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JaCVAM
1st International Workshop
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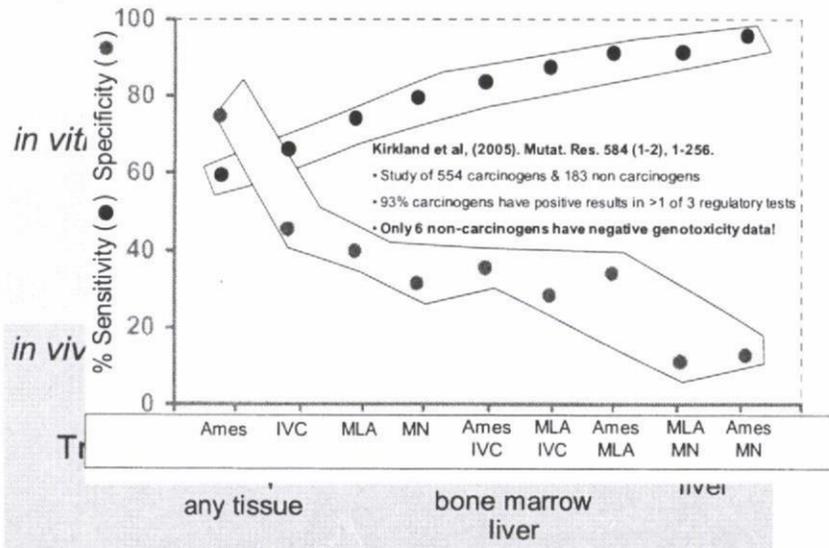
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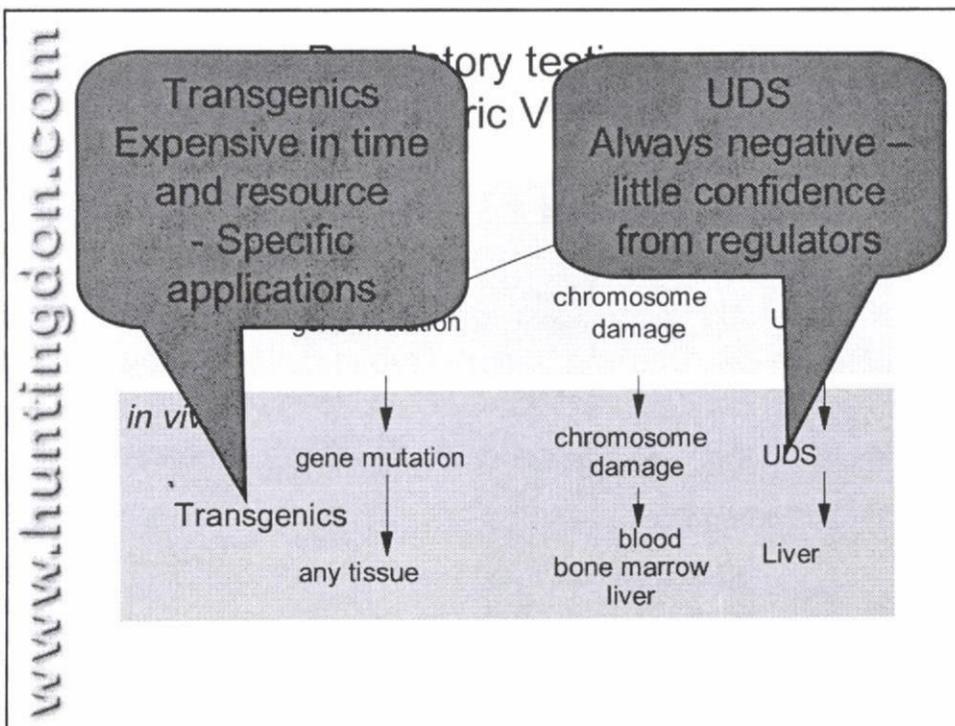
Content

- The Comet Assay – an overview
 - Past
 - Present
 - Future (?)

Is there a place for the Comet assay in the current test battery?

- Battery is there to identify hazard
- In vitro – do we need another assay?
 - General consensus was probably not
- In vivo – do we need another assay
 - General consensus is yes we do



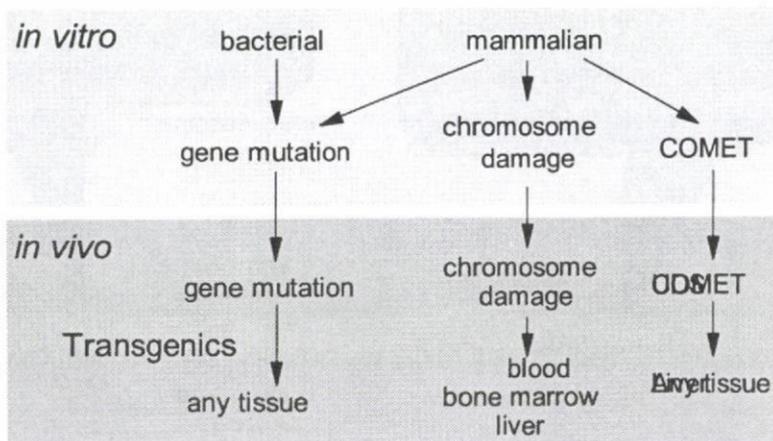


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So what about the COMET assay?

- Is it more specific than the other assays
 - In vivo? (11/13 non-carc were negative)
 - In vitro?
- In both cases we are in a position to take advantage of what we have learned about the other assays - especially sensible levels for the top dose

Regulatory testing: The theoretical ideal



Drug candidates with genotoxicity data sets triggering comet assay in vivo					
Drug	Available Results	Tissue(s) Comet assay	Comet Assay	Consequence for compound	
A	Ames	-	Liver, leukocytes (oral 3 and 24-hour sampling)	negative	ok from HA / EC for multiple dose clinical studies in patients
	MNT V79	-			
	CAT V79	+			
	CAT HuLy	-			
	MNT BM rat	-			
B	Ames	-	Liver (oral; 3 and 24-hour sampling)	negative	ok from HA/ EC for first administration to humans
	MNT V79	-			
	Comet HuLy	-			
	CAT V79	+			
	MNT BM rat	-			
C	Ames	+/-	Liver, leukocytes (sub-cut; 3 and 24-hour sampling)	negative	ok from HA / EC for first (topical) administration to humans
	MNT V79	+			
	CAT V79	+			
	CAT HuLy	+/-			
	MLA TK	+			
	HPRT CHO	-			
	Comet V79	+			
	MNT BM mouse	-			
	UDS liver rat	-			

Examples of drug candidates with tumour findings triggering comet assay in vivo

	Available results	Tumor target organ	Tissue and time Comet assay	Result of Comet Assay	Conclusion
D	Ames - MNT V79 - CAT V79 - HPRT V79 - MNT BM mouse - UDS liver rat -	Small intestine Rat	Jejunum, Liver (oral admin.) 3 and 24 hour sampling	negative	No unacceptable health risk (non-genotoxic mechanism, epigenetic mechanism)
E	Ames - MNT V79 - Comet V79 - CAT CHO - HPRT V79 - HPRT CHO - MNT BM rat -	Liver Rat	Liver (oral 3 and 24 hour sampling)	Negative	No unacceptable health risk (non-genotoxic mechanism, epigenetic mechanism)

Examples of drug candidates with data of comet assay in vivo using tissue of site of first contact

Drug	Available results	Tissue(s) investigated	Result	Outcome
F	Ames +/- MNT V79 (+) CAT HuLy - MNT BM mouse - DNA binding rat - MutaMouse -	Jejunum, liver (oral; 3 and 24-hour sampling)	negative	ok from health authorities to proceed into multiple dose clinical trials
G	Ames - MNT V79 + Comet V79 + CAT HuLy + MNT BM rat -	Stomach mucosa (oral; sampling time 3 hours)	Retarded DNA migration (reason: DNA-protein crosslinks)	Termination of development

Other Comments

- Lack of agreed criteria for collection, presentation and analysis of results
 - Uncertainty about toxicity
 - Large volume of published data – lots of false “positives”

 - But assay accepted in EU
- Andrew Smith (UK HSE)

- Large number of procedures (protocols)
- Variance in presenting results
- Little regulatory experience in evaluation.
- Common for “new” assays

Jon Battershill (DoH UK)

- Has important potential but many technical shortcomings
- Considered a tier 2 test for following up ICH battery or
- To exclude genotoxic MOA in positive carcinogenicity studies

David Jacobson-Kram FDA

JaCVAM trial

- It is clear that a lot of the concerns could be removed once there is an accepted OECD guideline
- The JaCVAM trial is aiming towards that goal
- International committee should ensure fast track through to acceptance

Summary of major points of the revisions

- S2A and S2B guidances merged into one
- Options provided for the test battery
 - Battery with in vitro mammalian cell assay
 - Battery without in vitro mammalian cell assay but two in vivo endpoints
- In vitro mammalian cell assay
 - Reduction in top concentration from 10 mM to 1 mM
 - Tightened acceptable cytotoxicity limits
 - No longer require testing of precipitating concentrations
- In vitro bacterial mutation assay no longer requires duplicate assay

Summary of major points of the revisions, continued

- Integration of genotoxicity endpoints into routine toxicology studies
 - Stringent criteria defined for acceptability of top dose
- Advice on choice of second in vivo genotoxicity endpoint
 - includes Comet assay, decreases emphasis on UDS assay)
- Provided advice on weight of evidence and data evaluation to determine relevance of positive findings

Timing of genotoxicity studies for Phase 1

- Option 1; two vitro tests only
 - in the past this led to many in vivo assays before FIM, to follow up positive in vitro results
- Option 2: one in vitro test (Ames) and two in vivo endpoints, preferably integrated into toxicology study

Benefits of revisions: The 3 R's

- No longer require concurrent positive controls in every in vivo assay
- Integration of genotoxicity into toxicology assays
- Reduction in “non-relevant” in vitro results will reduce number of follow-up in vivo assays

Benefits of revisions:

- Incorporates accumulated knowledge specific to testing of pharmaceuticals
- Takes advantage of new technologies
- More options in the test battery
- Reduction in delays caused by dealing with “non-relevant” in vitro positive genotoxicity results
- More efficient use of resources

Expected targets

- Finalization of text of draft guideline
- Ensure ultimate acceptance by groups represented by EWG
- Postal sign off for step 2
 - end December 2007
- Publication in the regions and regional consultation period
- Step 4 in June 2008 in Portland pending speed of Federal Register publication and comment period

HLS JaCVAM Data

- 3 unknown compounds supplied by JaCVAM panel

Tissues to date!

Tissue Type	Number of Males		Number of Females		Total Number	
	Vehicle	Positive	Vehicle	Positive	Vehicle	Positive
Bone Marrow	18	15	13	11	31	26
Liver	46	48	14	12	60	60
Stomach	40	41	5	4	45	45
Peripheral Blood	22	21	3	3	25	24
Kidney	23	19	5	5	28	24
Duodenum	17	13	5	5	22	18
Colon	18	13	5	4	23	17
Ileum	15	14	4	4	19	18
Jejunum	9	8	3	3	12	11
Testes	13	11			13	11
Lung	4	3	3	3	7	6
Bladder	3	1	1	1	4	2
Mammary Gland			2	2	2	2
Ovaries			1	1	1	1
			Number of Tissues		292	265
			Total Number of Tissues		557	

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Liver		Original Results	Repeated Results
Compound 7			
(%) Tail intensity fold increase over vehicle:		5	3
Tail Moment fold increase over vehicle:		7	3
Tail Length fold increase over vehicle:		2	1
Compound 8			
(%) Tail intensity fold increase over vehicle:		4	3
Tail Moment fold increase over vehicle:		5	3
Tail Length fold increase over vehicle:		2	2
Compound 9			
(%) Tail intensity fold increase over vehicle:		3	3
Tail Moment fold increase over vehicle:		5	3
Tail Length fold increase over vehicle:		2	1

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Stomach		Original Results	Repeated Results
Compound 7			
(%) Tail intensity fold increase over vehicle:		3	3
Tail Moment fold increase over vehicle:		3	3
Tail Length fold increase over vehicle:		2	1
Compound 8			
(%) Tail intensity fold increase over vehicle:		2	2
Tail Moment fold increase over vehicle:		2	3
Tail Length fold increase over vehicle:		2	2
Compound 9			
(%) Tail intensity fold increase over vehicle:		2	2
Tail Moment fold increase over vehicle:		2	2
Tail Length fold increase over vehicle:		1	2

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Bone Marrow

Compound 7

(%) Tail intensity fold increase over vehicle:	2
Tail Moment fold increase over vehicle:	2
Tail Length fold increase over vehicle:	1

Compound 8

(%) Tail intensity fold increase over vehicle:	2
Tail Moment fold increase over vehicle:	3
Tail Length fold increase over vehicle:	2

Compound 9

(%) Tail intensity fold increase over vehicle:	2
Tail Moment fold increase over vehicle:	2
Tail Length fold increase over vehicle:	1

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Kidney

Compound 7

(%) Tail intensity fold increase over vehicle:	6
Tail Moment fold increase over vehicle:	9
Tail Length fold increase over vehicle:	2

Compound 8

(%) Tail intensity fold increase over vehicle:	3
Tail Moment fold increase over vehicle:	4
Tail Length fold increase over vehicle:	1

Compound 9

(%) Tail intensity fold increase over vehicle:	5
Tail Moment fold increase over vehicle:	8
Tail Length fold increase over vehicle:	2

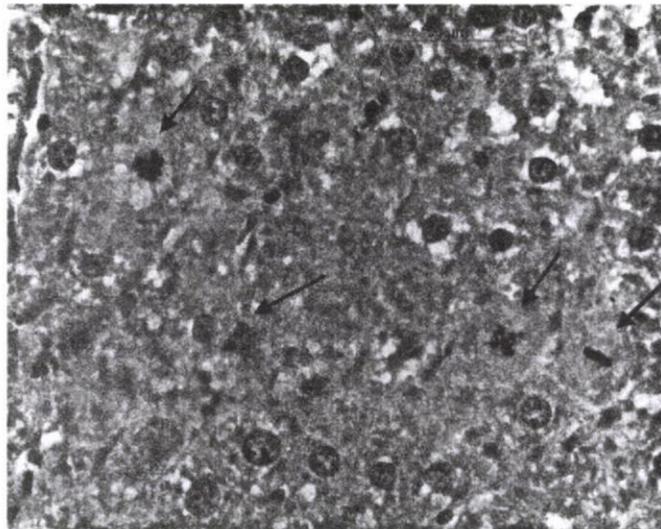
Colon

Compound 7

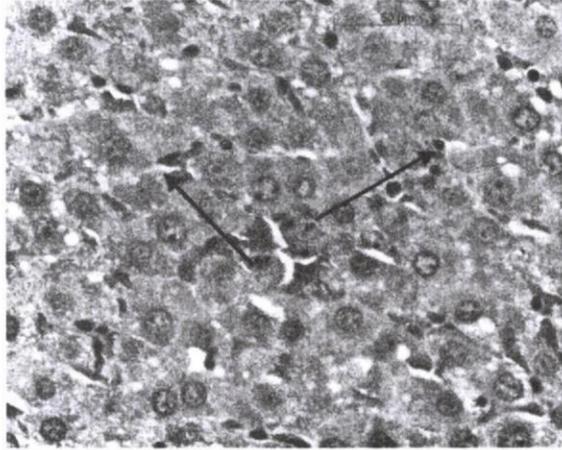
(%) Tail intensity fold increase over vehicle:	4
Tail Moment fold increase over vehicle:	5
Tail Length fold increase over vehicle:	2

Compound 8

(%) Tail intensity fold increase over vehicle:	3
Tail Moment fold increase over vehicle:	3
Tail Length fold increase over vehicle:	2



Animal No. 233 (Compound 7) Liver
Increased numbers of mitotic figures
(mitotic figures indicated by arrows).



Animal No. 432 (Compound 9) Liver
Increased apoptosis (apoptotic cells
indicated by arrows).

- Thank you and any questions?

JaCVAM 1st Int. Workshop
March 11, 2008; Tokyo

***In Vivo* Comet Assay:
Update on the On-Going Validation
Coordinated by JaCVAM**

**Yoshifumi Uno, D.V.M., Ph.D.
Mitsubishi Tanabe Pharma Co.
JEMS/MMS**

Introduction

An *in vivo* rodent alkaline Comet assay is practically used worldwide for detecting genotoxic chemicals, and it is expected as a second *in vivo* genotoxicity test in the revised ICH-S2 guidance.

The assay, however, has not been validated formally, and the international validation study is now on-going coordinated by JaCVAM. The purposes are to validate the *in vivo* Comet assay as a potential predictor of carcinogens and as an alternative follow-up assay to the more commonly used *in vivo* rodent UDS assay.

Organization

Validation Management Team (VMT)

M. Hayashi (Chair, JaCVAM/NIHS)
R. Corvi (ECVAM)
M. Honma (NIHS)
L. Schechtman (Consultant)
R. Tice (NIEHS)
Y. Uno (MTPC, JEMS/MMS)

Secretariat

H. Kojima (JaCVAM/NIHS)

Leading laboratory

Anpyo-Center (M. Nakajima, et al.)
BioReliance (formerly P. Escobar, et al.)
FDSC (K. Yamakage, et al.)
HLS (B. Burlinson, et al.)
Merck (R.D. Storer, et al.)

Consultation Team

N. Asano (Nitto Denko, JEMS/MMS)
D. Lovell (Univ. of Surrey)
T. Morita (NIHS)
N. Nakashima (PMDA)
Y. Ohno (JaCVAM/NIHS)
T. Omori (Kyoto Univ.)
YF. Sasaki (Hachinohe Nat.Coll.Tech.)

Local Committee in JPN

Mainly from JEMS/MMS members

Outline of *in vivo* Comet Assay Validation

The 1st Phase Validation Study on *in vivo* Comet Assay – almost finished

- **Purpose:** Optimization of study protocol
- **Test comp:** Ethyl methanesulfonate (EMS)
- **Solvent:** Physiological saline
- **Dose:** 0, 100, 200 mg/kg, p.o.
- **Animal:** CD(SD) rat, male, 5 animals/group
- **Administration:** Twice (21 or 24 hours interval)
- **Organ tested:** Liver, Glandular stomach
- **Cell preparation:** 3 hours after 2nd administration,
mincing for liver, scraping for stomach
- **Electrophoresis:** 300 mA, 15-40 min, below 10°C (2-20°C)
- **Image analysis:** Use of Comet IV, SYBR Gold staining
- **Cell observed:** 50 cells/slide, duplicate

The 2nd Phase Validation Study on *in vivo* Comet Assay – ongoing

- **Purpose:**
 - ✓ Investigate intra-/inter-laboratory validation with EMS
 - ✓ Decide acceptable variation criteria on Effect for the 3rd phase (main) validation studies
- **Test comp:** Three coded chemicals and EMS,
Each exp. for coded chemicals should include an EMS group, and thus
3 data of EMS/Lab. X 5 Labs. = 15 data available, then analyze validity
- **Solvent:** Directed by VMT
- **Dose:** One of three chemicals was directed by VMT,
Other two was decided in each laboratory
- **Others:** Same as the 1st phase validation study

The 3rd Phase (main) Validation Study on *in vivo* Comet Assay – now planning

- **Purpose:** Investigate predictive capacity of carcinogens
- **Test comp:** Coded “the number of X” chemicals
- **Participant:** 5 leading lab + selected lab*
 - * 15 lab hope to join, and selection process is ongoing
- **Method:** In accordance with our standard protocol
- **Schedule:** Begin at 3Q or 4Q/2008 (or 1Q/2009?)
Finish by the end of 2010 (tentative)

◆ **In addition...**

we will mix up existing data to evaluate the assay capacity

Steps of *in vivo* Comet Validation Studies (Based on ECVAM Validation Process)

- ✓ **Test definition**
Protocol optimization, Training ... 1st phase validation
- ✓ **Within-lab. variability**
3 exp. with EMS in each lab. ... 2nd phase validation
- ✓ **Transferability**
May be assessed with data from candidate facilities because they have done exp. based on our standard protocol without additional information
- ✓ **Between-lab. variability**
 - EMS data from 5 lab. ... 1st & 2nd phase validation
 - 3 coded chem. data from 5 lab. ... 2nd phase validation
- ✓ **Predictive capacity**
Exp. with “X” coded chem. in “Y” lab. ... 3rd phase (main) validation
- ✓ **Reproducibility**
Will be evaluated with data from 1st & 2nd (& 3rd?) phase validation

- ✓ **Applicability domain:** to be discussed
- ✓ **Minimum performance standards:** to be discussed
- ✓ **Independent peer review:** to be discussed

Blue: almost finished, Green: ongoing. Red: now planning

Data from Validation Study

Data in the 1st Phase Validation Study on *in vivo* Comet Assay

- Parameter: %DNA in tail, Tail length, Olive tail moment
- Data analysis:

Three key terms for data analysis are

- ✓ Endpoint: individual values for each parameter
- ✓ Estimate: a mean or median calculated with values of Endpoint in each animal
- ✓ Effect: difference (or ratio) of an average of Estimate between a control group and a treatment group.

Purpose of validation study analysis is to investigate how large variation exists among data from testing facilities, and Effect would be a preferable yardstick (criterion) to understand the variation of parameters among testing facilities.