

in each litter) in each group were selected as F1 parents on PNDs 21–25 to equalize the body weights among groups. The day on which F1 parental animals were selected was designated as 0 week of dosing for the F1 generation. The administration of DCBS in the diet was not suspended during PNDs 21–25. F1-selected rats were administered DCBS in the diet with the respective formulation for 10 weeks prior to the mating period and mated as described above. Administration of DCBS in the diet was continued throughout the mating, gestation, and lactation periods. On PND 26, F1 weanlings not selected for breeding and all F2 weanlings were necropsied.

#### 2.4. Mating procedures

Each female was mated with a single male of the same dosage group until copulation occurred or the mating period had elapsed. The mating periods for F0 and F1 animals were three weeks. During the mating period, daily vaginal smears were examined for the presence of sperm. The presence of sperm in the vaginal smear and/or a vaginal plug was considered evidence for successful mating. The day of successful mating was designated as day 0 of pregnancy. F1 females that did not mate during the 3-week mating period were cohoused with other males from the same group who had been proven to copulate. For F1 matings, cohabitation of siblings was avoided.

#### 2.5. Parental data

All adult rats were observed twice a day for clinical signs of toxicity, and body weights and food consumption were recorded weekly. For females exhibiting evidence of successful mating, body weight and food consumption were recorded on days 0, 7, 14, and 20 of pregnancy and days 0, 4, 7, 14, and 21 of lactation. Daily vaginal lavage samples of each F0 and F1 female were evaluated for estrous cyclicity throughout the 2-week pre-cohabitation period and during cohabitation until evidence of copulation was detected. Females having repeated 4–6 day estrous cycles were judged to have normal estrous cycles. After weaning of their pups, parental female rats were necropsied at the proestrous stage of the estrous cycle. For each female, the number of uterine implantation sites was recorded.

#### 2.6. Litter data

Once insemination was confirmed, female rats were checked at least three times daily at days 21–25 of pregnancy to determine the time of delivery. The females were allowed to deliver spontaneously and nurse their pups until PND 21 (the day of weaning). The day on which parturition was completed by 13:00 was designated as PND 0. Total litter size and the numbers of live and dead pups were recorded, and live pups were counted, sexed, examined grossly, and individually weighed on PNDs 0, 4, 7, 14, and 21. On PND 4, litters were randomly adjusted to eight pups comprising of four males and four females. No adjustment was made for litters of fewer than eight pups. Selected pups were assigned a unique number and limb tattooed on PND 4.

#### 2.7. Developmental landmarks

All F1 and F2 pups were observed daily for pinna unfolding on PNDs 1–4, incisor eruption beginning on PND 8, and eye opening beginning on PND 12. One male and one female F1 and F2 pup selected from each dam was evaluated for the surface righting reflex on PND 5, negative geotaxis reflex on PND 8, and mid-air righting reflex on PND 18 [16]. All F1 offspring were observed daily for male preputial separation beginning on PND 35 or female vaginal opening beginning on PND 25. Body weight of the respective F1 rats was recorded on the day of preputial separation or vaginal opening. The AGD was measured using calipers on PND 4 in all F1 and F2 pups, and the AGD per cube root of body weight ratio was calculated [17].

#### 2.8. Behavioral tests

Spontaneous locomotor activity was measured with a multi-channel activity monitoring system (Supermex; Muromachi Kikai Co. Ltd., Tokyo, Japan)

in 10 male and 10 female F1 rats selected from each group at 4 weeks of age. Rats were placed individually in transparent polycarbonate cages (27.6W × 44.5D × 20.4H cm, CL-0108-1, Clea Japan Inc., Tokyo, Japan) under an infrared sensor that detects thermal radiation from animals. Spontaneous motor activity was determined for 10 min intervals and for a total of 60 min.

A test in a water-filled multiple T-maze was conducted in 10 male and 10 female F1 rats selected from each group at 6 weeks of age. The apparatus was similar to that described by Biel [18]. The water temperature of the maze was kept 22–23 °C. As a preliminary swimming ability test, each rat was allowed to swim three times in a straight channel on the day before the maze trial, and then tested in the maze with three trials per day for the next consecutive three days. The elapsed time between entry into the water at the starting point and touching the goal ramp as well as the number of errors were recorded. To prevent exhaustion of the rats, no animal was allowed to remain in the water for more than 3 min in any trial.

#### 2.9. Termination/necropsy-adults

Parental rats were necropsied: males after the parturition of paired female, and females after weaning of their pups. Ages on the day of the scheduled terminal sacrifice were 19–20 weeks old in F0 males, 21–22 weeks old in F0 females, 18 weeks old in F1 males and 19–20 weeks old in F1 females. The proestrous stage of the estrous cycle was characterized by examination of the vaginal smears of female rats on the day of necropsy. A complete necropsy was performed on all rats found dead and those killed at the scheduled terminal sacrifice. Live rats were euthanized by exsanguination under ether anesthesia. The external surfaces of the rats were examined. The abdomen and thoracic cavities were opened, and a gross internal examination was performed. Weights of the brain, pituitary, thyroid, thymus, liver, kidney, spleen, adrenal, testis, epididymis, seminal vesicle (with coagulating glands and their fluids), ventral prostate, uterus and ovary were recorded. Weights of the thyroid and seminal vesicle were measured after fixation. Major organs were stored in 10% neutral buffered formalin. The testis and epididymis were fixed with Bouin's solution and preserved in 70% ethanol.

Histopathological evaluations in F0 and F1 adults were performed on the tissues specified below after fixation, paraffin embedding, and sectioning and staining with hematoxylin and eosin: the liver, pituitary, thymus, thyroid, kidney, spleen, adrenal, bone marrow, mesenteric lymph node, Peyer's patches, testis, epididymis, seminal vesicle, coagulating gland, ventral prostate, ovary, uterus, vagina and mammary gland of all males and females in the control and highest dose (4500 ppm) groups and of females with abnormal estrous cycles, of males and females without evidence of copulation or insemination and of females with abnormal delivery or totally dead pups in all groups. Any organs or tissues of F0 and F1 adults showing gross alterations were evaluated histopathologically.

In ten each F1 females of the control and highest dose groups, the primordial follicles were counted [19]. The right ovary was fixed in 10% neutral buffered formalin and then dehydrated and embedded in paraffin in a longitudinal orientation by routine procedures. Sections were cut serially at 5 µm and every 20th one was serially mounted on slides and stained with hematoxylin and eosin. About 40 sections per ovary were used to determine the primordial follicles.

#### 2.10. Termination/necropsy-pups

Following adjustment of litter size on PND 4, culled pups were euthanized by inhalation of carbon dioxide and subjected to a gross external and internal necropsy. No tissues from these pups were collected.

The weanlings not selected to become parents were euthanized and necropsied as described for the adults. Organ weights of one male and one female F1 and F2 weanling selected from each dam was measured as described above for adults. The weights of the pituitary and thyroid were not determined in weanlings. All pups found dead before weaning were also necropsied.

In all male and female F1 and F2 weanlings whose organs were collected, histopathological evaluations of the thymus, liver and spleen in the control and 4500 ppm groups were performed after fixation, paraffin embedding, and sectioning and staining with hematoxylin and eosin.

### 2.11. Hematological and blood biochemical parameters

On the day of the scheduled terminal sacrifice, blood samples were collected from the abdominal aorta of adult rats under ether anesthesia.

Hematological examinations were performed for 10 males and 10 females of F0 and F1 rats randomly selected from each group. Blood samples were analyzed for the following hematological parameters, using 2K-EDTA as an anticoagulant: white blood cell count (WBC) and differential leukocyte count.

Blood biochemical evaluations were performed for 10 males and 10 females of F0 and F1 rats randomly selected from each group. Serum samples obtained from centrifuged whole blood were analyzed for biochemistry parameters such as total protein, albumin and globulin.

### 2.12. Serum hormone levels

On the day of the scheduled terminal sacrifice, blood samples were collected from the abdominal aorta of adult rats. Eight males and eight proestrous females of the F0 and F1 generations from each group were selected randomly for blood collection. Hormone levels were determined by Panapharm Laboratories Co. Ltd. (Uto, Japan). Serum levels of testosterone, 5 $\alpha$ -dihydrotestosterone (DHT), luteinizing hormone (LH), and follicle stimulating hormone (FSH) in males, and estradiol, progesterone, LH, and FSH in females were measured. The testosterone, DHT, estradiol, and progesterone concentrations were measured using a double antibody kit (Diagnostic Products Corp., Los Angeles, CA or Diagnostic Systems Laboratories Inc., Webster, TX). Serum concentrations of LH and FSH were measured using (rat LH)[125I] and (rat FSH) [125I] assay systems (GE Healthcare Bio-Sciences Corp., Piscataway, NJ), respectively.

### 2.13. Sperm parameters

Sperm parameters were determined for all F0 and F1 male adults, except dead males, on the day of the scheduled terminal sacrifice. The right testis was used to count testicular homogenization-resistant spermatid heads. The right cauda epididymis was weighed and used for sperm analysis. Sperm motility was analyzed using a computer-assisted cell motion analyzer (TOX IVOS, Hamilton Thome Biosciences, Beverly, MA). The percentage of motile sperm and progressively motile sperm as well as their swimming speed and pattern were determined. After the recording of sperm motion, the cauda epididymal fluid was diluted and sperm were enumerated using a hemacytometer under a light microscope. A sperm count per gram of epididymal tissue was obtained by dividing the total count by the gram weight of the cauda epididymis. The sperm were stained with eosin and mounted on a slide glass. Two hundred sperm in each sample were examined under a light microscope, and the percentage of morphologically abnormal sperm was calculated.

### 2.14. Statistical analysis

Statistical analysis of offspring before weaning was carried out using the litter as the experimental unit.

Body weight, body weight gain, food consumption, length of estrous cycle, precoital interval, gestation length, numbers of implantations and pups delivered, delivery index, sperm parameters, hematological and blood chemical parameters, hormone levels, organ weight, organ/body weight ratio (relative organ weight), reflex response time, age displayed pinna unfolding, incisor eruption, and eye opening, age and body weight at sexual maturation, parameters of behavioral tests, AGD, AGD/cube root of body weight ratio, and the viability of pups were analyzed for statistical significance in the following way. Bartlett's test of homogeneity of variance was used to determine if the groups had equivalent variances. If the variances were equivalent, the groups were compared by one-way analysis of variance (ANOVA). If significant differences were found, Dunnett's multiple comparison test was performed. If the groups did not have equivalent variances, the Kruskal-Wallis test was used to assess the overall effects. Whenever significant differences were noted, pairwise comparisons were made by Mann-Whitney *U*-test. The incidence of pups with changes in clinical and gross internal observations, and reflex completion rate of pups were analyzed by Wilcoxon rank sum test. The number of primordial follicles in the control and highest dose groups was analyzed in the following way. Variance ratio was analyzed by *F*-test. Since the variance ratio was equivalent, the groups were compared by Student's *t*-test. The incidence of females with normal estrous cycles, copulation index, fertility index, gestation index, neonatal sex ratio, and completion rate of the reflex response were analyzed by Fisher's exact test.

The 0.05 level of probability was used as the criterion for significance.

## 3. Results

### 3.1. Clinical observations, body weight and food consumption during the pre-mating, mating, gestation, and lactation periods (F0 and F1)

There were no compound-related clinical signs of toxicity in either male or female F0 and F1 rats during the pre-mating, mating, gestation, or lactation periods. One F0 male at 80 ppm was euthanized in 11 weeks of dosing because of a moribund condition resulting from accidental injury in the home cage. One F1 female without any apparent clinical signs of toxicity died on day 5 of lactation in the control group, and no abnormal necropsy findings were found.

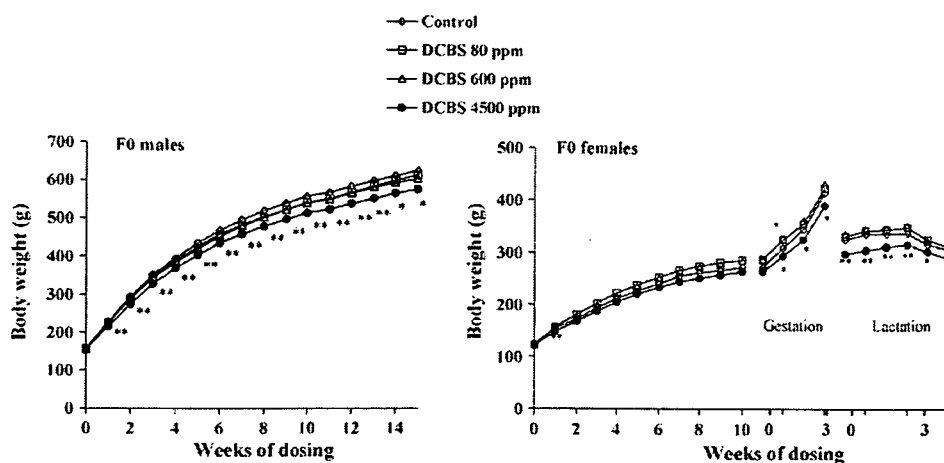


Fig. 1. Body weight of F0 males and females. \*Significantly different from the control.  $p < 0.05$ . \*\*Significantly different from the control.  $p < 0.01$ .

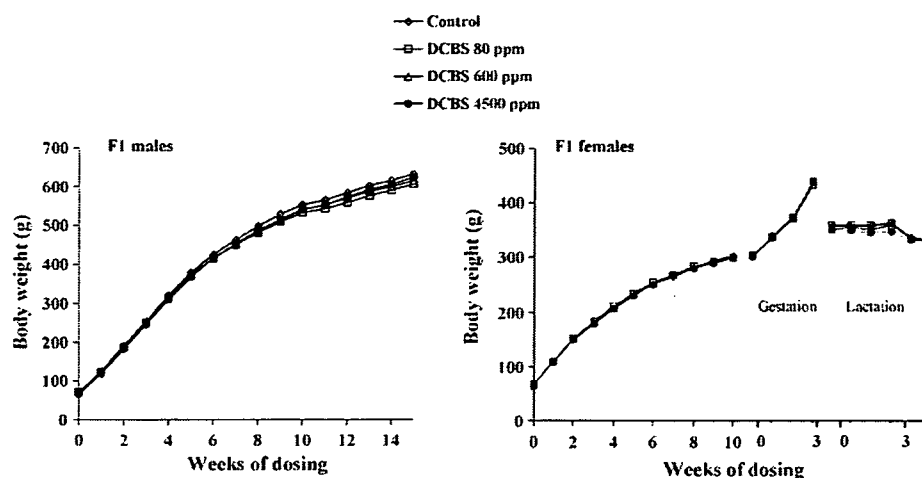


Fig. 2. Body weight of F1 males and females.

The body weights of F0 males and females during dosing are shown in Fig. 1. The body weight and body weight gain of male F0 rats were significantly lowered throughout the dosing period at 4500 ppm. At this dose, the body weight and body weight gain of F0 females were significantly reduced during the first week of dosing and throughout pregnancy and lactation. No compound-related changes in the body weight or body weight gain were noted in F0 males and females at 80 and 600 ppm.

Fig. 2 shows the body weights of F1 males and females during the dosing period. The body weight and body weight gain of F1 males and females exhibited no significant differences between the control and DCBS-treated groups.

There was a significant decrease in food consumption during weeks 1–8 and 13–14 of dosing in F0 males and during the first week of dosing and days 14–21 of lactation in F0 females at 4500 ppm. No significant changes in food consumption were observed in F0 rats of both sexes at 80 and 600 ppm (data not shown).

In F1 male rats, a significant decrease in food consumption was found during weeks 4–7 of dosing at 80 ppm, during week 6 of dosing at 600 ppm and during week 4 of dosing at 4500 ppm. No significant changes were observed in food consumption in F1 females at any dose (data not shown).

The mean daily intakes of DCBS were 5.2, 39 and 291 mg/kg bw in F0 males, 7.2, 54 and 416 mg/kg bw in F0 females, 5.9, 44 and 331 mg/kg bw in F1 males, and 7.4, 55 and 417 mg/kg bw in F1 females for 80, 600 and 4500 ppm, respectively.

### 3.2. Estrous cyclicity (F0 and F1 females)

Table 1 presents the estrous cyclicity of F0 and F1 females. All F0 females showed normal estrous cycles in all groups, and the length of the estrous cycles was not different between the control and DCBS-treated groups. Although one F1 female each in the control and 600 ppm groups displayed extended diestrous vaginal smears, no significant changes in the incidence of females having normal estrous cycles or length of the estrous cycles were observed.

### 3.3. Reproductive effects (F0 parents/F1 offspring and F1 parents/F2 offspring)

The reproductive and developmental parameters for F0 parent/F1 offspring are presented in Table 2. In F0 parent animals, all pairs in all groups copulated, although two females in the con-

Table 1  
Estrous cyclicity of F0 and F1 females

	DCBS (ppm)			
	0 (control)	80	600	4500
<b>F0 females</b>				
No. of females examined	24	24	24	24
Females with normal estrous cycles (%) <sup>b</sup>	100	100	100	100
Length of estrous cycles (days)	4.05 ± 0.16 <sup>a</sup>	4.01 ± 0.06	4.04 ± 0.15	4.01 ± 0.06
<b>F1 females</b>				
No. of females examined	24	24	24	24
Females with normal estrous cycles (%) <sup>b</sup>	95.8	100	95.8	100
Length of estrous cycles (days)	4.21 ± 0.34	4.05 ± 0.21	4.25 ± 1.08	4.07 ± 0.24

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Incidence of females with normal estrous cycles (%) = (no. of females with normal estrous cycles/no. of females examined) × 100.

Table 2  
Reproductive and developmental data for F0 parents/F1 offspring and F1 parents/F2 offspring

	DCBS (ppm)			
	0 (control)	80	600	4500
<b>F0 parents/F1 offspring</b>				
No. of pairs	24	24	24	24
Copulation index (%) <sup>b</sup>				
Male/female	100/100	100/100	100/100	100/100
Fertility index (%) <sup>c</sup>	91.7	100	100	100
No. of pregnant females	22	24	24	24
Precoital interval (days)	2.4 ± 1.2 <sup>a</sup>	2.8 ± 1.1	2.4 ± 1.0	2.4 ± 1.1
Gestation index (%) <sup>d</sup>	100	100	100	100
Gestation length (days)	22.1 ± 0.4	22.2 ± 0.4	22.0 ± 0.3	22.1 ± 0.3
No. of implantations	13.5 ± 2.1	13.9 ± 1.4	14.6 ± 1.3	13.2 ± 1.5
Delivery index (%) <sup>e</sup>	94.9	94.9	94.3	94.8
No. of pups delivered	12.8 ± 2.1	13.2 ± 1.6	13.8 ± 1.5	12.5 ± 1.7
No. of litters	22	24	24	24
Sex ratio of F1 pups <sup>f</sup>	0.528	0.554	0.506	0.525
<b>Viability index during lactation (%)<sup>g,h,i</sup></b>				
Day 0	99.0	99.3	99.7	99.0
Day 4	98.7	98.2	96.6	97.6
Day 21	100	99.0	99.5	99.5
<b>Male pup weight during lactation (g)</b>				
Day 0	6.9 ± 0.5	6.7 ± 0.6	6.7 ± 0.6	6.6 ± 0.7
Day 4	11.2 ± 1.1	10.5 ± 1.2	10.5 ± 1.4	10.3 ± 1.0*
Day 7	18.6 ± 1.8	18.1 ± 1.7	17.7 ± 2.5	16.7 ± 1.6**
Day 14	37.2 ± 3.6	36.8 ± 2.4	36.0 ± 4.0	33.6 ± 2.5**
Day 21	62.3 ± 5.6	62.2 ± 3.7	60.2 ± 6.3	55.3 ± 4.8**
<b>Female pup weight during lactation (g)</b>				
Day 0	6.5 ± 0.5	6.3 ± 0.5	6.3 ± 0.5	6.3 ± 0.6
Day 4	10.9 ± 1.3	10.1 ± 1.4	10.0 ± 1.2	9.9 ± 1.0*
Day 7	18.1 ± 1.9	17.1 ± 2.3	17.2 ± 2.3	16.2 ± 1.4**
Day 14	36.3 ± 3.5	34.8 ± 3.6	35.0 ± 4.0	32.8 ± 2.6**
Day 21	60.7 ± 5.2	58.5 ± 6.0	58.2 ± 6.5	53.7 ± 4.5**
<b>F1 parents/F2 offspring</b>				
No. of pairs	24	24	24	24
Copulation index (%) <sup>b</sup>				
Male/female	100/100	100/100	91.7/100	100/100
Fertility index (%) <sup>c</sup>	95.8	91.7	91.7	100
No. of pregnant females	23	22	22	24
Precoital interval (days)	2.7 ± 1.0	2.6 ± 1.4	2.6 ± 1.2	2.8 ± 1.7
Gestation index (%) <sup>d</sup>	100	100	95.5	100
Gestation length (days)	22.3 ± 0.4	22.2 ± 0.4	22.1 ± 0.4	22.1 ± 0.3
No. of implantations	14.1 ± 3.2	13.5 ± 3.7	13.0 ± 4.2	14.3 ± 2.1
Delivery index (%) <sup>e</sup>	90.4	92.9	88.9	91.3
No. of pups delivered	12.7 ± 3.6	12.6 ± 3.7	12.0 ± 4.2	13.0 ± 2.4
No. of litters	23	22	21	24
Sex ratio of F2 pups <sup>f</sup>	0.488	0.516	0.557	0.522
<b>Viability index during lactation (%)<sup>g,h,i</sup></b>				
Day 0	98.7	99.7	98.3	95.9
Day 4	95.9	94.2	93.1	88.4
Day 21	100 <sup>j</sup>	100	97.0	97.7 <sup>l</sup>
<b>Male pup weight during lactation (g)</b>				
Day 0	6.8 ± 0.9	6.7 ± 0.8	6.7 ± 0.5	6.7 ± 0.6
Day 4	11.0 ± 2.3	11.1 ± 2.6	10.0 ± 2.1	10.0 ± 1.4 <sup>l</sup>
Day 7	18.5 ± 2.7 <sup>j</sup>	18.4 ± 3.8	17.1 ± 2.8	15.9 ± 2.3 <sup>l,*</sup>
Day 14	37.1 ± 4.0 <sup>j</sup>	37.8 ± 6.3	35.5 ± 3.8	32.3 ± 4.1 <sup>l,**</sup>
Day 21	62.5 ± 7.0 <sup>j</sup>	63.4 ± 9.4	60.6 ± 5.6	53.5 ± 5.9 <sup>l,**</sup>

Table 2 (Continued)

	DCBS (ppm)			
	0 (control)	80	600	4500
Female pup weight during lactation (g)				
Day 0	6.5 ± 1.0	6.3 ± 0.7	6.3 ± 0.4 <sup>k</sup>	6.3 ± 0.7
Day 4	10.5 ± 2.3	10.5 ± 2.5	9.7 ± 2.0 <sup>k</sup>	9.5 ± 1.5 <sup>l</sup>
Day 7	17.6 ± 2.9 <sup>g</sup>	17.7 ± 3.8	16.3 ± 2.8 <sup>k</sup>	15.5 ± 2.2 <sup>l</sup>
Day 14	35.9 ± 4.1 <sup>j</sup>	36.6 ± 5.7	33.5 ± 4.9 <sup>k</sup>	31.7 ± 3.9 <sup>l,*</sup>
Day 21	59.6 ± 6.6 <sup>i</sup>	60.7 ± 8.5	56.3 ± 7.0 <sup>k</sup>	52.0 ± 5.7 <sup>l,**</sup>

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Copulation index (%) = (no. of animals with successful copulation/no. of animals paired) × 100.

<sup>c</sup> Fertility index (%) = (no. of females pregnant/no. of females with successful copulation) × 100.

<sup>d</sup> Gestation index (%) = (no. of females that delivered live pups/no. of pregnant females) × 100.

<sup>e</sup> Delivery index (%) = (no. of pups delivered/no. of implantations) × 100.

<sup>f</sup> Sex ratio = total no. of male pups/total no. of pups.

<sup>g</sup> Viability index on postnatal day 0 (%) = (no. of live pups on postnatal day 0/no. of pups delivered) × 100.

<sup>h</sup> Viability index on postnatal day 4 (%) = (no. of live pups on postnatal day 4/no. of live pups on postnatal day 0) × 100.

<sup>i</sup> Viability index on postnatal day 21 (%) = (no. of live pups on postnatal day 21/no. of live pups on postnatal day 4 after cull) × 100.

<sup>j</sup> Data were obtained from 22 litters because one female that died on day 5 of lactation was excluded from the data.

<sup>k</sup> Data were obtained from 20 litters because one female had no female pups.

<sup>l</sup> Data were obtained from 23 litters because one female that experienced a total litter loss on day 3 of lactation was excluded from the data.

\* Significantly different from the control,  $p < 0.05$ .

\*\* Significantly different from the control,  $p < 0.01$ .

control group did not become pregnant, and all pregnant females in all groups delivered live pups. There were no significant differences in the copulation index, fertility index, gestation index, pre-coital interval, gestation length, number of implantations, delivery index, number of F1 pups delivered, sex ratio of F1 pups, or viability of F1 pups during lactation between the control and DCBS-treated groups. No malformed F1 pups were found in any groups. A significantly lower body weight was observed in male and female F1 pups at 4500 ppm on PNDs 4, 7, 14 and 21.

The reproductive and developmental parameters for F1 parent/F2 offspring are also shown in Table 2. Two F1 males in the 600 ppm group did not copulate. One female in the control group and two females each in the 80 and 600 ppm groups did not become pregnant. One pregnant female in the 600 ppm group did not deliver. One dam in the control group died on day 5 of lactation, and her pups were euthanized. One dam experienced a total litter loss by PND 3 at 4500 ppm. No significant changes in the copulation index, fertility index, gestation index, pre-coital interval, gestation length, number of implantations, delivery index, number of F2 pups delivered, sex ratio of F2 pups, or viability of F2 pups during lactation were observed. Oligodactyly in one female of the control group and microphthalmia in one male at 80 ppm were observed. Body weights of F2 pups at 4500 ppm were significantly lowered on PNDs 7, 14 and 21 in males and PNDs 14 and 21 in females.

### 3.4. Developmental landmarks (F1 and F2)

Physical development of F1 and F2 pups is presented in Table 3. There was no significant difference in the age of male and female F1 and F2 pups that displayed pinna unfolding, or eye opening between the control and DCBS-treated groups. The completion of incisor eruption was delayed in male and female F1 pups at 80 ppm and in male and female F2 pups at 80 and

4500 ppm. The AGD and AGD per cube root of body weight ratio in male and female F1 and F2 pups were not significantly different between the control and DCBS-treated groups.

Reflex ontogeny in F1 and F2 pups is shown in Table 4. All male and female F1 pups in all groups completed the surface righting reflex on PND 5, negative geotaxis reflex on PND 8, and mid-air righting reflex on PND 18. In F1 pups, no significant difference was observed in the response time of the surface righting reflex or the negative geotaxis reflex between the control and DCBS-treated groups. Of the F2 pups, one female did not complete the surface righting reflex and one male did not complete the mid-air righting reflex at 80 ppm, one female did not complete the mid-air righting reflex at 600 ppm, and one female did not complete the negative geotaxis reflex at 4500 ppm; however, no significant difference was found between the control and DCBS-treated groups in the completion ratio and response time for these reflexes.

Table 5 presents data on sexual development in F1 rats. Although a significant delay in the age of preputial separation in males was noted at 4500 ppm, the body weight at the age of preputial separation was not significantly different between the control and DCBS-treated groups. In females, a significantly delayed age of vaginal opening and a higher body weight at the age of vaginal opening were found at 600 and 4500 ppm.

### 3.5. Behavioral effects (F1)

Spontaneous locomotor activity in 10 min intervals for a total of 60 min was not significantly different between the control and DCBS-treated groups in male and female F1 rats (data not shown).

Fig. 3 shows the results of the water filled T-maze test in F1 males and females. The pre-test swimming trials in the straight channel on the first day of the T-maze test revealed that all F1

Table 3  
Physical development in F1 and F2 pups

	DCBS (ppm)			
	0 (control)	80	600	4500
<b>F1 pups</b>				
No. of litters examined	22	23	24	24
Age at pinna unfolding (days)				
Male	2.7 ± 0.5 <sup>a</sup>	2.7 ± 0.5	2.9 ± 0.3	2.7 ± 0.5
Female	2.6 ± 0.6	2.6 ± 0.6	2.9 ± 0.4	2.7 ± 0.5
Age at incisor eruption (days)				
Male	10.2 ± 0.6	10.8 ± 0.6 <sup>**</sup>	10.3 ± 0.6	10.5 ± 0.4
Female	10.1 ± 0.6	10.7 ± 0.7 <sup>**</sup>	10.2 ± 0.7	10.2 ± 0.6
Age at eye opening (days)				
Male	14.5 ± 0.6	14.5 ± 0.5	14.7 ± 0.5	14.6 ± 0.5
Female	14.4 ± 0.6	14.5 ± 0.7	14.4 ± 0.4	14.5 ± 0.5
<b>AGD</b>				
Male pup AGD (mm)	5.60 ± 0.28	5.50 ± 0.28	5.51 ± 0.41	5.54 ± 0.28
Male pup AGD/(BW <sup>1/3</sup> )	2.51 ± 0.09	2.52 ± 0.08	2.52 ± 0.12	2.55 ± 0.09
Female pup AGD (mm)	3.02 ± 0.11	2.95 ± 0.14	2.99 ± 0.14	2.96 ± 0.14
Female pup AGD/(BW <sup>1/3</sup> )	1.36 ± 0.05	1.37 ± 0.06	1.39 ± 0.04	1.38 ± 0.04
<b>F2 pups</b>				
No. of litters examined	23	22	21	23
Age at pinna unfolding (days)				
Male	2.7 ± 0.8	2.7 ± 0.7	2.8 ± 0.6	2.7 ± 0.5
Female	2.7 ± 0.8	2.7 ± 0.8	2.8 ± 0.4 <sup>c</sup>	2.6 ± 0.6
Age at incisor eruption (days)				
Male	9.7 ± 0.7 <sup>b</sup>	10.6 ± 0.9 <sup>**</sup>	9.9 ± 0.6	10.3 ± 0.8 <sup>*</sup>
Female	9.8 ± 0.7 <sup>b</sup>	10.4 ± 0.8 <sup>*</sup>	10.0 ± 0.6 <sup>c</sup>	10.4 ± 0.9 <sup>*</sup>
Age at eye opening (days)				
Male	14.4 ± 0.7 <sup>b</sup>	14.6 ± 0.8	14.3 ± 0.7	14.6 ± 0.6
Female	14.3 ± 0.6 <sup>b</sup>	14.4 ± 0.8	14.4 ± 0.5 <sup>c</sup>	14.5 ± 0.7
<b>AGD</b>				
Male pup AGD (mm)	5.54 ± 0.51	5.60 ± 0.55	5.39 ± 0.56	5.47 ± 0.38
Male pup AGD/(BW <sup>1/3</sup> )	2.50 ± 0.12	2.53 ± 0.14	2.51 ± 0.12	2.55 ± 0.08
Female pup AGD (mm)	2.93 ± 0.19	2.91 ± 0.22	2.88 ± 0.19 <sup>c</sup>	2.85 ± 0.18
Female pup AGD/(BW <sup>1/3</sup> )	1.34 ± 0.04	1.34 ± 0.06	1.35 ± 0.03 <sup>c</sup>	1.35 ± 0.05

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Data were obtained from 22 litters because one dam that died on day 5 of lactation was excluded from the data.

<sup>c</sup> Data were obtained from 20 litters because one female had no female pups.

<sup>\*</sup> Significantly different from the control,  $p < 0.05$ .

<sup>\*\*</sup> Significantly different from the control,  $p < 0.01$ .

rats in each group could swim satisfactorily, and no significant changes in the elapsed time to traverse the straight channel were observed. In males, no significant differences were observed between the control and DCBS-treated groups in the elapsed time and number of errors in on days 2–4 of the T-maze test. In females, a significantly longer elapsed time at 600 and 4500 ppm and more errors at 4500 ppm were noted on day 2 of the T-maze test. There were no significant differences in the elapsed time or number of errors on days 3 and 4 of the T-maze test in female rats between the control and DCBS-treated groups.

### 3.6. Necropsy and histopathology (F0, F1 and F2)

There were no compound-related gross lesions or microscopic alterations in the reproductive organs of F0 and F1 males and females showing reproductive difficulties. No compound-

related gross lesions or remarkable microscopic alterations of tissues and organs, including the reproductive organs, were noted in F0 and F1 males and females in the highest dose group and dead animals before the scheduled terminal sacrifice. In the histopathological examinations of the ovary in F1 females, no significant difference was noted in the number of primordial follicles (mean ± S.D.) between the control (323 ± 57) and 4500 ppm (255 ± 109) groups. There were no compound-related gross lesions or microscopic alterations in male and female F1 and F2 pups, including pups that died before weaning (data not shown).

### 3.7. Organ weights (F0 adults)

The body weight at the scheduled terminal sacrifice was significantly lowered at 4500 ppm in males and females. Sig-

Table 4  
Reflex ontogeny in F1 and F2 pups

	DCBS (ppm)			
	0 (control)	80	600	4500
<b>F1 pups</b>				
No. of pups examined (male/female)	22/22	24/24	24/24	24/24
Surface righting reflex completion rate (%)				
Male/female	100/100	100/100	100/100	100/100
Surface righting reflex response time (s)				
Male	2.1 ± 1.6 <sup>a</sup>	1.5 ± 0.5	2.4 ± 2.3	1.8 ± 1.2
Female	2.8 ± 3.4	1.6 ± 0.6	1.9 ± 0.9	3.4 ± 3.9
Negative geotaxis reflex completion rate (%)				
Male/female	100/100	100/100	100/100	100/100
Negative geotaxis reflex response time (s)				
Male	14.5 ± 8.0	15.4 ± 8.2	13.8 ± 6.4	16.0 ± 7.5
Female	15.3 ± 6.8	14.1 ± 6.0	15.4 ± 6.2	18.3 ± 7.6
Mid-air righting reflex completion rate (%)				
Male/female	100/100	100/100	100/100	100/100
<b>F2 pups</b>				
No. of pups examined (male/female)	22/22	22/22	21/20	23/23
Surface righting reflex completion rate (%)				
Male/female	100/100	100/95.5	100/100	100/100
Surface righting reflex response time (s)				
Male	2.5 ± 1.6	2.2 ± 1.8	1.7 ± 0.5	2.1 ± 1.9
Female	2.6 ± 1.8	2.4 ± 2.0 <sup>b</sup>	2.5 ± 1.7	3.2 ± 4.5
Negative geotaxis reflex completion rate (%)				
Male/female	100/100	100/100	100/100	100/95.7
Negative geotaxis reflex response time (s)				
Male	15.3 ± 6.3	17.2 ± 7.4	14.4 ± 5.7	16.1 ± 4.9
Female	16.9 ± 7.2	14.0 ± 6.5	12.6 ± 8.1	16.0 ± 6.2 <sup>c</sup>
Mid-air righting reflex completion rate (%)				
Male/female	100/100	95.5/100	100/95.0	100/100

Surface righting reflex on postnatal day 5, negative geotaxis reflex on postnatal day 8 and mid-air righting reflex on postnatal day 18 were examined three times. Completion rate (%) = (number of animals showing all successful responses of three trials/number of animals examined) × 100.

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Data were obtained from 21 pups.

<sup>c</sup> Data were obtained from 22 pups.

Table 5  
Sexual development in F1 males and females

	DCBS (ppm)			
	0 (control)	80	600	4500
<b>Male preputial separation</b>				
No. of males examined	24	24	24	24
Age (days)	41.3 ± 1.6 <sup>a</sup>	41.4 ± 1.6	41.8 ± 1.6	42.8 ± 1.5 <sup>**</sup>
Body weight (g)	226.9 ± 20.3	226.5 ± 18.5	228.3 ± 17.0	229.6 ± 17.5
<b>Female vaginal opening</b>				
No. of females examined	24	24	24	24
Age (days)	29.6 ± 1.0	30.0 ± 1.7	31.2 ± 1.7 <sup>**</sup>	31.1 ± 1.3 <sup>**</sup>
Body weight (g)	104.6 ± 9.4	109.1 ± 10.6	112.1 ± 13.8 <sup>*</sup>	112.3 ± 9.1 <sup>*</sup>

<sup>a</sup> Values are given as the mean ± S.D.

<sup>\*</sup> Significantly different from the control,  $p < 0.05$ .

<sup>\*\*</sup> Significantly different from the control,  $p < 0.01$ .

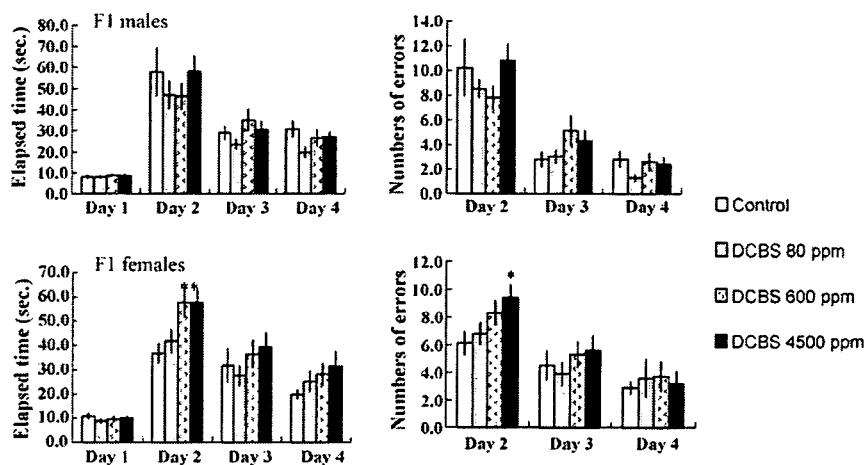


Fig. 3. Performance in water-filled multiple T-maze in F1 males and females. Each rat was allowed to swim in a straight channel on day 1, and then tested in the maze for the next consecutive three days (days 2–4). Values are given as the mean  $\pm$  S.E.M. \*Significantly different from the control,  $p < 0.05$ .

nificantly decreased absolute weights of the spleen and adrenal gland, and increased relative weights of the brain, thyroid, liver, kidney and testis were detected at 4500 ppm in males. A significant increase in the absolute weights of the brain at 80 and 600 ppm and the pituitary at 80 ppm, and decrease in the relative weight of the spleen at 80 and 600 ppm was observed in F0 females. Significantly decreased absolute weight of the spleen, and increased relative weights of the brain, kidney, and adrenal gland were found at 4500 ppm in females (data not shown).

### 3.8. Organ weights (F1 weanlings and adults)

The organ weights of male and female F1 weanlings are presented in Table 6. The body weight at the scheduled sacrifice was significantly lowered in males and females at 4500 ppm. The relative weights of the kidney at 80 ppm and the liver at 600 ppm were significantly higher in males. Significant decreases in the absolute weights of the brain, thymus, liver, kidney, adrenal gland, epididymis, and ventral prostate, and decrease in both the absolute and relative weights of the spleen, and increase in the relative weights of the brain, liver and testis were all observed at 4500 ppm in males. A significantly increased relative weight of the kidney at 80 ppm and decreased absolute weight of the ovary at 600 ppm was found in females. The absolute weights of the brain, thymus, liver, kidney, spleen, adrenal, ovary and uterus, and the relative weight of the spleen were significantly lowered at 4500 ppm in females. In this group, significantly higher relative weights of the brain and liver were also observed in females.

Table 7 shows the organ weights of male F1 adults at the scheduled terminal sacrifice. The absolute and relative weights of the thymus were significantly lower at 80 ppm in males. A significantly decreased absolute weight of the brain, decreased absolute and relative weights of the seminal vesicle, increased relative weight of the kidney, and increased absolute and relative weights of the liver were noted at 4500 ppm in males.

The organ weights of female F1 adults at the scheduled terminal sacrifice are shown in Table 8. The absolute weight of the

brain at 80 and 600 ppm, and the relative weights of the liver and kidney, and the absolute and relative weights of the adrenal gland at 4500 ppm were significantly increased.

### 3.9. Organ weights (F2 weanlings)

Table 9 presents the organ weights of male F2 weanlings. The body weight at sacrifice was significantly reduced at 4500 ppm. A significant decrease in the absolute and relative weight of the spleen was observed at 80 ppm. The relative weights of the liver and kidney were significantly higher at 600 ppm. At 4500 ppm, a significantly decreased absolute weight of the adrenal gland, decreased absolute and relative weights of the thymus and spleen, and increased relative weights of the brain, liver, and kidney were noted in males.

Table 9 also presents the organ weights of female F2 weanlings. A significant decrease in the body weight at sacrifice was found at 4500 ppm. The relative weight of the thymus was significantly lower at 80 ppm. Significantly increased relative weights of the liver and kidney, and reduced absolute and relative weights of the uterus were found at 600 ppm. At 4500 ppm, significantly decreased absolute weights of the brain and spleen, and absolute and relative weights of the thymus and uterus, and increased relative weights of the brain, liver and kidney were noted in females.

### 3.10. Hematological and blood biochemical parameters (F0 and F1 adults)

A significantly higher percent of lymphocytes was observed in male F0 adults at 4500 ppm and in female F1 adults at 600 ppm. In female F0 and male F1 adults, no significant difference was noted in the WBC or differential leukocyte count between the control and DCBS-treated groups. There were no significant changes in biochemistry parameters such as total protein, albumin and globulin in male and female F0 and F1 adult rats (data not shown).



Table 6  
Organ weight of male and female F1 weanlings

	DCBS (ppm)			
	0 (control)	80	600	4500
No. of male F1 weanlings examined	22	24	24	24
Body weight (g)	92.2 ± 8.0 <sup>a</sup>	88.9 ± 6.2	88.1 ± 9.1	78.2 ± 7.2 <sup>**</sup>
Brain (g)	1.69 ± 0.07 <sup>c</sup>	1.69 ± 0.05	1.70 ± 0.08	1.63 ± 0.07 <sup>**</sup>
	1.84 ± 0.13 <sup>d</sup>	1.91 ± 0.13	1.94 ± 0.15	2.09 ± 0.17 <sup>**</sup>
Thymus (mg)	359 ± 68	350 ± 53	365 ± 52	278 ± 37 <sup>**</sup>
	388 ± 52 <sup>d</sup>	393 ± 50	416 ± 61	357 ± 48
Liver (g)	4.16 ± 0.49	4.09 ± 0.37	4.09 ± 0.44	3.83 ± 0.51 <sup>*</sup>
	4.51 ± 0.26 <sup>d</sup>	4.60 ± 0.25	4.64 ± 0.15 <sup>*</sup>	4.89 ± 0.28 <sup>**</sup>
Kidney <sup>b</sup> (g)	1.023 ± 0.111	1.040 ± 0.079	1.003 ± 0.135	0.894 ± 0.069 <sup>**</sup>
	1.110 ± 0.073 <sup>d</sup>	1.171 ± 0.064 <sup>*</sup>	1.137 ± 0.079	1.146 ± 0.073
Spleen (mg)	394 ± 68	352 ± 68	356 ± 61	278 ± 41 <sup>**</sup>
	425 ± 52 <sup>d</sup>	395 ± 63	405 ± 59	357 ± 48 <sup>**</sup>
Adrenal <sup>b</sup> (mg)	25.5 ± 2.6	25.5 ± 3.3	25.0 ± 3.3	22.4 ± 2.9 <sup>**</sup>
	27.8 ± 2.9 <sup>d</sup>	28.7 ± 3.0	28.4 ± 3.1	28.8 ± 4.0
Testis <sup>b</sup> (mg)	561 ± 77	542 ± 64	541 ± 88	529 ± 88
	608 ± 61 <sup>d</sup>	610 ± 55	612 ± 58	677 ± 95 <sup>**</sup>
Epididymis <sup>b</sup> (mg)	78.6 ± 9.4	77.3 ± 9.5	75.9 ± 11.6	71.3 ± 10.2 <sup>*</sup>
	85.5 ± 9.8 <sup>d</sup>	86.9 ± 8.4	86.4 ± 11.3	91.3 ± 11.7
Ventral prostate (mg)	49.2 ± 9.8	47.5 ± 7.8	43.7 ± 10.1	42.3 ± 8.9 <sup>*</sup>
	53.3 ± 9.1 <sup>d</sup>	53.4 ± 7.8	49.6 ± 10.7	54.0 ± 9.5
No. of female F1 weanlings examined	22	24	24	24
Body weight (g)	85.9 ± 7.8	82.5 ± 6.4	82.8 ± 7.2	74.3 ± 6.7 <sup>**</sup>
Brain (g)	1.62 ± 0.07	1.65 ± 0.05	1.64 ± 0.05	1.57 ± 0.05 <sup>**</sup>
	1.90 ± 0.14 <sup>d</sup>	2.01 ± 0.17	2.00 ± 0.16	2.13 ± 0.16 <sup>**</sup>
Thymus (mg)	361 ± 77	327 ± 49	350 ± 60	281 ± 43 <sup>**</sup>
	418 ± 65 <sup>d</sup>	398 ± 60	423 ± 63	379 ± 56
Liver (g)	3.72 ± 0.44	3.52 ± 0.35	3.65 ± 0.40	3.43 ± 0.40 <sup>*</sup>
	4.33 ± 0.34 <sup>d</sup>	4.27 ± 0.22	4.41 ± 0.29	4.62 ± 0.31 <sup>**</sup>
Kidney <sup>b</sup> (g)	0.954 ± 0.108	0.967 ± 0.081	0.940 ± 0.114	0.850 ± 0.082 <sup>**</sup>
	1.110 ± 0.068 <sup>d</sup>	1.173 ± 0.055 <sup>**</sup>	1.133 ± 0.065	1.148 ± 0.088
Spleen (mg)	338 ± 58	323 ± 47	316 ± 53	249 ± 32 <sup>**</sup>
	392 ± 43 <sup>d</sup>	392 ± 54	382 ± 55	337 ± 49 <sup>**</sup>
Adrenal <sup>b</sup> (mg)	23.8 ± 2.6	24.5 ± 2.7	23.1 ± 2.9	21.5 ± 2.4 <sup>*</sup>
	27.8 ± 2.8 <sup>d</sup>	29.8 ± 3.6	27.9 ± 2.6	29.1 ± 3.6
Ovary <sup>b</sup> (mg)	23.2 ± 3.3	22.2 ± 3.4	20.5 ± 3.2 <sup>*</sup>	20.3 ± 3.2 <sup>*</sup>
	27.1 ± 3.3 <sup>d</sup>	27.0 ± 4.0	24.8 ± 4.3	27.5 ± 4.7
Uterus (mg)	58.2 ± 14.5	55.8 ± 7.6	62.1 ± 12.3	48.4 ± 11.8 <sup>*</sup>
	67.9 ± 15.8 <sup>d</sup>	67.9 ± 9.9	75.2 ± 14.1	65.0 ± 14.1

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Values are represented as the total weights of the organs of both sides.

<sup>c</sup> Absolute organ weight.

<sup>d</sup> Relative organ weight = organ weight (g or mg)/100 g body weight.

<sup>\*</sup> Significantly different from the control,  $p < 0.05$ .

<sup>\*\*</sup> Significantly different from the control,  $p < 0.01$ .

### 3.11. Serum hormone levels (F0 and F1 adults)

No significant changes in any serum hormone levels of male and female F0 adults were noted between the control and DCBS-treated groups (data not shown).

Serum hormone levels of male and female F1 adult rats are shown in Fig. 4. Although significantly higher levels of testosterone at 80 ppm and LH at 600 ppm were observed in F1 males, no significant changes were noted in any hormone levels in F1 males at 4500 ppm. There were no significant changes in any

Table 7  
Organ weight of male F1 adults

	DCBS (ppm)			
	0 (control)	80	600	4500
No. of male F1 adults examined	24	24	24	24
Body weight (g)	630.7 ± 74.7 <sup>a</sup>	605.1 ± 47.7	614.2 ± 52.5	622.6 ± 51.8
Brain (g)	2.26 ± 0.10 <sup>c</sup> 0.363 ± 0.038 <sup>d</sup>	2.29 ± 0.06 0.380 ± 0.028	2.26 ± 0.06 0.370 ± 0.030	2.21 ± 0.09* 0.356 ± 0.027
Pituitary gland (mg)	13.6 ± 1.4 2.17 ± 0.23 <sup>d</sup>	13.9 ± 1.3 2.30 ± 0.25	13.9 ± 1.1 2.27 ± 0.17	14.0 ± 1.6 2.26 ± 0.26
Thyroid <sup>b</sup> (mg)	24.9 ± 4.9 3.95 ± 0.66 <sup>d</sup>	23.3 ± 4.7 3.86 ± 0.78	23.8 ± 4.5 3.88 ± 0.69	24.6 ± 4.9 3.95 ± 0.67
Thymus (mg)	346 ± 116 54.8 ± 17.0 <sup>d</sup>	269 ± 54* 44.5 ± 8.9*	331 ± 83 53.9 ± 12.7	316 ± 62 50.9 ± 9.8
Liver (g)	20.80 ± 3.73 3.28 ± 0.29 <sup>d</sup>	19.69 ± 2.32 3.25 ± 0.19	21.19 ± 2.06 3.46 ± 0.28	22.82 ± 3.37* 3.65 ± 0.28**
Kidney <sup>b</sup> (g)	3.70 ± 0.52 0.586 ± 0.041 <sup>d</sup>	3.66 ± 0.23 0.606 ± 0.042	3.69 ± 0.36 0.602 ± 0.047	3.91 ± 0.43 0.629 ± 0.044**
Spleen (mg)	909 ± 129 145 ± 16 <sup>d</sup>	845 ± 141 139 ± 18	847 ± 124 138 ± 17	869 ± 162 139 ± 17
Adrenal <sup>b</sup> (mg)	60.5 ± 9.8 9.6 ± 1.5 <sup>d</sup>	60.3 ± 7.1 10.0 ± 1.0	61.8 ± 7.2 10.1 ± 1.3	61.3 ± 13.1 9.8 ± 2.0
Testis <sup>b</sup> (g)	3.60 ± 0.35 0.575 ± 0.062 <sup>d</sup>	3.61 ± 0.27 0.601 ± 0.073	3.60 ± 0.27 0.589 ± 0.066	3.78 ± 0.32 0.610 ± 0.062
Epididymis <sup>b</sup> (mg)	1348 ± 138 215 ± 24 <sup>d</sup>	1342 ± 67 223 ± 21	1327 ± 111 217 ± 22	1346 ± 118 217 ± 19
Seminal vesicle (g)	2.30 ± 0.23 0.368 ± 0.047 <sup>d</sup>	2.19 ± 0.28 0.364 ± 0.054	2.21 ± 0.22 0.362 ± 0.039	2.07 ± 0.26** 0.333 ± 0.045*
Ventral prostate (mg)	838 ± 174 133 ± 24 <sup>d</sup>	812 ± 181 134 ± 28	822 ± 190 134 ± 29	784 ± 168 127 ± 31

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Values are represented as the total weights of the organs of both sides.

<sup>c</sup> Absolute organ weight.

<sup>d</sup> Relative organ weight = organ weight (g or mg)/100 g body weight.

\* Significantly different from the control,  $p < 0.05$ .

\*\* Significantly different from the control,  $p < 0.01$ .

serum hormone levels of female F1 adults between the control and DCBS-treated groups.

### 3.12. Sperm parameters (F0 and F1 adults)

Table 10 shows the sperm parameters in F0 and F1 adult males. No significant changes in sperm counts, percentage of motile sperm and progressively motile sperm, swimming speed and pattern, or percentage of morphologically abnormal sperm were noted in F0 adults between the control and DCBS-treated groups. A significant decrease in the mean lateral head displacement was found at 4500 ppm in F1 males.

## 4. Discussion

A two-generation reproductive toxicity study was performed to further evaluate the potential effects of DCBS on reproduction and development in rats.

The deaths and clinical signs observed in the present study are not thought to be attributable to the administration of DCBS, because the incidences of deaths and clinical signs were very low and inconsistent across generations, and these occurrences are not uncommon in toxicological studies.

The decreased food consumption in F0 males and females at 4500 ppm was accompanied by decreases in the body weight and body weight gain. However, lowered food consumption in F1 males at 80, 600 and 4500 ppm was occasional, inconsistent, and unaccompanied by changes in body weight or body weight gain. It seems likely that DCBS adversely affects the body weight and food consumption in F0 rats at 4500 ppm, but not in F1 rats.

Although a few F0 and F1 adults showed reproductive difficulties, necropsy and the histopathology of reproductive organs revealed no evidence of reproductive failure in these rats. Two F1 females showing abnormal estrous cycles remained in diestrus for 10–11 days, suggesting they were pseudopregnant. No significant changes in reproductive indices were noted in any

Table 8  
Organ weight of female F1 adults

	DCBS (ppm)			
	0 (control)	80	600	4500
No. of female F1 adults examined	22	22	21	23
Body weight (g)	331.9 ± 32.5 <sup>a</sup>	331.2 ± 28.5	331.3 ± 23.1	330.2 ± 30.8
Brain (g)	2.08 ± 0.08 <sup>c</sup> 0.632 ± 0.056 <sup>d</sup>	2.17 ± 0.08 <sup>**</sup> 0.658 ± 0.056	2.15 ± 0.08 <sup>*</sup> 0.651 ± 0.043	2.08 ± 0.08 0.633 ± 0.060
Pituitary gland (mg)	15.9 ± 2.0 4.83 ± 0.73 <sup>d</sup>	16.1 ± 2.4 4.90 ± 0.79	15.8 ± 1.8 4.78 ± 0.52	16.1 ± 1.9 4.89 ± 0.66
Thyroid <sup>b</sup> (mg)	19.0 ± 3.9 5.72 ± 0.98 <sup>d</sup>	18.2 ± 2.7 5.51 ± 0.70	17.7 ± 3.5 5.35 ± 1.08	19.4 ± 4.1 5.89 ± 1.15
Thymus (mg)	251 ± 69 75.3 ± 18.4 <sup>d</sup>	212 ± 47 64.1 ± 14.2	261 ± 65 79.2 ± 20.2	211 ± 63 64.0 ± 18.7
Liver (g)	14.55 ± 1.66 4.39 ± 0.28 <sup>d</sup>	14.18 ± 2.14 4.28 ± 0.49	14.32 ± 1.49 4.33 ± 0.41	15.83 ± 2.11 4.81 ± 0.59 <sup>**</sup>
Kidney <sup>b</sup> (g)	2.37 ± 0.30 0.713 ± 0.046 <sup>d</sup>	2.39 ± 0.22 0.723 ± 0.040	2.40 ± 0.21 0.726 ± 0.063	2.53 ± 0.26 0.771 ± 0.080 <sup>**</sup>
Spleen (mg)	632 ± 73 191 ± 18 <sup>d</sup>	599 ± 63 181 ± 15	609 ± 80 184 ± 19	639 ± 115 194 ± 37
Adrenal <sup>b</sup> (mg)	70.0 ± 9.7 21.2 ± 3.2 <sup>d</sup>	73.5 ± 10.9 22.2 ± 3.1	73.4 ± 9.3 22.2 ± 3.0	77.5 ± 8.9 <sup>*</sup> 23.6 ± 3.2 <sup>*</sup>
Ovary <sup>b</sup> (mg)	110.6 ± 13.0 33.4 ± 2.9 <sup>d</sup>	109.1 ± 16.3 33.0 ± 4.5	108.5 ± 12.5 32.8 ± 3.2	108.2 ± 13.4 32.8 ± 3.3
Uterus (mg)	927 ± 191 280 ± 54 <sup>d</sup>	928 ± 128 283 ± 48	976 ± 185 295 ± 52	949 ± 192 288 ± 52

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Values are represented as the total weights of the organs of both sides.

<sup>c</sup> Absolute organ weight.

<sup>d</sup> Relative organ weight = organ weight (g or mg)/100 g body weight.

<sup>\*</sup> Significantly different from the control, *p* < 0.05.

<sup>\*\*</sup> Significantly different from the control. *p* < 0.01.

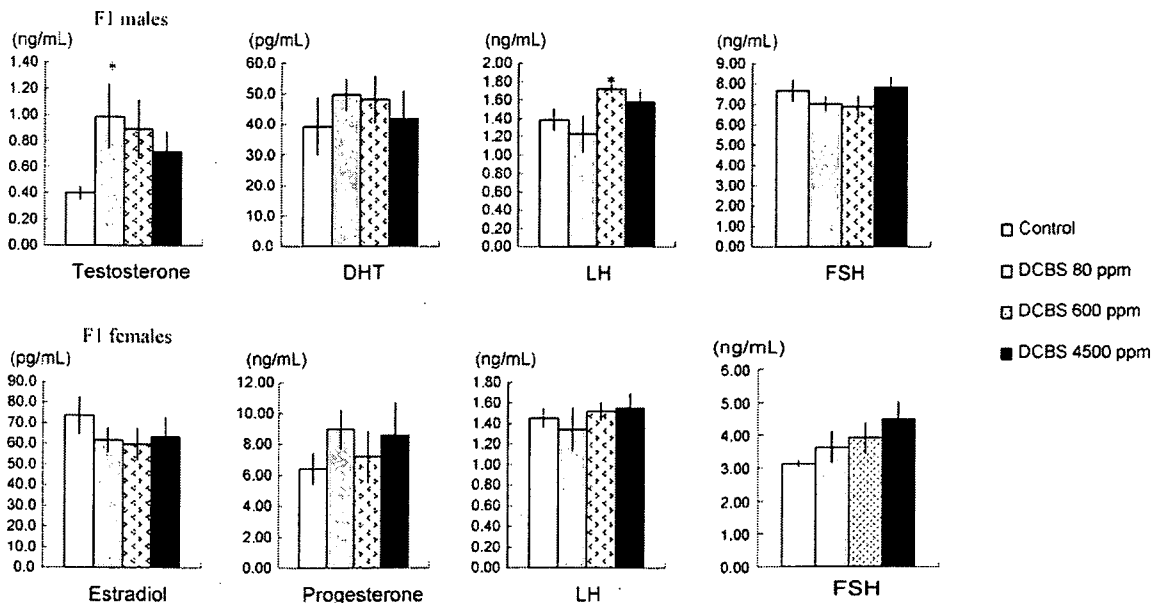


Fig. 4. Serum hormone levels in F1 males and females. The actual measurement of DHT was below the lower limit of quantification (<25.0 pg/mL) in one F1 male each in the control and 4500 ppm groups. The actual measurement of LH was below the lower limit of quantification (<0.80 ng/mL) in one F1 male and one F1 female in the 80 ppm group. Values are given as the mean ± S.E.M. \*Significantly different from the control, *p* < 0.05.

Table 9  
Organ weight of male and female F2 weanlings

	DCBS (ppm)			
	0 (control)	80	600	4500
No. of male F2 weanlings examined	22	22	21	23
Body weight (g)	90.8 ± 8.7 <sup>a</sup>	91.4 ± 13.1	89.1 ± 6.5	80.0 ± 7.8 <sup>**</sup>
Brain (g)	1.67 ± 0.08 <sup>c</sup> 1.85 ± 0.14 <sup>d</sup>	1.69 ± 0.09 1.87 ± 0.22	1.70 ± 0.08 1.92 ± 0.13	1.63 ± 0.09 2.05 ± 0.18 <sup>**</sup>
Thymus (mg)	355 ± 56 392 ± 58 <sup>d</sup>	325 ± 64 355 ± 47	361 ± 47 406 ± 48	283 ± 52 <sup>**</sup> 354 ± 55 <sup>*</sup>
Liver (g)	4.08 ± 0.47 4.49 ± 0.23 <sup>d</sup>	4.12 ± 0.66 4.50 ± 0.26	4.21 ± 0.44 4.72 ± 0.27 <sup>**</sup>	3.74 ± 0.54 4.66 ± 0.42 <sup>**</sup>
Kidney <sup>h</sup> (g)	1.006 ± 0.102 1.109 ± 0.037 <sup>d</sup>	1.009 ± 0.147 1.105 ± 0.066	1.022 ± 0.089 1.146 ± 0.049 <sup>*</sup>	0.923 ± 0.133 1.152 ± 0.117 <sup>*</sup>
Spleen (mg)	383 ± 61 422 ± 49 <sup>d</sup>	350 ± 83 <sup>*</sup> 381 ± 53 <sup>*</sup>	356 ± 46 400 ± 50	286 ± 52 <sup>**</sup> 357 ± 54 <sup>**</sup>
Adrenal <sup>h</sup> (mg)	25.5 ± 2.8 28.1 ± 2.8 <sup>d</sup>	24.1 ± 3.4 26.5 ± 2.6	24.2 ± 3.8 27.2 ± 4.3	22.7 ± 3.4 <sup>*</sup> 28.5 ± 4.2
Testis <sup>h</sup> (mg)	548 ± 106 602 ± 89 <sup>d</sup>	516 ± 103 563 ± 69	528 ± 82 590 ± 65	525 ± 98 653 ± 91
Epididymis <sup>h</sup> (mg)	79.5 ± 12.5 87.6 ± 10.6 <sup>d</sup>	72.9 ± 12.3 80.4 ± 12.6	72.7 ± 10.0 81.7 ± 10.6	71.3 ± 11.2 89.0 ± 10.0
Ventral prostate (mg)	50.9 ± 16.6 55.6 ± 15.0 <sup>d</sup>	44.6 ± 10.4 48.9 ± 9.9	47.0 ± 10.3 52.7 ± 10.1	42.6 ± 12.2 52.9 ± 13.1
No. of female F2 weanlings examined	22	22	20	23
Body weight (g)	83.6 ± 9.5	87.2 ± 10.8	82.4 ± 6.5	74.6 ± 7.9 <sup>**</sup>
Brain (g)	1.62 ± 0.08 1.96 ± 0.20 <sup>d</sup>	1.66 ± 0.07 1.92 ± 0.19	1.66 ± 0.05 2.03 ± 0.16	1.57 ± 0.07 <sup>*</sup> 2.11 ± 0.18 <sup>*</sup>
Thymus (mg)	364 ± 50 439 ± 63 <sup>d</sup>	326 ± 66 373 ± 56 <sup>**</sup>	348 ± 68 424 ± 80	283 ± 56 <sup>**</sup> 379 ± 63 <sup>**</sup>
Liver (g)	3.71 ± 0.47 4.44 ± 0.18 <sup>d</sup>	3.87 ± 0.50 4.44 ± 0.23	3.80 ± 0.37 4.61 ± 0.19 <sup>*</sup>	3.57 ± 0.46 4.78 ± 0.26 <sup>**</sup>
Kidney <sup>h</sup> (g)	0.915 ± 0.093 1.096 ± 0.046 <sup>d</sup>	0.983 ± 0.137 1.129 ± 0.085	0.960 ± 0.111 1.164 ± 0.083 <sup>**</sup>	0.885 ± 0.101 1.187 ± 0.061 <sup>**</sup>
Spleen (mg)	340 ± 63 407 ± 58 <sup>d</sup>	331 ± 55 380 ± 46	320 ± 46 389 ± 56	274 ± 40 <sup>**</sup> 370 ± 52
Adrenal <sup>h</sup> (mg)	23.6 ± 2.9 28.4 ± 3.3 <sup>d</sup>	23.3 ± 4.0 26.7 ± 3.3	22.2 ± 3.3 27.0 ± 4.0	21.6 ± 3.0 29.0 ± 3.9
Ovary <sup>h</sup> (mg)	22.0 ± 3.9 26.6 ± 5.2 <sup>d</sup>	22.5 ± 2.8 26.0 ± 3.0	20.9 ± 3.1 25.5 ± 4.4	21.4 ± 2.9 29.0 ± 4.3
Uterus (mg)	61.8 ± 18.9 73.3 ± 17.2 <sup>d</sup>	58.1 ± 11.9 67.0 ± 13.5	50.0 ± 10.0 <sup>*</sup> 60.7 ± 11.5 <sup>*</sup>	46.6 ± 12.9 <sup>**</sup> 62.3 ± 15.0 <sup>*</sup>

<sup>a</sup> Values are given as the mean ± S.D.

<sup>b</sup> Values are represented as the total weights of the organs of both sides.

<sup>c</sup> Absolute organ weight.

<sup>d</sup> Relative organ weight = organ weight (g or mg)/100 g body weight.

<sup>\*</sup> Significantly different from the control,  $p < 0.05$ .

<sup>\*\*</sup> Significantly different from the control,  $p < 0.01$ .

generation even at the highest dose of 4500 ppm. Our previous screening test revealed that DCBS given by gavage to rats from day 14 before mating to day 3 of lactation caused significant decreases in the gestation index, numbers of corpora lutea, implantations, pups born and pups born alive, live birth index, and viability index at 400 mg/kg bw per day [10]. This dose also

caused severe maternal toxicity and a total loss of pups until PND 4. No maternal or reproductive/developmental toxicity was detected at 100 mg/kg bw per day and below in our previous study. In the present feeding study, the mean daily intakes of DCBS were 416 and 417 mg/kg bw per day for the highest dose in F0 and F1 females, respectively. Consideration of these

Table 10  
Sperm parameters in F0 and F1 males

	DCBS (ppm)			
	0 (control)	80	600	4500
<b>F0 males</b>				
No. of animals	24	23	24	24
No. of testicular sperm ( $\times 10^6$ )				
Per testis	184.1 $\pm$ 29.3 <sup>a</sup>	187.7 $\pm$ 28.3	184.2 $\pm$ 32.7	180.8 $\pm$ 35.4
Per g testis	111.4 $\pm$ 13.2	110.7 $\pm$ 15.7	110.6 $\pm$ 17.1	106.1 $\pm$ 18.8
No. of epididymal sperm ( $\times 10^6$ )				
Per cauda	268.5 $\pm$ 47.6	276.2 $\pm$ 40.3	269.9 $\pm$ 56.8	263.7 $\pm$ 62.8
Per g cauda	856.4 $\pm$ 94.4	838.9 $\pm$ 99.4	850.3 $\pm$ 122.1	844.2 $\pm$ 191.3
Percent motile	88.1 $\pm$ 9.3	92.6 $\pm$ 8.2	93.2 $\pm$ 5.9	89.4 $\pm$ 10.2
Percent progressive	70.9 $\pm$ 17.4	77.3 $\pm$ 15.3	77.4 $\pm$ 12.1	70.5 $\pm$ 22.2
Mean path velocity ( $\mu\text{m/s}$ )	159.6 $\pm$ 20.8	159.8 $\pm$ 19.2	162.7 $\pm$ 22.0	156.8 $\pm$ 25.3
Straight line average velocity ( $\mu\text{m/s}$ )	112.1 $\pm$ 22.5	114.1 $\pm$ 20.0	116.1 $\pm$ 19.3	110.5 $\pm$ 29.2
Mean curvilinear velocity ( $\mu\text{m/s}$ )	365.7 $\pm$ 53.4	370.1 $\pm$ 42.5	372.3 $\pm$ 49.8	358.4 $\pm$ 56.3
Mean lateral head displacement ( $\mu\text{m}$ )	20.1 $\pm$ 1.1	19.9 $\pm$ 1.1	20.0 $\pm$ 1.3	19.9 $\pm$ 1.0
Mean beat cross frequency (Hz)	27.9 $\pm$ 1.5	27.4 $\pm$ 1.5	27.6 $\pm$ 2.2	28.3 $\pm$ 2.3
Mean straightness (%) <sup>b</sup>	69.3 $\pm$ 6.6	70.7 $\pm$ 5.7	71.0 $\pm$ 4.3	69.5 $\pm$ 8.6
Mean linearity (%) <sup>c</sup>	30.4 $\pm$ 2.8	30.7 $\pm$ 3.0	31.3 $\pm$ 2.5	30.6 $\pm$ 4.0
Total abnormal sperm ratio (%)	1.1 $\pm$ 0.6	1.2 $\pm$ 0.8	2.4 $\pm$ 3.5	2.0 $\pm$ 2.4
Tailless sperm (%)	1.0 $\pm$ 0.6	1.2 $\pm$ 0.8	2.2 $\pm$ 3.5	1.8 $\pm$ 2.0
<b>F1 males</b>				
No. of animals	24	24	24	24
No. of testicular sperm ( $\times 10^6$ )				
Per testis	194.5 $\pm$ 23.0 <sup>a</sup>	181.1 $\pm$ 21.3	186.3 $\pm$ 22.5	201.0 $\pm$ 33.3
Per g testis	115.3 $\pm$ 9.5	108.4 $\pm$ 14.3	111.1 $\pm$ 11.3	113.6 $\pm$ 15.0
No. of epididymal sperm ( $\times 10^6$ )				
Per cauda	273.6 $\pm$ 40.0	254.0 $\pm$ 40.4	256.2 $\pm$ 46.0	250.3 $\pm$ 55.4
Per g cauda	849.9 $\pm$ 69.4	821.5 $\pm$ 106.8	827.2 $\pm$ 93.3	807.0 $\pm$ 127.5
Percent motile	92.3 $\pm$ 5.0	92.9 $\pm$ 4.0	93.3 $\pm$ 5.6	93.0 $\pm$ 7.4
Percent progressive	81.8 $\pm$ 8.1	81.8 $\pm$ 4.9	83.9 $\pm$ 6.4	82.7 $\pm$ 8.2
Mean path velocity ( $\mu\text{m/s}$ )	175.2 $\pm$ 9.8	171.7 $\pm$ 11.2	172.4 $\pm$ 11.4	171.3 $\pm$ 13.9
Straight line average velocity ( $\mu\text{m/s}$ )	126.9 $\pm$ 10.2	123.9 $\pm$ 10.3	126.0 $\pm$ 10.5	125.7 $\pm$ 12.6
Mean curvilinear velocity ( $\mu\text{m/s}$ )	399.5 $\pm$ 19.8	391.5 $\pm$ 28.6	395.1 $\pm$ 28.6	393.6 $\pm$ 29.8
Mean lateral head displacement ( $\mu\text{m}$ )	21.3 $\pm$ 0.9	20.9 $\pm$ 0.8	20.8 $\pm$ 0.8	20.5 $\pm$ 1.0*
Mean beat cross frequency (Hz)	26.4 $\pm$ 1.6	26.8 $\pm$ 1.4	26.1 $\pm$ 1.6	27.0 $\pm$ 1.8
Mean straightness (%) <sup>b</sup>	72.5 $\pm$ 3.3	72.1 $\pm$ 2.7	73.3 $\pm$ 2.9	73.5 $\pm$ 2.8
Mean linearity (%) <sup>c</sup>	32.0 $\pm$ 2.1	31.9 $\pm$ 2.0	32.1 $\pm$ 1.8	32.2 $\pm$ 1.5
Total abnormal sperm ratio (%)	1.4 $\pm$ 1.3	1.1 $\pm$ 0.8	1.2 $\pm$ 1.7	1.6 $\pm$ 1.9
Tailless sperm (%)	1.3 $\pm$ 1.2	0.9 $\pm$ 0.8	1.0 $\pm$ 1.6	1.5 $\pm$ 1.8

<sup>a</sup> Values are given as the mean  $\pm$  S.D.

<sup>b</sup> Mean straightness (%) = straight line average velocity/mean path velocity  $\times$  100.

<sup>c</sup> Mean linearity (%) = straight line average velocity/mean curvilinear velocity  $\times$  100.

\* Significantly different from the control,  $p < 0.05$ .

findings suggests that the highest dose of DCBS in the present study may be very close to the dose that induces severe maternal and reproductive toxicity. However, the possibility remains that the difference in the degree of toxicity may be due to differences in administration method. There are some examples showing that gavage and feed administration result with differences in the toxicokinetics of chemicals [20,21]. Further studies are needed to clarify the relationship between maternal and reproductive/developmental toxicity.

Regarding developmental parameters, lowered body weights of male and female pre-weaning F1 and F2 pups were noted at 4500 ppm. These findings indicate that the dose level of

4500 ppm used in this study was potent enough to have adverse effects on the growth of pups. It is noted that there are strong correlations between developmental landmark parameters and pup body weight data, and that pup body weight data is consistently a more sensitive indicator of the developmental status of the offspring [22,23]. Although delayed completion of incisor eruption was noted in male and female F1 pups at 80 ppm and in male and female F2 pups at 80 and 4500 ppm, the delayed completion of incisor eruption was not dose-dependent and the difference from the control value was very slight. Therefore, it is unlikely that the delay of incisor eruption observed in the present study was compound-related or toxicologically significant. There were

no significant changes in indices of pre-weaning functional development in the DCBS-treated groups. The AGD is also a developmental landmark for the differentiation of the external genitalia and is commonly used as a hormonally sensitive parameter of sexual differentiation in rodents [24]. The AGD per cube root of body weight ratio provides a more appropriate adjustment when it is necessary to normalize AGD to body weight [17]. No changes were observed in the AGD per cube root of body weight ratio at any doses of DCBS in any generation. The data on the AGD indicate a lack of effect of DCBS on AGD. These findings on pre-weaning developmental parameters suggest that DCBS adversely affects the growth of offspring, but not the pre-weaning landmarks of development or reflex ontogeny. An increase in the frequency of fetuses with internal hydrocephalus was reported in rats given *N*-cyclohexyl-2-benzothiazolesulfenamide, a structurally similar compound, during organogenesis in rats [25]. However, no significantly increased incidence of pups with anomalies was detected even at the highest dose in the present and previous studies of DCBS [10,15]. Regarding post-weaning landmarks of development, delays of preputial separation at 4500 ppm and vaginal opening at 600 and 4500 ppm were observed in the present study. Although the body weight at the age of preputial separation was not different between the control and DCBS-treated groups, a higher body weight at the age of vaginal opening was found at 600 and 4500 ppm in females. Preputial separation and vaginal opening indicate the onset of sexual maturity, and the body weight is correlated with the occurrence of these events [23]. Ashby and Lefevre [26] described that delays in preputial separation can only be interpreted with confidence when they are not accompanied by losses in body weight, or when the expected delay in preputial separation due to a loss of body weight has been exceeded. They also noted that measurement of delays in preputial separation may be of value in cases of large delays, but delays of 1–2 days are difficult to interpret with confidence [26]. In the present study, the delay of preputial separation at 4500 ppm was slight (1.5 days) and was not accompanied by a change in body weight, and the age of preputial separation was within the range of the background control data (40.3–42.8 days) for the last seven years in the laboratory performed current study. It is likely that the delay in preputial separation at 4500 ppm is related to general delays in development. In female rats, the age at vaginal opening is the most commonly measured marker of puberty, and vaginal opening is an estrogen-dependent event that results from an increase in the blood estradiol levels [27]. Although the delay of female vaginal opening at 600 and 4500 ppm was slight (1.5–1.6 days), the age of vaginal opening was over the range of the background control data (29.6–31.0 days) for the last seven years in the laboratory performed present study. In the present study at 600 and 4500 ppm, the heavier body weight was noted at the completion of vaginal opening. Therefore, the possibility that the delay in vaginal opening may have toxicological meaning is not completely ruled out. Other hormone-dependent events including estrous cyclicity and AGD, as well as serum hormone levels at the scheduled terminal necropsy were not changed in the DCBS-treated groups. Moreover, DCBS did not affect the reproductive performance.

However, decreased weight of the uterus was found in F1 weanlings at 4500 ppm and F2 weanlings at 600 and 4500 ppm. It has been noted that variations in the weights of the reproductive organs, which are strongly dependent on endocrine status, can be considered a key parameter in the identification of endocrine effects [28–30]. These findings suggest that DCBS may have endocrine effects. Further studies are needed to clarify the effects of DCBS on endocrine endpoints.

Regarding the behavioral tests, the only significant change in the T-maze test was observed in females on day 2 of the test. Longer elapsed times at 600 and 4500 ppm and more errors at 4500 ppm were detected in females. There are behavioral functions not classically hormone-mediated and expressed by both sexes such as learning capacities, exploration activity, novelty seeking and anxiety levels that show both qualitative and quantitative differences in the two sexes [30]. The reduced activity, as well as the other effects on neuromuscular function, could be at least partially the result of lower body weight [31] and it has been found that light body weight caused worse performance in a learning task [32]. In the present study, the spontaneous activity, swimming ability in the straight channel and body weight at the time of the T-maze test was not different in F1 females between the control and DCBS-treated groups. Thus, it seems likely that DCBS may have transiently affected learning ability in the T-maze at the highest dose administered.

The changes in weight of the organs, such as the brain, thymus, kidney, and spleen that were observed at 80 and/or 600 ppm are not thought to be due to administration of DCBS, because changes occurred sporadically and not in a dose-dependent manner. The changes in the weights of the adrenal, thyroid, and male and female reproductive organs, except for the uterus, at 600 and/or 4500 ppm seem unlikely to be attributable to administration of DCBS because of inconsistent changes across ages, sexes and generations. No consistent DCBS-related effects on serum hormone levels or sperm parameters were also detected across generations. Decreased absolute weights and/or increased relative weights of the liver except for in female F0 adults, the spleen in F0 adults, and the brain and kidney in F0 and F1 adults and F1 and F2 weanlings at 4500 ppm seem to be due to secondarily lowered body weight, but not due to the direct effects of DCBS on the organs. Decreased absolute and/or relative weights of the thymus and spleen in the weanlings are supported by the results of our previous study in which atrophy of the thymus and spleen was observed at 400 mg/kg bw per day [10]. These findings may suggest that one of target systems of DCBS toxicity is the immune system in weanlings. In the present study, however, no DCBS-related histopathological changes were detected. The discrepancy in histopathological findings between the previous and present studies could be explained by a difference in the toxicokinetics of chemicals due to differences in administration method. No DCBS-related findings were found in the hematological and blood biochemical examinations. In general, the effects of DCBS on organ weights were more pronounced in weanlings than adults. These phenomena suggest that DCBS may be more toxic before weaning than after weaning, and this possibility is supported by the lowered body weight of pups during the pre-weaning period, but not post-weaning.

Table 11  
Summary of relevant findings in rat two-generation reproductive toxicity study of DCBS (80, 600 and 4500 ppm)

	F0			F1			F2				
	Male		Female	Male		Female	Male		Female		
	80	600	4500	80	600	4500	80	600	4500		
Lowered body weight			+			+			+		+
Decreased food consumption			+			+			+		+
Delayed vaginal opening								+	+		
Worse performance in water T-maze								+	+		
Reduced spleen weight						+			+		+
Reduced thymus weight						+			+		+
Reduced uterine weight									+		+

In conclusion, the results of the two-generation reproductive toxicity study described here provide a more comprehensive toxicity profile of DCBS than has been previously reported. Relevant findings obtained from the present rat two-generation reproductive toxicity study of DCBS are summarized in Table 11. The NOAEL in the present study is considered to be 80 ppm (5.2 mg/kg bw per day) in rats.

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ORIGINAL ARTICLE

## Evaluation of reproductive and developmental toxicity of the rubber accelerator N,N-dicyclohexyl-2-benzothiazolesulfenamide in rats

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**ABSTRACT** Male and female Crl:CD(SD) rats were fed a diet containing the rubber accelerator N,N-dicyclohexyl-2-benzothiazolesulfenamide (DCBS) at 0, 1500, 3000, 6000 or 10 000 p.p.m. (0, 83, 172, 343 or 551 mg/kg bw/day in males and 0, 126, 264, 476 or 707 mg/kg bw/day in females) for a total of 57 days beginning 16 days before mating in males, and a total of 61–65 days from 16 days before mating to day 21 of lactation in females. Body weight gains and food consumption were reduced in males at 6000 p.p.m. and higher and in females at 3000 p.p.m. and higher. The weights of the spleen at 6000 and 10 000 p.p.m. and of the thymus at 10 000 p.p.m. were decreased in females. No changes in estrous cyclicity, copulation index, fertility index, gestation index, delivery index, precoital interval or gestation length were observed at any dose of DCBS. Numbers of implantations at 6000 and 10 000 p.p.m. and pups delivered at 10 000 p.p.m. were reduced. There were no changes in the sex ratio or viability of pups. The body weights of male and female pups were lowered at 6000 p.p.m. and higher. Decreased weight of the spleen in weanlings was also observed in males at 1500 p.p.m. and higher and in females at 3000 p.p.m. and higher. The data indicate that DCBS possesses adverse effects on reproduction and development in rats.

**Key Words:** developmental toxicity, N, N-dicyclohexyl-2-benzothiazolesulfenamide, rat, reproductive toxicity, rubber accelerator

### INTRODUCTION

Sulfenamide accelerator compounds are widely used in the manufacture of automotive compartments and industrial rubber products such as tires, hoses, conveyer belts, bushings seals, gaskets and windshield wiper blades (EPA 2001). N,N-Dicyclohexyl-2-benzothiazolesulfenamide (DCBS, Fig. 1) is a sulfenamide accelerator. The annual production level of DCBS in Japan was approximately 1000 tons in 1990–1993 and 1900 tons in 2000–2003. Most of this amount was sold and handled domestically (OECD 2007). DCBS is used as an accelerator of vulcanization and is completely reacted in the vulcanizing process (OECD 2007). DCBS is regulated in Germany for use in articles that contact food, but is not regulated by the United States Food and Drug Administration for use in food contact applications (Flexsys 2000).

Exposure of workers handling sulfenamide accelerator materials is likely to be highest in the area of materials packaging. During material packout at the manufacturing site, and to a lesser degree during weigh-up activities at the consumer site, there is a possibility of skin and inhalation exposure. Although consumer exposure should be minimal, the most likely route of consumer exposure is skin contact with rubber or latex articles (EPA 2001).

Only up to 6% biodegradation has been determined for DCBS in a ready biodegradability test, and a measured log Kow value of 4.8 suggests that DCBS may have a high bioaccumulation potential (OECD 2007). The possibility of such a chemical compound entering biological systems has aroused great concern regarding its toxicological potential. Generally, biological effects of chemicals should be studied in laboratory animals to investigate their possible influences on human health, and the results of animal tests of chemical toxicity relevant to humans (Clayson & Krewski 1990). However, very little information on the toxicity of DCBS has been published. The toxic effects of DCBS have been briefly summarized by the European Chemical Bureau (2000) and US EPA (2001). It was reported that the oral LD50 values were 1077–10 000 mg/kg bw in rats, the oral NOAEL for 44-day repeated dose toxicity was higher than 100 mg/kg bw/day in rats, and no effects on reproduction were observed at doses up to 400 mg/kg bw/day in rats (EPA 2001). The oral LD50 value was 8500 mg/kg bw in male mice, and repeated daily inhalation exposure of male rats for 15 days at 2 h/day and 350–400 mg/m<sup>3</sup> caused mucous membrane irritation (Vorobera 1969).

The Japanese Government (MHW 1998) conducted toxicity studies for DCBS, including acute toxicity, *in vitro* genotoxicity and repeat dose toxicity combined with reproductive/developmental toxicity as a part of the Safety Examination of Existing Chemical Substances and Chemical Safety Programmes. These toxicity studies are summarized in the IUCLID Data Sets (EPA 2006), OECD Screening Information Data Sets (OECD 2007) and the Hazard Assessment Sheet (CERI 2002). We previously reported the results of a screening test for repeat dose toxicity combined with a reproductive/developmental toxicity in rats, where DCBS at 400 mg/kg bw/day had a deleterious effect on reproduction and development and caused a marked decrease in the number of live pups as well as a total loss of pups by postnatal day (PND) 4 (Ema *et al.* 2007). The primary effects may be on the gestation index for dams and live birth index for pups, both of which appear to be affected at multiple points along the female reproductive process. The viability of neonatal pups may also be affected. To examine the adverse effect of dietary DCBS on survival and growth of pups, a reproductive and developmental toxicity study was performed in rats given DCBS during an extended administration period up to the weaning of pups.

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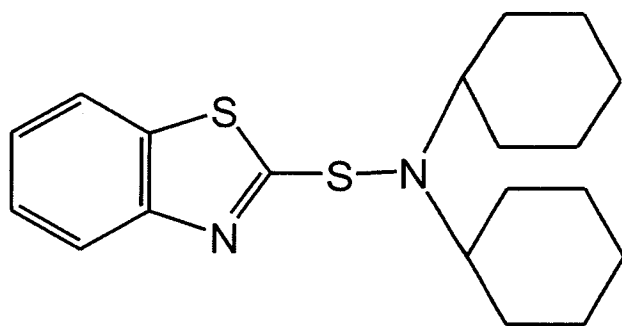


Fig. 1 Structural formula of N,N-dicyclohexyl-2-benzothiazolesulfenamide.

## MATERIALS AND METHODS

This study was performed in 2005–2006 at the Safety Research Institute for Chemical Compounds (Sapporo, Japan) in compliance with *Law for the Humane Treatment and Management of Animals* (Law no. 105, October 1, 1973, revised December 22, 1999, Revised Law no. 221; revised June 22, 2005, Revised Law no. 68), *Standards Relating to the Care, Management and Refinement of Laboratory Animals* (Notification no. 88 of the Ministry of the Environment, Japan, April 28, 2006) and *Fundamental Guidelines for Proper Conduct of Animal Experiment and Related Activities in the Testing Facility under the Jurisdiction of the Ministry of Health, Labour and Welfare* (Notification no. 0601005 of the Health Sciences Division, Ministry of Health, Labour and Welfare, Japan, June 1, 2006).

### Chemical and dosing

DCBS (CAS no. 4979-32-2) was obtained from Ouchishinko Chemical Industrial (Tokyo, Japan). DCBS in the form of off-white to tan granules is very slightly soluble in water and methanol but soluble in oil. Its melting point is 100–105°C, density is 1230 kg/m<sup>3</sup> and molecular weight is 347 (Flexsys 2000). DCBS (Lot no. 508001) used in this study was 99.7% pure and was kept in a sealed container under cool (1–8°C) and dark conditions. The purity and stability of the chemical were verified by analysis using high-performance liquid chromatography before and after the study. Rats were given dietary DCBS at a concentration of 0 (control), 1500, 3000, 6000 or 10 000 p.p.m. Males were fed a diet containing DCBS for a total of 57 days beginning 16 days before mating. Females were fed a diet containing DCBS for a total of 61–65 days from 16 days before mating to day 21 of lactation throughout the mating, gestation and lactation periods. Control rats were fed a basal diet only.

The dosage levels were determined based on the results of a previous study in rats that were given DCBS by gavage at 0, 6, 25, 100, or 400 mg/kg bw/day for a total of 44 days from 14 days before mating in males and a total of 40–51 days beginning 14 days before mating to day 3 of lactation throughout the mating and gestation periods in females (Ema *et al.* 2007). In that study, toxicologically significant changes were observed only at 400 mg/kg bw/day. Three of 10 females died during parturition. An increased incidence of females showing decreased locomotor activity, soil of the lower abdominal fur and reddish tears was observed. Decreased body weights were found in males and females. Decreased weight of the thymus in both sexes was noted. Decreases in the gestation

index, numbers of corpora lutea, implantations, pups born and pups born alive, live birth index and viability index were detected.

Dosed diet preparations were formulated by mixing DCBS into an appropriate amount of a powdered basal diet (CRF-1; Oriental Yeast, Tokyo, Japan) for each dietary concentration. Chemical analysis showed that DCBS in the diet was stable for at least 21 days at room temperature and the formulations were maintained in a room temperature for no more than 21 days. Generally, the diet was replaced once a week.

### Animals and housing conditions

Sprague–Dawley (CrI:CD[SD]) rats were used throughout this study. Rats of this strain were chosen because they are the most commonly used in reproductive and developmental toxicity studies and historical control data are available. Male and female rats at nine weeks of age were purchased from the Tsukuba Breeding Center (Charles River Laboratories Japan, Yokohama, Japan). The rats were acclimated to the laboratory for six days prior to the start of the experiment. Male and female rats found to be in good health were selected for use. Rats (F0) were randomly distributed into five groups of six males and six females each, and all animals were assigned a unique number and tattooed on the ear prior to the start of the experiment. Animals were housed individually in suspended aluminum/stainless steel cages except during the acclimation, mating and nursing periods. From day 17 of pregnancy to the day of weaning, individual dams and litters were reared using wood chips as bedding (White Flake; Charles River Laboratories Japan,).

Animals were reared on a basal diet or a diet containing DCBS and filtered tap water *ad libitum* and maintained in an air-conditioned room at 22 ± 3°C with a humidity of 50 ± 20% and a 12-h light (8:00–20:00)/dark (20:00–8:00) cycle. The room was ventilated 10–15 times/h.

### Observations

All rats were observed twice a day for clinical signs of toxicity. The body weight was recorded once a week for males and once a week during the pre-mating period, on days 0, 7, 14 and 20 of pregnancy, and on days 0, 4, 7, 14 and 21 of lactation for females. Food consumption was recorded once a week for males, and once a week during the pre-mating period, on days 0, 7, 14 and 20 of pregnancy and on days 0, 7, 14 and 21 of lactation for females.

Rats were euthanized by exsanguination under ether anesthesia. Males were euthanized at 17 weeks and females at 18 weeks on day 21 of lactation. The external surfaces of the rats were examined for abnormalities. The abdomen and thoracic cavities were opened and gross internal examination was performed. In females, the number of implantation sites was recorded. The brain, pituitary, thymus, thyroid, liver, kidney, spleen, adrenal gland, testis, epididymis, seminal vesicle, ventral prostate, ovary and uterus were weighed. The thyroid and seminal vesicle were weighed after fixation with 10% neutral buffered formalin.

Daily vaginal lavage samples from each female were evaluated for estrous cyclicity for two weeks of the pre-mating period. Females with repeated 4–6 day estrous cycles were judged to be normal. Each female rat was mated overnight with a single male rat of the same dosage group until copulation occurred. During the mating period, daily vaginal smears were examined for the presence of sperm. The presence of sperm in the vaginal smear and/or a vaginal plug was considered evidence of successful mating (day 0 of pregnancy). Copulated females were checked for signs of parturition three times a day on days 21–23 of pregnancy.

The females were allowed to deliver spontaneously and nurse their pups until PND 21. The day on which parturition was

completed by 13:00 was designated as PND 0. Total litter size and the numbers of live and dead pups were recorded. Live pups were counted, sexed, examined grossly and individually weighed on PND 0, 4, 7, 14 and 21. On PND 4, litters were randomly adjusted to eight pups comprised of four males and four females. No adjustment was made for litters with fewer than 8 pups. Selected pups were assigned a unique number and tattooed on a limb on PND 4. Unselected pups were necropsied on PND 4. Weanlings were necropsied on PND 21 and the brain, thymus, liver, spleen and uterus were weighed.

### Statistical analysis

Statistical analysis of the offspring was carried out using the litter as the experimental unit.

Body weight, body weight gain, food consumption, length of estrous cycle, pre-coital interval, gestation length, number of implantations and pups delivered, delivery index, organ weight, organ/body weight ratio (relative organ weight) and the viability of pups were analyzed for statistical significance in the following way. Bartlett's test of homogeneity of variance was used to determine if the groups had equivalent variances. If the variances were equivalent, the groups were compared by one-way analysis of variance

(ANOVA). If significant differences were found, Dunnett's multiple comparison test was performed. If the groups did not have equivalent variances, the Kruskal-Wallis test was used to assess the overall effects. Whenever significant differences were noted, pairwise comparisons were made by Mann-Whitney *U*-test. The incidence of females with normal estrous cycles, copulation index, fertility index, gestation index and neonatal sex ratio was analyzed by the  $\chi^2$  test or Fisher's exact test.

The 0.05 level of probability was used as the criterion for significance.

## RESULTS

### Clinical observations, body weight and food consumption (F0 males and females)

No deaths were found in F0 males and females. In males, there were no compound related clinical signs of toxicity at any doses. Hematuria and soil of perigenital fur were each observed at 10 000 p.p.m. in one female.

Table 1 shows body weight gain in F0 males and females during dosing. In males, body weight gain on days 0-7 of the dosing period at 6000 p.p.m. and higher was significantly lowered. In females,

**Table 1** Body weight gains of F0 parental male and female rats given N,N-dicyclohexyl-2-benzothiazolesulfenamide

Dose (p.p.m.)	0 (Control)	1500	3000	6000	10 000
No. males	6	6	6	6	6
Initial body weight (g)†	367 ± 7	367 ± 6	366 ± 7	366 ± 8	366 ± 7
Body weight gain during dosing period (g)†					
Days 0-7	48.0 ± 10.4	36.8 ± 14.5	36.3 ± 4.8	26.7 ± 8.5**	25.2 ± 6.5**
Days 7-14	38.2 ± 9.2	33.7 ± 13.4	34.5 ± 8.7	35.2 ± 5.6	29.5 ± 5.5
Days 14-21	21.7 ± 9.2	27.3 ± 7.1	24.3 ± 5.2	23.0 ± 12.6	21.8 ± 3.1
Days 21-28	26.8 ± 10.2	25.7 ± 8.3	22.3 ± 10.5	23.5 ± 6.7	25.2 ± 5.0
Days 28-35	21.2 ± 7.7	20.8 ± 6.5	28.5 ± 12.6	24.0 ± 3.7	19.2 ± 4.1
Days 35-42	14.8 ± 6.9	15.3 ± 6.5	20.3 ± 6.9	17.3 ± 6.3	20.5 ± 5.2
Days 42-49	13.8 ± 8.3	19.5 ± 4.2	13.5 ± 2.7	19.8 ± 5.5	17.2 ± 2.9
Days 49-56	14.8 ± 7.6	19.5 ± 6.3	16.7 ± 5.0	20.5 ± 6.2	17.0 ± 3.5
No. females	6	6	6	6	6
Initial body weight (g)†	238 ± 6	239 ± 7	237 ± 5	238 ± 6	237 ± 7
Body weight gain during pre-mating period (g)†					
Days 0-7	6.5 ± 7.7	8.8 ± 8.8	6.8 ± 4.6	-6.5 ± 9.7*	-19.3 ± 9.3**
Days 7-14	15.7 ± 8.5	16.3 ± 6.9	14.2 ± 5.9	12.2 ± 8.0	13.0 ± 8.7
Body weight gain during pregnancy (g)†					
Days 0-7	45.3 ± 6.5	42.5 ± 4.2	32.8 ± 5.4*	31.2 ± 8.6*	19.5 ± 12.0**
Days 7-14	38.3 ± 6.0	35.7 ± 5.4	36.8 ± 7.0	35.2 ± 6.2	31.2 ± 8.6
Days 14-20	76.7 ± 14.6	68.3 ± 4.3	75.8 ± 12.4	68.7 ± 7.7	62.5 ± 12.2
Body weight gain during lactation (g)†					
Days 0-4	28.0 ± 15.7	10.8 ± 24.3	28.0 ± 15.7	8.2 ± 8.7	-2.5 ± 14.6*
Days 4-7	6.5 ± 2.7	12.0 ± 9.0	10.0 ± 10.2	5.3 ± 7.3	0.5 ± 10.9
Days 7-14	1.3 ± 10.7	10.2 ± 7.3	4.2 ± 8.1	14.2 ± 11.1	6.0 ± 12.5‡
Days 14-21	-19.0 ± 14.7	-31.7 ± 9.9	-17.0 ± 8.3	-8.8 ± 9.8	2.6 ± 14.3‡*

\*Significantly different from the control,  $P < 0.05$ ; \*\*significantly different from the control,  $P < 0.01$ .

†Values are given as mean ± SD; ‡data were obtained from five females because one female was excluded (total litter loss on day 9 of lactation).

body weight gains were decreased on days 0–7 of the pre-mating period at 6000 p.p.m. and higher, on days 0–7 of pregnancy at 3000 p.p.m. and higher, and on days 0–4 of lactation at 10 000 p.p.m. Body weight gain on days 14–21 of lactation was significantly increased at 10 000 p.p.m.

In F0 males, food consumption was significantly decreased during the first week at 6000 p.p.m. and higher and during the second week at 10 000 p.p.m. In F0 females, food consumption was significantly decreased throughout the pre-mating, pregnancy and lactation periods at 6000 and 10 000 p.p.m., except on days 7–14 and 14–20 of pregnancy at 6000 p.p.m. A tendency towards decreased food consumption was observed on days 0–7 of pregnancy at 3000 p.p.m.

The mean daily intakes of DCBS were 83, 172, 343 and 551 mg/kg bw in F0 males, and 126, 264, 476 and 707 mg/kg bw in F0 females for 1500, 3000, 6000 and 10 000 p.p.m., respectively.

#### **Estrous cyclicity (F0 females)**

All F0 females showed normal estrous cycles in all groups, and the length of the estrous cycles was not significantly different between the control and DCBS-treated groups.

#### **Reproductive and developmental effects (F0 parents/F1 offspring)**

The reproductive and developmental parameters for F0 parents/F1 offspring are presented in Table 2. In F0 parent animals in all groups, all pairs copulated, all male and female rats were fertile and all females delivered live pups. All rats of all groups mated within four days. There were no significant differences between control and DCBS-treated groups in copulation index, fertility index, gestation index, pre-coital interval, gestation length, delivery index, sex ratio of F1 pups, or viability of F1 pups during lactation. Significantly lower numbers of implantations at 6000 and 10 000 p.p.m. and pups delivered at 10 000 p.p.m. were observed. Body weights of male pups were significantly lowered on PND 4, 7 and 21 at 6000 p.p.m. and on PND 7, 14 and 21 at 10 000 p.p.m. In female pups, significantly lower body weights were observed on PND 7, 14 and 21 at 6000 p.p.m. and higher. No malformed pups were detected in any groups.

#### **Necropsy and organ weights (F0 males and females)**

Atrophy of the thymus was found in two females at 10 000 p.p.m. No compound-related gross lesions of the reproductive organs were noted in F0 males and females. In males, significantly increased relative weights of the liver and kidney were observed at 10 000 p.p.m.

The organ weights of F0 females are shown in Table 3. The body weight at the scheduled terminal sacrifice was significantly lowered at 6000 and 10 000 p.p.m. The absolute weight of the ovary was significantly lowered at 10 000 p.p.m. Significantly increased relative weights were found for the pituitary at 3000 p.p.m., the liver at 6000 p.p.m., and the brain, kidney and adrenal gland at 10 000 p.p.m. The absolute and relative weights of the thymus at 10 000 p.p.m. and the spleen at 6000 p.p.m. and higher were significantly decreased.

#### **Necropsy and organ weights (F1 weanlings)**

No compound related gross lesions were observed in F1 weanlings.

The organ weights of F1 male weanlings are presented in Table 4. The body weight at the scheduled sacrifice was significantly reduced at 6000 and 10 000 p.p.m. The absolute weights of the brain at 6000 and 10 000 p.p.m. and the liver at 10 000 p.p.m. were also significantly reduced. The relative weights of the liver at

1500 and 6000 p.p.m. and of the brain at 10 000 p.p.m. were significantly increased. Significantly decreased absolute and relative weights of the spleen, except for the relative weight at 3000 p.p.m., were noted at 1500 p.p.m. and higher.

The organ weights of F1 female weanlings are presented in Table 5. The body weight at the scheduled sacrifice was significantly reduced at 6000 p.p.m. and higher. Significantly reduced absolute weights of the brain at 6000 and 10 000 p.p.m., the liver at 10 000 p.p.m., and the uterus at 3000 p.p.m. and 10 000 p.p.m. were also observed. The relative weight of the brain was significantly increased at 10 000 p.p.m. The absolute and relative weights of the spleen were significantly reduced at 3000 p.p.m. and higher.

## **DISCUSSION**

This study was designed to assess the effects of DCBS on continuous parameters such as body weight and food consumption, as well as endpoints for reproductive and developmental toxicity.

Significant decreases in body weight gain and food consumption were observed at 6000 p.p.m. and higher in F0 males and females. In females at 3000 p.p.m., body weight gain was significantly decreased during early pregnancy. Food consumption also decreased, but not significantly. The data indicate that changes in body weight gain were associated with changes in food consumption and that DCBS adversely affects body weight gain and food consumption at 6000 p.p.m. in male rats and 3000 p.p.m. in female rats. The higher relative weights of the liver and kidney at the highest dose in F0 males seem to be due to secondary effects of lowered body weight rather than direct effects of DCBS on the organs. More pronounced effects on organ weights were noted in females. Lower absolute and relative weights of the thymus at 10 000 p.p.m. and spleen at 6000 p.p.m. and higher were detected. In our previous study, histopathological examination revealed atrophy of the thymus and spleen at 400 mg/kg bw/day (Ema *et al.* 2007). Other changes in female organ weight such as the relative weights of the brain, pituitary, liver, kidney and adrenal gland, as well as the absolute weight of the ovary are unlikely to be due to the toxic effects of DCBS because the degree of changes was relatively small, no dose-dependency was shown and no changes were noted in absolute or relative weight. These findings suggest that the immune system may be a target of DCBS toxicity, and that female rats have a higher susceptibility to the toxicity of DCBS than male rats. These findings are consistent with our previous study (Ema *et al.* 2007). The higher susceptibility to DCBS toxicity in females may be explained by the stress of pregnancy and lactation. DCBS is likely to be not reproductively toxic in male rats because DCBS caused neither pathological changes in male reproductive organs nor changes in male reproductive parameters.

In our previous study, DCBS given by gavage to rats at 400 mg/kg bw/day from 14 days before mating to day 3 of lactation caused significant decreases in the gestation index, number of corpora lutea, implantations, pups born and pups born alive, live birth index and viability index (Ema *et al.* 2007). This dose also caused severe maternal toxicity and a total loss of pups by PND 4. No maternal or reproductive/developmental toxicity was detected at 100 mg/kg bw/day in our previous study. In the present study, no serious reproductive difficulties were noted even at the highest dose of 10 000 p.p.m., and necropsy of the reproductive organs revealed no evidence of reproductive failure. Although decreased numbers of implantations and pups delivered were noted at the highest dose, the viability of pups until weaning was not significantly decreased. In the present feeding study, the mean daily intakes of DCBS at the