

260 after injury than the immature neonatal brain. In addition to  
 261 regional differences, we should thus consider developmen-  
 262 tal age when trying to apply a regenerative therapy to  
 263 developing brain.

264 **Transplantation of neural stem cells**

265 Zheng et al. (2006) extracted multipotent astrocytic stem  
 266 cells from the subependymal zones of homozygous green  
 267 fluorescence protein (GFP) transgenic neonatal mice at  
 268 postnatal days 1–6. These cells are positive for glial  
 269 fibrillary acidic protein and negative for beta-III tubulin  
 270 and NeuN, markers of mature neurons (Laywell et al.  
 271 2000). The multipotent astrocytic stem cells were trans-  
 272 planted into Sprague-Dawley rat brains 24 h or 5 days after  
 273 hypoxia-ischemia on postnatal day 7 (Zheng et al. (2006).  
 274 GFP-positive cells successfully transformed into neurons  
 275 positive for beta-III tubulin and NeuN and were located  
 276 around the infarcted area, far from the injection site. This  
 277 observation was however not quantitative, and furthermore,  
 278 the authors did not describe any histological or behavioral  
 279 improvements.

280 In a study by Park et al (2002b), the clonal multipotent  
 281 neural precursor cell line (C17.2) was derived from the  
 282 external germinal layer of the neonatal mouse cerebellum;  
 283 these neural stem/progenitor cells were cultured with a  
 284 polymer scaffold (polyglycolic acid, PGA), for 4 days,  
 285 making a neural stem/progenitor cell-PGA complex. The  
 286 neural stem/progenitor cell-PGA matrix was transplanted  
 287 into the degenerated cavity of mouse brain, 7 days after  
 288 hypoxia-ischemia on postnatal day 7. The transplanted  
 289 neural stem cells not only differentiated into neurons,  
 290 oligodendrocytes, and astrocytes, but also made neural  
 291 networks with both donor-derived neurons and host-derived  
 292 neurons. This neuronal network extended through the  
 293 corpus callosum into the contralateral hemisphere. Al-  
 294 though Park et al (2002b) reported that neurological  
 295 function, as assessed by diminished unilateral rotation,  
 296 seemed to be improved, they did not quantitatively analyze  
 297 behavioral improvement.

298 **Exogenous enrichment of the milieu around the injury**

299 Another way to facilitate regeneration may be the enrichment  
 300 of the milieu around the injured region. Neurotrophic factors  
 301 play an important role in the development and maintenance of  
 302 specific populations of neurons in the central and peripheral  
 303 nervous system. Several neurotrophic factors have been  
 304 reported to help rescue neurons and repair neuronal injury.  
 305 These include nerve growth factor (NGF), brain-derived  
 306 neurotrophic factor (BDNF), glial-cell line-derived neuro-

trophic factor (GDNF), insulin-like growth factor, basic 307  
 fibroblast growth factor (bFGF), transforming growth factor 308  
 (TGF)-beta 3, neurotrophin-3 (NT-3), NT-4/5, and NT-6 309  
 (Gong et al. 1999; Hyman et al. 1994; Ikeda et al. 2000; 310  
 Kriegstein et al. 1996; Zawada et al. 1998). Although these 311  
 neurotrophic factors are also known to be upregulated after 312  
 hypoxia-ischemia in the neonatal rat (Ikeda et al. 2002), this 313  
 endogenous upregulation seems to be insufficient to support 314  
 neural differentiation from progenitor cells. 315

To date, bFGF-2 seems to be the most promising 316  
 candidate to facilitate neuroregeneration in neonatal bran 317  
 damage (Wagner et al. 1999). Monfils et al. (2006, 2005) 318  
 administered 50 ng/kg bFGF for 7 days after brain lesions 319  
 to the motor cortex of a neonatal rat and observed 320  
 significant neuronal recovery and synapse formation in 321  
 the treated group. This was accompanied with improved 322  
 forelimb movement. Although this injury is traumatic, 323  
 bFGF could be applied after hypoxic-ischemic injury with 324  
 or without another neurotrophic factor such as EGF 325  
 (Nakatomi et al. 2002). 326

Recently, Park et al. (2006b) transplanted a subclone of 327  
 neural stem cells, transduced with a retrovirus encoding 328  
 NT-3, into a site of unilateral hypoxic-ischemic brain 329  
 damage in neonatal mice. They observed a greater than 330  
 80% increase in neurons derived from neural stem cells 331  
 compared with those from non-transduced stem cells. 332  
 Donor-derived stem cells were shown to differentiate 333  
 successfully into cholinergic, gamma-aminobutyric-acid- 334  
 ergic, or glutamatergic subtypes of neuron. These findings 335  
 indicate the importance of adjusting the environment 336  
 around the injured area for successful stem cell therapy. 337

**Concluding remarks**

338  
 339 Although considerable potential with respect to prolifera-  
 340 tion and migration has been revealed in the immature and  
 341 developing brain, limited terminal differentiation may  
 342 hamper stem cell therapy. Aggressive efforts to adjust the  
 343 environment of the damaged part in which tissue regenera-  
 344 tion is occurring or more cautious stem cell transplantation  
 345 will probably be required.

**Acknowledgement** I thank Professor Dr. Alistair J. Gunn, Professor 346  
 of the Liggins Institute and Department of Paediatrics, New Zealand, 348  
 for expert suggestions and proofreading. 349

**References**

350  
 351 Back SA, Luo NL, Borenstein NS, Levine JM, Volpe JJ, Kinney HC  
 352 (2001) Late oligodendrocyte progenitors coincide with the  
 353 developmental window of vulnerability for human perinatal  
 354 white matter injury. *J Neurosci* 21:1302–1312

355 Back SA, Han BH, Luo NL, Chricton CA, Xanthoudakis S, Tam J, Arvin KL, Holtzman DM (2002) Selective vulnerability of late oligodendrocyte progenitors to hypoxia-ischemia. *J Neurosci* 22:455–463

359 Badawi N, Kurinczuk JJ, Keogh JM, Alessandri LM, O'Sullivan F, Burton PR, Pemberton PJ, Stanley FJ (1998a) Antepartum risk factors for newborn encephalopathy: the Western Australian case-control study. *BMJ* 317:1549–1553

363 Badawi N, Kurinczuk JJ, Keogh JM, Alessandri LM, O'Sullivan F, Burton PR, Pemberton PJ, Stanley FJ (1998b) Intrapartum risk factors for newborn encephalopathy: the Western Australian case-control study. *BMJ* 317:1554–1558

367 Daval JL, Pourie G, Grojean S, Lievre V, Strazielle C, Blaise S, Vert P (2004) Neonatal hypoxia triggers transient apoptosis followed by neurogenesis in the rat CA1 hippocampus. *Pediatr Res* 55:561–567

371 Felling RJ, Snyder MJ, Romanko MJ, Rothstein RP, Ziegler AN, Yang Z, Givogri MI, Bongarzone ER, Levison SW (2006) Neural stem/progenitor cells participate in the regenerative response to perinatal hypoxia/ischemia. *J Neurosci* 26:4359–4369

375 Francis F, Koulakoff A, Boucher D, Chafey P, Schaar B, Vinet MC, Friocourt G, McDonnell N, Reiner O, Kahn A, McConnell SK, Berwald-Netter Y, Denoulet P, Chelly J (1999) Doublecortin is a developmentally regulated, microtubule-associated protein expressed in migrating and differentiating neurons. *Neuron* 23:247–256

381 Gleeson JG, Lin PT, Flanagan LA, Walsh CA (1999) Doublecortin is a microtubule-associated protein and is expressed widely by migrating neurons. *Neuron* 23:257–271

384 Gluckman PD, Gunn AJ, Wyatt JS (2006) Hypothermia for neonates with hypoxic-ischemic encephalopathy. *N Engl J Med* 354:1643–1645

387 Gong L, Wyatt RJ, Baker I, Masserano JM (1999) Brain-derived and glial cell line-derived neurotrophic factors protect a catecholaminergic cell line from dopamine-induced cell death. *Neurosci Lett* 263:153–156

391 Hagberg H, Ichord R, Palmer C, Yager JY, Vannucci SJ (2002) Animal models of developmental brain injury: relevance to human disease. A summary of the panel discussion from the Third Hershey Conference on Developmental Cerebral Blood Flow and Metabolism. *Dev Neurosci* 24:364–366

396 Hankins GD, Speer M (2003) Defining the pathogenesis and pathophysiology of neonatal encephalopathy and cerebral palsy. *Obstet Gynecol* 102:628–636

399 Hayashi T, Iwai M, Ikeda T, Jin G, Deguchi K, Nagotani S, Zhang H, Sehara Y, Nagano I, Shoji M, Ikenoue T, Abe K (2005) Neural precursor cells division and migration in neonatal rat brain after ischemic/hypoxic injury. *Brain Res* 1038:41–49

403 Hyman C, Juhasz M, Jackson C, Wright P, Ip NY, Lindsay RM (1994) Overlapping and distinct actions of the neurotrophins BDNF, NT-3, and NT-4/5 on cultured dopaminergic and GABAergic neurons of the ventral mesencephalon. *J Neurosci* 14:335–347

407 Ikeda T, Xia XY, Xia YX, Ikenoue T, Han B, Choi BH (2000) Glial cell line-derived neurotrophic factor protects against ischemia/hypoxia-induced brain injury in neonatal rat. *Acta Neuropathol (Berl)* 100:161–167

411 Ikeda T, Koo H, Xia YX, Ikenoue T, Choi BH (2002) Bimodal upregulation of glial cell line-derived neurotrophic factor (GDNF) in the neonatal rat brain following ischemic/hypoxic injury. *Int J Dev Neurosci* 20:555–562

415 Ikeda T, Iwai M, Hayashi T, Nagano I, Shoji M, Ikenoue T, Abe K (2005) Limited differentiation to neurons and astroglia from neural stem cells in the cortex and striatum after ischemia/hypoxia in the neonatal rat brain. *Am J Obstet Gynecol* 193:849–856

Iwai M, Sato K, Omori N, Nagano I, Manabe Y, Shoji M, Abe K (2002) Three steps of neural stem cells development in gerbil dentate gyrus after transient ischemia. *J Cereb Blood Flow Metab* 22:411–419

Iwai M, Sato K, Kamada H, Omori N, Nagano I, Shoji M, Abe K (2003) Temporal profile of stem cell division, migration, and differentiation from subventricular zone to olfactory bulb after transient forebrain ischemia in gerbils. *J Cereb Blood Flow Metab* 23:331–341

Iwai M, Ikeda T, Hayashi T, Sato K, Nagata T, Nagano I, Shoji M, Ikenoue T, Abe K (2006) Temporal profile of neural stem cell proliferation in the subventricular zone after ischemia/hypoxia in the neonatal rat brain. *Neurol Res* 28:461–468

Jin K, Minami M, Lan JQ, Mao XO, Bateur S, Simon RP, Greenberg DA (2001) Neurogenesis in dentate subgranular zone and rostral subventricular zone after focal cerebral ischemia in the rat. *Proc Natl Acad Sci USA* 98:4710–4715

Kriegstein K, Maysinger D, Unsicker K (1996) The survival response of mesencephalic dopaminergic neurons to the neurotrophins BDNF and NT-4 requires priming with serum: comparison with members of the TGF-beta superfamily and characterization of the serum-free culture system. *J Neural Transm Suppl* 47:247–258

Laywell ED, Rakic P, Kukekov VG, Holland EC, Steindler DA (2000) Identification of a multipotent astrocytic stem cell in the immature and adult mouse brain. *Proc Natl Acad Sci USA* 97:13883–13888

Miller JT, Bartley JH, Wimborne HJ, Walker AL, Hess DC, Hill WD, Carroll JE (2005) The neuroblast and angioblast chemotactic factor SDF-1 (CXCL12) expression is briefly up regulated by reactive astrocytes in brain following neonatal hypoxic-ischemic injury. *BMC Neurosci* 6:63

Mishima K, Ikeda T, Aoo N, Takai N, Takahashi S, Egashira N, Ikenoue T, Iwasaki K, Fujiwara M (2005) Hypoxia-ischemic insult in neonatal rats induced slowly progressive brain damage related to memory impairment. *Neurosci Lett* 376:194–199

Monfils MH, Driscoll I, Vandenberg PM, Thomas NJ, Danka D, Kleim JA, Kolb B (2005) Basic fibroblast growth factor stimulates functional recovery after neonatal lesions of motor cortex in rats. *Neuroscience* 134:1–8

Monfils MH, Driscoll I, Kamitakahara H, Wilson B, Flynn C, Teskey GC, Kleim JA, Kolb B (2006) FGF-2-induced cell proliferation stimulates anatomical, neurophysiological and functional recovery from neonatal motor cortex injury. *Eur J Neurosci* 24:739–749

Morshead CM, Kooy D van der (1992) Postmitotic death is the fate of constitutively proliferating cells in the subependymal layer of the adult mouse brain. *J Neurosci* 12:249–256

Nakatomi H, Kuriu T, Okabe S, Yamamoto S, Hatano O, Kawahara N, Tamura A, Kirino T, Nakafuku M (2002) Regeneration of hippocampal pyramidal neurons after ischemic brain injury by recruitment of endogenous neural progenitors. *Cell* 110:429–441

Ong J, Plane JM, Parent JM, Silverstein FS (2005) Hypoxic-ischemic injury stimulates subventricular zone proliferation and neurogenesis in the neonatal rat. *Pediatr Res* 58:600–606

Ourednik V, Ourednik J, Flax JD, Zawada WM, Hutt C, Yang C, Park KI, Kim SU, Sidman RL, Freed CR, Snyder EY (2001) Segregation of human neural stem cells in the developing primate forebrain. *Science* 293:1820–1824

Parer JT, Ikeda T (2007) A framework for standardized management of intrapartum fetal heart rate patterns. *Am J Obstet Gynecol* 197(26):e1–e6

Park KI, Ourednik J, Ourednik V, Taylor RM, Aboody KS, Auguste KI, Lachyankar MB, Redmond DE, Snyder EY (2002a) Global gene and cell replacement strategies via stem cells. *Gene Ther* 9:613–624

Park KI, Teng YD, Snyder EY (2002b) The injured brain interacts reciprocally with neural stem cells supported by scaffolds to reconstitute lost tissue. *Nat Biotechnol* 20:1111–1117

- 485 Park KI, Hack MA, Ourednik J, Yandava B, Flax JD, Stieg PE, 508  
 486 Gullans S, Jensen FE, Sidman RL, Ourednik V, Snyder EY 509  
 487 (2006a) Acute injury directs the migration, proliferation, and 510  
 488 differentiation of solid organ stem cells: evidence from the effect 511  
 489 of hypoxia-ischemia in the CNS on clonal "reporter" neural stem 512  
 490 cells. *Exp Neurol* 199:156–178 513  
 491 Park KI, Himes BT, Stieg PE, Tessler A, Fischer I, Snyder EY (2006b) 514  
 492 Neural stem cells may be uniquely suited for combined gene 515  
 493 therapy and cell replacement: evidence from engraftment of 516  
 494 neurotrophin-3-expressing stem cells in hypoxic-ischemic brain 517  
 495 injury. *Exp Neurol* 199:179–190 518  
 496 Plane JM, Liu R, Wang TW, Silverstein FS, Parent JM (2004) Neonatal 519  
 497 hypoxic-ischemic injury increases forebrain subventricular zone 520  
 498 neurogenesis in the mouse. *Neurobiol Dis* 16:585–595 521  
 499 Qiu L, Zhu C, Wang X, Xu F, Eriksson PS, Nilsson M, Cooper-Kuhn 522  
 500 CM, Kuhn HG, Blomgren K (2007) Less neurogenesis and 523  
 501 inflammation in the immature than in the juvenile brain after 524  
 502 cerebral hypoxia-ischemia. *J Cereb Blood Flow Metab* 27:785–794 525  
 503 Riet JE van de, Vandenbussche FP, Le Cessie S, Keirse MJ (1999) 526  
 504 Newborn assessment and long-term adverse outcome: a system- 527  
 505 atic review. *Am J Obstet Gynecol* 180:1024–1029 528  
 506 Vannucci RC (1990) Experimental biology of cerebral hypoxia-ischemia: 529  
 507 relation to perinatal brain damage. *Pediatr Res* 27:317–326 530
- Volpe JJ (2000) *Neurology of the neonate*, 4th edn. Saunders, Philadelphia 508
- Wagner JP, Black IB, DiCicco-Bloom E (1999) Stimulation of neonatal and adult brain neurogenesis by subcutaneous injection of basic fibroblast growth factor. *J Neurosci* 19:6006–6016 510
- Yang Z, Levison SW (2006) Hypoxia/ischemia expands the regenerative capacity of progenitors in the perinatal subventricular zone. *Neuroscience* 139:555–564 513
- Yang Z, Covey MV, Bitel CL, Ni L, Jonakait GM, Levison SW (2007) Sustained neocortical neurogenesis after neonatal hypoxic-ischemic injury. *Ann Neurol* 61:199–208 516
- Zawada WM, Zastrow DJ, Clarkson ED, Adams FS, Bell KP, Freed CR (1998) Growth factors improve immediate survival of embryonic dopamine neurons after transplantation into rats. *Brain Res* 786:96–103 519
- Zhang RL, Zhang ZG, Zhang L, Chopp M (2001) Proliferation and differentiation of progenitor cells in the cortex and the subventricular zone in the adult rat after focal cerebral ischemia. *Neuroscience* 105:33–41 523
- Zheng T, Rossignol C, Leibovici A, Anderson KJ, Steindler DA, Weiss MD (2006) Transplantation of multipotent astrocytic stem cells into a rat model of neonatal hypoxic-ischemic encephalopathy. *Brain Res* 1112:99–105 527

UNCORRECTED PROOF

# Clinical Aspects of Type-1 Long-QT Syndrome by Location, Coding Type, and Biophysical Function of Mutations Involving the KCNQ1 Gene

Arthur J. Moss, MD\*; Wataru Shimizu, MD, PhD\*; Arthur A.M. Wilde, MD, PhD\*;  
Jeffrey A. Towbin, MD\*; Wojciech Zareba, MD, PhD; Jennifer L. Robinson, MS; Ming Qi, PhD;  
G. Michael Vincent, MD; Michael J. Ackerman, MD, PhD; Elizabeth S. Kaufman, MD;  
Nynke Hofman, MSc; Rahul Seth, MD; Shiro Kamakura, MD, PhD; Yoshihiro Miyamoto, MD, PhD;  
Ilan Goldenberg, MD; Mark L. Andrews, BBA; Scott McNitt, MS

**Background**—Type-1 long-QT syndrome (LQTS) is caused by loss-of-function mutations in the KCNQ1-encoded  $I_{Ks}$  cardiac potassium channel. We evaluated the effect of location, coding type, and biophysical function of KCNQ1 mutations on the clinical phenotype of this disorder.

**Methods and Results**—We investigated the clinical course in 600 patients with 77 different KCNQ1 mutations in 101 proband-identified families derived from the US portion of the International LQTS Registry (n=425), the Netherlands' LQTS Registry (n=93), and the Japanese LQTS Registry (n=82). The Cox proportional hazards survivorship model was used to evaluate the independent contribution of clinical and genetic factors to the first occurrence of time-dependent cardiac events from birth through age 40 years. The clinical characteristics, distribution of mutations, and overall outcome event rates were similar in patients enrolled from the 3 geographic regions. Biophysical function of the mutations was categorized according to dominant-negative (>50%) or haploinsufficiency ( $\leq$ 50%) reduction in cardiac repolarizing  $I_{Ks}$  potassium channel current. Patients with transmembrane versus C-terminus mutations (hazard ratio, 2.06;  $P<0.001$ ) and those with mutations having dominant-negative versus haploinsufficiency ion channel effects (hazard ratio, 2.26;  $P<0.001$ ) were at increased risk for cardiac events, and these genetic risks were independent of traditional clinical risk factors.

**Conclusions**—This genotype–phenotype study indicates that in type-1 LQTS, mutations located in the transmembrane portion of the ion channel protein and the degree of ion channel dysfunction caused by the mutations are important independent risk factors influencing the clinical course of this disorder. (*Circulation*. 2007;115:2481-2489.)

**Key Words:** electrocardiography ■ genetics ■ long-QT syndrome

The hereditary long-QT syndrome (LQTS) is characterized by prolonged ventricular repolarization on the ECG and arrhythmia-related syncope and sudden death.<sup>1</sup> Mutations in 1 or more of several ion channel genes are known to cause this disorder,<sup>2</sup> with mutations in the KCNQ1 gene causing the type-1 long-QT syndrome.<sup>3,4</sup> The KCNQ1 gene codes for the potassium channel protein responsible for the slow component of the delayed rectifier repolarizing current ( $I_{Ks}$ ). Mutations involving this gene result in reduction of the repolarizing  $I_{Ks}$  current and lengthening of the QT interval.<sup>3</sup>

## Clinical Perspective p 2489

Functional  $I_{Ks}$  channels result from the coassembly of 4 subunits into a tetrameric protein channel that is transported to the myocyte membrane. Each subunit contains 6 membrane-spanning domains (S1 to S6) flanked by amino (N)- and carboxyl (C)-terminus regions. Two distinct biophysical mechanisms mediate the reduced  $I_{Ks}$  current in patients with KCNQ1 mutations: (1) coassembly or trafficking defects in which mutant subunits are not transported

Received September 17, 2006; accepted March 2, 2007.

From the Cardiology Division (A.J.M., W.Z., J.L.R., I.G., M.L.A., S.M., R.S.) of the Department of Medicine and the Department of Pathology (M.Q.), University of Rochester School of Medicine and Dentistry, Rochester, NY; Division of Cardiology, Department of Internal Medicine (W.S., S.K.) and Laboratory of Molecular Genetics (Y.M.), National Cardiovascular Center, Suita, Japan; Departments of Clinical and Experimental Cardiology (A.A.M.W.) and Clinical Genetics (N.H.), Academic Medical Center, Amsterdam, the Netherlands; Department of Pediatrics, Baylor College of Medicine, Texas Children's Hospital, Houston (J.A.T.); School of Medicine, University of Utah, Salt Lake City (G.M.V.); Departments of Medicine, Pediatrics, and Molecular Pharmacology, Mayo Clinic College of Medicine, Rochester, Minn (M.J.A.); and Heart and Vascular Research Center, MetroHealth Campus of Case Western Reserve University, Cleveland, Ohio (E.S.K.).

\*The first 4 authors contributed equally to this work.

The online-only Data Supplement, consisting of references, is available with this article at <http://circ.ahajournals.org/cgi/content/full/CIRCULATIONAHA.106.665406/DC1>.

Correspondence to Arthur J. Moss, MD, Heart Research Follow-Up Program, Box 653, University of Rochester Medical Center, Rochester, NY 14642-8653. E-mail [heartajm@heart.rochester.edu](mailto:heartajm@heart.rochester.edu)

© 2007 American Heart Association, Inc.

*Circulation* is available at <http://www.circulationaha.org>

DOI: 10.1161/CIRCULATIONAHA.106.665406

Downloaded from [circ.ahajournals.org](http://circ.ahajournals.org) at National Cardiovascular Center on July 11, 2007

properly to the cell membrane and fail to incorporate into the tetrameric channel, with the net effect being a  $\leq 50\%$  reduction in channel function (haploinsufficiency)<sup>5</sup>; and (2) formation of defective channels involving mutant subunits with the altered channel protein transported to the cell membrane, resulting in a dysfunctional channel having  $> 50\%$  reduction in channel current (dominant-negative effect).<sup>6</sup>

Limited prior studies involving relatively small numbers of patients with type-1 LQTS have been reported with conflicting results on the relationship between various KCNQ1 mutations and the clinical outcome.<sup>7,8</sup> We hypothesized that the location, coding type, and functional effect of the channel mutation would have important influence on the phenotypic manifestations and clinical course of patients with this disorder. To test this hypothesis, we investigated the clinical aspects of a large cohort of subjects having a spectrum of KCNQ1 mutations categorized by their location, coding type, and type of biophysical ion channel dysfunction.

## Methods

### Study Population

The study population of 600 subjects with genetically confirmed KCNQ1 mutations was derived from 101 proband-identified families with the type-1 LQTS disorder. The proband in each family had QTc prolongation not due to a known cause. The subjects were drawn from the US portion of the International LQTS Registry (n=425), the Netherlands' LQTS Registry (n=93), and the Japanese LQTS Registry (n=82). All subjects or their guardians provided informed consent for the genetic and clinical studies.

### Phenotype Characterization

Routine clinical and ECG parameters were acquired at the time of enrollment in each of the registries. Follow-up was censored at age 41 years to avoid the influence of coronary disease on cardiac events. Measured parameters on the first recorded ECG included QT and R-R intervals in milliseconds, with QT corrected for heart rate by Bazett's formula. The QTc interval was expressed in its continuous form and categorized into 3 levels:  $< 500$ , 500 to 530, and  $> 530$  ms. Clinical data were collected on prospectively designed forms with information on demographic characteristics, personal and family medical history, ECG findings, therapy, and end points during long-term follow-up. LQTS-related cardiac events included syncope, aborted cardiac arrest, or unexpected sudden death without a known cause. Data common to all 3 LQTS registries involving genetically identified patients with type-1 genotype were electronically merged into a common database for the present study.

### Genotype Characterization

The KCNQ1 mutations were identified with the use of standard genetic tests performed in academic molecular-genetic laboratories including the Functional Genomics Center, University of Rochester Medical Center, Rochester, NY; Baylor College of Medicine, Houston, Tex; Mayo Clinic College of Medicine, Rochester, Minn; Boston Children's Hospital, Boston, Mass; Laboratory of Molecular Genetics, National Cardiovascular Center, Suita, Japan; and Department of Clinical Genetics, Academic Medical Center, Amsterdam, Netherlands.

Genetic alterations of the amino acid sequence were characterized by location and by the specific mutation (missense, splice site, in-frame insertions/deletions, nonsense, stop codon, and frameshift). The transmembrane region of the KCNQ1-encoded channel was defined as the coding sequence involving amino acid residues from 120 through 355 (S5-pore-S6 region 285 to 355), with the N-terminus region defined before residue 120 and the C-terminus region after residue 355. Nineteen study patients had intron mutations predicted to disrupt the canonical splice-site domains. Fifty-one

subjects died of sudden cardiac death at a young age but did not have genotype studies. These 51 subjects were assumed to have the same KCNQ1 mutation as other affected members of their respective family. Twelve subjects had 2 mutations, one in the KCNQ1 gene and a second mutation in another LQTS ion channel gene; these 12 subjects are described separately and are not included in any of the tables or outcome analyses. Subjects with Jervell and Lange-Nielsen syndrome with deafness and 2 KCNQ1 mutations as well as those with 1 known KCNQ1 mutation and congenital deafness are not included in the present study.

The biophysical function of the mutant channels was classified as having dominant-negative effect ( $> 50\%$  reduction in function) or haploinsufficiency ( $\leq 50\%$  reduction in function) on the basis of the following: (1) cellular expression studies for those with missense (n=21) and nonsense (n=2) mutations reported in the literature, with the functional information derived exclusively from heterologous expression studies; and (2) assumed loss of function for identified nonsense, splice site, in-frame deletion, and frameshift mutations (n=10) that have not yet been functionally characterized. Forty-one missense mutations and the 3 intron mutations that have not been functionally reported in cellular expression studies were categorized as unknown in terms of type of functional perturbation.

### Statistical Analysis

Differences in the univariate characteristics by specific groupings were evaluated by standard statistical methods. The primary end point was time to syncope, aborted cardiac arrest, or sudden death, whichever occurred first. The cumulative probability of a first cardiac event was assessed by the Kaplan-Meier method with significance testing by the log-rank statistic. The Cox proportional hazards survivorship model was used to evaluate the independent contribution of clinical and genetic factors to the first occurrence of time-dependent cardiac events from birth through age 40 years.<sup>9</sup> Stratified and unstratified Cox regression models, allowing for time-dependent covariates, were fit to estimate the adjusted hazard ratio of each factor as a predictor of first cardiac events. We observed that sex was not proportional as a function of age with crossover in risk at age 13 years on univariate Kaplan-Meier analysis. To relax the assumption of proportional hazards for sex over the entire age range, separate nonparametric baseline hazard functions were allowed for male and female subjects via the stratified Cox model; then, to summarize the sex effect, sex was modeled in an unstratified Cox model as a time-dependent covariate (via an interaction with time), allowing for different hazard ratios by sex before and after age 13 years.

Because almost all the subjects were first- and second-degree relatives of probands, the effect of lack of independence between subjects was evaluated in the Cox model with grouped jackknife estimates for family membership.<sup>10</sup> All grouped jackknife standard errors for the covariate risk factors fell within 3% of those obtained from the unadjusted Cox model, and therefore only the Cox model findings are reported.

Patients who died suddenly at a young age from suspected LQTS and who did not have an ECG for QTc measurement were identified in the Cox models as "QTc missing." Prespecified covariate interactions were evaluated. The influence of time-dependent  $\beta$ -blocker therapy (the age at which  $\beta$ -blocker therapy was initiated) on outcome was determined by adding this variable to the final Cox model containing the various covariates.

The authors had full access to and take full responsibility for the integrity of the data. All authors have read and agree to the manuscript as written.

## Results

### Total Study Population

The spectrum and number of KCNQ1 mutations by location, type of mutation, and functional effect are presented in Table 1, with the location frequency of the mutations presented diagrammatically in Figure 1. A total of 77 different KCNQ1

TABLE 1. KCNQ1 Mutations by Location and Coding, Type of Mutation, and Functional Effect

Location and Coding*	No. of Subjects†	Type of Mutation	Functional Effect‡
N-terminus			
M1V	1	Missense	Unknown
G57V	1	Missense	Unknown
Transmembrane			
W120C	2	Missense	Unknown
T144A	7	Missense	Unknown
A150fs/133 [del CT 451-452]	2	Frameshift	Haploinsufficiency
E160K	3	Missense	Unknown
G168R	44	Missense	Unknown
Y171X [513 C>G]	6	Nonsense	Haploinsufficiency
R174H	2	Missense	Unknown
A178P	5	Missense	Dominant-negative effect (a)
Y184S	18	Missense	Unknown
G185S	10	Missense	Unknown
G189E	2	Missense	Unknown
G189R	4	Missense	Dominant-negative effect (b)
R190Q	4	Missense	Haploinsufficiency (b, c)
L191fs/90 [del TGGCG 572-576]	8	Frameshift	Haploinsufficiency
R195fs/40 [del G 585]	2	Frameshift	Haploinsufficiency
S225L	13	Missense	Dominant-negative effect (d)
A226V	3	Missense	Unknown
R237P	1	Missense	Unknown
D242N	3	Missense	Unknown
R243C	13	Missense	Haploinsufficiency (e)
V254 mol/L	59	Missense	Dominant-negative effect (b, f)
R258C	1	Missense	Haploinsufficiency
R259C	1	Missense	Haploinsufficiency (g)
L266P	15	Missense	Unknown
G269D	35	Missense	Dominant-negative effect (h)
G269S	25	Missense	Haploinsufficiency (i)
L273F	6	Missense	Dominant-negative effect (a)
I274V	1	Missense	Unknown
S277L	3	Missense	Unknown
Y278H	2	Missense	Unknown
E284K	2	Missense	Unknown
G292D	3	Missense	Unknown
F296S	2	Missense	Unknown
G306R	2	Missense	Dominant-negative effect (b, j)
V310I	1	Missense	Unknown
T312I	14	Missense	Dominant-negative effect (a)
G314S	8	Missense	Dominant-negative effect (h, k, l, m)
Y315C	10	Missense	Dominant-negative effect (d, n)
Y315S	1	Missense	Dominant-negative effect (h, m)
D317G	3	Missense	Unknown
P320H	1	Missense	Unknown
T322 mol/L	2	Missense	Unknown
G325R	3	Missense	Unknown
delF340 [del CTT 1017-1019]	7	In-frame deletion	Haploinsufficiency
A341E	9	Missense	Dominant-negative effect (b)
A341V	20	Missense	Dominant-negative effect (o)

Downloaded from [circ.ahajournals.org](http://circ.ahajournals.org) at National Cardiovascular Center on July 11, 2007

TABLE 1. Continued

Location and Coding*	No. of Subjects†	Type of Mutation	Functional Effect‡
P343S	1	Missense	Dominant-negative effect (p)
A344A/sp [1032 G>A]	27	Splice site	Haploinsufficiency
A344V	17	Missense	Unknown
S349W	15	Missense	Unknown
L353P	4	Missense	Unknown
C-terminus			
Q357H	3	Missense	Unknown
R360G	3	Missense	Unknown
S373P	7	Missense	Unknown
K393N	10	Missense	Unknown
R397W	5	Missense	Unknown
P400fs/62 [Ins C 1201-1022]	6	Frameshift	Haploinsufficiency
P448fs/13 [Ins G 1344-1345]	11	Frameshift	Haploinsufficiency
I517T	3	Missense	Unknown
R518X [1552 C>T]	11	Nonsense	Haploinsufficiency (q)
M520R	3	Missense	Unknown
V524G	4	Missense	Unknown
Q530X [1588 C>T]	13	Nonsense	Haploinsufficiency (q)
R562 mol/L	2	Missense	Unknown
S566F	3	Missense	Unknown
I567S	6	Missense	Unknown
S571fs/20 [del C 1714]	3	Frameshift	Haploinsufficiency
R591C	5	Missense	Unknown
R591H	6	Missense	Haploinsufficiency (r)
R594Q	11	Missense	Haploinsufficiency (q)
D611Y	10	Missense	Haploinsufficiency (s)
A636fs/28 [del C 1909]	2	Frameshift	Haploinsufficiency
Intron			
IVS2+1 G>A	2	Splice site	Unknown
IVS4+5 G>A	2	Splice site	Unknown
IVS7+5 G>A	15	Splice site	Unknown

\*The numbers and letters refer to the amino acid coding of the mutant channel protein. The brackets contain the nucleotide code for deletions, frameshift, splice site, and nonsense mutations.

†Included in this table are 52 subjects who died suddenly at a young age. These subjects were from families with a known KCNQ1 mutation and were assumed to have their respective family mutation.

‡Dominant-negative effect is associated with >50% reduction whereas haploinsufficiency is associated with <50% reduction in ion channel repolarizing current. See text for details. Letters in parentheses refer to references that are available in the online-only Data Supplement.

mutations were identified. A majority of the mutations were localized to the S1 to S6 transmembrane domains (66%), and missense (single amino acid substitutions) accounted for 81% of all the mutations.

The phenotypic characteristics of patients enrolled in each of the 3 registries and by location and type of mutation are presented in Table 2. The clinical characteristics of the patients were similar among the 3 registries except for QTc duration and frequency of  $\beta$ -blocker use. The QTc interval was longer and cardiac events and  $\beta$ -blocker use were more frequent in patients with mutations in the transmembrane than in the C-terminus location.  $\beta$ -Blockers were used less frequently in patients from the Japanese registry than in patients from the other 2 registries. The frequency of first cardiac

events was higher in those with than without missense mutations. The clinical characteristics of the 19 subjects possessing intron mutations resembled those with transmembrane and missense mutations.

The QTc interval was significantly longer in the 12 patients with 2 mutations than in those with only single KCNQ1 mutations ( $570 \pm 70$  versus  $480 \pm 60$  ms;  $P < 0.01$ ). All 12 patients with 2 mutations experienced at least 1 cardiac event.

The cumulative probabilities of first cardiac event by location and type of mutation are presented in Figure 2A and 2B, respectively. Significantly higher event rates were found in subjects with transmembrane than C-terminus mutations and in those with than without missense mutations, with the most rapid increase in event rates occurring during ages 7 to

**# Subjects**

N-terminus: 2  
 Transmembrane: 452  
 C-terminus: 127

**Mutations in the KCNQ1 Channel**

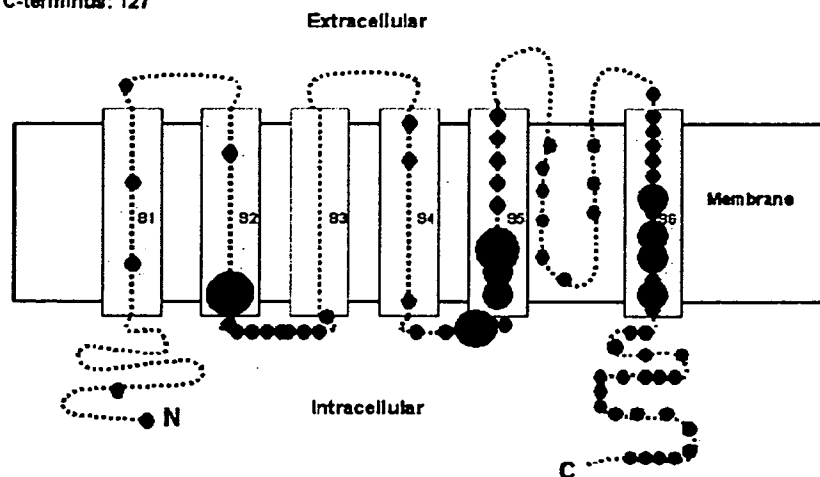


Figure 1. Frequency and location of 74 different mutations in the KCNQ1 potassium channel involving 581 subjects. The 19 subjects with 3 intron mutations are not included in this diagram. The  $\alpha$  subunit involves the N-terminus (N), 6 membrane-spanning segments, and the C-terminus portion (C). The size of the circles reflect the number of subjects with mutations at the respective locations, with the small circles indicating <15, medium-sized circles 15 to 30, and large circles >30 subjects.

20 years. In patients with transmembrane-localized mutations, the event rates for patients with mutations localized to the pore region (S5-pore-S6) were nearly identical to those with nonpore mutations (data not shown).

The findings from the Cox regression analysis for location and type of mutation are presented in Table 3. The clinical risk factors associated with first cardiac events involved males before age 13 years, females after age 13

**TABLE 2. Phenotypic Characteristics by Source of Subjects, Location of Mutation, and Type of Mutation**

Characteristics	Source of Subjects			Location of Mutation		Missense Mutation		Intron Mutation (n=19)
	United States (n=425)	Netherlands (n=93)	Japan (n=82)	Trans Membrane (n=452)	C-Terminus (n=127)	Yes (n=483)	No (n=98)	
Female, %	57	53	54	57	51	54	62	63
ECG at enrollment								
QTc††, ms	488±58	450±45	472±46	485±53	460±61	481±59	471±38	478±60
Therapy, %								
β-Blockers††	45	34	26	45	28	42	38	37
Pacemaker	2.4	0	0	1.5	2.4	1.4	3.1	0
Sympathectomy	0.5	0	0	0.4	0	0.4	0	0
Defibrillator	6.4	3.2	0	5.8	3.1	5.2	5.1	0
First cardiac event* †§, %								
Syncope‡ (n=200)	35	31	29	38	17	36	21	32
Aborted cardiac arrest (n=15)	1.9	1.1	7.3	2.9	0.8	2.5	2.0	5.3
Death (n=23)	4.0	5.5	1.2	4.0	3.1	4.2	2.0	5.3
Ever cardiac event, %								
Syncope‡§	35	31	31	39	17	37	21	33
Aborted cardiac arrest†	2.4	15	8.8	5.3	3.2	5.4	2.0	11
Death	11	14	2.4	10	6.3	11	4.1	26

Plus-minus values are mean±SD. Percentages >10 are rounded to a whole number. The 600 subjects in this table include 51 subjects who died suddenly at a young age, were from families with known KCNQ1 mutation, and were assumed to have the family mutation. Patients with intron mutations are categorized separately and are not included in the location or missense categories. Seven subjects with transmembrane mutations and 1 with C-terminus mutations had missing data about the date of the first cardiac event. Eight subjects with missense mutations had missing data about the date of the first cardiac event. Numbers in parentheses indicate the total number of specific first cardiac events from the 3 sources of patients.

\*First cardiac event was syncope, aborted cardiac arrest, or sudden death, whichever occurred first.

†P<0.01 for the comparison of characteristics among the 3 sources of subjects.

‡P<0.01 for the comparison of characteristics between the 2 locations of the mutations.

§P<0.01 for the comparison of characteristics between missense yes and no.



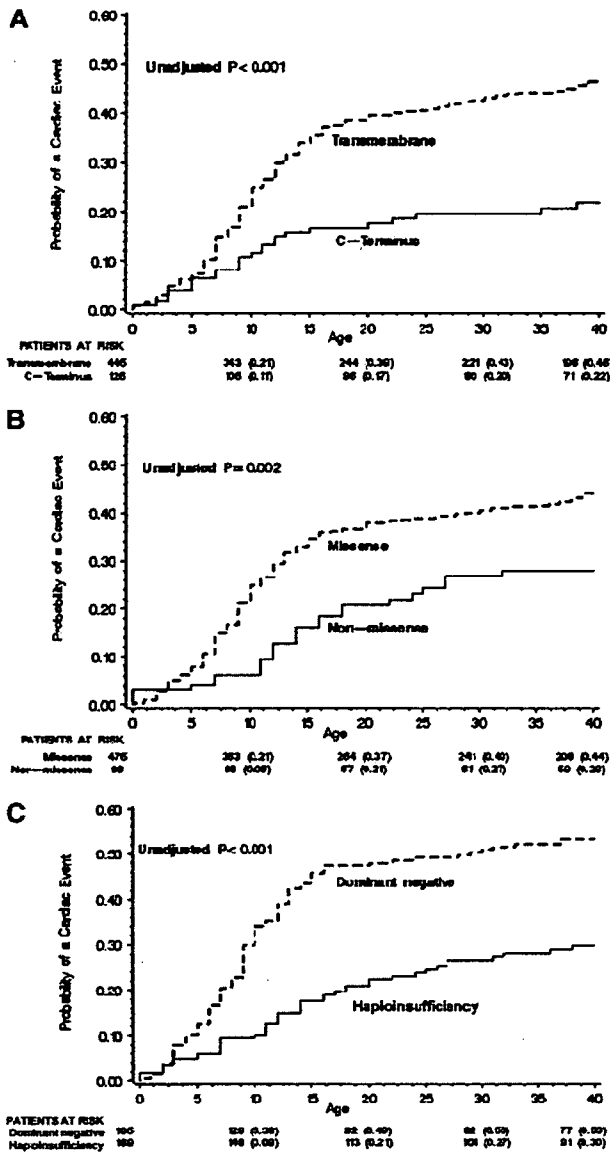


Figure 2. Kaplan-Meier estimate of the cumulative probability of a first cardiac event by location (A), type (B), and biophysical function of the mutation (C).

years, and longer QTc intervals. Mutations located in the transmembrane region of the channel made significant and independent contributions to the risk model, but missense mutations were not an independent risk factor. Three different intron mutations were present in 19 subjects from 4 families, and these intron mutations made a meaningful but nonsignificant contribution to the risk model. Prespecified interactions were investigated for their effect on cardiac events, and no significant interactions were found for transmembrane location by type of mutation, transmembrane location by QTc, or mutation type by QTc. Time-dependent  $\beta$ -blocker use was associated with a significant 74% reduction in the risk of first cardiac events ( $P < 0.001$ ).

TABLE 3. Cox Regression With Multiple Predictor Variables Including Location and Type of Mutations for First Cardiac Event

Variable	Hazard Ratio	95% CI	P
Netherlands:United States	1.15	0.74–1.78	0.55
Japan:United States	1.45	0.98–2.16	0.07
Male <13 y:female <13 y	1.72	1.25–2.38	<0.001
Female 13–40 y:male 13–40 y	2.27	1.30–3.96	<0.01
QTc 500–530 ms:QTc <500 ms	2.04	1.41–2.96	<0.001
QTc >530 ms:QTc <500 ms	3.25	2.25–4.69	<0.001
QTc missing*:QTc <500 ms	2.26	1.57–3.25	<0.001
Transmembrane:C-terminus	2.06	1.36–3.12	<0.001
Missense yes:no	1.33	0.86–2.05	0.20
Intron:C-terminus	2.45	0.98–6.11	0.06
Time-dependent $\beta$ -blocker use	0.26	0.14–0.49	<0.001

The Cox analysis involved 592 subjects with 445 transmembrane, 126 C-terminus, 2 N-terminus, and 19 intron mutations; 8 subjects were not included in this Cox analysis because of missing data about the date of their first cardiac event.

\*QTc missing category involves 47 subjects who died suddenly at a young age without a prior ECG.

**Biophysical Function and Outcome**

The clinical implications of disordered biophysical function of the mutant KCNQ1 channels were investigated in a subset of 356 subjects with known or suspected alteration in ion channel function (see Methods for functional categorization). The clinical characteristics of patients with dominant-negative and haploinsufficiency ion channel dysfunction are presented in Table 4. Patients with mutations having dominant-negative ion current effects had a longer QTc interval and a higher frequency of cardiac events than subjects with mutations resulting in haploinsufficiency. The cumulative probabilities of a first cardiac event by the biophysical function of the mutations are presented in Figure 2C. As shown in Table 5, patients with mutations having

TABLE 4. Phenotypic Characteristics by Biophysical Function of the KCNQ1 Mutations in 356 Subjects

Characteristics	Dominant-Negative Effect (n=187)	Haploinsufficiency (n=169)
Female, %	51	61
ECG at enrollment		
QTc,* ms	500±60	470±50
Therapy, %		
$\beta$ -Blockers	47	37
Pacemaker	1.1	4.1
Sympathectomy	0.5	0
Defibrillator	4.8	7.7
First cardiac event*, %	53	27
Syncope	45	22
Aborted cardiac arrest	2.1	3.0
Death	5.3	2.4

Percentages >10 are rounded to a whole number. Two subjects had missing data about the date of their first cardiac event.

\* $P < 0.01$ .

TABLE 5. Cox Regression With Multiple Predictor Variables Including Ion Channel Dysfunction for First Cardiac Events

Variable	Hazard Ratio	95% CI	P
Netherlands:United States	2.78	1.48-5.23	<0.01
Japan:United States	1.63	1.02-2.63	0.04
Male <13 y:female <13 y	1.94	1.29-2.91	<0.01
Female 13-40 y:male 13-40 y	1.95	0.99-3.87	0.06
QTc 500-530 ms:QTc <500 ms	1.88	1.18-2.99	<0.01
QTc >530 ms:QTc <500 ms	3.22	2.06-5.05	<0.001
QTc missing*:QTc <500 ms	2.07	1.29-3.33	<0.01
Dominant-negative:haploinsufficiency	2.26	1.56-3.25	<0.001
Time-dependent $\beta$ -blocker use	0.21	0.09-0.48	<0.001

The analysis involved 354 subjects with known or suspected ion channel dysfunction; 2 subjects were not included because of missing data about the date of their first cardiac event.

\*The QTc missing category involves 26 patients who died suddenly at a young age without a prior ECG.

dominant-negative functional effects experienced a significantly greater risk for cardiac events than those with haploinsufficiency (hazard ratio, 2.26; 95% CI, 1.56 to 3.25;  $P<0.001$ ) after adjustment for relevant covariates including QTc and gender effects by age group.  $\beta$ -Blocker use was associated with a significant 79% reduction in first cardiac events in this subset of patients. Because substantial collinearity exists for transmembrane mutations, missense mutations, and mutations with dominant-negative biophysical function, the individual effects of these 3 mutation parameters could not be ascertained reliably in the same Cox model.

## Discussion

The main results of the present study from 600 patients having a spectrum of KCNQ1 mutations derived from 3 LQTS registries are significantly higher cardiac event rates in patients with transmembrane mutations and in patients with mutations having a putative dominant-negative effect on the repolarizing  $I_{Ks}$  current. The effect of these genetically determined factors is independent of traditional clinical risk factors and of  $\beta$ -blocker therapy.

Since 1995, when the first 2 genes responsible for LQTS were identified,<sup>11,12</sup> molecular genetic studies have revealed a total of 9 forms of congenital LQTS caused by mutations in genes involving potassium channel (LQT-1, -2, -5, -6, and -7), sodium channel (LQT-3, -9), and calcium channel proteins (LQT-8) as well as a membrane-adaptor protein (LQT-4).<sup>2,13</sup> Genotype-phenotype studies have enabled us to stratify risk and to treat more specifically patients with LQT-1, LQT-2, and LQT-3 subtypes of this genetic disorder. LQT-1, the most common form of LQTS, accounts for  $\approx 50\%$  of genotyped patients<sup>4,14</sup> and has more variable expressivity and incomplete penetrance than the other forms.<sup>15</sup> Mutation location and knowledge of the functional effects of the mutation provide additional risk information beyond the clinical risk factors and the genotype, at least for LQT-1, and this information should contribute to improved risk stratification and more focused management of these higher-risk patients.

Mutations in KCNQ1 are responsible for defects in the slowly activating component of the delayed rectifier current  $I_{Ks}$ .<sup>16</sup> This current is the main repolarizing current at increased heart rate and is highly sensitive to catecholamines.<sup>3</sup> We speculate that  $I_{Ks}$  channels with transmembrane mutations might have reduced responsiveness to the regulatory  $\beta$ -adrenergic signaling of the ion-conduction pathway with more impairment of shortening of the QTc with exercise-related tachycardia than mutations in the C-terminus region.

Functional  $I_{Ks}$  channels result from the coassembly of 4 KCNQ1-encoded subunits. A mutated gene encodes a protein with aberrant function, and the presence of both normal and abnormal proteins in the ion channel contributes to a  $>50\%$  reduction in ion channel function (dominant-negative effect). An alternative mechanism of reduced repolarizing KCNQ1  $K^+$  current is the inability of mutated subunits to coassemble with normal gene products, such as occurs with a trafficking defect, resulting in a  $\leq 50\%$  reduction in channel function (haploinsufficiency). With only 1 exception,<sup>17</sup> this is the case for all studied truncating mutations leading to incomplete proteins. Our assumption that truncated proteins (based on frameshift nonsense mutations) lead to haploinsufficiency seems justified. The biophysical effect of missense mutations is unpredictable, and both haploinsufficiency and dominant-negative effects have been described. In the absence of reported biophysical studies, missense mutations were classified as unknown.

Previous attempts to identify a genotype-phenotype relationship for KCNQ1 mutations failed to reach consensus on the clinical outcome of the type and site of mutations.<sup>7,8</sup> Relatively small numbers and different ethnic background of the previously reported patients with the LQT-1 genotype might be responsible for the discrepant results. The present larger study allows us to demonstrate for the first time that the biophysical effect clearly affects the clinical outcome (ie, dominant-negative mutations are associated with a more severe phenotype than are mutations conferring haploinsufficiency [Figure 2C], even after adjustment for relevant covariates [Table 5]). The risk observed in 19 subjects with 3 different intron mutations was not quite significant ( $P=0.06$ ), possibly because of small numbers, but the magnitude of the risk effect was similar to the risk accompanying transmembrane mutations. Although these intron mutations produced splice-site alterations predicted to affect the transmembrane portion of the ion channel, we used a separate categorization of intron mutations in view of the limited understanding of the structural alterations and functional effects resulting from these exon-skipping intron mutations.

A few additional findings from this large genotype-phenotype study of type-1 LQTS patients emphasize high risk for first cardiac events during adolescence, a crossover in risk by sex at approximately age 13 years, and a lower rate of first cardiac events in the adult years than in the younger years. These findings are not especially new,<sup>18,19</sup> but the present study highlights their presence in type-1 LQTS.

## Study Limitations

The present study used the biophysical function of mutations reported in the literature in only a portion of the mutations

that were included (see references associated with Table 1 in the online-only Data Supplement). The published studies were from many different laboratories with the use of different cellular heterologous expression systems involving *Xenopus* oocytes and other cells at both room and physiological temperatures. Although such nonuniform testing may have contributed to some inconsistency in the categorized biophysical function, the finding of a significantly higher event rate in mutations with dominant-negative than with haploinsufficient effects (hazard ratio, 2.26;  $P < 0.001$ ) is unlikely to have resulted from the nonuniform testing. Unfortunately, we did not have the resources to perform such uniform testing in all 77 mutations presented in the present study.

Once a mutation was identified in KCNQ1, thorough genetic sequencing was not performed routinely in all the ion channel genes to look for second mutations. Thus, some of the patients included in the analysis may have had a second mutation in addition to the identified KCNQ1 mutation. It is estimated that  $\approx 10\%$  of genotype LQTS patients may carry a second mutation, and those with  $> 1$  mutation could contribute to some of the findings in our study. In addition, it is possible that some of the reported mutations (Table 1) are simply uncommon sequence mutations, but this is relatively unlikely because all the subjects in the present study were derived from families in which the proband had QTc prolongation not due to a known cause.

The outcome analyses included subjects from families with a known KCNQ1 mutation who died suddenly and unexpectedly at a young age and were classified as LQTS-related death with the same mutation that was present in the family. It is possible that a few of these subjects could have died from a non-LQTS cause or had an LQTS mutation different from the family mutation, but we think the error rate is likely to be small. The number of deaths and aborted cardiac arrest events is small, and there is insufficient power to evaluate the risk association of the genotype characteristics with these endpoint events in a multivariate time-dependent model.

### Conclusions

The present study confirms that in patients with type-1 LQTS, longer QTc intervals are associated with higher cardiac event rates and that male patients are generally younger than female patients at first cardiac events.<sup>20,21</sup> The new findings from the present study are that transmembrane mutations and mutations with dominant-negative functional effect adversely influence the outcome of this disorder independent of traditional clinical risk factors and  $\beta$ -blocker therapy. The present study was not designed to assess the effectiveness of different therapies in patients with KCNQ1 mutations. The findings presented do not provide justification for using specific genotype characteristics to identify patients for implanted defibrillator therapy.

### Note Added in Proof

After this article was accepted for publication, we noted the recent article by Tsuji et al, in which the A344A/sp [1032G>A] mutation that we categorized as haploinsufficient (Table 1) was reported to have a weak dominant-

negative effect.<sup>22</sup> We reran the KCNQ1 data recategorizing the 27 A344A/sp [1032 G>A] mutations as dominant-negative. Negligible changes occurred in the results as presented in Table 5 and Figure 2C; the hazard ratio for dominant-negative:haploinsufficiency (Table 5) was unchanged at 2.26 ( $P < 0.001$ ).

### Acknowledgment

We thank David J. Tester, Senior Research Technologist, Sudden Death Genomics Laboratory, Mayo Clinic College of Medicine, Rochester, Minn, for the detailed review and assistance he provided on the terminology and nomenclature for the annotated mutations presented in Table 1.

### Sources of Funding

This study was supported in part by (1) research grants HL-33843 and HL-51618 from the National Institutes of Health, Bethesda, Md (Dr Moss); (2) Ministry of Education, Culture, Sports, Science, and Technology Leading Project for Biosimulation and health sciences research grant (H18-Research on Human Genome-002) from the Ministry of Health, Labor, and Welfare, Japan (Dr Shimizu); (3) grant 2000.059 from the Nederlandse Hartstichting, Amsterdam, the Netherlands (Dr Wilde); and (4) research grants from the National Institutes of Health (HD42569), American Heart Association (Established Investigator Award), CJ Foundation for SIDS, and Dr Scholl Foundation (Dr Ackerman).

### Disclosures

Dr Ackerman is a consultant for Clinical Data (formerly Genaisance Pharmaceuticals) with respect to the FAMILION genetic test for cardiac ion channel mutations. The other authors report no conflicts.

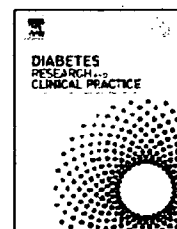
### References

- Moss AJ. Long QT syndrome. *JAMA*. 2003;289:2041–2044.
- Wilde AA, Bezzina CR. Genetics of cardiac arrhythmias. *Heart*. 2005; 91:1352–1358.
- Sanguinetti MC. Long QT syndrome: ionic basis and arrhythmia mechanism in long QT syndrome type 1. *J Cardiovasc Electrophysiol*. 2000;11:710–712.
- Tester DJ, Will ML, Haglund CM, Ackerman MJ. Compendium of cardiac channel mutations in 541 consecutive unrelated patients referred for long QT syndrome genetic testing. *Heart Rhythm*. 2005;2:507–517.
- Bianchi L, Priori SG, Napolitano C, Surewicz KA, Dennis AT, Memmi M, Schwartz PJ, Brown AM. Mechanisms of I(Ks) suppression in LQT1 mutants. *Am J Physiol*. 2000;279:H3003–H3011.
- Shalaby FY, Levesque PC, Yang WP, Little WA, Conder ML, Jenkins-West T, Blana MA. Dominant-negative KvLQT1 mutations underlie the LQT1 form of long QT syndrome. *Circulation*. 1997;96: 1733–1736.
- Zareba W, Moss AJ, Sheu G, Kaufman ES, Priori S, Vincent GM, Towbin JA, Benhorin J, Schwartz PJ, Napolitano C, Hall WJ, Keating MT, Qi M, Robinson JL, Andrews ML. Location of mutation in the KCNQ1 and phenotypic presentation of long QT syndrome. *J Cardiovasc Electrophysiol*. 2003;14:1149–1153.
- Shimizu W, Horie M, Ohno S, Takenaka K, Yamaguchi M, Shimizu M, Washizuka T, Aizawa Y, Nakamura K, Ohe T, Aiba T, Miyamoto Y, Yoshimasa Y, Towbin JA, Priori SG, Kamakura S. Mutation site-specific differences in arrhythmic risk and sensitivity to sympathetic stimulation in the LQT1 form of congenital long QT syndrome: multicenter study in Japan. *J Am Coll Cardiol*. 2004;44:117–125.
- Cox DR. Regression models and life-tables. *J Stat Soc [B]*. 1972;34: 187–220.
- Therneau TM, Grambsch PM. *Modeling Survival Data: Extending the Cox Model*. New York, NY: Springer-Verlag; 2000.
- Sanguinetti MC, Jiang C, Curran ME, Keating MT. A mechanistic link between an inherited and an acquired cardiac arrhythmia: HERG encodes the IKr potassium channel. *Cell*. 1995;81:299–307.
- Wang Q, Shen J, Li Z, Timothy K, Vincent GM, Priori SG, Schwartz PJ, Keating MT. Cardiac sodium channel mutations in patients with long QT

- syndrome, an inherited cardiac arrhythmia. *Hum Mol Genet.* 1995;4:1603–1607.
13. Vatta M, Ackerman MJ, Ye B, Makielski JC, Ughanze EE, Taylor EW, Tester DJ, Balijepalli RC, Foell JD, Li Z, Kamp TJ, Towbin JA. Mutant caveolin-3 induces persistent late sodium current and is associated with long-QT syndrome. *Circulation.* 2006;114:2104–2112.
  14. Splawski I, Shen J, Timothy KW, Lehmann MH, Priori S, Robinson JL, Moss AJ, Schwartz PJ, Towbin JA, Vincent GM, Keating MT. Spectrum of mutations in long-QT syndrome genes: KVLQT1, HERG, SCN5A, KCNE1, and KCNE2. *Circulation.* 2000;102:1178–1185.
  15. Shimizu W, Noda T, Takaki H, Nagaya N, Satomi K, Kurita T, Suyama K, Aihara N, Sunagawa K, Echigo S, Miyamoto Y, Yoshimasa Y, Nakamura K, Ohe T, Towbin JA, Priori SG, Kamakura S. Diagnostic value of epinephrine test for genotyping LQT1, LQT2, and LQT3 forms of congenital long QT syndrome. *Heart Rhythm.* 2004;1:276–283.
  16. Sanguinetti MC, Curran ME, Zou A, Shen J, Spector PS, Atkinson DL, Keating MT. Coassembly of K(V)LQT1 and minK (IsK) proteins to form cardiac I(Ks) potassium channel. *Nature.* 1996;384:80–83.
  17. Aizawa Y, Ueda K, Wu LM, Inagaki N, Hayashi T, Takahashi M, Ohta M, Kawano S, Hirano Y, Yasunami M, Kimura A, Hiraoka M. Truncated KCNQ1 mutant, A178fs/105, forms hetero-multimer channel with wild-type causing a dominant-negative suppression due to trafficking defect. *FEBS Lett.* 2004;574:145–150.
  18. Hobbs JB, Peterson DR, Moss AJ, McNitt S, Zareba W, Goldenberg I, Qi M, Robinson JL, Sauer AJ, Ackerman MJ, Benhorin J, Kaufman ES, Locati EH, Napolitano C, Priori SG, Towbin JA, Vincent GM, Zhang L. Risk of aborted cardiac arrest or sudden cardiac death during adolescence in the long-QT syndrome. *JAMA.* 2006;296:1249–1254.
  19. Moss AJ, Schwartz PJ, Crampton RS, Tzivoni D, Locati EH, MacCher J, Hall WJ, Weikamp L, Vincent GM, Garson A Jr. The long QT syndrome: prospective longitudinal study of 328 families. *Circulation.* 1991;84:1136–1144.
  20. Locati EH, Zareba W, Moss AJ, Schwartz PJ, Vincent GM, Lehmann MH, Towbin JA, Priori SG, Napolitano C, Robinson JL, Andrews M, Timothy K, Hall WJ. Age- and sex-related differences in clinical manifestations in patients with congenital long-QT syndrome: findings from the International LQTS Registry. *Circulation.* 1998;97:2237–2244.
  21. Zareba W, Moss AJ, Locati EH, Lehmann MH, Peterson DR, Hall WJ, Schwartz PJ, Vincent GM, Priori SG, Benhorin J, Towbin JA, Robinson JL, Andrews ML, Napolitano C, Timothy K, Zhang L, Medina A. Modulating effects of age and gender on the clinical course of long QT syndrome by genotype. *J Am Coll Cardiol.* 2003;42:103–109.
  22. Tsuji K, Akao M, Ishii TM, Ohno S, Makiyama T, Takenaka K, Doi T, Haruna Y, Yoshida H, Nakashima T, Kita T, Horie M. Mechanistic basis for the pathogenesis of long QT syndrome associated with a common splicing mutation in KCNQ1 gene. *J Mol Cell Cardiol.* 2007;42:662–669.

### CLINICAL PERSPECTIVE

Type-1 long-QT syndrome is caused by loss-of-function mutations in the KCNQ1-encoded  $I_{Ks}$  cardiac potassium channel. In the present study involving 600 patients having a spectrum of KCNQ1 mutations derived from 3 long-QT syndrome registries, we found that cardiac event rates are increased significantly in patients with mutations located in the transmembrane region of the potassium channel and in patients with mutations having a putative dominant-negative effect on the repolarizing  $I_{Ks}$  current. The effects of these genetically determined factors are independent of traditional clinical risk factors and of  $\beta$ -blocker therapy. Mutation location and knowledge of functional effects of the mutation provide additional risk information beyond the clinical risk factors and the genotype, at least for type-1 long-QT syndrome, and this information should contribute to improved risk stratification and more focused management of these higher-risk patients.

available at [www.sciencedirect.com](http://www.sciencedirect.com)Journal homepage: [www.elsevier.com/locate/diabres](http://www.elsevier.com/locate/diabres)

## Impaired flow-mediated vasodilatation and insulin resistance in type 2 diabetic patients with albuminuria

Hisashi Makino\*, Kentaro Doi, Aki Hiuge, Ayako Nagumo, Sadanori Okada, Yoshihiro Miyamoto, Masaaki Suzuki, Yasunao Yoshimasa

Department of Atherosclerosis and Diabetes, National Cardiovascular Center, 5-7-1 Fujishiro-dai, Suita City, Osaka 565-8565, Japan

### ARTICLE INFO

#### Article history:

Received 21 June 2007

Accepted 22 August 2007

Published on line 27 September 2007

#### Keywords:

Nitric oxide

Diabetic nephropathy

Endothelial dysfunction

Atherosclerosis

### ABSTRACT

An elevated urinary albumin excretion is associated with an increased risk of cardiovascular disease due to atherosclerosis, but the pathophysiological mechanism underlying this association is poorly understood. We studied 217 diabetic patients, that is, 121 normoalbuminuric patients, 71 microalbuminuric patients, and 25 macroalbuminuric patients. We evaluated flow-mediated dilatation of brachial artery (%FMD, one endothelial function marker associated with endogenous NO production), von Willebrand factor (vWF, endothelial activation marker), high-sensitive CRP (hsCRP, a low-grade inflammation marker), asymmetric dimethyl arginine (ADMA, an endogenous inhibitor of NO synthesis), and insulin sensitivity by steady-state plasma glucose method. %FMD was apparently decreased in microalbuminuric and macroalbuminuric patients compared with normoalbuminuric patients ( $p < 0.001$ ). Moreover, %FMD was significantly correlated with the degree of albuminuria ( $r = -0.38$ ,  $p < 0.05$ ). On the other hand, vWF and hsCRP did not show significant difference between normoalbuminuric patients and microalbuminuric patients. In diabetic patients with macroalbuminuria, ADMA was significantly elevated compared to those with normoalbuminuria. Insulin sensitivity was significantly associated with urinary albumin excretion rate. These results suggested that endothelial dysfunction which may be due to impaired NO production and insulin resistance underlie the association between diabetic nephropathy and atherosclerosis in diabetic patients.

© 2007 Elsevier Ireland Ltd. All rights reserved.

### 1. Introduction

Elevated urinary albumin excretion rate (UAER) is strongly associated with an increased risk of cardiovascular diseases, which is independent of conventional risk factors including hypertension, hyperlipidemia, and smoking, among individuals with and without type 2 diabetes [1,2]. This suggests that elevated UAER may be associated with atherosclerosis by the unidentified mechanism.

The endothelium plays a crucial role in the maintenance of vascular tone and structure, and endothelial dysfunction is a

key feature of atherosclerosis. Nitric oxide (NO) is one of the important endothelium-derived vasoactive mediators. NO is involved in a wide variety of regulatory mechanisms of cardiovascular system, including vascular tone and vascular structure [3].

Flow-mediated endothelium-dependent vasodilatation (FMD) method is based on the endothelial stimulus of increased shear stress (the tangential force on the vessel wall exerted by flowing blood). Increased shear stress is caused by post-ischemic hyperemia and elicits a slow  $Ca^{2+}$ -independent two to threefold increase in NO production [4,5]. Indeed,

\* Corresponding author. Tel.: +81 6 6833 5012; fax: +81 6 6833 9865.

E-mail address: [makinoh@hsp.ncvc.go.jp](mailto:makinoh@hsp.ncvc.go.jp) (H. Makino).

0168-8227/\$ – see front matter © 2007 Elsevier Ireland Ltd. All rights reserved.  
doi:10.1016/j.diabres.2007.08.014

Celemajer et al. reported that flow mediated vasodilatation was mainly blocked by *N*-monomethyl-*L*-arginine (an inhibitor of endothelial NO synthetase) [6].

To clarify the contribution of impaired NO production in vascular endothelium to the association between atherosclerotic disease and diabetic nephropathy, we examined FMD by ultrasonography. In addition, we measured asymmetric dimethyl arginine (ADMA), an endogenous NO synthesis inhibitor [3]. Since low-grade inflammation is another key feature of the pathophysiology of atherosclerosis [7], we further examined high-sensitive CRP, which is an inflammation marker, to investigate whether this feature is involved in the association between atherosclerotic disease and diabetic nephropathy.

It has recently been indicated that microalbuminuria and atherosclerosis are closely associated with insulin resistance [8–10], implying that insulin resistance may underlie these pathophysiological conditions although the causative relationship remains unknown. In the present study, we further examined insulin sensitivity in the type 2 diabetic patients with different stage of albuminuria and analyzed the correlation between insulin sensitivity and FMD, to investigate whether elevated UAER and endothelial dysfunction may be associated with insulin resistance.

## 2. Methods

### 2.1. Study subjects

We studied 217 patients with type 2 diabetes who were <75 years of age. Patients with a current acute illness (including clinically significant infectious disease) were excluded from this study. Twenty-four-hour urine collections were performed for two consecutive days to determine the stage of diabetic nephropathy. Creatinine clearance (Ccr) was calculated from the 24-h urine sample and serum creatinine levels. The patients were divided into three groups according to the UAER, as follows: normoalbuminuria (UAER <30 mg/day), microalbuminuria (30 ≤ UAER < 100 mg/day) and macroalbuminuria (UAER ≥ 300 mg/day). To exclude diabetic patients with nondiabetic kidney disease, we excluded patients with hematuria or abnormal urinary sediments. This study was conducted with the approval of National Cardiovascular Center Trust Ethics Committee, and patients gave written informed consent before participation.

### 2.2. Brachial artery flow-mediated dilatation

Using ultrasonography, arterial endothelium and smooth muscle function were measured by examining brachial artery responses to endothelium-dependent and endothelium-independent stimuli. Ultrasoundonographic measurements were carried out according to the method described by Celemajer et al. [6]. Brachial artery diameter was measured from B-mode ultrasound images using 10-MHz liner array transducer (ProSound SSD-5500; Aloka, Japan) while an ECG trace was simultaneously recorded. The right brachial artery was scanned in longitudinal sections 1–10 cm above elbow, after at least 15 min of rest in the supine position, the skin surface

was marked and the arm was kept in the same position during the study.

Baseline measurements of the diameter were carried out. Endothelium-dependent vasodilatation (flow-mediated dilatation) was determined by scans during reactive hyperemia. A pneumatic cuff placed around the forearm was inflated to 220 mmHg and was deflated after 4.5 min. The diameter of the brachial artery was scanned and recorded after dilation. After 10 min rest, the second control scan of the diameter was recorded. Then, sublingual glyceryl trinitrate spray (300 µg) was administered and 3.5 min later a final scan of the diameter was recorded.

Measurements of the vessel diameter were taken from the anterior to posterior "m" line (interface between the media and adventitia) at end-diastole, coincident with the R wave on a continuously recorded ECG. The diameters at four cardiac cycles were measured for each scan, and these results were averaged. Determinations of the FMD were carried out 45–60 s after the cuff release to measure a maximal diameter. Vasodilatation by reactive hyperemia or glyceryl trinitrate (NTG) was expressed as the percent change in diameter compared with the baseline values.

### 2.3. Insulin sensitivity test

Glucose utilization in response to insulin was evaluated with a newly modified steady-state plasma glucose (SSPG) method with octreotide acetate (Sandostatine; Novartis) after an overnight fasting period of 12 h [11]. Sandostatine (9.8-pmol bolus followed by a constant infusion of 73.5 pmol/h) and Humulin R insulin (45 pmol/kg bolus followed by a constant infusion at a rate of 4.62 pmol/(kg min); Eli Lilly) were infused intravenously for 120 min. Glucose in a final 12% solution containing KCl (0.5 µmol/(kg min)) was infused at a rate of 0.033 mmol/(kg min) (6 mg/(kg min)) through an antecubital vein via a constant infusion pump. Blood samples were drawn routinely at 0 and 120 min (9:00 and 11:00 a.m.) for the determination of glucose, insulin, and lipids. The value of glucose at 120 min (SSPG) was used as a marker of insulin sensitivity to glucose utilization. High SSPG levels showed peripheral insulin resistance.

Another marker of insulin resistance (IR) was estimated by calculating homeostasis model assessment (HOMA-IR) index ((fasting serum insulin (µU/ml) × fasting plasma glucose (mmol/l))/22.5) [12].

### 2.4. Measurement of vWF, hsCRP, and ADMA

vWF was determined in citrated plasma using a homemade enzyme-linked immunosorbent assay. Data are given as the percentage of pooled human plasma (set at 100%). Serum hsCRP concentration was determined by latex nephelometry method (SRL, Tokyo, Japan). Serum ADMA concentration was determined by high-performance liquid chromatography method (SRL, Tokyo, Japan).

### 2.5. Statistical analysis

Values are expressed as means ± S.D. Statistical analysis was performed by use of ANOVA followed by Scheffes' test. The

Table 1 – Characteristics of diabetic patients with normoalbuminuria, microalbuminuria, and overt nephropathy

Parameter	Stage of nephropathy		
	Normoalbuminuria	Microalbuminuria	Macroalbuminuria
n	121	71	25
Age (years)	62 ± 9	65 ± 8	66 ± 7
Men/women	76/45	34/37	12/13
Duration of diabetes (years)	12 ± 8	14 ± 8	18 ± 8*
BMI (kg/m <sup>2</sup> )	25.0 ± 3.7	25.1 ± 3.7	25.1 ± 3.9
SBP (mmHg)	128 ± 13	133 ± 15	141 ± 19*
DBP (mmHg)	74 ± 10	73 ± 9	76 ± 10
FBS (mmol/l)	7.4 ± 1.4	7.5 ± 1.5	7.5 ± 1.9
HbA1c (%)	8.3 ± 1.5	8.9 ± 1.7*	8.8 ± 1.4
HOMA-IR	1.62 ± 0.98	1.71 ± 2.06	2.29 ± 1.47
Total cholesterol (mmol/l)	4.86 ± 0.90	4.86 ± 0.90	4.73 ± 0.75
Serum creatinine (μmol/l)	70 ± 20	60 ± 20	110 ± 40
Urinary albumin (mg/day)	10 ± 7	85 ± 79**	583 ± 576**
Creatinine clearance (ml/s)	1.43 ± 0.52	1.50 ± 0.63	0.73 ± 0.43**
ACEI or ARB (yes/no)	36/85	24/47	11/14*
Statin (yes/no)	45/76	25/46	10/15
Current smoker (yes/no)	11/110	7/64	6/19

\* $p < 0.05$ , \*\* $p < 0.01$  vs. normoalbuminuria, mean ± S.D.

strength of correlation between variables was tested by linear correlation and multiple regression analysis.  $p < 0.05$  was considered to be statistically significant.

### 3. Results

#### 3.1. Patients characteristics

Table 1 shows the clinical characteristics of three groups. There was no significant difference in age, gender, BMI, FBS and total cholesterol among the three groups. HbA1c of diabetic patients with microalbuminuric patients was significantly higher than normoalbuminuric patients. Systolic blood pressure of macroalbuminuric patients was significantly higher than normo- and micro-albuminuric patients. Creatinine clearance was significantly decreased in macroalbuminuric patients compared with normo- and micro-albuminuric patients. There is no significant difference in rate of patients taking ACE/ARB between normo- and micro-albuminuric patients whereas the rate of patients taking ACE/ARB of macroalbuminuric patients were significantly large compared with other two groups. On the other hand, there is no significant difference in rate of patients taking statin among three groups.

#### 3.2. %FMD of diabetic patients

We studied the endothelial function by FMD using brachial artery echography. %FMD ( $\Delta$ hyperemia) of diabetic patients with microalbuminuria ( $4.5 \pm 3.7\%$ ) and macroalbuminuria ( $4.2 \pm 2.4\%$ ) was apparently decreased compared with those of diabetic patients with normoalbuminuria ( $6.6 \pm 3.7\%$ ) (Fig. 1A). Moreover, %FMD was significantly correlated with UAER in normo- and micro-albuminuric patients independent of age, HbA1c, and systolic blood pressure by multiple regression analysis ( $r = -0.38$ ,  $p < 0.05$ ) (Fig. 2). Dilatation of brachial artery by NTG ( $\Delta$ NTG) showed no difference among three groups (Fig. 1B).

#### 3.3. vWF, hsCRP, and ADMA of diabetic patients

We studied other atherosclerotic markers, that is, vWF, hsCRP, and ADMA. There was no significant difference of the levels of vWF and hsCRP between normoalbuminuric and microalbu-

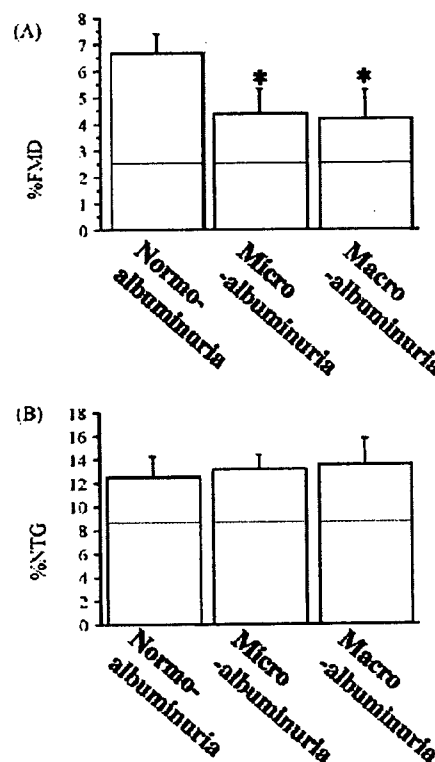
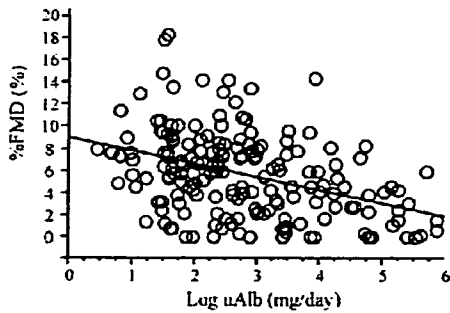


Fig. 1 – %FMD (A) and %NTG (B) in diabetic patients with normoalbuminuria, microalbuminuria and macroalbuminuria. Each value means (means ± S.D.), \* $p < 0.001$ .



**Fig. 2 – Correlation between degree of UAE and %FMD in normo- and micro-albuminuric diabetic patients. There was a significant correlation between both variables ( $r = -0.38, p < 0.05, n = 192$ ).**

minuric patients (Table 2). Although the levels of ADMA in microalbuminuric patients did not show significant difference compared with normoalbuminuric patients (Table 2), the levels of ADMA in macroalbuminuric patients were significantly elevated compared with normoalbuminuric patients (Table 2).

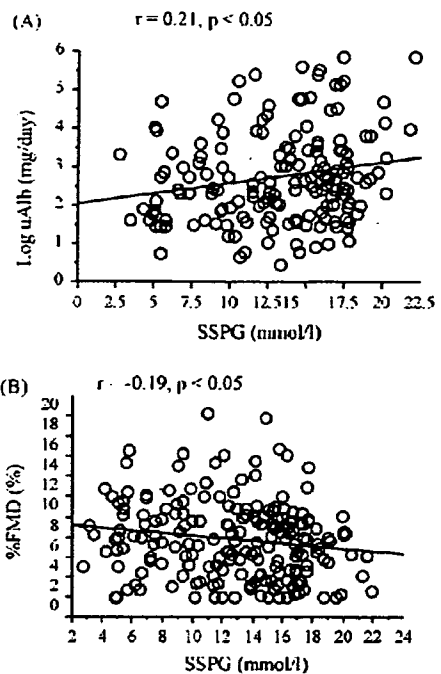
**3.4. Insulin sensitivity of diabetic patients**

We studied the insulin sensitivity by SSPG method. The levels of SSPG had weak but significant correlation with both %FMD ( $r = -0.175, p < 0.05$ ) and UAER ( $r = 0.181, p < 0.05$ ) independent of age, HbA1c, and systolic blood pressure (Fig. 3A, B).

**4. Discussions**

There were two main findings from this investigation in type 2 diabetic patients. First, diabetic micro- and macro-albuminuric patients showed significant reduction of %FMD compared with normoalbuminuric patients. This finding suggests that the endothelial dysfunction may account for the association between atherosclerosis and albuminuria in diabetic patients. Second, the level of SSPG was significantly associated with both UAER and %FMD. This finding suggests that insulin resistance may play a role in both atherosclerosis and nephropathy in type 2 diabetic patients.

In diabetic patients, %FMD is decreased compared with healthy control [13,14]. These reports indicated that diabetes mellitus is associated with endothelial dysfunction due to



**Fig. 3 – Correlation between SSPG and UAE (A), and correlation between SSPG and %FMD (B) in normo- and micro-albuminuric patients.**

impaired NO production. However the involvement of endothelial dysfunction in diabetic nephropathy has been unclarified. We demonstrated that microalbuminuric and macroalbuminuric patients showed significant decreased %FMD compared with normoalbuminuric patients. In contrast, there was no significant difference of vWF between normoalbuminuric patients and microalbuminuric patients. vWF is a product of vascular endothelial cell, and induces coagulation and platelet aggregation [15]. These findings suggest that endothelial dysfunction due to impaired NO production is specifically induced in micro- and macro-albuminuric patients. One recent report showed that coronary endothelium-dependent dilatation was impaired in a rat model of spontaneous albuminuria [16] supporting this hypothesis. It has been reported that renal NO production was decreased in rodent diabetic model [17]. This report suggests that decrease of NO production may play a role in the

**Table 2 – Parameters of atherosclerosis in diabetic patients with normoalbuminuria, microalbuminuria, and overt nephropathy**

Parameter	Stage of nephropathy		
	Normoalbuminuria	Microalbuminuria	Macroalbuminuria
von Willebrand factor (%)	147 ± 44	146 ± 44	143 ± 41
High-sensitive CRP (ng/ml)	976 ± 1401	951 ± 1110	1113 ± 1187
ADMA (nmol/ml)	0.45 ± 0.06	0.47 ± 0.07	0.55 ± 0.11*

\* $p < 0.001$  vs. normoalbuminuria, mean ± S.D.



progression of diabetic nephropathy as well as atherosclerosis. We investigated serum ADMA levels in diabetic patients. There was no significant difference of ADMA levels between normo- and micro-albuminuric patients, suggesting that the reduction of %FMD in microalbuminuric patients might not be resulted from the elevation of ADMA. However, in macro-albuminuric patients, ADMA level was significantly higher than normoalbuminuric patients. Vallance et al. reported that the level of ADMA was elevated in patients with chronic renal failure and suggested the involvement of this in coronary artery disease [18]. They indicate that the elevation of ADMA might be associated with atherosclerosis in patients with chronic renal disease [18]. Thus, this finding suggests that the elevation of ADMA might be associated with atherosclerotic change in diabetic patients with macroalbuminuria.

An association between chronic low-grade inflammation and development of atherosclerotic disease has been observed in basic and clinical studies [7,19-21]. Furthermore, diabetic patients have higher CRP levels than normal subjects, suggesting that chronic inflammation may contribute diabetic atherosclerotic complication [22]. An association between micro- and macro-albuminuria and inflammation has also been reported [23,24]. However, several other studies showed that inflammatory molecules were not associated with micro- and macro-albuminuria [25-27]. Thus the knowledge of this association is still controversial. Also we could not demonstrate the association between CRP and development of microalbuminuria in this study. Our data suggested that chronic low-grade inflammation might not be involved in the association between atherosclerosis and microalbuminuria. However, since this study was performed by cross-sectional analysis and other inflammatory marker was not measured, further study is necessary for demonstrating this hypothesis.

Insulin resistance has been reported to play an important role in the development and progression of atherosclerotic coronary disease [8,9]. Recently the association between insulin resistance and microalbuminuria was also reported [10]. Nakamura et al. demonstrated that administration of pioglitazone to diabetic patients attenuated UAER [28]. In this study, we showed that both the UAER and %FMD were significantly correlated to the level of SSPG. These findings suggest that insulin resistance may be involved in both the elevated urinary albumin excretion and endothelial dysfunction due to impaired NO production. However, HOMA-IR, another insulin sensitivity marker which reflects insulin sensitivity in both the liver and the periphery, did not show significant difference among three groups, suggesting that particularly peripheral insulin resistance may be important for the pathogenesis of atherosclerosis and diabetic nephropathy.

In summary, we showed that %FMD of micro- and macro-albuminuric patients was decreased compared with those of normoalbuminuric patients, without showing significant difference in other various atherosclerotic markers. Furthermore, the level of SSPG was significantly correlated to UAER and %FMD. These findings suggest that endothelial dysfunction which may be due to impaired NO production underlies the mechanism of association between elevated urinary albumin excretion and atherosclerosis in diabetic patients, and that peripheral insulin

resistance might be possibly involved in both diabetic nephropathy and atherosclerosis.

## Acknowledgement

This work was supported by the Research Grant for Cardiovascular Diseases (16C-2) from the Ministry of Health, Labour and Welfare.

## REFERENCES

- [1] S.F. Dinneen, H.C. Gerstein, The association of microalbuminuria and mortality in non-insulin dependent diabetes mellitus: a systematic overview of the literature, *Arch. Intern. Med.* 157 (1997) 1413-1418.
- [2] J.S. Yudkin, R.D. Forrest, C.A. Jackson, Microalbuminuria as predictor of vascular disease in non-diabetic subjects: Islington diabetes survey, *Lancet* 2 (1988) 530-533.
- [3] R.H. Boger, E.S. Ron, L-Arginine improves vascular function by overcoming the deleterious effects of ADMA, a novel cardiovascular risk factor, *Altern. Med. Rev.* 10 (2005) 14-23.
- [4] I. Fleming, R. Busse, Significant transduction of eNOS activation, *Cardiovasc. Res.* 43 (1999) 532-541.
- [5] R. Busse, I. Fleming, Pulsatile stretch and shear stress: physical stimuli determining the production of endothelium-derived relaxing factors, *J. Vasc. Res.* 35 (1998) 73-84.
- [6] D.S. Celemajer, K.E. Sorensen, V.M. Gooch, D.J. Spiegelhalter, O.I. Miller, I.D. Sullivan, et al., Non-invasive detection of endothelial dysfunction in children and adults at risk of atherosclerosis, *Lancet* 340 (1992) 1111-1115.
- [7] R. Ross, Atherosclerosis: an inflammatory disease, *N. Engl. J. Med.* 340 (1999) 115-126.
- [8] R.A. De Fronzo, E. Ferrannini, Insulin resistance: a multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia and atherosclerotic cardiovascular disease, *Diabetes Care* 14 (1991) 173-194.
- [9] S. Inchiostro, G. Bertoli, G. Zanette, Evidence of higher insulin resistance in NIDDM patients with ischemic heart disease, *Diabetologia* 37 (1994) 597-603.
- [10] M. Emoto, Y. Nishizawa, K. Maekawa, T. Kawagishi, K. Kogawa, Y. Hiura, et al., Insulin resistance in non-insulin-dependent diabetic patients with diabetic nephropathy, *Metabolism* 46 (1997) 1013-1018.
- [11] M. Suzuki, I. Takamizawa, K. Suzuki, A. Hiuge, T. Horio, Y. Yoshimasa, et al., Close association of endothelial dysfunction with insulin resistance and carotid wall thickening in hypertension, *Am. J. Hypertens.* 17 (2004) 228-232.
- [12] D.R. Matthews, J.P. Hosker, A.S. Rudenski, B.A. Naylor, D.F. Treacher, R.C. Turner, Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man, *Diabetologia* 28 (1985) 412-419.
- [13] N. Ihlemann, K.H. Stokholm, P.C. Eskildsen, Impaired vascular reactivity is present despite normal levels of von Willebrand factor in patients with uncomplicated type 2 diabetes, *Diabetes Med.* 19 (2002) 476-481.
- [14] R.V. Hogikyan, A.T. Galecki, B. Pitt, J.B. Halter, D.A. Greene, M.A. Supiano, Specific impairment of endothelium-dependent vasodilatation in subjects with type 2 diabetes independent of obesity, *J. Clin. Endocrinol. Metab.* 83 (1998) 1946-1952.

- [15] B.M. Ewenstein, Vascular biology of von Willebrand factor, in: G.V.R. Born, C.J. Schwartz (Eds.), *Vascular Endothelium*, Schattauer, Stuttgart, 1997, pp. 107-123.
- [16] S. Gschwend, S.J. Pinto-Siersma, H. Buikema, Y.M. Pinto, W.H. Van Gilst, A. Schulz, et al., Impaired coronary endothelial function in a rat model of spontaneous albuminuria, *Kidney Int.* 62 (2002) 181-191.
- [17] A. Erdely, G. Freshour, D.A. Maddox, J.L. Olson, L. Samsell, C. Baylis, Renal disease in rats with type 2 diabetes is associated with decreased renal nitric oxide production, *Diabetologia* 47 (2004) 1672-1676.
- [18] P. Vallance, A. Leone, A. Calver, J. Collier, S. Moncada, Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure, *Lancet* 339 (1992) 572-575.
- [19] J. Torzewski, M. Torzewski, D.E. Bowyer, M. Frohlich, W. Koenig, J. Waltenberger, et al., C-reactive protein frequently colocalizes with the terminal component complex in the intima of early atherosclerotic lesions of human coronary arteries, *Arterioscler. Thromb. Vasc. Biol.* 18 (1998) 1386-1392.
- [20] P.M. Ridker, M. Cushman, M.J. Stampfer, R. Tracy, C.H. Hennekens, Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men, *N. Engl. J. Med.* 336 (1997) 973-979.
- [21] F. Haverkate, S.G. Thompson, S.D.M. Pyke, J.R. Gallimore, M.B. Pepys, The European Concerted Action on Thrombosis and Disabilities Angina Pectoris Study Group, Production of C-reactive protein and risk of coronary events in stable and unstable angina, *Lancet* 349 (1997) 462-466.
- [22] E.S. Ford, Body mass index, diabetes, and C-reactive protein among U.S. adults, *Diabetes Care* 22 (1999) 1971-1977.
- [23] C.D.A. Stehouwer, M.A. Gall, J.W.R. Twisk, E. Knudsen, J.J. Emeis, H.H. Parving, Increased urinary albumin excretion, endothelial dysfunction, and chronic low-grade inflammation in type 2 diabetes. Progressive, interrelated, and independently associated with risk of death, *Diabetes* 51 (2002) 1157-1165.
- [24] K.E. Paisley, M. Beaman, J.E. Tooke, V. Mohamed-Ali, G.D.O. Lowe, A.C. Shore, Endothelial dysfunction and inflammation in asymptomatic proteinuria, *Kidney Int.* 63 (2003) 624-633.
- [25] B. Mirup, M. Demaat, P. Rossing, J. Gram, C. Kluft, J. Jespersen, Elevated fibrinogen and the relation to acute phase response in diabetic nephropathy, *Thromb. Res.* 81 (1996) 485-490.
- [26] M.A. Crook, P. Tutt, J.C. Pickup, Elevated serum sialic acid concentration in NIDDM and its relationship to blood pressure and retinopathy, *Diabetes Care* 16 (1993) 57-60.
- [27] O. Ortega, I. Rodriguez, P. Gallar, A. Carreno, M. Ortiz, A. Molina, et al., Significance of high C-reactive protein levels in pre-dialysis patients, *Nephrol. Dial. Transplant* 17 (2002) 1105-1109.
- [28] T. Nakamura, C. Ushiyama, S. Osada, M. Hara, N. Shimada, H. Koide, Pioglitazone reduces urinary podocyte excretion in type 2 diabetes patients with microalbuminuria, *Metabolism* 50 (2001) 1193-1196.