

physiological range when examined at the end of the surgical preparation as well as at the end of the experiment. The body temperature of each animal was maintained at $\sim 38^{\circ}\text{C}$ with a heating pad. AP was measured using a high-fidelity pressure transducer (Millar Instruments, Houston, TX) inserted from the right femoral artery to the aortic arch.

We isolated bilateral carotid sinuses from the systemic circulation by ligating the internal and external carotid arteries and other small branches originating from the carotid sinus region. Isolated carotid sinuses were filled with warmed physiological saline via catheters inserted through the common carotid arteries. Intra-CSP was controlled by a servo-controlled piston pump (model ET-126A, Labworks, Costa Mesa, CA). Bilateral vagal and aortic depressor nerves were sectioned at the neck to minimize reflexes from the cardiopulmonary region and from the aortic arch.

We exposed the left renal sympathetic nerve retroperitoneally and attached a pair of stainless steel wire electrodes (Bioflex wire AS633, Cooner Wire, Chatsworth, CA) to record SNA. The nerve bundle peripheral to the electrodes was tightly ligated and crushed to eliminate afferent signals from the kidney. The nerve and electrodes were secured with silicone glue (Kwik-Sil, World Precision Instruments, Sarasota, FL). The preamplified nerve signal was band-pass filtered at 150–1,000 Hz, full-wave rectified, and low-pass filtered with a cutoff frequency of 30 Hz to quantify the nerve activity.

With the rabbit in the prone position, the sacrum, left ankle, and knee were clamped with a custom-made apparatus to prevent body trunk and hindlimb movement during muscle stretch. The left triceps surae muscle, Achilles tendon, and calcaneus bone were exposed. The left triceps surae muscle was isolated from the surrounding tissue. The Achilles tendon was severed from the calcaneus bone and attached to a force transducer (Load Cell LUR-A-SA1, Kyowa Electronic Instruments, Tokyo, Japan). During muscle stretch, the other side of the force transducer was connected to a 5-kg weight via a pulley.

Protocols. To obtain operating pressure values, the carotid sinus baroreflex negative feedback loop was effectively closed by adjusting CSP to AP. Mean AP (and thus mean CSP) at steady state was treated as the operating pressure under control conditions. We then performed muscle stretch for 1 min while the carotid sinus baroreflex was effectively closed. Mean AP during the last 10 s of muscle stretch was treated as the operating pressure under muscle stretch conditions.

To estimate the baroreflex dynamic characteristics, CSP was assigned either high (+20 mmHg) or low (–20 mmHg) pressure values around the operating pressure according to a binary white noise sequence. The switching interval of the binary white noise signal was set at 500 ms so that the CSP power spectrum was fairly flat up to 1 Hz. We confirmed that the muscle stretch produced a sustained SNA increase for at least 7 min (58). To limit the maximum duration of muscle stretch within this time period, a 6-min CSP perturbation was performed twice using different binary sequences, and the two sets of data were pooled for analyses under both control and muscle stretch conditions. The order of control and muscle stretch conditions was randomized across the animals.

Data analysis. We recorded CSP, muscle tension, SNA, and AP at a sampling rate of 200 Hz using a 12-bit analog-to-digital converter. Data were stored on a dedicated laboratory computer system.

To estimate the neural arc transfer function of the carotid sinus baroreflex, we treated CSP as the input and SNA as the output of the system. In the peripheral arc transfer function, we treated SNA as the input and AP as the output of the system. In the total loop transfer function, we treated CSP as the input and AP as the output of the system. Data analysis was started from 90 s after the initiation of each trial to process the stationary portion of data without the effects of transition from closed-loop CSP waveform to open-loop binary white noise CSP input and the transition from nonstretch to stretch of muscle mechanoreceptors. The input-output data pairs were resampled at 10 Hz and segmented into 50%-overlapping bins of 1,024 points each. For each segment, a linear trend was subtracted, and a

Hanning window was applied. A fast Fourier transform was performed to obtain the frequency spectra of the input and output signals. The ensemble averages of input power spectral density [$S_{xx}(f)$], output power spectral density [$S_{yy}(f)$], and cross-spectral density between the input and output [$S_{xy}(f)$] were obtained over eight segments derived from two sets of data, where f represents frequency. Finally, we calculated the transfer function from input to output [$H(f)$] using the following equation (27):

$$H(f) = \frac{S_{xy}(f)}{S_{xx}(f)} \quad (1)$$

Hereinafter, we denote the modulus as the dynamic gain of the transfer function. To quantify the linear dependence between input and output signals in the frequency domain, we calculated a magnitude-squared coherence function [$\text{Coh}(f)$] using the following equation (27):

$$\text{Coh}(f) = \frac{|S_{xy}(f)|^2}{S_{xx}(f)S_{yy}(f)} \quad (2)$$

The coherence value ranges from zero to unity. Unity coherence indicates perfect linear dependence between input and output signals, whereas zero coherence indicates total independence between the two signals.

To facilitate an intuitive understanding of the transfer function, the step response corresponding to the transfer function was also calculated as follows. The system impulse response was derived from the inverse Fourier transform of $H(f)$. The step response was obtained from the time integral of the impulse response.

Statistical analysis. All data are presented as means \pm SD. Because the amplitude of SNA varied depending on recording conditions, such as the physical contact between the nerve and electrodes, SNA was presented in arbitrary units (au). Neural and peripheral arc transfer functions were normalized in each animal so that the average gain values below 0.03 Hz in the control trial became unity. To compare the transfer functions between two conditions, a transfer gain value at 0.01 Hz ($G_{0.01}$), 0.1 Hz ($G_{0.1}$), 0.5 Hz ($G_{0.5}$), and 1 Hz (G_1) were calculated. In the step response of the neural arc, the steady-state step response at 50 s (S_{50}), the negative peak value (S_{peak}), and the time to negative peak (T_{peak}) were calculated. The effects of muscle stretch on these parameters were examined using the paired t -test. Differences were considered significant when $P < 0.05$.

RESULTS

Figure 1 shows a typical time series of CSP, muscle tension, SNA, and AP under control (left) and muscle stretch (right) conditions. Although the same binary sequence was applied for two conditions in each animal, different binary sequences were applied for different animals to reduce possible systematic errors in system identification caused by a bias in whiteness specific to a selected binary sequence. The mean CSP during muscle stretch conditions (Fig. 1, right) was set higher than that during the control conditions (Fig. 1, left) to mimic the increase in the operating pressure during muscle stretch under baroreflex closed-loop conditions (i.e., the AP increase by muscle stretch increases the mean input pressure to the baroreceptors). Muscle stretch increased mean levels of SNA and AP compared with control conditions during the experiment (Table 1).

Figure 2 shows the transfer functions of the neural (left) and peripheral (right) arcs estimated under the control and muscle stretch conditions; gain plots (top), phase plots (middle), and $\text{Coh}(f)$ (bottom) are also presented. The thin and thick solid lines in Fig. 2 indicate control and muscle stretch conditions, respectively. In the neural arc, the dynamic gain increased as

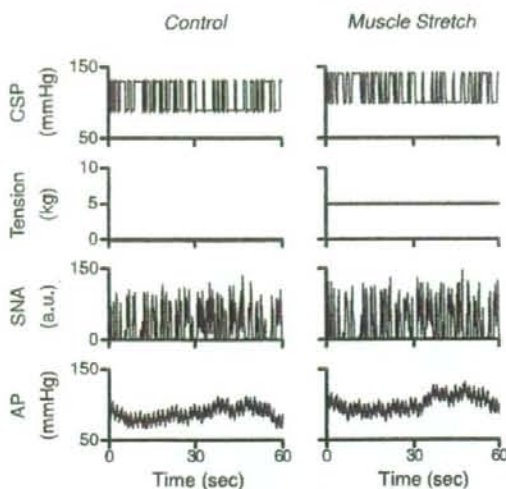


Fig. 1. Typical time series of intracarotid sinus pressure (CSP), muscle tension, sympathetic nerve activity (SNA; in arbitrary units (au)), and arterial pressure (AP) under control (left) and muscle stretch (right) conditions. CSP was perturbed according to a binary white noise sequence. Muscle stretch increased mean levels of SNA and AP under muscle stretch conditions compared with the control conditions.

the frequency of input modulation increased under both conditions, indicating derivative characteristics of the neural arc. Muscle stretch caused an approximately parallel upward shift of the gain plot. The phase approached $-\pi$ radians (-180°) at the lowest frequency (0.01 Hz) under both conditions, reflecting the negative feedback character of the baroreflex neural arc (i.e., an increase in CSP decreased SNA). Phase plots were nearly superimposed between the two conditions. Coherence

Table 1. Mean levels and CVs of CSP, SNA, and AP at 1, 2, 4, and 6 min under control and muscle stretch conditions

	Time			
	1 min	2 min	4 min	6 min
CSP				
Control	95 ± 18	96 ± 18	95 ± 18	96 ± 18
CV	13 ± 2	12 ± 3	11 ± 3	14 ± 2
Muscle stretch	114 ± 15*	115 ± 16*	113 ± 16*	114 ± 15*
CV	11 ± 2	11 ± 1	10 ± 2	12 ± 2
SNA				
Control	102 ± 4	99 ± 5	100 ± 4	99 ± 4
CV	46 ± 11	45 ± 9	43 ± 9	47 ± 9
Muscle stretch	133 ± 22*	129 ± 21*	127 ± 17*	126 ± 17*
CV	48 ± 11	47 ± 8	44 ± 9	49 ± 10
AP				
Control	90 ± 21	89 ± 20	88 ± 16	88 ± 18
CV	7 ± 2	6 ± 2	6 ± 2	6 ± 2
Muscle stretch	107 ± 26*	105 ± 22*	104 ± 15*	101 ± 15*
CV	7 ± 3	6 ± 3	6 ± 3	7 ± 2

Values are means ± SD; $n = 7$. CSP, carotid sinus pressure (in mmHg); SNA, sympathetic nerve activity (in %); AP, arterial pressure (in mmHg); CV, coefficient of variation. Mean and CV values were calculated from 30-s data ending at each time point. * $P < 0.05$ vs. control.

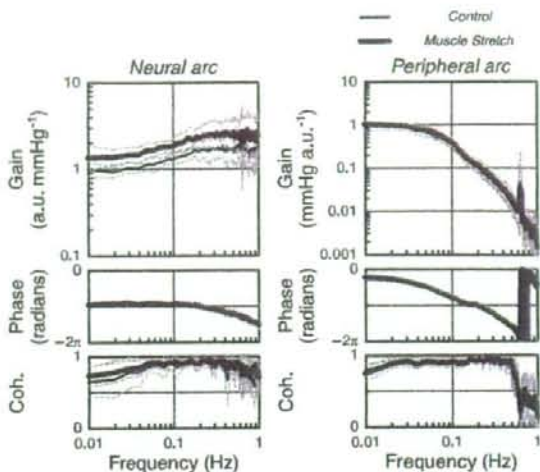


Fig. 2. Transfer functions of the neural (left) and peripheral (right) arcs under control and muscle stretch conditions. In the neural arc, the input was CSP and the output was SNA. In the peripheral arc, the input was SNA and the output was AP. The mean level of CSP input to the neural arc was set higher under muscle stretch conditions than under control conditions to mimic the physiological condition (i.e., baroreflex closed-loop conditions). Gain plots (top), phase plots (middle) and coherence (Coh) functions (bottom) are shown. Thin and thick solid lines indicate control and muscle stretch conditions, respectively. In the neural arc (left), muscle stretch caused an approximately parallel upward shift of the gain plot. Solid and dashed lines represent means and means ± SD values, respectively.

values did not differ between both conditions. In the peripheral arc, the dynamic gain decreased in the frequency range from 0.05 to 1 Hz as the frequency of input modulation increased under both conditions, indicating the low-pass characteristics of the peripheral arc. The phase approached 0 radians at the lowest frequency (0.01 Hz) under both conditions, reflecting the fact that an increase in SNA increased AP. The phase lagged with increasing frequency up to 1 Hz. The gain plot, phase plot, and Coh(f) did not differ between both conditions.

Table 2 summarizes gains of the transfer functions. In the neural arc, $G_{0.01}$, $G_{0.1}$, $G_{0.5}$, and G_1 were higher under muscle

Table 2. Gains of the transfer functions

	Control	Muscle Stretch
	Neural arc	
$G_{0.01}$, au/mmHg	1.01 ± 0.23	1.44 ± 0.56*
$G_{0.1}$, au/mmHg	1.30 ± 0.11	1.86 ± 0.37*
$G_{0.5}$, au/mmHg	1.77 ± 0.64	2.65 ± 1.08*
G_1 , au/mmHg	1.72 ± 0.66	2.72 ± 1.40*
Peripheral arc		
$G_{0.01}$, mmHg/au	1.08 ± 0.06	1.06 ± 0.20
$G_{0.1}$, mmHg/au	0.37 ± 0.09	0.42 ± 0.09
$G_{0.5}$, mmHg/au	0.02 ± 0.01	0.02 ± 0.01
G_1 , mmHg/au	0.004 ± 0.001	0.004 ± 0.002
Total loop		
$G_{0.01}$, mmHg/mmHg	1.08 ± 0.18	1.53 ± 0.63*
$G_{0.1}$, mmHg/mmHg	0.48 ± 0.12	0.81 ± 0.31*
$G_{0.5}$, mmHg/mmHg	0.04 ± 0.04	0.06 ± 0.04*
G_1 , mmHg/mmHg	0.006 ± 0.003	0.013 ± 0.013

Values are means ± SD; $n = 7$. $G_{0.01}$, $G_{0.1}$, $G_{0.5}$, and G_1 , dynamic gains at 0.01, 0.1, 0.5, and 1 Hz, respectively; au, arbitrary units. * $P < 0.05$ vs. control.

stretch compared with control conditions. In the peripheral arc, $G_{0.01}$, $G_{0.1}$, $G_{0.5}$, and G_1 were unchanged between control and muscle stretch conditions.

Figure 3 shows the total baroreflex loop transfer functions (CSP to AP) under control and muscle stretch conditions. The thin and thick solid lines in Fig. 3 indicate control and muscle stretch conditions, respectively. The dynamic gain decreased as the frequency of input modulation increased under both conditions, indicating low-pass characteristics. The dynamic gain under muscle stretch conditions was higher than that under control conditions in frequency from 0.01 to 0.5 Hz (Table 2). The phase plot and $\text{Coh}(f)$ did not differ between both conditions.

Figure 4 shows step responses of SNA corresponding to the transfer functions in the neural arc shown in Fig. 2. The initial drop in the SNA response as well as the steady-state response was augmented during muscle stretch (Table 3). T_{peak} did not differ between control and muscle stretch conditions (Table 3).

DISCUSSION

The key new findings of the present study are as follows. Muscle stretch increased the dynamic gain of the carotid sinus baroreflex neural arc as estimated by binary white noise input (Fig. 2). In contrast, the peripheral arc transfer function remained unchanged irrespective of the muscle stretch (Fig. 2). These results suggest that during muscle mechanoreflex activation, the dynamic SNA response to CSP perturbation is augmented.

System identification by the white noise approach. To identify the dynamic characteristics of arterial baroreflex function quantitatively, we described the carotid sinus baroreflex con-

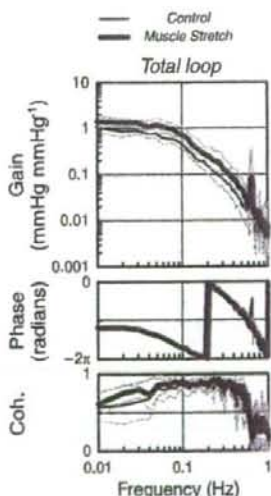


Fig. 3. Total loop transfer functions from CSP to AP under control and muscle stretch conditions. Gain plots (top), phase plots (middle) and coherence functions (bottom) are shown. Thin and thick solid lines indicate control and muscle stretch conditions, respectively. The dynamic gain decreased as the frequency of input modulation increased under both conditions, indicating low-pass characteristics. Muscle stretch caused an approximately parallel upward shift of the gain plot. Solid and dashed lines represent means and means \pm SD values, respectively.

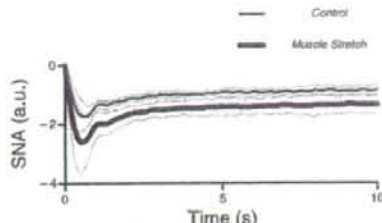


Fig. 4. Step responses corresponding to transfer functions of the neural arc obtained from Fig. 2, showing the SNA response to a 1-mmHg increase in input pressure. Thin and thick solid lines indicate control and muscle stretch conditions, respectively. The initial drop in the SNA response as well as the steady-state response was augmented by the muscle stretch. Solid and dashed lines represent means and means \pm SD values, respectively.

trol of SNA and AP in terms of system identification using the white noise technique. Compared with the traditional approach of testing dynamic properties of the physiological system with step and sine wave stimuli, the white noise approach has definite advantages, as follows (27). First, if a step stimulus is applied, we learn the response of the system to this step and have little notion of the response of the system to any other type of stimulus. If a sinusoidal pulse is applied, then we know the response of the system to such a stimulus and little else. The same applies for any other specific waveform. Theoretically speaking, the system is tested with every possible stimulus in the white noise approach. The white noise stimulus is a very rich stimulus. It should be emphasized that the white noise method is perfectly suited to the analysis of linear systems. As shown in Figs. 2 and 3, high coherence values close to unity indicate the validity of our method for system identification. Second, the identification of the physiological system through the white noise technique is largely unaffected by the types of contaminating noise usually present in such a system. Our study provides the first and quantitative description of the dynamic characteristics of the carotid sinus baroreflex during isolated activation of mechanosensitive afferents from skeletal muscle.

Effects of the muscle mechanoreflex on dynamic characteristics of the carotid sinus baroreflex. The effects of activation of afferents from skeletal muscle, such as those occurring during exercise, on the arterial baroreflex have been extensively studied (5, 13, 29, 42, 43, 49, 58, 59). These studies have demonstrated that the afferent input from muscle resets the baroreflex control of AP, heart rate, and SNA. However, the dynamic characteristics of the arterial baroreflex during isolated activation of muscle mechanosensitive afferents have never been analyzed. In the present study, muscle stretch increased dynamic gain in every frequency (Fig. 2 and Table

Table 3. Parameters of step responses

	Control	Muscle Stretch
S_{50} , a.u.	-1.05 ± 0.30	$-1.69 \pm 0.69^*$
S_{peak} , a.u.	-2.10 ± 0.50	$-3.08 \pm 1.45^*$
T_{peak} , s	0.63 ± 0.21	0.64 ± 0.20

Values are means \pm SD; $n = 7$. A step response is defined as a SNA response to a 1-mmHg change in input pressure. S_{50} , step response at 50 s; S_{peak} , negative peak response; T_{peak} , time to negative peak. * $P < 0.05$ vs. control.

2), whereas it did not affect the peripheral arc. These data are the first to provide quantitative evidence demonstrating that the dynamic SNA response to CSP perturbation is augmented during isolated activation of the muscle mechanoreflex. Although an increase in dynamic gain in the lowest frequency (0.01 Hz) was expected from the results of our previous studies showing an increase in static gain by muscle stretch (58, 59), the information was insufficient to perform a simulation study to examine the effects of muscle stretch on the closed-loop dynamic AP regulation (see *Physiological implications*). The present study extended our previous work by providing additional information on the dynamic interaction over a wide range of frequencies between 0.01 and 1 Hz in the carotid sinus baroreflex.

The static characteristics of the arterial baroreflex determine an operating point of the baroreflex system. Furthermore, the static characteristics described by a modeled sigmoid function provide the parameters of threshold, saturation, and maximal gain at the centering point. However, the static characteristics alone cannot provide the information on the changes over time in the response of the baroreflex system. On the other hand, dynamic analysis techniques such as transfer function analysis estimated by the white noise approach provide information on the stability and quickness of the system response. The dynamic SNA response to baroreceptor pressure input became greater as the frequency of input modulation increased, suggesting derivative characteristics (i.e., high-pass characteristics) of the baroreflex neural arc (Fig. 2, left, thin solid line). In contrast, the dynamic AP response to SNA became smaller as the frequency of SNA modulation increased, indicating low-pass characteristics of the baroreflex peripheral arc (Fig. 2, right, thin solid line). The total loop transfer function (CSP to AP) is determined by a product of the neural and peripheral arc transfer functions (Fig. 3, thin solid line). Therefore, the decreasing slope of dynamic gain in the total loop transfer function was shallower than that in the corresponding peripheral arc. In other words, the fast neural arc effectively compensates for the slow peripheral arc to accelerate dynamic AP regulation by the baroreflex negative-feedback loop (14). During muscle stretch, the dynamic gain in the neural arc was increased by ~50% in every frequency under study (Fig. 2 and Table 2), indicating that the derivative characteristics of the neural arc were maintained. As a result, the effect of the neural arc compensating for the slow AP response was preserved during the activation of muscle mechanoreflex (Fig. 3 and Table 2). Furthermore, the total loop dynamic gain was augmented during the muscle stretch due to the upward shift of the neural arc transfer function.

Because we used passive muscle stretch as the input for the muscle mechanoreflex, the physiological significance of the present results should be interpreted carefully. Several studies have examined the arterial baroreflex control of SNA during static and dynamic exercise. Static and heavy dynamic exercise resets the baroreflex control of SNA to higher SNA levels with an increase in its sensitivity (9, 11, 17, 32). On the other hand, mild to moderate dynamic exercise resets the baroreflex control of SNA without any change in its sensitivity (3, 24, 38). Because the muscle mechanoreflex is activated during mild to moderate dynamic exercise (4), our results indicate that the muscle mechanoreflex may contribute to increasing the baroreflex gain of SNA during mild to moderate dynamic exercise. In

addition to differences in the measured SNA (renal vs. muscle), analytic methods of baroreflex function, modes of mechanoreflex activation, and/or species between the present study and previous studies, the cardiopulmonary baroreflex should be taken into account. Charkoudian et al. (1) demonstrated that increasing central venous pressure via head-down tilt or saline infusion attenuated the baroreflex sensitivity in the control of SNA. The activation of cardiopulmonary baroreceptors induced by increasing central venous pressure may influence the arterial baroreflex control during dynamic exercise (37). In the present study, however, the cardiopulmonary baroreflex did not operate due to bilateral vagotomy.

Previous studies (7, 25) have suggested that the muscle mechanoreflex has a dominant role in pressor reflexes during muscle contraction in anesthetized or decerebrate cats. Although we believe that the mechanoreflex is one of the pressor reflexes during exercise, the functional importance of the muscle mechanoreflex in cardiovascular regulation during exercise in conscious conditions is debatable. Matsukawa et al. (28) recently reported that blockade of the muscle mechanoreflex by gadolinium did not alter AP responses to isometric exercise in conscious cats. Moreover, they found that gadolinium significantly diminished the pressor responses to passive muscle stretch in anesthetized cats. These observations suggest that, under the experimental design, the muscle mechanoreflex would not be activated during exercise or, even if it was activated, it has no functional importance in cardiovascular responses to exercise in conscious conditions. One criticism for the study is that there is always a possibility that changes in the central command in conscious conditions had compensated for the lack of muscle mechanoreflex. Further studies are needed to better understand the role of the muscle mechanoreflex on neural cardiovascular responses during exercise.

High-pass characteristics of the baroreflex neural arc. It is likely that the dynamic characteristics of the baroreflex neural arc actually reflect the intrinsic and synaptic properties of central nervous system neurons and neural circuits that transmit baroreceptor input. However, the central baroreceptor synapses are characterized as a low-pass filter (26). The difference between high-pass characteristics of the neural arc transfer gain and low-pass characteristics of the central baroreceptor synaptic transmission could be attributable to the difference of estimated frequency ranges. Frequency-dependent depression (FFD) of synaptic transmission in the baroreflex central pathways is the phenomenon that the probability of excitatory postsynaptic potentials progressively reduces as the frequency of afferent input increases beyond 1 Hz (2, 33). Although FFD and transfer gain should be discriminated in theory, interactions between FFD and transfer gain may occur when the modulation frequency of afferent fiber stimulation approached the frequency range of FFD. Indeed, Kawada et al. (23) found high-cut characteristics of the baroreflex neural arc in the frequency range above ~1 Hz. In the present study, the transfer gain was derived from 0.01 to 1 Hz. Whether the dynamic interaction between carotid sinus baroreflex and muscle mechanoreflex exists in the frequency range beyond 1 Hz awaits further studies.

Part of the high-pass characteristics in the baroreflex neural arc is attributable to the derivative nature observed in the baroreceptor transduction from CSP input to baroreceptor afferent nerve activity (i.e., mechanoneural transduction) (21).

However, we think there exists high-pass characteristics in the transduction from baroreceptor afferent input to efferent SNA, because the magnitude of high-pass characteristics slightly differs between cardiac and renal SNAs in response to the same baroreceptor pressure perturbation (18).

In an electrical circuit, we can design a high-pass filter only from low-pass filter elements using a feedback loop (Fig. 5). Although the main forward path of the baroreflex neural arc from afferent nerve activity to efferent SNA is considered to be the nucleus tractus solitarius, caudal ventrolateral medulla, and rostral ventrolateral medulla (53), there could be feedback connections between these areas. Therefore, it is possible that synaptic connection has basically low-pass characteristics, whereas the baroreflex neural arc reveals high-pass characteristics as a neural circuit. The speculation also needs to be verified experimentally in the future.

Physiological implications. Under physiological conditions, the baroreflex is closed as a negative feedback system. In the following discussion, we will focus on the effect of the augmentation of dynamic SNA modulation in the neural arc on the closed-loop dynamic AP regulation. Figure 6A illustrates a simulator consisting of the linear neural arc transfer function (H_N) and linear peripheral arc transfer function (H_P) followed by the nonlinear sigmoidal components (see the APPENDIX for details). A closed-loop AP response to a stepwise pressure perturbation (-40 mmHg) with pulsatile pressure was simulated, and the result is shown in Fig. 6B. Muscle stretch shortened the time to 95% of steady state by $\sim 33\%$ from 7.2 to 4.8 s (shaded and solid arrows in Fig. 6B). This result suggests that, under baroreflex closed-loop conditions, the rate of recovery in AP following a pressure perturbation occurs sooner when accompanied by the muscle mechanoreflex. Increasing the quickness of the negative-feedback system can be caused by augmentation and/or acceleration of the open-loop transfer function of the system. In our baroreflex open-loop experiment, S_{50} and S_{peak} in the step responses of SNA were

augmented by the muscle stretch (Fig. 4 and Table 3). On the other hand, T_{peak} did not differ between control and muscle stretch conditions (Fig. 4 and Table 3). These results suggest that the improvement in the quickness of the AP restoration via the baroreflex observed in the closed-loop simulation was induced by augmentation, rather than acceleration, of the dynamic SNA response in the neural arc. However, further experimental studies are needed to verify the simulation model.

Limitations. The present study has several limitations. First, we performed the experiment in anesthetized animals. Previous studies have suggested that any anesthetic could alter the baroreflex regulation in AP (54–56). The gain of the baroreflex is reported in the conscious state to be higher (~ 2 -fold) than in the anesthetized state. A previous study (52) suggested that α -chloralose anesthesia could alter the dynamic characteristics of the baroreflex regulation around the frequency of 5 Hz. However, the anesthesia was convenient for the elimination of the central command. Furthermore, we compared the baroreflex gain between muscle stretch and nonstretch conditions both under anesthesia. Therefore, a reasonable interpretation would be that the increased baroreflex gain is attributable to muscle stretch in this experiment.

Second, stretching of skeletal muscle provides a stimulus for the activation of mechanoreceptors that is different from that which occurs during muscle contraction. During contraction, mechanoreceptors are activated by a shortening of skeletal muscle and by compression of the receptors. Thus, mechanoreceptors may be stimulated in a very different manner during stretch, which would likely affect the magnitude of the corresponding reflex response. In addition, the level of muscle stretch used in our experiment was relatively high (50). The stretch may activate different afferents than contraction (8). Furthermore, the discharge profile of mechanosensitive afferents adapt during static muscle stretch (31). Accordingly, during the muscle stretch for 6 min in the present study, the firing level from the mechanoreceptors might have been

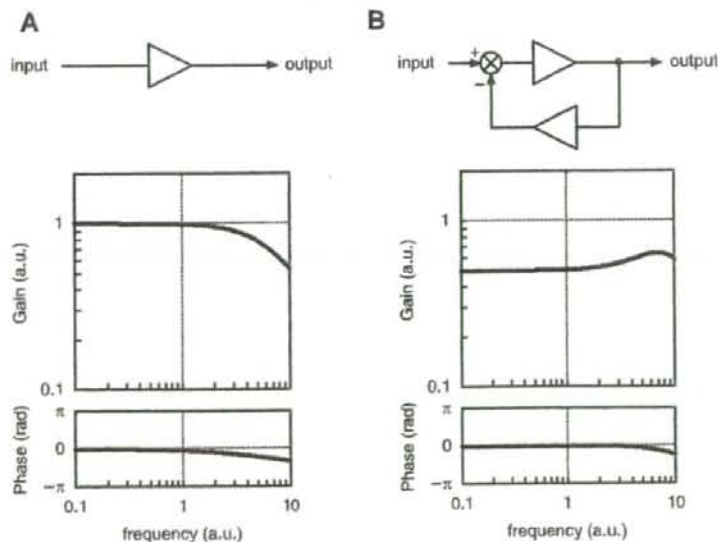


Fig. 5. An example that a circuit consisting of only low-pass elements yields high-pass characteristics as a circuit. A: block diagram of a single low-pass element (triangle) and its transfer function. Units for gain and frequency are arbitrary. B: block diagram of a circuit with a negative feedback loop with the same low-pass element (triangles). Because gain in the lower frequency range is attenuated more by the low-pass characteristics of the feedback path, the transfer function from input to output reveals high-pass characteristics.

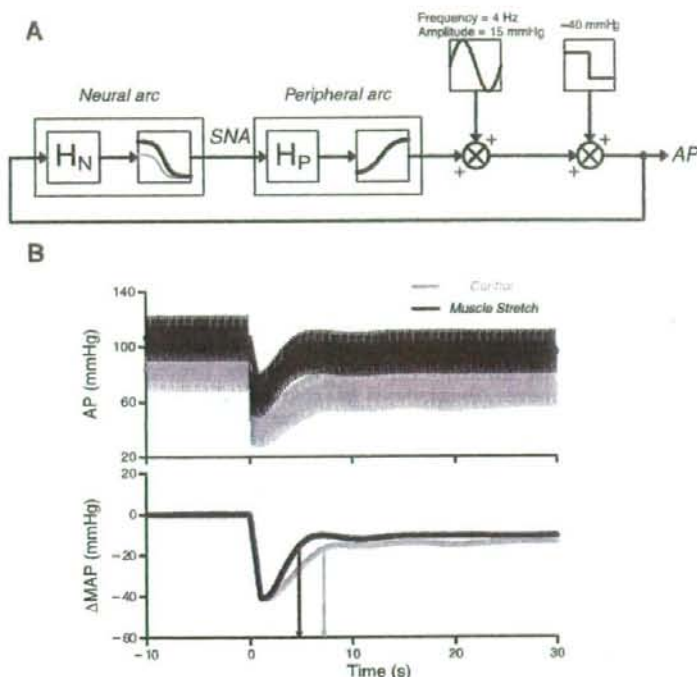


Fig. 6. *A*: simulator of the baroreflex system during activation of the muscle mechanoreflex. A stepwise perturbation with pulsatile pressure was applied to the baroreflex negative feedback system (see the APPENDIX for details). H_N , neural arc transfer function; H_P , peripheral arc transfer function. *B*: simulation results of the closed-loop AP response to the stepwise pressure perturbation (-40 mmHg). Muscle stretch shortened the time to 95% of steady state by $\sim 33\%$ (shaded and solid arrows). Shaded and solid thick lines indicate mean AP (MAP) resampled at 1 Hz. Δ MAP, change in MAP from baseline.

steadily diminishing. In fact, the increase in SNA and AP induced by muscle stretch gradually decreased from 90 s to 6 min after the initiation of the muscle stretch, which was used for data analysis (Table 1). However, SNA and AP remained significantly higher under muscle stretch conditions than control conditions over the protocol for 6 min. Thus, we believe that the mechanoreflex remained activated in this protocol. Further studies are required to elucidate the dynamic interactions between baroreflex and mechanoreflex induced by different modes of activation, such as cyclic activation of the mechanoreflex.

Third, the transfer function analysis is useful in identifying the linear input-output relationship of the baroreflex at a given operating point. However, the transfer function cannot characterize the nonlinear input-output relationship of the system. In the presence of nonlinear system behavior such as the baroreflex system, the transfer function analysis is partly compromised, indicating that the absolute output values of the nonlinear system to given input signals cannot be predicted accurately by the transfer function alone. Combining a linear transfer function with a nonlinear sigmoidal element would increase the accuracy to reproduce dynamic characteristics observed in the baroreflex neural arc (20, 22).

Finally, we measured renal SNA as a proxy of systemic sympathetic activity. SNAs to different organs may vary a lot. Although static and dynamic regulations of the baroreflex neural arc are similar among renal, cardiac, and muscle SNAs (15, 16, 18), whether this holds true during muscle stretch remains to be verified. Also, subsystems of the peripheral arc transfer function such as those relating car-

diac output and peripheral vascular resistance remain to be identified.

Conclusions. In conclusion, baroreflex open-loop transfer function analysis demonstrated that the activation of mechanosensitive afferents from skeletal muscles augmented the dynamic SNA response in the neural arc. This augmentation of the dynamic SNA response with maintained derivative characteristics of the neural arc may accelerate closed-loop AP regulation via the baroreflex.

APPENDIX

To simulate the closed-loop AP response to stepwise pressure perturbation (Fig. 6), we used the derivative-sigmoidal cascade model. The cascade model consists of a linear derivative filter followed by a nonlinear sigmoidal component (20, 22).

We modeled the sigmoidal nonlinearity in the baroreflex neural arc interacting with the muscle mechanoreflex by the following four-parameter logistic function with threshold according to a previous study (59):

$$y = \max \left\{ \frac{P_1}{1 + \exp[P_2(x - P_3)]} + P_4, \text{Th} \right\} \quad (A1)$$

where x and y are input (in mmHg) and output (in au) values. P_1 denotes the response range (in au), P_2 is the coefficient of gain, P_3 is the midpoint of the input range (in mmHg), P_4 is the minimum output value of the symmetric sigmoid curve (in au), and Th is a threshold value for the output (in au). The function $\max\{a, b\}$ gives the greater or equal value between a and b . We set $P_1 = 135$ au, $P_2 = 0.13$, $P_3 = 110$ mmHg, $P_4 = -40$ au, and $\text{Th} = 0$ au. Under muscle stretch conditions, the value of P_4 was changed to 5 au. These settings were determined based on the static interaction

between the baroreflex and muscle mechanoreflex obtained from previous studies (58, 59).

The sigmoidal nonlinearity in the peripheral arc was modelled by a four-parameter logistic function as follows:

$$z = \frac{Q_1}{1 + \exp[Q_2(y - Q_3)]} + Q_4 \quad (A2)$$

where y and z are input (in au) and output (in mmHg) values. Q_1 denotes the response range (in mmHg), Q_2 is the coefficient of gain, Q_3 is the midpoint of the input range (in au), and Q_4 is the minimum output value (in mmHg). We set $Q_1 = 120$ mmHg, $Q_2 = -0.05$, $Q_3 = 70$ au, and $Q_4 = 30$ mmHg under both conditions, according to a previous study (58).

The neural arc (H_N) and peripheral arc (H_P) linear transfer functions under control and muscle stretch conditions were obtained from Fig. 2. Because absolute values of the steady-state gains in the neural and peripheral arcs were determined by a sigmoid curve (Eqs. A1 and A2), the steady-state gains of H_N and H_P under both conditions were normalized to unity.

The input amplitude of the stepwise pressure perturbation was -40 mmHg. To mimic pulsatile pressure, we imposed a sinusoidal input on the output from the peripheral arc. The frequency and zero to peak amplitude of the sinusoidal input were 4 Hz and 15 mmHg, respectively (Fig. 6A). The closed-loop AP response was simulated up to 30 s (Fig. 6B).

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Upright Tilt Resets Dynamic Transfer Function of Baroreflex Neural Arc to Minify the Pressure Disturbance in Total Baroreflex Control

Atsunori KAMIYA¹, Toru KAWADA¹, Kenta YAMAMOTO², Masaki MIZUNO¹,
Shuji SHIMIZU¹, and Masaru SUGIMACHI¹

¹Department of Cardiovascular Dynamics, National Cardiovascular Centre Research Institute, Osaka, Japan; and ²Consolidated Research Institute for Advanced Science and Medical Care, Waseda University, Tokyo, 162-0041 Japan



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Atsunori KAMIYA¹, Toru KAWADA¹, Kenta YAMAMOTO², Masaki MIZUNO¹,
Shuji SHIMIZU¹, and Masaru SUGIMACHI¹

¹Department of Cardiovascular Dynamics, National Cardiovascular Centre Research Institute, Osaka, Japan; and ²Consolidated Research Institute for Advanced Science and Medical Care, Waseda University, Tokyo, 162-0041 Japan

Abstract: Maintenance of arterial pressure (AP) under orthostatic stress against gravitational fluid shift and pressure disturbance is of great importance. One of the mechanisms is that upright tilt resets steady-state baroreflex control to a higher sympathetic nerve activity (SNA). However, the dynamic feedback characteristics of the baroreflex system, a hallmark of fast-acting neural control, remain to be elucidated. In the present study, we tested the hypothesis that upright tilt resets the dynamic transfer function of the baroreflex neural arc to minify the pressure disturbance in total baroreflex control. Renal SNA and AP were recorded in ten anesthetized, vagotomized and aortic-denervated rabbits. Under baroreflex open-loop condition, isolated intracarotid sinus pressure (CSP) was changed according to a binary white noise sequence at operating pressure ± 20

mmHg, while the animal was placed supine and at 60° upright tilt. Regardless of the postures, the baroreflex neural (CSP to SNA) and peripheral (SNA to AP) arcs showed dynamic high-pass and low-pass characteristics, respectively. Upright tilt increased the transfer gain of the neural arc (resetting), decreased that of the peripheral arc, and consequently maintained the transfer characteristics of total baroreflex feedback system. A simulation study suggests that postural resetting of the neural arc would significantly increase the transfer gain of the total arc in upright position, and that in closed-loop baroreflex the resetting increases the stability of AP against pressure disturbance under orthostatic stress. In conclusion, upright tilt resets the dynamic transfer function of the baroreflex neural arc to minify the pressure disturbance in total baroreflex control.

Key words: baroreflex, blood pressure, sympathetic nervous system.

Since human beings are often under orthostatic stress, the maintenance of arterial pressure (AP) under orthostatic stress against gravitational fluid shift is of great importance. During standing, a gravitational fluid shift directed toward the lower part of the body would cause severe postural hypotension if not counteracted by compensatory mechanisms [1]. Arterial baroreflex has been considered to be the major compensatory mechanism [1–3], since denervation of baroreceptor afferents causes profound postural hypotension [4].

The baroreflex system consists of two subsystems: the neural arc that represents the input-output relationship between baroreceptor pressure and sympathetic nerve activity (SNA), and the peripheral arc that represents the relationship between SNA and systemic AP. Recently, we investigated the steady-state functional structure of these systems under orthostatic stress [5], and reported that upright tilt shifted the baroreflex peripheral arc to a lower AP for a given SNA. However, upright tilt reset the baroreflex neural arc to a higher steady state SNA. The resetting compensat-

ed for the blunted responsiveness of the peripheral arc and contributed to prevent postural hypotension [5].

In addition to the steady state characteristics [6, 7], the dynamic characteristics are other hallmark of the baroreflex system. It is because the system is a fast-acting neural control that quickly negative-feedback controls and stabilises AP against pressure disturbance in contrast to the slow-acting hormonal and humoral systems [8]. Earlier studies reported that the dynamic characteristics in supine position have a high-pass (fast) neural arc that may compensate for the low-pass (slow) peripheral arc to achieve rapid and stable AP regulation [8]. The importance of the dynamic characteristics in AP control increases under orthostatic stress that can cause postural hypotension. However, little is known about the dynamic characteristics of the baroreflex system in upright posture.

Because the gravitational body fluid shift decreases the effective circulatory blood volume [1, 9], we speculated that upright tilt may attenuate the dynamic transfer function from SNA to AP in the baroreflex peripheral arc.

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Correspondence should be addressed to: Atsunori Kamiya, Department of Cardiovascular Dynamics, National Cardiovascular Centre Research Institute, Osaka, 565-8565 Japan. Tel: +81-6-6833-5012, Fax: +81-6-6835-5403, E-mail: kamiya@ri.ncvc.go.jp

Moreover, if the upright tilt resets the dynamic characteristics of the neural arc in addition to resetting the steady state SNA reported previously [5], it would compensate for a blunted pressor response of the baroreflex peripheral arc and contribute to maintain the stability and quickness of the total baroreflex system. Accordingly, we hypothesized that upright tilt resets dynamic transfer function of baroreflex neural arc to minimize the pressure disturbance in total baroreflex control.

In the present study, we identified the transfer functions of two baroreflex subsystems (the neural and peripheral arcs) separately in 60° upright posture, while opening the baroreflex negative feedback loop by vascular isolation of carotid sinus regions [8]. In addition, by connecting the subsystem transfer functions in series and closing them, we investigated the dynamic transfer characteristics and the stability against pressure disturbance of total baroreflex arc system in upright posture.

MATERIAL AND METHODS

Animals were cared for in strict accordance with the Guiding Principles for the Care and Use of Animals in the Field of Physiological Science approved by the Physiological Society of Japan. Ten Japanese white rabbits weighing 2.4–3.3 kg were initially anesthetized by intravenous injection (2 ml/kg) of a mixture of urethane (250 mg/ml) and α -chloralose (40 mg/ml). Anesthesia was maintained by continuously infusing the anaesthetics at a rate of 0.33 ml/kg/h using a syringe pump (CFV-3200, Nihon Kohden, Tokyo). The rabbits were mechanically ventilated with oxygen-enriched room air. Bilateral carotid sinuses were isolated vascularly from systemic circulation by ligating the internal and external carotid arteries and other small branches originating from the carotid sinus regions. The isolated carotid sinuses were filled with warmed physiological saline pre-equilibrated with atmospheric air, through catheters inserted via the common carotid arteries. Intra-carotid sinus pressure (CSP) was controlled by a servo-controlled piston pump (model ET-126A, Labworks; Costa Mesa, CA). Bilateral vagal and aortic depressor nerves were sectioned in the middle of the neck region to eliminate reflexes from the cardiopulmonary region and the aortic arch. Systemic AP was measured using a high-fidelity pressure transducer (Millar Instruments; Houston, TX) inserted retrograde from the right common carotid artery below the isolated carotid sinus region. Body temperature was maintained at around 38°C with a heating pad.

The left renal sympathetic nerve was exposed retroperitoneally. A pair of stainless steel wire electrodes (Bioflex wire AS633, Cooner Wire) was attached to the nerve to record renal SNA. The nerve fibers peripheral to electrodes were ligated securely and crushed to eliminate afferent signals. The nerve and electrodes were covered

with a mixture of silicone gel (Silicon Low Viscosity, KWIK-SIL, World Precision Instrument, Inc., FL) to insulate and immobilize the electrodes. The preamplified SNA signal was band-pass filtered at 150–1,000 Hz. The nerve signal was full-wave rectified and low-pass filtered with a cutoff frequency of 30 Hz to quantify the nerve activity.

Protocols. Both protocols 1 and 2 were performed on each of eight animals. After the surgical preparation, the animal was maintained supine (0°) on a tilt bed. To stabilize the posture, the head was fixed full-frontal to the bed by strings, and the body and legs were rigged up in a clothes-like bag. Before performing protocols 1 and 2, we confirmed that the nerve activity measured in supine position was SNA. CSP was decreased stepwise from 100 mmHg to 40 mmHg in decrements of 20 mmHg, and then increased stepwise to 100 mmHg in increments of 20 mmHg. Each pressure step was maintained for 60 s. In all animals, a decrease in CSP increased SNA, whereas an increase in CSP decreased SNA (Fig. 1), indicating that the nerve activity recorded was SNA.

Protocol 1: The animal was placed supine. CSP was firstly matched with systemic AP to obtain the operating AP under the baroreflex closed-loop condition. After at least 5 minutes of stabilization, the SNA and AP were recorded for 10 min to obtain closed-loop baseline values. The data were stored on the hard disk of a dedicated laboratory computer system for analysis at a sampling rate of 200 Hz using a 12-bit analog-to-digital converter. The averaged AP over 10 min was defined as the operating AP in

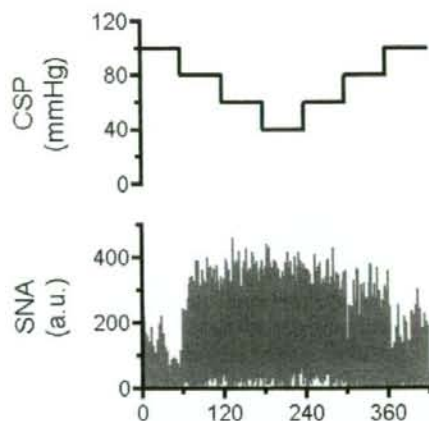


Fig. 1. Representative data of one rabbit in supine position, showing time series of carotid sinus pressure (CSP) and sympathetic nerve activity (SNA). CSP was decreased stepwise from 100 mmHg to 40 mmHg in decrements of 20 mmHg, and then increased stepwise to 100 mmHg in increments of 20 mmHg. Each pressure step was maintained for 60 s. A decrease in CSP increased SNA, whereas an increase in CSP decreased SNA, indicating that the nerve activity recorded was SNA. a.u., arbitrary unit.

supine position. Then, after at least 5 min of stabilization, CSP was randomly changed by 20 mmHg above or below the operating AP every 500 ms according to a binary white noise sequence for which the input power spectrum of CSP was reasonably flat up to 1 Hz [10]. The variables were recorded for a 10-min period and stored.

Protocol 2: CSP was firstly matched with systemic AP via a servo-controlled piston pump to obtain the actual operating pressure under baroreflex closed-loop conditions in supine and 60° upright postures. The animal was maintained supine for 10 min, and then tilted upright to 60° within 10 s by inclining the tilt bed to 60° and dropping the lower regions of the rabbit with the fulcrum set at the level of the carotid sinus. The 60° upright posture was maintained for 10 min for stabilization. Since the clothes-like bag stabilized the posture of the animal, there was no additional mechanical movement that reduced the quality of measurements. The position of the head remained almost fixed during the tilt to minimize vestibular stimulation. Thereafter, the average AP over the next 10 min was defined as the operating AP in upright tilt position. Then, after at least 5 min of stabilization, CSP was randomly changed according to a white noise sequence for 10 min as in protocol 1.

Data analysis. SNA signals were normalized by the following steps. First, the post-mortem noise level was assigned 0 arbitrary unit (a.u.). Second, SNA signals during the 10-min closed-loop baseline recording in protocol 1 (supine position) were averaged over 1 min, and assigned 100 a.u. Finally, the other SNA signals in all protocols were normalized to these values.

In both protocols 1 and 2, the transfer functions (gain and phase) and coherence function were calculated from CSP input to SNA in the baroreflex neural arc and from SNA input to AP in the baroreflex peripheral arc. The sig-

nals of CSP, SNA and AP were resampled at 10 Hz and segmented into 10 sets of 50% overlapping bins of 2¹⁰ data point each. The segment length was 102.4 s, which yielded the lowest frequency bound of 0.01 (0.0097) Hz. We subtracted a linear trend and applied a Hanning window for each segment. We then performed fast Fourier transform to obtain frequency spectra of the variables. We ensemble averaged the input power [$S_{xx}(f)$], output power [$S_{yy}(f)$], and cross power between them [$S_{yx}(f)$] over the 10 segments. Thereafter, we calculated the transfer function [$H(f)$] from input to output signals as follows,

$$H(f) = \frac{S_{yx}(f)}{S_{xx}(f)}$$

To quantify the linear dependence between input to output signals in the frequency domain, we calculated the magnitude-squared coherence function [$Coh(f)$] as follows:

$$Coh(f) = \frac{|S_{yx}(f)|^2}{S_{xx}(f)S_{yy}(f)}$$

The coherence value ranges from zero to unity. Unity coherence indicates a perfect linear dependence between input and output signals, whereas zero coherence indicates total independence of these two signals.

Statistic analysis. All data are presented as means \pm SD. Effects of upright tilt on baroreflex parameters were evaluated by repeated-measures analysis of variance. When the main effect was found to be significant, post hoc multiple comparisons were done using the Scheff's F-test to compare baroreflex controls between the supine and upright postures [11]. Differences were considered significant when $P < 0.05$.

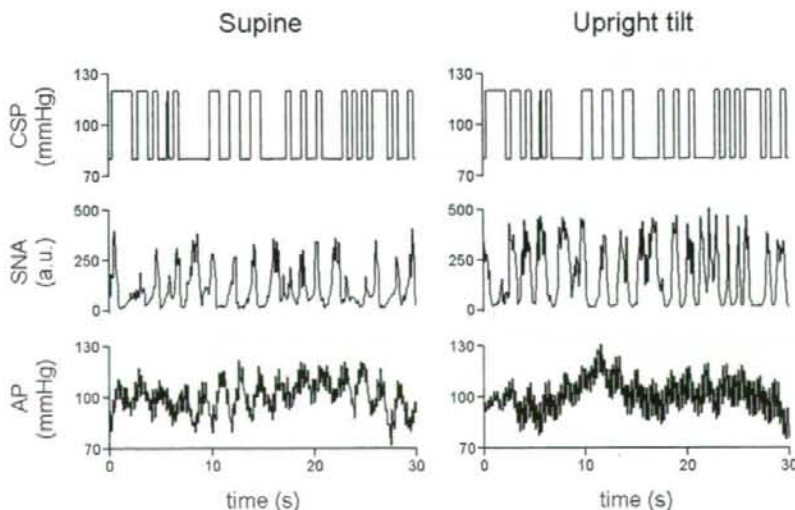


Fig. 2. Representative data of one rabbit in supine (left panels) and 60° upright tilt (right panels) positions, showing time series of carotid sinus pressure (CSP), sympathetic nerve activity (SNA) and systemic arterial pressure (AP) during CSP perturbation. CSP was changed according to a binary white noise signal with a switching interval of 500 ms. a.u., arbitrary unit.

RESULTS

Figure 2 shows the typical time series of CSP, SNA and AP derived in supine and 60° upright tilt positions in individual animal. CSP was perturbed according to a binary white noise sequence at 500-ms intervals. In both positions, SNA increased and decreased roughly in response to the decrease and increase in CSP, respectively. However, the SNA responses appeared higher in the upright tilt

than in the supine position. Data from all animals ($n = 8$) showed that the upright tilt increased the averaged SNA (175 ± 21 a.u.) during CSP perturbation compared with the supine position (96 ± 13 a.u.). Averaged AP during CSP perturbation was similar in supine (96 ± 13 mmHg) and in upright positions (103 ± 15 mmHg).

The baroreflex neural arc

Figure 3A shows the transfer function of baroreflex neural arc from CSP to SNA averaged from all animals. In both supine and upright tilt positions, the transfer gain increased as the frequency of CSP perturbation increased for the frequency range of 0.01 to 1 Hz. This shows dynamic high-pass characteristics, indicating that more rapid change of CSP results in greater response of SNA. Note that upright tilt increased the transfer gain for the whole frequency range observed (Table 1). In addition, upright tilt decreased the slope of gain increase. In both positions, the phase approached slightly above $-\pi$ radians at the lowest frequency reflecting negative feedback characters, and lagged as the frequency of CSP perturbation increased. The coherence was over 0.7 for the frequency range of 0.01 to 0.2 Hz. Upright tilt did not affect the phase or coherence. Figure 3B shows the step response of SNA corresponding to the transfer function shown in Fig. 3A. In both positions, the SNA response consisted of an initial decrease followed by partial recovery and then a steady state. Of note, upright tilt enhanced the initial decrease by 50%, and also decreased the steady-state SNA.

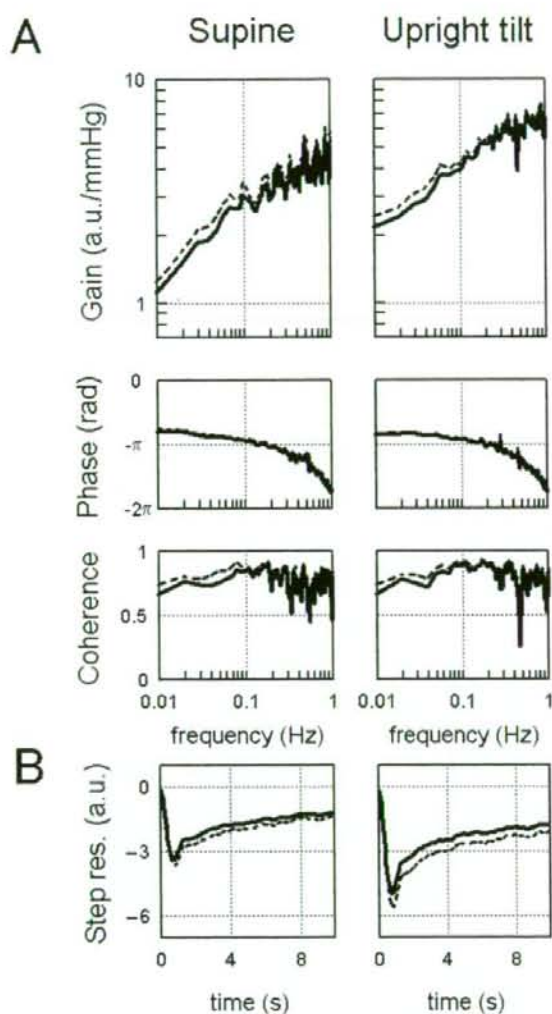


Fig. 3. A: The transfer function of the baroreflex neural arc from CSP to SNA averaged from all animals ($n = 8$) in supine (left panels) and 60° upright tilt (right panels) positions. The gain plots (top), phase plots (middle), and coherence function (bottom) are shown. Upright tilt increases the gain. **B:** Step responses (Step res.) derived from the transfer function corresponding to the transfer function shown in A. Upright tilt enhances the initial and steady-state responses. Solid line represents the mean values, and dashed line represents mean + SD in A and mean - SD in B. a.u., arbitrary unit.

Table 1. Transfer function of baroreflex neural arc (from CSP to SNA) in supine and upright tilt positions.

	Supine	Upright tilt
Gain (a.u./mmHg)		
0.01 Hz	1.11 ± 0.13	$2.14 \pm 0.41^*$
0.1 Hz	2.75 ± 0.43	$4.63 \pm 0.52^*$
0.3 Hz	3.69 ± 0.30	$5.08 \pm 0.42^*$
Phase (rad)		
0.01 Hz	-2.51 ± 0.15	-2.66 ± 0.09
0.1 Hz	-2.96 ± 0.08	-2.93 ± 0.06
0.3 Hz	-3.58 ± 0.14	-3.53 ± 0.12
Coherence		
0.01 Hz	0.67 ± 0.08	0.67 ± 0.07
0.1 Hz	0.84 ± 0.04	0.89 ± 0.02
0.3 Hz	0.77 ± 0.06	0.82 ± 0.03
Slope (dB/decade)		
0.01 Hz to 0.3 Hz	7.0 ± 0.4	$5.1 \pm 0.5^*$
Step response (a.u.)		
Initial response	-3.41 ± 0.21	$-4.99 \pm 0.62^*$
Steady-state level	-1.26 ± 0.18	$-1.80 \pm 0.32^*$

Values are mean \pm SD ($n = 10$). * $P < 0.05$; supine position vs. upright tilt.

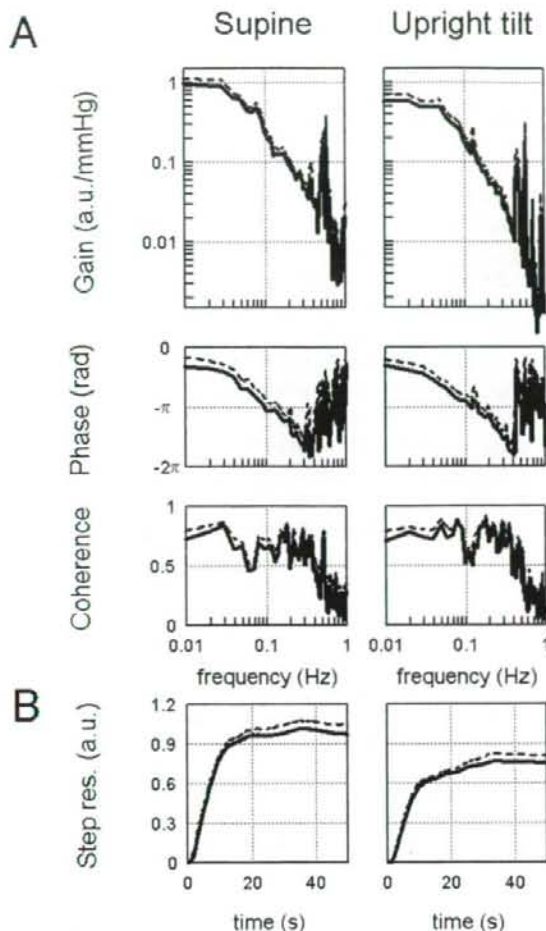


Fig. 4. A: The transfer function of the baroreflex peripheral arc from SNA to AP averaged from all animals ($n = 8$) in supine (left panels) and 60° upright tilt (right panels) positions. The gain plots (top), phase plots (middle), and coherence function (bottom) are shown. Upright tilt decreases the gain below the frequency of 0.1 Hz. **B:** Step responses (Step res.) derived from the transfer function corresponding to the transfer function shown in A. Upright tilt attenuates the response. Solid and dashed lines represent the mean and mean + SD values, respectively. a.u., arbitrary unit.

The baroreflex peripheral arc

Figure 4A shows the transfer function of the baroreflex peripheral arc from SNA to AP averaged from all animals. In both supine and upright tilt positions, the transfer gain decreased as the input frequency increased for the frequency range of 0.01 to 1 Hz, indicating low-pass characteristics. Upright tilt decreased the transfer gain between 0.01 and 0.1 Hz (Table 2). In both positions, the phase approached zero radian at the lowest frequency reflecting an increase in SNA with increased AP, and lagged as the in-

Table 2. Transfer function of baroreflex peripheral arc (from SNA to AP) in supine and upright tilt positions.

	Supine	Upright tilt
Gain (mmHg/au)		
0.01 Hz	0.97 ± 0.09	$0.63 \pm 0.06^*$
0.1 Hz	0.23 ± 0.03	$0.15 \pm 0.03^*$
0.3 Hz	0.04 ± 0.006	0.03 ± 0.003
Phase (rad)		
0.01 Hz	-0.79 ± 0.16	-0.69 ± 0.07
0.1 Hz	-2.83 ± 0.14	-2.58 ± 0.15
0.3 Hz	-4.74 ± 0.18	-4.63 ± 0.08
Coherence		
0.01 Hz	0.72 ± 0.07	0.71 ± 0.03
0.1 Hz	0.64 ± 0.08	0.62 ± 0.04
0.3 Hz	0.61 ± 0.08	0.68 ± 0.02
Step response (mmHg)		
Steady-state level	-0.97 ± 0.06	$-0.75 \pm 0.06^*$

Values are mean \pm SD ($n = 10$). * $P < 0.05$; supine position vs. upright tilt.

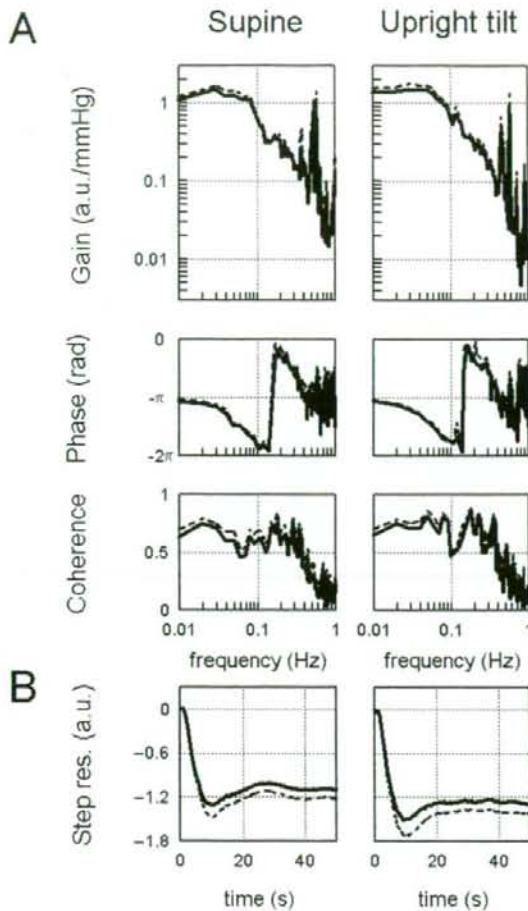
put frequency increased. The coherence was over 0.5 for the frequency range of 0.01 to 1 Hz. Upright tilt did not affect the phase or coherence. Figure 4B shows the step response of AP corresponding to the transfer function shown in Fig. 4A. In both positions, the AP response increased gradually to reach a steady state. Upright tilt decreased the steady-state AP.

The total baroreflex arc

Figure 5A shows the transfer function of the total baroreflex arc from CSP to AP averaged from all animals. In both supine and upright tilt positions, the transfer gain decreased as the input frequency increased for the frequency range from 0.01 to 1 Hz, indicating low-pass characteristics. Upright tilt did not affect the transfer gain (Table 3). In both positions, the phase approached $-\pi$ radians at the lowest frequency reflecting negative feedback attained by the total baroreflex loop, and lagged as the input frequency increased. The coherence was over 0.5 for the frequency range from 0.01 to 0.2 Hz. Upright tilt did not affect the phase or coherence. Figure 5B shows the step response of AP corresponding to the transfer function shown in Fig. 5A. In both positions, the AP response increased gradually to reach a steady state. Upright tilt did not affect the step response.

The right column of Table 3 shows a simulation of the total arc transfer function in the absence of resetting in the neural arc. The simulation was based on the neural arc transfer function in supine position and the peripheral arc transfer function in upright tilt position. Without the resetting, the upright tilt would decrease the transfer function gain and would attenuate the step response of AP at steady state, compared with the values in supine position and those in upright tilt position with resetting.

DISCUSSION



Arterial baroreflex is obviously a pivotal mechanism for maintaining AP under orthostatic stress against gravitational fluid shift and pressure disturbance [1, 2, 4], but the baroreflex function and its modulation in upright position are not fully understood. We previously reported that 60° upright tilt resets the steady-state characteristics of the baroreflex neural arc to a higher SNA [5]. However, the dynamic characteristics of the baroreflex system, which is a hallmark of fast-acting neural systems, in upright posture remain to be elucidated. Accordingly, in the present study, we identified the transfer function of the total baroreflex system and its two subsystems. The new major findings are that a 60° upright tilt increases the transfer gain of the baroreflex neural arc (CSP to SNA), decreases the transfer gain of the peripheral arc (SNA to AP), and as a result maintains the dynamic characteristics of the total baroreflex feedback system. These findings support our hypothesis that upright tilt resets dynamic transfer function of baroreflex neural arc to minimize the pressure disturbance in total baroreflex control. These results were not affected by the order of postures, since returning the ani-

Fig. 5. A: The transfer function of the total baroreflex arc from CSP to AP averaged from all animals ($n = 8$) in supine (left panels) and 60° upright tilt (right panels) positions. The gain plots (top), phase plots (middle), and coherence function (bottom) are shown. **B:** Step responses (Step res.) derived from the transfer function corresponding to the transfer function shown in A. The transfer function and step response are similar in the supine and upright tilt positions. Solid and dashed lines represent the mean and mean + SD values in A and mean - SD values in B, respectively. a.u., arbitrary unit.

Table 3. Transfer function of total baroreflex arc (from CSP to AP) in supine, upright tilt and simulated upright tilt positions.

	Supine	Upright tilt	Simulated upright tilt without resetting of the neural arc
Gain (a.u./mmHg)			
0.01 Hz	1.10 ± 0.12	1.38 ± 0.18	0.71 ± 0.18 [#]
0.1 Hz	0.63 ± 0.09	0.69 ± 0.12	0.41 ± 0.13 [#]
0.3 Hz	0.15 ± 0.03	0.15 ± 0.03	0.11 ± 0.04 [*]
Phase (rad)			
0.01 Hz	-3.33 ± 0.11	-3.29 ± 0.07	-3.21 ± 0.10
0.1 Hz	-5.76 ± 0.20	-5.55 ± 0.10	-5.51 ± 0.15
0.3 Hz	-1.98 ± 0.25	-1.87 ± 0.23	-1.91 ± 0.24
Coherence			
0.01 Hz	0.63 ± 0.06	0.65 ± 0.05	
0.1 Hz	0.60 ± 0.10	0.61 ± 0.06	
0.3 Hz	0.53 ± 0.07	0.55 ± 0.04	
Step response (mmHg)			
Steady-state level	-1.09 ± 0.11	-1.29 ± 0.12	-0.67 ± 0.11 [#]

Simulated transfer function in the absence of neural arc resetting is calculated from the neural arc transfer function in supine position and the peripheral arc transfer function in upright tilt position. Values are mean ± SD ($n = 10$). ^{*} $P < 0.05$; supine vs. simulated upright tilt, [#] $P < 0.05$; upright tilt vs. simulated upright tilt.

mal posture from 60° upright tilt to horizontal supine position restored the transfer functions to the magnitudes observed in the initial supine position (data not shown).

Little is known about the arterial baroreflex feedback system under orthostatic stress. Although earlier studies investigated the gains of baroreflex control of SNA [12–14], vascular resistance [15] and R-R interval [16], these gains are parts of the total baroreflex system, and thus are insufficient to explain the dynamics of the total arc of the baroreflex feedback system. In addition, no study has examined the phase function of baroreflex in the subsystems and the total system. Moreover, while earlier studies addressed baroreflex in relation to AP regulation under orthostatic stress, most of them evaluated the baroreflex in supine, and not orthostatic posture [14]. In the present study, we identified the transfer functions of the two baroreflex subsystems (the neural and peripheral arcs) in

upright posture independently using the baroreflex open-loop technique. Moreover, by connecting the subsystem transfer functions in series and closing them, we revealed the dynamic characteristics of the total baroreflex arc.

Our actual and simulation data indicated that resetting of the baroreflex neural arc in upright posture increases the transfer function gain of the total baroreflex arc. In our experiments, the 60° upright tilt reset and nearly doubled the transfer gain of the neural arc. Although the upright tilt decreased the transfer gain of the peripheral arc, resetting in the neural arc counteracted it and consequently preserved the dynamic transfer gain of the total baroreflex arc (1.4, Table 3). In a simulation of a situation where resetting in the neural arc is absent (Table 3), a 60° upright tilt would decrease the total arc transfer gain. These findings suggest that resetting of the neural arc (that is, baroreflex control of SNA) with dynamic characteristics plays an im-

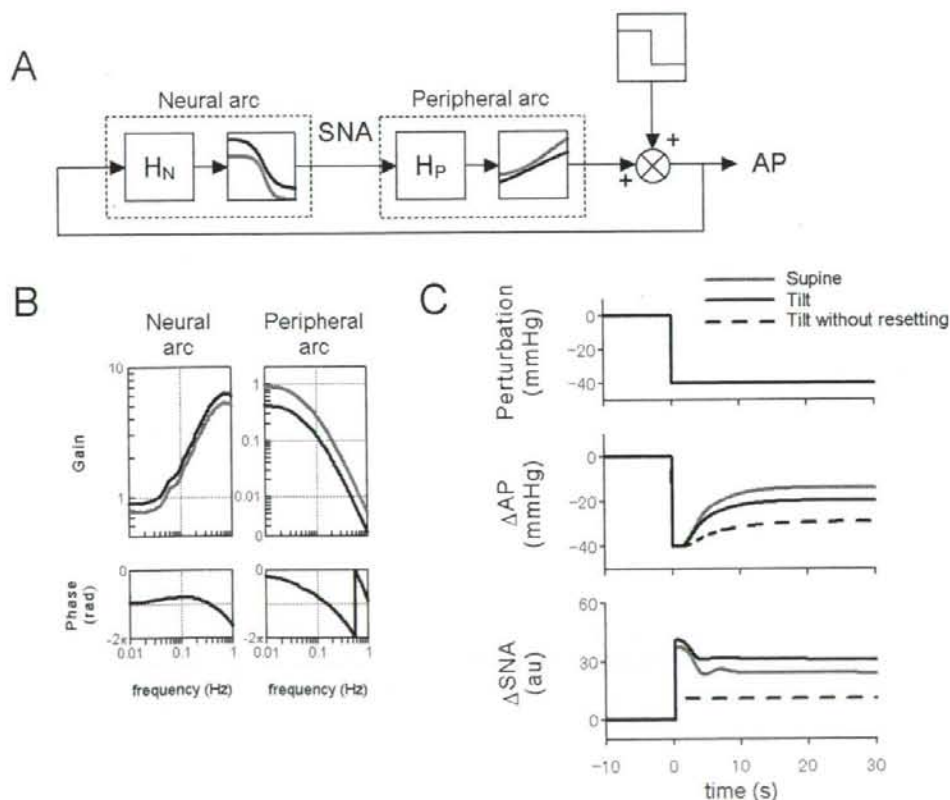


Fig. 6. A: Simulator of the baroreflex system during upright tilt. A stepwise perturbation was applied to the baroreflex negative feedback system (see APPENDIX for details). H_N , neural arc transfer function; H_P , peripheral arc transfer function. Nonlinear sigmoidal functions in the supine and upright tilt positions are shown by gray and black lines, respectively. **B:** Simulation results of integrated dynamic transfer function of linear-sigmoidal nonlinear cascade model in the neural (AP to SNA) and

peripheral (SNA to AP) arcs in the supine (gray lines) and upright tilt (black lines) positions. **C:** Simulation results of a closed-loop AP and SNA responses to the stepwise pressure perturbation (-40 mmHg). The resetting during upright tilt (black line) would enhance SNA excitation as compared with the supine position (gray line) to minimize a hypotension. Without the resetting in upright tilt, SNA responses would largely be attenuated to lead a hypotension.

portant role to maintain the dynamic transfer function of the total baroreflex system in upright posture.

A simulation of AP stability by baroreflex feedback control against pressure disturbance clearly suggests the importance of resetting of baroreflex neural arc in upright posture. Figure 6 shows the simulation of closed-loop baroreflex control of AP, when pressure disturbance was loaded to the peripheral cardiovascular compartment. According to an earlier study [17], we used the linear-sigmoidal nonlinear cascade model (Fig. 6A) to simulate the baroreflex dynamics. The result of simulation (Fig. 6B) was consistent with our *in vivo* findings that an upright tilt increased the dynamic transfer gain of the neural arc and decreased the dynamic gain of the peripheral arc. The simulation (Fig. 6C) shows that the baroreflex feedback system would minimize the pressure disturbance (40 mmHg) by 50% or more in supine (14 mmHg) and upright tilt (19 mmHg) positions. However, without the resetting of the neural arc in upright tilt, the residual pressure disturbance (29 mmHg) would persist and the velocity of pressure response would become slower (Fig. 6C). These findings suggest that dynamic resetting of the neural arc increases the stability and quickness in response of orthostatic AP against pressure disturbance in closed-loop condition of the total baroreflex arc. In addition, the simulation indicates that the resetting would enhance increases in SNA in response to pressure disturbance in upright tilt compared to supine position (Fig. 6C). Without the resetting in upright tilt, the SNA response would be greatly attenuated (Fig. 6C). This suggests that resetting of the neural arc has a critical role in activating SNA appropriately to prevent hypotension by pressure disturbance during orthostatic stress.

Some explanations for the changes in baroreflex peripheral arc in upright tilt posture may be postulated. First, since the gravitational fluid shift toward the lower part of body (i.e., abdominal vascular bed, lower limbs) during upright posture decreases the preload and effective circulatory blood volume [1, 9], it may attenuate the dynamic transfer function from SNA to AP. Our actual data revealed that upright tilt decreased the transfer gain, but not the transfer phase, of the baroreflex peripheral arc (Fig. 4A). Therefore, upright tilt would blunt the magnitude of AP response to SNA without delaying the response, as shown in the calculated step response (Fig. 4B). Next, increases in humoral factors (i.e., catecholamine, angiotensin II) during upright posture could reduce the dependency of vascular resistance on neural control. However, intravenous infusion of angiotensin II did not affect the transfer function of baroreflex peripheral arc [18]. Moreover, intravenous infusion of catecholamine had no effects on the transfer function from sympathetic stimulation to heart rate [19]. These studies are consistent with the predominance of sympathetic neural control on cardiovascular pressor function [20].

Limitations

The present study has several limitations. First, we excluded the efferent effect of vagally mediated arterial and cardiopulmonary baroreflexes that may affect baroreflex control of SNA. Second, we used an anesthetic agent that may attenuate the baroreflex peripheral arc by reducing the cardiac pumping function, and may affect the neural arc gain. Third, since we sectioned the aortic depressor nerves to open the baroreflex feedback loop, the total baroreflex gain may be lower than the physiological level. Fourth, since we measured only renal SNA, our findings have limited applicability to other SNA. Although static [10, 21] and dynamic [21] regulation of the baroreflex neural arc is similar in renal, cardiac and muscle (vasoconstrictor) SNAs in supine posture, whether this holds true during orthostatic stress remains to be verified.

Lastly, we used rabbits that are quadrupeds. Since humans spend most of their time in nearly 90° upright postures whereas rabbits do not, our findings have limited applicability to humans. However, Japanese White rabbits spend most of their time in 10–40° head-up postures, and frequently stand up to nearly 70°. Since the denervation of both carotid and aortic arterial baroreflexes is known to cause severe postural hypotension at 60° upright tilt in quadrupeds [4], this suggests that even in quadrupeds, arterial baroreflex has a very important function in the maintenance of AP under orthostatic stress. Accordingly, despite the difference in species, our findings may reflect, at least, the qualitative aspects of orthostatic baroreflex physiology in humans. Indeed, recent human studies have suggested that orthostatic stress (lower body negative pressure) enhances the SNA response to AP change [22, 23] and increases baroreflex control of SNA (assessed by the relation between spontaneous changes in diastolic AP and SNA) [12] under baroreflex closed-loop condition.

In conclusion, the transfer function identified in baroreflex open-loop condition showed that 60° upright tilt increases the transfer gain of the baroreflex neural arc, decreases the transfer gain of the peripheral arc, and as a result maintains the dynamic characteristics of the total baroreflex feedback system. Simulation study suggests that resetting of the neural arc increases the transfer gain of the total baroreflex arc and also increases the stability of orthostatic AP against pressure disturbance. These findings suggest that upright tilt resets the dynamic transfer function of the baroreflex neural arc to maintain total baroreflex stability.

APPENDIX

To simulate the closed-loop AP response to stepwise pressure perturbation (Fig. 6), we used the linear-sigmoidal nonlinear cascade model [17].

We modeled the sigmoidal nonlinearity in the baroreflex neural arc by a four-parameter logistic function with

threshold according to our previous study [5] using the following equation:

$$y = \frac{P_1}{1 + \exp[P_2(x - P_3)]} + P_4$$

where x and y are input (in mmHg) and output (in au) values. P_1 denotes the response range (in a.u.), P_2 is the coefficient of gain, P_3 is the midpoint of the input range (in mmHg), P_4 is the minimum output value of the symmetric sigmoid curve (in a.u.). We set $P_1 = 94$, $P_2 = 0.10$, $P_3 = 109$, $P_4 = 4$ in the supine position, and $P_1 = 112$, $P_2 = 0.09$, $P_3 = 109$, $P_4 = 29$ during upright tilt, according to our previous study [5].

The sigmoidal nonlinearity in the peripheral arc was modeled by a four-parameter logistic function using the following equation:

$$z = \frac{Q_1}{1 + \exp[Q_2(y - Q_3)]} + Q_4$$

where y and z are input (in a.u.) and output (in mmHg) values. Q_1 denotes the response range (in mmHg), Q_2 is the coefficient of gain, Q_3 is the midpoint of the input range (in a.u.), and Q_4 is the minimum output value (in mmHg). We set $Q_1 = 115$, $Q_2 = -0.04$, $Q_3 = 63$, $Q_4 = 50$ in the supine position, and $Q_1 = 82$, $Q_2 = -0.05$, $Q_3 = 88$, $Q_4 = 50$ during upright tilt, according to our previous study [5].

In rabbits, the transfer function of the baroreflex neural arc (baroreceptor pressure to SNA) approximates derivative characteristics in the frequency range below 0.8 Hz, and high-cut characteristics of frequencies above 0.8 Hz [17]. Therefore, according to our previous study [17], we modeled the neural arc transfer function (H_N) using the following equation:

$$H_N(f) = -K_N \frac{1 + \frac{f}{f_{c1}}}{\left(1 + \frac{f}{f_{c2}}\right)^2} \exp(-2\pi f j L)$$

where f and j represent the frequency (in Hz) and imaginary units, respectively; K_N is static gain (in a.u./mmHg); f_{c1} and f_{c2} ($f_{c1} < f_{c2}$) are corner frequencies (in Hz) for derivative and high-cut characteristics, respectively; and L is a pure delay (in s) that would represent the sum of delays in the synaptic transmission through the baroreflex central pathways and the sympathetic ganglion. The dynamic gain increases in the frequency range of f_{c1} to f_{c2} , and decreases above f_{c2} . In simulations showed in Fig. 6, we matched K_N to the actual data in the supine and upright tilt positions in this study. We also set f_{c1} , f_{c2} and L at 0.05, 0.8 and 0.2, respectively, according to the present and previous studies [17].

In addition, the transfer function of the baroreflex peripheral arc (SNA to AP) approximates a second-order low-pass filter with the dead time as follows:

$$H_P(f) = K_P \frac{1}{1 + 2\zeta \frac{f}{f_N} j - \left(\frac{f}{f_N}\right)^2} \exp(-2\pi f j L)$$

where f_N and ζ are the neutral frequency (in Hz) and damping ratio, respectively; and L is a pure delay (in s). In simulations showed in Fig. 6, we matched K_P to the actual data in the supine and upright tilt positions in this study. We also set f_N , ζ and L at 0.07, 1.4 and 1.0, respectively, according to the present and previous studies [17].

The input amplitude of the stepwise pressure perturbation was -40 mmHg (Fig. 5, A and C, top panel). The closed-loop AP (Fig. 5C, middle panel) and SNA (Fig. 5C, bottom panel) responses were simulated up to 30 s.

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Decoding rule from vasoconstrictor skin sympathetic nerve activity to nonglabrous skin blood flow in humans at normothermic rest

Atsunori Kamiya^{a,b,*}, Daisaku Michikami^{a,b}, Satoshi Iwase^{b,c}, Tadaaki Mano^{b,d}

^a Department of Cardiovascular Dynamics, National Cardiovascular Center Research Institute, Suita 565-8565, Japan

^b Research Institute of Environmental Medicine, Nagoya University, Nagoya 464-8601, Japan

^c Department of Physiology, Aichi Medical University, Aichi 480-1195, Japan

^d Gifu University of Medical Science, Gifu 501-3892, Japan