

measurements for glaucoma detection using optical coherence tomography. *Am J Ophthalmol* 139:44-55.

Miyamoto-Sato E, Ishizaka M, Horisawa K, Tateyama S, Takashima H, Fuse S, Sue K, Hirai N, Masuoka K, Yanagawa H (2005) Cell-free cotranslation and selection using in vitro virus for high-throughput analysis of protein-protein interactions and complexes. *Genome Res* 15:710-717.

Monemi S, Spaeth G, DaSilva A, Popinchalk S, Ilitchev E, Liebmann J, Ritch R, Heon E, Crick RP, Child A, Sarfarazi M (2005) Identification of a novel adult-onset primary open-angle glaucoma (POAG) gene on 5q22.1. *Hum Mol Genet* 14:725-733.

Moreland RJ, Dresser ME, Rodgers JS, Roe BA, Conaway JW, Conaway RC, Hanas JS (2000) Identification of a transcription factor IIIA-interacting protein. *Nucleic Acids Res* 28:1986-1993.

Moritz OL, Tam BM, Hurd LL, Peranen J, Deretic D, Papermaster DS (2001) Mutant *rab8* impairs docking and fusion of rhodopsin-bearing post-Golgi membranes and causes cell death of transgenic *Xenopus* rods. *Mol Biol Cell* 12:2341-2351.

Nachury MV, Loktev AV, Zhang Q, Westlake CJ, Peränen J, Merdes A, Slusarski DC, Scheller RH, Bazan JF, Sheffield VC, Jackson PK (2007) A core complex of BBS proteins cooperates with the GTPase Rab8 to promote ciliary membrane biogenesis. *Cell* 129:1201-13.

Park BC, Shen X, Samaraweera M, Yue BY (2006) Studies of optineurin, a glaucoma gene: Golgi fragmentation and cell death from overexpression of wild-type and mutant optineurin in two ocular cell types. *Am J Pathol* 169:1976-89.

Quigley HA (1996) Number of people with glaucoma worldwide. *Br J Ophthalmol* 80:389-393.

Quigley HA, Broman AT (2006) The number of people with glaucoma worldwide in 2010 and 2020. *Br J Ophthalmol* 90:262-267.

Rangaswamy NV, Shirato S, Kaneko M, Digby BI, Robson JG, Frishman LJ (2007) Effects of Spectral Characteristics of Ganzfeld Stimuli on the Photopic Negative Response (PhNR) of the ERG. *Invest Ophthalmol Vis Sci* 48:4818-28.

Rezaie T, Child A, Hitchings R, Brice G, Miller L, Coca-Prados M, Heon E, Krupin T, Ritch R, Kreutzer D, Crick RP, Sarfarazi M (2002) Adult-onset primary open-angle glaucoma caused by mutations in optineurin. *Science* 295:1077-1079.

Sahlender DA, Roberts RC, Arden SD, Spudich G, Taylor MJ, Luzio JP, Kendrick-Jones J, Buss F (2005) Optineurin links myosin VI to the Golgi complex and is involved in Golgi organization and exocytosis. *J Cell Biol* 169:285-95.

Sarfarazi M, Rezaie T (2003) Optineurin in primary open angle glaucoma. *Ophthalmol Clin North Am* 16:529-541.

Sato T, Mushiaki S, Kato Y, Sato K, Sato M, Takeda N, Ozono K, Miki K, Kubo Y, Tsuji A, Harada R, Harada A (2007) The Rab8 GTPase regulates apical protein localization in intestinal cells. *Nature* 448:366-9.

Senatorov V, Malyukova I, Fariss R, Wawrousek EF, Swaminathan S, Sharan SK, Tomarev S (2006) Expression of mutated mouse myocilin induces open-angle glaucoma in transgenic mice. *J Neurosci* 26:11903-14.

Stoilov I, Akarsu AN, Sarfarazi M (1997) Identification of three different truncating mutations in cytochrome P4501B1 (CYP1B1) as the principal cause of primary congenital glaucoma (Buphthalmos) in families linked to the GLC3A locus on chromosome 2p21. *Hum Mol Genet* 6:641-7.

Stone EM, Fingert JH, Alward WL, Nguyen TD, Polansky JR, Sunden SL, Nishimura D, Clark AF, Nystuen A, Nichols BE, Mackey DA, Ritch R, Kalenak JW, Craven ER, Sheffield VC (1997) Identification of a gene that causes primary open angle glaucoma. *Science* 275:668-70.

Tan O, Li G, Lu AT, Varma R, Huang D; Advanced Imaging for Glaucoma Study Group (2008) Mapping of macular substructures with optical coherence tomography for glaucoma diagnosis. *Ophthalmology* 115:949-56.

Wang L, Cioffi GA, Cull G, Dong J, Fortune B (2002) Immunohistologic evidence for retinal glial cell changes in human glaucoma. *Invest Ophthalmol Vis Sci* 43:1088-94.

Wässle H, Grünert U, Röhrenbeck J, Boycott BB (1989) Cortical magnification factor and the ganglion cell density of the primate retina. *Nature* 341:643-6.

Watanabe T, Raff MC (1988) Retinal astrocytes are immigrants from the optic nerve. *Nature* 332:834-7.

Willoughby CE, Chan LL, Herd S, Billingsley G, Noordeh N, Levin AV, Buys Y, Trope G, Sarfarazi M, Heon E (2004) Defining the pathogenicity of optineurin in juvenile open-angle glaucoma. *Invest Ophthalmol Vis Sci* 45:3122-3130.

Zerial M, McBride H (2001) Rab proteins as membrane organizers. *Nat Rev Mol Cell Biol* 2:107-117.

## FIGURE LEGENDS

### Figure 1.

*Development of transgenic mouse over expressing OPTN.*

**A**, Schematic diagram of the OPTN constructs used in this study. The green region corresponds to the OPTN protein. Positions of mutations and deletions are shown in red. The HA tag is marked by yellow color. The CAGGS region corresponds to the

chicken beta-actin promoter. **B**, Fundus photographs of normal, Wt and E50K Tg mouse eyes at 16 month. Curvature of the retinal vessels indicates the excavation of the area including the optic disc in E50K Tg mouse eye. **C**, Total expression of endogenous and mutant OPTN (red) in the retina of normal and E50K Tg mice at 16 month. Anti-OPTN antibody and Anti-HA antibody were used to detect endogenous and mutant OPTN respectively. Scale bar, 50  $\mu\text{m}$ .

## Figure 2.

*RGC loss and thinning of the retina thickness in the peripheral retina.*

**A**, staining of retina sections of 16 month old normal and Tg mice. Scale bar, 200  $\mu\text{m}$  (upper panel), 50  $\mu\text{m}$  (lower panel). **B**, Quantification of the RGC number and retina thickness of 16 month old normal and Tg mice (n=6). Only 50K Tg mice at 16 months showed significant RGC loss and thinning of retina (\*\* $p < 0.01$ ). **C**, Quantification of the RGC number and retina thickness during development of E50K Tg mice (n=6). Tg mice showed statistically significant RGC loss and thinning of the retina starting from 12 months of age. **D**, Impaired ERG in E50K Tg mice. The amplitude of the PhNR by E50K Tg mice decreased and removed the negative wave to the transient b-wave (arrow), suggesting RGC loss and other abnormality.

**Figure 3.**

*Histopathology of retina and optic nerve of 16 month old Wt and E50K Tg mouse eyes.*

**A**, Immunolabeling of the retina sections with calretinin, a specific marker for RGCs and amacrine cells. Synapse disruption was observed in the E50K Tg mouse retina (arrow). Scale bar, 20  $\mu\text{m}$ . **B**, Hematoxylin-eosin staining and immunostaining with antibodies against tubulin  $\beta$  III isoform in the optic nerve region. Significant thinning of the nerve fiber layer and the excavation of optic disc (arrow) was observed in E50K Tg mice. Scale bar, 100  $\mu\text{m}$ .

**Figure 4.**

*RGC degeneration in E50K Tg mice.*

**A**, Immunostaining of normal, Wt, and E50K Tg mouse whole retinas with antibodies against SMI32, a specific marker of large type RGCs. Scale bar, 500  $\mu\text{m}$ . White box indicate the location of photographs in lane B. **B**, Thinning of NFL, RGC loss. Scale bar, 50  $\mu\text{m}$ . **C**, RGC axon abnormality (arrows) was also observed. Scale bar, 50  $\mu\text{m}$ .

**Figure 5.**

*Glial cells death in E50K Tg mice.*

**A, B**, Flat mount retina of *E50K* Tg mice was double immunostained with SMI32 (red) and active caspase-3 (green) antibodies. Apoptotic cells were observed only in the whole mount retina of *E50K* Tg mice. Scale bar, 100  $\mu$ m. **C-E**, Flat mount retina of *E50K* Tg mice was also double immunostained with active caspase-3 (green) and GFAP (red) antibodies showing apoptosis of astrocytes. Scale bar, 50  $\mu$ m. **F**, Apoptotic astrocytes (green) in peripheral retina. **G**, Apoptotic astrocytes (green) in central retina. Scale bar, 100  $\mu$ m.

#### **Figure 6**

*IOP measurements for Wt and E50K Tg mice.*

**A**, impact-rebound tonometer and **B**, optical interferometry tonometer. Both methods gave normal IOP of 15 +/-1 mmHg for Tg mice at all ages examined (n=6).

#### **Figure 7**

*Disruption of OPTN-Rab8 interaction by E50K mutation.*

**A**, A diagram of cDNA constructs used in experiments to study protein-protein interaction. **B**, The protein-protein interaction of OPTN Wt and E50K with Rab8 Wt, T22N inactive form, and Q67L active form as measured in RGC-5 cells. Interaction of OPTN Wt and Q67L active form of Rab8 increased two and five times over Rab8 Wt

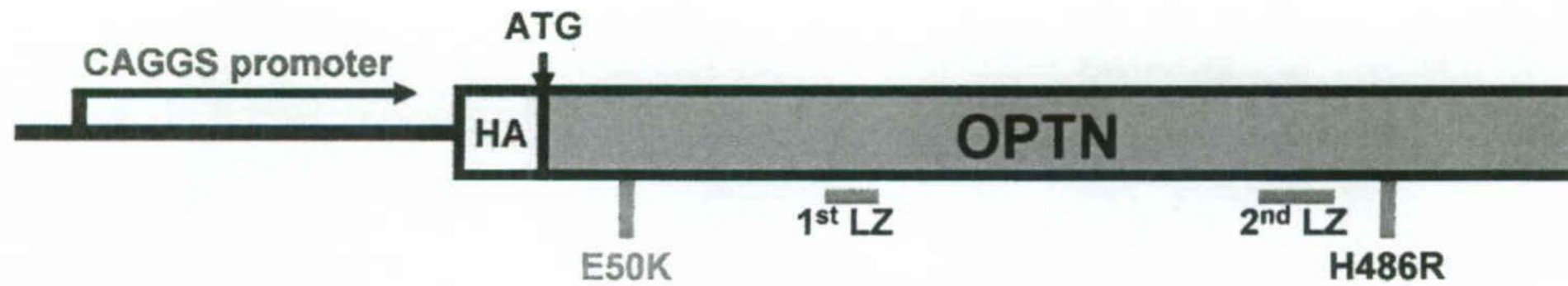


or T22N inactive form of Rab8 protein, respectively (\*\* $p < 0.01$ ). OPTN E50K did not show any interaction with any construct including the active form of Rab8 (n=6). **C**, Protein interaction of Wt or mutant OPTN E50K with Rab8 was measured by QCM technique. A sharp drop of QCM frequency was observed when control OPTN Wt was injected as guest sample, confirming the previous reports of OPTN-Rab8 interaction. Mutant OPTN E50K showed no interaction with Rab8. **D**, GST Pull-down assay to determine OPTN E50K-Rab8 interaction. The fusion protein GST-Rab8 was used for *in vitro* binding assay with purified OPTN Wt and OPTN E50K protein. For negative control OPTN Wt and OPTN E50K were reacted with GST alone (lane 1 & 2). *In vitro* translated OPTN Wt and OPTN E50K were analyzed by SDS-PAGE to show the protein size (lane 6 & 7). OPTN E50K showed significant loss of interaction with Rab8 compare with OPTN Wt (lane 3 & 4, graph). The illustration shows a diagram of the interaction experiment. **E**, Immunostaining the OPTN-Rab8 complex (green) with Golgi marker GM130 (red), indicate that these interactions take place adjacent to the Golgi network.

**Figure 8.**

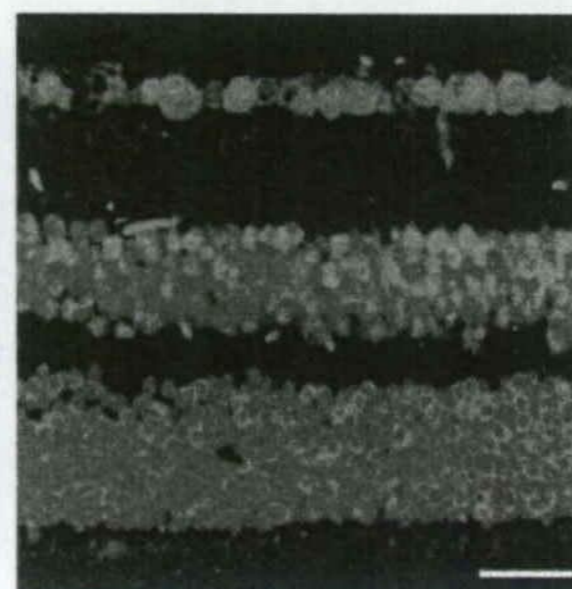
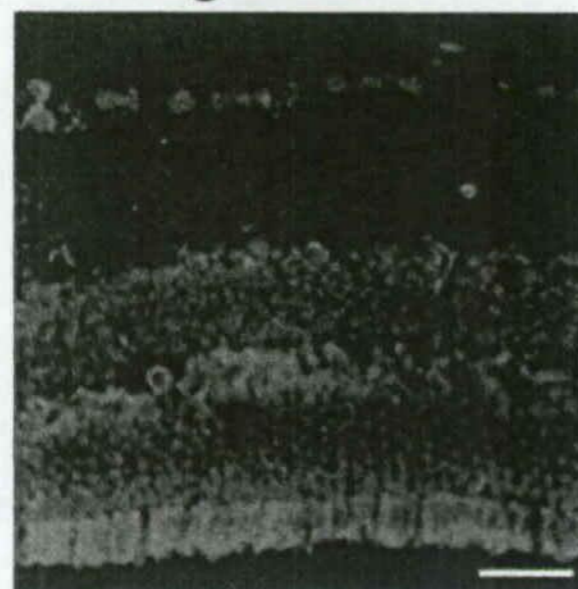
*NTG patient with OPTN E50K mutation.*

**A**, A pedigree of a NTG family with OPTN E50K mutation. The patients were diagnosed as NTG with glaucomatous optic neuropathy and visual field loss. **B**, Optical coherence tomography (Cirrus HD-OCT, Carl Zeiss Meditec, Dublin , CA ) and visual field test ( Humphrey Field Analyzer, Carl Zeiss Medic, Dublin, CA ) were shown on patient 2 and unrelated normal control. The retinal NFL thinning and glaucomatous visual field loss were observed in patients.

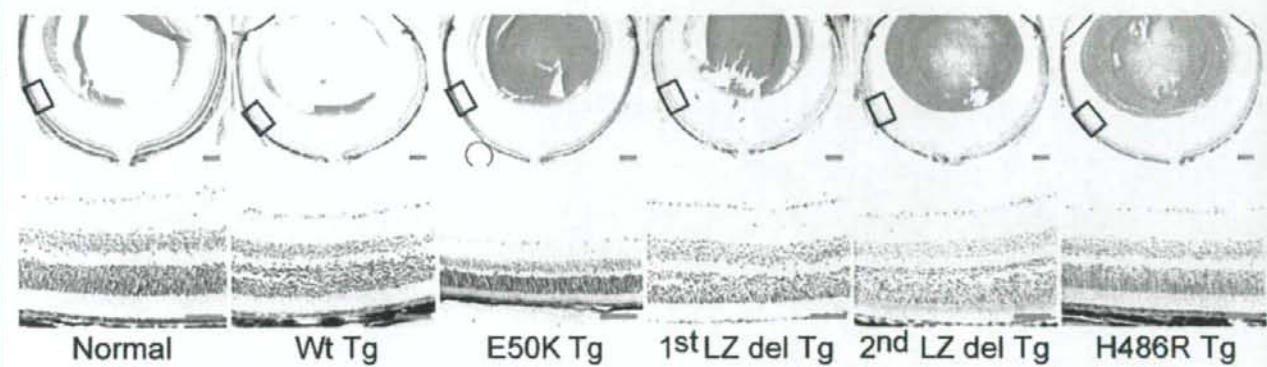
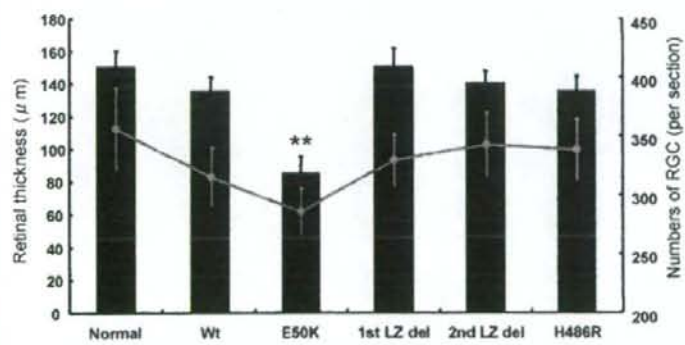
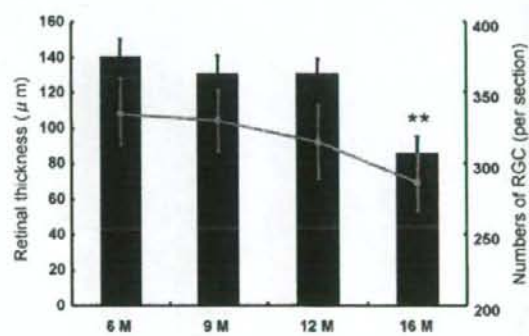
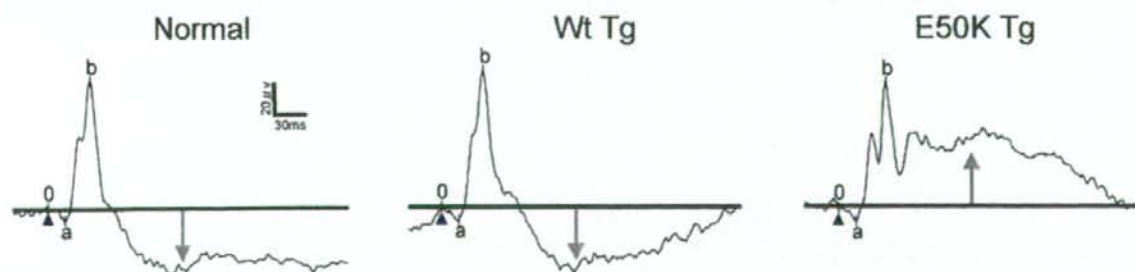
**A****B****C**

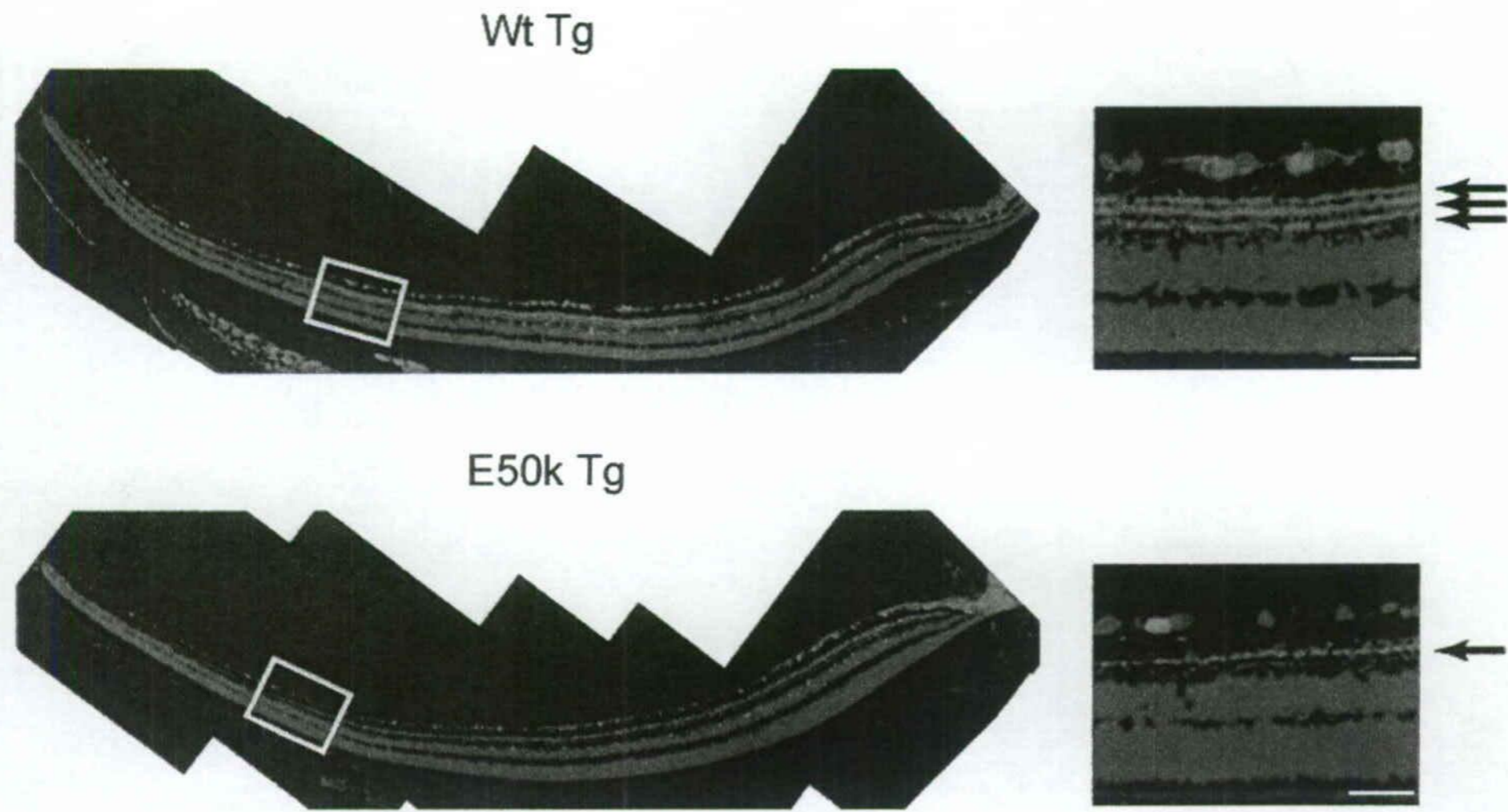
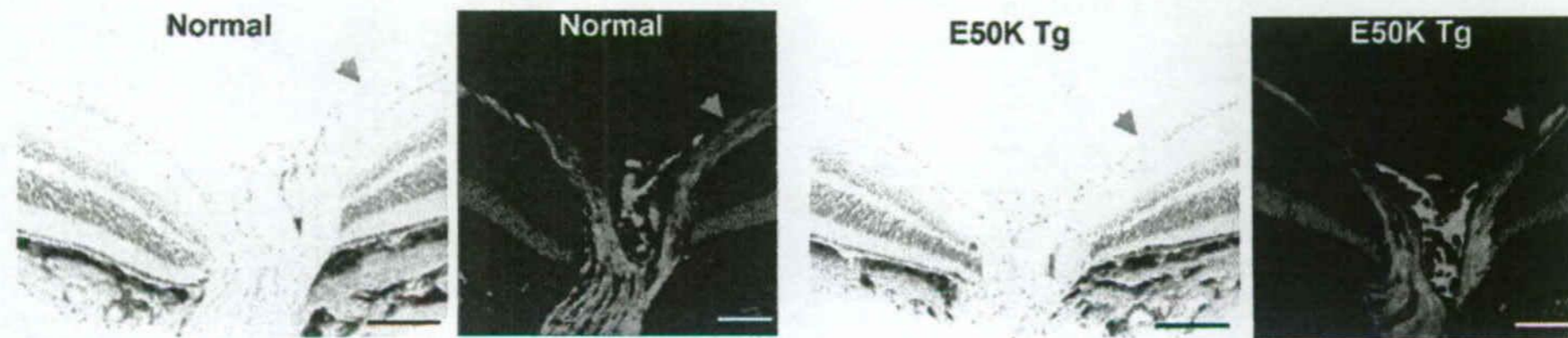
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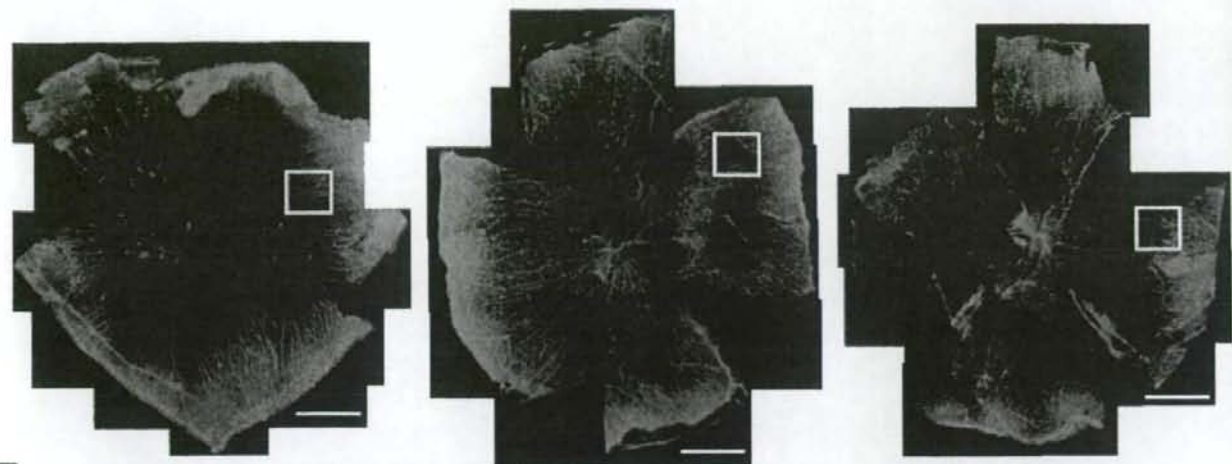
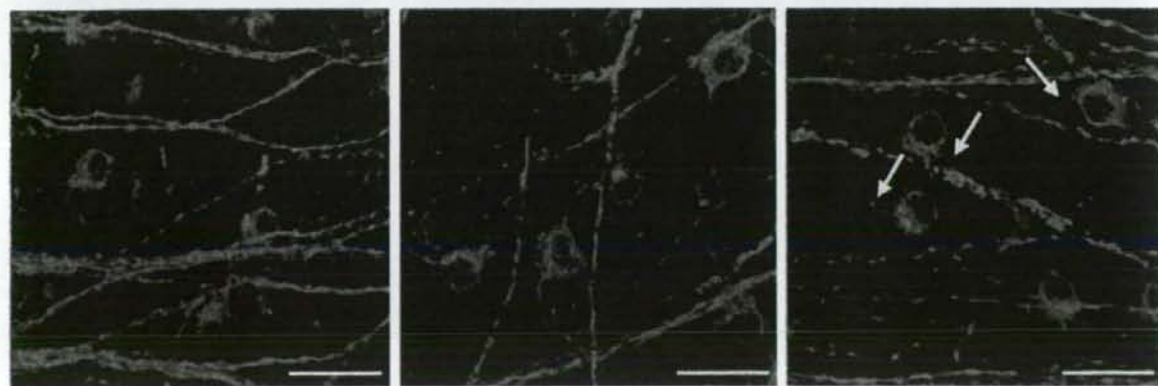
Mutant OPTN



Ganglion cell layer  
 Inner plexiform layer  
 Inner nuclear layer  
 Outer plexiform layer  
 Outer nuclear layer  
 Segments of rods and cones

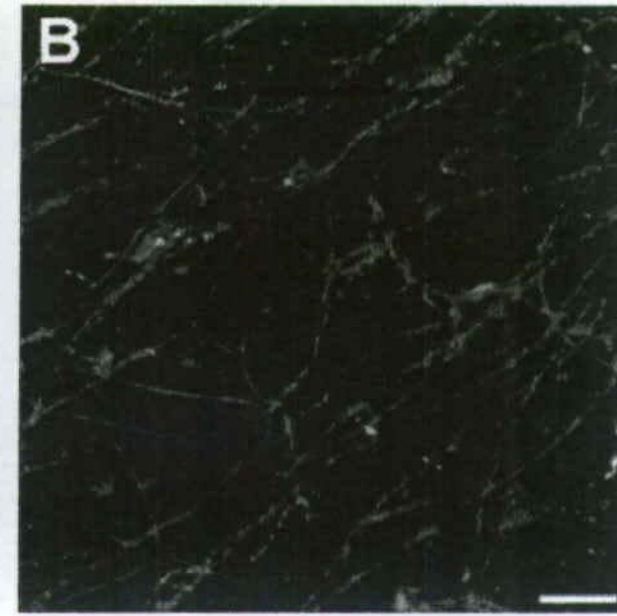
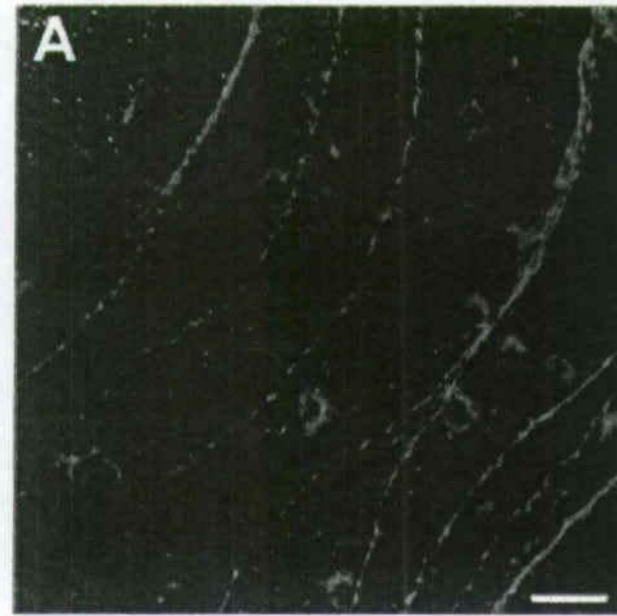
**A****B****C****D**

**A****B**

**A****normal****Wt Tg****E50K Tg****B****C**

Wt Tg

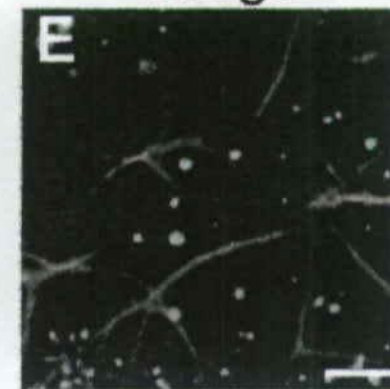
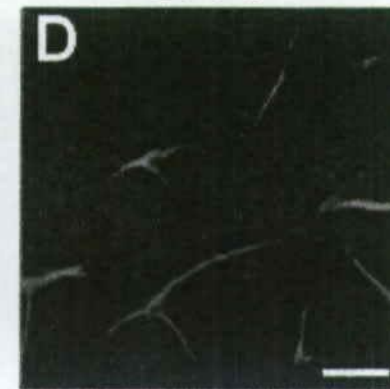
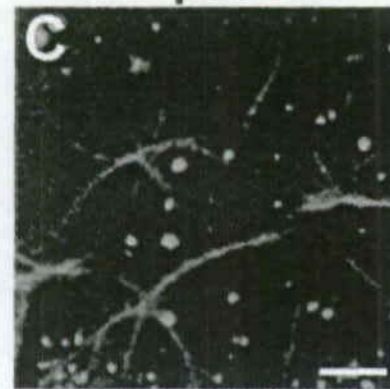
E50K Tg



Caspase-3

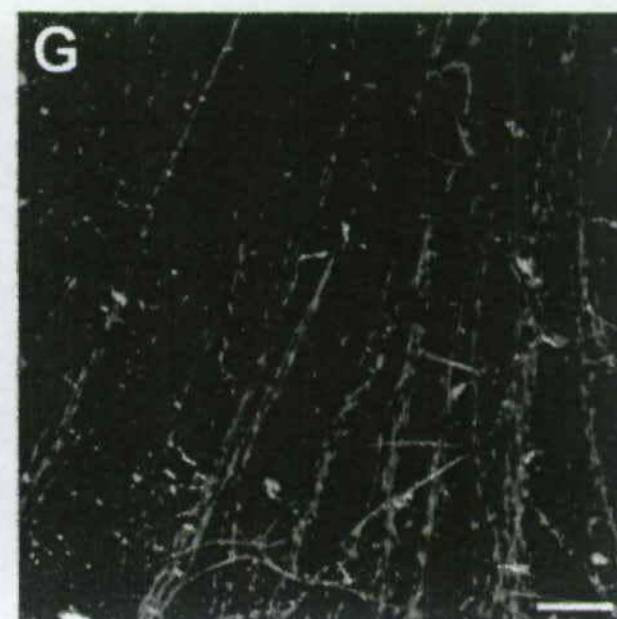
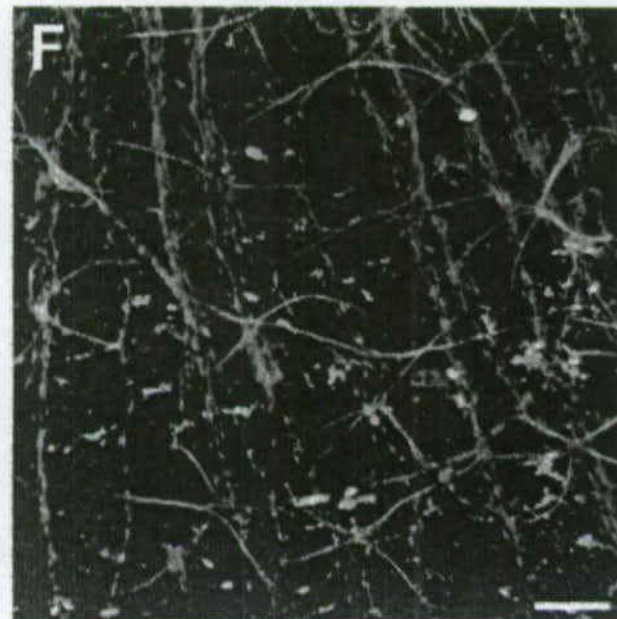
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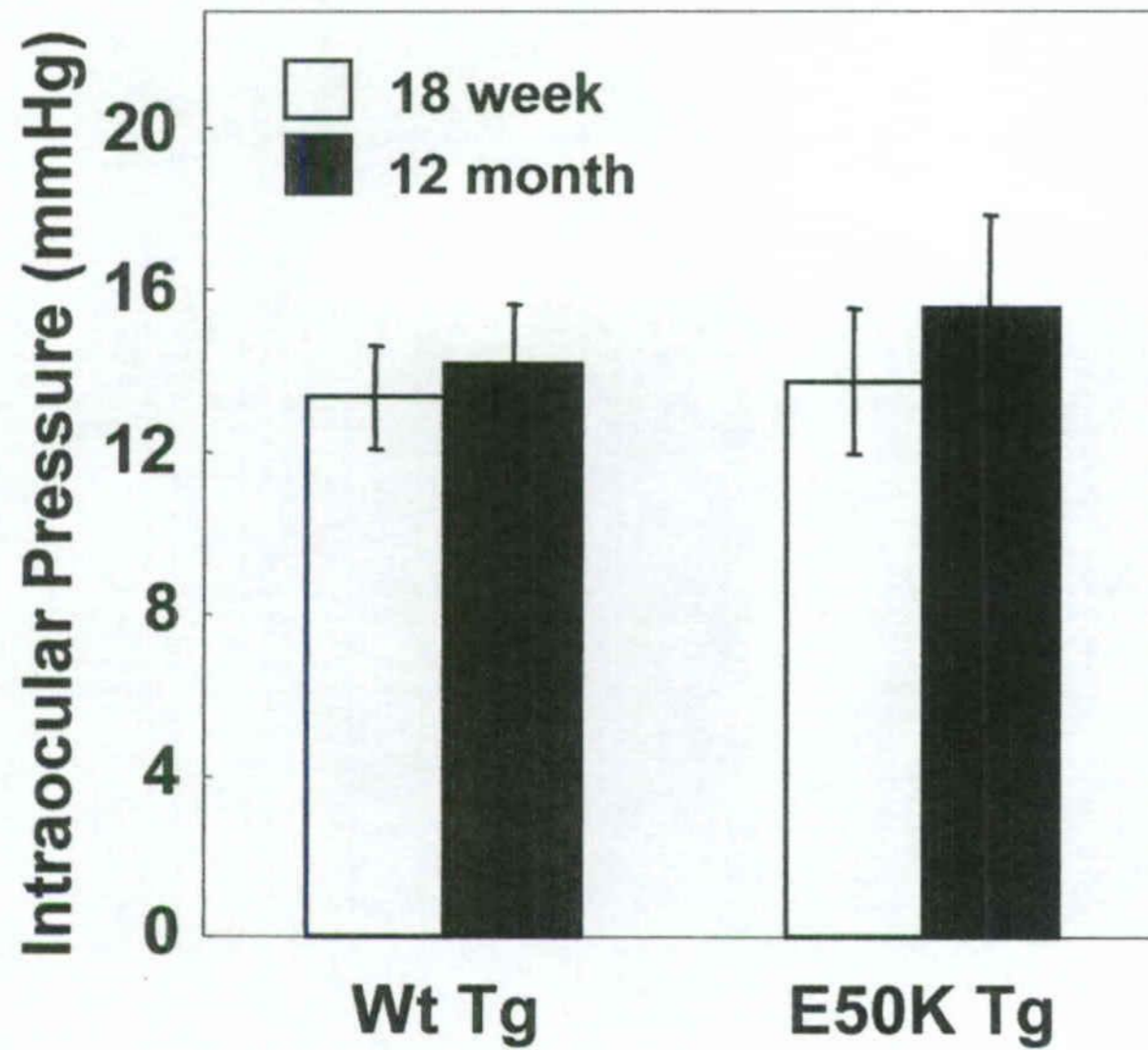
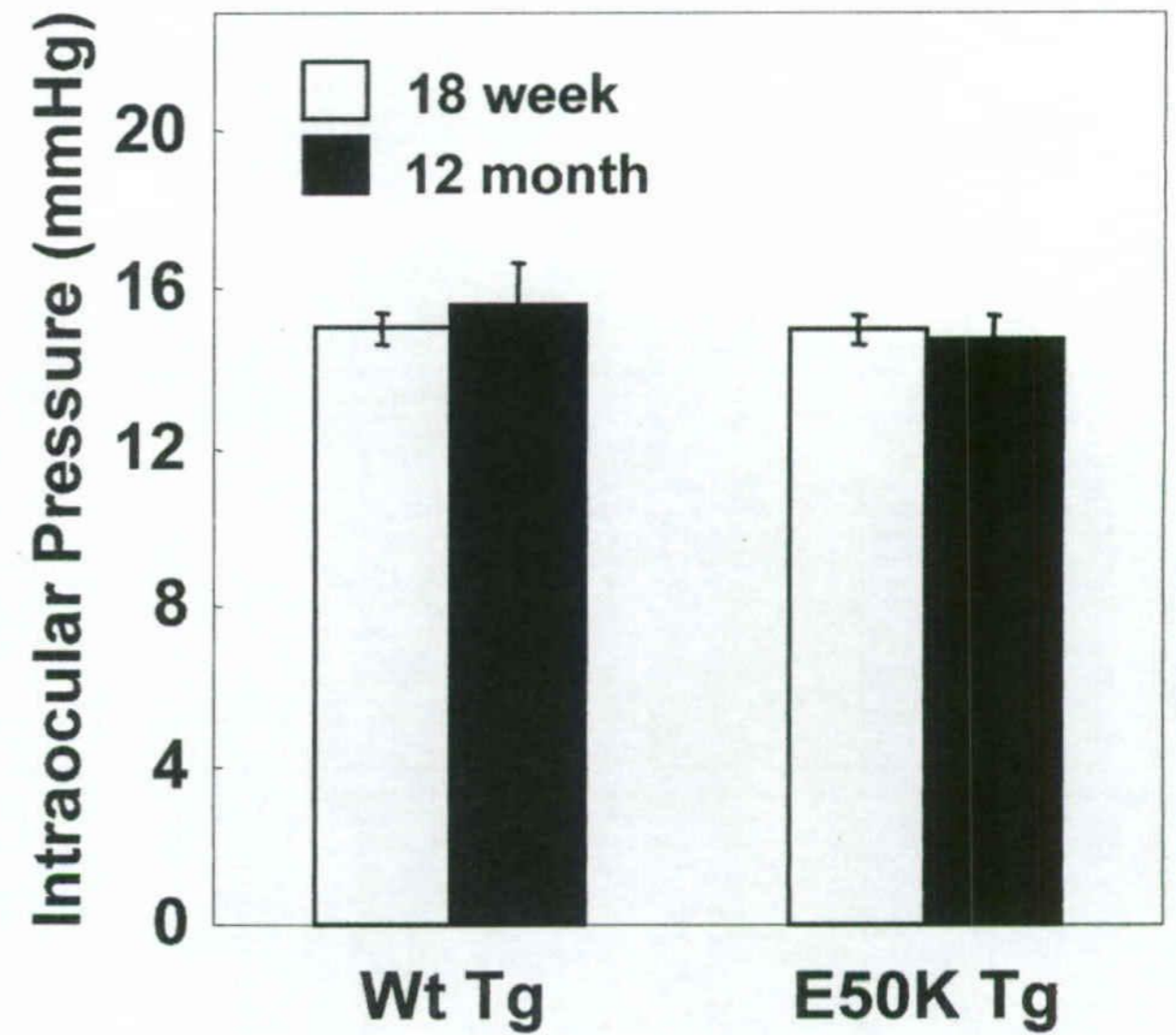
Merge



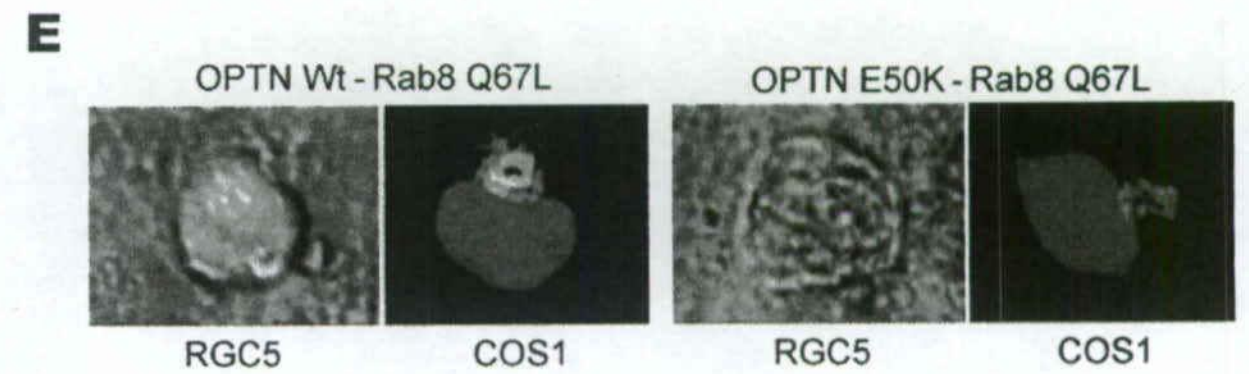
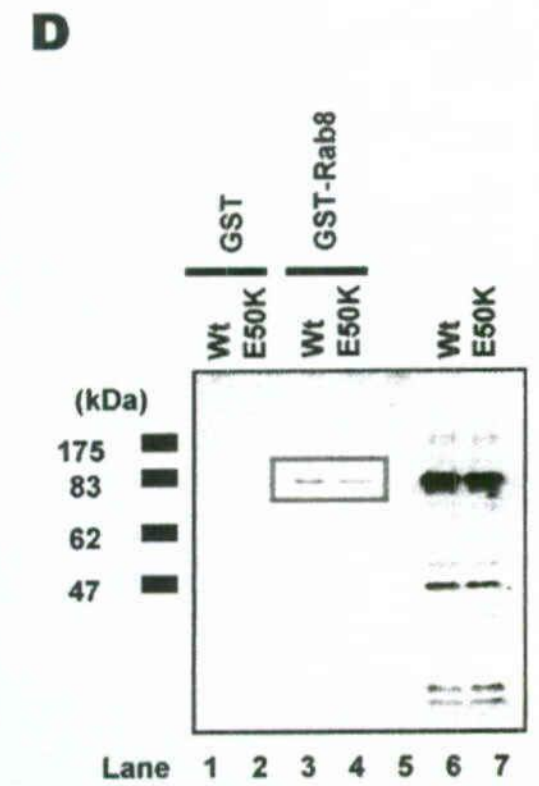
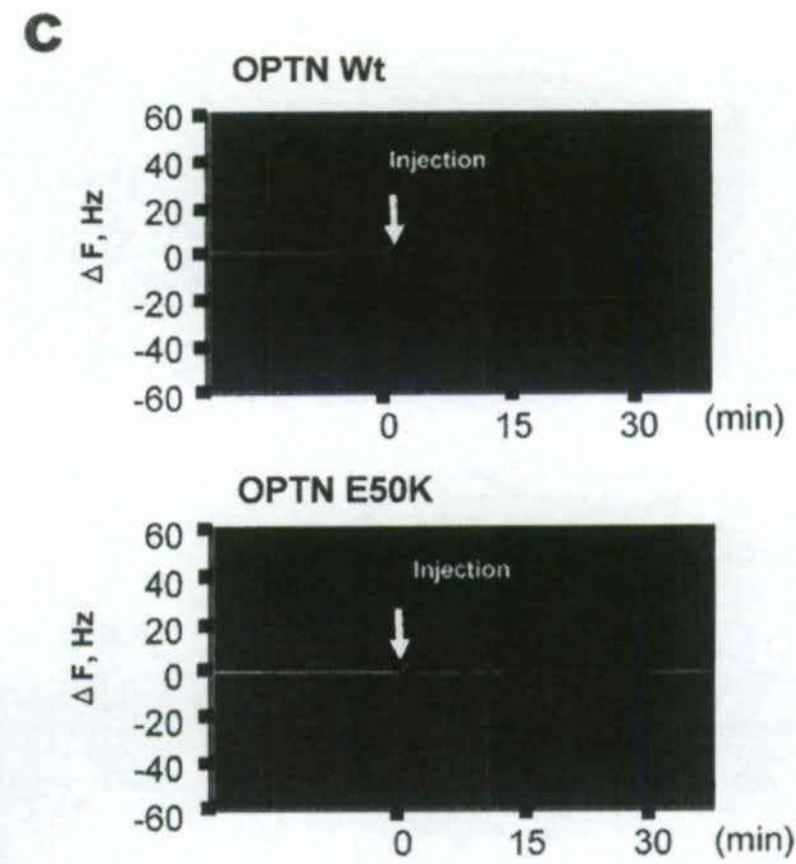
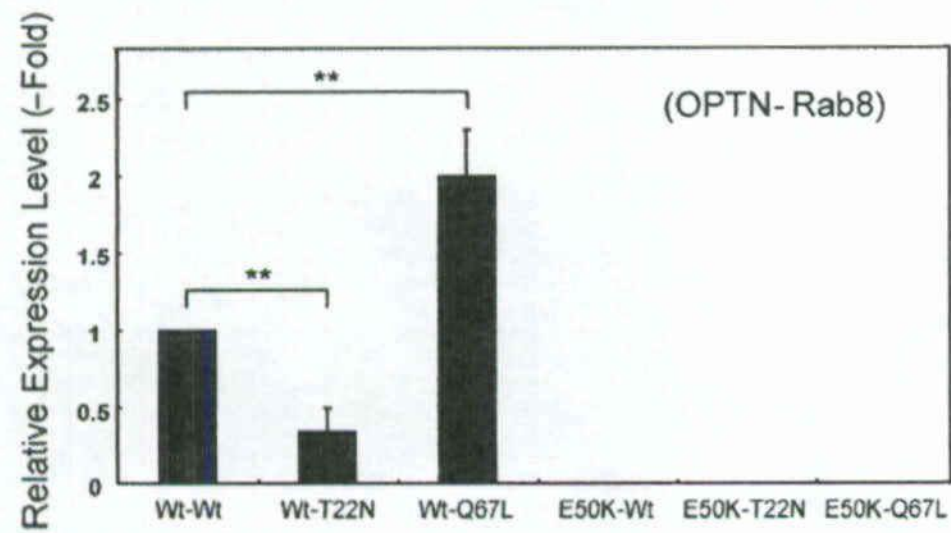
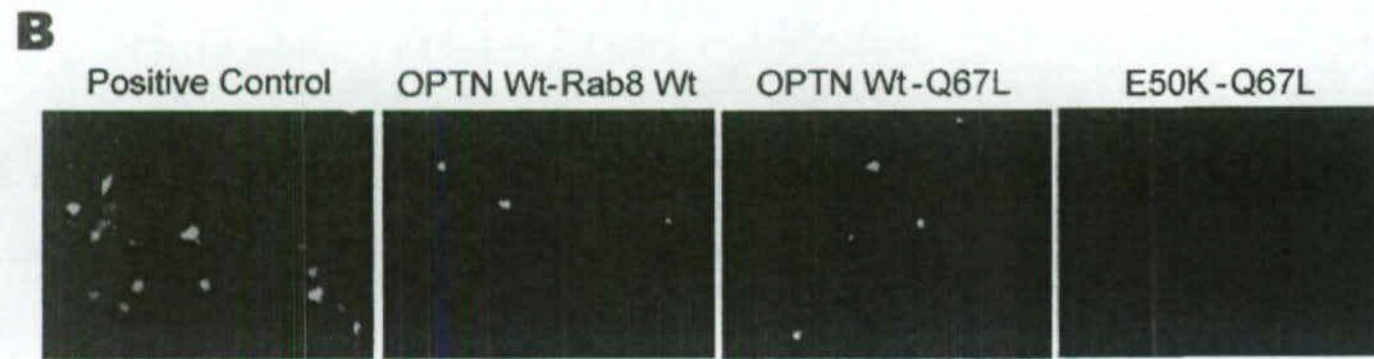
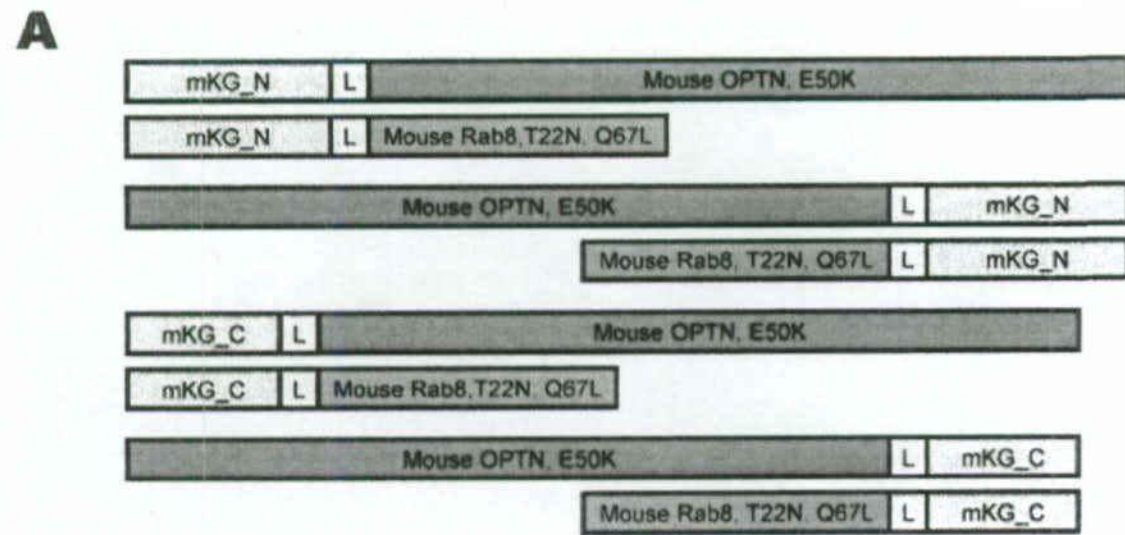
Peripheral

Central



**A****Impact Rebound Tonometer****B****Optical Interferometry Tonometer**



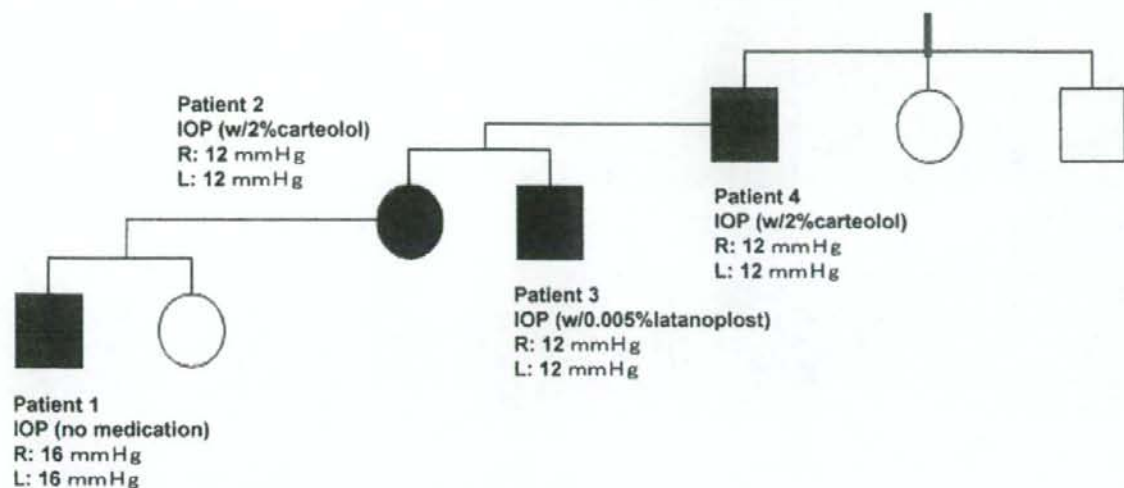
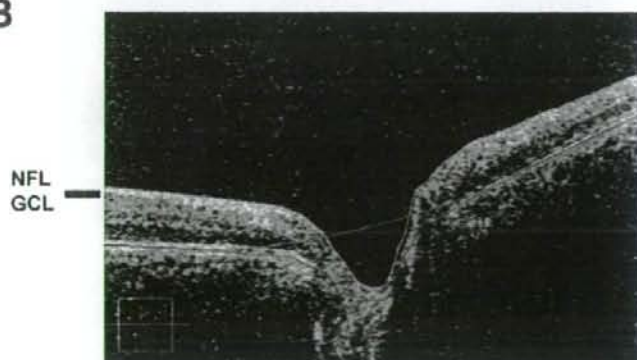


**A**

I

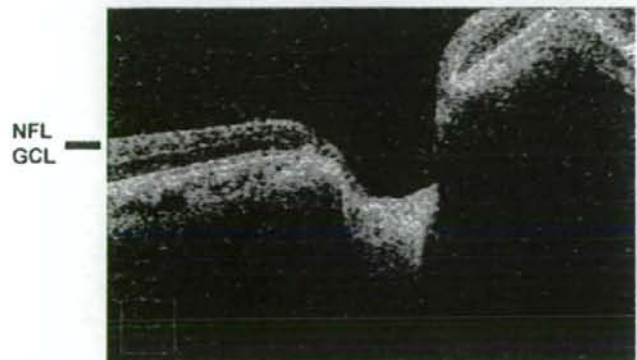
II

III

**B****Normal**

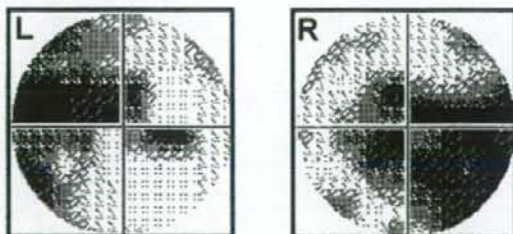
L

R

**Patient 2**

L

R



High Resolution Genome-wide Association Study  
on Advanced Wet-type Age-Related Macular Degeneration

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