XtalView (McRee, 1999). The final model was refined at a resolution of 2.1 Å, to a crystallographic *R* value of 0.213 and a free *R* value of 0.238. Detailed data collection and refinement statistics are summarized in Table 1. Atomic coordinates and structural factors have been deposited at the Protein Data Bank (PDB code:2Z2T).

3. Results

3.1. Anti-HIV-1 activity of SC34 and SC34EK

We examined the anti-HIV-1 activity of SC34 and SC34EK against not only HIV-1_{WT} but also T-20- and/or C34-resistant clones observed in vitro. SC34 and especially SC34EK that has aligned EK modification more effectively suppress HIV-1 infection compared to C34 and T-20 (Table 1). D36S/V38M substitutions in the gp41 region (HIV-$1_{D36S/V38M}$), and a five amino acid (FNSTW) deletion in the V4 region of gp120 (ΔV4) with L33S/N43K in the gp41 region (HIV-$1_{\Delta V4/L33S/N43K}$) were isolated in vitro (Fikkert et al., 2002; Rimsky et al., 1998). L33S was also selected during C34-resistant induction in vitro (Armand-Ugon et al., 2003). C34 and its derivatives effectively inhibit entry of these clones into the host cell. In particular, SC34EK maintained strong activity even against V38E containing clones, such as HIV-$1_{V38E/N42S}$ (Armand-Ugon et al., 2003), which showed cross-resistance to T-20, C34 and SC34. Reduction of activities by SC34 and SC34EK was moderate in HIV-$1_{\Delta FNSTW/L33S/N43K}$ that showed high level resistance to T-20 and C34. Next, we examined the antiviral activities of C34 derivatives against clones containing major primary mutations V38A and N43D, which are mutations frequently observed in T-20 resistant variants in vivo (Cabrera et al., 2006; Derdeyn et al., 2001; Menzo et al., 2004; Poveda et al., 2004; Poveda et al., 2002; Xu et al., 2005) (Table 1). SC34 reduced its antiviral activities against HIV-1_{N43D}, while SC34EK maintained its potent activity, indicating that when EK is bound with the complementary electrostatic interactions appropriately aligned SC34EK can effectively suppress the infection by various clones resistant to T-20 and C34 both in vitro and in vivo.

We further evaluated activities of SC34 and SC34EK against V3-substituted clones (Table 1). HIV-$1_{V3\text{-}ADA}$ uses mainly the CCR5 co-receptor for its entry into the host cells and has been reported to moderate T-20 resistance (≈10 fold), compared to the CXCR4 using strain of HIV-1, which shows higher susceptibility to fusion inhibitors (Reeves et al., 2002). As reported, the susceptibility of HIV-$1_{V3\text{-}ADA}$ to T-20 decreased, however, C34 and its derivatives maintained their activity against the same variant. Interestingly, in our experiments, HIV-$1_{V3\text{-}SF162}$, HIV-$1_{V3\text{-}CH1}$ and HIV-$1_{V3\text{-}CH2}$ also showed comparable susceptibility to T-20. These results indicate that sequence variations in the V3 region do not always correlate with the observed T-20 susceptibility and are not involved in the resistance to C34 and its derivatives.

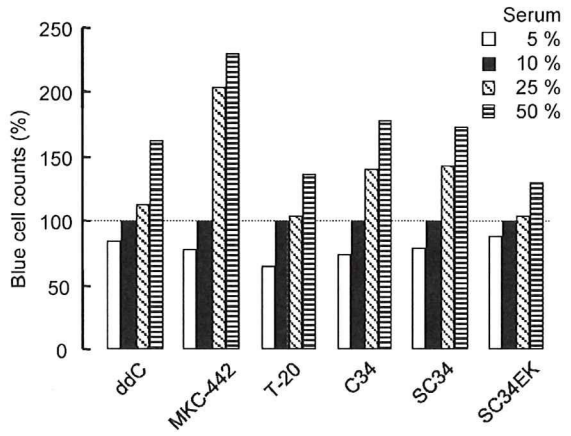

Fig. 3. Effect of FCS concentrations on anti-HIV-1 activity. Changes in the blue cell counts at various concentrations of FCS are shown. Blue cell counts at EC_{80} value in 10% FCS concentration (black bar) were used and set as 100%. White, black, hatched, and striped bars correspond to 5, 10, 25, and 50% FCS, respectively. Inhibitors for reverse transcriptase, ddC and MKC-442, and for fusion, T-20 were used as controls.

3.2. Amino acid sequence

Amino acid sequences of clinical isolates are shown in Fig. 2b. One isolate, HIV-1_{KT}, was obtained from a drug-naïve patient and the other three isolates (HIV-1_{411}, HIV-1_{412}, HIV-1_{A03}) were obtained from heavily drug-experienced patients. None of the patients had received T-20 therapy. Amino acid sequences of the N-HR were highly conserved within all HIV-1 clinical isolates with some small variations. In contrast, the N36 region of the two HIV-2 strains, EHO and ROD, was identical in both HIV-2 isolates. We found some variations in the amino acid sequences of the HIV-2 strains we isolated, as compared with the sequences deposited in the GenBank (accession number; M15390 and X05291 for HIV-2_{ROD}, and U272000 for HIV-2_{EHO}). Namely, we identified two different amino acids in the isolated HIV-2_{ROD}, V26L and

I157I/M (mixture of I and M), and one variation in the amino acid sequence of HIV-2_{EHO}, V45L. Except for I157M, other substitutions are observed in the majority of the HIV-2 strains, as reported in the HIV sequence database (Los Alamos National Laboratory: Los Alamos, NM, USA, http://www.hiv.lanl.gov). These substitutions might be introduced through different culture conditions, (e.g., host cells used for the propagation). We considered these substitutions as a polymorphism.

Sequence homology of the N36 region of the isolated HIV-1 strains was 31/36 (86%), including mutation D36G that is observed in the vast majority of HIV-1 strains (Kuiken et al., 2001). In contrast, those of the C34 region were relatively heterogeneous, 24/34 (71%) for HIV-1 and 12/34 (35%) for HIV-2. Sequence identity of the T-20 region (residues 117–152) in the HIV-1 strains was also variable 27/36 (75%), while in the HIV-2 strains the sequence identity was 15/36 (42%). These results indicate that even highly conserved two helical extracellular domain of the gp41 can allow polymorphisms.

3.3. Efficacy of the peptides against clinical isolates

To evaluate preclinical efficacy, we examined the antiviral activity of C34, SC34 and SC34EK against clinical isolates (Table 2). Replication of HIV-1_{NL4-3} and HIV-1_{KT}, a drug-naïve strain, was suppressed by all compounds tested. C34 showed decreased activity against HIV-$1_{IVR-A03}$, which was isolated from a heavily drug-exposed patient. SC34 also showed reduced susceptibility against three drug-experienced strains. However, it is difficult to conclude whether SC34 showed enhanced susceptibility against HIV-1_{KT} or reduced susceptibility against drug resistant strains. In contrast, T-20 and SC34EK suppressed the replication of all isolates tested to similar extents in EC_{50} values compared to HIV-1_{NL4-3} (Table 2), indicating that SC34EK with appropriately aligned EK residues effectively suppresses the replication of the clinical 3 isolates.

3.4. Anti-HIV-2 activity

To confirm the target specificity, we examined antiviral activities of SC34 and SC34EK against two HIV-2 strains, EHO and ROD. Compared to HIV-1_{NL4-3}, EHO and ROD contain 19 and 22 amino acid substitutions in the C34 region, respectively, and 15 amino acid substitutions in the N36 region, the anticipated site of binding of SC34 and SC34EK peptides (Fig. 2b). Like the parent peptide C34, both SC34 and SC34EK lost their potent activities (Table 3). Compared to HIV-1_{NL4-3}, 6 out of 19 residues in the C34 region of HIV-2_{EHO} and 7 out of 22 residues in the C34 region of HIV-2_{ROD} are located at positions *a*, *d*, and *e* that directly interact with the N36 binding surface. These substitutions in the N36 and C34 region in HIV-2 may be responsible for reduced anti-HIV-2 activities of the peptides derived from HIV-1. At present, we cannot conclude which amino acid substitutions are directly involved in the reduced susceptibility of the HIV-2 strain to the treatment with the peptide fusion inhibitor, and/or whether other regions besides the N36 and C34 regions might influence peptide susceptibility. However, our results indicate that SC34 and SC34EK maintain similar target specificity to the parent peptide, C34.

3.5. Effect of fetal calf serum (FCS) on anti-HIV-1 activity

To estimate the stability of the peptides in vivo, binding level of SC34EK, to serum components, (e.g., albumin) was examined. In this experiment, the antiviral activity in the presence of relatively high concentrations of fetal calf serum (FCS) was determined (Baba et al., 1993) (Fig. 3). EC_{80} values of the fusion inhibitors against HIV-1 replication in vitro were used. In the presence of 50% FCS, the activity of MKC-442 (I-EBU), a lipophilic non-nucleoside RT inhibitor, was reduced 2.3-fold compared with 10% FCS as described previously (Baba et al., 1993). However, the activities of SC34, SC34EK and T-20 were little influenced by serum components. Among the three, SC34EK was the least affected by the concentration of FCS.

We further examined the stability of peptide inhibitors in freshly prepared human sera ($n = 3$). After 1 h incubation of peptides in human sera (final concentration of 200 μM) at 37 °C, the anti-HIV-1 activity was examined using the MAGI assay. Comparable activities of all peptides tested were observed either with or without the incubation (data not shown). These results indicate that hydrophilic SC34EK likely retains its strong anti-HIV-1 activity in vivo, similarly to T-20, because of its low non-specific binding and protease cleavage in serum.

3.6. Peptide binding affinity

To clarify the mechanism of potent anti-HIV-1 activity observed with SC34EK, the binding affinity of SC34EK was evaluated by collecting the CD spectra using synthetic peptides. The CD spectra of equimolar mixtures of the N-HR and C-HR peptides showed spectrum minima at 208 and 222 nm, which indicate the presence of stable α-helical conformations. All combinations of peptides showed similar spectra at 25 °C, indicating that these peptides contained the same α-helicity (Fig. 4a), although the spectrum of C34 with N36 and N43D mutation ($N36_{N43D}$) indicated only weak α-helicity. These results indicate that N43D might reduce the stability of the conformation of the 6-helix bundle, thus decreasing the replication of HIV-1, whereas V38A does not. SC34EK formed stable 6-helix conformations with $N36_{V38A}$ and $N36_{N43D}$. Under these experimental conditions, wavelength-dependent spectra were similar with the exception of the spectrum of the $N36_{N43D}$/C34 complex. Thus, we analyzed thermal stabilities, defined as the midpoint of the thermal unfolding transition (T_m) values, of the potential 6-helix bundles of N-HR and C-HR peptides. T_m of N36/C34 was found to be 52.0 °C, while that of $N36_{V38A}$/C34 and $N36_{N43D}$/C34 decreased to 44.5 and 34.0 °C, respectively (Fig. 3b). In contrast, thermal stabilities of $N36_{V38A}$/SC34EK, $N36_{N43D}$/SC34EK and N36/SC34EK were much higher, 60.5, 56.0 and 69.5 °C, respectively. Thus, binding affinity of SC34EK to N-HR was stronger compared to that of C34. Alternatively, at the physiological temperature of 37 °C, only 60 and 40% of the α-helix content was observed in $N36_{V38A}$/C34 and $N36_{N43D}$/C34 mixtures, respectively, indicating that roughly half of C34 failed to form stably 6-helix bundle with the target N-HR harboring resistant mutations. Therefore, C34 reduces its anti-fusion activity exerted by dominant negative effect. In contrast, only 20% of the unfolded α-helix content was observed in SC34EK with mutated N36, which indicated that at 37 °C, binding of SC34EK to mutated N36 was comparable to that of C34 with wild-type N36 (Fig. 4b). Moreover, physiochemical properties of N-HR and SC34EK complexes, defined by T_m value, correlated well with their ability to inhibit HIV-1 fusion (Fig. 4c). These results suggest that the stability of the 6-helix complex, as judged by the binding stability (affinity), is directly correlated with the anti-HIV-1 activity.

3.7. Crystal structure of the N36/SC34EK complex

The crystal structure of the complex between SC34EK and the N-HR representative peptide N36 was resolved to a resolution of 2.1 Å (Table 4). In the asymmetric unit, a 6-helix bundle consisting of a central helix bundle of three N36 peptides surrounded by three SC34EK peptides was found. This arrangement is similar in the core structure of gp41 (Chan et al., 1997). Structural superimposition of the original gp41 core and the N36/SC34EK complex showed a good match, with an RMSD value of 0.59 for main-chain atoms

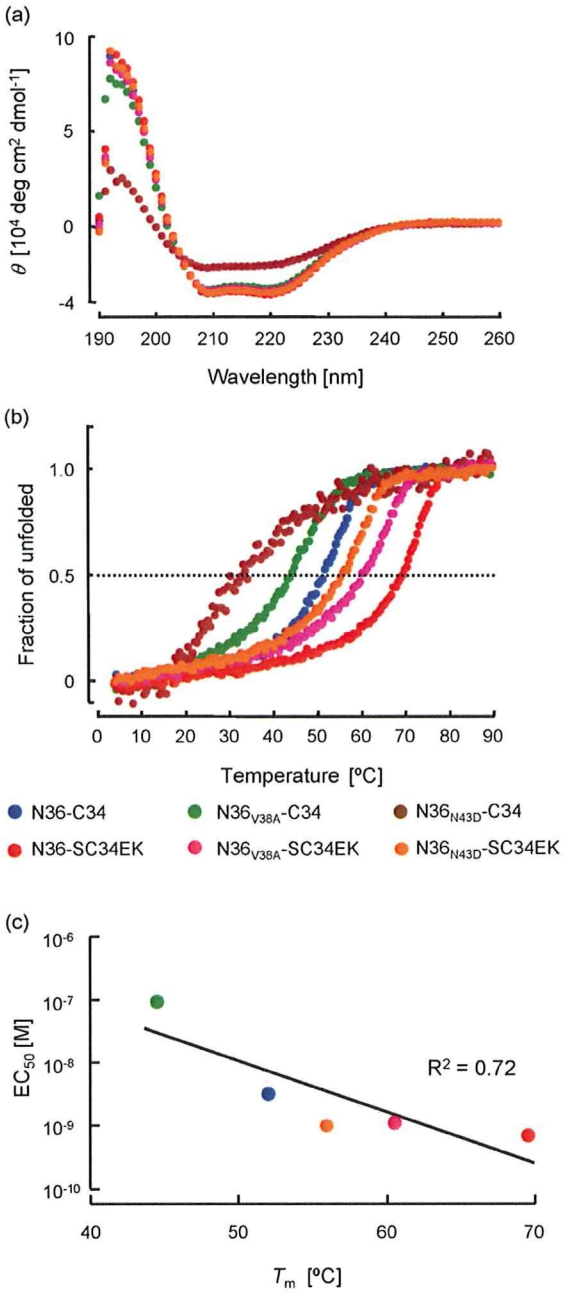

Fig. 4. CD analysis of peptide complex between resistant variants of N36 and C34 or SC34EK. (a) Wavelength-dependent CD spectra of the complexes in solution. The spectrum minima at 208 and 222 nm indicated the presence of stable α-helical conformations. (b) Thermal midpoint analysis was measured at 222 nm CD signal for the N and C peptide complexes. Final concentration of each peptide was 10 mM. The arrow indicates the physiological temperature of 37 °C. (c) The correlation between T_m (b) and EC_{50} values (Table 1). Colors of plots correspond to those in panels (a) and (b). Combination of $N36_{N43D}$ and C34 (EC_{50} > 100 nM) is excluded.

(Fig. 5a and b). Hydrophobic contacts between SC34EK and N36 with tryptophan rich domain (WRD) and leucine zipper were preserved for the original gp41 core. All introduced charged residues of the EK motif were directed toward the solvent (Fig. 5c). As a direct consequence of introducing the EK motifs, the ratio of surface area occupied by charged residues to the total surface area was increased from 35% in the original molecule to 60% in the N36/SC34EK complex. Importantly, it appeared that tight bonding, such as ion pairing or hydrogen bonding, was not present in the

Table 4
Crystallization, data collection and refinement statistics

Data collection	BL38B1 Spring-8
Temperature (K)	100
Space group	$P3_121$
Cell dimensions *a*, *b*, *c* (Å)	105.01, 105.01, 78.31
Resolution limits (Å)	90.00–2.10
Number of unique reflections	29,461
Average redundancy	7.53
Completeness (%)	99.7
R_{merge}[a]	0.122
Refinement statistics	
Refinements resolution range (Å)	20.00–2.20
R/R_{free}[b] (%)	0.213/0.238
The highest resolution shell (Å)	2.15–2.10
R/R_{free}[b] (%)	0.231/0.255
RMSD from ideal	
Bonds (Å)	0.010
Angles (°)	1.015
⟨*B*⟩ for atomic model[c] (Å²)	29.93
Ramachandran plot	
Most favored regions (%)	100

[a] $R_{merge} = \sum|I_h - \langle I_h\rangle| / \sum I_h$, where $\langle I_h\rangle$ is the average intensity of reflection *h* and symmetry-related reflections.
[b] *R* and $R_{free} = \sum||F_o| - |F_c|| / \sum|F_o|$ calculated for reflections of the working set and test (5%) set, respectively.
[c] ⟨*B*⟩ is the average temperature factor for all protein atoms.

side-chains of the residues of the EK motif. Electrostatic interaction may involve in constrained structure which provides the enhanced α-helicity observed (Fig. 4). This structural analysis demonstrated that the interaction between N36 and SC34EK retained the ability to form the 6-helix bundle structure despite the substitution of more than one third of the residues (13/34) in the sequence of SC34EK.

4. Discussion

In this study, we characterized a novel α-helical peptide, SC34EK that effectively inhibits replication of HIV-1 strains resistant to T-20 and C34. The activity was specific to HIV-1 and little influenced by serum components. We demonstrate that the potent anti-HIV-1 activity of SC34EK is derived from its high affinity to the N-HR region by the CD analysis. Further, we reveal that SC34EK binds to its target, N-HR in identical manner that C34 does by the structure analysis.

The structural analysis of the N36/SC34EK complex clearly demonstrated that the interaction between SC34EK and N36 peptides was maintained by hydrophobic contacts and that the EK motif was directed toward the solvent. The introduction of the EK residues increased the proportion of accessible surface area occupied by charged residues. Although tight bonding was not observed, a continuous electrostatic potential between the EK residues may serve to stabilize the helix bundle. Such helix stabilization, which might occur on the surface of the HIV-1 virion between SC34EK and the N36 region of gp41, could result in the high anti-HIV-1 activity. In this regard, SC34EK, containing an aligned EK motif, showed more potent anti-HIV-1 activity compared to SC34, which has one misaligned EK motif (Fig. 2a). Increasing the hydrophilic surface area may prevent aggregation of SC34EK as compared to parental peptide C34. Therefore, SC34EK might distribute into the various organs in the body without being trapped and destroyed in the reticular systems or having its activity reduced by non-specific binding to proteins (e.g., albumin) (Fig. 3).

We further demonstrate that SC34EK specifically binds to the target, N-HR of HIV-1, since it only exerted weak activity to two

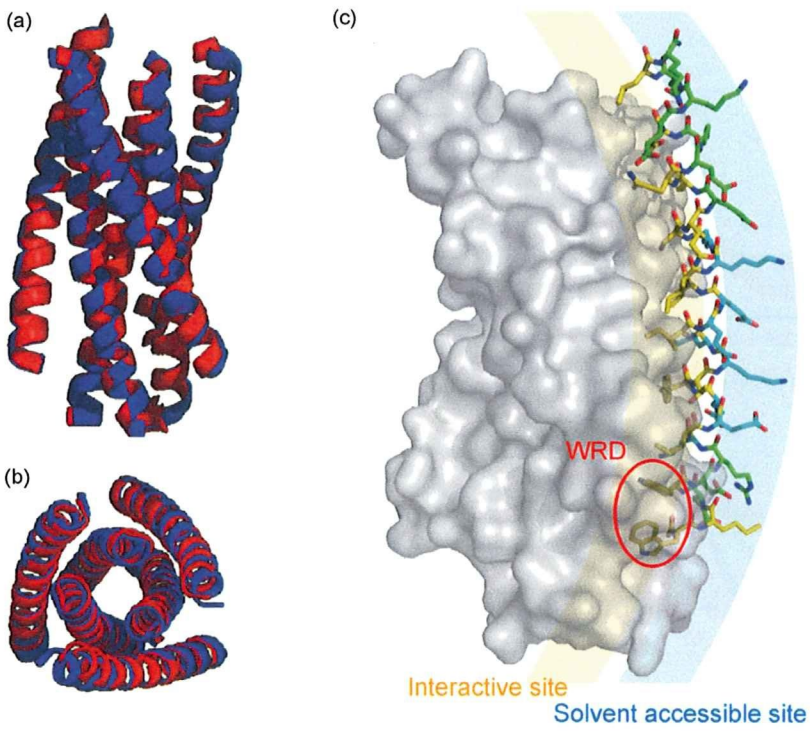

Fig. 5. Structure of the 6-helix bundle formed by N36 and SC34EK. (a and b) The gp41 core structure and N36/SC34EK complex are shown in red and blue, respectively. (c) Stick model representation of SC34EK. The stick model of SC34EK is shown, and three N36s in the core and two other surrounded SC34EK are represented in gray. SC34EK showed amphiphilic properties. The location of the N-terminal tryptophan rich domain (WRD) in SC34EK is indicated by a red circle. Original and introduced charged amino acids are indicated in green and blue, respectively.

HIV-2 strains that contain 15 amino acid substitutions in the N-HR compared to HIV-1 NL4-3 strain (Table 3 and Fig. 2b). These results suggest that to develop resistance to SC34EK, at least, certain mutations in not only the N-HR but also the C-HR are required to be introduced. This might delay emergence of resistant HIV-1 variants to SC34EK in vivo.

So far, some approaches for stabilizing α-helix structures through the introduction of artificial amino acids were reported for HIV-1 fusion inhibitors T-20 (Judice et al., 1997) and C34 (Sia et al., 2002), including an example of an amino acid containing terminal olefin-derived side chains, designed as a substrate for the ring-closing olefin metathesis (Blackwell et al., 2001) and an example of a hydrocarbon-stapled peptides (Phelan et al., 1997) Walensky et al. (2004) applied a hydrocarbon-stapled modification to generate peptides that bind to the BH3 helical domain of Bcl-2, an anti-apoptotic protein, and demonstrated that a synthesized peptide mimic that binds to the BH3 domain activates apoptosis in leukemic cells. However, all peptides exerted only moderate activity in vivo, although they showed efficient binding to the target proteins in vitro (Blackwell et al., 2001; Judice et al., 1997; Sia et al., 2002; Walensky et al., 2004). It is likely that during the formation of the 6-helix bundle and the fusion process, gp41 changes its conformation drastically, suggesting that a flexible conformation of the peptide may be required to preserve actual inhibition. Compared with tethered, constrained peptides, EK modification that facilitates electrostatic stabilization displays such flexibility while exhibiting enhanced α-helicity. Most recently, T290676, a 38 amino acid peptide, has been reported to suppress various fusion inhibitor-resistant strains of HIV-1 (Dwyer et al., 2007). Like SC34EK, T290676 is substituted with the charged and hydrophilic amino acids, glutamic acid (E) and arginine (R), at the solvent accessible site and shows potent anti-HIV-1 activity.

In conclusion, we have demonstrated that SC34EK selectively inhibits various HIV-1 strains, including T-20 resistant clones, through increased stability of the α-helix. The sequence of the solvent accessible site of α-helical peptides is replaceable and modifications of this sequence can regulate α-helicity with target specificity. Therefore, our approach of introducing the EK motif in the α-helical structure of the peptide inhibitor will help to generate future peptide inhibitors with high anti-HIV efficacy and potentially fewer adverse effects.

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Design of Peptide-based Inhibitors for Human Immunodeficiency Virus Type 1 Strains Resistant to T-20*[S]

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Enfuvirtide (T-20) is a fusion inhibitor that suppresses replication of human immunodeficiency virus (HIV) variants with multi-drug resistance to reverse transcriptase and protease inhibitors. It is a peptide derived from the C-terminal heptad repeat (C-HR) of HIV-1 gp41, and it prevents interactions between the C-HR and the N-terminal HR (N-HR) of gp41, thus interfering with conformational changes that are required for viral fusion. However, prolonged therapies with T-20 result in the emergence of T-20-resistant strains that contain primary mutations such as N43D in the N-HR of gp41 (where T-20 and C-HR bind) that help the virus escape at a fitness cost. Such variants often go on to acquire a secondary mutation, S138A, in the C-HR of gp41 region that corresponds to the sequence of T-20. We demonstrate here that the role of S138A is to compensate for the impaired fusion kinetics of HIV-1s carrying primary mutations that abrogate binding of T-20. To preempt this escape strategy, we designed a modified T-20 variant containing the S138A substitution and showed that it is a potent inhibitor of both T-20-sensitive and T-20-resistant viruses. Circular dichroism analysis revealed that the S138A provided increased stability of the 6-helix bundle. We validated our approach on another fusion inhibitor, C34. In this case, we designed a variant of C34 with the secondary escape mutation N126K and showed that it can effectively inhibit replication of C34-resistant HIV-1. These results prove that it is possible to design improved peptide-based fusion inhibitors that are efficient against a major mechanism of drug resistance.

HIV-1[2] entry into the target cells is mediated by two envelope glycoproteins, gp120 and gp41, that form a trimeric gp120·gp41 complex. After binding of gp120 to the CD4 receptor and CCR5 (or CXCR4) coreceptor on the surface of the target cell, the gp41 trimer forms an extended conformation of the three helices that allows a hydrophobic fusion peptide to be inserted into the target cell membrane, generating an intermediate that is anchored to both cellular and viral membranes. After this step, the gp41 is believed to start refolding to a more stable 6-helix bundle composed of the α-helical trimer of the N-terminal heptad repeat (N-HR) folded into an anti-parallel conformation with the three C-terminal heptad repeats (C-HR) (1, 2). This refolding brings the viral and cellular membranes together to catalyze fusion.

The transition of the extended intermediate to the 6-helix bundle can be inhibited by the addition of exogenous peptides derived from gp41 C-HR (Fig. 1*A*) that prevent the formation of the 6-helix bundle and inhibit the HIV-1 fusion with the target cells (3–6). T-20, a 36-amino acid peptide derived from C-HR, effectively suppresses *in vivo* replication of HIV-1 resistant to inhibitors of reverse transcriptase and protease (7, 8). However, HIV-1 variants resistant to T-20 have recently emerged carrying primary mutations in the Leu-33–Leu-45 region of the N-HR domain (9–15). Among them, V38A and N43D seem to be major primary mutations for T-20 resistance. Meanwhile, a secondary mutation at the C-HR region (S138A) has been reported to enhance T-20 resistance with an as yet undefined mechanism (9, 14, 15) (Fig. 1*B*).

The mechanism of resistance to C34, another C-HR peptide-based inhibitor of HIV fusion, has been the subject of multiple studies (13, 16). Because of a 22-amino acid overlap between the T-20 and C34 peptides (Fig. 1*B*), HIV-1 has developed primary mutations for C34 resistance *in vitro* at the identical Leu-33–Leu-45 region of the peptides. During *in vitro* selection of C34 resistance, we identified a mutation in the C-HR domain, N126K, that is also observed in some T-20-resistant clinical variants (10, 15, 17). We showed that N126K conferred resistance to C34 by compensating for the impaired intra-gp41 inter-

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[S] The on-line version of this article (available at http://www.jbc.org) contains supplemental Figs. 1 and 2 and Tables 1 and 2.

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[2] The abbreviations used are: HIV, human immunodeficiency virus; T-20, enfuvirtide; HR, heptad repeat; MAGI, multinuclear activation of galactosidase indicator; EC_{50}, 50% effective concentration; T_m, melting temperature; CD, circular dichroism; shRNA, short hairpin RNA; WT, wild-type.

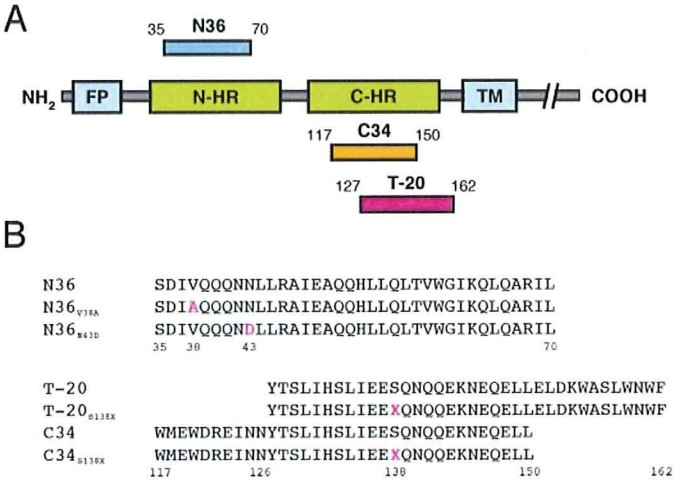

FIGURE 1. **Schematic view of gp41 and peptide sequence.** *A*, structure of HIV-1 gp41 and locations of N-HR or C-HR peptides (*FP*, fusion peptide; *TM*, transmembrane domain). *B*, amino acid sequences of peptides used in this study. Only the amino acid located at Ser-138 was substituted with all physiological amino acids (*X*), as Asn-126 lies outside of the amino acid sequence of T-20.

action by a primary mutation, I37K (13). N126K was initially identified in background of V38A, another primary mutation, for T-20 resistance *in vivo* (17). Baldwin *et al.* (17, 18) demonstrated a striking T-20-dependent replication phenotype in the V38A/N126K variant and proposed that T-20 acts as a safety pin to prevent premature formation of helical bundle, as N126K enhanced binding capacity of the introduced C-HR to N36 with V38A. Taken together, these studies suggest that mutations in the C-HR serve as secondary mutations.

In this study we show that the main role of secondary mutations that follow the appearance of primary mutations during treatment with peptide-based fusion inhibitors is to compensate for the impairment in replication kinetics that is caused by the primary mutations (supplemental Fig. 1). Based on this finding we hypothesized that analogs of T-20 carrying substitutions corresponding to secondary T-20 resistance mutations should be active against both wild-type and T-20-resistant viruses containing primary mutations. Indeed, our results confirmed our hypothesis and showed that T-20 with the S138A substitution ($T\text{-}20_{S138A}$) has a strong anti-HIV-1 activity even against T-20-resistant clones. Moreover, we demonstrate that this restoration is concomitant to improved binding of $C\text{-}HR_{S138A}$ to $N\text{-}HR_{N43D}$, suggesting that our approach utilizing the resistance-associated mutations to design peptides may provide useful broad insights into effective peptide-based therapies.

EXPERIMENTAL PROCEDURES

Cells and Viruses—MT-2 cells were grown in RPMI 1640 medium. 293T cells were grown in Dulbecco's modified Eagle's medium-based culture medium. HeLa-CD4-LTR-β-gal cells were kindly provided by M. Emerman through the AIDS Research and Reference Reagent Program, Division of AIDS, NIAID, National Institutes of Health (Bethesda, MD) and were used for the drug susceptibility assay as described previously (13, 19, 20). An HIV-1 infectious clone, pNL4–3 (21), was used for generation of HIV-1 variants.

Antiviral Agents—The peptides used in this study were synthesized as described previously (6).

Determination of Drug Susceptibility of HIV-1—The peptide sensitivity of infectious clones was determined by the multinuclear activation of galactosidase indicator (MAGI) assay as described previously (13). Briefly, the target cells (HeLa-CD4-LTR-β-gal; 10^4 cells/well) were plated in 96-well flat microtiter culture plates. On the following day the cells were inoculated with the HIV-1 clones (60 MAGI unit/well, giving 60 blue cells after 48 h of incubation) and cultured in the presence of various concentrations of drugs in fresh medium. Forty-eight hours after viral exposure, all the blue cells stained with 5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside (X-gal) were counted in each well. The activity of test compounds was determined as the concentration that blocked HIV-1 replication by 50% (50% effective concentration, (EC_{50}).

Generation of Recombinant HIV-1 Clones—Recombinant infectious HIV-1 clones, carrying various mutations, were generated as described previously (13). Each molecular clone was transfected into 293T cells with *Trans*IT® (Madison, WI). After 48 h, the supernatants were harvested and stored at −80 °C until use.

Circular Dichroism Spectroscopy—Each peptide (10 μM) was mixed with 10 mM phosphate-buffered saline, pH 7.4, and the data were collected using a Jasco spectrometer (Model J-710; Jasco, Tokyo, Japan) equipped with a thermoelectric temperature controller. The thermal stability was assessed by monitoring the change in the circular dichroism signal at 222 nm. The midpoint of the thermal unfolding transition (melting temperature, (T_m) of each complex was determined as described previously (6).

Viral Replication Kinetics Assay—MT-2 cells (10^5 cells/3 ml) were infected with each virus preparation (1000 MAGI unit) for 16 h. The infected cells were then washed and cultured in a final volume of 3 ml. The culture supernatants were harvested after infection on days 2–7, and the levels of p24 antigen were determined (22).

For each competitive HIV-1 replication assay, two infectious clones of interest that had been previously titrated were mixed and added to MT-2 cells (10^5 cells/3 ml) as described previously (13, 22) with minor modifications. To ensure that the two infectious clones being compared were of approximately equal infectivity, a fixed amount (500 MAGI unit) of one infectious clone was mixed with three different amounts (250, 500, and 1000 MAGI unit) of the other infectious clone. On day 1, one-third of the infected MT-2 cells were harvested and washed twice with phosphate-buffered saline, and the cellular DNA was extracted. The purified DNA was subjected to nested PCR and then direct DNA sequencing. The HIV-1 co-culture, which best approximated a 50:50 mixture on day 1, was further propagated. Every 3–4 days, the co-culture supernatant (100 μl) was transmitted to new uninfected MT-2 cells (5×10^5 cells/3 ml). The cells harvested at the end of each passage were subjected to direct sequencing, and the viral population change was determined.

Structure Modeling of gp41 S138A Mutant Core—The gp41 core model was built using the coordinates of crystal structure of the N36/C34 complex (23) (PDB code 1AIK). The coordi-

TABLE 1

Antiviral activity of T-20-derived peptides against T-20-resistant gp41 recombinant viruses

Anti-HIV activity was determined with the MAGI assay. The data shown are the mean values and S.D. that were obtained from the results of at least three independent experiments. Shown in parentheses are the -fold increases in resistance (increase in EC_{50} value) calculated by comparison to a reference virus. Increases of >10-fold are indicated in bold.

	EC_{50}			
	HIV-1_{WT}[a]	HIV-1_{V38A}	HIV-1_{N43D}	HIV-$1_{N43D/S138A}$
	nM			
T-20	2.4 ± 0.6	23 ± 8.2 (9.6)	49 ± 10 (20)	84 ± 16 (35)
Small				
T-20_{S138G}	1.3 ± 0.5 (0.5)	65 ± 8.8 (27)	141 ± 26 (59)	185 ± 68 (77)
T-20_{S138A}	0.6 ± 0.1 (0.3)	3.6 ± 1.7 (1.5)	3.5 ± 0.9 (1.5)	3.2 ± 1.0 (1.3)
Hydrophobic				
T-20_{S138V}	0.4 ± 0.2 (0.2)	31 ± 14 (13)	22 ± 3.5 (9.2)	23 ± 5.7 (9.6)
T-20_{S138L}	0.7 ± 0.1 (0.3)	13 ± 6 (5.4)	2.9 ± 0.7 (1.2)	2.2 ± 0.4 (0.9)
T-20_{S138I}	0.5 ± 0.1 (0.2)	4.9 ± 2 (2)	2.9 ± 0.8 (1.2)	2.4 ± 0.6 (1)
T-20_{S138M}	0.7 ± 0.2 (0.3)	4.4 ± 0.1 (1.8)	1.7 ± 0.5 (0.7)	1.2 ± 0.4 (0.5)
T-20_{S138P}	446 ± 167 (186)	>1000 (>416)	>1000 (>416)	>1000 (>416)
Nucleophilic				
T-20_{S138T}	0.9 ± 0.2 (0.4)	39 ± 8.5 (16)	161 ± 35 (67)	124 ± 43 (52)
Aromatic				
T-20_{S138F}	9.4 ± 2.6 (4)	203 ± 89 (85)	393 ± 119 (164)	478 ± 116 (200)
T-20_{S138Y}	25 ± 9 (10)	516 ± 223 (215)	>1000 (>416)	>1000 (>416)
T-20_{S138W}	29 ± 14 (12)	>1000 (>416)	>1000 (>416)	>1000 (>416)
Amide				
T-20_{S138N}	19 ± 4 (8)	>1000 (>416)	>1000 (>416)	>1000 (>416)
T-20_{S138Q}	34 ± 11 (14)	>1000 (>416)	>1000 (>416)	>1000 (>416)
Acidic				
T-20_{S138D}	210 ± 94 (88)	>1000 (>416)	>1000 (>416)	>1000 (>416)
T-20_{S138E}	283 ± 80 (118)	>1000 (>416)	>1000 (>416)	>1000 (>416)
Basic				
T-20_{S138H}	210 ± 85 (88)	>1000 (>416)	>1000 (>416)	>1000 (>416)
T-20_{S138K}	708 ± 145 (295)	>1000 (>416)	>1000 (>416)	>1000 (>416)
T-20_{S138R}	362 ± 114 (150)	>1000 (>416)	>1000 (>416)	>1000 (>416)

[a] To improve the replication kinetics, D36G mutation, observed in the majority of HIV-1 strains, was introduced into the NL4-3 background used in this study (reference virus).

nates of the water molecules were removed. Additionally, the hydrogen atoms were placed in optimal positions and refined by the energy minimization with the AMBER9 program (24) using the FF99 force field. Ser-138 in the gp41 core model was replaced with alanine (replacement of -OH with -H), and the positions of the hydrogen atoms were refined as described above. The S138A mutant core model (N36/$C34_{S138A}$ complex) was further optimized by the energy minimization using the FF99 force field with the restraints on each of the three residues of N and C termini and the backbone atoms. The restraint weight was 5.0 kcal/mol Å^2.

RESULTS

Effect of Amino Acid Substitutions at 138 on Antiviral Activities—We chemically synthesized peptide analogs of T-20 with all natural amino acid substitutions at the 138 position (T-20_{S138X}) and evaluated them for their ability to inhibit three major T-20-resistant clones using the MAGI assay (13) (Table 1). The results indicated that only T-20_{S138A} inhibited replication of T-20-resistant clones as efficiently as the wild-type clone. Substitution to glycine enhanced T-20 activity, but unlike T-20_{S138A}, T-20_{S138G} reduced its activity against T-20-resistant clones by ~2–3-fold as compared with the parental peptide, T-20. Substitutions to hydrophobic amino acids leucine, isoleucine, and methionine maintained their anti-HIV-1 activity; however, those to valine reduced anti-HIV-1 activity to T-20-resistant clones. The proline substitution drastically decreased the anti-HIV-1 activity of the peptide inhibitors.

Nucleophilic amino acid at position 138 of T-20 (T-20_{S138T}) showed similar profiles. Conversely, aromatic and amide substitutions reduced the anti-HIV-1 activity of T-20 against HIV-1_{WT}- and T-20-resistant clones. Other amino acid substitutions, especially acidic and basic amino acids, decreased the anti-HIV-1 inhibitory activity even against HIV-1_{WT}. These results suggest that smaller hydrophobic (Ala > Leu, Ile) or more flexible (Met > Thr) residues are preferred in this position. Furthermore, the α-helical structure is important for the interaction, as a mutation to proline which is expected to disrupt the helix (25) resulted in an inactive T-20 analog.

Circular Dichroism—To clarify the mechanism by which the substitutions at Ser-138 influence the antiviral activity of T-20 derivatives, we examined the binding affinities of these peptides to N-HR using circular dichroism (CD) analysis (Fig. 2). CD spectra reveal the presence of stable α-helical structure of the 6-helix bundle that is a requisite for biological activity and is thought to be mechanistically and thermodynamically correlated with HIV-1 fusion (26). Therefore, CD spectra typically at 222 nm indicate interaction of N-HR (N36) and C-HR (T-20 or C34). Because T-20 does not interact significantly *in vitro* with the N36 peptide, which is derived from amino acids 35–70 of N-HR, we used a derivative of C34, a peptide that overlaps with T-20 and also inhibits HIV fusion by the same mechanism. The C34 derivative contained the analogous T-20 substitutions described above (Fig. 1*B*). Consistent with antiviral activities, a mixture of N36 and $C34_{S138P}$ or $C34_{S138W}$ showed no apparent or reduced α-helicity, respectively. For binding with $N36_{V38A}$

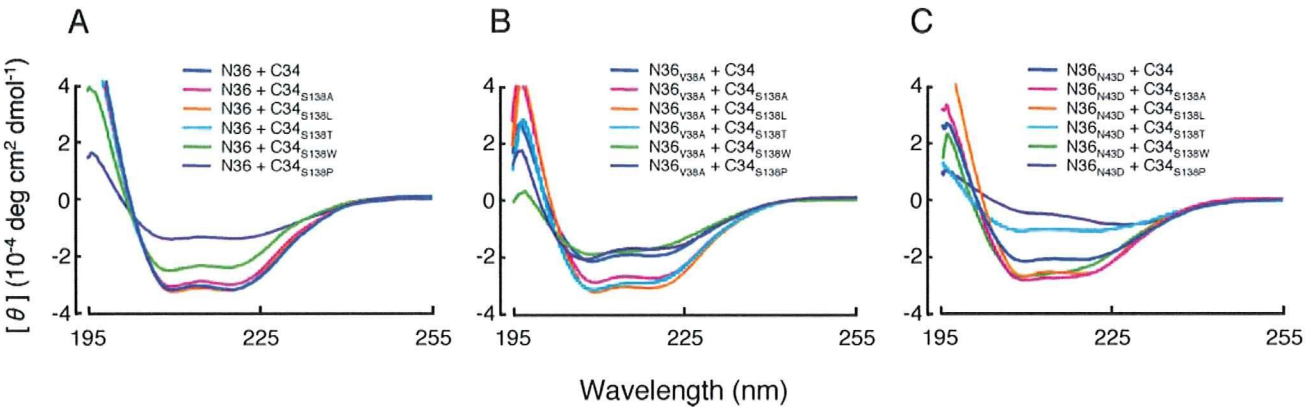

FIGURE 2. **CD spectra of $C34_{S138X}$ complexes with N36 (*A*), $N36_{V38A}$ (*B*), and $N36_{N43D}$ (*C*) are shown.** Equimolar amounts (10 μM) of the N- and C-HR peptides were incubated at 37 °C for 30 min in phosphate-buffered saline. The CD spectra of each mixture were then collected at 25 °C using a Jasco (Model J-710) spectropolarimeter.

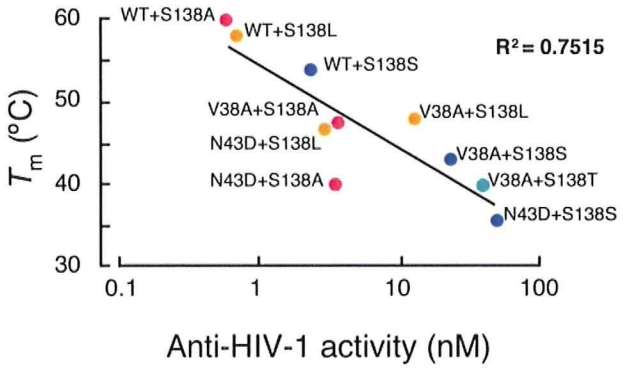

FIGURE 3. **Correlation of T_m values of complexes formed from N36 and C34 peptides (Fig. 2) and anti-HIV-1 activities of T-20_{S138X} (Table 1).**

or $N36_{N43D}$, sufficient α-helicity at 25 °C was observed only in $C34_{S138A}$, $C34_{S138L}$, and $C34_{S138T}$ or $C34_{S138A}$, $C34_{S138L}$, and $C34_{S138W}$, respectively (Fig. 2, *A–C*).

To determine the thermal stability of the helical complexes formed from the N36 and C34 peptides, we measured the melting temperature (T_m) of each complex (supplemental Table 1). The sigmoidal transition of the CD signal at 222 nm correlates with the thermal stability of the helical complexes formed from the N36 and C34 peptides, which in turn are indicative of the binding affinity of these peptides. The melting temperature (T_m) indicating the 50% disruption of 6-helix bundle was comparatively evaluated. Complexes of N36 and C34 containing the S138A or S138L substitutions (N36/$C34_{S138A}$ or N36/$C34_{S138L}$) showed high thermal stability, comparable with that of the wild-type N36/C34 complex. Similarly, the addition of the S138A or S138L also improved the thermal stability of the $N36_{N43D}$/C34 complex. These results reveal a striking correlation between the thermal stability and the anti-HIV-1 activity of the complexes (R^2 = 0.75, Fig. 3). The low T_m value of the complex formed from $N36_{N43D}$ and C34 suggests that virus containing the N43D mutation shows high resistance to T-20, likely due to less favorable thermodynamics that are expected to drive the formation of the 6-helix bundles containing T-20 inhibitor.

Antiviral Activity of Substituted C34 at Ser-138—To confirm that binding of C34 to N-HR is indeed representative of T-20 binding to N-HR, we examined the anti-HIV-1 activities of

TABLE 2

Antiviral activity of $C34_{N126K}$ peptides against C34-resistant gp41 recombinant viruses

Anti-HIV activity was determined by the MAGI assay. The data shown are the mean values and S.D. that were obtained from the results of at least three independent experiments. Shown in parentheses are the -fold increases in resistance (increase in EC_{50} value) calculated by comparison to a reference virus. The increase of >10-fold is indicated in bold.

	EC_{50}	
	HIV-1_{WT} [a]	HIV-$1_{\Delta V4/I37K/N126K/L204I}$ [b]
	nM	
C34	1.6 ± 0.35	114 ± 29 (**71**)
$C34_{N126K}$	0.95 ± 0.22 (0.6)	1.1 ± 0.5 (0.7)

[a] To improve the replication kinetics, the D36G mutation, observed in majority of HIV-1 strains, was introduced into the NL4-3 background used in this study (reference virus).

[b] C34-resistant HIV-1 was constructed with the reference virus as described (13). ΔV4 indicates 5 amino acids deletion (FNSTW) in the V4 region of gp120.

C34-derived peptides that have S138A substitutions. The $C34_{S138A}$ and $C34_{S138L}$ peptides showed potent anti-HIV-1 activities, similar to T-20_{S138A} and T-20_{S138L} (supplemental Table 2). Based on these findings, we conclude that the stability of complexes comprised of modified C34s and N36s containing T-20 resistance mutations offers a good measure of the binding affinity of T-20_{S138X} to N-HR.

Antiviral Activity of C34 with N126K—We have recently identified another mutation at the N-HR of gp41 (N126K) during exposure of HIV-1 to C34 *in vitro* (13). The N126K has been occasionally observed after prolonged T-20-containing therapy (10, 15). Here we have confirmed that the $C34_{N126K}$ peptide can also suppress a C34-resistant clone containing several mutations: I37K/N126K/L204I (Table 2). Therefore, peptides designed to have compensatory mutations seem to have potent antiviral activity. However, because residue 126 is located outside the amino acid sequence of T-20 (Fig. 1*B*), we could not examine the effect of N126K substitution on T-20 activity.

Replication Kinetics of Ser-138-substituted HIV-1—To evaluate the effect of Ser-138 substitutions on viral replication, we constructed molecular clones introducing several Ser-138 and determined their replication kinetics by measuring p24 gag antigen production in the culture supernatant. Single nucleotide changes to the TCA codon for Ser-138 may generate 4 amino acid substitutions, Ala, Thr, Leu, Pro, and Trp. As expected, the compensative substitution, S138A, in the T-20

resistance mutation N43D background enhanced replication kinetics of the N43D-containing clone as shown in supplemental Fig. 1. However, in the WT background the S138A appeared to decrease production of p24 as compared with HIV-1_{WT} (Fig. 4). Other substitutions also reduced their replication kinetics. Interestingly, the S138W substitution did not show measurable p24 production. Syncytia induction and single cycle replication kinetics of the Ser-138-substituted HIV-1 were also examined (supplemental Fig. 2). Sizes of syncytia of each virus formed in the MAGI cells (supplemental Fig. 2, *panels A–E*) were associated with p24-normalized single-cycle infectivities (supplemental Fig. 2, *panel F*) and multicycle replication kinetics (Fig. 4). These results suggest that substitutions at Ser-138 are not likely to appear in the absence of T-20 therapy or the emergence of N43D mutation.

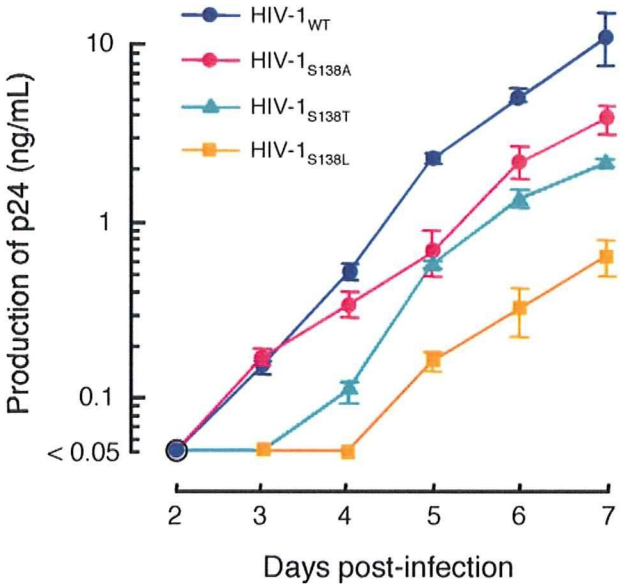

FIGURE 4. **Replication kinetics of HIV-1_{S138X} variants (*X*, any natural amino acid).** HIV-1_{S138A} (*bright red circles*) showed replication kinetics comparable with those seen for HIV-1_{WT} (*blue circles*). Replication of HIV-1_{S138T} (*emerald green triangles*) was reduced, somewhat surprisingly, as both threonine and serine are β-hydroxy amino acids, albeit with different hydrophobicity and torsional flexibility. HIV-1_{S138L} (*orange squares*) also showed reduced replication kinetics. Note that HIV-1_{S138P} and HIV-1_{S138W} failed to replicate (data not shown). Results shown are representative of three independent experiments. An identical order of replication kinetics was observed. Productions of p24 antigen on days 4–7 between HIV-1_{WT} and HIV-1_{S138A} were significant (*t* test, $p < 0.05$).

Structure Modeling—The side chain of amino acid 138 (Ser or Ala) closely contacts with the hydrophobic pocket formed by Leu-44 and Leu-45 in the N-HR. The mutation from Ser to Ala increases hydrophobicity and may help to stabilize the N-HR/C-HR complex related with the potency of the HIV-1 fusion inhibitors (Fig. 5). Larger hydrophobic substitutions such as S138W, S138L, or S138I are likely to sterically interfere with efficient packing of the N-HR and C-HR helices. Similarly, introduction of charged residues at this region of the interface would also disrupt the hydrophobic environment and result in destabilized helix bundles, consistent with the biochemical and virological findings (Figs. 2–4 and Table 1).

Based on crystallographic studies (27, 28), we observe that the T-20 resistance N43D mutation should affect interactions between helices in the 6-helix bundle. Specifically, residue 46 of N-HR is proximal to residue Glu-137 of the C-HR helix of another molecule in the 6-helix bundle. We believe that this increase in proximal negative charges and juxtaposition of Asp-36 next to Glu-137 may destabilize the formation of the 6-helix bundle in a way that results in reduced efficiency of fusion and reduced replication kinetics. Increase of the hydrophobic interactions by introduction of the S138A mutation should help overcome the negative effects of the N43D mutation.

DISCUSSION

In this study we demonstrate that by introducing a secondary resistance mutation into the sequence of peptide-fusion inhibitors such as C34 and T-20, we can suppress efficiently replication of wild-type and of fusion inhibitor-resistant HIV-1. Our circular dichroism analysis revealed that C-HR-based fusion inhibitors that carry secondary resistance mutations can form tight 6-helix bundles with N-HR that contains primary resistance mutations responsible for T-20 resistance. A similar approach has been applied for the development of short hairpin RNA (shRNA) sequences that inhibit HIV-1 replication (29). The synthesized shRNA with mutations that confers resistance to the parental shRNA effectively suppressed replications of shRNA resistant HIV-1 but not wild-type HIV-1. Therefore, it is possible to gain valuable insights from the resistance information and directly apply it to design new peptides or oligonucleotides in the case of shRNA that preempt the viral escape mechanism and suppress resistant variants. Moreover, this strategy should not result in more adverse effect than those that might be obtained during use of the original peptide or oligonucleotide reagents.

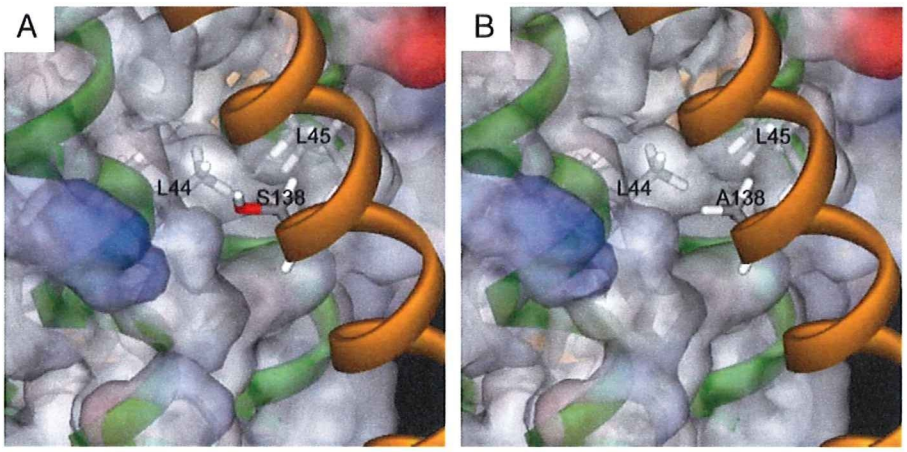

FIGURE 5. **Structure of gp41 at the region near position 138 in the C-HR.** *A*, crystal structure of the N36/C34 complex (PDB code 1AIK). *B*, computational structure modeling of the S138A mutant (N36/$C34_{S138A}$ complex). N-HR and C-HR helices are colored *green* and *orange*, respectively. The van der Waals surface of only N-HR is shown and colored according to the electrostatic potential.

Recently we (6, 30, 31) and others (5) reported that hydrophilic amino

acid substitutions stabilized the α-helix of C-HR peptides and increased their binding affinity to N-HR, thus providing potent anti-HIV activity. This property may be one of the key attributes of the recently developed potent peptide inhibitors, SC34EK (6, 30), T-20EK (31), or T-2429 (5), that have been reported to efficiently inhibit T-20 resistant variants. However, the S138A substitution on T-20 in the present study had little effect on the random coil structure, as judged by CD (data not shown), indicating that $T\text{-}20_{S138A}$ increases its binding affinity not by simply enhancing the α-helicity of this region (5, 6). Our approach of introducing substitutions selected on the basis of the mutation(s) that appears in resistant viruses significantly improved the affinity with N-HR. This approach may complement the effects of enhancing helical stability and may help generate more potent and effective fusion inhibitors for resistant HIV-1 variants.

Other methods have also been employed to improve the potency of HIV fusion inhibitors. For example, T-1249 is a peptide that is based on the T-20 sequence and has improved binding properties (32, 33). It contains 17 changes compared with T-20 (3 additional residues and 14 substitutions to increase the α-helicity/binding affinity according to amino acid sequences of HIV-2 and simian immunodeficiency virus). T-2635 is another efficient peptide fusion inhibitor that was recently developed and is also modified extensively (19 substitutions in 38 amino acids) (5). Also, SC34EK is an electrostatically constrained peptide that also suppresses replication of T-20-resistant variants, and it required 12 substitutions in the original C34 inhibitor (6, 30). Hence, it is possible to improve the potency of existing peptide inhibitors through intense modeling and iterative testing in *in vitro* studies that could lead to the design and synthesis of improved peptide drugs. However, the approach we followed in the design of the $T\text{-}20_{S138A}$ inhibitor is considerably simpler and involves a smaller number of sequence changes (1 residue changed, compared with 19 and 12 in the cases of T-2635 and SC34EK, respectively; see above). It takes advantage of information obtained from the viral evolution under drug pressure and uses the resistance information to design improved inhibitors. In addition, we believe that this approach may be applicable to other targets even when the interactions do not involve helical bundles or detailed information on related systems is not available. Importantly, whenever possible, a combination of the two approaches would likely generate even more effective peptide inhibitors that can suppress replication of resistant variants.

α-Helical structure is a significant factor not only in HIV-1 fusion but also in other examples of protein-protein interactions. Peptide-based drugs have to overcome multiple obstacles, including poor oral bioavailability, less permeability into the target cells, and high cost. Several modifications, such as using arginine-rich peptide tags (34, 35), and chemical treatments (36) have been used to overcome the cell permeability problem. At any rate, peptide-based reagents can be an important tool in the discovery and validation of novel therapeutic targets through *in vitro* experiments. For example, it has been shown that the function of a target protein can be inhibited by designing synthetic peptides that have the amino acid sequence of a domain which is important for the protein function. In such cases the peptides may act as decoys that have antagonistic/agonistic or competitive effects, leading to inhibition of the protein function. Similarly, screening through peptide sequences of proteins may be useful for the identification of functionally important domains that could become future targets for peptide-based or small molecule-based drug development.

In this study we designed peptides tailored to suppress T-20-resistant HIV-1 strains. To our knowledge, this is the first report of direct application of resistance information in drug design and may be applicable to other, unrelated systems. For example, a BH3 domain of the anti-apoptotic protein Bcl-2 has been targeted by an α-helical domain mimic peptide (37, 38). The resulting hydrocarbon-stapled peptide, $SAHB_A$, penetrates into cells via endocytosis pathway and inhibits the function of Bcl-2, inducing apoptosis in transplanted leukemia cells in mice. However, during prolonged therapy with such peptides, leukemic cells could develop resistance to the peptides through substitutions in the Bcl-2 region in the selection process for survival reminiscent of HIV-1. One can envision that our strategy of using mutational resistance information to overcome drug resistance might help in the design of substituted peptides that suppress the resistant variants more efficiently, thus contributing to broader applications of successful peptide-based therapies.

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研究成果の刊行に関する一覧表

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The Novel CXCR4 Antagonist KRH-3955 Is an Orally Bioavailable and Extremely Potent Inhibitor of Human Immunodeficiency Virus Type 1 Infection: Comparative Studies with AMD3100▽

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The previously reported CXCR4 antagonist KRH-1636 was a potent and selective inhibitor of CXCR4-using (X4) human immunodeficiency virus type 1 (HIV-1) but could not be further developed as an anti-HIV-1 agent because of its poor oral bioavailability. Newly developed KRH-3955 is a KRH-1636 derivative that is bioavailable when administered orally with much more potent anti-HIV-1 activity than AMD3100 and KRH-1636. The compound very potently inhibits the replication of X4 HIV-1, including clinical isolates in activated peripheral blood mononuclear cells from different donors. It is also active against recombinant X4 HIV-1 containing resistance mutations in reverse transcriptase and protease and envelope with enfuvirtide resistance mutations. KRH-3955 inhibits both SDF-1α binding to CXCR4 and Ca^{2+} signaling through the receptor. KRH-3955 inhibits the binding of anti-CXCR4 monoclonal antibodies that recognize the first, second, or third extracellular loop of CXCR4. The compound shows an oral bioavailability of 25.6% in rats, and its oral administration blocks X4 HIV-1 replication in the human peripheral blood lymphocyte-severe combined immunodeficiency mouse system. Thus, KRH-3955 is a new promising agent for HIV-1 infection and AIDS.

The chemokine receptors CXCR4 and CCR5 serve as major coreceptors of human immunodeficiency virus type 1 (HIV-1), along with CD4 as a primary receptor for virus entry (2, 15, 18, 19). SDF-1α, which is a ligand for CXCR4, blocks the infection of CXCR4-utilizing X4 HIV-1 strains (7, 34). On the other hand, ligands for CCR5 such as RANTES inhibit CCR5-utilizing R5 HIV-1 (10). These findings made chemokines, chemokine derivatives, or small-molecule inhibitors of chemokine receptors attractive candidates as a new class of anti-HIV-1 agents. Many CCR5 antagonists have been developed as anti-HIV-1 drugs. These include TAK-779 (Takeda Pharmaceutical Company) (5), TAK-652 (6), TAK-220 (45), SCH-C (Schering-Plough) (43), SCH-D (vicriviroc) (42), GW873140 (aplaviroc; Ono Pharmaceutical/Glaxo Smith Kline) (28), and UK-427,857 (maraviroc; Pfizer Inc.) (17). Of these, maraviroc was approved by the U.S. FDA in 2007 for the treatment of R5 HIV-1 in treatment-experienced adult patients, combined with other antiretroviral treatment. Several classes of CXCR4 antagonists have also been reported. The bicyclam AMD3100 showed antivirus activity against many X4 and some R5X4 HIV strains in peripheral blood mononuclear cells (PBMCs) but not against R5 strains (16, 40). The pharmacokinetics and antiviral activity of this compound were also evaluated in humans (21, 22). T22, [Tyr-5,12, Lys-7]polyphemusin II, which is an 18-mer peptide derived from horseshoe crab blood cells, was reported to specifically inhibit X4 HIV-1 strains (30). Studies on the pharmacophore of T140 (a derivative of T22) led to the identification of cyclic pentapeptides (46).

In 2003, we reported that KRH-1636 is a potent and selective CXCR4 antagonist and inhibitor of X4 HIV-1 (23). Although the compound was absorbed efficiently from the rat duodenum, it has poor oral bioavailability. Continuous efforts to find more potent CXCR4 antagonists that are bioavailable when administered orally allowed us to develop KRH-3955 by a combination of chemical modification of the lead compound and biological assays. In this report, we describe the results of a preclinical evaluation of KRH-3955, including its in vitro anti-HIV-1 activity, its in vivo efficacy in the human peripheral blood lymphocyte (hu-PBL)-severe combined immunodeficiency (SCID) mouse model, and its pharmacokinetics in rats in comparison with those of AMD3100.

MATERIALS AND METHODS

Compounds. The synthesis and purification of KRH-3955, *N,N*-dipropyl-*N'*-[4-({[(1*H*-imidazol-2-yl)methyl][(1-methyl-1*H*-imidazol-2-yl)methyl]amino}methyl)benzyl]-*N'*-methylbutane-1,4-diamine tri-(2*R*,3*R*)-tartrate, were carried out by Kureha Corporation. The chemical structure of KRH-3955 is shown in Fig. 1. The CXCR4 antagonist AMD3100 and zidovudine (AZT) were obtained from Sigma. Saquinavir was obtained

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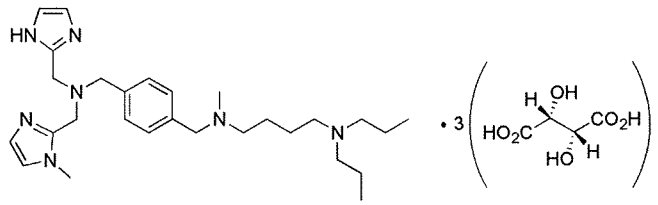

FIG. 1. Chemical structure of KRH-3955.

from the NIH AIDS Research and Reference Reagent Program, NIAID, Bethesda, MD. AMD070 and SCH-D were synthesized at Kureha Corporation.

Cells. Molt-4 no. 8 cells (24) were maintained in RPMI 1640 medium (Invitrogen, Carlsbad, CA) supplemented with 10% fetal bovine serum (Sigma, St. Louis, MO) and antibiotics (50 ng/ml penicillin, 50 ng/ml streptomycin, and 100 ng/ml neomycin; Invitrogen), which is referred to as RPMI medium. Chemokine receptor-expressing human embryonic kidney 293 (HEK293) cells (ATCC CRL-1573) and Chinese hamster ovary (CHO) cells (ATCC CCL-61) were maintained in minimal essential medium or F-12 (Invitrogen) supplemented with 10% fetal bovine serum and antibiotics (50 ng/ml penicillin, 50 ng/ml streptomycin, and 100 ng/ml neomycin). PBMCs from HIV-1-seronegative healthy donors were isolated by Ficoll-Hypaque density gradient (Lymphosepal; IBL, Gunma, Japan) centrifugation (31) and grown in RPMI medium supplemented with recombinant human interleukin-2 (rhIL-2; Roche, Mannheim, Germany) at 50 U/ml.

Viruses. Viral stocks of HIV-1_{NL4-3}, HIV-1_{JR-CSF}, and HIV-$1_{89.6}$ were each produced in the 293T cell line by transfection with HIV-1 molecular clone plasmids pNL4-3 (1), pYK-JRCSF (25), and p89.6 (11), respectively, by the calcium phosphate method. The 50% tissue culture infective dose was determined by an end-point assay with PBMC cultures activated with immobilized anti-CD3 monoclonal antibody (MAb) (33, 51). Subtype B HIV-1 primary isolates 92HT593, 92HT599 (N. Hasley), and 91US005 (B. Hahn) and AZT-resistant HIV-1 (A018) (D. D. Richman) (26) were obtained from the AIDS Research and Reference Reagent Program, Division of AIDS, NIAID, NIH. These clinical isolates were propagated in the activated PBMCs prepared as described above.

Anti-HIV-1 assays. Human PBMCs activated with immobilized anti-CD3 MAb (OKT-3; ATCC, Manassas, VA) in RPMI medium for 3 days were infected with various HIV-1 strains, including primary clinical isolates, at a multiplicity of infection of 0.001. After 3 h of adsorption, the cells were washed and cultured in RPMI medium supplemented with rhIL-2 (50 U/ml) in the presence or absence of the test compounds. Amounts of HIV-1 capsid (p24) antigen produced in the culture supernatants were measured by an enzyme-linked immunosorbent assay kit (ZeptoMetrix Corp., Buffalo, NY) 7 to 10 days after infection. The cytotoxicities of the compounds were tested on the basis of the viability and proliferation of the activated PBMCs, as determined with Cell Proliferation Kit II (XTT) from Roche (36).

Susceptibility of multidrug-resistant HIV-1 to CXCR4 antagonists was also measured by using recombinant viruses in a single replication cycle assay (9, 49). HIV-1 resistance test vectors (RTVs) contain the entire protease (PR) coding region and the reverse transcriptase (RT) coding region, from amino acid 1 to amino acid 305, amplified from patient plasma and a luciferase expression cassette inserted in the *env* region. The RTVs in this study contain patient-derived PR and RT sequences that possess mutations associated with resistance to PR, RT, or both PR and RT. Env-pseudotyped viruses were produced by cotransfecting 293 cells with RTV plasmids and expression vectors encoding the Env protein of well-characterized X4-tropic laboratory strain HXB2, NL4-3, or NL4-3 containing the Q40H enfuvirtide (T20) resistance mutation introduced by site-direct mutagenesis. The virus stocks were harvested 2 days after transfection and used to infect U87 $CD4^+$ cells (kind gifted from N. Landau, NYU School of Medicine) expressing CXCR4 in 96-well plates, with serial dilutions of CXCR4 antagonists. Target cells were lysed, and luciferase activity was measured to assess virus replication in the presence and absence of inhibitors. Drug concentrations required to inhibit virus replication by 50% (IC_{50}) were calculated.

Immunofluorescence. Molt-4 cells or CXCR4-expressing HEK293 cells were treated with various concentrations of KRH-3955 or AMD3100 in RPMI medium or phosphate-buffered saline containing 1% bovine serum albumin and 0.05% NaN_3 (fluorescence-activated cell sorting [FACS] buffer). In washing experiments, cells were washed with RPMI medium or FACS buffer. The cells were Fc blocked with 2 mg/ml normal human immunoglobulin G (IgG) in FACS buffer and then stained directly with mouse MAbs 12G5-phycoerythrin (PE) and 44717-PE (R&D Systems, Inc., Minneapolis, MN) or rat MAb A145-fluorescein isothiocyanate (FITC) and indirectly with MAb A80. The A145 and A80 MAbs were produced in ascitic fluid of BALB/c nude mice, and IgG fractions were obtained from ascitic fluid by gel filtration chromatography with Superdex G200 (Amersham Pharmacia). Goat anti-rat IgG (heavy and light chains) labeled with FITC was purchased from American Corlex (47). After washing, the cells were analyzed on a FACScalibur (BD Biosciences, San Jose, CA) flow cytometer with CellQuest software (BD Biosciences).

DNA construction and transfection. Chemokine receptor-expressing CHO cells were generated as reported previously (23). Human CXCR4 cDNA was cloned into the pcDNA3.1 vector. Mutations were introduced by using the QuikChange II site-directed mutagenesis kit (Stratagene, La Jolla, CA). All constructs were verified by DNA sequencing and transfected into 293 cells by using the Lipofectamine reagent (Invitrogen) (48). Stable transfectants were selected in the presence of 400 μg/ml G418 (Invitrogen). The COOH-terminal intracellular domain of CXCR4 (residues 308 to 352) was deleted in all mutants and the wild type. This deletion has no influence on HIV-1 infection or on SDF-1α binding and signaling but abolishes ligand-induced endocytosis (3).

Ligand-binding assays. Chemokine receptor-expressing CHO cells (5×10^6/0.2 ml per well) were cultured in a 24-well microtiter plate. After 24 h of incubation at 37°C, the culture medium was replaced with binding buffer (RPMI medium supplemented with 0.1% bovine serum albumin). Binding reactions were performed on ice in the presence of ^{125}I-labeled chemokines (final concentration of 100 pmol/liter; PeproTech Inc., Rocky Hill, NJ) and various concentrations of test compounds. After washing away of unbound ligand, cell-associated radioactivity was counted with a scintillation counter as described previously (23).

CXCR4-mediated Ca^{2+} signaling. Fura2-acetoxymethyl ester (Dojindo Laboratories, Kumamoto, Japan)-loaded CXCR4-expressing CHO cells were incubated in the absence or presence of various concentrations of KRH-3955 or AMD3100. Changes in intracellular Ca^{2+} levels in response to SDF-1α (1 μg/ml) were determined by using a fluorescence spectrophotometer as described previously (30).

Detection of KRH-3955 in blood after oral administration. The plasma concentration-time profile of R-176211 (distilled water was used as a vehicle), the free form of KRH-3955, was examined after a single oral administration of KRH-3955 at a dose of 10 mg/kg or intravenous administration at a dose of 10 mg/kg to male Sprague-Dawley rats (CLEA, Kanagawa, Japan). R-176211 in plasma was measured by liquid chromatography-tandem mass spectrometry. Pharmacokinetic parameters were calculated by using WinNonlin Professional (ver. 3.1; Pharsight Co.).

Infection of hu-PBL–SCID mice. Two groups of C.B-17 SCID mice (CLEA, Kanagawa, Japan) were administered a single dose of either KRH-3955 or tartrate (2% glucose solution was used as the vehicle) as a control orally (p.o.) and fed for 2 weeks. These mice were then engrafted with human PBMCs (1×10^7 cells/animal intraperitoneally [i.p.]) and after 1 day were infected i.p. with 1,000 infective units of X4 HIV-1_{NL4-3}. IL-4 (2 μg per animal) was administrated i.p. on days 0 and 1 after PBMC engraftment to enhance X4 HIV-1 infection. After 7 days, human lymphocytes were collected from the peritoneal cavities and spleens of the infected mice and cultured in vitro for 4 days in RPMI medium supplemented with 20 U/ml rhIL-2. HIV-1 infection was monitored by measuring p24 levels in the culture supernatant. We used a selected donor whose PBMCs could be engrafted at an efficiency of >80% in C.B-17 SCID mice. Usually, 5×10^5 to 10×10^5 human $CD4^+$ T cells can be recovered from each hu-PBL–SCID mouse. Mice with no or low recovery of human $CD4^+$ T cells at the time of analysis were omitted. For ex vivo cultures, we used a quarter of the cells recovered from a mouse. The protocols for the care and use of the hu-PBL–SCID mice were approved by the Committee on Animal Research of the University of the Ryukyus before initiation of the present study.

RESULTS

Anti-HIV-1 activities of KRH-3955 in activated PBMCs. The inhibitory activity of KRH-3955 against X4 HIV-1 (NL4-3), R5X4 HIV-1 (89.6), and R5 HIV-1 (JR-CSF) was examined in activated human PBMCs from two different donors. KRH-3955 inhibited the replication of both X4 and R5X4 HIV-1 in activated PBMCs with 50% effective concentrations (EC_{50}) of 0.3 to 1.0 nM but did not affect R5 HIV-1 replication, even at concentration of up to 200 nM (Table 1). In contrast, the CCR5 antagonist SCH-D (vicriviroc) inhibited R5 HIV-1 rep-

TABLE 1. Anti-HIV-1 activity of KRH-3955 in activated PBMCs[a]

Virus	Donor	EC_{50} (nM)[b]					
		KRH-3955	AMD3100	AMD070	SCH-D	AZT	SQV
NL4-3	A	1.1	41	35	>1,000	11	9.0
X4	B	0.33	15	15	>1,000	8.0	29
89.6	A	0.38	44	55	>1,000	7.4	9.9
R5X4	B	ND[c]	ND	ND	ND	ND	ND
JR-CSF	A	>200	>200	>200	0.37	0.96	2.6
R5	B	>200	>200	>200	1.2	6.2	8.0
A018H (X4) (pre-AZT)	C	1.4	38	ND	ND	1.9	ND
A018G (X4) (post-AZT)	C	1.3	32	ND	ND	87,000	ND

[a] PBMCs from two different donors were used in each assay. Anti-HIV-1 activity was determined by measuring the p24 antigen level in culture supernatants.
[b] Assays were carried out in triplicate wells. The average of two to four experiments is shown.
[c] ND, not determined.

lication but inhibited neither X4 nor R5X4 HIV-1 replication (Table 1). The anti-HIV activity of KRH-3955 against the 89.6 virus from donor B was not determined because the virus did not replicate enough for calculation of the anti-HIV activity of KRH-3955 and other drugs. Notably, the anti-HIV-1 activity of KRH-3955 was much higher than that of AMD3100, a well-known X4 HIV-1 inhibitor, or AMD070, the other X4 inhibitor that is bioavailable when administered orally. KRH-3955 also inhibited the replication of clinical isolates of X4 HIV-1 (92HT599) and R5X4 HIV-1 (92HT593) with EC_{50} ranging from 4.0 to 4.2 nM (data not shown). Although both KRH-3955 and AMD3100 were effective against at least some R5X4 HIV-1 strains in activated PBMCs, neither KRH-3955 nor AMD3100 inhibited the infection of CD4/CCR5 cells by R5 or R5X4 HIV-1, even at a concentration of 1,660 nM (data not shown). Importantly, the 50% cytotoxic concentration of KRH-3955 in activated PBMCs (donor A) was 57 μM, giving a high therapeutic index (51,818) in the case of NL4-3 infection, which was higher than that of AZT (8,000 in the case of donor A). These results indicate that the compound is a selective inhibitor of HIV-1 that can utilize CXCR4 as a coreceptor. Since a CXCR4 antagonist should be used in combination with a CCR5 antagonist in a clinical setting, we next examined whether the combined use of both antagonists efficiently blocks mixed infection with X4 and R5 HIV-1. Combination of KRH-3955 and SCH-D at 4 plus 4 nM and 20 plus 20 nM blocked the replication of 50:50 mixtures of NL4-3 and JR-CSF by 91 and 96%, respectively (data not shown). Thus, KRH-3955 is a highly potent and selective inhibitor of X4 HIV-1.

Anti-HIV-1 activities of KRH-3955 in activated PBMCs from different donors. It has been observed that the anti-HIV-1 activity of compounds in PBMCs varies from donor to donor. Therefore, the anti-HIV-1 activity of KRH-3955 against X4 HIV-1 was examined in activated PBMCs from eight different donors. The levels of p24 antigen in NL4-3-infected cultures ranged from 17 to 120 ng/ml (Table 2). KRH-3955 inhibited the replication of NL4-3 with EC_{50} ranging from 0.23 to 1.3 nM and with EC_{90} ranging from 2.7 to 3.5 nM (Table 2), demonstrating that the anti-HIV-1 activity of KRH-3955 was independent of the PBMC donor.

Anti-HIV-1 activities of KRH-3955 against drug-resistant HIV-1 strains. To further assess the efficacy of KRH-3955, we used a single-cycle assay to evaluate the activity of KRH-3955 against a panel of recombinant viruses that express an X4-tropic envelope protein (HXB2) but contain PR and RT sequences containing a wide variety of mutations associated with resistance to PR inhibitors (PIs), nucleoside RT inhibitors (NRTIs), and non-NRTIs (NNRTIs). This assessment was also performed with recombinant viruses that express an X4-tropic envelope protein (NL4-3) that contains the Q40H mutation and displays resistance to T20 (an entry inhibitor). The results of these experiments demonstrate that both KRH-3955 and AMD3100 inhibited the infection of CD4/CXCR4 cells by these recombinant drug-resistant viruses, including viruses resistant to PIs, NRTIs, or NNRTIs; multidrug-resistant viruses; and T20-resistant viruses (Table 3). We also observed that KRH-3955 inhibited the replication of A018G, a highly AZT-resistant strain, in activated PBMCs with an EC_{50} of 1.3 nM (Table 1).

KRH-3955 selectively inhibits ligand binding to CXCR4. To investigate whether KRH-3955 specifically blocks ligand binding to CXCR4, the inhibitory effect of the compound on chemokine binding to CHO cells expressing CXCR4, CXCR1, CCR2b, CCR3, CCR4, or CCR5 was determined. KRH-3955 efficiently inhibited SDF-1α binding to CXCR4 in a dose-dependent manner (Fig. 2 and 3b), and the IC_{50} for SDF-1α binding was 0.61 nM, which is similar to its EC_{50} against HIV-1. Similar results were obtained when we used a Molt-4 T cell line as the CXCR4-expressing target cell (Fig. 3a). Interestingly, the inhibitory activity of AMD3100 against SDF-1α binding was much weaker than its anti-HIV-1 activity (Fig. 3), suggesting that the binding sites of these two compounds are different. In contrast, the compound did not affect the binding

TABLE 2. Anti-HIV-1 activity of KRH-3955 against NL4-3 infection of PBMCs from eight different donors

Donor	p24 level (ng/ml)	EC_{50} (nM)	EC_{90} (nM)
1	31	1.30	3.2
2	25	1.20	3.2
3	17	1.20	3.3
4	40	0.70	2.9
5	120	0.77	2.9
6	58	1.50	3.5
7	49	0.23	2.7
8	53	1.00	3.0
Mean ± SD	49 ± 32	0.99 ± 0.40	3.1 ± 0.30