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3 shown in Fig. 1. All components of the stereotaxic apparatus consisted of
4
5 non-magnetic materials that fixed the head position of the rats during data
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7 acquisition. The RF coil was designed to have an impedance of 50Ω at a resonance
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9 frequency of 127.76 MHz. An additional single-turn surface coil of 62 mm diameter
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11 was also developed for better homogeneity, and was used for a single slice of
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13 DSC-MRI. The RF power had to be reduced in these coils below that of the standard
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15 human head coil because of the diameter of the small coil. The transmission signal
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17 was therefore attenuated to 20 dB, which allowed the use of automated scanner
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19 software including the calibration of the RF transmission power and receiver gains.
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21 All rats were fixed on the stereotaxic apparatus. They were placed at the center of
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23 the gantry and oriented with the cranio-caudal axis perpendicular to a static
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25 magnetic field. Their heads were positioned inside the coil toward the cranio-caudal
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27 direction.
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38 T_1W images were obtained with a conventional two-dimensional fast spin
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40 echo (2D-FSE) sequence. The repetition time (TR) was 1500 ms [10]. The echo time
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42 (TE) was fixed at 14 msec. The echo train length (ETL) was 3. The field of view (FOV)
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44 was set at $40 \times 30 \text{ mm}^2$, the slice thickness at 1.5 mm, the slice gap at 0.5 mm, the
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46 number of excitations (NEX) at 10, and the band width (BW) at 31.3 kHz. The
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48 acquired matrix (256×160) was interpolated, and null pixels were added in k-space to
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50 produce square matrices of 256×256 . The acquisition time was 10 min 3 sec.
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T_2W images were obtained with a 2D-FSE and the following imaging parameters: TR 4100 ms, TE 128 ms, ETL 14, FOV 40×30 mm, slice thickness 1.5 mm, slice gap 0.5 mm, NEX 8, BW 31.3 kHz, acquired matrix 256×160, zero-filled to 256×256, phase direction ventral-dorsal, and an acquisition time of 11 min 2 sec.

TOF-MRA was performed by use of a three-dimensional flow-compensated spoiled gradient recalled (3D-SPGR) sequence prepared with magnetization transfer and with TR 53 ms, TE 5.5 ms, flip angle (FA) 45 degrees, BW 16 kHz, FOV 8×6 cm, slice thickness 0.2 mm, 1 acquired slab of 512×512×64, a voxel resolution 0.156 × 0.156 × 0.2 mm³, NEX 1, and an acquisition time of 21 min 46 sec.

DSC images were obtained following the intravenous administration of Gd-DTPA to the T_2^* -weighted gradient echo dynamic images. A bolus of Gd-DTPA (0.1 mmol/kg) was injected manually into the tail vein with a 22-gauge catheter via one meter of polyethylene tubing (PE50, internal diameter: 0.58 mm / outer diameter: 0.965 mm, Becton Dickinson and Company, Franklin Lakes, NJ, USA), and was followed by an additional administration of saline (1.0 ml). A multi-shot EPI with the number of shots at 2 was employed for improving EPI distortion and temporal resolution. Imaging parameters were TR 142 ms, TE 22.1 ms, FA 20°, FOV 40×40 mm, and a matrix size of 64×64, leading to a pixel size of 0.625×0.625 mm². The slice thickness was 3 mm in single slice around the hypothalamus (HT). The temporal resolution was 0.284 sec per image, and the acquisition time was 1 min 15

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3 sec. This assessment was repeated three times at intervals of 40 min and 10 min,
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5 corresponding to the 1st-2nd and 2nd-3rd scans, respectively.
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8 9 10 *Data analysis*

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12 All MRI images were reconstructed on the same workstation provided for the
13 GE Signa 3-Tesla scanner as used for the clinical programs. Then the images were
14 transferred to a Linux workstation. Lastly, data analysis was carried out by use of
15 in-house and commercial software program.
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23 To evaluate the T₁W and T₂W image quality, we calculated the
24 contrast-to-noise ratio (CNR) with an inter-tissue method [25-27] as follows: $CNR =$
25 $(\pi/2)^{1/2} (SI_a - SI_b) / SI_{air}$, where SI_{air} represent air-mean-signal-intensity, SI_a and SI_b
26 represent the signal intensities of tissue a and tissue b , respectively.
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35 Angiograms were created by generation of the partial maximum intensity
36 projection (MIP) with a commercial software programs (Virtual Place Liberty (VPL),
37 AZE Co. Ltd. Tokyo, Japan). Visible middle cerebral artery (MCA) branches and left
38 to right differences of MCA were carefully investigated.
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47 For the DSC image slice section containing the internal carotid artery (ICA)
48 and/or MCA, a series of images were carefully observed. A region of interest (ROI)
49 was selected in the MCA region from which the AIF was obtained. Attention was
50 paid, with the help of other anatomic information. For avoiding the susceptibility
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3 artifacts caused by air in the trachea, the area of the arterial circle of Willis was
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5 excluded in the definition of the AIF. The anterior cerebral artery was also excluded,
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7 because of possible susceptibility effects attributed to the venous blood. A Gaussian
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9 filter of 1.1 mm full width at half maximum (FWHM) was applied to all dynamic
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11 images. The time-versus-signal-intensity curves (TICs) were converted to the
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13 Gd-DTPA concentration according to Eq. (1) given in the Appendix. Functional
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15 mapping images of the mean transit time (MTT), cerebral blood volume (CBV), and
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17 cerebral blood flow (CBF) were carried out with the deconvolution method [28]. The
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19 theory is described in detail in the Appendix. For ROI analysis, images of 64×64
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21 matrix size were converted to 256×256 with use of a sinc interpolation function.
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32 Results

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35 T_1W and T_2W images reconstructed with a spatial resolution of 0.156×0.188
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37 $\times 1.5$ mm are shown in Fig. 2. White matter could be discriminated from cortical and
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39 deep-gray-matter regions. Locations of small anatomic features such as the caudate
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41 putamen (CPu); striatum, the CC, and the hippocampus (HC) could also be identified
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43 in both T_1W and T_2W anatomic images. The CNR between the HC and CC was 15.6
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45 and 9.8, corresponding to the T_1W and T_2W images shown in Fig.2. The CNR
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47 between the HC and IC was 23.2 and 13.6 respectively, in which underestimation
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49 may exist in CNR attributed to the contamination of signal from the globus pallidus.
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3 Results for MIP images obtained with contrast-free TOF-MRA are shown in
4 Fig. 3. Coronal MIP images around the HT of 5 mm thickness are shown in Fig.3 (A).
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6 In this figure, the slice section contained ICAs and MCA. The MCA, the cortical
7 branches in both the left and right hemispheres, can be identified. It is important to
8 note that the anatomic structure of both cortical MCA arteries is different between
9 the right and left hemispheres. The ROI for the AIF was selected in the MCA region
10 indicated by the arrows in Fig.3 (A) and (B).
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22 A typical example of a DSC-MRI image is shown in Fig.4. Distortion of the
23 DSC-MRI images is visible in Fig.4(A) toward the phase direction. The magnified
24 area in dynamic images around the MCA region, which is shown as a rectangle in
25 Fig.4(A), is displayed in Fig.4(B). There were several pixels which indicated temporal
26 changes in pixel contrast as a function of time, and these were reflected by Gd-DTPA
27 negative enhancement. The pixel signal intensity varied as shown in Fig.4(C), and
28 the curve shown was employed for estimating the AIF. Figure 5 shows the TIC in
29 this area together with the TIC for the whole brain region, obtained from each of the
30 three scans. The curves were visually reproducible in terms of shape, height and
31 width of the curves around the peak, as well as the tail height at end of the scan. It
32 should also be noted that the baseline before each injection of Gd-DTPA was
33 consistent, even though the 2nd and the 3rd curves should have been affected by the
34 previous injection of Gd-DTPA.
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Functional mapping images of CBF, CBV, and MTT calculated according to the theory described in the Appendix are shown in Figure 6. Images obtained from this sequential assessment appeared to be reasonably clear, although slightly noisy, and were consistent among the scans. The absolute CBF (mean \pm S.D.) in the cortical-gray-matter area (mainly the parietal cortex (Pt)) was 24.04 ± 2.88 , 17.75 ± 3.34 , and 31.87 ± 7.27 ml/g/min, corresponding to the first, second, and third scans, respectively. The Pt-to-HT CBF ratio was 46.7%, 51.5%, and 43.0%, corresponding to the first, second, and third scans, respectively. The CBV was 0.49 (0.44), 0.50 (0.35), and 0.47 (0.41) ml/ml in the Pt (the HT) region, corresponding to the first, second, and third scans, respectively. The MTT in the same regions was 1.22 (2.51), 1.72 (2.29), and 0.92 (2.16) seconds, corresponding to each of the three scans. Among the three injections, the absolute MTTs (mean \pm S.D.) were 1.29 ± 0.40 sec in the Pt and 2.32 ± 0.17 sec in the HT region.

Discussion

This study demonstrated that our system of a human whole-body 3-Tesla MRI, fitted with an in-house solenoid coil developed for small animals, can provide morphologic and functional images of the rat brain in vivo. The quality of T₁-W and T₂-W images obtained with a scan duration of approximately 10 min was better than those obtained in previous studies in which 1.5-Tesla clinical MRI scanners [3] were employed. The neocortex and large subcortical structures, such as the Thal and HC, are readily recognized in their topographic relationship to the CC, the

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3 ventricular system, and the subarachnoid space [2]. In T_1 -W images, the
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5 cerebrospinal fluid (CSF)-containing spaces are visible as hypo-intense, and the
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7 arterial flow showed a signal loss caused by the so-called flow void effects, which is
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9 typically seen in spin echo sequence [2]. In T_2 -W images, the CSF showed a bright
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11 white intensity (prolonged T_2 relaxation time), whereas the myelinated white matter
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13 showed black intensity (short T_2 relaxation time). The white-matter tracts such as
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15 the IC and CC were clearly visible in both T_1 -W and T_2 -W images, with better quality
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17 compared with previous reports employing 1.0- and 1.5-Tesla clinical MRI scanners
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28 MRA images also clearly showed the structure of distal MCA branches. The
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30 MRA findings of inter-hemispheric differences with regards to MCA in the SD rat
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32 were also consistent with a previous report on Wistar rats [29], which indicated the
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34 left-to-right asymmetrical structure in three out of 10 Wistar rats using a 7-Tesla
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36 MRI scanner dedicated to the small animal imaging [29]. Our MRA images are
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38 superior to those in previous work [11] that employed a clinical 3-Tesla MRI scanner
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40 and that only showed the major cerebral arteries and the carotid arteries because
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42 they focused mainly on validating occlusion models [11]. The superior quality of our
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44 MRA images can largely be attributed to the type of RF coil we used. Ours is a
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46 three-turn solenoid coil that covers only the cerebral area, whereas the previous work
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48 [11] employed a single-turn coil (diameter of 6.4 cm and length of 10 cm) that covered
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50 the whole head, including the brain and the neck. Additionally, the prolonged
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3 acquisition period in our study (almost twenty minutes) versus the significantly
4 shorter acquisition period in the previous study [11] (almost four minutes) may have
5 been a factor that led to higher-quality MRA images.
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12 An important point with regard to this study is that DSC-MRI images of
13 reasonable quality can also be obtained with a clinical MRI scanner at 3 Teslas on
14 rats. In addition, we were able to extract the AIF from the rat brain, which is an
15 important accomplishment. Selection of an ROI in the MCA region successfully
16 provided the AIF. Spatial distortion or a susceptibility artifact was not visible in our
17 observation. Signal changes were obtained during DSC-MRI following a single dose
18 of Gd-DTPA. During this study, the dedicated transmitting and receiving RF coils
19 were considered essential for obtaining a reasonable SNR. Our study was performed
20 with the same sequence and the same dose rate (0.1 mmol/kg) of Gd-DTPA that are
21 commonly used in clinical examinations. Moreover, it was performed with high (1.17
22 mm³)-resolution dynamic imaging.
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42 The quantitative images of CBF were consistent with a previous report on the
43 use of [¹⁴C] iodoantipyrine [30]. Namely, the Pt-to-HT contrast in CBF was 43-52 %
44 in this study, which is close to the values reported by Bloom et al. [30] of 44-58 %.
45 Although the absolute CBF and CBV values in our study were different from
46 previous ones [13, 30, 31], unsure scaling factors of each ones were canceled out in
47 calculating the MTT with Eq.(5) [32] (shown also in the Appendix). An analogous
48 value for the MTT was represented in previous work [31, 33]. In the theoretical
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3 deconvolution step, the level of plasma gadolinium which was not influenced after
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5 previous injection by calculated subtraction with peak to base line with $(S(0)/S(t))$ of
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7 Eq.(1) (Appendix).
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12 We noticed that the absolute CBF and CBV values were overestimated,
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14 which suggests that some limitations apply, such as the partial volume effect (PVE)
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16 caused by insufficient spatial resolution as compared with the anatomic structure of
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18 the MCA. Detection of AIF with repeat injection was performed (Fig.5, right row).
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20 The major MCA diameter was approximately 0.5 mm at the maximum as evaluated
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22 from Fig.3, which suggested that the measured AIF is largely influenced by the PVE
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24 [24]. Also, the differences of absolute value may be reflected by the fact that a
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26 non-linear relationship exists between the signal intensity and the contrast agent
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28 concentration. Previous reports proposed methods for the non-linearity correction
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30 for brain tissue [34] and AIF [35]. Further studies are needed to confirm the accuracy
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32 and the reproducibility [36]. Image distortion caused by dielectric effects [37] and/or
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34 EPI distortion [38] are also additional error sources, and should be investigated
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36 systematically.
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48 Along with the improved quality of acquired original dynamic images, the
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50 mapping image quality would also improve. For better detection of the dynamic
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52 susceptibility contrast that is caused by T_2^* signals, the optimization of TE, FA, and
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54 the acquisition matrix should be investigated. We speculate that the multi-channel
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56 phased array coil and parallel imaging techniques would reduce the level of
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3 distortion. The DSC-MRI in this study was obtained only for a single slice. Further
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5 careful attention is needed to multi-slice imaging in order to minimize the inflow
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7 effects [35]. In our study, the contrast concentration $C(t)$ curves including AIF's
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9 varied slightly among the three injections. A sophisticated injector system which
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11 need to be MR-compatible, may improve the variation.
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18 A dedicated high-magnetic-field scanner equipped with a dedicated small
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20 bore is the optimal device for small-animal imaging. However, such systems are not
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22 commonly available. The system developed in this study might serve as a low-cost
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24 solution or an alternative. The use of the present system provides an opportunity for
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26 using the same imaging platform available for clinical studies for small-animal
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28 imaging [7]. This would allow us understanding the pathophysiological status from
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30 MRI signals using animal models with various diseases. More importantly,
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32 optimizing several scan parameters which have been limited in clinical patients may
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34 be performed easily on small animals with this system. In particular, the
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36 reproducibility of CBF assessment with DSC-MRI, which has been reported the
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38 limitation in clinical studies [19, 36], may be improved by performance of a
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40 systematic evaluation of each scan parameter when this system is used on small
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42 animals rather than clinical patients. With the addition of a high strength insert
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44 gradient coil [39] which allow for thinner slices and much faster read-out, the system
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46 performance can be enhanced and the spatial resolution with an acceptable SNR can
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48 be improved. The use of adapting coils can be an effective solution for those who
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3 operate MR scanners for human subjects and intend to gain experience [40] in
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5 pre-clinical research.
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10 **Conclusion**
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13 In this pre-clinical study on rats, reasonable image quality for morphologic
14 information was obtained with T₁-W, T₂-W, and contrast-free TOF-MRA images,
15 accomplished by use of a human whole-body 3-Tesla MRI scanner and newly
16 developed solenoid coil. In DSC-MRI, it visualized transient signal changes with a
17 single dose of Gd-DTPA, and with the same sequences which have been commonly
18 used in clinical examinations. A human whole-body 3-Tesla MRI scanner and
19 dedicated coil make it possible to detect the AIF in the MCA region of Wistar rats.
20 High-resolution DSC-MRI was accomplished with a clinical scanner, but the spatial
21 resolution with an acceptable SNR was insufficient for the rat brain. Although there
22 might be some remaining issues with regard to AIF, we have shown the potential of
23 DSC-MRI in our study.
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42 **Appendix**
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44 *Calculation of functional mapping images from DSC-MRI*
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47 The observed TIC $S(t)$ was converted to a time-versus-concentration curve
48 (TCC) $C(t)$ by the following equation [16, 36]
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$$C(t) = k \cdot \int R_2^*(t) = -k \cdot \ln(S(0)/S(t)) / TE, \quad (1)$$

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3 where $\Delta R2^*$ is the change in the T_2^* relaxation rate and k is a constant. In this
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5 study, it was assumed that $k=1$. $S(0)$ is the precontrast (baseline) signal and $S(t)$ is
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7 the measured signal at time t . The next step was to fit this first-pass period of TCC to
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9 a gamma-variate function:

$$10 \quad C(t)=a(t-b)^k \exp(-(t-b)/d), \quad (2)$$

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12 where a , b , c , and d were determined by a nonlinear least-square fitting. To minimize
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14 the effects of the re-circulation of the contrast agent, data were neglected in the fit if
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16 these concentrations were less than 50 % of the maximum after the peak of the TCC.
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18 The fitted tissue TCC $Ct(t)$ was deconvolved by the fitted AIF $C_{AIF}(t)$ by use of a
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20 singular value decomposition with a block-circulant deconvolution matrix (b-SVD)
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22 method [28] and according to the equation

$$23 \quad CBF \cdot R(t) = Ct(t) \otimes^{-1} C_{AIF}(t), \quad (3)$$

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25 where \otimes^{-1} represents the deconvolution operator, and $R(t)$ is a residue function
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27 representing the tissue response to an instantaneous bolus. $CBF \cdot R(t)$ was
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29 estimated by deconvolving $Ct(t)$ by $C_{AIF}(t)$ using b-SVD, and then CBF was
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31 determined as the maximum value of the obtained $CBF \cdot R(t)$.
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46 The CBV was calculated as follows:

$$47 \quad CBV = \int_0^{\infty} C_i(t)dt / \int_0^{\infty} C_{AIF}(t)dt. \quad (4)$$

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5 **Acknowledgments**
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8 The authors would like to express our appreciation to reviewers, Editors and
9 Editorial assistant of Radiological Physics and Technology for their invaluable advice
10 to improve our manuscript. This study was supported by a grant for research on
11 Advanced Medical Technology from the Ministry of Health, Labour and Welfare,
12 Japan. We would like to thank the VPL released by AZE Ltd. (Tokyo, Japan) and the
13 software library by the Oxford University Centre for Functional MRI of the Brain.
14 We are grateful to the staffs at the National Cardiovascular Center for their
15 invaluable contribution and efforts. Last but not least, our thanks go to Miss Atra
16 Ardekani (a summer intern from McGill University in Montreal, Quebec, Canada).
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3 **Figure Captions**
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7 **Figure 1**

8 The head fixation system for small animals, fitted with an MRI coil for RF
9 transmission and reception. The coil is typically a three-turn solenoid; it can also be a
10 single-turn coil, as shown on the top left of this figure.
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16 **Figure 2**

17 T₁-W images (top) and T₂-W images (bottom) obtained from an SD rat, with the
18 following anatomic locations as indicated: the somato-sensory cortex (SS), corpus
19 callosum (CC), parietal cortex (Pt), external capsule (EC), caudate putamen (CPu),
20 internal capsule (IC), hippocampus (HC), thalamus (Thal), auditory cortex (Au),
21 amygdala (AM), hypothalamus (HT), internal carotid arterial system (ICAs),
22 trigeminal nerve (TN), interpeduncular cistern (IPC), dorsal third ventricle (D3V),
23 and the lateral ventricle (LV). These slice sections resemble the distance of -1.5 mm
24 (left) and -3.5 mm (right), respectively, in appearance from the Bregma.
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34 **Figure 3**

35 Partial maximum-intensity-projection maps of contrast-free time-of-flight MRA
36 obtained for a Sprague-Dawley rat. Maps represent areas around the Thal (A), right
37 hemisphere (B), and left hemisphere (C).
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42 Note: the white arrows indicate the selected ROI level (as described in the
43 Discussion). AZACA: Azygos anterior cerebral artery.
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47 **Figure 4**

48 Typical time-frame images obtained from Gd-bolus tracking T₂*-weighted dynamic
49 images (after the first injection). A white rectangle was placed in the MCA area (A),
50 and its magnified images were visualized as (a), (b), (c), (d), and (e) in (B). TIC within
51 the pixels is indicated by white arrows in (B), and the data were then plotted in a
52 graph (C). Typical data points were extracted from the first injection at the times of
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3 9.2 (a), 16.4 (b), 18.1 (c), 20.1 (d), and 33.9 (e) sec. In (A), the phase direction is shown
4 as a white arrow.
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9 **Figure 5**

10 TIC dynamic changes in the whole brain are shown in the first injection (A), the
11 second injection (B), and the third injection (C). For each injection, a one-pixel ROI
12 was selected and observed, as shown for the first (D), second (E) and third (F)
13 injections, respectively. The negative enhancement changes in the rate from peak to
14 base line on signal intensity were 16.8%(A), 10.0 %(B), and 10.3 %(C) at the whole
15 brain (average 12.3, SD 3.9). The fraction of peak-to-base at selected ROIs were
16 19.2 %(D), 14.5 %(E), and 16.9 %(F) (average 16.9, SD 2.3), for the first, second, and
17 third injections, respectively.
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27 **Figure 6**

28 CBF maps using the AIF from MCA, pictured in a gray scale from 0 to 4 ml/g/min, in
29 the first column on the left. MTT maps are shown in the next column, followed by the
30 CBV maps with 0 to 2 ml/g/min in the third and last column. Deconvolution was
31 carried out pixel by pixel with b-SVD and then smoothed to 0.15625×0.15625 mm
32 in-plane resolution from acquired voxel resolution of a $0.625 \times 0.625 \times 3$ mm (1.17
33 mm^3) in DSC-MRI.
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