

1 The quantitative analysis of the bilayer thickness and internal density fluctuation is possible, while
 2 the maximum diameter of the vesicle (ca. 240 nm) by far exceeds the maximum resolution of our SAXS
 3 experiments. The theoretical descriptions of scattering functions from a multilamellar stack of bilayers
 4 can be summarized as follows.⁵⁷ Assuming monodisperse bilayers, the scattered intensity $I(q)$ can be
 5 written as the product of the form factor $P(q)$ of the bilayers and the structure factor $S(q)$ describing their
 6 spatial arrangement similarly to that of globular particle systems:
 7

$$8 \quad I(q) \propto P(q)S(q) \quad (6)$$

9 When the thickness of the bilayer is sufficiently smaller than the lengthscale of the remaining two other
 10 dimensions as well as the radius of curvature of the bilayer, the form factor $P(q)$ in eq.(6) can be
 11 replaced with the product of the so-called Lorentz factor, $1/q^2$, and the thickness form factor or
 12 scattering function, $P_1(q)$.⁵⁸
 13

$$14 \quad P(q) = (2\pi A/q^2)P_1(q) \quad (7)$$

15 where A is the area of the basal plane. Substituting $P_1(q)$ for $P(q)$, $I(q)$ in eq.(6) can be rewritten in the
 16 convenient form
 17

$$18 \quad I(q) \propto P_1(q)S(q)/q^2 \quad (8)$$

19 The thickness scattering function $P_1(q)$ is the cosine transformation of the thickness distance distribution
 20 function (thickness PDF) $p_1(r)$. Therefore, we can use a similar method to a conventional indirect
 21 Fourier transformation (IFT)⁵⁹ to deduce $p_1(r)$ from the experimental $P_1(q)$.⁵⁸
 22

$$23 \quad P_1(q) = 2 \int_0^\infty p_1(r) \cos(qr) dr \quad (9)$$

24 In the case of the absence of interlamellar interference peaks and their signature (no bilayer stacking
 25 as to be expected for vesicle solutions), we can assume $S(q) = 1$ in eq.(6). Figure 2c presents the
 26 calculated thickness PDF, $p_1(r)$, of the vesicle designed for Hb encapsulation. We have already shown
 27 that the surface modification with low density poly(ethylene glycol) (PEG) is quite efficient for long-
 28 term storage of the HBV, inhibiting intervesicular aggregation and fusion.^{48,49} However, due to the low
 29 density of PEG for surface modification of the vesicle as the concentration of DSPE-PEG is as low as
 30 0.3 mol% of the total lipids, we were not able to observe the effect of PEG in $p_1(r)$. Nevertheless, $p_1(r)$
 31 deduced without using any geometrical model reveals the thickness of the bilayer excluding PEG layer
 32 to be ca. 5.8 nm, which is read out from the distance (r -value) where $p_1(r)$ goes to zero, as highlighted in
 33 Figure 2c. Recently, we studied static structure and molecular dynamics in PEG-lipid (DSPE-PEG)
 34 micellar solutions with different molecular weights of PEG, M_{PEG} , by means of SAXS and dielectric
 35 relaxation spectroscopy (DRS).⁶¹ The thickness of the hydrated PEG layer was estimated to be 8-9 nm
 36 for $M_{PEG} = 5000$. Due to different density and configuration, the extension of polymer chains in solution
 37

38 may not necessarily be identical in the micelle and on the surface of the vesicle, it is apparent that the
 39 actual total bilayer thickness involving the surface PEG layer must be far greater than 5.8 nm.

40 The positive-negative-positive behavior of $p_1(r)$ when going from $r = 0$ to higher- r values is directly
 41 connected to the occurrence of positive and negative internal electron density layers within the bilayer.
 42 Theoretically, $p_1(r)$ is given by the convolution square of the electron density profile $\Delta\rho(r)$ perpendicular
 43 to the midplane of the bilayers, which is given by

$$44 \quad p_1(r) = 2 \int_0^\infty \Delta\rho(r') \Delta\rho(r' + r) dr' \quad (10)$$

45 To extract more intuitive pictures of the internal structure of the bilayer, we performed a deconvolution
 46 procedure of the experimental $p_1(r)$ into $\Delta\rho(r)$ using a convolution square-root technique.⁶²⁻⁶⁴ In Figure
 47 2d, we display $\Delta\rho(r)$ obtained from the deconvolution analysis of $p_1(r)$ shown in Figure 2c. The
 48 innermost (the smallest- r) negative density part apparently reflects hydrophobic chain of the lipids. The
 49 following plateau having nearly zero electron density fluctuation, or an identical electron density with
 50 that of solvent water including salt, indicates localized cholesterol molecules on the
 51 hydrophobic/hydrophilic interface.⁶⁵ A terminal hydroxyl group is expected to be oriented toward the
 52 hydrophilic site of the lipids. The positive density layer seen in $1.7 \leq r/\text{nm} \leq 2.9$ is attributed to the
 53 hydrophilic headgroup of the lipids. We note that the forward SAXS intensity is generally very sensitive
 54 not only to the structure but also to the temperature dependent contrast. The pronounced difference in the
 55 forward intensities and different positions of the minimum in $I(q)$ at different temperatures as shown in
 56 Figure 2b sensitively reflect different contrast as a function of temperature. IFT and deconvolution
 57 analyses⁶²⁻⁶⁴ given in Figures 2c and 2d demonstrate the stable bilayer structure, which does not
 58 significantly depend on temperature.

59 **Hierarchical structures of HBV and the solution state of the encapsulated Hbs.** The HBVs are
 60 hierarchically organized cellular-type artificial oxygen carrier that encapsulates a concentrated Hb
 61 solution in phospholipid vesicles, one HBV particle containing ca. 30000 Hb molecules. Figure 3 shows
 62 the SAXS intensity, $I(q)$, of the concentrated HBV dispersion ([Hb] = 10 g dL⁻¹, [solutes] = 16 g dL⁻¹;
 63 volume fraction, ca. 40%). We successfully measured more than five orders of magnitude of the
 64 scattered intensity (ca. 3.5×10^5 time-different maximum and minimum intensities), covering the
 65 extremely wide q -range of 0.06 - 10 nm⁻¹, which was quite essential for exploring the internal structures
 66 of HBV at the molecular level.

67 First, we checked the concentration dependence of $I(q)$ by diluting the dense stock HBV dispersion
 68 ([Hb] = 10 g dL⁻¹) with saline solution, and found virtually no concentration dependence of the shape of
 69 the scattering functions. This means that the effect of inter-particle interactions or static structure factor

$\Delta I(q)$, which is given as the Fourier transform of the total correlation function $[g(r)-1]$ and whose first peak position is expected to be $q = \sim 0.025 \text{ nm}^{-1}$ when hard-sphere (HS) interaction and a 120 nm radius are assumed, is not visible on the experimental $I(q)$ in our accessible q -range ($q > 0.06 \text{ nm}^{-1}$). Consistent with the DLS results, the forward intensity of the HbV dispersion exhibits a typical feature of to some extent polydisperse spherical particles. In Figure 3, we added a theoretical scattering function calculated by accounting for the **averaged diameter** ($\sim 240 \text{ nm}$) and the size distribution obtained by DLS (green line, adjusted in height to the experimental intensity). Suppose that the HbV has homogeneous electron density distribution inside the particle, $I(q)$ should rapidly decrease proportional to q^{-4} , according to the theoretical curve for homogeneous spheres. However, we clearly observed an enhanced excess scattering in the regime $2 \leq q/\text{nm}^{-1} \leq 10$. This high- q excess component manifests the internal electron density fluctuations of the HbV particle, reflecting the shorter lengthscale structures of the encapsulated Hb solution and the lipid bilayer.

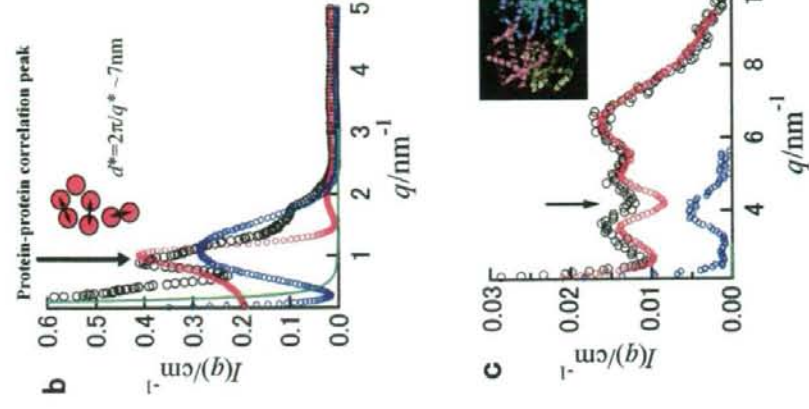
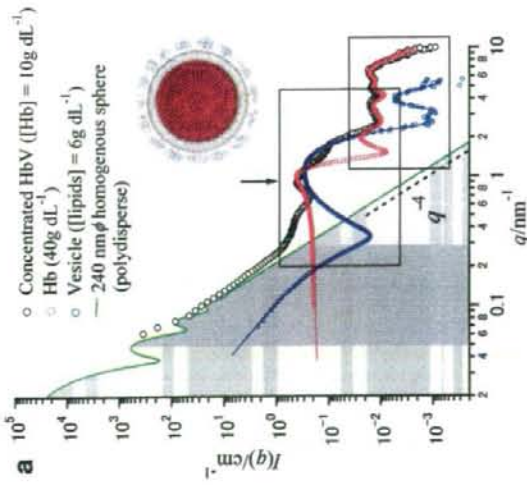


Figure 3: X-ray scattered intensity $I(q)$ in a log-log plot of the concentrated HbV dispersion ($[Hb] = 10 \text{ g} \cdot \text{dL}^{-1}$) at 25°C in $0.06 \leq q/\text{nm}^{-1} \leq 10$ is shown on absolute scale. A schematic picture of HbV is inserted into the panel a as well as a simulated scattering curve of a solution of polydisperse spheres according to the DLS results. In the panels b and c, the enlarged views of the high- q range are displayed in a lin-lin plot to visualize the significant features of $I(q)$ for protein-protein correlation peak and microscopic intramolecular structure of the encapsulated Hbs. Molecular structure of Hb is inserted in the panel c.

1 If we carefully look at the two specific q -ranges indicated by the two squares placed in Figure 3a,
 2 which are magnified in Figures 3b and 3c, we are able to access the information on the protein-protein
 3 interactions in the confined space of the aqueous phase of the HBV and the microscopic intra molecular
 4 structure of Hb molecules. As shown in Figures 3b and 3c, we found that the high- q part of $I(q)$ of the
 5 HBV is well approximated by a superposition of the experimental scattering functions of the
 6 concentrated Hb solution with identical concentration (ca. 38 g dL^{-1}) and of the phospholipids bilayer
 7 membrane (empty vesicles). Generally, for globular colloidal particle systems like proteins and micelles,
 8 the position, height, and width of the first peak of $S(q)$ strongly depends on the interparticle interactions
 9 determined e.g. by the radius, volume fraction, effective charge of the particles, a screening effect of
 10 electrostatic interaction, surface adhesion, and so forth.^{53, 61, 66-69} The details of the concentration and
 11 temperature dependence of the Hb-Hb interactions in relation to the biological functions of Hb will be
 12 reported elsewhere. Importantly, we found that the protein-protein correlation peak position as well as
 13 the height and width of the first peak of $S(q)$ of the encapsulated Hbs are almost identical to those of a
 14 normal Hb solution with identical concentration (Figures 3b). The finding suggests that confinement
 15 effects into a ca. 240 nm space are not very significant for the interparticle interaction potential of Hbs.

16 Figure 3c confirms the perfect coincidence of the scattering functions for the encapsulated and
 17 normal (bulk) Hbs in $2 \leq q/\text{nm}^{-1} \leq 10$, where the small difference between those of HbV and the normal
 18 Hb solution seen in $3 \leq q/\text{nm}^{-1} \leq 5$, highlighted by an arrow, can clearly be explained in terms of the
 19 contribution from the encapsulating vesicle. This implies unbiased intramolecular structures of Hb
 20 before and after its encapsulation into HbV.

21 **Particle diffusion dynamics in a concentrated HbV dispersion.** One of the most significant
 22 specificities of the HbV compared to any other conventional liposomal products is its extremely high
 23 concentration ($[\text{Hb}] = 10 \text{ g dL}^{-1}$; $[\text{lipids}] = 6 \text{ g dL}^{-1}$). This comes essentially from the requirement for
 24 achieving an oxygen-carrying capacity comparable to that of blood. As a result, the HbV dispersion for
 25 practical medical use is totally turbid due to high concentration and the sub-micron particle size. DLS
 26 techniques are often successfully used for observing diffusion processes of particle systems, thus being
 27 efficient for characterizing microscopic motion and inter particle interactions of particles. However,
 28 conventional DLS techniques that rely on a singly scattered signal are no longer applicable for such a
 29 dense, turbid system because of multiple scattering. In addition, absorption of hemoprotein is also a
 30 specific problem for the HBV system to be overcome. Therefore, we needed to efficiently reduce the
 31 scattering volume of the sample to observe microscopic motions of the HBV particles without diluting
 32 the samples. To overcome the obstacles, we used a modified thin-layer cell dynamic light-scattering
 33 (TC-DLS) technique.^{62, 63} As shown in the inset of Figure 4a, the concentrated HBV ($[\text{Hb}] = 10 \text{ g dL}^{-1}$)

dispersion inserted into the thin-layer cell ($50 \mu\text{m}$ thick) is transparent as if it were a red cellophane
 sheet. The opposite side can be seen through the TC window, as 'HBV' written on a paper is readable
 without mistiness, and thus DLS experiments in the forward direction become possible. The total
 scattering angle in air was fixed to 41° for all the experiments, which corresponds to 29° in water,
 resulting in $q = 6.75 \times 10^3 \text{ nm}^{-1}$.

Figure 4 shows the concentration dependence of the normalized intensity correlation functions,
 $[g_2(\tau) - 1]/\beta$, of the concentrated HBV dispersions measured at $q = 6.75 \times 10^3 \text{ nm}^{-1}$, which is lower than
 an anticipated HBV-HbV correlation peak position $q \sim 0.025 \text{ nm}^{-1}$ (See Figure 5). With increasing
 concentration from $[\text{Hb}] = 1.0 \text{ g dL}^{-1}$ to 5.0 g dL^{-1} , the short-time decay of $g_2(\tau)$ becomes faster, as
 expected for the corrective diffusion of all repulsively interacting hard-sphere (HS) systems. At the same
 time, we found the stretch of the long-time tail of $g_2(\tau)$, which may seem to indicate the occurrence of
 the gradual slowdown of the long-time behavior. With the further increase to $[\text{Hb}] = 10 \text{ g dL}^{-1}$, in turn,
 the short-time decay of $g_2(\tau)$ becomes slower, which is opposite to what is anticipated for a repulsively
 interacting system, and may imply the existence of weak attractive (adhesive) potential between HBVs.
 We may need to perform further experimental works to seek to understand the details of the interaction
 potentials between the HBVs. Nevertheless, the present TC-DLS data confirm that $g_2(\tau)$ for the most
 concentrated stock HBV dispersion ($[\text{Hb}] = 10 \text{ g dL}^{-1}$) rapidly converges to the baseline (unity) showing
 single-step behavior, which demonstrates, from a viewpoint of the particle diffusion dynamics, that the
 HBV dispersion is an ergodic system. The finding well supports good dispersion stability and proved
 long term preservation, owing to the applied surface modification of the HBV with poly(ethylene glycol)
 (PEG).

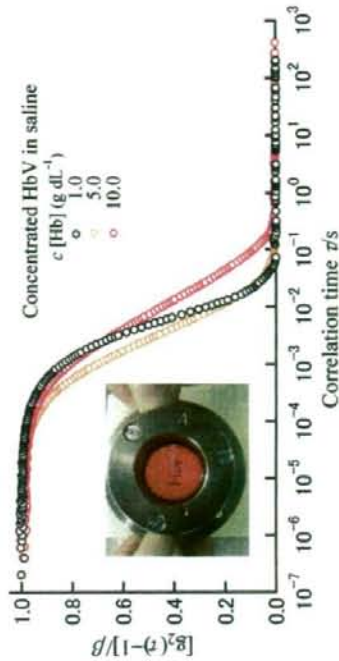


Figure 4: The normalized intensity correlation functions, $[g^{(2)}(r)-1]/\beta$, of the concentrated dispersions of the HbV at $[Hb] = 1.0, 5.0,$ and 10 g dL^{-1} , as obtained by a modified thin-layer cell dynamic light scattering (TC-DLS) technique at $q = 6.75 \times 10^3 \text{ nm}^{-1}$.

The effect of plasma substitutes on HbV-HbV interactions. Since the HbV does not contribute to colloid osmotic pressure (COP) of blood, upon anticipated actual medical treatments, the HbV must be dispersed with aqueous solution of plasma substitutes (water-soluble polymers) to adjust COP. Recently, Sakai et al. reported the rheological properties of the HbV dispersion ($[Hb] = 10 \text{ g dL}^{-1}$) in a series of plasma substitute solutions with various molecular structures and molecular weights, such as recombinant human serum albumin (rHSA), dextran (DEX), modified fluid gelatin (MFG), and hydroxyethyl starch (HES).³² They found that among these polymers, only HbV suspended in rHSA behaves nearly as a Newtonian fluid. In contrast, other plasma substitutes, such as HES, DEX, and MFG, induce non-Newtonian behavior of the HbV dispersions in particular for high molecular weight HES. Microchannel flow experiments indicated that this is due to flocculation of HbV in the presence of plasma substitutes. To gain insights into the mechanism of the HbV flocculation, we use different DLS techniques in this section.

Simulated static structure factors with depletion interaction potential. The depletion interaction potential²⁶⁻²⁸ for hard-sphere having the diameter σ in the presence of nonadsorbing polymers with the radius of gyration R_g can be approximated in the range of $\sigma < r < \sigma + 2R_g$ as²⁹

$$u(r) = -\eta_p^{(R)} k_B T \frac{3}{2} \frac{1 + \xi}{\xi^3} \left(\frac{r}{\xi} - 1 - \xi \right)^3 \quad (11)$$

where r is the center-of-mass to center-of-mass distance between the two particles, $\xi = 2R_g/\sigma$ is the size ratio, corresponding to the dimensionless range of the depletion attraction, $\eta_p^{(R)}$ is the reservoir polymer volume fraction, which can be calculated from the volume fraction of the large particles ϕ and the size ratio ξ .³⁰ From Eq. 11, one can easily find that the depth of the depletion potential is closely related to the two key parameters of ξ and $\eta_p^{(R)}$; the stronger attractive force between the two large particles is induced for smaller ξ and larger $\eta_p^{(R)}$. We simulated the pair correlation functions, $g(r)$, and the resulting static structure factors, $S(q)$, for the model systems of a very diluted HbV suspended in a series of plasma substitutes using depletion interaction potential model and SMSA closure relation.^{70,71}

In Figure 5, we present $g(r)$ and $S(q)$ calculated assuming the diameter $\sigma = 240 \text{ nm}$ and volume fraction $\phi = 0.001$ to account for the HbV. To give ξ , the radius of gyration of the plasma substitutes determined by SAXS at $c = 1.0 \text{ g dL}^{-1}$ (unpublished results) was used, 5.5 nm (MFG), 4.96 nm (DEX), 2.84 nm (rHSA), 5.96 nm (HES₇₀), 6.83 nm (HES₁₃₀), and 12.95 nm (HES₂₇₀). The solution structures

of these plasma substitutes, involving the Flory radius, screening length, persistence length, and length-scale-dependent fractal dimensions of the polymer chains will be reported elsewhere. We selected half of the original concentration of commercially available plasma substitutes; 2.5 g dL^{-1} (rHSA), 5.0 g dL^{-1} (DEX), 2.0 g dL^{-1} (MFG), 3.0 g dL^{-1} (HES₇₀), 3.0 g dL^{-1} (HES₁₃₀), and 3.0 g dL^{-1} (HES₂₇₀). In these calculations, we rely on the depletion potential and SMSA closure. According to the theoretical phase diagram,^{30,31} the selected concentration of DEX seems to be already in the two phase region even at such a dilute concentration of the vesicle (to distinguish, $S(q)$ and $g(r)$ for DEX are given using a dashed line), while theoretically, others are still in one-phase fluid region.

In terms of the increased osmotic compressibility and coordination number manifested respectively in the $S(q)$ and $g(r)$, we estimated the effect of the different plasma substitutes on the flocculation tendency of the HbV particles to be in the order of $DEX > MFG > HES_{670} > HES_{70} > HES_{130} > rHSA$. We will examine the validity of this prediction using DLS technique in the next sub-section.

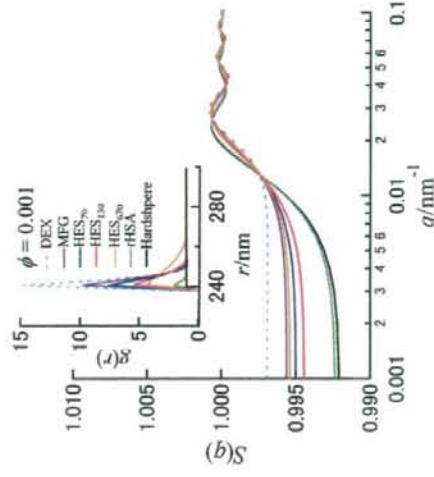
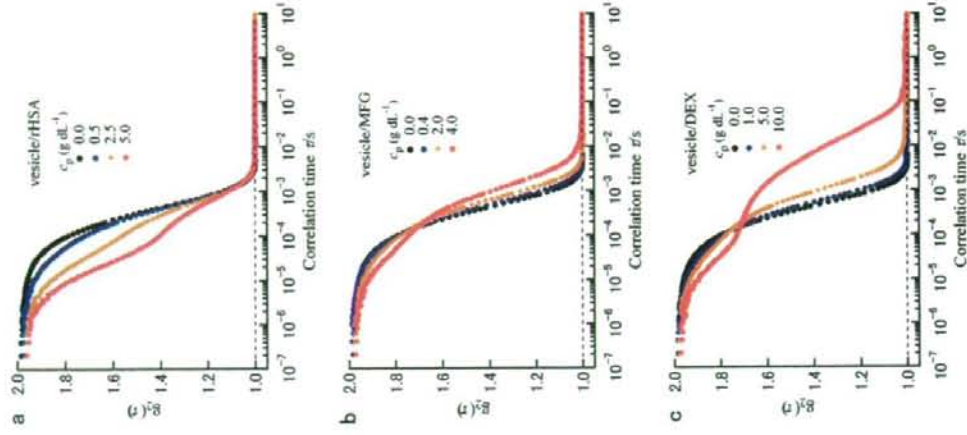


Figure 5: Simulated static structure factors $S(q)$ and pair correlation functions $g(r)$ of the model systems accounting for a dilute HbV suspended in the series of plasma substitutes. $g(r)$ and $S(q)$ were calculated using a depletion interaction potential for monodisperse hard-spheres in the presence of nonadsorbing polymers and SMSA closure relation. We chose the diameter $\sigma = 240 \text{ nm}$ and volume fraction $\phi = 0.001$ to account for HbV in a dilute suspension, and the diameter $2R_g$ for small particles (polymers), where radius of gyration of the plasma substitutes determined by SAXS at $c = 1.0 \text{ g dL}^{-1}$ (unpublished results) was used for R_g . We assumed the situation that HbVs are suspended in plasma substitutes having half of the

original concentration, i.e. 2.5 g dL⁻¹ (rHSA), 5.0 g dL⁻¹ (DEX), 2.0 g dL⁻¹ (MFG), 3.0 g dL⁻¹ (HES₇₀), 3.0 g dL⁻¹ (HES₁₃₀), and 3.0 g dL⁻¹ (HES₆₇₀).

Effects of plasma substitutes on the HBV diffusion dynamics in a diluted dispersion. To start with the simplest case, we first tried to investigate variation of the particle diffusion dynamics of the vesicle (without Hb encapsulation) in a diluted suspension in the presence of plasma substitutes and its polymer-concentration dependence. Figure 6 shows the intensity correlation functions, $g_2(t)$, of a diluted vesicle ([lipids] = 0.012 g dL⁻¹) dispersed in various plasma substitute solutions of rHSA, DEX, MFG, HES₇₀, HES₁₃₀, and HES₆₇₀, measured at $q = 0.223 \text{ nm}^{-1}$. The concentrations of these plasma substitutes are varied from 10% to 100% of their original concentrations, while fixing vesicle concentration. Figure 7a shows $g_2(t)$ of the vesicle/rHSA systems. With increasing rHSA concentration, two distinct decays become clearly visible. The faster decay is apparently attributed to the diffusion of rHSAs, and the slower one is that of the vesicles. Importantly, we observed that $g_2(t)$ at different rHSA concentrations almost simultaneously converges to the baseline at about the same point, demonstrating that a significant slowdown of the vesicle diffusion is not induced by rHSA.

The appearance of two distinct decays is common for all polymers. However, in contrast to the case of rHSA, we found that, as can be seen from Figures 6b-e, all other plasma substitutes cause a pronounced slowdown of the vesicle translational diffusion, and the effect becomes significantly stronger with increasing polymer concentration. In Figure 7, we display the intensity distribution functions $D_i(R_{ij})$ of vesicle dispersions calculated using ORT⁵⁶ from the same DLS data shown in Figure 6a-f, where the data at highest plasma substitute concentrations were chosen. Figure 7 underlines that rHSA does not cause the vesicle flocculation, but other polymers do. In terms of $D_i(R_{ij})$, the efficacy of the plasma substitutes on the vesicle flocculation are estimated to be in the order of DEX > HES₆₇₀ > MFG > HES₁₃₀ > HES₇₀ > rHSA, instead of DEX > MFG > HES₆₇₀ > HES₁₃₀ > HES₇₀ > rHSA that is predicted from the depletion potential model. In light of the results, the tendency can broadly be explained with depletion interaction, but there exist some singularity of HES series; (i) the influence of HES₆₇₀ is considerably stronger than expected and (ii) the effect of HES₁₃₀ is stronger than that for HES₇₀, which is opposite to the theoretical prediction. These are likely to be due to unideality of the systems involving the distribution of molecular weights and the complexity of the polymer chain structure.



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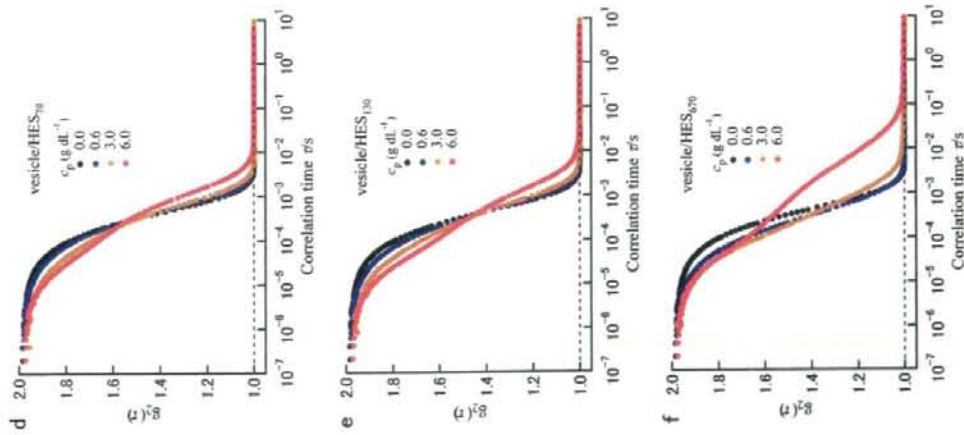


Figure 6: The intensity correlation functions, $g_2(t)$, of a 0.012 g dL^{-1} vesicle suspended in various plasma substitute solutions of rHSA, DEX, MFG, HES₇₀, HES₁₃₀, and HES₆₇₀, measured at $q = 0.223 \text{ nm}^{-1}$. The concentrations of the plasma substitutes are adjusted to 10%, 50%, and 100% of their original concentration by dilution with saline solution.

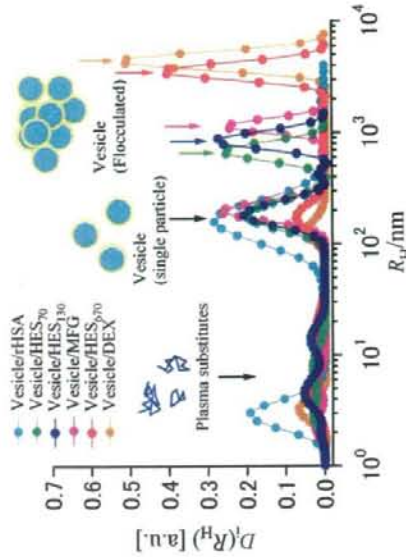


Figure 7: Effect of plasma substitutes on vesicle flocculation in a dilute suspension. The intensity distribution functions, $D_i(R_{H_i})$, of a 0.012 g dL^{-1} vesicle suspended in various plasma substitute solutions of rHSA, DEX, MFG, HES₇₀, HES₁₃₀, and HES₆₇₀ as a function of hydrodynamic radius, R_{H_i} . $D_i(R_{H_i})$ was calculated from the intensity correlation functions, $g_2(t)$, presented in Figure 6 using ORT procedure.⁵⁶

In Figure 8, a partial phase diagram of HBV/polymer mixtures as a function of ϕ_{HBV} , $\eta_p^{(R)}$, and ξ is presented together with a typical theoretical phase diagram of a colloid-polymer mixture ($\xi = 0.08$),²⁷ where the reservoir polymer volume fractions $\eta_p^{(R)}$ is estimated for different plasma substitutes at half of their original concentrations; 2.5 g dL^{-1} (HSA), 5.0 g dL^{-1} (DEX), 2.0 g dL^{-1} (MFG), 3.0 g dL^{-1} (HES₇₀), 3.0 g dL^{-1} (HES₁₃₀), and 3.0 g dL^{-1} (HES₆₇₀), using the radius of gyration R_g determined with SAXS data at 1.0 g dL^{-1} (unpublished data). The filled and empty circles represent single liquid and two phases, respectively. In practice, other conditions, e.g., polydispersity and surface charge of the large particle as well as non-ideality of polymer structure especially at high $\eta_p^{(R)}$ may affect the phase behavior. Nevertheless, the figure imply that the phase separation of the HBV suspension in the presence of the

plasma substitutes can be understood based on the depletion interaction potential model for hard-sphere mixed with non-adsorbing polymers.

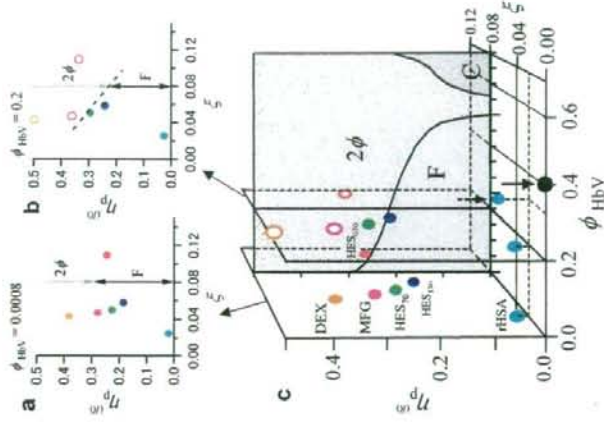
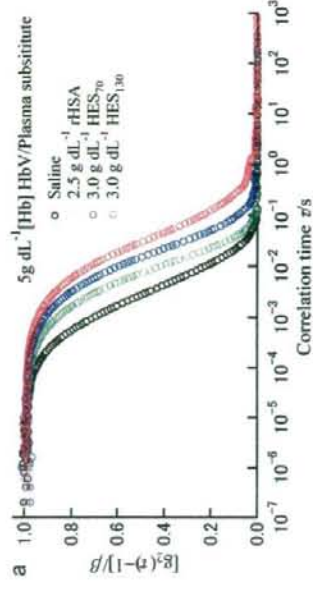


Figure 8: Partial phase diagram of HBV in the presence of plasma substitutes as a function of HBV volume fraction ϕ_{HBV} , reservoir polymer volume fractions $\eta_p^{(R)}$, and the polymer to HBV size ratio ξ , shown together with a theoretical phase diagram of a colloid-polymer mixture ($\xi = 0.08$).²⁷ Panels a and b represent 2-D phase diagrams at $\phi_{\text{HBV}} = 0.0008$ and $\phi_{\text{HBV}} = 0.2$, respectively, and c presents a 3-D diagram as a function of ϕ_{HBV} , $\eta_p^{(R)}$, and ξ . F, C, and 2ϕ on the theoretical phase diagram indicate single phase fluid, crystal, and two phase regions, respectively. For $\phi_{\text{HBV}} = 0.0008$ and 0.2, the concentration of plasma substitutes was fixed to half of their original ones, i.e., 2.5 g dL⁻¹ (rHSA; light blue), 5.0 g dL⁻¹ (DEX; orange), 2.0 g dL⁻¹ (MFG; pink), 3.0 g dL⁻¹ (HES₇₀; right green), 3.0 g dL⁻¹ (HES₁₀₀; blue), and 3.0 g dL⁻¹ (HES₇₀; red). Filled and empty circles represent single phase fluid and two-phase, respectively. Arrows highlight the stock HBV dispersion ($\phi_{\text{HBV}} = 0.4$) and a currently major applicatory HBV/rHSA mixture ($\phi_{\text{HBV}} = 0.32$) used for preclinical tests.

The effects of plasma substitutes on the HBV diffusion dynamics in a dense dispersion. We used TC-DLS to study concentrated HBV/plasma substitute mixtures. In Figures 9a and 9b, we present $g_2(t)$ of the concentrated HBV ($[\text{Hb}] = 5 \text{ g dL}^{-1}$, [lipids] = 3 g dL⁻¹) and the vesicle without Hb encapsulation ([lipids] = 3 g dL⁻¹) suspended in saline solution and in a series of different plasma substitute solutions of rHSA, HES₇₀, and HES₁₀₀. In the presence of these plasma substitutes, the decay of $g_2(t)$ becomes apparently slower, indicating the induced depletion forces by plasma substitute (Figures 9a). As shown in Figure 9b, we confirmed that with no reference to Hb-encapsulation, $g_2(t)$ exhibits a similar trend.

In Figure 9c, we compare the TC-DLS result on the concentrated HBV ($[\text{Hb}] = 10 \text{ g dL}^{-1}$) suspended in 5 g dL⁻¹ rHSA solution to that of the dense stock HBV dispersion ($[\text{Hb}] = 10 \text{ g dL}^{-1}$) in saline solution. We note that rHSA is currently a leading candidate for the co-injected plasma substitute, and most pre-clinical tests of the HBV were carried out using rHSA.⁸⁹ These two samples exhibit similar single-step behavior of $g_2(t)$ and a rapid decrease to the baseline. This observation provides evidence for ergodicity of these two samples as a nature of single fluid phase, which supports the results obtained in the previous rheological study.



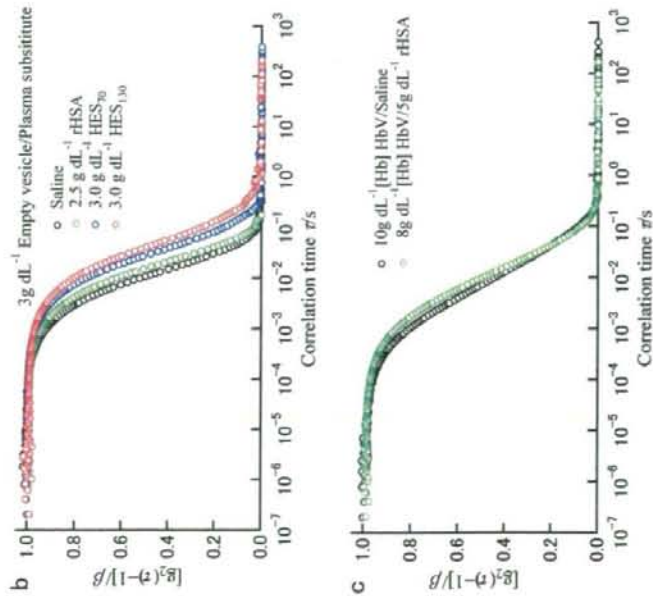


Figure 9: The intensity correlation functions, $g_2(t)$, of the concentrated dispersions of HbV and empty uni-lamellar vesicles in saline and plasma substitute solutions; $g_2(t)$ of the concentrated HbV dispersions at $[Hb] = 1.0, 5.0, \text{ and } 10 \text{ g dL}^{-1}$ (a), the HbV ($[Hb] = 5.0 \text{ g dL}^{-1}$) suspended in 2.5 g dL^{-1} rHSA, 3.0 g dL^{-1} HES₇₀, and 3.0 g dL^{-1} HES_{130} solutions (b), the 3.0 g dL^{-1} vesicle in the same series of plasma substitutes (c), and the stock HbV dispersion ($[Hb] = 10 \text{ g dL}^{-1}$) and the applicatory HbV ($[Hb] = 8 \text{ g dL}^{-1}$) in a 5 g dL^{-1} rHSA dispersion in actual medical applications.}

Effects of large-molecular-weight HES on the particle motions of HbV. If the HbV is suspended at $[Hb] = 5 \text{ g dL}^{-1}$ in high concentration solutions of HES₆₇₀, MFG, or DEX, the occurrence of HbV flocculation is apparent. HbV exhibits gradual sedimentation and phase separation, as a red blood cell (RBC), does in normal blood. Note that no hemolysis was observed in these flocculating systems. To

explore the arrested dynamics in the flocculating dispersion, we employed brute-force⁴⁴ and echo DLS⁴⁵ experiments. The multi-speckle averaging can be achieved in the brute-force technique by step-by-step altering the sample configurations and performing repeated short time measurements. For a long-time part of ($\tau > 10 \text{ s}$), the sample cell was precisely rotated to explore many independent sample geometries during one evolution of the rotation to observe echo peaks. The time averaging can essentially be identical with the ensemble averaging.

In Figure 10, we display the field correlation function, $g_1(\tau)$, of the concentrated HbV ($[Hb] = 5 \text{ g dL}^{-1}$, [lipids] = 3 g dL^{-1}) dispersed in a series of 3 g dL^{-1} HES solutions. The data of HbV/HES₆₇₀ dispersion were measured with the brute-force and echo DLS techniques in the TC configuration, and other $g_1(\tau)$ functions for HbV in saline and in the lower-molecular-weight HES solutions (HES_{130}} and HES₇₀) are converted from the same $g_2(t)$ data already shown in Figure 9a. In contrast to the rapid single-step relaxation behavior and apparently ergodic nature of the HbV/saline, HbV/HES₇₀, and HbV/HES_{130} systems, the HbV/HES₆₇₀ dispersion shows a significant slowdown of the particle diffusion dynamics and looks like nonergodic within our time window (up to $3 \times 10^4 \text{ sec}$). The process is characterized by the two-step behavior of $g_1(t)$, showing a faster decay with a small amplitude of ca. 0.15, and the following second slower decay. This observation demonstrates that the short-range particle diffusion is highly restricted in a flocculating system due to the depletion (polymer-induced adhesive) interaction between the HbV particles. The slower decay appears to reflect the slow dynamics of the large HbV flocculation in a HbV rich-phase of the phase separated dispersion, which certainly involves an aging effect during the echo experiment. Although we did not perform echo measurements for the HbV/DEX and HbV/MFG dispersions, repeated short-time DLS measurements with rotating the sample cell gave strong fluctuations of the intercept (coherence factor). This is a clear signature of the non-ergodicity or gradual phase-separation of these samples.}

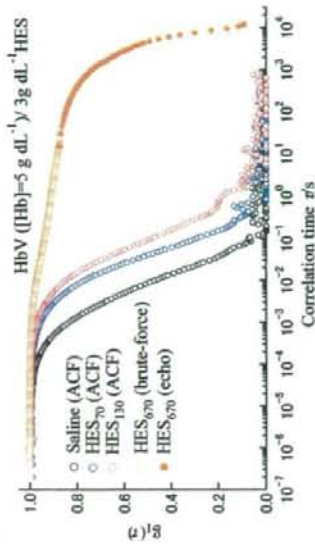


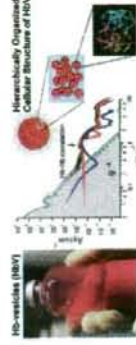
Figure 10; The field correlation functions, $g_2(t)$, of the concentrated HbV ($[Hb] = 5 \text{ g dL}^{-1}$, $[Lipids] = 3 \text{ g dL}^{-1}$) dispersed in saline and in a series of HES solutions. The $g_2(t)$ of the HbV/HES₇₀ was measured using brute-force and echo DLS techniques in the thin layer cell configuration. The $g_2(t)$ data on the HbV in saline and in the lower-molecular-weight HES solutions (HES₇₀ and HES₁₀₀) were converted from the $g_2(t)$ data shown in Figure 9a.

Conclusions

We have investigated static structure and dynamics of HbV, a cellular-type artificial oxygen carrier functionalized as a transfusion alternative. DLS data on a diluted HbV dispersion evaluated by optimized regularization technique (ORT) yielded an averaged hydrodynamic diameter of 238 nm and a 20 nm standard deviation in volume distribution. The SAXS data on the vesicle for Hb encapsulation confirmed its uni-lamellar structure as designed, and the IFT analysis provided the thickness (ca. 5.8 nm) and internal electron density profiles of the bilayer membrane. Consistent with the DLS data, the forward SAXS intensity of the HbV dispersion exhibits a feature of polydisperse spherical particles having an averaged radius of ca 120 nm, while its high q -part manifests the internal density fluctuations reflecting the structure of the encapsulated Hb solution. We found that the position, height, and width of static structure factor of the encapsulated Hbs are quite similar to those in bulk Hb solution at identical protein concentration. The finding demonstrates that confinement into an inner aqueous phase of the vesicle (ca. 240 nm ϕ) does not significantly affect the interparticle interaction of Hbs. TC-DLS combined with the brute-force and echo technique enables us to overcome the interference from multiple scattering of a turbid sample and thus makes it possible to observe collective diffusion of a concentrated HbV dispersion in the presence of various plasma substitutes (water-soluble polymers used to maintain colloid osmotic pressure of blood in medical treatments) without dilution. It should also be mentioned that this technique is perfectly non-invasive, i.e. no external force is applied to the system like in rheology. During the duration of these experiments, the HbV dispersions were perfectly stable. Actually, we confirmed that the dispersion stability of HbV and the oxygen binding function of encapsulated HbV were preserved for over two years at room temperature.⁴⁸ In contrast to the rapid single-step relaxation behavior and apparently ergodic nature of the HbV/saline, HbV/HES₇₀, and HbV/HES₁₀₀ systems, the HbV/HES₆₀ dispersion shows two-step behavior of $g_2(t)$, which looks like nonergodic within our time window, a faster decay with a small amplitude and the following second slower decay indicate that the short-range particle diffusion is strongly restricted in a flocculating system due to the depletion interaction between the HbV particles, while the slow mode appears to reflect the slow dynamics of the

HbV flocculation in a HbV rich-phase of the phase separated dispersion. We note that as we confirmed in the previous rheological study,³⁷ such a HbV flocculation dissociates rapidly under a shear flow and is completely reversible.

Despite a controversy on the mechanism of liposome flocculation, our DLS results, simulated structure factor, and the phase behavior, demonstrate that the underlying mechanism of the HbV flocculation in the presence of plasma substitutes is due to depletion interaction; exclusion of polymer molecules from the region closely spaced HbV particles induces an effective attractive potential between the HbVs, increasing the overall disorder of the system. At the same time, a pending question about a significantly weaker effect of rHSA on HbV flocculation and suspension viscosity enhancement than those induced by other polymers is clearly answered. This is attributed to a compact native structure of the protein, which efficiently reduces the reservoir polymer volume fraction, and thus suppresses the strength of the depletion forces. **The significance of these entropically driven phenomena is highlighted by the tunable suspension rheology of HbVs using different combinations of plasma substitute solutions. High viscosity fluid is occasionally advantageous for sustaining peripheral blood flow, giving shear stress on the vascular wall to facilitate the production of vasorelaxation factors.** The results are not only implicative in the fields of soft-condensed matter physics and bio-chemistry, but of interdisciplinary importance for a new class of forthcoming medical applications.



Acknowledgement The authors thank Dr. Dominique Emi (Inselhospital Hospital, Univ. of Berne), Dr. Amy G. Tsai (Univ. of San Diego), Dr. Masahiko Takaori (East Takarazuka Sato Hospital), and Dr. Koichi Kobayashi (Keio Univ.) for their cooperation and meaningful discussions. This work was supported in part by Health Sciences Research Grants (Research on Publicly Essential Drugs and Medical Devices, H18-Soyaku-Ippan-022), from the Ministry of Health, Labour and Welfare, Japan (H.S., E.T.), Grants-in-Aid for Scientific Research from Japan Society for the Promotion of Science (B19300164) (H.S.), and Young Researchers Empowerment Project of Shinshu University (T.S.) from the Ministry of Education, Culture, Sports, Science and Technology (MEXT), Japan. M.M. and O.G. acknowledge financial support by the European Marie-Curie Research and Training Network on Arrested Matter under Grant No. MRTN-CT-2003-504712. The rHSA, HES₁₃₀, and MFG used in this study were gifts respectively from Nipro Co., Fresenius Kabi A.G., and B. Braun.

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人工赤血球 京都に製造設備 オキシジェニクス

び役であるヘモグロビン
を「リボソーム」と呼ぶ
脂質のカプセルで包んで
作る製剤。二年程度の長
期保存が可能で、血液型
を問わず輸血の代わりに
利用できる。主に災害な
ど緊急時の代替血液とし
て有効とみられている。

パイオベンチャーのオ
キシジェニクス(東京・
海、大村孝男社長)は、
赤血球の代わりに酸素を
運ぶ「人工赤血球」の製
造設備を同社京都研究所
(京都市)内に新設した。

〇七年春にも開始する
治験で使う人工赤血球は
すべて同施設で製造す
る。治験が順調に進み、
対象となる患者が大幅に
増える「第二相後期」に
達した段階で、ニプロに
製造委託する。それ以降、
同施設は研究開発用の新
製剤製造に転用する。

ニプロ、人工血液事業参入

バイオVβと業務提携

ニプロは人工血液事業 ける人工血液の製造を受 人工血液は直径二百ミ
に乗り出す。人工血液を 託する。臨床試験(治験) (γは十億分の一)位の
共同研究中のバイオベン に使う高品質の人工血液 微小カプセルに、約三週
チャーのオキシエニク を一〇〇八年から製造。 間の使用期限が切れた輸
ス(東京・港)と業務提携 臨床試験の進ちよく状況 血用の血液から採取した
携し、手術などの際に血 に応じ、段階的に量産化 有効成分を封入したも
液の代わりに「輸血」で を進める。 の。欧米で臨床試験が進

め、災害などの緊急備
用に全国で需要が見込め
るといふ。

オキシエニクスは人

んでいるが、まだ製品化
には至っていない。二年
程度の長期保存が可能
で、血液型も問わないた

工血液を自社で製造し、
来年春から日欧で臨床試
験を始める。
有効性の確認が順調に
進み、対象患者が大幅に

増える後期の臨床試験に
着手するタイミングで、
ニプロの製造に切り替え
る。一五年の製品化を目
指す。

○七年春から人工血液を自社製造して日欧で治験(臨床試験)を開始。安全性などの確認が進んで対象患者が増えるとみられる二〇〇八年からニプロに製造を委託する計画。

この人工血液は「人工酸素運搬体」と呼び、赤血球の代わりに酸素を運ぶ。約三週間の使用期限が切れた輸血用血液から有効成分を抽出し、直径二百五十μ(μは十億分の一)の脂質の微小カプセルに包んで作る。

約二年の長期保存が可能。血液型も問わない。災害などの緊急備蓄の需要が見込まれている。

人工血液事業化
ニプロと提携
オキシエニクス
バイオベンチャーのオキシエニクスは二〇
キシエニクス(東京・
湾、大村孝男社長)とニ
プロは二十三日、人工血
液の事業化を目指して薬
務提携すると発表した。
オキシエニクスは二〇
欧米メーカーが先行し
て治験を進めているが、
製品化には至っていない。国内ではテルモが〇
七年春に治験を始める計
画を発表している。

☆オキシエニクス

酸薬運搬体製造で二プロと提携

バイオベンチャーのオキシエニクスは23日、二プロと人工酸薬運搬体「オキシエニキヤリア(OXY-0301)」の製造で業務提携した。両社は2004年に3年間の共同開発契約を結んだが、今回の製造契約締結で、二プロは07年からOXY-0301のGMP(医薬品の製造管理および品質管理に関する基準)準拠製造プラントの設計を開始し、08年から同治療薬の製造、早期の大規模製造と実用化を目指す。

(人事)

◇日本ケミコア(6月29日付予定)〔役員異動〕(新任取締役候補)社外取締役(非常勤)・茂原敏明(新任監査役)社外監査役(非常勤)。

アポットなど3社

「ホクナリンテープ」表面に品名表示

アポットジャパン、マルホ、日東電工は22日、貼付型の気管支拡張剤「ホクナリンテープ」(一般名ヒソプロテロール)の表面に、安全確保を目的として、製品名を表記したと発表した。テープ表面に製品名表示がないと、患者、医療従事者などが、何の薬剤を貼り付けたのか分からないという問題点が指摘されていた。今回、テープ表面に製品名を表示することで、その問題に対応した。

(日本ケミコア)

電子公告など定款変更を提案へ

日本ケミコアは22日の取締役会で、電子公告ができるようにするためのなどの定款変更議案を、6月29日開催予定の第74回定時株主総会に付議することを決めた。公告閲覧の利便性向上や株主総会事務処理の効率化などが主な目的。また取締役会の機動的、効率的運営を図るため、取締役全員の同意があり、監査役が異議を述べないときに限り、取締役会の書面決議を可能とする内容も盛り込まれている。

高橋剛(退任予定監査役)山中徹(監査役(非常勤))

◇アクロネット(5月23日付)〔役員人事〕

代表取締役社長・相松尚

〔開催〕日本能率協会は6月26日(月)午前10時、大阪市北区の同協会関西地域事業部研修室で、「第11回医薬品GMP教育訓練担当者養成コース」を開催する。ファーマサイブスイコマ代表の相澤邦平氏が、①GMP教育訓練にかかわる法規事項などの概要②GMPの歴史とSOPの情報③情報の効果的活用―などについて講演する。参加費は同協会会員3万6750円、非会員4万2000円で、5月31日(水)までの申し込みは、それぞれ3万1500円、3万7800円の割引価格(いずれも税込み)。問い合わせ・申し込みは、同協会関西地域事業部(Tel.06-147977-2052、FAX06-147977-2051)へ。

(本号9頁)

●わかかもと製薬 2006年3月期決算概況

【連結】	当 期	売上高	営業利益	経常利益	単位：百万円、%	
					当期利益	当期利益
		10,185(1.5)	673(△8.6)	744(△12.3)	448(△8.0)	
次期中間期予想		5,000	—	320	200	
次期通期予想		10,200	—	670	420	
【単体】						
当 期		10,142(1.5)	650(△10.0)	731(△13.6)	441(△8.1)	
次期中間期予想		4,980	—	310	195	
次期通期予想		10,150	—	655	410	
配 当 異 議		中間 2.50円	期末 2.50円		年間 5.00円	
次期中間期予想		中間 2.50円	期末 2.50円		年間 5.00円	

輸血不足を解決するか

人工“酸素”運搬体(人工赤血球)

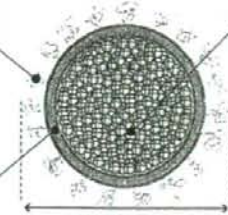
テルモなど日欧米で研究開発進む

人工酸素運搬体が人工血液の可能性を上げようとしている。

我々ヒトという生物は、全

オキシジェニクスの「Oxygen Carrier」の構造と特徴

・PEG表面修飾による物理的安定度向上
(長期保存可)
・血液成分との相互作用回避
(血中滞留性の延長)



・ヒトHbと同等の高いHb濃度
ウィルスを除去した高純度精製Hbの内包
赤血球と類似構造による副作用の回避
脱酸素保存による化学的安定度向上
(長期保存可)

・混合リン脂質型合成脂質の使用により血小板活性化回避

粒径約250nm

身に約60兆個と言われる細胞と、それを中枢制御する脳というスーパーコンピュータから成り立っている。全身の臓器、組織など全ての生命活動の維持には膨大なエネルギーを必要とし、そのエネルギーは基本的には、細胞内で糖などの呼吸基質と酸素との酸化反応で生み出されている。細胞呼吸と呼ばれる代謝である。

体内に取り込まれた、エネルギーを作り出す元となる酸素を運搬する細胞は血液全体の45%の量である赤血球であり、酸素が結合するのは赤血球にあるヘモグロビンというタンパク質のヘムという真っ赤な分子の真ん中にある鉄分である。各臓器で、酸素はこの鉄から離れ、細胞に存在する間、解糖系、クエン酸回路を経てミトコンドリアの電子伝達系に取り込まれてエネルギー生産に使われるのだ。それだけに手術などの治療に用いられる血液はまさに余命の源となる。

ところが世界的に見て、輸血用の血液は、「AIDS」等の問題もあって供給不足の状況にあるとされる。

厚生労働省の資料によれば、日本において、日本赤十字社が製造・供給している輸血用血液製剤はすべて国内の献血で賄われており、その年間確保量は約200万ℓ。その内の約半分が赤血球製剤という。輸血用製剤の有効期限は全血製剤と赤血球製剤で21日間(3週間)、血小板製剤が採血後72時間とされている。この赤血球製剤の使用期限の限界があることから、現在の献血システムで救急救命用途や災害用備蓄として賄いきれていないという現状がある。

”日本発”の世界的人工血液技術

そこで開発が進められているのが、酸素を運搬するといふ、生命にとって重要な機能を備えた「人工血液」＝「人工赤血球」である。欧米の数

JAPAN AIR GASES

わたしたちが提供するのは、
社会になくてはならない産業ガス。



産業ガスの供給、ガス応用機器の供給、そして各種付加価値サービスの提供。
最先端科学から、生活に密着した身近なものまで、私たちの業務はさまざまな形で
皆さまの暮らしを支えています。世界をリードする2大産業・医療ガスグループ
「エア・リキードグループ(株)」[BOCグループ(株)]。その、最先端技術・インフラが
利用できるという私たちだけの強みを最大限に活かし、これからもずっと、世界中から評価を受ける
「イノベーション」を提供していきます。常に皆さまのニーズを第一に考えながら…。

ジャパン・エア・ガズ株式会社 〒135-0062 東京都江東区東横1丁目9番1号 <http://www.japanairgases.co.jp/>

社のベンチャー企業、日本ではテルモ、そしてパイオベンチャーのオキシジェニクスが研究・開発に携わっている。ヒトや牛のヘモグロビンを

使って造られて来たこれまでの「人工血液」は、「ヘモグロビン」が実際の赤血球とは異なる「ヘモグロビン」そのものは、その大きさ、形状から血管壁の閉塞を通過してしまい、NO（一酸化窒素）やCOと結合してしまふ」など実用段階での安全性が確認されないことから、実用化されていくのが現状らしい。

身体におけるガスの働きを創薬に応用する。

慶應義塾大学医学部

医学教室 末松誠教授

前述のオキシジェニクス（本社・東京都港区）は早稲田大学理工学部と慶應義塾大学医学部の共同研究による技術シーズの事業化を目的に、02年12月に設立されたベンチャー企業である。本社の他、京都と横浜に研究所を持ち、慶應義塾大学医学部信濃町キャンパス（東京都新宿区）総合医学研究棟に基礎研究センターを置く。人工酸素運搬体（酸素治療薬）「Oxygen Carrier」の研究開発、用途開発事業を中心に、体内におけるGassの働き「Gass Biology」をベースとした創薬開発事業も進めている。この「Gass Biology」

そこで日本のオキシジェニクス、テルモの世界的に見て、2社のみが開発したのが、酸素運搬体であるヘモグロビンをリポソームという脂質の二重膜で内包する技術である。テルモとオキシジェニクスとはその技術に違いがあるらしい。人工酸素運搬体の特長といえば、例えばオキシジェニクスの「Oxygen Carrier」の場合、室温での長期保存安定性があることとされ、粒径250nmに設計されていることから10〜100nmとされる血管壁の閉塞を通過しない。特に、他社海外製

品を研究テーマとして数年前から取り組んで来たのが、同社の設立にも関わり、現在、技術顧問となっている慶應義塾大学医学部医学教室の末松誠教授である。

末松教授の研究テーマである「Gass Biology」の「Gass」とは主に酸素、そしてNO（一酸化窒素）とCO（一酸化炭素）のことを指す。まず、NOに関して研究開発したものについては、「ヘモグロビンは1分子当たり、 α 1、 α 2、 β 1、 β 2の4つのポケットがあり、2つの α のポケットをNOで満たした（NOで安定させた） α NOを持つたヘモグロビンは酸素運搬能力が高（酸素の放出量が多）くなり、解糖系の代謝が活発になる。また、血管拡張作用を持つATP（アデノシン三リン酸）の放出を活発に

品の修飾型ヘモグロビンと比較してNOやCOとの相互作用を引き起こさない適切なサイズ（250nm）に設計されているものであるという。さらに250nmという粒径は8000nmのヒト赤血球と比較した場合、狭窄血管への酸素輸送が可能で、ガン、虚血性疾患治療に使われる「酸素治療薬」としての使用の可能性も高いという。

医薬品業界ではDDS（ドラッグデリバリーシステム）、またGDS（ジーンデリバリーシステム）といわれるのが一般的だが、「GII GAS」

し、血流を流れやすくする。ヘモグロビンは局所の酸素濃度を感知しながら各種の代謝物を活発に出し入れするメタボリックサイトとして作用することが明らかになっているわけだ。虚血性疾患、肝障害など、低酸素状態になっているところにおけるNOを処理（付加）した赤血球の効果がラットを使った実験により確認できている（末松教授）と説明する。またCOについては、輸血して保存されている血液は4℃でも代謝が起きているがCOを付加することで代謝が抑えられることを確認しており、輸血の保存に役立つという。

研究開発に使うNOは溶剤を使い、直接ガスのNOを使うものではないが、COはガスディレーターから購入したものを使っているようだ。

で、ガスデリバリーシステムという概念もあり、酸素を代表とする、生体内に存在する各種ガスの働きを追求し、ガスバイオロジーをベースとした創薬研究にも取り組む動きがここに来て出て来たと言え



るだろう。特に、治療薬として酸素が活用されるのは注目すべきテーマだ。尚、同社はGMP製造設備を整えた京都研究所で製造された製剤を用いて来年には臨床試験（治験）を開始する計画である。

命を守る最前線で。
健やかな暮らしを願う
心の中に。
いつも星医療酸器は
あなたといたい。

30年以上にわたる医療用ガスの提供サービス。その中で私たちは、常に「命」と「暮らし」に真正面から向き合い、自らの可能性を切り拓いてきました。そして今、高齢化・情報化が進む社会に即応するべく、新しい事業の開拓・確立へ、社会との接点は、ますます広がりつつあります。



株式会社 星医療酸器

〒121-0836 東京都足立区入谷7-11-12
TEL 03-3899-8855 FAX 03-3899-5661
URL <http://ns1.hosi.co.jp/>

人工赤血球

実用化に向けて最終段階

「人工赤血球」が体の全細胞の呼吸に必要な充分量の酸素を運搬する

日本の献血・輸血システムは世界的水準にあり、国民の医療と健康福祉に多大の貢献をしている。献血血液の厳重な検査により、安全性のさらなる向上が図られている。他方、「人工赤血球」の開発が厚生労働省の積極的な支援も受けて急速に進められている。その特徴は、1.血液型がないので、ただちに投与可能。2.ウイルスなど感染源がない。3.室温で2年間安定なので災害用に備蓄して救急救命に使用できる。4.大量投与しても体内の代謝系を経て分解排泄される。などである。また、5.原料確保のための遺伝子組換え技術によるヘモグロビン、アルブミンの供給技術も利用可能になってきている。思いきった国策の採用とあわせ、国民の健康福祉と安全の確保に賢くべき貢献をもたらすことになろう。

実現が最も近いとされる人工赤血球が、土田英復 早稲田大学教授らの開発した「ヘモグロビン小胞体」だ。酸素結合タンパク質「ヒトヘモグロビン」約3万個を脂質二分子膜で包んだ直径250nmの小球粒子で、ヘモグロビン分子単体の毒性をささぎっている。小林祐一慶應義塾大学医学部教授ほか、国内外の研究機関との共同で、動物投与と試験による安全度と効能の確認を進めている。大量出血によるショック状態にヘモグロビン小胞体を投与すると、酸素供給が速やかに回復し救命できること、微小なヘモグロビン小胞体は血管中に均一に分散し、赤血球の到達が困難な末梢組織まで酸素供給できることがわかった。富良野での安全性も確認済みで、2006年内に臨床試験を申請する予定だ。

'06.6.11

トピック

■ オキシジェニクスがニプロと

人工酸素運搬体の製造で契約

バイオベンチャーのオキシジェニクス（東京都港区）は5月23日、ニプロとの間で、赤血球の代用品となる人工酸素運搬体「Oxygen Carrier（開発コード=OXY-0301）」の将来製造に関する業務契約を締結したと発表した。

同社は、ニプロと2004年から3年間の共同開発契約を結び、製造技術開発に着手。5月にミニスケールの治験薬製造プラントを稼働させ、ニプロが量産化に向けたスケールアップの検討を進めることとなった。

今回の契約によりニプロは、08年か

らOXY-0301の治験薬の製造を開始する。同社は、研究・開発に関する情報交換などをさらに強化し、今後、海外戦略についても検討していく。

同社は、早稲田大理工学部と慶応大医学部の共同研究による技術シーズを事業化するため、02年12月に設立された。