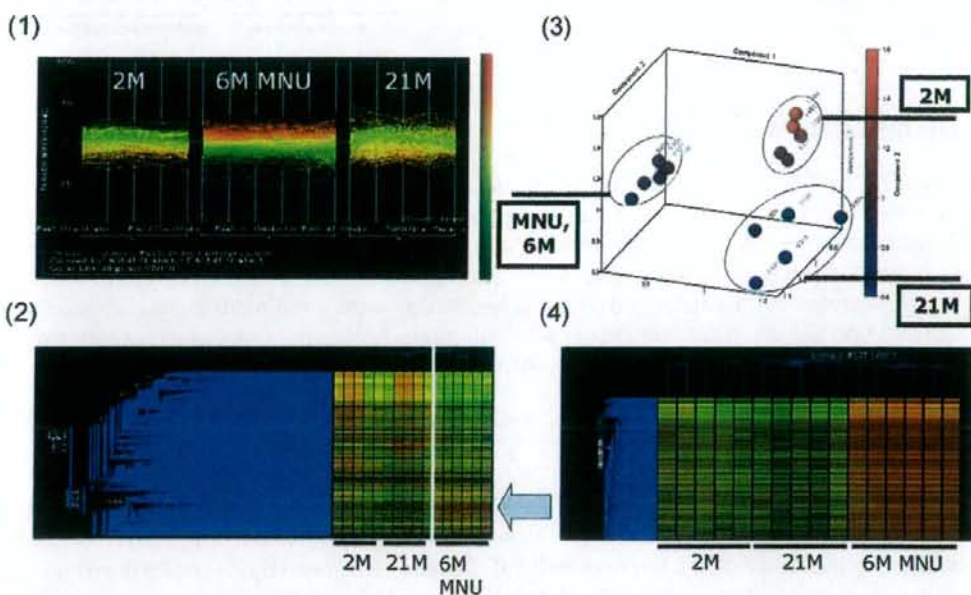


1 The effect of age on experimental animals during toxicological experiments is one of
 2 the interesting factors associated with toxicological endpoints. Two groups of linear con-
 3 figurations represented by five mice each for the 2-month-old and 21-month-old groups are
 4 shown in Figure 3.8a. These two different age groups show different expression diversities;
 5 the aged group (21 months old) shows a much wider range of expression intensities than
 6 the younger (2 months old) group. Furthermore, the order of gene expression intensity
 7 between the two groups is reversed. The wider expression intensities of the aged group
 8 suggest that senescent changes appear stochastically and are probabilistically different
 9 from one individual to another. Note that the wider expression intensities do not indicate
 10 differences in the quality of individual animals but the different probabilistic responses
 11 of individual animals during aging. These individual differences are also observed at the
 12 cellular level, if each cellular function is affected by gene expression (Bahar *et al.*, 2006).



34 **Figure 3.8** (a) Age-related gene expression profiles determined from the bone marrow of
 35 2-month-old and 21-month-old mice are shown in the line configuration. See text. (Six gene
 36 expression profiles of bone marrow obtained from mice 6 months after treatment with a single
 37 dose of methyl-nitroso-urea (MNU) at 50 mg kg⁻¹ b.w.) (b) Two-dimensional dendrographic
 38 diagram of the same expressed genes from the bone marrow of 2-month-old and 21-month-
 39 old mice. (Six gene expression profiles of bone marrow obtained from mice 6 months after
 40 treatment with a single dose of MNU. Arrow indicates the gene cluster specifically up-regulated
 41 in the MNU-treated groups; an expanded view of the profiles is shown in (d).) (c) PCA of
 42 age control group, five mice each for the 2-month-old and 21-month-old groups, is shown in
 43 the three-dimensional contribution scores for components #1, #2 and #3, which discriminate
 44 between the clusters from the 2-month-old and 21-month-old groups. Six gene expression
 45 profiles of bone marrow obtained from mice 6 months after MNU treatment belong to another
 46 separate cluster. (d) Expanded view of profiles indicated by the arrow in (b) showing the gene
 cluster specifically up-regulated in the MNU-treated groups.

1 Such differences between the two age groups cannot be defined using a dendrogram, as
2 observed in the 10 columns on the left of Figure 3.8b, in which the aged group and the
3 young group are not clearly separated. The reason that these different age groups are not
4 clearly separated in the dendrogram is because of the relatively small number of genes
5 with weak expressions that may define these different age groups. To pinpoint the possible
6 responsible genes that define both groups, PCA was applied. Results show that there are
7 discriminant components defined by components #1, #2 and #3, as shown in Figure 3.8c.

8 When one compares the gene expression profiles of these two age groups with those of
9 mice treated with methyl-nitrosourea (MNU), a direct genophilic leukemogenic compound,
10 the expression profiles of six bone marrow tissues from MNU-treated mice are clearly
11 defined in the six columns on the right in Figure 3.8d. The discrimination of the clusters
12 shown in the three-dimensional expressions is clearly separated in Figure 3.8c. The genes
13 responsible for these discriminations can be autogenerated (data not shown).

17 3.8 Radiation-specific Probabilistic Union Genes after Subtracting 18 Age-specific Gene Expressions

19
20 As mentioned previously, regardless of the difference between spontaneous and radiation-
21 induced myelogenous leukemias, it took nearly a lifetime for both leukemias to develop
22 fatally. Furthermore, radiation-induced myelogenous leukemias express stochastically di-
23 vergent profiles, as observed in the line configuration in Figure 3.5. Therefore, each gene
24 expression profile from a total of six radiation-induced myelogenous leukemia cases was
25 analyzed by PCA to elucidate each unique gene list for nontreated mice and compared with
26 the gene expression profiles of bone marrow cells from five 21-month-old nontreated mice.
27 To obtain the union gene list, the gene expression profile of the 21-month-old group was
28 compared separately with each individual expression profile of radiation-induced myel-
29 ogenous leukemias by PCA followed by the selection of genes with a contribution score
30 of over 1.0 from components #2 or #3 and #6 from the result of PCA (Table 3.1). In
31 this PCA, the 287 union genes obtained were subtracted by 45 genes that overlapped with
32 another 249 union genes analyzed by the PCA combination between the profile of the 21-
33 month-old bone marrow group and that of spontaneous myelogenous leukemias (data not
34 shown). Consequently, the final number of union genes obtained was 128 genes after the
35 subtraction of 114 expression sequence tags (ESTs), which were generated in an unsuper-
36 vised manner; however, most of them showed gene functions consistent with the response
37 to radiation exposure. Specifically, five radiation-damage-related genes, including *Hus1*,
38 *Eef1a2*, *Vegfc*, 13 cell cycle/cell-growth-related genes, and 12 apoptosis/cell-death-related
39 genes were observed. Notably, 42 tumorigenesis-related genes (i.e. 33%) were observed
40 in which the down-regulation of tumor suppressor genes and the up-regulation of tumor
41 promoter genes were commonly observed. The expressions of genes for cytoskeleton and
42 cell adhesion molecules and those of genes for oxidative stress and inflammatory cytokines
43 were also observed and included in the list; the expressions of these genes are also con-
44 sidered consequences of the xenobiotic responses to radiation exposure, because these
45 genes were not included in the list of genes associated with spontaneous myelogenous
46 leukemia (data not shown). As a reference to the functions of the autogenerated genes,

Table 3.1 Union gene list for radiation-induced leukemias

Affymetrix systemic name	Common name	Genbank ID	Description
1415874_at	<i>Spry1</i>	NM.011896	sprouty homolog 1 (<i>Drosophila</i>)
1416001_a.at	<i>Cot1</i>	NM.028071	coactosin-like 1 (<i>Dictyostelium</i>)
1417097_at	<i>Nrbf1</i>	NM.025297	nuclear receptor binding factor 1
1417194_at	<i>Sod2</i>	NM.013671	superoxide dismutase 2, mitochondrial
1417602_at	<i>Per2</i>	AF035830	period homolog 2 (<i>Drosophila</i>)
1417623_at	* <i>Slc12a2</i>	BG069505	solute carrier family 12, member 2
1417851_at	<i>Cxcl13</i>	AF030636	chemokine (C-X-C motif) ligand 13
1418062_at	<i>Eef1a2</i>	NM.007906	eukaryotic translation elongation factor 1 alpha 2
1418094_s.at	<i>Car4</i>	NM.007607	carbonic anhydrase 4
1418450_at	<i>Isr</i>	NM.012043	immunoglobulin superfamily containing leucine-rich repeat
1418547_at	<i>Tfpi2</i>	NM.009364	tissue factor pathway inhibitor 2
1418597_at	<i>Top3a</i>	NM.009410	topoisomerase (DNA) III alpha
1418666_at	<i>Ptx3</i>	NM.008987	pentaxin related gene
1418697_at	* <i>Temt</i>	NM.009349	thioether S-methyltransferase
1418712_at	<i>Cdc42ep5</i>	NM.021454	CDC42 effector protein (Rho GTPase binding) 5
1418713_at	<i>Pcbd</i>	NM.025273	6-pyruvoyl-tetrahydropterin synthase/dimerization cofactor of hepatocyte nuclear factor 1 alpha (TCF1)
1418764_a.at	<i>Bpnt1</i>	BB412311	bisphosphate 3'-nucleotidase 1, metal-dependent lithium-inhibited phosphomonoesterase protein family
1419196_at	<i>Hamp1/Hepc</i>	NM.032541	hepcidin antimicrobial peptide
1419353_at	<i>Dpm1</i>	NM.010072	dolichol-phosphate (beta-D) mannosyltransferase 1
1419365_at	<i>Pex11a</i>	NM.011068	adaptor-related protein complex 3, sigma 2 subunit
1419417_at	<i>Vegfc</i>	NM.009506	vascular endothelial growth factor C
1419561_at	* <i>Ccl3</i>	NM.011337	chemokine (C-C motif) ligand 3

Table 3.1 (Continued)

Affymetrix systemic name		Common name	Genbank ID	Description
1419664_at		<i>Srr</i>	BC011164	serine racemase
1419714_at		<i>Pdcd1lg1</i>	NM_021893	programmed cell death 1 ligand 1
1419967_at		<i>Seh1l</i>	AW540070	SEH1-like
1419970_at		<i>Slc35a5</i>	C86506	solute carrier family 35, member A5
1420034_at		<i>Ppp2r2d</i>	AU019644	protein phosphatase 2, regulatory subunit B, delta isoform (AU019644 Mouse eight-cell stage embryo cDNA <i>Mus musculus</i> cDNA clone J0520E06 3-, mRNA sequence)
1420052_x.at	*	<i>Psmb1</i>	C81484	proteasome (prosome, macropain) subunit, beta type 1
1420090_at	*	<i>Raf1</i>	AA990557	v-raf-1 leukemia viral oncogene 1
1420688_a.at		<i>Sgce</i>	NM_011360	sarcoglycan, epsilon
1420843_at	*	<i>Ptprf</i>	BF235516	protein tyrosine phosphatase, receptor type, F
1420872_at	*	<i>Gucy1b3</i>	BF472806	guanylate cyclase 1, soluble, beta 3
1421251_at		<i>Zfp40</i>	NM_009555	zinc finger protein 40
1421462_a.at		<i>Lepre1</i>	NM_019783	leprecan 1
1421619_at	*	<i>Kcnh3</i>	NM_010601	potassium voltage-gated channel, subfamily H (eag-related), member 3
1422025_at	*	<i>Mitf</i>	NM_008601	microphthalmia-associated transcription factor
1422218_at		<i>P2rx7</i>	NM_011027	purinergic receptor P2X, ligand-gated ion channel, 7
1423070_at		<i>Rpl21</i>	BG922742	general transcription factor III A
1423259_at		<i>ldb4</i>	BB121406	inhibitor of DNA binding 4
1423499_at		<i>Sncaip</i>	AK017012	synuclein, alpha interacting protein (synphilin)
1423677_at		<i>Fkbp9</i>	AF279263	FK506 binding protein 9
1424041_s.at		<i>C1s</i>	BC022123	complement component 1, s subcomponent
1424228_at		<i>Polr3h</i>	AK019868	polymerase (RNA) III (DNA directed) polypeptide H
1424295_at		<i>Dppa3</i>	AY082485	<i>Mus musculus</i> stella mRNA, complete cds
1424322_at		<i>Apex2</i>	AB072498	apurinic/aprimidinic endonuclease 2

(Continued)

Table 3.1 (Continued)

Affymetrix systemic name	Common name	Genbank ID	Description
1424586_at	<i>Ehbp1</i>	AF424697	EH domain binding protein 1
1424651_at	<i>BC021611</i>	BC021611	hypothetical protein LOC257633
1424893_at	<i>Ndel1</i>	BC021434	nuclear distribution gene E-like homolog 1 (<i>A. nidulans</i>)
1425198_at	<i>Ptpn2</i>	BG076152	protein tyrosine phosphatase, non-receptor type 2
1425278_at	<i>Ube4a</i>	BC021406	ubiquitination factor E4A, UFD2 homolog (<i>S. cerevisiae</i>)
1425366_a.at	<i>Hus1</i>	AF076845	Hus1 homolog (<i>S. pombe</i>)
1425555_at	* <i>Crk7/Crkr5</i>	BG070845	Cdc2-related kinase, arginine/serine-rich (RIKEN cDNA 1810022J16 gene)
1425597_a.at	<i>Qk</i>	AW060288	quaking
1425608_at	<i>Dusp3/VHR</i>	BC016269	dual specificity phosphatase 3 (vaccinia virus phosphatase VH1-related)
1425750_a.at	<i>Jak3</i>	L40172	Janus kinase 3
1425865_a.at	<i>Lig3</i>	U66057	ligase III, DNA, ATP-dependent
1425918_at	<i>Egln3</i>	BC022961	EGL nine homolog 3 (<i>C. elegans</i>)
1427558_s.at	* <i>Alg12</i>	AJ429133	asparagine-linked glycosylation 12 homolog (yeast, alpha-1,6-mannosyltransferase)
1427595_at	<i>Acac</i>	BE650741	acetyl-Coenzyme A carboxylase alpha
1427833_at	<i>Spi16/mBM17</i>	U96702	serine protease inhibitor 16
1427843_at	<i>Cebpb</i>	AB012278	CCAAT/enhancer binding protein (C/EBP), beta
1428386_at	<i>Acsl3</i>	AK012088	acyl-CoA synthetase long-chain family member 3
1430148_at	<i>Rab19</i>	BM241400	RAB19, member RAS oncogene family
1430391_a.at	<i>Siat8d</i>	AK003690	sialyltransferase 8 (alpha-2,8-sialyltransferase) D
1430483_a.at	<i>Tmem79</i>	AK010144	transmembrane protein 79 (RIKEN cDNA 2310042N02 gene)
1430651_s.at	<i>Zfp191</i>	AI504586	zinc finger protein 191
1431066_at	<i>Fut11</i>	BB626220	fucosyltransferase 11

Table 3.1 (Continued)

Affymetrix systemic name		Common name	Genbank ID	Description
1432072_at	*	<i>Kif2a</i>	AK016720	kinesin family member 2A
1432115_a.at		<i>Pign</i>	AK014165	phosphatidylinositol glycan, class N
1433509_s.at		<i>Reep1</i>	BQ174328	receptor accessory protein 1(D6Ert253e)
1433992_at	*	<i>Apxl</i>	BQ176992	apical protein, <i>Xenopus laevis</i> -like
1434349_at		<i>Vars2l</i>	AV258022	valyl-tRNA synthetase 2-like
1434369_a.at		<i>Cryab</i>	AV016515	crystallin, alpha B
1435132_at		<i>Disp1</i>	A1505698	dispatched homolog 1 (<i>Drosophila</i>)
1435557_at	*	<i>Fhod1</i>	AV298805	formin homology 2 domain containing 1
1435962_at		<i>Rps6</i>	BG089974	ribosomal protein S6 (Transcribed sequence with strong similarity to protein sp:P10660 (<i>H. sapiens</i>) R56_HUMAN 40S ribosomal protein S6)
1436429_at		<i>Zfp606</i>	BB198855	zinc finger protein 606 (BB198855 RIKEN full-length enriched, 0 day neonate thymus <i>Mus musculus</i> cDNA clone A430007N09 3-, mRNA sequence)
1436521_at		<i>Slc36a2</i>	A1596194	solute carrier family 36 (proton/amino acid symporter), member 2
1436623_at		<i>Entpd7</i>	AV381133	ectonucleoside triphosphate diphosphohydrolase 7 (RIKEN cDNA 2900026G05 gene)
1436682_at		<i>Tmsb10</i>	AW259435	thymosin, beta 10 (up29e07.x1 NCI.CGAP_Mam2 <i>Mus musculus</i> cDNA clone IMAGE:2655780 3' similar to gb:S54005 THYMOSIN BETA-10 (HUMAN), mRNA sequence)
1436895_at		<i>Centd1</i>	BB182934	centaurin, delta 1
1436904_at		<i>Thrap1</i>	BB667559	hypothetical protein D030023K18

(Continued)

Table 3.1 (Continued)

Affymetrix systemic name	Common name	Genbank ID	Description
1436993_x.at	<i>Pfn2</i>	BB560492	profilin 2 (BB560492 RIKEN full-length enriched, 10 days neonate olfactory brain <i>Mus musculus</i> cDNA clone E530111B09 3' similar to AL096719 <i>Homo sapiens</i> mRNA; cDNA DKFZp566N043 (from clone DKFZp566N043), mRNA sequence.)
1437059.at	<i>Sox21</i>	BB046776	SRY-box containing gene 21 (BB046776 RIKEN full-length enriched, 11 days embryo <i>Mus musculus</i> cDNA clone 6230417M22 3-, mRNA sequence)
1437106.at	<i>Jarid1a</i>	BM246184	jumonji, AT rich interactive domain 1A (Rbp2 like) (K0734F05-3 NIA Mouse Hematopoietic Stem Cell (Lin-/c-Kit-/Sca-1-) cDNA Library (Long) <i>Mus musculus</i> cDNA clone NIA:K0734F05 IMAGE:30076864 3-, mRNA sequence)
1437123.at	<i>Mmrn2</i>	BB038352	multimerin 2
1437307.at	<i>Senp8</i>	BG069815	SUMO/sentrin specific protease family member 8
1437473.at	<i>Maf</i>	AV284857	avian musculoaponeurotic fibrosarcoma (v-maf) AS42 oncogene homolog (RIKEN cDNA A230108G15 gene)
1437789.at	<i>Birc6</i>	BB527646	baculoviral IAP repeat-containing 6 (BB527646 RIKEN full-length enriched, 15 days embryo head <i>Mus musculus</i> cDNA clone D930041P14 3-, mRNA sequence)
1437863.at	<i>Bche</i>	BB667762	butyrylcholinesterase (BB667762 RIKEN full-length enriched, adult male liver tumor <i>Mus musculus</i> cDNA clone C730038G20 3-, mRNA sequence)

Table 3.1 (Continued)

Affymetrix systemic name	Common name	Genbank ID	Description
1438463_x.at	<i>Zdhhc6</i>	AV142865	AV142865 <i>Mus musculus</i> C57BL/6J 10–11 day embryo <i>Mus musculus</i> cDNA clone 2810427C08, mRNA sequence.
1438825_at	<i>Calm3</i>	AV047570	calmodulin 3 (Similar to calmodulin – rabbit (tentative sequence) (LOC384465), mRNA (AV047570 <i>Mus musculus</i> adult C57BL/6J testis <i>Mus musculus</i> cDNA clone 1700069D17, mRNA sequence))
1438857_x.at	<i>Irak1/pelle-like</i>	BB058253	Irak1 (interleukin-1 receptor-associated kinase 1)/pelle-like (BB058253 RIKEN full-length enriched, 2 days neonate sympathetic ganglion <i>Mus musculus</i> cDNA clone 7120478B17 3- similar to U56773 <i>Mus musculus</i>
1439247_at	<i>Dock10</i>	BB763030	dedicator of cytokinesis 10 (BB763030 RIKEN full-length enriched, B16 F10Y cells <i>Mus musculus</i> cDNA clone G370018M23 3-, mRNA sequence)
1440180_x.at	<i>Zbtb3</i>	AV258279	zinc finger and BTB domain containing 3 (AV258279 RIKEN full-length enriched, adult male testis (BNN132) <i>Mus musculus</i> cDNA clone 4923101A10 3', mRNA sequence)
1440871_at	<i>Baiap1</i>	AI835038	BAI1-associated protein 1
1441272_at	* <i>Matr3</i>	BI249188	matrin 3 (602994742F1 NCI.CGAP_Mam5 <i>Mus musculus</i> cDNA clone IMAGE:5150530 5-, mRNA sequence)
1442100_at	<i>Inpp5f</i>	BB619843	inositol polyphosphate-5-phosphatase F

(Continued)

Table 3.1 (Continued)

Affymetrix systemic name	Common name	Genbank ID	Description
1443229_at	Atad2	AV319821	ATPase family, AAA domain containing 2 (RIKEN cDNA 2610509G12 gene (AV319821 RIKEN full-length enriched mouse cDNA library, C57BL/6J testis male 13 days embryo <i>Mus musculus</i> cDNA clone 6030413117 3-, mRNA))
1443493_at	Dhx37	BB766805	DEAH (Asp-Glu-Ala-His) box polypeptide 37
1443952_at	Nr1d1	BI525006	nuclear receptor subfamily 1, group D, member 1 (602924093F1 NCI.CGAP_Lu33 <i>Mus musculus</i> cDNA clone IMAGE:5056607 5-, mRNA sequence)
1445195_at	C77631	C77631	expressed sequence C77631 (Mouse 3.5-dpc blastocyst cDNA <i>Mus musculus</i> cDNA clone J0035A08 3' similar to Mouse T-cell receptor (TCR V-alpha 16.1) gene exons 1-2, mRNA, mRNA sequence)
1447753_at	Cdc37l	BB391093	cell division cycle 37 homolog (<i>S. cerevisiae</i>)-like (BB391093 RIKEN full-length enriched, 0 day neonate cerebellum <i>Mus musculus</i> cDNA clone C230073C03 3-, mRNA sequence)
1447897_x_at	Anapc11	AV019615	AV019615 <i>Mus musculus</i> 18-day embryo C57BL/6J <i>Mus musculus</i> cDNA clone 1190010L24, mRNA sequence.
1448169_at	Krt1-18	NM_010664	keratin complex 1, acidic, gene 18
1448443_at	Serpini1	NM_009250	serine (or cysteine) proteinase inhibitor, clade I, member 1
1448986_x_at	*	Dnase2a	NM_010062 deoxyribonuclease II alpha

Table 3.1 (Continued)

Affymetrix systemic name	Common name	Genbank ID	Description
1449481_at	Slc25a13	BC016571	solute carrier family 25 (mitochondrial carrier; adenine nucleotide translocator), member 13
1449493_at	InsI5	NM_011831	insulin-like 5
1449700_at	Igbbp1	C81413	immunoglobulin (CD79A) binding protein 1
1449789_x_at	Ly6g6c	AV088850	lymphocyte antigen 6 complex, locus G6C (<i>Mus musculus</i> tongue C57BL/6J adult <i>Mus musculus</i> cDNA clone 2310040E07, mRNA sequence)
1449851_at	* Per1	AF022992	period homolog 1 (<i>Drosophila</i>)
1450046_at	Tmem59/O RF18	NM_019801	transmembrane protein 59 thymic dendritic cell-derived factor 1
1450135_at	Fzd3	AU043193	frizzled homolog 3 (<i>Drosophila</i>)
1450173_at	Ripk2	NM_138952	receptor (TNFRSF)-interacting serine-threonine kinase 2
1450199_a_at	Stab1	NM_138672	stabilin 1
1450208_a_at	Elmo1	NM_080288	engulfment and cell motility 1, ced-12 homolog (<i>C. elegans</i>)
1450296_at	* Klrb1a	NM_010737	killer cell lectin-like receptor subfamily B member 1A
1450297_at	Il6	NM_031168	interleukin 6
1450424_a_at	Il18bp	AF110803	interleukin 18 binding protein
1451541_at	Bcs1l	BC019781	RIKEN cDNA 1700112N14 gene
1451583_a_at	BC025076	BC025076	hypothetical protein LOC216829 membrane magnesium transporter 2
1451592_at	P42pop	AF364868	Myb protein P42POP
1451768_a_at	Slc20a2	AF196476	solute carrier family 20, member 2
1451950_a_at	Cd80	D16220	CD80 antigen
1451996_at	Bbp	AF353993	beta-amyloid binding protein precursor
1452253_at	Crim1	AK018666	cysteine-rich motor neuron 1
1452905_at	Gtl2	AV015833	GTL2, imprinted maternally expressed untranslated mRNA

(Continued)

Table 3.1 (Continued)

Affymetrix systemic name	Common name	Genbank ID	Description
1453055_at	Sema6d	BB462688	sema domain, transmembrane domain (TM), and cytoplasmic domain, (semaphorin) 6D
1453227_at	Rhobtb3	BG801497	Rho-related BTB domain containing 3
1453481_at	Zdhhc2	BB342242	zinc finger, DHHC domain containing 2
1453690_at	* Mpp7	AV292557	membrane protein, palmitoylated 7 (MAGUK p55 subfamily member 7) (RIKEN full-length enriched, 6 days neonate head <i>Mus musculus</i> cDNA clone 5430426E14 3', mRNA sequence)
1454414_at	* Btbd7	AK017755	BTB (POZ) domain containing 7
1455158_at	Itga3	BI664675	integrin alpha 3
1455297_at	SPIN-2	BG070258	Similar to Spindlin-like protein 2 (SPIN-2) (LOC278240), mRNA
1455404_at	Jph2	BG870711	junctionophilin 2
1455717_s.at	* Daam2	BM206030	dishevelled associated activator of morphogenesis 2
1455985_x.at	Shmt2	AV213251	serine hydroxymethyltransferase 2 (mitochondrial) (AV213251 RIKEN full-length enriched, ES cells <i>Mus musculus</i> cDNA clone 2410126G07 3', mRNA sequence)
1456975_at	Taok1	BM238077	TAO kinase 1 (RIKEN cDNA 2810468K05 gene)
1457040_at	Lgi2	BE947711	leucine-rich repeat LGI family, member 2
1457311_at	Camk2a	AW490258	calcium/calmodulin-dependent protein kinase II alpha
1457451_at	Acvr2	BB199213	activin receptor IIA
1458047_at	Tnfsf13b	BB667811	tumor necrosis factor (ligand) superfamily, member 13b
1458381_at	Clic5	BB028501	chloride intracellular channel 5
1458641_at	Braf	BM217816	Braf transforming gene

Table 3.1 (Continued)

Affymetrix systemic name	Common name	Genbank ID	Description
1459597_at	Mtpn	BG074849	myotrophin
1459868_x.at	Il11ra1	AV313111	interleukin 11 receptor, alpha chain 1 (RIKEN full-length enriched, adult male thymus <i>Mus musculus</i> cDNA clone 5830408C01 3' similar to X74953 <i>M. musculus</i> ETL-2 mRNA, mRNA sequence.)
1460170_at	Ext2	NM_010163	exostoses (multiple) 2
1460666_a.at	Ebf3	NM_010096	early B-cell factor 3

*: overlapped in both lists of union genes for radiation-induced and spontaneous leukemias.

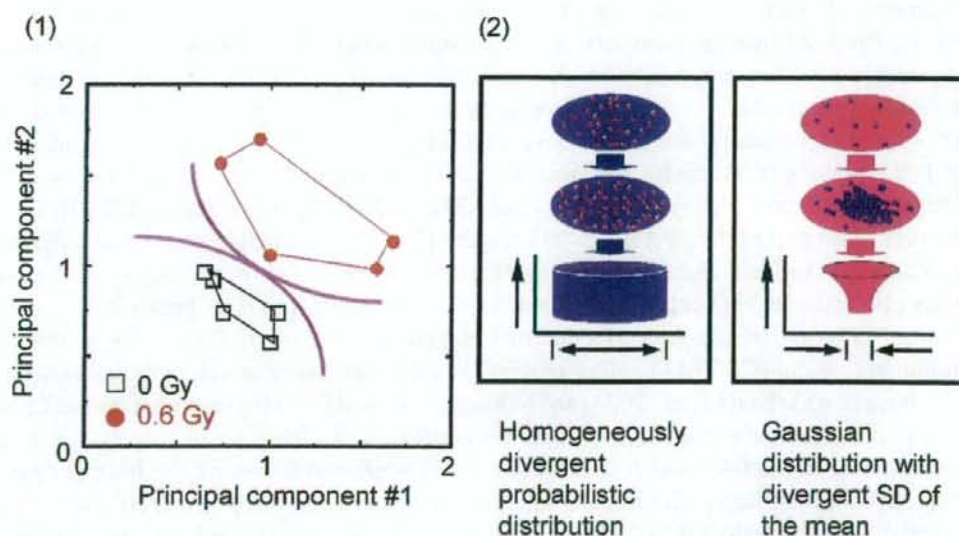
10 genes that represent the 128 genes mentioned above are described as follows: *Eef1a2* (eukaryotic translation elongation factor 1 alpha 2), expressed in tumors of the ovary, breast and lung (Amiri *et al.*, 2007; Tomlinson *et al.*, 2007) and plays a role in the resistance to apoptosis induced by oxidative stress (Chang and Wang, 2007); *Top3a* (topoisomerase (DNA) III alpha), required for accurate DNA replication (Oh *et al.*, 2002) and related to telomere–telomere recombination (Tsai *et al.*, 2006), cancer, and aging (Laursen *et al.*, 2003); *Ppp2r2d* (protein phosphatase 2, regulatory subunit B, delta isoform), expression biomarker for blast crisis (Liu *et al.*, 2007; Neviani *et al.*, 2007); *Leprel* (leprecan 1), basement-membrane-associated proteoglycan that functions in growth suppression and is a potential suppressor gene (Wassenhove-McCarthy and McCarthy, 1999); *Idb4* (inhibitor of DNA binding4), promotes neuronal stem cell proliferation (Yun *et al.*, 2004) and induction of leukemia cell apoptosis (Yu *et al.*, 2005); *Hus1* (hydroxyurea sensitive1), DNA damage checkpoint (Harris *et al.*, 2006), required for telomere maintenance and functions as Rad9–Hus1–Rad1 checkpoint; *Dusp3/VHR* (dual-specificity phosphatase 3 (vaccinia virus phosphatase VH1-related)), induces expression of cyclin D1 in breast cancer (Hao *et al.*, 2007) and arrests the cell cycle in VHR (Rahmouni *et al.*, 2006); *Igbp1/alpha 4* (immunoglobulin (CD79A) binding protein 1), apoptosis inhibitor via dephosphorylation of c-Jun and p53 (Kong *et al.*, 2004) and biomarker for acute myelogenous leukemia (Cruse *et al.*, 2005; Bhargava *et al.*, 2007); *Iga3* (integrin alpha 3), inhibitor of caspase 3 activity (Manohar *et al.*, 2004), which is up-regulated in adenocarcinoma of the lung (Boelens *et al.*, 2007), esophagus (Hourihan *et al.*, 2003) and stomach (Varis *et al.*, 2002); *Il11ra1* (Interleukin 11 receptor alpha 1), functions in carcinogenesis associated with up-regulation of PI3K and p44/p42 MAPK in gastric cancer (Nakayama *et al.*, 2007) and colon cancer (Yoshizaki *et al.*, 2006) associated with STAT3 in the prostate cancer (Zurita *et al.*, 2004), and constitutively activated in myeloma/B-CLL (Tsimanis *et al.*, 2001). The functions of these genes may satisfy the characteristics of cluster-specific gene expression profiles linked to radiation-induced leukemias.

Among the 149 genes, 21 genes, including *raf1*, *Mitf*, and *Crkrs*, overlap in both lists of union genes for both radiation-induced and spontaneous leukemias (see asterisks in

1 Table 3.1). These overlapped genes observed in both union gene lists imply that not all the
 2 radiation-specific union genes (i.e. 'stochastically necessary genes') are always required
 3 for the development of radiation-induced myelogenous leukemias, but a combination of
 4 the 'stochastically necessary genes' in the list is required in addition to the common
 5 leukemogenic genes. An essential rule for the combination of the 'stochastically necessary
 6 genes' for radiation-induced leukemogenicity is not yet identified.

9 3.9 New Risk Evaluation Strategy Using Gene Expression Profiles

11 These toxicological endpoints obtained by gene chip and microarray technologies can be
 12 used to evaluate their quantitative and qualitative differences in gene expression for risk
 13 evaluation by PCA. Figure 3.9a shows sample expression data between the two groups,
 14 one for bone marrow tissues exposed to 0.6 Gy and the other for nonirradiated controls,
 15 which is shown by two-dimensional expressions from PCA components #1 and #2. No
 16 observed effect level (NOEL) or no observed adverse effect level (NOAEL) can be statisti-
 17 cally calculated by the 95 % confidence areas of the two clusters, so that the differences
 18 between the two groups can be considered as the risk evaluation parameters. Furthermore,
 19 when one attempts to evaluate the possible differences between the two groups by gene
 20 chip and microarray methods, one can use any component(s) with low differential contribu-
 21 tion. However, when one attempts to evaluate homogeneity, one should evaluate possible



40 **Figure 3.9** (a) Sample gene expression clusters associated with the bone marrow after 0.6 Gy
 41 whole-body irradiation and those associated with the control are shown in the two-dimensional
 42 principal component diagram plotted along the contribution factors. Areas of each cluster are
 43 statistically defined along with the confidence levels of NOEL and/or NOAEL. See text. (b)
 44 Two types of data distribution pattern: homogeneously divergent probabilistic distribution on
 45 the left and divergent distribution due to Gaussian distribution with the error of the mean on
 46 the right.

consistencies not only for the major component, but also for other components with low contribution, and determine whether the differences can be ignored. Because minor components may sometimes play an important toxicological role, careful examination of the genomic repertoire is required to determine whether the expressions of responsible genes are identical in a case-by-case manner.

Lastly, it is very important to recognize the characteristics of each data point. When one calculates NOEL or NOAEL, one should note the different characteristics and diversity of data between the two types, such as whether the data show probabilistic homogeneously diverse distribution or a Gaussian normal equivalent distribution due to error/deviation, as shown on the left and right sides of Figure 3.9b. Interestingly, data from developmental toxicology and the growth parameters tend to show the latter convergent distribution. However, data from toxicological changes in relation to senescence tend to show the former scattered distribution.

In this chapter, murine spontaneous and radiation-induced myelogenous leukemias were used as experimental models to discuss an essential principle of data-mining in toxicogenomics. The examinations were focused only on the bone marrow. A possible reason for the predictability observed in tissues other than the bone marrow may be attributable to the characteristics of radiation-induced tissue injury, of which the general rule may be stochastic but generally applicable to other tissues. In the case of chemicals, the predictability of the results of a single tissue examination may be limited owing to the possible tissue-specific interactions of such test chemicals.

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Percellome Projectによる毒性トランスクリプトミクスの新しい試み

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Special Review

Percellome Projectによる毒性トランスクリプトミクスの新しい試み

Percellome Project as a New Approach to Toxicology Transcriptomics

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身の回りの物質の毒性(有害性)を予測し、その被害を未然に防ぐのが毒性学の役割である。この精度向上を目指したトキシコゲノミクス研究を実施する際に、マイクロアレイなどから細胞1個当たりのmRNAコピー数を得るPercellome法を開発した。90化合物のマウス肝初期応答データを採取し終え、新たな対象(反復投与、胎児毒性、吸入毒性、多臓器連携)を加えたPercellome Projectを展開している。

key words

トキシコゲノミクス, 分子毒性学, 遺伝子発現カスケード, 標準化, Percellome法, 3次元多層(Millefeuille)データ

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はじめに

医薬品, 食品, 化粧品, 生活関連用品など, 身の回りの物質が我々の身体に取り込まれた際に生じる可能性のある毒性(有害性)を予測し, それらの使用に際しての被害を未然に防ぐのが毒性学の役割である¹⁾(図1)。具体的には, 人々の安全を確保するために使用法(用途)や使用量(残留量)を制限したり, 場合によっては禁止したりするための科学的根拠を提供するが, その際, 人の身代わりとして実験動物を用いる場合が多い。このような毒性学の精度向上の一環として, 従来からの毒性研究(毒性症候学, 毒性病理学, など)に加えてのトキシコゲノミクス(Toxicogenomics)研究が進められている。

トキシコゲノミクスでは, 物質が生体に及ぼす影響をトランスクリプトームとして観測・解析する。その際, ①分子毒性学を構築し種差や個体差の問題, 複合暴露の問題などを解決するためには, 遺伝子発現カスケードの全容解明を目指す必要がある, ②形態学的に変化が現れた段階のトランスクリプトームは, 遺伝子発現カスケードの最終段階に過ぎない, ③形態変化の現れないごく初期段階を含む遺伝子発現カスケードを描出するためにははまとまった量のデータの蓄積が必須である。との観点から, 筆者らは, マイクロアレイや定量PCRから細胞1個当たりのmRNAコピー数を得るPercellome手法と, そのデータ解析のための3次元多

層(Millefeuille)システムを開発・実用化した。遺伝子発現量が共通の尺度, すなわち“コピー数/細胞”で表現されることから, 検体間, 実験間, マイクロアレイのバージョン間, 異なったプラットフォーム間, などのデータ比較が直接的に行えるようになり, 数年かけて蓄積したデータの有機的活用が可能となった。現在, 90種類の化学物質によるマウス肝の初期応答データを採取し終えたところである。新たな対象(反復投与, 胎児毒性, 吸入毒性, 多臓器連携)を加えたPercellome Projectの概要を紹介する。

I. Percellome法: 細胞1個当たりのmRNA絶対量を得る方法

原理は単純である。サンプルの細胞数を計測し, 外部標準mRNA(スパイクRNA)を細胞1個当たり決まった分子数だけそのサンプルに添加し, そしてRNA抽出, 測定に移る。サンプルのRNAの測定値を, スパイクRNAの値を基準に, 細胞1個当たりのコピー数に換算する。実際には細胞数を直接計測するのが困難なことが多いため, その代替指標として細胞核内のゲノムDNA量を用いる^{1), 2)}。定量性・直線性の検証にはLBM標準サンプル(肝[L]と脳[B]を100:0, 75:25, 50:50, 25:75および0:100に混合した5サンプルから成るセット)を用いる。なお, スパイクRNAは, 5種類の枯草菌遺伝子のmRNAを濃度公比3で混合したカクテル(dose-graded spike cocktail; GSC)として用意した。高精度を要求されるDNA定量法は手作業プロトコルおよび自動ロボット(PerkinElmer JANUS)のプロトコルを準備

注1 環境への配慮も含まれる。



図1. 毒性学の対象

毒性学は、身の回りの物質が引き起こす障害を予測し、その発生を未然に防ぐことを目的としている。トキシコゲノミクス（毒性ゲノミクス）は、最先端の網羅的遺伝子発現解析技術を用いて、従来の毒性学の予測の精度を著しく向上、迅速化させることで、国民の健康安全の確保にさらに貢献することを目指している。

中である。カクテルとも共同研究ベースで供給可能である（連絡先：kanno@nihs.go.jp）。また、ERCC（The External RNA Control Consortium）と連絡をとるとともに、国際的標準化への関与を深めるため平成18年度厚労科研費「医薬品などの有効性・安全性評価に資する遺伝子発現解析の国際的標準化に関わる研究（H18-特別-指定-023）」を立ち上げた。現在、この他にシックハウス症候群を考慮した低用量域での吸入毒性トキシコゲノミクス、1匹のマウスから多臓器を採取しそれらの連携状況をトランスクリプトームから解析する多臓器トキシコゲノミクスを開始し、特徴的な遺伝子について組織内の発現分布を *in situ* ハイブリダイゼーションで確認する作業を並行している。また、下記の3次元データをweb公開するサーバを整備し、一部の化合物から3次元多層（Millefeuille）データを順次閲覧可能とした（<http://toxicomics.nihs.go.jp/db/>）。

II. 3次元多層（Millefeuille）データシステム：生物系研究者に優しいデータ可視化と解析

医薬品を含む毒性既知の90化合物について単回経口投与後のトランスクリプトームデータを取得して、初期応答遺伝子カスケードを解析するための基盤データベースを構築した。現在、第二段階として反復暴露データ集積を開始し

た。データは、用量軸、時間軸、および遺伝子発現軸から成る3次元表示により、遺伝子発現の用量および時間に依存した変化を1枚の曲面として表すことで可視的に変化を判別しやすいように配慮した（図2）。これにより、コンピュータが選び出した遺伝子クラスターの中身を確認する際、特に、mRNAの合成分解のスピードなどの知見から生物学的にありえないパターン（用量軸の方向にも時間軸の方向にもジグザグな変化など）を排除する際に威力を発揮している。

1つの実験から排出されるGeneChip約50枚のデータを一括処理する能力を持ったPerceLLome自動換算・データ品質管理（QC）に関わるソフトウェアに加えて、3次元多層（Millefeuille）データに最適化した、発現パターン類似性による候補遺伝子検索、およびそれを発展させた教師無しクラスタリング³⁾を中心とした解析システム（MF System, MFシリーズ、開発：相崎 健一）を独自に実用化し、開発継続中である（図3）。これらにより、データQCはその日のうちに、基本的な発現情報検索から全遺伝子の教師無しクラスタリングまでを3日間で完遂できるものとなっている。

この基本解析を用いて、発現パターンによって分類された候補遺伝子リストが多数生成される。一部の幸運な例ではただちに新規と思われる毒性関連反応を見いだすことができた。またそうでない場合のための1つの補強手段とし