

Novel Cardiac Precursor-Like Cells from Human Menstrual Blood-Derived Mesenchymal Cells

Naoko Hida, Nobuhiro Nishiyama, Shunichiro Miyoshi, Shinichiro Kira, Kaoru Segawa, Taro Uyama, Taisuke Mori, Kenji Miyado, Yukinori Ikegami, ChangHao Cui, Tohru Kiyono, Satoru Kyo, Tatsuya Shimizu, Teruo Okano, Michiie Sakamoto, Satoshi Ogawa and Akihiro Umezawa

Stem Cells 2008;26:1695-1704; originally published online Apr 17, 2008;

DOI: 10.1634/stemcells.2007-0826

This information is current as of August 19, 2008

**Updated Information
& Services**

including high-resolution figures, can be found at:
<http://www.StemCells.com/cgi/content/full/26/7/1695>

Supplementary Material

Supplementary material can be found at:
<http://www.StemCells.com/cgi/content/full/2007-0826/DC1>

 **AlphaMed Press**

日程	班	氏名	テーマ	プレゼンテーション(60)	レポート(15)	質疑(10)	出席(15)	質問(5)	時間	合計
		青山剛三		58		5	15	5		83
		阿部舞美		60	15	7	15	5		102
		池水有里		57	15	7	15	5		99
		上村悦央		58	15	8	15	5		101
		浦上信太郎		60	15	10	15	5		105
		岡村真弥		60	15	10	15	5		105
		英村美帆		60	15	10	15	5		105
		川口明日香		60	15	10	15	5		105
		川口佳保理		55	15	10	15	5		100
		川崎友美		55	15	7	15	5		89
		黒田京子		58	15	10	15	5		103
		後藤祐美		60	15	7	15	5		102
		小林友幸		60	15	8	15	5		103
		酒井博生		60	15	7	15	5		99
		清水友理		57	15	8	15	5		101
		社領美紀		58	15	8	15	5		101
		須河内昭成		60	15	10	15	5		105
		千田百合絵		60	15	10	15	5		105
		西原千尋		60	15	8	15	5		88
				50	15	5	15	5		90

- A 菌周疾患と骨
 B 破骨細胞と骨吸収の調節メカニズム
 C 遺伝子と菌周疾患
 D 骨芽細胞と骨形成の調節メカニズム
 E 骨リモデリング
 F 遺伝子と歯科疾患
 G 菌と菌周組織の再生について
 H 細胞外マトリックス

Establishment of an immortalized human extravillous trophoblast cell line by retroviral infection of E6/E7/hTERT and its transcriptional profile during hypoxia and reoxygenation

HIROKO OMI¹, AIKOU OKAMOTO^{1,2}, TAKASHI NIKAI³, MITSUYOSHI URASHIMA⁴,
RIE KAWAGUCHI¹, NAGAYOSHI UMEHARA¹, KENTARO SUGIURA¹,
MISATO SAITO¹, TOHRU KIYONO⁵ and TADAO TANAKA¹

Departments of ¹Obstetrics and Gynecology, ²Gene Therapy, Institute of DNA Medicine,
³Department of Pathology, ⁴Division of Clinical Research and Development, The Jikei University,
School of Medicine, 3-25-8 Nishishinbashi, Minato-ku, Tokyo 105-8461; ⁵Virology Division,
National Cancer Center Research Institute, 5-1-1 Tsukiji, Chuo-ku, Tokyo 104-045, Japan

Received September 26, 2008; Accepted October 29, 2008

DOI: 10.3892/ijmm_00000121

Abstract. Investigation into the function of human trophoblasts has been largely restricted by a lack of suitable cell models. We aimed to produce normal human trophoblast cell lines with a long lifespan and to provide an ideal *in vitro* cell model. Primary human trophoblast cells were derived from a placenta that had undergone elective abortion at the 7th week of gestation. The cells were immortalized by infection with retroviral expression vectors containing the type 16 human papillomaviruses E6 and E7 in combination with human telomerase reverse transcriptase (hTERT). Characterization of the cell line was performed by immunocytochemistry using a panel of antibodies, Western blotting, real-time RT-PCR, an invasion assay, gelatin zymography, karyotype analysis and a nude mouse assay. Gene expression profiles under hypoxia (1% O₂, 1 h) and subsequent reoxygenation (20% O₂, 6 h) were analyzed using cDNA microarray. Immunocytochemistry revealed an extravillous trophoblastic phenotype by positive staining for hCG β , cytokeratin 7, HLA-G and CD9. A transwell insert invasion assay showed the invasiveness of this cell line and gelatin zymography detected the secretion of MMP-2 and MMP-9. Karyotype analysis exhibited an almost normal chromosomal number which ranged from 46 to 48 and the cells showed no tumorigenicity in a nude mouse assay. Forty-three genes showing reversible up- or down-regulation during hypoxia were detected using an oligonucleotide array.

This newly immortalized cell line, HChEpC1b, is a useful model for the study of extravillous trophoblast function.

Introduction

In humans, cells outside the morula form the trophoblast and differentiate into trophoblasts. These cells form the fetal compartment of the placenta during pregnancy. After the initial phase of implantation, human trophoblasts differentiate along either the villous or the extravillous trophoblast pathway. Multinucleate syncytiotrophoblasts, which form the epithelial layer of the villi by cell fusion, are involved in the exchange of gas and the nutrients between the mother and the fetus. Mononuclear extravillous cytotrophoblasts invade deep into the decidua, the myometrium and the uterine spiral arteries (1). Clinically, many pregnancy-associated conditions result from abnormal functioning of trophoblasts, such as abortion, intrauterine growth retardation and pre-eclampsia (2).

Investigation into the function of human trophoblasts has been largely restricted by the short life span of primary cultured trophoblasts *in vitro*. There are two major mechanisms that cause the limited life span of primary cultured cells. One is the telomere-based replication senescence (3,4) and the other is telomere-independent senescence, which is thought to be controlled by the Rb/p16 (5) and p53 pathways (6). Since HPV16 E6 and E7 are known to inhibit p53 and Rb function, respectively, we introduced E6/E7 and human telomerase reverse transcriptase (hTERT) into primary human trophoblast cells to immortalize them.

We obtained one cell line, HChEpC1b, that retained an extravillous phenotype and acquired immortality. This cell line expressed molecular markers for human extravillous trophoblasts and it demonstrated invasiveness and metalloproteinase production; nevertheless, it did not show tumorigenic potency. There have been a number of trophoblast cell lines that have gained a long life span spontaneously (7-12), or by transduction of the SV40 T antigen (7,13-17), HPV16 E6/E7 (18,19) and hTERT (20). However, this is the

Correspondence to: Dr Tadao Tanaka, Department of Obstetrics and Gynecology, The Jikei University, School of Medicine, 3-25-8 Nishishinbashi, Minato-ku, Tokyo 105-8461, Japan
E-mail: tanaka3520@jikei.ac.jp

Key words: trophoblast, immortalization, human papillomavirus, E6, E7, human telomerase reverse transcriptase, hypoxia, oligonucleotide microarray

first study of an immortalized trophoblast cell line produced by retroviral infection of HPV16 E6/E7 in combination with hTERT and this newly established cell line may provide a useful model for the study of extravillous trophoblast function as well as that of other cell lines.

Materials and methods

Preparation of human trophoblast primary cultures. A human placenta was obtained from a patient that had undergone a legal termination of a normal pregnancy at 7 weeks of gestation with informed consent. This study was approved by the Ethics Committee of the Jikei University. After removal of all attached decidua and blood clots, chorionic villi were gently shaken in normal saline solution. The fallen cells and tissue fragments were collected and suspended in RPMI-1640 (Sigma-Aldrich, Tokyo, Japan) supplemented with 10% fetal bovine serum (Invitrogen Corp., Carlsbad, CA, USA), 50 IU/ml penicillin, 50 µg/ml streptomycin, and 100 mg/ml neomycin (Invitrogen Corp) and were cultured in a collagen I coated culture dish (Asahi Techno Glass, Tokyo, Japan). Two weeks later, colonies were collected using cloning rings with 0.05% trypsin and 0.53 mM EDTA (Invitrogen Corp) and were transferred to a 24-well culture dish. When the colonies became confluent, the cells were transferred to a 6-well culture dish. Three clones with epithelioid phenotypes [human chorionic epithelium cell-1 (HChEpC1) a-c] were selected for the next immortalization step.

Vector construction and retroviral transduction of E6/E7 and hTERT. Construction of retroviral vector plasmids, pCMSCV puro-hTERT, pCLXSN-16E6E7, has been described previously (21). Production of recombinant retroviruses was carried out, as previously described (22). Briefly, a retroviral vector and packaging construct, pCL-10A1, was co-transduced into 293T cells using TransIT-293 (Mirus Bio Corp., Madison, WI, USA) according to the manufacturer's instructions. The culture fluid was harvested at 48 to 72 h post-transduction. The titer of the recombinant viruses was $>1 \times 10^5$ drug-resistant colony-forming U/ml on HeLa cells. HChEpC1 cells seeded on 24-well dishes were inoculated with a 0.5 ml aliquot of the culture fluid in the presence of polybrene (4 µg/ml). Following inoculation with viruses, the cells were grown without drug selection as mock-infected cells and stopped growing within two weeks. To achieve different combinations of retroviral infections, the cells were serially infected with MSCVpuro-hTERT and LXSN-16E6E7 at passage 2.

Cell culture. The infected HChEpC1 cells were cultured in RPMI-1640 supplemented with 10% FBS, 50 U/ml penicillin G, 50 µg/ml streptomycin, and 100 mg/ml neomycin and were incubated in 5% CO₂ air at 37°C. Serial passages were made in one to four splits. One clone, HChEpC1b, was selected for further characterization because it had the highest purity of trophoblast cells as proved by its uniformly positive staining for hCGβ. The population doubling time (DT) was calculated as follows: $DT = T \times \log_2 / \log (N_T / N_0)$, where N_0 is the initial number of cells and N_T is the number of cells harvested after T h culture.

The choriocarcinoma cell lines JAR and JEG-3 and the fibroblast cell line Hs795P1 were obtained from ATCC and propagated in RPMI-1640, MEM (Invitrogen Corp) and DMEM (Invitrogen Corp) respectively, supplemented with 10% FBS.

Karyotyping. Cells in the exponential phase were arrested by adding 0.03 µg/ml of colchicine for 6 h, before being treated with hypotonic solution (75 mM KCl, pH 8.0) and fixed twice in Carnoy's fixative (methanol/acetic acid). The G-band pattern was obtained using Giemsa staining and the chromosomes of 20 metaphases were analyzed.

Nude mouse assay. Single cell suspensions (10^7 cells) of HChEpC1b cells or JAR cells were subcutaneously inoculated into the right flank of 5 female mice (balb/c nu/nu) each. These mice were observed for up to 3 months and examined for tumor growth.

Immunocytochemistry. HChEpC1b cells were cultured on collagen I coated culture slides (BD Biosciences, Bedford, MA, USA) for two days, washed once with phosphate-buffered saline (PBS, pH 7.4) and fixed with 100% ethanol (room temperature) for 30 min. After three 5-min hydration periods in distilled water (room temperature) and two 10-min periods of washing in 0.1% Tween in PBS (4°C), the cells were incubated overnight at 4°C with the primary antibodies diluted in Antibody Diluent (Dako, Glostrup, Denmark) as described in Table I. The cells were washed three times with 0.1% Tween in PBS (4°C) and incubated with EnVision Labelled Polymer, Peroxidase (Dako) for 1 h at room temperature. After washing three times with 0.1% Tween in PBS (4°C), visualization was developed according to the manufacturer's instructions using diaminobenzidine (DAB) as the substrate. The cells were counterstained with hematoxylin, dehydrated and mounted for light microscopy.

Real-time reverse transcriptase polymerase chain reaction (real-time RT-PCR). Total RNA were isolated from subconfluent cultured cells using the RNeasy mini kit (Qiagen, Gmbh Hilden, Germany). Reverse transcription was performed according to the manufacturer's protocol (Takara RNA PCR kit, Takara, Shiga, Japan) with 1 µg of total RNA. For real-time PCR, each target was amplified on the same plate with GAPDH as the reference using the Taq Man Universal PCR master mix and the ABI PRISM 7700 sequence detection system (Applied Biosystems, Foster City, USA) and the relative mRNA amounts and range were determined. The primer/probe sets were purchased from Taq Man gene expression assays (Applied Biosystems, Assay IDs are listed in Table II). Relative mRNA levels were calculated by the comparative C_T method described in ABI User Bulletin 2.

Western blotting. Subconfluent HChEpC1b, JAR, JEG-3 and Hs795P1 cells were scraped from culture dishes, washed three times with cold PBS, and then incubated in the RIPA buffer [10 mM Tris-HCl (pH 7.5), 150 mM NaCl, 2 mM EDTA, 1% NP-40, 1% sodium deoxycholate, 0.1% SDS, complete mini (Roche, Basel, Switzerland)] for 30 min on ice. After centrifugation, the supernatants were subjected to Western blot

Table I. Antibodies used for immunocytochemistry and Western blotting.

Epitope	Clone name	Host	Isotype	Company	Cat. no.	Nation	Dilution
Cytokeratin 7	OVTL 12-30	Mouse	IgG1 κ	Acris Antibodies	DM057	Germany	1:50
HLA class I	W6/32HL	Mouse	IgG2a	Chemicon International	CBL139	USA	1:50
HLA-G	87G	Mouse	IgG2a	Exbio	10-437-C100	Czech	1:100
HLA-G	MEM-G/1	Mouse	IgG1	Serotec	MCA2043	UK	1:50
CD9	Polyclonal	Rabbit	IgG	Santa Cruz Biotechnology	sc-9148	USA	1:50
CD9	M-L13	Mouse	IgG1 κ	BD Biosciences	555370	USA	1:50
Vimentin	V9	Mouse	IgG1	Chemicon International	MAB3400	USA	1:200
E-cadherin	Polyclonal	Rabbit	IgG	Santa Cruz Biotechnology	sc-7870	USA	1:50
hCG β	Polyclonal	Rabbit	IgG	Dako	A0231	USA	1:50
hPL	Polyclonal	Rabbit	IgG	Lab Vision	RB-9067-P	UK	1:50
PLAP	8B6	Mouse	IgG2a	Chemicon International	CBL207	USA	1:50

Table II. Details of assay ID for real-time PCR.

Gene	Assay ID
Cytokeratin 7	Hs00818825_m1
HLA-G	Hs00918802_m1
CD9	Hs00233521_m1
HPL	Hs01862611_g1
PLAP	Hs01654626_s1
Vimentin	Hs00185584_m1
Integrin α -1	Hs00235006_m1
Integrin α -6	Hs00173952_m1
Integrin B-1	Hs00559595_m1
Integrin B-4	Hs00236216_m1

analysis. Total proteins (20 μ g) per sample was electrophoresed on a 4-20% SDS-polyacrylamide gel and transferred to a PDVF membrane. After washing in TBST (0.1% Tween in Tris-buffered saline), the membrane was incubated with primary antibodies (anti-cytokeratin 7, 1:500; anti-E-cadherin, 1:500; anti-vimentin, 1:1000) for 1 h at 37°C, before being washed in TBST and incubated with secondary HRP-conjugated goat anti mouse or rabbit IgG goat antibodies (Santa Cruz Biotechnology, Santa Cruz, CA, USA) for 1 h at 37°C. The antigen-antibody complexes were visualized using DAB.

Gelatin zymography. The gelatinolytic activities of matrix metalloproteinase-2 (MMP-2) and MMP-9 were assayed using gelatin zymography. Subconfluent cells were rinsed and cultured with serum-free medium for three days. The harvested medium was mixed 1:1 with 2X sample buffer [125 mM Tris-HCl (pH 6.8), 4% SDS, 20% glycerol] and was then applied to gels for electrophoresis without boiling under nonreducing conditions in a 10% acrylamide gel containing 1 mg/ml gelatin (Invitrogen Corp). After electrophoresis, the gels were washed at room temperature for 1 h in a buffer consisting of 50 mM Tris-HCl (pH 7.5) and

2.5% Triton X-100 to remove SDS and were then incubated overnight in a buffer consisting of 50 mM Tris-HCl (pH 7.6), 150 mM NaCl, 10 mM CaCl₂ and 0.05% NaN₃ at 37°C. The gels were stained with 0.25% Coomassie Brilliant Blue R-250 in 50% methanol and 5% acetic acid for 60 min and destained in 5% methanol and 7% acetic acid. MMP-2 and MMP-9 were visualized as clear bands against a dark background at 72 and 92 kDa, respectively.

In vitro invasion assay. Invasion assays were carried out using BD Matrigel Invasion Chambers, 24-well plate, 8-mm pore size (BD Biosciences). Cells were seeded at a density of 25,000 cells/well in 500 μ l culture medium. Following incubation periods of 24 and 48 h, the filter inserts were removed and washed twice with PBS and the upper side of the filter was cleaned with a cotton tipped swab. For assessment of the number of invaded cells, the filters were fixed and stained with 0.2% crystal violet in 4% formaldehyde. The total number of cells that had invaded onto the underside of the filter was counted manually under a light microscope.

Hypoxia-reoxygenation treatment. HChEpC1b cells were incubated under normoxic conditions (20% O₂) and then were subjected to hypoxia (1% O₂) for 1 h and subsequent reoxygenation under normoxic conditions (20% O₂) for 6 h. At the end of each culture period, cells were scraped from culture dishes and were centrifuged and snap-frozen. All samples were stored at -80°C before analysis and total RNA were isolated using the RNeasy mini kit (Qiagen).

Microarray analysis. Human genome-wide gene expression was examined using the Human Genome U133 Plus 2.0 Array (HG-U133 Plus 2.0 Set: GeneChip, Affymetrix, Santa Clara, CA, USA), which contains almost 45,000 probe sets, representing >39,000 transcripts derived from ~33,000 well-substantiated human genes (<http://www.affymetrix.com/products/arrays/specific/hgu133.affx>). Double-stranded cDNA was synthesized and the cDNA was subjected to *in vitro* transcription in the presence of biotinylated nucleotide triphosphates. Ten micrograms of the biotinylated cRNA was hybridized with a probe array for 16 h at 45°C and the

Table III. Expression of marker molecules for trophoblasts by immunocytochemistry.

	HChEpC1b	JAR	JEG-3	Hs795PI
Cytokeratin 7 ^a	+	+	+	-
HLA-G	+	-	+	-
CD9	+	+	+	+
Vimentin ^a	+	-	-	+
E-cadherin ^a	-	+	+	-
hCG	+	+	+	-
hPL	+	+	+	-
PLAP	-	+	+	-

^aSame results were confirmed by Western blotting.

hybridized biotinylated cRNA was stained with streptavidin-phycoerythrin and was then scanned with a Gene Array Scanner. The fluorescence intensity of each probe was quantified using the GeneChip Analysis Suite 5.0 software (Affymetrix). The expression level of a single RNA strand was set as the average fluorescence intensity among the intensities obtained from 11-paired (perfect-matched and single nucleotide-mismatched) probes consisting of 25-mer oligonucleotides. If the intensities of mismatched probes were very high, gene expression was judged to be absent even if a high average fluorescence was obtained with the Microarray Analysis Suite 5.0 program (MAS5.0). The data was processed with Affimetrix's default parameters, except for scaling (Target Intensity 1000), without normalization procedures to calculate the level of gene expression as the signal.

Statistics. Assuming the changes are reversible, the detection p-value at normoxic conditions (P1) should equal to that at subsequent reoxygenation (P3). Therefore, we deleted genes for which the value P3/P1 was outside of the range 0.75-1.25. Then, we selected genes of which $P2 \times 2 / (P1 + P3) > 2$, suggesting up-regulation by hypoxia (P2 was the detection p-value under hypoxic conditions). On the other hand, genes for which $P2 \times 2 / (P1 + P3) < 0.5$, were defined as down-regulated. The analysis was performed with Excel (Microsoft Office Excel 2007).

Results

Immortalization of HChEpC cells. After transduction of hTERT and HPV16 E6/E7, the clone HChEpC1b demonstrated continuous growth for >80 passages, which is equivalent to 160 population doublings at a split ratio of 1:4. The population doubling time was ~24 h. As no indication of cellular crisis throughout this period was observed, the cells were considered to be virtually immortalized.

Extravillous trophoblast phenotype of HChEpC1b. Immunocytochemistry was performed in JAR and JEG-3 cells, which are choriocarcinoma cell lines, and Hs795PI, which is the

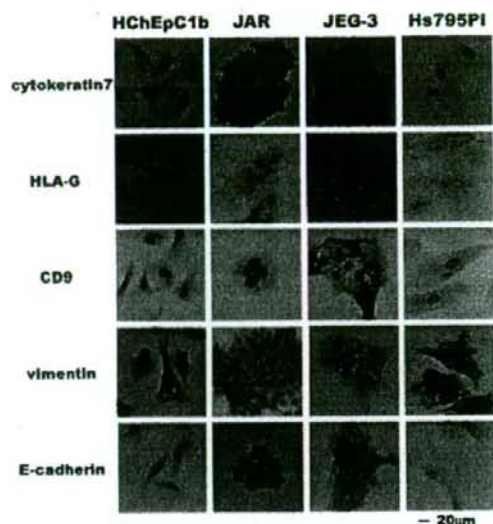


Figure 1. Immunocytochemistry of HChEpC1b, JAR, JEG-3 and Hs795PI (each column, left to right), stained by cytokeratin 7, HLA-G, CD9, vimentin and E-cadherin (each row, top to bottom) at passage 75.

fibroblast cell line that was derived from the term placenta. HChEpC1b showed positive staining for cytokeratin 7, HLA-G, CD9, hCG, hPL, and vimentin, but was negative for E-cadherin and PLAP (Fig. 1 and Table III). As discussed in the international workshop on trophoblast cell lines (23), the minimum characteristic requirements for an extravillous trophoblast cell line are the positive expression of cytokeratin 7, HLA-G and CD9. By immunocytochemistry, HChEpC1b showed expression of all of these extravillous markers. Some of these results were confirmed by Western blotting using the same antibodies.

Real-time RT-PCR demonstrated the same expression profiles as the immunological studies for CK7, HLA-G, CD7 and vimentin (Fig. 2). The ability of these cells to produce hPL and PLAP was low compared to that of the choriocarcinoma cells. The integrin expression profile revealed an extravillous trophoblastic phenotype.

The transwell invasion assay showed the invasiveness of HChEpC1b (Fig. 3A) and gelatin zymography detected the secretion of MMP-2 and MMP-9 (Fig. 3B). These results fulfill the biological criteria of extravillous trophoblasts suggested in the IFPA Workshop (24).

Growth potential, karyotyping and transformed properties. G-banding staining of HChEpC1b was conducted at passage 6, 14, 25 and 75. Karyotype analysis exhibited almost normal chromosomal numbers ranging from 46 to 48, but also contained minor structural alterations, probably associated with p53 inactivation (Table IV).

No tumors developed in nude mice after injection with HChEpC1b after a 3-month period. Therefore, immortalization of HChEpC1b was not accompanied by tumorigenic transformation.

Table IV. Doubling time and karyotype of HChEpC1b.

Passage	Doubling time (h)	Chromosome no.	Karyotype
6	21.3	47	47,XY,+20[18]/ 47,idem,-14,+mar[2]
14	ND	46-47	47,XY,+20[2]/ 47,idem,-14,+mar[12]/ 46,XY[6]
25	24	46-48	47,XY,+20[7]/ 47,idem,-14,+mar[6]/ 46,idem,der(4;16)(p10;q10)[1]/ 48,idem,+6[1]/ 47,idem,i(14)(q10)[1]/ 47,XY,+14[1]/ 46,XY[3]
75	23.9	46-47	46,XY,add(11)(q13or21),-13,+20[14]/ 46,idem,+14,der(14;21)(q10;q10)[1]/ 47,idem,+der(1;2)(p10;q10),add(14)(p10),add(14)(q22)[1]/ 46,idem,t(13;16)(p10;p10)[1]/ 46,idem,add(18)(q12)[1]/ 47,idem,+mar1[1]/ 46,idem,-18,+mar2[1]

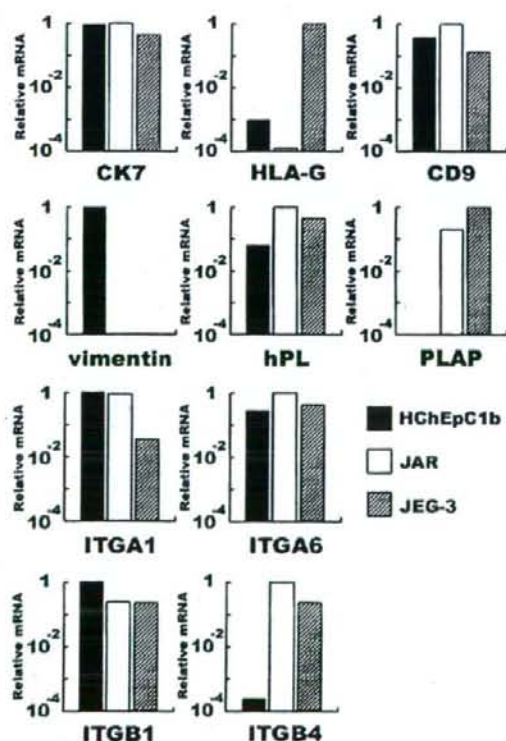


Figure 2. Real-time RT-PCR to detect the mRNA expression of molecular markers for trophoblast characterization.

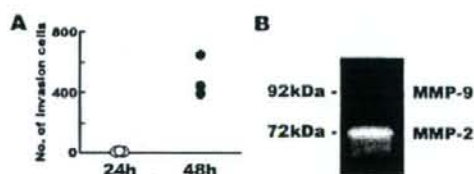


Figure 3. (A) A transwell insert invasion assay to show the invasiveness of HChEpC1b. The number of cells indicates the total count of cells that invaded through the insert membrane, which is coated with Matrigel Matrix, toward the underside of the membrane. (B) Gelatin zymography showing the secretion of MMP-9 and MMP-2.

Responsiveness to hypoxia reoxygenation treatment. The up-regulated and down-regulated genes that showed reversible expression in hypo-normo oxidative conditions are shown in Tables V and VI, respectively.

Discussion

In the present study, we established the cell line HChEpC1b, that acquired immortality after HPV16 E6/E7 and hTERT introduction. This cell line expressed molecular markers for human extravillous trophoblasts, including cytokeratin 7, HLA-G and CD9. Study of the biology of trophoblasts has been limited by the lack of suitable cell models that represent this unique cell population *in vitro*. In the past, choriocarcinoma cell lines, such as BeWo (25), JEG (26) and JAR (27) were the only immortal cell lines available and have been used extensively to study placental functions regardless of limitations related to their malignant origins. Recently, several cell lines have been established by transfection with viral

Table V. Up-regulated genes showing reversible expression in hypo-normo oxidative conditions.

UniGene ID_Affymetrix	Gene title
Hs.427236	MyoD family inhibitor domain containing
Hs.431081	Ubiquitin-specific peptidase 53
Hs.29725	Hypothetical protein FLJ13197
Hs.435991	Chromosome 4 open reading frame 16
Hs.469970	Splicing factor, arginine/serine-rich 4
Hs.522074	TSC22 domain family, member 3
Hs.631772	Solute carrier family 25 (mitochondrial carrier; ornithine transporter) member 15
---	Zinc finger protein 252
Hs.533499	Membrane-associated DNA binding protein
Hs.492348	SLAM family member 6
Hs.469840	Cytoplasmic linker-associated protein 1
Hs.369430	Peptidylglycine α -amidating monooxygenase
Hs.436121	Coiled-coil domain containing 112
Hs.459790	Vacuolar protein sorting 13 homolog A (<i>S. cerevisiae</i>)
Hs.302085	Prostaglandin I2 (prostacyclin) synthase /// prostaglandin I2 (prostacyclin) synthase
Hs.157818	Potassium voltage-gated channel, shaker-related subfamily, β member 1
Hs.413801	Proteasome (prosome, macropain) activator subunit 4
Hs.512152	HLA-G histocompatibility antigen, class I, G
Hs.124565	Solute carrier family 23 (nucleobase transporters), member 3
Hs.441124	Apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like 3F

Table VI. Down-regulated genes showing reversible expression in hypo-normo oxidative conditions.

UniGene ID_Affymetrix	Gene title
Hs.368912	Dipeptidyl-peptidase 4 (CD26, adenosine deaminase complexing protein 2)
Hs.357128	Molybdenum cofactor synthesis 1 /// molybdenum cofactor synthesis 1
Hs.533499	Membrane associated DNA binding protein
Hs.617709	Chromosome 1 open reading frame 136
Hs.12929	Chromosome 17 open reading frame 80
Hs.470907	Adenylate kinase 2
Hs.471040	Hypothetical protein FLJ38973
Hs.529577	U2-associated SR140 protein
Hs.478000	Muscleblind-like (<i>Drosophila</i>)
Hs.502182	Brain-derived neurotrophic factor
Hs.443301	Centrosomal protein 63 kDa
Hs.321176	Transcribed locus, weakly similar to XP_850647.1 PREDICTED: similar to LINE-1 reverse transcriptase homolog [<i>Canis familiaris</i>]
Hs.28780	Zinc finger protein 449
Hs.554740	Fanconi anemia, complementation group B
Hs.340623	Hypothetical protein LOC644873
Hs.569809	Rho GTPase activating protein 27
Hs.461860	Nucleoporin 214 kDa
Hs.452398	MRNA; cDNA DKFZp564E143 (from clone DKFZp564E143)
Hs.16004	Chromosome 10 open reading frame 76
Hs.612891	Hypothetical protein FLJ21272
Hs.59159	Chromodomain helicase DNA binding protein 9
Hs.397001	CDNA clone IMAGE:4304686
Hs.512682	Carcinoembryonic antigen-related cell adhesion molecule 1 (biliary glycoprotein)

oncogenes. The most commonly used gene was the early region of SV40 containing the small and large T-antigens that bind to and inactivate p53 and Rb. Although the expression of the T-antigen extends the life span of the cells, the onset of cellular senescence is inevitable. This is associated with a reduction of telomere length. Other commonly used viral genes include HPV16 E6 and E7, which inhibit the functions of p53 and Rb respectively. E6 has been reported to activate telomerase (28), which can stabilize the chromosome profile and avoid an immortalization-related crisis. Therefore, we infected human trophoblast cells with retroviral expression vectors containing type 16 human papillomaviruses E6 and E7 in combination with hTERT to obtain immortalized trophoblast cells showing approximately diploid chromosomes.

Although cytokeratin-positive, vimentin-negative phenotypes are regarded as markers of trophoblast lineage (24), HChEpC1b expressed both cytokeratin and vimentin. Coexpression of cytokeratin and vimentin by extravillous trophoblasts was reported earlier (29), which is a characteristic similar to that of some cancer cells of epithelial origin (30-32), as part of an epithelial-mesenchymal transition (33,34). On the other hand, E6 is known to up-regulate vimentin (35) and another trophoblast cell line immortalized by E6/E7, TEV-1, was reported to express vimentin. From our results, it is difficult to distinguish whether the origin of HChEpC1b was the vimentin-positive extravillous trophoblasts or the vimentin negative trophoblasts that displayed the mesenchymal phenotype after E6 introduction. To answer this problem, further study is needed to compare the effects of E6 on each trophoblast population.

We obtained 43 genes that showed reversible up- or down-regulation during hypoxia. There are several interesting up-regulated genes including that for the MyoD family inhibitor domain containing protein isoform p40, which is involved in repair of muscle injury in ischemic micro-environments (36); prostaglandin I₂ synthase, which is a potent vasodilator and inhibitor of platelet aggregation and plays an important role in the predisposition to lung cancer (37); human leukocyte antigen G (HLA-G) molecule, which plays an important role in immune response regulation and has been implicated in the inhibition of the cytolytic function of natural killer and cytotoxic T cells (38); and apolipoprotein B mRNA editing enzyme, a catalytic polypeptide-like 3C, which plays roles in growth or cell cycle control (39). Among the interesting down-regulated genes, there were adenylate kinase 2, which is localized in the mitochondrial intermembrane space and may play a role in apoptosis (40); BDNF brain-derived neurotrophic factor, which plays a role in the regulation of stress response and in the biology of mood disorders (41); and carcinoembryonic antigen-related cell adhesion molecule 1, which plays roles in the differentiation and arrangement of tissue three-dimensional structure, angiogenesis, apoptosis, tumor suppression, metastasis and the modulation of innate and adaptive immune responses (42). In future, we should confirm the expression of these genes by real-time RT-PCR and further examination should focus on a functional assay of these candidate genes to assess whether they play important roles during hypoxia.

In summary, we established a newly immortalized trophoblast cell line via infection with HPV16 E6/E7 in

combination with hTERT and this cell line is a useful model for the study of extravillous trophoblast function.

Acknowledgements

This study was supported in part by Grants-in-Aid from the Ministry of Health, Labor and Welfare of Japan.

References

- Benirschke K, Kaufmann P and Baergen RN: Pathology of the Human Placenta. 5th edition, Springer, New York, 2005.
- Redman CW and Sargent IL: Latest advances in understanding preeclampsia. *Science* 308: 1592-1594, 2005.
- Allsopp RC, Vaziri H, Patterson C, et al: Telomere length predicts replicative capacity of human fibroblasts. *Proc Natl Acad Sci USA* 89: 10114-10118, 1992.
- Saretzki G, Sitte N, Merkel U, Wurm RE and von Zglinicki T: Telomere shortening triggers a p53-dependent cell cycle arrest via accumulation of G-rich single stranded DNA fragments. *Oncogene* 18: 5148-5158, 1999.
- Kiyono T, Foster SA, Koop JI, McDougall JK, Galloway DA and Klingelutz AJ: Both Rb/p16INK4a inactivation and telomerase activity are required to immortalize human epithelial cells. *Nature* 396: 84-88, 1998.
- Rheinwald JG, Hahn WC, Ramsey MR, et al: A two-stage, p16(INK4A)- and p53-dependent keratinocyte senescence mechanism that limits replicative potential independent of telomere status. *Mol Cell Biol* 22: 5157-5172, 2002.
- Graham CH, Hawley TS, Hawley RG, et al: Establishment and characterization of first trimester human trophoblast cells with extended lifespan. *Exp Cell Res* 206: 204-211, 1993.
- Ho CK, Chiang H, Li SY, Yuan CC and Ng HT: Establishment and characterization of a tumorigenic trophoblast-like cell line from a human placenta. *Cancer Res* 47: 3220-3224, 1987.
- Ho CK, Li SY, Yu KJ, Wang CC, Chiang H and Wang SY: Characterization of a human tumorigenic, poorly differentiated trophoblast cell line. *In Vitro Cell Dev Biol Anim* 30A: 415-417, 1994.
- Morgan M, Kniss D and McDonnell S: Expression of metalloproteinases and their inhibitors in human trophoblast continuous cell lines. *Exp Cell Res* 242: 18-26, 1998.
- Rong-Hao L, Luo S and Zhuang LZ: Establishment and characterization of a cytotrophoblast cell line from normal placenta of human origin. *Hum Reprod* 11: 1328-1333, 1996.
- Zdravkovic M, Aboagye-Mathiesen G, Guimond MJ, Hager H, Ebbesen P and Lala PK: Susceptibility of MHC class I expressing extravillous trophoblast cell lines to killing by natural killer cells. *Placenta* 20: 431-440, 1999.
- Chou JY: Establishment of clonal human placental cells synthesizing human chorionic gonadotropin. *Proc Natl Acad Sci USA* 75: 1854-1858, 1978.
- Choy MY and Manyonda IT: The phagocytic activity of human first trimester extravillous trophoblast. *Hum Reprod* 13: 2941-2949, 1998.
- Khoo NK, Bechberger JF, Shepherd T, et al: SV40 Tag transformation of the normal invasive trophoblast results in a premalignant phenotype. I. Mechanisms responsible for hyperinvasiveness and resistance to anti-invasive action of TGFbeta. *Int J Cancer* 77: 429-439, 1998.
- Lei KJ, Gluzman Y, Pan CJ and Chou JY: Immortalization of virus-free human placental cells that express tissue-specific functions. *Mol Endocrinol* 6: 703-712, 1992.
- Lewis MP, Clements M, Takeda S, et al: Partial characterization of an immortalized human trophoblast cell-line, TCL-1, which possesses a CSF-1 autocrine loop. *Placenta* 17: 137-146, 1996.
- Feng HC, Choy MY, Deng W, et al: Establishment and characterization of a human first-trimester extravillous trophoblast cell line (TEV-1). *J Soc Gynecol Investig* 12: e21-e32, 2005.
- Shih I, Wang T, Wu T, Kurman RJ and Gearhart JD: Expression of Mel-CAM in implantation site intermediate trophoblastic cell line, IST-1, limits its migration on uterine smooth muscle cells. *J Cell Sci* 111 (Pt 17): 2655-2664, 1998.
- Wang YL, Qiu W, Feng HC, et al: Immortalization of normal human cytotrophoblast cells by reconstitution of telomeric reverse transcriptase activity. *Mol Hum Reprod* 12: 451-460, 2006.

21. Kyo S, Nakamura M, Kiyono K, *et al.*: Successful immortalization of endometrial glandular cells with normal structural and functional characteristics. *Am J Pathol* 163: 2259-2269, 2003.
22. Naviaux RK, Costanzi E, Haas M and Verma IM: The pCL vector system: rapid production of helper-free, high-titer, recombinant retroviruses. *J Virol* 70: 5701-5705, 1996.
23. Shiverick KT, King A, Frank H, Whitley GS, Cartwright JE and Schneider H: Cell culture models of human trophoblast II: trophoblast cell lines - a workshop report. *Placenta* 22 (Suppl A): S104-S106, 2001.
24. Morrish DW, Whitley GS, Cartwright JE, Graham CH and Caniggia I: In vitro models to study trophoblast function and dysfunction - a workshop report. *Placenta* 23 (Suppl A): S114-S118, 2002.
25. Pattillo RA and Gey GO: The establishment of a cell line of human hormone-synthesizing trophoblastic cells in vitro. *Cancer Res* 28: 1231-1236, 1968.
26. Kohler PO and Bridson WE: Isolation of hormone-producing clonal lines of human choriocarcinoma. *J Clin Endocrinol Metab* 32: 683-687, 1971.
27. Husa RO, Story MT and Pattillo RA: Regulation of human chorionic gonadotropin (hCG) secretion by serum and dibutyl cyclic AMP in malignant trophoblast cells in vitro. *J Clin Endocrinol Metab* 40: 401-405, 1975.
28. Gewin L, Myers H, Kiyono T and Galloway DA: Identification of a novel telomerase repressor that interacts with the human papillomavirus type-16 E6/E6-AP complex. *Genes Dev* 18: 2269-2282, 2004.
29. Aboagye-Mathiesen G, Laugesen J, Zdravkovic M and Ebbesen P: Isolation and characterization of human placental trophoblast subpopulations from first-trimester chorionic villi. *Clin Diagn Lab Immunol* 3: 14-22, 1996.
30. Azumi N and Battifora H: The distribution of vimentin and keratin in epithelial and nonepithelial neoplasms. A comprehensive immunohistochemical study on formalin- and alcohol-fixed tumors. *Am J Clin Pathol* 88: 286-296, 1987.
31. McNutt MA, Bolen JW, Gown AM, Hammar SP and Vogel AM: Coexpression of intermediate filaments in human epithelial neoplasms. *Ultrastruct Pathol* 9: 31-43, 1985.
32. Ramaekers FC, Haag D, Kant A, Moesker O, Jap PH and Vooijs GP: Coexpression of keratin - and vimentin-type intermediate filaments in human metastatic carcinoma cells. *Proc Natl Acad Sci USA* 80: 2618-2622, 1983.
33. Christiansen JJ and Rajasekaran AK: Reassessing epithelial to mesenchymal transition as a prerequisite for carcinoma invasion and metastasis. *Cancer Res* 66: 8319-8326, 2006.
34. Ferretti C, Bruni L, Dangles-Marie V, Pecking AP and Bellet D: Molecular circuits shared by placental and cancer cells, and their implications in the proliferative, invasive and migratory capacities of trophoblasts. *Hum Reprod Update* 13: 121-141, 2007.
35. Duffy CL, Phillips SL and Klingelutz AJ: Microarray analysis identifies differentiation-associated genes regulated by human papillomavirus type 16 E6. *Virology* 314: 196-205, 2003.
36. Yun Z, Lin Q and Giaccia AJ: Adaptive myogenesis under hypoxia. *Mol Cell Biol* 25: 3040-3055, 2005.
37. Stearman RS, Grady MC, Nana-Sinkam P, Varella-Garcia M and Geraci MW: Genetic and epigenetic regulation of the human prostacyclin synthase promoter in lung cancer cell lines. *Mol Cancer Res* 5: 295-308, 2007.
38. Castelli EC, Mendes-Junior CT, Viana de Camargo JL and Donadi EA: HLA-G polymorphism and transitional cell carcinoma of the bladder in a Brazilian population. *Tissue Antigens* 72: 149-157, 2008.
39. Kock J and Blum HE: Hypermutation of hepatitis B virus genomes by APOBEC3G, APOBEC3C and APOBEC3H. *J Gen Virol* 89: 1184-1191, 2008.
40. Lee HJ, Pyo JO, Oh Y, *et al.*: AK2 activates a novel apoptotic pathway through formation of a complex with FADD and caspase-10. *Nat Cell Biol* 9: 1303-1310, 2007.
41. Licinio J and Wong ML: Brain-derived neurotrophic factor (BDNF) in stress and affective disorders. *Mol Psychiatry* 7: 519, 2002.
42. Gray-Owen SD and Blumberg RS: CEACAM1: contact-dependent control of immunity. *Nat Rev Immunol* 6: 433-446, 2006.

Regulation of MyD88-Dependent Signaling Events by S Nitrosylation Retards Toll-Like Receptor Signal Transduction and Initiation of Acute-Phase Immune Responses⁷

Takeshi Into,^{1*} Megumi Inomata,¹ Misako Nakashima,¹ Ken-ichiro Shibata,²
Hans Häcker,³ and Kenji Matsushita¹

Department of Oral Disease Research, National Institute for Longevity Sciences, National Center for Geriatrics and Gerontology, 36-3, Gengo, Morioka, Obu, Aichi 474-8522, Japan¹; Laboratory of Oral Molecular Microbiology, Department of Oral Pathobiological Science, Hokkaido University Graduate School of Dental Medicine, Sapporo 060-8586, Japan²; and Department of Infectious Diseases, St. Jude Children's Research Hospital, 332 North Lauderdale Street, Memphis, Tennessee 38105³

Received 7 August 2007/Returned for modification 23 September 2007/Accepted 28 November 2007

Nitric oxide (NO) has been thought to regulate the immune system through S nitrosylation of the transcriptional factor NF- κ B. However, regulatory effects of NO on innate immune responses are unclear. Here, we report that NO has a capability to control Toll-like receptor-mediated signaling through S nitrosylation. We found that the adaptor protein MyD88 was primarily S nitrosylated, depending on the presence of endothelial NO synthase (eNOS). S nitrosylation at a particular cysteine residue within the TIR domain of MyD88 resulted in slight reduction of the NF- κ B-activating property. This modification could be restored by the antioxidant glutathione. Through S nitrosylation, NO could negatively regulate the multiple steps of MyD88 functioning, including translocation to the cell membrane after LPS stimulation, interaction with TRAF, binding to TRAF6, and induction of I κ B α phosphorylation. Interestingly, glutathione could reversibly neutralize such NO-derived effects. We also found that an acute febrile response to LPS was precipitated in eNOS-deficient mice, indicating that eNOS-derived NO exerts an initial suppressive effect on inflammatory processes. Thus, NO has a potential to retard induction of MyD88-dependent signaling events through the reversible and oxidative modification by NO, by which precipitous signaling reactions are relieved. Such an effect may reflect appropriate regulation of the acute-phase inflammatory responses in living organisms.

It is increasingly becoming evident that nitric oxide (NO) regulates a broad spectrum of protein functions through S nitrosylation, a posttranscriptional modification that forms S-nitrosothiol by covalent addition to cysteine residues of an NO moiety (14, 42, 43). Through S nitrosylation, NO is thought to exert a physiological inhibitory effect on nuclear factor κ B (NF- κ B) (25, 32, 33, 39), the major transcriptional factor family deeply associated with regulation of the immune system through transcription of a wide range of genes, including cytokines, adhesion molecules, antimicrobial molecules, and antiapoptotic molecules (10, 13, 24). S nitrosylation of NF- κ B inhibits its DNA binding, promoter activity, and subsequent transcription (25, 33). It has been known that S nitrosylation targets a particular cysteine residue of the NF- κ B p50 and p65 subunits located in the N-terminal DNA binding loop within the Rel homology domain (25, 32, 33). This residue is conserved in other NF- κ B subunits, including p52, p100, p105, and c-Rel, and other Rel homology domain-containing molecules. Upstream of NF- κ B, I κ B kinase β (IKK β), a catalytic subunit of the I κ B (inhibitor of NF- κ B) kinase complex, also undergoes S nitrosylation, resulting in reduction of its kinase function on phosphorylation of I κ B (39). Such reduction of the

IKK β function leads to reduced I κ B ubiquitinylation and proteasomal degradation, resulting in NF- κ B inhibition (14, 32, 39).

Toll-like receptors (TLRs) are the central innate immune sensors for a broad array of pathogen-associated molecular patterns, ranging from bacterial constituents to viral genomes (2, 35). TLRs initiate early processes of proinflammatory immune responses that help to strengthen the processes of innate and adaptive immunity (2, 20), in which NF- κ B plays many important roles (13, 24). TLRs utilize MyD88, a Toll/interleukin-1 receptor (IL-1R) homology (TIR) domain-containing adaptor molecule, to activate the NF- κ B pathway through IL-1R-associated kinases (IRAKs) and tumor necrosis factor (TNF) receptor-associated factor 6 (TRAF6) (1). It has been thought that TLR agonistic molecules, such as lipopolysaccharide (LPS), can regulate NO generation through upregulation of expression of all NO synthase (NOS) isoforms through NF- κ B activation (4, 9, 32). TLR stimulation can directly activate an antimicrobial property through inducible NOS (iNOS) expression and NO generation in macrophages (46). NO generation is a general feature of immune cells, including neutrophils, monocytes, macrophages, dendritic cells, and NK cells, as well as other cells, including endothelial cells, epithelial cells, and fibroblasts (4), all of which express multiple members of the TIR family. However, it has remained obscure whether generated NO exerts any regulatory effects on TLR signaling or subsequent processes of innate immune responses.

There has been an accumulation of biochemical evidence indicating that TLR signaling components, including IKK β

*Corresponding author. Mailing address: Section of Oral Infection Control, Department of Oral Disease Research, National Institute for Longevity Sciences, National Center for Geriatrics and Gerontology, 36-3 Gengo, Morioka, Obu, Aichi 474-8522, Japan. Phone: 81-562-44-5651, ext. 5004, Fax: 81-562-46-8083, E-mail: into@nri.go.jp.
Published ahead of print on 17 December 2007.

and NF- κ B, might be regulated by S nitrosylation. S nitrosylation inhibits the kinase activity of apoptosis signal regulation kinase 1 (ASK1) through inhibition of its binding to substrates (38). ASK1 is known as an important regulator of the TRAF6-p38 mitogen-activated protein kinase (MAPK) pathway downstream of TLR4 and is also involved in modulation of both the NF- κ B and apoptotic pathways downstream of TLR2 (19, 34). Caspase-1 was recently found to be involved in TLR2- and TLR4-mediated signal transduction of the MyD88-dependent pathway through the cleavage of the TIR domain-containing adaptor protein TIRAP (also known as Mal) (37). Caspase-1 also undergoes S nitrosylation at a cysteine residue within the enzymatic active site, suppressing its proteolytic activity (6, 31). Thus, it is possible that NO provides regulatory effects on the multiple steps of TLR-mediated innate immune signaling through S nitrosylation. In this study, we therefore designed experiments to determine the effect of S nitrosylation on TLR signaling. We further investigated how S nitrosylation affects TLR-initiated immune responses in vivo. We report here that S nitrosylation controls TLR signaling through redox-sensitive and reversible suppression of the MyD88 pathway, which facilitates appropriate control of acute-phase inflammatory responses in vivo.

MATERIALS AND METHODS

Reagents and cell culture. *N*^G-Monomethyl-L-arginine monoacetate (L-NMMA), S-nitrosoglutathione (GSNO), glutathione (GSH), N-ethylmaleimide, coumestrolin A, N-acetyl L-cysteine (NAC), ascorbic acid, and diphenylpicrylhydrazolium (DPPH) were obtained from Sigma-Aldrich. SNAP (S-nitroso-N-acetyl-L-penicillamine) was purchased from Cayman Chemical. ODO (1H-[1,2,4]oxadiazolo[4,3-a]quinoxalin-1-one) and KT5823 were obtained from Calbiochem. Preparation of TLR ligands, including highly purified *Escherichia coli* LPS, *Salmonella* LPS, Pam₂CSK₆, macrophage-activating lipopeptide 2 (MALP-2), and *Salmonella enterica* serovar Typhimurium flagellin, was as described previously (18). Recombinant human IL-1 β was from R&D Systems. Human aortic endothelial cells (HAECs) and human embryonic kidney 293 (HEK293) cells were maintained as described previously (18). HEK293 cells stably expressing human TLR4, MD2, and CD14 (293-TLR4 cells) and HEK293 cells stably expressing human TLR2 and CD14 (293-TLR2 cells) were obtained from InvivoGen.

Mice. eNOS-deficient (eNOS^{-/-}) mice and endothelial-NOS (eNOS)-deficient (eNOS^{-/-}) mice were from The Jackson Laboratories. C57BL/6J control (wild-type) mice were obtained from Japan SLC. All mice were kept under specific pathogen-free conditions. Male mice between 6 and 10 weeks of age were used for all of experiments. All animal protocols were approved by the National Institute for Longevity Sciences Animal Experimentation Committee at the National Center for Geriatrics and Gerontology (Aichi, Japan).

For LPS-induced acute lung injury, anesthetized mice received *Escherichia coli* LPS dissolved in pyrogen-free phosphate-buffered saline (PBS) containing 1 mg/ml Evans Blue intratracheally immediately after mechanical ventilation. After 30 min of administration, lung was excised and then lysed for immunoblot analysis. The febrile responses in mice treated with *E. coli* LPS were tested according to a protocol described previously (45, 49). Mice ($n = 6$) were maintained at a neutral ambient temperature of 31°C and challenged by intraperitoneal (i.p.) injection of 5 mg LPS/kg of body weight dissolved in pyrogen-free PBS. A high dose of LPS (more than 50 mg/kg) was fatal within 90 min in eNOS mice. A colonic thermocouple was inserted and fixed to the base of the tail with adhesive tape. The change in temperature was monitored at 5-min intervals during a period of 120 min after LPS administration. All of the tests were performed at the temperature of 31°C. After 2 h or 12 h of LPS administration, 2 ml of PBS was injected into the abdominal cavity of each mouse. Then, fluids were collected and centrifuged for assessment of cytokine production by an enzyme-linked immunosorbent assay (ELISA). Preparation of peritoneal macrophages was as described previously (28).

Plasmids. The DNA construct encoding 3 \times Flag-tagged MyD88 fused to the B subunit of the bacterial DNA gyrase (MyD88-GyrB) was as described previously (11). Plasmids encoding human MyD88 and TIRAP were kind gifts from Margaret K. Oeffermann (Emory University School of Medicine). The cDNAs of

N-terminal Flag-tagged and Myc-tagged MyD88, Myc-tagged TIRAP, and IRAK-1 were amplified by PCR and cloned into the pcDNA3.1 vector (Invitrogen). The construct encoding human TLR2 was as described previously (17). Constructs encoding mutated Flag-MyD88 were obtained using a QuikChange II site-directed mutagenesis kit (Stratagene) according to the manufacturer's instructions.

Protein purification. Recombinant Flag-MyD88 proteins were prepared using a FLAG M purification kit (Sigma-Aldrich) from HEK293 cells stably expressing Flag-MyD88 constructs, according to the manufacturer's instructions. Purity of recombinant proteins was confirmed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE), followed by silver staining and immunoblotting with anti-Flag antibody.

Detection of S-nitrosylated proteins. To detect S-nitrosylated MyD88 from lung lysates from wild-type mice and eNOS^{-/-} mice, we referred to the protocol described by Jailrey et al. (21). Several experiments were performed using a NitroGlo nitrosylation detection kit (PerkinElmer) according to the manufacturer's instructions. Lung lysates from wild-type and eNOS^{-/-} mice were subjected to the biotin switching S-nitrosylation assay, and then biotinylated proteins were purified on streptavidin-agarose. Purified proteins eluted by 2-mercaptoethanol were detected by immunoblotting with anti-MyD88 antibody.

The quantitative measurement of S-nitrosylated recombinant MyD88 by ELISA was performed as follows. Briefly, recombinant Flag-MyD88 (150 μ g) was treated with or without SNAP for 30 min at 37°C in the dark. Then, the free sulfides of Flag-MyD88 were blocked with 4 mM methylmethanethiosulfonate for 15 min. After purification by using Micro Bio-Spin chromatography columns (Bio-Rad Laboratories), Flag-MyD88 was reacted with 25 mM ascorbate to be completely denitrosylated. Free sulfides were then labeled with a biotin-conjugated maleimide, using a biotin labeling kit (SH; Dojindo Laboratories) according to the manufacturer's instructions. The diluents of biotinylated Flag-MyD88 proteins dissolved in Tris-buffered saline (pH 7.2) were stabilized in the wells of immobilizer streptavidin plates (Nunc). Flag-MyD88 proteins in the wells were detected by using anti-Flag antibody and a secondary antibody conjugated with horseradish peroxidase. Colorimetric reaction was detected by absorbance on a spectrophotometer at 450 nm. Results were expressed as means \pm standard deviations (SD) of three determinations.

Photolysis of S-nitrosylated proteins. Mouse lung lysates were exposed for 3 min to a UV-visible light mercury vapor lamp according to a protocol recently described (8). The samples were then subjected to the biotin switch technique as described above.

Luciferase reporter assay. 293-TLR2 cells were transiently transfected with wild-type MyD88-GyrB or MyD88-GyrB mutants, each with a cysteine residue replaced with a serine residue, together with 50 ng of an NF- κ B (5 \times) luciferase reporter plasmid (pNF- κ B-Luc; Stratagene) and 5 ng of an internal control luciferase reporter plasmid (pRL-TK; Promega) and incubated for 16 h. At 6 h before the end of incubation, cells were treated with or without 250 μ M SNAP. Cells were then stimulated with 100 ng/ml Pam₂CSK₆ for 6 h. HEK293 cells stably expressing MyD88-GyrB were transfected with pNF- κ B-Luc and pRL-TK. After 24 h of incubation, cells were stimulated with coumestrolin A in the presence and absence of 250 μ M SNAP. The dual luciferase activity was measured as described previously (19).

Immunoblot analysis of IRAK-1 and I κ B α . HAECs were stimulated with 10 ng/ml of LPS for 0 to 90 min. HEK293 cells stably expressing MyD88-GyrB were stimulated with 1 μ M coumestrolin for 20 min. Cells were lysed in the presence of protease inhibitor and phosphatase inhibitor cocktails (Roche) at 4°C. Cell lysates or lysates from the mouse lungs were separated by SDS-PAGE, followed by immunoblot analyses using anti-IRAK-1, anti-I κ B α , and phosphorylation-specific anti-I κ B α (Ser32/Ser36) antibodies (Cell Signaling Technology).

RNA extraction and reverse transcription-PCR. Total RNA was isolated from mouse peritoneal macrophages stimulated with 100 ng/ml LPS and 10 ng/ml gamma interferon, and transcripts were quantified by real-time quantitative reverse transcription-PCR on a LightCycler ST300 system (Roche). All values were normalized to the level of β -actin mRNA. The primer sets used are as follows: for mouse macrophage inflammatory protein 2 (MIP-2), 5'-ATCCAG AGT TTAGTGTGACGC-3' (sense) and 5'-AAGGCAAACTTTTTCACCC AA-3' (antisense); for mouse IL-6, 5'-CCACGGCTTCCTCTAC-3' (sense) and 5'-AGTGCATCACTGTGTTC-3' (antisense); and for mouse β -actin, 5'-AA ATCGTGGTGTGACATCAAA-3' (sense) and 5'-AAGGAAGCTGTGAAAAG AGC-3' (antisense).

Cytokine ELISA. Concentrations of human IL-6, mouse MIP-2, and mouse IL-6 were determined using a Cytoset ELISA kit (Biosource) according to the manufacturer's instructions.

Subcellular fractionation. Subcellular fractionation of HEK293 cells stably expressing Flag-MyD88 and 293-TLR4 cells stably expressing Flag-MyD88-GyrB

was performed using a ProteoExtract subcellular proteome extraction kit (Calbiochem) according to the manufacturer's instructions. This kit enables extraction of different subcellular fractions of the cytoplasm, plasma membrane, nuclei, and cytoskeleton from mammalian cells. 293-TLR4 cells stably expressing Flag-MyD88-GyrB were maintained in serum-free Dulbecco's modified Eagle's medium containing 5% PANEXIN H cell growth supplement (PAN Biotech GmbH) to avoid nonspecific cell activation by animal serum components. The whole-cell lysate was obtained using NuPAGE LDS sample buffer containing 2-mercaptoethanol. Each fraction was mixed with NuPAGE LDS sample buffer and boiled for 5 min, followed by SDS-PAGE and immunoblot analyses using anti-Flag (Sigma), anti-MyD88 (Santa Cruz Biotechnology), anti-IRAK-1, and anti-vimentin (BD Biosciences) antibodies.

Blue native PAGE and immunoprecipitation. HEK293 cells stably expressing Flag-MyD88 were treated with SNAP for 1 h, washed twice with PBS, and lysed with HEPES buffer (pH 7.2) containing 1% Triton X-100, 1% Nonidet-P40, and proteinase inhibitor cocktail (Roche). Cell lysates were separated by blue native PAGE according to the protocol provided by Invitrogen and then immunoblotted with anti-Flag antibody. HEK293 cells transiently transfected combinationally with Flag-MyD88 and Myc-TRAP or Flag-MyD88 and IRAK-1 were treated with GSNO or GSH for 1 h. HEK293 cells stably expressing 3 \times Flag-MyD88-GyrB were stimulated with 1 μ M canomycin A for 20 min. Cells were lysed with HEPES buffer (pH 7.2) containing 1% Triton X-100 and proteinase inhibitor cocktail (Roche). Clarified cell lysates were immunoprecipitated with anti-Flag antibody, followed by immunoblot analysis using anti-Flag, anti-Myc, and anti-TRAF6 (StressGen) antibodies. All experiments were performed at least three times, and representative results are shown.

Immunofluorescent cell staining. HAECs were fixed at -20°C with methanol, and double immunostaining was then carried out with anti- β -actin monoclonal antibody (Santa Cruz Biotechnology) and Alexa 488-conjugated immunoglobulin G secondary antibody (Invitrogen) and then with anti-MyD88 rabbit polyclonal antibody (Santa Cruz Biotechnology) and Alexa 564-conjugated immunoglobulin G secondary antibody (Invitrogen). Cell nuclei were also stained with 2.5 μ g/ml of Hoechst 33342 for 30 min.

Statistical analysis. Probability (*P*) values were calculated by Student's *t* test and analysis of variance and were considered significant at 0.05 or 0.01.

RESULTS

TLR signaling components are S-nitrosylated *in vivo*. To examine whether TLR signal components are physiologically S-nitrosylated *in vivo*, we detected S-nitrosylated proteins in the lung lysates from wild-type and eNOS $^{-/-}$ mice by utilizing the biotin switch method (21). Interestingly, this method facilitated the detection of MyD88 as an S-nitrosylated protein (Fig. 1A). To exclude the possibility that the detection of S-nitrosylated MyD88 is an experimental artifact, we utilized the biotin switch method combined with photolysis of S-nitrosylation, which has recently been reported as a useful method for confirming the specificity of S-nitrosylation (8). The detectable S-nitrosylated MyD88 protein was reduced after exposure of the samples to a UV lamp (Fig. 1B), suggesting that the result is not false positive. Thus, our result at least suggests that MyD88 is potentially S-nitrosylated in addition to other signaling molecules, including NF- κ Bs, ASK1, and caspase-1.

We further examined the details of S-nitrosylated MyD88 *in vitro* by utilizing a quantitative method for detecting S-nitrosylation of recombinant proteins. We could detect S-nitrosylation of recombinant MyD88, which increased, accompanied by an increase in the concentration of the NO donor SNAP (Fig. 1C). It has been known that S-nitrosylated proteins are reversibly denitrosylated by antioxidants or oxidoreductases, by which substantial protein functions are restored (14, 42). Indeed, the detectable S-nitrosylated MyD88 protein was reduced when the NO donor-treated protein was reacted with ascorbate, HgCl₂, or GSH (Fig. 1C). We further determined the site of S-nitrosylation because the modification is effected

toward particular cysteine residues (14). Mammalian MyD88 contains a total of nine cysteine residues: one in a short linker region and the other eight in the TIR domain (Fig. 1D). These residues are thought not to be involved in the formation of intramolecular disulfide bonds. Among vertebrates, all cysteine residues are highly conserved (data not shown). We prepared recombinant MyD88 proteins of nine individual mutants, each with one of the nine cysteine residues replaced with a serine residue. We found that the degrees of S-nitrosylation of Cys113 and Cys216 were significantly reduced compared with those of wild-type MyD88 (Fig. 1E). These cysteine residues partially fulfill the predictive site of the S-nitrosylation "acid-base motif" that comprises flanking acidic and basic residues (14) (Fig. 1F). Interestingly, the cysteine residue equivalent of Cys216 is conserved even in invertebrates, while others are not (data not shown). Among other TIR domain-containing adaptor molecules, only TRAP and SIGIRR have a cysteine residue corresponding to the position of Cys216 (Fig. 1G).

To determine the requirement of cysteine residues for functioning of MyD88, we utilized MyD88 fused to the β subunit of the bacterial DNA gyrase (MyD88-GyrB). The *Sreptomyces*-derived bivalent antibiotic coumermycin binds GyrB with a stoichiometry of 1:2, acting as a natural dimerizer of GyrB (7). Although overexpressed MyD88 is known to reveal TLR stimulation-independent nonspecific activation of downstream signaling through self-dimerization (11, 36), MyD88-GyrB does not reveal such nonspecific activation unless cells are exposed to TLR stimulation or coumermycin treatment (11). We prepared GyrB-fused wild-type MyD88 and MyD88 mutants, each with one of the nine cysteine residues replaced with a serine residue, and examined the NF- κ B-activating properties in the TLR2 ligand Pam₃CSK₄-stimulated HEK293 cells stably expressing TLR2. None of the cysteine replacement mutants abrogated the NF- κ B-activating property of MyD88 (Fig. 1H). However, the Cys216Ser mutant significantly increased the activity compared with that of wild-type MyD88 (Fig. 1H). Additionally, similar results were found in the cells treated with SNAP (Fig. 1H). Thus, it is possible that Cys216 of MyD88 mediates the suppressive effect of NO.

S-nitrosylation alters MyD88-mediated signaling events. We next explored how S-nitrosylation of signaling components alters TLR signaling events. To examine this *in vivo*, we utilized an animal model of acute lung injury induced by intratracheal administration of LPS. We investigated degradation of IRAK-1 and I κ B α , hallmarks of MyD88-dependent and IKK β -dependent signaling events, in lungs 30 min after LPS administration. Interestingly, degradation of IRAK-1 and I κ B α was apparently promoted in eNOS $^{-/-}$ mice compared with that in wild-type mice (Fig. 2A). We also examined whether NO alters degradation of IRAK-1 and I κ B α in cultivated vascular endothelial cells. In HAECs, LPS induced degradation of IRAK-1 and I κ B α within 30 min after stimulation (Fig. 2B). Degradation of IRAK-1 and I κ B α was promoted and occurred within 15 min after stimulation when endogenous NO was predepleted by the L-arginine analog L-NMMA (Fig. 2B). In addition, IRAK-1 degradation was also promoted when confluent HAECs were maintained in culture media without the eNOS activator vascular endothelial growth factor or the phosphatidylinositol 3-kinase inhibitor LY294002 (data not shown). In contrast to these results, degradation was delayed

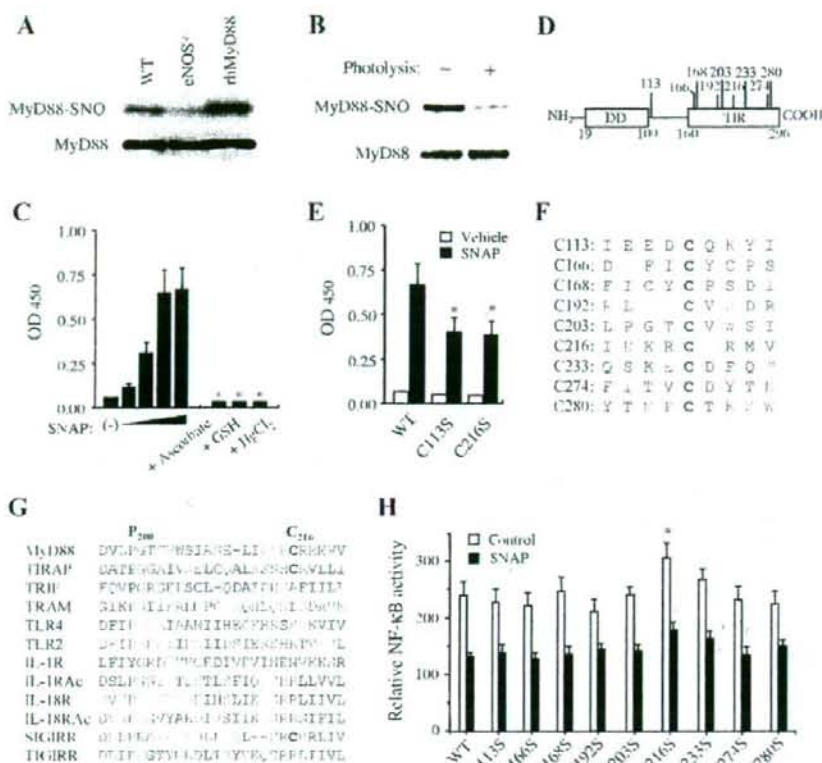


FIG. 1. S-nitrosylation of MyD88. (A) Lung lysates from wild-type (WT) and eNOS^{-/-} mice and SNAP-treated recombinant human MyD88 (rhMyD88) were subjected to the biotin switching S-nitrosylation assay, and then biotinylated proteins were purified on streptavidin-agarose. Purified proteins were detected by immunoblotting with anti-MyD88 antibody (upper). MyD88 proteins in lung lysates and rhMyD88 were also shown as loading controls (lower). (B) Mouse lung lysates were exposed for 3 min to a UV-visible light mercury vapor lamp. The samples were then subjected to the biotin switch method. (C) A recombinant Flag-MyD88 protein was treated with or without SNAP (100, 200, 500, and 1,000 μ M) for 30 min. For the denitrosylation study, SNAP-treated proteins were incubated with 1 mM ascorbic acid, 1 mM GSH, or 1 mM HgCl₂ for 5 min before the blockade of free thiols by methylmethanethioniosulfonate. Then, S-nitrosylated residues of MyD88 were switched into biotins and proteins were fixed on streptavidin-coated plates, followed by ELISA with anti-Flag antibody. Each value is the mean \pm SD ($n = 3$). See text for details. (*, $P < 0.01$ for comparison with the group of 500 μ M SNAP). (D) Schematic of human MyD88. (E) Recombinant Flag-MyD88 wild-type proteins and mutants with each cysteine residue replaced with a serine residue were treated with or without 500 μ M SNAP for 30 min. Then, S-nitrosylated MyD88 fixed on streptavidin-coated plates was detected by ELISA using anti-Flag antibody. Each value is the mean \pm SD ($n = 3$). See text for details. (*, $P < 0.01$ for comparison with the wild-type group). (F) Sequence alignment of the region around nine cysteine residues of MyD88. (G) Sequence alignment of human TIR domain-containing molecules. The regions corresponding to that around the residues of Pro200, a critical residue for TIR-TIR interaction, and Cys216 of MyD88 are shown. (H) HEK293 cells stably expressing TLR2 were transiently transfected with GyrB-fused wild-type MyD88 or mutant MyD88 with each cysteine residue replaced with a serine residue together with the NF- κ B-driven luciferase gene and incubated for 16 h. At 6 h before the end of incubation, cells were treated with 200 μ M SNAP. Cells were stimulated with 100 ng/ml Pam₂CSK₄ for 6 h, and then luciferase activity was measured. Each value is the mean \pm SD ($n = 3$). (*, $P < 0.05$ for comparison with the wild-type group).

and residual proteins were observed even at 45 min after stimulation when cells were pretreated with SNAP (Fig. 2B). The effect of NO was not altered in the presence of the guanylate cyclase inhibitor ODQ or the cyclic-GMP-dependent protein kinase inhibitor KT5823 (data not shown). Notably, LPS-induced degradation of IRAK-1 and I κ B α in HAECs was prevented by the irreversible thiol modification by *N*-ethylmaleimide (Fig. 2C), implying that the effect of NO on the signaling events depends on modification of cysteine residues.

To test whether NO alters the MyD88-dependent signal events, we utilized the MyD88-GyrB construct. Under coumer-

mycin treatment of cells stably expressing MyD88-GyrB, MyD88-GyrB undergoes dimerization and mimics TLR-triggered typical MyD88-dependent functions, such as the activation of MAPKs and IKKs and secretion of proinflammatory cytokines (11). We found that coumermycin-dependent induction of NF- κ B activation in HEK293 cells was suppressed by pretreatment with cells with NO donors (Fig. 3A). Coumermycin could induce phosphorylation of I κ B α at Ser32 and Ser36, the target residues of IKK β involved in degradation of I κ B α (51), but SNAP pretreatment could suppress induction of the response (Fig. 3B). SNAP also suppressed coumermycin-induced phos-

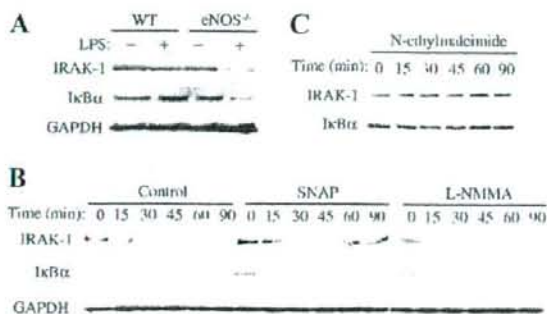


FIG. 2. NO suppresses LPS-induced degradation of IRAK-1 and I κ B α . (A) Anesthetized wild-type (WT) and eNOS^{-/-} mice intratracheally received *E. coli* LPS and mechanical ventilation. After 30 min of administration, lung was excised and then lysed for immunoblotting with anti-IRAK-1, anti-I κ B α , and anti-GAPDH antibodies. (B) HAECs pretreated with 1 mM L-NMMA for 12 h or 0.25 mM SNAP for 1 h were stimulated with 10 ng/ml *E. coli* LPS for the indicated periods. The expression levels of IRAK-1 and I κ B α were determined by immunoblot analysis. (C) HAECs pretreated with 0.1 mM N-ethylmaleimide for 10 min were stimulated with 10 ng/ml *E. coli* LPS for the indicated periods. The expression levels of IRAK-1 and I κ B α were determined by immunoblot analysis.

phorylation of MAPKs (data not shown). The coumermycin-dependent dimerization of MyD88-GyrB induced interaction with TRAF6, consistent with a previous study (10), and we found that this interaction was reduced by SNAP treatment (Fig. 3C).

We found that NO and alteration of Cys113 and Cys216 residues of MyD88 did not alter the interaction of overexpressed MyD88 with IRAK-1 in HEK293 cells (Fig. 3D). However, NO clearly attenuated TLR4 stimulus (LPS)-dependent induction of MyD88-IRAK-1 interaction (Fig. 3E), suggesting that NO targets upstream signaling events of IRAK-1. It has been known that the recruitment of MyD88 to TLR2 or TLR4 is mediated by binding of the sorting adaptor TIRAP to the membrane phosphatidylinositol 4,5-bisphosphate, followed by interaction of MyD88 with TIRAP through TIR-TIR interaction (23). We examined whether NO affects the interaction of MyD88 with TIRAP. Overexpressed MyD88 interacted with TIRAP in HEK293 cells (Fig. 3F). We found that treatment of the cells with the NO donor GSNO attenuated the interaction (Fig. 3F). We further investigated whether S-nitrosylation of residues 113 and 216 is involved in MyD88-TIRAP interaction. Alteration of Cys residues did not affect the interaction (Fig. 3G). However, SNAP-induced attenuation of the interaction was reduced in the Cys216 mutant (Fig. 3G).

In HAECs, MyD88 was enriched with filamentous cytoskeletal structures and partly colocalized with β -actin (Fig. 4A). Additionally, a large part of MyD88 stably expressed in HEK293 cells was found in the cytoskeletal fraction (Fig. 4B). These findings are consistent with results of a previous study showing that MyD88 associates with β -actin in HeLa cells (22). Cytoskeletal MyD88 was separated from IRAK-1, which was found only in the cytoplasm (Fig. 4B), suggesting that MyD88 is maintained as an inactive state in cytoskeleton. We found that SNAP treatment altered such cytoskeletal localization of MyD88 into the cytoplasm (Fig. 4C). We further investigated

the subcellular localization of MyD88 after TLR4 stimulation in 293-TLR4/MD2-CD14 cells stably expressing Flag-MyD88-GyrB. After LPS treatment, a part of MyD88 was transported to the cytoplasmic membrane from the cytoskeleton (Fig. 4D). However, SNAP treatment retarded such LPS-induced transportation of MyD88 (Fig. 4D). Although native PAGE analysis revealed that MyD88 formed a protein complex (more than 480 kDa) in HEK293 cells, SNAP treatment resulted in reduction in the size of the complex to approximately 450 kDa or 250 kDa, accompanied by an increase in concentration (Fig. 4E). Moreover, higher concentrations of SNAP further altered the complex to render a monomer (approximately 35 kDa) (Fig. 4E), implying disruption of functional MyD88 protein complex by NO.

Thus, NO has a capability to obstruct the MyD88 signaling pathway through disruption of the multiple steps of protein interactions.

NO reversibly suppresses the MyD88 signaling events. S-nitrosylated proteins are known to undergo denitrosylation, by which regulatory effects of NO are conferred to control protein functions. We therefore investigated how S-nitrosylation and denitrosylation affect MyD88-mediated signaling events. For this purpose, we utilized GSH because GSH had a capability to denitrosylate MyD88 (Fig. 1C). We found that GSH restored NO-induced impaired interaction of MyD88 with TIRAP (Fig. 4F). Furthermore, the NO-induced cytoplasmic localization of cytoskeletal MyD88 was restored by treatment of the cells with GSH (Fig. 4G). Thus, these results suggest that S-nitrosylation alters the MyD88 pathway, and antioxidants or oxidoreductases restore such NO-derived actions, probably through denitrosylation.

NO reversibly suppresses TLR-mediated cellular responses. HAECs responded to multiple bacterial TLR agonistic molecules, Pam₃CSK₄ (for TLR1/TLR2), MALP-2 (for TLR2/TLR6), LPS (for TLR4), flagellin (for TLR5), and IL-1 β , all of which are known to activate MyD88-dependent signaling to induce production of the NF- κ B-driven chemokine IL-8 after stimulation for 3 h (Fig. 5A). Predepletion of endogenous NO by L-NMMA resulted in a significant increase in IL-8 production induced by each stimulator (Fig. 5A), indicating that endogenous NO has a suppressive effect on the TLR-mediated cellular response. Furthermore, IL-8 production by each stimulator was suppressed in the presence of SNAP (Fig. 5A). The effect of NO donors was not altered in the presence of ODQ or KT5823 (data not shown). We investigated whether such a suppressive effect of NO can be restored because S-nitrosylation is a reversible protein modification. However, it is difficult to examine the effects of antioxidants or oxidoreductases because the TLR signaling pathway is greatly affected by reactive oxygen species generated from NADPH oxidases (27, 34, 48). Indeed, treatment of cells with ascorbic acid, GSH, NAC, or the NADPH oxidase inhibitor DPI greatly impaired LPS-induced IL-8 production in HAECs (Fig. 5B). We therefore attempted to address whether the effect of NO is transient or persistent. For this purpose, HAECs were pretreated with SNAP for 1 h and washed two times to remove the NO donor. Then, at various times afterwards, the cells were stimulated with LPS and IL-8 production was measured. NO suppression of IL-8 production was gradually neutralized or restored in a

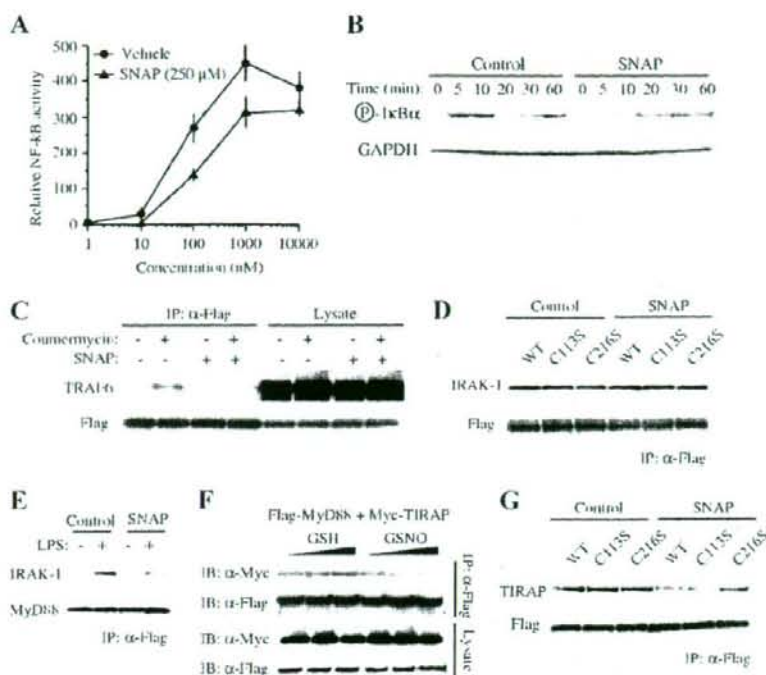


FIG. 3. Effects of NO on TLR-mediated signaling events. (A) HEK293 cells stably expressing MyD88-GyrB were transiently transfected with an NF- κ B-driven luciferase gene and incubated for 24 h. Cells were pretreated with or without 0.25 mM SNAP for 1 h and then treated with 1 μ M coumestrolin for 3 h. Then, luciferase activity was measured. Each value is the mean \pm SD ($n = 3$). (B) HEK293 cells stably expressing MyD88-GyrB were pretreated with or without 0.25 mM SNAP for 1 h and then treated with 1 μ M coumestrolin for the indicated periods. The phosphorylation of I κ B α at Ser32/Ser36 was detected by immunoblot analysis. (C) HEK293 cells stably expressing MyD88-GyrB were pretreated with or without 0.25 mM SNAP for 1 h and then treated with 1 μ M coumestrolin for 20 min. Then, cell lysates were immunoprecipitated (IP) with anti-Flag antibody, followed by immunoblotting with anti-Flag and anti-TRAF6 antibodies. (D) HEK293 cells transiently expressing Flag-tagged wild-type (WT) or Cys residue (113 or 216) replacement MyD88 together with IRAK-1 were treated with 500 μ M SNAP for 1 h. Then, cell lysates were immunoprecipitated with anti-Flag antibody, followed by immunoblotting with anti-Flag and anti-IRAK-1 antibodies. (E) 293-TLR4/MD2-CD14 cells stably expressing Flag-MyD88-GyrB were treated with or without 500 μ M SNAP for 1 h and then stimulated with 100 ng/ml LPS for 20 min. Then, cell lysates were immunoprecipitated with anti-Flag antibody, followed by immunoblotting with anti-IRAK-1 and anti-Flag antibodies. (F) HEK293 cells transiently expressing Flag-tagged MyD88 together with Myc-tagged TIRAP were treated with GSH or GSNO (0, 100, and 500 μ M) for 1 h. Then, cell lysates were immunoprecipitated with anti-Flag antibody, followed by immunoblotting (IB) with anti-Flag and anti-Myc antibodies. (G) HEK293 cells transiently expressing Flag-tagged wild-type or Cys residue (113 or 216) replacement MyD88 together with Myc-tagged TIRAP were treated with 500 μ M SNAP for 1 h. Then, cell lysates were immunoprecipitated with anti-Flag antibody, followed by immunoblotting with anti-Flag and anti-IRAK-1 antibodies.

time-dependent manner, although the restrictive effect continues for several hours (Fig. 5C).

NO suppresses acute-phase immune responses to LPS in vivo. To explore how NO regulation of MyD88-dependent signaling reflects innate immune or proinflammatory responses in vivo, we utilized a popular animal model of sepsis induced by i.p. administration of LPS. We first investigated the cytokine responses as the major hallmark of innate immune responses. MIP-2 is known as one of the early LPS-responsive genes, the mRNA expression of which indeed showed a rapid rise and reached a peak within 1 h after LPS stimulation in mouse peritoneal macrophages (Fig. 6A). In contrast, IL-6 is known as a late LPS-responsive gene, the expression of which showed a gradual rise and reached a peak more than 4 h after stimulation (Fig. 6A). We determined the amounts of MIP-2 and IL-6 produced in the abdominal cavity 2 h after LPS administration in wild-type, eNOS^{-/-}, and iNOS^{-/-} mice. Interest-

ingly, eNOS^{-/-} mice exhibited the most intensive production of MIP-2 (Fig. 6B). On the other hand, the most prominent production of IL-6 was observed in iNOS^{-/-} mice (Fig. 6C). In contrast to these results, there was no significant difference in the amounts of MIP-2 and IL-6 production when the fluids were collected 12 h after LPS administration (data not shown). Thus, eNOS and iNOS at least exert a suppressive effect on early cytokine responses in vivo.

We further examined LPS-induced febrile response as a hallmark of acute-phase responses of inflammation. LPS is known to act as a pyrogen to induce TLR4-dependent polyphasic fever (44). The major initiator of LPS fever is generated prostaglandin E₂, which stimulates thermoregulatory neurons and elevates body core temperature (3). LPS can directly induce prostaglandin E₂ generation through the MyD88-dependent signaling pathway (47). The first-phase febrile response in eNOS^{-/-} mice occurred significantly earlier than that in wild-

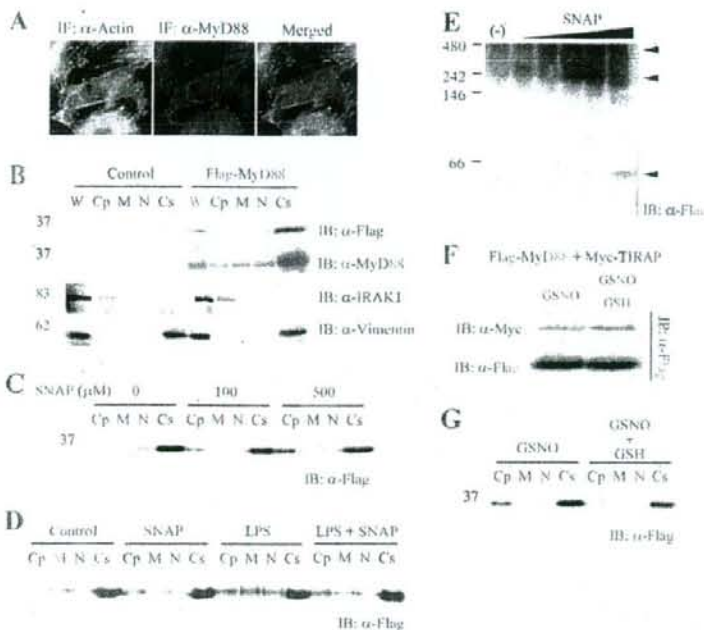


FIG. 4. Reverse of the suppressive effect of NO by GSH. (A) IIAECs were fixed and stained immunofluorescently (IF) with anti- β -actin antibody (green, left), anti-MyD88 antibody (red, middle), and Hoechst33342 (blue, right). (B) Parental HEK293 cells and HEK293 cells stably expressing Flag-MyD88 were fractionated into the cytoplasm (Cp), cytoplasmic membrane (M), nucleus (N), and cytoskeleton (Cs). Whole-cell lysates (W) were also obtained. The fractions were assessed by immunoblotting (IB) with anti-Flag, anti-MyD88, anti-IRAK-1, and anti-vimentin antibodies. (C) HEK293 cells stably expressing Flag-MyD88 were treated with the indicated concentration of SNAP for 1 h and fractionated into each fraction. The fractions were assessed by immunoblotting with anti-Flag antibody. (D) 293-TLR4/MD2-CD14 cells stably expressing Flag-MyD88-GyrB were treated with or without 500 μ M SNAP for 1 h and then stimulated with 100 ng/ml LPS for 20 min. Cells were then fractionated into each fraction. The fractions were assessed by immunoblotting with anti-Flag antibody. (E) HEK293 cells stably expressing Flag-MyD88 were treated with or without SNAP (10, 50, 125, 250, and 500 μ M) for 1 h. Then, cell lysates were assessed by blue native PAGE and immunoblotting with anti-Flag antibody. (F) HEK293 cells transiently expressing Flag-MyD88 and Myc-tagged TIRAP were treated with 500 μ M GSN0 for 1 h and then with or without 500 μ M GSH for 15 min. Then, cell lysates were immunoprecipitated with anti-Flag antibody, followed by immunoblotting with anti-Flag and anti-Myc antibodies. (G) HEK293 cells stably expressing Flag-MyD88 were treated with the 500 μ M GSN0 for 1 h and then with or without 500 μ M GSH for 15 min. The cells were fractionated into each fraction, followed by immunoblotting with anti-Flag antibody.

type or *iNOS*^{-/-} mice (Fig. 6D), indicating that NO from eNOS suppresses the initiation of the response. However, eNOS deficiency did not alter the magnitude of febrile response compared with that for wild-type mice, suggesting that the suppressive effect of NO is not persistent. On the other hand, a transient decrease in fever was found in wild-type and eNOS^{-/-} mice at about 70 min after LPS administration but not in *iNOS*^{-/-} mice (Fig. 6D), indicating that NO from *iNOS* suppresses promotion of the response. Thus, these results suggest that NO generated from eNOS and *iNOS* exerts a suppressive effect on acute-phase inflammatory responses to LPS *in vivo*, probably through S nitrosylation.

DISCUSSION

Our findings imply that MyD88-dependent signaling events are affected by S nitrosylation, by which innate immune signal transduction might be reduced in living organisms. The effect of NO is transient and is restored by antioxidants or oxidoreductases, in which protein denitrosylation plays an important role. Although the physiological significance of such reg-

ulation of TLR signal transduction is unsettled, NO is likely to retard signaling cascades through S nitrosylation, by which rapid and precipitous signaling reactions may be initially or inductively relieved. Such an effect may reflect an adequate regulation of acute-phase inflammatory responses, leading to limitation of the degree of inflammation and resolution of inflammation.

We found that the suppressive effect of NO on TLR-mediated cellular responses was transient and degraded in a time-dependent manner (Fig. 5C). The specificity of NO regulation may be conferred by the spatial regulation of S nitrosylation within or between proteins and the stimulus-coupled temporal regulation through denitrosylation (14). Signal transduction by ligand-receptor interactions is thought to trigger denitrosylation, restoring substantial protein functions. For example, reduction of the functions of caspase-3 and IKK β by S nitrosylation is restored by FasL-1 as interaction and TNF- α -TNFR interaction, respectively (30, 39). Thus, it is possible that TLR ligation-dependent protein denitrosylation also facilitates the restoration of NO suppression although the mechanism of denitrosylation has been poorly studied. Protein denitrosyla-

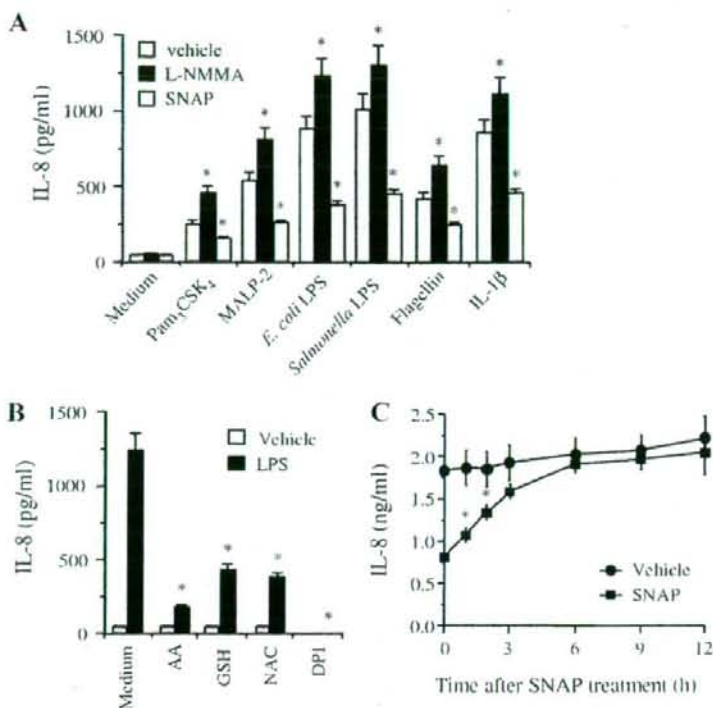


FIG. 5. NO reversibly suppresses TLR-mediated cellular responses. (A) HAECS pretreated with 1 mM L-NMMA for 12 h or 0.25 mM SNAP for 1 h were stimulated with 1 μ g/ml Pam₃CSK₄, 100 nM MALP-2, 10 ng/ml *E. coli* LPS, 10 ng/ml *Salmonella* LPS, 5 μ g/ml flagellin, and 10 ng/ml IL-1 β for 3 h. Then, production of IL-8 was determined by ELISA. Each value is the mean \pm SD ($n = 3$). (*, $P < 0.01$ for comparison with the vehicle group). (B) HAECS were stimulated with 10 ng/ml *E. coli* LPS for 3 h in the presence or absence of 1 mM ascorbic acid (AA), 1 mM GSH, 1 mM NAC, or 20 μ M DPI. Then production of IL-8 was determined by ELISA. Each value is the mean \pm SD ($n = 3$). (*, $P < 0.01$ for comparison with the vehicle group). (C) HAECS were treated with 0.25 mM SNAP for 1 h and then at various times afterwards stimulated with 10 ng/ml *E. coli* LPS for 3 h. Then, production of IL-8 was determined by ELISA. Each value is the mean \pm SD ($n = 3$). (*, $P < 0.01$ for comparison with the vehicle group).

tion is catalyzed by antioxidants or oxidoreductases, including ascorbic acid, thioredoxin-thioredoxin reductase, superoxide dismutase GSH, and GSNO reductase (14, 15, 50, 53). S-nitrosylated MyD88 can be denitrosylated in the presence of ascorbic acid and GSH in vitro (Fig. 1C). Although it is still unclear how TLR ligation activates cellular redox activity, LPS has a potential to activate cellular redox activity and transition of GSH into GSNO (41). More details of TLR-mediated protein S-nitrosylation and denitrosylation should be investigated in future studies.

TLR ligation can initiate recruitment of MyD88 to the receptor complex through TIR-TIR interaction. In the case of TLR2 and TLR4, the sorting adaptor TIRAP is essentially required to recruit MyD88 (23). MyD88 then dissociates from the receptor complex and recruits IRAK-1 (and IRAK-4) through death domain (DD)-DD interaction, inducing TRAF6-mediated signaling events and ubiquitin ligation to IRAK-1 or I κ B α , followed by proteasomal degradation (1). Nevertheless, how MyD88 can be initially controlled to be recruited to TLRs has remained unclear. We found that a large part of cellular MyD88 existed in the cytoskeleton and associated with β -actin (Fig. 4A and B), wherein MyD88 formed a complex dissociated from IRAK-1 (Fig. 4B).

Our finding suggests that MyD88 preferentially interacts with the cytoskeleton as an inactive form, followed by release into the cytoplasm and recruitment to TLRs after ligation-dependent actin rearrangement. Indeed, inhibition of actin rearrangement by cytochalasin D suppresses LPS-induced signal transduction and cytokine production (5). In addition, cytochalasin D also alters TIRAP recruitment to the cytoplasmic membrane (23). NO restriction of MyD88 function may be achieved through disruption of the protein complex and dissociation of MyD88 from the actin cytoskeleton to the cytoplasm (Fig. 4A to E). NO also reduces the interaction of MyD88 with TIRAP (Fig. 3). These effects may ultimately result in mitigated potential for the ligation-dependent recruitment of MyD88 to TLRs.

We found that S-nitrosylation of MyD88 plays some roles in NO modulation of TLR signal transduction. Interestingly, eight of the nine cysteine residues of MyD88 are concentrated in the TIR domain, but the C-terminal DD contains no cysteine residue, suggesting that cysteine modification affects TIR-TIR interaction but not DD-DD interaction. Indeed, NO attenuated the interaction of MyD88 with TIRAP but not that with IRAK-1 (Fig. 3). This result is supported by the result found by Xiong et al. (52) showing that SNAP treatment did

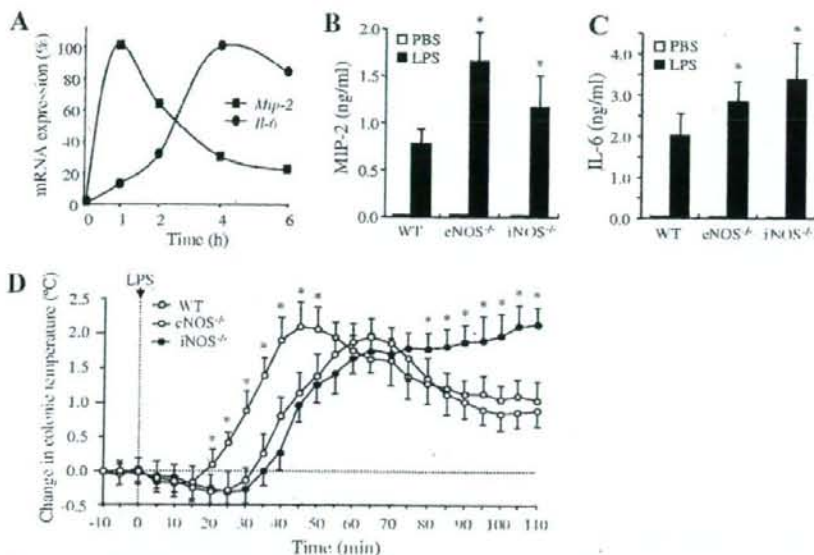


FIG. 6. Roles of eNOS and iNOS in early innate immune responses in vivo. (A) Peritoneal macrophages from wild-type mice were stimulated with 100 ng/ml LPS and 10 ng/ml gamma interferon for the indicated periods. Then, expression levels of mRNAs of *Mip-2* and *Il-6* were determined by quantitative PCR. Percent mRNA expression was calculated by taking the maximum values of mRNA levels of *Mip-2* and *Il-6* as 100%. (B, C) Wild-type (WT), eNOS^{-/-}, and iNOS^{-/-} mice were i.p. treated with LPS. After 2 h, PBS was injected into the abdominal cavity and fluids were collected for measurement of the amounts of MIP-2 (B) and IL-6 (C) by ELISA. Each value is the mean \pm SD ($n = 6$). (*, $P < 0.01$ for comparison with wild-type mice). (D) Wild-type, eNOS^{-/-}, and iNOS^{-/-} mice were i.p. treated with LPS. The colonic temperature was monitored at 5-min intervals during a period of 10 min before and 120 min after LPS administration. Each value is the mean \pm SD ($n = 6$). (*, $P < 0.01$ for comparison with wild-type mice).

not affect the interaction of MyD88 with IRAK-1 in mouse macrophages. Although S-nitrosylation of the Cys216 residue of MyD88 may participate in the NO regulation of TLR signal transduction, it is likely that this modification does not have a dominant effect, because Cys216 was not essential for activation of downstream signaling (Fig. 1G). NO modification of Cys216 may yield a slight structural change in the base of the TIR domain, resulting in slightly reduced interaction with a counterpart TIR domain of TIRAP. Alternatively, NO may antagonize other reversible modifications, such as palmitoylation or disulfide bonding to a counterpart molecule, leading to transient impairment of MyD88 functioning. Also, other unknown mechanisms for MyD88 regulation may be negatively influenced by S-nitrosylation.

We found that eNOS and iNOS differentially regulate LPS-induced acute-phase immune responses in vivo (Fig. 6). Although the amount of NO derived from eNOS is comparatively small, NO is steadily generated from endothelial cells as a vasodilatory gas that continually maintains an antiproliferative and antiapoptotic environment in vasculatures (16). Simultaneously, eNOS increases the amounts of cellular S-nitrosylated proteins and circulating NO donors by nitrosylating GSH and albumin (19). Such functions of NO from eNOS may systemically reduce cellular reactivity to a TLR stimulus to maintain a weak tolerance, which may lead to prevention of a rapid rise of inflammation. In contrast, NO from iNOS is generated in large quantities and exerts a strong antimicrobial action, although NO from eNOS also has an antimicrobial property (4,

29). The large amount of NO derived from iNOS is thought to disrupt cellular signaling cascades, resulting in anti-inflammatory or immunosuppressive effects. Such functions of NO from iNOS may regionally reduce cellular reactivity to TLR recognition of pathogens to initiate an inducible tolerance, which may transiently prevent promotion of excess inflammatory responses, although excess NO production ultimately results in nitrosative stress and apoptotic cell death (12, 26).

Our study proposes that TLR signal transduction involves an oxidative protein modification by NO and its redox regulation. NO may exert other effects, such as activation of cyclic-GMP-dependent signaling, on TLR signaling events, but such effects may not be dominant, at least in the acute-phase innate immune responses. Further investigations will be necessary to clarify more details about the relationship between such NO regulation and physiological or pathophysiological innate immune responses.

ACKNOWLEDGMENTS

We thank Margaret K. Olfenbutten (Emory University School of Medicine) for providing DNA constructs of human MyD88 and TIRAP.

This work was supported by grants-in-aid for Scientific Research on Priority Areas (19041079 to T. Inoh) and for Young Scientists (B) (18791363 to T. Inoh), provided by the Ministry of Education, Culture, Sports, Science and Technology, Japan.

REFERENCES

1. Akira, S., and K. Takeda. 2004. Toll-like receptor signalling. *Nat. Rev. Immunol.* 4:499–511.

2. Akira, S., S. Uematsu, and O. Takeuchi. 2006. Pathogen recognition and innate immunity. *Cell* 124:783-801.
3. Blatteis, C. M., E. Schic, and S. Li. 2000. Pyrogen sensing and signaling: old views and new concepts. *Clin. Infect. Dis.* 31(S1):168-177.
4. Bogdan, C. 2001. Nitric oxide and the immune response. *Nat. Immunol.* 2:907-916.
5. Dai, Q., and S. B. Pruetz. 2006. Ethanol suppresses LPS-induced Toll-like receptor 4 clustering, reorganization of the actin cytoskeleton, and associated TNF- α production. *Alcohol Clin. Exp. Res.* 30:1436-1444.
6. Dinarello, S. J., H. Haseler, V. Nels, and A. M. Zeller. 1997. Suppression of apoptosis by nitric oxide via inhibition of interleukin-1 β -converting enzyme (ICE)-like and cysteine protease protein (CPP)-32-like proteases. *J. Exp. Med.* 185:601-607.
7. Farrar, M. A., L. Allierol, and R. M. Peroutter. 1996. Activation of the Raf-1 kinase cascade by c-myc-induced dimerization. *Nature* 383:178-181.
8. Forrester, M. L., M. W. Foster, and J. S. Stamler. 2007. Assessment and application of the biotin switch technique for examining protein S-nitrosylation under conditions of pharmacologically induced oxidative stress. *J. Biol. Chem.* 282:13977-13983.
9. Forstermann, U., J. P. Boissel, and H. Kleinert. 1998. Expressional control of the 'constitutive' isoforms of nitric oxide synthase (NOS I and NOS III). *FASEB J.* 12:773-790.
10. Ghosh, S., and M. Karin. 2002. Missing pieces in the NF- κ B puzzle. *Cell* 109:881-96.
11. Hacker, H., V. Redecke, B. Blagoev, I. Kratchmarova, L. C. Hsu, G. G. Wang, M. P. Kamts, E. Raz, H. Wagner, G. Hacker, M. Mann, and M. Karin. 2006. Specificity in Toll-like receptor signaling through distinct effector functions of TRAF3 and TRAF6. *Nature* 439:204-207.
12. Hara, M., R. N. Agrawal, S. F. Kim, M. B. Cascio, M. Fujimuro, Y. Ozeki, M. Takahashi, J. H. Cheah, S. K. Tankou, L. D. Hester, C. D. Ferris, S. D. Hayward, S. H. Snyder, and A. Sawa. 2002. S-nitrosylated GAPDH initiates apoptotic cell death by nuclear translocation following Shb1 binding. *Nat. Cell Biol.* 7:665-674.
13. Hayden, M. S., A. P. West, and S. Ghosh. 2006. NF- κ B and the immune response. *Oncogene* 25:6758-6780.
14. Hess, D. T., A. Matsumoto, S. O. Kim, H. E. Marshall, and J. S. Stamler. 2005. Protein S-nitrosylation: preview and parameters. *Nat. Rev. Mol. Cell Biol.* 6:150-166.
15. Hoffmann, J., J. Haendeler, A. M. Zeiber, and S. Dinarello. 2001. TNF α and oxLDL reduce protein S-nitrosylation in endothelial cells. *J. Biol. Chem.* 276:41383-41387.
16. Igarashi, L. J. 2002. Nitric oxide as a unique signaling molecule in the vascular system: a historical overview. *J. Physiol. Pharmacol.* 53:503-514.
17. Into, T., J. Dohkan, M. Inomata, M. Nakashima, K. Shiluta, and K. Matsushita. 2007. Synthesis and characterization of a dipalmitoylated lipospike derived from pathogenic lipoproteins of *Mycobacterium pneumoniae*. *Infect. Immun.* 75:2253-2259.
18. Into, T., Y. Kanno, J. Dohkan, M. Nakashima, M. Inomata, K. Shibata, C. J. Lowenstein, and K. Matsushita. 2007. Pathogen recognition by Toll-like receptor 2 activates Weibel-Palade body exocytosis in human aortic endothelial cells. *J. Biol. Chem.* 282:8134-8141.
19. Into, T., and K. Shiluta. 2005. Apoptosis signal-regulating kinase 1-mediated sustained p38 mitogen-activated protein kinase activation regulates mycoplasma lipoprotein- and staphylococcal peptidoglycan-triggered Toll-like receptor 2 signaling pathways. *Cell. Microbiol.* 7:1305-1317.
20. Iwasaki, A., and R. Medzhitov. 2004. Toll-like receptor control of the adaptive immune responses. *Nat. Immunol.* 5:987-995.
21. Jaffrey, S. R., H. Erdjument-Bromage, C. D. Ferris, P. Tempst, and S. H. Snyder. 2001. Protein S-nitrosylation: a physiological signal for neuronal nitric oxide. *Nat. Cell Biol.* 3:193-197.
22. Jamin, F., K. Burns, J. Tschopp, T. E. Martin, and S. Fakan. 1998. Ultrastructural distribution of the death-domain-containing MyD88 protein in HeLa cells. *Exp. Cell Res.* 243:67-75.
23. Kagan, J. C., and R. Medzhitov. 2006. Phosphoinositide-mediated adaptor recruitment controls Toll-like receptor signaling. *Cell* 125:943-955.
24. Karin, M., and F. R. Greten. 2005. NF- κ B linking inflammation and immunity to cancer development and progression. *Nat. Rev. Immunol.* 5:749-759.
25. Kelleher, Z. T., A. Matsumoto, J. S. Stamler, and H. E. Marshall. 2007. NOS2 regulation of NF- κ B by S-nitrosylation of p65. *J. Biol. Chem.* 282:30667-30672.
26. Kim, K. M., P. K. Kim, Y. G. Kwon, S. K. Bai, W. D. Nam, and Y. M. Kim. 2002. Regulation of apoptosis by nitrosative stress. *J. Biochem. Mol. Biol.* 35:127-133.
27. Laroux, F., S. X. Romero, L. Weizler, P. Engel, and C. Terhorst. 2005. MyD88 controls phagocytic NADPH oxidase function and killing of gram-negative bacteria. *J. Immunol.* 175:5596-5600.
28. Lawrence, T., M. Bebbien, G. Y. Liu, V. Nizet, and M. Karin. 2005. IKK α limits macrophage NF- κ B activation and contributes to the resolution of inflammation. *Nature* 434:1138-1143.
29. MacKicking, J., Q. W. Xie, and C. Nathan. 1997. Nitric oxide and macrophage function. *Annu. Rev. Immunol.* 15:323-350.
30. Mannick, J. B., A. Hausladen, L. Liu, D. T. Hess, M. Zeng, Q. X. Miao, L. S. Kane, A. J. Gow, and J. S. Stamler. 1999. Fas-induced caspase denitrosylation. *Science* 284:651-654.
31. Mannick, J. B., C. Schonhoff, N. Papeta, P. Ghafourfar, M. Szibor, K. Fang, and B. Gaston. 2001. S-Nitrosylation of mitochondrial caspases. *J. Cell Biol.* 154:1111-1116.
32. Marshall, H. E., D. T. Hess, and J. S. Stamler. 2004. S-nitrosylation: physiological regulation of NF- κ B. *Proc. Natl. Acad. Sci. USA* 101:8841-8842.
33. Marshall, H. E., and J. S. Stamler. 2001. Inhibition of NF- κ B by S-nitrosylation. *Biochemistry* 40:1688-1693.
34. Matsuzawa, A., K. Saegusa, T. Noguchi, C. Sadamitsu, H. Nishitoh, S. Nagai, S. Koyasu, K. Matsumoto, K. Takeda, and H. Ichijo. 2005. ROS-dependent activation of the TRAF6-ASK1-p38 pathway is selectively required for TLR4-mediated innate immunity. *Nat. Immunol.* 6:587-592.
35. Medzhitov, R., P. Preston-Hurlbert, and C. A. Janeway, Jr. 1997. A human homologue of the Drosophila Toll protein signals activation of adaptive immunity. *Nature* 388:394-397.
36. Medzhitov, R., P. Preston-Hurlbert, E. Kopp, A. Stadler, C. Chen, S. Ghosh, and C. A. Janeway, Jr. 1998. MyD88 is an adaptor protein in the b1001-H1-1 receptor family signaling pathways. *Mol. Cell* 2:253-258.
37. Mignin, S. M., E. Palsom-McDermott, A. Doune, C. Jefferies, E. Pinteaux, K. Banaban, C. Murphy, P. Moynagh, M. Yamamoto, S. Akira, N. Rothwell, D. Golenbock, K. A. Fitzgerald, and L. A. O'Neill. 2007. NF- κ B activation by the Toll-H1-1 receptor domain protein MyD88 adaptor-like is regulated by caspase-1. *Proc. Natl. Acad. Sci. USA* 104:3372-3377.
38. Park, H. S., J. W. Yu, J. H. Cho, M. S. Kim, S. H. Huh, K. Ryon, and E. J. Choi. 2004. Inhibition of apoptosis signal-regulating kinase 1 by nitric oxide through a thiol redox mechanism. *J. Biol. Chem.* 279:7584-7590.
39. Reynaert, N., L. K. Kleiss, S. H. Korn, N. Vos, A. S. Guila, E. F. Wouters, A. van der Vliet, and Y. M. Janssens-Heineiner. 2004. Nitric oxide represses inhibitory κ B kinase through S-nitrosylation. *Proc. Natl. Acad. Sci. USA* 101:8945-8950.
40. Richardson, G., and N. Benjamin. 2002. Potential therapeutic uses for S-nitrosothiols. *Clin. Sci. (London)* 102:99-105.
41. Rubin, D. B., G. Iteznik, E. A. Weiss, and P. R. Young. 2000. Non-protein thiols flux to S-nitrosothiols in endothelial cells: an LPS redox signal. *Shock* 14:200-207.
42. Stamler, J. S., S. Lamas, and F. C. Fang. 2001. Nitrosylation: The prototypic redox-based signaling mechanism. *Cell* 106:675-683.
43. Stamler, J. S., D. I. Simon, J. A. Osborne, M. E. Mullins, O. Jaraki, T. Michel, D. J. Singel, and J. Loscalzo. 1992. S-nitrosylation of proteins with nitric oxide: synthesis and characterization of biologically active compounds. *Proc. Natl. Acad. Sci. USA* 89:444-448.
44. Steiner, A. A., S. Chakravarti, A. Y. Rudaya, M. Herkenham, and A. A. Romanovsky. 2006. Bacterial lipopolysaccharide fever is initiated via Toll-like receptor 4 on hematopoietic cells. *Blood* 107:4000-4002.
45. Steiner, A. A., A. Y. Rudaya, A. I. Ivanov, and A. A. Romanovsky. 2004. Febrile signaling to the brain does not involve nitric oxide. *Br. J. Pharmacol.* 141:1204-1213.
46. Thoma-Uzynski, S., S. Stenger, O. Takeuchi, M. T. Ochoa, M. Engele, P. A. Sieling, P. F. Barnes, M. Rollinghoff, P. L. Boleskel, M. Wagner, S. Akira, M. V. Nurgard, J. T. Belisle, P. J. Godowski, R. R. Bloom, and R. L. Modlin. 2001. Induction of direct antimicrobial activity through mammalian Toll-like receptors. *Science* 291:1544-1547.
47. Uematsu, S., M. Matsumoto, K. Takeda, and S. Akira. 2002. Lipopolysaccharide-dependent prostaglandin E₂ production is regulated by the glutathione-dependent prostaglandin E₂ synthase gene induced by the Toll-like receptor 4 MyD88/NF- κ B pathway. *J. Immunol.* 168:5811-5816.
48. Ulevano, M., S. Dusi, D. Lissandrini, R. Badolati, P. Marzi, L. Gibaldi, E. Borroni, A. Calleri, M. Donini, A. Plebani, L. Notarangelo, T. Musso, and S. Sozzani. 2004. Toll receptor-mediated regulation of NADPH oxidase in human dendritic cells. *J. Immunol.* 173:5749-5756.
49. Wang, W., A. Mirra, B. Paole, S. Falk, M. S. Lucia, S. Fayal, and R. Schrier. 2004. Endothelial nitric oxide synthase-deficient mice exhibit increased susceptibility to endotoxin-induced acute renal failure. *Am. J. Physiol. Renal Physiol.* 287:F1044-F1048.
50. West, M. B., B. G. Hill, Y. T. Xuan, and A. Bhatnagar. 2006. Protein glutathiolation by nitric oxide: an intracellular mechanism regulating redox protein modification. *FASEB J.* 20:1715-1717.
51. Woroniez, J. D., X. Gao, Z. Cao, M. Roth, and D. V. Goeddel. 1997. I κ B kinase β NF- κ B activation and complex formation with I κ B kinase α and NIK. *Science* 278:866-869.
52. Xiong, H., C. Zhu, F. Li, R. Hegazi, K. He, M. Babyatsky, A. J. Bauer, and S. E. Plevy. 2004. Inhibition of interleukin-12 p40 transcription and NF- κ B activation by nitric oxide in murine macrophages and dendritic cells. *J. Biol. Chem.* 279:10776-10783.
53. Yamawaki, H., J. Haendeler, and B. C. Berk. 2003. Thioresosin: a key regulator of cardiovascular homeostasis. *Circ. Res.* 93:1029-1033.