初診時視力右(1.2)、左(1.2)。両眼前眼部炎症と硝子体混濁がみられた。蛍光眼底造影では両眼網膜血管からシダ状の蛍光漏出がみられた。その後も眼炎症発作を繰り返し、口腔内アフタ性潰瘍とあわせてベーチェット病と診断された。著明な視力低下を伴う眼炎症発作を繰り返すためコルヒチン内服を開始したが、その後も頻発する眼炎症発作とともに腸管ベーチェット病を発症。インフリキシマブ治療を開始した。その後は大きな眼炎症発作はなく経過し、眼外症状も落ちついている。

D.E. 考察と結論

小児ベーチェット病2症例における眼炎症 所見は成人発症例とほぼ同じであり、視力低下 をきたす重篤なものであった。陰部潰瘍はみら れなかった。インフリキシマブ治療が発作抑制 に有効であった。今後、副作用の発現や効果の 減弱など長期的な有用性についてはさらに経 過観察が必要である。

F. 健康危険情報 特記事項なし。

G. 研究発表

- 1. 論文発表 なし
- ,2. 学会発表 なし
- H. 知的財産権の出願、登録状況 なし

図1. 症例1経過

症例1 経過

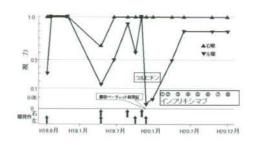
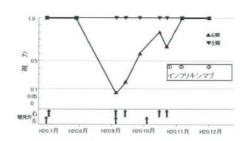


図2. 症例2経過

症例2 経過



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The role of streptococcal hypersensitivity in the pathogenesis of Behçet's Disease

Behcet's disease (BD) is still considered as a mysterious multisystemic disorder characterized by recurrent involvement of muco-cutaneous, ocular, intestinal, vascular and/or nervous system organs. In this review, we would like to highlight and discuss several important advances in our understanding of the pathogenesis of BD based on the intrinsic genetic factors including HLA-B51 and MICA expression and extrinsic triggering factors. As one of the extrinsic triggering factors, we focused on the hypersensitivity against oral streptococci which might be acquired through the innate immune mechanism. It was found that HLA-B51 restricted CD8 T cell response was clearly correlated with the target tissues expressing MICA*009 by stress in active BD patients with HLA-B51 as the intrinsic factors. Bes-1 gene and HSP-65 derived from oral S. sanguinis, which is the uncommon serotype (KTH-1, strain BD113-20), are supposed to play important roles as an extrinsic factor in BD pathogenesis. The peptides of the Bes-1 gene are highly homologous with the retinal protein Brn3b and moreover, the Bes-1 peptides were homologous with HSP-65 derived from microorganisms in association with the counterpart human HSP-60, which appeared reactively in the patients. HSP-65/60 also has high homologies with the respective T cell epitope of BD patients. Although HSP-65/60 and the peptides of Bes-1 gene were found to stimulate PBMCs from BD patients in the production of pro-inflammatory Th1 type cytokines, some homologous peptides of HSP-65 with T cell epitopes were found to reduce IL-8, IL-12 and TNF-α produced from PBMCs of active BD patients. The findings might be correlated with the clinically therapeutic effects for BD patients with severe uveitis, who were led to immunotolerance by the peptide of human HSP-60 (336-351), as previously reported. Then, the pathogenesis of BD was discussed referring to intrinsic genetic factors and extrinsic triggering factors in aspects of streptococcal hypersensitivity, which might be acquired through the innate immune mechanisms. The BD symptoms were thought to be due to vascular reactions as immune responses in correlation with monocyte expressed streptococcal agents.

Key words: Behçet's disease, Streptococcus sanguinis, heat shock protein, Bes-1 gene, vascular reaction

Behçet's disease (BD) [1] or Adamantiades-Behçet's disease [2] is a chronic multisystemic inflammatory disorder characterized by the recurrent involvement of muco-cutaneous (oral and genital ulceration, erythema nodosum (EN)-like eruption, acne-like eruption, etc.), ocular, vascular, digestive and/or nervous system organs. Although the actual etiology of BD is still unclear, the pathogenesis has become generally clearer by investigation of the epidemiology, clinical manifestations and basic etiological research based on the intrinsic genetic and extrinsic triggering factors and the immunological findings [3-13]. The genetic predisposition is included as one of the intrinsic factors, because more than 60% of BD patients are associated with HLA-B51 [3-6]. As one of the

extrinsic triggering factors, an unhygienic oral condition may be suspected, because periodontitis, decayed teeth, chronic tonsillitis, etc. are frequently noted in the oral cavity of BD patients [9, 10]. The proportion of Streptococcus sanguinis (S. sanguinis), which was previously recognized as a species of the genus Streptococcus named "S. sanguis" [11-13], was significantly higher in the oral bacterial flora of BD patients than those of healthy and disease controls (table 1). The S. sanguis isolated from BD patients was different from reference ATCC strains in DNA homology and sugar constituents. Here, we call these clinical isolates S. sanguinis, although they may include not only S. sanguinis but also S. oralis, because the chemiluminescence of neutrophils obtained from BD patients was in-

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creased in correlation with the proportion of S. sanguinis in the oral flora. The strains isolated as S. sanguinis were serologically different from various strain types from healthy controls, but the DNA-DNA homology was shown as S. oralis and S. sanguinis-like species [12, 13]. Then, S. sanguinis was identified as an uncommon serotype KTH-1 (so-called BD113-20 strain) by its bacterial and enzymatic properties [12, 13] Most patients tend to acquire delayed type hypersensitivity against streptococci in their oral flora, and as previously demonstrated, much stronger cutaneous reactions than the general pathergy test when intracutaneous injections and/or prickle tests were carried out using various killed bacteria and/or their cell wall antigens, including streptococci, enterococci, staphylococci, etc. (table 2) [9, 10, 14-16]. In vitro experiments, IL-6 and INF-γ were significantly produced from PBMCs of BD patients in stimulation by KTH-1 antigens [17]. The serum-antibody titers against streptococci were also elevated in BD patients [18]. The 65kDa of a heat shock protein (HSP-65) derived from oral bacteria, including S. sanguinis, can be detected along with counterpart human HSP-60 which appears reactively in the sera and lesions of BD patients. The peptides of HSP-65 derived from the bacteria show considerable homology with those of the human HSP-60 [19-21].

Although Sakane et al. [22] have reviewed the general clinical manifestations and pathogenesis of BD, based on data since 1972 from the Research Committee for Behçet's Disease organized by Japanese Ministry of Health, Labour and Welfare, our Research Committee, newly organized in 2001, revised the diagnostic criteria established in 1987. In the new criteria, in 2003, we included hypersensitivity skin reactions against streptococci in the diagnosis as one of the references and the levels of disease severity of BD patients, as introduced by Suzuki et al. [23]. Hence, we would mainly like to discuss the role of abnormally hypersensitive immune reactions against oral streptococci as one of the extrinsic triggering factors in connection with the intrinsic factors in the pathogenesis of BD.

HLA genotyping of BD and streptococcal infection

HLA-B51 is supposed to be a highly associated genetic marker of BD patients from many different ethnic groups, including European, Mediterranean and Asian people [3, 5, 24, 25] and

Abbreviations: APCs: antigen presenting cells; BD: Behçet's disease; CTLs: cytotoxic T lymphocytes; DNA: deoxyribonucleic acid; EN: erythema nodosum; HHV: human herpes virus; HIs: healthy individuals; HLA: human leukocyte antigen; HSP-65: 65 kDa of heat shock protein; HSV: herpes simplex virus; IL: interleukin; IFN-γ: interferon-γ; MBL: mannose-binding lectin; MICA: major histocompatibility complex class I chain-related gene A; mRNA: messenger ribonucleic acid; NK cell: natural killer cell; PBMCs: peripheral blood mononuclear cells; PCR: polymerase chain reaction; rCTB: recombinant cholera toxin B subunit; sIL-2R: soluble IL-2 receptor; S. sanguinis: Streptococcus sanguinis; Th1 cell: T helper I cell; TNF-α: tumor necrosis factor-α

BD has several unique epidemiological features which seem to go from Southern Europe to Japan along "the old Silk Route" [3, 5, 6, 25]. Although the HLA-B51 phenotype is important as an intrinsic factor for BD patients, and HLA-B51-transgenic mice show enhanced neutrophil function, because the HLA-B51 gene presents endogenous peptides to CD8 T cells (cytotoxic T lymphocytes; CTLs), these mice do not express BD symptoms [24]. The appearance of BD lesions is not considered to be directly correlated with HLA-B51 in the immunological background of patients, but it was recently found that HLA-B51-restricted CTLs played some roles in BD pathogenesis in correlation with the stressed target tissues expressing major histocompatibility complex class I-related gene A (MICA) [26, 27]. When the transmembrane-MICA located nearly at the HLA-B51 gene is preferentially expressed on epithelial and endothelial cells by stress, they seem to be candidates for the HLA-B51-restricted CTLs response [27]. These findings are based on the following; in HLA-B51 positive BD patients in the acute phase, MICA-transmembrane peptides derived from the amino acid sequence of MICA*009, which is in strong linkage disequilibrium with HLA-B51 [28], were significantly detected as targets of T cell responses. MICA expression was lost after the BD-related symptoms disappeared and the MICA-induced T cell response was also inhibited by anti-HLA class I antibodies and by CD8 T cell depletion. MICA expressed on the stressed epithelium and endothelium are considered to be the ligand for activating natural killer (NK) cells with the NKG2D molecule and CD8 T cells as CTLs [27]. BD lesional reactions might be accelerated by inflammatory cytokines and chemokines secreted from CTLs and NK cells [29, 30]. Regarding NK cell activation, inhibitory CD34/NKG2A and activating CD94/NKG2C molecules are alternatively expressed on NK, CD4 +CD8 +T cells. indicating an imbalance in cytotoxic activity in BD patients [31], although the function of NK cells is supposed to be down-regulated in the active stage and to be up-regulated in the remission stage of BD patients [32].

It is considered that the HSP-65/60 derived from microorganisms including S. sanguinis and from human tissues, which is detected in the oral mucosal and skin lesions of BD patients [19, 20], also becomes a stress-inducible factor in connection with MICA*009 expression. Generally, antigen presenting cells (APCs), which produce IL-12 in correlation with Th1 type immune-reactions, are thought to be activated in BD patients with HLA-B51 in the active stage. as indicated by Yasuoka et al. [27]. However, we have recently obtained interesting results that PBMCs from BD patients without the HLA-B51 gene can be significantly stimulated by S. sanguinis antigen in the expression of IL-12p40 mRNA and in the increasing of protein levels in connection with IL-12p70 (70 kDa composed of p35 and p40 subunits), compared to those of patients with HLA-B51 [33]. It has been suggested that the antibacterial host response in T type immunity mediated by IL-12 is much stronger in HLA-B51-negative BD patients, though the precise findings will be discussed later again.

Hypersensitivity against S. sanguinis

Generally, oral health is impaired in BD patients [9, 10, 12, 13], which seems to be associated with disease severity

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Table 1. Oral bacterial flora in BD patients, disease controls and healthy controls. Subjects: 22 BD patients with oral aphthous ulcerations (age-mean: 35.8) and 10 healthy controls and 8 disease controls including Vogt-Harada disease, sarcoidosis, herpes simplex infection, etc. who were similarly aged to the BD patients. Sampling: Supragingival plaque was taken from the lower first and second premolars or first molar after clinical examinations. Cotton swab specimens were obtained from the surface of the tongue dorsum and buccal mucosa. Each sample was incubated on TYC agar with 5% sucrose and Mitis-Salivarius agar for Streptococcus species at 37 °C for 2 days. For identification of streptococcus species, API-STREP system and confirming tests were used and the total viable count was calculated as percentage, as described by Isogai et al. [13]. The proportion of S. sanguinis (S. sanguis) was significantly higher in the oral bacterial flora of BD patients than controls

Bacteria	% prominent flora (mean ± SE) of plaque from				
	Patients with BD (n = 22)	Healthy controls (n = 10)	Disease controls (n = 8)		
Gram-positive bacteria	66.7 ± 3.6	69.0 ± 6.2	56.0 ± 6.2		
Streptococcus	53.3±4.1****	48.1 ± 4.6	39.0 ± 5.7		
S. sanguis	26.7±3.7*,***	9.4 ± 0.6	7.5 ± 2.3		
S. salivarius	7.4 ± 1.4	6.6 ± 2.4	6.0 ± 1.9		
S. mitis	$14.9 \pm 2.1 \ (**), \ (****)$	25.9 ± 4.5	24.0 ± 3.6		
S. mutans	4.1 ± 1.1*,***	0.2 ± 0.1	1.5 ± 0.6		
Other streptococci	$0.2 \pm 0.1(*)$	6.0 ± 2.5	< 0.1		
Enterococcus	0.25 ± 0.11	0.01 ± 0.01	0.02 ± 0.01		
Staphylococcus	0.26 ± 0.19	0.02 ± 0.01	< 0.001		
Lactobacillus	1.6 ± 0.6	0.38 ± 0.02	0.19 ± 0.11		
Eubacterium	0.36 ± 0.17	0.23 ± 0.28	0.17 ± 0.08		
Gram-positive bacteria	33.2 ± 3.5	29.2 ± 7.1	44.0 ± 6.2		
Bacteroides	16.5 ± 2.2*	6.4 ± 1.9	24.1 ± 5.1		
Black pigmented Bacteroides	3.1 ± 0.7**	0.9 ± 0.2	2.9 ± 1.4		
Fusobacterium	$2.6 \pm 0.6 (****)$	1.9 ± 0.7	9.3 ± 3.8		
Veillonella	3.3 ± 1.1	7.8 ± 3.1	3.1 ± 1.2		
Enterobacteriaceae	< 0.001	ND	ND		
Others	7.5 ± 1.1	6.8 ± 2.8	4.6 ± 1.3		
Molds	0.15 ± 0.08	ND	< 0.01		

^{*}Higher %, P < 0.01 vs. healthy controls. ** Higher %, P < 0.05 vs. healthy controls. ***Higher %, P < 0.01 vs. disease controls. ****Higher %, P < 0.05 vs. disease controls.

Table 2. The skin tests by bacterial antigens and saline for BD patients and healthy controls. Each 0.01 mL of the bacterial antigens (bacteria vaccines: $1 \times 10^9 \text{ org./mL}$. Hollister-Stier Lab., USA) was intracutaneously injected in 84 BD patients and 10 healthy controls. The reaction was observed at 15 min. and 48 hours after injection. The erythematous skin reactions by streptococcal antigens were significantly stronger than those by other antigens in BD patients [9, 10]

Bacterial vaccines	Behçet's	syndrome	Normal	controls
imetos apontentano	15 min	48 h	15 min	48 h
S. pyogenes	7 ± 8	41 ± 15	11 ± 11	8 ± 6
S. viridans	8 ± 7	46 ± 11	4 ± 4	2 ± 3
S. non-hemolyticus	7 ± 7	35 ± 14	5 ± 5	3 ± 4
S. faecalis	10 ± 8	40 ± 19	12 ± 11	0
Pneumococcus	16 ± 10	32 ± 14	8 ± 9	3 ± 4
E. coli	6 ± 7	16 ± 11	2 ± 1	1 ± 2
H. influenzae	11 ± 10	15 ± 15	9 ± 13	11 ± 12
Sta. aureus	8 ± 7	12 ± 13	3 ± 7	0 ± 1
Sta. epidemidis	5 ± 7	6 ± 7	2 ± 7	1 ± 2
Prot. vulgaris	10 ± 16	28 ± 13	6 ± 6	17 ± 4
Pseud. aeruginosa	1 ± 1	22 ± 11	2 ± 3	5 ± 0
SK-SD (50 U/mL)		9 ± 12		20 ± 9
Saline		2 ± 4		0 ± 1

The numbers denote mean ± SD(mm) of Length + width/2 erythemas.

disease controls. (***) lower %, P < 0.01 vs. healthy controls. (***) lower %, P < 0.05 vs. healthy controls. (***) lower %, P < 0.01 vs. disease controls. (****) Lower %, P < 0.05 vs. disease controls. ND: not detected.

[34]. It is not clear that the predisposition of the patients is correlated with streptococcal infection, but the uncommon oral S. sanguinis serotypes are significantly increased in BD patients compared with healthy and disease controls, as previously described [12, 13]. The antibodies against S. sanguinis in sera from BD patients showed cross reactivity with some synthetic peptides of HSP-65 derived from S. sanguinis [35, 36]. The patients show strong delayed type cutaneous hypersensitivity reactions against streptococcal antigens in skin tests and sometimes BD symptoms were provoked by skin injection of the antigens [9, 10, 14-16]. Because aphthous ulceration can be also induced by a prick with streptococcal antigens on the oral mucous membrane of a BD patient [10], the appearance of aphthous ulceration is considered to be based on a hypersensitive reaction against S. sanguinis, which may penetrate traumatically into the oral membrane of BD patients. Isogai et al. [36] demonstrated that symptoms mimicking BD appeared in germ-free mice when S. sanguinis from BD patients was inoculated into their oral tissue which was damaged by heat shock and/or mechanical stress. This report suggests that immunization with S. sanguinis through the oral membrane route elicits BD-like symptoms in the animal model, as is seen in BD patients who carry S. sanguinis as the pathogenic microorganism in their oral cavity. We tried to find PCR targeting Bes-1 gene in BD lesions using 2 distinct primer sets (peptides, 229-243 and 373-385) encoding S. sanguinis (serotype KTH-1), which was prepared by Yoshikawa et al. [37]. Bes-1 DNA was present in various muco-cutaneous lesions including oral and genital ulcerations and EN-like lesions and the PCR-in situ hybridization revealed that Bes-1 DNA was expressed in the cytoplasm of inflammatory infiltrated monocytes adhering the vascular walls in muco-cutaneous lesions (figure 1A and B) [38]. These infiltrated monocytes may express streptococcal antigens on the cell membrane because they were detected by immunofluorescence with anti-streptococcal antibodies, as previously reported (figure 2 A-C) [10, 15]. In contrast, we failed to detect the DNA of HSV-1, HSV-2, cytomegalovirus, HHV-6 and HHV-7 in the lesions by PCR [39], although HSV infection has been speculated as etiologically important since the report of H. Behçet [1]. However, animal models infected by HSV have been also demonstrated to mimic BD like symptoms [40]. Interestingly, the amino acid sequence of the peptides of Bes-1 (229-243) and 373-385) shows more than 60% similarity to the human intraocular ganglion peptide, Brn-3b which is a subfamily of POU (pit-Oct Unc) domain factors containing Brn-3a and Brn-3c [41]. The peptide of Bes-1 (229-243) was also found to be correlated with the peptide of HSP-60 (336-351) [35]. Recently it has been found that the peptide of Bes-1 (337-385) stimulated the production of IFN-y and IL-12 from PBMCs of BD patients, although cellular proliferation was not observed [42]. These results suggest that Bes-1 derived from oral S. sanguinis might be an inducer for the retinal and neural involvement possible in BD patients.

HSP-65 derived from microorganisms and human HSP-60

HSPs, which scavenge denatured intracellular proteins, are supposed to be induced by microorganisms and mamma-

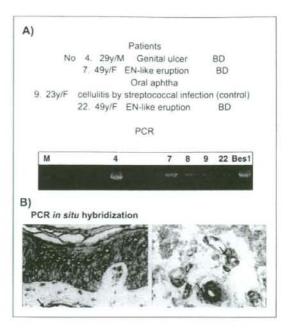


Figure 1. Bes-1 gene expression in the muco-cutaneous lesions of patients with Behçet's disease (BD) [38]. A) Three of 11 BD patients were positive for Bes-1 DNA in the lesions including aphthous and genital ulcerations and erythema nodosum (EN)-like eruption by amplified polymerase chain reaction (PCR) using the primers: Bes-1-1 (5'-TAATAACCCTGACCAAGCCTA-3') and Bes-1-2 (5'-CCCTTTCAAAAGTCATAAATC-3') encoding S. sanguinis. B) In these positive lesions, Bes-1 DNA was also detected in the cytoplasm of monocytes adhering to the vascular walls and infiltrated around the vessels by PCR in situ hybridization.

lian tissues under a variety of stressful conditions [43] and they may be involved in the pathogenesis of some autoimmune diseases [44]. In BD patients, the serum levels of IgA antibodies to mycobacterial HSP-65, which cross-reacts with selected strains of S. sanguinis, are increased significantly [45, 46]. HSPs taken up by APCs are thought to stimulate T cells directly, the monocytes expressing HSP-60 led T cells to undergo apoptosis after IFN-y production [47] and the presence of HSP-60 was also detected in various lesions of BD patients [46, 48, 49]. On the other hand, 4 peptides of HSP-65 (111-125, 154-172, 219-233 and 311-326) derived from S. sanguinis were recognized as immuno-dominant agents for T cell and B cell responses and they showed 50-80% homology to the counterpart human HSP-60, as shown in figure 3 [20, 48-50]. The 4 peptides of HSP-65 were shown to significantly stimulate and undergo CD4 and CD8 T cell apoptosis in PBMCs from BD patients and HSP-60 also seemed to stimulate them [46. 47]. On the contrary, the other two peptides of HSP-65 (21-35 and 401-415) corresponding to the peptide of human HSP-60 (425-441), are reported not to stimulate PB-MCs from BD patients and healthy individuals (HIs) [43]. The peptide of HSP-60 (336-351) was also identified to be highly homologous to the T cell epitope [43, 45-52]. Whole

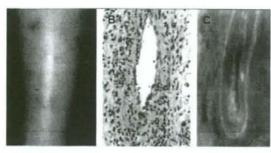


Figure 2. The vascular findings of EN-like eruption of a BD patient. A) A clinical finding of EN-like eruptions of a BD patient. B) A magnified histological view of the vascular reaction with inflammatory cells including monocytes and a few neutrophils in the EN-like eruption (Hematoxylin-eosin stain, × 400). C) An immunofluorescent finding of monocytes adhering to the vascular wall by anti-streptococcal group D antibody (Difco Co., USA). The finding suggests monocytes expressed streptococcal antigen at the vascular wall [10, 15]. Photos: Fukushima Medical University School of Medicine, Department of Dermatology.

HSP-60 is, however, suspected to increase vascular endothelial growth factor (VEGF) which activates, impairs and proliferates vascular endothelial cells [53] and which may lead to thrombophebitis and vasculitis, by damaging endothelial cells in BD patients. Although the term of "vasculitis" has been frequently used in BD lesions, Jorrizo et al. [54] previously reported that the real vasculitis, exhibiting "necrotizing vasculitis", was rarely seen in the EN-like eruptions of BD patients, and in most cases, the vascular reaction is surrounded by monocytes and a few neutrophils, as we demonstrated in figure 2B. This is so-called "lymphocytic vasculitis" seen histologically, as recently described by other authors [55, 56]. It is observed, however, that the serum levels of soluble(s) adhesion molecules, such as s-selectins and s-intercellular adhesion molecule-1, are elevated [57, 58] and also the expression of VEGF is increased in the presence of HSP in the lesions of BD [59].

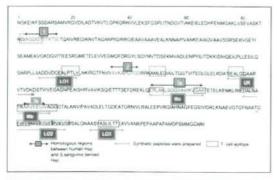


Figure 3. Newly synthesized peptides of HSP-65 derived from Streptococcus. sanguinis (S. sanguinis) (KTH-1, BD113-20 strain). There are 10 peptides homologous with T cell epitopes including 1, UK, LO1, 3a(IIIa), 3b(IIIb), LO2 and LO3 and they are also highly homologous with human HSP-60 peptides. The peptide of UK(311-326) corresponds to the peptide of human HSP-60 (336-351). (Lin and Oguma, 2003).

The subcutaneous administration of HSP peptides to mice has been shown to induce uveitis with vascular impairment [60].

On the other hand, it is of interest that the HSP-60 peptide (336-351), linked to the recombinant cholera toxin B subunit (rCTB), reduced the uveitis induced by whole HSP-60. although the peptide without adjuvant is reported to induce uveitis in Lewis strain rats [60, 61]. Recently, a therapeutic trial with the peptide conjugated with rCTB was given orally to BD patients with recurrent uveitis and successful results were obtained, as 5 of 8 patients had no relapse of the uveitis, no side-effects were present and 2 of the remaining 3 patients had improved recurrent oral ulceration. folliculitis, EN-like eruptions, and genital ulcers [62]. In those patients with control of uveitis and extra-articular manifestations, a lack of the peptide-specific CD4 T cell population, a decrease in expression of Th1 type cells (CCR5, CXCR3) and a reduction of IFN-γ, TNF-α, CCR7 T cells and co-stimulatory molecules (CD40 and CD28) were described in comparison to BD patients with relapse of disease [62]. These findings may suggest immunotoleration in active BD patients. It is hypothesized that CTLs play a role in BD pathogenesis by targeting a self antigen selectively expressed in the affected tissues. In BD patients with active disease, the endogenously generated MICA transmembrane-peptide by autoreactive CTLs is present [27] and the excessive inflammatory responses might be induced by extrinsic factors correlated with S. sanguinis and other organisms, including Helicobactor pylori, mycoplasma fermentas, etc. [63-65]. Neutrophilic hyperfunction and a cross-reactive autoimmune response between microbial and human HSPs are proposed to be correlated with the hyperreactivity against microorganisms, including S. sanguinis, seen in BD patients [9, 10, 14-17]. These HSPs presented by APCs can directly stimulate αβT and γδ T cells, which play important roles in oral mucosal immunity as the first defense against microorganisms. It is thought that $V\gamma 9\delta 2^+$ T cells, a major subset of $\gamma \delta$ T cells in PBMCs, recognize antigens produced by bacteria and that innate and adaptive immune responses are influenced by secreting IFN-y, towards a Th1 profile [20, 66, 67]. These γδ T cells seemed to be elevated in PBMCs and in the muco-cutaneous lesions of BD patients [47, 66]. The second major subset, γδ1+ T cell, is enriched in the mucosa and the antigens are presented by APCs with stressinducible MICA and MICB. The γδ T cells, which highly express CD29 and CD69, produce IFN-γ and TNF-α from stimulation by HSP-65/60 in the peripheral blood and in the lesions of BD patients with active disease [20, 47, 66]. These activated APCs and γδ T cells might activate αβ T cells by their secretion of sIL-2R, IFN-γ, TNF-α and also high levels of other cytokines, IL-1α, IL-6, IL-8, IL-15, etc., which are detected in the sera of BD patients [67-69]. In the active stage of BD patients, IL-12 is also produced as a sign of an advanced Th1 type reaction. The gene polymorphism in the promoter region regarding a 4 bp insertion within IL-12p40 was significantly higher in HLA-B51 negative BD patients than HLA-B51 positive patients and HIs. The expression of IL-12p40 mRNA and protein levels in conjugation with IL-12p70 induction were also significantly increased in PBMCs from BD patients without HLA-B51 by stimulation with S. sanguinis antigen, as previously described [33]. It has been recently found that expression of IL-23, which is composed of a shared p40

subunit of IL-12 and p19 subunit of IL-23, was also increased with IL-12 in EN-like lesions of BD patients [70].

HSPs and BD pathogenesis

Although antibodies against the HSP peptides derived from bacteria including S. sanguinis are found in sera of BD patients [35, 36], HSP specific antibodies and T cells are considered to play a complicated role in the pathogenesis of human autoimmune diseases [71]. HSPs might trigger both innate and adaptive immune mechanisms in BD. On the other hand, the therapeutic approaches involving HSP immunomodulation may be available as "oral toleration" using the peptide of HSP (336-351) linked to rCTB for BD patients with advanced uveitis, as demonstrated by Stanford et al. [62]. Then, we tried to analyze HSP-65 derived from S. sanguinis to find homologous peptides to T cell epitopes of BD patients, and some peptides were found to be highly homologous with T cell epitopes in the correlation with human HSP-60, as indicated in figure 3. Attempts have been made to find out how the newly synthesized homologous peptides influence proinflammatory cytokine production from PBMCs of active BD patients. The peptides, LO1 (249-264), IIIa (365-384), IIIb (395-413), LO2 (480-499), LO3 (504-518) and UK (311-326), corresponding to the peptide of human HSP-60 (336-351), were applied to lead immuno-toleration for activated CTLs of BD patients in vitro. PBMCs from 7 active BD patients and 5 HIs were incubated with and without these peptides and 7 days after incubation IL-8, IL-12, IFN-γ and TNF-α were measured and compared with those from PBMCs of active BD patients incubated without the peptides as controls. Although IL-12 and IL-8 were actively produced from PBMCs in active BD patients, even though they were not stimulated, a significant reduction of inflammatory cytokines was found by some kinds of the peptides. The 5 peptides, LO1, LO2, LO3, IIIb and UK significantly reduced IL-12 production and also LO1, IIIa and IIIb significantly inhibited IL-8 production, except for LO2 and LO3 (figure 4A and B). On the other hand, the cytokines from PBMCs of HIs were significantly increased on stimulation by the peptides. In order to understand the suppressive mechanisms of the cytokine production in PBMCs from active BD patients, we tried to find the binding sites of the peptides on monocytes by cDNA chips (Gene Chip; Human Genome) using NOMO-1 cells (human macrophage cell line) activated by S. sanguinis antigen and they were incubated with the peptides. It was found that although the expression of IL-8, IL-16, IL-13R and IL-17R was decreased after incubation with LO1 and UK, respectively, LO2 did not decrease IL-8 production. CD58 (lymphocyte function-associated antigen-3) molecule and/or FK506 binding protein were highly expressed on the cell membrane by LO1 and UK [72]. It is considered that activated CTLs of BD patients might lead to apoptosis and/or dysfunction of lymphoid cells by the binding of LO1 and UK on the cell-receptors.

Toll-like receptor (TLR) expression in innate immunity

Regarding the recognition system for microorganism antigens in humans, 10 members of the TLR family are supposed to act as innate immune receptors by binding particular structures present on bacteria, viruses, fungi, etc. [73]. Although, generally, TLRs are weakly detectable in various human tissues with varying levels, the TLR expression of the organs involved in immune responses and exposed to the environment, is found to be significantly stronger [74]. Our BD Research Group have already found the expressions of TLR-2 (recognize: bacterial lipoprotein, zymosan, lipopolysaccharide (LPS), lipoteicholic acid of microbial antigen, etc.) and TLR-4 (LPS, HSP-65/60, etc.) on PB-MCs and their presence has also been recognized in intestinal lesions by immunohistology (not yet published in the English literature) [75]. TLR-3 [ds RNA] and TLR-6 (mycoplasma, staphylococci, etc.) are also reported to have enhanced expression on the neutrophils and monocytes of BD patients, when stimulated by HSP-60 and S. sanguinis antigens [76]. In oral ulcer lesions, expression of TLR-9 (unmethylated CpG DNA, bacteria and virus) has recently been found [77]. These findings suggest that the innate immune system contributes to the acquisition of hypersensitivity against oral S. sanguinis as the extrinsic factor in the pathogenesis of BD.

Complement system in innate immunity

It is generally accepted that the compliment system is accelerated in relation to chemokine and neutrophilic activation [78, 79]. In BD lesions, deposits of complement C3 with immunoglobulins are frequently detectable by immunofluorescent techniques [15, 56]. With respect to the complement system of BD patients, the titer of serum complement is generally high in the inactive stage but decreases in the active stage, although levels of the mannose-binding lectin (MBL) pathway of complement is reported to be decreased [80]. The MBL pathway is considered to play an important role in innate immunity. It is thought to be a C-type serum lectin secreted by the liver, which binds to mannose and N-acetyl-glucosamine oligosaccharides on the surfaces of yeast, bacteria and viruses [81]. The reaction serves as the initiator of the third pathway of the complement system, independently from antibodies. Ficolin (FCN) is a soluble protein that binds to carbohydrate on the microbial cells and 3 different types of FCN are detected. FCN 1 and 2 genes are located in chromosome 9q34 and the FCN 3 gene is assigned to chromosome 1. FCN 2 binds to lipoteicholic acid on the cell wall constituent in all Gram-positive bacteria and activate immune cells, to produce proinflammatory cytokines [78, 82]. Recently, we have found that novel FCN 2 gene single nucleotide polymorphisms (SNPs) were identified in the promoter regions as well as in the exon regions. The MBL genetic polymorphisms might be involved in immune responses to streptococcus infections in BD patients, because a relationship between MBL gene mutations and microbiological factors is suspected in the lesional immune reaction of BD patients [83]. Although a significant difference was not present in the genotype allele frequencies of MBL gene SNPs between BD patients and HIs, the allele frequencies of FCN2 gene SNPs were significantly recognized in the promoter regions (-557 and -64 sites) among HLA-B51 positive BD patients [84]. The findings suggest the possibility that the FCN gene of the MBL pathway in the complement system contributes to innate immununity in BD patients with the HLA-B51 haplotype.

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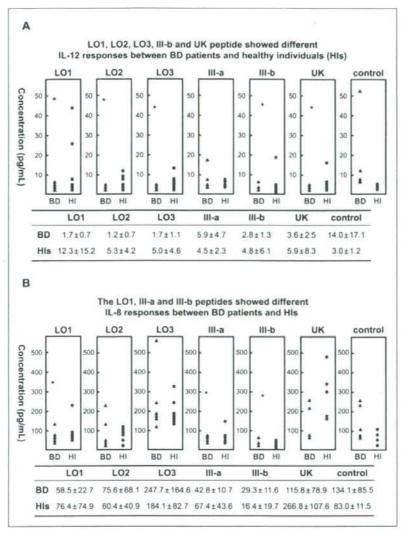


Figure 4. Effects of the peptides of T cell epitope as shown in *figure 3*. A) IL-12 was highly produced by PBMCs from active BD patients without stimulation (Control). IL-12 production from BD patients was significantly reduced by incubation with LO1, LO2, LO3, III-b and UK (*). On the other hand, IL-12 production from PBMCs of HIs was elevated by these peptides. B) LO1, III-a and III-b significantly reduced IL-8 production from PBMCs of BD patients (*). On the other hand, IL-8 production was accelerated from PBMCs of HIs by these peptides.

How do the muco-cutaneous symptoms appear in BD patients?

BD symptoms are characterized by vascular involvements, histologically showing swollen endothelial cells of the micro-arteries, infiltrated by inflammatory monocytes and a few neutrophils, a so-called "vascular reaction" seen in EN-like eruptions (figure 2B) and other lesions [15, 54-56]. The strong hypersensitivity reaction against S. sanguinis agents [9, 10, 14-17] which might be caused by APCs through the innate immune mechanism, can be suspected as the extrinsic triggering factor in the pathogenesis of BD. In the treatment by antibiotics for the involvement of oral

S. sanguinis, minocycline, which reduces not only the growth of streptococci but also suppresses IL-1β and IL-6 production from inflamed T cells, was especially clinically effective for aphthous ulceration, acne-like eruption and EN-like lesions in BD patients [10]. Other studies also showed that combination therapy, colchicine and benzathine penicillin, were effective to suppress BD symptoms, compared to colchicine monotherapy [85, 86]. Muncu et al. [86] and others [6-8] have already reviewed the role of infectious agents in the pathogenesis of BD, but we also dare to propose the hypothesis that after the Bes-1 gene is taken into the cytoplasm of APCs (figure 1A and B) through the TLRs in the oral cavity, the APCs, which are expressing

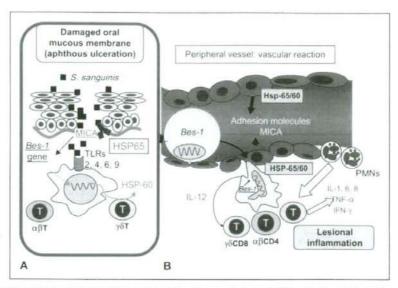


Figure 5. Hypothesis of the mechanisms in the appearance of various lesions of BD patients. A) The antigen presenting cells (APCs) (macrophages and/or dendritic cells) immunized by S. sanguinis agents through TLRs in the oral cavity might be carried to the peripheral regions. B) If the APCs in the blood flow adhered to the impaired and/or MICA and adhesion molecules expressed endotherial cells of vascular walls, the immunological reaction might appear as a BD lesion.

the streptococcal antigen as seen in figure 2C, produce HSP-65. If these APCs are carried in the blood flow to the impaired and/or MICA expressed endothelium of the vessels in correlation with HSP-65/60, VEGF, adhesion molecules, etc., BD lesions might be induced with "vascular reaction" and/or "lymphocytic vasculitis" as the immunological reaction by the APCs expressing the S. sanguinis antigen (figure 5).

Conclusion

The pathogenesis of BD was discussed, including aspects of the intrinsic genetic factors and with oral bacteria antigens as one of the extrinsic triggering factors. HLA-B51 restricted CTL was found to target the MICA expressed organs by stress in correlation with HSP-65/60 derived from oral bacteria, including S. sanguinis. The immune responses which are based on a Th1 type reaction with chemotaxis to the bacterial agents are considered to correlate with various BD symptoms, histologically exhibiting "vascular reaction" and/or "lymphocytic vasculitis".

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