

innate antiviral responses during acute infection. Thus, some HCV infections may be "silent" because they minimally activate the TLR and/or IFN cellular defense systems. This would have obvious selective advantage for the virus and could contribute to the establishment of chronic infection. Alternatively, when a virus enters cells in a "noisy" fashion, it has a poor chance of establishing chronic infection because the innate antiviral responses would quickly shut down virus replication. Finally, stimulation of the IFN system by the HCV core protein may be required to balance the anti-IFN functions of other HCV proteins such as E2 (Taylor et al., 1999), NS5A (Gale et al., 1997; Polyak et al., 2001), and NS3 (Foy et al., 2003) during certain stages of the HCV replication cycle.

#### **EXTRACELLULAR VERSUS INTRACELLULAR EFFECTS OF CORE**

HCV core is found within infected cells as well as in patient serum (Kashiwakuma et al., 1996; Widell et al., 2002). Extracellular core protein likely affects the modulation of T cell function, TLR signaling and DC function as described above. Thus, it is important to consider the contribution of extracellular and intracellular core protein to the biological activity in question. Indeed, CD81 engagement by the HCV envelope glycoprotein E2 inhibits NK and T cell cytotoxic function and signal transduction (Crotta et al., 2002; Tseng and Klimpel, 2002; Wack et al., 2001), and induced pro-inflammatory chemokine expression in hepatocytes (Balasubramanian et al., 2003). Thus, immune function may be altered as cells "sample" the microenvironment through HCV-host interactions that are limited to molecules on the cell surface, such as the HCV core-TLR or core-C1qR interaction. Moreover, these extracellular HCV-host interactions may also contribute to HCV pathogenesis.

#### **HCV CORE AND PATHOGENESIS**

Recent work has demonstrated that the HCV core protein may also participate in the pathogenesis of liver disease. Development of fibrosis is characterized histologically with infiltration of inflammatory lymphocytes, hepatocellular apoptosis, and Kupffer cell activation. HSC proliferate and undergo and become highly activated, which involves secretion of large amounts of extracellular matrix proteins (Bataller and Brenner, 2005). Despite a large body of literature from clinical and animal studies on fibrosis development, very little is known about how HCV causes fibrosis.

A recent study found that addition of recombinant core protein to activated human hepatic stellate cells (HSC) stimulated intracellular signaling pathways, while viral transduction of HCV core into HSCs caused increased cell proliferation (Bataller et al., 2004). Interestingly, the HSC response appeared to differ between core and other HCV non-structural proteins. The data suggest that HCV core and non-structural proteins can modulate the activity of HSC, which may contribute to fibrosis. This study also reinforces the notion that HCV proteins can have intracellular as well as extracellular effects on a variety of cells.

A second important point from the study of Bataller et al., (Bataller and Brenner, 2005), is that HCV proteins including core induce oxidative stress on HSC which is involved in HSC activation. Indeed, antioxidant therapy reduces the effects of HCV proteins on HSCs. This finding is in line with the current thinking that oxidative stress is central to induction of fibrosis in many model systems. HCV core induced oxidative stress also affects mitochondrial physiology.

#### **HCV CORE AND MITOCHONDRIAL DYSFUNCTION**

Expression of HCV core protein in transgenic mice and in cell culture induces oxidative stress. It has been shown that core protein localizes to mitochondria, between the mitochondrial outer membrane and ER (Moriya et al., 1998; Moriya et al., 2001a; Okuda et al., 2002; Schwer et al., 2004; Suzuki et al., 2005), as described in the targeting section above. Core protein expression and mitochondrial localization inhibits electron transport at complex I, increases complex I reactive oxygen species (ROS) production, decreases mitochondrial glutathione, and increases mitochondrial permeability transition in response to exogenous oxidants such as alcohol (Korenaga et al., 2005; Okuda et al., 2002; Wen et al., 2004). These effects are associated with increased hepatocyte apoptosis in the presence of HCV core protein, ethanol and cytochrome P4502E1. Like the case with HSC, core and ethanol metabolism effects on apoptosis can be prevented with antioxidants (Otani et al., 2005).

#### **HEPATITIC STEATOSIS AND HEPATOCARCINOGENESIS**

Evidence has been accumulating that HCV core protein is directly involved in pathogenesis (Giannini and Brechot, 2003; McLauchlan, 2000). As shown in Table 1, many cellular proteins, which interact with core protein have been identified. Several studies have suggested that the core protein plays a crucial role for hepatocarcinogenesis (Chang et al., 1998; Moriya et al., 1998; Ray et al., 1996).

Recent studies have highlighted steatosis as a basis of HCV-associated HCC (Lerat et al., 2002; Moriya et al., 1998). Steatosis, which is an accumulation of fat deposits in hepatocytes, is one of the histological features of chronic hepatitis C (Bach et al., 1992; Lefkowitz, 2003). In vitro studies have shown that HCV core protein associates with cellular lipid droplets, via direct interaction with apolipoprotein A2 (Barba et al., 1997; Shi et al., 2002). The mice transgenic for HCV core gene have been shown to develop steatosis and hepatocellular carcinoma (HCC) (Moriya et al., 1998; Moriya et al., 1997b). Steatosis in the core-transgenic mice is age-dependent and characterized by the appearance of micro- and macro-vesicular lipid droplets (Moriya et al., 1998). Lerat et al. have confirmed that transgenic mice expressing the whole genome of HCV also develops steatosis and HCC (Lerat et al., 2002).



**Table 1.** Cellular proteins that bind to the HCV core protein. The list contains cellular proteins with various cellular functions that interact with HCV core. The interaction of HCV core with these cellular proteins may have pathogenic implications. Please refer to the text for details.

Core-Interacting protein	Function	Reference
Apolipoprotein AII	lipid metabolism	Sabile et al., 1999; Shi et al., 2002
CAP-Rf	RNA helicase	You et al., 1999
complement receptor gC1qR	T-cell response	Kittlesen et al., 2000
cyclin-dependent kinase 7	cell cycle	Ohkawa et al., 2004
DEAD box protein	RNA helicase	Mamiya and Worman, 1999
DEAD box protein 3	RNA helicase	Owsianka and Patel, 1999
heterogeneous nuclear ribonucleoprotein K	transcriptional control	Hsieh et al., 1998
JAK1/2	signal transduction	Hosui et al., 2003
lymphotoxin- $\beta$ receptor	cytotoxicity	Chen et al., 1997
p53	transcriptional control	Otsuka et al., 2000
p73	transcriptional control	Alisi et al., 2003
proteasome activator PA28 $\gamma$	protein stability	Morishi et al., 2003
retinoid X receptor $\alpha$	transcriptional control	Tsutsumi et al., 2002
Smad3	transcriptional control	Cheng et al., 2004
Sp110b	transcriptional control	Watashi et al., 2003
STAT3	cell transformation	Yoshida et al., 2002
TAFII28	transcriptional control	Otsuka et al., 2000
Tumor necrosis factor receptor 1	apoptosis	Zhu et al., 2001
14-3-3 protein	signal transduction	Aoki et al., 2000

Although the molecular mechanisms of steatosis caused by the core protein is still unclear, the core protein may alter lipid metabolism by interacting with cellular proteins involved in lipid accumulation and storage in hepatocytes (Barba et al., 1997; Sabile et al., 1999; Shi et al., 2002). The concentration of carbon 18 monosaturated fatty acids were increased in the livers of the core-transgenic mice and chronic hepatitis C patients, suggesting that HCV core affects a specific pathway in lipid metabolism (Moriya et al., 2001b). Nonetheless, transgenic mouse lines established by other groups did not show either steatosis nor HCC (Kawamura et al., 1997; Pasquinelli et al., 1997). These discrepancies suggest that not only the viral proteins but also other factors are involved in hepatocarcinogenesis. These discrepancies may be due to differences in genetic backgrounds of the mice and expression levels of the viral proteins.

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## REFERENCES

- Aizaki, H., Nagamori, S., Matsuda, M., Kawakami, H., Hashimoto, O., Ishiko, H., Kawada, M., Matsuura, T., Hasumura, S., Matsuura, Y., et al. (2003). Production and release of infectious hepatitis C virus from human liver cell cultures in the three-dimensional radial-flow bioreactor. *Virology* 314, 16-25.
- Alisi, A., Giambartolomei, S., Cupelli, F., Merlo, P., Fontemaggi, G., Spaziani, A., and Balsano, C. (2003). Physical and functional interaction between HCV core protein and the different p73 isoforms. *Oncogene* 22, 2573-2580.
- Anthony, D. D., Yonkers, N. L., Post, A. B., Asaad, R., Heinzl, F. P., Lederman, M. M., Lehmann, P. V., and Valdez, H. (2004). Selective impairments in dendritic cell-associated function distinguish hepatitis C virus and HIV infection. *J Immunol* 172, 4907-4916.
- Aoki, H., Hayashi, J., Moriyama, M., Arakawa, Y., and Hino, O. (2000). Hepatitis C virus core protein interacts with 14-3-3 protein and activates the kinase Raf-1. *J Virol* 74, 1736-1741.
- Bach, N., Thung, S. N., and Schaffner, F. (1992). The histological features of chronic hepatitis C and autoimmune chronic hepatitis: a comparative analysis. *Hepatology* 15, 572-577.
- Balasubramanian, A., Ganju, R. K., and Groopman, J. E. (2003). Hepatitis C virus and HIV envelope proteins collaboratively mediate interleukin-8 secretion through activation of p38 MAP kinase and SHP2 in hepatocytes. *J Biol Chem* 278, 35755-35766.
- Banks, L., Pim, D., and Thomas, M. (2003). Viruses and the 26S proteasome: hacking into destruction. *Trends Biochem Sci* 28, 452-459.
- Barba, G., Harper F, Harada T, Kohara M, Goulinet S, Matsuura Y, Eder G, Schaff Z, Chapman MJ, Miyamura T, and C. B. (1997). Hepatitis C virus core protein shows a cytoplasmic localization and associates to cellular lipid storage droplets. *Proc Natl Acad Sci USA* 175, 740-744.



- Bataller, R., and Brenner, D. A. (2005). Liver fibrosis. *J Clin Invest* 115, 209-218.
- Bataller, R., Paik, Y. H., Lindquist, J. N., Lemasters, J. J., and Brenner, D. A. (2004). Hepatitis C virus core and nonstructural proteins induce fibrogenic effects in hepatic stellate cells. *Gastroenterology* 126, 529-540.
- Baumert, T. F., Ito, S., Wong, D. T., and Liang, T. J. (1998). Hepatitis C virus structural proteins assemble into viruslike particles in insect cells. *J Virol* 72, 3827-3836.
- Baumert, T. F., Vergalla, J., Satoi, J., Thomson, M., Lechmann, M., Herion, D., Greenberg, H. B., Ito, S., and Liang, T. J. (1999). Hepatitis C virus-like particles synthesized in insect cells as a potential vaccine candidate. *Gastroenterology* 117, 1397-1407.
- Blight, K. J., Kolykhalov, A. A., and Rice, C. M. (2000). Efficient initiation of HCV RNA replication in cell culture. *Science* 290, 1972-1974.
- Bosman, C., Valli, M. B., Bertolini, L., Serafino, A., Boldrini, R., Marcellini, M., and Carloni, G. (1998). Detection of virus-like particles in liver biopsies from HCV-infected patients. *Res Virol* 149, 311-314.
- Bradley, D., McCaustland, K., Krawczynski, K., Spelbring, J., Humphrey, C., and Cook, E. H. (1991). Hepatitis C virus: buoyant density of the factor VIII-derived isolate in sucrose. *J Med Virol* 34, 206-208.
- Bukh, J., Pietschmann, T., Lohmann, V., Krieger, N., Faulk, K., Engle, R. E., Govindarajan, S., Shapiro, M., St Claire, M., and Bartenschlager, R. (2002). Mutations that permit efficient replication of hepatitis C virus RNA in Huh-7 cells prevent productive replication in chimpanzees. *Proc Natl Acad Sci USA* 99, 14416-14421.
- Chang, J., Yang, S. H., Cho, Y. G., Hwang, S. B., Hahn, Y. S., and Sung, Y. C. (1998). Hepatitis C virus core from two different genotypes has an oncogenic potential but is not sufficient for transforming primary rat embryo fibroblasts in cooperation with the H-ras oncogene. *J Virol* 72, 3060-3065.
- Chang, S. C., Yen, J. H., Kang, H. Y., Jang, M. H., and Chang, M. F. (1994). Nuclear localization signals in the core protein of hepatitis C virus. *Biochem Biophys Res Commun* 205, 1284-1290.
- Chen, C. M., You, L. R., Hwang, L. H., and Lee, Y. H. W. (1997). Direct interaction of hepatitis C virus core protein with the cellular lymphotoxin-beta receptor modulates the signal pathway of the lymphotoxin-beta receptor. *J Virol* 71, 9417-9426.
- Cheng, P. L., Chang, M. H., Chao, C. H., and Lee, Y. H. (2004). Hepatitis C viral proteins interact with Smad3 and differentially regulate TGF-beta/Smad3-mediated transcriptional activation. *Oncogene* 23, 7821-7838.
- Cristofari, G., Ivanyi-Nagy, R., Gabus, C., Boulant, S., Lavergne, J. P., Penin, F., and Darlix, J. L. (2004). The hepatitis C virus Core protein is a potent nucleic acid chaperone that directs dimerization of the viral (+) strand RNA *in vitro*. *Nucleic Acids Res* 32, 2623-2631.

- Crotta, S., Stilla, A., Wack, A., D'Andrea, A., Nuti, S., D'Oro, U., Mosca, M., Filliponi, F., Brunetto, R. M., Bonino, F., et al. (2002). Inhibition of natural killer cells through engagement of CD81 by the major hepatitis C virus envelope protein. *J Exp Med* 195, 35-41.
- Delhem, N., Sabile, A., Gajardo, R., Podevin, P., Abadie, A., Blaton, M. A., Kremsdorf, D., Beretta, L., and Brechot, C. (2001). Activation of the interferon-inducible protein kinase PKR by Hepatocellular carcinoma derived-Hepatitis C virus core protein. *Oncogene* 20, 5836-5845.
- Dolganiuc, A., Kodys, K., Kopasz, A., Marshall, C., Do, T., Romics, L., Jr., Mandrekar, P., Zapp, M., and Szabo, G. (2003). Hepatitis C virus core and nonstructural protein 3 proteins induce pro- and anti-inflammatory cytokines and inhibit dendritic cell differentiation. *J Immunol* 170, 5615-5624.
- Dolganiuc, A., Oak, S., Kodys, K., Golenbock, D. T., Finberg, R. W., Kurt-Jones, E., and Szabo, G. (2004). Hepatitis C core and nonstructural 3 proteins trigger toll-like receptor 2-mediated pathways and inflammatory activation. *Gastroenterology* 127, 1513-1524.
- Duesberg, U., von dem Bussche, A., Kirschning, C., Miyake, K., Sauerbruch, T., and Spengler, U. (2002). Cell activation by synthetic lipopeptides of the hepatitis C virus (HCV)-core protein is mediated by toll like receptors (TLRs) 2 and 4. *Immunol Lett* 84, 89-95.
- Fan, Z., Yang, Q. R., Twu, J. S., and Sherker, A. H. (1999). Specific *in vitro* association between the hepatitis C viral genome and core protein. *J Med Virol* 59, 131-134.
- Finley, D., Ciechanover, A., and Varshavsky, A. (2004). Ubiquitin as a central cellular regulator. *Cell* 116, S29-32, 22 p following S32.
- Foy, E., Li, K., Wang, C., Sumpter, R., Jr., Ikeda, M., Lemon, S. M., and Gale, M., Jr. (2003). Regulation of interferon regulatory factor-3 by the hepatitis C virus serine protease. *Science* 300, 1145-1148.
- Gale, M. J., M.J. Korth, N.M. Tang, S.L. Tan, D.A. Hopkins, T.E. Dever, S.J. Polyak, D.R. Gretch, and Katze, M. G. (1997). Evidence that hepatitis C virus resistance to interferon is mediated through repression of the PKR protein kinase by the nonstructural 5A protein. *Virology* 230, 217-227.
- Giannini, C., and Brechot, C. (2003). Hepatitis C virus biology. *Cell Death Differ* 10 Suppl 1, S27-38.
- Gonzalez-Peralta, R. P., Fang, J. W., Davis, G. L., Gish, R., Tsukiyama-Kohara, K., Kohara, M., Mondelli, M. U., Lesniewski, R., Phillips, M. I., Mizokami, M., and et al. (1994). Optimization for the detection of hepatitis C virus antigens in the liver. *J Hepatol* 20, 143-147.
- Gowans, E. J. (2000). Distribution of markers of hepatitis C virus infection throughout the body. *Seminars In Liver Disease* 20, 85-102.
- Heller, T., Saito, S., Auerbach, J., Williams, T., Moreen, T. R., Jazwinski, A., Cruz, B., Jeurkar, N., Sapp, R., Luo, G., and Liang, T. J. (2005). An *in vitro* model of hepatitis C virion production. *Proc Natl Acad Sci USA* 102, 2579-2583.



- Hershko, A., and Ciechanover, A. (1998). The ubiquitin system. *Annu Rev Biochem* 67, 425-479.
- Hertzog, P. J., O'Neill, L. A., and Hamilton, J. A. (2003). The interferon in TLR signaling: more than just antiviral. *Trends Immunol* 24, 534-539.
- Hijikata, M., Kato, N., Ootsuyama, Y., Nakagawa, M., and Shimotohno, K. (1991). Gene mapping of the putative structural region of the hepatitis C virus genome by *in vitro* processing analysis. *Proc Natl Acad Sci USA* 88, 5547-5551.
- Hope, R. G., and McLauchlan, J. (2000). Sequence motifs required for lipid droplet association and protein stability are unique to the hepatitis C virus core protein. *J. Gen. Virol.* 8, 1913-1925.
- Hope, R. G., Murphy, D. J., and McLauchlan, J. (2002). The domains required to direct core proteins of hepatitis C virus and GB virus-B to lipid droplets share common features with plant oleosin proteins. *J Biol Chem* 277, 4261-4270.
- Hosui, A., Ohkawa, K., Ishida, H., Sato, A., Nakanishi, F., Ueda, K., Takehara, T., Kasahara, A., Sasaki, Y., Hori, M., and Hayashi, N. (2003). Hepatitis C virus core protein differently regulates the JAK-STAT signaling pathway under interleukin-6 and interferon-gamma stimuli. *J Biol Chem* 278, 28562-28571.
- Hsieh, T. Y., Matsumoto, M., Chou, H. C., Schneider, R., Hwang, S. B., Lee, A. S., and Lai, M. M. C. (1998). Hepatitis C virus core protein interacts with heterogeneous nuclear ribonucleoprotein K. *J Biol Chem* 273, 17651-17659.
- Hussy, P., Langen, H., Mous, J., and Jacobsen, H. (1996). Hepatitis C virus core protein: carboxy-terminal boundaries of two processed species suggest cleavage by a signal peptide peptidase. *Virology* 224, 93-104.
- Iwasaki, A., and Medzhitov, R. (2004). Toll-like receptor control of the adaptive immune responses. *Nat Immunol* 5, 987-995.
- Kaito, M., Watanabe, S., Tsukiyama, K. K., Yamaguchi, K., Kobayashi, Y., Konishi, M., Yokoi, M., Ishida, S., Suzuki, S., and Kohara, M. (1994). Hepatitis C virus particle detected by immunoelectron microscopic study. *J Gen Virol.* 75, 1755-1756.
- Kanto, T., Hayashi, N., Takehara, T., Hagiwara, H., Mita, E., Naito, M., Kasahara, A., Fusamoto, H., and Kamada, T. (1994). Buoyant density of hepatitis C virus recovered from infected hosts: two different features in sucrose equilibrium density-gradient centrifugation related to degree of liver inflammation. *Hepatology* 19, 296-302.
- Kanto, T., Inoue, M., Miyatake, H., Sato, A., Sakakibara, M., Yakushijin, T., Oki, C., Itose, I., Hiramatsu, N., Takehara, T., et al. (2004). Reduced numbers and impaired ability of myeloid and plasmacytoid dendritic cells to polarize T helper cells in chronic hepatitis C virus infection. *J Infect Dis* 190, 1919-1926.
- Kashiwakuma, T., Hasegawa, A., Kajita, T., Takata, A., Mori, H., Ohta, Y., Tanaka, E., Kiyosawa, K., Tanaka, T., Tanaka, S., et al. (1996). Detection of hepatitis C virus specific core protein in serum of patients by a sensitive fluorescence enzyme immunoassay (FEIA). *J Immunol Methods* 190, 79-89.

- Kato, T., Miyamoto, M., Furusaka, A., Date, T., Yasui, K., Kato, J., Matsushima, S., Komatsu, T., and Wakita, T. (2003). Processing of hepatitis C virus core protein is regulated by its C-terminal sequence. *J Med Virol* 69, 357-366.
- Kawamura, T., Furusaka, A., Koziel, M. J., Chung, R. T., Wang, T. C., Schmidt, E. V., and Liang, T. J. (1997). Transgenic expression of hepatitis C virus structural proteins in the mouse. *Hepatology* 25, 1014-1021.
- Kim, W. R. (2002). The burden of hepatitis C in the United States. *Hepatology* 36, S30-34.
- Kittleson, D. J., Chianese-Bullock, K. A., Yao, Z. Q., Braciale, T. J., and Hahn, Y. S. (2000). Interaction between complement receptor gC1qR and hepatitis C virus core protein inhibits T-lymphocyte proliferation. *J Clin Invest* 106, 1239-1249.
- Klein, K. C., Dellos, S., and Lingappa, J. R. (2005). Identification of residues in the hepatitis C virus core protein that are critical for capsid assembly in a cell-free system. *J Virol* 79, in press.
- Klein, K. C., Polyak, S. J., and Lingappa, J. R. (2004). Unique features of hepatitis C virus capsid formation revealed by de novo cell-free assembly. *J Virol* 78, 9257-9269.
- Koike, K., Moriya, K., Ishibashi, K., Matsuura, Y., Suzuki, T., Saito, I., Iino, S., Kurokawa, K., and Miyamura, T. (1995). Expression of hepatitis C virus envelope proteins in transgenic mice. *J Gen Virol* 76, 3031-3038.
- Korenaga, M., Okuda, M., Otani, K., Wang, T., Li, Y., and Weinman, S. A. (2005). Mitochondrial dysfunction in hepatitis C. *J Clin Gastroenterol* 39, S162-166.
- Kunkel, M., Lorinczi, M., Rijnbrand, R., Lemon, S. M., and Watowich, S. J. (2001). Self-assembly of nucleocapsid-like particles from recombinant hepatitis C virus core protein. *J Virol* 75, 2119-2129.
- Kunkel, M., and Watowich, S. J. (2002). Conformational changes accompanying self-assembly of the hepatitis C virus core protein. *Virology* 294, 239-245.
- Lanford, R. E., Notvall, L., Chavez, D., White, R., Frenzel, G., Simonsen, C., and Kim, J. (1993). Analysis of hepatitis C virus capsid, E1, and E2/NS1 proteins expressed in insect cells. *Virology* 197, 225-235.
- Lefkowitz, J. H. (2003). Hepatobiliary pathology. *Curr Opin Gastroenterol* 19, 185-193.
- Lerat, H., Honda, M., Beard, M. R., Loesch, K., Sun, J., Yang, Y., Okuda, M., Gosert, R., Xiao, S. Y., Weinman, S. A., and Lemon, S. M. (2002). Steatosis and liver cancer in transgenic mice expressing the structural and nonstructural proteins of hepatitis C virus. *Gastroenterology* 122, 352-365.
- Lindenbach, B. D., Evans, M. J., Syder, A. J., Wolk, B., Tellinghuisen, T. L., Liu, C. C., Maruyama, T., Hynes, R. O., Burton, D. R., McKeating, J. A., and Rice, C. M. (2005). Complete replication of hepatitis C virus in cell culture. *Science* 309, 623-626.
- Lingappa, J. R., Hill, R. L., Wong, M. L., and Hegde, R. S. (1997). A multistep, ATP-dependent pathway for assembly of human immunodeficiency virus capsids in a cell-free system. *J Cell Biol* 136, 567-581.



- Lingappa, J. R., Martin, R. L., Wong, M. L., Ganem, D., Welch, W. J., and Lingappa, V. R. (1994). A eukaryotic cytosolic chaperonin is associated with a high molecular weight intermediate in the assembly of hepatitis B virus capsid, a multimeric particle. *J Cell Biol* 125, 99-111.
- Lingappa, J. R., Newman, M. A., Klein, K. C., and Doohar, J. E. (2005). Comparing capsid assembly of primate lentiviruses and hepatitis B virus using cell-free systems. *Virology* 333, 114-123.
- Lo, S. Y., Masiarz, F., Hwang, S. B., Lai, M. M., and Ou, J. H. (1995). Differential subcellular localization of hepatitis C virus core gene products. *Virology* 213, 455-461.
- Lohmann, V., Korner, F., Koch, J., Herian, U., Theilmann, L., and Bartenschlager, R. (1999). Replication of subgenomic hepatitis C virus RNAs in a hepatoma cell line. *Science* 285, 110-113.
- Lorenzo, L. J., Duenas-Carrera, S., Falcon, V., Acosta-Rivero, N., Gonzalez, E., de la Rosa, M. C., Menendez, I., and Morales, J. (2001). Assembly of truncated HCV core antigen into virus-like particles in *Escherichia coli*. *Biochem Biophys Res Commun* 281, 962-965.
- Lu, W., and Ou, J. H. (2002). Phosphorylation of hepatitis C virus core protein by protein kinase A and protein kinase C. *Virology* 300, 20-30.
- Lu, W., Strohecker, A., and Ou Jh, J. H. (2001). Post-translational modification of the hepatitis C virus core protein by tissue transglutaminase. *J Biol Chem* 276, 47993-47999.
- Maillard, P., Krawczynski, K., Nitkiewicz, J., Bronnert, C., Sidorkiewicz, M., Gounon, P., Dubuisson, J., Faure, G., Crainic, R., and Budkowska, A. (2001). Nonenveloped nucleocapsids of hepatitis C virus in the serum of infected patients. *J Virol* 75, 8240-8250.
- Majeau N., Gagne V., Boivin A., Bolduc M., Majeau J.A., Ouellet D., and Leclerc D. (2004). The N-terminal half of the core protein of hepatitis C virus is sufficient for nucleocapsid formation. *J Gen Virol*. 85, 971-81.
- Mamiya, N., and Worman, H. J. (1999). Hepatitis C virus core protein binds to a DEAD box RNA helicase. *J Biol Chem* 274, 15751-15756.
- Matsuura, Y., Harada, T., Makimura, M., Sato, M., Aizaki, H., Suzuki, T., and Miyamura, T. (1994). Characterization of HCV structural proteins expressed in various animal cells. *Intervirology* 37, 114-118.
- McLauchlan, J. (2000). Properties of the hepatitis C virus core protein: a structural protein that modulates cellular processes. *Journal Of Viral Hepatitis* 7, 2-14.
- McLauchlan, J., Lemberg, M. K., Hope, G., and Martoglio, B. (2002). Intramembrane proteolysis promotes trafficking of hepatitis C virus core protein to lipid droplets. *Embo J* 21, 3980-3988.
- Miller, K., McArdle, S., Gale, M. J., Jr., Geller, D. A., Tenoever, B., Hiscott, J., Gretch, D. R., and Polyak, S. J. (2004). Effects of the hepatitis C virus core protein on innate cellular defense pathways. *J Interferon Cytokine Res* 24, 391-402.

- Miyamoto, H., Okamoto, H., Sato, K., Tanaka, T., and Mishiro, S. (1992). Extraordinarily low density of hepatitis C virus estimated by sucrose density gradient centrifugation and the polymerase chain reaction. *J Gen Virol*.
- Moriishi, K., Okabayashi, T., Nakai, K., Moriya, K., Koike, K., Murata, S., Chiba, T., Tanaka, K., Suzuki, R., Suzuki, T., et al. (2003). Proteasome activator PA28gamma-dependent nuclear retention and degradation of hepatitis C virus core protein. *J Virol* 77, 10237-10249.
- Moriya, K., Fujie, H., Shintani, Y., Yotsuyanagi, H., Tsutsumi, T., Ishibashi, K., Matsuura, Y., Kimura, S., Miyamura, T., and Koike, K. (1998). The core protein of hepatitis C virus induces hepatocellular carcinoma in transgenic mice. *Nature Medicine* 4, 1065-1067.
- Moriya, K., Fujie, H., Yotsuyanagi, H., Shintani, Y., Tsutsumi, T., Matsuura, Y., Miyamura, T., Kimura, S., and Koike, K. (1997a). Subcellular localization of hepatitis C virus structural proteins in the liver of transgenic mice. *Jpn J Med Sci Biol* 50, 169-177.
- Moriya, K., Nakagawa, K., Santa, T., Shintani, Y., Fujie, H., Miyoshi, H., Tsutsumi, T., Miyazawa, T., Ishibashi, K., Horie, T., et al. (2001a). Oxidative stress in the absence of inflammation in a mouse model for hepatitis C virus-associated hepatocarcinogenesis. *Cancer Res* 61, 4365-4370.
- Moriya, K., Todoroki, T., Tsutsumi, T., Fujie, H., Shintani, Y., Miyoshi, H., Ishibashi, K., Takayama, T., Makuuchi, M., Watanabe, K., et al. (2001b). Increase in the concentration of carbon 18 monounsaturated fatty acids in the liver with hepatitis C: analysis in transgenic mice and humans. *Biochem Biophys Res Commun* 281, 1207-1212.
- Moriya, K., Yotsuyanagi, H., Shintani, Y., Fujie, H., Ishibashi, K., Matsuura, Y., Miyamura, T., and Koike, K. (1997b). Hepatitis C virus core protein induces hepatic steatosis in transgenic mice. *J Gen Virol* 78, 1527-1531.
- Murakami, H., Akbar, S. M., Matsui, H., Horiike, N., and Onji, M. (2004a). Decreased interferon-alpha production and impaired T helper 1 polarization by dendritic cells from patients with chronic hepatitis C. *Clin Exp Immunol* 137, 559-565.
- Murakami, K., Ishii, K., Yoshizaki, S., Aizaki, H., Tanaka, K., Shoji, I., Sata, T., Suzuki, T., Bartenschlager, R., and Miyamura, T. (2004b). Assembly of HCV-like particles in three-dimensional cultures. Paper presented at: 11th international symposium on HCV and Related viruses (Heidelberg, Germany).
- Naganuma, A., Nozaki, A., Tanaka, T., Sugiyama, K., Takagi, H., Mori, M., Shimotohno, K., and Kato, N. (2000). Activation of the interferon-inducible 2'-5' oligoadenylate synthetase gene by hepatitis C virus core protein. *J Virol* 74, 8744-8750.
- Ohkawa, K., Ishida, H., Nakanishi, F., Hosui, A., Ueda, K., Takehara, T., Hori, M., and Hayashi, N. (2004). Hepatitis C virus core functions as a suppressor of cyclin-dependent kinase-activating kinase and impairs cell cycle progression. *J Biol Chem* 279, 11719-11726.



- Okamoto, K., Moriishi, K., Miyamura, T., and Matsuura, Y. (2004). Intramembrane proteolysis and endoplasmic reticulum retention of hepatitis C virus core protein. *J Virol* 78, 6370-6380.
- Okuda, M., Li, K., Beard, M. R., Showalter, L. A., Scholle, F., Lemon, S. M., and Weinman, S. A. (2002). Mitochondrial injury, oxidative stress, and antioxidant gene expression are induced by hepatitis C virus core protein. *Gastroenterology* 122, 366-375.
- Otani, K., M. Korenaga, M.R. Beard, K. Li, T. Qian, L.A. Showalter, A.K. Singh, T. Wang, and Weinman, S. A. (2005). Hepatitis C Virus Core Protein, Cytochrome P450 2E1, and Alcohol Produce Combined Mitochondrial Injury and Cytotoxicity in Hepatoma Cells. *Gastroenterology* 128, 96-107.
- Otsuka, M., Kato, N., Lan, K. H., Yoshida, H., Kato, J., Goto, T., Shiratori, Y., and Omata, M. (2000). Hepatitis C virus core protein enhances p53 function through augmentation of DNA binding affinity and transcriptional ability. *J Biol Chem* 275, 34122-34130.
- Owsianka, A. M., and Patel, A. H. (1999). Hepatitis C virus core protein interacts with a human DEAD box protein DDX3. *Virology* 257, 330-340.
- Pasquinelli, C., J.M. Shoenberger, J. Chung, K. Chang, L.G., G., M. Selby, K. Berger, R. Lesniewski, M. Houghton, and Chisari, F. V. (1997). Hepatitis C virus core and E2 protein expression in transgenic mice. *Hepatology* 25, 719-727.
- Pawlotsky, J. M. (2003). Hepatitis C virus genetic variability: pathogenic and clinical implications. *Clin Liver Dis* 7, 45-66.
- Pellerin, M., Lopez-Aguirre, Y., Penin, F., Dhumeaux, D., and Pawlotsky, J. M. (2004). Hepatitis C virus quasispecies variability modulates nonstructural protein 5A transcriptional activation, pointing to cellular compartmentalization of virus-host interactions. *J Virol* 78, 4617-4627.
- Pietschmann, T., G. Koutsoudakis, S. Kallis, T. Kato, S. Fong, T. Wakita, and Bartenschlager, R. (2004). Chimeric hepatitis C virus infectious in cell culture. Paper presented at: 11th International Symposium on Hepatitis C Virus and Related Viruses (Heidelberg, Germany).
- Pietschmann, T., Lohmann, V., Kaul, A., Krieger, N., Rinck, G., Rutter, G., Strand, D., and Bartenschlager, R. (2002). Persistent and transient replication of full-length hepatitis C virus genomes in cell culture. *J Virol* 76, 4008-4021.
- Polyak, S. J., Khabar, K. S., Paschal, D. M., Ezelle, H. J., Duverlie, G., Barber, G. N., Levy, D. E., Mukaida, N., and Gretch, D. R. (2001). Hepatitis C virus nonstructural 5A protein induces interleukin-8, leading to partial inhibition of the interferon-induced antiviral response. *J Virol* 75, 6095-6106.
- Ray, R. B., Lagging, L. M., Meyer, K., and Ray, R. (1996). Hepatitis C virus core protein cooperates with ras and transforms primary rat embryo fibroblasts to tumorigenic phenotype. *J Virol* 70, 4438-4443.
- Realini, C., Jensen, C. C., Zhang, Z., Johnston, S. C., Knowlton, J. R., Hill, C. P., and Rechsteiner, M. (1997). Characterization of recombinant REGalpha, REGbeta, and REGgamma proteasome activators. *J Biol Chem* 272, 25483-25492.

- Sabile, A., Perlemuter, G., Bono, F., Kohara, K., Demaugre, F., Kohara, M., Matsuura, Y., Miyamura, T., Brechot, C., and Barba, G. (1999). Hepatitis C virus core protein binds to apolipoprotein AII and its secretion is modulated by fibrates. *Hepatology* 30, 1064-1076.
- Sansonno, D., Lauletta, G., and Dammacco, F. (2004). Detection and quantitation of HCV core protein in single hepatocytes by means of laser capture microdissection and enzyme-linked immunosorbent assay. *J Viral Hepat* 11, 27-32.
- Santolini, E., Migliaccio, G., and La, M. N. (1994). Biosynthesis and biochemical properties of the hepatitis C virus core protein. *J Virol* 68, 3631-3641.
- Sarobe, P., Lasarte, J. J., Zabaleta, A., Arribillaga, L., Arina, A., Melero, I., Borrás-Cuesta, F., and Prieto, J. (2003). Hepatitis C virus structural proteins impair dendritic cell maturation and inhibit *in vivo* induction of cellular immune responses. *J Virol* 77, 10862-10871.
- Scheffner, M., Huibregtse, J. M., Vierstra, R. D., and Howley, P. M. (1993). The HPV-16 E6 and E6-AP complex functions as a ubiquitin-protein ligase in the ubiquitination of p53. *Cell* 75, 495-505.
- Schwer, B., Ren, S., Pietschmann, T., Kartenbeck, J., Kaehlcke, K., Bartenschlager, R., Yen, T. S., and Ott, M. (2004). Targeting of hepatitis C virus core protein to mitochondria through a novel C-terminal localization motif. *J Virol* 78, 7958-7968.
- Shi, S. T., Polyak, S. J., Hong, T., Taylor, D. R., Gretch, D. R., and Lai, M. M. C. (2002). Hepatitis C Virus NS5A colocalizes with the Core Protein on Lipid Droplets and Interacts with Apolipoproteins. *Virology* 292, 198-210.
- Shih, C. M., Chen, C. M., Chen, S. Y., and Lee, Y. H. (1995). Modulation of the trans-suppression activity of hepatitis C virus core protein by phosphorylation. *J Virol* 69, 1160-1171.
- Shimizu, Y. K., Feinstone, S. M., Kohara, M., Purcell, R. H., and Yoshikura, H. (1996). Hepatitis C virus: detection of intracellular virus particles by electron microscopy. *Hepatology* 23, 205-209.
- Shimoike, T., Mimori, S., Tani, H., Matsuura, Y., and Miyamura, T. (1999). Interaction of hepatitis C virus core protein with viral sense RNA and suppression of its translation. *J Virol* 73, 9718-9725.
- Shindo, M., Di, B. A. M., Silver, J., Limjoco, T., Hoofnagle, J. H., and Feinstone, S. M. (1994). Detection and quantitation of hepatitis C virus RNA in serum using the polymerase chain reaction and a colorimetric enzymatic detection system. *J Virol Methods* 48, 65-72.
- Shoukry, N. H., Cawthon, A. G., and Walker, C. M. (2004). Cell-Mediated Immunity and the Outcome of Hepatitis C Virus Infection. *Annu Rev Microbiol* 58, 391-424.
- Suzuki, R., Matsuura, Y., Suzuki, T., Ando, A., Chiba, J., Harada, S., Saito, I., and Miyamura, T. (1995). Nuclear localization of the truncated hepatitis C virus core protein with its hydrophobic C terminus deleted. *J Gen Virol* 76, 53-61.



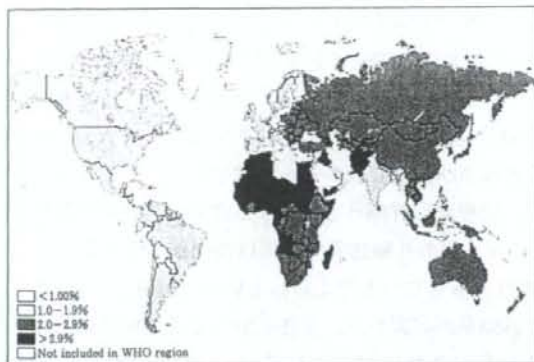
- Suzuki, R., Sakamoto, S., Tsutsumi, T., Rikimaru, A., Tanaka, K., Shimoike, T., Moriishi, K., Iwasaki, T., Mizumoto, K., Matsuura, Y., et al. (2005). Molecular determinants for subcellular localization of hepatitis C virus core protein. *J Virol* 79, 1271-1281.
- Suzuki, R., Suzuki, T., Ishii, K., Matsuura, Y., and Miyamura, T. (1999). Processing and functions of Hepatitis C virus proteins. *Intervirology* 42, 145-152.
- Suzuki, R., Tamura, K., Li, J., Ishii, K., Matsuura, Y., Miyamura, T., and Suzuki, T. (2001). Ubiquitin-mediated degradation of hepatitis C virus core protein is regulated by processing at its carboxyl terminus. *Virology* 280, 301-309.
- Suzuki, T., Y. Matsuura, T. Harada, R. Suzuki, I. Saito, and Miyamura, T. (1996). Molecular basis of subcellular localization of HCV core protein. *Liver* 16, 221-224.
- Tanahashi, N., Yokota, K., Ahn, J. Y., Chung, C. H., Fujiwara, T., Takahashi, E., DeMartino, G. N., Slaughter, C. A., Toyonaga, T., Yamamura, K., et al. (1997). Molecular properties of the proteasome activator PA28 family proteins and gamma-interferon regulation. *Genes Cells* 2, 195-211.
- Taylor, D. R., Shi, S. T., Romano, P. R., Barber, G. N., and Lai, M. M. C. (1999). Inhibition of the interferon-inducible protein kinase PKR by HCV E2 protein. *Science* 285, 107-110.
- Tseng, C. T., and Klimpel, G. R. (2002). Binding of the hepatitis C virus envelope protein E2 to CD81 inhibits natural killer cell functions. *J Exp Med* 195, 43-49.
- Tsubouchi, E., Akbar, S. M., Horiike, N., and Onji, M. (2004a). Infection and dysfunction of circulating blood dendritic cells and their subsets in chronic hepatitis C virus infection. *J Gastroenterol* 39, 754-762.
- Tsubouchi, E., Akbar, S. M., Murakami, H., Horiike, N., and Onji, M. (2004b). Isolation and functional analysis of circulating dendritic cells from hepatitis C virus (HCV) RNA-positive and HCV RNA-negative patients with chronic hepatitis C: role of antiviral therapy. *Clin Exp Immunol* 137, 417-423.
- Tsutsumi, T., Suzuki, T., Shimoike, T., Suzuki, R., Moriya, K., Shintani, Y., Fujie, H., Matsuura, Y., Koike, K., and Miyamura, T. (2002). Interaction of hepatitis C virus core protein with retinoid X receptor alpha modulates its transcriptional activity. *Hepatology* 35, 937-946.
- Wack, A., Soldaini, E., Tseng, C., Nuti, S., Klimpel, G., and Abrignani, S. (2001). Binding of the hepatitis C virus envelope protein E2 to CD81 provides a co-stimulatory signal for human T cells. *Eur J Immunol* 31, 166-175.
- Wakita, T., Kato, T., Date, T., and Miyamoto, M. (2004). Infectious virus production from hepatitis C virus RNA replicating cells. Paper presented at: 11th international symposium on HCV and Related viruses (Heidelberg, Germany).
- Wakita, T., Pietschmann, T., Kato, T., Date, T., Miyamoto, M., Zhao, Z., Murthy, K., Habermann, A., Krausslich, H. G., Mizokami, M., et al. (2005). Production of infectious hepatitis C virus in tissue culture from a cloned viral genome. *Nat Med* 11, 791-796.

- Watashi, K., Hijikata, M., Tagawa, A., Doi, T., Marusawa, H., and Shimotohno, K. (2003). Modulation of retinoid signaling by a cytoplasmic viral protein via sequestration of Sp110b, a potent transcriptional corepressor of retinoic acid receptor, from the nucleus. *Mol Cell Biol* 23, 7498-7509.
- Weihofen, A., Binns, K., Lemberg, M. K., Ashman, K., and Martoglio, B. (2002). Identification of signal peptide peptidase, a presenilin-type aspartic protease. *Science* 296, 2215-2218.
- Wen, F., Abdalla, M. Y., Aloman, C., Xiang, J., Ahmad, I. M., Walewski, J., McCormick, M. L., Brown, K. E., Branch, A. D., Spitz, D. R., et al. (2004). Increased prooxidant production and enhanced susceptibility to glutathione depletion in HepG2 cells co-expressing HCV core protein and CYP2E1. *J Med Virol* 72, 230-240.
- Wertheimer, A. M., Bakke, A., and Rosen, H. R. (2004). Direct enumeration and functional assessment of circulating dendritic cells in patients with liver disease. *Hepatology* 40, 335-345.
- Widell, A., Molnégren, V., Pieksma, F., Calmann, M., Peterson, J., and Lee, S. R. (2002). Detection of hepatitis C core antigen in serum or plasma as a marker of hepatitis C viraemia in the serological window-phase. *Transfus Med* 12, 107-113.
- Yap, S. H., Willems, M., Van den Oord, J., Habets, W., Middeldorp, J. M., Hellings, J. A., Nevens, F., Moshage, H., Desmet, V., and Fevery, J. (1994). Detection of hepatitis C virus antigen by immuno-histochemical staining: a histological marker of hepatitis C virus infection. *J Hepatol* 20, 275-281.
- Yasui, K., Wakita, T., Tsukiyama-Kohara, K., Funahashi, S. I., Ichikawa, M., Kajita, T., Moradpour, D., Wands, J. R., and Kohara, M. (1998). The native form and maturation process of hepatitis C virus core protein. *J Virol* 72, 6048-6055.
- Yoshida, T., Hanada, T., Tokuhisa, T., Kosai, K., Sata, M., Kohara, M., and Yoshimura, A. (2002). Activation of STAT3 by the hepatitis C virus core protein leads to cellular transformation. *J Exp Med* 196, 641-653.
- You, L. R., Chen, C. M., Yeh, T. S., Tsai, T. Y., Mai, R. T., Lin, C. H., and Lee, Y. H. (1999). Hepatitis C virus core protein interacts with cellular putative RNA helicase. *J Virol* 73, 2841-2853.
- Zhong, J., Gastaminza, P., Cheng, G., Kapadia, S., Kato, T., Burton, D. R., Wieland, S. F., Uprichard, S. L., Wakita, T., and Chisari, F. V. (2005). Robust hepatitis C virus infection *in vitro*. *Proc Natl Acad Sci USA* 102, 9294-9299.
- Zhu, N., Ware, C. F., and Lai, M. M. (2001). Hepatitis C virus core protein enhances FADD-mediated apoptosis and suppresses TRADD signaling of tumor necrosis factor receptor. *Virology* 283, 178-187.

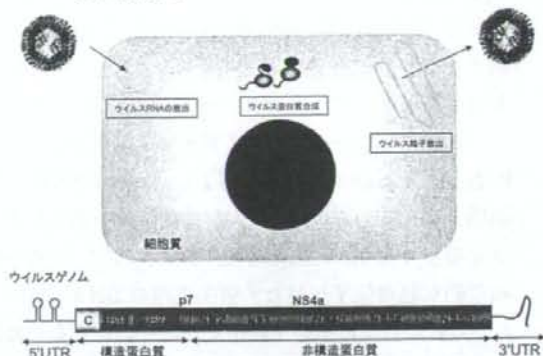


# C型肝炎ウイルスの発揮する腫瘍原性

## I. 世界におけるC型肝炎ウイルス感染者の分布

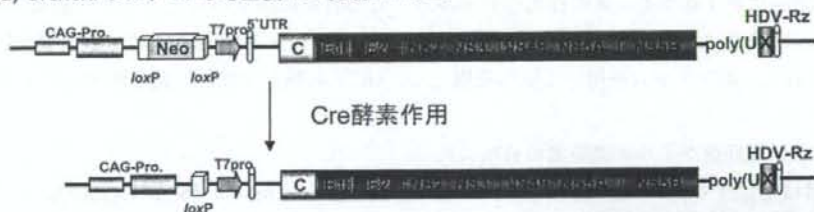


## II. C型肝炎ウイルスの生活環とウイルスゲノム構造

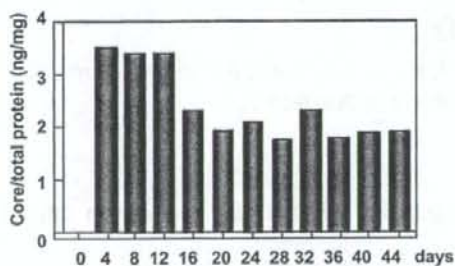


## III. C型肝炎ウイルスによる腫瘍原性亢進

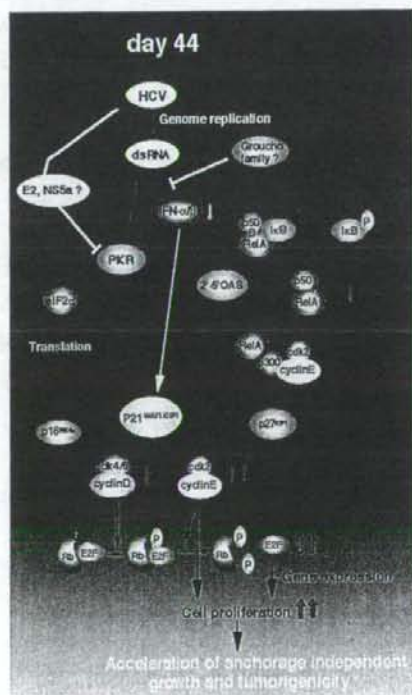
### (a) Cre/loxPシステムによる全長HCV発現システム



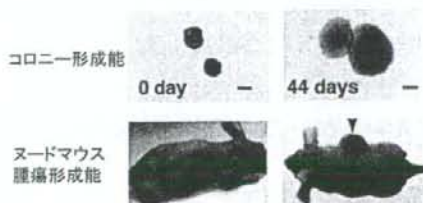
### (b) HCVの持続発現(コア蛋白質)



### (d) HCV発現継代細胞で活性化するシグナル経路



### (c) HCV発現継代細胞の腫瘍原性亢進



肝炎の原因ウイルスは、1970年代までにA型およびB型肝炎ウイルスが発見されたが、同時にこれらのウイルスでは引き起こされない血液媒介性の肝炎、いわゆる非A非B型肝炎の存在が明らかとなった。非A非B型肝炎の病原体の分離には困難を極めたが、唯一の動物実験系であるチンパンジーを用いた感染実験でフラビウイルスに近縁のRNAウイルスである事が予測され、1989年に米国カイロン社のホートン博士らが遺伝子クローニングに成功してC型肝炎ウイルス（HCV）ゲノムの構造と塩基配列が同定された。この遺伝子産物を抗原として世界中の患者血液が調べられた結果、現在全世界では約1億2千万人の抗体陽性者がいると推定される（図Ⅰ）。我が国でも抗体陽性者は200万人前後いると推定されているが、さらに問題となっているのはHCVが持続感染を成立しやすく、肝硬変、肝癌へと進展する点で、日本の肝癌患者さんの多くはHCV抗体陽性である。HCVはRNAウイルスであり細胞質で主に複製増殖すると同時に、典型的な癌遺伝子は持たず宿主細胞のDNAに組み込まれる事もないと考えられている（図Ⅱ）。このようなHCVがいかに細胞を癌化するかを解析するためには感染系が必要だが、現在までのところ肝癌細胞以外には確立されておらず腫瘍原性解析は困難な状況である。これに対して私達は、HCV全長遺伝子をCre/loxPシステムを用いてどんな細胞でも必要な時に発現できるユニットを構築し腫瘍原性解析を可能にしている（図Ⅲ-a）。本ユニットでHCV蛋白質をスイッチング発現すると持続的に発現が維持されるので慢性感染時の肝細胞におけるウイルスの作用のモデル系として容易に解析が可能となる（図Ⅲ-b）。継代44日以降には細胞の腫瘍原性が亢進し（図Ⅲ-c）、標的経路の1つはcdk-Rb-E2F経路である事を明らかにした（図Ⅲ-d）。さらに、本システムを用いて抗体を樹立し、HCVの新たな標的経路を解明している。

#### I. 世界におけるC型肝炎ウイルス感染者の分布

HCV感染者は全世界で約1億2千万人とされ（*Lancet Infect. Dis.*, 5: 558-567, 2005）、エジプトなど一部の地域では高い抗体陽性率を示す。

#### II. C型肝炎ウイルスの生活環とウイルスゲノム構造

肝臓細胞におけるHCVの複製の場は細胞質であり、ウイルス遺伝子はプラス1本鎖RNA（約9.6kb）でウイルス蛋白質は1本のポリプロテインから切断されて機能を持つ。

#### III. C型肝炎ウイルスによる腫瘍原性亢進

(a) Cre/loxPシステムを用いた全長HCV遺伝子発現システム（*J. Biol. Chem.*, 279 (15): 14531-14541, 2004）

HCV遺伝子は、Cre酵素で遺伝子発現後、リボザイム配列で正確に切断される。

(b) (a)のユニットを持つ細胞株でのHCVコア蛋白質発現（ELISAによる定量）

(c) HCV発現継代細胞はコロニー形成能やヌードマウス腫瘍形成能が亢進している。

(d) HCV発現継代細胞で活性化されるシグナル経路

共同研究者：勲東京都医学研究機構 小原道法博士

（熊本大学大学院医学薬学研究部・感染症阻止学寄附講座 小原恭子）



## Q6

最近口内炎がよく出来ます。肝臓が悪いことと関係があるのでしょうか？

**患者背景** 55歳 男性, AST 55 IU/L, ALT 85 IU/L, HCV 抗体 (+)

### A

C型肝炎ウイルスの持続感染者の方には、治りにくい口内炎が出来ることがよくあります。扁平苔癬という名前の病気で、長い間炎症が続き、自然に治ることはほとんどありません。無症状のこともあります。食事の際に口の中の粘膜が“しみる”というのが特徴的な症状です。扁平苔癬は、やがて口腔癌（口の中の癌）に変化することもありますので、必ず専門医での診察が必要となります。なお、C型慢性肝炎で扁平苔癬を合併している方が、インターフェロン治療を受ける場合は、扁平苔癬が悪化しインターフェロン治療が行えない場合があります。したがって、インターフェロン治療を受ける予定の方は、口の中に自覚症状がなくても専門医での診察を受けておくようにしましょう。

## AのPOINT

### 1. 肝外病変とは

肝外病変

C型肝炎ウイルス (HCV) は、肝障害のみならず種々の臓器障害の原因にもなっていることが明らかにされている。HCV は肝臓以外の疾患を引き起こし (肝外病変と呼ぶ)、その代表的な疾患としてクリオグロブリン血症、膜性増殖性糸球体腎炎、晩発性皮膚ポルフィリン症、シェーグレン症候群、悪性リンパ腫、筋炎、心筋障害、扁平苔癬、口腔癌、糖尿病、間質性肺炎、モーレン角膜潰瘍、関節リウマチ、慢性甲状腺炎などがある。HCV キャリア (HCV 持続感染者) が少なくとも1つ以上の肝外病変を有する率は、retrospective study では74% (1,202/1,604人)、prospective study では38% (122/321人) と報告されている。

HCV キャリア

肝外病変についての知識は医療従事者の中でも必ずしも十分に認識されているとはいえ、また引き起こされる病態も多岐にわたるため、臓器別診療体制の傾向が強い医療施設では、各診療科間の十分な連携のもとでHCV キャリアの経過観察が必要である。

### 2. 扁平苔癬とHCV

扁平苔癬

慢性の角化異常を伴う炎症性疾患である扁平苔癬とHCV感染に関する報告は多数認められる (表6-1)。その感染率に差があるのは、各国でのHCV感染率の地域差、人種あるいは研究対象の選択の差等に影響していると考えられる。扁平苔癬の発症には、肝疾患の程度やウイルス側因子 (HCV RNA量やHCV genotype) は関与しない。皮膚あるいは口腔粘膜上皮内において、HCV増殖が証明

表 6-1 扁平苔癬患者における HCV 感染率

国	報告者	報告年	対象人数			LP 患者		
			(口腔扁平苔癬のみ)	(皮膚扁平苔癬のみ)	(口腔皮膚扁平苔癬のみ)	HCV 抗体陽性率	HCV RNA 陽性率 (%)	コントロール HCV 抗体陽性率 (%)
日本	Nagao et al.	1995	45 (45/0/0)			62	60	コントロールなし
	Tanei et al.	1995	45 (28/8/9)			37.8	未施行	6.7% (3 of 45)
イタリア	Divano et al.	1992	46 (0/46/0)			32.6	未施行	コントロールなし
	Rebora et al.	1992	50 (50/0/0) (group 1 : without CLD)			4	未施行	コントロールなし
		29 (29/0/0) (group 2 : with CLD)		65	未施行	コントロールなし		
		46 (46/0/0) (group 3 : with or without CLD)		24	未施行	コントロールなし		
	Gandolfo et al.	1994	105 (105/0/0)			9.5	未施行	コントロールなし
	Carrozo et al.	1996	70 (70/0/0)			27.1	21.4	4.3% (3 of 70)
Mignogna et al.	1998	263 (263/0/0)			28.8	未施行	3% (3 of 100)	
	2002	600 (600/0/0)			27.5	未施行	コントロールなし	
スペイン	Bagan et al.	1994	187 (187/0/0)			15	未施行	コントロールなし
	Sanchez-Perez et al.	1996	78 (22/22/34)			20	16.7	2.4% (2 of 82)
	Bagan et al.	1998	100 (100/0/0)			23	未施行	5% (5 of 100)
フランス	Cribier et al.	1994	52 (4/48/0)			3.8	未施行	2.6% (3 of 112)
	Dupin et al.	1997	102 (102/0/0)			4.9	未施行	4.5% (14 of 306)
	Dupond et al.	1998	28 (28/0/0)			28.6	17.9	コントロールなし
イギリス	Ingafou et al.	1998	55 (55/0/0)			0	未施行	0% (0 of 110)
	Tucker et al.	1999	45 (13/32/0)			0	0	3.1% (1 of 32)
スコットランド	Roy et al.	2000	6 (6/0/0)			0	0	コントロールなし
USA	Bellman et al.	1995	30 (0/30/0)			23	16.7	4.8% (2 of 41)
	Chuang et al.	1999	22 (0/22/0)			55	未施行	25% (10 of 40) (control 1)
	Beairst et al.	2001	24 (0/24/0)			17	未施行	0.17% (255 of 149,756) (control 2)
	Chainal-Wu et al.	2001	31 (31/0/0)			45	未施行	5% (1 of 20)
	Eisen	2002	195 (195/0/0)			0	未施行	コントロールなし
	ドイツ	Imhof et al.	1997	84 (22/62/0)			16	14
Grote et al.	1998	24 (24/0/0)			4.2	未施行	コントロールなし	
オランダ	van der Meij et al.	2000	55 (55/0/0)			0	未施行	コントロールなし
トルコ	Ilter et al.	1998	75 (0/75/0)			0	未施行	0% (0 of 75)
	Kirtak et al.	2000	73 (27/46/0)			6.8	未施行	1.36% (1 of 73)
	Erkek et al.	2001	54 (0/54/0)			12.9	9.3	3.7% (2 of 54)
ネパール	Garg et al.	2002	64 (14/35/15)			0	未施行	0% (不明)
ナイジェリア	Daramola et al.	2002	57 (0/55/2)*			15.8	未施行	25% (6 of 24) (control A) 0% (0 of 24) (control B)
ブラジル	Figueiredo et al.	2002	68 (63/0/5)			8.8	未施行	0.6% (6 of 898)

CLD: 慢性肝疾患, control 1: 乾癬, control 2: ボランティア供血者, control A: 扁平苔癬以外の皮膚疾患, control B: 正常, \*著者が Dr. Daramola に関わった

(Nagao et al. J Gastroenterol Hepatol 2004; 19: 1101-1113)

され (*in situ* hybridization 法あるいは RT-PCR 法), HCV 特異的な T 細胞の関与が証明されている。

HCV 関連の肝外病変に対する治療として、インターフェロンやリバビリンによる治療効果が確認されている。現在までに、効果の認められた疾患として、膜性増殖性糸球体腎炎、クリオグロブリン血症、悪性リンパ腫、晩発性皮膚ポルフィリン症、扁平苔癬、モーレン角膜潰瘍などがある。難治性の HCV 関連の膜性増殖性糸球体腎炎やクリオグロブリン血症に対しては、インターフェロンならびにリバビリン併用療法が有効な治療法であると考えられ、期待される治療であるともいえる。しかしながら、必ずしも肝外病変に対するインターフェロン治療が有効な治療とはいえ、逆に肝外症状が悪化し、インターフェロン治療を中止せざるをえないケースも存在することやインターフェロンを中止すると再発することが問題視されている。特にびらん型の口腔扁平苔癬を合併する C 型慢性肝炎例に、

IFN 治療



● 症 状

インターフェロン療法を行うと皮膚や口腔病変が増悪する傾向があり、インターフェロン投与はきわめて慎重に行うことが肝要である。したがって、肝外病変に対するインターフェロン治療には注意が必要である。C型肝炎に対するインターフェロン治療前には、全身のスクリーニングと専門医への紹介を怠ってはならない。

口腔外科専門医

口腔粘膜病変に対する診断は、口腔外科専門医に紹介することを推奨する。ただし、口腔外科専門医名や口腔外科専門医施設は公表されていないのが現状であるが、2005年7月7日付けの口腔外科専門医の指定研修機関は [http://www.jsoms.or.jp/2\\_03\\_3.html](http://www.jsoms.or.jp/2_03_3.html) で閲覧可能である。なお、口腔外科専門医であってもHCVと扁平苔癬について精通しているとは限らない。

〈佐田 通夫〉

# Q55

キャリアの血液に触れたのですが、どのように対処したらよいのでしょうか？

患者背景 キャリアの家族

## A

B型肝炎ウイルスに汚染した場合の積極的な感染予防策として、高力価 HBs 抗体含有免疫グロブリン (HBIG) と B型肝炎ウイルスのワクチンがあります。しかし現在、C型肝炎ウイルス感染予防のためのワクチンはありません。C型肝炎ウイルスを含む血液に触れた方は、まず接触直後に C型肝炎ウイルス抗体検査と肝機能検査を受け、その後 1 週間後、2 週間後の 2 回を目安として C型肝炎ウイルス RNA 定性検査を受けながら、約 6 か月間経過をみます。万一感染したことがわかったときには、インターフェロンを投与することによりウイルス駆除が可能であり、慢性化を防止できることがわかっています。

なお、肝炎ウイルスの感染を避けるためには、感染している人の血液になるべく触れないことが大切です。①歯ブラシ、カミソリなどを他人と共用しない、②ほかの人の血液に触るときは、ゴム手袋を着ける、③注射器や注射針を共用して、覚醒剤や麻薬等の注射をしない、④口うつしで乳幼児に食べ物を与えない、⑤入れ墨やピアスをするときは、清潔な器具であることを必ず確かめる、⑥よく知らない相手との性行為にはコンドームを使用するといったことを守りましょう。また、現在日本で行われている医療行為（歯科医療を含む）で、肝炎ウイルスに感染する可能性はまれと考えられています。また、理髪店においてもカミソリを共用しなければ、感染することはまれです。しかし、長期間にわたって血液透析を受けている方では、施設内での感染が発生しており、医療機関における感染予防が重要な問題となっています。

## AのPOINT

### 1. 感染予防

#### 感染予防

B型肝炎ウイルス (HBV) も C型肝炎ウイルス (HCV) も日常生活（食事や入浴）で感染することはないが、血液による汚染を介して感染する。HBV の感染予防対策としては、2つの方法が存在する。

#### HBIG

#### a. 高力価 HBs 抗体含有免疫グロブリン (HBIG) の投与

HBV に対して免疫を持たない人が、HBV を含む血液に汚染した場合には（汚染血液による針刺し事故、汚染血液による縫合針による誤刺、垂直感染予防時）、48 時間以内に HBIG を投与し、必要に応じて HBs ワクチン接種を併用する。

#### HBs ワクチン

#### b. HBs ワクチンの接種

ワクチンの接種対象は、HBV キャリア妊婦からの出生児、血液に触れる機会の多い HBV キャリアの家族や医療従事者などである。HBs ワクチンによる HBs 抗体獲得率は約 90% 以上に及ぶ。