

**Fig. 3** Computed tomography showed a round mass in the stomach (*arrow*) (a). FDG-PET shows hypermetabolism in the stomach (*arrow*) (b). Pathological examination of specimens obtained by surgery from the stomach (H&E) (c) revealed findings similar to those of pulmonary carcinoma obtained from the lung at surgery, confirming a metastatic poorly differentiated squamous cell carcinoma



studies have reported metastatic tumors of the gastrointestinal tract and the use of FDG-PET in gastrointestinal metastases [17, 18]. Studies have reported on 124 patients with gastrointestinal metastasis from lung cancer, 106 of whom were diagnosed on the occasion of serious complications such as perforation [3, 19]. Among our patients with lung cancer and MFH, one patient with vomiting after CR for chemotherapy, two patients with increased levels of CEA, and one patient with gastrointestinal bleeding underwent FDG-PET for the detection of recurrent lesions, which showed FDG uptake for the detection of gastrointestinal tract and other recurrent lesions. Thus, FDG-PET enabled the detection of metastatic tumors before the development of serious complications, and CT demonstrated well the local anatomy of the metastatic tumor of the gastrointestinal tract and other recurrent lesions.

FDG-PET imaging provided valuable information for the diagnosis of gastrointestinal metastasis, and these observations suggest that this imaging modality is also useful for the detection of metastatic tumors of the gastrointestinal tract, thereby enabling the prevention of complications such as intestinal obstruction, and massive bleeding, caused by metastatic tumors of the gastrointestinal tract.

## References

- Abrams HL, Spiro R, Goldstein N. Metastasis in carcinoma: analysis of 1000 autopsied cases. *Cancer* 1950;3:74–85.
- Menuck LS, Amberg JR. Metastatic disease involving the stomach. *Am J Dig Dis* 1975;20:903–13.
- Garwood RA, Sawyer MD, Ledesma EJ, Foley E, Claridge JA. A case and review of bowel perforation secondary to metastatic lung cancer. *Am Surg* 2005;71:110–6.
- Kostakoglu L, Agress H, Golodsmith FS. Clinical role of FDG PET in evaluation of cancer patients. *Radiographics* 2003;23:315–40.
- Rohren EM, Turkington TG, Coleman RE. Clinical applications of PET in oncology. *Radiology* 2004;231:305–31.
- van Kouwen MC, Oyen WJ, Negengast FM, Jansen JB, Drenth JP. FDG-PET scanning in the diagnosis of gastrointestinal cancers. *Scan J Gastroenterol Suppl* 2004;241:85–92.
- Gutman F, Albeirni JL, Wartski M, Vilain D, LeStanc E, Sarandi F, et al. Incidental colonic focal lesion detected by FDG PET/CT. *Am J Roentgenol* 2005;185:495–500.
- Hustrinx R. PET imaging in assessing gastrointestinal tumors. *Radiol Clin North Am* 2004;42:1123–39.
- Meyers MA, McSweeney J. Secondary neoplasms of the bowel. *Radiology* 1972;105:1–11.
- Hinoshita E, Nakahashi H, Wakasugi K, Kaneko S, Hamatake M, Sugimachi K. Duodenal metastasis from large cell carcinoma of the lung: report of a case. *Surg Today* 1999; 29:799–802.
- De Castro CA, Dockerty MB, Mayo CW. Metastatic tumors of the small intestines. *Surg Gynecol Obstet* 1957;105:159–65.
- Nussbaum M, Grossman M. Metastasis to the esophagus causing gastrointestinal bleeding. *Am J Gastroenterol* 1976;66:467–72.
- McAteer D, Wallis F, Couper G, Norton M, Welch A, Bruce D, et al. Evaluation of F18-FDG positron emission tomography in gastric and esophageal carcinoma. *Br J Radiol* 1999;72:525–9.
- Chen J, Cheong JH, Yun MJ, Kim J, Lim JS, Hyung WJ, et al. Improvement in preoperative staging of gastric adenocarcinoma with positron emission tomography. *Cancer* 2005;103:2283–90.

15. Choi MY, Lee KM, Chung JK, Lee DS, Jeong JM, Park JG, et al. Correlation between by FDG PET in postoperative patients with recurrent colorectal cancer. *Ann Nucl Med* 2005;19:123–9.
16. Kato H, Miyazaki T, Nakajima M, Fukuchi M, Manda R, Kuwano H. Value of positron emission tomography in the diagnosis of recurrent oesophageal carcinoma. *Br J Surg* 2004;91:1004–9.
17. Alibazoglu H, Alibazoglu B, Ali A, La Monica G. False-negative FDG-PET imaging in a patient with metastatic melanoma and ileal intussusception. *Clin Nucl Med* 1999; 24:129.
18. Tatlidil R, Mandelker NM. FDG-PET in the detection of gastrointestinal metastases in melanoma. *Melanoma Res* 2001;11:297–301.
19. Yamamoto M, Matsuzaki K, Kusumoto H, Uchida H, Mine H, Kabashima A, et al. Gastric metastasis from lung carcinoma: case report. *Hepatogastroenterology* 2002;49:363–5.



## Platelet-derived growth factor-BB amplifies $\text{PGF}_{2\alpha}$ -stimulated VEGF synthesis in osteoblasts: Function of phosphatidylinositol 3-kinase

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### Abstract

We have reported that prostaglandin  $\text{F}_{2\alpha}$  ( $\text{PGF}_{2\alpha}$ ) stimulates the synthesis of vascular endothelial growth factor (VEGF) via p44/p42 mitogen-activated protein (MAP) kinase in osteoblast-like MC3T3-E1 cells. In addition, we recently showed that phosphatidylinositol 3 (PI3)-kinase activated by platelet-derived growth factor-BB (PDGF-BB) negatively regulates the interleukin-6 synthesis in these cells. In the present study, we investigated the effect of PDGF-BB on the  $\text{PGF}_{2\alpha}$ -induced VEGF synthesis in MC3T3-E1 cells. PDGF-BB, which alone did not affect the levels of VEGF, significantly enhanced the  $\text{PGF}_{2\alpha}$ -stimulated VEGF synthesis. The amplifying effect of PDGF-BB was dose dependent in the range between 10 and 70 ng/ml. LY294002 or wortmannin, specific inhibitors of PI3-kinase, which by itself failed to affect the  $\text{PGF}_{2\alpha}$ -stimulated VEGF synthesis, significantly suppressed the amplification by PDGF-BB. PD98059, a specific inhibitor of MEK1/2, suppressed the amplification by PDGF-BB of the  $\text{PGF}_{2\alpha}$ -stimulated VEGF synthesis similar to the levels of  $\text{PGF}_{2\alpha}$  with PD98059. PDGF-BB itself induced the phosphorylation of p44/p42 MAP kinase in these cells, and the effects of PDGF-BB and  $\text{PGF}_{2\alpha}$  on the phosphorylation of p44/p42 MAP kinase were additive. Moreover, LY294002 had little effect on the phosphorylation of p44/p42 MAP kinase induced by  $\text{PGF}_{2\alpha}$  with PDGF-BB. These results strongly suggest that  $\text{PGF}_{2\alpha}$ -stimulated VEGF synthesis is amplified by PI3-kinase-mediated PDGF-BB signaling in osteoblasts, and that the effect is exerted at a point downstream from p44/p42 MAP kinase.

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### 1. Introduction

Platelet-derived growth factor (PDGF) is a mitogenic polypeptide, which mainly acts on connective tissue cells [1,2]. PDGF occurs as five different isoforms [2]. PDGF isoforms were initially isolated from human platelets, but have been shown to be synthesized and released from a variety of cell types including osteosarcoma and osteoblasts [1,3,4]. It is well recognized that bone

metabolism is strictly regulated by two functional cells, osteoblasts and osteoclasts, responsible for bone formation and bone resorption, respectively [5]. As for bone metabolism, PDGF-BB is a potent stimulator of osteoblast proliferation and induces bone resorption [4]. It is recognized that PDGF, released during platelet aggregation, plays a crucial role in fracture healing as a systemic factor and that PDGF also regulates bone remodeling as a local factor [4]. PDGF receptor has an intrinsic protein tyrosine kinase activity and associates with SH-2 domain-containing substrates such as phospholipase C- $\gamma$  and phosphatidylinositol 3-kinase (PI3-kinase) [1]. We have previously reported that

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PDGF-BB activates phosphatidylcholine-hydrolyzing phospholipase D via tyrosine kinase activation, resulting in protein kinase C activation in osteoblast-like MC3T3-E1 cells [6]. In addition, we recently showed that PI3-kinase negatively regulates the PDGF-BB-stimulated interleukin-6 synthesis in these cells [7]. However, the exact role of PDGF-BB in osteoblasts has not yet been fully clarified.

Bone remodeling carried out by osteoclasts and osteoblasts is accompanied with new capillaries extending [8,9]. During bone remodeling, capillary endothelial cells provide the microvasculature. Therefore, it is well recognized that the activities of osteoblasts, osteoclasts, and capillary endothelial cells are closely coordinated and regulate bone metabolism [10]. These functional cells are considered to influence one another via humoral factors as well as by direct cell-to-cell contact. Vascular endothelial growth factor (VEGF) is an angiogenic growth factor displaying high specificity for vascular endothelial cells [11]. VEGF that is synthesized and secreted from a variety of cell types such as vascular smooth muscle cells, increases capillary permeability and stimulates proliferation of endothelial cells [11]. As for bone metabolism, it has been shown that inactivation of VEGF causes complete suppression of blood vessel invasion concomitant with impaired trabecular bone formation and expansion of hypertrophic chondrocyte zone in mouse tibial epiphyseal growth plate [12]. Accumulating evidence suggests that osteoblasts among bone cells synthesize and secrete VEGF in response to various physiological agents [11,13–15]. We have previously reported that PGF<sub>2α</sub> (prostaglandin F<sub>2α</sub>) stimulates VEGF synthesis in MC3T3-E1 cells, and that among the mitogen-activated protein (MAP) kinase superfamily [16], p44/p42 MAP kinase plays as a positive regulator in the VEGF synthesis [17]. Taking these findings into account, it is currently speculated that VEGF secreted from osteoblasts may play an important role in the regulation of bone metabolism [10,18]. However, the mechanism behind VEGF synthesis and its synthesis in osteoblasts is not fully understood.

In the present study, we investigated the effect of PDGF-BB on the PGF<sub>2α</sub>-stimulated VEGF synthesis in osteoblast-like MC3T3-E1 cells. We here show that PDGF-BB up-regulates PGF<sub>2α</sub>-stimulated VEGF synthesis via PI3-kinase in these cells.

## 2. Materials and methods

### 2.1. Materials

PGF<sub>2α</sub> was purchased from Sigma Chemical Co. (St. Louis, MO). PDGF-BB and mouse VEGF enzyme immunoassay kit were purchased from R&D Systems,

Inc. (Minneapolis, MN). LY294002, wortmannin and PD98059 were obtained from Calbiochem–Novabiochem Co. (La Jolla, CA). Phospho-specific PI3-kinase antibodies, PI3-kinase antibodies, phospho-specific p44/p42 MAP kinase antibodies and p44/p42 MAP kinase antibodies were purchased from Cell Signaling Technology (Beverly, MA). ECL Western blotting detection system was purchased from Amersham Biosciences (Piscataway, NJ). Other materials and chemicals were obtained from commercial sources. PGF<sub>2α</sub> was dissolved in ethanol. LY294002, wortmannin and PD98059 were dissolved in dimethyl sulfoxide (DMSO). The maximum concentration of ethanol or DMSO was 0.1%, which did not affect the assay for VEGF or the Western blotting analysis.

### 2.2. Cell culture

Cloned osteoblast-like MC3T3-E1 cells derived from newborn mouse calvaria [19] were maintained as previously described [20]. Briefly, the cells were cultured in  $\alpha$ -minimum essential medium ( $\alpha$ -MEM) containing 10% fetal calf serum (FCS) at 37 °C in a humidified atmosphere of 5% CO<sub>2</sub>/95% air. The cells were seeded into 35-mm (5 × 10<sup>4</sup>) or 90-mm (5 × 10<sup>5</sup>) diameter dishes in  $\alpha$ -MEM containing 10% FCS. After 5 days, the medium was exchanged for  $\alpha$ -MEM containing 0.3% FCS. The cells were used for experiments after 48 h.

### 2.3. VEGF assay

The cultured cells were stimulated by various dose of PGF<sub>2α</sub> in 1 ml of  $\alpha$ -MEM containing 0.3% FCS for the indicated periods. When indicated, the cells were pretreated with LY294002, wortmannin or PD98059 for 60 min. The conditioned medium was collected at the end of the incubation, and the VEGF concentration was measured by ELISA kit.

### 2.4. Analysis of Western blotting

The cultured cells were stimulated by PGF<sub>2α</sub> and/or PDGF-BB in  $\alpha$ -MEM containing 0.3% FCS for the indicated periods. The cells were washed twice with phosphate-buffered saline and then lysed, homogenized and sonicated in a lysis buffer containing 62.5 mM Tris/HCl, pH 6.8, 2% sodium dodecyl sulfate (SDS), 50 mM dithiothreitol and 10% glycerol. The cytosolic fraction was collected as a supernatant after centrifugation at 125,000 × *g* for 10 min at 4 °C. SDS-polyacrylamide gel electrophoresis (PAGE) was performed by Laemmli [21] in 10% polyacrylamide gel. A Western blotting analysis was performed as described previously [17] by using phospho-specific p44/p42 MAP kinase antibodies, p44/p42 MAP kinase antibodies, phospho-specific PI3-kinase antibodies or PI3-kinase antibodies with

peroxidase-labeled antibodies raised in goat against rabbit IgG being used as second antibodies. The peroxidase activity on PVDG membrane was visualized on X-ray film by means of the ECL Western blotting detection system. When indicated, the cells were pretreated with various doses of LY294002 or wortmannin for 60 min.

### 2.5. Determination

The absorbance of enzyme immunoassay samples was measured at 450 nm with EL 340 Bio Kinetic Reader (Bio-Tek Instruments, Inc., Winooski, VT). The densitometric analysis was performed using Molecular Analyst/Macintosh (Bio-Rad Laboratories, Hercules, CA).

### 2.6. Statistical analysis

The data were analyzed by ANOVA followed by the Bonferroni method for multiple comparisons between pairs and a  $p < 0.05$  was considered significant. All data are presented as the mean  $\pm$  S.E.M. of triplicate determinations. Each experiment was repeated three times with similar results.

## 3. Results

### 3.1. Effect of PDGF-BB on PGF<sub>2 $\alpha$</sub> -induced VEGF synthesis in MC3T3-E1 cells

PDGF-BB, which alone had no effect on the VEGF levels, significantly enhanced the PGF<sub>2 $\alpha$</sub> -stimulated VEGF synthesis in a time-dependent manner in osteoblast-like MC3T3-E1 cells (Fig. 1). The effect of PDGF-BB was dose-dependent in the range between 10 and 70 ng/ml (Fig. 2). The maximum effect of PDGF-BB on the VEGF synthesis was observed at 70 ng/ml, which caused about 16 times enhancement of the PGF<sub>2 $\alpha$</sub> -effect.

### 3.2. Effect of LY294002 or wortmannin on the amplification by PDGF-BB of the PGF<sub>2 $\alpha$</sub> -induced VEGF synthesis in MC3T3-E1 cells

We have previously reported that PI3-kinase activated by PDGF-BB limits the PDGF-BB-stimulated IL-6 synthesis in osteoblast-like MC3T3-E1 cells [7]. In order to investigate whether the amplifying effect of PDGF-BB on the PGF<sub>2 $\alpha$</sub> -stimulated VEGF synthesis is via activation of PI3-kinase in MC3T3-E1 cells, we next examined the effect of LY294002, a specific inhibitor of PI3-kinase [22], on the synthesis of VEGF. LY294002, which by itself did not suppress the PGF<sub>2 $\alpha$</sub> -induced VEGF synthesis, significantly reduced the enhancement

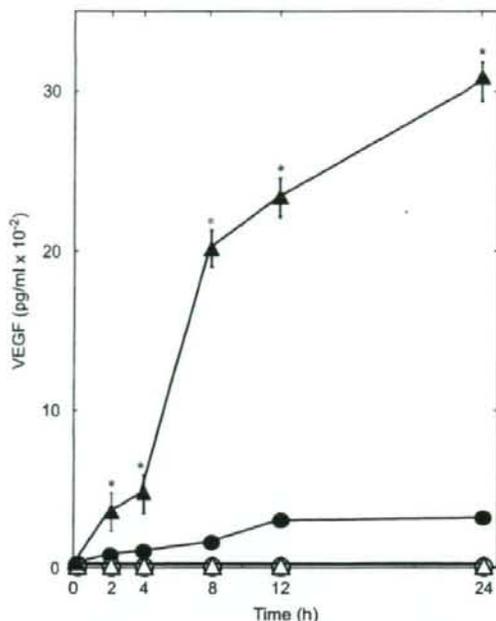


Fig. 1. Effect of PDGF-BB on the PGF<sub>2 $\alpha$</sub> -stimulated VEGF synthesis in MC3T3-E1 cells. The cultured cells were pretreated with 30 ng/ml PDGF-BB (▲, △) or vehicle (●, ○) for 60 min, and then stimulated by 10  $\mu$ M PGF<sub>2 $\alpha$</sub>  (●, ▲) or vehicle (○, △) for the indicated periods. Each value represents the mean  $\pm$  SEM of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \* $p < 0.05$ , compared to the value of PGF<sub>2 $\alpha$</sub>  alone.

by PDGF-BB of PGF<sub>2 $\alpha$</sub> -induced VEGF synthesis (Table 1). The effect of LY294002 was dose dependent in the range between 1 and 50  $\mu$ M (data not shown).

Wortmannin, another PI3-kinase inhibitor [23], as well as LY294002, also suppressed the amplification by PDGF-BB of the VEGF synthesis without affecting the PGF<sub>2 $\alpha$</sub> -stimulated synthesis (Table 2). The effect of wortmannin was dose dependent between 10 and 30  $\mu$ M (data not shown). We also examined the effects of LY294002 or wortmannin on the PDGF-BB-induced phosphorylation of PI3-kinase in these cells. Neither LY294002 nor wortmannin inhibited PDGF-BB-induced phosphorylation of PI3-kinase in these cells (Fig. 3).

### 3.3. Effect of PD98059 on the amplification by PDGF-BB of the PGF<sub>2 $\alpha$</sub> -induced VEGF synthesis in MC3T3-E1 cells

In a previous study [17], we showed that PGF<sub>2 $\alpha$</sub>  stimulates VEGF synthesis through p44/p42 MAP kinase activation in osteoblast-like MC3T3-E1 cells. In order to clarify whether the up-regulating effect of PDGF-BB on PGF<sub>2 $\alpha$</sub> -induced VEGF synthesis is due to p44/p42 MAP kinase activation in MC3T3-E1 cells, we

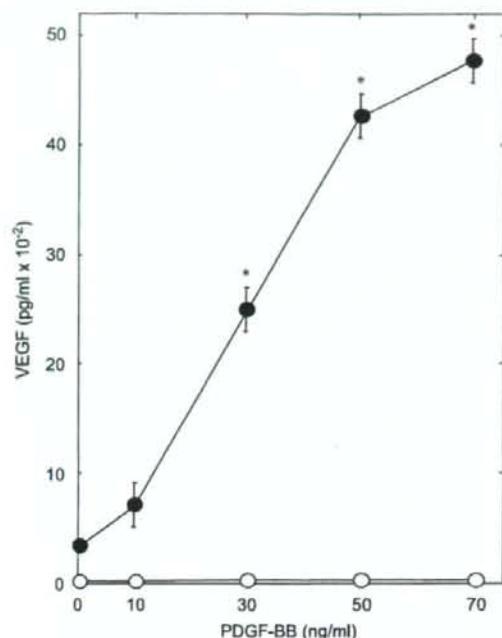


Fig. 2. Dose dependent effect of PDGF-BB on the PGF<sub>2α</sub>-stimulated VEGF synthesis in MC3T3-E1 cells. The cultured cells were pretreated with various doses of PDGF-BB for 60 min, and then stimulated by 10 μM PGF<sub>2α</sub> (●) or vehicle (○) for 24 h. Each value represents the mean ± SEM of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \**p* < 0.05, compared to the value of PGF<sub>2α</sub> alone.

Table 1

Effect of LY294002 on the enhancement by PDGF-BB of the PGF<sub>2α</sub>-stimulated VEGF synthesis in MC3T3-E1 cells

LY294002	PDGF-BB	PGF <sub>2α</sub>	VEGF (pg/ml)
–	–	–	15 ± 5
–	–	+	306 ± 27*
–	+	–	20 ± 5
–	+	+	3919 ± 157**
+	–	–	10 ± 5
+	–	+	440 ± 45
+	+	–	23 ± 10
+	+	+	1381 ± 88***

The cultured cells were pretreated with 50 μM LY294002 or vehicle for 60 min and then incubated by 30 ng/ml PDGF-BB or vehicle for 60 min. The cells were stimulated by 10 μM PGF<sub>2α</sub> or vehicle for 48 h. Each value represents the mean ± SEM of triplicate determinations. Similar results were obtained with two additional and different cell preparations.

\**p* < 0.05, compared to the control.

\*\**p* < 0.05, compared to the value of PGF<sub>2α</sub> alone.

\*\*\**p* < 0.05, compared to the value of PGF<sub>2α</sub> with PDGF-BB pretreatment.

Table 2

Effects of wortmannin or PD98059 on the enhancement by PDGF-BB of the PGF<sub>2α</sub>-stimulated VEGF synthesis in MC3T3-E1 cells

Inhibitor	PDGF-BB	PGF <sub>2α</sub>	VEGF (pg/ml)
–	–	–	20 ± 5
–	–	+	370 ± 28*
–	+	–	15 ± 10
–	+	+	3010 ± 139**
Wortmannin	–	–	20 ± 5
Wortmannin	–	+	462 ± 45
Wortmannin	+	–	18 ± 10
Wortmannin	+	+	406 ± 36***
PD98059	–	–	18 ± 5
PD98059	–	+	114 ± 20**
PD98059	+	–	15 ± 8
PD98059	+	+	122 ± 25***

The cultured cells were pretreated with 30 nM wortmannin, 30 μM PD98059 or vehicle for 60 min and then incubated by 30 ng/ml PDGF-BB or vehicle for 60 min. The cells were stimulated by 10 μM PGF<sub>2α</sub> or vehicle for 24 h. Each value represents the mean ± SEM of triplicate determinations. Similar results were obtained with two additional and different cell preparations.

\**p* < 0.05, compared to the control.

\*\**p* < 0.05, compared to the value of PGF<sub>2α</sub> alone.

\*\*\**p* < 0.05, compared to the value of PGF<sub>2α</sub> with PDGF-BB pretreatment.

examined the effect of PD98059, a specific inhibitor of the upstream kinase that activates p44/p42 MAP kinase [24], on the enhancement by PDGF-BB. PD98059, which by itself had no effect on the basal levels of VEGF, significantly reduced the enhancement by PDGF-BB of PGF<sub>2α</sub>-induced VEGF synthesis to the levels of the PGF<sub>2α</sub> with PD98059 (Table 2). The effect of PD98059 was dose dependent in the range between 1 and 30 μM (data not shown).

#### 3.4. Effect of PDGF-BB in combination with PGF<sub>2α</sub> on the phosphorylation of p44/p42 MAP kinase in MC3T3-E1 cells

We further investigated whether PDGF-BB could enhance the phosphorylation of p44/p42 MAP kinase induced by PGF<sub>2α</sub> in osteoblast-like MC3T3-E1 cells, and whether LY294002 could abolish it in osteoblast-like MC3T3-E1 cells. PDGF-BB itself induced the phosphorylation of p44/p42 MAP kinase in these cells, and that the effects of PDGF-BB and PGF<sub>2α</sub> on the phosphorylation of p44/p42 MAP kinase were additive (Fig. 4). In addition, LY294002 had little effect on the phosphorylation of p44/p42 MAP kinase induced by PGF<sub>2α</sub> with PDGF-BB (Fig. 4).

#### 4. Discussion

In the present study, we showed that PDGF-BB, which alone did not affect the levels of VEGF,

significantly amplified the PGF<sub>2α</sub>-stimulated VEGF synthesis in osteoblast-like MC3T3-E1 cells. We next investigated the mechanism of PDGF-BB behind the amplification. In our previous study [7], we reported

that PDGF-BB-activated PI3-kinase negatively regulates the interleukin-6 synthesis in MC3T3-E1 cells. We investigated whether or not PI3-kinase functions in the PDGF-BB-enhanced VEGF synthesis in osteoblast-like MC3T3-E1 cells. LY294002, a specific inhibitor of PI3-kinase [22], markedly suppressed the amplification by PDGF-BB of PGF<sub>2α</sub>-induced VEGF synthesis while the PGF<sub>2α</sub>-induced VEGF synthesis was not inhibited by LY294002. Thus, these results suggest that PI3-kinase is implicated in the PDGF-BB-induced potentiation. In addition, wortmannin, another PI3-kinase inhibitor [23], reduced the PDGF-BB-effect in the VEGF synthesis without suppressing the PGF<sub>2α</sub>-induced VEGF synthesis as well as LY294002. We found that neither LY294002 nor wortmannin could inhibit PDGF-BB-induced phosphorylation of PI3-kinase in these cells. Both LY294002 and wortmannin are competitive inhibitors of PI3-kinase [22,23]. It is well recognized that PI3-kinase is activated by its phosphorylation and that LY294002 is a highly selective PI3-kinase inhibitor which blocks PI3-kinase-dependent phosphorylation of its target proteins [22]. Namely, LY294002 and wortmannin are not inhibitors of the phosphorylation of PI3-kinase. Thus, it is reasonable that the effects of these inhibitors on PI3-kinase-mediated PDGF-BB signaling are exerted at a point downstream of the phosphorylation of PI3-kinase in MC3T3-E1 cells. Indeed, we have previously reported that the phosphorylation of Akt induced by PDGF-BB is significantly inhibited by both LY294002 and wortmannin [25]. Therefore, it is sure that these inhibitors attenuate the PDGF-BB-activating PI3-kinase/Akt signaling in these cells. On the other hand, the amplification by PDGF-BB of PGF<sub>2α</sub> was observed during 4 and 8 h in the time course study. It is possible that PDGF-BB-stimulated other protein synthesis might be involved in the PDGF-BB effect. Therefore, taking our results into account as a whole, it is most likely that PDGF-BB activates the PI3-kinase pathway, resulting in the potentiation of PGF<sub>2α</sub>-induced VEGF synthesis in MC3T3-E1 cells.

It is well recognized that the MAP kinase superfamily mediates intracellular signaling of extracellular agonists and plays a pivotal role in cellular functions including

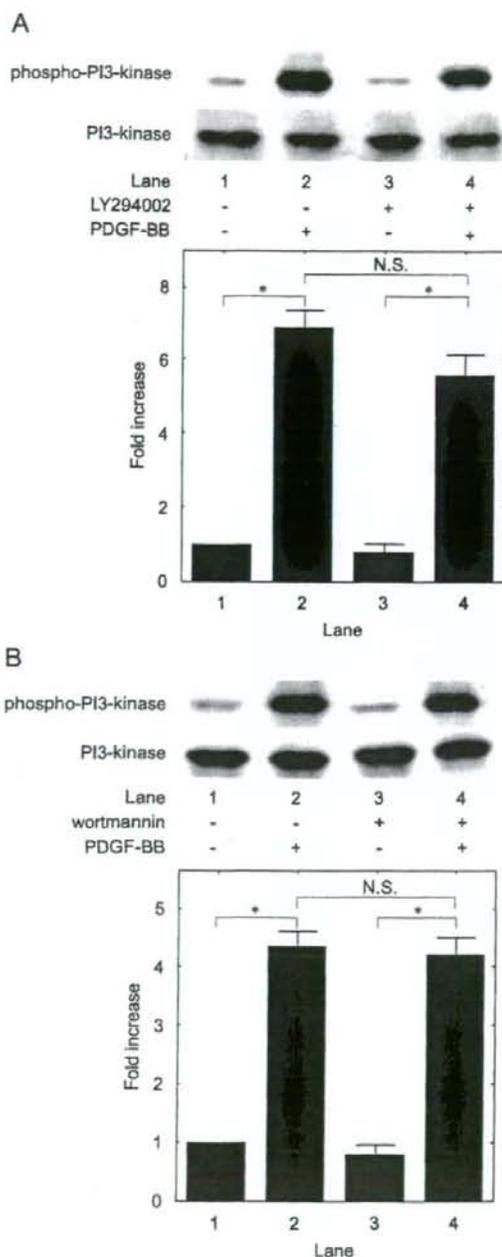


Fig. 3. Effects of LY294002 or wortmannin on the phosphorylation of PI3-kinase induced by PDGF-BB in MC3T3-E1 cells. The cultured cells were pretreated with 50  $\mu$ M LY294002 (A), 30 nM wortmannin (B) or vehicle for 60 min, and then stimulated by 30 ng/ml PDGF-BB or vehicle for 60 min. The extracts of the cells were subjected to SDS-PAGE with a subsequent Western blotting analysis with antibodies against phospho-specific PI-3 kinase or PI-3 kinase. The histogram shows quantitative representations of the levels of PDGF-BB-induced phosphorylation obtained from a laser densitometric analysis of three independent experiments. Each value represents the mean  $\pm$  SEM of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \* $p$  < 0.05, compared to the value of control. N.S., no statistical significance.

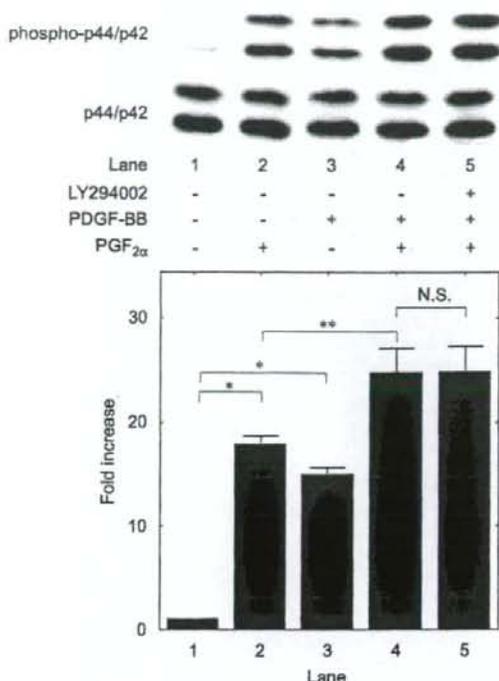


Fig. 4. Effect of PDGF-BB in combination with PGF<sub>2α</sub> on the phosphorylation of p44/p42 MAP kinase in MC3T3-E1 cells. The cultured cells were pretreated with 50 μM LY294002 or vehicle for 60 min, and then incubated by 30 ng/ml PDGF-BB or vehicle for 60 min. The treated cells were stimulated by 10 μM PGF<sub>2α</sub> or vehicle for 30 min. The extracts of cells were subjected to SDS-PAGE with subsequent Western blotting analysis with antibodies against phospho-specific p44/p42 MAP kinase or p44/p42 MAP kinase. The histogram shows quantitative representations of the levels of PDGF-BB- and/or PGF<sub>2α</sub>-induced phosphorylation obtained from a laser densitometric analysis of three independent experiments. Each value represents the mean ± SEM of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \**p* < 0.05, compared to the value of control. \*\**p* < 0.05, compared to the value of PGF<sub>2α</sub> alone. N.S., no statistical significance.

proliferation, differentiation, and apoptosis in a variety of cells [16]. Three major MAP kinase, p44/p42 MAP kinase, p38 MAP kinase, and SAPK/JNK are known as central elements used by mammalian cells to transduce the diverse messages [16]. In our previous study [17], we have shown that p44/p42 MAP kinase acts as a positive regulator in PGF<sub>2α</sub>-induced VEGF synthesis in MC3T3-E1 cells. Therefore, we tried to clarify the relationship between PDGF-BB-effect and p44/p42 MAP kinase in the PGF<sub>2α</sub>-stimulated VEGF synthesis in these cells. The amplification by PDGF-BB of the PGF<sub>2α</sub>-stimulated VEGF synthesis was suppressed by PD98059, a specific inhibitor of MEK1/2 [23], similar to the levels of PGF<sub>2α</sub> with PD98059. Moreover, we found that PDGF-

BB itself induced the phosphorylation of p44/p42 MAP kinase in these cells, and that the effects of PDGF-BB and PGF<sub>2α</sub> on the phosphorylation of p44/p42 MAP kinase were additive. These results suggest that PDGF-BB activates p44/p42 MAP kinase independently from PGF<sub>2α</sub> in these cells. PDGF-BB as well as PGF<sub>2α</sub> induced the phosphorylation of p44/p42 MAP kinase, however, VEGF synthesis was only observed by PGF<sub>2α</sub>, not by PDGF-BB. Our results indicate that PDGF-BB-induced p44/p42 MAP kinase activation alone is not sufficient for VEGF synthesis in osteoblasts, which is quite different from PGF<sub>2α</sub>-induced p44/p42 MAP kinase activation. In addition, the increase in VEGF by PDGF-BB and PGF<sub>2α</sub> was not additive but synergistic. We also found that LY294002 had little effect on the phosphorylation of p44/p42 MAP kinase induced by PGF<sub>2α</sub> with PDGF-BB. Therefore, it is probable that the enhancement by PDGF of PGF<sub>2α</sub>-induced VEGF synthesis through PI3-kinase is exerted at a point downstream from p44/p42 MAP kinase in these cells. As a whole, it is most likely that PDGF-BB independently activates p44/p42 MAP kinase and PI3-kinase in osteoblasts, and the latter is involved in the enhancement of PGF<sub>2α</sub>-induced VEGF synthesis at a point downstream from p44/p42 MAP kinase.

The expansion of capillary network providing microvasculature is an essential process of bone remodeling [10]. Since VEGF is a specific mitogen of vascular endothelial cells [11], it is recognized that VEGF secreted by osteoblasts plays a crucial role as an intercellular mediator between osteoblasts and vascular endothelial cells in bone metabolism. Moreover, it has been reported that VEGF is involved in trabecular bone formation and expansion of the hypertrophic chondrocyte zone in epiphyseal growth plate of mouse [12], supporting the significance of VEGF in bone metabolism. On the other hand, the mitogenic activities of PDGF and its release by platelets suggest a pivotal role in wound healing and bone fracture repair [26]. Therefore, our present findings lead us to speculate that PDGF-BB-enhanced VEGF synthesis from osteoblasts plays a pivotal role in the process of bone remodeling via up-regulating the proliferation of capillary endothelial cells. Further investigations are required to elucidate the precise role of PDGF-BB in bone metabolism.

In conclusion, our present results strongly suggest that PI3-kinase signaling pathway activated by PDGF-BB amplifies PGF<sub>2α</sub>-induced VEGF synthesis in osteoblasts, and that the effect is exerted at a point downstream from p44/p42 MAP kinase.

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## References

- [1] C.H. Heldin, B. Westermark, Mechanism of action and in vivo role of platelet-derived growth factor, *Physiol. Rev.* 79 (1999) 1283–1316.
- [2] C.H. Heldin, U. Eriksson, A. Ostman, New members of the platelet-derived growth factor family of mitogens, *Arch. Biochem. Biophys.* 398 (2002) 284–290.
- [3] C.H. Heldin, A. Johnsson, S. Wennergren, C. Wernstedt, C. Betsholtz, B. Westermark, A human osteosarcoma cell line secretes a growth factor structurally related to a homodimer of PDGF A-chains, *Nature* 319 (1986) 511–514.
- [4] E. Canalis, S. Rydziel, Platelet-derived growth factor and the skeleton, *Principles Bone Biol.* 2 (2002) 817–824.
- [5] P.J. Nijweide, E.H. Burger, J.H.M. Feyen, Cells of bone: proliferation, differentiation, and hormonal regulation, *Physiol. Rev.* 86 (1986) 855–886.
- [6] O. Kozawa, A. Suzuki, Y. Watanabe, J. Shinoda, Y. Oiso, Effect of platelet-derived growth factor on phosphatidylcholine-hydrolyzing phospholipase D in osteoblast-like cells, *Endocrinology* 136 (1995) 4473–4478.
- [7] Y. Hanai, H. Tokuda, T. Ohta, R. Matsushima-Nishiwaki, S. Takai, O. Kozawa, Phosphatidylinositol 3-kinase/Akt auto-regulates PDGF-BB-stimulated interleukin-6 synthesis in osteoblasts, *J. Cell. Biochem.* 99 (2006) 1564–1571.
- [8] C.T. Brighton, R.M. Hunt, Early histological and ultrastructural changes in medullary fracture callus, *Am. J. Bone Joint Surg.* 73 (1981) 832–847.
- [9] A.M. Parfitt, Osteonal and hemi-osteonal remodeling: the spatial and temporal framework for signal traffic in adult human bone, *J. Cell. Biochem.* 55 (1994) 273–286.
- [10] A. Erlebacher, E.H. Filvaroff, S.E. Gitelman, R. Derynck, Toward a molecular understanding of skeletal development, *Cell* 80 (1995) 371–378.
- [11] N. Ferrara, T. Davis-Smyth, The biology of vascular endothelial growth factor, *Endocr. Rev.* 18 (1997) 4–25.
- [12] H.P. Gerber, T.H. Vu, A.M. Ryan, J. Kowalski, Z. Werb, N. Ferrara, VEGF couples hypertrophic cartilage remodeling, ossification and angiogenesis during endochondral bone formation, *Nat. Med.* 5 (1999) 623–628.
- [13] D.L. Goad, J. Rubin, H. Wang, A.H. Tashjian Jr., C. Patterson, Enhanced expression of vascular endothelial growth factor in human SaOS-2 osteoblast-like cells and murine osteoblasts induced by insulin-like growth factor I, *Endocrinology* 137 (1996) 2262–2268.
- [14] D.S. Wang, K. Yamazaki, K. Nohtomi, K. Shizume, K. Ohsumi, M. Shibuya, H. Demura, K. Sato, Increase of vascular endothelial growth factor mRNA expression by 1,25-dihydroxyvitamin D<sub>3</sub> in human osteoblast-like cells, *J. Bone Miner. Res.* 11 (1996) 472–479.
- [15] J.M. Schallaeppi, S. Gutzwiller, G. Finlenzeller, B. Fournier, 1,25-dihydroxyvitamin D<sub>3</sub> induces the expression of vascular endothelial growth factor in osteoblastic cells, *Endocr. Res.* 23 (1997) 213–229.
- [16] C. Widmann, S. Gibson, M.B. Jarpe, G.L. Johnson, Mitogen-activated protein kinase: conservation of a three-kinase module from yeast to human, *Physiol. Rev.* 79 (1999) 143–180.
- [17] H. Tokuda, A. Harada, K. Hirade, H. Matsuno, H. Ito, K. Kato, Y. Oiso, O. Kozawa, Incadronate amplifies prostaglandin F<sub>2α</sub>-induced vascular endothelial growth factor in osteoblasts: enhancement of MAP kinase activity, *J. Biol. Chem.* 278 (2003) 18930–18937.
- [18] E. Zelzer, B.R. Olsen, Multiple roles of vascular endothelial growth factor (VEGF) in skeletal development, growth, and repair, *Curr. Top. Dev. Biol.* 65 (2005) 169–187.
- [19] H. Sudo, H. Kodama, Y. Amagai, S. Yamamoto, S. Kasai, In vitro differentiation and calcification in a new clonal osteogenic cell line derived from newborn mouse calvaria, *J. Cell. Biol.* 96 (1983) 191–198.
- [20] O. Kozawa, H. Tokuda, M. Miwa, J. Kotoyori, Y. Oiso, Cross-talk regulation between cyclic AMP production and phosphoinositide hydrolysis induced by prostaglandin E<sub>2</sub> in osteoblast-like cells, *Exp. Cell Res.* 198 (1992) 130–134.
- [21] U.K. Laemmli, Cleavage of structural proteins during the assembly of the head of bacteriophage T4, *Nature* 227 (1970) 680–685.
- [22] C.J. Vlahos, W.F. Matter, K.Y. Hui, R.F. Brown, A specific inhibitor of phosphatidylinositol 3-kinase, 2-(4-morpholinyl)-8-phenyl-4H-1-benzopyran-4-one (LY294002), *J. Biol. Chem.* 269 (1994) 5721–5728.
- [23] A. Arcaro, M.P. Wymann, Wortmannin is a potent phosphatidylinositol 3-kinase inhibitor: the role of phosphatidylinositol 3,4,5-trisphosphate in neutrophil responses, *Biochem. J.* 296 (1993) 297–301.
- [24] D.R. Alessi, A. Cuenda, P. Cohen, D.T. Dudley, A.R. Saltiel, PD98059 is a specific inhibitor of the activation of mitogen-activated protein kinase in vitro and in vivo, *J. Biol. Chem.* 270 (1995) 27489–27494.
- [25] Y. Hanai, H. Tokuda, H. Ohta, R. Matsushima-Nishiwaki, S. Takai, O. Kozawa, Phosphatidylinositol 3-kinase/Akt auto-regulates PDGF-BB-stimulated interleukin-6 synthesis in osteoblasts, *J. Cell. Biochem.* 99 (2006) 1564–1571.
- [26] E. Canalis, S. Rydziel, Platelet-derived growth factor and the skeleton, *Principles of Bone Biology*, second ed., 2002, pp. 817–824.

## Negative Regulation by p70 S6 Kinase of FGF-2-Stimulated VEGF Release Through Stress-Activated Protein Kinase/c-Jun N-Terminal Kinase in Osteoblasts

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**ABSTRACT:** To clarify the mechanism of VEGF release in osteoblasts, we studied whether p70 S6 kinase is involved in basic FGF-2-stimulated VEGF release in osteoblast-like MC3T3-E1 cells. In this study, we show that p70 S6 kinase activated by FGF-2 negatively regulates VEGF release through SAPK/JNK in osteoblasts.

**Introduction:** Vascular endothelial growth factor (VEGF) plays an important role in bone metabolism. We have previously reported that fibroblast growth factor-2 (FGF-2) stimulates the release of VEGF through p44/p42 mitogen-activated protein (MAP) kinase and stress-activated protein kinase/c-Jun N-terminal kinase (SAPK/JNK) in osteoblast-like MC3T3-E1 cells and that FGF-2-activated p38 MAP kinase negatively regulates VEGF release. However, the mechanism behind VEGF release in osteoblasts is not precisely known.

**Materials and Methods:** The levels of VEGF released from MC3T3-E1 cells were measured by enzyme immunoassay. The phosphorylation of each protein kinase was analyzed by Western blotting. To knock down p70 S6 kinase in MC3T3-E1 cells, the cells were transfected with siRNA to target p70 S6 kinase.

**Results:** FGF-2 time-dependently induced the phosphorylation of p70 S6 kinase. Rapamycin significantly enhanced the FGF-2-stimulated VEGF release and VEGF mRNA expression. The FGF-2-induced phosphorylation of p70 S6 kinase was suppressed by rapamycin. Rapamycin markedly enhanced the FGF-2-induced phosphorylation of SAPK/JNK without affecting the phosphorylation of p44/p42 MAP kinase or p38 MAP kinase. SP600125, a specific inhibitor of SAPK/JNK, suppressed the amplification by rapamycin of the FGF-2-stimulated VEGF release similar to the levels of FGF-2 with SP600125. Finally, downregulation of p70 S6 kinase by siRNA significantly enhanced the FGF-2-stimulated VEGF release and phosphorylation of SAPK/JNK.

**Conclusions:** These results strongly suggest that p70 S6 kinase limits FGF-2-stimulated VEGF release through self-regulation of SAPK/JNK, composing a negative feedback loop, in osteoblasts.

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**Key words:** p70 S6 kinase, fibroblast growth factor-2, vascular endothelial growth factor, osteoblasts, mitogen-activated protein kinase

### INTRODUCTION

IT IS WELL known that osteoblasts synthesize basic fibroblast growth factor (FGF)-2, and FGF-2 is embedded in bone matrix.<sup>(1,2)</sup> FGF-2 expression in osteoblasts is detected during fracture repair.<sup>(3)</sup> Bone metabolism is strictly regulated by osteoblasts and osteoclasts, which are responsible for bone formation and bone resorption, respectively.<sup>(4)</sup> Therefore, it is thought that FGF-2 plays a pivotal role in fracture healing, bone remodeling, and osteogenesis.<sup>(5)</sup> We have previously reported that FGF-2 autophos-

phorylates FGF receptors 1 and 2 among four structurally related high affinity receptors in osteoblast-like MC3T3-E1 cells.<sup>(6)</sup> In addition, we reported that FGF-2 stimulates induction of heat shock protein 27 in these cells.<sup>(7)</sup>

Bone remodeling carried out by osteoclasts and osteoblasts is accompanied by angiogenesis and capillary outgrowth.<sup>(8,9)</sup> During bone remodeling, capillary endothelial cells provide the microvasculature. It is well recognized that the activities of osteoblasts, osteoclasts, and capillary endothelial cells are closely coordinated and regulate bone metabolism.<sup>(10)</sup> These functional cells are considered to influence one another through humoral factors and by direct cell-to-cell contact. Vascular endothelial growth factor

The authors state that they have no conflicts of interest.

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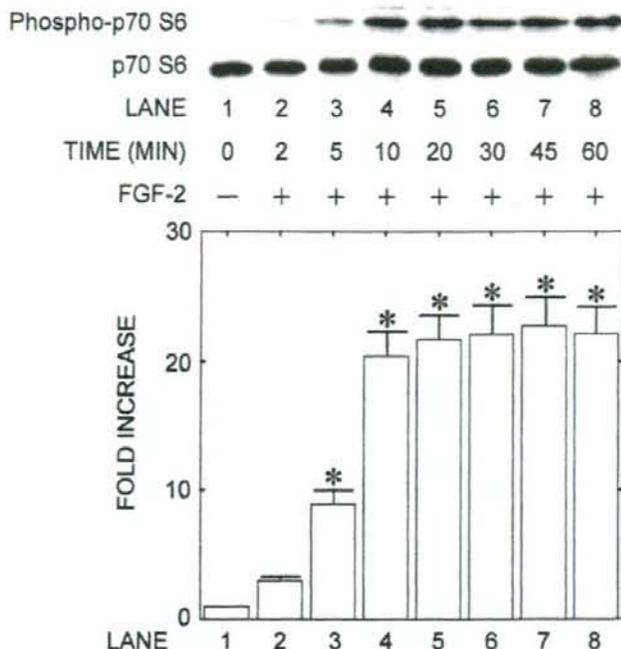


FIG. 1. Effect of FGF-2 on the phosphorylation of p70 S6 kinase in MC3T3-E1 cells. The cultured cells were stimulated by 70 ng/ml FGF-2 for the indicated periods. The extracts of cells were subjected to SDS-PAGE with subsequent Western blotting analysis with antibodies against phospho-specific p70 S6 kinase or p70 S6 kinase. The histogram shows quantitative representations of the levels of FGF-2-induced phosphorylation obtained from laser densitometric analysis of three independent experiments. Each value represents the mean  $\pm$  SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \* $p < 0.05$  compared with the value of control.

(VEGF) is an angiogenic growth factor displaying high specificity for vascular endothelial cells.<sup>(11)</sup> VEGF, produced and secreted from a variety of cell types, increases capillary permeability and stimulates proliferation of endothelial cells.<sup>(11)</sup> As for bone metabolism, an inactivation of VEGF reportedly causes complete suppression of blood vessel invasion concomitant with impaired trabecular bone formation and expansion of hypertrophic chondrocyte zone in mouse tibial epiphyseal growth plate.<sup>(12)</sup> Evidence is accumulating that osteoblasts among bone cells produce and secrete VEGF in response to various physiological agents.<sup>(11,13-15)</sup> We have previously reported that FGF-2 stimulates VEGF release in MC3T3-E1 cells and that the release is positively regulated by p44/p42 mitogen-activated protein (MAP) kinase and stress-activated protein kinase/c-Jun N-terminal kinase (SAPK/JNK)<sup>(16,17)</sup> among the MAP kinase superfamily.<sup>(18)</sup> Based on these findings, it is currently recognized that VEGF secreted from osteoblasts may play an important role in the regulation of bone metabolism.<sup>(10,19)</sup> However, the mechanism behind VEGF synthesis in osteoblasts and its release from these cells is not fully understood.

p70 S6 kinase is a mitogen-activated serine/threonine kinase required for cell proliferation and G1 cell cycle progression.<sup>(20)</sup> As for osteoblasts, it has been shown that fluoroaluminate induces an increase in p70 S6 kinase phosphorylation.<sup>(21)</sup> In our previous study,<sup>(22)</sup> we reported that p70 S6 kinase plays a positive regulator in bone morphogenetic protein-4-stimulated release of VEGF in osteoblast-like MC3T3-E1 cells. In addition, we recently showed that

p38 MAP kinase, a member of the MAP kinase superfamily, functions at a point upstream from p70 S6 kinase in the release of VEGF in these cells.<sup>(23)</sup> However, the exact role of p70 S6 kinase in osteoblasts still remains unclear.

In this study, we investigated the involvement of p70 S6 kinase in the FGF-2-stimulated VEGF release in osteoblast-like MC3T3-E1 cells. We show that p70 S6 kinase activated by FGF-2 negatively regulates VEGF release through SAPK/JNK in these cells.

## MATERIALS AND METHODS

### Materials

FGF-2 and mouse VEGF enzyme immunoassay kits were purchased from R&D Systems (Minneapolis, MN, USA). Rapamycin and SP600125 were obtained from Calbiochem-Novabiochem Co. (La Jolla, CA, USA). Phospho-specific p70 S6 kinase antibodies, p70 S6 kinase antibodies, phospho-specific p44/p42 MAP kinase antibodies, p44/p42 MAP kinase antibodies, phospho-specific p38 MAP kinase antibodies, p38 MAP kinase antibodies, phospho-specific SAPK/JNK antibodies, and SAPK/JNK antibodies were purchased from Cell Signaling (Beverly, MA, USA). ECL Western blotting detection system was purchased from Amersham Biosciences (Piscataway, NJ, USA). Control short interfering RNA (siRNA; Silencer Negative Control no. 1 siRNA) or p70 S6 kinase siRNA (Silencer Pre-designed siRNA, 75849, 75755, and 75942) was purchased from Ambion (Austin, TX, USA). siLentFect was pur-

chased from Bio-Rad (Hercules, CA, USA). Trizol reagent was purchased from Invitrogen (Carlsbad, CA, USA). Omniscript Reverse Transcriptase Kit was purchased from QIAGEN (Hilden, Germany). Fast-Start DNA Master SYBR Green I was purchased from Roche Diagnostics (Mannheim, Germany). Other materials and chemicals were obtained from commercial sources. Rapamycin or SP600125 was dissolved in dimethyl sulfoxide (DMSO). The maximum concentration of DMSO was 0.1%, which did not affect the assay for VEGF or Western blot analysis.

#### Cell culture

Cloned osteoblast-like MC3T3-E1 cells derived from newborn mouse calvaria<sup>(24)</sup> were maintained as previously described.<sup>(25)</sup> Briefly, the cells were cultured in  $\alpha$ -MEM containing 10% FCS at 37°C in a humidified atmosphere of 5% CO<sub>2</sub>/95% air. The cells were seeded into 35- (5 × 10<sup>4</sup>) or 90-mm-diameter (5 × 10<sup>5</sup>) dishes in  $\alpha$ -MEM containing 10% FCS. After 5 days, the medium was exchanged for  $\alpha$ -MEM containing 0.3% FCS. The cells were used for experiments after 48 h.

#### siRNA transfection

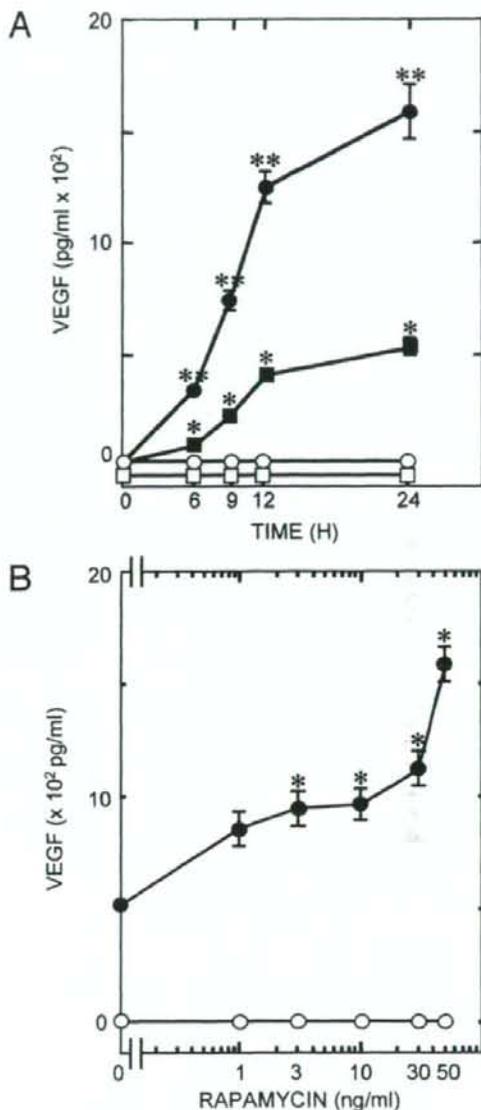
To knock down p70 S6 kinase in MC3T3-E1 cells, the cells were transfected with control siRNA (Silencer Negative Control no. 1 siRNA) or p70 S6 kinase siRNA (Silencer Predesigned siRNA, 75849, 75755, and 75942; Ambion) using the siLentFect (Bio-Rad) according to the manufacturer's protocol. In brief, the cells were seeded in a 35-mm-diameter (1 × 10<sup>5</sup>) dish in  $\alpha$ -MEM containing 10% FCS and subcultured for 48 h. After that, the cells were incubated at 37°C for 48 h with 250 nM siRNA-siLentFect complexes.

#### VEGF assay

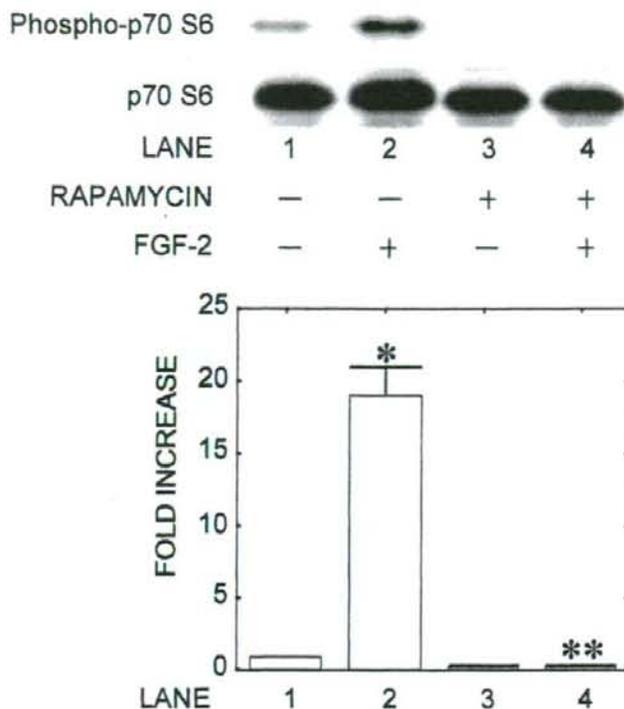
The cultured cells were stimulated by various doses of FGF-2 in 1 ml of  $\alpha$ -MEM containing 0.3% FCS for the indicated periods. When indicated, the cells were pretreated with rapamycin or SP600125 for 60 minutes. The conditioned medium was collected at the end of the incubation, and the VEGF concentration was measured by ELISA kit.

#### Western blot analysis

The cultured cells were stimulated by FGF-2 in  $\alpha$ -MEM containing 0.3% FCS for the indicated periods. The cells were washed twice with PBS, lysed, homogenized, and sonicated in a lysis buffer containing 62.5 mM Tris/HCl, pH 6.8, 2% SDS, 50 mM dithiothreitol, and 10% glycerol. The cytosolic fraction was collected as a supernatant after centrifugation at 125,000g for 10 minutes at 4°C. SDS-PAGE was performed according to Laemmli<sup>(26)</sup> in 10% polyacrylamide gel. Western blotting analysis was performed as described previously<sup>(27)</sup> using phospho-specific p70 S6 kinase antibodies, p70 S6 kinase antibodies, phospho-specific p44/p42 MAP kinase antibodies, p44/p42 MAP kinase antibodies, phospho-specific p38 MAP kinase antibodies, p38 MAP kinase antibodies, phospho-specific SAPK/JNK antibodies, or SAPK/JNK antibodies, with peroxidase-labeled antibodies raised in goat against rabbit IgG being used as second



**FIG. 2.** Effect of rapamycin on the FGF-2-stimulated VEGF release in MC3T3-E1 cells. (A) The cultured cells were pretreated with 10 ng/ml rapamycin (circle symbols) or vehicle (square symbols) for 60 minutes and stimulated by 70 ng/ml FGF-2 (solid symbols) or vehicle (open symbols) for the indicated periods. \**p* < 0.05 compared with the control. \*\**p* < 0.05 compared with the value of FGF-2 alone. (B) The cultured cells were pretreated with various doses of rapamycin for 60 minutes and stimulated by 70 ng/ml FGF-2 (●) or vehicle (○) for 24 h. \**p* < 0.05 compared with the value of FGF-2 alone. Each value represents the mean ± SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations.



**FIG. 3.** Effect of rapamycin on the FGF-2-induced phosphorylation of p70 S6 kinase in MC3T3-E1 cells. The cultured cells were pretreated with 10 ng/ml rapamycin or vehicle for 60 minutes and stimulated by 30 ng/ml FGF-2 or vehicle for 10 minutes. The extracts of cells were subjected to SDS-PAGE with subsequent Western blotting analysis with antibodies against phospho-specific p70 S6 kinase or p70 S6 kinase. The histogram shows quantitative representations of the levels of FGF-2-induced phosphorylation obtained from laser densitometric analysis of three independent experiments. Each value represents the mean  $\pm$  SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \* $p < 0.05$  compared with the control. \*\* $p < 0.05$  compared with the value of FGF-2 alone.

antibodies. Peroxidase activity on the polyvinylidene difluoride (PVDF) sheet was visualized on X-ray film by means of the electrochemiluminescence (ECL) Western blotting detection system.

#### Determination

The absorbance of enzyme immunoassay samples was measured at 450 nm with EL 340 Bio Kinetic Reader (Bio-Tek Instruments, Winooski, VT, USA). The densitometric analysis was performed using Molecular Analyst/Macintosh (Bio-Rad Laboratories).

#### Real-time RT-PCR

The cultured cells were pretreated with rapamycin and/or SP600125 and stimulated by FGF-2 for the indicated time period. Total RNA was isolated and transcribed into cDNA using Trizol reagent and Omniscript Reverse Transcriptase Kit. Real-time PCR was performed using a Light Cycler system (Roche Diagnostics) in capillaries and Fast-Start DNA Master SYBR Green I provided with the kit. Sense and antisense primers were synthesized based on the report of Simpson et al.<sup>(28)</sup> for mouse VEGF mRNA and GAPDH mRNA. The amplified products were determined by melting curve analysis and agarose electrophoresis. VEGF mRNA levels were normalized with those of GAPDH mRNA.

#### Statistical analysis

The data were analyzed by ANOVA followed by the Bonferroni method for multiple comparisons between pairs, and  $p < 0.05$  was considered significant. All data are presented as the mean  $\pm$  SE of triplicate determinations. Each experiment was repeated three times with similar results.

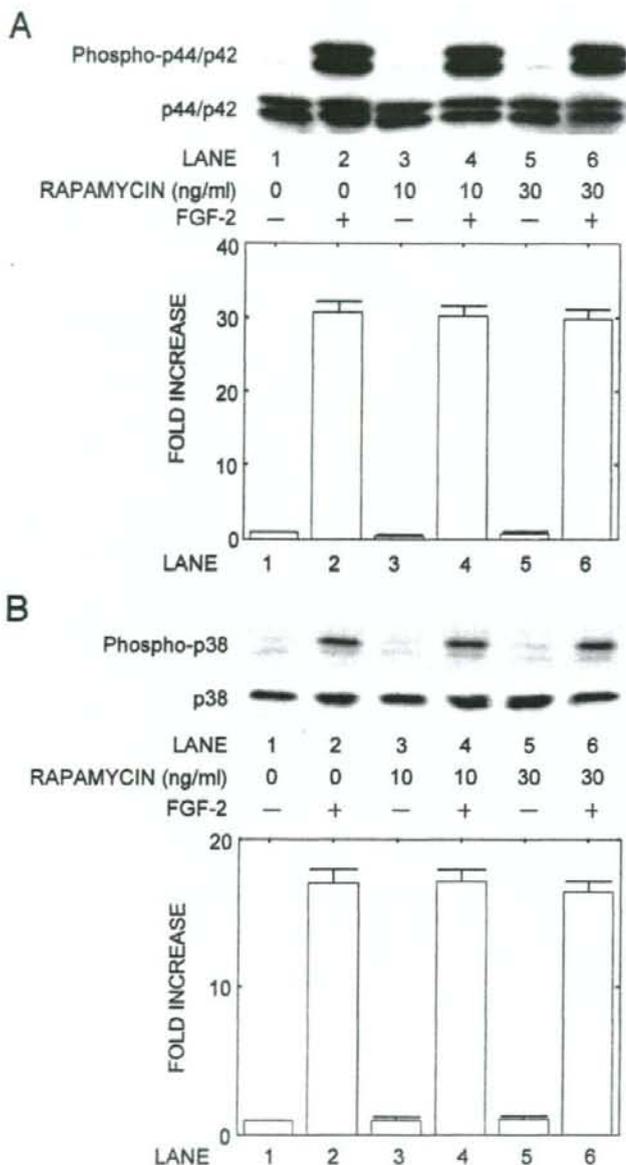
## RESULTS

#### Effect of FGF-2 on the phosphorylation of p70 S6 kinase in MC3T3-E1 cells

To study whether FGF-2 activates p70 S6 kinase in osteoblast-like MC3T3-E1 cells, we examined the effect of FGF-2 on the phosphorylation of p70 S6 kinase. The stimulation of FGF-2 time-dependently induced the phosphorylation of p70 S6 kinase (Fig. 1). The maximum effect was observed at 20 minutes after the stimulation of FGF-2.

#### Effect of rapamycin on the FGF-2-stimulated VEGF release in MC3T3-E1 cells

To clarify whether p70 S6 kinase is involved in the FGF-2-induced release of VEGF in MC3T3-E1 cells, we examined the effect of rapamycin, a specific inhibitor of p70 S6



**FIG. 4.** Effects of rapamycin on the FGF-2-induced phosphorylation of p44/p42 MAP kinase or p38 MAP kinase in MC3T3-E1 cells. (A) The cultured cells were pretreated with various doses of rapamycin for 60 minutes and stimulated by 30 ng/ml FGF-2 or vehicle for 20 minutes. The extracts of cells were subjected to SDS-PAGE with subsequent Western blotting analysis with antibodies against phospho-specific p44/p42 MAP kinase or p44/p42 MAP kinase. (B) The cultured cells were pretreated with various doses of rapamycin for 60 minutes and stimulated by 30 ng/ml FGF-2 or vehicle for 10 minutes. The extracts of cells were subjected to SDS-PAGE with subsequent Western blotting analysis with antibodies against phospho-specific p38 MAP kinase or p38 MAP kinase. The histogram shows quantitative representations of the levels of FGF-2-induced phosphorylation obtained from laser densitometric analysis of three independent experiments. Each value represents the mean  $\pm$  SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations.

kinase,<sup>(29,30)</sup> on the FGF-2-induced release of VEGF. Rapamycin, which alone did not affect the basal levels of VEGF, significantly enhanced the FGF-2-induced release of VEGF in a time-dependent manner (Fig. 2A). The amplifying effect of rapamycin was dose dependent in a range between 1 and 50 ng/ml (Fig. 2B). Rapamycin at 50 ng/ml caused ~200% enhancement in the FGF-2 effect.

#### *Effect of rapamycin on the FGF-2-stimulated VEGF mRNA expression in MC3T3-E1 cells*

To clarify whether the enhancement of FGF-2-stimulated VEGF release by rapamycin is mediated through transcriptional events, we examined the effect of rapamycin on the FGF-2-induced VEGF mRNA expres-

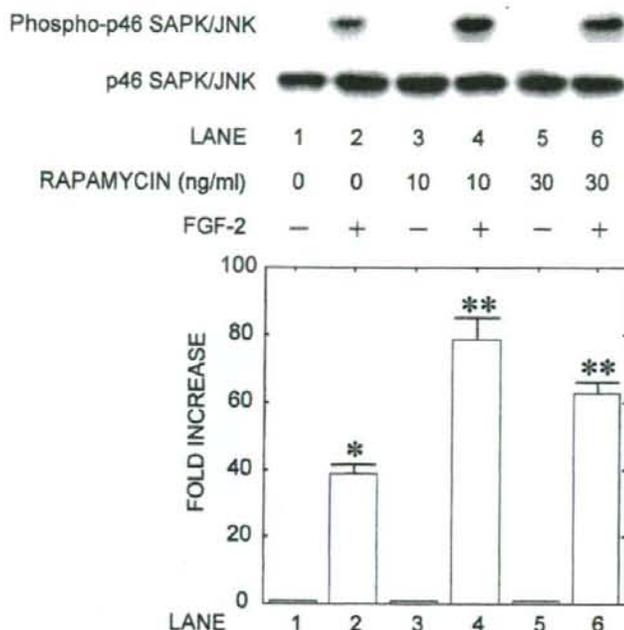


FIG. 5. Effect of rapamycin on the FGF-2-induced phosphorylation of SAPK/JNK in MC3T3-E1 cells. The cultured cells were pretreated with various doses of rapamycin for 60 minutes and stimulated by 30 ng/ml FGF-2 or vehicle for 20 minutes. The extracts of cells were subjected to SDS-PAGE with subsequent Western blotting analysis with antibodies against phospho-specific SAPK/JNK or SAPK/JNK. The histogram shows quantitative representations of the levels of FGF-2-induced phosphorylation obtained from laser densitometric analysis of three independent experiments. Each value represents the mean  $\pm$  SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \* $p < 0.05$  compared with the control. \*\* $p < 0.05$  compared with the value of FGF-2 alone.

TABLE 1. EFFECT OF SP600125 ON THE ENHANCEMENT BY RAPAMYCIN OF THE FGF-2-STIMULATED VEGF RELEASE IN MC3T3-E1 CELLS

SP600125	Rapamycin	FGF-2	VEGF (pg/ml)
-	-	-	<7.8
-	-	+	523 $\pm$ 40*
-	+	-	<7.8
-	+	+	1586 $\pm$ 121†
+	-	-	<7.8
+	-	+	111 $\pm$ 12†
±	±	-	<7.8
±	±	±	120 $\pm$ 15‡

The cultured cells were pretreated with 50  $\mu$ M SP600125 or vehicle for 60 minutes and incubated with 50 ng/ml rapamycin or vehicle for 60 minutes. The cells were stimulated by 70 ng/ml FGF-2 or vehicle for 24 h thereafter. Each value represents the mean  $\pm$  SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations.

\*  $p < 0.05$  compared with the control.

†  $p < 0.05$  compared with the value of FGF-2 alone.

‡  $p < 0.05$  compared with the value of FGF-2 with rapamycin pretreatment.

sion by real-time PCR. We found that rapamycin significantly upregulated the FGF-2-induced VEGF mRNA expression 6 h after stimulation (3.5  $\pm$  0.3-fold for 70 ng/ml FGF-2 alone compared with the control; 5.2  $\pm$  0.3-fold for 70 ng/ml FGF-2 with 50 ng/ml rapamycin compared the control). These results suggest that the amplifying effect of rapamycin is mediated, at least in part, by upregulation of VEGF mRNA expression in osteoblast-like MC3T3-E1 cells.

#### Effects of rapamycin on the FGF-2-induced phosphorylation of p70 S6 kinase in MC3T3-E1 cells

We next examined the effect of rapamycin on the FGF-2-induced phosphorylation of p70 S6 kinase. Rapamycin almost completely attenuated the FGF-2-induced phosphorylation of p70 S6 kinase in a range between 10 and 50 ng/ml (Fig. 3).

#### Effects of rapamycin on the FGF-2-induced phosphorylation of p44/p42 MAP kinase, p38 MAP kinase, or SAPK/JNK in MC3T3-E1 cells

To study whether the amplifying effect of rapamycin on the FGF-2-stimulated VEGF release was through activation of p44/p42 MAP kinase, p38 MAP kinase, or SAPK/JNK in MC3T3-E1 cells, we examined the effect of rapamycin on the phosphorylation of p44/p42 MAP kinase, p38 MAP kinase, or SAPK/JNK induced by FGF-2. Rapamycin failed to affect the FGF-2-induced phosphorylation of p44/p42 MAP kinase (Fig. 4A) or p38 MAP kinase (Fig. 4B). However, the FGF-2-induced phosphorylation of SAPK/JNK was markedly enhanced by rapamycin (Fig. 5). Rapamycin at 10 ng/ml caused ~100% enhancement in the FGF-2 effect.

#### Effects of SP600125 on the amplification by rapamycin of the FGF-2-induced VEGF release and phosphorylation of SAPK/JNK in MC3T3-E1 cells

SP600125, a specific SAPK/JNK inhibitor,<sup>(31)</sup> which by itself did not affect the basal levels of VEGF, significantly

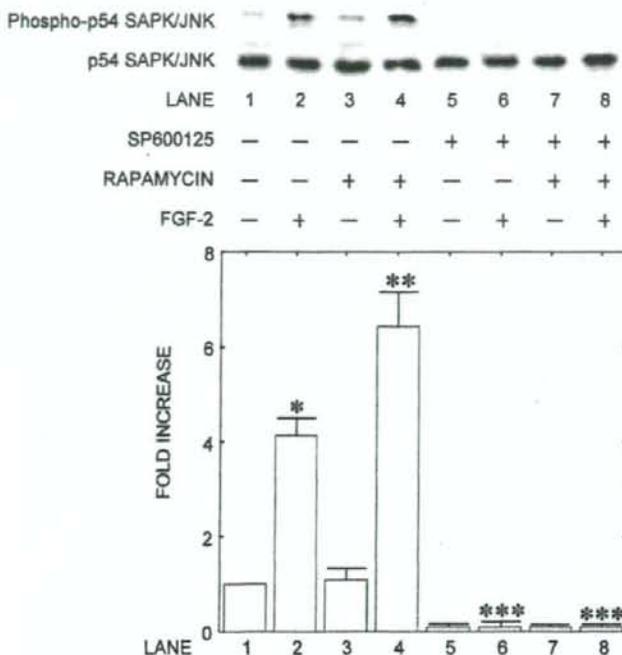


FIG. 6. Effect of SP600125 on the enhancement by rapamycin of the FGF-2-induced phosphorylation of SAPK/JNK in MC3T3-E1 cells. The cultured cells were pretreated with 50  $\mu$ M SP600125 or vehicle for 60 minutes and incubated by 30 ng/ml rapamycin or vehicle for 60 minutes. The cells were stimulated by 30 ng/ml FGF-2 or vehicle for 20 minutes. The extracts of cells were subjected to SDS-PAGE with subsequent Western blotting analysis with antibodies against phospho-specific SAPK/JNK or SAPK/JNK. The histogram shows quantitative representations of the levels of FGF-2-induced phosphorylation obtained from laser densitometric analysis of three independent experiments. Each value represents the mean  $\pm$  SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \* $p$  < 0.05 compared with the control. \*\* $p$  < 0.05 compared with the value of FGF-2 alone. \*\*\* $p$  < 0.05 compared with the value of FGF-2 with rapamycin pretreatment.

reduced the enhancement by rapamycin of FGF-2-induced VEGF release (Table 1). The enhanced levels by rapamycin of FGF-2-induced VEGF release were reduced by SP600125 similar to the levels by FGF-2 with SP600125 treatment. In addition, SP600125 almost completely suppressed both the FGF-2-induced phosphorylation of SAPK/JNK and its enhancement by rapamycin (Fig. 6).

#### Effects of SP600125 on the upregulation by rapamycin of the FGF-2-stimulated VEGF mRNA expression in MC3T3-E1 cells

To clarify whether the suppressive effect by SP600125 of FGF-2-stimulated VEGF release is mediated through transcriptional events, we examined the effect of SP600125 in the presence or absence of rapamycin on the FGF-2-stimulated VEGF mRNA expression by real-time PCR. We found that SP600125 significantly downregulated the FGF-2-induced VEGF mRNA expression 2 h after stimulation ( $5.0 \pm 0.9$ -fold for 70 ng/ml FGF-2 alone compared with the control;  $3.2 \pm 0.4$ -fold for 70 ng/ml FGF-2 with 10  $\mu$ M SP600125 compared with the control;  $3.0 \pm 0.3$ -fold for 70 ng/ml FGF-2 with 10  $\mu$ M SP600125 and 50 ng/ml rapamycin compared with the control). These results suggest that the suppressive effect of SP600125 is mediated, at least in part, by downregulation of VEGF mRNA expression in osteoblast-like MC3T3-E1 cells.

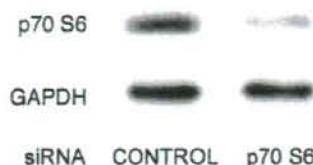
#### Effect of p70 S6 kinase-siRNA on the FGF-2-stimulated VEGF release in MC3T3-E1 cells

To confirm the results from rapamycin, we examined the effects of p70 S6 kinase downregulation by p70 S6 kinase siRNA on the VEGF release and the phosphorylation of SAPK/JNK induced by FGF-2 in osteoblast-like MC3T3-E1 cells. We found that p70 S6 kinase siRNA (250 nM) successfully reduced the p70 S6 kinase levels compared with those of control siRNA determined by the band intensity of Western blot analysis (Fig. 7A). The FGF-2-induced levels of VEGF release in p70 S6 kinase-downregulated cells were truly enhanced compared with those in control siRNA-transfected cells. Downregulation of p70 S6 kinase caused ~180% enhancement in the FGF-2 effect (Table 2). FGF-2 significantly enhanced the phosphorylation levels of SAPK/JNK in the p70 S6 kinase-downregulated cells (Fig. 7B).

## DISCUSSION

In this study, we found that FGF-2 time-dependently induced the phosphorylation of p70 S6 kinase in osteoblast-like MC3T3-E1 cells, using phospho-specific p70 S6 kinase (Thr389) antibodies. It is generally recognized that the activity of p70 S6 kinase is regulated by multiple phosphorylation events.<sup>(20)</sup> It has been shown that phosphorylation at Thr389 among the phosphorylation most strongly correlates with p70 S6 kinase activity.<sup>(20)</sup> Thus, it is probable that

A



B

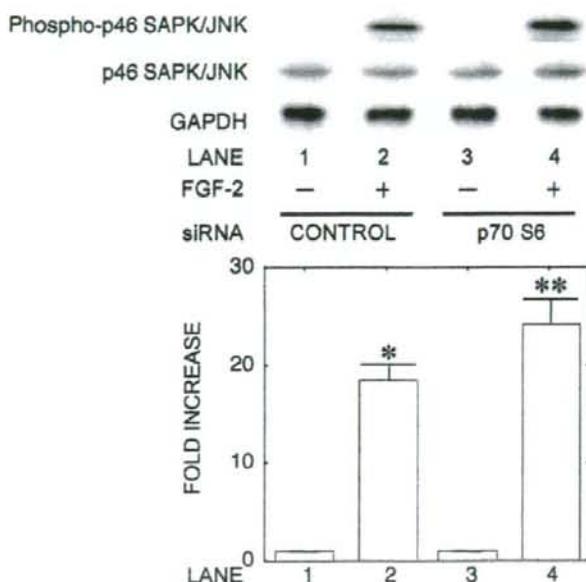


FIG. 7. Effect of p70 S6 kinase-siRNA in MC3T3-E1 cells and effect of p70 S6 kinase downregulation on the FGF-2-induced-phosphorylation of SAPK/JNK in MC3T3-E1 cells. The cells were transfected with 250 nM of control siRNA or p70 S6 kinase siRNA by using the siLentFect. (A) Cells were incubated at 37°C for 48 h with siRNA-siLentFect complexes and subsequently harvested for preparation of Western blot analysis. (B) The cultured cells transfected with control siRNA or p70 S6 kinase siRNA were stimulated by 70 ng/ml FGF-2 or vehicle for 20 minutes and subsequently harvested for preparation of Western blot analysis. The histogram shows quantitative representations of the levels of FGF-2-induced phosphorylation obtained from laser densitometric analysis of three independent experiments. Each value represents the mean  $\pm$  SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations. \* $p < 0.05$  compared with the value of vehicle. \*\* $p < 0.05$  compared with the value of FGF-2 with control siRNA transfection.

FGF-2 induces the activation of p70 S6 kinase in osteoblast-like MC3T3-E1 cells. To the best of our knowledge, this is probably the first report showing the FGF-2-induced p70 S6 kinase activation in osteoblasts.

We have previously reported that FGF-2 stimulates the release of VEGF in osteoblast-like MC3T3-E1 cells.<sup>(14)</sup> Therefore, we studied whether p70 S6 kinase functions in FGF-2-stimulated VEGF release in osteoblast-like MC3T3-E1 cells. Rapamycin, a specific inhibitor of p70 S6 kinase,<sup>(29,30)</sup> markedly enhanced the FGF-2-stimulated release of VEGF and expression of VEGF mRNA in MC3T3-E1 cells. We confirmed that the FGF-2-induced phosphorylation of p70 S6 kinase was truly attenuated by rapamycin. In addition, downregulation of p70 S6 kinase by siRNA markedly enhanced FGF-2-stimulated VEGF release in these cells. These results strongly suggest that FGF-2-stimulated VEGF release is reduced by the activation of

p70 S6 kinase. Therefore, it is probable that FGF-2 activates the p70 S6 kinase pathway, resulting in negatively regulating the release of VEGF. We speculate that p70 S6 kinase signaling activated by FGF-2 limits the FGF-2-induced over-release of VEGF in osteoblast-like MC3T3-E1 cells. As far as we know, this finding is the first report to show that the activation of p70 S6 kinase leads to negative feedback regulation of VEGF release in osteoblasts.

We have previously shown that FGF-2 induces the phosphorylation of p44/p42 MAP kinase, p38 MAP kinase, and SAPK/JNK in MC3T3-E1 cells.<sup>(16,17)</sup> It is generally recognized that the MAP kinase superfamily mediates intracellular signaling of extracellular agonists and plays a crucial role in cellular functions including proliferation, differentiation, and apoptosis in a variety of cells.<sup>(18)</sup> Three major MAP kinases, p44/p42 MAP kinase, p38 MAP kinase, and SAPK/JNK, are known as central elements used by mam-

TABLE 2. EFFECT OF p70 S6 KINASE (S6K) siRNA ON THE FGF-2-STIMULATED VEGF RELEASE IN MC3T3-E1 CELLS

siRNA	FGF-2	Time (h)	VEGF (pg/ml)
Control	-	4	<7.8
Control	+	4	244 ± 32*
S6K	-	4	<7.8
S6K	+	4	558 ± 73 <sup>†</sup>
Control	-	9	<7.8
Control	+	9	681 ± 106*
S6K	-	9	<7.8
S6K	+	9	1269 ± 226 <sup>†</sup>

The cultured cells were transfected with control siRNA or p70 S6 kinase siRNA by using the siLentFect according to the manufacturer's protocol. The cells were stimulated by 70 ng/ml FGF-2 or vehicle for the indicated time periods. Each value represents the mean ± SE of triplicate determinations. Similar results were obtained with two additional and different cell preparations.

\*  $p < 0.05$  compared with the value of vehicle.

<sup>†</sup>  $p < 0.05$  compared with the value of FGF-2 with control siRNA transfection.

malian cells to transduce the diverse messages.<sup>(18)</sup> In our previous studies,<sup>(16,17)</sup> we showed that p44/p42 MAP kinase and SAPK/JNK act as positive regulators in FGF-2-induced VEGF release. On the other hand, FGF-2-activated p38 MAP kinase negatively regulates VEGF release. These findings lead us to speculate that there is cross-talk regulation between p70 S6 kinase and these MAP kinases in FGF-2-stimulated VEGF release in these cells. However, rapamycin<sup>(29,30)</sup> failed to affect the phosphorylation of p44/p42 MAP kinase and p38 MAP kinase. Therefore, it seems unlikely that the p70 S6 kinase signaling pathway affects FGF-2-stimulated release of VEGF through the amplification of activities of p44/p42 MAP kinase and p38 MAP kinase in osteoblast-like MC3T3-E1 cells. On the contrary, we showed that the phosphorylation levels of FGF-2-induced SAPK/JNK were strengthened by rapamycin and p70 S6 kinase siRNA transfection. In addition, the amplification by rapamycin of the FGF-2-stimulated VEGF release and mRNA expression was suppressed by SP600125, a specific inhibitor of SAPK/JNK,<sup>(31)</sup> similar to the levels of FGF-2 with SP600125. Furthermore, we showed that SP600125 almost completely reduced the rapamycin-enhanced phosphorylation of SAPK/JNK and FGF-2-induced phosphorylation of SAPK/JNK. Taking our findings into account, it is probable that the p70 S6 kinase signal pathway affects the FGF-2-stimulated VEGF release through upregulation of SAPK/JNK in osteoblast-like MC3T3-E1 cells.

The p70 S6 kinase pathway is recognized to play a crucial role in various cellular functions, especially cell cycle progression.<sup>(20)</sup> These results indicate that the p70 S6 kinase pathway in osteoblasts has an important role in the control of the production of VEGF, one of the key regulators of bone metabolism. Bone remodeling carried out by osteoclasts and osteoblasts is accompanied by angiogenesis and capillary outgrowth.<sup>(32)</sup> Because VEGF is a specific mitogen of vascular endothelial cells,<sup>(11)</sup> these results lead us to speculate that FGF-2-activated p70 S6 kinase signaling acts as a negative regulator of the microvasculature development in bone. Thus, the p70 S6 kinase pathway in osteo-

blasts might be considered to be a new candidate as a molecular target of bone resorption concurrent with various bone diseases. On the contrary, we have previously shown that p70 S6 kinase acts as a positive regulator in bone morphogenetic protein-4-stimulated release of VEGF in MC3T3-E1 cells.<sup>(22)</sup> The physiological significance of a regulatory mechanism by p70 S6 kinase in osteoblasts still remains unclear. Further study is required to clarify the exact roles of p70 S6 kinase in osteoblasts and bone metabolism.

In conclusion, these results strongly suggest that p70 S6 kinase plays an important role in the regulation of FGF-2-stimulated VEGF release in osteoblasts and may serve as a negative feedback mechanism to prevent oversynthesizing of VEGF through SAPK/JNK in these cells.

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#### REFERENCES

- Baylink DJ, Finkleman RD, Mohan S 1993 Growth factor to stimulate bone formation. *J Bone Miner Res* 8:S565-S572.
- Hurley MM, Ahreu C, Harrison JR, Lichtler AC, Raisz LG, Kream BE 1993 Basic fibroblast growth factor inhibits type I collagen gene expression in osteoblastic MC3T3-E1 cells. *J Biol Chem* 268:5588-5593.
- Bolander ME 1992 Regulation of fracture repair by growth factors stimulate tyrosine kinase activity *in vivo*. *Proc Soc Exp Biol Med* 200:165-170.
- Nijweide PJ, Burger EH, Feyen JHM 1986 Cells of bone: Proliferation, differentiation, and hormonal regulation. *Physiol Rev* 66:855-886.
- Mari PJ 2003 Fibroblast growth factor signaling controlling osteoblast differentiation. *Gene* 316:23-32.
- Suzuki A, Shinoda J, Kanda S, Oiso Y, Kozawa O 1996 Basic fibroblast growth factor stimulates phosphatidylcholine-hydrolyzing phospholipase D in osteoblast-like cells. *J Cell Biochem* 63:491-499.
- Kozawa O, Niwa M, Matsuno H, Ishisaki A, Kato K, Uematsu T 2001 Stimulatory effect of basic fibroblast growth factor on induction of heat shock protein 27 in osteoblasts: Role of protein kinase C. *Arch Biochem Biophys* 388:237-242.
- Brighton CT, Hunt RM 1991 Early histological and ultrastructural changes in medullary fracture callus. *J Bone Joint Surg Am* 73:832-847.
- Parfitt AM 1994 Osteonal and hemi-osteonal remodeling: The spatial and temporal framework for signal traffic in adult human bone. *J Cell Biochem* 55:273-286.
- Erlebacher A, Filvaroff EH, Gitelman SE, Derynck R 1995 Toward a molecular understanding of skeletal development. *Cell* 80:371-378.
- Ferrara N, Davis-Smyth T 1997 The biology of vascular endothelial growth factor. *Endocr Rev* 18:4-25.
- Gerber HP, Vu TH, Ryan AM, Kowalski J, Werb Z, Ferrara N 1999 VEGF couples hypertrophic cartilage remodeling, ossification and angiogenesis during endochondral bone formation. *Nat Med* 5:623-628.
- Goad DL, Rubin J, Wang H, Tashjian AH Jr, Patterson C 1996 Enhanced expression of vascular endothelial growth fac-

- tor in human SaOS-2 osteoblast-like cells and murine osteoblasts induced by insulin-like growth factor I. *Endocrinology* **137**:2262-2268.
14. Wang DS, Yamazaki K, Nohtomi K, Shizume K, Ohsumi K, Shibuya M, Demura H, Sato K 1996 Increase of vascular endothelial growth factor mRNA expression by 1,25-dihydroxyvitamin D<sub>3</sub> in human osteoblast-like cells. *J Bone Miner Res* **11**:472-479.
  15. Schalaepi JM, Gutzwiller S, Finlenzeller G, Fournier B 1997 1,25-dihydroxyvitamin D<sub>3</sub> induces the expression of vascular endothelial growth factor in osteoblastic cells. *Endocr Res* **23**:213-229.
  16. Tokuda H, Kozawa O, Uematsu T 2000 Basic fibroblast growth factor stimulates vascular endothelial growth factor release in osteoblasts: Divergent regulation by p44/p42 mitogen-activated protein kinase and p38 mitogen-activated protein kinase. *J Bone Miner Res* **15**:2371-2379.
  17. Tokuda H, Hirade K, Wang X, Oiso Y, Kozawa O 2003 Involvement of SAPK/JNK in basic fibroblast growth factor-induced VEGF release in osteoblasts. *J Endocrinol* **177**:101-107.
  18. Widmann C, Gibson S, Jarpe MB, Johnson GL 1999 Mitogen-activated protein kinase: Conservation of a three-kinase module from yeast to human. *Physiol Rev* **79**:143-180.
  19. Zelzer E, Olsen BR 2005 Multiple roles of vascular endothelial growth factor (VEGF) in skeletal development, growth, and repair. *Curr Top Dev Biol* **65**:169-187.
  20. Pullen N, Thomas G 1997 The modular phosphorylation and activation of p70s6k. *FEBS Lett* **410**:78-82.
  21. Susa M, Standke GJ, Jeschke M, Rohner D 1997 Fluoroaluminate induces pertussis toxin-sensitive protein phosphorylation: differences in MC3T3-E1 osteoblastic and NIH3T3 fibroblastic cells. *Biochem Biophys Res Commun* **235**:680-684.
  22. Kozawa O, Matsuno H, Uematsu T 2001 Involvement of p70 S6 kinase in bone morphogenetic protein signaling: Vascular endothelial growth factor release by bone morphogenetic protein-4 in osteoblasts. *J Cell Biochem* **81**:430-436.
  23. Tokuda H, Hatakeyama D, Shibata T, Akamatsu S, Oiso Y, Kozawa O 2003 p38 MAP kinase regulates BMP-4-stimulated VEGF release via p70 S6 kinase in osteoblasts. *Am J Physiol Endocrinol Metab* **284**:E1202-E1209.
  24. Sudo H, Kodama H, Amagai Y, Yamamoto S, Kasai S 1983 In vitro differentiation and calcification in a new clonal osteogenic cell line derived from newborn mouse calvaria. *J Cell Biol* **96**:191-198.
  25. Kozawa O, Tokuda H, Miwa M, Kotoyori J, Oiso Y 1992 Cross-talk regulation between cyclic AMP production and phosphoinositide hydrolysis induced by prostaglandin E<sub>2</sub> in osteoblast-like cells. *Exp Cell Res* **198**:130-134.
  26. Laemmli LK 1970 Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* **227**:680-685.
  27. Kato K, Ito H, Hasegawa K, Inaguma Y, Kozawa O, Asano T 1996 Modulation of the stress-induced synthesis of hsp27 and alpha B-crystallin by cyclic AMP in C6 rat glioma cells. *J Neurochem* **66**:946-950.
  28. Simpson DA, Freney S, Boyle C, Stitt AW 2000 Retinal VEGF mRNA measured by SYBR Green I fluorescence: A versatile approach to quantitative PCR. *Mol Vis* **6**:178-183.
  29. Price DJ, Grove JR, Calvo V, Avruch J, Bierer BE 1992 Rapamycin-induced inhibition of the 70-kilodalton S6 protein kinase. *Science* **257**:973-977.
  30. Kuo CJ, Chung J, Fiorentino DF, Flanagan WM, Blenis J, Crabtree GR 1992 Rapamycin selectively inhibits interleukin-2 activation of p70 S6 kinase. *Nature* **358**:70-73.
  31. Bennett BL, Sasaki DT, Murray BW, O'Leary EC, Sakata ST, Xu W, Leisten JC, Motiwala A, Pierce S, Satoh Y, Bhagwat SS, Manning AM, Anderson DW 2001 SP600125, an anthracycline inhibitor of Jun N-terminal Kinase. *Proc Natl Acad Sci USA* **98**:13681-13686.

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特 集 運動器不安定症と高齢者の転倒  
—その概念と今後の展開

運動器不安定症と今後の展開

原 田 敦\*

要旨：運動器不安定症は運動機能を原因とする転倒危険因子の総称と考えられ、その診療が充実して転倒危険因子の評価と可逆的因子への介入が展開できれば、高齢者の自立を支える大きな基盤となる。それによって転倒そのものが予防されることは過去のエビデンスからみて確実性が高い。最終目標は、骨折、なかでも大腿骨頸部/転子部骨折の予防実現で、そのエビデンスは現状では骨粗鬆症薬剤やヒッププロテクターに限られており、転倒予防プログラムなど運動器不安定症診療による骨折予防が今後の課題である。さらに医療経済的妥当性が証明されることが最終的には必要と思われる。

はじめに

高齢期を迎えると成人期のある時点から潜在的に進行していた筋量や骨量の減少がいつそう進んで顕在化すると筋肉減少症(sarcopenia)や骨量減少(osteopenia)に陥る。その結果として、運動の量と質は徐々に低下して行動範囲は狭まり、骨折や脳卒中などの特別な急性疾患発症がなくとも次第に“動けない”状態に近づく。この過程で日常生活動作(activity of daily life; ADL)のレベルは、スポーツや社交などの拡大日常生活動作(advanced ADL; AADL)から交通機関の利用、買い物、料理、家事、電話などの手段的ADL(instrumental ADL; IADL)、食事、排尿・排便、入浴、歩行、起立、寝返りなどの基本的ADL(basic ADL; BADL)へと低下していく。手段的ADLがこなせなくなる頃から介護を要する状態とみなされ、基本的ADLも自立できない状態となると、準

寝たきり・寝たきり(日常生活自立でランクA~C)である。

このような一連の運動の量と質の低下は、加齢とともに進み、それだけで独立して高齢期の転倒リスクは上昇するとされる。しかしながら、そこに運動機能に悪影響を及ぼす運動器疾患や神経疾患、一過性意識障害をもたらすような循環器疾患、目を通じた補正を妨げるような視覚障害、転倒を誘発させる薬剤使用が重くなると、内的因子による転倒リスクはいつそう高くなり、さらに段差や不適な照明などの不良な物的環境が外的因子として転倒リスクを独立して上昇させるという構図が考えられている(図1)<sup>1)</sup>。

さて、運動器不安定症は、運動機能の低下をきたす疾患を基礎として、「高齢化により、バランス能力および移動歩行能力の低下が生じ、閉じこもり、転倒リスクが高まった状態」という定義とされている。すなわち、運動器不安定症は、前述したような転倒の多種多様な病因のうち、以前は易転倒性などと呼ばれていた運動器疾患が原因で運動機能が低下する病態を新しくまとめた概念と考えられる。つまり、転倒の原因として代表的な症

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Unstable locomotorium disease and its future  
Key words : Fall, Fracture, Prevention