Table 3. Clinical summary of MADD patients.

Patient	Age	Gender	Clinical feature	Serum CK (IU/L)	CoQ <sub>10</sub> level (μg/g muscle)	Treatment	Clinical course
4	5 mo	F	Muscle weakness, hepatomegaly	55	NA	NA	NA
5	6 mo	М	Muscle weakness, hepatomegaly	2000-4000	NA	L-carnitine riboflavin	Normal development after treatment
6	11 mo	M	Vomiting, hypertrophic cardiomyopathy	128-618	24.1	L-carnitine	Died at 2 years of age due to pulmonary alveolar bleeding
7	13 y 4 mo	F	Progressive muscle weakness	127	32.3	L-carnitine riboflavin	No muscle weakness at present

NA, not available. Normal range: CK, 57-197 IU/L; CoQ10, 32.1 ± 6.8 (mean ± standard deviation).

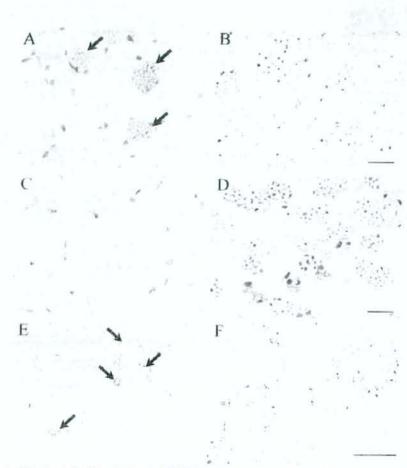


FIGURE 4. Routine histochemical staining of patients with LSM. Ragged-red-like fibers are seen on modified Gomori trichrome staining (A) and numerous lipid droplets are seen with oil-red-O (B) in a PCD patient, but not in an MADD patient (C, D). Rimmed vacuoles are seen in myofibers of patients with NLSDM (E), in addition to the characteristic numerous lipid droplets predominantly in type 1 fibers (F). Bar = 20 μm

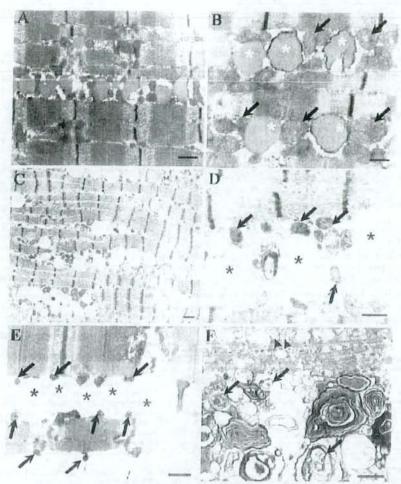


FIGURE 5. Electron microscopy findings. In a patient with PCD, markedly increased lipid droplets and increased numbers of mitochondria are seen (A). On higher magnification, lipid droplets (asterisks) are seen next to mitochondria (arrows) (B). In a patient with MADD, note the increase in the number of lipid droplets (C) and the proximity of these droplets (asterisks) with mitochondria (arrows) (D), findings similar to those for PCD. In a patient with NLSDM, there is also a marked increase in the amount of lipid (asterisks) deposited within the myofibers; however, note that mitochondria appear pyknotic (arrows) (E). In this patient, numerous autophagic vacuoles (arrows) in close proximity to lipid deposition (arrowheads) are also observed (F). Bar 1 μm in (A), (C), and (F). Bar 0.5 μm in (B), (D), and (E).

are necessary to fully understand the mechanism of the disease.

CPT II deficiency has been reported to show increased lipid droplets in muscle. However, in all patients whose samples were suitable for biochemical assays, normal enzymatic activity was seen. Furthermore, in our series, we had 7 patients with CPT II deficiency, but none showed lipid droplet accumulation on muscle pathology (data not shown)

This suggests that lipid storage may not be a common pathological feature of CPT II deficiency, although analysis of a larger population of patients with CPT II deficiency would be needed to further support this contention.

In spite of an extensive genetic survey of known causative genes for LSM, we did not find mutations in 76% of the patients. One possible explanation for the absence of mutations in these patients is that

they may have secondary LSM, as intramuscular lipid content is known to be secondarily increased under a variety of conditions, including diabetes, renal disease, iatrogenic conditions, gastrointestinal disturbance, elevated plasma fatty acid levels, and high dietary fat intake. 18 The fact that 2 patients were taking antiepileptic drugs could be supportive of this notion. In the majority of patients without mutation. conditions associated with a secondary increase in muscle lipids were not seen. In addition, 2 patients had pathological features that differed from the rest of the patients in the form of lipid droplets almost exclusively in type 2 fibers, in contrast to the preference of lipid accumulation in type 1 fibers in all others, indicating the probability of a common pathomechanism, at least in these 2 cases.

Among the 28 patients without mutations in known causative genes, 5 had a positive family history and/or consanguinity (Table 1), suggesting that these individuals are likely to have primary genetic lipid disorders, rather than secondary LSM and the presence of additional yet-to-be-identified causative genes for LSM. Further analysis on biochemical analyses of accumulated metabolites and extended study of candidate genes involved in lipid metabolism will be helpful in the genetic diagnosis of LSM patients.

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## REFERENCES

- Akiyama M, Sakai K, Ogawa M, McMillan JR, Sawamura D, Shimizu H. Novel duplication mutation in the patatin domain of adipose triglyceride lipase (PNPLA2) in neutral lipid storage disease with severe myopathy. Muscle Nerve 2007;36:856– 859.
- Deufel T, Wieland OH. Sensitive assay of carnitine palmitoyl transferase activity in tissue homogenates with a modified spectrophotometric method for enzymatic carnitine determination. Clin Chim Acta 1983;135:247–251.
- Di Mauro S, Trevisan C, Hays A. Disorders of lipid metabolism in muscle. Muscle Nerve 1980;3:369–388.
- Fischer J, Lefvre C, Morava E, Mussini JM, Laforêt P, Negre-Salvayre A, et al. The gene encoding adipose triglyceride lipase (PNPLA2) is mutated in neutral lipid storage disease with myopathy. Nat Genet 2007;39:28–30.
- Folch JM. Lees M. Stanlex, GHS. A simple method for the isolation and purification of total lipids from animal tissues. J Biol Chem 1957;226:497–509.
- Freeman FE, Goodmann SI. Defects of electron transfer flavoprotein and electron transfer flavoprotein—ubiquinone ox-

- idoreductase: glutaric aciduria type Il. In: Scriver CR, Beaudet AL, Sly WS, Valle D, editors. The metabolic and molecular bases of inherited disease. New York: McGraw-Hill; 2001, p 2357–2365.
- Gempel K, Topaloglu H, Talim B, Schneiderat P, Schoser BG, Hans VH, et al. The myopathic form of coenzyme Q10 deficiency is caused by mutations in the electron-transferringflavoprotein dehydrogenase (ETFDH) gene. Brain 2007;130: 2037-2044.
- Haemmerle G, Zimmermann R, Strauss JG, Kratky D, Riederer M, Knipping G, et al. Hormone-sensitive lipase deficiency in mice causes diglyceride accumulation in adipose tissue, muscle, and testis. J Biol Chem 2002;277:4806–4815
- Lass A, Zimmermann R, Haemmerle G, Riederer M, Schoiswohl G, Schweiger M, et al. Adipose triglyceride lipase-mediated lipolysis of cellular fat stores is activated by CGI-58 and defective in Chanarin-Dorfman syndrome. Cell Metab 2006; 3:309–319.
- Lefèvre C, Jobard F, Caux F, Bouadjar B, Karaduman A, Heilig R, et al. Mutations in CGI-58, the gene encoding a new protein of the esterase/lipase/thioesterase subfamily, in Chanarin-Dorfman syndrome. Am J Hum Genet 2001;69: 1002-1012.
- Massuishi T, Hirata K, Terasawa K, Kato H, Yoshino M, Ohtaki E, et al. Successful carnitine treatment in two siblings having lipid storage myopathy with hypertrophic cardiomyopathy. Neuropediatrics 1985;16:6–12.
- Nezu J, Tamai I, Oku A, Ohashi R, Yabuuchi H, Hashimoto N, et al. Primary systemic carnitine deficiency is caused by mutations in a gene encoding sodium ion-dependent carnitine transporter. Nat Genet 1999;21:91–94.
- Ohashi Y, Hasegawa Y, Murayama K, Ogawa M, Hasegawa T Kawai M, et al. A new diagnostic test for VLCAD deficiency using immunohistochemistry. Neurology 2004;22:62:2209–2213.
- Ohkuma A, Nonaka I, Malicdan MCV, Noguchi S, Nomura K, et al. Distal lipid storage myopathy due to PNLPA2 mutations. Neuromuscul Disord. 2008;18:671–674.
- Olsen RK, Olpin SE, Andresen BS, Miedzybrodzka ZH, Pourfarzam M, Merinero B, et al. ETFDH mutations as a major cause of riboflavin-responsive multiple acyl-CoA dehydrogenation deficiency. Brain 2007;130:2045–2054.
- Sambrook J, Russell DW, Molecular cloning: a laboratory manual. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press; 2001.
- Scaglia F, Longo N. Primary and secondary alterations of neonatal carnitine metabolism. Semin Perinatol 1999;23:152– 161.
- Schrauwen-Hinderling VB, Hesselink MK, Schrauwen P, Kooi ME. Intramyocellular lipid content in human skeletal muscle. Obesity 2006;14:357–367.
- Stockler S, Radner H, Karpf EF. Hauer A, Ebner F. Symmetric hypoplasia of the temporal cerebral lobes in an infant with glutaric aciduria type II (multiple acyl-coenzyme A dehydrogeniase deficiency). J Pediatr 1994;124:601–604.
- Takanashi J, Fujii K, Sugita K, Kohno Y. Neuroradiologic findings in glutaric aciduria type II. Pediatr Neurol 1999;20: 142–145.
- Vockley J, Whiteman DA. Defects of mitochondrial beta-oxidation: growing group of disorders. Neuromuscul Disord 2002;12:295–246.
- Wu X, Prasad PD, Leibach FH, Ganapathy V. cDNA sequence, transport function, and genomic organization of human OCTN2, a new member of the organic cation transporter family. Biochem Biophys Res Commun 1998;246589–595.
- Zimmermann R, Strauss JG, Haemmerle G, Schoiswohl G, Birner-Gruenberger R. Riederer M, et al. Fat mobilization in adipose tissue is promoted by adipose triglyceride lipase. Science 2004;306:1388–1386.