「乱用薬物による神経毒性・依存症に対する診断・予防及び治療法」

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乱用薬物による神経毒性・依存症に対する診断・予防及び治療法に関する研究

平成19年度 総括研究報告書 別 冊

Establishment of the Methods of Diagnosis, Therapy and Prophylaxis for Dependence and Neurotoxicity induced by Drugs of Abuse.

REPRINTS BOOK

Annual Report

Research on Pharmaceutical and Medical Safety
Supported by Grant from the Ministry of Health, Labour and Welfare,
Japan in 2007

(Chief; Toshitaka Nabeshima)

平成 20 年 3 月

主任研究者 鍋島俊隆

総括報告書をまとめるに当たり、本年度の研究成果報告として分担研究者の皆さまより、 発表論文や刊行物の別刷りをお預かりしました。

150 報を超える数の報告が集まりましたが、その中でも特に本研究の成果として顕著であり、公にひろくお伝えしたい発表について、別冊の形にまとめました。

本来であればすべての成果をご報告申し上げるところではございますが、予算が十分ではないので、その中のいくつかを選ばせていただきました。巻末に全論文のタイトル一覧を掲載しております。

我々の成果を新しい知見への萌生としていただければ本懐に存じます。

平成20年4月吉日

2. 平成19年度総括報告 別冊

<u>乱用薬物による神経毒性・依存症に対する診断・予防及び治療法に関する研究</u>
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(名城大学薬学部)
Yan, Y., Yamada, K., Niwa, M., Nagai, T., Nitta, A. and Nabeshima, T.·····1
Enduring vulnerability to reinstatement of methamphetamine-seeking behavior in glial cell line-derived
neurotrophic factor mutant mice.
FASEB J. 21 1994-2004 (2007)
Mizoguchi, H., Yamada, K., Niwa, M., Mouri, A., Mizuno, T., Noda, Y., Nitta, A., Itohara, S., Banno, Y. and
Nabeshima, T.······12
Reduction of methamphetamine-induced sensitization and reward in matrix metalloproteinase-2 and -9
deficient mice. J. Neurochem. 100, 1579-1588 (2007)
J. Wedi Ochem. 100, 1010 1000 (2001)
Wang, D., Noda, Y., Zhou, Y., Nitta, A., Furukawa, H. and Nabeshima, T.·····22
Synergistic effect of galantamine with risperidone on impairment of social interaction in phencyclidine-treated
mice as a schizophrenic animal model.
Neuropharmacology, 52, 1179-1187 (2007)
Wang, D., Noda, Y., Zhou, Y., Nitta, A., Furukawa H. and Nabeshima, T.·····31
Synergistic effect of combined treatment with risperidone and galantamine on phencyclidine-induced
impairment of latent visuospatial learning and memory: role of nAChR activation-dependent increase of
dopamine D1 receptor-mediated neurotransmission.
Neuropharmacology, 53, 379-389 (2007)
乱用薬物への渇望(薬物検索行動)の再燃の脳内機序解明およびその治療薬開発に関する研究
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脳内 endocannabinoid の異常性に基づくマウスの知的機能障害
が ferrus (2007)
2 4 1311 pm 2 1 1300 01 (2001)

分担研究者:鈴木 勉	
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M. Narita, Y. Nagumo, M. Miyatake, D. Ikegami, F.	K. Kurahashi and T. Suzuki·····44
Implication of protein kinase \boldsymbol{C} in the orexin-induce	ed elevation of extracellular dopamine levels and rewarding
effect.	
	Eur. J. Neurosci. 25,1537–1545 (2007)
乱用薬物に共通の依存症発症因子に対する新規(Ca ²⁺ 動態調節剤の治療への応用
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	, and Ohkuma S.····53
Up-regulation of L-type high voltage-gated calcium	n channel subunits by sustained exposure to 1,4- and
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	J Neurochem. 103, 2518–2528 (2007)
Shibasaki M, Katsura M, Torigoe F, Honda T, Sun	nimoto A, Tsujimura A, and Ohkuma S.·····64
Increase in diazepam binding inhibitor expression b	y sustained morphine exposure is mediated via m-opioid
receptors in primary cultures of mouse cerebral con	rtical neurons. <i>J Neurosci Res.</i> , 85, 2971–2980,(2007)
乱用薬物に共通の治療薬の開発ならびに新規標的	遺伝子の検討
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	to, K., Seishima, M., Shen, L., Noda, Y., Furukawa, S. and
Nabeshima, T.····	74
An inducer for glial cell line–derived neurotrophic $\boldsymbol{\theta}$	actor and tumor necrosis factor- $lpha$ protects against
methamphetamine-induced rewarding effects and se	ensitization.
	Biol. Psychiatry 61, 890-901 (2007)
Niwa, M., Nitta, A., Yamada, Y., Nakajima, A., Sa	ito, K., Seishima, M., Noda, Y. and Nabeshima, T.····86
Tumor necrosis factor- α and its inducer inhibit m	orphine-induced rewarding effects and sensitization.
	Biol. Psychiatry 62, 658-668(2007)
Niwa, M., Nitta, A., Mizoguchi, H., Ito, Y., Noda,	Y., Nagai, T. and Nabeshima, T97
A novel molecule 'shati' is involved in methamphe	tamine-induced hyperlocomotion, sensitization, and
conditioned place preference.	J. Neurosci. 27,7604-7615 (2007)

乱用薬物による依存形成における視床下部神経ペプチドの役割

依存性薬物の精神神経毒性と標的分子への作用機序の解明
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Uhl, GR., Drgon, T., Liu, QR., Johnson, C., Walther, D., Ujike, H., Komiyama, T., Harano, M., Sekine,
Y., Inada, T., Ozaki, N., Iyo, M., Iwata, N., Yamada, M., Sora, I., Chen, CK., Liu, HC. and Lin, SK.
109
Genome-wide association for methamphetamine dependence: convergent results from two samples.
Arch Gen Psychiatry. in press
Kishimoto M, Ujike H, Motohashi Y, Tanaka Y, Okahisa Y, Kotaka T, Harano M, Inada T, Yamada M,
Komiyama T, Hori T, Sekine Y, Iwata N, Sora I, Iyo M, Ozaki N, Kuroda S120
The dysbindin gene (DTNBP1) is associated with methamphetamine psychosis.
Biol Psychiatry. 63,191–196(2008)
The glycine transporter 1 gene (GLYT1) is associated with methamphetamine-use disorder. Am J Med Genet B Neuropsychiatr Genet.5;147(1)54-58(2007)
Fukushima S, Shen H, Ikeda K, Hata H, Ohara A, Ohmi K, Kobayashi H, Numachi Y, Hall F, Uhl GR, Sora I
Methamphetamine-induced locomotor activity and sensitization in dopamine transporter and vesicular
monoamine transporter 2 double mutant mice. <i>Psychopharmacology</i> , 193(1), 55-62, (2007)
<u>覚せい剤投与によるドパミン神経傷害に及ぼす mithramycin の効果</u> 分担研究者:伊豫雅臣 (千葉大学大学院医学研究院・精神医学,)
Hashimoto K, Ishima T, Fujita Y, Matsuo M, Kobashi T, Takahagi M, Tsukada H, Iyo M139
Phencyclidine-induced cognitive deficits in mice are improved by subsequent subchronic administration of the
novel selective a7 nicotinic receptor agonist SSR180711.
Biol Psychiatry. 63,92–97(2008)
Hashimoto K, Tsukada H, Nishiyama S, Fukumoto D, Kakiuchi T, Iyo M.······147
Protective effects of minocycline on the reduction of dopamine transporters in the striatum after

Biol Psychiatry. 61,577-581(2007)

administration of methamphetamine: A PET study in conscious monkeys.

Phencyclidine-induced cognitive deficits in mice are improved by subsequent subc	152
Phencyclidine-induced cognitive deficits in mice are improved by subsequent subclique fluvoxamine: role of sigma-1 receptors.	
	acology 32,514-521(2007)
乱用薬物による易再発性精神病様状態および依存症の予防・治療法開発に関	する研究
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Kaneko Y, Kashiwa A, Ito T, Ishii S, Umino A, Nishikawa T	160
Selective serotonin reuptake inhibitors, fluoxetine and paroxetine, attenuate thee:	
behavioral sensitization induced by methamphetamine.	
	acol 32,658-664(2007)
Numachi Y, Yoshida S, Yamashita M, Fujiyama K, Toda S, Matsuoka H, Kajii Y,	Nishikawa T.····167
Altered EphA5 mRNA expression in rat brain with a single methamphetamine trea	
-	Lett. 424, 116-121 (2007)
薬物依存における薬物再使用危険度評価尺度の開発と候補治療薬の探索	
—————————————————————————————————————	
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池田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記 祐士, 池田和彦, 天野直二, 五味渕隆志, 斎藤正彦編)	
池田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記 祐士, 池田和彦, 天野直二, 五味渕隆志, 斎藤正彦編)	
池田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記 祐士, 池田和彦, 天野直二, 五味渕隆志, 斎藤正彦編)	念論文集(坂口正道, 岡崎 (<i>/書店,東京</i> ,135-141(2007)
池田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位:松下正明先生古稀記 祐士,池田和彦,天野直二,五味渕隆志,斎藤正彦編) 中山	念論文集(坂口正道, 岡崎 / <i>書店,東京</i> ,135-141(2007)
 池田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記 祐士, 池田和彦, 天野直二, 五味渕隆志, 斎藤正彦編) 中止 Takamatsu Y, Yamamoto H, Hagino Y, Markou A, Ikeda K. The selective serotonin reuptake inhibitor paroxetine, but not fluvoxamine, decre 	念論文集(坂口正道, 岡崎 / <i>書店,東京</i> ,135-141(2007)
 池田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記祐士,池田和彦,天野直二,五味渕隆志,斎藤正彦編) 中止 Takamatsu Y, Yamamoto H, Hagino Y, Markou A, Ikeda K. The selective serotonin reuptake inhibitor paroxetine, but not fluvoxamine, decre conditioned place preference in mice. 	念論文集(坂口正道, 岡崎 / <i>書店,東京</i> ,135-141(2007)
連田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記 祐士, 池田和彦, 天野直二, 五味渕隆志, 斎藤正彦編) 「Takamatsu Y, Yamamoto H, Hagino Y, Markou A, Ikeda K	念論文集(坂口正道, 岡崎 (書店,東京,135-141(2007)) ———————————————————————————————————
 池田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記祐士,池田和彦,天野直二,五味渕隆志,斎藤正彦編) <i>中山</i> Takamatsu Y, Yamamoto H, Hagino Y, Markou A, Ikeda K. The selective serotonin reuptake inhibitor paroxetine, but not fluvoxamine, decreconditioned place preference in mice. 	念論文集(坂口正道, 岡崎 (書店,東京,135-141(2007)) ———————————————————————————————————
施田和隆(書籍) 薬物依存における再使用抑制薬の探索精神医学の方位: 松下正明先生古稀記祐士,池田和彦, 天野直二, 五味渕隆志, 斎藤正彦編) 「中山、 Takamatsu Y, Yamamoto H, Hagino Y, Markou A, Ikeda K The selective serotonin reuptake inhibitor paroxetine, but not fluvoxamine, decreconditioned place preference in mice. A Ogai Y, Haraguchi A, Kondo A, Ishibashi Y, Umeno M, Kikumoto H, Hori T, Kon Asukai N, Senoo E, Ikeda K Development and validation of the stimulant relapse risk scale for drug abusers in	念論文集(坂口正道, 岡崎 /書店,東京,135-141(2007)

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依存性薬物の分子標的としてのモノアミントランスポーター
<i>日本薬理学雑誌</i> 130,450-454(2007)
薬物依存の新たな治療開発のための候補分子の探索 Dysbindin 遺伝子の解析
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Inada T, Ozaki N, Iwata N, Harano M, Komiyama T, Yamada M, Sora I, Ujike H, Hata A, Sawa A, Iyo M.
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Identification of functional polymorphisms in the promoter region of the human PICK1 gene and their
association with methamphetamine psychosis.

Am J Psychiatry. 164,1105·1114 (2007)

Enduring vulnerability to reinstatement of methamphetamine-seeking behavior in glial cell line-derived neurotrophic factor mutant mice

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Genetic factors are considered to play ABSTRACT an important role in drug dependence/addiction including the development of drug dependence and relapse. With the use of a model of drug self-administration in mutant mice, several specific genes and proteins have been identified as potentially important in the development of drug dependence. In contrast, little is known about the role of specific genes in enduring vulnerability to relapse, a clinical hallmark of drug addiction. Using a mouse model of reinstatement, which models relapse of drug-seeking behavior in addicts, we provide evidence that a partial reduction in the expression of the glial cell line-derived neurotrophic factor (GDNF) potentiates methamphetamine (METH) self-administration, enhances motivation to take METH, increases vulnerability to drug-primed reinstatement, and prolongs cue-induced reinstatement of extinguished METH-seeking behavior. In contrast, there was no significant difference in novelty responses, METH-stimulated hyperlocomotion and locomotor sensitization, food-reinforced operant behavior and motivation, or reinstatement of food-seeking behavior between GDNF heterozygous knockout mice and wild-type littermates. These findings suggest that GDNF may be associated with enduring vulnerability to reinstatement of METH-seeking behavior and a potential target in the development of therapies to control relapse.-Yan, Y., Yamada, K., Niwa, M., Nagai, T., Nitta, A., Nabeshima, T. Enduring vulnerability to reinstatement of methamphetamine-seeking behavior in glial cell line-derived neurotrophic factor mutant mice. FASEB J. 21, 1994-2004 (2007)

Key Words: GDNF mutant mice · METH self-administration · relapse

GENETIC FACTORS ARE CONSIDERED TO PLAY an important role in drug dependence/addiction and alcoholism (1-7). In animal models, vulnerability to self-administration and reinstatement in the taking of different addictive substances has been suggested to share common genetic determinants (8). By using a

model of drug self-administration in mutant mice, several specific genes or proteins have been identified as potentially involved in the development of drug dependence (9–14). However, a good model of relapse in mutant mice has yet to be established. Thus, few lines of direct evidence have been obtained for an association between specific genes and vulnerability to relapse of drug-seeking behavior, which is a major challenge in the clinical treatment of addiction (15, 16).

Glial cell line-derived neurotrophic factor (GDNF) was originally purified from a rat glioma cell-line supernatant as a trophic factor for embryonic midbrain dopamine neurons (17). As a potential therapeutic agent for the treatment of Parkinson's disease, GDNF has been widely tested (18, 19). It is well established that dopaminergic transmission in the cortico-limbic system is crucial for the development of drug dependence/addiction (20-25). Given that GDNF is considered an important modulator for dopaminergic neuronal function (17, 26), it is reasonable to postulate that GDNF may be involved in drug addiction. Although direct evidence of a clinical association between GDNF and drug dependence/addiction has yet to be obtained, GDNF has been identified in the development of drug dependence in animal models (27–30). Manipulations that modulate GDNF content in the brain affected cocaine-induced conditioned place preference and cocaine or ethanol self-administration in rats (27-30). GDNF (+/-) heterozygous knockout mice [GDNF (+/-) mice] showed greater morphine, cocaine, and methamphetamine (METH) conditioned place preference (27, 31). However, the role of GDNF in vulnerability to relapse of drug-seeking behavior remains unclear. Using animal models of drug selfadministration and relapsing behavior recently established in our laboratory (32, 33), which represent drug-taking and relapse of drug-seeking behavior in

doi: 10.1096/fj.06-7772com

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addicts (34), we provided evidence that a partial loss of GDNF expression not only facilitated the acquisition of METH self-administration, resulted in an upward shift in the dose-response curve, and increased motivation to take METH, but also led to increased vulnerability to METH-primed reinstatement and enduring cue-induced reinstatement of extinguished drug-seeking behavior.

MATERIALS AND METHODS

Subjects and drugs

The generation of GDNF knockout mice was described elsewhere (35). GDNF (-/-) homozygous knockout mice die shortly after birth, but GDNF (+/-) mice are viable. After genomic DNA was purified from a 0.5-1.0 cm segment of tail, the mice were genotyped by PCR utilizing three sets of primers selective of the neomycin cassette: primer 1 (5'-GAC TGG CTT GGT TCT TTG CAT GCA TCC -3'); primer 2 (5'-ACC AAA GAA CGG AGC CGG TTG GCG C-3'), and primer 3 (5'-GAG AGG AAT CGG CAG GCT GCA GCT G-3'). To characterize the influence of the GDNF expression on the operant behavior, a colony of GDNF (+/-) mice was employed in the present study. In this colony, the levels of GDNF expression in corticolimbic areas of the brain are reduced to 54-66% of those in wild-type littermates, at the age of 8 wk (Supplemental Fig. 1). Wild-type littermates were used as a control of the GDNF (+/-) mice. GDNF (+/-) and wildtype mice were bred locally in the Laboratory Animal Center, Nagoya University Graduate School of Medicine in Japan. Male GDNF (+/-) and wild-type mice were 8-wk-old and weighed 25-30 g at the beginning of the experiments. All mice were kept in a regulated environment (23 \pm 0.5°C; 50 \pm 0.5% humidity) with a reversed 12-h light/dark cycle (lights on at 9:00 AM). Both water and food were available ad libitum throughout the experiments unless otherwise noted. All procedures followed the National Institutes of Health Guidelines for the Care and Use of Laboratory Animals and were approved by the Nagoya University School of Medicine Animal Care and Use Committee.

METH hydrochloride (Dainippon Pharmaceutical Ltd, Osaka, Japan) was dissolved in sterile saline and self-administered at a dose of 0.1 mg/kg/infusion over 5 s (infusion volume=2.1 µl). The unit dose for METH self-administration is based on our previous report (32).

Food-reinforced operant behavior and reinstatement of food-seeking behavior

Food-reinforced operant behavior and motivation

Food-reinforced operant behavior and motivation were tested in standard mouse operant conditioning chambers as described previously (32). Briefly, the chamber was equipped with two nose-poke sensors (ENV-313M, Med Associates) in two holes, two cue-lamps in and above each hole, and a food pellet dispenser (ENV-203-20, Med Associates, Georgia, VT, USA) connected to a rectangular opening (2.25 cm × 2.25 cm) between the two holes. The bottom of the opening was 5 mm above the chamber floor and was equidistant from the holes. A house light was located at the top of the chamber opposite the holes. During the tests for food-reinforced operant behavior and motivation, one hole was defined as active, and the other, as inactive. Nose-poke responses in the active hole resulted in the delivery of a single food pellet

(dustless precision pellets 20 mg, A Holton Industries Co., Frenchtown, NJ, USA) to the opening by the dispenser (ENV302M, Med Associates) and inactivation of the cue-lamp and hole-lamp for 5 s followed by a 5 s timeout period. Nose-poke responses in the active hole during the timeout period and in the inactive hole had no programmed consequences but were recorded by the software MED-PC for Windows (Med Associates).

Naive GDNF (+/-) and wild-type mice (n=7 for each)genotype) were deprived of food for 20 h (water remained available ad libitum throughout the experiments). From the next day, both genotypes were daily subjected to nose-poke responding for food pellets in the standard operant chambers as mentioned above. During this phase, the mice were returned to their home cages and given unlimited amounts of food for 2 h immediately after each session of nose-poke responding for food pellets. The daily 3 h sessions of foodreinforced nose-poke responding in GDNF (+/-) and wildtype mice were initially performed under a fixed ratio (FR) 1 schedule. Once the mice showed stable nose-poke responding for food pellets (deviations of <15% of the mean of active responses in 3 consecutive training sessions), the reinforcement schedule was changed to FR2 until the same criterion as above was achieved. The same groups of mice were then subjected to nose-poke responding for food pellets under a progressive ratio (PR) schedule, in which the number of active nose-poke responses required to obtain a single food pellet escalates according to the following series: 1, 2, 4, 6, 9, 12, 15, 20, 25, 32, 40, 50, 62, 77, 95, 118, 145,178, 219, 268, 328, 402, 492, 603, 737, etc. (36). This "breaking point," expressed as the final ratio (the number of active nose-poke responses needed to earn the last single food pellet), reflects the intensity of motivation for earning food pellets. Each session under the PR schedule lasted for 5 h or until mice failed to respond within 1 h. After 2-4 daily sessions, GDNF (+/-) and wild-type mice demonstrated stable active nosepoke responses for food pellets (deviations of <15% of the mean of total active responses in 2 consecutive sessions).

Extinction and reinstatement of food-seeking behavior

During this phase, both food and water were available ad libitum in the home cages. After the test for motivation to take food pellets under the PR schedule, the same groups of GDNF (+/-) and wild-type mice were then subjected to 6-10 daily 3 h sessions of extinction. Throughout the extinction session, the house light was on. The food-associated cueand hole-lamps, and the system that delivers food pellets were turned off. Therefore, the nose-poke responses into the previously active hole resulted in neither the delivery of food pellets nor food-associated cues (cue- and hole-lamps). Once the mice met the criterion of extinction (<15 active responses or 25% of active responses in the stable phase of self-administration in 2 consecutive sessions), they were subjected to a 3 h session of the food-priming reinstatement test under the same conditions as in the extinction sessions (without either food-associated cues or the delivery of food pellets). As priming of food pellets, 12 food pellets were placed into the rectangular opening between the 2 holes before the food-priming reinstatement test. Nose-poke responses in the previously active or inactive hole were counted as active and inactive, respectively.

After the food-primed reinstatement test, the same groups of mice were subjected to 3–6 daily 3 h sessions of extinction immediately, and 3 months, after withdrawal from responding for food pellets. Once the mice met the extinction criterion as mentioned above, they were subjected to daily 3 h sessions of food-associated cue-induced reinstatement tests immediately, and 3 months, after the withdrawal. The food-

associated cue-induced reinstatement tests were performed under the same conditions as the food-reinforced operant behavioral test under the FR2 schedule, except that there was no delivery of food pellets after the nose-poke responses in a previously active hole. Nose-poke responses in the previously active or inactive hole were counted as active and inactive, respectively.

Surgery and apparatus for METH self-administration

Catheter implantation

New groups of naive GDNF (+/-) and wild-type mice were deprived of food for 20 h (water remained available ad libitum) and then trained to make nose-poke responses under the FR 1 schedule for food pellets in the operant chambers as mentioned above, except that both nose-poke holes were defined as active. Once a mouse had earned 30 food pellets, the session for nose-poke training ended (for 2-8 h, no difference between GDNF (+/-) and wild-type mice). After the training session, the mice were returned to their home cages, where both food and water were available ad libitum throughout the subsequent experiments. Two days later, the mice were anesthetized with pentobarbital sodium (50 mg/kg ip). Indwelling catheters were constructed of microsilicone tubing (inner diameter, 0.50 mm; outer diameter, 0.7 mm; IMG, Imamura Co., Ltd., Tokyo, Japan) and polyethylene tubing (inner diameter, 0.50 mm; outer diameter, 0.8 mm). Incisions were made on the skin of the head and ventral neck, and the right jugular vein was externalized. The end of the catheter was inserted into the jugular vein via a small incision and was secured to the vein and surrounding tissue with silk sutures. The exit port of the catheter passed subcutaneously to the top of the skull where it was attached to a modified 24-gauge cannula, which was secured to the mouse's skull with quick self-curing acrylic resin (Shofu Inc., Tokyo, Japan). To extend catheter patency, the catheters were flushed immediately after surgery, and in the morning and evening of the following days, with 0.03 ml of an antibiotic solution of cefmetazole sodium (20.0 mg/ml; Sankyo Co., Ltd., Tokyo, Japan) dissolved in heparinized saline (70 U/ml; Leo Pharmaceutical Products, Ltd., Tokyo, Japan). The patency of the catheter was usually confirmed once a week before operant behavior tests by infusion of a pentobarbital sodium solution (6.0 mg/ml, 0.15 ml/mouse) into the jugular vein. If the mice could not be knocked down within 5 s, the corresponding data were excluded from the statistical analysis.

Apparatus for METH self-administration

METH self-administration was conducted in standard mouse operant conditioning chambers (ENV-307A, Med Associates) located within ventilated sound attenuation cubicles as described previously (32). Briefly, the chambers were equipped with nose-poke sensors (ENV-313M, Med Associates) in two holes located on one side of the chamber 1.0 cm above the floor, cue- and hole-lamps located, respectively, above and in each hole, and a red house light located on the top of the chamber opposite the holes. During the self-administration, one hole was defined as active, and the other, as inactive. Nose-poke responses in the active hole resulted in activation of the infusion pump (PHM-100, Med Associates) and inactivation of the cue-lamp and hole-lamp. Nose-poke responses in the inactive hole, and in the active hole during the timeout period, had no programmed consequences but were recorded. The components of the infusion line were connected to each other from the injector to the exit port of the mouse's catheter by joint FEP tubing (inner diameter=0.25 mm; outer diameter=0.55 mm; Eicom Co., Ltd., Japan), which was encased in steel spring leashes (Instech, Plymouth Meeting, PA). Swivels were suspended above the chamber. One pump/ syringe set was used for each chamber located inside of the cubicle. The infusion pump/syringe set was outside of the chambers but inside of the cubicles.

METH self-administration and reinstatement of METH-seeking behavior

Outline

After recovering from the surgery to implant the catheter, GDNF (+/-) (n=28) and wild-type mice (n=26) were subjected to METH self-administration, extinction, and reinstatement of extinguished METH-seeking behavior according to the workflow shown in **Table 1**. During METH self-administration, nose-poke responses in the active hole resulted in an infusion of METH at a dose of 0.1 mg/kg/

TABLE 1. Workflow for METH self-administration and reinstatement

		Number (n)		
Step	Experiment	Wild-type	GDNF (+/-)	
1	Training for self-administration	26 (-14)*	28 (-10)*	
2.1	Dose-response	6	9 (-2)*	
2.2	Motivation	6	9 (-2)*	
3	Re-training for self-administration	12	14	
4	Extinction	7	8	
$\hat{5}$	METH-primed reinstatement	7	8	
6	Extinction	7	8	
7	1st cue-induced reinstatement	7	8	
8	Withdrawal until 3 months after the re-training	7	8	
9	Extinction	7	8	
10	2 nd cue-induced reinstatement	7	8	
11	Withdrawal until 6 months after the re-training	7	8	
12	Extinction	7	8	
13	3 rd cue-induced reinstatement	7	8	

^{*}The number in the brackets indicates the number of mice excluded from the statistical analysis because of a failure of catheter patency during the METH self-administration, or death from infection before completion of the 3rd cue-induced reinstatement.

infusion over 5 s (infusion volume=2.1 µl) followed by a 5 s timeout period. Nose-poke responses in the inactive hole, and in the active hole during the timeout period, had no programmed consequences but were recorded.

Acquisition of METH self-administration under an FR schedule

METH self-administration was initially under the FR1 schedule. Once the mice could make a minimum of 60% nose-poke responses in the active hole and received no >10 infusions of METH <2 consecutive sessions (at least for 4 sessions), the METH reinforcement schedule was changed to FR2. Under the FR2 schedule, the mice gradually acquired stable METH self-administration behavior (deviations of <15% of the mean of active responses in 3 consecutive training sessions). After acquiring stable self-administration behavior, GDNF (+/-) and wild-type mice were each counterbalance-separated into two subgroups. One subgroup of GDNF (+/-) and wild-type mice were subjected to the test for dose responses. The others were subjected to METH self-administration under the PR schedule.

Dose responses for METH self-administration under an FR2 schedule

After acquiring stable self-administration behavior, one subgroup of GDNF (+/-) and wild-type mice were subjected to METH self-administration under the FR2 schedule of reinforcement in the dose range 0.003-0.1 mg/kg/infusion from the higher to lower dose. Each mouse was subjected to two to four daily 3 h sessions of METH self-administration at one dose until it demonstrated stable active nose-poke responses (deviations of <15% of the mean of total active responses in 2 consecutive sessions).

Motivation for METH self-administration under a PR schedule

After stable self-administration behavior was acquired, the other subgroup of GDNF (+/-) and wild-type mice were subjected to METH self-administration under the PR schedule. The "breaking point" is defined as the final ratio (the number of active nose-poke responses needed to earn the last infusion of METH) and reflects the intensity of motivation for taking the drug tested. Each session lasted for 5 h or until mice failed to respond within 1 h. Each mouse was subjected to two to five sessions of METH self-administration. Both genotypes of mice demonstrated stable active nose-poke responses for METH infusion (as described in the section of dose response) during the two to five sessions.

Extinction

After the self-administration under the FR or PR schedule, the two subgroups of GDNF (+/-) and wild-type mice were subjected to METH (0.1 mg/kg/infusion) self-administration under the FR2 schedule until both genotypes showed stable (as described above) active nose-poke responses once again, and took approximately the same amount of METH. The mice were then subjected to 6-10 daily 3 h sessions of extinction before the METH-primed reinstatement test or 3-6 daily 3 h sessions of extinction before the cue-induced reinstatement test until they met the extinction criterion (<15 active responses or 25% of active responses in the stable phase of self-administration in 2 consecutive sessions). Throughout the extinction session, the house light was on. The METH-associated cue- and hole-lamps, and the pump for METH infusion, were turned off. Therefore, nose-poke responses into the previously active hole resulted in neither an infusion of METH nor METH-associated cues (cue- and hole-lamps, and pump noise for METH infusion).

METH-primed reinstatement

Once the extinction criterion was met, the GDNF (+/-) and wild-type mice were firstly subjected to a 3 h session of the operant test 30 min after the injection (ip) of saline as a control for the METH-primed reinstatement. From the next day, the mice were consecutively subjected to METH-primed reinstatement tests 30 min after the intraperitoneal injection with increasing doses of METH (0.2, 0.4, 1.0, 1.5, or 3.0 mg/kg, each dose for 1 daily 3 h session). The METH-primed reinstatement tests were conducted under the same conditions as in the extinction sessions in which neither METH infusions nor METH-associated cues were available after nose-poke responses into a previously active hole. Nose-poke responses in the previously active or inactive hole were counted as active and inactive, respectively.

Cue-induced reinstatement

Once the extinction criterion was met, the same groups of mice were subjected to the cue-induced reinstatement tests immediately, 3 months, and 6 months after withdrawal from METH self-administration. The cue-induced reinstatement tests were conducted under the same conditions as the METH self-administration under the FR2 schedule, except that METH was unavailable throughout the testing session. Nose-poke responses in the previously active or inactive hole were counted as active and inactive, respectively.

Data analysis

All data are \pm se. A one- or two-way ANOVA with (or without) repeated measures was performed for the difference in locomotor activity and nose-poke responses between the two genotypes of mice during the self-administration training, dose-response function, and METH-primed and cue-induced reinstatement of drug-seeking behavior, followed post hoc by the Bonferroni/Dunn test. The Mann-Whitney test was used to analyze the breaking points under the PR schedule, whereas Student's t test was used to analyze the other two sets of data. In all cases, a significant difference was set at P < 0.05.

RESULTS

Food-reinforced operant behavior and motivation of GDNF (+/-) and wild-type mice

Naive GDNF (+/-) and wild-type mice were trained to make nose-poke responses for food reinforcement under the FR and PR schedules in daily 3 h sessions. GDNF (+/-) mice did not show any significant difference from wild-type littermates in either active or inactive responses under the FR schedule of food reinforcement (Fig. 1A). Also, there was no significant difference in the number of training sessions to acquire stable operant behavior between GDNF (+/-) and wild-type mice (Fig. 1B). Furthermore, GDNF (+/-) and wild-type mice showed similar breaking points under the PR schedule (Fig. 1C). These findings indicate that a partial loss of GDNF expression does not

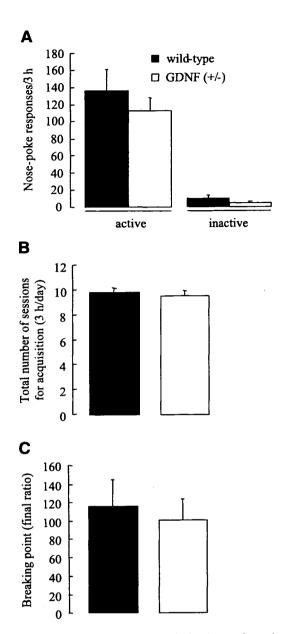


Figure 1. Food-reinforced operant behavior and motivation in GDNF (+/-) and wild-type mice. A) Active and inactive nose-poke responses for food reinforcement in a 3 h session under FR2 schedule during stable phase. B) Total number of training sessions needed to acquire stable active nose-poke responses for food reinforcement under FR schedule. C) Breaking points (final ratio) for food reinforcement under PR schedule. Data are mean \pm SE. n=7 for each genotype.

affect food-reinforced operant behavior and motivation in mice.

Reinstatement of food-seeking behavior in GDNF (+/-) and wild-type mice

To evaluate the reinstatement of food-seeking behavior in the mutant animals, the same groups of GDNF (+/-) and wild-type mice were subjected to extinction training after the tests for the food-reinforced operant behavior. After similar daily 3 h sessions of extinction

training, GDNF (+/-) and wild-type mice achieved the extinction criterion (Fig. 2A; $F_{(3,48)}=27.83$; P<0.001). However, the priming of food pellets failed to reinstate food-seeking behavior in either genotype (Fig. 2A). The same groups of mice were then subjected to extinction training once again. Once the extinction criterion was met, the food-associated cue-induced reinstatement tests were conducted immediately and 3 months after the end of tests for the food-reinforced operant behavior. Food-associated cues reliably trig-

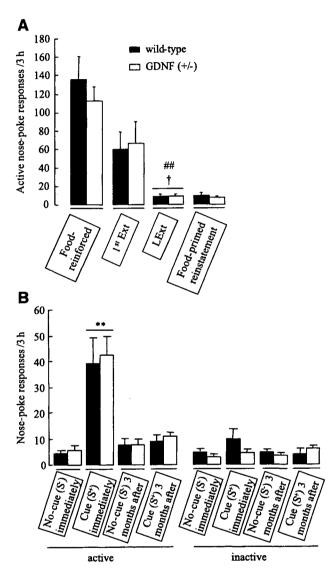


Figure 2. Extinction and reinstatement of food-seeking behavior in GDNF(+/-) and wild-type mice. A) Active nose-poke responses during stable phase of food-reinforced operant responding, extinction training, and food-primed reinstatement. B) Nose-poke responses in food-associated cue-induced reinstatement tests immediately, and 3 months, after withdrawal from food-reinforced operant behavior; Data are mean \pm se. n=7 for each genotype. **P<0.01 vs. Food-reinforced in same genotype; *P<0.05 vs. 1st Ext in same genotype; **P<0.01 vs. No-cue (S⁻) in same genotype. Food-reinforced, stable food-reinforced operant behavior; 1st Ext, first session of extinction; LExt, last session of extinction; No-cue (S⁻), control for cue-induced reinstatement (without food-associated cues and food pellets); Cue (S⁺), food-associated cue-induced reinstatement.

gered reinstatement of food-seeking behavior in both GDNF (+/-) and wild-type mice immediately after withdrawal (Fig. 2B; $F_{(1,24)}=33.44$; P<0.001). However, no significant difference in cue-induced reinstatement behavior was observed between GDNF (+/-) and wild-type mice (Fig. 2B). Importantly, the food cue-induced reinstatement of food-seeking behavior disappeared within the period of a 3 month withdrawal in both genotypes of animals (Fig. 2B). These findings suggest that reinstatement of food-seeking behavior in both genotypes of animals is weak or transient and that the partial loss of GDNF expression does not affect extinction behavior, reinstatement of food-seeking behavior, or duration of food-associated cue-induced reinstatement behavior in mice.

Facilitated acquisition of METH self-administration behavior in GDNF (+/-) mice

To investigate whether a partial loss of GDNF expression affects drug self-administration behavior in animals, separate groups of GDNF (+/-) and wild-type mice were subjected to METH self-administration training. GDNF (+/-) mice took less time than wild-type littermates to acquire stable METH self-administration behavior (Fig. 3A, P<0.01). However, there was no significant difference in total METH intake during the period of METH self-administration training between wild-type (30.7±2.0 mg/kg) and GDNF $(28.5\pm1.5 \text{ mg/kg})$ mice (Fig. 3B). In the early phase of METH self-administration under the FR1 schedule (Fig. 3C, day 1-4), neither genotype could discriminate active (METH-associated) from inactive (without METH infusion) nose-poke responses. Accordingly, there was no significant difference in active nose-poke responses for METH self-administration between

GDNF (+/-) and wild-type mice. The mice gradually demonstrated stable METH self-administration behavior under the FR2 schedule. Accordingly, they could discriminate active from inactive nose-poke responses to METH reinforcement (Fig. 3C, the last day 1–6 (L1-L6), P<0.001). However, there was no significant difference in active nose-poke responses for METH-taking between wild-type and GDNF (+/-) mice. These findings indicate that GDNF (+/-) mice are capable of METH self-administration.

Upward shifted dose responses and increased motivation to take METH in GDNF (+/-) mice

GDNF (+/-) and wild-type mice showed significantly different dose responses to self-administer METH (Fig. 4A; $F_{(1,55)} = 12.43$, P < 0.001). In the dose range of 0.01-0.03 mg/kg/infusion, the number of active nosepoke responses for METH-taking was significantly higher in GDNF (+/-) mice than in wild-type littermates (P < 0.05 and 0.001). There was no significant difference in active nose-poke responses to self-administer METH at 0.1 and 0.003 mg/kg/infusion between GDNF (+/-) and wild-type mice. When saline was substituted for METH, no significant difference was observed in self-administration behavior between GDNF (+/-) and wild-type mice. The upward shift of the dose-response function for METH self-administration in GDNF (+/-) mice suggests that the partial loss of GDNF expression may increase sensitivity to METH self-administration in mutant animals during the maintenance phase.

To further support this idea, the other subgroups of GDNF (+/-) and wild-type mice were subjected to METH (0.1 mg/kg/infusion) self-administration under the PR schedule. GDNF (+/-) mice demonstrated a

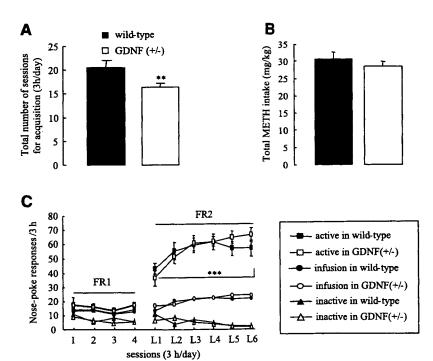
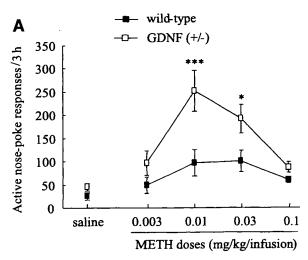
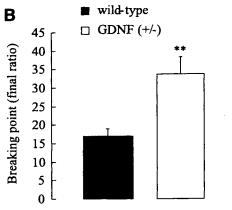


Figure 3. Acquisition of stable METH (0.1 mg/ kg/infusion) self-administration behavior in GDNF (+/-) and wild-type mice. A) Total number of training sessions needed to acquire stable METH self-administration behavior for GDNF (+/-) and wild-type mice. **P < 0.01 vs. wildtype littermates. B) Total METH intake during period of METH self-administration training for GDNF (+/-) and wild-type mice. C) Nose-poke responses and number of METH infusions during first 4 sessions (session 1-4) and last 6 sessions (session L1-L6) of METH self-administration under FR1 and FR2 schedules of reinforcement for GDNF (+/-) and wild-type mice. ***P < 0.001 vs. inactive nose-poke responses in same genotype; Data are mean \pm se. n = 12-18for each genotype.





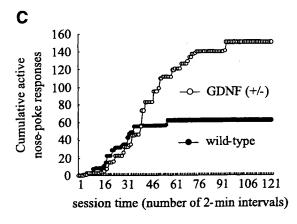


Figure 4. Dose responses and motivation for METH self-administration in GDNF (+/-) and wild-type mice. A) Dose-response function for METH self-administration under an FR2 schedule of reinforcement. B) Breaking points (final ratio) under a PR schedule of reinforcement. C) Representative curves for cumulative active nose-poke responses for METH-taking under PR schedule in mutant animals. Data are mean \pm se. n = 6-9 for each genotype. *P < 0.05, **P < 0.01, ***P < 0.001 vs. wild-type littermates.

significantly increased breaking point compared with wild-type littermates (Fig. 4B; P<0.01), suggesting that the partial loss of GDNF expression leads to greater motivation to take METH. Representative curves for the two genotypes of animals are illustrated in Fig. 4C.

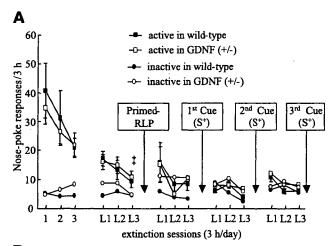
Increased vulnerability to METH-primed reinstatement of drug-seeking behavior in GDNF (+/-) mice

We further investigated the performance of GDNF (+/-) mice in the reinstatement of extinguished METH-seeking behavior. After once again acquiring stable METH self-administration behavior, during which wild-type and GDNF (+/-) mice had taken similar amounts of METH (43.9±3.2 and 50.1±4.1 mg/kg, respectively), the two genotypes were exposed to extinction training for 6-10 daily 3 h sessions. There was no significant difference between GDNF (+/-)and wild-type mice in the number of nose-poke responses into previously active holes [METH-associated; Fig. 5A, session 1-3 and the last session 1-3 (L1-L3)] or the number of extinction training sessions to achieve the extinction criterion (data not shown). During the last two extinction sessions, neither group of animals could discriminate active (previously associated with METH self-administration) from inactive (previously without METH self-administration) nose-poke responses, similar to the early stage of METH self-administration. These findings suggested that in GDNF (+/-) and wild-type mice, purposely active nose-poke responses acquired during METH self-administration had been extinguished.

Once the mice met the extinction criterion (Fig. 5A), a drug-primed reinstatement test was carried out 30 min after treatment with either saline or a different dose of METH. Wild-type and GDNF (+/-) mice showed different active nose-poke responses (Fig. 5B; $F_{(1,77)} = 12.72$; P < 0.001), although there was no significant difference in inactive nose-poke responses between GDNF (+/-) and wild-type mice. In wild-type littermates, both lower (0.2 and 0.4 mg/kg) and higher (1.5 and 3.0 mg/kg) doses of METH-priming failed to reinstate drug-seeking behavior. However, a moderate dose of METH (1.0 mg/kg) reliably triggered the reinstatement behavior. In contrast, both lower and moderate doses of METH (0.4 mg/kg and 1.0 mg/kg) reliably triggered the reinstatement of extinguished drug-seeking behavior in GDNF (+/-) mice (P < 0.05), although higher doses of METH did not evoke the reinstatement behavior. The leftward shift of the doseresponse curve for METH-primed reinstatement behavior suggests that the partial loss of GDNF expression may affect vulnerability to the reinstatement of extinguished METH-seeking behavior in mice.

Prolonged persistence of cue-induced reinstatement of drug-seeking behavior in GDNF (+/-) mice

To investigate the enduring vulnerability to cue-induced reinstatement of METH-seeking behavior in GDNF (+/-) mice, the same groups of mice were subjected to three to six extinction sessions, followed by three cue-induced reinstatement tests, which were conducted immediately, 3 months, and 6 months after METH withdrawal. During the extinction sessions, neither genotype showed any significant difference in



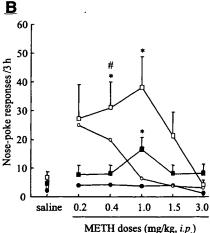


Figure 5. Nose-poke responses during extinction training (A) and METH-primed reinstatement of drug-seeking behavior (B; Primed-RLP) in GDNF (+/-) and wild-type mice. In A, data are from first 3 daily 3 h sessions (indicated by 1-3) and last 3 daily 3 h sessions (indicated by L1-L3) during 6-10 extinction training sessions before METH-primed reinstatement test (Primed-RLP), and last 3 daily 3 h sessions (indicated by L1-L3) during 3-6 sessions of extinction training before cue-induced reinstatement tests (1st -3rd Cue (S^+). Data are mean \pm se. n = 7-8 for each genotype. $^{\ddagger}P < 0.01$ vs. first session of extinction in same genotype. * $P < 0.05 \ vs$. saline treatment in same genotype. * $P < 0.05 \ vs$. wild-type littermates during same reinstatement test. Primed-RLP, METH-primed reinstatement; 1st Cue (S⁺), first test for METH-associated cue-induced reinstatement immediately after withdrawal; 2nd Cue (S+), second test for METH-associated cue-induced reinstatement 3 months after withdrawal; 3rd Cue (S+), third test for METH-associated cue-induced reinstatement 6 months after withdrawal.

nose-poke responses or in the number of training sessions needed to achieve the extinction criterion (Fig. 5A). Once the extinction criterion was achieved, the mice were subjected to cue-induced reinstatement tests. GDNF (+/-) and wild-type mice initially demonstrated a cue-triggered reinstatement of METH-seeking behavior (Fig. 6; 1^{st} test; P < 0.001). Importantly, there was a clear tendency for GDNF (+/-) mice to show more active nose-poke responses than wild-type littermates when exposed to the METH-associated cues (Fig. 6; $F_{(1, 26)} = 3.99$; P = 0.056). With a prolonged with-

drawal, GDNF (+/-) and wild-type mice showed significantly different responses (Fig. 6, $F_{(1, 150)}=26.1$; P<0.001). In wild-type littermates, the cue-induced reinstatement behavior was still observed 3 months after the withdrawal (Fig. 6; $2^{\rm nd}$ test; P<0.05) but disappeared after a 6 month withdrawal ($3^{\rm rd}$ test). In contrast, GDNF (+/-) mice maintained the cue-induced reinstatement behavior even after a 6-month withdrawal (Fig. 6; $3^{\rm rd}$ test; P<0.01). In addition, there was no significant difference in inactive nose-poke responses during any of the tests for cue-induced reinstatement behavior between GDNF (+/-) and wild-type mice.

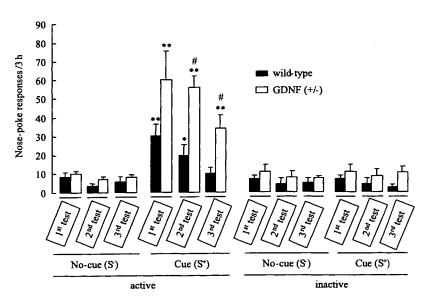
DISCUSSION

With the use of mouse models of METH self-administration and reinstatement of METH-seeking behavior, the present study demonstrated that a partial loss of GDNF expression resulted in a facilitated acquisition of METH self-administration behavior, upward shifted dose responses and enhanced motivation to take METH, increased vulnerability to drug-primed reinstatement, and prolonged cue-induced reinstatement of extinguished METH-seeking behavior. In contrast, there was no significant difference in food-reinforced operant behavior and motivation, locomotor activity, or novelty responses between the two genotypes of animals. These findings may provide evidence that GDNF is associated with vulnerability to relapse of METH-seeking behavior.

Acquisition and maintenance of METH self-administration behavior

It has been documented that GDNF (+/-) mice demonstrate increased morphine, cocaine, and METHconditioned place preference (27, 31). In the present study, GDNF (+/-) mice took a shorter period of time to acquire stable METH self-administration behavior, with an upward shift of dose responses to METH-taking, compared with wild-type littermates. Furthermore, GDNF (+/-) mice showed greater motivation (breaking point) for METH self-administration. In contrast, there was no significant difference in food-reinforced operant behavior (including acquisition and maintenance) and motivation between GDNF (+/-) and wild-type mice. Thus, GDNF (+/-) mice may represent a phenotype susceptible to the rewarding and reinforcing effects of addictive drugs although the precise molecular mechanisms underlying this phenomenon remain unclear. It has been documented that the extracellular dopamine concentration, FosB levels, and deltaFosB expression are elevated in the nucleus accumbens and striatum of GDNF (+/-) mice as compared with wild-type littermates (37). In addition, it is well established that GDNF is an important modulator for dopaminergic neuronal function (e.g., refs. 17, 26). Thus, it seems reasonable to postulate that

Figure 6. Active and inactive nose-poke responses during cue-induced reinstatement of METH-seeking behavior in GDNF (+/-) and wild-type mice. Tests for cue-induced reinstatement were conducted immediately (without withdrawal, 1st test), 3 months (2nd test), and 6 months (3rd test) after withdrawal from METH self-administration. Data are mean \pm se. n = 7-8for each genotype. *P < 0.05, **P < 0.01 vs. No-cue (S⁻) groups in same reinstatement test of same genotype. $^*P < 0.05$ vs. wild-type littermates in same reinstatement test. No-cue (S⁻): control for reinstatement test (without either METH-associated cues or METH infusion). Cue (S+): METH-associated cue-induced reinstatement (with METH-associated cues but no METH infusion).



the reduced GDNF content causes an increase in the extracellular dopamine concentration, FosB levels, and deltaFosB expression, leading to greater morphine-cocaine-, or METH-conditioned place preference (27, 31, 37), facilitated acquisition of METH self-administration behavior, upward shifted dose responses, and enhanced motivation to take METH (in the present study).

It seems unlikely that the differences in acquisition of self-administration behavior, dose responses, and motivation to take METH between GDNF (+/-) and wild-type mice are due to nonspecific physiological adaptations or compensatory effects during the development of mutant animals. Firstly, in our colony of GDNF (+/-) mice, the levels of GDNF expression in cortico-limbic areas of the brain were reduced 34-46%(see Supplemental Fig. 1). This is consistent with previous reports that striatal GDNF contents are reduced in GDNF (+/-) mice (37, 38). Secondly, both cocaine-conditioned place preference and cocaine- or ethanol-reinforced self-administration are reduced by an increased level of GDNF in the animal brain (27, 28, 30). Thirdly, cocaine-conditioned place preference and ethanol self-administration are potentiated by a decrease in the amount of GDNF in the brain through local delivery of anti-GDNF neutralizing antibodies (27, 30). A previous report has demonstrated an impairment of water-maze learning in GDNF (+/-) mice (39). In the present study, there was no significant difference in the acquisition of, retention of, and motivation for food-reinforced operant behavior between GDNF (+/-) and wild-type mice. Thus, it seems difficult to explain the alterations in METH-reinforced self-administration behavior and motivation, based on the learning and memory deficits in GDNF (+/-) mice. It has been suggested that responses to novelty in animals are associated with the propensity for drug self-administration (40). Given that there was no significant difference in exploratory behavior in the open field test, locomotor activities during habituation to the testing box environment, and locomotor responses to METH between GDNF (+/-) and wild-type mice (see Supplemental Figs. 2, 3), it is unlikely that differences in novelty responses or locomotor activity during METH self-administration contribute to alterations in METH-reinforced self-administration and motivation between GDNF (+/-) and wild-type mice.

Vulnerability to METH-primed reinstatement of drug-seeking behavior in GDNF (+/-) mice

No significant difference was observed in active or inactive nose-poke responses (Fig. 5A), and the number of extinction training sessions needed to achieve the extinction criterion (data not shown) between GDNF (+/-) and wild-type mice during the period of extinction training. This phenomenon indicates that a primary reinforcer (METH) or secondary reinforcer (METH-associated cues) may be necessary for the effects of GDNF on the development of METH-reinforced self-administration behavior. Indeed, GDNF (+/-) mutant mice showed a leftward and upward shifted dose-response curve for reinstatement of extinguished drug-seeking behavior after a priming injection of the primary reinforcer METH, whereas neither genotype showed reinstatement of food-seeking behavior after priming with food pellets. It has been shown that drug-primed reinstatement and drug self-administration share similar anatomical neural substrates (cortico-limbic system) and neural transmission (dopamine) in the brain (22, 41). Thus, possible mechanisms underlying enhanced METH-reinforced self-administration and motivation may contribute to the vulnerability to METH-primed reinstatement behavior in GDNF (+/-) mice. It seems unlikely that the vulnerability to METH-primed reinstatement behavior is due to the different experiences of METH self-administration. First, wild-type littermates took longer to acquire stable METH self-administration behavior than GDNF (+/-) mice (Fig. 5A), whereas there was no significant difference in active nose-poke responses for METH during the early and stable phases of METH selfadministration between GDNF (+/-) and wild-type mice (Fig. 3C). Second, there was no significant difference in total METH intake before the test for METHprimed reinstatement between wild-type and GDNF (+/-) mice $(43.9\pm3.2 \text{ and } 50.1\pm4.1 \text{ mg/kg}, \text{ respec-}$ tively). In addition, there was no significant difference in novelty seeking behavior and in METH-stimulated hyperlocomotion and locomotor sensitization between GDNF (+/-) and wild-type mice (see Supplemental Figs. 2, 3). This phenomenon is consistent with a previous report that the effects of acute and repeated treatment of cocaine on locomotor activity are similar between GDNF (+/-) and wild-type mice (37). For similar reasons to those mentioned above, it is unlikely that vulnerability to METH-primed reinstatement behavior in GDNF (+/-) mice reflects nonspecific increases in motor activity or novelty responses.

Prolonged cue-induced reinstatement of METHseeking behavior in GDNF (+/-) mice

In the present study, GDNF (+/-) mice demonstrated a stronger and more persistent cue-induced reinstatement of extinguished METH-seeking behavior than did wild-type littermates. Moreover, cue-induced reinstatement behavior in GDNF (+/-) mice could be observed even after a 6 month withdrawal when the cue-induced reinstatement in wild-type littermates had disappeared. In contrast, there was no significant difference in the transient cue-induced reinstatement of food-seeking behavior between GDNF (+/-) and wildtype mice (Fig. 2B). The more severe and persistent cue-induced reinstatement of extinguished METHseeking behavior in GDNF (+/-) mice suggests that the partial loss of GDNF expression may lead to vulnerability to and persistence of cue-induced reinstatement of drug-seeking behavior, without affecting food-seeking behavior. It has been reported that striatal synaptic plasticity is crucial for the formation of an addictive habit or cue-controlled drug-seeking behavior (42–44) and that deltaFosB, once expressed, persists in the brain for a relatively long period of time in the absence of further drug exposure and acts as a sustained molecular switch for addiction (45-47). Thus, the enduring vulnerability to cue-induced reinstatement may be attributable to higher levels of deltaFosB in striatal brain areas of GDNF (+/-) mice (37). In addition, the occurrence of drug-seeking behavior after a delay of several weeks in rats (48) seems inconsistent with our finding that cue-induced reinstatement of drug-seeking behavior was reduced with the time of withdrawal from METH self-administration in GDNF (+/-) and wild-type mice. This discrepancy may be because the cue-induced reinstatement behavior in the present study was examined after repeated cycles of extinction training (a with-subjects design), since repeated extinction training decreases the propensity for a relapse of extinguished drug-seeking behavior (49).

The present series of experiments demonstrated an association between specific genes or proteins, for

example, the expression of the GDNF gene, and vulnerability to relapse of drug-seeking behavior, suggesting that GDNF may be critically involved in the acquisition and maintenance of METH self-administration, vulnerability to METH-primed reinstatement, and persistent cue-induced reinstatement of extinguished drug-seeking behavior. In line with previous reports (27-30) and our present findings, GDNF may be a potential target of therapeutic agents not only for the prevention of drug dependence but also for the control of relapse of drug-seeking behavior.

We thank Dr. L. Shen and Dr. S. Furukawa for kindly providing the GDNF (+/-) heterozygous knockout mice. This study was supported in part by a Grant-in-Aid for Scientific Research and Special Coordination Funds for Promoting Science and Technology, Target-Oriented Brain Science Research Program, from the Ministry of Education, Culture, Sports, Science and Technology of Japan; by a Grant-in-Aid for Health Science Research on Regulatory Science of Pharmaceuticals and Medical Devices, and Comprehensive Research on Aging and Health from the Ministry of Health, Labor and Welfare of Japan; by a Grant-in-Aid for Scientific Research (B) and Young Scientists (A); in part by the 21st Century Center of Excellence Program "Integrated Molecular Medicine for Neuronal and Neoplastic Disorders" from the Ministry of Education, Culture, Sports, Science and Technology of Japan; and by the Brain Research Center from the 21st Century Frontier Research Program funded by the Ministry of Science and Technology, Republic of Korea.

REFERENCES

- 1. Tsuang, M. T., Lyons, M. J., Doyle, T., Eisen, S. A., Goldberg, J., True, W., Lin, N., Toomey, R., and Eaves, L. (1998) Cooccurrence of abuse of different drugs in men: the role of drug-specific and shared vulnerabilities. Arch. Gen. Psychiatry 55, 967-972
- Prescott, C. A., and Kendler, K. S. (1999) Genetic and environment contributions to alcohol abuse and dependence in a populated-based sample of male twins. Am. J. Psychiatry 156,
- Nestler, E. J. (2000) Genes and addiction. Nat. Genet. 26, 277-281
- Kendler, K. S., Jacobson, K. C., Prescott, C. A., and Neale, M. C. (2003) Specificity of genetic and environmental risk factors for use and abuse/dependence of cannabis, cocaine, hallucinogens, sedatives, stimulants, and opiates in male twins. Am. J. Psychiatry 160, 687-695
- Uhl, G. R., and Grow, R. W. (2004) The burden of complex
- genetics in brain disorders. Arch. Gen. Psychiatry 61, 223-229 Volkow, N. D., and Li, T. K. (2004) Drug addiction: the neurobiology of behaviour gone awry. Nat. Rev. Neurosci. 5,
- Kreek, M. J., Nielsen, D. A., Butelman, E. R., LaForge, K. S. (2005) Genetic influences on impulsivity, risk taking, stress responsivity and vulnerability to drug abuse and addiction. Nat. Neurosci. 8, 1450-1457
- Le, D. A., Li, Z., Funk, D., Shram, M., Li, T. K., and Shaham, Y. (2006) Increased vulnerability to nicotine self-administration and relapse in alcohol-naive offspring of rats selectively bred for high alcohol intake. J. Neurosci. 26, 1872-1879
- Picciotto, M. R., Zoli, M., Rimondini, R., Lena, C., Marubio, L. M., Pich, E. M., Fuxe, K., and Changeux, J. P. (1998) Acetylcholine receptors containing the beta2 subunit are involved in the reinforcing properties of nicotine. Nature 391, 173-177
- Rocha, B. A., Scearce-Levie, K., Lucas, J. J., Hiroi, N., Castanon, N., Crabbe, J. C., Nestler, E. J., and Hen, R. (1998) Increased

- vulnerability to cocaine in mice lacking the serotonin-1B receptor. Nature 393, 175-178
- Chiamulera, C., Epping-Jordan, M. P., Zocchi, A., Marcon, C., Cottiny, C., Tacconi, S., Corsi, M., Orzi, F., and Conquet, F. (2001) Reinforcing and locomotor stimulant effects of cocaine are absent in mGluR5 null mutant mice. *Nat. Neurosci.* 4, 873–874
- Caine, S. B., Negus, S. S., Mello, N. K., Patel, S., Bristow, L., Kulagowski, J., Vallone, D., Saiardi, A., and Borrelli, E. (2002) Role of dopamine D2-like receptors in cocaine self-administration: studies with D2 receptor mutant mice and novel D2 receptor antagonists. J. Neurosci. 22, 2977–2988
- Elmer, G. I., Pieper, J. O., Rubinstein, M., Low, M. J., Grandy, D. K., and Wise, R. A. (2002) Failure of intravenous morphine to serve as an effective instrumental reinforcer in dopamine D2 receptor knock-out mice. J. Neurosci. 22, 1-6
- Szumlinski, K. K., Dehoff, M. H., Kang, S. H., Frys, K. A., Lominac, K. D., Klugmann, M., Rohrer, J., Griffin, W., III, Toda, S., Champtiaux, N. P., et al. (2004) Homer proteins regulate sensitivity to cocaine. Neuron 43, 401-413
- Hyman, S. E. (2005) Addiction: a disease of learning and memory. Am. J. Psychiatry 162, 1414-1422
- Kalivas, P. W., and Volkow, N. D. (2005) The neural basis of addiction: a pathology of motivation and choice. Am. J. Psychiatry 162, 1403-1413
- Lin, L. F., Doherty, D. H., Lile, J. D., Bektesh, S., and Collins, F. (1993) GDNF: a glial cell line-derived neurotrophic factor for midbrain dopaminergic neurons. Science 260, 1130-1132
- Choi-Lundberg, D. L., Lin, Q., Chang, Y. N., Chiang, Y. L., Hay, C. M., Mohajeri, H., Davidson, B. L., and Bohn, M. C. (1997) Dopaminergic neurons protected from degeneration by GDNF gene therapy. Science 275, 838-841
- 19. Mandel, R. J., Spratt, S. K., Snyder, R. O., and Leff, S. E. (1997) Midbrain injection of recombinant adeno-associated virus encoding rat glial cell line-derived neurotrophic factor protects nigral neurons in a progressive 6-hydroxydopamine-induced degeneration model of Parkinson's disease in rats. Proc. Natl. Acad. Sci. U. S. A. 94, 14083-14088
- Nestler, E. J., and Aghajanian, G. K. (1997) Molecular and cellular basis of addiction. Science 278, 58-63
- Koob, G. F., Sanna, P. P., and Bloom, F. E. (1998) Neuroscience of addiction. *Neuron* 21, 467-476
- Self, D. W. (2004) Regulation of drug-taking and -seeking behaviors by neuroadaptations in the mesolimbic dopamine system. *Neuropharmacology* 47, 242–255
- Mizoguchi, H., Yamada, K., Mizuno, M., Mizuno, T., Nitta, A., Noda, Y., and Nabeshima, T. (2004) Regulations of methamphetamine reward by extracellular signal-regulated kinase 1/2/ ets-like gene-1 signaling pathway via the activation of dopamine receptors. Mol. Pharmacol. 65, 1293-1301
- Nagai, T., Yamada, K., Yoshimura, M., Ishikawa, K., Miyamoto, Y., Hashimoto, K., Noda, Y., Nitta, A., and Nabeshima, T. (2004)
 The tissue plasminogen activator-plasmin system participates in the rewarding effect of morphine by regulating dopamine release. Proc. Natl. Acad. Sci. U. S. A. 101, 3650-3655
- Stuber, G. D., Wightman, R. M., and Carelli, R. M. (2005) Extinction of cocaine self-administration reveals functionally and temporally distinct dopaminergic signals in the nucleus accumbens. *Neuron* 46, 661-669
- Tomac, A., Lindqvist, E., Lin, L. F., Ogren, S. O., Young, D., Hoffer,
 B. J., and Olson, L. (1995) Protection and repair of the nigrostriatal dopaminergic system by GDNF in vivo. *Nature* 373, 335–339
- Messer, C. J., Eisch, A. J., Carlezon, W. A. Jr., Whisler, K., Shen, L., Wolf, D. H., Westphal, H., Collins, F., Russell, D. S., and Nestler, E. J. (2000) Role for GDNF in biochemical and behavioral adaptations to drugs of abuse. *Neuron* 26, 247-257
- Green-Sadan, T., Kinor, N., Roth-Deri, I., Geffen-Aricha, R., Schindler, C. J., and Yadid, G. (2003) Transplantation of glial cell line-derived neurotrophic factor-expressing cells into the striatum and nucleus accumbens attenuates acquisition of cocaine self-administration in rats. Eur. J. Neurosci. 18, 2093–2098
- Green-Sadan, T., Kuttner, Y., Lublin-Tennenbaum, T., Kinor, N., Boguslavsky, Y., Margel, S., and Yadid, G. (2005) Glial cell line-derived neurotrophic factor-conjugated nanoparticles suppress acquisition of cocaine self-administration in rats. Exp. Neurol. 194, 97-105
- He, D. Y., McGough, M. N., Ravindranathan, A., Jeanblanc, J., Logrip, M. L., Phamluong, K., Janak, P. H., and Ron, D. (2005)

- Glial cell line-derived neurotrophic factor mediates the desirable actions of the anti-addiction drug ibogaine against alcohol consumption. J. Neurosci. 25, 619–628
- Niwa, M., Nitta, A., Yamada, Y., Nakajima, A., Saito, K., Seishima, M., Shen, L., Noda, Y., Furukawa, S., and Nabeshima, T. (In press) An inducer for glial cell line-derived neurotrophic factor and tumor necrosis factor-f_c protects against methamphetamineinduced rewarding effects and sensitization. *Biol. Psychiatry*
- Yan, Y., Nitta, A., Mizoguchi, H., Yamada, K., and Nabeshima, T. (2006) Relapse of methamphetamine seeking behavior demonstrated by a reinstatement procedure involving self-administration. Behav. Brain. Res. 168, 137-143
- Yan, Y., Yamada, K., Nitta, A., and Nabeshima, T. (2006)
 Transient drug-primed but persistent cue-induced reinstatement of extinguished methamphetamine-seeking behavior in mice. Behav. Brain Res. 177, 261-268
- 34. Epstein, D. H., Preston, K. L., Stewart, J., and Shaham, Y. (2006) Toward a model of drug relapse: an assessment of the validity of the reinstatement procedure. *Psychopharmacology* 189, 1-16
- Pichel, J. G., Shen, L., Sheng, H. Z., Granholm, A. C., Drago, J., Grinberg, A., Lee, E. J., Huang, S. P., Saarma, M., et al. (1996) Defects in enteric innervation and kidney development in mice lacking GDNF. Nature 382, 73-76
- Roberts, D. C., and Bennett, S. A. (1993) Heroin self-administration in rats under a progressive ratio schedule of reinforcement. Psychopharmacology 111, 215-218
- Airavaara, M., Planken, A., Gaddnas, H., Piepponen, T. P., Saarma, M., and Ahtee, L. (2004) Increased extracellular dopamine concentrations and FosB/DeltaFosB expression in striatal brain areas of heterozygous GDNF knockout mice. Eur. J. Neurosci. 20, 2336-2344
- Griffin, W. C., III, Boger, H. A., Granholm, A. C., and Middaugh, L. D. (2006) Partial deletion of glial cell line-derived neurotrophic factor (GDNF) in mice: Effects on sucrose reward and striatal GDNF concentrations. *Brain Res.* 1068, 257–260
- Gerlai, R., McNamara, A., Choi-Lundberg, D. L., Armanini, M., Ross, J., Powell-Braxton, L., and Phillips, H. S. (2001) Impaired water maze learning performance without altered dopaminergic function in mice heterozygous for the GDNF mutation. Eur. J. Neurosci. 14, 1153-1163
- Piazza, P. V., Deminiere, J.-M., Le, Moal. M., and Simon, H. (1989) Factors that predict vulnerability to amphetamine selfadministration. Science 245, 1511-1513
- Zavala, A. R., Weber, S. M., Rice, H. J., Alleweireldt, A. T., and Neisewander, J. L. (2003) Role of the prelimbic subregion of the medial prefrontal cortex in acquisition, extinction, and reinstatement of cocaine-conditioned place preference. *Brain. Res.* 990, 157-164
- Gerdeman, G. L., Partridge, J. G., Lupica, C. R., and Lovinger, D. M. (2003) It could be habit forming: drugs of abuse and striatal synaptic plasticity. *Trends Neurosci.* 26, 184-192
- 43. Everitt, B. J., and Robbins, T. W. (2005) Neural systems of reinforcement for drug addiction: from actions to habits to compulsion. *Nat. Neurosci.* 8, 1481-1489
- Vanderschuren, L. J., Di Ciano, P., and Everitt, B. J. (2005) Involvement of the dorsal striatum in cue-controlled cocaine seeking. J. Neurosci. 25, 8665–8670
- Nestler, E. J., Barrot, M., and Self, D. W. (2001) DeltaFosB: a sustained molecular switch for addiction. *Proc. Natl. Acad. Sci.* U. S. A. 98, 11042–11046
- Colby, C. R., Whisler, K., Steffen, C., Nestler, E. J., and Self, D. W. (2003) Striatal cell type-specific overexpression of DeltaFosB enhances incentive for cocaine. J. Neurosci. 23, 2488–2493
- 47. Ulery, P. G., Rudenko, G., and Nestler, E. J. (2006) Regulation of DeltaFosB stability by phosphorylation. *J. Neurosci.* 26, 5131-5149
- Lu, L., Grimm, J. W., Hope, B. T., Shaham, Y. (2004) Incubation of cocaine craving after withdrawal: a review of preclinical data. Neuropharmacology. 47 (Suppl), 214–226
- Sutton, M. A., Schmidt, E. F., Choi, K. H., Schad, C. A., Whisler, K., Simmons, D., Karanian, D. A., Monteggia, L. M., Neve, R. L., Self, D. W. (2003) Extinction-induced upregulation in AMPA receptors reduces cocaine-seeking behaviour. *Nature* 421, 70-75

Received for publication November 29, 2006. Accepted for publication February 1, 2007.

Reduction of methamphetamine-induced sensitization and reward in matrix metalloproteinase-2 and -9-deficient mice

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Abstract

Matrix metalloproteinases (MMPs) and their inhibitors (TIMPs) function to remodel the pericellular environment. Their activation and regulation are associated with synaptic physiology and pathology. Here, we investigated whether MMP-2 and MMP-9 are involved in the rewarding effects of and sensitization to methamphetamine (METH) in animals, in which the remodelling of neural circuits may play a crucial role. Repeated METH treatment induced behavioural sensitization, which was accompanied by an increase in MMP-2 and MMP-9 activity in the brain. In MMP-2- and MMP-9-deficient mice [MMP-2-(-/-)) and MMP-9-(-/-)], METH-induced behavioural sensitization and conditioned place preference, a measure of the rewarding effect, as well as METH-increased dopamine

release in the nucleus accumbens (NAc) were attenuated compared with those in wild-type mice. In contrast, infusion of purified human MMP-2 into the NAc significantly potentiated the METH-increased dopamine release. The [³H]dopamine uptake into striatal synaptosomes was reduced in wild-type mice after repeated METH treatment, but METH-induced changes in [³H]dopamine uptake were significantly attenuated in MMP-2-(-/-) and MMP-9-(-/-) mice. These results suggest that both MMP-2 and MMP-9 play a crucial role in METH-induced behavioural sensitization and reward by regulating METH-induced dopamine release and uptake in the NAc.

Keywords: dopamine, drug dependence, matrix metalloproteinases, methamphetamine.

J. Neurochem. (2007) 100, 1579-1588.

Drug dependence is a complex phenomenon with important psychological and social causes and consequences, which result from adaptations in specific brain circuits caused by repeated exposure to drugs of abuse (Berke and Hyman 2000; Yamada et al. 2005). It has been proposed that cellular and molecular mechanisms for drug dependence involve processes similar to those operating in other forms of synaptic plasticity such as learning and memory (Shippenberg and Heidbreder 1995; Mizoguchi et al. 2004). Methamphetamine (METH), a typical drug of abuse, has both acute and long-lasting effects on psychomotor behaviours. The effects of METH are associated with an increase in extracellular dopamine levels in the brain through redistri-

Received September 2, 2006; revised manuscript received September 21, 2006; accepted October $16,\,2006.$

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Abbreviations used: DAT, dopamine transporter; ECM, extracellular matrix; Fc, frontal cortex; GFAP, glial fibrillary acidic protein; METH, methamphetamine; MMP, matrix metalloproteinase; NAc, nucleus accumbens; NeuN, neuron-specific nuclear antigen; SDS-PAGE, sodium dodecyl sulfate – polyacrylamide gel electrophoresis; TIMP, tissue inhibitors of MMP; tPA, tissue plasminogen activator.

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