There was no difference in mRNA expression of the HSL gene in WAT and BAT between the groups (Fig. 3). On the other hand, mRNA expression of the HSL gene in the liver was low in the OVX group and in groups after treatment with a low dose of isoflavone, compared with the sham-operated group. A high dose of isoflavone tended to maintain the mRNA expression of the HSL gene in the OVX rats at the same level as that in the sham-operated rats.

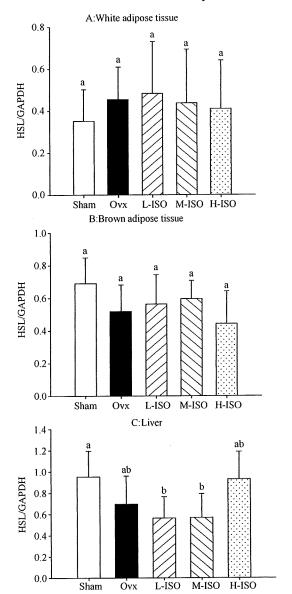


FIG. 3. Effect of isoflavone on HSL mRNA expression in white adipose tissue (A), brown adipose tissue (B), and liver (C). Animal groups comprised the sham-operated controls (Sham), ovariectomized controls (OVX), and OVX rats treated with different doses of isoflavone (L-ISO, M-ISO, and H-ISO). The means with the different letters differ (P<0.05).

There were no differences in mRNA expression of PPAR α in WAT and liver (Fig. 4). In contrast, the expression of PPAR α in BAT of rats treated with a high dose of isoflavones was significantly higher than those in the other groups.

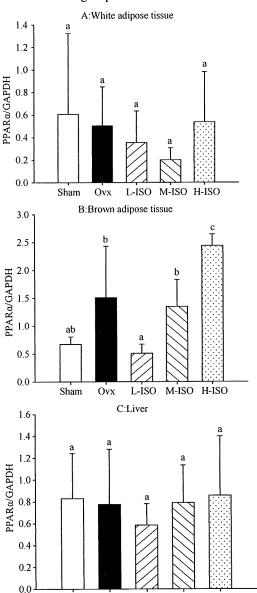


FIG. 4. Effect of isoflavone on PPARα mRNA expression in white adipose tissue (A), brown adipose tissue (B), and liver (C). Animal groups comprised sham-operated controls (Sham), ovariectomized controls (OVX), ovariectomized rats treated with different doses of isoflavone (L-1SO, M-ISO, and H-ISO). The means with the different letters differ (P<0.05).

L-ISO

M-ISO

Ovx

The expression of PPARγ in WAT, BAT, and liver in the OVX group tended to be higher than that in the

sham-operated group (Fig. 5). Isoflavone slightly suppressed the up-regulation of PPARy mRNA

expression. However, this effect was not statistically significant.

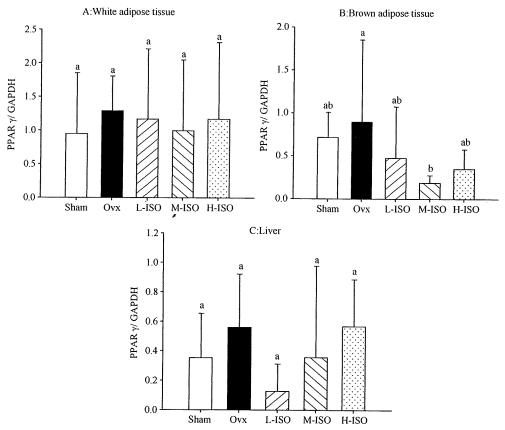


Fig. 5. Effect of isoflavone on PPARγ mRNA expression in white adipose tissue (A), brown adipose tissue (B), and liver (C). Animal groups comprised the sham-operated controls (Sham), ovariectomized controls (OVX), OVX rats treated with different doses of isoflavone (L-ISO, M-ISO, and H-ISO).

DISCUSSION

The relationship between dietary isoflavones, body weight, and fat mass was explored in this study. Isoflavones affected body and fat weights in OVX rats fed on a high-fat diet. Adding different concentrations of isoflavone in the diet (0.68-2.00 g/kg diet) significantly decreased the body weight. Furthermore, the weight of adipose tissue was also significantly lower in the isoflavone group than in the OVX group (P<0.05). Although no significant effect of isoflavone on food intake was observed, food efficiency was significantly lower in OVX rats treated with isoflavone than in OVX rats not treated with isoflavone (P < 0.05). indicating that food consumption does not reflect the change in the amount of adipose tissue. However, isoflavone inhibited food utilization and adipose deposition. Naaz et al.[10] and Kim et al.[11] reported that the soy isoflavone genistein decreases body weight and adipose deposition in OVX mice. Ali et al.[12] reported that isoflavones significantly reduces

weight gain and fat deposition in obese and lean male rats. Another research also demonstrated that soy isoflavone prevents body fat accretion and bone loss in OVX mice^[13]. Epidemiological surveys and nutrition intervention studies indicated that genistein has beneficial effects on obesity in human beings^[14-15], suggesting that an isoflavone-rich diet improves lipid metabolism and has anti-obesity properties.

Serum total cholesterol and adipocytokines, such as leptin and adiponectin, were significantly higher in the OVX rats than those in the sham-operated rats (Table 3). Leptin affects energy homeostasis by inhibiting food intake and stimulating energy expenditure. However, obesity is generally accompanied by a high serum leptin concentration, which is termed as leptin resistance. Isoflavone treatment reduced serum total cholesterol and leptin, suggesting that isoflavones prevents body fat gain, in part, by inhibiting leptin resistance in OVX rats. D'Eon et al. [16] reported that 17β-estradiol reduces the serum levels of leptin and adiponectin in OVX mice,

suggesting that the effects of isoflavones on lipid metabolism in animals after OVX animals might, in part, be exhibited *via* an estrogenic pathway.

Despite the beneficial effects of dietary isoflavone on metabolic abnormalities and obesity, studies on the effect of isoflavone on the expression of lipid metabolism-related genes in OVX rats are limited. Therefore, we performed RT-PCR to reveal the possible mechanism underlying the effects of isoflavone on lipid metabolism in OVX rats with diet-induced obesity.

FAS is one of the primary lipogenic enzymes and plays a central role in lipogenesis of mammals and converts dietary calories into a stored form of energy. The activity of FAS has a positive correlation with body fat mass. It was reported that inhibition of the FAS gene expression can sharply decrease the body fat in mice^[17]. In this study, the change in FAS gene expression in liver and adipose tissues varied. FAS mRNA expression in the liver was higher in OVX rats than in sham-operated rats, and was significantly inhibited after treatment with isoflavone (P<0.05), demonstrating that estrogen deficiency induces body weight gain with fatty acid synthesis in the liver and these changes were improved after treatment with isoflavone. On the other hand, mRNA expression of the FAS gene in WAT and BAT of the OVX group was significantly lower than that in the sham-operated group (P<0.05), which is consistent with the previously reported data in OVX mice^[1]. It appears that down-regulation of FAS may occur in adipose tissue of rodents after OVX. Further study is needed to clarify this mechanism.

HSL is an intracellular, neutral lipase, and the rate-limiting enzyme in lipolysis^[18]. HSL is expressed in various tissues and plays an important role in lipid metabolism, including neutral cholesteryl hydrolase^[19-20]. Palin *et al.*^[21] showed that 17β-estradiol at high doses up-regulates HSL expression. However, a low dose of 17β-estradiol does not significantly alter the HSL expression in subcutaneous abdominal adipocytes isolated from a woman. Although Banz et al. [5] reported that soy protein enhances the expression of HSL mRNA in the liver of obese male Zucker diabetic fatty (ZDF) rats. There is no report concerning the direct influence of isoflavone on HSL-gene expression in OVX rats. In this study, the expression of HSL mRNA was lower in the liver of OVX rats treated with a low dose of isoflavones than in sham-operated rats. However, the gene expression tended to restore to the level in the sham-operated group following treatment with a high dose of isoflavones, which is consistent with the previously reported data^[21], suggesting that the decreased lipolysis in liver is, in part, responsible for the body weight gain in OVX rats, and isoflavone intake inhibits this change.

PPARs belong to the nuclear steroid receptor super-family. PPARa is mainly expressed in the liver, heart, skeletal muscles, kidney, and intestine. PPARy is mainly expressed whereas PPAR-delta (PPAR8 is widely distributed in adipose tissue and the immune system^[22]. Of the three PPAR isoforms known to date, PPARα and PPARy play an important role in the regulation of whole-body and cellular metabolism^[23]. The activation of PPARα increases fatty acid β-oxidation and insulin sensitization, whereas blood and liver lipid concentrations are typically reduced. Ovariectomy decreases expression of the PPARB gene, and estrogen treatment can stimulate the expression of this gene^[24]. On the other hand, PPARB controls the expression of genes involved in adipocyte differentiation and lipogenesis. The increased adipocyte differentiation attributing to PPARy activation often results in weight gain because of increased body fat; resulting in the paradoxical effect of PPARy agonist treatment i.e., increased fat mass along with improvement in other metabolic lipid variables.

It has been demonstrated that isoflavone-containing soy extracts or soy isoflavone can activate PPAR-mediated gene expression and induce metabolic changes suggestive of a PPAR agonist-like effect of soy isoflavone^[25]. Similarly, Sujong *et al.*^[4] reported that genistein up-regulates PPAR α and PPAR γ gene expression in the liver of C57BL/6J mice fed on a high-fat diet. It has been shown that soflavone enhances PPAR α and PPAR γ mRNA expression in WAT of rats^[26].

Expressions of the PPAR α and PPAR γ gene in WAT, BAT, and liver of OVX mice were examined in this study. The expression of PPAR α in liver and WAT tended to be lower in the OVX rats treated with a low dose of isoflavone than in sham-operated rats, whereas treatment of OVX rats with a high dose of isoflavone tended to up-regulate the gene expression (Fig. 4). Meanwhile the expression of PPAR γ in liver and BAT in OVX rats tended to be higher than that in sham-operated rats, and treatment with isoflavone improved these changes. However, dose response was not observed (Fig. 5), suggesting that OVX increases lipogenesis while decreases lipolysis, and isoflavone improves lipid metabolism in the liver of OVX rats.

In summary, isoflavone is a type of phytoestrogen that can inhibit body weight gain and obesity induced by OVX with a high-fat diet by decreasing food utilization and modulating the gene expression related to lipid metabolism. These effects of isoflavones may involve not only the reduction of the expression of lipogenic genes related to fatty acid synthesis and adipocyte differentiation but also increase the expression of lipolysis-related genes in the liver of OVX rats.

364

ACKNOWLEDGEMENTS

The authors are grateful to Dr. Jian WU (Nisshin OilliO Group Co. Ltd.) for helpful discussion. We also thank Dr. Kazuhiko Yamada, Dr. Miki Miyoshi, and Dr. Nobuo Yoshiike (National Institute of Health and Nutrition) for arrangement of the research program.

REFERENCES

- Kamei Y, Suzuki M, Miyazaki H, et al. (2005). Ovariectomy in mice decreases lipid metabolism-related gene expression in adipose tissue and skeletal muscle with increased body fat. J Nutr Sci Vitaminol (Tokyo) 51, 110-117.
- Genazzani A R, Gambacciani M (2006). Effect of climacteric transition and hormone replacement therapy on body weight and body fat distribution. Gymecol Endocrinol 22, 145-150.
- 3. Dixon R A (2004). Phytoestrogens. Annu Rev Plant Biol 55, 225-261.
- Kim S, Sohn I, Lee Y S, et al. (2005). Hepatic gene expression profiles are altered by genistein supplementation in mice with diet-induced obesity. J Nutr 135, 33-41.
- Banz W J, Davis J, Peterson R, et al. (2004). Gene expression and adiposity are modified by soy protein in male Zucker diabetic fatty rats. Obesity Res 12, 1907-1913.
- Bhathena S J, Velasquez M T (2002). Beneficial role of dietary phytoestrogens in obesity and diabetes. Am J Clin Nutr 76, 1191-1201.
- Goodman-Gruen D, Kritz-Silverstein D (2001). Usual dietary isoflavone intake is associated with cardiovascular disease risk factors in postmenopausal women. J Nutr 131, 1202–1206.
- Goodman-Gruen D, Kritz-Silverstein D (2003). Usual dietary isoflavone intake and body composition in postmenopausal women. *Menopause* 10, 427-432.
- Reeves P G, Nielsen F H, Fahey G C (1993). AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition adhoc writing committee on the reformulation of the AIN-76A rodent diet. J Nutr 123, 1939-1951.
- 10.Naaz A, Yellayi S, Zakroczymski M A, et al. (2003). The soy isoflavone geniatein decreases adipose deposition in mice. *Endocrinology* 144, 3315-3320.
- 11.Kim H K, Nelson-Dooley C, Della-Fera M A, et al. (2006). Genistein decreases food intake, body weight, and fat pad weight and causes adipose tissue apoptosis in ovariectomized female mice. J Nutr 136, 409-414.
- 12. Ali A A, Velasquez M T, Hansen C T, et al. (2004). Effects of soybean isoflavones, probiotics, and their interactions on lipid

- metabolism and endocrine system in an animal model of obesity and diabetes. *J Nutr Biochem* **15**, 583-590.
- 13. Wu J, Wang X, Chiba H, et al. (2004). Combined intervention of soy isoflavone and moderate exercise prevents body fat elevation and bone loss in ovariectomized mice. Metabolism 53, 942-948.
- 14. Yamori Y (2004). Worldwide epidemic of obesity: hope for Japanese diets. Clin Exp Pharmacol Physiol 31, S2-4.
- 15. Mori M, Aizawa T, Tokoro M, et al. (2004) Soy isoflavone tablets reduce osteoporosis risk factors and obesity in middle-aged Japanese women. Clin Exp Pharmacol Physiol 31, S39-41.
- 16.D'Eon T M, Souza S C, Aronovitz M, et al. (2005). Estrogen regulation of adiposity and fuel partitioning. J Biol Chem 280, 35983-35991.
- 17.McInnes K J, Corbould A, Simpson E R, et al. (2006). Regulation of adenosine 5', monophosphate-activated protein kinase and lipogenesis by androgens contributes to visceral obesity in an estrogen-deficient state. Endocrinology 147, 5907-5913.
- Kraemer F B, Shen W J (2006). Hormone-sensitive lipase knockouts. Nutr Metab (Lond) 3, 12.
- Yeaman S J (2004). Hormone-sensitive lipase new roles for an old enzyme. Biochem J 379, 11-22.
- Kraemer F B, Shen W J (2002). Hormone-sensitive lipase: control of intracellular tri-(di-)acylglycerol and cholesteryl ester hydrolysis. J Lipid Res 43, 1585-1594.
- 21. Palin S L, McTernan P G, Anderson L A, et al. (2003). 17Beta-estradiol and anti-estrogen ICI:compound 182,780 regulate expression of lipoprotein lipase and hormone-sensitive lipase in isolated subcutaneous abdominal adipocytes. Metabolism 52, 383-388.
- 22. Braissant O, Foufelle F, Scotto C (1996). Differential expression of peroxisome proliferator-activated receptors (PPARs): tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat. *Endocrinology* 137, 354-366.
- 23. Schoonjans K, Staels B, Auwerx J (1996). The peroxisome proliferator activated receptors (PPARs) and their effects on lipid metabolism and adipocyte differentiation. *Biochim Biophys Acta* 1302, 93-109.
- 24. Campbell S E, Mehan K A, Tunstall R J (2003). 17β-Estradiol upregulates the expression of peroxisome proliferator-activated receptor-alpha and lipid oxidative genes in skeletal muscle. J Mol Endocrinol 31, 37-45.
- 25. Mezei O, Banz W J, Steger R W, et al. (2003). Soy isoflavones exert antidiabetic and hypolipidemic effects through the PPAR pathways in obese Zucker rats and murine RAW 264.7 cells. J Nutr 133, 1238-1243.
- 26. Kawakami Y, Tsurugasaki W, Nakamura S, et al. (2005). Comparison of regulative functions between dietary soy isoflavones aglycone and glucoside on lipid metabolism in rats fed cholesterol. J Nutr Bio 16, 205-212.

(Received November 7, 2007 Accepted June 19, 2008)

