

recently reported that S1P, a lipid mediator, contributes to the regulation of migration of pathogenic CD4 T and mast cells from the systemic immune compartment into the large intestine and inhibition of S1P-mediated pathway results in the inhibition of development of allergic diarrhea (205).

Celiac Disease

Celiac disease is another related disorder of mucosal immunity, which is characterized by small-intestinal mucosal injury in response to the dietary ingestion of gluten (29). Gluten is a proline- and glutamine-rich protein that is found in wheat, rye, and barley. Pathologic features of celiac disease include increased numbers of IELs and less extensive villous atrophy and crypt hypertrophy (29). IELs from patients with celiac disease preferentially include NK-like cells, which recognize stress-induced MICA molecules expressed on IECs (29). This process has been considered to be mediated by IL-15. IL-15 induces increased expression of MICA and its receptor, NKG2D, and up-regulates gluten-specific CTL activity in the small intestine (206). Gluten-free diet currently is the only accepted therapy for celiac disease, and various immunological approaches such as blocking IL-15 and treatment with IL-10 have been examined (29). However, in terms of quality of life, it is doubtful that this treatment will be effective with so many potential side effects. Thus, it is essential to identify the gluten-specific pathogenic immunocompetent cell population for the development of novel strategies to selectively delete the specific pathogenic populations.

ACKNOWLEDGMENTS

We thank all members of our laboratories for their assistance with the preparation of this chapter. The experimental results included in this chapter were supported by U.S. Public Health grant U19 AI 28147; MSM 0021620812 from the Czech Republic; Core Research for Evolutional Science and Technology (CREST) of the Japan Science and Technology Corporation (JST), the Ministry of Education, Science, Sports, and Culture, and the Ministry of Health and Welfare in Japan.

REFERENCES

1. Cone RA. Mucus. *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:35–48.
2. Furuse M, Tsukita S. Claudins in occluding junctions of humans and flies. *Trends Cell Biol*. 2006;16:181–188.
3. Vijay-Kumar M, Gewirtz AT. Role of epithelium in mucosal immunity. *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:423–434.
4. Lehrer RI, Bevins CL, Ganz T. Defensins and other antimicrobial peptides and proteins. *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:73–94.
5. Russell MW, Bobek LA, Brock JH, et al. Innate Humoral Defense Factors. *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:73–94.
6. Kunisawa J, McGhee J, Kiyono H. Mucosal S-IgA enhancement: development of safe and effective mucosal adjuvants and mucosal antigen delivery vehicles. In: *Mucosal Immune Defense: Immunoglobulin A*. Kaetzel C, ed. New York: Kluwer Academic/Plenum Publishers. 2007:346–389.
7. Chaby R, Garcia-Verdugo I, Espinassous O, et al. Interactions between LPS and lung surfactant proteins. *J Endotoxin Res*. 2005;11:181–185.
8. Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science*. 2004;303:1532–1535.
9. Kaetzel CS. The polymeric immunoglobulin receptor: bridging innate and adaptive immune responses at mucosal surfaces. *Immunol Rev*. 2005;206:83–99.
10. Lencer WI, Blumberg RS. A passionate kiss, then run: exocytosis and recycling of IgG by FcRn. *Trends Cell Biol*. 2005;15:5–9.
11. Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell*. 2006;124:783–801.
12. Lee MS, Kim YJ. Signaling pathways downstream of pattern-recognition receptors and their cross talk. *Annu Rev Biochem*. 2007;76:447–480.
13. Cario E, Podolsky DK. Toll-like receptor signaling and its relevance to intestinal inflammation. *Ann NY Acad Sci*. 2006;1072:332–338.
14. Kelly D, Conway S, Aminov R. Commensal gut bacteria: mechanisms of immune modulation. *Trends Immunol*. 2005;26:326–333.
15. Lotz M, Gutle D, Walther S, et al. Postnatal acquisition of endotoxin tolerance in intestinal epithelial cells. *J Exp Med*. 2006;203:973–984.
16. Lee J, Mo JH, Kataoka K, et al. Maintenance of colonic homeostasis by distinctive apical TLR9 signalling in intestinal epithelial cells. *Nat Cell Biol*. 2006;8:1327–1336.
17. Schenk M, Mueller C. Adaptations of intestinal macrophages to an antigen-rich environment. *Semin Immunol*. 2007;19:84–93.
18. Kuwata H, Matsumoto M, Altarashi K, et al. L_xBNS inhibits induction of a subset of Toll-like receptor-dependent genes and limits inflammation. *Immunity*. 2006;24:41–51.
19. Uematsu S, Jiang MH, Chevrier N, et al. Detection of pathogenic intestinal bacteria by Toll-like receptor 5 on intestinal CD11c⁺ lamina propria cells. *Nat Immunol*. 2006;7:868–874.
20. Munn DH, Sharma MD, Lee JR, et al. Potential regulatory function of human dendritic cells expressing indoleamine 2,3-dioxygenase. *Science*. 2002;297:1867–1870.
21. Watanabe T, Kitani A, Murray PJ, et al. Nucleotide binding oligomerization domain 2 deficiency leads to dysregulated TLR2 signaling and induction of antigen-specific colitis. *Immunity*. 2006;25:473–485.
22. Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, et al. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell*. 2004;118:229–241.
23. Hysi P, Kabesch M, Moffatt MF, et al. NOD1 variation, immunoglobulin E and asthma. *Hum Mol Genet*. 2005;14:935–941.
24. Nigo YI, Yamashita M, Hirahara K, et al. Regulation of allergic airway inflammation through Toll-like receptor 4-mediated modification of mast cell function. *Proc Natl Acad Sci U S A*. 2006;103:2286–2291.
25. Kunisawa J, Takahashi I, Kiyono H. Intraepithelial lymphocytes: their shared and divergent immunological behaviors in the small and large intestine. *Immunol Rev*. 2007;215:136–153.
26. Cepek KL, Shaw SK, Parker CM, et al. Adhesion between epithelial cells and T lymphocytes mediated by E-cadherin and the $\alpha\beta\gamma$ integrin. *Nature*. 1994;372:190–193.
27. Cheroutre H. Starting at the beginning: new perspectives on the biology of mucosal T cells. *Annu Rev Immunol*. 2004;22:217–246.
28. Lefrancois L. Cytotoxic T cells of the mucosal immune system. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:559–564.

29. Kagnoff MF. Celiac disease: pathogenesis of a model immunogenetic disease. *J Clin Invest.* 2007;117:41–49.
30. Staton TL, Habtezion A, Winslow MM, et al. CD8⁺ recent thymic emigrants home to and efficiently repopulate the small intestine epithelium. *Nat Immunol.* 2006;7:482–488.
31. Lambbolez F, Kronenberg M, Cheroutre H. Thymic differentiation of TCR β^+ CD8 α^+ IELs. *Immunol Rev.* 2007;215:178–188.
32. Ishikawa H, Naito T, Iwanaga T, et al. Curriculum vitae of intestinal intraepithelial T cells: their developmental and behavioral characteristics. *Immunol Rev.* 2007;215:154–165.
33. Kunisawa J, Kurashima Y, Higuchi M, et al. Sphingosine 1-phosphate dependence in the regulation of lymphocyte trafficking to the gut epithelium. *J Exp Med.* 2007;204:2335–2348.
34. Ishikawa H, Kanamori Y, Hamada H, et al. Development and function of organized gut-associated lymphoid tissues. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:385–406.
35. Saito H, Kanamori Y, Takemori T, et al. Generation of intestinal T cells from progenitors residing in gut cryptopatches. *Science.* 1998;280:275–278.
36. Bendelac A, Savage PB, Teitton L. The biology of NKT cells. *Annu Rev Immunol.* 2007;25:297–336.
37. Mayer L, Blumberg RS. Role of epithelial cells in mucosal antigen presentation. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:435–450.
38. Treiner E, Duban L, Bahram S, et al. Selection of evolutionarily conserved mucosal-associated invariant T cells by MR1. *Nature.* 2003;422:164–169.
39. Croxford JL, Miyake S, Huang YY, et al. Invariant V α 19 γ T cells regulate autoimmune inflammation. *Nat Immunol.* 2006;7:987–994.
40. Bienzenstock J, McDermott MR. Bronchus- and nasal-associated lymphoid tissues. *Immunol Rev.* 2005;206:22–31.
41. Kiyono H, Fukuyama S. NALT versus Peyer's-patch-mediated mucosal immunity. *Nat Rev Immunol.* 2004;4:699–710.
42. Kunisawa J, Fukuyama S, Kiyono H. Mucosa-associated lymphoid tissues in aerodigestive tract: their shared and divergent traits and their importance to the orchestration of mucosal immune system. *Curr Mol Med.* 2005;5:557–572.
43. Debertin AS, Tscherning T, Tonjes H, et al. Nasal-associated lymphoid tissue (NALT): frequency and localization in young children. *Clin Exp Immunol.* 2003;134:503–507.
44. Neutra MR, Kraehenbuhl JP. Cellular and molecular basis for antigen transport across epithelial barriers. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:111–132.
45. Kerneis S, Bogdanov A, Kraehenbuhl JP, et al. Conversion by Peyer's patch lymphocytes of human enterocytes into M cells that transport bacteria. *Science.* 1997;277:949–952.
46. Brandtzaeg P, Johansen FE. Mucosal B cells: phenotypic characteristics, transcriptional regulation, and homing properties. *Immunol Rev.* 2005;206:32–63.
47. Cebray JJ, Jiang HO, Boiko NV, et al. The role of mucosal microbiota in the development, maintenance, and pathologies of the mucosal immune system. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:335–368.
48. Fujishashi K, McGhee J. Th1/Th2/Th3 cells for regulation of mucosal immunity, tolerance, and inflammation. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:539–558.
49. Wagner N, Lohler J, Kunkel EJ, et al. Critical role for β 7 integrins in formation of the gut-associated lymphoid tissue. *Nature.* 1996;382:366–370.
50. Youngman K, Lazarus N, Butcher EC. Lymphocyte homing: chemokines and adhesion molecules in T cell and IgA plasma cell. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:667–680.
51. Johansson C, Kelsall BL. Phenotype and function of intestinal dendritic cells. *Semin Immunol.* 2005;17:284–294.
52. Iwasaki A. Mucosal dendritic cells. *Annu Rev Immunol.* 2007;25:381–418.
53. Cook DN, Prosser DM, Forster R, et al. CCR6 mediates dendritic cell localization, lymphocyte homeostasis, and immune responses in mucosal tissue. *Immunity.* 2000;12:495–503.
54. Fagarasan S, Shinkura R, Kamata T, et al. Alymphoplasia (aly)-type nuclear factor kappaB-inducing kinase (NIK) causes defects in secondary lymphoid tissue chemokine receptor signaling and homing of peritoneal cells to the gut-associated lymphoid tissue system. *J Exp Med.* 2000;191:1477–1486.
55. Fukuda K, Yoshida H, Sato T, et al. Mesenchymal expression of Foxl1, a winged helix transcriptional factor, regulates generation and maintenance of gut-associated lymphoid organs. *Dev Biol.* 2003;255:278–289.
56. Veiga-Fernandes H, Coles MC, Foster KE, et al. Tyrosine kinase receptor RET is a key regulator of Peyer's patch organogenesis. *Nature.* 2007;446:547–551.
57. Fagarasan S, Muramatsu M, Suzuki K, et al. Critical roles of activation-induced cytidine deaminase in the homeostasis of gut flora. *Science.* 2002;298:1424–1427.
58. Kweon MN, Yamamoto M, Rennert PD, et al. Prenatal blockage of lymphotxin beta receptor and TNF receptor p55 signaling cascade resulted in the acceleration of tissue genesis for isolated lymphoid follicles in the large intestine. *J Immunol.* 2005;174:4365–4372.
59. Brayden DJ, Jepson MA, Baird AW. Keynote review: intestinal Peyer's patch M cells and oral vaccine targeting. *Drug Discov Today.* 2005;10:1145–1157.
60. Tellebaum R, Schubert W, Gunther I, et al. The M cell as a portal of entry to the lung for the bacterial pathogen *Mycobacterium tuberculosis*. *Immunity.* 1999;10:641–650.
61. Weaver CT, Harrington LE, Mangan PR, et al. Th17: an effector CD4 T cell lineage with regulatory T cell ties. *Immunity.* 2006;24:677–688.
62. Mangan PR, Harrington LE, O'Quinn DB, et al. Transforming growth factor- β induces development of the T(H)17 lineage. *Nature.* 2006;441:231–234.
63. Bettelli E, Carrier Y, Gao W, et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature.* 2006;441:235–238.
64. Ward RL, Greenberg HB, Estes MK. Viral gastroenteritis vaccines. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:887–904.
65. Schmid DS, Rouse BT. Respiratory viral vaccines. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:923–936.
66. Strober W, Fagarasan S, Lycke N. IgA B cell development. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:583–616.
67. Mega J, Bruce MG, Beagley KW, et al. Regulation of mucosal responses by CD4 $^+$ T lymphocytes: effects of anti-L3T4 treatment on the gastrointestinal immune system. *Int Immunol.* 1991;3:793–805.
68. Kawanishi H, Ozato K, Strober W. The proliferative response of cloned Peyer's patch T cells to syngeneic and allogeneic stimuli. *J Immunol.* 1985;134:3586–3591.
69. Zan H, Cerutti A, Dramitinos P, et al. CD40 engagement triggers switching to IgA1 and IgA2 in human B cells through induction of endogenous TGF- β : evidence for TGF- β but not IL-10-dependent direct $Sp1 \rightarrow S\alpha$ and sequential $Sp1 \rightarrow S\gamma, S\gamma \rightarrow S\alpha$ DNA recombination. *J Immunol.* 1998;161:5217–5225.
70. Benson EB, Strober W. Regulation of IgA secretion by T cell clones derived from the human gastrointestinal tract. *J Immunol.* 1988;140:1874–1882.
71. Lebman DA, Lee FD, Coffman RL. Mechanism for transforming growth factor β and IL-2 enhancement of IgA expression in lipopolysaccharide-stimulated B cell cultures. *J Immunol.* 1990;144:952–959.
72. Sonoda E, Matsumoto R, Hitoshi Y, et al. Transforming growth factor β induces IgA production and acts additively with interleukin 5 for IgA production. *J Exp Med.* 1989;170:1415–1420.

73. Rousset F, Garcia E, Banchereau J. Cytokine-induced proliferation and immunoglobulin production of human B lymphocytes triggered through their CD40 antigen. *J Exp Med.* 1991;173:705–710.
74. Defrance T, Vanbervliet B, Briere F, et al. Interleukin 10 and transforming growth factor β cooperate to induce anti-CD40-activated naive human B cells to secrete immunoglobulin A. *J Exp Med.* 1992;175:671–682.
75. Cazac BB, Roes J. TGF- β receptor controls B cell responsiveness and induction of IgA *in vivo*. *Immunity.* 2000;13:443–451.
76. Jung S, Rajewsky K, Radbruch A. Shutdown of class switch recombination by deletion of a switch region control element. *Science.* 1993;259:984–987.
77. Harriman GR, Bradley A, Das S, et al. IgA class switch in $I \alpha$ exon-deficient mice. Role of germline transcription in class switch recombination. *J Clin Invest.* 1996;97:477–485.
78. Honjo T, Nagaoka H, Shinkura R, et al. AID to overcome the limitations of genomic information. *Nat Immunol.* 2005;6:655–661.
79. Shikina T, Hiroi T, Iwatsuki K, et al. IgA class switch occurs in the organized nasopharynx-gut-associated lymphoid tissue, but not in the diffuse lamina propria of airways and gut. *J Immunol.* 2004;172:6259–6264.
80. Fagarasan S, Kinoshita K, Muramatsu M, et al. *In situ* class switching and differentiation to IgA-producing cells in the gut lamina propria. *Nature.* 2001;413:639–643.
81. Macpherson AJ, Lamarre A, McCoy K, et al. IgA production without μ or δ chain expression in developing B cells. *Nat Immunol.* 2001;2:625–631.
82. Iwasato T, Arakawa H, Shimizu A, et al. Biased distribution of recombination sites within S regions upon immunoglobulin class switch recombination induced by transforming growth factor β and lipopolysaccharide. *J Exp Med.* 1992;175:1539–1546.
83. Brandtzæg P, Surjan L, Jr., Berdal P. Immunoglobulin-producing cells in clinically normal, hyperplastic and inflamed human palatine tonsils. *Acta Otolaryngol Suppl.* 1979;360:211–215.
84. Craig SW, Cebray JJ. Peyer's patches: an enriched source of precursors for IgA-producing immunocytes in the rabbit. *J Exp Med.* 1971;134:188–200.
85. Cyster JG. Chemokines, sphingosine-1-phosphate, and cell migration in secondary lymphoid organs. *Annu Rev Immunol.* 2005;23:127–159.
86. Farstad IN, Halstensen TS, Kvale D, et al. Topographic distribution of homing receptors on B and T cells in human gut-associated lymphoid tissue: relation of L-selectin and integrin $\alpha 4\beta 7$ to naive and memory phenotypes. *Am J Pathol.* 1997;150:187–199.
87. Agace WW. Tissue-tropic effector T cells: generation and targeting opportunities. *Nat Rev Immunol.* 2006;6:682–692.
88. Iwata M, Hirakiyama A, Eshima Y, et al. Retinoic acid imprints gut-homing specificity on T cells. *Immunity.* 2004;21:527–538.
89. Mora JR, Iwata M, Eksteen B, et al. Generation of gut-homing IgA-secreting B cells by intestinal dendritic cells. *Science.* 2006;314:1157–1160.
90. Quiding-Jarbrink M, Nordstrom I, Granstrom G, et al. Differential expression of tissue-specific adhesion molecules on human circulating antibody-forming cells after systemic, enteric, and nasal immunizations. A molecular basis for the compartmentalization of effector B cell responses. *J Clin Invest.* 1997;99:1281–1286.
91. Mestecky J, Moro I, Kerr MA, et al. Mucosal Immunoglobulins. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:153–182.
92. Kutte WH, Koopman WJ, Conley ME, et al. Production of predominantly polymeric IgA by human peripheral blood lymphocytes stimulated *in vitro* with mitogens. *J Exp Med.* 1980;152:1424–1429.
93. Kilian M, Russell MW. Microbial evasion of IgA function. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:291–303.
94. Halpern MS, Koshland ME. Novel subunit in secretory IgA. *Nature.* 1970;228:1276–1278.
95. Sorensen V, Sundvold V, Michaelsen TE, et al. Polymerization of IgA and IgM: roles of Cys309/Cys414 and the secretory tailpiece. *J Immunol.* 1999;162:3448–3455.
96. Monteiro RC, Van De Winkel JG. IgA Fc receptors. *Annu Rev Immunol.* 2003;21:177–204.
97. Shibuya A, Honda S. Molecular and functional characteristics of the Fca/ μ R, a novel Fc receptor for IgM and IgA. *Springer Semin Immunopathol.* 2006;28:377–382.
98. Moura IC, Centelles MN, Arcos-Fajardo M, et al. Identification of the transferrin receptor as a novel immunoglobulin (Ig)A1 receptor and its enhanced expression on mesangial cells in IgA nephropathy. *J Exp Med.* 2001;194:417–425.
99. McGhee JR, Mestecky J, Elson CO, et al. Regulation of IgA synthesis and immune response by T cells and interleukins. *J Clin Immunol.* 1989;9:175–199.
100. Murray PD, McKenzie DT, Swain SL, et al. Interleukin 5 and interleukin 4 produced by Peyer's patch T cells selectively enhance immunoglobulin A expression. *J Immunol.* 1987;139:2669–2674.
101. Beagley KW, Eldridge JH, Lee F, et al. Interleukins and IgA synthesis. Human and murine interleukin 6 induce high rate IgA secretion in IgA-committed B cells. *J Exp Med.* 1989;169:2133–2148.
102. Ramsay AJ, Husband AJ, Ramsshaw IA, et al. The role of interleukin-6 in mucosal IgA antibody responses *in vivo*. *Science.* 1994;264:561–563.
103. Fujihashi K, McGhee JR, Lue C, et al. Human appendix B cells naturally express receptors for and respond to interleukin 6 with selective IgA1 and IgA2 synthesis. *J Clin Invest.* 1991;88:248–252.
104. Fujihashi K, Taguchi T, Aicher WK, et al. Immunoregulatory functions for murine intraepithelial lymphocytes: gamma/delta T cell receptor-positive (TCR $^+$) T cells abrogate oral tolerance, while α/β TCR $^+$ T cells provide B cell help. *J Exp Med.* 1992;175:695–707.
105. Jang MH, Kweon MN, Iwatsuki K, et al. Intestinal villous M cells: an antigen entry site in the mucosal epithelium. *Proc Natl Acad Sci U S A.* 2004;101:6110–6115.
106. Mach J, Hsieh T, Hsieh D, et al. Development of intestinal M cells. *Immunol Rev.* 2005;206:177–189.
107. Ebert EC. Proliferative responses of human intraepithelial lymphocytes to various T-cell stimuli. *Gastroenterology.* 1989;97:1372–1381.
108. Rescigno M, Urbano M, Valzasina B, et al. Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. *Nat Immunol.* 2001;2:361–367.
109. Vazquez-Torres A, Jones-Carson J, Baumler AJ, et al. Extraintestinal dissemination of *Salmonella* by CD18-expressing phagocytes. *Nature.* 1999;401:804–808.
110. Niess JH, Br S, Gu X, et al. CX3CR1-mediated dendritic cell access to the intestinal lumen and bacterial clearance. *Science.* 2005;307:254–258.
111. Huang FP, Platt N, Wykes M, et al. A discrete subpopulation of dendritic cells transports apoptotic intestinal epithelial cells to T cell areas of mesenteric lymph nodes. *J Exp Med.* 2000;191:435–444.
112. Bos NA, Kroese FG, Cebray JJ. B-1 cells and the mucosal immune system. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:655–666.
113. Kunisawa J, Kiyono H. A marvel of mucosal T cells and secretory antibodies for the creation of first lines of defense. *Cell Mol Life Sci.* 2005;62:1308–1321.
114. Snider DP, Liang H, Switzer I, et al. IgA production in MHC class II-deficient mice is primarily a function of B-1 cells. *Int Immunol.* 1999;11:191–198.
115. Macpherson AJ, Gatto D, Sainsbury E, et al. A primitive T cell-independent mechanism of intestinal mucosal IgA responses to commensal bacteria. *Science.* 2000;288:2222–2226.
116. Kunisawa J, Kurashima Y, Gohda M, et al. Sphingosine 1-phosphate regulates peritoneal B-cell trafficking for subsequent intestinal IgA production. *Blood.* 2007;109:3749–3756.
117. Russell MW, Kilian M. Biological activities of IgA. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:267–289.
118. Gunningham-Rundles C. Immunodeficiency and mucosal immunity. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienzenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:1145–1158.
119. Underdown BJ. Passive immunization: systemic and mucosal. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W,

- Bienenschock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:841–851.
120. Murphy BR. Mucosal immunity to viruses. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenschock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:799–814.
 121. Eichelberger M, Allan W, Zijlstra M, et al. Clearance of influenza virus respiratory infection in mice lacking class I major histocompatibility complex-restricted CD8⁺ T cells. *J Exp Med*. 1991;174:875–880.
 122. Graham BS, Bunton LA, Wright PF, et al. Role of T lymphocyte subsets in the pathogenesis of primary infection and rechallenge with respiratory syncytial virus in mice. *J Clin Invest*. 1991;88:1026–1033.
 123. Lehner T, Bergmeier LA. Mucosal infection and immune responses to simian immunodeficiency virus. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenschock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:1179–1197.
 124. Smith PD, Wahl SM. Immunobiology of mucosal HIV-1 infection. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenschock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:1999–1211.
 125. Savage DC. Mucosal Microbiota. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenschock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:19–33.
 126. Macpherson AJ, Geuking MB, McCoy KD. Immune responses that adapt the intestinal mucosa to commensal intestinal bacteria. *Immunology*. 2005;115:153–162.
 127. Hooper LV, Gordon JI. Commensal host-bacterial relationships in the gut. *Science*. 2001;292:1115–1118.
 128. Macpherson AJ, Uhr T. Induction of protective IgA by intestinal dendritic cells carrying commensal bacteria. *Science*. 2004;303:1662–1665.
 129. Caramalho I, Lopes-Carvalho T, Ostler D, et al. Regulatory T cells selectively express toll-like receptors and are activated by lipopolysaccharide. *J Exp Med*. 2003;197:403–411.
 130. Kiyono H, McGhee JR, Wannemuehl MJ, et al. Lack of oral tolerance in C3H/HeJ mice. *J Exp Med*. 1982;155:605–610.
 131. Krieg AM. Therapeutic potential of Toll-like receptor 9 activation. *Nat Rev Drug Discov*. 2006;5:471–484.
 132. Mowat AM, Faria AM, Weiner HL. Oral tolerance: physical basis and clinical applications. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenschock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005:487–538.
 133. Alpan O, Rudomen G, Matzinger P. The role of dendritic cells, B cells, and M cells in gut-oriented immune responses. *J Immunol*. 2001;166:4843–4852.
 134. Spahn TW, Fontana A, Faria AM, et al. Induction of oral tolerance to cellular immune responses in the absence of Peyer's patches. *Eur J Immunol*. 2001;31:1278–1287.
 135. Fujihashi K, Dohi T, Renner PD, et al. Peyer's patches are required for oral tolerance to proteins. *Proc Natl Acad Sci U S A*. 2001;98:3310–3315.
 136. Shi HN, Grusby MJ, Nagler-Anderson C. Orally induced peripheral nonresponsiveness is maintained in the absence of functional Th1 or Th2 cells. *J Immunol*. 1999;162:5143–5148.
 137. Hirahara K, Hisatsune T, Nishijima K, et al. CD4⁺ T cells anergized by high dose feeding establish oral tolerance to antibody responses when transferred in SCID and nude mice. *J Immunol*. 1995;154:6238–6245.
 138. Sakaguchi S, Ono M, Setoguchi R, et al. Foxp3⁺ CD25⁺ CD4⁺ natural regulatory T cells in dominant self-tolerance and autoimmune disease. *Immunol Rev*. 2006;212:8–27.
 139. Izcue A, Coombes JL, Powrie F. Regulatory T cells suppress systemic and mucosal immune activation to control intestinal inflammation. *Immunol Rev*. 2006;212:256–271.
 140. Grohmann U, Orabona C, Fallarino F, et al. CTLA-4 Ig regulates tryptophan catabolism in vivo. *Nat Immunol*. 2002;3:1097–1101.
 141. Ono M, Shimizu J, Miyachi Y, et al. Control of autoimmune myocarditis and multiorgan inflammation by glucocorticoid-induced TNF receptor family-related protein^{high}, Foxp3⁺ expressing CD25⁺ and CD25⁻ regulatory T cells. *J Immunol*. 2006;176:4748–4756.
 142. Chen W, Jin W, Hardegen N, et al. Conversion of peripheral CD4⁺ CD25⁻ naive T cells to CD4⁺ CD25⁺ regulatory T cells by TGF- β induction of transcription factor Foxp3. *J Exp Med*. 2003;198:1875–1886.
 143. Chen Y, Kuchroo VK, Inobe J, et al. Regulatory T cell clones induced by oral tolerance: suppression of autoimmune encephalomyelitis. *Science*. 1994;265:1237–1240.
 144. Roncarolo MG, Gregori S, Battaglia M, et al. Interleukin-10-secreting type I regulatory T cells in rodents and humans. *Immunol Rev*. 2006;212:28–50.
 145. Papadakis KA, Landers C, Prehn J, et al. CC chemokine receptor 9 expression defines a subset of peripheral blood lymphocytes with mucosal T cell phenotype and Th1 or T-regulatory 1 cytokine profile. *J Immunol*. 2003;171:159–165.
 146. Gianfrani C, Levings MK, Sartirana C, et al. Gliadin-specific type I regulatory T cells from the intestinal mucosa of treated celiac patients inhibit pathogenic T cells. *J Immunol*. 2006;177:4178–4186.
 147. Mowat AM, Lamont AG, Bruce MG. A genetically determined lack of oral tolerance to ovalbumin is due to failure of the immune system to respond to intestinally derived tolerogen. *Eur J Immunol*. 1987;17:1673–1676.
 148. Chang CC, Ciubatariu R, Manavalan JS, et al. Tolerization of dendritic cells by T(S) cells: the crucial role of inhibitory receptors ILT3 and ILT4. *Nat Immunol*. 2002;3:237–243.
 149. Gleissner CA, Zastrow A, Klingenberg R, et al. IL-10 inhibits endothelial-dependent T cell costimulation by up-regulation of ILT3/4 in human vascular endothelial cells. *Eur J Immunol*. 2007;37:177–192.
 150. Rufián M, Kawamoto Y, Nakashima I, et al. Essential roles of CD8⁺ CD122⁺ regulatory T cells in the maintenance of T cell homeostasis. *J Exp Med*. 2004;200:1123–1134.
 151. McGechie MJ, Bak-Jensen KS, Chen Y, et al. TGF- β and IL-6 drive the production of IL-17 and IL-10 by T cells and restrain TH-17 cell-mediated pathology. *Nat Immunol*. 2007;8:1390–1397.
 152. Awasthi A, Carrier Y, Peron JP, et al. A dominant function for interleukin 27 in generating interleukin 10-producing anti-inflammatory T cells. *Nat Immunol*. 2007;8:1380–1389.
 153. Stuhmofe JS, Silver JS, Laurence A, et al. Interleukins 27 and 6 induce STAT3-mediated T cell production of interleukin 10. *Nat Immunol*. 2007;8:1363–1371.
 154. Coombes JL, Siddiqui KR, Arancibia-Carcamo CV, et al. A functionally specialized population of mucosal CD103⁺ DCs induces Foxp3⁺ regulatory T cells via a TGF- β and retinoic acid-dependent mechanism. *J Exp Med*. 2007;204:1757–1764.
 155. Sun CM, Hall JA, Blank RB, et al. Small intestine lamina propria dendritic cells promote de novo generation of Foxp3⁺ Treg cells via retinoic acid. *J Exp Med*. 2007;204:1775–1785.
 156. Benson MJ, Pino-Lagos K, Roseblatt M, et al. All-trans retinoic acid mediates enhanced T reg cell growth, differentiation, and gut homing in the face of high levels of co-stimulation. *J Exp Med*. 2007;204:1765–1774.
 157. Mucida D, Park Y, Kim G, et al. Reciprocal Th17 and regulatory T cell differentiation mediated by retinoic acid. *Science*. 2007;317:256–260.
 158. Ke Y, Pearce K, Lake JP, et al. $\gamma\delta$ T lymphocytes regulate the induction and maintenance of oral tolerance. *J Immunol*. 1997;158:3610–3618.
 159. Fujihashi K, Dohi T, Kweon MN, et al. $\gamma\delta$ T cells regulate mucosally induced tolerance in a dose-dependent fashion. *Int Immunopharmacol*. 1999;11:1907–1916.
 160. Viney JL, Mowat AM, O'Malley JM, et al. Expanding dendritic cells in vivo enhances the induction of oral tolerance. *J Immunol*. 1998;160:5815–5825.
 161. Ito T, Yang M, Wang YH, et al. Plasmacytoid dendritic cells prime IL-10-producing T regulatory cells by inducible costimulator ligand. *J Exp Med*. 2007;204:105–115.
 162. Gillies M, Liu YJ. Generation of human CD8 T regulatory cells by CD40 ligand-activated plasmacytoid dendritic cells. *J Exp Med*. 2002;195:695–704.
 163. Allez M, Brimnes J, Dotan I, et al. Expansion of CD8⁺ T cells with regulatory function after interaction with intestinal epithelial cells. *Gastroenterology*. 2002;123:1516–1526.

164. Cruickshank SM, McVay LD, Baumgart DC, et al. Colonic epithelial cell mediated suppression of CD4 T cell activation. *Gut*. 2004;53:678–684.
165. Westendorf AM, Bruder D, Hansen W, et al. Intestinal epithelial antigen induces CD4⁺ T cells with regulatory phenotype in a transgenic autoimmune mouse model. *Ann NY Acad Sci*. 2006;1072:401–406.
166. Ostroukhova M, Seguin-Devaux C, Oriss TB, et al. Tolerance induced by inhaled antigen involves CD4⁺ T cells expressing membrane-bound TGF- β and FOXP3. *J Clin Invest*. 2004;114:28–38.
167. Oriss TB, Ostroukhova M, Seguin-Devaux C, et al. Dynamics of dendritic cell phenotype and interactions with CD4⁺ T cells in airway inflammation and tolerance. *J Immunol*. 2005;174:854–863.
168. Kutteh WH, Mestecky J, Wira CR. Mucosal immunity in the human female reproductive tract. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;1631–1646.
169. Anderson DF, Pudney J. Human male genital tract immunity and experimental models. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;1647–1660.
170. Peppoloni S, Ruggiero P, Contorno M, et al. Mutants of the *Escherichia coli* heat-labile enterotoxin as safe and strong adjuvants for intranasal delivery of vaccines. *Experi Rev Vaccines*. 2003;2:285–293.
171. Elson CO, Dertzbaugh MT. Mucosal Adjuvants. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;967–986.
172. Lycke N. Targeted vaccine adjuvants based on modified cholera toxin. *Curr Mol Med*. 2005;5:591–597.
173. Borsigky S, Kretschmer K, Becker PD, et al. The mucosal adjuvant macrophage-activating lipopeptide-2 directly stimulates B lymphocytes via the TLR2 without the need of accessory cells. *J Immunol*. 2005;174:6308–6313.
174. Michalek SM, O'Hagan D, Childers NK, et al. Antigen delivery systems I: Nonliving microparticles, liposomes, and immune stimulating complexes (ISCOMs). In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;987–1008.
175. Jepson MA, Clark MA, Hirst BH. M cell targeting by lectins: a strategy for mucosal vaccination and drug delivery. *Adv Drug Deliv Rev*. 2004;56:511–525.
176. Lambkin I, Pinilla C, Hamashin C, et al. Toward targeted oral vaccine delivery systems: selection of lectin mimetics from combinatorial libraries. *Pharm Res*. 2003;20:1258–1266.
177. Higgins LM, Lambkin I, Donnelly G, et al. In vivo phage display to identify M cell-targeting ligands. *Pharm Res*. 2004;21:695–705.
178. Nochi T, Yuki Y, Matsumura A, et al. A novel M cell specific carbohydrate-targeted mucosal vaccine effectively induces antigen-specific immune responses. *J Exp Med*. 2007 (in press).
179. Wu Y, Wang X, Csernitski KL, et al. M cell-targeted DNA vaccination. *Proc Natl Acad Sci U S A*. 2001;98:9318–9323.
180. Kunisawa J, Nakagawa S, Mayumi T. Pharmacotherapy by intracellular delivery of drugs using fusogenic liposomes: application to vaccine development. *Adv Drug Deliv Rev*. 2001;52:177–186.
181. Curtiss R. 3rd. Antigen delivery system II: development of live recombinant attenuated bacterial antigen and DNA vaccine delivery vector vaccines. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;1009–1038.
182. Rosenthal KL. Recombinant live viral vectors as vaccines for mucosal immunity. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;1039–1052.
183. Mason HS, Chikwamba R, Santi L, et al. Transgenic plants for mucosal vaccines. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;1053–1060.
184. Cunningham-Rundles C. Immunodeficiency and mucosal immunity. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;1145–1158.
185. Meng G, Wei X, Wu X, et al. Primary intestinal epithelial cells selectively transfer R5 HIV-1 to CCR5⁺ cells. *Nat Med*. 2002;8:150–156.
186. Brenchley JM, Price DA, Schacker TW, et al. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nat Med*. 2006;12:1365–1371.
187. Kozlowski PA, Jackson S. Serum IgA subclasses and molecular forms in HIV infection: selective increases in monomer and apparent restriction of the antibody response to IgA1 antibodies mainly directed at env glycoproteins. *AIDS Res Hum Retroviruses*. 1992;8:1773–1780.
188. Kaul R, Trabattoni D, Bwayo JJ, et al. HIV-1-specific mucosal IgA in a cohort of HIV-1-resistant Kenyan sex workers. *Aids*. 1999;13:23–29.
189. Elson CO, Cong Y, McCracken VJ, et al. Experimental models of inflammatory bowel disease reveal innate, adaptive, and regulatory mechanisms of host dialogue with the microbiota. *Immunol Rev*. 2005;206:260–276.
190. Powrie F, Uhlig H. Animal models of intestinal inflammation: clues to the pathogenesis of inflammatory bowel disease. *Novartis Found Symp*. 2004;263:164–174; discussion 174–168, 211–168.
191. Rath HC, Wilson KH, Sartor RB. Differential induction of colitis and gastritis in HLA-B27 transgenic rats selectively colonized with *Bacteroides vulgatus* or *Escherichia coli*. *Infect Immun*. 1999;67:2969–2974.
192. Gionchetti P, Rizzello F, Lammers KM, et al. Antibiotics and probiotics in treatment of inflammatory bowel disease. *World J Gastroenterol*. 2006;12:3306–3313.
193. Kraus TA, Cheifetz A, Toy L, et al. Evidence for a genetic defect in oral tolerance induction in inflammatory bowel disease. *Inflamm Bowel Dis*. 2006;12:82–88.
194. Kaser A, Nieuwenhuis EE, Strober W, et al. Natural killer T cells in mucosal homeostasis. *Ann NY Acad Sci*. 2004;1029:154–168.
195. Vercelli D. Mechanisms of the hygiene hypothesis—molecular and otherwise. *Curr Opin Immunol*. 2006;18:733–737.
196. Cohn L, Elias JA, Chupp GL. Asthma: mechanisms of disease persistence and progression. *Annu Rev Immunol*. 2004;22:789–815.
197. Kweon M, Kiyono H. Allergic diseases in the gastrointestinal tract. In: *Mucosal Immunology*, 3rd ed. Mestecky J, Lamm ME, Strober W, Bienenstock J, McGhee JR, Mayer L, eds. San Diego: Academic Press. 2005;1351–1360.
198. Popescu FD. New asthma drugs acting on gene expression. *J Cell Mol Med*. 2003;7:475–486.
199. Brownell J, Casale TB. Anti-IgE therapy. *Immunol Allergy Clin North Am*. 2004;24:551–568, v.
200. Takagi H, Hiroi T, Yang L, et al. A rice-based edible vaccine expressing multiple T cell epitopes induces oral tolerance for inhibition of Th2-mediated IgE responses. *Proc Natl Acad Sci U S A*. 2005;102:17525–17530.
201. Hawrylowicz CM. Regulatory T cells and IL-10 in allergic inflammation. *J Exp Med*. 2005;202:1459–1463.
202. Lewkowich IP, Herman NS, Schleifer KW, et al. CD4⁺CD25⁺ T cells protect against experimentally induced asthma and alter pulmonary dendritic cell phenotype and function. *J Exp Med*. 2005;202:1549–1561.
203. Schnyder-Candrian S, Togbe D, Couillin I, et al. Interleukin-17 is a negative regulator of established allergic asthma. *J Exp Med*. 2006;203:2715–2725.
204. Bisset LR, Schmid-Grendelmeier P. Chemokines and their receptors in the pathogenesis of allergic asthma: progress and perspective. *Curr Opin Pulm Med*. 2005;11:35–42.
205. Kurashima Y, Kunisawa J, Higuchi M, et al. Sphingosine 1-phosphate-mediated trafficking of pathogenic Th2 and mast cells for the control of food allergy. *J Immunol*. 2007;179:1577–1585.
206. Hue S, Mention JJ, Monteiro RC, et al. A direct role for NKG2D/MICA interaction in villous atrophy during celiac disease. *Immunity*. 2004;21:367–377.