

Fig. 1. Typical time series of intracarotid sinus pressure (CSP), muscle tension, sympathetic nerve activity [SNA; in arbitrary units (au)], and arterial pressure (AP) under control (left) and muscle stretch (right) conditions. CSP was perturbed according to a binary white noise sequence. Muscle stretch increased mean levels of SNA and AP under muscle stretch conditions compared with the control conditions.

the frequency of input modulation increased under both conditions, indicating derivative characteristics of the neural arc. Muscle stretch caused an approximately parallel upward shift of the gain plot. The phase approached  $-\pi$  radians  $(-180^\circ)$  at the lowest frequency (0.01~Hz) under both conditions, reflecting the negative feedback character of the baroreflex neural arc (i.e., an increase in CSP decreased SNA). Phase plots were nearly superimposed between the two conditions. Coherence

Table 1. Mean levels and CVs of CSP, SNA, and AP at 1, 2, 4, and 6 min under control and muscle stretch conditions

	Time			
	I min	2 min	4 min	6 min
		CSP		
Control CV Muscle stretch CV	95±18 13±2 114±15* 11±2	96±18 12±3 115±16* 11±1	95±18 11±3 113±16* 10±2	96±18 14±2 114±15* 12±2
		SNA		
Control CV Muscle stretch CV	102±4 46±11 133±22* 48±11	99±5 45±9 129±21* 47±8	100±4 43±9 127±17* 44±9	99±4 47±9 126±17* 49±10
		AP		
Control CV Muscle stretch	90±21 7±2 107±26* 7±3	89±20 6±2 105±22* 6±3	88±16 6±2 104±15* 6±3	88±18 6±2 101±15* 7±2

Values are means ± SD; n = 7. CSP, carotid sinus pressure (in mmHg); SNA, sympathetic nerve activity (in %); AP, arterial pressure (in mmHg); CV, coefficient of variation. Mean and CV values were calculated from 30-s data ending at each time point. \*P < 0.05 vs. control.

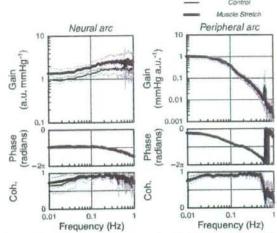


Fig. 2. Transfer functions of the neural (left) and peripheral (right) arcs under control and muscle stretch conditions. In the neural arc, the input was CSP and the output was SNA. In the peripheral arc, the input was SNA and the output was AP. The mean level of CSP input to the neural arc was set higher under muscle stretch conditions than under control conditions to mimie the physiological condition (i.e., baroreflex closed-loop conditions). Gain plots (top), phase plots (middle) and ocherence (Coh) functions (bottom) are shown. Thin and thick solid lines indicate control and muscle stretch conditions, respectively. In the neural arc (left), muscle stretch caused an approximately parallel upward shift of the gain plot. Solid and dashed lines represent means and means ± SD values, respectively.

values did not differ between both conditions. In the peripheral arc, the dynamic gain decreased in the frequency range from 0.05 to 1 Hz as the frequency of input modulation increased under both conditions, indicating the low-pass characteristics of the peripheral arc. The phase approached 0 radians at the lowest frequency (0.01 Hz) under both conditions, reflecting the fact that an increase in SNA increased AP. The phase lagged with increasing frequency up to 1 Hz. The gain plot, phase plot, and Coh(f) did not differ between both conditions.

Table 2 summarizes gains of the transfer functions. In the neural arc,  $G_{0.01}$ ,  $G_{0.1}$ ,  $G_{0.5}$ , and  $G_1$  were higher under muscle

Table 2. Gains of the transfer functions

	Control	Muscle Stretch
Neural arc	C Accel Historia	CONTRACTOR
Gunt, au/mmHg	$1.01 \pm 0.23$	1.44±0.56*
$G_{n+1}$ au/mmHg	$1.30 \pm 0.11$	1.86±0.37*
Gn.5, au/mmHg	$1.77 \pm 0.64$	2.65±1.08*
G <sub>1</sub> , au/mmHg	1.72±0.66	2.72±1.40°
Peripheral arc		
Gnot, mmHg/au	$1.08 \pm 0.06$	$1.06 \pm 0.20$
Go i, mmHg/au	$0.37 \pm 0.09$	$0.42 \pm 0.09$
Go.5, mmHg/au	$0.02 \pm 0.01$	$0.02 \pm 0.01$
G <sub>1</sub> , mmHg/au	$0.004 \pm 0.001$	$0.004 \pm 0.002$
Total loop		
Goot, mmHg/mmHg	$1.08 \pm 0.18$	1.53±0.63*
Gn.t. mmHg/mmHg	$0.48 \pm 0.12$	$0.81\pm0.31$ *
Gu.s. mmHg/mmHg	$0.04 \pm 0.04$	$0.06 \pm 0.04 *$
G <sub>1</sub> , mmHg/mmHg	$0.006 \pm 0.003$	$0.013 \pm 0.013$

Values are means  $\pm$  SD; n = 7.  $G_{0.01}$ ,  $G_{0.1}$ ,  $G_{0.5}$ , and  $G_{1}$ , dynamic gains at 0.01, 0.1, 0.5, and 1 Hz, respectively; au, arbitrary units. \*P < 0.05 vs. control.

stretch compared with control conditions. In the peripheral arc,  $G_{0.01}$ ,  $G_{0.1}$ ,  $G_{0.5}$ , and  $G_{1}$  were unchanged between control and muscle stretch conditions.

Figure 3 shows the total baroreflex loop transfer functions (CSP to AP) under control and muscle stretch conditions. The thin and thick solid lines in Fig. 3 indicate control and muscle stretch conditions, respectively. The dynamic gain decreased as the frequency of input modulation increased under both conditions, indicating low-pass characteristics. The dynamic gain under muscle stretch conditions was higher than that under control conditions in frequency from 0.01 to 0.5 Hz (Table 2). The phase plot and Coh(f) did not differ between both conditions.

Figure 4 shows step responses of SNA corresponding to the transfer functions in the neural arc shown in Fig. 2. The initial drop in the SNA response as well as the steady-state response was augmented during muscle stretch (Table 3). T<sub>peak</sub> did not differ between control and muscle stretch conditions (Table 3).

#### DISCUSSION

The key new findings of the present study are as follows. Muscle stretch increased the dynamic gain of the carotid sinus baroreflex neural arc as estimated by binary white noise input (Fig. 2). In contrast, the peripheral arc transfer function remained unchanged irrespective of the muscle stretch (Fig. 2). These results suggest that during muscle mechanoreflex activation, the dynamic SNA response to CSP perturbation is augmented.

System identification by the white noise approach. To identify the dynamic characteristics of arterial baroreflex function quantitatively, we described the carotid sinus baroreflex con-

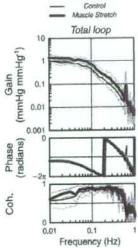


Fig. 3. Total loop transfer functions from CSP to AP under control and muscle stretch conditions. Gain plots (top), phase plots (middle) and coherence functions (bottom) are shown. Thin and thick solid lines indicate control and muscle stretch conditions, respectively. The dynamic gain decreased as the frequency of input modulation increased under both conditions, indicating low-pass characteristics. Muscle stretch caused an approximately parallel upward shift of the gain plot. Solid and dashed lines represent means and means ± SD values, respectively.

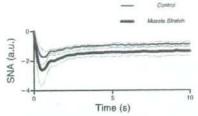


Fig. 4. Step responses corresponding to transfer functions of the neural arc obtained from Fig. 2, showing the SNA response to a 1-mmHg increase in input pressure. Thin and thick solid lines indicate control and muscle stretch conditions, respectively. The initial drop in the SNA response as well as the steady-state response was augmented by the muscle stretch. Solid and dashed lines represent means and means  $\pm$  SD values, respectively.

trol of SNA and AP in terms of system identification using the white noise technique. Compared with the traditional approach of testing dynamic properties of the physiological system with step and sine wave stimuli, the white noise approach has definite advantages, as follows (27). First, if a step stimulus is applied, we learn the response of the system to this step and have little notion of the response of the system to any other type of stimulus. If a sinusoidal pulse is applied, then we know the response of the system to such a stimulus and little else. The same applies for any other specific waveform. Theoretically speaking, the system is tested with every possible stimulus in the white noise approach. The white noise stimulus is a very rich stimulus. It should be emphasized that the white noise method is perfectly suited to the analysis of linear systems. As shown in Figs. 2 and 3, high coherence values close to unity indicate the validity of our method for system identification. Second, the identification of the physiological system through the white noise technique is largely unaffected by the types of contaminating noise usually present in such a system. Our study provides the first and quantitative description of the dynamic characteristics of the carotid sinus baroreflex during isolated activation of mechanosensitive afferents from skeletal muscle.

Effects of the muscle mechanoreflex on dynamic characteristics of the carotid sinus baroreflex. The effects of activation of afferents from skeletal muscle, such as those occurring during exercise, on the arterial baroreflex have been extensively studied (5, 13, 29, 42, 43, 49, 58, 59). These studies have demonstrated that the afferent input from muscle resets the baroreflex control of AP, heart rate, and SNA. However, the dynamic characteristics of the arterial baroreflex during isolated activation of muscle mechanosensitive afferents have never been analyzed. In the present study, muscle stretch increased dynamic gain in every frequency (Fig. 2 and Table

Table 3. Parameters of step responses

	Control	Muscle Stretch
S <sub>50</sub> , au	-1.05±0.30	-1.69±0.69*
S <sub>peak</sub> , au	-2.10±0.50	-3.08±1.45*
T <sub>peak</sub> , s	0.63±0.21	0.64±0.20

Values are means  $\pm$  SD; n=7. A step response is defined as a SNA response to a 1-mmHg change in input pressure.  $S_{50}$ , step response at 50 s;  $S_{peak}$ , negative peak response;  $T_{peak}$ , time to negative peak. \*P < 0.05 vs. control.

2), whereas it did not affect the peripheral arc. These data are the first to provide quantitative evidence demonstrating that the dynamic SNA response to CSP perturbation is augmented during isolated activation of the muscle mechanoreflex. Although an increase in dynamic gain in the lowest frequency (0.01 Hz) was expected from the results of our previous studies showing an increase in static gain by muscle stretch (58, 59), the information was insufficient to perform a simulation study to examine the effects of muscle stretch on the closed-loop dynamic AP regulation (see *Physiological implications*). The present study extended our previous work by providing additional information on the dynamic interaction over a wide range of frequencies between 0.01 and 1 Hz in the carotid sinus haroreflex.

The static characteristics of the arterial baroreflex determine an operating point of the baroreflex system. Furthermore, the static characteristics described by a modeled sigmoid function provide the parameters of threshold, saturation, and maximal gain at the centering point. However, the static characteristics alone cannot provide the information on the changes over time in the response of the baroreflex system. On the other hand, dynamic analysis techniques such as transfer function analysis estimated by the white noise approach provide information on the stability and quickness of the system response. The dynamic SNA response to baroreceptor pressure input became greater as the frequency of input modulation increased, suggesting derivative characteristics (i.e., high-pass characteristics) of the baroreflex neural arc (Fig. 2, left, thin solid line). In contrast, the dynamic AP response to SNA became smaller as the frequency of SNA modulation increased, indicating lowpass characteristics of the baroreflex peripheral arc (Fig. 2, right, thin solid line). The total loop transfer function (CSP to AP) is determined by a product of the neural and peripheral arc transfer functions (Fig. 3, thin solid line). Therefore, the decreasing slope of dynamic gain in the total loop transfer function was shallower than that in the corresponding peripheral arc. In other words, the fast neural arc effectively compensates for the slow peripheral arc to accelerate dynamic AP regulation by the baroreflex negative-feedback loop (14). During muscle stretch, the dynamic gain in the neural arc was increased by ~50% in every frequency under study (Fig. 2 and Table 2), indicating that the derivative characteristics of the neural arc were maintained. As a result, the effect of the neural are compensating for the slow AP response was preserved during the activation of muscle mechanoreflex (Fig. 3 and Table 2). Furthermore, the total loop dynamic gain was augmented during the muscle stretch due to the upward shift of the neural arc transfer function.

Because we used passive muscle stretch as the input for the muscle mechanoreflex, the physiological significance of the present results should be interpreted carefully. Several studies have examined the arterial baroreflex control of SNA during static and dynamic exercise. Static and heavy dynamic exercise resets the baroreflex control of SNA to higher SNA levels with an increase in its sensitivity (9, 11, 17, 32). On the other hand, mild to moderate dynamic exercise resets the baroreflex control of SNA without any change in its sensitivity (3, 24, 38). Because the muscle mechanoreflex is activated during mild to moderate dynamic exercise (4), our results indicate that the muscle mechanoreflex may contribute to increasing the baroreflex gain of SNA during mild to moderate dynamic exercise. In

addition to differences in the measured SNA (renal vs. muscle), analytic methods of baroreflex function, modes of mechanoreflex activation, and/or species between the present study and previous studies, the cardiopulmonary baroreflex should be taken into account. Charkoudian et al. (1) demonstrated that increasing central venous pressure via head-down tilt or saline infusion attenuated the baroreflex sensitivity in the control of SNA. The activation of cardiopulmonary baroreceptors induced by increasing central venous pressure may influence the arterial baroreflex control during dynamic exercise (37). In the present study, however, the cardiopulmonary baroreflex did not operate due to bilateral vagotomy.

Previous studies (7, 25) have suggested that the muscle mechanoreflex has a dominant role in pressor reflexes during muscle contraction in anesthetized or decerebrate cats. Although we believe that the mechanoreflex is one of the pressor reflexes during exercise, the functional importance of the muscle mechanoreflex in cardiovascular regulation during exercise in conscious conditions is debatable. Matsukawa et al. (28) recently reported that blockade of the muscle mechanoreflex by gadolinium did not alter AP responses to isometric exercise in conscious cats. Moreover, they found that gadolinium significantly diminished the pressor responses to passive muscle stretch in anesthetized cats. These observations suggest that, under the experimental design, the muscle mechanoreflex would not be activated during exercise or, even if it was activated, it has no functional importance in cardiovascular responses to exercise in conscious conditions. One criticism for the study is that there is always a possibility that changes in the central command in conscious conditions had compensated for the lack of muscle mechanoreflex. Further studies are needed to better understand the role of the muscle mechanoreflex on neural cardiovascular responses during exercise.

High-pass characteristics of the baroreflex neural arc. It is likely that the dynamic characteristics of the baroreflex neural arc actually reflect the intrinsic and synaptic properties of central nervous system neurons and neural circuits that transmit baroreceptor input. However, the central baroreceptor synapses are characterized as a low-pass filter (26). The difference between high-pass characteristics of the neural arc transfer gain and low-pass characteristics of the central baroreceptor synaptic transmission could be attributable to the difference of estimated frequency ranges. Frequency-dependent depression (FFD) of synaptic transmission in the baroreflex central pathways is the phenomenon that the probability of excitatory postsynaptic potentials progressively reduces as the frequency of afferent input increases beyond 1 Hz (2, 33). Although FDD and transfer gain should be discriminated in theory, interactions between FDD and transfer gain may occur when the modulation frequency of afferent fiber stimulation approached the frequency range of FDD. Indeed, Kawada et al. (23) found high-cut characteristics of the baroreflex neural arc in the frequency range above ~1 Hz. In the present study, the transfer gain was derived from 0.01 to 1 Hz. Whether the dynamic interaction between carotid sinus baroreflex and muscle mechanoreflex exists in the frequency range beyond 1 Hz awaits further studies.

Part of the high-pass characteristics in the baroreflex neural arc is attributable to the derivative nature observed in the baroreceptor transduction from CSP input to baroreceptor afferent nerve activity (i.e., mechanoneural transduction) (21). However, we think there exists high-pass characteristics in the transduction from baroreceptor afferent input to efferent SNA, because the magnitude of high-pass characteristics slightly differs between cardiac and renal SNAs in response to the same baroreceptor pressure perturbation (18).

In an electrical circuit, we can design a high-pass filter only from low-pass filter elements using a feedback loop (Fig. 5). Although the main forward path of the baroreflex neural arc from afferent nerve activity to efferent SNA is considered to be the nucleus tructus solitarius, caudal ventrolateral medulla, and rostral ventrolateral medulla (53), there could be feedback connections between these areas. Therefore, it is possible that synaptic connection has basically low-pass characteristics, whereas the baroreflex neural arc reveals high-pass characteristics as a neural circuit. The speculation also needs to be

verified experimentally in the future.

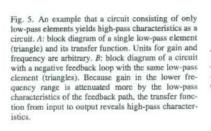
Physiological implications. Under physiological conditions, the baroreflex is closed as a negative feedback system. In the following discussion, we will focus on the effect of the augmentation of dynamic SNA modulation in the neural arc on the closed-loop dynamic AP regulation. Figure 6A illustrates a simulator consisting of the linear neural arc transfer function (HN) and linear peripheral arc transfer function (HP) followed by the nonlinear sigmoidal components (see the APPENDIX for details). A closed-loop AP response to a stepwise pressure perturbation (-40 mmHg) with pulsatile pressure was simulated, and the result is shown in Fig. 6B. Muscle stretch shortened the time to 95% of steady state by ~33% from 7.2 to 4.8 s (shaded and solid arrows in Fig. 6B). This result suggests that, under baroreflex closed-loop conditions, the rate of recovery in AP following a pressure perturbation occurs sooner when accompanied by the muscle mechanoreflex. Increasing the quickness of the negative-feedback system can be caused by augmentation and/or acceleration of the open-loop transfer function of the system. In our baroreflex open-loop experiment, S50 and Speak in the step responses of SNA were

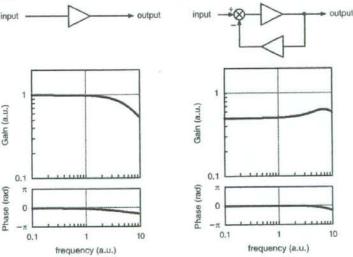
A

augmented by the muscle stretch (Fig. 4 and Table 3). On the other hand,  $T_{\rm peak}$  did not differ between control and muscle stretch conditions (Fig. 4 and Table 3). These results suggest that the improvement in the quickness of the AP restoration via the baroreflex observed in the closed-loop simulation was induced by augmentation, rather than acceleration, of the dynamic SNA response in the neural arc. However, further experimental studies are needed to verify the simulation model.

Limitations. The present study has several limitations. First, we performed the experiment in anesthetized animals. Previous studies have suggested that any anesthetic could alter the baroreflex regulation in AP (54–56). The gain of the baroreflex is reported in the conscious state to be higher (~2-fold) than in the anesthetized state. A previous study (52) suggested that α-chloralose anesthesia could alter the dynamic characteristics of the baroreflex regulation around the frequency of 5 Hz. However, the anesthesia was convenient for the elimination of the central command. Furthermore, we compared the baroreflex gain between muscle stretch and nonstretch conditions both under anesthesia. Therefore, a reasonable interpretation would be that the increased baroreflex gain is attributable to muscle stretch in this experiment.

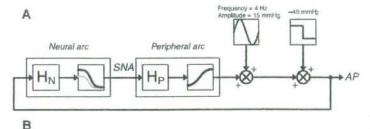
Second, stretching of skeletal muscle provides a stimulus for the activation of mechanoreceptors that is different from that which occurs during muscle contraction. During contraction, mechanoreceptors are activated by a shortening of skeletal muscle and by compression of the receptors. Thus, mechanoreceptors may be stimulated in a very different manner during stretch, which would likely affect the magnitude of the corresponding reflex response. In addition, the level of muscle stretch used in our experiment was relatively high (50). The stretch may activate different afferents than contraction (8). Furthermore, the discharge profile of mechanosensitive afferents adapt during static muscle stretch (31). Accordingly, during the muscle stretch for 6 min in the present study, the firing level from the mechanoreceptors might have been





B

AJP-Heart Circ Physiol • VOL 295 • SEPTEMBER 2008 • www.ajpheart.org



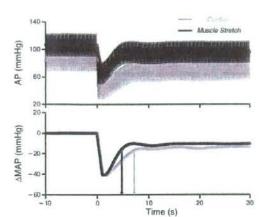


Fig. 6. A: simulator of the baroreflex system during activation of the muscle mechanoreflex. A stepwise perturbation with pulsatile pressure was applied to the baroreflex negative feedback system (see the ΑΡΡΕΝDΙΧ for details). H<sub>N</sub>, neural arc transfer function, H<sub>P</sub>, peripheral arc transfer function. B: simulation results of the closed-loop AP response to the stepwise pressure perturbation (-40 mmHg). Muscle stretch shortened the time to 95% of steady state by -33% (shaded and solid arrows). Shaded and solid thick lines indicate mean AP (MAP) resampled at 1 Hz. ΔMAP, change in MAP from baseline.

steadily diminishing. In fact, the increase in SNA and AP induced by muscle stretch gradually decreased from 90 s to 6 min after the initiation of the muscle stretch, which was used for data analysis (Table 1). However, SNA and AP remained significantly higher under muscle stretch conditions than control conditions over the protocol for 6 min. Thus, we believe that the mechanoreflex remained activated in this protocol. Further studies are required to elucidate the dynamic interactions between baroreflex and mechanoreflex induced by different modes of activation, such as cyclic activation of the mechanoreflex.

Third, the transfer function analysis is useful in identifying the linear input-output relationship of the baroreflex at a given operating point. However, the transfer function cannot characterize the nonlinear input-output relationship of the system. In the presence of nonlinear system behavior such as the baroreflex system, the transfer function analysis is partly compromised, indicating that the absolute output values of the nonlinear system to given input signals cannot be predicted accurately by the transfer function alone. Combining a linear transfer function with a nonlinear sigmoidal element would increase the accuracy to reproduce dynamic characteristics observed in the baroreflex neural arc (20, 22).

Finally, we measured renal SNA as a proxy of systemic sympathetic activity. SNAs to different organs may vary a lot. Although static and dynamic regulations of the barore-flex neural arc are similar among renal, cardiac, and muscle SNAs (15, 16, 18), whether this holds true during muscle stretch remains to be verified. Also, subsystems of the peripheral arc transfer function such as those relating car-

diac output and peripheral vascular resistance remain to be identified.

Conclusions. In conclusion, baroreflex open-loop transfer function analysis demonstrated that the activation of mechanosensitive afferents from skeletal muscles augmented the dynamic SNA response in the neural arc. This augmentation of the dynamic SNA response with maintained derivative characteristics of the neural arc may accelerate closed-loop AP regulation via the baroreflex.

#### APPENDIX

To simulate the closed-loop AP response to stepwise pressure perturbation (Fig. 6), we used the derivative-sigmoidal cascade model. The cascade model consists of a linear derivative filter followed by a nonlinear sigmoidal component (20, 22).

We modeled the sigmoidal nonlinearity in the baroreflex neural arc interacting with the muscle mechanoreflex by the following fourparameter logistic function with threshold according to a previous study (59):

$$y = \max \left\{ \frac{P_1}{1 + \exp[P_2(x - P_3)]} + P_4, \text{ Th} \right\}$$
 (A1)

where x and y are input (in mmHg) and output (in au) values.  $P_1$  denotes the response range (in au),  $P_2$  is the coefficient of gain,  $P_3$  is the midpoint of the input range (in mmHg),  $P_4$  is the minimum output value of the symmetric sigmoid curve (in au), and Th is a threshold value for the output (in au). The function  $\max\{a,b\}$  gives the greater or equal value between a and b. We set  $P_1 = 135$  au,  $P_2 = 0.13$ ,  $P_3 = 110$  mmHg,  $P_4 = -40$  au, and Th = 0 au. Under muscle stretch conditions, the value of  $P_4$  was changed to 5 au. These settings were determined based on the static interaction

between the baroreflex and muscle mechanoreflex obtained from previous studies (58, 59).

The sigmoidal nonlinearity in the peripheral arc was modelled by a four-parameter logistic function as follows:

$$z = \frac{Q_1}{1 + \exp[Q_2(y - Q_3)]} + Q_4 \tag{A2}$$

where y and  $\varepsilon$  are input (in au) and output (in mmHg) values.  $Q_1$  denotes the response range (in mmHg),  $Q_2$  is the coefficient of gain,  $Q_3$  is the midpoint of the input range (in au), and  $Q_4$  is the minimum output value (in mmHg). We set  $Q_1 = 120$  mmHg,  $Q_2 = -0.05$ ,  $Q_3 = 70$  au, and  $Q_4 = 30$  mmHg under both conditions, according to a previous study (58).

The neural arc  $(H_N)$  and peripheral arc  $(H_P)$  linear transfer functions under control and muscle stretch conditions were obtained from Fig. 2. Because absolute values of the steady-state gains in the neural and peripheral arcs were determined by a sigmoid curve  $(Eqs.\ AI)$  and A2), the steady-state gains of  $H_N$  and  $H_P$  under both conditions were normalized to unity.

The input amplitude of the stepwise pressure perturbation was -40 mmHg. To mimic pulsatile pressure, we imposed a sinusoidal input on the output from the peripheral arc. The frequency and zero to peak amplitude of the sinusoidal input were 4 Hz and 15 mmHg, respectively (Fig. 6A). The closed-loop AP response was simulated up to 30 s (Fig. 6B).

#### GRANTS

This work was supported by Ministry of Health, Labour and Welfare of Japan Health and Labour Sciences Research Grant for Research on Advanced Medical Technology, Health and Labour Sciences Research Grant for Research on Medical Devices for Analyzing, Supporting and Substituting the Function of Human Body, and Health and Labour Sciences Research Grants H18-Inyo-Ippan-023 and H18-Nano-Ippan-003; the Industrial Technology Research Grant Program of the New Energy and Industrial Technology Development Organization of Japan; and Ministry of Education, Culture, Sports, Science and Technology Grant-In-Aid for Scientific Research 18591992.

#### REFERENCES

- Charkoudian N, Martin EA, Dinenno FA, Eisenach JH, Dietz NM, Joyner MJ. Influence of increased central venous pressure on baroreflex control of sympathetic activity in humans. Am J Physiol Heart Circ Physiol 287: H1658-H1662, 2004.
- Chen CY, Horowitz JM, Bonham AC. A presynaptic mechanism contributes to depression of autonomic signal transmission in NTS. Am J Physiol Heart Circ Physiol 277: H1350–H1360, 1999.
- Fadel PJ, Ogoh S, Watenpaugh DE, Wasmund W, Olivencia-Yurvati A, Smith ML, Raven PB. Carotid buroreflex regulation of sympathetic nerve activity during dynamic exercise in humans. Am J Physiol Heart Circ Physiol 280: H1383–H1390, 2001.
- Gallagher KM, Fadel PJ, Smith SA, Norton KH, Querry RG, Olivencia-Yurvati A, Raven PB. Increases in intramuscular pressure raise arterial blood pressure during dynamic exercise. J Appl Physiol 91: 2351–2358, 2001.
- Gallagher KM, Fadel PJ, Stromstad M, Ide K, Smith SA, Querry RG, Raven PB, Secher NH. Effects of exercise pressor reflex activation on carotid baroreflex function during exercise in humans. J Physiol 533: 871-880, 2001.
- Gallagher KM, Fadel PJ, Stromstad M, Ide K, Smith SA, Querry RG, Raven PB, Secher NH. Effects of partial neuromuscular blockade on carotid baroreflex function during exercise in humans. J Physiol 533: 861–870, 2001.
- Hayes SG, Kaufman MP. Gadolinium attenuates exercise pressor reflex in cats. Am J Physiol Heart Circ Physiol 280: H2153–H2161, 2001.
- Hayes SG, Kindig AE, Kaufman MP. Comparison between the effect of static contraction and tendon stretch on the discharge of group III and IV muscle afferents. J Appl Physiol 99: 1891–1896, 2005.
- Ichinose M, Saito M, FuJii N, Ogawa T, Hayashi K, Kondo N, Nishiyasu T. Modulation of the control of muscle sympathetic nerve activity during incremental leg cycling. J Physiol 586: 2753–2766, 2008.
- Ichinose M, Saito M, Kondo N, Nishiyasu T. Time-dependent modulation of arterial baroreflex control of muscle sympathetic nerve activity

- during isometric exercise in humans. Am J Physiol Heart Circ Physiol 290: H1419-H1426, 2006.
- Ichinose M, Saito M, Wada H, Kitano A, Kondo N, Nishiyasu T. Modulation of arterial baroreflex control of muscle sympathetic nerve activity by muscle metaboreflex in humans. Am J Physiol Heari Circ Physiol 286: H701–H707, 2004.
- Ichinose M, Saito M, Wada H, Kitano A, Kondo N, Nishiyasu T. Modulation of arterial baroreflex dynamic response during muscle metaboreflex activation in humans. J Physiol 544: 939–948, 2002.
- Iellamo F, Legramante JM, Raimondi G, Peruzzi G. Baroreflex control
  of sinus node during dynamic exercise in humans: effects of central
  command and muscle reflexes. Am J Physiol Heart Circ Physiol 272:
  H1157-H1164, 1997.
- Ikeda Y, Kawada T, Sugimachi M, Kawaguchi O, Shishido T, Sato T, Miyano H, Matsuura W, Alexander J Jr, Sunagawa K. Neurai are of baroreflex optimizes dynamic pressure regulation in achieving both stability and quickness. Am J Physiol Heart Circ Physiol 271: H882–H890, 1996.
- Kamiya A, Kawada T, Yamamoto K, Michikami D, Ariumi H, Miyamoto T, Shimizu S, Uemura K, Aiba T, Sunagawa K, Sugimachi M. Dynamic and static baroreflex control of muscle sympathetic nerve activity (SNA) parallels that of renal and cardiac SNA during physiological change in pressure. Am J Physiol Heart Circ Physiol 289: H2641–H2648, 2005.
- Kamiya A, Kawada T, Yamamoto K, Michikami D, Ariumi H, Miyamoto T, Uemura K, Sugimachi M, Sunagawa K. Muscle sympathetic nerve activity averaged over 1 minute parallels renal and cardiac sympathetic nerve activity in response to a forced baroreceptor pressure change. Circulation 112: 384–386, 2005.
- Kamiya A, Michikami D, Fu Q, Nilmi Y, Iwase S, Mano T, Suzumura A. Static handgrip exercise modifies arterial baroreflex control of vascular sympathetic outflow in humans. Am J Physiol Regul Integr Comp Physiol 281: R1134–R1139, 2001.
- Kawada T, Shishido T, Inagaki M, Tatewaki T, Zheng C, Yanagiya Y, Sugimachi M, Sunagawa K. Differential dynamic baroreflex regulation of cardiac and renal sympathetic nerve activities. Am J Physiol Heart Circ Physiol 280: H1581–H1590, 2001.
- Kawada T, Shishido T, Inagaki M, Zheng C, Yanagiya Y, Uemura K, Sugimachi M, Sunagawa K. Estimation of baroreflex gain using a baroreflex equilibrium diagram. Jpn J Physiol 52: 21–29, 2002.
- Kawada T, Uemura K, Kashihara K, Kamiya A, Sugimachi M, Sunagawa K. A derivative-sigmoidal model reproduces operating pointdependent baroreflex neural arc transfer characteristics. Am J Physiol Heart Circ Physiol 286: H2272–H2279, 2004.
- Kawada T, Yamamoto K, Kamiya A, Ariumi H, Michikami D, Shishido T, Sunagawa K, Sugimachi M. Dynamic characteristics of carotid sinus pressure-nerve activity transduction in rabbits. *Jpn J Physiol* 55: 157–163, 2005.
- Kawada T, Yanagiya Y, Uemura K, Miyamoto T, Zheng C, Li M, Sugimachi M, Sunagawa K. Input-size dependence of the baroreflex neural arc transfer characteristics. Am J Physiol Heart Circ Physiol 284: H404-H415, 2003.
- Kawada T, Zheng C, Yanagiya Y, Uemura K, Miyamoto T, Inagaki M, Shishido T, Sugimachi M, Sunagawa K. High-cut characteristics of the baroreflex neural are preserve baroreflex gain against pulsatile pressure. Am J Physiol Heart Circ Physiol 282: H1149-H1156, 2002.
- Keller DM, Fadel PJ, Ogoh S, Brothers RM, Hawkins M, Olivencla-Yurvati A, Raven PB. Carotid baroreflex control of leg vasculature in exercising and non-exercising skeletal muscle in humans. J Physiol 561: 283–293, 2004.
- Leshnower BG, Potts JT, Garry MG, Mitchell JH. Reflex cardiovascular responses evoked by selective activation of skeletal muscle ergoreceptors. J Appl Physiol 90: 308–316, 2001.
- Liu Z, Chen CY, Bonham AC. Frequency limits on aortic baroreceptor input to nucleus tractus solitarii. Am J Physiol Heart Circ Physiol 278: H577–H585, 2000.
- Marmarelis PZ, Marmarelis VZ. The white noise method in system identification. In: Analysis of Physiological Systems. New York: Plenum, 1978. p. 131–221.
- Matsukawa K, Nakamoto T, Inomoto A. Gadolinium does not blunt the cardiovascular responses at the onset of voluntary static exercise in cats: a predominant role of central command. Am J Physiol Heart Circ Physiol 292: H121-H129, 2007.

- McIlveen SA, Hayes SG, Kaufman MP. Both central command and exercise pressor reflex reset carotid sinus baroreflex. Am J Physiol Heart Circ Physiol 280: H1454–H1463, 2001.
- Melcher A, Donald DE. Maintained ability of carotid baroreflex to regulate arterial pressure during exercise. Am J Physiol Heart Circ Physiol 241: H838–H849, 1981.
- Mense S, Stahnke M. Responses in muscle afferent fibres of slow conduction velocity to contractions and ischaemia in the cat. J Physiol 342: 383–397, 1983.
- Miki K, Yoshimoto M, Tanimizu M. Acute shifts of baroreflex control of renal sympathetic nerve activity induced by treadmill exercise in rats. J Physiol 548: 313–322, 2003.
- Miles R. Frequency dependence of synaptic transmission in nucleus of the solitary tract in vitro. J Neurophysiol 55: 1076–1090, 1986.
- Mohrman DE, Heller LJ. Regulation of arterial pressure. In: Cardiovascular Physiology (4th ed.). New York: McGraw-Hill, 1997, p. 158–230.
- Norton KH, Boushel R, Strange S, Saltin B, Raven PB. Resetting of the carotid arterial baroreflex during dynamic exercise in humans. J Appl Physiol 87: 332–338, 1999.
- Ogoh S, Fisher JP, Dawson EA, White MJ, Secher NH, Raven PB. Autonomic nervous system influence on arterial baroreflex control of heart rate during exercise in humans. J Physiol 566: 599-611, 2005.
- Ogoh S, Fisher JP, Fadel PJ, Raven PB. Increases in central blood volume modulate carotid baroreflex resetting during dynamic exercise in humans. J Physiol 581: 405–418, 2007.
- Ogoh S, Fisher JP, Raven PB, Fadel PJ. Arterial buroreflex control of muscle sympathetic nerve activity in the transition from rest to steady-state dynamic exercise in humans. Am J Physiol Heart Circ Physiol 293: H2202-H2209. 2007.
- Ogoh S, Wasmund WL, Keller DM, AOY, Gallagher KM, Mitchell JH, Raven PB. Role of central command in carotid baroreflex resetting in humans during static exercise. J Physiol 543: 349–364, 2002.
- Papelier Y, Escourrou P, Gauthier JP, Rowell LB. Carotid baroreflex control of blood pressure and heart rate in men during dynamic exercise. J Appl Physiol 77: 502–506, 1994.
- Papelier Y, Escourrou P, Helloco F, Rowell LB, Muscle chemoreflex alters carotid sinus baroreflex response in humans. J Appl Physiol 82: 577–583, 1997.
- Potts JT, Hand GA, Li J, Mitchell JH. Central interaction between carotid baroreceptors and skeletal muscle receptors inhibits sympathoexcitation. J Appl Physiol 84: 1158–1165, 1998.
- Potts JT, Li J. Interaction between carotid baroreflex and exercise pressor reflex depends on baroreceptor afferent input. Am J Physiol Heart Circ Physiol 274: H1841–H1847. 1998.
- Potts JT, Mitchell JH. Rapid resetting of carotid baroreceptor reflex by afferent input from skeletal muscle receptors. Am J Physiol Heart Circ Physiol 275: H2000–H2008, 1998.
- Potts JT, Shl XR, Raven PB. Carotid baroreflex responsiveness during dynamic exercise in humans. Am J Physiol Heart Circ Physiol 265: H1928–H1938, 1993.

- Querry RG, Smith SA, Stromstad M, Ide K, Raven PB, Secher NH. Neural blockade during exercise augments central command's contribution to carotid baroreflex resetting. Am J Physiol Heart Circ Physiol 280: H1635–H1644, 2001.
- Rowell LB, O'Leary DS. Reflex control of the circulation during exercise: chemoreflexes and mechanoreflexes. J Appl Physiol 69: 407–418, 1990.
- Sato T, Kawada T, Inagaki M, Shishido T, Takaki H, Sugimachi M, Sunagawa K. New analytic framework for understanding sympathetic baroreflex control of arterial pressure. Am J Physiol Heart Circ Physiol 276: H2251–H2261, 1999.
- Smith SA, Querry RG, Fadel PJ, Gallagher KM, Stromstad M, Ide K, Raven PB, Secher NH. Partial blockade of skeletal muscle somatosensory afferents attenuates baroreflex resetting during exercise in humans. J Physiol 551: 1013–1021, 2003.
- Stebbins CL, Brown B, Levin D, Longhurst JC. Reflex effect of skeletal muscle mechanoreceptor stimulation on the cardiovascular system. J Appl Physiol 65: 1539–1547, 1988.
- Sugimachi M, Imaizumi T, Sunagawa K, Hirooka Y, Todaka K, Takeshita A, Nakamura M. A new method to identify dynamic transduction properties of aortic baroreceptors. Am J Physiol Heart Circ Physiol 258: H887–H895, 1990.
- Suzuki S, Ando S, Imalzumi T, Takeshita A. Effects of anesthesia on sympathetic nerve rhythm: power spectral analysis. J Auton Nerv Syst 43: 51–58, 1993.
- Terul N, Masuda N, Saeki Y, Kumada M. Activity of barosensitive neurons in the caudal ventrolateral medulla that send axonal projections to the rostral ventrolateral medulla in rabbits. *Neurosci Len* 118: 211–214, 1990.
- Vatner SF, Braunwald E. Cardiovascular control mechanisms in the conscious state. N Engl J Med 293: 970–976, 1975.
- Vatner SF, Franklin D, Braunwald E. Effects of anesthesia and sleep on circulatory response to carotid sinus nerve stimulation. Am J Physiol 220: 1249–1255, 1971.
- Vatner SF, Franklin D, Van Citters RL, Braunwald E. Effects of carotid sinus nerve stimulation on blood-flow distribution in conscious dogs at rest and during exercise. Circ Res 27: 495–503, 1970.
- Wray DW, Fadel PJ, Keller DM, Ogoh S, Sander M, Raven PB, Smith ML. Dynamic carotid baroreflex control of the peripheral circulation during exercise in humans. J Physiol 559: 675–684, 2004.
- Yamamoto K, Kawada T, Kamiya A, Takaki H, Miyamoto T, Sugimachi M, Sunagawa K. Muscle mechanoreflex induces the pressor response by resetting the arterial baroreflex neural arc. Am J Physiol Heart Circ Physiol 286: H1382-H1388, 2004.
- Yamamoto K, Kawada T, Kamiya A, Takaki H, Sugimachi M, Sunagawa K. Static interaction between muscle mechanoreflex and arterial baroreflex in determining efferent sympathetic nerve activity. Am J Physiol Heart Cire Physiol 289: H1604–H1609, 2005.

# Upright Tilt Resets Dynamic Transfer Function of Baroreflex Neural Arc to Minify the Pressure Disturbance in Total Baroreflex Control

Atsunori KAMIYA<sup>1</sup>, Toru KAWADA<sup>1</sup>, Kenta YAMAMOTO<sup>2</sup>, Masaki MIZUNO<sup>1</sup>, Shuji SHIMIZU<sup>1</sup>, and Masaru SUGIMACHI<sup>1</sup>

<sup>1</sup>Department of Cardiovascular Dynamics, National Cardiovascular Centre Research Institute, Osaka, Japan; and <sup>2</sup>Consolidated Research Institute for Advanced Science and Medical Care, Waseda University, Tokyo ,162-0041 Japan

## Reprinted from The Journal of Physiological Sciences

Volume 58, Number 3, pp. 189–198, 2008 http://jps.physiology.jp/ doi:10.2170/physiolsci.RP004308

Published by The Physiological Society of Japan

# Upright Tilt Resets Dynamic Transfer Function of Baroreflex Neural Arc to Minify the Pressure Disturbance in Total Baroreflex Control

Atsunori KAMIYA<sup>1</sup>, Toru KAWADA<sup>1</sup>, Kenta YAMAMOTO<sup>2</sup>, Masaki MIZUNO<sup>1</sup>, Shuji SHIMIZU<sup>1</sup>, and Masaru SUGIMACHI<sup>1</sup>

<sup>1</sup>Department of Cardiovascular Dynamics, National Cardiovascular Centre Research Institute, Osaka, Japan; and <sup>2</sup>Consolidated Research Institute for Advanced Science and Medical Care, Waseda University, Tokyo ,162-0041 Japan

Abstract: Maintenance of arterial pressure (AP) under orthostatic stress against gravitational fluid shift and pressure disturbance is of great importance. One of the mechanisms is that upright tilt resets steady-state baroreflex control to a higher sympathetic nerve activity (SNA). However, the dynamic feedback characteristics of the baroreflex system, a hallmark of fast-acting neural control, remain to be elucidated. In the present study, we tested the hypothesis that upright tilt resets the dynamic transfer function of the baroreflex neural arc to minify the pressure disturbance in total baroreflex control. Renal SNA and AP were recorded in ten anesthetized, vagotomized and aortic-denervated rabbits. Under baroreflex open-loop condition, isolated intracarotid sinus pressure (CSP) was changed according to a binary white noise sequence at operating pressure  $\pm$  20

mmHg, while the animal was placed supine and at 60° upright tilt. Regardless of the postures, the baroreflex neural (CSP to SNA) and peripheral (SNA to AP) arcs showed dynamic highpass and low-pass characteristics, respectively. Upright tilt increased the transfer gain of the neural arc (resetting), decreased that of the peripheral arc, and consequently maintained the transfer characteristics of total baroreflex feedback system. A simulation study suggests that postural resetting of the neural arc would significantly increase the transfer gain of the total arc in upright position, and that in closed-loop baroreflex the resetting increases the stability of AP against pressure disturbance under orthostatic stress. In conclusion, upright tilt resets the dynamic transfer function of the baroreflex neural arc to minify the pressure disturbance in total baroreflex control.

Key words: baroreflex, blood pressure, sympathetic nervous system.

Since human beings are often under orthostatic stress, the maintenance of arterial pressure (AP) under orthostatic stress against gravitational fluid shift is of great importance. During standing, a gravitational fluid shift directed toward the lower part of the body would cause severe postural hypotension if not counteracted by compensatory mechanisms [1]. Arterial baroreflex has been considered to be the major compensatory mechanism [1–3], since denervation of baroreceptor afferents causes profound postural hypotension [4].

The baroreflex system consists of two subsystems: the neural arc that represents the input-output relationship between baroreceptor pressure and sympathetic nerve activity (SNA), and the peripheral arc that represents the relationship between SNA and systemic AP. Recently, we investigated the steady-state functional structure of these systems under orthostatic stress [5], and reported that upright tilt shifted the baroreflex peripheral arc to a lower AP for a given SNA. However, upright tilt reset the baroreflex neural arc to a higher steady state SNA. The resetting compensat-

ed for the blunted responsiveness of the peripheral arc and contributed to prevent postural hypotension [5].

In addition to the steady state characteristics [6, 7], the dynamic characteristics are other hallmark of the barore-flex system. It is because the system is a fast-acting neural control that quickly negative-feedback controls and stabilises AP against pressure disturbance in contrast to the slow-acting hormonal and humoral systems [8]. Earlier studies reported that the dynamic characteristics in supine position have a high-pass (fast) neural arc that may compensate for the low-pass (slow) peripheral arc to achieve rapid and stable AP regulation [8]. The importance of the dynamic characteristics in AP control increases under orthostatic stress that can cause postural hypotension. However, little is known about the dynamic characteristics of the baroreflex system in upright posture.

Because the gravitational body fluid shift decreases the effective circulatory blood volume [1, 9], we speculated that upright tilt may attenuate the dynamic transfer function from SNA to AP in the baroreflex peripheral arc.

Received on Mar 18, 2008; accepted on May 9, 2008; released online on May 13, 2008; dol:10.2170/physiolsci.RP004308
Correspondence should be addressed to: Atsunori Kamiya, Department of Cardiovascular Dynamics, National Cardiovascular Centre Research Institute, Osaka, 565-8565 Japan. Tel: +81-6-6833-5012, Fax: +81-6-6835-5403, E-mail: kamiya@ri.ncvc.go.jp

Moreover, if the upright tilt resets the dynamic characteristics of the neural arc in addition to resetting the steady state SNA reported previously [5], it would compensate for a blunted pressor response of the baroreflex peripheral arc and contribute to maintain the stability and quickness of the total baroreflex system. Accordingly, we hypothesized that upright tilt resets dynamic transfer function of baroreflex neural arc to minify the pressure disturbance in total baroreflex control.

In the present study, we identified the transfer functions of two baroreflex subsystems (the neural and peripheral arcs) separately in 60° upright posture, while opening the baroreflex negative feedback loop by vascular isolation of carotid sinus regions [8]. In addition, by connecting the subsystem transfer functions in series and closing them, we investigated the dynamic transfer characteristics and the stability against pressure disturbance of total baroreflex arc system in upright posture.

#### MATERIAL AND METHODS

Animals were cared for in strict accordance with the Guiding Principles for the Care and Use of Animals in the Field of Physiological Science approved by the Physiological Society of Japan. Ten Japanese white rabbits weighing 2.4-3.3 kg were initially anesthetized by intravenous injection (2 ml/kg) of a mixture of urethane (250 mg/ml) and α-chloralose (40 mg/ml). Anesthesia was maintained by continuously infusing the anaesthetics at a rate of 0.33 ml/kg/h using a syringe pump (CFV-3200, Nihon Kohden, Tokyo). The rabbits were mechanically ventilated with oxygen-enriched room air. Bilateral carotid sinuses were isolated vascularly from systemic circulation by ligating the internal and external carotid arteries and other small branches originating from the carotid sinus regions. The isolated carotid sinuses were filled with warmed physiological saline pre-equilibrated with atmospheric air, through catheters inserted via the common carotid arteries. Intra-carotid sinus pressure (CSP) was controlled by a servo-controlled piston pump (model ET-126A, Labworks; Costa Mesa, CA). Bilateral vagal and aortic depressor nerves were sectioned in the middle of the neck region to eliminate reflexes from the cardiopulmonary region and the aortic arch. Systemic AP was measured using a high-fidelity pressure transducer (Millar Instruments; Houston, TX) inserted retrograde from the right common carotid artery below the isolated carotid sinus region. Body temperature was maintained at around 38°C with a heating pad.

The left renal sympathetic nerve was exposed retroperitoneally. A pair of stainless steel wire electrodes (Bioflex wire AS633, Cooner Wire) was attached to the nerve to record renal SNA. The nerve fibers peripheral to electrodes were ligated securely and crushed to eliminate afferent signals. The nerve and electrodes were covered with a mixture of silicone gel (Silicon Low Viscosity, KWIK-SIL, World Precision Instrument, Inc., FL) to insulate and immobilize the electrodes. The preamplified SNA signal was band-pass filtered at 150–1,000 Hz. The nerve signal was full-wave rectified and low-pass filtered with a cutoff frequency of 30 Hz to quantify the nerve activity.

Protocols. Both protocols 1 and 2 were performed on each of eight animals. After the surgical preparation, the animal was maintained supine (0°) on a tilt bed. To stabilize the posture, the head was fixed full-frontal to the bed by strings, and the body and legs were rigged up in a clothes-like bag. Before performing protocols 1 and 2, we confirmed that the nerve activity measured in supine position was SNA. CSP was decreased stepwise from 100 mmHg to 40 mmHg in decrements of 20 mmHg, and then increased stepwise to 100 mmHg in increments of 20 mmHg. Each pressure step was maintained for 60 s. In all animals, a decrease in CSP increased SNA, whereas an increase in CSP decreased SNA (Fig. 1), indicating that the nerve activity recorded was SNA.

Protocol 1: The animal was placed supine. CSP was firstly matched with systemic AP to obtain the operating AP under the baroreflex closed-loop condition. After at least 5 minutes of stabilization, the SNA and AP were recorded for 10 min to obtain closed-loop baseline values. The data were stored on the hard disk of a dedicated laboratory computer system for analysis at a sampling rate of 200 Hz using a 12-bit analog-to-digital converter. The averaged AP over 10 min was defined as the operating AP in

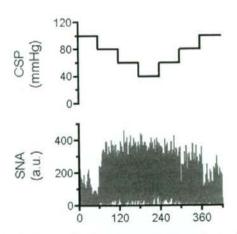


Fig. 1. Representative data of one rabbit in supine position, showing time series of carotid sinus pressure (CSP) and sympathetic nerve activity (SNA). CSP was decreased stepwise from 100 mmHg to 40 mmHg in decrements of 20 mmHg, and then increased stepwise to 100 mmHg in increments of 20 mmHg. Each pressure step was maintained for 60 s. A decrease in CSP increased SNA, whereas an increase in CSP decreased SNA, indicating that the nerve activity recorded was SNA, a.u., arbitrary unit.

supine position. Then, after at least 5 min of stabilization, CSP was randomly changed by 20 mmHg above or below the operating AP every 500 ms according to a binary white noise sequence for which the input power spectrum of CSP was reasonably flat up to 1 Hz [10]. The variables were recorded for a 10-min period and stored.

Protocol 2: CSP was firstly matched with systemic AP via a servo-controlled piston pump to obtain the actual operating pressure under baroreflex closed-loop conditions in supine and 60° upright postures. The animal was maintained supine for 10 min, and then tilted upright to 60° within 10 s by inclining the tilt bed to 60° and dropping the lower regions of the rabbit with the fulcrum set at the level of the carotid sinus. The 60° upright posture was maintained for 10 min for stabilization. Since the clotheslike bag stabilized the posture of the animal, there was no additional mechanical movement that reduced the quality of measurements. The position of the head remained almost fixed during the tilt to minimize vestibular stimulation. Thereafter, the average AP over the next 10 min was defined as the operating AP in upright tilt position. Then, after at least 5 min of stabilization, CSP was randomly changed according to a white noise sequence for 10 min as in protocol 1.

Data analysis. SNA signals were normalized by the following steps. First, the post-mortem noise level was assigned 0 arbitrary unit (a.u.). Second, SNA signals during the 10-min closed-loop baseline recording in protocol 1 (supine position) were averaged over 1 min, and assigned 100 a.u. Finally, the other SNA signals in all protocols were normalized to these values.

In both protocols 1 and 2, the transfer functions (gain and phase) and coherence function were calculated from CSP input to SNA in the baroreflex neural arc and from SNA input to AP in the baroreflex peripheral arc. The signals of CSP, SNA and AP were resampled at 10 Hz and segmented into 10 sets of 50% overlapping bins of  $2^{10}$  data point each. The segment length was 102.4 s, which yielded the lowest frequency bound of 0.01 (0.0097) Hz. We subtracted a linear trend and applied a Hanning window for each segment. We then performed fast Fourier transform to obtain frequency spectra of the variables. We ensemble averaged the input power [Sxx(f)], output power [Syy(f)], and cross power between them [Syx(f)] over the 10 segments. Thereafter, we calculated the transfer function [H(f)] from input to output signals as follows,

$$H(f) = \frac{Syx(f)}{Sxx(f)}$$

To quantify the linear dependence between input to output signals in the frequency domain, we calculated the magnitude-squared coherence function [Coh(f)] as follows:

$$Coh(f) = \frac{\left|Syx(f)\right|^2}{Sxx(f)Syy(f)}$$

The coherence value ranges from zero to unity. Unity coherence indicates a perfect linear dependence between input and output signals, whereas zero coherence indicates total independence of these two signals.

Statistic analysis. All data are presented as means  $\pm$  SD. Effects of upright tilt on baroreflex parameters were evaluated by repeated-measures analysis of variance. When the main effect was found to be significant, post hoc multiple comparisons were done using the Scheff's F-test to compare baroreflex controls between the supine and upright postures [11]. Differences were considered significant when P < 0.05.

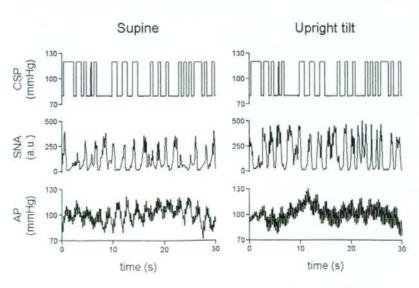


Fig. 2. Representative data of one rabbit in supine (left panels) and 60° upright tilt (right panels) positions, showing time series of carotid sinus pressure (CSP), sympathetic nerve activity (SNA) and systemic arterial pressure (AP) during CSP perturbation. CSP was changed according to a binary white noise signal with a switching interval of 500 ms. a.u., arbitrary unit.

#### RESULTS

Figure 2 shows the typical time series of CSP, SNA and AP derived in supine and 60° upright tilt positions in individual animal. CSP was perturbed according to a binary white noise sequence at 500-ms intervals. In both positions, SNA increased and decreased roughly in response to the decrease and increase in CSP, respectively. However, the SNA responses appeared higher in the upright tilt

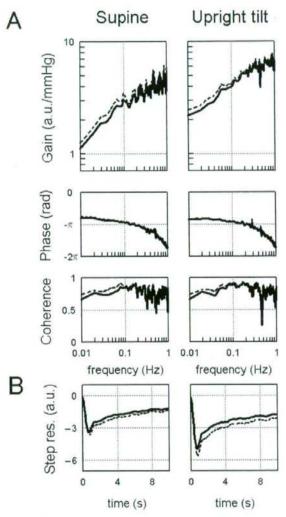


Fig. 3. A: The transfer function of the baroreflex neural arc from CSP to SNA averaged from all animals (n=8) in supine (left panels) and  $60^{\circ}$  upright tilt (right panels) positions. The gain plots (top), phase plots (middle), and coherence function (bottom) are shown. Upright tilt increases the gain. B: Step responses (Step res.) derived from the transfer function corresponding to the transfer function shown in A. Upright tilt enhances the initial and steady-state responses. Solid line represents the mean values, and dashed line represents mean + SD in A and mean – SD in B. a.u., arbitrary unit.

than in the supine position. Data from all animals (n=8) showed that the upright tilt increased the averaged SNA  $(175\pm21~a.u.)$  during CSP perturbation compared with the supine position  $(96\pm13~a.u.)$ . Averaged AP during CSP perturbation was similar in supine  $(96\pm13~mmHg)$  and in upright positions  $(103\pm15~mmHg)$ .

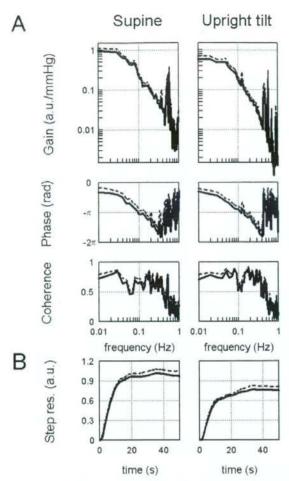
#### The baroreflex neural arc

Figure 3A shows the transfer function of baroreflex neural arc from CSP to SNA averaged from all animals. In both supine and upright tilt positions, the transfer gain increased as the frequency of CSP perturbation increased for the frequency range of 0.01 to 1 Hz. This shows dynamic high-pass characteristics, indicating that more rapid change of CSP results in greater response of SNA. Note that upright tilt increased the transfer gain for the whole frequency range observed (Table 1). In addition, upright tilt decreased the slope of gain increase. In both positions, the phase approached slightly above -π radians at the lowest frequency reflecting negative feedback characters, and lagged as the frequency of CSP perturbation increased. The coherence was over 0.7 for the frequency range of 0.01 to 0.2 Hz. Upright tilt did not affect the phase or coherence. Figure 3B shows the step response of SNA corresponding to the transfer function shown in Fig. 3A. In both positions, the SNA response consisted of an initial decrease followed by partial recovery and then a steady state. Of note, upright tilt enhanced the initial decrease by 50%, and also decreased the steady-state SNA.

Table 1. Transfer function of baroreflex neural arc (from CSP to SNA) in supine and upright tilt positions.

	Supine	Upright tilt
Gain (a.u./mmHg)		
0.01 Hz	$1.11 \pm 0.13$	$2.14 \pm 0.41*$
0.1 Hz	$2.75 \pm 0.43$	$4.63 \pm 0.52$ *
0.3 Hz	$3.69 \pm 0.30$	$5.08 \pm 0.42$ *
Phase (rad)		
0.01 Hz	$-2.51 \pm 0.15$	$-2.66 \pm 0.09$
0.1 Hz	$-2.96 \pm 0.08$	$-2.93 \pm 0.06$
0.3 Hz	$-3.58 \pm 0.14$	$-3.53 \pm 0.12$
Coherence		
0.01 Hz	$0.67 \pm 0.08$	$0.67 \pm 0.07$
0.1 Hz	$0.84 \pm 0.04$	$\boldsymbol{0.89 \pm 0.02}$
0.3 Hz	$0.77 \pm 0.06$	$0.82 \pm 0.03$
Slope (dB/decade)		
0.01 Hz to 0.3 Hz	$7.0 \pm 0.4$	$5.1 \pm 0.5^*$
Step response (a.u.)		
Initial response	$-3.41 \pm 0.21$	$-4.99 \pm 0.62$
Steady-state level	$-1.26 \pm 0.18$	$-1.80 \pm 0.32$

Values are mean  $\pm$  SD (n = 10). \*P < 0.05; supine position vs. upright tilt.



**Fig. 4.** A: The transfer function of the baroreflex peripheral arc from SNA to AP averaged from all animals (n=8) in supine (left panels) and  $60^\circ$  upright tilt (right panels) positions. The gain plots (top), phase plots (middle), and coherence function (bottom) are shown. Upright tilt decreases the gain below the frequency of 0.1 Hz. **B**: Step responses (Step res.) derived from the transfer function corresponding to the transfer function shown in A. Upright tilt attenuates the response. Solid and dashed lines represent the mean and mean + SD values, respectively. a.u., arbitrary unit.

#### The baroreflex peripheral arc

Figure 4A shows the transfer function of the baroreflex peripheral arc from SNA to AP averaged from all animals. In both supine and upright tilt positions, the transfer gain decreased as the input frequency increased for the frequency range of 0.01 to 1 Hz, indicating low-pass characteristics. Upright tilt decreased the transfer gain between 0.01 and 0.1 Hz (Table 2). In both positions, the phase approached zero radian at the lowest frequency reflecting an increase in SNA with increased AP, and lagged as the in-

Table 2. Transfer function of baroreflex peripheral arc (from SNA to AP) in supine and upright tilt positions.

	Supine	Upright tilt
Gain (mmHg/au)		
0.01 Hz	$0.97 \pm 0.09$	$0.63 \pm 0.06^{\circ}$
0.1 Hz	$0.23 \pm 0.03$	$0.15 \pm 0.03^*$
0.3 Hz	$0.04 \pm 0.006$	$0.03 \pm 0.003$
Phase (rad)		
0.01 Hz	$-0.79 \pm 0.16$	$-0.69 \pm 0.07$
0.1 Hz	$-2.83 \pm 0.14$	$-2.58 \pm 0.15$
0.3 Hz	$-4.74 \pm 0.18$	$-4.63 \pm 0.08$
Coherence		
0.01 Hz	$0.72 \pm 0.07$	$0.71 \pm 0.03$
0.1 Hz	$0.64 \pm 0.08$	$\textbf{0.62} \pm \textbf{0.04}$
0.3 Hz	$0.61 \pm 0.08$	$0.68 \pm 0.02$
Step response (mmH	lg)	
Steady-state level	$-0.97 \pm 0.06$	$-0.75 \pm 0.06$ *

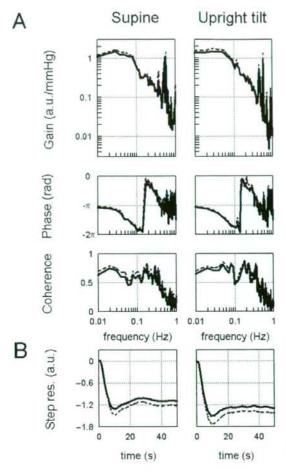
Values are mean  $\pm$  SD (n =10). \*P < 0.05; supine position vs. upright tilt.

put frequency increased. The coherence was over 0.5 for the frequency range of 0.01 to 1 Hz. Upright tilt did not affect the phase or coherence. Figure 4B shows the step response of AP corresponding to the transfer function shown in Fig. 4A. In both positions, the AP response increased gradually to reach a steady state. Upright tilt decreased the steady-state AP.

### The total baroreflex arc

Figure 5A shows the transfer function of the total baroreflex arc from CSP to AP averaged from all animals. In both supine and upright tilt positions, the transfer gain decreased as the input frequency increased for the frequency range from 0.01 to 1 Hz, indicating low-pass characteristics. Upright tilt did not affect the transfer gain (Table 3). In both positions, the phase approached  $-\pi$  radians at the lowest frequency reflecting negative feedback attained by the total baroreflex loop, and lagged as the input frequency increased. The coherence was over 0.5 for the frequency range from 0.01 to 0.2 Hz. Upright tilt did not affect the phase or coherence. Figure 5B shows the step response of AP corresponding to the transfer function shown in Fig. 5A. In both positions, the AP response increased gradually to reach a steady state. Upright tilt did not affect the step response.

The right column of Table 3 shows a simulation of the total arc transfer function in the absence of resetting in the neural arc. The simulation was based on the neural arc transfer function in supine position and the peripheral arc transfer function in upright tilt position. Without the resetting, the upright tilt would decrease the transfer function gain and would attenuate the step response of AP at steady state, compared with the values in supine position and those in upright tilt position with resetting.



#### DISCUSSION

Arterial baroreflex is obviously a pivotal mechanism for maintaining AP under orthostatic stress against gravitational fluid shift and pressure disturbance [1, 2, 4], but the baroreflex function and its modulation in upright position are not fully understood. We previously reported that 60° upright tilt resets the steady-state characteristics of the baroreflex neural arc to a higher SNA [5]. However, the dynamic characteristics of the baroreflex system, which is a hallmark of fast-acting neural systems, in upright posture remain to be elucidated. Accordingly, in the present study, we identified the transfer function of the total baroreflex system and its two subsystems. The new major findings are that a 60° upright tilt increases the transfer gain of the baroreflex neural arc (CSP to SNA), decreases the transfer gain of the peripheral arc (SNA to AP), and as a result maintains the dynamic characteristics of the total baroreflex feedback system. These findings support our hypothesis that upright tilt resets dynamic transfer function of baroreflex neural arc to minify the pressure disturbance in total baroreflex control. These results were not affected by the order of postures, since returning the ani-

Fig. 5. A: The transfer function of the total baroreflex arc from CSP to AP averaged from all animals (n=8) in supine (left panels) and  $60^\circ$  upright tilt (right panels) positions. The gain plots (top), phase plots (middle), and coherence function (bottom) are shown. B: Step responses (Step res.) derived from the transfer function corresponding to the transfer function shown in A. The transfer function and step response are similar in the supine and upright tilt positions. Solid and dashed lines represent the mean and mean + SD values in A and mean – SD values in B, respectively, a.u., arbitrary unit.

Table 3. Transfer function of total baroreflex arc (from CSP to AP) in supine, upright tilt and simulated upright tilt positions.

	Supine	Upright tilt	Simulated upright tilt withou resetting of the neural arc
Gain (a.u./mmHg)			
0.01 Hz	$1.10 \pm 0.12$	$1.38 \pm 0.18$	$0.71 \pm 0.18*#$
0.1 Hz	$0.63 \pm 0.09$	$0.69 \pm 0.12$	$0.41 \pm 0.13*#$
0.3 Hz	$0.15 \pm 0.03$	$0.15 \pm 0.03$	$0.11 \pm 0.04$ *
Phase (rad)			
0.01 Hz	$-3.33 \pm 0.11$	$-3.29 \pm 0.07$	$-3.21 \pm 0.10$
0.1 Hz	$-5.76 \pm 0.20$	$-5.55 \pm 0.10$	$-5.51 \pm 0.15$
0.3 Hz	$-1.98 \pm 0.25$	$-1.87 \pm 0.23$	$-1.91 \pm 0.24$
Coherence			
0.01 Hz	$0.63 \pm 0.06$	$0.65 \pm 0.05$	
0.1 Hz	$0.60 \pm 0.10$	$0.61 \pm 0.06$	
0.3 Hz	$0.53 \pm 0.07$	$0.55 \pm 0.04$	
Step response (mmHg)			
Steady-state level	$-1.09 \pm 0.11$	$-1.29 \pm 0.12$	$-0.67 \pm 0.11*$ #

Simulated transfer function in the absence of neural arc resetting is calculated from the neural arc transfer function in supine position and the peripheral arc transfer function in upright tilt position. Values are mean  $\pm$  SD (n = 10). \*P < 0.05; supine vs. simulated upright tilt, #P < 0.05; upright tilt vs. simulated upright tilt.

mal posture from 60° upright tilt to horizontal supine position restored the transfer functions to the magnitudes observed in the initial supine position (data not shown).

Little is known about the arterial baroreflex feedback system under orthostatic stress. Although earlier studies investigated the gains of baroreflex control of SNA [12–14], vascular resistance [15] and R-R interval [16], these gains are parts of the total baroreflex system, and thus are insufficient to explain the dynamics of the total arc of the baroreflex feedback system. In addition, no study has examined the phase function of baroreflex in the subsystems and the total system. Moreover, while earlier studies addressed baroreflex in relation to AP regulation under orthostatic stress, most of them evaluated the baroreflex in supine, and not orthostatic posture [14]. In the present study, we identified the transfer functions of the two baroreflex subsystems (the neural and peripheral arcs) in

upright posture independently using the baroreflex openloop technique. Moreover, by connecting the subsystem transfer functions in series and closing them, we revealed the dynamic characteristics of the total baroreflex arc.

Our actual and simulation data indicated that resetting of the baroreflex neural arc in upright posture increases the transfer function gain of the total baroreflex arc. In our experiments, the 60° upright tilt reset and nearly doubled the transfer gain of the neural arc. Although the upright tilt decreased the transfer gain of the peripheral arc, resetting in the neural arc counteracted it and consequently preserved the dynamic transfer gain of the total baroreflex arc (1.4, Table 3). In a simulation of a situation where resetting in the neural arc is absent (Table 3), a 60° upright tilt would decrease the total arc transfer gain. These findings suggest that resetting of the neural arc (that is, baroreflex control of SNA) with dynamic characteristics plays an im-

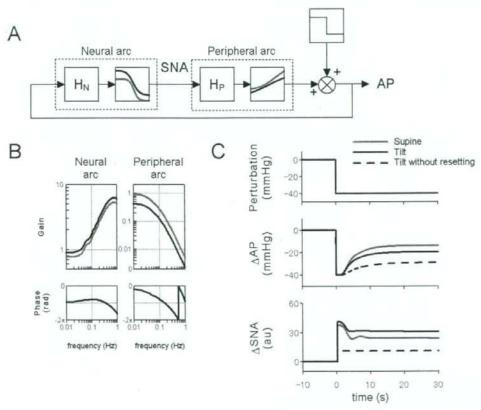


Fig. 6. A: Simulator of the baroreflex system during upright tilt. A stepwise perturbation was applied to the baroreflex negative feedback system (see APPENDIX for details). H<sub>N</sub>, neural arc transfer function; H<sub>P</sub> peripheral arc transfer function. Nonlinear sigmoidal functions in the supine and upright tilt positions are shown by gray and black lines, respectively. B: Simulation results of integrated dynamic transfer function of linear-sigmoidal nonlinear cascade model in the neural (AP to SNA) and

peripheral (SNA to AP) arcs in the supine (gray lines) and upright tilt (black lines) positions. **C**: Simulation results of a closed-loop AP and SNA responses to the stepwise pressure perturbation (–40 mmHg). The resetting during upright tilt (black line) would enhance SNA excitation as compared with the supine position (gray line) to minify a hypotension. Without the resetting in upright tilt, SNA responses would largely be attenuated to lead a hypotension.

portant role to maintain the dynamic transfer function of the total baroreflex system in upright posture.

A simulation of AP stability by baroreflex feedback control against pressure disturbance clearly suggests the importance of resetting of baroreflex neural arc in upright posture. Figure 6 shows the simulation of closed-loop baroreflex control of AP, when pressure disturbance was loaded to the peripheral cardiovascular compartment. According to an earlier study [17], we used the linear-sigmoidal nonlinear cascade model (Fig. 6A) to simulate the baroreflex dynamics. The result of simulation (Fig. 6B) was consistent with our in vivo findings that an upright tilt increased the dynamic transfer gain of the neural arc and decreased the dynamic gain of the peripheral arc. The simulation (Fig. 6C) shows that the baroreflex feedback system would minify the pressure disturbance (40 mmHg) by 50% or more in supine (14 mmHg) and upright tilt (19 mmHg) positions. However, without the resetting of the neural arc in upright tilt, the residual pressure disturbance (29 mmHg) would persist and the velocity of pressure response would become slower (Fig. 6C). These findings suggest that dynamic resetting of the neural arc increases the stability and quickness in response of orthostatic AP against pressure disturbance in closed-loop condition of the total baroreflex arc. In addition, the simulation indicates that the resetting would enhance increases in SNA in response to pressure disturbance in upright tilt compared to supine position (Fig. 6C). Without the resetting in upright tilt, the SNA response would be greatly attenuated (Fig. 6C). This suggests that resetting of the neural arc has a critical role in activating SNA appropriately to prevent hypotension by pressure disturbance during orthostatic stress.

Some explanations for the changes in baroreflex peripheral arc in upright tilt posture may be postulated. First, since the gravitational fluid shift toward the lower part of body (i.e., abdominal vascular bed, lower limbs) during upright posture decreases the preload and effective circulatory blood volume [1, 9], it may attenuate the dynamic transfer function from SNA to AP. Our actual data revealed that upright tilt decreased the transfer gain, but not the transfer phase, of the baroreflex peripheral arc (Fig. 4A). Therefore, upright tilt would blunt the magnitude of AP response to SNA without delaying the response, as shown in the calculated step response (Fig. 4B). Next, increases in humoral factors (i.e., catecholamine, angiotensin II) during upright posture could reduce the dependency of vascular resistance on neural control. However, intravenous infusion of angiotensin II did not affect the transfer function of baroreflex peripheral arc [18]. Moreover, intravenous infusion of catecholamine had no effects on the transfer function from sympathetic stimulation to heart rate [19]. These studies are consistent with the predominance of sympathetic neural control on cardiovascular pressor function [20].

#### Limitations

The present study has several limitations. First, we excluded the efferent effect of vagally mediated arterial and cardiopulmonary baroreflexes that may affect baroreflex control of SNA. Second, we used an anesthetic agent that may attenuate the baroreflex peripheral arc by reducing the cardiac pumping function, and may affect the neural arc gain. Third, since we sectioned the aortic depressor nerves to open the baroreflex feedback loop, the total baroreflex gain may be lower than the physiological level. Fourth, since we measured only renal SNA, our findings have limited applicability to other SNA. Although static [10, 21] and dynamic [21] regulation of the baroreflex neural arc is similar in renal, cardiac and muscle (vasoconstrictor) SNAs in supine posture, whether this holds true during orthostatic stress remains to be verified.

Lastly, we used rabbits that are quadrupeds. Since humans spend most of their time in nearly 90° upright postures whereas rabbits do not, our findings have limited applicability to humans. However, Japanese White rabbits spend most of their time in 10-40° head-up postures, and frequently stand up to nearly 70°. Since the denervation of both carotid and aortic arterial baroreflexes is known to cause severe postural hypotension at 60° upright tilt in quadrupeds [4], this suggests that even in quadrupeds, arterial baroreflex has a very important function in the maintenance of AP under orthostatic stress. Accordingly, despite the difference in species, our findings may reflect, at least, the qualitative aspects of orthostatic baroreflex physiology in humans. Indeed, recent human studies have suggested that orthostatic stress (lower body negative pressure) enhances the SNA response to AP change [22, 23] and increases baroreflex control of SNA (assessed by the relation between spontaneous changes in diastolic AP and SNA) [12] under baroreflex closed-loop condition.

In conclusion, the transfer function identified in baroreflex open-loop condition showed that 60° upright tilt increases the transfer gain of the baroreflex neural arc, decreases the transfer gain of the peripheral arc, and as a result maintains the dynamic characteristics of the total baroreflex feedback system. Simulation study suggests that resetting of the neural arc increases the transfer gain of the total baroreflex arc and also increases the stability of orthostatic AP against pressure disturbance. These findings suggest that upright tilt resets the dynamic transfer function of the baroreflex neural arc to maintain total baroreflex stability.

#### **APPENDIX**

To simulate the closed-loop AP response to stepwise pressure perturbation (Fig. 6), we used the linear-sigmoidal nonlinear cascade model [17].

We modeled the sigmoidal nonlinearity in the baroreflex neural arc by a four-parameter logistic function with threshold according to our previous study [5] using the following equation:

$$y = \frac{P_1}{1 + \exp[P_2(x - P_3)]} + P_4$$

where x and y are input (in mmHg) and output (in au) values.  $P_1$  denotes the response range (in a.u.),  $P_2$  is the coefficient of gain,  $P_3$  is the midpoint of the input range (in mmHg),  $P_4$  is the minimum output value of the symmetric sigmoid curve (in a.u.). We set  $P_1 = 94$ ,  $P_2 = 0.10$ ,  $P_3 = 109$ ,  $P_4 = 4$  in the supine position, and  $P_1 = 112$ ,  $P_2 = 0.09$ ,  $P_3 = 109$ ,  $P_4 = 29$  during upright tilt, according to our previous study [51].

The sigmoidal nonlinearity in the peripheral arc was modeled by a four-parameter logistic function using the following equation:

$$z = \frac{Q_1}{1 + \exp[Q_2(y - Q_1)]} + Q_4$$

where y and z are input (in a.u.) and output (in mmHg) values.  $Q_1$  denotes the response range (in mmHg),  $Q_2$  is the coefficient of gain,  $Q_3$  is the midpoint of the input range (in a.u.), and  $Q_4$  is the minimum output value (in mmHg). We set  $Q_1 = 115$ ,  $Q_2 = -0.04$ ,  $Q_3 = 63$ ,  $Q_4 = 50$  in the supine position, and  $Q_1 = 82$ ,  $Q_2 = -0.05$ ,  $Q_3 = 88$ ,  $Q_4 = 50$  during upright tilt, according to our previous study [5].

In rabbits, the transfer function of the baroreflex neural arc (baroreceptor pressure to SNA) approximates derivative characteristics in the frequency range below  $0.8~{\rm Hz}$ , and high-cut characteristics of frequencies above  $0.8~{\rm Hz}$  [17]. Therefore, according to our previous study [17], we modeled the neural arc transfer function  $(H_N)$  using the following equation:

$$H_{N}(f) = -K_{N} \frac{1 + \frac{f}{f_{c1}} f}{\left(1 + \frac{f}{f_{c2}}\right)^{2}} \exp(-2\pi f j L)$$

where f and f represent the frequency (in Hz) and imaginary units, respectively;  $K_N$  is static gain (in a.u./mmHg);  $f_{c1}$  and  $f_{c2}$  ( $fc_1 < f_{c2}$ ) are corner frequencies (in Hz) for derivative and high-cut characteristics, respectively; and L is a pure delay (in s) that would represent the sum of delays in the synaptic transmission through the baroreflex central pathways and the sympathetic ganglion. The dynamic gain increases in the frequency range of  $f_{c1}$  to  $f_{c2}$  and decreases above  $f_{c2}$ . In simulations showed in Fig. 6, we matched  $K_N$  to the actual data in the supine and upright tilt positions in this study. We also set  $f_{c1}$ ,  $f_{c2}$  and L at 0.05, 0.8 and 0.2, respectively, according to the present and previous studies [17].

In addition, the transfer function of the baroreflex peripheral arc (SNA to AP) approximates a second-order low-pass filter with the dead time as follows:

$$H_p(f) = K_p \frac{1}{1 + 2\varsigma \frac{f}{f_N} j - \left(\frac{f}{f_N}\right)^2} \exp(-2\pi f j L)$$

where  $f_N$  and  $\zeta$  are the neutral frequency (in Hz) and damping ratio, respectively; and L is a pure delay (in s). In simulations showed in Fig. 6, we matched  $K_P$  to the actual data in the supine and upright tilt positions in this study. We also set  $f_N$ ,  $\zeta$  and L at 0.07, 1.4 and 1.0, respectively, according to the present and previous studies [17].

The input amplitude of the stepwise pressure perturbation was -40 mmHg (Fig. 5, A and C, top panel). The closed-loop AP (Fig. 5C, middle panel) and SNA (Fig. 5C, bottom panel) responses were simulated up to 30 s.

This study was supported by the research project promoted by Ministry of Health, Labour and Welfare in Japan (#H18-nano-ippan-003), the Grants-in-Aid for Scientific Research promoted by Ministry of Education, Culture, Sports, Science and Technology in Japan (#18591992, #20390462) and the Industrial Technology Research Grant Program from New Energy and Industrial Technology Development Organization of Japan.

#### REFERENCES

- Rowell LB. Human cardiovascular control. New York: Oxford Univ. Press, 1993.
- Eckberg DL, Sleight P. Human baroreflexes in Health and Disease. New York: Oxford Univ. Press, 1992.
- Persson P, Kirchheim H. Baroreceptor reflexes: integrative functions and clinical aspects. Berlin: Springer-Verlag, 1991.
- Sato T, Kawada T, Sugimachi M, Sunagawa K. Bionic technology revitalizes native baroreflex function in rats with baroreflex failure. Circulation. 2002;106:730-4
- Kamiya A, Kawada T, Yamamoto K, Michikami D, Ariumi H, Uemura K, et al. Resetting of the arterial baroreflex increases orthostatic sympathetic activation and prevents postural hypotension in rabbits. J Physiol. 2005;566:237-46.
- Sato T, Kawada T, Inagaki M, Shishido T, Takaki H, Sugimachi M, et al. New analytic framework for understanding sympathetic baroreflex control of arterial pressure. Am J Physiol. 1999;276:H2251-61.
- Yamamoto K, Kawada T, Kamiya A, Takaki H, Miyamoto T, Sugimachi M, et al. Muscle mechanoreflex induces the pressor response by resetting the arterial baroreflex neural arc. Am J Physiol. 2004;286:H1382-8.
- Ikeda Y, Kawada T, Sugimachi M, Kawaguchi O, Shishido T, Sato T, et al. Neural arc of baroreflex optimizes dynamic pressure regulation in achieving both stability and quickness. Am J Physiol. 1996;271:H882-90.
- Sagawa K, Maughan L, Suga H, Sunagawa K. Cardiac contraction and the pressure-volume relationship. New York: Oxford Univ Press, 1988.
- Kawada T, Shishido T, Inagaki M, Tatewaki T, Zheng C, Yanagiya Y, et al. Differential dynamic baroreflex regulation of cardiac and renal sympathetic nerve activities. Am J Physiol Heart Circ Physiol. 2001;280:H1581-90.
- 11. Glantz SA. Primer of Biostatistics (4th ed). New York: McGraw-Hill, 1997.
- Ichinose M, Saito M, Fujii N, Kondo N, Nishiyasu T. Modulation of the control of muscle sympathetic nerve activity during severe orthostatic stress. J Physiol. 2006;576:947-58.
- Fu Q, Shook RP, Okazaki K, Hastings JL, Shibata S, Conner CL, et al. Vasomotor sympathetic neural control is maintained during sustained upright posture in humans. J Physiol (Lond). 2006;577:679-87.
- Mosqueda-Garcia R, Furlan R, Fernandez-Violante R, Desai T, Snell M, Jaral Z, et al. Sympathetic and baroreceptor reflex function in neurally mediated syncope evoked by tilt. J Clin Invest. 1997;99:2736-44.
- Cooper VL, Hainsworth R. Carotid baroreceptor reflexes in humans during orthostatic stress. Exp Physiol. 2001;86:677-81.
- 16. Cooke WH, Hoag JB, Crossman AA, Kuusela TA, Tahvanainen KU, Eckberg DL

### A. KAMIYA et al.

- Human responses to upright tilt: a window on central autonomic integration. J Physiol. 1999;517:617-28.
- Kawada T, Yanagiya Y, Uemura K, Miyamoto T, Zheng C, Li M, et al. Input-size dependence of the baroreflex neural arc transfer characteristics. Am J Physiol Heart Circ Physiol. 2003;284:H404-15.
- Kashihara K, Takahashi Y, Chatani K, Kawada T, Zheng C, Li M, et al. Intravenous angiotensin II does not affect dynamic baroreflex characteristics of the neural or peripheral arc. Jpn J Physiol. 2003;53:135-43.
- Kawada T, Miyamoto T, Miyoshi Y, Yamaguchi S, Tanabe Y, Kamiya A, et al. Sympathetic neural regulation of heart rate is robust against high plasma catecholamines. J Physiol Sci. 2006;56:235-45.
- Minson J, Chalmers J, Kapoor V, Cain M, Caon A. Relative importance of sympathetic nerves and of circulating adrenaline and vasopressin in mediating

- hypertension after lesions of the caudal ventrolateral medulla in the rat. J Hypertens, 1986;4:273-81.
- Kamiya A, Kawada T, Yamamoto K, Michikami D, Ariumi H, Miyamoto T, et al. Muscle sympathetic nerve activity averaged over 1 minute parallels renal and cardiac sympathetic nerve activity in response to a forced baroreceptor pressure change. Circulation. 2005;112:384-6.
- Ichinose M, Saito M, Ogawa T, Hayashi K, Kondo N, Nishiyasu T. Modulation of control of muscle sympathetic nerve activity during orthostatic stress in humans. Am J Physiol Heart Circ Physiol. 2004;287:H2147-53.
- Ichinose M, Saito M, Kitano A, Hayashi K, Kondo N, Nishiyasu T. Modulation of arterial baroreflex dynamic response during mild orthostatic stress in humans. J Physiol. 2004;557:321-30.