

に最も進化した大脳皮質部位の一つである前頭極の個体発生過程に影響する可能性が示唆された。統合失調症をはじめ、一卵性双生児での一致率が 50-60%に留まるような精神疾患の病態と成因の解明には、若年者を対象として縦断研究も含んだ“発達双生児法”による大規模研究を進めることが今後必要であろう。

F. 研究発表

1. 論文発表

・齋藤治, 臺弘: 統合失調症の瞬間意識仮説. Schizophrenia Frontier (メディカルレビュー社), Vol. 8 No. 4; 257-261, 2008

・齋藤治: 妄想発生の脳メカニズム. ころの科学, 126号, pp. 18-22, 2006

・岡本長久: 難治性うつ病への対応. 医学の歩み, 219: 955-962, 2006

・岡本長久: うつ病の薬物療法-最新の知見を踏まえて-. 調剤と情報, 686-691, 2006

2. 学会発表

・Okamoto N, Saitoh O, Yamashita F, Tatemichi N, Imamura A, Ohnishi T, and Anami K: An MRI study of neurodevelopmental risk factors for psychosis: a relationship between discordant birth weight and morphological brain development in healthy monozygotic twins. Schizophrenia Res, 86 (2006) S77-8.

(presented at the 5th International Conference on Early Psychosis, Birmingham, UK, October 4-6, 2006)

・長房裕子, 佐藤真由美, 山下典生, 森崎洋平, 伊藤暢厚, 坂本広太, 岡本長久: 「うつ病における疾患特異的 SPECT 所見の検討」, 第 103 回日本精神神経学会総会、2007

年 5 月, 高知

G. 知的財産権の出願・登録状況

1. 特許取得

無し。

2. 実用新案登録

無し。

3. その他

無し。

厚生労働科学研究費補助金（こころの健康科学研究事業）
（総合）研究報告書

双生児法による精神疾患の病態解明に関する研究
精神疾患における内因性レトロウイルスの関与についての検討

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研究要旨：統合失調症の発症に関して内在性レトロウイルス（HERV）やLINE-1（L1）とDNAメチル化に注目し、網羅的探索を行った。脳での潜在的転写活性を持つ期待の高いHERV 4つを同定した。L1の同定には効率の高い画期的方法の開発が必須である。

A. 研究目的：環境要因→DNAメチル化→HERV・L1→病像形成遺伝子⇒発症。模式的に示した作業仮説を検証することが1つの区切りとなる目標である。この分担課題では、ヒトゲノムの全配列を探索して、転写活性能を持っているHERV及びL1を可能なかぎり数多く同定することが目的となった。

B. 研究方法：HERV及び(昨年度の)L1のスクリーニングにヒトdbESTを利用した。照合元のQueryには延べ98種類のHERV配列又はL1.3完全長（約6 kb）を用いた。得られたESTをBLASTにより該当するローカスに配置して選出の指標にした。更にL1に関しては、今年度の報告書に記したようにゲノム配列からのアプローチも行った。抽出した候補ローカスは従来の方法で検証を行った（一次検証）。

（倫理面への配慮）

データベース解析と特定不能な1例の核酸を材料とした基礎的実験であるため、特に倫理面の問題は無いと判断した。

C. 研究結果：転写活性能を保持している可能性が高い4つのHERVローカスを同定した：HERV-K102（1q21-q22）、HERV-H（22q12）、HERV-P3b（1p36.13）、HERV-K9（5p15.33）。L1のESTデータベーススクリーニングから、転写活性能保持型L1候補を254ローカスに絞り込んだ。そのうち染色体2番上のL1ローカスと染色体15番に局在するL1ローカスが一次検証の対象となった。が、二カ所に設定されたプライマーによるRT-PCRのいずれにも検出されなかった。ゲノム配列からの探索では146個のL1が抽出された。同様に、解析

した計403クローンに検出されたローカスはわずか7つだった。しかし、サブファミリーの分布はゲノムPCRとRT-PCRで異なっていた。また、同一配列が複数のクローンに検出される頻度にも有意な差が認められた。

D. 考察：潜在的転写活性を持つHERV 4つを同定した。期待したよりはるかに少ない。L1に至っては一次検証の段階で壁にぶつかった。母集団が大き過ぎる。反復配列解析の困難さを改めて痛感した。また、EST配列の質が低いことも問題である。検証法の開発・Queryの選び方が課題になる。

E. 結論：転写活性能をもつHERVおよびL1の探索・同定を試みた。困難な壁があり、Queryの選び方の工夫と検証法の開発が必須である。

F. 研究発表

1. 論文発表

1. Shen H et al. Tissue-specificity in methylation and expression of human genes coding for neuro-peptides and their receptors, and of a human endogenous retrovirus K family. J Hum Genet 51 (5): 440-450, 2006/2. Shimabukuro M et al. Haloperidol treatment induces tissue- and sex-specific changes in DNA methylation. Behav Brain Funct 2: 37, 2006/3. Shimabukuro M et al. J Psychiat Res 41 (12): 1042-1046, 2007

2. 学会発表：神山聡子ら、ゲノム解析による活性LINE1の同定. 15th 日精・行遺医

G. 知的財産権の出願・登録状況

1. 特許取得：なし/2. 実用新案登録：なし/3. その他：なし

研究成果の刊行一覧表（平成 17～19 年度）

加藤 忠史

氏名	タイトル	雑誌／書籍名	巻	頁	年
Kakiuchi C, Ishiwata M, Nanko S, Ozaki N, Iwata N, Umekage T, Tochigi M, Kohda K, Sasaki T, Imamura A, Okazaki Y, Kato T.	Up-regulation of ADM and SEPX1 in the lymphoblastoid cells of monozygotic twins discordant for schizophrenia.	Am J Med Genet B Neuropsychiatr Genet.			2007
Kato T, Kakiuchi C, Iwamoto K	Comprehensive gene expression analysis in bipolar disorder	Can J Psychiatry	J 52 (12)	763-71	2007
McGowan PO, Kato T	Epigenetics in mood disorders	Environmental Health and Preventive Medicine			in press
加藤忠史	気分障害のエピジェネティクス	分子精神医学	8	38-44	2008
Kuratomi G, Iwamoto K, Bundo M, Kusumi I, Kato N, Iwata N, Ozaki N, Kato T	Aberrant DNA methylation associated with bipolar disorder identified from discordant monozygotic twins.	Molecular Psychiatry			2007
Chihiro Kakiuchi, Mizuho Ishiwata, Akiko Hayashi, Tadafumi Kato	XBP1 induces WFS1 through an endoplasmic reticulum stress response element-like motif in SH-SY5Y cells	Journal of Neurochemistry	of 97	545-555	2006

大木 秀一

氏名	タイトル	雑誌／書籍名	巻	頁	年
Syuichi Ooki	Population-Based Database of Twin Research Multiples in Childhood of Ishikawa Prefecture, Japan	Genetics	9(6)	832-837	2006

大野 裕

氏名	タイトル	雑誌／書籍名	巻	頁	年
Yamagata S, Takahashi Y, Kijima N, Maekawa H, Ono Y, Ando J	Genetic and environmental etiology of effortful control.	Twin Res Hum Genet	8(4)	300-306	2005

岡崎 祐士

氏名	タイトル	雑誌／書籍名	巻	頁	年
谷井久志、井上顕、西村幸香、梶木直美、貝谷久宣、佐々木司、岡崎祐士	パニック障害の遺伝子探索	脳と精神の医学	18(1)	1-7	2007
Shimabukuro M, Sasaki T, Imamura A, Tsujita T, Fuke C, Umekage T, Tochigi M, Hiramatsu K, Miyazaki T, Oda T, Sugimoto J, Jinno Y, Okazaki	Global hypomethylation of peripheral leukocyte DNA in male patients with schizophrenia: A potential link between epigenetics and schizophrenia.	Journal of Psychiatric Research			2006

笠井 清登

氏名	タイトル	雑誌／書籍名	巻	頁	年
Kasai K*, Yamasue H*, Gilbertson MW, Shenton ME, Lasko NB, Rauch SL, Pitman RK (*equal contribution)	Evidence for acquired pregenual anterior cingulate gray matter loss from a twin study of combat-related post-traumatic stress disorder	Biol Psychiatry	63	550-558	2008
Yamasue H, Abe O, Suga M, Yamada H, Inoue H, Tochigi M, Rogers MA, Aoki S, Kato N, Kasai K	Gender-common and -specific neuroanatomical basis of human anxiety-related personality traits	Cereb Cortex	18	46-52	2008
Yamasue H, Ishijima M, Abe O, Sasaki T, Yamada H, Suga M, Rogers MA, Minowa I, Someya T, Kurita H, Aoki S, Kato N, Kasai K	Neuroanatomy in monozygotic twins with Asperger's disorder discordant for comorbid depression	Neurology	65	491-492	2005

陣野 吉広

氏名	タイトル	雑誌／書籍名	巻	頁	年
Shen H, Nakamura A, Sugimoto J, Sakumoto N, Oda T, Jinno Y, Okazaki Y	Tissue-specificity of methylation and expression of human genes coding for neuropeptides and their receptors, and of a human endogenous retrovirus K family.	J Hum Genet	51(5)	440-450	2006

資 料

Up-Regulation of *ADM* and *SEPX1* in the Lymphoblastoid Cells of Patients in Monozygotic Twins Discordant for Schizophrenia

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The contribution of genetic factors to schizophrenia is well established and recent studies have indicated several strong candidate genes. However, the pathophysiology of schizophrenia has not been totally elucidated yet. To date, studies of monozygotic twins discordant for schizophrenia have provided insight into the pathophysiology of this illness; this type of study can exclude inter-individual variability and confounding factors such as effects of drugs. In this study we used DNA microarray analysis to examine the mRNA expression patterns in the lymphoblastoid (LB) cells derived from two pairs of monozygotic twins discordant for schizophrenia. From five independent replicates for each pair of twins, we selected five genes, which included *adrenomedullin* (*ADM*) and *selenoprotein X1* (*SEPX1*), as significantly changed in both twins with schizophrenia. Interestingly, *ADM* was previously reported to be up-regulated in both the LB cells and plasma of schizophrenic patients, and *SEPX1* was included in the list of genes up-regulated in the peripheral blood cells of schizophrenia patients by microarray analysis. Then, we performed a genetic association study of schizophrenia in the Japanese population and examined the copy number variations, but observed no association. These findings suggest the possible role of *ADM* and

SEPX1 as biomarkers of schizophrenia. The results also support the usefulness of gene expression analysis in LB cells of monozygotic twins discordant for an illness. © 2007 Wiley-Liss, Inc.

KEY WORDS: adrenomedullin; selenoprotein; DNA microarray; gene expression; genetic association study

Please cite this article as follows: Kakiuchi C, Ishiwata M, Nanko S, Ozaki N, Iwata N, Umekage T, Tochigi M, Kohda K, Sasaki T, Imamura A, Okazaki Y, Kato T. 2007. Up-Regulation of *ADM* and *SEPX1* in the Lymphoblastoid Cells of Patients in Monozygotic Twins Discordant for Schizophrenia. *Am J Med Genet Part B* 9999:1–8.

INTRODUCTION

Genetic factors in schizophrenia have been shown by family, twin, and adoption studies. A higher concordance rate of schizophrenia in monozygotic twins (41–79%) compared with that in dizygotic twins (0–14%) especially supports the contribution of genetic factors in schizophrenia [Shih et al., 2004]. As the risk genes for schizophrenia, a balanced translocation disrupting disrupted schizophrenia-1 (*DISC1*) [Millar et al., 2000] and a chromosomal deletion at 22q11 [Bassett and Chow, 1999] are well established. As common variants associated with schizophrenia, dystrobrevin-binding protein 1 (*DTNBP1*) [Straub et al., 2002] and neuregulin 1 (*NRG1*) [Stefansson et al., 2002], which were identified from linkage analysis, were reported. However, the association of *DTNBP1* haplotype with schizophrenia is not consistent among studies [Mutsuddi et al., 2006]. Further studies to identify the molecular pathology of this illness are needed.

In addition to the traditional genetic approaches, an additional strategy to identify the genetic basis of endophenotypes of schizophrenia is becoming popular. In this approach, endophenotypes, measurable biological variables associated with genetic risk of schizophrenia, are first identified; then their genetic basis is studied [Braff et al., 2007]. Many established endophenotypes, such as eye tracking abnormality [Holzman et al., 1977], ventricular enlargement [Reveley et al.,

Grant sponsor: Japanese^{Q2} Ministry of Health and Labor; Grant number: H17-KOKORO-general-009; Grant sponsor: The Ministry of Education, Culture, Sports, Science and Technology (MEXT); Grant number: 16659307.

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Received 20 May 2007; Accepted 6 September 2007

DOI 10.1002/ajmg.b.30643

Published online 00 Month 2007 in Wiley InterScience (www.interscience.wiley.com)

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1982], reduced hippocampal volume [Suddath et al., 1990], hypofrontality [Berman et al., 1992], and neuropsychological measures [Goldberg et al., 1995], were validated by the study of monozygotic twins discordant for schizophrenia.

In an attempt to identify molecular endophenotypes, biochemical differences in blood between the monozygotic twins discordant for schizophrenia have been investigated. These studies showed some differences between twins: plasma haptoglobin levels [Vander Putten et al., 1996], DNA methylation status [Tsujita et al., 1998; Petronis et al., 2003], soluble interleukin-2 receptors (SIL-2Rs) [Rapaport et al., 1993], mRNA expression level of a certain transcript [Friedhoff et al., 1995], retrovirus [Deb-Rinker et al., 1999], catecholamine levels [Walker et al., 2002], DNA stability [Nguyen et al., 2003], and lipid metabolism [Tsang et al., 2006]. On the other hand, no difference was found for viral nucleic acids [Sierra-Honigmann et al., 1995], platelet monoamine oxidase activity [Reveley et al., 1983], and genomic sequences [Polymeropoulos et al., 1993; Vincent et al., 1998; McDonald et al., 2003]. If a robust difference between discordant twins is well validated, such a finding will become a clue to identify the cause of this difficult illness [Kato et al., 2005a].

To identify the genes differentially expressed between the twins, one may use peripheral blood cells. However, this method is hampered by the fact that most of the patients are under treatment with drugs such as antipsychotics, which potentially affect the gene expression patterns. One possible method to avoid these confounding factors is to use the lymphoblastoid (LB) cells. Gene expression patterns in LB cells can be assessed with minimum inter-individual variability [Cheung et al., 2003], and the effect of drugs may be avoided or reduced by culturing the cells.

We previously performed DNA microarray analysis and examined the mRNA expression pattern using LB cells of monozygotic twins discordant for bipolar disorder. On the basis of our findings, we suggested the possible contribution of the endoplasmic reticulum stress response pathway to the pathophysiology of the illness [Kakiuchi et al., 2003]. Recently, Matigian et al. [2007] also performed DNA microarray analysis in three pairs of monozygotic twins discordant for bipolar disorder and found that genes related to the WNT signaling pathway were altered in patients. Several other groups have also applied the similar strategy to other illnesses such as autism and rheumatoid arthritis [Haas et al., 2006; Hu et al., 2006].

In this study, we used DNA microarray analysis to examine the mRNA expression pattern in the cells of two pairs of monozygotic twins discordant for schizophrenia. Because one of the problems in this strategy is lack of statistical analysis due to small sample size, we performed five independent experiments for each pair of twins. The expression of five genes commonly was shown to be altered in both of the twins, and two genes survived after the exclusion of three immunoglobulin-related genes. Interestingly, both of the final genes, *adrenomedullin* (*ADM*) and *selenoprotein X1* (*SEPX1*), had been reported to be up-regulated in the cells or plasma of schizophrenic patients. We further tried to identify the genetic basis of up-regulation of *ADM* and *SEPX1* levels in schizophrenia by a case-control association analysis of schizophrenia in the Japanese population. Because copy number variation (CNV) was reported to exist around these loci, CNV was also examined.

MATERIALS AND METHODS

Subjects

For the DNA microarray analysis, two pairs of monozygotic twins discordant for schizophrenia (SZ twins) were recruited.

The SZ twins A were 54-year-old males, and SZ twins B were 24-year-old females, who were previously reported elsewhere [Kunugi et al., 2003].

The SZ twins A were diagnosed by the consensus of two senior psychiatrists after independent unstructured interviews. Their family history was obtained from interviews of the twins. They had two healthy sisters, and their parents did not have major mental disorders. The affected twin of this pair (SZ-twin-A1) graduated from a university and worked as an office worker for 2 years. At age 25, he developed disorganized behavior and thought, accompanied by excitation. He also had non-systematic delusion of persecution and auditory hallucination. He was hospitalized in a psychiatric ward for 3 months. After the first episode, he was admitted to psychiatric hospitals 13 times. He began to develop negative symptoms and changed jobs several times because of interpersonal problems. He married at age 32, but divorced 1 year later. After that, he could not continue to work and lived alone, supported by social welfare. His diagnosis according to the International Classification of Diseases, Revision 10 (ICD-10) was schizophrenia, disorganized type. He was also diagnosed to have diabetes mellitus. He had been treated with 150 mg of clocapramine hydrochloride, a typical antipsychotic, and 3 mg of trihexyphenidyl hydrochloride, as an antiparkinsonian drug. It is not known whether his diabetes is a side effect of these drugs. His co-twin had been working at a company for 30 years and had been married. He was not diagnosed to have any major mental disorders or personality disorders. He did not have diabetes.

The proband of SZ twins B was diagnosed by the consensus of two senior psychiatrists after independent unstructured interviews. The diagnosis of the proband according to the Diagnostic and Statistical Manual of Mental Disorders, Fourth Edition (DSM-IV; American Psychiatric Association) was schizophrenia. Her co-twin was interviewed with the use of the schedule for affective disorders and schizophrenia (SADS), which revealed no current or past history of affective disorders or psychotic disorders. Their mother was interviewed and found to be healthy. Their father was also healthy, according to the available information. The symptoms of the proband is minutely described elsewhere [Kunugi et al., 2003]. In brief, the proband's symptoms began around the age of 15, with delusion of persecution. After that, she developed auditory hallucination and negative symptoms.

For the case-control association study, the genomic DNA derived from peripheral blood cells of 223 patients with schizophrenia (45.7 ± 14.9 years old, 129 males and 94 females) and 364 controls (50.4 ± 12.5 years old, 184 males and 180 females) in the Japanese population were analyzed. They were diagnosed according to the DSM-IV criteria. Controls were selected from students, nurses, office workers, and doctors in participating institutes, and their friends. A senior psychiatrist interviewed the controls and found no major mental disorders. Only a subset of the controls were interviewed with the use of a structured interview, the mini-international neuropsychiatric interview (M.I.N.I.) [Sheehan^{Q3}, 1998 #61]. In the Japanese population, no significant population stratification has been repeatedly reported in several studies [Kakiuchi et al., 2003; Arinami et al., 2005; Shimizu et al., 2006].

For the quantitative genomic polymerase chain reaction (PCR), we used genomic DNA derived from LB cells of the two pairs of discordant SZ twins, 46 Japanese unrelated schizophrenia patients (38.6 ± 14.6 years old, 18 males and 28 females), and 11 controls (56.3 ± 11.0 years old, 8 males and 3 females), and 13 schizophrenia patients (55.0 ± 9.9 years old, 9 males and 4 females) obtained from NIMH Genetics Initiative Pedigrees. Written informed consent was obtained from all subjects. The ethics committees of the Brain Science

Institute (RIKEN) and participating institutes approved the study.

Cell Culture

The lymphocytes derived from peripheral blood were transformed by Epstein-Barr (EB) virus and cultured with the use of standard techniques as described before [Kato et al., 2002]. For mRNA quantification by DNA microarray analysis, we extracted the RNA from frozen cells, and thawed and recultured the cells. The culture of the cells and mRNA extraction were performed independently five times for each pair of twins.

DNA Microarray

DNA microarray experiments were performed as described previously with the use of an Affymetrix HU133A chip (Affymetrix, Santa Clara, CA) [Kakiuchi et al., 2006]. We used 5 µg of total RNA for reverse-transcription into cDNA, and biotin-labeled cRNA was synthesized from the cDNA. After testing the integrity of the samples by the Test2Chip (Affymetrix), fragmented cRNA was applied to the HU133A chip. The hybridization signal on the chip was scanned and subjected to image analysis (Affymetrix).

Analysis of DNA Microarray Data

The microarray raw data were processed by MAS5.0 (Affymetrix) and robust multiarray average (RMA) methods [Irizarry et al., 2003], and analyzed with the use of GeneSpring software (SiliconGenetics, Redwood, CA). Data were normalized by the median value. Genes expressed differently in each pair of twins were selected by the following criteria: (1) the genes were called as present in all samples (five samples of affected twin and five samples of control co-twin); (2) both the parametric test and the non-parametric test showed a significant difference ($P < 0.05$) between the five cultures in a patient and five cultures in the co-twin by both normalization methods (MAS5.0 and RMA). Then, the genes commonly changed to the same trend in both SZ twins A and SZ twins B according to these four statistical comparisons: MAS5 and RMA, parametric and non-parametric.

Genetic Association Studies

We selected five SNPs (rs7944706, rs6484148, rs6484147, rs4597056, rs726102) for *ADM* according to the linkage disequilibrium (LD) map database on SNPbrowser™ (Applied Biosystems, Foster City, CA). Although a previous report in the Japanese population hypothesized a possible role of dinucleotide repeat in the 4 kb downstream of *ADM* in the pathophysiology of hypertension, this microsatellite marker was not associated with plasma ADM concentration [Ishimitsu et al., 2001]; thus, this marker was not selected for the analysis. We selected three SNPs (rs9928312, rs9934331, rs1003904) for *SEPX1*, because their TaqMan probes were commercially available and they are polymorphic in Japanese according to the LD map database on HapMap projects accessed with the SNPbrowser™ software. We performed genotyping by TaqMan probes and ABI7900HT according to the protocol recommended by the manufacturer (Applied Biosystems). Assessment of LD patterns by the standardized disequilibrium coefficient (D') and squared correlation coefficient (r^2), and analysis of haplotypic distribution, and frequencies were performed with the use of the COCAPHASE programs (<http://portal.litbio.org/Registered/Option/unphased.html>). Global significance was calculated by the random permutation test (10,000 times).

Quantification of Genome Copy Number

The copy number of *ADM* and *SEPX1* was analyzed by the real-time PCR method with the use of SYBR/GREEN dye

(Applied Biosystems) as described elsewhere [Kato et al., 2005b]. *MLC1* (megalencephalic leukoencephalopathy with subcortical cysts gene 1) was used as a single copy control gene and the copy number of *ADM* was calculated as a relative ratio to *MLC1*. A minimum of three probes for *ADM* was used. For quality control, a gene on the X chromosome [phosphofructo-2-kinase (*PF2K*)] was also examined by SYBR/GREEN dye, and separation between males and females was confirmed. Sequences of primers and probes for these analyses will be provided upon request.

RESULTS

Microarray Analysis in the Cells of Monozygotic Twins Discordant for Schizophrenia

By the criteria described above, five genes were identified (Table I). Among the up-regulated genes in schizophrenia, two genes (GenBank accession nos. L06101 and Z00008) were immunoglobulin-related genes, and *CD200* (GenBank accession no. AF063591) was also a member of the immunoglobulin superfamily (OMIM 155970). This result possibly reflects transformation of a subset of B-cells by the EB virus rather than a difference in disease state. Surprisingly, both of the finally listed genes [*ADM* (GenBank accession no. NM_001124) and *SEPX1* (GenBank accession no. NM_016332)] have been reported to be altered in schizophrenia. The mRNA expression of *ADM* was reported to be up-regulated in the LB cells derived from schizophrenia patients, and the plasma ADM level was significantly higher in schizophrenic patients than in controls [Zoroglu et al., 2002; Huang et al., 2004; Yilmaz et al., 2007]. *SEPX1* was included in the list of genes up-regulated in the peripheral blood cells of schizophrenia by microarray analysis [Glatt et al., 2005]. Interestingly, the expressions of both genes were up-regulated in all the studies, which was the same trend shown in this study. These results suggested that *ADM* and *SEPX1* were strong candidate genes for schizophrenia.

Association Analysis of *ADM* and *SEPX1* in Schizophrenia

If up-regulation of *ADM* and *SEPX1* is a risk factor for schizophrenia, genetic variations of these genes may contribute to the illness. Thus, we also performed association analysis of *ADM* and *SEPX1* in schizophrenia in the Japanese population. We examined the genotype of five SNPs for *ADM* and three SNPs for *SEPX1*. LD patterns for *ADM* and *SEPX1*, as measured by D' and r^2 , are shown in Figure 1. No significant association was observed in single SNPs (Table II) and haplotypes (Table III) for *ADM*, and in single SNPs for *SEPX1* (Table II).

Quantification of Genome Copy Number

In addition to sequence variations, CNVs may also contribute to the up-regulation of *ADM* and *SEPX1*. Indeed, CNVs were reported for the loci of both genes [*ADM* (RP11-79E12) and *SEPX1* (RP11-451K7 and Variation_5329), <http://projects.tcag.ca/variation/>]. The CNV may cause altered mRNA expression and may confound the results of association analysis. Thus, we quantified the copy number of *ADM* and *SEPX1* genes by the real-time PCR method in two pairs of discordant SZ twins, 46 Japanese unrelated schizophrenia patients, and 11 controls, and from genetic information for 13 schizophrenia patients obtained from NIMH Genetics Initiative Pedigrees. However, we observed no loss or gain of the genome in the tested loci (data not shown).

TABLE I. The Result of DNA Microarray Analysis in the Lymphoblastoid Cells of Monozygotic Twins Discordant for Schizophrenia

Probe ID	Genbank	Symbol	SZ twin1				SZ twin2							
			MAS		RMA		MAS		RMA					
			FC	P(P)	P(non-P)	FC	P(P)	P(non-P)	FC	P(P)	P(non-P)			
Up-regulation														
211641_x_at	L06101		1.428	0.042	0.028	1.273	0.022	0.009	1.380	0.030	0.028	1.277	0.011	0.016
217977_at	NM_016332	SEPX1	1.183	0.015	0.047	1.175	0.012	0.016	1.124	0.044	0.028	1.129	0.011	0.028
202912_at	NM_001124	ADM	1.622	0.011	0.028	1.564	0.005	0.009	1.966	0.002	0.009	1.922	0.004	0.009
216517_at	Z00008		1.684	0.022	0.028	1.683	0.038	0.028	2.076	0.000	0.009	1.839	0.001	0.009
Down-regulation														
209583_s_at	AF063591	CD200	0.755	0.016	0.009	0.750	0.019	0.009	0.603	0.003	0.016	0.704	0.005	0.016

FC, fold change; P(P/non-P), *P*-value calculated by parametric/non-parametric test using GeneSpring software.

DISCUSSION

In this study, we demonstrated that mRNA expressions of *ADM* and *SEPX1* were up-regulated in the LB cells of the two patients with schizophrenia compared with their healthy co-twins. This observation is consistent with the previous reports examined in unrelated patients and controls. Genetic association studies of *ADM* and *SEPX1* for schizophrenia in the Japanese population, however, did not support the association of SNPs in these genes with schizophrenia. Further, we did not observe CNVs in these genes.

ADM is a potent vasodilator peptide consisting of 52 amino acids (OMIM103275), which was initially identified from pheochromocytoma [Kitamura et al., 1993]. *ADM* is synthesized by many tissues including the central nervous system and is known to bind to calcitonin receptor-like receptor. The reported roles of *ADM* are variable, such as dilation of blood vessels and increase in urine output. *ADM* is also abundantly expressed in the central nervous system, especially in the thalamus, hypothalamus, and pituitary gland, and it regulates neuroendocrine response to stress [Taylor and Samson, 2004]. Intracerebroventricular administration of *ADM* is known to affect water intake and salt appetite. A probably reactive increase of *ADM* in plasma is reported in some diseases such as heart failure, renal diseases, septic shock, and diabetes mellitus [Beltowski and Jamroz, 2004]. This increased level in plasma was first reported in patients with schizophrenia [Zoroglu et al., 2002]. This observation might reflect reactive up-regulation associated with some somatic condition associated with schizophrenia. However, elevated mRNA levels also were reported in LB cells of schizophrenia patients [Huang et al., 2004], which suggested that increase of *ADM* is intrinsic rather than reactive. In this study, *ADM* mRNA level was increased in the affected co-twins. Thus, intrinsic increase of *ADM* may be related to the pathophysiology of schizophrenia.

SEPX1 is one of the selenoproteins, which includes selenocystein, and is abundant in liver, leucocytes, and pancreas (OMIM 606216). The function of *SEPX1* has not been clarified; however, interestingly, selenium-binding protein1 (*SELENBP1*), which also binds to selenium, was demonstrated to be up-regulated in both the brain and the peripheral blood leukocytes in patients with schizophrenia, and was suggested to be a candidate biomarker of schizophrenia [Glatt et al., 2005]. In the list of genes up-regulated in peripheral blood cells in this report, *SEPX1* was also included. In the present study, *SEPX1* mRNA level was also increased in the affected co-twins. Thus, the up-regulation of *SEPX1* may play a role in the pathophysiology of schizophrenia. Geographical analysis showed that low selenium in soil and food might be associated with schizophrenia [Brown, 1994]. At deficiency selenium is preferentially retained in the brain compared with other organs, and several studies have shown that selenium deficiency is associated with mood [Benton, 2002]. A possible role of selenium transport has been proposed in schizophrenia [Berry, 1993]. Thus, the roles of selenium metabolism in pathophysiology of schizophrenia may merit further study.

Although linkage with schizophrenia and presence of CNVs around the *ADM* and *SEPX1* loci [Yamada et al., 2004; Moon et al., 2006; Redon et al., 2006] prompted us to perform an association study, no association was found. This result suggests that up-regulation of *ADM* and *SEPX1* might be a phenomenon secondary to schizophrenia. However, in the association study, we studied only 223 schizophrenic patients and 364 control subjects. The number of the subjects and the number of SNPs examined are not large enough to totally exclude a possible association between schizophrenia and the SNPs of *SEPX1* and *ADM*. In addition, the result should be treated with caution, because there was a significant difference in gender between patients with schizophrenia and controls ($P < 0.05$).

TABLE II. The Result^{Q4} of Case-Control Studies in Japanese Population

		Genotype			HWE	P -value	Allele		P -value
ADM									
rs7944706		A/A	A/G	G/G			A	G	
	CT	50	176	138	0.606		276	452	
	SZ	35	102	86	0.604	0.748	172	274	0.823
rs6484148		C/C	C/T	T/T			C	T	
	CT	43	166	155	0.887		252	476	
	SZ	25	84	114	0.121	0.117	134	312	0.106
rs6484147		C/C	C/T	T/T			C	T	
	CT	43	166	155	0.887		252	476	
	SZ	25	84	114	0.121	0.117	134	312	0.106
rs4597056		C/C	C/T	T/T			C	T	
	CT	157	163	44	0.865		477	251	
	SZ	114	84	25	0.121	0.160	312	134	0.116
rs726102		A/A	A/G	G/G			A	G	
	CT	42	165	157	0.892		249	479	
	SZ	25	84	114	0.121	0.147	134	312	0.140
SEPX1									
rs9928312		A/A	A/G	G/G			A	G	
	CT	45	175	144	0.464		265	463	
	SZ	27	102	94	0.934	0.820	156	290	0.622
rs9934331		C/C	C/G	G/G			C	G	
	CT	103	176	85	0.559		382	346	
	SZ	61	109	53	0.752	0.969	231	215	0.821
rs1003904		A/A	A/G	G/G			A	G	
	CT	158	150	56	0.044		466	262	
	SZ	78	105	40	0.653	0.129	261	185	0.060

CT, control; SZ, schizophrenia; HWE, Hardy-Weiberg equilibrium. P values are calculated by Fisher's exact test.

With regard to endophenotypes of schizophrenia, mainly psychophysiological, neurocognitive, and neuroimaging findings have been proposed [Gottesman and Gould, 2003]. Relatively few studies focused on blood analysis in schizophrenia. Altered mRNA levels in LB cells were reported for *ADM* [Huang et al., 2004] and *PDLIM5* [Iwamoto et al., 2004]. Alterations in peripheral blood leukocytes mRNA were reported for *SELENBP1* and other candidate genes [Glatt et al., 2005], mitochondria-related transcripts [Whatley et al., 1998; Mehler-Wex et al., 2006], dopamine receptors [Ilani et al., 2001; Kwak et al., 2001; Zvara et al., 2005; Boneberg et al., 2006], alpha 7-nicotinic acetylcholine receptor subunit (*CHRNA7*) [Perl et al., 2006], and transforming growth factor beta receptor II (*TGFB2*) [Numata et al., 2007]. Although none of these candidate mRNA markers in blood cells has been established, it is promising that two genes detected in this study have already been reported in the literature. *ADM* and *SEPX1* are a promising target of further research of biomarkers of schizophrenia.

After our previous report of gene expression analysis in monozygotic twins discordant for bipolar disorder [Kakiuchi et al., 2003], we used the same approach in LB cells [Haas et al.,

2006; Hu et al., 2006; Matigian et al., 2007] or different tissues [Zhou et al., 2005; Cutting and Snowden, 2006; Sarkijarvi et al., 2006]. The present results that two previously reported genes were identified in the twins supported the validity of this methodology. It has been difficult to apply statistical analysis to a limited number of twin samples. Thus, in this study, we performed five independent experiments for each pair of twins. Although it is difficult to prove the validity of this method, it is possible that this extensive analysis enabled the successful selection of these two genes.

In this study, the two pairs of twins discordant for schizophrenia did not have other family history. Thus, the dysregulation of genes in the affected twin is not due to a heritable factor such as a genetic polymorphism, but rather to some environmental or epigenetic effect. Thus, lack of association of the two genes with schizophrenia may be reasonable.

Although we focused on *ADM* and *SEPX1* in this study, the change in *CD200* might also be potentially interesting, because several studies reported that the immune system in schizophrenics may be involved in its susceptibility [Nawa and Takei, 2006]. Moreover, *CD200* has a unique expression pattern that is expressed on B-cells and neurons [Wright et al., 2001].

TABLE III. Haplotype Analysis of ADM in Japanese SZ Samples

Haplotype	SZ	CT	χ^2	P-value	Global P-value
ADM					
A-T-T-C-G	168 (0.380)	270 (0.381)	0.00184	0.965	
G-C-C-T-A	132 (0.298)	238 (0.336)	1.76	0.184	
G-T-T-C-G	142 (0.321)	200 (0.282)	1.94	0.162	0.345

SZ, schizophrenia; CT, control.

Global P-value was calculated by a random permutation test (10,000 times) with the use of COCAPHASE program. Only haplotypes that were verified at least once were analyzed.

A

ADM

	D'	rs7944706	rs6484148	rs6484147	rs4597056	rs726102
r2	rs7944706	1				
	rs6484148	0.3172	1			
	rs6484147	0.3172	1	1		
	rs4597056	0.3093	1	1	1	
	rs726102	0.3114	1	1	1	1

B

SEPX1

	D'	rs9928312	rs9934331	rs1003904
r2	rs9928312	1	0.4592	
	rs9934331	0.6233	1	
	rs1003904	0.2071	0.5721	1

Fig. 1. Intermarker linkage disequilibrium pattern for *ADM* (A) and *SEPX1* (B). The standardized disequilibrium coefficient (D') and squared correlation coefficient (r^2) calculated by the COCAPHASE program are shown for Japanese control samples.

CD200 is expressed in developing neuronal cell bodies and axons [Morris and Beech, 1987]. Thus, *CD200* may be a promising target for further study.

In conclusion, we demonstrated the possible pathological contribution of *ADM* and *SEPX1* to schizophrenia and the usefulness of LB cells of monozygotic twins discordant for schizophrenia.

ACKNOWLEDGMENTS

Data and biomaterials of the NIMH pedigrees were collected in four projects that participated in the NIMH Bipolar Disorder Genetics Initiative. From 1991 to 1998, the principal investigators and co-investigators were Indiana University, Indianapolis, IN, U01 MH46282, J. Nurnberger, M. Miller, and E. Bowman; Washington University, St. Louis, MO, U01 MH46280, T. Reich, A. Goate, and J. Rice; Johns Hopkins University, Baltimore, MD U01 MH46274, J. R. DePaulo, Jr., S. Simpson, and C. Stine; NIMH Intramural Research Program, Clinical Neurogenetics Branch, Bethesda, MD, E. Gershon, D. Kazuba, and E. Maxwell. The authors are grateful to all the subjects who participated in this study. The authors thank Research Resource Center (RRC) of Brain Science Institute, RIKEN, for technical assistance. Funding of this study was provided by a Grant-in-Aid from the Japanese Ministry of Health and Labor (H17-KOKORO-general-009), and a Grant-in-Aid for Exploratory Research (16659307) from The Ministry of Education, Culture, Sports, Science and Technology (MEXT); these agencies had no further role in study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the paper for publication. Authors CK and TK designed the study and wrote the first draft of the manuscript. Author CK performed the experiments and the data analysis. Author MI performed the experiments. Authors SN, NO, NI, TU, MT, KK, TS, AI, and YO contributed to the samples collection and clinical evaluation. All authors approved the final manuscript. The authors declare no conflict of interest.

REFERENCES

- Arinami T, Ohtsuki T, Ishiguro H, Ujike H, Tanaka Y, Morita Y, Mineta M, Takeichi M, Yamada S, Imamura A, et al. 2005. Genomewide high-density SNP linkage analysis of 236 Japanese families supports the existence of schizophrenia susceptibility loci on chromosomes 1p, 14q, and 20p. *Am J Hum Genet* 77(6):937–944.
- Bassett AS, Chow EW. 1999. 22q11 deletion syndrome: A genetic subtype of schizophrenia. *Biol Psychiatry* 46(7):882–891.
- Beltowski J, Jamroz A. 2004. Adrenomedullin—what do we know 10 years since its discovery? *Pol J Pharmacol* 56(1):5–27.
- Benton D. 2002. Selenium intake, mood and other aspects of psychological functioning. *Nutr Neurosci* 5(6):363–374.
- Berman KF, Torrey EF, Daniel DG, Weinberger DR. 1992. Regional cerebral blood flow in monozygotic twins discordant and concordant for schizophrenia. *Arch Gen Psychiatry* 49(12):927–934.
- Berry T. 1993. Possible involvement of selenium transport protein in a subtype of schizophrenia. *Biol Psychiatry* 34(6):422–423.
- Boneberg EM, von Seyditz E, Propster K, Watzl H, Rockstroh B, Illges H. 2006. D3 dopamine receptor mRNA is elevated in T cells of schizophrenic patients whereas D4 dopamine receptor mRNA is reduced in CD4+T cells. *J Neuroimmunol* 173(1–2):180–187.
- Braff DL, Freedman R, Schork NJ, Gottesman ^{???} II. 2007. Deconstructing schizophrenia: An overview of the use of endophenotypes in order to understand a complex disorder. *Schizophr Bull* 33(1):21–32.
- Brown JS Jr. 1994. Role of selenium and other trace elements in the geography of schizophrenia. *Schizophr Bull* 20(2):387–398.
- Cheung VG, Conlin LK, Weber TM, Arcaro M, Jen KY, Morley M, Spielman RS. 2003. Natural variation in human gene expression assessed in lymphoblastoid cells. *Nat Genet* 33(3):422–425.
- Cutting RJ, Snowden JA. 2006. Myeloma in monozygotic twin. *Br J Haematol* 134(6):646.
- Deb-Rinker P, Klempner TA, O'Reilly RL, Torrey EF, Singh SM. 1999. Molecular characterization of a MSR-like sequence identified by RDA from monozygotic twin pairs discordant for schizophrenia. *Genomics* 61(2):133–144.
- Friedhoff AJ, Miller JC, Basham DA. 1995. A subtracted probe derived from lymphocytes of twins discordant for schizophrenia hybridizes to selective areas of rat brain. *Biol Psychiatry* 37(2):127–131.
- Glatt SJ, Everall IP, Kremen WS, Corbeil J, Sasik R, Khanlou N, Han M, Liew CC, Tsuang MT. 2005. Comparative gene expression analysis of blood and brain provides concurrent validation of SELENBP1 up-regulation in schizophrenia. *Proc Natl Acad Sci USA* 102(43):15533–15538.
- Goldberg TE, Torrey EF, Gold JM, Bigelow LB, Ragland RD, Taylor E, Weinberger DR. 1995. Genetic risk of neuropsychological impairment in schizophrenia: A study of monozygotic twins discordant and concordant for the disorder. *Schizophr Res* 17(1):77–84.
- Gottesman ^{???} II, Gould TD. 2003. The endophenotype concept in psychiatry: Etymology and strategic intentions. *Am J Psychiatry* 160(4):636–645.
- Haas CS, Creighton CJ, Pi X, Maine I, Koch AE, Haines GK, Ling S, Chinnaiyan AM, Holoshitz J. 2006. Identification of genes modulated in rheumatoid arthritis using complementary DNA microarray analysis of lymphoblastoid B cell lines from disease-discordant monozygotic twins. *Arthritis Rheum* 54(7):2047–2060.

- Holzman PS, Kringlen E, Levy DL, Proctor LR, Haberman SJ, Yasillo NJ. 1977. Abnormal-pursuit eye movements in schizophrenia. Evidence for a genetic indicator. *Arch Gen Psychiatry* 34(7):802–805.
- Hu VW, Frank BC, Heine S, Lee NH, Quackenbush J. 2006. Gene expression profiling of lymphoblastoid cell lines from monozygotic twins discordant in severity of autism reveals differential regulation of neurologically relevant genes. *BMC Genomics* 7:118.
- Huang CH, Chen ML, Tsai YL, Tsai MT, Chen CH. 2004. Elevated adrenomedullin mRNA in lymphoblastoid cells from schizophrenic patients. *Neuroreport* 15(9):1443–1446.
- Ilani T, Ben-Shachar D, Strous RD, Mazor M, Sheinkman A, Kotler M, Fuchs S. 2001. A peripheral marker for schizophrenia: Increased levels of D3 dopamine receptor mRNA in blood lymphocytes. *Proc Natl Acad Sci USA* 98(2):625–628.
- Irizarry RA, Hobbs B, Collin F, Beazer-Barclay YD, Antonellis KJ, Scherf U, Speed TP. 2003. Exploration, normalization, and summaries of high density oligonucleotide array probe level data. *Biostatistics* 4(2):249–264.
- Ishimitsu T, Hosoya K, Tsukada K, Minami J, Futoh Y, Ono H, Ohru M, Hino J, Kangawa K, Matsuoka H. 2001. Microsatellite DNA polymorphism of human adrenomedullin gene in normotensive subjects and patients with essential hypertension. *Hypertension* 38(1):9–12.
- Iwamoto K, Bundo M, Washizuka S, Kakiuchi C, Kato T. 2004. Expression of HSPF1 and LIM in the lymphoblastoid cells derived from patients with bipolar disorder and schizophrenia. *J Hum Genet* 49(5):227–231.
- Kakiuchi C, Iwamoto K, Ishiwata M, Bundo M, Kasahara T, Kusumi I, Tsujita T, Okazaki Y, Nanko S, Kunugi H, et al. 2003. Impaired feedback regulation of XBP1 as a genetic risk factor for bipolar disorder. *Nat Genet* 35(2):171–175.
- Kakiuchi C, Ishiwata M, Hayashi A, Kato T. 2006. XBP1 induces WFS1 through an endoplasmic reticulum stress response element-like motif in SH-SY5Y cells. *J Neurochem* 97(2):545–555.
- Kato T, Ishiwata M, Nagai T. 2002. Mitochondrial calcium response in human transformed lymphoblastoid cells. *Life Sci* 71(5):581–590.
- Kato T, Iwamoto K, Kakiuchi C, Kuratomi G, Okazaki Y. 2005a. Genetic or epigenetic difference causing discordance between monozygotic twins as a clue to molecular basis of mental disorders. *Mol Psychiatry* 10(7):622–630.
- Kato T, Iwayama Y, Kakiuchi C, Iwamoto K, Yamada K, Minabe Y, Nakamura K, Mori N, Fujii K, Nanko S, et al. 2005b. Gene expression and association analyses of LIM (PDLIM5) in bipolar disorder and schizophrenia. *Mol Psychiatry* 10(11):1045–1055.
- Kitamura K, Kangawa K, Kawamoto M, Ichiki Y, Nakamura S, Matsuo H, Eto T. 1993. Adrenomedullin: A novel hypotensive peptide isolated from human pheochromocytoma. *Biochem Biophys Res Commun* 192(2):553–560.
- Kunugi H, Urushibara T, Murray RM, Nanko S, Hirose T. 2003. Prenatal underdevelopment and schizophrenia: A case report of monozygotic twins. *Psychiatry Clin Neurosci* 57(3):271–274.
- Kwak YT, Koo MS, Choi CH, Sunwoo I. 2001. Change of dopamine receptor mRNA expression in lymphocyte of schizophrenic patients. *BMC Med Genet* 2:3.
- Matigian N, Windus L, Smith H, Filippich C, Pantelis C, McGrath J, Mowry B, Hayward N. 2007. Expression profiling in monozygotic twins discordant for bipolar disorder reveals dysregulation of the WNT signalling pathway. *Mol Psychiatry* 12(4):447–457.
- McDonald P, Lewis M, Murphy B, O'Reilly R, Singh SM. 2003. Appraisal of genetic and epigenetic congruity of a monozygotic twin pair discordant for schizophrenia. *J Med Genet* 40(2):E16.
- Mehler-Wex C, Duvigneau JC, Hartl RT, Ben-Shachar D, Warnke A, Gerlach M. 2006. Increased mRNA levels of the mitochondrial complex I 75-kDa subunit. A potential peripheral marker of early onset schizophrenia? *Eur Child Adolesc Psychiatry* 15(8):504–507.
- Millar JK, Wilson-Annan JC, Anderson S, Christie S, Taylor MS, Semple CA, Devon RS, Clair DM, Muir WJ, Blackwood DH, et al. 2000. Disruption of two novel genes by a translocation co-segregating with schizophrenia. *Hum Mol Genet* 9(9):1415–1423.
- Moon HJ, Yim SV, Lee WK, Jeon YW, Kim YH, Ko YJ, Lee KS, Lee KH, Han SI, Rha HK. 2006. Identification of DNA copy-number aberrations by array-comparative genomic hybridization in patients with schizophrenia. *Biochem Biophys Res Commun* 344(2):531–539.
- Morris RJ, Beech JN. 1987. Sequential expression of OX2 and Thy-1 glycoproteins on the neuronal surface during development. An immunohistochemical study of rat cerebellum. *Dev Neurosci* 9(1):33–44.
- Mutsuddi M, Morris DW, Waggoner SG, Daly MJ, Scolnick EM, Sklar P. 2006. Analysis of high-resolution HapMap of DTNBP1 (Dysbindin) suggests no consistency between reported common variant associations and schizophrenia. *Am J Hum Genet* 79(5):903–909.
- Nawa H, Takei N. 2006. Recent progress in animal modeling of immune inflammatory processes in schizophrenia: Implication of specific cytokines. *Neurosci Res* 56(1):2–13.
- Nguyen GH, Bouchard J, Boselli MG, Tolstol LG, Keith L, Baldwin C, Nguyen NC, Schultz M, Herrera VL, Smith CL. 2003. DNA stability and schizophrenia in twins. *Am J Med Genet Part B* 120B(1):1–10.
- Numata S, Ueno SI, Iga JI, Yamauchi K, Hongwei S, Hashimoto R, Takeda M, Kunugi H, Itakura M, Ohmori T. 2007. TGFBR2 gene expression and genetic association with schizophrenia. *J Psychiatr Res* 41(4):307–314.
- Perl O, Strous RD, Dranikov A, Chen R, Fuchs S. 2006. Low levels of alpha7-nicotinic acetylcholine receptor mRNA on peripheral blood lymphocytes in schizophrenia and its association with illness severity. *Neuropsychobiology* 53(2):88–93.
- Petronis A, Gottesman II, Kan P, Kennedy JL, Basile VS, Paterson AD, Popendikyte V. 2003. Monozygotic twins exhibit numerous epigenetic differences: Clues to twin discordance? *Schizophr Bull* 29(1):169–178.
- Polymeropoulos MH, Xiao H, Torrey EF, DeLisi LE, Crow T, Merrill CR. 1993. Search for a genetic event in monozygotic twins discordant for schizophrenia. *Psychiatry Res* 48(1):27–36.
- Rapaport MH, Torrey EF, McAllister CG, Nelson DL, Pickar D, Paul SM. 1993. Increased serum soluble interleukin-2 receptors in schizophrenic monozygotic twins. *Eur Arch Psychiatry Clin Neurosci* 243(1):7–10.
- Redon R, Ishikawa S, Fitch KR, Feuk L, Perry GH, Andrews TD, Fiegler H, Shaperon MH, Carson AR, Chen W, et al. 2006. Global variation in copy number in the human genome. *Nature* 444(7118):444–454.
- Reveley AM, Reveley MA, Clifford CA, Murray RM. 1982. Cerebral ventricular size in twins discordant for schizophrenia. *Lancet* 1(8271):540–541.
- Reveley MA, Reveley AM, Clifford CA, Murray RM. 1983. Genetics of platelet MAO activity in discordant schizophrenic and normal twins. *Br J Psychiatry* 142:560–565.
- Sarkijarvi S, Kuusisto H, Paalavuo R, Levula M, Airla N, Lehtimäki T, Kaprio J, Koskenvuo M, Elovaara I. 2006. Gene expression profiles in Finnish twins with multiple sclerosis. *BMC Med Genet* 7:11.
- Shih RA, Belmonte PL, Zandi PP. 2004. A review of the evidence from family, twin and adoption studies for a genetic contribution to adult psychiatric disorders. *Int Rev Psychiatry* 16(4):260–283.
- Shimizu H, Iwayama Y, Yamada K, Toyota T, Minabe Y, Nakamura K, Nakajima M, Hattori E, Mori N, Osumi N, et al. 2006. Genetic and expression analyses of the STOP (MAP6) gene in schizophrenia. *Schizophr Res* 84(2–3):244–252.
- Sierra-Honigsmann AM, Carbone KM, Yolken RH. 1995. Polymerase chain reaction (PCR) search for viral nucleic acid sequences in schizophrenia. *Br J Psychiatry* 166(1):55–60.
- Stefansson H, Sigurdsson E, Steinthorsdottir V, Bjornsdottir S, Sigmundsson T, Ghosh S, Brynjolfsson J, Gunnarsdottir S, Ivarsson O, Chou TT, et al. 2002. Neuregulin 1 and susceptibility to schizophrenia. *Am J Hum Genet* 71(4):877–892.
- Straub RE, Jiang Y, MacLean CJ, Ma Y, Webb BT, Myakishev MV, Harris-Kerr C, Wormley B, Sadek H, Kadambi B, et al. 2002. Genetic variation in the 6p22.3 gene DTNBP1, the human ortholog of the mouse dysbindin gene, is associated with schizophrenia. *Am J Hum Genet* 71(2):337–348.
- Suddath RL, Christison GW, Torrey EF, Casanova MF, Weinberger DR. 1990. Anatomical abnormalities in the brains of monozygotic twins discordant for schizophrenia. *N Engl J Med* 322(12):789–794.
- Taylor MM, Samson WK. 2004. A possible mechanism for the action of adrenomedullin in brain to stimulate stress hormone secretion. *Endocrinology* 145(11):4890–4896.
- Tsang TM, Huang JT, Holmes E, Bahn S. 2006. Metabolic profiling of plasma from discordant schizophrenia twins: Correlation between lipid signals and global functioning in female schizophrenia patients. *J Proteome Res* 5(4):756–760.
- Tsujita T, Niikawa N, Yamashita H, Imamura A, Hamada A, Nakane Y, Okazaki Y. 1998. Genomic discordance between monozygotic twins discordant for schizophrenia. *Am J Psychiatry* 155(3):422–424.
- Vander Putten DM, Torrey EF, Larive AB, Merrill CR. 1996. Plasma protein variations in monozygotic twins discordant for schizophrenia. *Biol Psychiatry* 40(6):437–442.

- Vincent JB, Kalsi G, Klempan T, Tatuch Y, Sherrington RP, Breschel T, McInnis MG, Brynjolfsson J, Petursson H, Gurling HM, et al. 1998. No evidence of expansion of CAG or GAA repeats in schizophrenia families and monozygotic twins. *Hum Genet* 103(1):41–47.
- Walker EF, Bonsall R, Walder DJ. 2002. Plasma hormones and catecholamine metabolites in monozygotic twins discordant for psychosis. *Neuropsychiatry Neuropsychol Behav Neurol* 15(1):10–17.
- Whatley SA, Curti D, Das Gupta F, Ferrier IN, Jones S, Taylor C, Marchbanks RM. 1998. Superoxide, neuroleptics and the ubiquinone and cytochrome b5 reductases in brain and lymphocytes from normals and schizophrenic patients. *Mol Psychiatry* 3(3):227–237.
- Wright GJ, Jones M, Puklavec MJ, Brown MH, Barclay AN. 2001. The unusual distribution of the neuronal/lymphoid cell surface CD200 (OX2) glycoprotein is conserved in humans. *Immunology* 102(2):173–179.
- Yamada K, Iwayama-Shigeno Y, Yoshida Y, Toyota T, Itokawa M, Hattori E, Shimizu H, Yoshikawa T. 2004. Family-based association study of schizophrenia with 444 markers and analysis of a new susceptibility locus mapped to 11q13.3. *Am J Med Genet Part B* 127B(1):11–19.
- Yilmaz N, Herken H, Cicek HK, Celik A, Yurekli M, Akyol O. 2007. Increased levels of nitric oxide, cortisol and adrenomedullin in patients with chronic schizophrenia. *Med Princ Pract* 16(2):137–141.
- Zhou X, Tan FK, Xiong M, Arnett FC, Feghali-Bostwick CA. 2005. Monozygotic twins clinically discordant for scleroderma show concordance for fibroblast gene expression profiles. *Arthritis Rheum* 52(10): 3305–3314.
- Zoroglu SS, Herken H, Yurekli M, Uz E, Tutkun H, Savas HA, Bagci C, Ozen ME, Cengiz B, Cakmak EA, et al. 2002. The possible pathophysiological role of plasma nitric oxide and adrenomedullin in schizophrenia. *J Psychiatr Res* 36(5):309–315.
- Zvara A, Szekeres G, Janka Z, Kelemen JZ, Cimmer C, Santha M, Puskas LG. 2005. Over-expression of dopamine D2 receptor and inwardly rectifying potassium channel genes in drug-naive schizophrenic peripheral blood lymphocytes as potential diagnostic markers. *Dis Markers* 21(2):61–69.

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Evidence for Acquired Pregenual Anterior Cingulate Gray Matter Loss from a Twin Study of Combat-Related Posttraumatic Stress Disorder

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Background: Controversy exists over the nature and origin of reduced regional brain volumes in posttraumatic stress disorder (PTSD). At issue is whether these reductions represent preexisting vulnerability factors for developing PTSD upon traumatic exposure or acquired PTSD signs due to the traumatic stress that caused the PTSD or the chronic stress of having the disorder (or both). We employed a case-control design in monozygotic twin pairs discordant for combat exposure to address the preexisting versus acquired origin of brain morphometric abnormalities in PTSD.

Methods: We used voxel-based morphometry to search for gray matter density reductions in magnetic resonance imaging (MRI) data obtained in a previous study of combat-exposed Vietnam veteran twins with ($n = 18$) versus without ($n = 23$) PTSD and their "high-risk" versus "low-risk" (respectively) identical combat-unexposed cotwins.

Results: Compared with the combat-exposed twins without PTSD, the combat-exposed twins with PTSD showed significant gray matter density reductions in four predicted brain regions: right hippocampus, pregenual anterior cingulate cortex (ACC), and left and right insulae. There was a significant PTSD Diagnosis \times Combat Exposure interaction in pregenual ACC in which combat-exposed PTSD twins had lower gray matter density than their own combat-unexposed cotwins as well as than the combat-exposed twins without PTSD and their cotwins.

Conclusions: The results point to gray matter volume diminutions in limbic and paralimbic structures in PTSD. The pattern of results obtained for pregenual ACC suggests that gray matter reduction in this region represents an acquired sign of PTSD consistent with stress-induced loss.

Key Words: Anterior cingulate gyrus, brain, magnetic resonance imaging, monozygotic twins, posttraumatic stress disorder

Several structural magnetic resonance imaging (MRI) studies employing anatomic segmentation have found lower gray matter volumes in the hippocampus in posttraumatic stress disorder (PTSD) stemming from various traumatic events (1). One segmentation study found diminished gray matter volumes in pregenual anterior cingulate cortex (ACC) and subcallosal cortex but not dorsal ACC (2), whereas another did find dorsal ACC reduction (3). Decreased pregenual ACC activation in response to trauma-related stimuli is a prominent functional neuroimaging finding in PTSD (4–5).

The technique of voxel-based morphometry (VBM) allows an automated examination of structural brain differences using statistical parametric mapping (SPM) techniques. The validity of the VBM technique for assessing regional gray matter density compared with conventional region of interest measurements has been confirmed in several previous studies (6–9). Employing VBM, the first authors found reduced dorsal ACC gray matter

density in victims of an urban terrorist attack with PTSD (10). Another recent study that employed VBM found gray matter density reduction in pregenual ACC but not dorsal ACC, although manual segmentation did not confirm volumetric reduction in the former structure (11). That study also found gray matter density reduction in left insula. Yet another recent PTSD VBM study found gray matter density reductions in hippocampus, pregenual ACC, and insula (12).

Controversy exists over the nature and origin of reduced regional brain volume in PTSD. Thus far the debate has focused on the hippocampus (13). At issue is whether reduced volume represents an acquired PTSD sign, for example, is due to the traumatic stress that caused the PTSD or the chronic stress of having PTSD (or both) or is a preexisting vulnerability factor for developing PTSD upon traumatic exposure. We have been employing a case-control design in monozygotic twin pairs discordant for combat exposure in Vietnam to address the preexisting versus acquired origin of biological abnormalities found in PTSD (14). In a structural MRI study that manually traced the outlines of the right and left hippocampus, we found that lower total hippocampal volume constituted a "familial" vulnerability factor for PTSD because it was found in both the combat-exposed twins with PTSD and their "high-risk" combat-unexposed cotwins whose hippocampal volumes were lower than those of the combat-exposed twins without PTSD and their "low-risk," combat-unexposed cotwins (15). (Note that the term "familial" includes both heredity and shared environment, i.e., environmental experiences that both members of a twin pair have had in common.)

In this study, we applied a VBM analysis to MRI data from the same twin sample to conduct a search throughout the entire brain for regional gray matter structural differences. On the basis of the published structural imaging studies just reviewed, we

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Received April 5, 2007; revised June 19, 2007; accepted June 26, 2007.

0006-3223/08/\$34.00
doi:10.1016/j.biopsych.2007.06.022

BIOL PSYCHIATRY 2008;63:550–556
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predicted lower gray matter density in combat-exposed twins with PTSD compared with combat-exposed twins without PTSD in the following regions: hippocampus, dorsal ACC, pregenual ACC, subcallosal cortex, and insula. In an attempt to clarify the origin of any such differences, we used the data from the combat-unexposed cotwins. Gray matter diminution that confers familial vulnerability to PTSD would be expected in the high-risk, compared with the low-risk, combat-unexposed twins. In contrast, diminution that reflects acquired damage in PTSD would be expected to be manifest in a Diagnosis \times Combat Exposure interaction in which the combat-exposed twins with PTSD had lower gray matter density than their own high-risk, combat-unexposed cotwins as well as the combat-exposed twins without PTSD and their low-risk cotwins.

Methods and Materials

Subjects

The strategy for subject ascertainment and recruitment has been presented elsewhere (16). The sample was described in detail in the report of our previous hippocampus manual tracing study (15). This study analyzed the same MRI scans from the same subjects. In the previous study, one combat-exposed twin with PTSD and his combat-unexposed cotwin were removed from the analysis because the former was an extreme, asymmetrical outlier for manually traced hippocampal volume. This subject and his cotwin were included in the current study. (Exclusion of this pair did not alter the conclusions.) The protocol was approved by the institutional review board of the Manchester, New Hampshire, VA Medical Center. All subjects had previously given written informed consent before participation after a complete description of the procedures.

MRI Data Preanalysis

The MRI acquisition techniques were described in the previous report (15). The methods used to analyze these data in the present study were similar to those reported elsewhere (10). Image analysis was performed using ANALYZE PC 3.0 (Mayo Foundation, Rochester, Minnesota) and SPM 99 software (Wellcome Department of Cognitive Neurology, Institute of Neurology, London, United Kingdom) running in MATLAB 6.1 (Mathworks, Sherborn, Massachusetts). In ANALYZE, image data were resampled using an algorithm to make them isotropic, with the sides measuring .9375 mm, and then stored. Resampled images were first spatially normalized into the standard MNI152 template (17,18). Normalized images were then segmented into gray matter, white matter, cerebrospinal fluid, and skull and scalp compartments using an automated, operator-independent process (19). The segmentation step also incorporated an image density nonuniformity correction to address image density variations caused by various positions of cranial structures within the MRI head coil (20). The spatially normalized segments of the gray matter were smoothed with a 12-mm full-width at half-maximum isotropic Gaussian kernel to accommodate individual variability in sulcal and gyral anatomy. For medial temporal regions (e.g., hippocampus), a 4-mm smoothing kernel was used instead, as has been recommended (21). By smoothing the data, the partial volume effect was used to create a spectrum of gray matter densities. Gray matter density is equivalent to the weighted average of the gray matter voxels located in the volume defined by the smoothing kernel. Because previous studies have shown a fair correlation between regional gray matter density determined by VBM and structural volumes measured by conven-

tional, manual tracing (7,9,22), the regional gray matter density can be considered to represent the local volume of gray matter.

Statistical Analyses

Demographic and psychometric data were analyzed by means of a mixed model that treated Diagnosis (in the combat-exposed twin) as a between-pairs fixed effect, Combat Exposure as a within-pairs fixed effect (repeated measure), and pairs as a random effect (16). This model analysis yields a t statistic. Gray matter density was estimated on a voxel-by-voxel basis using SPM 99 (23). Contrasts were made between the 18 combat-exposed twins with PTSD and the 23 combat-exposed twins without PTSD, and separately between their high- and low-risk cotwins, using independent t tests, adjusted for individual intracranial volume. Diagnosis \times Exposure interactions were evaluated by means of a mixed, multigroup (Diagnosis), conditions (Combat Exposure), and covariates (intracranial volume) model in which one twin pair was treated as though one subject with two conditions. In this analysis, 82 covariates were entered corresponding to 82 images ($(18 + 23) \times 2$). For each of the foregoing analyses, a statistical parametric map (SPM) of the t statistic (SPM(t)) was created, and the SPM(t) values were transformed to the normal distribution (SPM(z)). The statistical significance threshold was set at $p < .05$ corrected for multiple comparisons using the False Discovery Rate (FDR) (24).

The anatomic locations of peak coordinates were initially defined using the latest version of Talairach Daemon Client (25). These localizations were then confirmed by visually inspecting the coordinates overlaid on the mean structural image of the study sample. For peaks located within predicted brain regions, small volume correction was applied using the following a priori volume approximations from the literature: hippocampus 3.5 mL each side; insula 8 mL each side; dorsal ACC 10 mL bilaterally; pregenual ACC 5 mL bilaterally; and subcallosal cortex 5 mL bilaterally (total volume of predicted brain regions = 43 mL). For peaks located outside predicted regions, correction for whole brain was employed. Because the predictions were directional, namely, lower gray matter density in combat-exposed subjects and PTSD pairs, the tests were one-tailed, and only results in the predicted direction(s) are reported.

Results

Demographics and Psychometrics. Group mean age, Combat Severity Score (26), Total Clinician-Administered PTSD Scale (CAPS) score (27), Number of potentially traumatic lifetime noncombat events (16), Total Michigan Alcoholism Screening Test (MAST) score (28), and Symptom Checklist-90-Revised (SCL-90-R) Depression Scale score (29), along with statistical analyses, are presented in Table 1. It may be seen that age was similar among subject groups. Combat-exposed twins with PTSD had more severe combat exposure than combat-exposed twins without PTSD. As expected by virtue of selection, the former had greater combat-related symptom severity on the CAPS. Twin pairs with PTSD (i.e., twin pairs in which the combat-exposed twin was diagnosed with current, combat-related PTSD) reported more potentially traumatic lifetime noncombat events than non-PTSD pairs (i.e., twin pairs in which the combat-exposed twin was diagnosed with neither current nor past combat-related PTSD). Combat-exposed twins also reported more potentially traumatic lifetime noncombat events than their combat-unexposed cotwins. Combat-exposed twins with PTSD had more severe alcoholism histories than the other three groups. Combat-exposed twins with PTSD also reported more depression than

Table 1. Group Means (SD) of Combat-Exposed Vietnam Veterans with and without Posttraumatic Stress Disorder (PTSD) and Their Combat-Unexposed, Identical Cotwins

	PTSD Pairs ^a		Non-PTSD Pairs ^b		Mixed Model						Independent t Tests			
	Exposed (n = 18)	Unexposed (High Risk) (n = 18)	Exposed (n = 23)	Unexposed (Low Risk) (n = 23)	Diagnosis		Exposure		Interaction		Exposed		Unexposed	
					t 39	p	t 40	p	t 39	p	t 39	p	t 39	p
Age (years) ^c	52.8 (3.4)	52.8 (3.4)	51.8 (2.3)	51.8 (2.3)	—	—	—	—	—	—	1.1	.27	1.1	.27
CAPS ^d	73.3 (16.9)	—	6.2 (7.3)	—	—	—	—	—	—	—	17.2	<.001	—	—
Combat Severity ^e	7.7 (2.1)	—	3.5 (2.6)	—	—	—	—	—	—	—	5.6	<.001	—	—
Traumatic Events ^f	8.1 (2.6)	5.2 (3.7)	5.1 (4.0)	4.2 (3.0)	2.6	.01	2.4	.02	1.3	.20	2.8	.009	1.0	.34
MAST ^g	19.1 (17.6)	6.4 (10.2)	2.4 (4.5)	2.5 (4.0)	4.5	<.001	2.8	.007	2.8	.007	4.4	<.001	1.7	.10
Depression ^h	2.5 (.9)	.3 (.5)	.6 (.7)	.4 (.5)	4.6	<.001	5.3	<.001	6.9	<.001	7.4	<.001	.7	.51

^aAs determined by the presence of current combat-related PTSD in the combat-exposed twin.

^bAs determined by the absence of current or past combat-related PTSD in the combat-exposed twin.

^cAs of October 1, 2000.

^dClinician-Administered PTSD Scale (range 0–136).

^e18-item measure (range 0–18).

^fNumber of potentially traumatic lifetime noncombat events.

^gMichigan Alcoholism Screening Test (range 0–25).

^hSymptom Check List-90-Revised Depression Subscale (range 0–4).

the other three groups. The Pearson Correlation Coefficient between total CAPS score and SCL-90-R Depression was large ($r = .86$) in the PTSD pairs but negligible ($r = -.06$) in the non-PTSD pairs.

Gray Matter Density

Table 2 presents the results of the contrasts between combat-exposed twins with versus without PTSD. For the sake of a complete exposition of these data, all results statistically significant at the liberal threshold of uncorrected $p < .001$ with spatial extent of $k > 10$ voxels are shown. Of the seven peaks that met this threshold, four were located in predicted brain regions, namely, right hippocampus, pregenual ACC, right midinsula, and left anterior insula (shown in Figure 4). Each of these four peaks also met the statistical significance threshold of $p < .05$ with small volume correction for the a priori size of the structure (as shown in the second column of Table 2). No voxels in nonpredicted brain regions met the threshold of $p < .05$ corrected for whole brain in these, or any other, analyses.

At the right hippocampus, pregenual ACC, left anterior insula, and right midinsula (shown in Table 2, within the 18 PTSD combat veterans, we examined the correlations between gray matter density and total CAPS score, as well as SCL-90-R Depression score. Because these analyses involved single voxels, the

Table 2. Loci Showing Gray Matter Density Reductions in Combat-Exposed Twins with versus without Combat-Related Posttraumatic Stress Disorder That Were Significant at Uncorrected $p < .001$

z	p_{sv}	k	[x y z]	Brain Region
4.50	.001	248	[44 -2 -14]	Right midinsula
4.47		145	[-62 -56 -4]	Left middle temporal gyrus
4.39	.001	27	[34 -28 -16]	Right hippocampus
3.95	.005	236	[-36 10 -4]	Left anterior insula
3.71	.004	185	[0 46 10]	Pregenual anterior cingulate cortex
3.52		35	[-46 42 -14]	Left inferior frontal gyrus
3.47		22	[-64 -44 10]	Left superior temporal gyrus

k, cluster size; p_{sv} , significance level with small volume correction on the basis of the a priori size of the structure; [x y z] = Montréal Neurological Institute coordinates of peak voxel.

Predicted areas appear in boldface.

significance threshold was $p < .05$ uncorrected. None of these correlations were significant. We also performed the same correlations for CAPS A (reexperiencing), B (avoidance/numbing), and C (hyperarousal) symptom cluster subscores; for these analyses we applied a Bonferroni correction to the significance threshold, namely, $p < .017$ (.05/3). The only significant correlations were between symptom cluster B (reexperiencing) and gray matter density in pregenual ACC ($r = -.57$, $p = .008$), left anterior insula ($r = -.53$, $p = .0130$), and right midinsula ($r = -.59$, $p = .006$).

The contrasts between the high-risk, combat-unexposed cotwins of the combat-exposed twins with PTSD versus the low-risk, combat-unexposed cotwins of the combat-exposed twins without PTSD did not identify any voxels that met the statistical significance threshold of $p < .05$, even with small volume corrections. The only statistically significant Diagnosis \times Exposure interaction was found in pregenual ACC (8 50 12), $z = 3.32$, $k = 16$, $p = .02$ corrected for the a priori size of this structure. The location of this cluster is shown in Figure 2. Scatterplots of individual subjects' values at the [8 50 12] pregenual ACC locus are shown in Figure 3. There were no significant correlations between gray matter density in the 18 PTSD combat veterans minus gray matter density in their combat-unexposed cotwins, and total CAPS score, SCL-90-R Depression, or (with Bonferroni corrections) any of the CAPS symptom cluster subscores.

The mixed model and t test analyses that yielded the statistically significant results described earlier were repeated entering the following possibly confounding variables into the respective models as covariates: age, combat severity (in the exposed twin), number of potentially traumatic lifetime noncombat events, MAST score, and SCL-90-R Depression score. To control for a possibly confounding role of childhood physical or sexual abuse, the data were re-analyzed deleting pairs within which either member had such a history. All these results are shown in Table 3.

Discussion

Of the seven loci at which combat-exposed twins with PTSD had lower gray matter density than combat-exposed twins without PTSD at a liberal threshold of $p < .001$, four were located in predicted brain regions, namely, right hippocampus, pre-

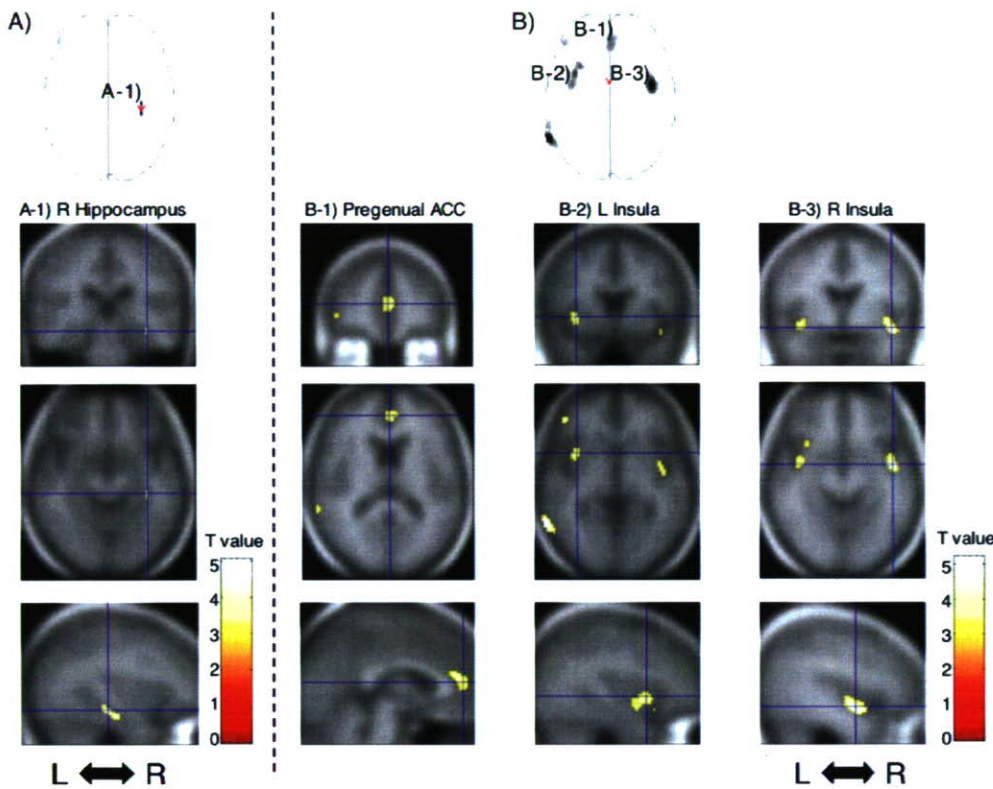


Figure 1. Brain regions showing diminution in gray matter density in combat-exposed twins with posttraumatic stress disorder (PTSD) versus those without PTSD. **(A)** Statistical parametric mapping (SPM) analysis with 4-mm Gaussian smoothing kernel revealed statistically significant reduced gray matter density shown in the axial projection. **(A-1)** Regional gray matter density reduction in the right hippocampus is rendered onto orthogonal slices of the averaged magnetic resonance image of the present study's subjects. **(B)** SPM analysis with 12-mm Gaussian smoothing kernel revealed statistically significant reduced gray matter density shown in the axial projections. Regional gray matter density reductions in the following areas are rendered: **(B-1)** pregenual anterior cingulate cortex; **(B-2)** left insula; **(B-3)** right insula. L, left hemisphere; R, right hemisphere; ACC, anterior cingulate cortex.

genual ACC, and left anterior and right midinsula, even though the predicted brain regions occupy less than 10% of total gray matter volume. This regional specificity supports the validity of our results and implicates limbic and paralimbic structures as the major sites of gray matter density reductions in combat-related PTSD. Gray matter reductions in pregenual ACC and both insulae significantly correlated only with the cluster B “reexperiencing” symptoms of PTSD.

The ACC, especially its pregenual (or “affective”) division, and insula are components of the anterior “paralimbic belt,” are strongly interconnected to each other and to the amygdala, and are highly involved in emotional aspects of brain function (30–32). Impaired pregenual ACC function is one of the most

robust neuroimaging findings in PTSD (4–5). A neurocircuitry model of PTSD posits that the ventromedial prefrontal cortex, including pregenual ACC, inhibits the expression of classically conditioned fear responses by the amygdala (33). Thus impairment in this brain region might be expected to most affect the DSM-IV symptoms that are putatively most closely related to conditioned fear, namely, the cluster B symptoms (especially B.4 and B.5, that is, intense psychological distress (and/or) physiological reactivity on exposure to internal or external cues that symbolize or resemble an aspect of the traumatic event). To the extent that diminished structure implies diminished function, reduced pregenual ACC gray matter density is consistent with this neurocircuitry model.

Table 3. Results Adjusted for Potentially Confounding Variables

Unadjusted	Age	Combat Severity ^a	Traumatic Events ^b	MAST Score ^c	Childhood Abuse ^d	SCL-90-R Depression ^e	Brain Region
<i>z</i> (<i>p</i> _{svcl})	<i>z</i> (<i>p</i> _{svcl})	<i>z</i> (<i>p</i> _{svcl})	<i>z</i> (<i>p</i> _{svcl})	<i>z</i> (<i>p</i> _{svcl})	<i>z</i> (<i>p</i> _{svcl})	<i>z</i> (<i>p</i> _{svcl})	
Combat-Exposed Twins: PTSD vs. non-PTSD							
4.50 (.001)	4.43 (.001)	3.23 (.05)	3.98 (.005)	3.39 (.02)	3.19 (.04)	2.07 (.49)	Right midinsula [44 –2 –14]
4.39 (.001)	4.36 (.001)	3.05 (.11)	4.11 (.004)	4.63 (<.001)	3.56 (.04)	1.52 (.99)	Right hippocampus [34 –28 –16]
3.95 (.005)	3.98 (.005)	3.32 (.03)	3.06 (.07)	3.26 (.04)	3.63 (.01)	2.15 (.44)	Left anterior insula [–36 10 –4]
3.71 (.004)	3.59 (.009)	2.50 (.18)	2.79 (.08)	2.40 (.14)	2.92 (.03)	.93 (.99)	Pregenual anterior cingulate cortex [0 46 10]
Diagnosis × Combat Exposure Interaction							
3.32 (.02)	3.39 (.02)	3.32 (.02) ^f	2.76 (.10)	2.77 (.10)	3.42 (.01)	1.81 (.53)	Pregenual anterior cingulate cortex [8 50 12]

*p*_{svcl} significance level with small volume correction; PTSD, posttraumatic stress disorder.

^a18-item measure (range 0–18).

^bNumber of potentially traumatic lifetime noncombat events.

^cMichigan Alcoholism Screening Test (range 0–25).

^dDeleting six PTSD pairs and five non-PTSD pairs within which either twin had a history of childhood abuse.

^eDepression subscale of Symptom Check List 90—Revised.

^fCovariate is value in combat-exposed twin.

Diagnosis x Exposure Interaction

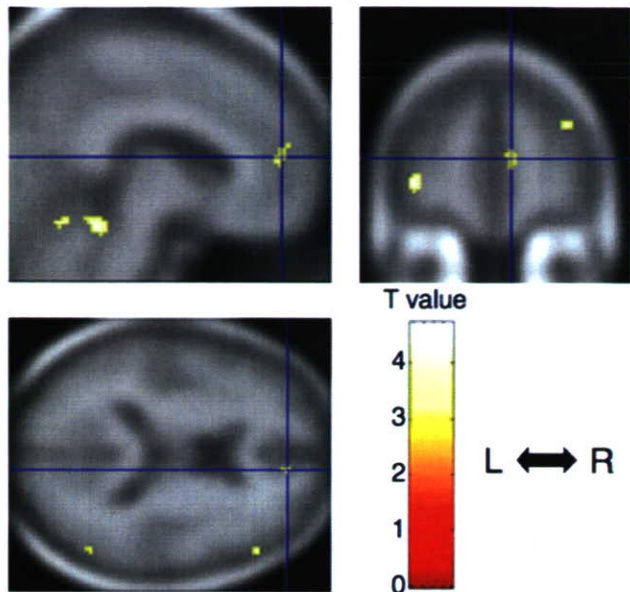


Figure 2. Brain region showing posttraumatic stress disorder Diagnosis × Combat Exposure interaction. Regional interaction for gray matter density in pregenual anterior cingulate cortex is rendered onto orthogonal slices of the averaged magnetic resonance image of the study subjects. Crosshairs indicate the peak coordinate of the interaction [8 50 12]. L, left hemisphere; R, right hemisphere.

Functional neuroimaging studies of the hippocampus in PTSD are less common, but they, too, support impairment in this brain region (34–36). Hippocampal impairment may contribute to PTSD by reducing the ability to construct declarative narratives that bind the affect associated with the traumatic event (37), by the ability to recognize safe contexts (33), or by other unknown mechanisms. The reduced gray matter density found in bilateral insulae is paradoxical in light of studies that have generally found hyperactivity in this brain region in PTSD (38) and other anxiety conditions (39). One model of anterior insula function posits that this structure detects the difference between an observed and expected body state and generates an interoceptive prediction signal that triggers anxiety (39). A structurally compromised insula may be less inhibited in generating such signals in PTSD, but this is in the realm of speculation.

The most interesting result from our study is the significant Diagnosis × Exposure interaction in the pregenual ACC, with combat-exposed PTSD twins having lower gray matter density than their own combat-unexposed cotwins and than the combat-exposed twins without PTSD and their cotwins, supporting the inference that pregenual ACC gray matter reduction is an acquired sign of PTSD. In animals, exposure to chronic stress has been shown to damage not only the hippocampus in rodents (40) and primates (41) but also the ACC in rodents (42,43) and primates (44). It has been hypothesized that such damage may provide a basis for structural changes observed in PTSD (42,45). A recent study of mentally healthy persons that used automated segmentation found that those who reported early life stressors had smaller ACCs than those who did not (46). However, causal inferences are difficult to draw from the cross-sectional study of nontwins.

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When the Diagnosis × Exposure interaction at [8 50 12] was adjusted for MAST score, its statistical significance level was reduced to corrected $p = .10$, which falls short of statistical significance. We did not obtain data regarding recent alcohol consumption. This is a limitation considering that imaging findings related to alcohol may be more sensitive to recent as opposed to more remote intake. On the other hand, the likelihood that increased alcohol use by the PTSD veterans accounts for the gray matter density reduction in their ACCs is diminished by the consideration that if the PTSD subjects studied here had consumed enough alcohol to damage their brains, evidence for this should have been found in other brain regions known to be affected by alcohol, including superior, motor, and other areas of the frontal cortex and cerebellum (47,48), none of which (except for a small cluster in left inferior frontal cortex) showed volumetric reduction in the PTSD compared with the non-PTSD combat veterans at even the liberal threshold of uncorrected $p < .001$. Thus the specificity of volumetric diminution to our predicted brain regions argues against a global effect such as alcohol-induced brain damage. Finally, a recent manual tracing study found comparably (and significantly) reduced ACC volume in subgroups of PTSD veterans with and without a history of lifetime alcohol abuse or dependence, in comparison to non-PTSD veterans (49).

When the Diagnosis × Exposure interaction in pregenual ACC was adjusted for number of potentially traumatic lifetime noncombat events, its statistical significance level was also reduced to corrected $p = .10$. This means we cannot be fully confident that stressful events other than military combat do not account for the reduced ACC gray matter density in the PTSD veterans. However, even if such events did contribute, this would still not be inconsistent with stress-induced diminution of this structure. When the Diagnosis × Exposure interaction in pregenual ACC was adjusted for depression, it was no longer significant. This is not surprising given the high association between depression and PTSD in our sample, in which self-

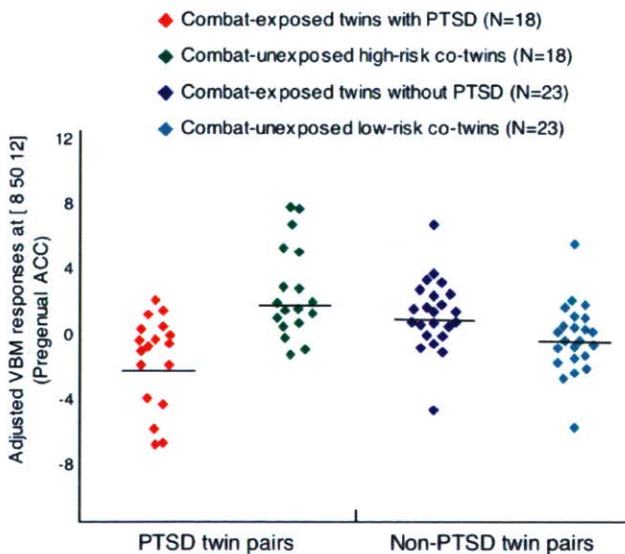


Figure 3. Scatterplots of individual subjects' adjusted voxel-based morphometry responses. Shown at the site of the posttraumatic stress disorder Diagnosis × Combat Exposure interaction in pregenual anterior cingulate cortex [8 50 12]. Means are represented by solid horizontal lines drawn on each group's distribution.

reported depression appears to have been acquired along with PTSD, making the two likely facets of the same posttraumatic psychopathology.

However, because the most salient, common difference in our study was the presence of combat-related PTSD in the former, and because, as noted earlier, the observed effects remained significant or nearly significant after considering the contributions of several important potentially confounding variables, it is reasonable to attribute this lower gray matter density to the presence of combat-related PTSD.

Combat-exposed twins with PTSD also had lower gray matter density than combat-exposed twins without PTSD in right hippocampus and left anterior and right midinsula, as well as at another site within pregenual ACC, replicating previous studies. These results could not be explained by group differences in age, combat severity, number of potentially traumatic lifetime non-combat events, alcoholism, or child abuse. Unfortunately, the analyses that included the data from the combat-unexposed cotwins were unable to shed light on the origin of these gray matter reductions in the combat-exposed twins with PTSD, because they failed to yield either a significant difference between high- and low-risk combat-unexposed cotwins (which would support a pretrauma vulnerability factor) or a significant Diagnosis \times Exposure interaction (which would support an acquired abnormality). Finally, these results were unable to replicate previously reported segmentation and voxel-based morphometric findings of gray matter reduction in subcallosal cortex and dorsal ACC.

In the same twin sample studied here, we previously found manual tracing evidence that diminished hippocampal volume represents a pretrauma vulnerability factor for PTSD, rather than an acquired PTSD sign (15). In contrast, our VBM results suggest that diminished volume in pregenual ACC is acquired as a result of the combat exposure that led to PTSD, the PTSD itself, or both. We have no ready explanation as to why diminutions in these two structures should have different origins. As noted earlier, however, the origin of gray matter density reduction in the pregenual ACC site other than the one that showed the significant interaction could not be explicated by our data; it is possible that it represents a PTSD vulnerability factor. Additional techniques that may help to clarify this uncertainty in future studies include cortical parcellation (segmentation) and magnetic resonance spectroscopy.

It has been suggested that VBM may not detect very small, localized gray matter volume reductions because false-negative VBM findings may arise from the changes in the shape or displacement of structures in the course of spatial normalization (7). Additionally, VBM may be biased against finding group differences in areas that are spatially complex (50). Inversely, we cannot rule out the possibility that the abnormalities detected by VBM in our study reflected group differences in the shape of brain structures rather than their volume (11), although even shape differences may have functional consequences. The failure of VBM to find a significant hippocampal gray matter reduction in the high- versus low-risk, combat-unexposed cotwins contrasts with our positive result in the same sample using manual segmentation of hippocampus (15) suggests that the latter technique may be more sensitive to reduced volume in this structure than the voxel-based approach. Similarly, we are unable to rule out the possibility that subtle group differences in other brain regions in this study remained below the sensitivity of VBM or the detection power conferred by our sample.

KK and HY contributed equally to this work.

This work was supported by Special Coordination Funds for Promoting Science and Technology from the Ministry of Education, Culture, Sports, Science and Technology of the Government of Japan to KK; a U.S. Department of Veterans Affairs Merit Review Grant to Dr. MWG; U.S. Public Health Service (USPHS) Grant No. R01MH54636 to RKP; and USPHS Grant No. K05MH01110 to MES. Professor Karl Friston provided helpful suggestions regarding the SPM analyses. The authors thank H. Matsuda, T. Ohnishi, M. Macklin, S. Williston, L. Paulus, and H. Croteau for assistance. The U.S. Department of Veterans Affairs provided financial support for the development and maintenance of the Vietnam Era Twin (VET) Registry. Through their support of the VET Registry, numerous other U.S. organizations also provided invaluable assistance, including the Department of Defense; National Personnel Records Center, National Archives and Records Administration; Internal Revenue Service; National Institutes of Health; National Opinion Research Center; National Research Council, National Academy of Sciences; and Institute for Survey Research, Temple University. We acknowledge the continued cooperation and participation of the members of the VET Registry and their families, without whose contribution this research would not have been possible.

There are no conflicts of interest to disclose for any of the authors. This study was wholly federally funded and did not involve treatment, medications, or devices.

1. Kitayama N, Vaccarino V, Kutner M, Weiss P, Bremner JD (2005): Magnetic resonance imaging (MRI) measurement of hippocampal volume in posttraumatic stress disorder: A meta-analysis. *J Affect Disord* 88:79–86.
2. Rauch SL, Shin LM, Segal E, Pitman RK, Carson MA, McMullin K, *et al.* (2003): Selectively reduced regional cortical volumes in post-traumatic stress disorder. *Neuroreport* 14:913–916.
3. Kitayama N, Quinn S, Bremner JD (2006): Smaller volume of anterior cingulate cortex in abuse-related posttraumatic stress disorder. *J Affect Disord* 90:171–174.
4. Bremner JD (2006): Traumatic stress: Effects on the brain. *Dialogues Clin Neurosci* 8:445–461.
5. Milad MR, Rauch SL, Pitman RK, Quirk GJ (2006): Fear extinction in rats: Implications for human brain imaging and anxiety disorders. *Biol Psychol* 2006;73:61–71.
6. Vargha-Khadem F, Watkins KE, Price CJ, Ashburner J, Alcock KJ, Connelly A, *et al.* (1998): Neural basis of an inherited speech and language disorder. *Proc Natl Acad Sci U S A* 95:12695–12700.
7. Wright IC, Ellison ZR, Sharma T, Friston KJ, Murray RM, McGuire PK (1999): Mapping of grey matter changes in schizophrenia. *Schizophr Res* 35:1–14.
8. Suzuki M, Nohara S, Hagino H, Kurokawa K, Yotsutsuji T, Kawasaki Y, *et al.* (2002): Regional changes in brain gray and white matter in patients with schizophrenia demonstrated with voxel-based analysis of MRI. *Schizophr Res* 55:41–54.
9. Luders E, Gaser C, Jancke L, Schlaug G (2004): A voxel-based approach to gray matter asymmetries. *Neuroimage* 22:656–664.
10. Yamasue H, Kasai K, Iwanami A, Ohtani T, Yamada H, Abe O, *et al.* (2003): Voxel-based analysis of MRI reveals anterior cingulate gray-matter volume reduction in posttraumatic stress disorder due to terrorism. *Proc Natl Acad Sci U S A* 100:9039–9043.
11. Corbo V, Clément MH, Armory JL, Pruessner JC, Brunet A (2005): Size vs. shape differences: Contrasting voxel-based and volumetric analyses of the anterior cingulate cortex in individuals with PTSD. *Biol Psychiatry* 58:119–124.
12. Chen S, Xia W, Li L, Liu J, He Z, Zhang Z, *et al.* (2006): Gray matter density reduction in the insula in fire survivors with posttraumatic stress disorder: A voxel-based morphometric study. *Psychiatry Res* 146:65–72.
13. Sapolsky RM (2001): Atrophy of the hippocampus in posttraumatic stress disorder: How and when? *Hippocampus* 11:90–91.
14. Pitman RK, Gilbertson MW, Gurvits TV, May FS, Lasko NB, Metzger LJ, *et al.* (2006): Clarifying the origin of biological abnormalities in PTSD