

が少ない結果が得られた⁸⁴⁾。さらに、Stage 1 大うつ病患者に対するリチウム治療の前後で DEX/CRH 負荷試験を行ったが、リチウムに対する反応者と非反応者で負荷後の血中コルチゾールや ACTH 濃度に差はみられなかった⁸⁵⁾。しかし、その後の研究で、Stage 1 大うつ病患者でリチウム併用前の DEX/CRH 試験における血中コルチゾール/ACTH 濃度比がリチウムに対する非反応者で反応者よりも高いことを報告した⁸⁶⁾。なお、DEX/CRH 試験における HPA 系の過大反応は抗うつ薬による改善と共に通常正常化するが^{69,70)}、リチウム併用後は、治療効果の有無にかかわらず HPA 系の反応はむしろより大きくなる(すなわちむしろ非抑制となる)点は、DEX/CRH 試験の実施上注意を要する⁸⁵⁾。

c. 甲状腺ホルモン併用(保険適用外)

T₃による効果増強について、8試験(292例の Stage 1 の大うつ病および双極性うつ病患者。ただし、大部分は大うつ病)のメタ解析が報告された⁶⁾。絶対ベネフィット増加は 23.2% (95% 信頼区間 4.5~41.9%)、治療効果発現必要症例数は 4.3 人であった。しかし、そのうち 4 つの RCT の結果を解析すると、T₃による効果増強は有意とはいえず、治療効果発現必要症例数は 12.5 人であった。このメタ解析は、T₃による効果増強について、無作為化対照試験がさらに必要であることを指摘した。

メタ解析にも含まれていた研究のうち、示唆に富む 2 研究を紹介する。Joffe らは、Stage 1 の非精神病性大うつ病 50 例を対象に、偽薬あるいは T₃ 37.5 μg/日、炭酸リチウム 900 mg/日を抗うつ薬に併用して 2 週間の RCT を行った⁸⁷⁾。反応率は偽薬群 19%、T₃ 群 59%、リチウム群 53% であり、T₃ とリチウムの効果は有意であった。さらに、Joffe と Singer は、Stage 1 の大うつ病患者(ただし、抗うつ薬の治療期間は 3 週間であり不十分)を対象に T₃ 37.5 μg/日と L-サイロキシン(T₄) 150 μg/日の 3 週間の二重盲検比較試験を行った⁸⁸⁾。T₃ に対する反応率は 59% であり、T₄ の 19% に比べて有意に優れていた。

以上のように、T₃ の難治性うつ病に対する効果の方が多く報告されているが、T₄ の Stage 2 難治性うつ病に対する有効性もオープン試験で報告されている⁸⁹⁾。双極性うつ病 12 例、大うつ病 5 例を対象として、抗うつ薬に T₄ を平均 482 μg/日と極めて高用量追加したところ、8 週間で 8 例 (47%) が反応かつ寛解した(双極性うつ病では 50%、大うつ病では 60% が寛解)。高用量の T₄ で、free T₄ 濃度は正常値の上限の平均 1.5 倍となったが、発汗、振戦以外に重篤な副作用はなかったという。筆者らは、Stage 2 の難治性うつ病患者に対する T₄ 併用の効果を報告したが¹¹⁾、最長 11 年までの長期経過観察後の最終診断に基づいて T₄ の有効性を再解析したところ、有効率は大うつ病で 12.5% (8 例中 1 例)、双極性うつ病で 67% (6 例中 4 例) であり、双極性で高い傾向がみられた⁵⁵⁾。

d. ドパミンアゴニスト併用(保険適用外)

難治性うつ病に対するドパミンアゴニストの効果については前方視的なオープン試験、症例報告、後方視的調査が 10 編ほど報告されている⁵⁵⁾。残念ながら、難治性うつ病に対する偽薬との二重盲検比較試験は行われていないので、ドパミンアゴニストの効果のエビデンス・レベルは低い。難治性大うつ病を対象とした研究ではないが、プラミベキソールの非精神病性大うつ病に対する二重盲検比較試験が偽薬と fluoxetine を対照として行われた⁹⁰⁾。プラミベキソール (1 mg) と fluoxetine (20 mg) はともに偽薬と比べて有意にうつ症状を改善した⁹⁰⁾。したがって、ドパミンアゴニストは抗うつ作用を有することから、抗うつ薬に併用した時の効果は、増強効果というよりはむしろ併用効果である。

筆者らは Stage 2 の難治性うつ病患者に対するドパミンアゴニスト(プロモクリプチン)併用の効果を報告したが¹¹⁾、最長 11 年までの長期経過観察後の最終診断に基づいてドパミンアゴニストの有効性を再解析したところ、有効率は大うつ病では 62.5% (15 例中 9 例)、双極性うつ病では 62.5%

(8例中5例)であり、ドパミンアゴニストは両病型で有効であった⁵⁵⁾。

e. 非定型抗精神病薬併用(保険適用外)

ドパミンアゴニストの結果と一見矛盾した効果のようにも思われるが、抗うつ薬と非定型抗精神病薬の併用が難治性うつ病の治療に有効であることが報告されている。Barbeeらは、後方視的調査で、Stage 1以上の非精神病性大うつ病に対する非定型抗精神病薬併用の有効率を、オランザピン57%、リスベリドン50%、クエチアピン33%、ziprasidone 10%と報告した⁹¹⁾。2006年にわが国でも発売されたアリピプラゾールのStage 1以上の非精神病性大うつ病に対する併用効果も、最近多くのオープン試験で検討され有効性が報告された(引用は最新の文献のみとする)⁹²⁾。

非定型抗精神病薬のうちオランザピン併用は大規模な二重盲検比較試験でStage 2の非精神病性大うつ病に有効であることが報告された⁹³⁾。8週間のfluoxetine治療に対する非反応者605例をオランザピン/fluoxetine併用群、fluoxetine継続群、オランザピン単独群の3群に無作為割り付けし8週間試験したところ、併用群はそれぞれの単独群に比べて有意に改善し、反応率、寛解率ともに有意に優れていた⁹³⁾。

一方、オランザピン/fluoxetine併用療法がStage 2の難治性非精神病性大うつ病に有効ではないという大規模な二重盲検比較試験(500例)も報告された⁹⁴⁾。SSRI非反応者のうち、ノルトリプチリンで7週間治療してやはり非反応であった症例をオランザピン/fluoxetine併用群、fluoxetine単独群、オランザピン単独群、ノルトリプチリン継続群の4群で8週間比較したところ、反応率は4群に差はみられなかった⁹⁴⁾。しかし、オランザピンによる抗うつ薬の効果増強を期待するのであれば、むしろオランザピン/ノルトリプチリン併用群をオランザピン単独群、ノルトリプチリン継続群と比較すべきだったのではないか。この点が、上記の研究⁹³⁾との方法上の大きな相違点である。

f. ピンドロール併用(保険適用外)

1990年代より、 β 遮断薬であるとともに、セロトニン1A/1D受容体アンタゴニストであるピンドロールとセロトニン再取り込み阻害薬(SSRIとクロミプラミン)の併用がセロトニン再取り込み阻害薬の効果を増強するのではないかと注目され、多くの臨床試験が行われてきた。実験的にピンドロールは前シナプスの受容体を遮断し、SSRIとの併用でシナプス間隙セロトニン濃度をさらに上昇させることから両者の併用効果が期待された。しかし、最近の二重盲検比較試験では、セロトニン再取り込み阻害薬非反応者に対してピンドロール併用は無効であることが報告されている⁹⁵⁾。

g. 反復性経頭蓋磁気刺激法

repetitive transcranial magnetic stimulation (TMS) (rTMS) (保険適用外)

大うつ病に対するrTMS群と偽治療(Sham)群の二重盲検比較試験がいくつか行われた。初期の研究では、Stage 1あるいはStage 2の難治性大うつ病を対象として左前頭前野へのrTMSを行ったが、Sham群との差がみられないか⁹⁶⁾。Sham群との差がみられても改善度・寛解率は軽度であり⁹⁷⁾、あまり有望とはいえない結果であった。しかし、最近報告された2試験では、Stage 2難治性うつ病(主に大うつ病だが、20~30%の双極性うつ病を含む)を対象としたSham群との二重盲検比較試験でrTMS群は有意に高い反応率(44% vs 8%⁹⁸⁾; 61% vs 6%⁹⁹⁾あるいは寛解率(36% vs 0%⁹⁸⁾)を示した。初期の研究との施行方法の違いは、Rossiniらの試験では左前頭前野へのrTMSでより強い強度(100% motor threshold, MT)を用いたことであり、初期の研究^{96, 97)}で採用された左前頭前野への80% MTのrTMSと比べても100% MTのrTMSは有意に有効であることを報告した⁹⁹⁾。また、Fitzgeraldらの試験では、以上の3試験とはrTMSの刺激部位が異なり、右前頭前野への低頻度rTMSと左前頭前野への高頻度rTMSを組み合わせた治療を行っ

た⁹⁸⁾、いずれも抗うつ薬あるいは気分安定薬に併用してrTMSを施行した。最近の2試験では双極性うつ病を対象に含んでいるが、躁転はみられなかった。

h. 迷走神経刺激 vagus nerve stimulation (VNS) (国内未承認)

新しいうつ病治療法としてVNSが最近注目されている。Stage 2の非精神病性難治性大うつ病と非精神病性難治性双極性うつ病(双極性うつ病は全対象の10.4%で、気分安定薬あるいは抗うつ薬による2種類の十分な治療に非反応と定義された)合わせて210例に対して、抗うつ薬あるいは気分安定薬に併用して、VNS群と偽治療(Sham)群との10週間の二重盲検比較試験が行われた¹⁰⁰⁾。VNS群の反応率はSham群と差はみられなかった。なお、大うつ病と双極性うつ病では反応率の差はみられず、病型にかかわらずVNSは難治性うつ病に有効とはいえない。

i. 脳深部刺激療法 deep brain stimulation (DBS) (国内未承認)

4種類以上の抗うつ薬に難治性の非精神病性大うつ病6例(5例ではECTにも難治性)に対して膝下野(Brodmann 25野, subgenual cingulate)に電極を置き、高頻度電気刺激を慢性的に加えたところ、治療開始2カ月後に6例中5例で反応がみられ、治療開始6カ月後では6例中4例が反応し、3例で寛解が得られた¹⁰¹⁾。Maybergらは難治性うつ病では膝下野の高活動が認められ(非難治性大うつ病の研究では低活動が報告されているが⁷³⁾、DBSによりこの部位の活動が低下し、同時に他の脳部位の活動が刺激されることが、抗うつ効果と関連することを示唆している¹⁰¹⁾。

j. lamotrigine 併用

Stage 2の難治性非精神病性大うつ病症例へのlamotrigine併用の有効性がオープン試験で報告された¹⁰²⁾。本章-2で紹介したように、現在わが国で抗てんかん薬として申請中のlamotrigineは双極性うつ病に有効性であることはエビデンスと

なっているが²⁶⁾、難治性大うつ病への効果も期待できる。

k. 精神療法

大うつ病の治療において精神療法は必須であり、これまで紹介した種々の薬物・身体療法は精神療法と併用して行われるべきものである。小精神療法³⁾の他、認知行動療法、対人関係療法などが行われている他に、大うつ病について深く理解することを助ける心理教育も精神療法的である。多くの二重盲検比較試験では偽薬に対する大うつ病の反応率が26~49%¹⁰³⁾(Janicak, 2001)と報告されているが、このことは単に偽薬効果を意味するだけでなく、精神科医による関わりが薬物療法なしでも治療的に有効であることを示していると筆者らは考える。

難治性大うつ病の薬物・心理的治療についてのStimpsonらによる系統的レビューは、難治性大うつ病に対する精神療法の効果についての十分なRCTはないと評価した⁴³⁾。その後、Stimpsonらが指摘した限界を踏まえたうえで、McPhersonらは組み入れ基準をゆるめて12の試験(ほとんどが認知行動療法、うち4つが比較試験)についての系統的レビューを発表した¹⁰⁴⁾。多くの研究はStage 1の大うつ病を対象としているが、難治性うつ病の定義が十分ではないこと、臨床試験としての質の高いものがないことなどが指摘され、やはり難治性うつ病に対する精神療法の効果に関するエビデンスが欠如していることが明らかになった。今後、質の高いRCTによって精神療法の効果が難治性うつ病で検討されることが必要である。

8 難治性双極性うつ病に対する治療

本章-2でも紹介したが、双極性うつ病の治療ガイドラインはまだ流動的であり、双極性うつ病の第一選択薬として非定型抗精神病薬が注目される一方、抗うつ薬の有効性については疑問視する意見もある²¹⁾。双極性うつ病に対する第一選択薬

のラインアップが確立されないかぎり、難治性双極性うつ病の定義も完成されないであろう。定義が確立されていないということは、その治療に関する研究もまだ十分にされていない、あるいはできないということの意味する。

以上のような現状であるが、難治性双極性うつ病の治療に関する少数の研究が発表されている。気分安定薬と1種類の抗うつ薬併用による十分な治療に反応しない難治性双極性うつ病に対して、オープン試験でアリピプラゾール併用(全例で気分安定薬に併用。一部症例で抗うつ薬・非定型抗精神病薬も併用)の有効性が報告された²⁵⁾。同様の定義の難治性双極性うつ病に対する lamotrigine とリスペリドン、inositol のオープン試験(3群あるいは任意の2群に無作為割り付けのため、一次的解析は2群間の比較となる)が気分安定薬・抗うつ薬に併用して最近行われた²⁴⁾。回復率(完全寛解に相当)に統計学的に有意な差はみられなかったが、lamotrigine 群でより症状改善する傾向がみられた²⁴⁾。

1種類の抗うつ薬に非反応(大うつ病の分類では Stage 1 に該当)の双極 II 型障害のうつ病患者(リチウムカルプロ酸を服用中)に対してプラミベキソール(最終与薬量 0.375~4.5 mg/日、平均 1.7 mg/日)が有効であることが偽薬との二重盲検比較試験で報告された(反応率はプラミベキソール群 60%、偽薬群 9%)¹⁰⁵⁾。同じ年に、別のグループも難治性双極性うつ病(定義は気分安定薬に併用して2種類以上の抗うつ薬による十分な治療に非反応であること、約 2/3 は双極 I 型)に対して、気分安定薬に併用してプラミベキソール(平均最高用量 1.7 mg/日)と偽薬の二重盲検比較試験を行い、プラミベキソールの有効性(反応率はプラミベキソール群 67%、偽薬群 20%)を報告した²³⁾。

本章-5で紹介したように、難治性大うつ病の一部は偽性単極性うつ病であると筆者らは報告した⁵⁵⁾。躁・軽躁病相が出現するまで偽性単極性うつ病(潜在性双極性うつ病)症例は、双極性うつ病の治療ガイドラインとはまったく異なる大うつ病の治療ガイドラインに準じた治療を受けることに

なり、当然治療抵抗性(むしろ偽性治療抵抗性というべきかもしれない)となりうる可能性がある。したがって、難治性の潜在性双極性うつ病を疑う症例では、双極性うつ病の治療ガイドラインに準じて治療することが、難治性の解決につながると思われる。偽性単極性うつ病(潜在性双極性うつ病)と大うつ病の鑑別が、症状レベル(困難と思われる)あるいは生物学的マーカーによって可能になるまでは、この問題は常に一定の割合(非難治性で 12.5%¹⁴⁾、難治性で 29%⁵⁵⁾)で起こりうることを常に認識すべきである。

9 予後

a. 筆者らの長期予後研究：難治性うつ病は寛解するか？

これまで Stage 2 の難治性うつ病の研究は少なかったが、最近徐々に Stage 2 の難治性うつ病の治療に関する大規模な比較試験が発表されてきている^{93, 94, 100)}。一方、Stage 2 の難治性うつ病の長期予後についてはまったく報告されてこなかった。「そもそも難治性うつ病のうつ病相は寛解するのだろうか？ 寛解するとしたらいつ、どのような治療で寛解するのか？」という臨床的疑問に答えるために、筆者らは 1995 年に調査した難治性うつ病症例¹¹⁾のうち、1年以上経過を追うことができた 26 症例(双極性うつ病 5 例、大うつ病 21 例)を最長 7 年間当科で診療を継続し、診断・重症度・薬物療法について長期転帰調査を行い報告した⁴²⁾。その後観察期間をさらに 11 年間まで延長して、診断・重症度・寛解に寄与した薬物療法他、病型別の各種増強治療の効果、発症時の病相、遺伝負因について報告した⁵⁵⁾。

長期経過観察期間中に、観察開始時の単極性うつ病 21 例のうち 5 例(28.6%)が双極性障害に移行したが、気分障害以外の病名に変更となる症例はなかった⁵⁵⁾。1995 年の調査開始時の大うつ病エピソードは、1995 年の調査時点で既に平均 5.1 年と慢性に続いていた¹¹⁾。うつ病相が自然に寛解することは Kraepelin の教科書¹⁵⁾にも詳細に記載されているが、Kraepelin は 14 年間うつ病が統

いた症例も紹介しており¹⁵⁾、実際に難治性うつ病が寛解するかどうかを確認することは重要である。1995年調査時の難治性うつ病相のうち、完全寛解(症状が消失し、機能も回復した状態)した症例を調査したところ、最終診断が大うつ病の症例では15例中9例が完全寛解となった(残り6例は最終観察時は軽症であった)⁵⁵⁾。一方、最終診断が双極性うつ病の症例では、11例中9例が完全寛解となった(残りの症例は最終観察時に軽症1例、中等症1例であった)。寛解までのうつ病相期間は、大うつ病で平均5.6年、双極性うつ病で平均4.8年であった。

b. 筆者らの長期予後研究：難治性うつ病の再発と治療薬

筆者らの研究では、大うつ病の寛解例9例中4例(3例は患者自身の判断で中断)、双極性うつ病の寛解例9例中6例(5例は患者自身の判断で中断)は薬物治療中止となったが、薬物療法中止後経過を十分に追跡できた症例では大うつ病で2例、双極性で4例再発した⁵⁵⁾。再発率が高率であることから、両病型において薬物療法は寛解後も長期に継続すべきであると思われる。1995年調査時点の難治性うつ病相の寛解に寄与したと思われる治療を調査したところ、寛解した大うつ病9例全例でドパミンアゴニストが有効であり、1例でリチウム併用も有効であった。寛解した双極性うつ病9例においても、6例でドパミンアゴニストが、4例でリチウムが、2例で甲状腺ホルモン(L-サイロキシン)が有効であり、寛解に寄与したと考えられた⁵⁵⁾。自然史的な観察で薬剤の効果を評価したため、自然寛解との区別をできない点为本研究の限界点であるが、これらの結果を今後RCTで確認する必要がある。

c. Dunner らの長期予後研究

筆者らの報告と同時期に、Dunner らが難治性の大うつ病あるいは双極性うつ病患者124人を2年間経過観察し、報告した¹⁶⁾。彼らの報告によると、種々の作用機序の抗うつ薬や気分安定薬、抗精神病薬、リチウム、ホルモン、ECTなどの treat-

ment as usual(従来型の経験論的な治療)を行っても持続的な寛解に至る症例はほとんどなく、観察開始1年後に寛解していた患者は112例中4例であり、そのうち2年後まで寛解が持続した症例はわずか1例であった¹⁶⁾。彼らの報告を読むと失望するかもしれないが、筆者らの追跡調査と同様により長期に観察を続けることによって、あるいは、彼らが試みていない治療法の導入によって、今後もっと高い寛解率が得られると期待する。

本稿では実際の症例を呈示しなかったが、個々の難治性うつ病症例は実に多様である。すべてのうつ病患者が回復することがわれわれ精神科医の目標であるが、なかなか目標に達しないことが多い。そのような時に、まず鑑別診断・併存疾患・性格要因を再検討しなくてはならないし、本稿で指摘したように偽性単極性うつ病の可能性も考慮に入れる必要がある。大うつ病という診断が間違いないとしても、心理社会的要因が大きくて、環境調整や精神療法的関わり工夫が功を奏する場合もある。さらに、難治性かなと思っても、他の抗うつ薬への変更で劇的に回復する症例もしばしばみられる。診断から治療まで実に多くのハードルがあり、そのハードルを1つ1つ確実にクリアしていくことが難治性うつ病の診療では肝要である。そして、たくさんあるハードルの存在を常に忘れないことと、そのハードルの乗り越え方を知っていることが、ゴールに到達する条件でもある。このようなハードルにたとえられる診断・治療上の多くの難治化要因を科学的に明らかにし、論文として発表し、精神医学の知識として蓄積することはわれわれの務めである。多数の難治性うつ病研究と称する研究はあるが、真に難治性うつ病(Stage 2以上)を対象とした研究は数えるほどしかないのが現状であり、難治性うつ病を十分に定義するだけで多くの研究が可能となる。今後、Stage 2以上(可能ならばStage 5)の難治性うつ病にできるだけ焦点を絞って、多角的に研究成果を蓄積していくことが求められている。

【文献】

- 1) 井上 猛, 北市雄士, 小山 司: 治療抵抗性うつ病の治療戦略とその作用機序. 日本精神神経学雑誌 106: 1016-23, 2004
- 2) Thase ME, Rush AJ: Treatment-resistant depression. In: Bloom FE, Kupfer DJ (eds), *Psychopharmacology: The Fourth Generation of Progress*. Raven Press, New York, pp1081-1097, 1995
- 3) 笠原 嘉: 治療 一般的事項. In: 笠原 嘉, 松下正明, 岸本英爾(編), *感情障害—基礎と臨床—*. 朝倉書店, pp46-347, 1999
- 4) 井上 猛, 小山 司: 難治性うつ病の治療. わが国における現状と治療アルゴリズム. *精神医学* 39: 6-14, 1997
- 5) Schatzberg AF, Cole JO, Elliot GR: Recent views on treatment-resistant depression. In: Halbreich U, Feinberg SS (eds), *Psychosocial Aspects of Nonresponse to Antidepressant Drugs*. American Psychiatric Press, Washington DC, pp93-109, 1986
- 6) Aronson R, Offman HJ, Joffe RT, et al: Triiodothyronine augmentation in the treatment of refractory depression: a meta-analysis. *Arch Gen Psychiatry* 53: 842-848, 1996
- 7) Bauer M, Döpfmer S: Lithium augmentation in treatment-resistant depression: meta-analysis of placebo-controlled studies. *J Clin Psychopharmacol* 19: 427-434, 1999
- 8) Dombrowski AY, Mulsant BH, Haskett RF, et al: Predictors of remission after electroconvulsive therapy in unipolar major depression. *J Clin Psychiatry* 66: 1043-1049, 2005
- 9) Sackeim HA, Prudic J, Devanand DP, et al: The impact of medication resistance and continuation pharmacotherapy on relapse following response to electroconvulsive therapy in major depression. *J Clin Psychopharmacol* 10: 96-104, 1990
- 10) Sharma V, Khan M, Smith A: A closer look at treatment resistant depression: is it due to a bipolar diathesis? *J Affect Disord* 84: 251-257, 2005
- 11) 井上 猛, 泉 剛, 本間裕士, 他: 抗うつ薬に治療抵抗性うつ病の実態とその治療戦略—自験例における調査結果と治療抵抗性うつ病の段階的治療に関する試案—. *日本精神神経学雑誌* 98: 329-342, 1996
- 12) Thase ME, Rush AJ: When at first you don't succeed: sequential strategies for antidepressant nonresponders. *J Clin Psychiatry* 58(Suppl 13): 23-29, 1997
- 13) Souery D, Lipp O, Massat I, et al: The characterization and definition of treatment-resistant mood disorders. In: Amsterdam JD, Hornig M, Nierenberg AA (eds), *Treatment-Resistant Mood Disorders*. Cambridge University Press, Cambridge, pp3-29, 2001
- 14) Akiskal HS, Maser JD, Zeller PJ, et al: Switching from 'unipolar' to bipolar II. An 11-year prospective study of clinical and temperamental predictors in 559 patients. *Arch Gen Psychiatry* 52: 114-123, 1995
- 15) Kraepelin E: *Psychiatrie. Ein Lehrbuch für Studierende und Ärzte*, achten Auflage. Johann Amrosius Barth, Leipzig, 1913(西丸四方, 西丸甫夫(訳): *躁うつ病とてんかん*. みすず書房, 1986)
- 16) Dunner DL, Rush AJ, Russell JM, et al: Prospective, long-term, multicenter study of the naturalistic outcomes of patients with treatment-resistant depression. *J Clin Psychiatry* 67: 688-695, 2006
- 17) Sachs GS: Treatment-resistant bipolar depression. *Psychiatr Clin North Am* 19: 215-236, 1996
- 18) Sachs GS, Thase ME, Otto MW, et al: Rationale, design, and methods of the systematic treatment enhancement program for bipolar disorder(STEP-BD). *Biol Psychiatry* 53: 1028-1042, 2003
- 19) American Psychiatric Association: *Practice Guideline for the Treatment of Psychiatric Disorders*. Compendium. American Psychiatric Pub, 2004(佐藤光源, 樋口輝彦, 井上新平(監訳): *米国精神医学会治療ガイドラインコンペンディウム*. 医学書院, 2006)
- 20) Yatham LN, Calabrese JR, Kusumakar V: Bipolar depression: criteria for treatment selection, definition of refractoriness, and treatment options. *Bipol Disord* 5: 85-97, 2003
- 21) 井上 猛, 田中輝明, 北市雄士, 他: 双極性障害での抗うつ薬の使い方. *精神科治療学* 20: 1141-1149, 2005
- 22) 中村 純: 双極性障害. In: 本橋伸高(編), *気分障害の薬物治療アルゴリズム*. じほう, pp55-63, 2003
- 23) Goldberg JF, Burdick KE, Endick CJ: Preliminary randomized, double-blind, placebo-controlled trial of pramipexole added to mood stabilizers for treatment-resistant bipolar depression. *Am J Psychiatry* 161: 564-566, 2004
- 24) Nierenberg AA, Ostacher MJ, Calabrese JR, et al: Treatment-resistant bipolar depression: a STEP-BD equipose randomized effectiveness trial of antidepressant augmentation with lamotrigine, inositol, or risperidone. *Am J Psychiatry* 163: 210-216, 2006
- 25) Kemp DE, Gilmer WS, Fleck J, et al: Aripiprazole augmentation in treatment-resistant bipolar depression: Early response and development of akathisia. *Prog Neuropsychopharmacol Biol Psychiatry* 31: 574-577, 2007
- 26) Calabrese JR, Bowden CL, Sachs GS et al: A double-blind placebo-controlled study of lamotrigine monotherapy in outpatients with bipolar I depression. *J Clin Psychiatry* 60: 79-88, 1999
- 27) Tohen M, Vieta E, Calabrese J, et al: Efficacy of olanzapine and olanzapine-fluoxetine combination in the treatment of bipolar I depression. *Arch Gen Psychiatry* 60: 1079-1088, 2003
- 28) Calabrese JR, Keck PE Jr, Macfadden W, et al: A randomized, double-blind, placebo-controlled trial of quetiapine in the treatment of bipolar I or II depression. *Am J Psychiatry* 162: 1351-1360, 2005.
- 29) Thase ME, Macfadden W, Weisler RH, et al: Efficacy of quetiapine monotherapy in bipolar I and II depression: a double-blind, placebo-controlled study (the BOLDER II study). *J Clin Psychopharmacol* 26: 600-609, 2006
- 30) Davis LL, Bartolucci A, Petty F: Divalproex in the treatment of bipolar depression: a placebo-controlled study. *J Affect Disord* 85: 259-266, 2005
- 31) Yatham LN, Kennedy SH, O'Donovan C, et al: Canadian Network for Mood and Anxiety Treatments

- (CANMAT) guidelines for the management of patients with bipolar disorder: update 2007. *Bipolar Disord* 8: 721-739, 2006
- 32) Gijsman HJ, Geddes JR, Rendell JM, et al: Antidepressants for bipolar depression: a systematic review of randomized, controlled trials. *Am J Psychiatry* 161: 1537-1547, 2004
 - 33) Nelsen MR, Dunner DL: Clinical and differential diagnostic aspects of treatment-resistant depression. *J Psychiatr Res* 29: 43-50, 1995
 - 34) O'Reardon JP, Amsterdam JD: Overview of treatment-resistant depression and its management. In: Amsterdam JD, Hornig M, Nierenberg AA(eds), *Treatment-Resistant Mood Disorders*. Cambridge University Press, Cambridge, pp30-45, 2001
 - 35) Sackeim HA: The definition and meaning of treatment-resistant depression. *J Clin Psychiatry* 62(Suppl 16): 10-17, 2001
 - 36) Licht RW, Qvitzau S: Treatment strategies in patients with major depression not responding to first-line sertraline treatment. A randomised study of extended duration of treatment, dose increase or mianserin augmentation. *Psychopharmacology* 161: 143-151, 2002
 - 37) Halpern JK, Glassman AH: Adequate tricyclic treatment: Defining the tricyclic nonresponder. In: Roose SP, Glassman AH(eds), *Treatment Strategies for Refractory Depression*. American Psychiatric Press, Washington DC, pp11-32, 1990
 - 38) Adli M, Baethge C, Heinz A, et al: Is dose escalation of antidepressants a rational strategy after a medium-dose treatment has failed? A systematic review. *Eur Arch Psychiatry Clin Neurosci* 255: 387-400, 2005
 - 39) Inoue T, Tsuchiya K, Miura J, et al: Bromocriptine treatment of tricyclic and heterocyclic antidepressant-resistant depression. *Biol Psychiatry* 40: 151-153, 1996
 - 40) Poirier MF, Boyer P: Venlafaxine and paroxetine in treatment-resistant depression. Double-blind, randomised comparison. *Br J Psychiatry* 175: 12-16, 1999
 - 41) Nolen WA, van de Putte JJ, Dijken WA, et al: Treatment strategy in depression. I. Non-tricyclic and selective reuptake inhibitors in resistant depression: a double-blind partial crossover study on the effects of oxaprotiline and fluvoxamine. *Acta Psychiatr Scand* 78: 668-675, 1988
 - 42) Inoue T, Nakagawa S, Kitaichi Y, et al: Long-term outcome of antidepressant-refractory depression: The relevance of unrecognized bipolarity. *J Affect Disord* 95: 61-67, 2006
 - 43) Stimpson N, Agrawal N, Lewis G: Randomised controlled trials investigating pharmacological and psychological interventions for treatment-refractory depression. Systematic review. *Br J Psychiatry* 181: 284-294, 2002
 - 44) Papakostas GI, Petersen T, Worthington JJ, et al: A pilot, open study of sertraline in outpatients with treatment-resistant depression (TRD) or with a history of TRD who responded but later relapsed. *Int Clin Psychopharmacol* 18: 293-296, 2003
 - 45) 井上 猛, 小山 司: 三環系・四環系抗うつ薬に治療抵抗性うつ病の定義の妥当性に関する研究—第二選択の三環系・四環系抗うつ薬への変更は有効な治療戦略か—。 *臨床精神医学* 26: 1603-1607, 1997
 - 46) Lam RW, Wan DD, Cohen NL, et al: Combining antidepressants for treatment-resistant depression: a review. *J Clin Psychiatry* 63: 685-693, 2002
 - 47) Möller HJ: Therapieresistenz auf Antidepressiva: Risikofaktoren und Behandlungsmöglichkeiten. *Nervenarzt*. 62: 658-669, 1991
 - 48) Parker GB, Malhi GS, Crawford JG, et al: Identifying "paradigm failures" contributing to treatment-resistant depression. *J Affect Disord* 87: 185-191, 2005
 - 49) 山下 格: *精神医学ハンドブック* 第3版. 日本評論社, 2000
 - 50) 笠原 嘉, 木村 敏: うつ状態の臨床分類に関する研究. *日本精神神経学雑誌* 77: 715-735, 1975
 - 51) 古川壽亮: DSMとICDの歴史. In: 古川壽亮, 神庭重信(編), *精神科診察診断学エビデンスからナラティブへ*. 医学書院, pp275-285, 2003
 - 52) 松浪克文: うつ病の概念を考える: 「神経症性うつ病」という概念の行方. *精神科治療学* 17: 969-978, 2002
 - 53) Nelson JC, Mazure CM, Jatlow PI: Characteristics of desipramine-refractory depression. *J Clin Psychiatry* 55: 12-19, 1994
 - 54) Roose SP, Glassman AH, Walsh BT, et al: Tricyclic nonresponders: phenomenology and treatment. *Am J Psychiatry* 143: 345-348, 1986
 - 55) 井上 猛, 北市雄士, 田中輝明, 他: 難治性うつ病に対するドパミン関連薬剤の効果と安全性. *脳と精神の医学* 18: 35-43, 2007
 - 56) Chan CH, Janicak PG, Davis JM, et al: Response of psychotic and nonpsychotic depressed patients to tricyclic antidepressants. *J Clin Psychiatry* 48: 197-200, 1987
 - 57) Glassman AH, Perel JM, Shostak M, et al: Clinical implications of imipramine plasma levels for depressive illness. *Arch Gen Psychiatry* 34: 197-204, 1977
 - 58) Liebowitz MR, Quitkin FM, Stewart JW, et al: Antidepressant specificity in atypical depression. *Arch Gen Psychiatry* 45: 129-137, 1988
 - 59) Fava M, Uebelacker LA, Alpert JE, et al: Major depressive subtypes and treatment response. *Biol Psychiatry* 42: 568-576, 1997
 - 60) Petersen T, Gordon JA, Kant A, et al: Treatment resistant depression and axis I co-morbidity. *Psychol Med* 31: 1223-1229, 2001
 - 61) Fava M: Diagnosis and definition of treatment-resistant depression. *Biol Psychiatry* 53: 649-659, 2003
 - 62) Petersen T, Hughes M, Papakostas GI, et al: Treatment-Resistant Depression and Axis II Comorbidity. *Psychother Psychosom* 71: 269-274, 2002
 - 63) Papakostas GI, Petersen T, Iosifescu DV, et al: Axis III disorders in treatment-resistant major depressive disorder. *Psychiatry Res* 118: 183-188, 2003
 - 64) Goodwin FK, Jamison KR: *Manic-Depressive Illness*. Oxford University Press, New York, 1990.
 - 65) Ghaemi SN, Ko JY, Goodwin FK: The bipolar spectrum and the antidepressant view of the world. *J Psychiatr Pract* 7: 287-297, 2001

- 66) Kusumi I, Suzuki K, Sasaki Y, et al: Treatment response in depressed patients with enhanced Ca mobilization stimulated by serotonin. *Neuropsychopharmacology* 23: 690-696, 2000
- 67) Suzuki K, Kusumi I, Sasaki Y, et al: Serotonin-induced platelet intracellular calcium mobilization in various psychiatric disorders: is it specific to bipolar disorder? *J Affect Disord* 64: 291-296, 2001
- 68) Ising M, Kunzel HE, Binder EB, et al: The combined dexamethasone/CRH test as a potential surrogate marker in depression. *Prog Neuropsychopharmacol Biol Psychiatry* 29: 1085-1093, 2005
- 69) Nikisch G, Mathe AA, Czernik A, et al: Long-term citalopram administration reduces responsiveness of HPA axis in patients with major depression: relationship with S-citalopram concentrations in plasma and cerebrospinal fluid (CSF) and clinical response. *Psychopharmacology* 181: 751-756, 2005
- 70) Ising M, Horstmann S, Kloiber S, et al: Combined dexamethasone/corticotropin releasing hormone test predicts treatment response in major depression-A potential biomarker? *Biol Psychiatry* 62: 47-54, 2007
- 71) Mulert C, Juckel G, Brunnermeier M, et al: Prediction of treatment response in major depression: integration of concepts. *J Affect Disord* 98: 215-225, 2007
- 72) Fagiolini A, Kupfer DJ: Is treatment-resistant depression a unique subtype of depression? *Biol Psychiatry* 53: 640-648, 2003
- 73) Drevets WC, Bogers W, Raichle ME: Functional anatomical correlates of antidepressant drug treatment assessed using PET measures of regional glucose metabolism. *Eur Neuropsychopharmacol* 12: 527-544, 2002
- 74) Mayberg HS, Brannan SK, Mahurin RK, et al: Cingulate function in depression: a potential predictor of treatment response. *Neuroreport* 8: 1057-1061, 1997
- 75) Mayberg HS, Brannan SK, Tekell JL, et al: Regional metabolic effects of fluoxetine in major depression: serial changes and relationship to clinical response. *Biol Psychiatry* 48: 830-843, 2000
- 76) Little JT, Ketter TA, Kimbrell TA, et al: Bupropion and venlafaxine responders differ in pretreatment regional cerebral metabolism in unipolar depression. *Biol Psychiatry* 57: 220-228, 2005
- 77) Brody AL, Saxena S, Silverman DH, et al: Brain metabolic changes in major depressive disorder from pre- to post-treatment with paroxetine. *Psychiatry Res* 91: 127-139, 1999
- 78) Shah PJ, Glabus MF, Goodwin GM, et al: Chronic, treatment-resistant depression and right fronto-striatal atrophy. *Br J Psychiatry* 180: 434-440, 2002
- 79) Videbeck P, Ravnkilde B: Hippocampal volume and depression: a meta-analysis of MRI studies. *Am J Psychiatry* 161: 1957-1966, 2004
- 80) Sackeim HA, Prudic J, Devanand DP, et al: A prospective, randomized, double-blind comparison of bilateral and right unilateral electroconvulsive therapy at different stimulus intensities. *Arch Gen Psychiatry* 57: 425-434, 2000
- 81) Stein G, Bernadt M: Lithium augmentation therapy in tricyclic-resistant depression. A controlled trial using lithium in low and normal doses. *Br J Psychiatry* 162: 634-640, 1993
- 82) Ontiveros A, Fontaine R, Elie R: Refractory depression: the addition of lithium to fluoxetine or desipramine. *Acta Psychiatr Scand* 83: 188-192, 1991
- 83) Nelson JC, Mazure CM: Lithium augmentation in psychotic depression refractory to combined drug treatment. *Am J Psychiatry* 143: 363-366, 1986
- 84) Bschor T, Canata B, Muller-Oerlinghausen B, et al: Predictors of response to lithium augmentation in tricyclic antidepressant-resistant depression. *J Affect Disord* 64: 261-265, 2001
- 85) Bschor T, Adli M, Baethge C, et al: Lithium augmentation increases the ACTH and cortisol response in the combined DEX/CRH test in unipolar major depression. *Neuropsychopharmacology* 27: 470-478, 2002
- 86) Bschor T, Baethge C, Adli M, et al: Association between response to lithium augmentation and the combined DEX/CRH test in major depressive disorder. *J Psychiatr Res* 37: 135-143, 2003
- 87) Joffe RT, Singer W, Levitt AJ, et al: A placebo-controlled comparison of lithium and triiodothyronine augmentation of tricyclic antidepressants in unipolar refractory depression. *Arch Gen Psychiatry* 50: 387-393, 1993
- 88) Joffe RT, Singer W: A comparison of triiodothyronine and thyroxine in the potentiation of tricyclic antidepressants. *Psychiatry Res* 32: 241-251, 1990
- 89) Bauer M, Hellweg R, Graf KJ, et al: Treatment of refractory depression with high-dose thyroxine. *Neuropsychopharmacology* 18: 444-455, 1998
- 90) Corrigan MH, Denahan AQ, Wright CE, et al: Comparison of pramipexole, fluoxetine, and placebo in patients with major depression. *Depression Anxiety* 11: 58-65, 2000
- 91) Barbee JG, Conrad EJ, Jamhour NJ: The effectiveness of olanzapine, risperidone, quetiapine, and ziprasidone as augmentation agents in treatment-resistant major depressive disorder. *J Clin Psychiatry* 65: 975-981, 2004
- 92) Pae CU, Patkar AA, Jun TY, Lee C, et al: Aripiprazole augmentation for treatment of patients with inadequate antidepressants response. *Depress Anxiety* 24: 522-526, 2007
- 93) Thase ME, Corya SA, Osuntokun O, et al: A randomized, double-blind comparison of olanzapine/fluoxetine combination, olanzapine, and fluoxetine in treatment-resistant major depressive disorder. *J Clin Psychiatry* 68: 224-236, 2007
- 94) Shelton RC, Williamson DJ, Corya SA, et al: Olanzapine/fluoxetine combination for treatment-resistant depression: a controlled study of SSRI and nortriptyline resistance. *J Clin Psychiatry* 66: 1289-1297, 2005
- 95) Perry EB, Berman RM, Sanacora G, et al: Pindolol augmentation in depressed patients resistant to selective serotonin reuptake inhibitors: a double-blind, randomized, controlled trial. *J Clin Psychiatry* 65: 238-243, 2004
- 96) Boutros NN, Gueorguieva R, Hoffman RE, et al: Lack

- of a therapeutic effect of a 2-week sub-threshold transcranial magnetic stimulation course for treatment-resistant depression. *Psychiatry Res* 113: 245-254, 2002
- 97) Berman RM, Narasimhan M, Sanacora G, et al: A randomized clinical trial of repetitive transcranial magnetic stimulation in the treatment of major depression. *Biol Psychiatry* 47: 332-337, 2000
- 98) Fitzgerald PB, Benitez J, de Castella A, et al: A randomized, controlled trial of sequential bilateral repetitive transcranial magnetic stimulation for treatment-resistant depression. *Am J Psychiatry* 163: 88-94, 2006
- 99) Rossini D, Lucca A, Zanardi R, et al: Transcranial magnetic stimulation in treatment-resistant depressed patients: a double-blind, placebo-controlled trial. *Psychiatry Res* 137: 1-10, 2005
- 100) Rush AJ, Marangell LB, Sackeim HA, et al: Vagus nerve stimulation for treatment-resistant depression: a randomized, controlled acute phase trial. *Biol Psychiatry* 58: 347-354, 2005
- 101) Mayberg HS, Lozano AM, Voon V, et al: Deep brain stimulation for treatment-resistant depression. *Neuron* 45: 651-660, 2005
- 102) Barbee JG, Jamhour NJ: Lamotrigine as an augmentation agent in treatment-resistant depression. *J Clin Psychiatry* 63: 737-741, 2002
- 103) Janicak PG, Davis JM, Preskorn SH: Principles and Practice of Psychopharmacotherapy, 3rd ed. Lippincott Williams & Wilkins, Philadelphia, 2001
- 104) McPherson S, Cairns P, Carlyle J, et al: The effectiveness of psychological treatments for treatment-resistant depression: a systematic review. *Acta Psychiatr Scand* 111: 331-340, 2005
- 105) Zarate CA Jr, Payne JL, Singh J, et al: Pramipexole for bipolar II depression: a placebo-controlled proof of concept study. *Biol Psychiatry* 56: 54-60, 2004

(井上 猛・小山 司)

Prior Neonatal Isolation Reduces Induction of NGF mRNA and Decreases GDNF mRNA in the Hippocampus of Juvenile and Adult Rodents Subjected to Immobilization Stress

KI-ICHIRO KAWANO,¹ SHIGERU MORINOBU,^{1*} TAKUYA SAWADA,¹ SEIICHI TSUJI,¹ KAORI ERABI,¹ MANABU FUCHIKAMI,¹ TOSHIRO KOZURU,¹ SHIGETO YAMAWAKI,¹ KAZUE HISAOKA,² AND MINORU TAKEBAYASHI²

¹Division of Frontier Medicine, Department of Psychiatry and Neurosciences, Graduate School of Medical Sciences, Hiroshima University, 1-2-3 Kasumi, Minami-ku, Hiroshima, 734-8551, Japan

²Division of Clinical Research, Department of Psychiatry, National Hospital Organization, Kure Medical Center, 3-1 Aoyama, Kure, 737-0023, Japan

KEY WORDS neonatal isolation; immobilization; NGF; GDNF; NT-3; hippocampus

ABSTRACT Numerous studies have demonstrated that early adverse experiences are associated with the development of susceptibility to stress later in life. Although it is known that early experience of adversity, such as neonatal isolation, maternal separation, and low maternal care, enhances the activity of the hypothalamo-pituitary-adrenal axis in rodents, the detailed mechanism underlying stress susceptibility induced by early adversity remains to be elucidated. Since neurotrophins have been shown to have a neuroprotective effect, we examined the influence of repeated neonatal isolation on expression of nerve growth factor (NGF), glia cell-derived neurotrophic factor (GDNF), and neurotrophin-3 mRNA in the hippocampus of juvenile and adult rats subsequently exposed to immobilization stress, using real-time quantitative PCR and *in situ* hybridization. Neonatal isolation did not affect the basal hippocampal expression of these neurotrophin mRNAs in either juvenile or adult rats not subsequently exposed to immobilization. Similarly, there was a significant interaction between neonatal isolation and immobilization that affected the expression of NGF and GDNF mRNAs. Neonatal isolation attenuated the induction of NGF mRNA in both groups of rats and decreased GDNF mRNA in juvenile rats in response to immobilization. The decreased induction of NGF mRNA and reduced GDNF mRNA in response to immobilization was found in the CA3 pyramidal cell layer and dentate gyrus granular cell layer in the hippocampus of adult rats that had been subjected to neonatal isolation. These findings suggest that susceptibility to stress arising from prior neonatal isolation might be a result of decreased neuroprotective support through NGF and GDNF. *Synapse* 62:259–267, 2008. © 2008 Wiley-Liss, Inc.

INTRODUCTION

Numerous preclinical studies have revealed that exposure to either early adversity and/or long-term stress in adulthood can lead to alterations in hippocampal structure and function. For example, chronic adulthood stresses, such as forced immobilization or restraint, have been reported to induce a significant decrease in the dendritic length and the number of branch points in hippocampal CA3 pyramidal neurons (Vyas et al., 2002), and to atrophy of the apical dendrites of CA3 pyramidal cells in the rat hippocampus (Magarinos et al., 1995, 1996; Watanabe et al., 1992).

Similarly, chronic parental separation induces a significant decrease in the length of the dendrites of hippocampal CA1 pyramidal neurons, and a significant reduction of granular cell spine density in the hippocampal dentate gyrus (DG) in the juvenile degus

*Correspondence to: Shigeru Morinobu, MD, PhD, Division of Frontier Medicine, Department of Psychiatry and Neurosciences, Graduate School of Medical Sciences, Hiroshima University, 1-2-3 Kasumi minami-ku, Hiroshima 724-8551, Japan. E-mail: smorib@hiroshima-u.ac.jp

Received 9 February 2007; Accepted 30 September 2007

DOI 10.1002/syn.20487

Published online in Wiley InterScience (www.interscience.wiley.com).

(Poeggel et al., 2003). In addition, there is a significant reduction of mossy fiber density in the hippocampus of adult rats subjected to repeated episodes of maternal separation (Huot et al., 2002). Moreover, juvenile rats subjected to repeated episodes of maternal separation experienced a significant decrease in the volume and total number of BrdU-positive cells within the DG as well as a significant increase in the number of TUNEL-positive cells within this region of the brain (Lee et al., 2001). Mild prenatal stress has been shown to decrease synaptic density in the hippocampus of affected offspring (Hayashi et al., 1998). Taken together, these morphological findings suggest that various types of stress affect synaptic structure. However, the precise mechanism by which exposure to stress induces morphological changes in the hippocampus remains to be elucidated.

Neurotrophic factors and growth factors play a critical role in the differentiation and maintenance of neurons (Folli et al., 1996; Maisonpierre et al., 1990), suggesting that these factors may be involved in stress-induced synaptic remodeling. A significant reduction of brain-derived neurotrophic factor (BDNF) mRNA was found in the hippocampus of adult rats subjected to single and repeated episodes of maternal separation (MacQueen et al., 2003; Roceri et al., 2002). In contrast, the level of BDNF protein in adult rats subjected to repeated episodes of maternal separation was higher than that in adult rats subjected to neonatal brief handling despite a similar level of BDNF mRNA in these 2 groups (Greisen et al., 2005). Furthermore, the level of BDNF mRNA in the hippocampus of offspring of mothers that exhibited a high degree of maternal care was significantly higher than that in offspring of mothers that exhibited a low degree of maternal care (Liu et al., 2000). In contrast, a single episode of maternal separation was reported to increase the level of nerve-growth factor (NGF) mRNA in the developing rat hippocampus (Cirulli et al., 1998, 2000). Recently, we demonstrated that repeated episodes of neonatal isolation led to a significant decrease in the level of insulin-like growth factor (IGF) 1 receptor mRNA and protein in the hippocampus of the corresponding adult rats (Erabi et al., 2007). Although the influences of early adverse experiences on the expression of neurotrophic and growth factors in the hippocampus has been recognized, further studies focusing on other factors such as NGF, glia cell-derived neurotrophic factor (GDNF), and neurotrophin-3 (NT-3), are required to elucidate the involvement of these factors in the morphological changes in response to early adversity.

In this report, we examined the effect of repeated episodes of neonatal isolation on the levels of NGF, GDNF, and NT-3 mRNA in the hippocampus of juvenile (28-day-old) and adult (90-day-old) rats. In addition,

epidemiological studies of mental disorders have suggested that early adverse experiences increase the susceptibility to mental disorders in response to stress. Therefore, we also examined the levels of these neurotrophic factors in response to a single episode of immobilization stress (SIS) in juvenile and adult animals that had been previously exposed to repeated episodes of neonatal isolation.

MATERIALS AND METHODS

Animals

Pregnant female Sprague-Dawley rats were purchased from Charles River Japan (Yokohama, Japan). The rats were housed individually in a breeding colony at constant room temperature ($23^{\circ}\text{C} \pm 2^{\circ}\text{C}$) and humidity (60%) with a 12 h/12 h light/dark cycle (lights on 8 AM to 8 PM). Food and water were provided ad libitum. The litters were culled to 12 pups on postnatal day 1 (PN day 1).

Neonatal isolation paradigm

The mothers and pups of the nonisolated group were left undisturbed until weaning. Neonatal isolation was conducted according to the method of Kehoe and Bronzino (1999). Pups were isolated from the dam, nest, and siblings and placed in individual round containers for 1 h per day on PN days 2–9. All litters were weaned on PN day 21, separated on the basis of sex, and maintained with ad libitum access to food and water. Only the male rats were subjected to the following experimental procedure. All animal procedures were conducted in accordance with the Guiding Principles on Animal Experimentations in Research Facilities for Laboratory Animal Science Hiroshima University and approved by Hiroshima University Animal Care Committee.

Single immobilization stress paradigm

On PN day 28 (juvenile) or 90 (adult), half of both the isolated and nonisolated rats were subjected to a single restraint stress (SRS) for 2 h. The immobilization stress was conducted as described previously (Morinobu et al., 2003; Suenaga et al., 2004). Briefly, rats were immobilized in clear polyethylene disposable cone bags (Asahikasei, Tokyo, Japan), sized so that rats were equally immobilized. Animals were sacrificed by decapitation after completion of the immobilization stress. On PN day 28 or 90, isolated and nonisolated rats (sham) were sacrificed by decapitation. The hippocampus was isolated, immediately frozen, stored at -70°C , and used for real-time quantitative PCR analysis. In this procedure, the lateral choroid plexus was carefully removed from the hippocampus. For *in situ* hybridization, brains were

TABLE I. Primer and probe sequences used for real time PCR

NGF	Forward primer	AGCCACTGGACTAACTTCAGC
	Reverse primer	GGGACTGGGGCTC
	TaqMan probe	TTCCCTTGACACAGCCCTCCGC
GDNF	Forward primer	ACTCCCTCGGCCACCG
	Reverse primer	ATAATCTTCGGGCATATTGGAGTC
	TaqMan probe	CGTGCCCTTCGCGCTGACC
NT-3	Forward primer	GAGGCACCCAGAGAACCAGA
	Reverse primer	TTGCAATCATCGGCTGGA
	TaqMan probe	CAGGGAGAGGCCACAGGTCAGAA

Forward primer: 5' to 3'; Reverse primer: 5' to 3'; TaqMan probe: 5' [FAM] to 3' [TAMRA].

removed rapidly, immediately frozen, and stored at -70°C .

Four groups of both juvenile and adult rats [subjected to neonatal isolation followed by SRS (NI + SRS); neonatal isolation alone (NI); SRS alone (SRS); or sham treatment (Sham)] were used for real-time quantitative PCR. No more than two rats per experimental group were from any single litter. A total of 24 juvenile and 51 adult rats were subjected to real-time quantitative PCR analysis and a different set of rats was used for each experiment (real-time quantitative PCR and hybridization).

Real-time quantitative PCR

Total RNA was extracted with an RNAqueous Phenol-free Total RNA Isolation Kit (Ambion, Austin, TX). After treatment with RNase-free DNase I (Takara, Kusatsu, Japan), real-time quantitative PCR was performed with an ABI PRISM 7700 sequence detection system (Applied Biosystems, Foster City, CA) to quantify relative mRNA levels in samples. Real-time quantitative PCR was performed to amplify the genes encoding NGF, GDNF, and NT-3. The primers and TaqMan hybridization probe were designed using Primer Express software (Applied Biosystems). Table I shows the sequences and associated fluorescent dye of each PCR primers and TaqMan probe. The TaqMan probes, which were designed to hybridize to the PCR products, were labeled with a fluorescent reporter dye at the 5' end and a quenching dye at the 3' end. PCR was carried out using the TaqMan Universal PCR Master Mix (Applied Biosystems). All standards and samples were assayed in triplicate. Each plate contained the same standard. Thermal cycling was initiated with an initial denaturation at 50°C for 2 min and 95°C for 10 min. After this initial step, 40 cycles of PCR (heating at 95°C for 15 s and 60°C for 1 min) were performed. The PCR assay for glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was performed using TaqMan Rodent GAPDH Control Reagents (Applied Biosystems). The PCR assays on the test samples were performed simultaneously with the assays on the standard samples (rat brain tissue), which allowed us to construct a standard curve. The estimated relative expression

of GAPDH and NGF, GDNF, or NT-3 mRNAs in the test samples were calculated using this standard curve and the ratio of the relative expression of NGF, GDNF, or NT-3 mRNAs was calculated relative to the expression of GAPDH.

Hybridization paradigm

Analysis of NGF and GDNF expression by *in situ* hybridization was conducted according to the method of Wetmore et al. (1992) with a minor modification. Oligonucleotide probes complementary to parts of the mRNAs encoding NGF and GDNF were synthesized and labeled with DIG at the 5' end. The following antisense DNA oligonucleotides were used: NGF: 5'-CTG CGG GCT CTG CGG AGG GCT GTG TCA AGG GAA TGC TGA AGT TTA GTC CA-3' (Nosrat et al., 1997) and GDNF: 5'-AGC CAC CAT CAA AAG ACT GAA AAG GTC ACC AGA TAA ACA AGC GGC GGC A-3' (Lin et al., 1993).

Fresh-frozen coronal brain sections ($15\ \mu\text{m}$) through the hippocampus were prepared using a cryostat, thaw-mounted onto slides (Matsunami, Kishiwada, Japan) and fixed with 4% paraformaldehyde for 5 min. Sections were hybridized for 18 h at 42°C in a humidified chamber with at least 0.5 ng probe/slide in a mixture of $4 \times \text{SSC}$ ($1 \times \text{SSC} = 0.15\ \text{M NaCl}/0.015\ \text{M sodium citrate}$), 50% formamide, $1 \times \text{Denhardt's solution}$ (0.02% each of polyvinylpyrrolidone, bovine serum albumin, and Ficoll), 1% sarcosyl, 0.02 M phosphate buffer (pH 7.0), 10% dextran sulfate, 1 mg/ml of yeast RNA, and 200 mM dithiothreitol. Following hybridization, slides were rinsed for $5 \times 15\ \text{min}$ at 55°C in $2 \times \text{SSC}$, and allowed to reach room temperature. The sections were then incubated at 4°C overnight with an antidigoxigenin-alkaline phosphatase conjugate (Roche, Basel, Switzerland). After the incubation, the sections were extensively washed in PBT containing 0.2% BSA and 4 mM Levamisol, then washed in the reaction buffer (100 mM NaCl, 100 mM TrisHCl at pH 9.5, 50 mM MgCl_2 , 0.1% Tween 20, and 4 mM Levamisol), and incubated with chromagen containing nitroblue tetrazolium salt (337 mg/ml) and 5-bromo-4-chloro-3-indoyl phosphate (175 mg/ml) according to the manufacturer's protocol (Roche). After color development (DIG labeling is visualized as a blue reaction product under light microscopy), slides were photographed under light microscopy (Model BZ-8000) (Keyence, Osaka, Japan). The mean density of sections was measured in the CA1 and CA3 regions ($400\ \mu\text{m} \times 400\ \mu\text{m}$), and granule cell layer of the DG ($400\ \mu\text{m} \times 200\ \mu\text{m}$) using the NIH Scion Image analysis program (Figs. 3a and 4a).

Statistical analysis

Results were expressed as mean \pm SEM. The results of real-time quantitative PCR were analyzed

Synapse DOI 10.1002/syn

by two-way analysis of variance (ANOVA) (neonatal isolation \times single immobilization stress), and posthoc comparisons were performed using Scheffe's test. For hybridization analyses, the results of experiments containing two groups of rats were analyzed by the Mann-Whitney U test. Significance was set at $P < 0.05$.

RESULTS

We first performed real-time quantitative PCR analysis to measure NGF, GDNF, and NT-3 mRNA in RNA isolated from the hippocampus of juvenile and adult rats subjected to either neonatal isolation followed by SIS (NI + SIS), neonatal isolation alone (NI), SIS alone (SIS) or sham treatment. In both juvenile and adult rats, two-way ANOVA showed no significant effect of NI [juvenile: $F(1, 23) = 2.652$, $P = 0.119$; adult: $F(1, 23) = 4.257$, $P = 0.051$] on the level of NGF mRNA, but revealed a significant effect of SIS [juvenile: $F(1, 23) = 7.093$, $P = 0.015$; adult: $F(1, 23) = 4.926$, $P = 0.037$], and a significant interaction between NI and SIS [juvenile: $F(1, 23) = 12.437$, $P = 0.002$; adult: $F(1, 23) = 6.605$, $P = 0.017$] (Figs. 1a and 2a). Posthoc analysis revealed that the levels of NGF mRNA in the SIS group was significantly higher ($P < 0.05$) than those in the Sham or NI + SIS group for both juveniles and adults (Figs. 1a and 2a). However, there was no significant difference between the level of NGF mRNA in the NI + SIS and Sham groups. In situ hybridization analysis demonstrated that the level of NGF mRNA in the CA3 pyramidal cell layer ($P = 0.022$) and the DG granular cell layer ($P = 0.002$) of the NI + SIS group was significantly lower than that in the SIS group in adulthood (Figs. 3a and 3b).

In the analyses of GDNF mRNA in juvenile rats, we observed no significant effect of SIS on the level of GDNF mRNA [$F(1, 23) = 0.877$, $P = 0.360$], but we did observe a significant effect of NI [$F(1, 23) = 8.865$, $P = 0.007$]. We found the trend toward significant interaction between NI and SIS [$F(1, 23) = 4.328$, $P = 0.051$] (Fig. 1b). Posthoc analysis revealed that the level of GDNF mRNA ($P < 0.05$) in the NI + SIS group was significantly lower than that in the SIS group (Fig. 1b). In adulthood, we observed no significant effect of NI on the level of GDNF mRNA [$F(1, 23) = 0.017$, $P = 0.935$], but we did observe a significant effect of SIS [$F(1, 23) = 8.172$, $P = 0.009$], as well as a significant interaction between NI and SIS [$F(1, 23) = 12.143$, $P = 0.002$] (Fig. 2b). Posthoc analysis revealed that the level of GDNF mRNA ($P < 0.05$) in the NI + SIS group was significantly lower than that in the NI group (Fig. 2b). The levels of GDNF mRNA in the Sham group also tended to be lower than that in the NI group, although the difference did not reach statistical significance ($P = 0.096$).

Synapse DOI 10.1002/syn

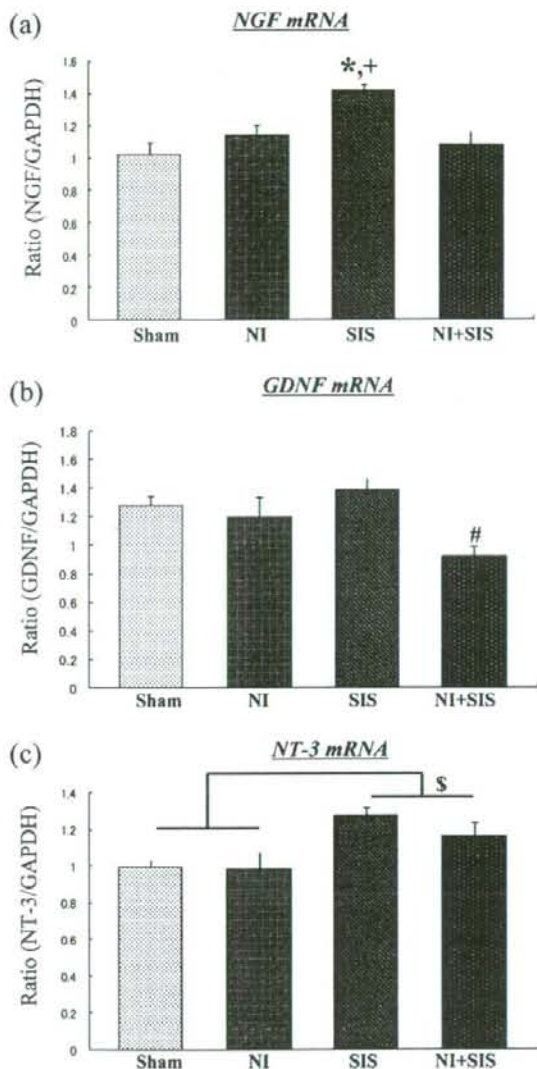


Fig. 1. Expression of (a) NGF, (b) GDNF, and (c) NT-3 mRNA in the hippocampus of juvenile rats subjected to sham treatment (Sham), neonatal isolation alone (NI), single immobilization stress (SIS), and neonatal isolation followed by a single immobilization stress (NI + SIS). Results are expressed as the ratio of the concentration of the target neurotrophin to that of GAPDH (target neurotrophin/GAPDH) percentage of Sham levels. The mean \pm SEM ($n = 6$) is shown. * $P < 0.05$ compared with sham, $^{\#}P < 0.05$ compared with SIS, $^{\$}P < 0.05$ compared with NI + SIS, $^{\$}P < 0.05$ compared with Sham and NI (two-way ANOVA followed by Scheffe's test).

In situ hybridization analysis demonstrated that the level of GDNF mRNA in the CA1 ($P = 0.004$) and CA3 ($P = 0.025$) pyramidal cell layers and the DG ($P = 0.010$) granular cell layer of the NI + SIS group was significantly lower than that in the NI group (Fig. 4).

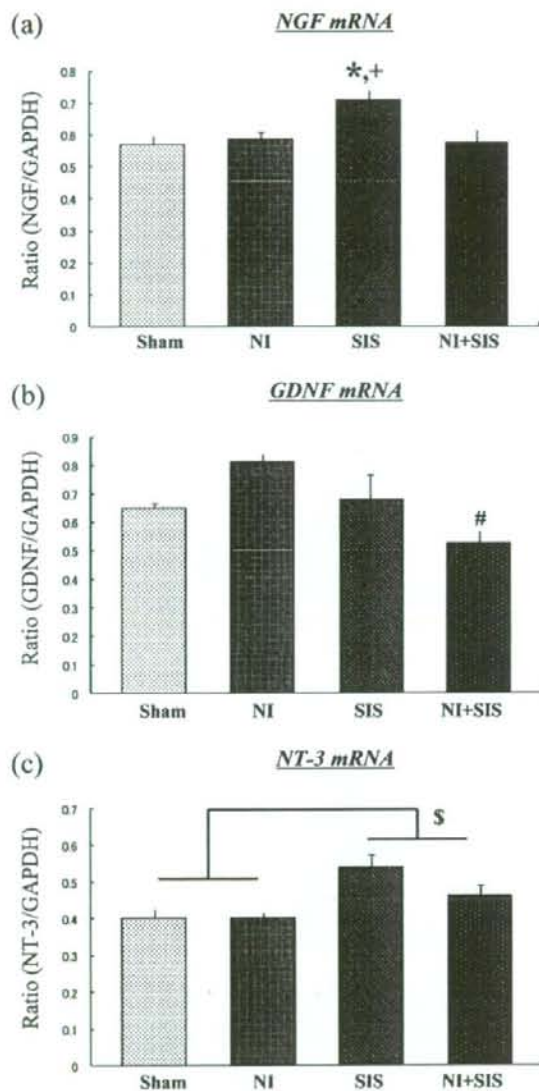


Fig. 2. Expression of (a) NGF, (b) GDNF, and (c) NT-3 mRNA in the hippocampus of adult rats subjected to sham treatment (Sham), neonatal isolation alone (NI), single immobilization stress alone (SIS), and neonatal isolation followed by a single immobilization stress (NI + SIS). Results are expressed as the ratio of the concentration of the target neurotrophin to that of GAPDH (target neurotrophin/GAPDH) percentage of Sham levels. The mean \pm SEM ($n = 6-8$) is shown. * $P < 0.05$ compared with sham, # $P < 0.05$ compared with NI, S $P < 0.05$ compared with NI + SIS, * $P < 0.05$ compared with Sham and NI (two-way ANOVA followed by Scheffe's test).

In both juvenile and adult rats, two-way ANOVA showed no significant effect of NI on the level of NT-3 mRNA [juvenile: $F(1, 23) = 1.084$, $P = 0.309$; adult: $F(1, 23) = 2.539$, $P = 0.125$], but did show a significant effect of SIS [juvenile: $F(1, 23) = 13.285$, $P =$

0.001; adult: $F(1, 23) = 15.136$, $P = 0.001$], but no significant interaction between NI and SIS [juvenile: $F(1, 23) = 0.611$, $P = 0.443$; adult: $F(1, 23) = 2.319$, $P = 0.141$] (Figs. 1c and 2c). Posthoc analysis revealed that the level of NT-3 mRNA in rats subjected to SIS was significantly higher ($P < 0.05$) than that in juvenile and adult rats not subjected to SIS.

DISCUSSION

The results of the present study demonstrate that repeated episodes of neonatal isolation do not affect the basal levels of NGF, GDNF, or NT-3 mRNA in the hippocampus of juvenile or adult rats not subsequently subjected to immobilization stress. However, we did observe a significant effect of NI on the basal level of GDNF mRNA in juvenile rats. The neonatal isolation paradigm used in this study has been reported to alter plasma corticosterone responses to restraint stress in juveniles and adults (Erabi et al., 2007; McCormick et al., 2002). This paradigm has also been reported to alter the levels of insulin-like growth factor-I (IGF-1) receptor and IGF binding protein-2 in the hippocampus in response to adulthood restraint stress (Erabi et al., 2007). Thus, it is likely that this neonatal isolation paradigm can affect the expression of neurotrophins in response to immobilization stress later in life.

Two studies conducted by Crelli et al. (1998, 2000) examined the influence of early adverse experiences on the expression of NGF in the rat hippocampus. The former study showed that a single brief episode of maternal separation (45 min) on PN day 3 significantly increased the level of NGF mRNA in the hippocampus of 3-day-old rats (Cirulli et al., 1998). Similarly, the latter study indicated that a single 1-h episode of maternal separation on PN day 9 or a single 3-h episode of maternal separation on PN day 16 significantly increased NGF mRNA in the DG (Cirulli et al., 2000). However, we found no significant influence of neonatal isolation on expression of NGF mRNA in the hippocampus of 28- or 90-day-old rats, and it may be that the effects of early adverse experience may no longer manifest at PN day 28. Further studies are necessary to determine whether there is a specific period of time during which neonatal isolation can affect the levels of NGF mRNA in the hippocampus. In contrast to studies using NGF (Cirulli et al., 1998, 2000), there has been no previous study examining the influence of early adversity on the basal levels of GDNF and NT-3 mRNA in the adult rat hippocampus. We have shown here that the level of neither GDNF nor NT-3 mRNA is altered in the hippocampus of 28- or 90-day-old rats that had been subjected to repeated episodes of neonatal isolation. In contrast to what has been reported for BDNF (Roceri et al., 2002, 2004), we did not find any change in the basal level of

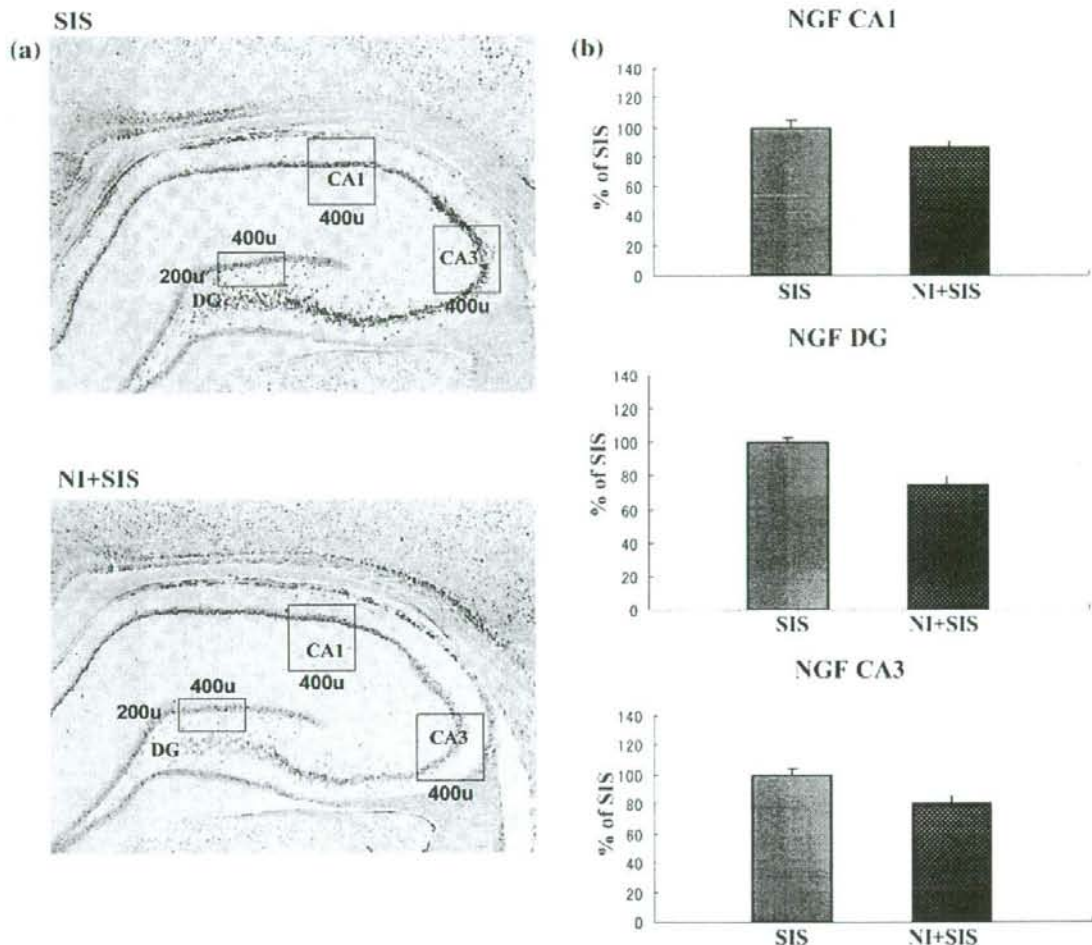


Fig. 3. Analysis of NGF mRNA expression by *in situ* hybridization in the adult rat hippocampus. (a) NGF mRNA expression in the rat hippocampus. SIS, single immobilization stress, NI + SIS, neonatal isolation followed by a single immobilization stress. (b) Mean density of NGF mRNA in the CA1 and CA3 pyramidal cell layers, and the dentate gyrus granular cell layer in the hippocam-

pus of adult rats subjected to single immobilization stress (SIS) and neonatal isolation followed by a single immobilization stress (NI + SIS). Results are expressed as the percentage of SIS levels. The mean \pm SEM (SIS: $n = 7$; NI + SIS: $n = 6$) is shown. * $P < 0.05$ compared with SIS (Mann-Whitney U test). CA, cornu ammonis; DG, dentate gyrus.

NGF, GDNF, or NT-3 mRNA in the hippocampus of juvenile or adult rats that had been subjected solely to repeated episodes of neonatal isolation.

In this study, we also observed that exposure of rats to neonatal isolation abrogated the increase in the level of NGF mRNA that subsequently occurs in the same rats exposed to a single episode of immobilization as juveniles or adults. Furthermore, the exposure of rats to neonatal isolation decreased the level of GDNF mRNA in response to a single immobilization in juvenile rats. Recently, maternal separation (from Day PN 2 to PN day 14) has been shown to affect the level of NGF and NT-3 in the hippocampus of rats in response to single swim stress on PN day 35 and PN

day 60 (Faure et al., 2006). Faure et al., (2006) demonstrated that adolescent and adult rats subjected to the swim stress expressed significantly higher levels of NGF and NT-3 in the hippocampus if they had been previously subjected to repeated episodes of maternal separation. Unfortunately, since they did not evaluate the influence of swim stress on the levels of NGF and NT-3 in the hippocampus of adult rats that had not been subjected to maternal separation as neonates, it is difficult to determine the effect of repeated episodes of maternal separation on the response of NGF and NT-3 to swim stress during adulthood.

Whereas subjection of adult rats to a single 8-h episode of immobilization stress leads to a significant

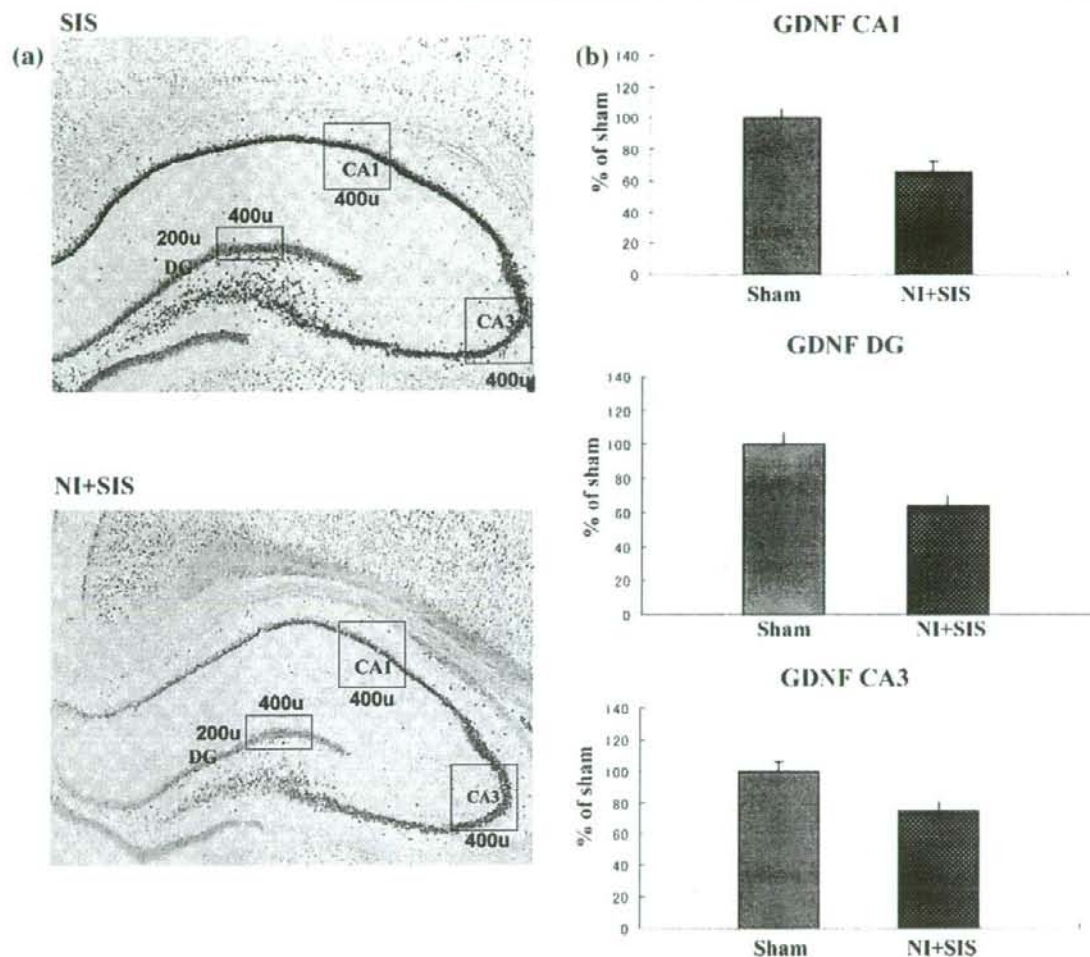


Fig. 4. Analysis of GDNF mRNA expression by in situ hybridization in the adult rat hippocampus. (a) GDNF mRNA expression in the rat hippocampus. NI, neonatal isolation; NI + SIS, neonatal isolation followed by a single immobilization stress. (b) Mean density of GDNF mRNA in the CA1 and CA3 pyramidal cell layers, and the dentate gyrus granular cell layer in the hippocampus of adult

rats subjected to neonatal isolation (NI) and neonatal isolation followed by a single immobilization stress (NI + SIS). Results are expressed as the percentage of the SIS levels. The mean \pm SEM (NI: $n = 7$; NI + SIS: $n = 6$) is shown. * $P < 0.05$ compared with NI (Mann-Whitney U test). CA, cornu ammonis; DG, dentate gyrus.

reduction in the level of NGF in the hippocampus (Ueyama et al., 1997), traumatic brain injury (Grundy et al., 2001), subchronic cold stress (Foreman et al., 1993), immobilization stress (Smith and Clazza, 1996; Smith et al., 1995), and glucocorticoids (Mocchetti et al., 1996; Sun et al., 1993) significantly increase the expression of NGF in the rat hippocampus. These data indicating that stress-induced activation of the hypothalamo-pituitary-adrenal (HPA) axis upregulates the expression of NGF in the hippocampus, are consistent with the present findings. On the basis of the functional properties of NGF (Semkova and Kriegstein, 1999; Sofroniew et al., 2001; Tabakman

et al., 2005), the induction of NGF in response to stress may be associated with neuroprotective responses. As such, the decreased response of NGF induction in both juvenile and adult rats and the decreased level of GDNF in juvenile, but not adult, rats in response to immobilization stress in the adult rats subjected to neonatal isolation may be, at least in part, involved in the stress susceptibility in later life due to neonatal isolation.

It is unclear how neonatal isolation attenuates the increased expression of NGF and GDNF in response to adulthood immobilization stress. Since activation of the HPA axis increases NGF expression in the rat

brain (Mocchetti et al., 1996; Sun et al., 1993), it is hypothesized that the decreased response of the HPA axis to adulthood immobilization may account for the decreased induction of NGF in the rats subjected to neonatal isolation. However, we have recently reported that the corticosterone response following a single 2-h restraint stress in adult rats previously subjected to neonatal isolation was significantly enhanced compared with the effect in adult rats not previously subjected to neonatal isolation (Erabi et al., 2007). On the basis of our recent finding, it is unlikely that the HPA axis is involved in attenuation of the induction of NGF in adult rats that had been previously subjected to neonatal isolation.

Early adverse experiences have been suggested to alter epigenetic programming through DNA methylation and histone acetylation (Weaver et al., 2004). Weaver et al. (2004) demonstrated that low levels of maternal care markedly induced cytosine methylation in the promoter of the glucocorticoid receptor (GR). This increased methylation inhibited the stimulation of GR mRNA expression in response to restraint stress in adult rats, due to inhibition of the binding of NGF1A to the GR promoter. In analogy, the induction of cytosine methylation in transcription-factor binding regions of the promoters of NGF or GDNF genes following neonatal isolation might lead to a reduction of the induction of NGF and GDNF in response to immobilization stress in adult rats. Further studies examining the effect of neonatal isolation on cytosine methylation in the promoters of these genes are needed to elucidate the mechanism of the inhibition of gene induction.

In summary, repeated episodes of neonatal isolation inhibit the induction of NGF in the hippocampus of juvenile and adult rats, and decrease the levels of GDNF in the hippocampus of juvenile rats in response to a SIS, but have no significant effect on the basal expression of these mRNAs in the absence of immobilization stress. These findings suggest that the susceptibility to stress derived from experience of neonatal isolation might play a role in decreased neuroprotective support in response to subsequent exposure to stress later in life through the down regulation of NGF induction and decreased expression of GDNF.

REFERENCES

- Cirulli F, Micera A, Allea E, Aloe L. 1998. Early maternal separation increases NGF expression in the developing rat hippocampus. *Pharmacol Biochem Behav* 59:853-858.
- Cirulli F, Allea E, Antonelli A, Aloe L. 2000. NGF expression in the developing rat brain: Effects of maternal separation. *Brain Res Dev Brain Res* 123:129-134.
- Erabi K, Morinobu S, Kawano K, Tsuji S, Yamawaki S. 2007. Neonatal isolation changes the expression of IGF-1R and IGFBP-2 in the hippocampus in response to adulthood restraint stress. *Int J Neuropsychopharmacol* 10:369-381.
- Faure J, Uys JDK, Marals L, Stein DJ, Daniels WMU. 2007. Early maternal separation followed by later stressors leads to dysregulation of the HPA-axis and increases in hippocampal NGF and NT-3 levels in adult male. *Metab Brain Dis* 22:183-195.
- Folli F, Ghidella S, Bonfanti L, Kahn CR, Merighi A. 1996. The early intracellular signaling pathway for the insulin/insulin-like growth factor receptor family in the mammalian central nervous system. *Mol Neurobiol* 13:155-183.
- Foreman PJ, Tagliatela G, Angelucci L, Turner CP, Perez-Polo JR. 1993. Nerve growth factor and p75NGFR factor receptor mRNA change in rodent CNS following stress activation of the hypothalamic-pituitary-adrenocortical axis. *J Neurosci Res* 36:10-18.
- Greisen MH, Altar CA, Bolwig TG, Whitehead R, Wortwein G. 2005. Increased adult hippocampal brain-derived neurotrophic factor and normal levels of neurogenesis in maternal separation rats. *J Neurosci Res* 79:772-778.
- Grundly PL, Patel N, Harbuz MS, Lightman SL, Sharples PM. 2001. Glucocorticoids modulate the NGF mRNA response in the rat hippocampus after traumatic brain injury. *Brain Res* 892:386-390.
- Hayashi A, Nagaoka M, Yamada K, Ichitani Y, Miake Y, Okado N. 1998. Maternal stress induces synaptic loss and developmental disabilities of offspring. *Int J Dev Neurosci* 16:209-216.
- Huot RL, Plotsky PM, Lenox RH, McNamara RK. 2002. Neonatal maternal separation reduces hippocampal mossy fiber density in adult Long Evans rats. *Brain Res* 950:52-63.
- Kehoe P, Bronzino JD. 1999. Neonatal stress alters LTP in freely moving male and female adult rats. *Hippocampus* 9:651-658.
- Lee HJ, Kim JW, Yim SV, Kim MJ, Kim SA, Kim YJ, Kim CJ, Chung JH. 2001. Fluoxetine enhances cell proliferation and prevents apoptosis in dentate gyrus of maternally separated rats. *Mol Psychiatry* 6:725-728.
- Lin LF, Doherty DH, Lile JD, Bektess S, Collins F. 1993. GDNF: A glial cell line-derived neurotrophic factor for midbrain dopaminergic neurons. *Science* 260:1130-1132.
- Liu D, Diorio J, Day JC, Francis DD, Meaney MJ. 2000. Maternal care, hippocampal synaptogenesis and cognitive development in rats. *Nat Neurosci* 3:799-806.
- MacQueen GM, Ramakrishnan K, Ratnasigan R, Chen B, Young LT. 2003. Desipramine treatment reduces the long-term behavioural and neurochemical sequelae of early-life maternal separation. *Int J Neuropsychopharmacol* 6:391-396.
- Magarinos AM, McEwen BS. 1995. Stress-induced atrophy of apical dendrites of hippocampal CA3c neurons: Comparison of stressors. *Neuroscience* 69:83-88.
- Magarinos, McEwen BS, Flugge G, Fuchs E. 1996. Chronic psychosocial stress causes apical dendritic atrophy of hippocampal CA3 pyramidal neurons in subordinate tree shrews. *J Neurosci* 16:3534-3540.
- Maisonpierre PC, Belluscio L, Friedman B, Alderson RF, Wiegand SJ, Furth ME, Lindsay RM, Yancopoulos GD. 1990. NT-3, BDNF, and NGF in the developing rat nervous system: Parallel as well as reciprocal patterns of expression. *Neuron* 5:501-509.
- McCormick CM, Kehoe P, Mallinson K, Cecchi L, Frye CA. 2002. Neonatal isolation alters stress hormone and mesolimbic dopamine release in juvenile rats. *Pharmacol Biochem Behav* 73:77-85.
- Mocchetti I, Spiga G, Hayes VY, Isackson PJ, Colangelo A. 1996. Glucocorticoids differentially increase nerve growth factor and basic fibroblast factor expression in the rat brain. *J Neurosci* 16:2141-2148.
- Morinobu S, Fujimaki K, Kawano K, Tanaka K, Takahashi J, Ohkawa M, Yamawaki S, Kato N. 2003. Influence of immobilization stress on the expression and phosphatase activity of protein phosphatase 2A in the rat brain. *Biol Psychiatry* 54:1060-1066.
- Nosrat CA, Fried K, Lindskog S, Olson L. 1997. Cellular expression of neurotrophin mRNAs during tooth development. *Cell Tissue Res* 290:569-580.
- Poeggel G, Helmcke C, Abraham A, Schwabe T, Friedrich P, Braun K. 2003. Juvenile emotional experience alters synaptic composition in the rodent cortex, hippocampus, and lateral amygdala. *Proc Natl Acad Sci USA* 100:16137-16142.
- Roceri M, Hendriks W, Racagni G, Ellenbroek BA, Riva MA. 2002. Early maternal deprivation reduces the expression of BDNF and NMDA receptor subunits in rat hippocampus. *Mol Psychiatry* 7:609-616.
- Roceri M, Cirulli F, Pessina C, Peretto P, Racagni G, Riva MA. 2004. Postnatal repeated maternal deprivation produces age-dependent changes of brain-derived neurotrophic factor expression in selected rat brain regions. *Biol Psychiatry* 55:708-714.
- Semkova I, Krieglstein J. 1999. Neuroprotection mediated via neurotrophic factors and induction of neurotrophic factors. *Brain Res Brain Res Rev* 30:176-188.
- Smith MA, Clzsa G. 1996. Stress-induced changes in brain-derived neurotrophic factor expression are attenuated in aged Fischer 344/N rats. *Neurobiol Aging* 17:859-864.

- Smith MA, Makino S, Kvetnansky R, Post RM. 1995. Stress and glucocorticoids affect the expression of brain-derived neurotrophic factor and neurotrophin-3 mRNAs in the hippocampus. *J Neurosci* 15:1768-1777.
- Sofroniew MV, Howe CL, Mobley WC. 2001. Nerve growth factor signaling, neuroprotection, and neural repair. *Annu Rev Neurosci* 24:1217-1281.
- Suenaga T, Morinobu S, Kawano K, Sawada T, Yamawaki S. 2004. Influence of immobilization stress on the levels of CaMKII and phospho-CaMKII in the rat hippocampus. *Int J Neuropsychopharmacol* 7:299-309.
- Sun FY, Costa E, Mochetti I. 1993. Adrenal steroids mediate the increases of hippocampal nerve growth factor biosynthesis following bicuculline convulsions. *Neuropharmacol* 8:219-225.
- Tabakman P, Jiang H, Shahar I, Arien-Zakay H, Levine RA, Lazarovici P. 2005. Neuroprotection by NGF in the PC12 in vitro OGD model: Involvement of mitogen-activated protein kinases and gene expression. *Ann N Y Acad Sci* 1053:84-96.
- Ueyama T, Kawai Y, Nemoto K, Sekimoto M, Tone S, Senba E. 1997. Immobilization stress reduced the expression of neurotrophins and their receptors in the rat brain. *Neurosci Res* 28:103-110.
- Vyas A, Mitra R, Rao S, Chatterji S. 2002. Chronic stress induces contrasting patterns of dendritic remodeling in hippocampal and amygdaloid neurons. *J Neurosci* 22:6810-6818.
- Watanabe Y, Gould E, McEwen BS. 1992. Stress induces atrophy of apical dendrites of hippocampal CA3 pyramidal neurons. *Brain Res* 588:341-345.
- Weaver IC, Cervoni N, Champagne FA, D'Alessio AC, Sharma S, Seckl JR, Dymov S, Szyf M, Meaney MJ. 2004. Epigenetic programming by maternal behavior. *Nat Neurosci* 7:847-854.
- Wetmore C, Bygdeman M, Olson L. 1992. High and low affinity NGF receptor mRNAs in human fetal spinal cord and ganglia. *Neuroreport* 3:689-692.

Regular Article

'Insistence on recovery' as a positive prognostic factor in Japanese stroke patients

Seiji Hama, MD,^{1*} Hidehisa Yamashita, MD,² Toyohiko Kato, CP,³ Masaya Shigenobu, MD,¹ Atsuko Watanabe, MD,¹ Megumi Sawa, CP,¹ Kaoru Kurisu, MD,⁴ Shigeto Yamawaki, MD² and Tamotsu Kitaoka, MD¹

¹Department of Rehabilitation, Nishi-Hiroshima Rehabilitation Hospital, ²Department of Neurosurgery and ³Department of Psychiatry and Neuroscience, Graduate School of Biomedical Science Hiroshima University, Hiroshima and ⁴Department of Teacher Education, Kinki University, Osaka, Japan

Aim: The present study used two-step analyses to examine the effect of acceptance of disability or 'insistence on recovery' in Japanese stroke patients: first on their functional improvement and second, on their psychological symptoms.

Methods: Disability was assessed using functional independence measurements (FIM), examining the stage of acceptance of disability by observation using Fink's theory (from shock to defensive retreat, acknowledgement, and acceptance/change stage), and estimation of insistence on recovery (on a scale of 1–4) by observation. The differences over time and the effects on the improvement in their FIM were then assessed. Depression was measured using the Zung Self-rating Depression Scale (SDS); apathy was measured using the Apathy Scale (AS), and the correlation with the acceptance stage or insistence on recovery was analyzed.

Results: The acceptance stage and functional improvement progressed significantly, but insistence

on recovery did not change significantly during hospitalization. Multiple regression indicated that the insistence on recovery score (but not the acceptance stage) was a good predictor of the degree of improvement in FIM (FIM gain per week) in the elderly group. Post-hoc testing showed that the SDS or AS score decreased from the first stage to the fourth stage (but increased at the third stage) of acceptance; whereas for insistence on recovery score, the SDS and AS scores decreased as insistence on recovery score changed from 1 to 3, and then increased as insistence on recovery score changed from 3 to 4.

Conclusions: The appropriate level of insistence on recovery reduced depression and apathy, resulting in enhanced improvement of disability after a stroke in elderly stroke patients.

Key words: acceptance of disability, elderly, functional independence measurement, insistence on recovery, stroke.

STROKE HAS BEEN described as a condition with a unique epidemiological profile, with high incidence and mortality rates, and in which a large proportion of survivors have significant but varying

degrees of residual disability.^{1,2} There are two types of improvement that occur after a stroke: neurological improvement and improvement in functional abilities or performance.¹ Neurological recovery depends upon the mechanism of the stroke and the location of the lesion. By contrast, the improvement of functional abilities, such as activities of daily living (ADL), depends upon the environment in which the stroke patient is placed and how much training and motivation there is for the patient to learn to become independent again in terms of self-care and mobility. The ability to perform ADL can improve through

*Correspondence: Seiji Hama, MD, Nishi-Hiroshima Rehabilitation Hospital, Miyake 265, Saeki-ku, Hiroshima 731-5143, Japan. Email: shama@hiroshima-u.ac.jp

¹Present address: Department of Neurosurgery, Graduate School of Biomedical Science Hiroshima University, Kasumi 1-2-3, Minami-ku, Hiroshima 734-8551, Japan.

Received 1 November 2006; revised 10 December 2007; accepted 13 March 2008.

acceptance and training in the presence or absence of natural neurological recovery.¹⁻⁴ Stroke rehabilitation must therefore restore optimal physical, psychological, social, and vocational function to enable the patient to become a productive participant in the community.

Emotional responses to stroke have traditionally been thought to follow a natural course of evolution: an initial state of significant distress or depression that, over time and as a result of some active process of 'working through', resolves to a condition of acceptance and relative emotional harmony.^{5,6} This process was thought to be a mourning process or acceptance of disability. Disabled stroke patients who refuse to accept their impairment, eventually realize they are sick but continue to think that they will soon get well.^{5,6} They manifested this attitude through direct verbal report or inferred from behavior continuously and repeatedly (this attitude was similar to whining, and we termed this emotional status as 'insistence on recovery'). Insistence on recovery is thought to be a sign of denial and thus is regarded as an irrational belief. Denial is generally found in psychiatric illness, but it is also found in the mourning process among disabled stroke patients.⁶ It was traditionally thought that patients with insistence on recovery, whose only goal is recovery, can be motivated to do any work perceived as aiding recovery; but are not motivated to learn to function as a disabled person, and so fail to gain the maximum benefit from rehabilitation services.⁵⁻⁸ Thus insistence on recovery is considered a maladaptive status and is thought to be a target for psychotherapeutic intervention.⁵⁻⁸ We have, however, experienced many stroke disabled patients who say repeatedly that they will recover someday, and who participate in rehabilitation and gain independence in ADL in the rehabilitation service. The question arises whether insistence on recovery is an irrational or rational belief. Although there is much popular and professional literature attesting to the veracity of stages of acceptance of disability, the empirical data do not support such a contention.⁵ Moreover, insistence on recovery is regarded as counterproductive to maximizing functional abilities and enhancing quality of life,⁵⁻⁸ and no previous paper has examined the nature of 'insistence on recovery'.

The aim of the present study was to evaluate the effect of the stage of acceptance of disability or insistence on recovery on functional independence in stroke patients. Aging is associated with a high incidence of physical impairment, functional disability

and depression.^{9,10} Therefore, we divided stroke inpatients into two age groups: middle-aged and elderly. Moreover, depression and apathy are common neuropsychiatric consequences of a stroke, and can be examined using self-reported tests: the Zung Self-rating Depression Scale (SDS) and the Apathy Scale (AS). To clarify the psychopathological aspects of insistence on recovery after a stroke, we also examined the correlation between insistence on recovery score and the SDS or AS score.

METHODS

Study design

This study consisted of two parts: first, the changes in 'acceptance of disability' and 'insistence on recovery' over time were estimated, and we examined the effect of these factors on functional recovery after a stroke. Second, we examined the psychopathological symptoms (depression and apathy) using self-reporting scales, and estimated the association between depression and 'acceptance of disability' or 'insistence on recovery' after a stroke.

Patients

The approval of institutional ethics committees was obtained for this prospective study. Informed consent for functional or psychological measurements, including acceptance of disability, was obtained on admission from all patients or from those authorized to give consent on their behalf. The subjects for the first study were 231 patients with hemorrhagic and occlusive stroke without subarachnoid hemorrhage, diagnosed on computed tomography (CT) and/or magnetic resonance imaging (MRI), who were admitted to the Nishi-Hiroshima Rehabilitation Hospital <3 months after suffering a stroke (range, 10-109 days; mean, 44 ± 21 days), and who were hospitalized for more than 1 month. The patients were divided into two age groups at admission: the middle-aged group (95 patients, age 40-65 years); and the elderly group (136 patients, aged ≥ 66 years).

For the second study the subjects, who completed the SDS and AS scales, included 237 patients after excluding patients with (i) a history of major psychiatric illness, such as major depression, bipolar disorder, schizophrenia, or schizoaffective disorder; (ii) subarachnoid hemorrhage; (iii) emergency discharge