

- [18] Nistico L, Buzzetti R, Pritchard LE, Van der Auwera B, Giovannini C, Bosi E, et al. The CTLA-4 gene region of chromosome 2q33 is linked to, and associated with, type 1 diabetes. Belgian Diabetes Registry. *Hum Mol Genet* 1996;5:1075–1080.
- [19] Oaks MK, Hallett KM, Penwell RT, Stauber EC, Warren SJ, Tector AJ. A native soluble form of CTLA-4. *Cell Immunol* 2000;201:144–153.
- [20] Fan LY, Tu XQ, Cheng QB, Zhu Y, Feltens R, Pfeiffer T, et al. Cytotoxic T lymphocyte associated antigen-4 gene polymorphisms confer susceptibility to primary biliary cirrhosis and autoimmune hepatitis in Chinese population. *World J Gastroenterol* 2004;10:3056–3059.
- [21] Bittencourt PL, Palacios SA, Farias AQ, Abrantes-Lemos CP, Cancado EL, Carrilho FJ, et al. Analysis of major histocompatibility complex and CTLA-4 alleles in Brazilian patients with primary biliary cirrhosis. *J Gastroenterol Hepatol* 2003;18:1061–1066.
- [22] Donaldson P, Veeramani S, Baragiotta A, Floreani A, Venturi C, Pearce S, et al. Cytotoxic T-lymphocyte-associated antigen-4 single nucleotide polymorphisms and haplotypes in primary biliary cirrhosis. *Clin Gastroenterol Hepatol* 2007;5:755–760.
- [23] Parks CG, Pandey JP, Dooley MA, Treadwell EL, St Clair EW, Gilkeson GS, et al. Genetic polymorphisms in tumor necrosis factor (TNF)-alpha and TNF-beta in a population-based study of systemic lupus erythematosus: associations and interaction with the interleukin-1alpha-889 C/T polymorphism. *Hum Immunol* 2004;65:622–631.
- [24] Correa PA, Gomez LM, Cadena J, Anaya JM. Autoimmunity and tuberculosis. Opposite association with TNF polymorphism. *J Rheumatol* 2005;32:219–224.
- [25] Mitchell SA, Grove J, Spurkland A, Boberg KM, Fleming KA, Day CP, et al. Association of the tumour necrosis factor alpha-308 but not the interleukin 10-627 promoter polymorphism with genetic susceptibility to primary sclerosing cholangitis. *Gut* 2001;49:288–294.
- [26] Gordon MA, Oppenheim E, Camp NJ, di Giovine FS, Duff GW, Gleeson D. Primary biliary cirrhosis shows association with genetic polymorphism of tumour necrosis factor alpha promoter region. *J Hepatol* 1999;31:242–247.
- [27] Jones DE, Watt FE, Grove J, Newton JL, Daly AK, Gregory WL, et al. Tumour necrosis factor-alpha promoter polymorphisms in primary biliary cirrhosis. *J Hepatol* 1999;30:232–236.
- [28] Tanaka A, Quaranta S, Mattalia A, Coppel R, Rosina F, Manns M, et al. The tumor necrosis factor-alpha promoter correlates with progression of primary biliary cirrhosis. *J Hepatol* 1999;30:826–829.
- [29] Donaldson PT. TNF gene polymorphisms in primary biliary cirrhosis: a critical appraisal. *J Hepatol* 1999;31:366–368.
- [30] Donaldson P, Agarwal K, Craggs A, Craig W, James O, Jones D. HLA and interleukin 1 gene polymorphisms in primary biliary cirrhosis: associations with disease progression and disease susceptibility. *Gut* 2001;48:397–402.
- [31] Leslie EM, Deeley RG, Cole SP. Multidrug resistance proteins: role of P-glycoprotein, MRP1, MRP2, and BCRP (ABCG2) in tissue defense. *Toxicol Appl Pharmacol* 2005;204:216–237.
- [32] Sarkadi B, Homolya L, Szakaes G, Varadi A. Human multidrug resistance ABCB and ABCG transporters: participation in a chemoinnate defense system. *Physiol Rev* 2006;86:1179–1236.
- [33] Lam P, Wang R, Ling V. Bile acid transport in sister of P-glycoprotein (ABCB11) knockout mice. *Biochemistry* 2005;44:12598–12605.
- [34] Kimura Y, Selmi C, Leung PS, Mao TK, Schauer J, Watnik M, et al. Genetic polymorphisms influencing xenobiotic metabolism and transport in patients with primary biliary cirrhosis. *Hepatology* 2005;41:55–63.
- [35] Zein CO, Beatty K, Post AB, Logan L, Debanne S, McCullough AJ. Smoking and increased severity of hepatic fibrosis in primary biliary cirrhosis: a cross validated retrospective assessment. *Hepatology* 2006;44:1564–1571.
- [36] Gershwin ME, Selmi C, Worman HJ, Gold EB, Watnik M, Utts J, et al. Risk factors and comorbidities in primary biliary cirrhosis: a controlled interview-based study of 1032 patients. *Hepatology* 2005;42:1194–1202.
- [37] Karlsten TH, Lie BA, Frey Frosli K, Thorsby E, Broome U, Schrupf E, et al. Polymorphisms in the steroid and xenobiotic receptor gene influence survival in primary sclerosing cholangitis. *Gastroenterology* 2006;131:781–787.
- [38] Banales JM, Arenas F, Rodriguez-Ortigosa CM, Saez E, Uriarte I, Doctor RB, et al. Bicarbonate-rich choleresis induced by secretin in normal rat is taurocholate-dependent and involves AE2 anion exchanger. *Hepatology* 2006;43:266–275.
- [39] Lardner A. The effects of extracellular pH on immune function. *J Leukoc Biol* 2001;69:522–530.
- [40] Prieto J, Qian C, Garcia N, Diez J, Medina JF. Abnormal expression of anion exchanger genes in primary biliary cirrhosis. *Gastroenterology* 1993;105:572–578.
- [41] Medina JF, Martinez A, Vazquez JJ, Prieto J. Decreased anion exchanger 2 immunoreactivity in the liver of patients with primary biliary cirrhosis. *Hepatology* 1997;25:12–17.
- [42] Prieto J, Garcia N, Marti-Climent JM, Penuelas I, Richter JA, Medina JF. Assessment of biliary bicarbonate secretion in humans by positron emission tomography. *Gastroenterology* 1999;117:167–172.
- [43] Salas JT, Banales JM, Sarvide S, Recalde S, Ferrer A, Uriarte I, et al. Ae2_g,b deficient mice develop antimitochondrial antibodies and other features resembling primary biliary cirrhosis. *Gastroenterology* 2008;134:1482–1493.

Original Article

Correlation between *CFH* Y402H and *HTRA1* rs11200638 genotype to typical exudative age-related macular degeneration and polypoidal choroidal vasculopathy phenotype in the Japanese populationNorimoto Gotoh MD,^{1,2} Ryo Yamada MD,² Hideo Nakanishi MD,^{1,2} Masaaki Saito MD,³ Tomohiro Iida MD,³ Fumihiko Matsuda PhD^{2,4} and Nagahisa Yoshimura MD¹¹Department of Ophthalmology and ²Center for Genomic Medicine, Kyoto University Graduate School of Medicine, Kyoto, and³Department of Ophthalmology, Fukushima Medical University School of Medicine, Fukushima, Japan, and ⁴Centre National de Génotypage, Evry, France

ABSTRACT

Background: Typical exudative age-related macular degeneration (AMD) and polypoidal choroidal vasculopathy (PCV) are two of the major macular diseases found in Asians. Although genomic studies have shown a contribution by *CFH* and LOC387715/*HTRA1* polymorphisms to the development of these two diseases, the correlation of the clinical phenotypes to these genotypes has not been determined in Asian patients.

Methods: The prevalence of the *CFH* Y402H and *HTRA1* rs11200638 genotypes was determined in 116 patients with typical exudative AMD and in 204 patients with PCV. Potential correlations of these polymorphisms were tested retrospectively and cross-sectionally for bilaterality of the disease, final visual acuity and the greatest linear dimension of the choroidal neovascular (CNV) lesion.

Results: There was no significant difference in the incidence of *CFH* Y402H ($P=0.598$) and *HTRA1* rs11200638 ($P=0.290$) between eyes with typical exudative AMD and with PCV. There was a significant association between the lesion size and *HTRA1* rs11200638. For eyes with typical AMD, the size of the lesion ($6363 \pm 2837 \mu\text{m}$) was significantly larger in the high-risk homozygous group (AA), than in the low-risk homozygous group (GG) ($3866 \pm 1947 \mu\text{m}$; $P=0.0003$). The same tendency was observed for the size of the lesion in PCV cases (homozy-

gous group: $6347 \pm 2673 \mu\text{m}$, non-risk homozygous group: $4405 \pm 2066 \mu\text{m}$, $P=1.3 \times 10^{-5}$).

Conclusions: A common genetic background may exist between typical exudative AMD and PCV patients. Among the patients with these two clinical entities, those with a homozygous *HTRA1* rs11200638 risk allele had larger CNV lesions.

Key words: age-related macular degeneration, choroidal vasculopathy, genetic analysis.

INTRODUCTION

Macular diseases are the most common cause of visual impairment in the elderly in developed countries.^{1,2} The impact of macular diseases is similar for Caucasians and Asians, but the prevalence of the different types of macular disease differs between the two populations.^{3,4} In Asians, typical exudative age-related macular degeneration (AMD) and polypoidal choroidal vasculopathy (PCV) account for around 90% of all cases of senile macular disease. Of these, the percentage of eyes with PCV is about 30–50%, and geographic atrophy (GA) is quite rare in Asians.^{3,5} Although the pathophysiology of typical AMD and PCV has been believed to be different,⁶ there is an overlap in the clinical findings for both entities, at least in type I choroidal neovascularization (CNV).⁷

Genetic studies of Caucasian patients with AMD have detected two disease-contributing polymorphisms.

■ Correspondence: Dr Nagahisa Yoshimura, Department of Ophthalmology, Kyoto University Graduate School of Medicine, Shogoinkawaharacho 54, Sakyo, Kyoto 606-8507, Japan. Email: nagaeyef@kuhp.kyoto-u.ac.jp

Received 14 November 2007, accepted 6 June 2008

© 2008 The Authors
Journal compilation © 2008 Royal Australian and New Zealand College of Ophthalmologists

Complement factor H (*CFH*) on chromosome 1q32 has been identified by many studies as a major AMD-susceptible gene polymorphism in Caucasians.⁸⁻¹² In addition, biochemical and pathological studies have shown that a *CFH* Y402H polymorphism may be involved in AMD.^{13,14}

Chromosome 10q26 has also been found to carry another important AMD susceptibility locus. Several studies have used refined linkage disequilibrium (LD) mapping and case-control association studies to probe the most susceptible alleles, *LOC387715* rs10490924 and *HTRA1* rs11200638, for AMD.¹⁵⁻¹⁸ In Asians, replication studies of cases of typical AMD have been reported for *CFH* Y402H.¹⁹⁻²⁴ In those with typical AMD, the contribution from *CFH* Y402H was found to be less strong than that for Caucasians. This is due to the much lower frequency of the risk allele, Y402H, that is seen in <6% of Asians compared with >35% of Caucasians.¹⁹⁻²⁵ The relationship of *LOC387715* rs10490924 and *HTRA1* rs11200638 polymorphisms to typical AMD and PCV has been investigated in Asians.^{15,26-30} Several studies have shown an almost complete LD in the two SNPs,^{15,29,30} with both showing an identical and very strong positive association not only with typical AMD but also with PCV.³⁰

Positive correlations have been reported between the phenotypes of Caucasian patients with exudative AMD and the polymorphic genotypes. Goverdhan *et al.* suggested that the risk allele *CFH* Y402H is significantly more prevalent in individuals with predominantly classic CNV, and that those who are homozygous for the *CFH* Y402H genotype tend to have the poorest visual acuity after the photodynamic therapy.³¹ Chen *et al.* reported that the *HTRA1* rs11200638 homozygote odds ratios of wet AMD and GA are significantly greater in bilateral than in unilateral cases.³² In Asians, we were unable to find any studies on the phenotype-genotype correlations.

Thus, the purpose of this study was to determine whether a significant correlation exists between the two major disease-associated polymorphisms, *CFH* Y402H and *HTRA1* rs11200638, and the clinical genotypes in cases of typical AMD and PCV in the Japanese population.

METHODS

Subjects

From April 2005 to December 2006, we studied 116 Japanese patients with typical AMD and 204 patients with PCV at the Center for Macular Diseases, Department of Ophthalmology, Kyoto University Hospital, and at the Department of Ophthalmology, Fukushima Medical University Hospital. All of the patients signed a written informed consent form (Table 1). Although the medical histories of the patients differed, all patients were being treated for exudative lesions. All patients had a complete ophthalmological examination including fluorescein and indocyanine green angiography with a scanning laser ophthalmoscope (HRA2, Heidelberg, Germany).

The inclusion criteria for PCV were the presence of reddish-orange, spheroidal, polypoidal structures detected

Table 1. Characteristics of study population

Variable	tAMD	PCV
Number	116	204
Age (mean)	76.1	73.1
SD	8.27	7.70
Gender (% male)	74.1%	71.5%

PCV, polypoidal choroidal vasculopathy; SD, standard deviation; tAMD, typical exudative age-related macular degeneration.

by slit-lamp biomicroscopy of the macula with a contact lens, and the presence of a branching vascular network of choroidal vessels with terminal aneurysmal dilations detected by indocyanine green angiography. The clinical presentation and angiographic findings were used to exclude secondary CNV diseases, for example, angioid streaks, degenerative myopia, idiopathic CNV and presumed ocular histoplasmosis syndrome.

The visual acuities of both eyes at the final examination were converted to logarithm of the minimum angle of resolution (logMAR) units. Whether the disease was monocular or binocular was also recorded. Before the photodynamic therapy, the greatest linear dimension (GLD) of the CNV lesion was measured on the angiographic images obtained by the HRA2 and the Heidelberg Eye Explorer software (Heidelberg).^{33,34}

Genotyping

CFH Y402H rs1016670 (dbSNP build 126) was genotyped using the Taqman SNP assay (Applied Biosystems, Foster City, CA, USA). *HTRA1* rs11200638 was genotyped by direct sequencing with the following PCR primer sets: 5'-CACGTGTGAAGGATTCTATTTCGAA-3' and 5'-CGTCCTTCAAACCTAATGGAACCTT-3'. The sequencing reactions were performed by the Dye Terminator method with an ABI PRISM 3730 DNA Analyzer (Applied Biosystems). The alignment of the sequences, SNP discovery and genotyping were performed with Genalys (<http://www.software.cng.fr/docs/genalys.html>).³⁵

Statistical analyses

The deviations of the allelic distributions and bilaterality were assessed with the χ^2 -test. The visual acuity of the poorer eye at the final visit and the lesion size were analysed with a one-way ANOVA. *P*-values obtained by these analyses were corrected for multiple testing using the Bonferroni correction by Statistica (StatSoft, Tulsa, OK, USA). The power calculation was made with the R public open software and its tools (<http://www.r-project.org/>).

RESULTS

CFH Y402H polymorphism and phenotypes

The genotype and allelic distributions for *CFH* Y402H are given in Table 2. The frequency of the risk allele C was 0.129

Table 2. Distribution of single nucleotide polymorphisms *CFH* Y402H and *HTRA1* rs11200638 in Japanese patients with tAMD and PCV

	tAMD (116 cases)		PCV (204 cases)	
	<i>CFH</i> Y402H	<i>HTRA1</i> rs11200638	<i>CFH</i> Y402H	<i>HTRA1</i> rs11200638
Genotype	CC/CT/TT	AA/GA/GG	CC/CT/TT	AA/GA/GG
Count	4/22/90	48/47/21	2/43/159	73/88/43
Allele	C/T	A/G	C/T	A/G
Count	30/202	143/89	47/361	234/174
Bilaterality				
Monocular	CC/CT/TT 1/16/64	AA/GA/GG 29/37/15	CC/CT/TT 1/31/119	AA/GA/GG 50/71/30
Binocular	CC/CT/TT 3/6/26	AA/GA/GG 19/10/6	CC/CT/TT 1/12/40	AA/GA/GG 23/17/13

PCV, polypoidal choroidal vasculopathy; tAMD, typical exudative age-related macular degeneration.

in typical AMD patients and 0.115 in PCV patients. This difference in the frequency of the C allele was not significant ($P = 0.598$).

We next determined whether a correlation between the risk allele of *CFH* Y402H and the clinical presentation was significant. Six patients with the risk of homozygous CC were studied, and analyses were made between patients having at least one C allele and those having a non-risk TT genotype. Among the patients with typical AMD, those with the C allele showed a tendency to be binocularly affected in 34.6%, and those with the TT genotype in 28.9% (Table 2).

The visual acuity of patients with typical AMD with the C allele was 0.71 ± 0.50 , and that for the TT genotype was 0.94 ± 0.55 (logMAR system). In patients with PCV, the correlation between the presence of the C allele and bilaterality (28.9%) and the TT genotype (25.2%) was not significant. The visual acuity for PCV patients with the C allele was 0.70 ± 0.57 and 0.64 ± 0.53 with the TT genotype. Although the differences for bilaterality and visual acuities between the groups was not statistically significant ($P > 0.055$), a power calculation indicated that the number of participants was too small to evaluate the negative findings for either bilaterality or visual acuity.

The GLD of the patients with typical AMD with the C allele was $4976 \pm 2349 \mu\text{m}$ and that for the TT genotype was $5381 \pm 2679 \mu\text{m}$ ($P = 0.491$). The GLD of patients with PCV with the C allele was $5376 \pm 2323 \mu\text{m}$ and that for the TT genotype was $5219 \pm 2598 \mu\text{m}$ ($P = 0.715$). Because the association between GLD and the *CFH* Y402H variant in our sample was not significant, we calculated that we would have to analyse almost 1000 patients in order to achieve a power of 0.8 ($\alpha = 0.05$).

HTRA1 rs11200638 polymorphism and phenotype

The genotype and allelic frequencies for *HTRA1* rs11200638 are given in Table 2. The frequency of the risk allele A of rs11200638 was 0.616 in typical AMD patients and 0.574 in

PCV patients. This difference was not significant ($P = 0.290$).

We studied whether there was any significant correlation between the risk allele of rs11200638 and the clinical phenotype. In patients with typical AMD, there was a greater tendency for the AA genotype to be found in patients binocularly affected (39.6%) than in patients with the GG genotypes (28.6%). In patients with PCV, an association of the AA genotype with bilaterality (31.5%) or the GG genotype (30.2%) was not significant (Table 2). A power calculation showed that because of the small number of participants, a statistical evaluation of the genetic influence for both bilaterality and visual acuity could not be done.

In typical AMD, the average GLD size in the patients with GG ($3866 \pm 1947 \mu\text{m}$) was significantly smaller than that in those with AA genotype ($6363 \pm 2837 \mu\text{m}$; $P = 0.0003$, Fig. 1). In PCV, the GLD was significantly smaller for the GG group ($4405 \pm 2066 \mu\text{m}$) than for the AA group ($6347 \pm 2673 \mu\text{m}$; $P = 1.3 \times 10^{-5}$, Fig. 1).

Only one patient was found to have double-risk polymorphisms, namely, CC genotype in *CFH* Y402H and AA genotype in *HTRA1* rs11200638. This patient was diagnosed with typical AMD with a GLD size of $5250 \mu\text{m}$ in the one eye that was affected. No statistical analysis was possible.

DISCUSSION

We investigated whether a significant correlation exists between two disease-associated polymorphisms with AMD and PCV that are present in both Caucasians and Asians. PCV was first described by Yannuzzi,³⁶ who called it, idiopathic PCV. After a decade of many studies, the detail characteristics of PCV were finally established.⁶ These studies showed that PCV is a distinct clinical entity that is more common in individuals with pigmented skin including Asians. In addition, PCV is a different clinical entity than typical CNV that is found in patients with AMD, and indocyanine green angiograms are necessary to make a definitive diagnosis.⁶ Recently, this condition has been referred to as polypoidal CNV, and classified as PCV with

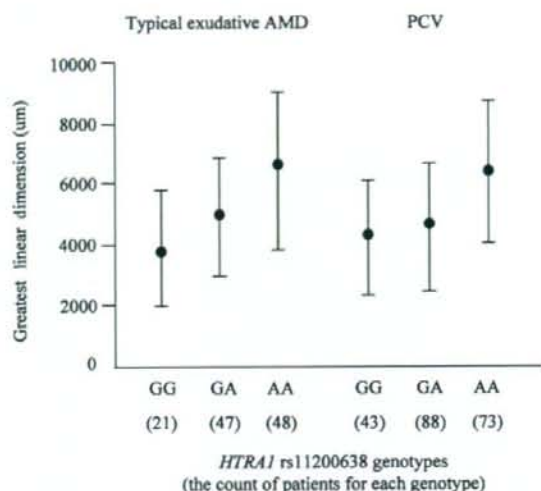


Figure 1. Correlations between the greatest linear dimension (GLD) in patients with typical exudative age-related macular degeneration (AMD) and patients with polypoidal choroidal vasculopathy (PCV) with the *HTRA1* rs11200638 genotypes. The circles represent mean lesion size and error bars represent standard deviation in patients carrying each genotype. Patients with typical AMD and the AA genotype had a significantly larger lesion size ($6363 \pm 2837 \mu\text{m}$) than the other genotypes (GA, $4830 \pm 2159 \mu\text{m}$; GG, $3866 \pm 1947 \mu\text{m}$; $P = 0.0003$) as well as in the patients with PCV (AA $6347 \pm 2673 \mu\text{m}$, GA, $4762 \pm 2316 \mu\text{m}$ and GG, $4405 \pm 2066 \mu\text{m}$; $P = 1.3 \times 10^{-5}$).

type I CNV.⁷ Although many pathologic studies have been reported on whether PCV is abnormal choroidal vessels, intra-Bruch CNV, or a mixture of both is still undetermined.³⁷⁻⁴²

Our results showed that the frequency of the risk allele of *CFH* Y402H was not significantly different in patients with typical AMD and PCV. For *CFH* Y402H, Johnson *et al.* reported that individuals who are homozygous for the *CFH* Y402H risk allele have elevated levels of C-reactive protein in the choroid.¹³ Lommatzsch *et al.* reported that a pathologic examination of classic CNV or occult CNV revealed that the subjects were either heterozygous or homozygous for the *CFH* Y402H risk allele.¹⁴ In addition, in several studies in which no functional experiments were performed, more significant associations were reported in the haplotypes with other SNPs in both Caucasian and Asian populations.^{12,20,21,24} In our study, we only reported data of *CFH* Y402H, and thus further studies are needed to be able to clearly define the contribution of other SNPs or haplotypes in *CFH*.

The frequency of the risk allele of *HTRA1* rs11200638 was not significantly different in typical AMD and PCV. We genotyped rs10490924, but a complete LD led us to analyse the clinical association only with rs11200638 (data not shown). In a previous study on Japanese patients, it was concluded that rs11200638 made a greater contribution to

typical AMD than to PCV.³⁰ However, we recalculated the reported genotype and allele data using the same method as the original article and found that no significant difference existed between typical AMD and PCV in the two *HTRA1* polymorphisms.³⁰

For *CFH* Y402H and *HTRA1* rs11200638, it may well be that patients with typical AMD and PCV have the same genetic background. It has been reported that PCV can co-exist with typical AMD.^{3,43}

We examined the potential contribution of genetic factors to clinical presentation. However, because the number of participants was too small to establish a high-enough power to determine the statistical significance of the findings, only positive associations could be considered. We obtained a statistically significant correlation between the frequency of the polymorphisms and the lesion size, particularly for the GLD and the *HTRA1* rs11200638 polymorphisms. To measure the size of the CNV lesion, we chose the GLD as this was clearly defined in the treatment of age-related macular degeneration with photodynamic therapy (TAP) study,³³ and therefore the measurements for typical AMD and PCV was less biased, especially in Asians who tend to have haemorrhagic or fibrovascular pigment epithelial detachments.

There were two limitations of this study. The border of the lesion tended to be not well demarcated in some Japanese patients, and we did not measure the lesion size in these patients. Additionally, the number of subjects was limited because this was a retrospective and cross-sectional examination, and a patient's consent to participate was required for the genomic analyses.

In summary, our results indicate a possible common genetic background of patients with typical AMD and PCV. We also noted a potential association between the disease-risk allele *HTRA1* rs11200638 polymorphism and larger lesion size, which was observed in both typical AMD and PCV patients.

ACKNOWLEDGEMENTS

This study was supported in part by the Ministry of Education, Culture, Sports, Science and Technology of Japan and the Japan National Society for the Prevention of Blindness.

REFERENCES

- Attebo K, Mitchell P, Smith W. Visual acuity and the causes of visual loss in Australia. The Blue Mountains Eye study. *Ophthalmology* 1996; **103**: 357-64.
- Bird AC. The Bowman lecture. Towards an understanding of age-related macular disease. *Eye* 2003; **17**: 457-66.
- Maruko I, Iida T, Saito M, Nagayama D, Saito K. Clinical characteristics of exudative age-related macular degeneration in Japanese patients. *Am J Ophthalmol* 2007; **144**: 15-22.
- Sho K, Takahashi K, Yamada H *et al.* Polypoidal choroidal vasculopathy: incidence, demographic features, and clinical characteristics. *Arch Ophthalmol* 2003; **121**: 1392-6.

5. Oshima Y, Ishibashi T, Murata T, Tahara Y, Kiyohara Y, Kubota T. Prevalence of age related maculopathy in a representative Japanese population: the Hisayama study. *Br J Ophthalmol* 2001; **85**: 1153-7.
6. Yannuzzi LA, Ciardella A, Spaide RF, Rabb M, Freund KB, Orlock DA. The expanding clinical spectrum of idiopathic polypoidal choroidal vasculopathy. *Arch Ophthalmol* 1997; **115**: 478-85.
7. Iranmanesh R, Eandi CM, Peiretti E *et al*. The nature and frequency of neovascular age-related macular degeneration. *Eur J Ophthalmol* 2007; **17**: 75-83.
8. Edwards AO, Ritter R 3rd, Abel KJ, Manning A, Panhuysen C, Farrer LA. Complement factor H polymorphism and age-related macular degeneration. *Science* 2005; **308**: 421-4.
9. Hageman GS, Anderson DH, Johnson LV *et al*. A common haplotype in the complement regulatory gene factor H (HF1/CFH) predisposes individuals to age-related macular degeneration. *Proc Natl Acad Sci USA* 2005; **102**: 7227-32.
10. Haines JL, Hauser MA, Schmidt S *et al*. Complement factor H variant increases the risk of age-related macular degeneration. *Science* 2005; **308**: 419-21.
11. Klein RJ, Zeiss C, Chew EY *et al*. Complement factor H polymorphism in age-related macular degeneration. *Science* 2005; **308**: 385-9.
12. Li M, Atmaca-Sonmez P, Othman M *et al*. CFH haplotypes without the Y402H coding variant show strong association with susceptibility to age-related macular degeneration. *Nat Genet* 2006; **38**: 1049-54.
13. Johnson PT, Betts KE, Radeke MJ, Hageman GS, Anderson DH, Johnson LV. Individuals homozygous for the age-related macular degeneration risk-conferring variant of complement factor H have elevated levels of CRP in the choroid. *Proc Natl Acad Sci USA* 2006; **103**: 17456-61.
14. Lommatzsch A, Hermans P, Weber B, Pauleikhoff D. Complement factor H variant Y402H and basal laminar deposits in exudative age-related macular degeneration. *Graefes Arch Clin Exp Ophthalmol* 2007; **245**: 1713-16.
15. Dewan A, Liu M, Hartman S *et al*. HTRA1 promoter polymorphism in wet age-related macular degeneration. *Science* 2006; **314**: 989-92.
16. Jakobsdottir J, Conley YP, Weeks DE, Mah TS, Ferrell RE, Gorin MB. Susceptibility genes for age-related maculopathy on chromosome 10q26. *Am J Hum Genet* 2005; **77**: 389-407.
17. Rivera A, Fisher SA, Fritsche LG *et al*. Hypothetical LOC387715 is a second major susceptibility gene for age-related macular degeneration, contributing independently of complement factor H to disease risk. *Hum Mol Genet* 2005; **14**: 3227-36.
18. Kanda A, Chen W, Othman M *et al*. A variant of mitochondrial protein LOC387715/ARMS2, not HTRA1, is strongly associated with age-related macular degeneration. *Proc Natl Acad Sci USA* 2007; **104**: 16227-32.
19. Gotoh N, Yamada R, Hiratani H *et al*. No association between complement factor H gene polymorphism and exudative age-related macular degeneration in Japanese. *Hum Genet* 2006; **120**: 139-43.
20. Mori K, Gehlbach PL, Kabasawa S *et al*. Coding and noncoding variants in the CFH gene and cigarette smoking influence the risk of age-related macular degeneration in a Japanese Population. *Invest Ophthalmol Vis Sci* 2007; **48**: 5315-19.
21. Okamoto H, Umeda S, Obazawa M *et al*. Complement factor H polymorphisms in Japanese population with age-related macular degeneration. *Mol Vis* 2006; **12**: 156-8.
22. Uka J, Tamura H, Kobayashi T *et al*. No association of complement factor H gene polymorphism and age-related macular degeneration in the Japanese population. *Retina* 2006; **26**: 985-7.
23. Lau LJ, Chen SJ, Cheng CY *et al*. Association of the Y402H polymorphism in complement factor H gene and neovascular age-related macular degeneration in Chinese patients. *Invest Ophthalmol Vis Sci* 2006; **47**: 3242-6.
24. Chen LJ, Liu DT, Tam PO *et al*. Association of complement factor H polymorphisms with exudative age-related macular degeneration. *Mol Vis* 2006; **12**: 1536-42.
25. Grassi MA, Fingert JH, Scheetz TE *et al*. Ethnic variation in AMD-associated complement factor H polymorphism p.Tyr402His. *Hum Mutat* 2006; **27**: 921-5.
26. Lu F, Hu J, Zhao P *et al*. HTRA1 variant increases risk to neovascular age-related macular degeneration in Chinese population. *Vision Res* 2007; **47**: 3120-3.
27. Mori K, Horie-Inoue K, Kohda M *et al*. Association of the HTRA1 gene variant with age-related macular degeneration in the Japanese population. *J Hum Genet* 2007; **52**: 636-41.
28. Tanimoto S, Tamura H, Ue T *et al*. A polymorphism of LOC387715 gene is associated with age-related macular degeneration in the Japanese population. *Neurosci Lett* 2007; **414**: 71-4.
29. Yoshida T, DeWan A, Zhang H *et al*. HTRA1 promoter polymorphism predisposes Japanese to age-related macular degeneration. *Mol Vis* 2007; **13**: 545-8.
30. Kondo N, Honda S, Ishibashi K, Tsukahara Y, Negi A. LOC387715/HTRA1 variants in polypoidal choroidal vasculopathy and age-related macular degeneration in a Japanese population. *Am J Ophthalmol* 2007; **144**: 608-12.
31. Goverdhan SV, Hannan S, Newsom RB, Luff AJ, Griffiths H, Lotery AJ. An analysis of the CFH Y402H genotype in AMD patients and controls from the UK, and response to PDT treatment. *Eye* 2007; **22**: 849-54.
32. Chen H, Yang Z, Gibbs D *et al*. Association of HTRA1 polymorphism and bilaterality in advanced age-related macular degeneration. *Vision Res* 2008; **48**: 690-4.
33. Bressler NM. Photodynamic therapy of subfoveal choroidal neovascularization in age-related macular degeneration with verteporfin: two-year results of 2 randomized clinical trials-report 2. *Arch Ophthalmol* 2001; **119**: 198-207.
34. Schmidt-Erfurth U, Miller JW, Sickenberg M *et al*. Photodynamic therapy with verteporfin for choroidal neovascularization caused by age-related macular degeneration: results of retreatments in a phase 1 and 2 study. *Arch Ophthalmol* 1999; **117**: 1177-87.
35. Takahashi M, Matsuda F, Margetic N, Lathrop M. Automated identification of single nucleotide polymorphisms from sequencing data. *J Bioinform Comput Biol* 2003; **1**: 253-65.
36. Yannuzzi LA. Idiopathic polypoidal choroidal vasculopathy. Presented at the Macula Society Meeting, February 5, 1982, Miami, FL.
37. Kuroiwa S, Tateiwa H, Hisatomi T, Ishibashi T, Yoshimura N. Pathological features of surgically excised polypoidal choroidal

- vasculopathy membranes. *Clin Experiment Ophthalmol* 2004; **32**: 297–302.
38. Lafaut BA, Aisenbrey S, Van den Broecke C, Bartz-Schmidt KU, Heimann K. Polypoidal choroidal vasculopathy pattern in age-related macular degeneration: a clinicopathologic correlation. *Retina* 2000; **20**: 650–4.
 39. MacCumber MW, Dastgheib K, Bressler NM *et al.* Clinicopathologic correlation of the multiple recurrent serosanguineous retinal pigment epithelial detachments syndrome. *Retina* 1994; **14**: 143–52.
 40. Okubo A, Sameshima M, Uemura A, Kanda S, Ohba N. Clinicopathological correlation of polypoidal choroidal vasculopathy revealed by ultrastructural study. *Br J Ophthalmol* 2002; **86**: 1093–8.
 41. Rosa RH Jr, Davis JL, Eifrig CW. Clinicopathologic reports, case reports, and small case series: clinicopathologic correlation of idiopathic polypoidal choroidal vasculopathy. *Arch Ophthalmol* 2002; **120**: 502–8.
 42. Terasaki H, Miyake Y, Suzuki T, Nakamura M, Nagasaka T. Polypoidal choroidal vasculopathy treated with macular translocation: clinical pathological correlation. *Br J Ophthalmol* 2002; **86**: 321–7.
 43. Chen Y, Wen F, Sun Z, Wu D. Polypoidal choroidal vasculopathy coexisting with exudative age-related macular degeneration. *Int Ophthalmol* 2008; **28**: 119–23.

Lung cancer susceptibility locus at 5p15.33

James D McKay^{1,25}, Rayjean J Hung^{1,2,25}, Valerie Gaborieau^{1,25}, Paolo Boffetta¹, Amelie Chabrier¹, Graham Byrnes¹, David Zaridze³, Anush Mukeria³, Neonilia Szeszenia-Dabrowska⁴, Jolanta Lissowska⁵, Peter Rudnai⁶, Eleonora Fabianova⁷, Dana Mates⁸, Vladimir Bencko⁹, Lenka Foretova¹⁰, Vladimir Janout¹¹, John McLaughlin^{2,12}, Frances Shepherd¹³, Alexandre Montpetit¹⁴, Steven Narod¹⁵, Hans E Krokan¹⁶, Frank Skorpen¹⁶, Maiken Bratt Elvestad¹⁶, Lars Vatten¹⁶, Inger Njolstad¹⁷, Tomas Axelsson¹⁸, Chu Chen¹⁹, Gary Goodman¹⁹, Matt Barnett¹⁹, Melissa M Loomis¹⁹, Jan Lubiński²⁰, Joanna Matyjasik²⁰, Marcin Lener²⁰, Dorota Oszutowaska²⁰, John Field²¹, Triantafillos Liloglou²¹, George Xinarianos²¹, Adrian Cassidy²¹, EPIC Study²⁴, Diana Zelenika²², Anne Boland²², Marc Delepine²², Mario Foglio²², Doris Lechner²², Fumihiko Matsuda²², Helene Blanche²³, Ivo Gut²², Simon Heath²², Mark Lathrop^{22,23} & Paul Brennan¹

We carried out a genome-wide association study of lung cancer (3,259 cases and 4,159 controls), followed by replication in 2,899 cases and 5,573 controls. Two uncorrelated disease markers at 5p15.33, rs402710 and rs2736100 were detected by the genome-wide data ($P = 2 \times 10^{-7}$ and $P = 4 \times 10^{-6}$) and replicated by the independent study series ($P = 7 \times 10^{-5}$ and $P = 0.016$). The susceptibility region contains two genes, *TERT* and *CLPTM1L*, suggesting that one or both may have a role in lung cancer etiology.

We and others have recently reported a susceptibility locus for lung cancer in gene region 15q25, an area that includes a cluster of nicotinic acetylcholine receptor genes¹⁻³. In order to identify further susceptibility gene loci, we genotyped an additional 1,291 cases and 1,561 controls from three further studies (Toronto case-control study, HUNT2/Tromsø cohort study and CARET cohort study) for a total of 3,259 cases of lung cancer and 4,159 controls with genome-wide

data (Table 1 and Supplementary Methods online). After exclusion of subjects because of genotyping quality or evidence of non-European ancestry (Supplementary Methods and Supplementary Fig. 1 online), we analyzed under a log-additive model 315,194 SNPs for 2,971 lung cancer cases and 3,746 controls, adjusting for age, sex and country (Supplementary Fig. 2 online). Using principal-component analysis (Supplementary Methods) to adjust for population stratification, we found only minor differences in the estimates of risk and significance (Supplementary Table 1 online).

Eight SNPs exceeded the genome-wide significance level of 5×10^{-7} (Supplementary Fig. 2b and Supplementary Table 1). Seven of these are located at 15q25.1, the locus previously reported as being associated with lung cancer¹⁻³, with the most prominent association with rs1051730 ($P = 1 \times 10^{-15}$). The eighth SNP, rs402710, is located at 5p15.33 ($P = 2 \times 10^{-7}$), indicating a potentially new susceptibility locus for lung cancer. Three additional SNPs in the 5p15.33 region showed evidence of association $P < 5 \times 10^{-6}$ (Supplementary Table 1). Two of these, rs31489 and rs401681, were in strong linkage disequilibrium (LD) with rs402710 ($r^2 > 0.680$) in the 3,746 controls genotyped on the Illumina platform. In contrast, rs2736100 showed relatively little LD with rs402710 ($r^2 = 0.026$) (Supplementary Fig. 3 online).

We subsequently genotyped rs402710 and rs2736100 using Taqman in an additional 2,899 lung cancer cases and 5,573 controls from four separate studies (Table 1 and Supplementary Methods). These included the EPIC cohort study, the Liverpool case-control study, the Szczecin lung cancer study and, uniquely for rs402710 because of limited DNA availability, additional cases and controls from the CARET cohort study. This independent sample provided evidence for replication of the initial finding for both variants ($P = 7 \times 10^{-5}$ for rs402710 and $P = 0.016$ for rs2736100). A combined association using all 5,870 cases and 9,319 controls with correction for the 315,194 comparisons in the genome-wide analysis yielded P values of 4×10^{-6} for rs402710 and 0.02 for rs2736100. The estimated allelic odds ratio (OR) in the replication series was more modest than that of the initial GWA series, subject to the 'winner's curse'. The more conservative OR in replication series is the preferred estimate.

More detailed information on the association between lung cancer and the SNPs rs402710 and rs2736100 is presented in Figure 1. The

¹International Agency for Research on Cancer (IARC), Lyon 69008, France. ²Samuel Lunenfeld Research Institute, Toronto M5T 3L9, Canada. ³Institute of Carcinogenesis, Cancer Research Centre, Moscow 115478, Russia. ⁴Department of Epidemiology, Institute of Occupational Medicine, Lodz 90950, Poland. ⁵The M. Skłodowska-Curie Memorial Cancer Center and Institute of Oncology, Warsaw 02781, Poland. ⁶National Institute of Environmental Health, Budapest 1097, Hungary. ⁷Specialized Institute of Hygiene and Epidemiology, Banska Bystrica 97556, Slovakia. ⁸Institute of Public Health, Bucharest 050463, Romania. ⁹Charles University in Prague, First Faculty of Medicine, Institute of Hygiene and Epidemiology, Prague 2 12800, Czech Republic. ¹⁰Department of Cancer Epidemiology and Genetics, Masaryk Memorial Cancer Institute, Brno 65653, Czech Republic. ¹¹Palacky University, Olomouc 77515, Czech Republic. ¹²Cancer Care Ontario, Toronto M5G 2L7, Canada. ¹³Princess Margaret Hospital, Ontario Cancer Institute, Toronto M5G 2M9, Canada. ¹⁴McGill University and Genome Quebec Innovation Centre, Montreal H3A 1A4, Canada. ¹⁵Women's College Research Institute, Toronto M5G 1N8, Canada. ¹⁶Norwegian University of Science and Technology, Trondheim 7489, Norway. ¹⁷Institute of Community Medicine, University of Tromsø, Tromsø 9037, Norway. ¹⁸Uppsala University, Department of Medical Sciences, SNP Technology Platform, Academic Hospital, Uppsala 751 85, Sweden. ¹⁹Fred Hutchinson Cancer Research Center, Seattle, Washington 98109, USA. ²⁰Pomeranian Medical University, Department of Genetics and Pathology, International Hereditary Cancer Center, Szczecin 70 115, Poland. ²¹Roy Castle Lung Cancer Research Programme, University of Liverpool Cancer Research Centre, Liverpool L3 9TA, UK. ²²Commissariat à l'Énergie Atomique, Institut Genomique, Centre National de Genotypage, Evry 91000, France. ²³Fondation Jean Dausset-CEPH, Paris 75010, France. ²⁴A full list of authors appears at the end of this paper. ²⁵These authors contributed equally to this work. Correspondence should be addressed to P.B. (brennan@iarc.fr).

Received 21 May; accepted 10 September; published online 2 November 2008; doi:10.1038/ng.254

Table 1 Description of the seven studies contributing to the genome-wide and replication analysis

Study	No. of subjects on ILLUMINA		No. of subjects passing QC		Location	Study design
	Cases	Controls	Cases	Controls		
Genome-wide association studies						
Central Europe	1,968	2,598	1,841	2,441	Romania, Hungary, Poland, Russia, Slovakia, Czech Republic	Case control
Toronto	438	709	330	500	Greater Toronto area (Canada)	Case control
HUNT2/Tromsø	433	433	403	412	North Trøndelag County (Norway)	Cohort
CARET	420	419	397	393	and Tromsø city in Tromsø County (Norway)	Cohort
Total	3,259	4,159	2,971	3,746	United States	Cohort
Replication studies						
EPIC	-	-	1,213	2,591	Sweden, Netherlands, UK, France, Germany, Spain, Italy, Norway, Denmark, Greece	Cohort
Szczecin	-	-	908	1,037	Poland	Case control
CARET2	-	-	363	1,128	United States	Cohort
Liverpool	-	-	415	817	UK	Case control
Total	-	-	2,899	5,573		
Total overall			5,870	9,319		

For quality control (Supplementary Methods), we excluded samples with call rate <95%, sex discrepancy or non-European ancestry. We also excluded non-expected duplicates and first-degree relatives from the final analysis.

risk-associated allele was the more common allele of rs402710 and the less common allele of rs2736100. The association with rs402710 was prominent in never-smokers ($P = 0.01$), ex-smokers ($P = 0.0007$) and current smokers ($P = 0.0001$), and there was no evidence of any heterogeneity by study, histology, age or sex. There was no apparent geographical heterogeneity in the allele frequencies of rs402710. Adjustment for smoking exposure (pack years) had no effect on the observed association with a smoking-adjusted OR per allele of 1.19 (1.12–1.26). We also investigated rs402710 in the context of smoking intensity among controls and did not observe any association between number of cigarettes consumed per day and rs402710 ($P = 0.74$). The effects observed with rs2736100 were similar, with the associations for the less common (risk) allele being largely comparable to those for rs402710.

Several lines of evidence suggest that the associations observed with rs402710 and rs2736100 are independent. We found little LD between rs402710 and rs2736100 using all available controls. After incorporation of either one of these SNPs into the logistic regression, the association with the other remained significant, and there was no change in the risk estimate (OR per allele for rs402710 = 1.17 ($P = 2 \times 10^{-8}$) with adjustment for rs2736100 and OR per allele for rs2736100 = 1.11 ($P = 0.0004$) with adjustment for rs402710). Second, when cases and controls were compared for the number of risk alleles for rs402710 and rs2736100, there was an increasing trend with increasing number of risk alleles ($P = 2 \times 10^{-13}$) reaching an OR of 1.65 (1.34–2.02) for those who were homozygous for both risk variants (Supplementary Table 2 online). Finally, when we imputed genotypes

(Supplementary Methods) at 5p15.33 in the 2,971 cases of lung cancer and 3,746 controls with genome-wide data, we did not identify any SNPs more strongly associated with risk than rs402710 (Supplementary Table 3 online). The top 11 imputed SNPs ($P < 0.0001$) were genotyped subsequently in the cases and controls of central European ancestry (Supplementary Methods) and comparison of haplotype frequencies from this direct genotyping indicated that the prevalence of two distinct haplotypes differed between cases and controls (Supplementary Table 4 online). One haplotype carried the minor allele of rs402710 and eight additional SNPs in high LD ($r^2 > 0.644$) with rs402710, and the second haplotype tagged the minor allele of both rs2736100 and a second SNP rs2736098. Nevertheless, the possibility remains that rs402710 and rs2736100, although only weakly associated with each other, are in LD with one or more causal variants in this region.

The 5p15.33 locus contains two known genes: the *TERT* (human telomerase reverse transcriptase) gene and the *CLPTMIL* (alias *CCR9*; cleft lip and palate transmembrane 1 like) gene. There is no clear evidence to suggest that rs2736100 or rs402710 are themselves causative alleles. The rs2736100 variant is located in intron 1 of *TERT*, and rs402710 is located in a region of high LD that includes the proximal and putative promoter regions of *TERT*; as well as the entire coding region of the *CLPTMIL* gene (Supplementary Fig. 3). Current knowledge of the role of these genes would seem to implicate *TERT* as the more plausible candidate. *TERT* is the reverse transcriptase component of telomerase⁴, making it essential for telomerase enzyme production and maintenance of

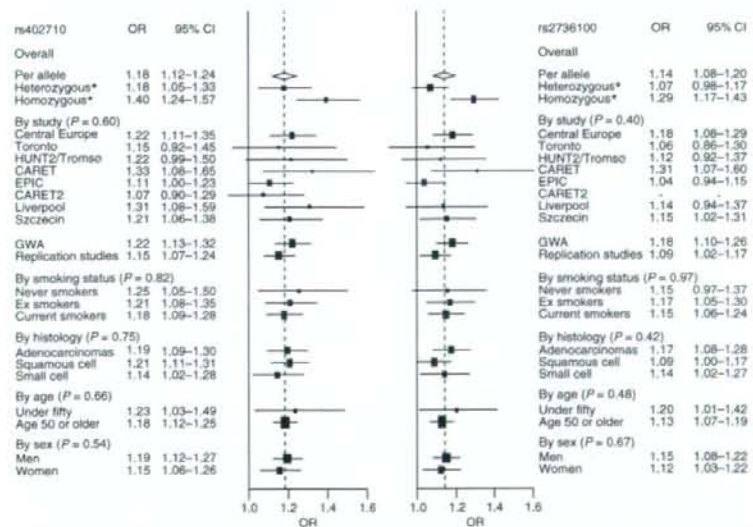


Figure 1 Forest plot representing lung cancer risk and the two variants in the 5p region (rs402710 and rs2736100). Apart from the odds ratios for heterozygous and homozygous effect (*), odds ratios and 95% confidence intervals are derived from the per-allele model. All models are adjusted for age, sex and country. The overall OR is shown by the broken vertical line. P values are from heterogeneity tests.

telomeres⁵. The telomerase enzyme is responsible for telomere regeneration, and up to 90% of human tumor samples, including lung cancer⁶, show telomerase activity, indicating that regeneration of telomeres is a vital step for most forms of carcinogenesis⁷. *TERT* expression is actively present in germ cells, although is found in very low levels for most types of normal cells⁸. Activation of the *TERT* promoter seems to be a key step in synthesis of the *TERT* protein and resulting telomerase activity⁹. Such activity may be measured with the telomeric repeat amplification protocol (TRAP) and has been associated with both lung cancer progression and prognosis^{6,10,11}. Inhibitors of *TERT* are clearly of much interest for potential chemoprevention and treatment of cancer, although their development has so far been unsuccessful⁶. DNA resequencing has shown that there is little common genetic variation in the *TERT* coding region which, along with its high conservation between species, implies that the gene itself is under strong evolutionary restraint¹². Rare mutations in the *TERT* coding sequence have been implicated in dyskeratosis congenita¹³, an autosomal dominant syndrome characterized by bone marrow abnormalities, but also pulmonary fibrosis and increased risk of some cancers¹⁴.

The other gene in this region, *CLPTMIL*, named for its similarity to a gene implicated in susceptibility to cleft lip palate, was identified through screening for cisplatin (CDDP) resistance-related genes and was found to be upregulated in CDDP-resistant ovarian tumor cell lines and to induce apoptosis in CDDP-sensitive cells¹⁵. The *CLPTMIL* gene is well conserved and expressed in various tissues, including lung tissue. On the basis of these properties, it could be hypothesized that *CLPTMIL* induces apoptosis of lung cells under genotoxic exposures such as tobacco carcinogen-related stress.

In summary, we have identified a new susceptibility locus for lung cancer that comprises two potential candidate genes: *TERT*, an essential component of telomerase production and of carcinogenesis, and *CLPTMIL*, which may induce apoptosis. The nature of the causative alleles remains unclear. Further studies to identify the causal genetic variants and elucidate their function will aid our understanding of the etiology of lung cancer.

Note: Supplementary information is available on the Nature Genetics website.

The full list of authors and affiliations is as follows:

Paolo Vineis^{26,27}, Françoise Clavel-Chapelon²⁸, Domenico Palli²⁹, Rosario Tumino³⁰, Vittorio Krogh³¹, Salvatore Panico³², Carlos A González³³, José Ramón Quirós³⁴, Carmen Martínez³⁵, Carmen Navarro^{36,37}, Eva Ardanaz³⁸, Nerea Larrañaga³⁹, Kay Tee Kham⁴⁰, Timothy Key⁴¹, H Bas Bueno-de-Mesquita⁴², Petra H M Peeters⁴³, Antonia Trichopoulos⁴⁴, Jakob Linseisen⁴⁵, Heiner Boeing⁴⁶, Göran Hallmans⁴⁷, Kim Overvad⁴⁸, Anne Tjønneland⁴⁹, Merethe Kumle⁵⁰ & Elio Riboli²⁷

²⁶Servizio di Epidemiologia dei Tumori, Università di Torino and CPO-Piemonte, Turin 10126, Italy. ²⁷Department of Epidemiology and Public Health, Imperial College, London SW7, UK. ²⁸INSERM, E3N-EPIC Group Institut Gustave Roussy, Villejuif 94805, France. ²⁹Molecular and Nutritional Epidemiology Unit, Cancer Research and Prevention Institute (ISPO), Florence 50139, Italy. ³⁰Cancer Registry and Histopathology Unit, Azienda Ospedaliera "Civile M.P. Arezzo", Ragusa 97100, Italy. ³¹Istituto Nazionale dei Tumori, Milan 20133, Italy. ³²Dipartimento di Medicina Clinica e Sperimentale, Università di Napoli, Federico II, Naples 80131, Italy. ³³Servicio de Epidemiología y registro del Cáncer, Instituto Catalán de Oncología, Barcelona 08907, Spain. ³⁴Jefe Sección Información Sanitaria, Consejería de Servicios Sociales, Principado de Asturias, Oviedo 33001, Spain. ³⁵Escuela Andaluza de Salud Pública, Granada 18011, Spain. ³⁶Epidemiology Department, Murcia Health Council, Murcia 18011, Spain. ³⁷CIBER Epidemiología y Salud Pública (CIBERESP), Barcelona 8003, Spain. ³⁸Registro de Cáncer de Navarra, Instituto de Salud Pública, Gobierno de Navarra, Pamplona 31003, Spain. ³⁹Subdirección de Salud Pública de Gipuzkoa, Gobierno Vasco, San Sebastian 20113, Spain. ⁴⁰MRC Dunn Human Nutrition Unit, Cambridge CB2 0XY, UK. ⁴¹Cancer Research UK, University of Oxford, Oxford OX3 7XP, UK. ⁴²Centre for Food and Health, National Institute of Public Health and the Environment (RIVM), Bilthoven 3720 BA, The Netherlands. ⁴³Julius Center for Health Sciences and Primary Care, Department of Epidemiology, University of Utrecht, Utrecht 3508 GA, The Netherlands. ⁴⁴Department of Hygiene and Epidemiology, University of Athens, Athens 11527, Greece. ⁴⁵Division of Clinical Epidemiology, German Cancer Research Centre, Heidelberg 69120, Germany. ⁴⁶Department of Epidemiology, Deutsches Institut für Ernährungsforschung, Potsdam-Rehbrücke 14558, Germany. ⁴⁷Department of Public Health and Clinical Medicine, University of Umeå, Umeå 90187, Sweden. ⁴⁸Department of Epidemiology and Social Medicine, Aarhus University, Aarhus 8000, Denmark. ⁴⁹The Danish Cancer Society, Institute of Cancer Epidemiology, Copenhagen 2100, Denmark. ⁵⁰Institute of Community Medicine, University of Tromsø, Tromsø 9037, Norway.

ACKNOWLEDGMENTS

P. Brennan and M. Lathrop designed the study. J.D.M., R.J.H., V.G., M.B.E., A.B. and H. Blanche coordinated the preparation and inclusion of all biological samples. J.D.M., S.H. and V.G. undertook the statistical analysis. Bioinformatics analysis was undertaken by E.M., M.F. and S.H. D.Z., D.L. and I.G. coordinated the genotyping of the central Europe samples. A.M. and R.I.H. coordinated the genotyping of the Toronto samples. J.D.M., D.Z., M.D., A.C., T.A. and H.E.K. coordinated the genotyping of the other studies. All other coauthors coordinated the initial recruitment and management of the studies. M. Lathrop obtained financial support for genotyping of the central Europe study; P. Brennan, R.I.H. and H.E.K. obtained financial support for genotyping of the other studies. P. Brennan and J.D.M. drafted the manuscript with substantial contributions from R.I.H. and M. Lathrop. All authors contributed to the final paper. The authors thank all of the participants who took part in this research and the funders and support and technical staff who made this study possible. Support for the central Europe, HUNT2/Tromsø and CARET genome-wide studies and follow-up genotyping was provided by Institut National du Cancer, France. Support for the HUNT2/Tromsø genome-wide study was also provided by the European Community (Integrated Project DNA repair, grant no. LSHG-CT-2005-512113), the Norwegian Cancer Association and the Functional Genomics Programme of Research Council of Norway. Funding for the Toronto genome-wide study was provided by the Ontario Institute of Cancer Research. Funding for the Szczecin/Poland replication study was provided by European Community program "Marie-Curie Host Fellowships for the Transfer of Knowledge," grant no. MTKD-CT-2004-510114. Additional funding for study coordination, genotyping of replication studies and statistical analysis was provided by the US National Cancer Institute (R01 CA092039).

Published online at <http://www.nature.com/naturegenetics/>
Reprints and permissions information is available online at <http://npg.nature.com/reprintsandpermissions/>

- Hung, R.J. *et al.* *Nature* **452**, 633–637 (2008).
- Amos, C.I. *et al.* *Nat. Genet.* **40**, 616–622 (2008).
- Thorgerirsson, T. *et al.* *Nature* **452**, 638–642 (2008).
- Weinrich, S.L. *et al.* *Nat. Genet.* **17**, 498–502 (1997).
- Greider, C.W. *et al.* *Cell* **43**, 405–413 (1985).
- Lantuejoul, S. *et al.* *Int. J. Cancer* **120**, 1835–1841 (2007).
- Hanahan, D. & Weinberg, R.A. *Cell* **100**, 57–70 (2000).
- Greider, C.W. *et al.* *Nature* **337**, 331–337 (1989).
- Janknecht, R. *FEBS Lett.* **564**, 9–13 (2004).
- Mavrogiannou, E. *et al.* *Clin. Chem.* **53**, 53–61 (2007).
- Wu, T.C. *et al.* *Lung Cancer* **41**, 163–169 (2003).
- Savage, S.A. *et al.* *Hum. Mutat.* **26**, 343–350 (2005).
- Armanios, M. *et al.* *Proc. Natl. Acad. Sci. USA* **102**, 15960–15964 (2005).
- Garcia, C.K. *et al.* *Nucleic Acids Res.* **35**, 7406–7416 (2007).
- Yamamoto, K. *et al.* *Biochem. Biophys. Res. Commun.* **280**, 1148–1154 (2001).

NOTE

Subclinical Hypercortisolism in Hospitalized Patients with Type 2 Diabetes Mellitus

TAKAO TANIGUCHI, AKIHIRO HAMASAKI* AND MOTOZUMI OKAMOTO

Department of Internal Medicine, Ohtsu Red Cross Hospital, Ohtsu, Japan

*Department of Diabetes and Clinical Nutrition, Kyoto University Faculty of Medicine, Kyoto, Japan

Abstract. Pre(sub)clinical Cushing's disease is a recently described entity defined by the autonomous secretion of ACTH and the absence of a cushingoid appearance. We screened 77 hospitalized patients with diabetes mellitus for subclinical hypercortisolism and detected pre(sub)clinical Cushing's disease in 2 (2.6%) of them. In both patients, transsphenoidal surgery was performed and a microadenoma was removed. Their metabolic clearance rate of glucose measured by a glucose clamp study, an index of insulin sensitivity, significantly improved after surgery. Our results indicate that screening for subclinical hypercortisolism in diabetic patients might be useful, as surgery improves glucose tolerance and insulin sensitivity.

Key words: Subclinical hypercortisolism, Pre(sub)clinical Cushing's disease, Diabetes mellitus, Insulin sensitivity, Glucose clamp study

(Endocrine Journal 55: 429–432, 2008)

THE prevalence of subclinical hypercortisolism in patients with diabetes mellitus in Western countries is higher than previously believed [1–3], but it is not known whether this is the case in Japan. Screening and detection of subclinical hypercortisolism in diabetic patients might be useful because surgery may improve glucose tolerance.

Pre(sub)clinical Cushing's disease is a recently described entity defined by the autonomous secretion of ACTH and the absence of a cushingoid appearance [4], and the diagnostic criteria were recently published in Japan. We screened hospitalized patients with diabetes mellitus for subclinical hypercortisolism. In patients in whom pre(sub)clinical Cushing's disease was detected, transsphenoidal surgery was performed. Insulin sensitivity was assessed with the glucose clamp technique before and after surgery.

Methods

The subjects were recruited from 135 consecutive patients admitted to Ohtsu Red Cross Hospital for control of diabetes mellitus from September 2005 to September 2006. Exclusion criteria were cushingoid appearance, type 1 diabetes, diabetes secondary to other causes, alcoholism, renal failure, depression, and acute illness. A total of 77 patients (47 males and 30 females, age 61 ± 13 years, BMI 25.0 ± 4.2 , HbA1c $10.4 \pm 2.1\%$) were studied. None of the patients displayed moon face, buffalo hump, central obesity, striae cutis, skin atrophy, ecchymosis, or weakness associated with proximal muscle wasting. The study protocol was approved by the hospital ethics committee, and informed consent was obtained from all patients.

After at least 36 h after admission, the serum cortisol concentration was measured between 2300 and 2400 h (midnight cortisol). Although the cut-off value of midnight cortisol for screening is $2.5 \mu\text{g/dl}$ according to the diagnostic criteria for pre(sub)clinical Cushing's disease (Guidelines of Diagnosis and Treatment of Pre(sub)clinical Cushing's Disease by the Research Committee of the Ministry of Health, Labor, and Wel-

Received: September 6, 2007

Accepted: October 20, 2007

Correspondence to: Takao TANIGUCHI, M.D., Ph.D., Department of Internal Medicine, Ohtsu Red Cross Hospital, 1-1-35, Nagara, Ohtsu, Shiga, 520-8511, Japan

fare of Japan, 2006), we set the cut-off value as 5 µg/dl because the diurnal rhythm of cortisol is insufficient in diabetic patients [5]. Patients with a midnight cortisol level of more than 5.0 µg/dl underwent a 0.5 mg overnight dexamethasone suppression test (DST). Patients in whom the plasma cortisol levels were greater than 3 µg/dl after the 0.5 mg overnight DST were regarded as positive according to the diagnostic criteria, and basal ACTH and cortisol levels were measured. Patients with ACTH levels above 10 pg/ml underwent dynamic magnetic resonance imaging (MRI) of the pituitary. Abdominal computed tomography would have been performed in patients with ACTH levels of less than 10 pg/ml, however none of the patients had such low levels.

After improvement of glycemic control by medical treatment and stabilization for 4 to 6 months, transsphenoidal surgery was performed and a microadenoma was identified and removed in patients diagnosed with pre(sub)clinical Cushing's disease.

A euglycemic hyperinsulinemic glucose clamp study was performed 1 week before and 4 to 6 weeks after surgery as described earlier [6] with slight modification [7]. Serum C-peptide values were measured 6 min after intravenous administration of 1 mg glucagon before and after surgery.

Results

Of the 77 patients, 27 patients had a midnight cortisol level of more than 5.0 µg/dl. Among these 27, there were 7 in whom plasma cortisol levels were greater than 3 µg/dl after the 0.5 mg overnight DST. One patient declined further study. Basal ACTH and cortisol levels were measured in the other 6 patients, all of whom had ACTH levels above 10 pg/ml. Dynamic MRI of the pituitary revealed cystic lesions in two of the patients, and the findings in the other 4 patients were normal.

The endocrinologic data of the two patients with positive MRI findings are shown in Table 1. Basal ACTH and cortisol levels were normal, and the peak/basal ratio of plasma ACTH after a CRH stimulation test was above 1.5. Serum cortisol was not suppressed by an overnight low-dose (0.5 mg) DST, but was suppressed by an overnight high-dose (8 mg) DST. The central-peripheral ratio of ACTH measured by bilateral simultaneous sampling of the cavernous sinuses after

Table 1. Endocrinologic data of the patients with positive MRI findings

	Patient 1	Patient 2
Age, sex	72, M	47, M
Midnight F (µg/dl)	5.8	7.5
Basal ACTH (pg/ml)	32.6	22.1
Basal F (µg/dl)	17.3	10.6
0.5 mg DST-F (µg/dl)	10.1	5.7
8 mg DST-F (µg/dl)	2.4	1.4
peak/basal ACTH ratio post-CRH	4.1	3.9
c/p ACTH ratio post-CRH	7.3	418

peak/basal ACTH ratio post-CRH, peak/basal ratio of plasma ACTH after a CRH stimulation test; c/p ACTH ratio post-CRH, central-peripheral ratio of ACTH measured by bilateral simultaneous sampling of the cavernous sinuses after stimulation with 100 µg CRH

Table 2. Diabetologic data of the two patients with pre(sub)clinical Cushing's disease.

	Patient 1	Patient 2
BMI		
pre op	26.6	25.4
post op	25.1	23.5
HbA1c (%)		
on 1st admission	9.6	12.9
pre op	7.0	5.2
post op	6.4	4.7
MCR-G (ml/kg/min)		
pre op	2.9	5.1
post op	4.0	7.0
CPR 6 min. after glucagon test (ng/ml)		
pre op	5.4	5.3
post op	5.2	4.8
treatment		
pre op	Insulin 20 U/day	OHA
post op	Insulin 6 U/day → OHA	Diet only

After improvement of glycemic control by medical treatment and stabilization for 4 to 6 months, transsphenoidal surgery was performed. A euglycemic hyperinsulinemic glucose clamp study was performed 1 week before and 4 to 6 weeks after surgery. Hydrocortisone replacement therapy was stopped 2.5 weeks after surgery. Post-op, post-operation; MCR-G, metabolic clearance rate of glucose measured by a euglycemic hyperinsulinemic glucose clamp study; CPR; serum C-peptide

stimulation with 100 µg CRH was above 3.0. Both patients were diagnosed with pre(sub)clinical Cushing's disease based on the diagnostic criteria. Transsphenoidal surgery was performed and a microadenoma was identified and removed in both patients. The pathologic diagnosis based on immunohistochemistry was an

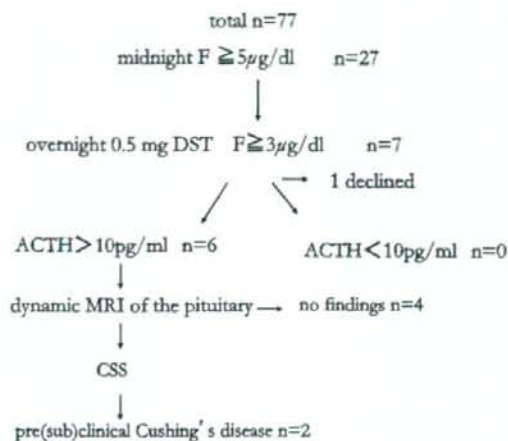


Fig. 1. Study design.

F; serum cortisol, DST; dexamethasone suppression test, MRI; magnetic resonance imaging

ACTH-secreting adenoma.

Patient 2 had transient hypoglycemia postoperatively. In both patients, however, the ACTH and cortisol response after stimulation with CRH 2 weeks after surgery was normal, consistent with preoperative pre(sub)clinical Cushing's disease without severe suppression of the residual pituitary gland, and hydrocortisone replacement therapy was stopped. Serum cortisol was suppressed by an overnight low-dose (0.5 mg) DST.

The diabetologic data of the two patients are shown in Table 2. Neither patient had a family history of diabetes. Glucose tolerance improved after surgery in both patients.

Notably, the metabolic clearance rate of glucose measured by a glucose clamp study, an index of insulin sensitivity, significantly improved after surgery. There was no significant difference between the serum C-peptide values 6 min after intravenous administration of 1 mg of glucagon before and after surgery. Patient 1 required mixed insulin at 20 U/day to control blood glucose levels before surgery; the dose was reduced to 6 U/day after surgery, and the patient is currently under good control on oral hypoglycemic agents. Patient 2 was able to stop taking an oral hypoglycemic agent after surgery.

The blood pressure of Patients 1 and 2 was 140/75 mmHg and 132/74 mmHg preoperatively, and 125/70 and 110/70 mmHg postoperatively, respectively.

Patient 1 was taking antihypertensive drugs, the dosages of which were not changed either before or after surgery. Although bone mineral density was not measured, osteoporosis was observed in the thoracic spine in a lateral view chest X-ray in Patient 2, which improved 8 months after surgery.

Discussion

The prevalence of subclinical hypercortisolism in diabetic patients in Western countries is reported to be 2% to 9.4%, depending on the population studied and the diagnostic criteria [1-3]. In the present study, we detected 2 patients with pre(sub)clinical Cushing's disease (2.6%) out of 77 hospitalized diabetic patients. Our findings suggest that the prevalence of subclinical hypercortisolism in diabetic patients in Japan might be higher than expected, although further studies with larger numbers of patients are required to determine the actual prevalence.

Because glucose tolerance and insulin sensitivity improved after surgery in both patients, screening for subclinical hypercortisolism in diabetic patients might be useful. This is the first report of pre(sub)clinical Cushing's disease with the insulin sensitivity and secretion evaluated by a euglycemic hyperinsulinemic glucose clamp study and glucagon test before and after surgery. It is unlikely that the improved insulin sensitivity was the result of ameliorated glucose toxicity, because a preoperative glucose clamp test was performed after glycemic control had been stabilized for 4 to 6 months.

The mechanism of insulin resistance in pre(sub)clinical Cushing's disease without clinical features of Cushing's disease remains speculative. One possible explanation is that sensitivity to cortisol may differ among tissues. Another possibility is that differences in the activity of 11 β HSD-1, an enzyme that generates active cortisol from inactive cortisone, may cause differences in glucocorticoid actions among various tissues [8].

There are some limitations in this study. The possibility that midnight cortisol was increased due to venipuncture cannot be excluded. In this sense, screening by midnight salivary cortisol is promising [9], although this is currently not clinically available in Japan. Although various approaches have been studied, there is no single screening test that can detect all cases of

subclinical hypercortisolism. Combination of a 0.5-mg overnight DST and midnight cortisol might be desirable [10], although the risk of causing hyperglycemia in uncontrolled diabetic patients is not negligible. We performed DST in patients screened by midnight cortisol after fair glycemic control was achieved.

The detection rate of a pituitary adenoma in Cushing's disease by dynamic MRI is approximately 60%. Indeed, in the present two cases of pre(sub)clinical Cushing's disease, microadenoma was found apart from the cysts detected by preoperative MRI. In 4 cases with negative MRI findings, the plasma cortisol levels after the 0.5-mg overnight DST were between 3

and 5 $\mu\text{g/dl}$, and basal ACTH levels were between 44.5 and 65.5 pg/ml. Thus, ectopic ACTH syndrome or adrenogenic preclinical Cushing's syndrome is unlikely. All four cases had hypertension. Pre(sub)clinical Cushing's disease cannot be excluded, but the patients were reluctant to undergo further studies including sampling of the cavernous sinuses. Therefore, we might have underestimated the prevalence of pre(sub)clinical Cushing's disease in the present study.

In conclusion, screening for subclinical hypercortisolism in diabetic patients might be useful, as glucose tolerance and insulin sensitivity improve after surgery.

References

1. Leibowitz G, Tsur A, Chayen SD, Salameh M, Raz I, Cerasi E, Gross DJ (1996) Pre-clinical Cushing's syndrome: an unexpected frequent cause of poor glycemic control in obese diabetic patients. *Clin Endocrinol* 44: 717-722.
2. Catargi B, Rigalleau V, Poussin A, Ronci-Chaix N, Bex V, Vergnot V, Gin H, Roger H, Tabarin A (2003) Occult Cushing's syndrome in type-2 diabetes. *J Clin Endocrinol Metab* 88: 5808-5813.
3. Chiodini I, Torlontano M, Scillitani A, Arosio M, Bacci S, Di Lembo S, Epaminonda P, Augello G, Enrini R, Ambrosi B, Adda G, Trischitta V (2005) Association of subclinical hypercortisolism with type 2 diabetes mellitus: a case-control study in hospitalized patients. *Eur J Endocrinol* 157: 837-844.
4. Takao T, Mimoto T, Yamamoto M, Hashimoto K (2001) Preclinical Cushing disease. *Arch Intern Med* 161: 892-893.
5. Cameron OG, Thomas B, Hariharan M, Greden JF (1987) Hypercortisolism in diabetes mellitus. *Diabetes Care* 10: 662-664.
6. DeFronzo RA, Tobin JD, Andres R (1979) Glucose clamp technique: a method for quantifying insulin secretion and resistance. *Am J Physiol* 237: E214-E223.
7. Taniguchi T, Okamoto M, Ueno H, Tanaka J (2001) Use of the insulin sensitivity index obtained from oral glucose tolerance test in Japanese subjects. *Diabetes Res Clin Pract* 54:143-54144.
8. Reimond G, Allasino B, Bovio S, Paccotti P, Angeli A, Terzolo M (2005) Evaluation of the effectiveness of midnight serum cortisol in the diagnostic procedures for Cushing's syndrome. *Eur J Endocrinol* 153: 803-809.
9. Yaneva M, Mosnier-Pudar H, Dugue MA, Grabar S, Fulla Y, Bertaga X (2004) Midnight salivary cortisol for the initial diagnosis of Cushing's syndrome for various causes. *J Clin Endocrinol Metab* 89: 3345-3351.
10. Tomlinson JW, Draper N, Mackie J, Johnson AP, Holder G, Wood P, Stewart PM (2002) Absence of Cushingoid phenotype in a patient with Cushing's disease due to defective cortisone to cortisol conversion. *J Clin Endocrinol Metab* 87: 57-62.

Original Article

Effects of Pitavastatin on Lipid Profiles and High-Sensitivity CRP in Japanese Subjects with Hypercholesterolemia: Kansai Investigation of Statin for Hyperlipidemic Intervention in Metabolism and Endocrinology (KISHIMEN) Investigators

Hiroyuki Koshiyama^{1,8}, Ataru Taniguchi², Kiyoshi Tanaka³, Shinji Kagimoto⁴, Yoshio Fujioka⁵, Kenichi Hirata⁶, Yoshio Nakamura⁶, Akane Iwakura⁶, Kyoko Hara⁶, Taizo Yamamoto⁷, Akira Kuroe², Michihiro Ohya², Shimpei Fujimoto², Yoshiyuki Hamamoto^{1,8}, Sachiko Honjo¹, Hiroki Ikeda¹, Koichiro Nabe¹, Kinsuke Tsuda⁹, Nobuya Inagaki⁸, Yutaka Seino^{2,8}, and Noriaki Kume¹⁰

¹Center for Diabetes & Endocrinology, The Tazuke Kofukai Foundation Medical Research Institute Kitano Hospital, Osaka, Japan

²Division of Diabetes and Clinical Nutrition, Kansai Electric Power Hospital, Osaka, Japan

³Department of Nutrition, Kyoto Women's University, Kyoto, Japan

⁴Department of Endocrinology and Metabolism, Kamo Hospital, Kyoto, Japan

⁵Division of Cardiovascular and Respiratory Medicine, Department of Internal Medicine, Kobe University Graduate School of Medicine, Kobe, Japan

⁶Division of Diabetes & Endocrinology, Department of Internal Medicine, Hyogo Prefectural Amagasaki Hospital, Hyogo, Japan

⁷Department of Endocrinology and Diabetes, Kyoto-Katsura Hospital, Kyoto, Japan

⁸Department of Diabetes & Clinical Nutrition, Graduate School of Medicine, Kyoto University, Kyoto, Japan

⁹Graduate School of Human Science, Kyoto University, Kyoto, Japan

¹⁰Department of Cardiovascular Medicine, Graduate School of Medicine, Kyoto University, Kyoto, Japan

Aim: The effect of pitavastatin on high-sensitivity C-reactive protein (hs-CRP) has not been reported, yet, in humans. We, therefore, investigated the effects of pitavastatin on lipid profiles and hs-CRP in Japanese subjects with hypercholesterolemia.

Methods: The subjects were 178 Japanese with hypercholesterolemia, including 103 (58%) with type 2 diabetes. Pitavastatin (1–2 mg/day) was administered for 12 months. Serum low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C), remnant-like particle cholesterol (RLP-C), triglycerides (TG) and hs-CRP levels were measured for 12 months.

Results: Serum LDL-C and RLP-C levels were significantly decreased by 30.3% and 22.8%, respectively. Serum TG levels were decreased by 15.9% in subjects with basal TG levels above 150 mg/dl. Serum HDL-C levels were significantly increased. The administration of pitavastatin reduced serum hs-CRP levels by 34.8%. No serious adverse events were observed, including changes in glycosylated hemoglobin levels of diabetic patients.

Conclusion: These results suggest that pitavastatin significantly improves lipid profiles and reduces proinflammatory responses, without adverse effects, in Japanese subjects with hypercholesterolemia, including those with diabetes mellitus.

J Atheroscler Thromb, 2008; 15:345-350.

Key words; Inflammation, Diabetes, LDL-C, HDL-C

Address for correspondence: Hiroyuki Koshiyama, Center for Diabetes & Endocrinology, The Tazuke Kofukai Foundation Medical Research Institute Kitano Hospital, Osaka, 530-8480, Japan

E-mail: h-koshiyama@kitano-hp.or.jp

Received: March 7, 2008

Accepted for publication: July 22, 2008

Introduction

The 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (statins) are well known to reduce circulating low-density lipoprotein cholesterol (LDL-C) levels by inhibiting *de novo* cholesterol synthesis in the liver and thereby inducing the

expression of hepatic LDL receptors^{1,2}). However, in clinical trials, the overall reductions in cardiovascular events following statins appear to occur much earlier and to a greater extent than expected from the levels of LDL-C lowering alone^{3,4}; therefore, it has been considered that statins have pleiotropic effects independent of the reduction in circulating LDL-C levels^{3,4}, including anti-inflammatory effects⁵⁻⁸.

Numerous studies have suggested that low-grade inflammation has a pivotal role in atherosclerosis^{5,9,10}. Prospective studies have indicated that high-sensitivity C-reactive protein (hs-CRP) is an important risk factor for atherosclerotic cardiovascular disease^{5,9}. Ridker et al. demonstrated that slight elevation in hs-CRP could lead to the evolution of atherosclerosis in humans^{5,9}. We previously demonstrated that hs-CRP is associated with insulin resistance and fibrinogen levels in non-obese Japanese type 2 diabetic patients¹¹. Although few studies have investigated the effect of statins on hs-CRP in humans, some conflicting reports exist. Several studies indicated that statins reduce hs-CRP levels^{5,10,12,13}; however, one study failed to demonstrate the inhibitory effect of a statin on hs-CRP levels in diabetic subjects¹⁴.

Pitavastatin is a synthetic strong statin, whose molecular structure is similar to atorvastatin and rosuvastatin¹⁵. There has been a single report about the effect of pitavastatin on lipid profiles in humans¹⁶. Although several studies demonstrated that pitavastatin, as well as other lipophilic statins, has anti-inflammatory effects *in vitro*^{6-8,17}, the effects of pitavastatin on inflammatory markers, including hs-CRP, have not been reported in humans *in vivo* to the best of our knowledge.

In the present study, therefore, we explored the effects of pitavastatin on lipid profiles as well as hs-CRP levels in Japanese subjects with hypercholesterolemia, including those with diabetes mellitus.

Patients and Methods

This study was a 12-month, multi-center, prospective, open-label study. Japanese patients, who met the following inclusion criteria: serum total-cholesterol (TC) ≥ 220 mg/dL, and triglycerides (TG) < 400 mg/dL, were recruited. A total of 209 Japanese subjects were enrolled, of which 31 were excluded because they did not follow the protocol. As a result, 178 cases were investigated. Pitavastatin in a dose of 1–2 mg/day was administered [1 mg/day in 44 cases (25%) and 2 mg/day in 134 cases (75%)]. Before the administration of pitavastatin, no lipid-lowering medications had been administered in 111 cases (62%), whereas other

lipid-lowering drugs had been prescribed in 67 cases (38%: 27 pravastatin, 18 atorvastatin, 10 simvastatin, 8 fluvastatin, 1 rosuvastatin, 2 bezafibrate and 1 fenofibrate cases), all of which were withdrawn at least one week before the administration of pitavastatin.

Blood samples were obtained at the beginning and 3, 6 and 12 months after the administration of pitavastatin. Serum TC, LDL-C, TG, HDL-C, glucose, glycosylated hemoglobin (A1C), aspartate aminotransferase (AST), alanine aminotransferase (ALT), creatinine, and creatinine phosphokinase (CK) were measured by standard techniques. Remnant-like particle-cholesterol (RLP-C) and hs-CRP were measured by immunoaffinity gel (JIMRO, Japan) and N Latex CRP II (Dade Behring Marburg GmbH, Marburg, Germany), respectively.

Continuous variables are shown as the mean \pm S.E.M. when the distribution was normal. Statistically significant differences among groups were analyzed by paired *t* test. When the distribution was skewed, statistically significant differences among groups were analyzed by Wilcoxon signed-rank test. The JMP Software computer program (Version 5.0 for Windows; SAS Institute Inc., Cary, NC, USA) was used for all statistical analyses. *P* values 0.05 were considered significant.

This study was approved by the local ethics committee, and informed consent was obtained from all participants before the study.

Results

Basal characteristics of the 178 patients are shown in Table 1 [age: 62.0 ± 0.9 years, 83 men (47%) and 95 women (53%)]. The mean body mass index (BMI) and fasting glucose levels of the subjects were 24.5 kg/m² and 126.0 mg/dL, respectively. The participants in this study included 103 cases (58%) of type 2 diabetes, 62 cases (35%) of hypertension, 7 cases (4%) of peripheral arterial disease, 12 cases (7%) of cerebral infarction, and 32 cases (18%) of coronary heart disease. Serum LDL-C levels were significantly decreased by 32.6%, 31.0% and 30.3% after 3, 6 and 12 months, respectively (Fig. 1A). Serum TG levels were significantly decreased by 17.7% and 15.9% after 3 and 12 months, respectively, in subjects whose basal TG levels were more than 150 mg/dL, although serum TG levels were not significantly changed in overall subjects (Fig. 1B). Serum HDL-C levels were significantly increased by 3.1%, 5.9% and 2.6% after 3, 6 and 12 months, respectively (Fig. 1C). In subjects whose basal HDL-C levels were below 40 mg/dL, HDL-C levels were increased by 16.2%, 22.4% and 19.0% after 3, 6

Table 1. Basal characteristics of subjects.

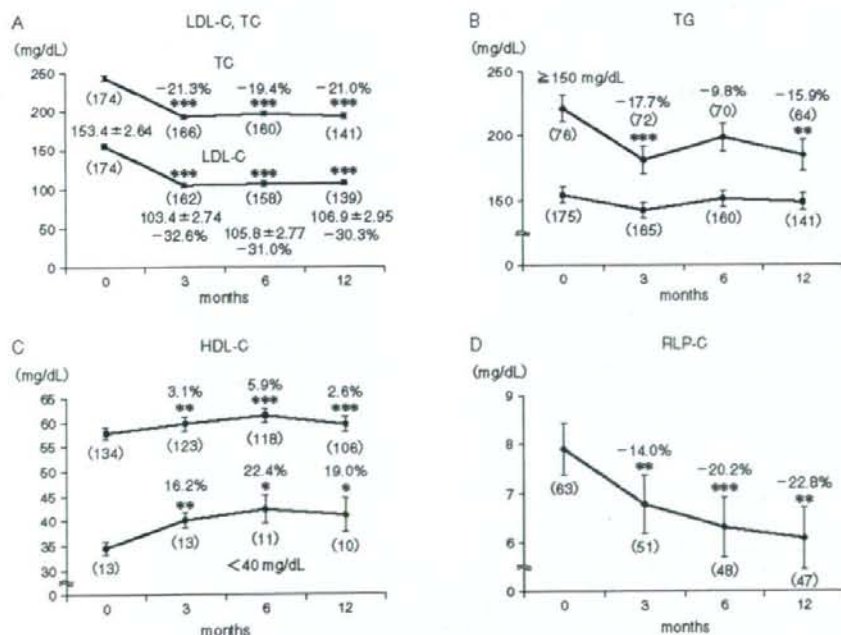
Age	62.0 ± 0.9 (178)
Male %	47% (83)
BMI (kg/m ²)	24.5 ± 0.3 (174)
LDL-C (mg/dL)	153.4 ± 3.3 (174)
HDL-C (mg/dL)	57.8 ± 1.4 (134)
TG (mg/dL)	154.7 ± 5.5 (175)
TC (mg/dL)	242.2 ± 3.6 (174)
RLP-C (mg/dL)	7.9 ± 0.5 (63)
A1C (%)	7.2 ± 0.2 (139)
PPG (mg/dL)	126.0 ± 3.9 (116)
hs-CRP (mg/L)	0.69 (0.33-1.36) (31)
Diabetes mellitus %	58% (103)
Hypertension %	35% (62)
Peripheral arterial disease %	4% (7)
Cerebral infarction %	7% (12)
Coronary heart disease %	18% (32)

Data are presented as the mean ± S.E.M. Numbers of subjects are shown in parentheses. Abbreviations: TC: total cholesterol, FPG: fasting plasma glucose.

and 12 months, respectively (Fig. 1C). Furthermore, serum RLP-C levels were significantly decreased by 14.0%, 20.2% and 22.8% after 3, 6 and 12 months, respectively (Fig. 1D).

Serum hs-CRP levels were significantly decreased in 31 subjects after 12 months (median: 0.69 mg/L to 0.45 mg/L, -34.8%, $p < 0.01$, Fig. 2A). Pitavastatin similarly decreased hs-CRP levels in subjects with diabetes mellitus (median: 0.59 mg/L to 0.36 mg/L, -39.0%, $p < 0.05$, Fig. 2B).

There was no significant correlation between changes in LDL-C and hs-CRP levels even after transformation of hs-CRP values into a logarithm after 12 months ($r = 0.124$, $p = 0.39$). We also found no significant correlation between changes in hs-CRP and changes in serum TG or serum RLP-C levels during the study (data not shown) and there were no serious adverse events. In a subgroup of diabetic patients, there was a slight and statistically insignificant decrease in A1C after pitavastatin treatment for 12 months (7.0% to 6.9%, $n = 80$, $p = 0.07$).

**Fig. 1.** Effect of pitavastatin on lipid levels in all subjects.

A: Effect of pitavastatin on total cholesterol (TC) and LDL cholesterol (LDL-C) levels. B: Effect of pitavastatin on triglyceride (TG) levels in all subjects and in subjects with basal TG levels of more than 150 mg/dL. C: Effect of pitavastatin on HDL cholesterol (HDL-C) levels. Data in subjects with basal HDL-C levels less than 40 mg/dL are indicated separately. D: Effect of pitavastatin on remnant-like particle cholesterol (RLP-C) levels.

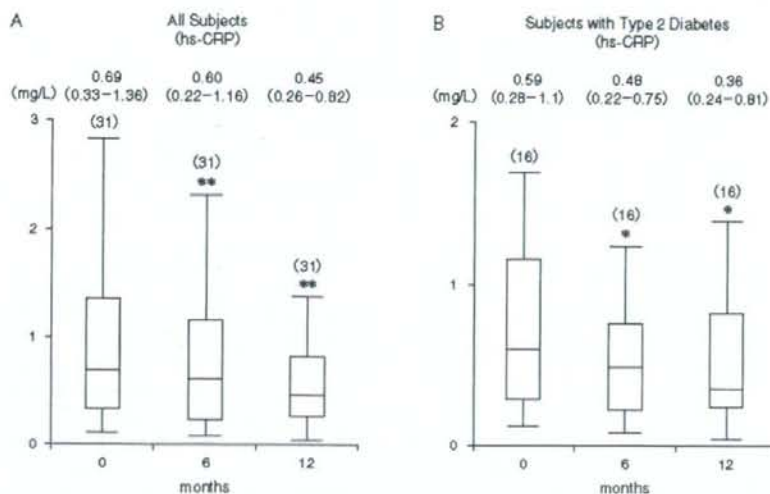


Fig. 2. Effect of pitavastatin on hs-CRP levels in all patients (panel A) and patients with type 2 diabetes (panel B) after 12 months.

Data are presented as the mean \pm S.E.M. Numbers of subjects are shown in parentheses. * $p < 0.05$ vs. basal values, ** $p < 0.01$ vs. basal values.

Discussion

The present study demonstrated that pitavastatin improved serum lipid profiles in Japanese subjects with hypercholesterolemia: a decrease in LDL-C, an increase in HDL-C, a decrease in RLP-C, and a decrease in TG among those with higher basal TG levels. The improvement in lipid profiles was compatible with a recent report on pitavastatin in Japanese subjects¹⁶, although, in the present study, LDL-C levels before the administration of pitavastatin were lower (153 ± 3 mg/dL) than in the previous report (183 ± 21 mg/dL)¹⁶.

Like other statins, pitavastatin has been indicated to have pleiotropic effects on vascular cells *in vitro*. In cell culture experiments, for example, pitavastatin is reported to reduce osteopontin expression in rat vascular smooth muscle cells¹⁷, suppress interleukin-8 (IL-8), monocyte chemoattractant protein-1 and endothelin-1 expression, enhance endothelial nitric oxide synthase expression⁷, and inhibit CRP-induced IL-8 production⁸, thus showing its direct anti-inflammatory and anti-atherogenic effect. However, there have been no reports about anti-inflammatory effects of pitavastatin *in vivo* in humans.

The present study showed that pitavastatin lowered hs-CRP in patients with hypercholesterolemia. A recent study in Korea has shown that pitavastatin

reduced mean hs-CRP levels from 24.6 to 16.5 mg/L by 8-week treatment¹⁸. In the present study, pitavastatin, at similar doses, significantly reduced hs-CRP levels in hypercholesterolemic subjects, including type 2 diabetes, with lower basal hs-CRP levels. These findings have been supported by previous reports which indicated anti-inflammatory effects of pitavastatin *in vitro*^{6,8}. There have been conflicting reports about the effects of statins on hs-CRP *in vivo* in diabetic subjects; one report did not show a significant decrease in hs-CRP after atorvastatin¹⁴. Therefore, it is possible that diabetic patients are resistant to the inhibitory effects of statins on hs-CRP, although diabetic patients showed a significant decrease in hs-CRP after atorvastatin in a recent paper¹⁹. The present as well as the previous study¹⁸, also demonstrated that pitavastatin, a new synthetic lipophilic strong statin, lowered hs-CRP levels in hypercholesterolemic subjects, including type 2 diabetes. There was no significant correlation between pitavastatin-induced decreases in hs-CRP and LDL-C, in the present study with pitavastatin, as previously reported with other statins^{9,10,12,19}. We also found no significant correlation between pitavastatin-induced decrement in hs-CRP and those in RLP-C in this study cohort. Thus, it is suggested that pitavastatin may have direct anti-inflammatory effects which are independent of improved lipid profiles. A previous study showed that non-responders,

whose LDL-C levels were above 100 mg/dL after atorvastatin treatment, tender to be resistant to hs-CRP reduction by statin treatment¹⁹; however, the present study did not show such a tendency but hs-CRP reduction to be independent of LDL-C levels after pitavastatin treatment (data not shown).

There were no serious adverse events in the present study, including A1C changes in subjects with diabetes mellitus. In contrast, atorvastatin has been reported to result in a slight but significant increase of A1C in Caucasian diabetic subjects, as shown by the Collaborative Atorvastatin Diabetes Study (CARDS)²⁰. It has been shown that Japanese subjects with type 2 diabetes are mainly insulin-deficient rather than insulin-resistant²¹. In addition, Asians have higher plasma levels of statins than Caucasians²². Therefore, it is possible to speculate that Japanese subjects with type 2 diabetes may be more prone to worsening glycemic control due to, if any, adverse effects of statins on insulin secretion; however, the present study indicated that pitavastatin did not significantly worsen glycemic control in Japanese diabetic subjects.

This study is limited by the fact that it was an uncontrolled study with a moderate number of subjects, especially a relatively small number with hs-CRP measurement. It is to be noted, however, that the subjects showed lower basal hs-CRP levels before pitavastatin treatment (median: 0.69 mg/L) than those in previous studies of Caucasians (median: about 3 mg/L)^{5,9}. In the Pravastatin Inflammation/CRP Evaluation (PRINCE) study, baseline hs-CRP levels were the major determinants of the change in hs-CRP levels after pravastatin¹², indicating that anti-inflammatory effects of statins would be more pronounced in subjects with higher hs-CRP levels. It is intriguing that pitavastatin was able to reduce hs-CRP even in subjects with lower basal hs-CRP levels in the present study. Previous large scale cardiovascular event outcome studies with other statins have been conducted in Caucasians with high basal hs-CRP levels; therefore, it is of clinical importance to investigate whether pitavastatin can similarly reduce cardiovascular events among Japanese subjects with lower basal hs-CRP values than Caucasians^{10, 23, 24}.

Conclusions

It is concluded that pitavastatin improves lipid profiles, including the reduction of RLP-C levels, and that it decreases hs-CRP levels independently of improved lipid profiles, without any adverse effects, in Japanese subjects with hypercholesterolemia, including type 2 diabetes.

Acknowledgements

The authors are very grateful to Mr. Kazunobu Kasai for his assistance with data analyses.

References

- 1) Goldstein JL, Kita T, Brown MS: Defective lipoprotein receptors and atherosclerosis. Lessons from an animal counterpart of familial hypercholesterolemia. *N Engl J Med*, 1983; 309:288-296
- 2) Kume N, Kita T, Mikami A, Yokode M, Ishii K, Nagan o Y, Kawai C: Induction of mRNA for low-density lipoprotein receptors in heterozygous Watanabe heritable hyperlipidemic rabbits treated with CS-514 (pravastatin) and cholestyramine. *Circulation*, 1989; 79:1084-1090
- 3) Koshiyama H: The impact of statin treatment on diabetic patients. *Curr Opin Investig Drugs*, 2003; 4:395-400
- 4) Liao JK: Clinical implications for statin pleiotropy. *Curr Opin Lipidol*, 2005; 16:624-629
- 5) Ridker PM, Rifai N, Pfeiffer MA, Sacks FM, Moye LA, Goldman S, Flaker GC, Braunwald E: Inflammation, pravastatin, and the risk of coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events (CARE) Investigators. *Circulation*, 1998; 98:839-844
- 6) Inoue I, Goto S, Mizotani K, Awata T, Matsunaga T, Kawai S, Nakagima T, Hokari S, Komada T, Katayama S: Lipophilic HMG-CoA reductase inhibitor has an anti-inflammatory effect: reduction of mRNA levels for interleukin-1 β , interleukin-6, cyclooxygenase-2, and p^{22phox} by regulation of peroxisome proliferator-activated receptor α (PPAR α) in primary endothelial cells. *Life Sci*, 2000; 67:863-876
- 7) Morikawa S, Takabe W, Mataka C, Kanke T, Itoh T, Wada Y, Izumi A, Saito Y, Hamakua T, Kodama T: The effect of statins on mRNA levels of genes related to inflammation, coagulation, and vascular constriction in HUVEC. Human umbilical vein endothelial cells. *J Atheroscler Thromb*, 2002; 9:178-183
- 8) Kibayashi E, Urakaze M, Kobashi C, Kishida M, Takata M, Sato A, Yamazaki K, Kobayashi M: Inhibitory effect of pitavastatin (NK-104) on the C-reactive protein-induced interleukin-8 production in human aortic endothelial cells. *Clin Sci (Lond)*, 2005; 108:515-521
- 9) Ridker PM, Rifai N, Rose L, Buring JE, Cook NR: Comparison of C-reactive protein and low-density lipoprotein cholesterol levels in the prediction of first cardiovascular events. *N Engl J Med*, 2002; 347:1557-1565
- 10) Ridker PM, Rifai N, Clearfield M, Downs JR, Weis SE, Miles JS, Gotto AM Jr, Air Force/Texas Coronary Atherosclerosis Prevention Study Investigators: Measurement of C-reactive protein for the targeting of statin therapy in the primary prevention of acute coronary events. *N Engl J Med*, 2001; 344:1959-1965
- 11) Taniguchi A, Nagasaka S, Fukushima M, Sasaki M, Okumura T, Yoshii S, Watanabe T, Ogura M, Yamadori N, Nin K, Kuroe A, Yamada Y, Seino Y, Nakai Y: C-reactive protein and insulin resistance in non-obese Japanese type

- 2 diabetic patients. *Metabolism*, 2002; 51:1578-1581
- 12) Albert MA, Danielson E, Rifai N, Ridker PM: PRINCE Investigators. Effect of statin therapy on C-reactive protein levels: the pravastatin inflammation/CRP evaluation (PRINCE): a randomized trial and cohort study. *JAMA*, 2001; 286:64-70
- 13) Jialal I, Stein D, Balis D, Grundy SM, Adams-Huet B, Devaraj S: Effect of hydroxymethyl glutaryl coenzyme A reductase inhibitor therapy on high sensitive C-reactive protein levels. *Circulation*, 2001; 103:1933-1935
- 14) Economides PA, Caselli A, Tiani B, Khaodhilar L, Horton ES, Veves A: The effects of atorvastatin on endothelial function in diabetic patients and subjects at risk for type 2 diabetes. *J Clin Endocrinol Metab*, 2004; 89:740-747
- 15) Hayashi T, Yokote K, Saito Y, Iguchi A: Pitavastatin: efficacy and safety in intensive lipid lowering. *Expert Opin Pharmacother*, 2007; 8:2315-2327
- 16) Yoshitomi Y, Ishii T, Kaneki M, Tsujibayashi T, Sakurai S, Nagakura C, Miyauchi A: Efficacy of a low dose of pitavastatin compared with atorvastatin in primary hyperlipidemia: results of a 12-week, open label study. *J Atheroscler Thromb*, 2006; 13:108-113
- 17) Takemoto M, Kitahara M, Yokote K, Asaumi S, Take A, Saito Y, Mori S: NK-104, a 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitor, reduces osteopontin expression by rat aortic smooth muscle cells. *Br J Pharmacol*, 2001; 133:83-88
- 18) Lee SH, Chung N, Kwan J, Kim DI, Kim WH, Kim CJ, Kim HS, Park SH, Seo HS, Shin DG, Shin YW, Shim WJ, Ahn TH, HO Yun K, Yoon MH, Cha KS, Choi SW, Han SW, Hyon MS: Comparison of the efficacy and tolerability of pitavastatin and atorvastatin: an 8-week, multicenter, randomized, open-label, dose-titration study in Korean patients with hypercholesterolemia. *Clin Ther*, 2007; 29:2365-2373
- 19) Yamada S, Yanagawa T, Sasamoto K, Araki A, Miyao M, Yamanouchi T: Atorvastatin lowers plasma low-density lipoprotein cholesterol and C-reactive protein in Japanese type 2 diabetic patients. *Metabolism*, 2006; 55:67-71
- 20) Colhoun HM, Betteridge DJ, Durrington PN, Hitman GA, Neil HA, Livingstone SJ, Thomason MJ, Mackness MI, Charlton-Menys V, Fuller JH; CARDS investigators: Primary prevention of cardiovascular disease with atorvastatin in type 2 diabetes in the Collaborative Atorvastatin Diabetes Study (CARDS): multicentre randomised placebo-controlled trial. *Lancet*, 2004; 364:685-696
- 21) Koshiyama H, Taniguchi A, Inagaki N, Seino Y: Is the concept of "cardiometabolic risk" more useful than "metabolic syndrome"? *Diabet Med*, 2007; 24:571
- 22) Liao JK: Safety and efficacy of statins in Asians. *Am J Cardiol*, 2007; 99:410-414
- 23) Chan AW, Bhatt DL, Chew DP, Reginelli J, Schneider JP, Topol EJ, Ellis SG: Relation of inflammation and benefit of statins after percutaneous coronary interventions. *Circulation*, 2003; 107:1750-1756
- 24) Ridker PM, Cannon CP, Morrow D, Ridker PM, Cannon CP, Morrow D, Rifai N, Rose LM, McCabe CH, Pfeffer MA, Braunwald E: Pravastatin or Atorvastatin Evaluation and Infection Therapy-Thrombolysis in Myocardial Infarction 22 (PROVE IT-TIMI22) Investigators: C-reactive protein level and outcomes after statin therapy. *N Engl J Med*, 2005; 352:20-28