

**Figure 1** Structures of HCV replicon plasmids. The pRep-Feo expressed a chimeric reporter protein of firefly luciferase (Fluc) and neomycin phosphotransferase (GenBank accession No. AB119282).<sup>14,20</sup> The pRep-Fluc expressed the Fluc protein. The pRep-BSD expressed the blasticidin S (BSD) resistance gene. pT7, T7 promoter; 5'UTR, HCV 5'-untranslated region;  $\Delta$ C, truncated HCV core region (nt. 342–377); neo, neomycin phosphotransferase gene; EMCV, encephalomyocarditis virus; NS3, NS4, NS5A and NS5B, genes that encode HCV non-structural proteins; 3'UTR, HCV 3'-untranslated region.

HCV-directed siRNA *in vivo* may be effective in silencing viral protein expression in the liver. Here, we report that HCV replication was suppressed *in vitro* by recombinant retrovirus and adenovirus vectors expressing short hairpin RNA (shRNA) and that the delivery of the adenovirus vector to mice *in vivo* specifically inhibited viral protein synthesis in the liver.

## Methods

### Cells and cell culture

Huh7 and Retro Pack PT67 cells (Clontech, Palo Alto, CA, USA) were maintained in Dulbecco's modified minimal essential medium (Sigma, St. Louis, MO, USA) supplemented with 10% fetal calf serum at 37°C under 5% CO<sub>2</sub>. To maintain cell lines carrying the HCV replicon, G418 (Wako, Osaka, Japan) was added to the culture medium to a final concentration of 500  $\mu$ g/mL.

### HCV replicon constructs and transfection

HCV replicon plasmids, pRep-Feo, pRep-Fluc and pRep-BSD were constructed from a virus, HCV-N strain, genotype 1b.<sup>21</sup> The pRep-Feo expressed a chimeric reporter protein of firefly luciferase (Fluc) and neomycin phosphotransferase.<sup>14,20</sup> The pRep-Fluc and the pRep-BSD expressed the Fluc and blasticidin S (BSD) resistance genes, respectively (Fig. 1). The replicon RNA synthesis and the transfection protocol have been described previously.<sup>22</sup>

### Synthetic siRNA and siRNA-expression plasmid

The design and construction of HCV-directed siRNA vectors have been described.<sup>14</sup> Briefly, five siRNA targeting the 5'-UTR of HCV RNA were tested for their efficiency to inhibit HCV replication, and the most effective sequence, which targeted nucleotide position of 331 through 351, was used in the present study. To construct shRNA-expressing DNA cassettes, oligonucleotide inserts were synthesized that contained the loop sequence (5'-TTC AAG AGA-

3') flanked by sense and antisense siRNA sequences (Fig. 2a). These were inserted immediately downstream of the human U6 promoter. To avoid a problem in transcribing shRNA because of instability of the DNA strands arising from the tight palindrome structure, several C-to-T point mutations, which retained completely the silencing activity of the shRNA, were introduced into the sense strand of the shRNA sequences (referred to as 'm').<sup>23</sup> A control plasmid, pUC19-shRNA-Control, expressed shRNA directed towards the Machado-Joseph disease gene, which is a mutant of ataxin-3 gene and is not normally expressed. We have previously described the sequence specific activity of the shRNA-Control.<sup>24</sup>

Prior to construction of the virus vectors, we tested silencing efficiency of five shRNA constructs of different lengths that covered the target sequence (Fig. 2a). The shRNA-HCV-19, shRNA-HCV-21 and shRNA-HCV-27 had target sequences of 19, 21 and 27 nucleotides, respectively. Transfection of these shRNA constructs into Huh7/pRep-Feo showed that shRNA with longer target sequences had better suppressive effects (Fig. 2b). Therefore, we used shRNA-HCV-27m (abbreviated as shRNA-HCV) in the following study.

### Recombinant retrovirus vectors

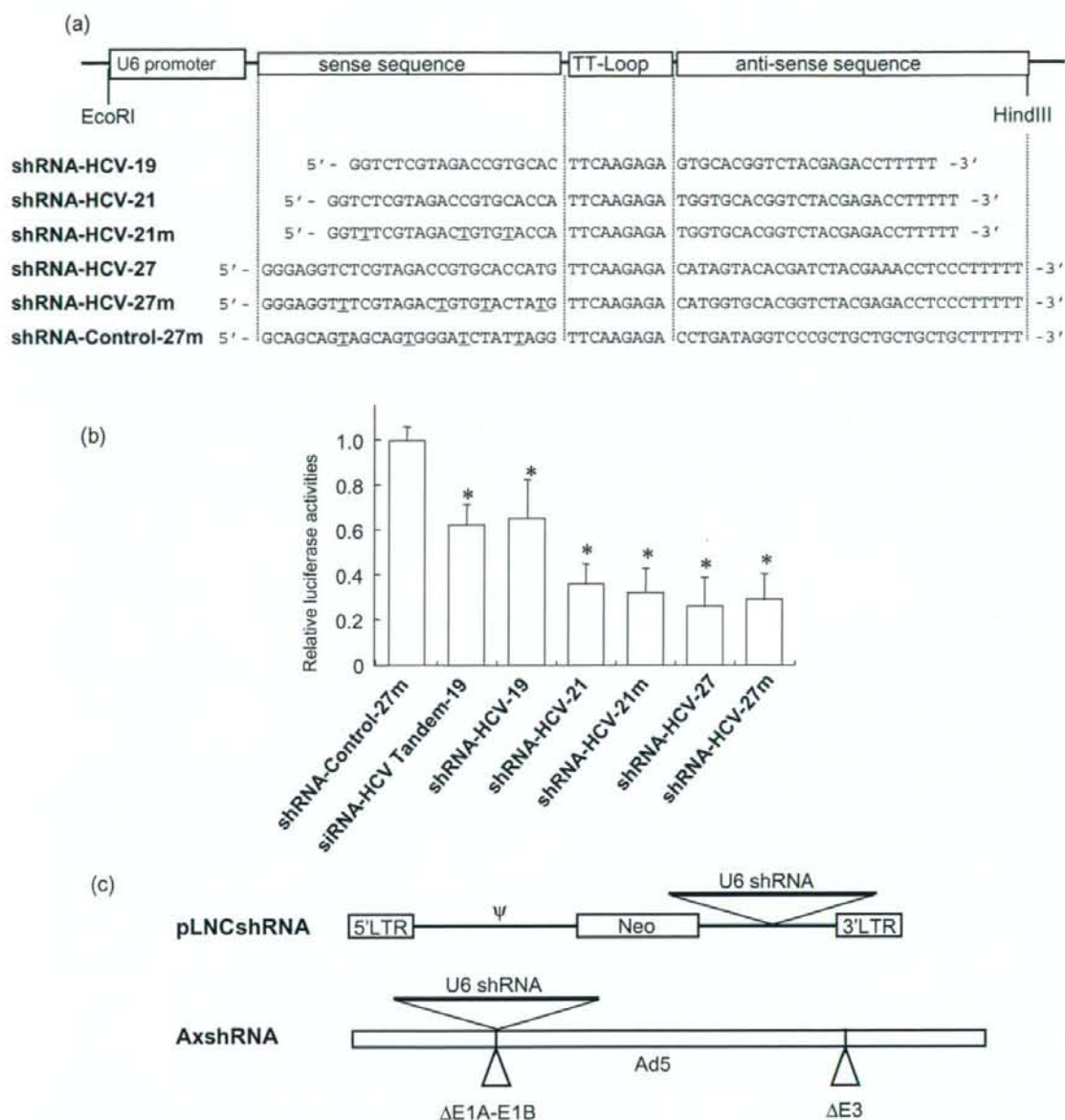
The U6-shRNA expression cassettes were inserted into the *StuI*/*HindIII* site of a retrovirus vector, pLNCX2 (Clontech) to construct pLNCshRNA-HCV and pLNCshRNA-Control (Fig. 2c). The plasmids were transfected into the packaging cells, Retro Pack PT67. The culture supernatant was filtered and added onto Huh7 cells with 4  $\mu$ g/mL of polybrene. Huh7 cell lines stably expressing shRNA were established by culture in the presence of 500  $\mu$ g/mL of G418.

### Recombinant adenovirus

Recombinant adenoviruses expressing shRNA were constructed using an Adenovirus Expression Vector Kit (Takara, Otsu, Japan). The U6-shRNA expression DNA cassette was inserted into the *SmaI* site of pAxcw to construct pAxshRNA-HCV and pAxshRNA-Control. The adenoviruses were propagated according to the manufacturer's protocol (AxshRNA-HCV and AxshRNA-Control; Fig. 2c). A 'multiplicity of infection' (MOI) was used to standardize infecting doses of adenovirus. The MOI stands for the ratio of infectious virus particles to the number of cells being infected. An MOI = 1 represents equivalent dose to introduce one infectious virus particle to every host cell that is present in the culture.

### Plasmids for assays of interferon responses

pISRE-TA-Luc (Invitrogen, Carlsbad, CA, USA) contained five copies of the consensus interferon stimulated response element (ISRE) motifs upstream of the Fluc gene. pTA-Luc (Invitrogen), which lacks the enhancer element, was used for background determination. The pcDNA3.1 (Invitrogen) was used as an empty vector for mock transfection. pRL-CMV (Promega, Madison, WI, USA), which expresses the *Renilla* luciferase protein, was used for normalization of transfection efficiency.<sup>25</sup> A plasmid, pEGFPneo (Invitrogen), was used to monitor percentages of transduced cells.



**Figure 2** Structure of shRNA-expression constructs and shRNA sequences. (a) Structure of shRNA-expression cassette and shRNA sequences. TT-Loop, the loop sequence. The shRNA-Control was directed toward an unrelated target, Machado-Joseph disease gene. Underlined letters indicate C-to-T point mutations in the sense strand. (b) The shRNA-expression plasmids were transfected into Huh7/pRep-Feo cells, and internal luciferase activities were measured at 48 h of transfection. Each assay was done in triplicate, and the values are displayed as mean + SD. \* $P < 0.05$ . (c) pLNCshRNA, structure of a recombinant retrovirus expressing shRNA.  $\Psi$ , the retroviral packaging signal sequence. AxshRNA, structure of a recombinant adenovirus expressing shRNA.



### Real-time RT-PCR analysis

Total cellular RNA was extracted from cultured cells or liver tissue using ISOGEN (Nippon Gene, Tokyo, Japan). Total cellular RNA (2 µg) was used to generate cDNA from each sample using the SuperScript II reverse-transcriptase (Invitrogen). The mRNA expression levels were measured using the Light Cycler PCR and detection system (Roche, Mannheim, Germany) and Light Cycler Fast Start DNA Master SYBR Green 1 mix (Roche).

### Luciferase assays

Luciferase activity was measured using a luminometer, Lumat LB9501 (Promega) and the Bright-Glo Luciferase Assay System (Promega) or the Dual-Luciferase Reporter Assay System (Promega).

### Northern and western hybridization

Total cellular RNA was separated by denaturing agarose-formaldehyde gel electrophoresis, and transferred to a nylon membrane. The membrane was hybridized with a digoxigenin-labeled probe specific for the full-length replicon sequence, and subsequently with a probe specific for beta-actin. The signals were detected by chemiluminescence reaction using a Digoxigenin Luminescent Detection Kit (Roche), and visualized by Fluoro-Imager (Roche). For the western blotting, 10 µg of total cell lysate was separated on NuPAGE 4.12% Bis-TrisGel (Invitrogen), and blotted onto an Immobilon PVDF Membrane (Roche). The membrane was incubated with monoclonal antibodies specific for HCV-NSSA (BioDesign, Saco, ME, USA), NS4A (Virogen, Watertown, MA, USA), or beta-actin (Sigma), and detected by a chemiluminescence reaction (BM Chemiluminescence Blotting Substrate; POD, Roche).

### Transient-replication assays

A replicon, pRep-Fluc, was transfected into cells and the luciferase activities of the cell lysates were measured serially. To correct the transfection efficiency, each value was divided by the luciferase activity at 4 h after the transfection.

### Stable colony formation assays

Cells were transfected with a replicon, pRep-BSD, and were cultured in the presence of 150 µg/mL of BSD (Invitrogen). BSD-resistant cell colonies appeared after ~3 weeks of culture, and were counted.

### HCV-JFH1 virus cell culture

An *in-vitro* transcribed HCV-JFH1 RNA<sup>26</sup> was transfected into Huh7.5.1 cells.<sup>27</sup> Naive Huh7.5.1 cells were subsequently infected by the culture supernatant of the JFH1-RNA transfected Huh7.5.1 cells, and subjected to siRNA or drug treatments. Replication levels of HCV-RNA were quantified by the realtime RT-PCR by using primers that targeted HCV-NSSB region, HCV-JFH1 sense: 5'-TCA GAC AGA GCC TGA GTC CA-3', and HCV-JFH1 anti-sense: 5'-AGT TGC TGG AGG GCT TCT GA-3'.

### Mice and adenovirus infection

Transgenic mice, CN2-29, inducibly express mRNA for the HCV structural proteins (genotype 1b, nucleotides 294–3435) by the Cre/loxP switching system.<sup>28</sup> The transgene does not contain full-length HCV 5'-UTR, but shares the target sequence of the shRNA-HCV. Although the transgenic mouse CN2 has been previously reported as expressing higher levels of the viral proteins, the expression levels of the viral core protein in the CN2-29 mice are modest and similar to that in the liver of HCV patients. Thus, we chose CN2-29 mice in the present study.

The mice were infected with AxshRNA-HCV or controls (AxshRNA-Control or AxCAw1) in combination with AxCAN-Cre, which expressed Cre recombinase. Three days after the infection, the mice were killed and HCV core protein in the liver was measured as described below. The BALB/c mice were maintained in the Animal Care Facility of Tokyo Medical and Dental University, and transgenic mice were in the Tokyo Metropolitan Institute of Medical Science. Animal care was in accordance with institutional guidelines. The review board of the university approved our experimental animal studies and all experiments were approved by the institutional animal study committees.

### Measurement of HCV core protein in mouse liver

The amounts of HCV core protein in the liver tissue from the mice was measured by a fluorescence enzyme immunoassay (FEIA)<sup>29</sup> with a slight modification. Briefly, the 5F11 monoclonal anti-HCV-core antibody was used as the first antibody on the solid phase, and the 5E3 antibody conjugated with horseradish peroxidase was the second antibody. This FEIA can detect as little as 4 pg/mL of recombinant HCV-core protein. Contents of the HCV core protein in the liver samples were normalized by the total protein contents and expressed as pg/mg total protein.

### Immunohistochemical staining

Liver tissue was frozen with optimal cutting temperature (OTC) compound (Tissue Tek; Sakura Finetechnical, Tokyo, Japan). The sections (8 µm thick) were fixed with a 1:1 solution of acetone : methanol at -20°C for 10 min and then washed with phosphate-buffered saline (PBS). Subsequently, the sections were incubated with the IgG fraction of an anti-HCV core rabbit polyclonal antibody (RR8)<sup>28</sup> in blocking buffer or antialbumin rabbit polyclonal antibody (Dako Cytomation, Glostrup, Denmark) in PBS overnight at 4°C. The sections were incubated with secondary antibody, Alexa-antirabbit IgG (Invitrogen) or TRITIC-antirabbit IgG (Sigma), for 2 h at room temperature. Fluorescence was observed using a fluorescence microscope.

### Statistical analyses

Statistical analyses were performed using Student's *t*-test; *P*-values of less than 0.05 were considered to be statistically significant.



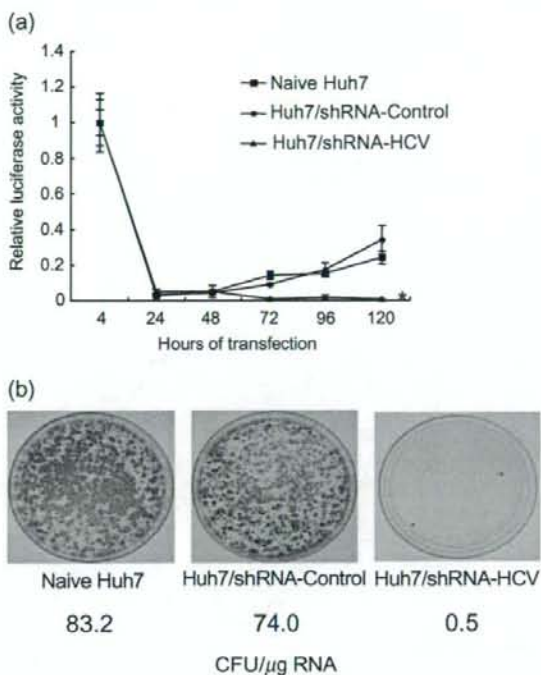
## Results

### Retrovirus transduction of shRNA can protect from HCV replication

Retrovirus vectors propagated from pLNCshRNA-HCV and pLNCshRNA-Control were used to infect Huh7 cells, and cell lines were established that constitutively express shRNA-HCV and shRNA-Control (Huh7/shRNA-HCV and Huh7/shRNA-Control, respectively). There were no differences in the cell morphology or growth rate between shRNA-transduced and non-transduced Huh7 cells (data not shown). The HCV replicon, pRep-Fluc, was transfected into Huh7/shRNA-HCV, Huh7/shRNA-Control and naive Huh7 cells by electroporation. In Huh7/shRNA-Control and naive Huh7 cells, the initial luciferase activity at 4 h decreased temporarily, which represents decay of the transfected replicon RNA, but increased again at 48 h and 72 h, which demonstrate *de novo* synthesis of the HCV replicon RNA. In contrast, transfection into Huh7/shRNA-HCV cells resulted in a decrease in the initial luciferase activity, reaching background by 72 h (Fig. 3a). Similarly, transfection of the replicon, pRep-BSD, into Huh7 cells and BSD selection yielded numerous BSD-resistant colonies in the naive Huh7 (832 colonies) and Huh7/shRNA-Control cell lines (740 colonies), while transfection of Huh7/shRNA-HCV, which expressed shRNA-HCV, yielded obviously fewer colonies (five colonies), indicating reduction of colony forming units by  $\sim 10^2$  (Fig. 3b). There was no difference in shape, growth or viability between cells expressing the shRNA or not. These results indicated that cells expressing HCV-directed shRNA following retrovirus transduction acquired resistance to HCV replication.

### Effect of recombinant adenoviruses expressing shRNA on *in vitro* HCV replication

We investigated subsequently the effects of recombinant adenovirus vectors expressing shRNA. AxshRNA-HCV and AxshRNA-Control were used separately to infect Huh7/pRep-Feo cells, and the internal luciferase activities were measured sequentially (Fig. 4a). AxshRNA-HCV caused continuous suppression of HCV RNA replication. Six days postinfection, the luciferase activities fell to background levels. In contrast, the luciferase activities of the Huh7/pRep-Feo cells infected with AxshRNA-Control did not show any significant changes compared with untreated Huh7/pRep-Feo cells (Fig. 4a). The dimethylthiazol carboxymethoxyphenyl sulfophenyl tetrazolium (MTS) assay showed no significant difference between cells that were infected by recombinant adenovirus and uninfected cells (Fig. 4b). In the northern blotting analysis, the cells were harvested 6 days after infection with the adenovirus at an MOI of 1. Feo-replicon RNA of 9.6 kb, which was detectable in the untreated Huh7/pRep-Feo cells and in the cells infected with AxshRNA-Control, diminished substantially following infection with the AxshRNA-HCV (Fig. 4c). Densitometries showed that the intracellular levels of the replicon RNA in the Huh7/pRep-Feo cells correlated well with the internal luciferase activities. Similarly in the western blotting, cells were harvested 6 days after infection with adenovirus. Levels of the HCV NS4A and NS5A proteins that were translated from the HCV replicon decreased following infection with the AxshRNA-HCV

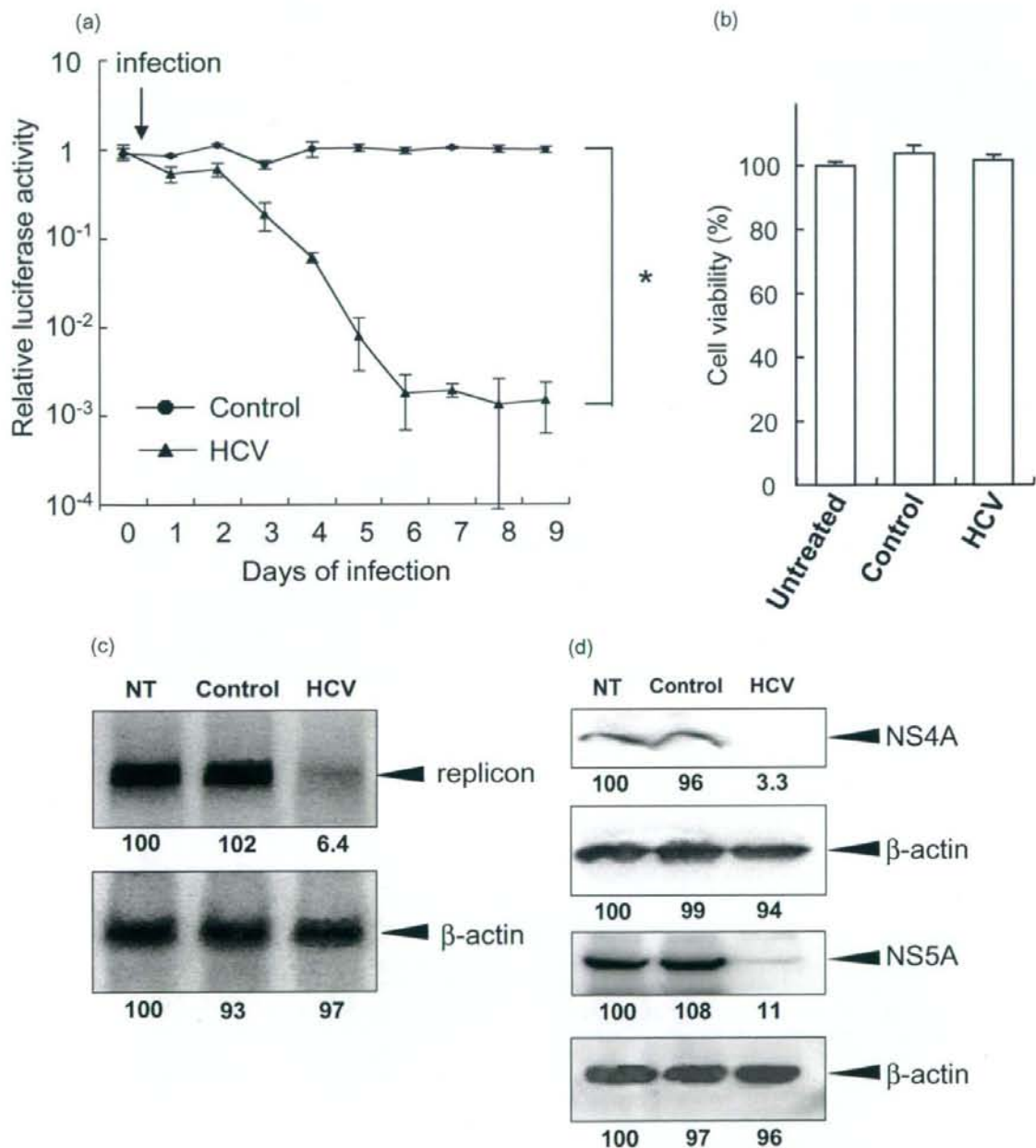


**Figure 3** HCV replication can be inhibited by shRNA-HCV which was stably transfected into cells. Huh7/shRNA-HCV and Huh7/shRNA-Control stably express shRNA-HCV or shRNA-Control, respectively, following retroviral transduction. (a) Transient replication assay. An HCV replicon RNA, pRep-Fluc, was transfected into naive Huh7, Huh7/shRNA-HCV and Huh7/shRNA-Control cells. Luciferase activities of the cell lysates were measured serially at the times indicated, and the values were plotted as ratios relative to luciferase activities at 4 h. The luciferase activities at 4 h represent transfected replicon RNA. The data are mean  $\pm$  SD. An asterisk denotes a *P*-value of less than 0.001 compared with the corresponding value of the naive Huh7 cells. (b) Stable colony formation assay. The HCV replicon, pRep-BSD, was transfected into naive Huh7, Huh7/shRNA-HCV and Huh7/shRNA-Control cells. The cells were cultured in the presence of blasticidin S (BSD) in the medium for  $\sim 3$  weeks, and the BSD-resistant colonies were counted. These assays were repeated twice. The colony-forming units per microgram RNA (CFU/ $\mu$ g RNA) are shown at the bottom.

(Fig. 4d). These results indicated that the decrease in luciferase activities was due to specific suppressive effects of shRNA on expression of HCV genomic RNA and the viral proteins, and not due to non-specific effects caused by the delivery of shRNA or to toxicity of the adenovirus vectors.

### Absence of interferon-stimulated gene responses by siRNA delivery

It has been reported that double-stranded RNA may induce interferon-stimulated gene (ISG) responses which cause instability of mRNA, translational suppression of proteins and apoptotic cell

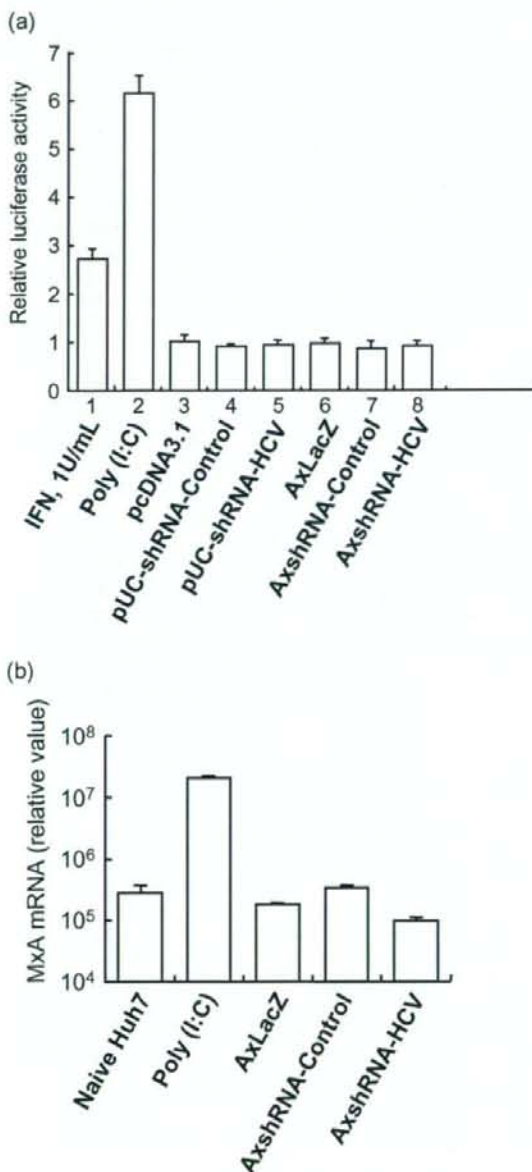


death.<sup>18,30,31</sup> Therefore, we examined the effects of the shRNA-expressing plasmids and adenoviruses on the activation of ISG expression in cells. The ISRE-reporter plasmid, pISRE-TA-Luc, and a control plasmid, pGFPneo, were transfected into Huh7 cells

with plasmid pUC19-shRNA-HCV or pUC19-shRNA-Control, or adenovirus, AxshRNA-HCV or AxshRNA-Control, and the ISRE-mediated luciferase activities were measured. On day 2, the ISRE-luciferase activities did not significantly change in cells in which



**Figure 4** Effect of a recombinant adenovirus expressing shRNA on HCV replicon. (a) Huh7/pRep-Feo cells were infected with AxshRNA-HCV or shRNA-Control at a multiplicity of infection (MOI) of 1. The cells were harvested, and internal luciferase activities were measured on day 0 though day 9 after adenovirus infection. Each assay was done in triplicate, and the value is displayed as a percentage of no treatment and as mean  $\pm$  SD. An asterisk indicates a *P*-value of less than 0.05. (b) Dimethylthiazol carboxymethoxyphenyl sulfophenyl tetrazolium (MTS) assay of Huh7/pRep-Feo cells. Cells were infected with indicated recombinant adenoviruses at an MOI of 1. The assay was done at day 6 of infection. Error bars indicate mean  $\pm$  SD. (c) Northern blotting. The upper panel shows replicon RNA, and the lower panel shows beta-actin mRNA. (d) Western blotting. Total cell lysates were separated on NuPAGE gel, blotted and incubated with monoclonal anti-NS4A or anti-NS5A antibodies. The membrane was re-blotted with antibeta-actin antibodies. NT, untreated Huh7/pRep-Feo cells; Control, cells infected with AxshRNA-Control; HCV, cells treated with AxshRNA-HCV. In panels (b) and (c), cells were harvested on day 6 after adenovirus infection at an MOI of 1.

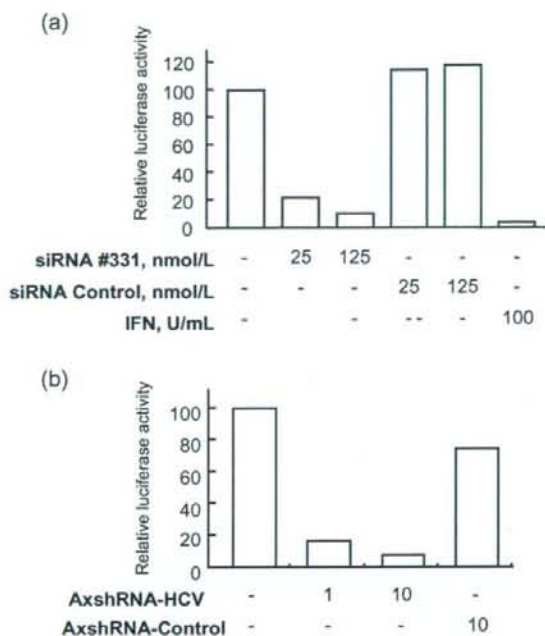


**Figure 5** Interferon-stimulated gene responses by transfection of siRNA vectors. (a) Huh7 cells were seeded at  $5 \times 10^4$  per well in 24-well plates on the day before transfection. As a positive control, 200 ng of pSRE-TA-Luc, or pTA-Luc, 1 ng of pRL-CMV, were transfected into a well using FuGENE-6 Transfection Reagent (Roche), and the cells were cultured with 1 U/mL of interferon (IFN) in the medium (lane 1). Lanes 3–5: 200 ng of pSRE-TA-Luc or pTA-Luc, and 1 ng of pRL-CMV were cotransfected with (lane 2) 300 ng of poly (I : C), or 200 ng of plasmids (lane 3) pcDNA3.1, (lane 4) pUC19-shRNA-Control or (lane 5) pUC19-shRNA-HCV. Lanes 6–8: 200 ng of pSRE-TA-Luc or pTA-Luc, and 1 ng of pRL-CMV were transfected, and MOI = 1 of adenoviruses, (lane 6) AxLacZ, which expressed the beta-galactosidase (LacZ) gene under control of the chicken beta-actin (CAG) promoter as a control, (lane 7) AxshRNA-Control or (lane 8) AxshRNA-HCV were infected. Dual luciferase assays were performed at 48 h after transfection. The Fluc activity of each sample was normalized by the respective Rluc activity, and the respective pTA luciferase activity was subtracted from the pSRE luciferase activity. The experiment was done in triplicate, and the data are displayed as means  $\pm$  SD. (b) Huh7 cells were infected with indicated recombinant adenoviruses, AxLacZ, AxshRNA-Control and AxshRNA-HCV. RNA was extracted from each sample at day 6, and mRNA expression levels of an interferon-inducible MxA protein were quantified by the real-time RT-PCR analysis. Primers used were as follows: human MxA sense, 5'-CGA GGG AGA CAG GAC CAT CG-3'; human MxA antisense, 5'-TCT ATC AGG AAC AAC ATT TT-3'; human beta-actin sense, 5'-ACA ATG AAG ATC AAG ATC ATT GCT CCT CCT-3'; and human beta-actin antisense, 5'-TTT GCG GTG GAC GAT GGA GGG GCC GGA CTC-3'.

negative- or positive-control shRNA plasmids was transfected. (Fig. 5a). Similarly, the expression levels of an interferon-inducible MxA protein did not significantly change by transfection of shRNA-expression vectors (Fig. 5b). These results demonstrate that the shRNA used in the present study lack induction of the ISG responses both in the form of the expression plasmids and the adenovirus vectors.

#### Effect of siRNA and shRNA adenoviruses on HCV-JFH1 cell culture

The effects of HCV-targeted siRNA- and shRNA-expressing adenoviruses were confirmed by using HCV-JFH1 virus cell culture system. Transfection of the siRNA #331<sup>14</sup> into HCV-infected Huh7.5.1 cells resulted in substantial decrease of intracellular HCV RNA, while a control siRNA showed no effect (Fig. 6a). Similarly, infection of AxshRNA-HCV into Huh7.5.1/HCV-JFH1 cells specifically suppressed expression of HCV RNA (Fig. 6b).



**Figure 6** Effects of an siRNA and adenovirus expressing shRNA on HCV-JFH1 cell culture. (a) The siRNA #331, the siRNA-Control<sup>14</sup>, (b) AxshRNA-HCV or AxshRNA-Control were, respectively, transfected or infected onto HCV-JFH1-infected Huh7.5.1 cells. Seventy-two hours of the transfection or infection, expression level of HCV-RNA was quantified by real-time RT-PCR. The assays were repeated twice, and consistent results were obtained. IFN, recombinant interferon- $\alpha$  2b.

### Suppression of HCV-IRES-mediated translation *in vivo* by adenovirus expressing shRNA

The effects of the shRNA expression on the expression of the viral structural proteins *in vivo* were investigated using conditional HCV cDNA-transgenic mice, CN2-29.<sup>28</sup> Adenoviruses, AxshRNA-HCV, AxshRNA-Control or AxCAw1 were injected into CN2-29 mice in combination with AxCANCre, an adenovirus expressing Cre DNA recombinase. The mice were killed on the fourth day after the injection, and the hepatic expression of the HCV core protein was measured. The expressed amounts of the core protein were  $143.0 \pm 56.2$  pg/mg and  $108.5 \pm 42.4$  pg/mg in AxCAw1 and AxshRNA-Control-infected mice, respectively, and the expressed amount was significantly lower in mice injected with AxshRNA-HCV ( $28.7 \pm 7.0$  pg/mg,  $P < 0.05$ , Fig. 7a). Similarly, the induced expression of HCV core protein was not detectable by immunohistochemistry in AxshRNA-HCV infected liver tissue (Fig. 7c). Staining of a host cellular protein, albumin, was not obviously different between the liver infected with AxCAw1, AxshRNA-HCV and AxshRNA-Control (Fig. 7d). The expression levels of two ISG, IFN- $\beta$  and Mx1, in the liver tissue were not significantly different between individuals with

and without injection of the adenovirus vectors (Fig. 7b). These results indicate specific shRNA silencing of HCV structural protein expression in the liver.

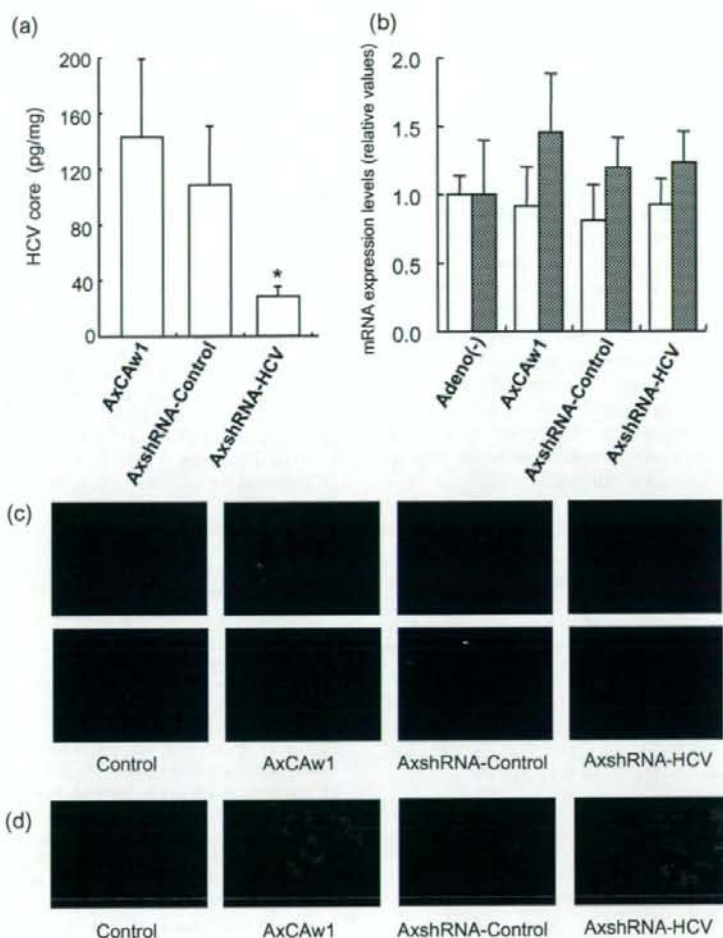
### Discussion

The requirements to achieve a high efficiency using RNAi are: (i) selection of target sequences that are the most susceptible to RNAi; (ii) persistence of siRNA activity; and (iii) efficient *in vivo* delivery of siRNA to cells. We have used an shRNA sequence that was derived from a highly efficient siRNA (siRNA331), and constructed a DNA-based shRNA expression cassette that showed competitive effects with the synthetic siRNA (Fig. 2).<sup>14</sup> The shRNA-expression cassette does not only allow extended half-life of the RNAi, but also enables use of gene-delivery vectors, such as virus vectors. As shown in the results, a retrovirus vector expressing shRNA-HCV could stably transduce cells to express HCV-directed shRNA, and the cells acquired protection against HCV subgenomic replication (Fig. 3). An adenovirus vector expressing shRNA-HCV resulted in suppression of HCV subgenomic and protein expression by around three logs to almost background levels (Fig. 4). Consistent results were obtained by using an HCV cell culture (Fig. 6). More importantly, we have demonstrated *in-vivo* effects on viral protein expression in the liver using a conditional transgenic mouse model (Fig. 7). These results suggest that efficient delivery of siRNA could be effective against HCV infection *in vivo*.

An obstacle to applying siRNA technology to treat virus infections is that viruses are prone to mutate during their replication.<sup>32</sup> HCV continuously produces mutated viral strains to escape immune defense mechanisms. Even in a single patient, the circulating HCV population comprises a large number of closely related HCV sequence variants called quasispecies. Therefore, siRNA targeting the protein-coding sequence of the HCV genome, which have been reported by others,<sup>15-19</sup> may vary considerably among different HCV genotypes, and even among strains of the same genotype.<sup>33</sup> Our shRNA sequence targeted the 5'-UTR of HCV RNA, which is the most conserved region among various HCV isolates.<sup>33</sup> In addition, the structural constraints on the 5'-UTR, in terms of its requirement to direct internal ribosome entry and translation of viral proteins, might not permit the evolution of escape mutations. Our preliminary results have shown that the siRNA-HCV suppressed replication of an HCV genotype 2a replicon<sup>34</sup> to the same extent as the HCV 1b replicon.

Although the siRNA techniques rely on a high degree of specificity, several studies report siRNA-induced non-specific effect that may result from induction of ISG responses.<sup>18,31</sup> These effects may be mediated by activation of double-strand RNA-dependent protein kinase, toll-like receptor 3,<sup>35</sup> or possibly by a recently identified RNA helicase, RIG-I.<sup>36</sup> It remains to be determined whether these effects are generally induced by every siRNA construct. Sledz *et al.* have reported that transfection of two siRNA induced cellular interferon responses,<sup>37</sup> while Bridge *et al.* report that shRNA-expressing plasmids induced an interferon response but transfection of synthetic siRNA did not.<sup>31</sup> Speculatively, these effects on the interferon system might be construct dependent. Our shRNA-expression plasmids and adenoviruses did not activate ISG responses *in vitro* (Fig. 5a,b) or *in vivo* (Fig. 7b). We have preliminarily detected phosphorylated PKR (P-PKR) by western





**Figure 7** Effects of a recombinant adenovirus expressing shRNA on HCV core protein expression in CN2-29 transgenic mice. CN2-29 transgenic mice were administered with  $1 \times 10^8$  PFU of AxCANCre combined with  $6.7 \times 10^8$  PFU of AxshRNA-HCV, AxshRNA or AxCaW1. The mice were killed on day 4 after injection. (a) Quantification of HCV core protein in liver. Liver tissues were homogenized and used to determine the amount of HCV core protein. Each assay was done in triplicate, and the values are displayed as mean  $\pm$  SD. Asterisk indicates *P*-value of less than 0.05. (b) Expression levels of mouse interferon-beta (white bars) and Mx1 (shaded bars) mRNA in the mouse liver tissue were quantified by the real-time RT-PCR analyses. Primers used were as follows: mouse interferon-beta sense, 5'-ACA GCC CTC TCC ATC AAC TA-3'; mouse interferon-beta antisense, 5'-CCC TCC AGT AAT AGC TCT TC-3'; mouse Mx1 sense, 5'-AGG AGT GGA GAG GCA AAG TC-3'; mouse Mx1 antisense, 5'-CAC ATT GCT GGG GAC TAC CA-3'; mouse beta-actin sense, 5'-ACT CCT ATG TGG GTG ACG AG-3'; mouse beta-actin antisense, 5'-ATA GCC CTC GTA GAT GGG CA-3'. Adeno (-) denotes mice without adenovirus administration. (c) Immunofluorescence microscopy of HCV core protein in the liver tissue. Liver sections of mice were stained using rabbit anticore polyclonal antibody and normal rabbit IgG as a negative control. The upper photographs were obtained at 400x magnification, and the lower photographs were at 1000x. (d) Immunofluorescence microscopy of albumin in liver. Liver sections from the mice were fixed and stained using rabbit antialbumin antibody and normal rabbit IgG as a negative control.

blotting, and found no apparent increase of P-PKR (data not shown). These results indicate that these target sequences and structures are of sufficient specificity to silence the target gene without eliciting non-specific interferon responses.

Beside the canonical action of siRNA, a sequence-specific cleavage of target mRNA, the siRNA could act as a micro-RNA

that suppresses translational initiation of mRNA,<sup>38</sup> or it could mediate transcriptional gene silencing.<sup>39</sup> Regarding our *in vivo* experiments, it was difficult to differentially analyze the effect of siRNA at individual sites of action because post-translational effect of siRNA concomitantly destabilizes target mRNA, which leads to apparent decrease of mRNA transcripts.



Efficiency and safety of gene transfer methods are the key determinants of the clinical success of gene therapy and an unresolved problem. There are several reports of delivery of siRNA or siRNA-expression vectors to cells *in vivo*;<sup>12,40,41</sup> however, gene delivery methods that are safe enough to apply to clinical therapeutics are currently under development. Adenovirus vectors are one of the most commonly used carriers for human gene therapies.<sup>42–44</sup> Our present results demonstrate that the adenoviral delivery of shRNA is effective in blocking HCV replication *in vitro* and virus protein expression *in vivo*. Adenovirus vectors have several advantages of efficient delivery of transgene both *in vitro* and *in vivo* and natural hepatotropism when administered *in vivo*. The AxshRNA-HCV specifically blocked expression of HCV structural proteins in a conditional transgenic mouse expressing those proteins. The current adenovirus vectors may cause inflammatory reactions in the target organ,<sup>45</sup> however, and produce neutralizing antibodies which make repeated administration difficult. These problems may be overcome by the improved constructs of virus vectors with attenuated immunogenicity or by the development of non-viral carriers for gene delivery.<sup>46</sup>

In conclusion, our results demonstrate the effectiveness and feasibility of the siRNA expression system. The efficiency of adenovirus expressing shRNA that target HCV suggests that delivery and expression of siRNA in hepatocytes may eliminate the virus and that this RNA-targeting approach might provide a potentially effective future therapeutic option for HCV infection.

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## References

- Alter MJ. Epidemiology of hepatitis C. *Hepatology* 1997; **26**: 62S–65S.
- Hadziyannis SJ, Sette H Jr, Morgan TR *et al.* Peginterferon-alpha2a and ribavirin combination therapy in chronic hepatitis C: a randomized study of treatment duration and ribavirin dose. *Ann. Intern. Med.* 2004; **140**: 346–55.
- Fire A, Xu S, Montgomery M, Kostas S, Driver S, Mello C. Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. *Nature* 1998; **391**: 806–11.
- Elbashir SM, Harborth J, Lendeckel W, Yalcin A, Weber K, Tuschl T. Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells. *Nature* 2001; **411**: 494–8.
- Coburn GA, Cullen BR. Potent and specific inhibition of human immunodeficiency virus type 1 replication by RNA interference. *J. Virol.* 2002; **76**: 9225–31.
- Jacque JM, Triques K, Stevenson M. Modulation of HIV-1 replication by RNA interference. *Nature* 2002; **418**: 435–8.
- Gitlin L, Karelsky S, Andino R. Short interfering RNA confers intracellular antiviral immunity in human cells. *Nature* 2002; **418**: 430–4.
- Ge Q, Filip L, Bai A, Nguyen T, Eisen HN, Chen J. Inhibition of influenza virus production in virus-infected mice by RNA interference. *Proc. Natl. Acad. Sci. USA* 2004; **101**: 8676–81.
- Wang C, Pflugheber J, Sumpter R Jr *et al.* Alpha interferon induces distinct translational control programs to suppress hepatitis C virus RNA replication. *J. Virol.* 2003; **77**: 3898–912.
- Klein C, Bock CT, Wedemeyer H *et al.* Inhibition of hepatitis B virus replication *in vivo* by nucleoside analogues and siRNA. *Gastroenterology* 2003; **125**: 9–18.
- Konishi M, Wu CH, Wu GY. Inhibition of HBV replication by siRNA in a stable HBV-producing cell line. *Hepatology* 2003; **38**: 842–50.
- McCaffrey AP, Meuse L, Pham TT, Conklin DS, Hannon GJ, Kay MA. RNA interference in adult mice. *Nature* 2002; **418**: 38–9.
- Shlomai A, Shaul Y. Inhibition of hepatitis B virus expression and replication by RNA interference. *Hepatology* 2003; **37**: 764–70.
- Yokota T, Sakamoto N, Enomoto N *et al.* Inhibition of intracellular hepatitis C virus replication by synthetic and vector-derived small interfering RNAs. *EMBO Rep.* 2003; **4**: 602–8.
- Kapadia SB, Brideau-Andersen A, Chisari FV. Interference of hepatitis C virus RNA replication by short interfering RNAs. *Proc. Natl. Acad. Sci. USA* 2003; **100**: 2014–18.
- Kronke J, Kittler R, Buchholz F *et al.* Alternative approaches for efficient inhibition of hepatitis C virus RNA replication by small interfering RNAs. *J. Virol.* 2004; **78**: 3436–46.
- Randall G, Grakoui A, Rice CM. Clearance of replicating hepatitis C virus replicon RNAs in cell culture by small interfering RNAs. *Proc. Natl. Acad. Sci. USA* 2003; **100**: 235–40.
- Seo MY, Abrignani S, Houghton M, Han JH. Letter to the editor: small interfering RNA-mediated inhibition of hepatitis C virus replication in the human hepatoma cell line Huh-7. *J. Virol.* 2003; **77**: 810–12.
- Wilson JA, Jayasena S, Khvorov A *et al.* RNA interference blocks gene expression and RNA synthesis from hepatitis C replicons propagated in human liver cells. *Proc. Natl. Acad. Sci. USA* 2003; **100**: 2783–8.
- Guo JT, Bichko VV, Seeger C. Effect of alpha interferon on the hepatitis C virus replicon. *J. Virol.* 2001; **75**: 8516–23.
- Tanabe Y, Sakamoto N, Enomoto N *et al.* Synergistic inhibition of intracellular hepatitis C virus replication by combination of ribavirin and interferon-alpha. *J. Infect. Dis.* 2004; **189**: 1129–39.
- Maekawa S, Enomoto N, Sakamoto N *et al.* Introduction of NS5A mutations enables subgenomic HCV-replicon derived from chimpanzee-infectious HC-J4 isolate to replicate efficiently in Huh-7 cells. *J. Viral. Hepat.* 2004; **11**: 394–403.
- Miyagishi M, Sumimoto H, Miyoshi H, Kawakami Y, Taira K. Optimization of a siRNA-expression system with an improved hairpin and its significant suppressive effects in mammalian cells. *J. Gene Med.* 2004; **6**: 715–23.
- Li Y, Yokota T, Matsumura R, Taira K, Mizusawa H. Sequence-dependent and independent inhibition specific for mutant ataxin-3 by small interfering RNA. *Ann. Neurol.* 2004; **56**: 124–9.
- Kanazawa N, Kurosaki M, Sakamoto N *et al.* Regulation of hepatitis C virus replication by interferon regulatory factor-1. *J. Virol.* 2004; **78**: 9713–20.
- Wakita T, Pietschmann T, Kato T *et al.* Production of infectious hepatitis C virus in tissue culture from a cloned viral genome. *Nat. Med.* 2005; **11**: 791–6.
- Zhong J, Gastaminza P, Cheng G *et al.* Robust hepatitis C virus infection in vitro. *Proc. Natl. Acad. Sci. USA* 2005; **102**: 9294–9.
- Wakita T, Taya C, Katsume A *et al.* Efficient conditional transgene expression in hepatitis C virus cDNA transgenic mice mediated by the Cre/loxP system. *J. Biol. Chem.* 1998; **273**: 9001–6.
- Kashiwakuma T, Hasegawa A, Kajita T *et al.* Detection of hepatitis C virus specific core protein in serum of patients by a sensitive fluorescence enzyme immunoassay (FEIA). *J. Immunol. Methods* 1996; **28**: 79–89.

- 30 Baglioni C, Nilsen TW. Mechanisms of antiviral action of interferon. *Interferon* 1983; **5**: 23–42.
- 31 Bridge A, Pebernard S, Ducreaux A, Nicoulaz A, Iggo R. Induction of an interferon response by RNAi vectors in mammalian cells. *Nat. Genet.* 2003; **34**: 263–4.
- 32 Carmichael GG. Silencing viruses with RNA. *Nature* 2002; **418**: 379–80.
- 33 Okamoto H, Okada S, Sugiyama Y *et al.* Nucleotide sequence of the genomic RNA of hepatitis C virus isolated from a human carrier: comparison with reported isolates for conserved and divergent regions. *J. Gen. Virol.* 1991; **72**: 2697–704.
- 34 Kato T, Date T, Miyamoto M *et al.* Efficient replication of the genotype 2a hepatitis C virus subgenomic replicon. *Gastroenterology* 2003; **125**: 1808–17.
- 35 Alexopoulou L, Holt AC, Medzhitov R, Flavell RA. Recognition of double-stranded RNA and activation of NF- $\kappa$ B by Toll-like receptor 3. *Nature* 2001; **413**: 732–8.
- 36 Yoneyama M, Kikuchi M, Natsukawa T *et al.* The RNA helicase RIG-I has an essential function in double-stranded RNA-induced innate antiviral responses. *Nat. Immunol.* 2004; **5**: 730–7.
- 37 Sledz C, Holko M, de Veer M, Silverman R, Williams B. Activation of the interferon system by short-interfering RNAs. *Nat. Cell. Biol.* 2003; **5**: 834–9.
- 38 Doench JG, Petersen CP, Sharp PA. siRNAs can function as miRNAs. *Genes Dev.* 2003; **17**: 438–42.
- 39 Morris KV. siRNA-mediated transcriptional gene silencing: the potential mechanism and a possible role in the histone code. *Cell. Mol. Life Sci.* 2005; **62**: 3057–66.
- 40 Xia H, Mao Q, Paulson HL, Davidson BL. siRNA-mediated gene silencing in vitro and in vivo. *Nat. Biotechnol.* 2002; **20**: 1006–10.
- 41 Zender L, Hutker S, Liedtke C *et al.* Caspase 8 small interfering RNA prevents acute liver failure in mice. *Proc. Natl. Acad. Sci. USA* 2003; **100**: 7797–802.
- 42 Akli S, Caillaud C, Vigne E *et al.* Transfer of a foreign gene into the brain using adenovirus vectors. *Nat. Genet.* 1993; **3**: 224–8.
- 43 Bajocchi G, Feldman SH, Crystal RG, Mastrangeli A. Direct in vivo gene transfer to ependymal cells in the central nervous system using recombinant adenovirus vectors. *Nat. Genet.* 1993; **3**: 229–34.
- 44 Davidson BL, Allen ED, Kozarsky KF, Wilson JM, Roessler BJ. A model system for in vivo gene transfer into the central nervous system using an adenoviral vector. *Nat. Genet.* 1993; **3**: 219–23.
- 45 Yang Y, Wilson JM. Clearance of adenovirus-infected hepatocytes by MHC class I-restricted CD4+ CTLs in vivo. *J. Immunol.* 1995; **155**: 2564–70.
- 46 Fleury S, Driscoll R, Simeoni E *et al.* Helper-dependent adenovirus vectors devoid of all viral genes cause less myocardial inflammation compared with first-generation adenovirus vectors. *Basic Res. Cardiol.* 2004; **99**: 247–56.



# Targeting Lipid Metabolism in the Treatment of Hepatitis C Virus Infection

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Recently, microdomains of organelle membranes rich in sphingomyelin and cholesterol (called "lipid rafts") have been considered to act as a scaffold for the hepatitis C virus (HCV) replication complex. Using the HCV cell culture system, we investigated the effect of myriocin, a sphingomyelin synthesis inhibitor, on HCV replication. We also investigated the combined effect of myriocin with interferon (IFN) and myriocin with simvastatin. Myriocin suppressed replication of both a genotype 1b subgenomic HCV replicon (Huh7/Rep-Feo) and genotype 2a infectious HCV (JFH-1 HCV) in a dose-dependent manner (for subgenomic HCV-1b, maximum of 79% at 1000 nmol/L; for genomic HCV-2a, maximum of 40% at 1000 nmol/L). Combination treatment with myriocin and IFN or myriocin and simvastatin attenuated HCV RNA replication synergistically in Huh7/Rep-Feo cells. Our data demonstrate that the sphingomyelin synthesis inhibitor strongly suppresses replication of both the subgenomic HCV-1b replicon and the JFH-1 strain of genotype 2a infectious HCV, indicating that lipid metabolism could be a novel target for HCV therapy.

Hepatitis C virus (HCV) is a major etiologic agent of liver diseases, affecting 170 million people worldwide [1]. Fifty-five percent to 85% of acute infections become persistent [2], and at least 20% of patients with chronic HCV infection progress to cirrhosis within 20 years [3]. With therapeutic advances, including the recent combination of pegylated interferon (IFN) plus ribavirin, half of patients can achieve a sustained virologic response [4]. However, the remaining half cannot clear the virus, demonstrating a strong need for HCV-specific therapies.

Positive-strand RNA viruses replicate intracellularly on certain membrane structures, including the endoplasmic reticulum [5], the Golgi apparatus [6], endo-

somes, and lysosomes [7]. During replication, RNA viruses form distinct replication complexes made of several membrane compartments and viral proteins [8]. In HCV, the membranous web (consisting of vesicles in a membranous matrix) has been described in the cellular matrix of HCV replicon-harboring cells [9, 10]. This membranous web is considered to be the HCV replication complex, consisting of viral and host proteins.

Recent studies suggest that the HCV replication complexes are formed on lipid rafts (which are detergent-insoluble microdomains of intracellular vesicular membranes rich in cholesterol and sphingolipid) [11–13]. It has been reported that viral nonstructural proteins and both positive- and negative-sense HCV RNAs were localized distinctively in a fraction of lipid rafts when subgenomic HCV replicon cells were subjected to membrane flotation analysis [12]. On the other hand, recent studies have demonstrated that agents related to lipid metabolism affect the replication of genotype 1 HCV. Leu et al. [14] reported that polyunsaturated fatty acids exerted strong anti-HCV activity on a subgenomic HCV-1b replicon. Moreover, 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (statins), which prevent cholesterol synthesis, have been shown to suppress replication of ge-

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nomic and subgenomic HCV-1b replicons [15, 16]. Even though the precise mechanism has not been defined, these agents may attenuate HCV replication through the destruction of lipid rafts, according to their pharmacological actions. If this is the mechanism, sphingomyelin, the remaining and essential component of lipid rafts, might play a role in HCV replication. With this in view, recent studies have demonstrated that a sphingomyelin synthesis inhibitor attenuated the replication of a subgenomic HCV-1b replicon in cultured cells [17] and the replication of genomic HCV-1 in a chimeric mouse model [18]. However, investigation of anti-HCV activity in these agents has been limited to genotype 1 HCV, and the combined effect of these agents has not been determined. If they do not target the HCV structure itself but exert their antiviral activity through destruction of the host's lipid raft, it would be plausible to speculate that they might be effective irrespective of the viral isolate, and the combined effect of these agents might be additive or synergistic.

In the present study, we investigated the role played by the sphingomyelin synthesis pathway and the mevalonate pathway in HCV replication, using a subgenomic HCV-1b replicon and the particle-producing cell culture HCV 2a model of JFH-1 HCV [19].

## MATERIALS AND METHODS

**Cell culture and HCV replicon.** The human hepatoma cell lines Huh7 and Huh7.5.1 [20] were maintained in Dulbecco's modified Eagle's medium (Sigma) supplemented with 10% fetal calf serum at 37°C in 5% CO<sub>2</sub>. The subgenomic HCV replicon used was derived from Rep-Feo (genotype 1b) [21, 22], and a full-length genomic HCV RNA was derived from genotype 2a JFH-1 HCV [19]. Subgenomic or genomic HCV RNA was synthesized from replicon cDNA-harboring plasmids (pRep-Feo and pJFH-1) by means of T7 polymerase (RiboMax Large Scale RNA Production System; Promega) and transfected into these cells. For the subgenomic replicon, cell lines stably expressing the replicon were established (Huh7/Rep-Feo) in the presence of 500 µg/mL G418.

**Reporter plasmids and luciferase assay.** pSRE-TA-Rluc expressing the *Renilla* luciferase reporter gene under control of the IFN-stimulated response element (ISRE) was constructed by replacing the firefly luciferase gene with the *Renilla* luciferase gene of pSRE-TA-Luc, purchased from Invitrogen. Luciferase activity was quantified using the Bright-Glo or Dual-Luciferase assay system (both from Promega) and a luminometer (AB-2250; ATTO). Assays were performed in triplicate, and the results were expressed as mean ± SD percentages of the control values. QuantiLum recombinant luciferase (Promega) was used as the positive control for the analysis.

**Reagents.** The reagents used included myriocin (Biomol), IFN-α 2b (Santa Cruz Biotechnology), phytosphingosine hydrochloride (Sigma), 2-hydroxypropyl-β-cyclodextrin (2-HP-β-CyD; Sigma), and simvastatin (Cosmobio).

**Northern blotting.** Total cellular RNA was extracted from cells by means of Isogen (Wako). The RNA was separated by denaturing agarose-formaldehyde gel electrophoresis and transferred to a membrane from a NorthernMax kit (Ambion). The membrane was hybridized with a digoxigenin-labeled probe that was specific for the nonstructural replicon sequence. The signals were detected in a chemiluminescence reaction by using a digoxigenin detection kit (Roche) and were visualized by using an LAS-1000 imaging system (Fuji Film).

**Western blotting.** Ten micrograms of total cell lysate was separated using NuPAGE 4%–12% Bis-Tris gel (Invitrogen) and was blotted onto an Immobilon polyvinylidene difluoride membrane (Roche). The membrane was incubated with an anti-core monoclonal antibody (MAB; Affinity Bioreagents), an anti-NS3 MAB (Virogen), an anti-NS5A MAB (gift from Burckstummer, Robert Koch Institute), or a anti-β-catenin MAB (Sigma). Detection was done in a chemiluminescence reaction (ECL; Amersham).

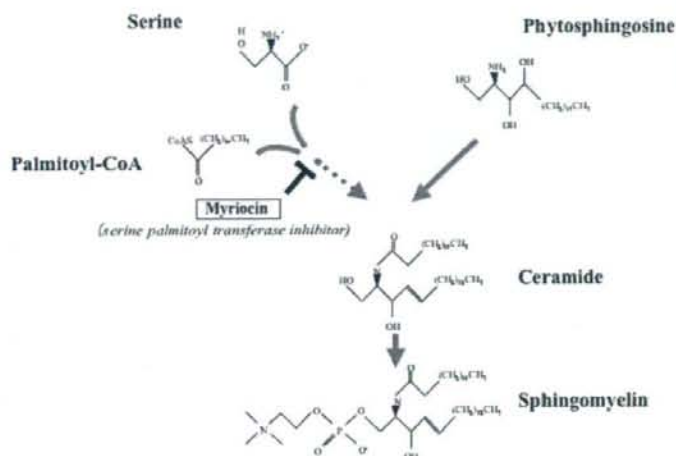
**Dimethylthiazol carboxymethoxyphenyl sulfophenyl tetrazolium (MTS) assays.** To evaluate cytotoxicity, MTS assays were performed using a CellTiter 96 Aqueous One Solution Cell Proliferation Assay (Promega), in accordance with the manufacturer's instructions.

**Thin-layer chromatography (TLC).** The lipid fraction of cells treated with myriocin was extracted using the method of Bligh and Dyer [23], and total lipids from the cells treated with myriocin were extracted with 3 mL of chloroform. The extracts were spotted onto silica gel TLC plates (Merck) and were chromatographed with chloroform-methanol-water (65:25:4 [vol/vol/vol]). The plate was visualized with a molybdenum spray.

**Real-time reverse-transcription polymerase chain reaction (RT-PCR).** TaqMan RT-PCR targeting the 5' untranslated region was used for the quantitation of intracellular genomic JFH-1 HCV RNA. The sequences of the sense and antisense primers and the TaqMan probe were 5'-TGCGGAACCGGTGAGTACA-3', 5'-CTTAAGTTTAGGATTCGTGCTCAT-3', and 5'-(FAM)CACCTATCAGGCAGTACCACAAGGCC(TAMRA)-3', respectively. The method has been described elsewhere [24].

**Short interfering RNA (siRNA) analysis.** The sequence encoding the LCB1 subunit of serine palmitoyltransferase (SPT) was selected as the target for siRNA (sense, 5'-AACAA-CAUCGUUUCAGGUCCUTT-3'; antisense, 5'-AGGGCCUG-AAACGAUGUUGTT-3'). siRNA targeting enhanced green fluorescent protein (GFP) was used as the negative control (sense, 5'-CUUACGCUGAGUACUUCGATT-3'; antisense, 5'-UCG-AAGUACUCAGCGUAAATT-3'). (Underlined letters indicate deoxyribonucleotides.)





**Figure 1.** The sphingomyelin synthesis pathway. Serine palmitoyltransferase catalyzes the first committed step of sphingomyelin biosynthesis from serine and palmitoyl-coenzyme A (CoA). Myriocin inhibits the catalyzing activity of serine palmitoyltransferase. Phytosphingosine is known to work as a precursor of ceramide in both mammalian and fungal cells.

**Statistical analyses.** Statistical analyses were performed using Student's *t* test; statistically significant differences were defined as those for which  $P < .05$ .

## RESULTS

**Specific suppression of the replication of a subgenomic HCV-1b replicon by an inhibitor of sphingomyelin synthesis.** To clarify the role played by the sphingomyelin synthesis pathway in HCV replication, we added myriocin, a specific inhibitor of SPT that catalyzes the first committed step of sphingomyelin biosynthesis (figure 1), to the medium of Huh7/Rep-Feo cells. The luciferase activity, reflecting replication of the subgenomic HCV-1b replicon, dropped to 37% and 21% of the control at myriocin concentrations of 100 and 1000 nmol/L, respectively (figure 2A, upper panel), but myriocin did not cause toxicity to the cultured cells (figure 2A, lower panel). The result indicates that the decrease in HCV replication is due to a specific suppressive effect of myriocin and not to the cytotoxicity of myriocin. Northern hybridization analysis also demonstrated a substantial reduction of the subgenomic HCV replicon RNA in Huh7/Rep-Feo cells treated with myriocin in a dose-dependent manner (figure 2B). Similarly, Western blot analysis demonstrated a decrease in HCV NS5A after treatment with myriocin (figure 2C).

**No enhancement of ISRE promoter activity after myriocin treatment.** To determine whether the effect of myriocin in suppressing the subgenomic HCV replicon was associated with the activation of IFN-stimulated genes, the ISRE-*Renilla* luciferase plasmid was transfected into Huh7/Rep-Feo cells, and these cells were cultured with various concentrations of myriocin. As a positive control for the enhancement of ISRE reporter

activity, the ISRE-*Renilla* luciferase-transfected cells were cultured with IFN. Myriocin had no significant effect on ISRE promoter activity, whereas IFN significantly up-regulated ISRE activity (figure 2D, upper panel). In contrast, firefly luciferase activity in the Huh7/Rep-Feo cells, reflecting HCV replication, was inhibited by both IFN and myriocin in a dose-dependent manner (figure 2D, lower panel). These results demonstrate that the action of myriocin on HCV replication is independent of the IFN pathway.

**Decrease in the sphingomyelin content of Huh7 cells after myriocin treatment.** To clarify whether myriocin really inhibits the biosynthesis of sphingomyelin in Huh7 cells, we treated Huh7 cells with 100 nmol/L myriocin and analyzed the change in the cellular phospholipid composition by TLC. As demonstrated in figure 2E, the cellular sphingomyelin content decreased after myriocin treatment, but no significant change was observed in other cellular phospholipids.

**Restoration of HCV replication by addition of phytosphingosine.** To confirm that suppression of HCV RNA replication was due to depletion of sphingomyelin, we incubated replicon cells with phytosphingosine, a precursor of ceramide in mammalian and fungal cells, in the presence of myriocin. Treatment with phytosphingosine restored HCV replication in a dose-dependent manner (figure 2F, upper panel). On the other hand, phytosphingosine by itself did not have any effect on HCV replication (figure 2F, lower panel). This result indicates that inhibition of HCV replication was the direct result of depletion of sphingomyelin.

**Suppression of HCV replication by knocking down SPT with siRNA.** Next, we determined whether inhibition of SPT expression suppresses HCV replication by knocking down SPT with siRNA. As demonstrated in the upper panel of

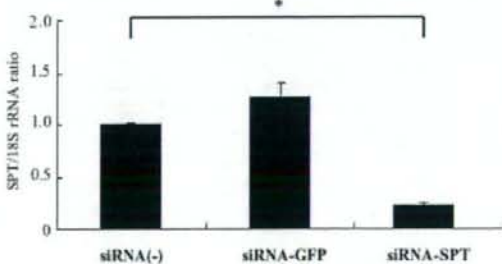
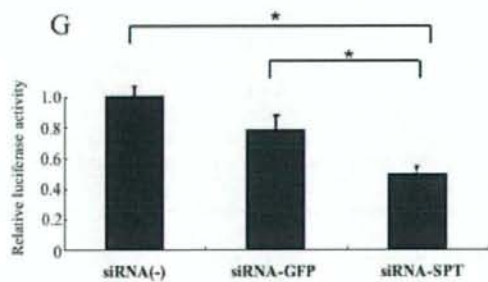
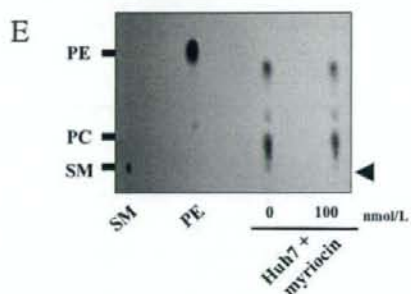
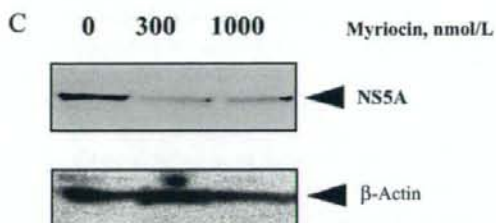
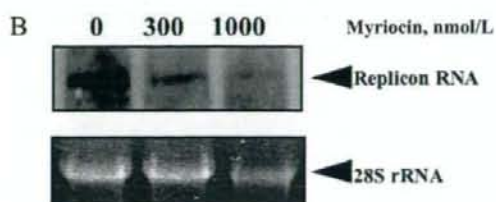
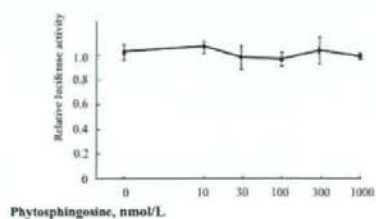
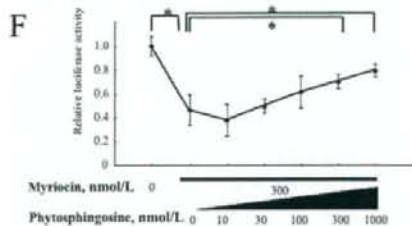
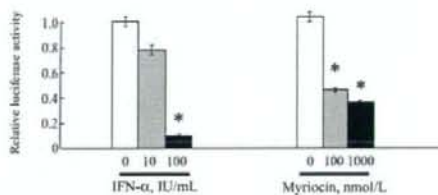
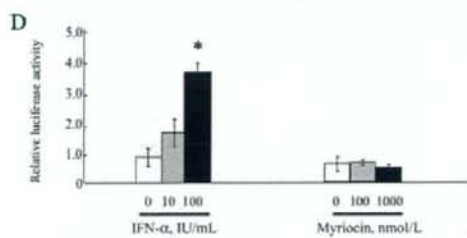
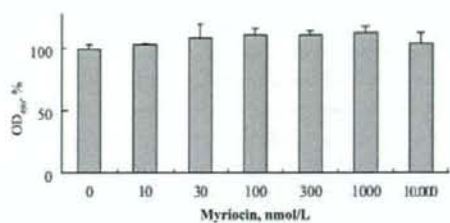
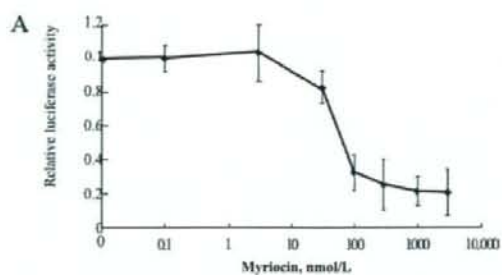




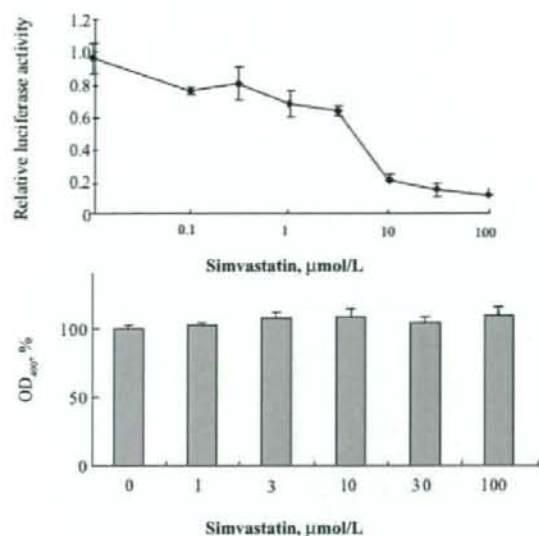
figure 2G, HCV replication was suppressed significantly by siRNA targeting SPT compared with no siRNA or siRNA targeting GFP (negative control). We confirmed with real-time PCR that the siRNA targeting SPT significantly decreased expression of SPT mRNA (figure 2G, lower panel). This result indicates that the SPT enzyme plays an important role in HCV replication.

**Inhibition of the replication of a subgenomic HCV-1b replicon by an HMG-CoA reductase inhibitor (simvastatin).** HMG-CoA reductase inhibitors have been reported to suppress replication of subgenomic and genomic HCV-1b replicons [15, 16]. Because cholesterol is another important component of lipid rafts, it may be speculated that depletion of cholesterol by HMG-CoA reductase inhibitors disrupts the lipid raft, affecting the ability of the HCV replicon to replicate in Huh7 cells. To confirm the effect of HMG-CoA reductase inhibitors on the subgenomic HCV-1b replicon, we examined the effect of simvastatin by means of Huh7/Rep-Feo cells. Cultures of Huh7/Rep-Feo cells with simvastatin at concentrations of 0–100  $\mu\text{mol/L}$  showed a dose-dependent reduction of the subgenomic HCV-1b replicon (figure 3, upper panel). The MTS assay showed that treatment with simvastatin had no toxic effect on Huh7/Rep-Feo cells in the dose range used (figure 3, lower panel). These results demonstrated that simvastatin specifically suppressed replication of a subgenomic HCV-1b replicon. However, because recent studies showed that statins suppress HCV replication through inhibition of geranylgeranylation of certain proteins rather than inhibition of cholesterol synthesis [15], we also

examined the effect on HCV replication of 2-HP- $\beta$ -CyD, an agent known to deplete cholesterol directly from membranes. As demonstrated in figure 4A, 2-HP- $\beta$ -CyD also suppressed HCV replication without cytotoxicity. To confirm that 2-HP- $\beta$ -CyD did not inhibit firefly luciferase activity nonspecifically rather than by suppressing HCV RNA, we incubated recombinant firefly luciferase with various concentrations of 2-HP- $\beta$ -CyD in the culture medium, and the medium was subjected to luciferase analysis. As demonstrated in figure 4B, 2-HP- $\beta$ -CyD did not affect luciferase activity. These results indicate that cholesterol itself plays an important role in HCV replication.

**Synergistic inhibitory effects of myriocin with IFN, simvastatin with IFN, and myriocin with simvastatin.** We carried out the following assay to determine whether myriocin and IFN have a synergistic inhibitory effect on HCV replication. Huh7/Rep-Feo cells were treated with combinations of myriocin and IFN at various concentrations. The relative dose-inhibition curves of IFN were plotted for each fixed concentration of myriocin (0, 30, 100, and 300 nmol/L). As demonstrated in the upper panel of figure 5A, the curves shifted to the left with increasing concentrations of myriocin, demonstrating the synergy of the 2 drugs against the subgenomic HCV-1b replicon. Isobologram analysis also confirmed the synergy (figure 5A, lower panel). To determine whether this synergistic effect was associated with up-regulation of the IFN-stimulated gene responses, we investigated the combined effect of myriocin and IFN on ISRE activity. As demonstrated in figure 5B (upper panel, right), myriocin did not enhance the ISRE-*Renilla* luciferase activity induced by IFN, but

**Figure 2.** Specific inhibition of the replication of a subgenomic hepatitis C virus (HCV) genotype 1b replicon by myriocin. *A*, Inhibition of HCV replicon replication by myriocin. By use of Huh7/Rep-Feo cells expressing a selectable chimeric luciferase reporter Feo gene, the intracellular replication level of an HCV replicon was quantified on the basis of luciferase activity [22, 25]. Huh7/Rep-Feo cells were cultured with various concentrations of myriocin. After 96 h of treatment, the luciferase assay was performed, as described in Materials and Methods (upper panel). In the dimethylthiazol carboxymethoxyphenyl sulfophenyl tetrazolium (MTS) assay, Huh7/Rep-Feo cells were cultured with various concentrations of myriocin for 96 h (lower panel). Data are means  $\pm$  SDs of triplicates from 2 independent experiments. *B*, Northern hybridization. Huh7/Rep-Feo cells were cultured with various concentrations of myriocin and harvested at 96 h after administration. Ten micrograms of total cellular RNA was electrophoresed in each lane. The membrane containing the HCV replicon RNA was hybridized using a digoxigenin-labeled probe specific for the replicon sequence (upper panel), and 28S human ribosomal RNA (rRNA) was used as an internal control (lower panel). Lane 1, no myriocin; lane 2, 300 nmol/L myriocin; lane 3, 1000 nmol/L myriocin. *C*, Western blotting. Ten micrograms of total cellular protein was electrophoresed in each lane. Anti-NS5A monoclonal antibody was used as the primary antibody to detect HCV proteins (upper panel), and  $\beta$ -actin was used as an internal control (lower panel). Lane 1, no myriocin; lane 2, 300 nmol/L myriocin; and lane 3, 1000 nmol/L myriocin. *D*, No enhancement of interferon (IFN)-stimulated response element (ISRE) promoter activity by myriocin. To investigate whether the effect of myriocin was associated with the activation of IFN-stimulated genes, the ISRE-*Renilla* luciferase plasmid was transfected into Huh7/Rep-Feo cells in the presence of myriocin. The upper panel demonstrates the ISRE-*Renilla* luciferase activity at 48 h after transfection. The lower panel demonstrates the firefly luciferase activity of the Huh7/Rep-Feo cells, reflecting HCV replication. Data are means  $\pm$  SDs of triplicates from 2 independent experiments. \* $P < .05$ . *E*, Decrease in the sphingomyelin (SM) content of Huh7 cells after myriocin treatment. The change in the cellular phospholipid content was analyzed by thin-layer chromatography. Huh7 cells were cultured alone or with 100 nmol/L myriocin for 96 h. PC, phosphatidylcholine; PE, phosphatidylethanolamine. *F*, Restoration of the HCV replication that was suppressed by myriocin after the addition of phytosphingosine. Huh7/Rep-Feo cells were cultured with myriocin alone or with various concentrations of phytosphingosine. The luciferase assay was performed after 72 h of treatment (upper panel). Huh7/Rep-Feo cells were also cultured with phytosphingosine alone as indicated for 72 h (lower panel). Data are means  $\pm$  SDs of triplicates from 2 independent experiments. \* $P < .05$ . *G*, Suppression of HCV replication by knocking down of serine palmitoyltransferase (SPT) with short interfering RNA (siRNA). Huh7/Rep-Feo cells were transfected with 10 nmol/L siRNA oligonucleotides targeting the LCB1 subunit of SPT or control siRNA targeting green fluorescent protein (GFP). The luciferase activity of the HCV replicon was measured 72 h after transfection (upper panel). SPT mRNA expression at 72 h after siRNA transfection was analyzed by real-time polymerase chain reaction. The SPT mRNA level was measured relative to 18S rRNA (lower panel). Values are shown as ratios to negative control levels and as the means  $\pm$  SDs of triplicates from 2 independent experiments. siRNA(-), no siRNA. \* $P < .05$ .

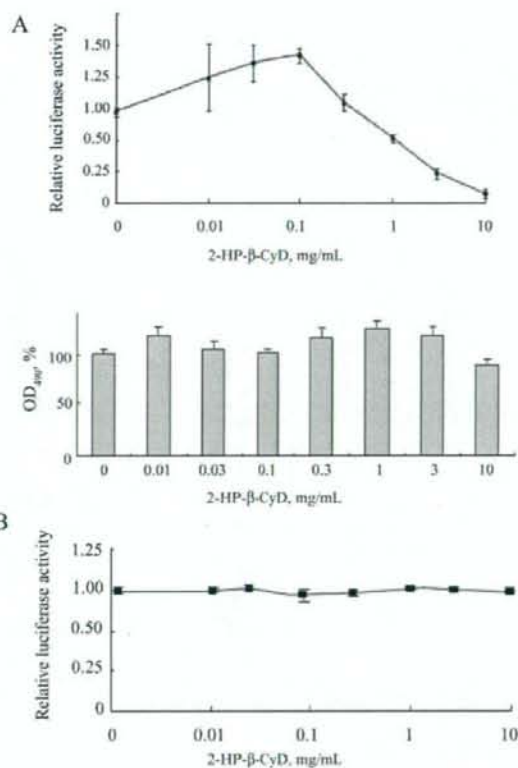


**Figure 3.** Inhibition of replication of a subgenomic hepatitis C virus genotype 1b replicon by simvastatin. Huh7/Rep-Feo cells were cultured with various concentrations of simvastatin, and the luciferase assay was performed after 48 h of treatment (*upper panel*). The dimethylthiazol carboxymethoxyphenyl sulfophenyl tetrazolium assay was performed after Huh7/Rep-Feo cells were cultured with various concentrations of simvastatin for 48 h (*lower panel*). Data are means  $\pm$  SDs of triplicates from 2 independent experiments.

it significantly enhanced IFN-induced suppression of the firefly luciferase activity reflecting HCV replication (*lower panel, right*). This demonstrated that the synergistic effect was not caused by up-regulation of the IFN-stimulated genes. We also assessed the synergy of simvastatin with IFN and of myriocin with simvastatin. In each case, the 2 drugs showed synergistic effects at the concentrations indicated (figure 5C and 5D). In all cases, the MTS reduction values at the drug concentrations used in this assay did not show any significant decrease (data not shown). These results indicate that the synergistic effects on HCV replication of IFN with myriocin, IFN with simvastatin, and myriocin with simvastatin were exerted through their pharmacological effects and were not due to the augmentation of cytotoxicity.

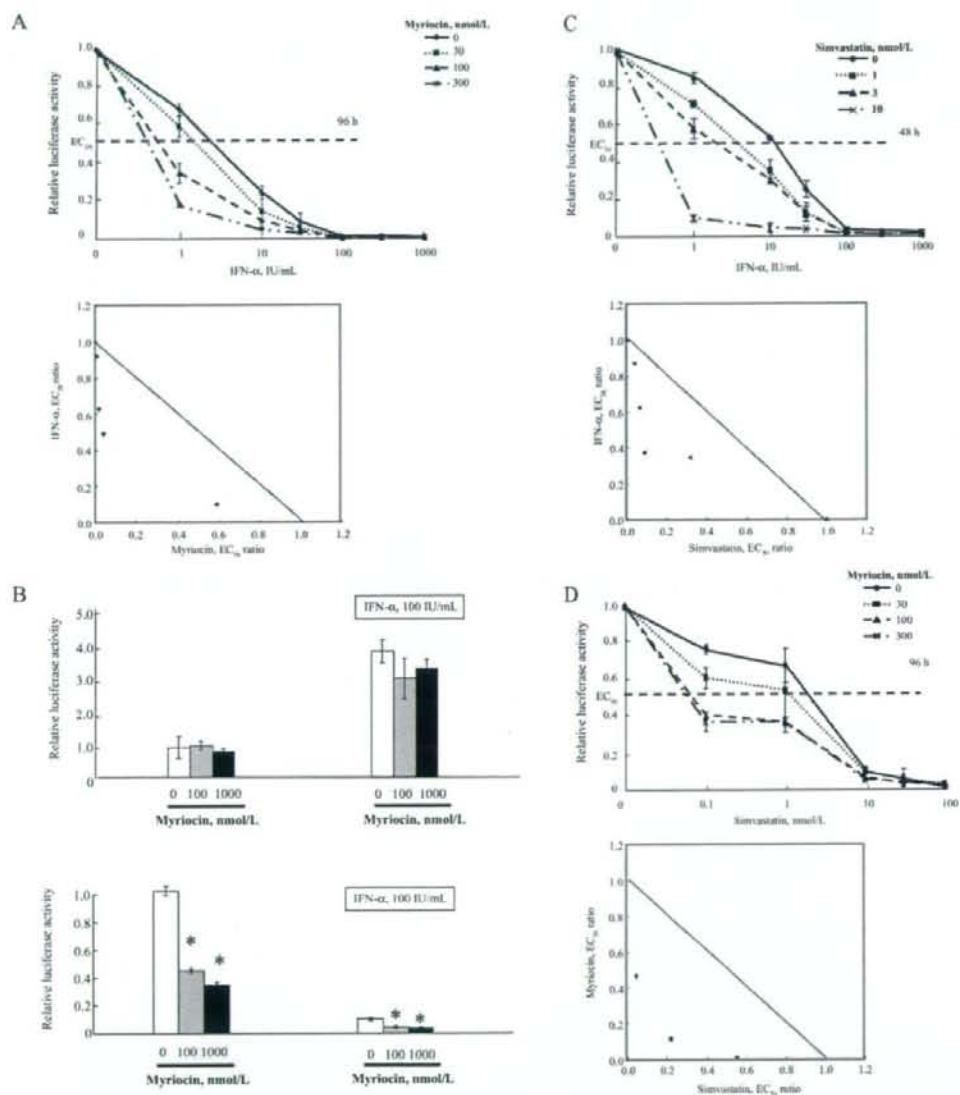
**Suppression of JFH-1 HCV replication by myriocin and simvastatin.** The experiments described thus far were done using the subgenomic HCV-1b replicon system. Recently, Wakita et al. [19] established an infectious HCV model in cultured cells. This system, known as the JFH-1 system and based on genotype 2a HCV, secretes viral particles into the medium, and the medium is infectious for chimpanzees. This JFH-1 system completely mimics HCV infection *in vivo* and is considered more suitable for analyzing the effect of drugs. Therefore, we

examined the effect of myriocin and simvastatin using the JFH-1 system. Huh7.5.1/JFH-1 HCV cells were cultured for 96 h with 1000 nmol/L myriocin, 10 μmol/L simvastatin, 1000 IU/mL IFN, and a combination of 1000 nmol/L myriocin and 10 μmol/L simvastatin. The intracellular JFH-1 HCV RNA titer was analyzed using real-time RT-PCR. As demonstrated in figure 6A, intracellular JFH-1 HCV RNA treated with myriocin or simvastatin decreased to 60% of control in 96 h, demonstrating that the inhibitory effect of myriocin and simvastatin on replication was not restricted to the subgenomic HCV-1b replicon. When both agents were used in combination, JFH-1 HCV RNA also



**Figure 4.** Inhibition of replication of a subgenomic hepatitis C virus genotype 1b replicon by 2-hydroxypropyl- $\beta$ -cyclodextrin (2-HP- $\beta$ -CyD). *A*, Huh7/Rep-Feo cells cultured with various concentrations of 2-HP- $\beta$ -CyD for 48 h. The luciferase assay was performed after 48 h of treatment (*upper panel*). The dimethylthiazol carboxymethoxyphenyl sulfophenyl tetrazolium assay was performed after Huh7/Rep-Feo cells were cultured with various concentrations of 2-HP- $\beta$ -CyD for 48 h (*lower panel*). Data are means  $\pm$  SDs of triplicates from 2 independent experiments. *B*, Recombinant firefly luciferase incubated with various concentrations of 2-HP- $\beta$ -CyD in the culture medium at 37°C for 48 h. The medium was collected and subjected to luciferase analysis. Data are means  $\pm$  SDs of triplicates from 2 independent experiments.

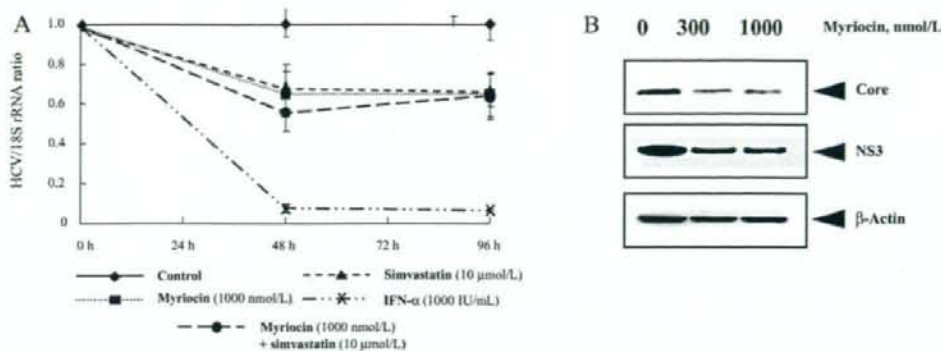




**Figure 5.** Synergistic inhibitory effects of myriocin with interferon (IFN), simvastatin with IFN, and myriocin with simvastatin. **A**, Synergistic inhibitory effect of myriocin with IFN on hepatitis C virus replication. Huh7/Rep-Feo cells were treated with combinations of myriocin and IFN at various concentrations. The upper panel shows the relative dose-inhibition curves of IFN plotted for each fixed concentration of myriocin (0, 30, 100, and 300 nmol/L). The lower panel shows the isobologram analysis for the combination of myriocin with IFN. **B**, IFN-stimulated response element (ISRE) promoter activity induced by a combination of myriocin with IFN. Huh7/Rep-Feo cells transfected with ISRE-*Renilla* luciferase were cultured with various concentrations of myriocin alone (left) or with 100 IU/mL IFN (right). The upper panel demonstrates the ISRE-*Renilla* luciferase activity at 48 h after transfection. The lower panel demonstrates the firefly luciferase activity of the Huh7/Rep-Feo cells, reflecting hepatitis C virus (HCV). Data are means  $\pm$  SDs of triplicates from 2 independent experiments. \* $P < .05$ . **C**, Synergistic inhibitory effect of simvastatin with IFN on HCV replication. **D**, Synergistic inhibitory effect of simvastatin and myriocin on HCV replication.

decreased to almost 60% of the control at 48 and 96 h after treatment. However, no evident synergistic inhibitory effect was observed (figure 6A). To clarify the inhibitory effect of myriocin on JFH-1 HCV, we performed Western blot analysis for JFH-1

HCV proteins. As demonstrated in figure 6B, a substantial decrease in the core and NS3 proteins of JFH-1 HCV was observed 96 h after treatment with myriocin, confirming the RT-PCR results (figure 6B).



**Figure 6.** Suppression of JFH-1 hepatitis C virus (HCV) replication by myriocin and simvastatin. *A*, Cells containing JFH-1 HCV treated for 96 h with 1000 nmol/L myriocin, 10 μmol/L simvastatin, 1000 IU/mL IFN, or a combination of 1000 nmol/L myriocin and 10 μmol/L simvastatin. The cells were collected at 48 and 96 h, and the JFH-1 HCV RNA level relative to 18S rRNA was analyzed by real-time polymerase chain reaction. Values are shown as the ratios to negative control values (cells receiving no treatment) and as means ± SDs. *B*, Western blotting. Cells containing JFH-1 HCV were treated with 300 or 1000 nmol/L of myriocin and harvested at 96 h after administration. Ten micrograms of total cellular protein was electrophoresed in each lane. Anti-core monoclonal antibody (MAb) and anti-NS3 MAb were used as the primary antibodies to detect JFH-1 HCV proteins. β-Actin was detected as an internal control. Lane 1, no myriocin; lane 2, 300 nmol/L myriocin; and lane 3, 1000 nmol/L myriocin.

## DISCUSSION

In the present study, we demonstrated that the sphingomyelin synthesis inhibitor myriocin suppressed not only replication of a subgenomic HCV-1b replicon but also replication of the JFH-1 strain of infectious genotype 2a HCV. We also demonstrated that simvastatin suppressed replication of both a subgenomic HCV-1b replicon and JFH-1 HCV. When a subgenomic HCV-1b replicon was used, the anti-HCV activity of both myriocin and simvastatin was enhanced synergistically with IFN. Moreover, when myriocin and simvastatin were used together, their anti-HCV activity was enhanced synergistically.

What is the mechanism by which myriocin suppresses viral replication? Because myriocin is a specific inhibitor of SPT, which catalyzes the first committed step of sphingomyelin biosynthesis, we speculated that myriocin exerts its action by inhibiting production of downstream substrates, especially sphingomyelin. The findings that siRNA targeted against SPT decreased HCV replication and that HCV replication was restored by addition of phytosphingosine, a precursor of sphingomyelin, demonstrated that the effect was specific to SPT activity. Moreover, the fact that treatment of Huh7 cells with myriocin did not enhance the ISRE promoter activity indicated that the inhibitory effects of myriocin were independent of those of IFN. It is known that intracellular replication of most RNA viruses occurs on certain membrane structures—including the endoplasmic reticulum, the Golgi apparatus, endosomes, and lysosomes—by making replication complexes at these sites [5–7]. For HCV, it has been reported by several groups that *in vitro* replication activity is located in the membrane fractions of cultured cells [26–28]. In addition, newly synthesized HCV RNA and the nonstructural proteins in replicon cells were colocalized in detergent-resistant

membrane structures, most likely lipid rafts [18]. Caveolin-2, a lipid raft protein, was also shown to colocalize with the non-structural proteins [18]. According to these findings, the HCV replication complex machinery is considered to form on a lipid raft. Therefore, because sphingomyelin is the major component of the lipid raft, it is plausible to speculate that myriocin disrupted lipid raft formation and inhibited HCV replication.

Cholesterol is another major component of lipid rafts and might also be targeted for anti-HCV therapy. Because cholesterol is synthesized in the mevalonate pathway, an inhibitor of the pathway might act to disrupt lipid rafts. In accordance with this concept, statins, which are HMG-CoA reductase inhibitors, already have been reported to suppress the replication of genomic and subgenomic HCV-1b replicons [15, 16]. In the present study, we also confirmed that simvastatin suppressed replication of a subgenomic HCV-1b replicon without toxicity. Moreover, we showed for the first time that the suppressive effect was also observed in an infectious HCV-2a model of JFH-1 HCV. Meanwhile, recent studies found that the effect of statins was attributable to inhibition of geranylgeranylation rather than depletion of cholesterol, because addition of geranylgeraniol rescued HCV suppression induced by statins [15]. However, although geranylgeranylation might play a role in HCV regulation, the importance of cholesterol itself has not yet been determined. To clarify further the role played by cholesterol in HCV replication, we investigated the effect of 2-HP-β-CyD, which is known to deplete cholesterol directly from cells. As demonstrated in figure 4, specific suppression of HCV replication by 2-HP-β-CyD indicated the importance of cholesterol itself for HCV replication. It is unlikely that these agents suppressed replication of the subgenomic replicon through inhibi-



tion of encephalomyocarditis virus internal ribosome entry site (EMCV-IRES) activity, because they also significantly suppressed replication of a full-length genomic HCV (JFH-1 HCV) that does not include EMCV-IRES (figure 6A; data for 2-HP- $\beta$ -CyD not shown).

Although we observed an inhibitory effect of myriocin and simvastatin on both the subgenomic HCV-1b replicon and JFH-1 HCV, there was a difference in efficacy between the 2 HCV systems; the subgenomic HCV-1b replicon was more sensitive to and was more strongly inhibited by either agent alone or in combination, compared with JFH-1 HCV. This result was unexpected, because we had speculated that these agents might be effective irrespective of the viral isolate if these agents targeted not the virus itself but rather host factors, such as lipid rafts. However, there are several differences between these 2 systems, and we cannot directly compare the results. In particular, the subgenomic HCV replicon lacks viral structural proteins and has only an HCV RNA intracellular replication step, whereas JFH-1 HCV includes all steps of the HCV life cycle. We do not know the precise target of the agents, and further studies are still needed.

Is it really possible to use these agents in clinical HCV treatment? Especially because statins have been used in the treatment of hyperlipidemia for many years worldwide with proven safety, it would be ideal if we could use statins as one therapeutic application for anti-HCV therapy. Most recently, O'Leary et al. [29] undertook a human pilot study and treated 10 patients with atorvastatin for 12 weeks; they reported that there was no statistically significant change in HCV RNA levels compared with pretreatment levels. The reason for the discrepancy between *in vitro* and *in vivo* findings is unknown. However, as also discussed by O'Leary et al., the most plausible explanation for this discrepancy is that the plasma concentrations of atorvastatin after a conventionally approved dose were unlikely to reach those found to be effective in cell culture medium. According to their calculations, to inhibit HCV RNA replication the plasma atorvastatin concentration should be 3 logs higher than that achieved by a conventional dose. However, even though it would be difficult to inhibit HCV RNA replication with statins alone, a clinical antiviral effect might be still achieved if statins were used in combination with IFN (or myriocin), because a synergistic effect was observed in our *in vitro* study. To determine the synergistic effect *in vivo*, however, further clinical trials are needed. On the other hand, although promising *in vitro*, myriocin has not yet been used for human clinical diseases, and its safety has not been established. However, in chimeric mice, the plasma myriocin concentration equivalent to culture medium effectively inhibited HCV RNA replication, and drug toxicity was not observed at this concentration [30]. This finding suggested the possibility that myriocin could be used *in vivo*, although further studies are needed.

In conclusion, we have demonstrated that inhibition of the sphingomyelin synthesis pathway and the mevalonate pathway

both effectively suppressed HCV replication *in vitro*, indicating that lipid metabolism could be an important target for new anti-HCV therapies.

## References

1. Global surveillance and control of hepatitis C. Report of a WHO consultation organized in collaboration with the Viral Hepatitis Prevention Board, Antwerp, Belgium. *J Viral Hepat* 1999; 6:35–47.
2. Hoofnagle JH. Hepatitis C: the clinical spectrum of disease. *Hepatology* 1997; 26:15S–20S.
3. Seeff LB. Natural history of chronic hepatitis C. *Hepatology* 2002; 36: S35–46.
4. Fried MW, Shiffman ML, Reddy KR, et al. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. *N Engl J Med* 2002; 347: 975–82.
5. Wengler G, Nowak T, Castle E. Description of a procedure which allows isolation of viral nonstructural proteins from BHK vertebrate cells infected with the West Nile flavivirus in a state which allows their direct chemical characterization. *Virology* 1990; 177:795–801.
6. Shi ST, Schiller JJ, Kanjanahaluethai A, Baker SC, Oh JW, Lai MM. Colocalization and membrane association of murine hepatitis virus gene 1 products and de novo-synthesized viral RNA in infected cells. *J Virol* 1999; 73:5957–69.
7. van der Meer Y, Snijder EJ, Dobbe JC, et al. Localization of mouse hepatitis virus nonstructural proteins and RNA synthesis indicates a role for late endosomes in viral replication. *J Virol* 1999; 73:7641–57.
8. Ahlquist P, Noueiry AO, Lee WM, Kushner DB, Dye BT. Host factors in positive-strand RNA virus genome replication. *J Virol* 2003; 77:8181–6.
9. Egger D, Wolk B, Gosert R, et al. Expression of hepatitis C virus proteins induces distinct membrane alterations including a candidate viral replication complex. *J Virol* 2002; 76:5974–84.
10. Moradpour D, Gosert R, Egger D, Penin F, Blum HE, Bienz K. Membrane association of hepatitis C virus nonstructural proteins and identification of the membrane alteration that harbors the viral replication complex. *Antiviral Res* 2003; 60:103–9.
11. Gao L, Aizaki H, He JW, Lai MM. Interactions between viral nonstructural proteins and host protein hVAP-33 mediate the formation of hepatitis C virus RNA replication complex on lipid raft. *J Virol* 2004; 78: 3480–8.
12. Aizaki H, Lee KJ, Sung VM, Ishiko H, Lai MM. Characterization of the hepatitis C virus RNA replication complex associated with lipid rafts. *Virology* 2004; 324:450–61.
13. Mannova P, Fang R, Wang H, et al. Modification of host lipid raft proteome upon hepatitis C virus replication. *Mol Cell Proteomics* 2006; 5: 2319–25.
14. Leu GZ, Lin TY, Hsu JT. Anti-HCV activities of selective polyunsaturated fatty acids. *Biochem Biophys Res Commun* 2004; 318:275–80.
15. Kapadia SB, Chisari FV. Hepatitis C virus RNA replication is regulated by host geranylgeranylation and fatty acids. *Proc Natl Acad Sci USA* 2005; 102:2561–6.
16. Ikeda M, Abe K, Yamada M, Dansako H, Naka K, Kato N. Different anti-HCV profiles of statins and their potential for combination therapy with interferon. *Hepatology* 2006; 44:17–25.
17. Sakamoto H, Okamoto K, Aoki M, et al. Host sphingolipid biosynthesis as a target for hepatitis C virus therapy. *Nat Chem Biol* 2005; 1:333–7.
18. Shi ST, Lee KJ, Aizaki H, Hwang SB, Lai MM. Hepatitis C virus RNA replication occurs on a detergent-resistant membrane that cofractionates with caveolin-2. *J Virol* 2003; 77:4160–8.
19. Wakita T, Pietschmann T, Kato T, et al. Production of infectious hepatitis C virus in tissue culture from a cloned viral genome. *Nat Med* 2005; 11:791–6.
20. Zhong J, Gastaminza P, Cheng G, et al. Robust hepatitis C virus infection *in vitro*. *Proc Natl Acad Sci USA* 2005; 102:9294–9.

21. Nakagawa M, Sakamoto N, Enomoto N, et al. Specific inhibition of hepatitis C virus replication by cyclosporin A. *Biochem Biophys Res Commun* 2004; 313:42-7.
22. Tanabe Y, Sakamoto N, Enomoto N, et al. Synergistic inhibition of intracellular hepatitis C virus replication by combination of ribavirin and interferon- $\alpha$ . *J Infect Dis* 2004; 189:1129-39.
23. Bligh EG, Dyer WJ. A rapid method of total lipid extraction and purification. *Can J Biochem Physiol* 1959; 37:911-7.
24. Martell M, Gomez J, Esteban JI, et al. High-throughput real-time reverse transcription-PCR quantitation of hepatitis C virus RNA. *J Clin Microbiol* 1999; 37:327-32.
25. Yokota T, Sakamoto N, Enomoto N, et al. Inhibition of intracellular hepatitis C virus replication by synthetic and vector-derived small interfering RNAs. *EMBO Rep* 2003; 4:602-8.
26. Schmidt-Mende J, Bieck E, Hugle T, et al. Determinants for membrane association of the hepatitis C virus RNA-dependent RNA polymerase. *J Biol Chem* 2001; 276:44052-63.
27. Pietschmann T, Lohmann V, Rutter G, Kurpanek K, Bartenschlager R. Characterization of cell lines carrying self-replicating hepatitis C virus RNAs. *J Virol* 2001; 75:1252-64.
28. Lai VC, Dempsey S, Lau JY, Hong Z, Zhong W. In vitro RNA replication directed by replicase complexes isolated from the subgenomic replicon cells of hepatitis C virus. *J Virol* 2003; 77:2295-300.
29. O'Leary JG, Chan JL, McMahon CM, Chung RT. Atorvastatin does not exhibit antiviral activity against HCV at conventional doses: a pilot clinical trial. *Hepatology* 2007; 45:895-8.
30. Umehara T, Sudoh M, Yasui F, et al. Serine palmitoyltransferase inhibitor suppresses HCV replication in a mouse model. *Biochem Biophys Res Commun* 2006; 346:67-73.