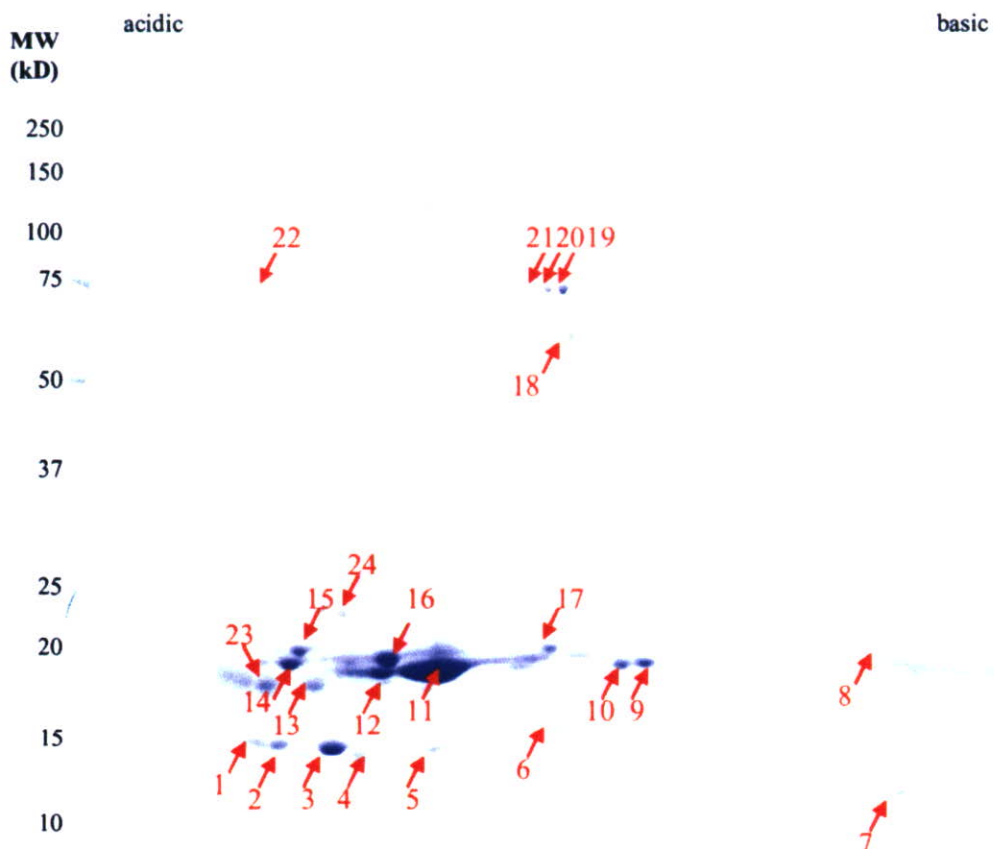


図 23 ラット尿中タンパク質の2次元電気泳動解析



(MALDI-TOF マス解析による同定結果)

spot No.	MS	MS/MS
1	nd	nd
2	nd	nd
3	nd	AY327506
4	nd	prostatic steroid-binding protein chain C3 precursor
5	nd	nd
6	nd	urinary protein 2 precursor
7	nd	nd
8	RATVPSP	RATVPSP
9	alpha-2u-globulin, cain A	alpha-2u-globulin, cain A (precursor)
10	alpha-2u-globulin, cain A	alpha-2u-globulin, cain A (precursor)
11	alpha-2u-globulin, cain A	alpha-2u-globulin, cain A (precursor)
12	alpha-2u-globulin, cain A	alpha-2u-globulin, cain A (precursor)
13	nd	urinary protein 2 precursor
14	alpha-2u-globulin (precursor)	alpha-2u-globulin (precursor)
15	alpha-2u-globulin (precursor)	alpha-2u-globulin (precursor)
16	nd	alpha-2u-globulin, cain A (precursor)
17	nd	alpha-2u-globulin, cain A (precursor)
18	Alpha-amylase	Alpha-amylase
19	Albumin	nd
20	Albumin	serum albumin precursor
21	Albumin	nd
22	Albumin	nd
23	nd	nd
24	alpha-2u-globulin, cain A	nd

図 24 抗体結合磁気ビーズによる高発現タンパク質の除去

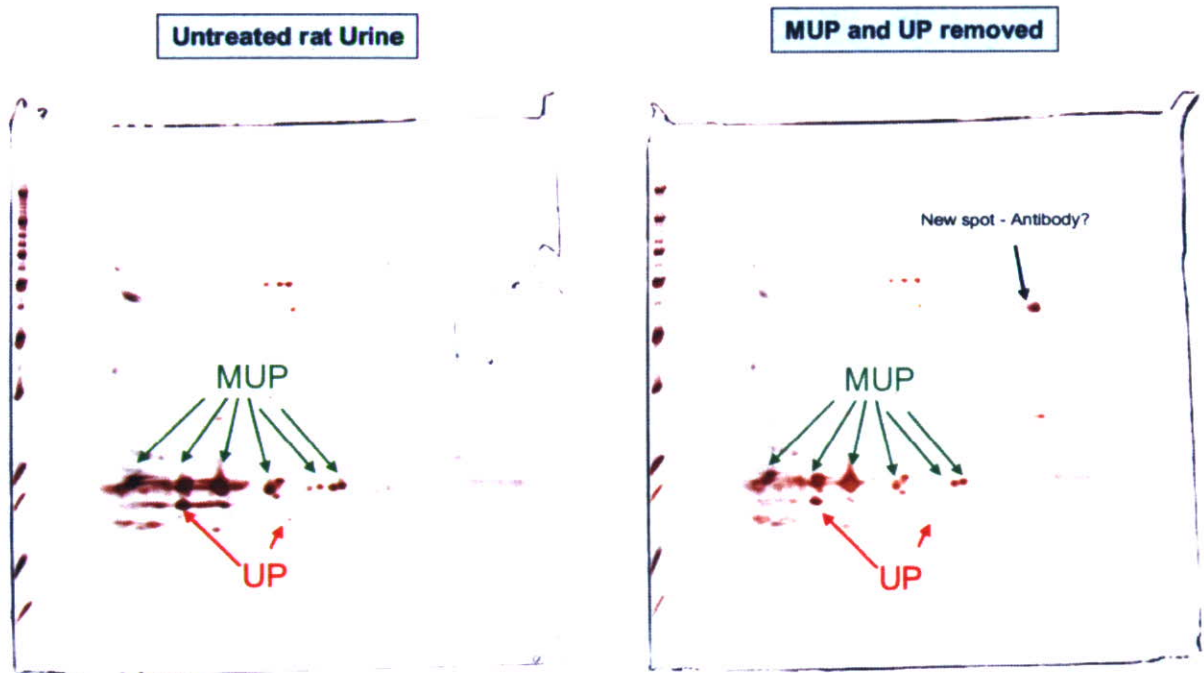


表1 ヒトブロード肝サイトゾル中でMASCOT検索により同定されたタンパク質

NAME	ID	Description
CPM8_HUMAN	(P31327)	Carbonic anhydrase 8 [amino acid] mitochondrial precursor (EC 6.3.4.18)
ADH9_HUMAN	(P00278)	Alcohol dehydrogenase beta chain (EC 1.1.1.1)
ADH4_HUMAN	(P00279)	Alcohol dehydrogenase alpha 1 chain (EC 1.1.1.1)
ALDO5_HUMAN	(P00882)	Fruktose-1,6-bisphosphatase 5 (EC 4.1.3.13) (Liver-type aldolase)
ADH8_HUMAN	(P07327)	Alcohol dehydrogenase alpha chain (EC 1.1.1.1)
ADH9_HUMAN	(P00278)	Alcohol dehydrogenase beta chain (EC 1.1.1.1)
G3P2_HUMAN	(P04408)	Glycerinaldehyd-3-phosphat dehydrogenase, liver (EC 1.2.1.12) (GAPDH)
FAPD1_HUMAN	(P07148)	Fatty acyl-CoA synthetase, liver (L-FAPD)
GSTA1_HUMAN	(P07383)	Glutathione S-transferase A1 (EC 2.3.1.18) (GST A1) (MA subunit 1) (GST-meridian) (GSTA1-1)
ACTA1_HUMAN	(P00709)	Actin, cytoplasmic 1 (beta-actin)
ACTA2_HUMAN	(P07295)	Actin, cardiac muscle isoform (Alpha-actin-2)
HBB_HUMAN	(P00817)	Hemoglobin beta subunit (Hemoglobin beta chain) (beta-globin)
GS1A2_HUMAN	(P09210)	Glycylglycine transferase A2 (EC 2.3.1.18) (GT2) (MA subunit 2) (GST-gamma 2) (GSTA2-2)
HBA_HUMAN	(P00805)	Hemoglobin alpha subunit (Hemoglobin alpha chain) (alpha-globin)
FTCD_HUMAN	(Q52944)	Ferrocenyltetrahydrocyclohexadienone (Formyl transferase-associated protein) (FTCD) (LCHC1)
DNS2_HUMAN	(P00877)	Glutamate dehydrogenase 1, mitochondrial precursor (EC 1.4.1.3) (GDH)
GS1A1_HUMAN	(Q14722)	Glycylglycine transferase A1 (EC 2.3.1.18) (GT1) (MA subunit 1) (GST-gamma 1) (GSTA1-1)
PRDX1_HUMAN	(Q08930)	Peroxisidase 1 (EC 1.11.1.18) (Thiol oxidase peroxidase 2) (Thiol oxidase-dependent peroxidase)
PRDX2_HUMAN	(P25041)	Peroxisidase 2 (EC 1.11.1.18) (Alkyl hydroperoxidase 2) (EC-type peroxidase) (t-Cys PRX)
EPAS1_HUMAN	(P09104)	Epoetin receptor 1 (EPOR) (Epoetin receptor) (A-1) (EPOR-1)
ASBY_HUMAN	(P00888)	Argininosuccinyl synthetase (EC 6.3.4.8) (Citrullinase-argininase)
DCDC2_HUMAN	(Q71414)	Dystroglycan 2 (EC 1.1.1.90) (Dys) (Dysglycyl synthetase)
ARID1_HUMAN	(P00889)	Artemin 1 (EC 3.3.2.1) (Type I artemin) (Liver-type artemin)
ECHM1_HUMAN	(P30242)	Enoyl-CoA hydratase, mitochondrial precursor (EC 4.2.1.17) (Enoyl-CoA acyl-CoA hydratase)
FABP_HUMAN	(P00277)	Fatty acid synthase (EC 2.3.1.85) (Fatty acid synthase) (Fatty acid synthase)
AL1A_HUMAN	(P00332)	Aldehyde dehydrogenase 1 (EC 1.2.1.30) (ALDH1) (ALDH1) (Aldehyde dehydrogenase family 1)
STXA1_HUMAN	(Q08202)	Alcohol sulfotransferase (EC 2.3.2.3) (Hydroxyphenyl sulfotransferase) (STX1)
HMCB2_HUMAN	(P08166)	Hydroxyethylglutaryl-CoA synthase, mitochondrial precursor (EC 2.3.2.10)
IDIC_HUMAN	(Q78874)	Isoform dehydrogenase (NADP) cytoplasmic (EC 1.1.1.42) (Oxidoreductase dehydrogenase)
PEBP_HUMAN	(P30889)	Protein-binding protein (PEBP) (Protein binding protein) (MCPBP)
ECDC1_HUMAN	(P04112)	Enoyl-CoA hydratase (EC 4.2.1.17) (Enoyl-CoA hydratase)
PP2A_HUMAN	(P02937)	Protein phosphatase 2A (EC 3.1.3.1) (PP2A) (Serine/threonine phosphatase)
AKC1_HUMAN	(Q08483)	Adenylate cyclase family 1 member C1 (EC 1.1.1.1) (G-protein-coupled receptor dehydrogenase)
CHSD_HUMAN	(P10092)	Chondroitinase (EC 3.1.1.1) (CHSD) (Chondroitinase)
ALDH2_HUMAN	(P00911)	Aldehyde dehydrogenase, mitochondrial precursor (EC 1.2.1.3) (ALDH2) (ALDH2)
CHST1_HUMAN	(P00912)	Chondroitinase (EC 3.1.1.1) (CHST1) (Chondroitinase)
MBD_HUMAN	(P03042)	Methyl-binding domain protein (Methyl-binding domain protein)
TH1_HUMAN	(P24722)	Thymidylate synthase, mitochondrial precursor (EC 2.3.1.2)
LDNA_HUMAN	(P09133)	Lactonase (EC 1.1.1.37) (LDNA) (Lactonase) (LDNA)
ANKK1_HUMAN	(P08123)	AnkK1 (AnkK1) (Lipocalin V) (P84) (P70) (P74) (C) (Lipocalin-like-20)
QSOX1_HUMAN	(Q14788)	Quercetin oxidase, cytoplasmic isoform 1 (QSOX1) (Quercetin oxidase)
QSOX2_HUMAN	(Q14789)	Quercetin oxidase, cytoplasmic isoform 2 (QSOX2) (Quercetin oxidase)
AMT2_HUMAN	(Q01484)	Amiloride (B) (Amiloride B) (Amiloride, non-stimulatory)
THM2_HUMAN	(P27260)	Thymidylate synthase, mitochondrial precursor (EC 2.3.1.2) (Thymidylate synthase)
BYHE1_HUMAN	(Q08911)	By-heparinase (EC 3.1.1.1) (By-heparinase) (By-heparinase)
IQGAP2_HUMAN	(Q12874)	IQ motif-containing G-actin-binding protein 2 (IQGAP2) (IQGAP2)
HBO1_HUMAN	(P00819)	Hemoglobin gamma-1 chain (Hemoglobin gamma-1 chain) (Hb F gamma 1)
MLL3_HUMAN	(Q08924)	Mixed-lineage leukemia 3 protein 3 homolog
HEB1_HUMAN	(P00829)	Hemoglobin beta chain (Hemoglobin beta chain) (beta-globin)
ZNF108_HUMAN	(Q08272)	Zinc finger protein 108 homolog (ZNF108) (Zinc finger protein 47)
TEAD1_HUMAN	(Q08103)	Tead1
UDPA1_HUMAN	(Q07131)	UDP-glucose 4-epimerase (EC 5.2.1.8) (UDP-glucose pyrophosphorylase)
ADH7_HUMAN	(P00284)	Alcohol dehydrogenase class IV member 7 (EC 1.1.1.1) (Alcohol dehydrogenase)
FXR1_HUMAN	(P08124)	Farnesyl transferase 1 (EC 2.3.1.17) (Farnesyl transferase)
ACBL1_HUMAN	(Q08133)	Long-chain fatty-acyl-CoA ligase 1 (EC 6.3.1.3) (Long-chain acyl-CoA synthetase 1)
PAB3_HUMAN	(Q12107)	Poly(A)-binding protein 3 (PAB3) (Poly(A)-binding protein 3)
THSD1_HUMAN	(Q08109)	Thrombospondin 1 (EC 1.1.1.1) (Thrombospondin 1)
ATP5A1_HUMAN	(P08047)	ATP synthase F0 complex subunit 1 (EC 3.6.3.14)
LYRC1_HUMAN	(Q08105)	Lysine-specific proteinase 1 (EC 3.4.21.1) (Lysine-specific proteinase 1) (LSP1)
SH3BP1_HUMAN	(Q00738)	SH3 domain protein 1 (EC 1.1.1.1) (SH3BP1) (SH3BP1)
NCOA2_HUMAN	(Q14688)	Nuclear coactivator 2 (NcoA2) (Nuclear coactivator 2) (Nuclear coactivator 2)
BPTN_HUMAN	(Q08938)	BPTN (BPTN) (BPTN) (BPTN)
QSOX1_HUMAN	(Q14788)	Quercetin oxidase, cytoplasmic isoform 1 (QSOX1) (Quercetin oxidase)
QSOX2_HUMAN	(Q14789)	Quercetin oxidase, cytoplasmic isoform 2 (QSOX2) (Quercetin oxidase)
QSOX3_HUMAN	(Q14790)	Quercetin oxidase, cytoplasmic isoform 3 (QSOX3) (Quercetin oxidase)
QSOX4_HUMAN	(Q14791)	Quercetin oxidase, cytoplasmic isoform 4 (QSOX4) (Quercetin oxidase)
QSOX5_HUMAN	(Q14792)	Quercetin oxidase, cytoplasmic isoform 5 (QSOX5) (Quercetin oxidase)
QSOX6_HUMAN	(Q14793)	Quercetin oxidase, cytoplasmic isoform 6 (QSOX6) (Quercetin oxidase)
QSOX7_HUMAN	(Q14794)	Quercetin oxidase, cytoplasmic isoform 7 (QSOX7) (Quercetin oxidase)
QSOX8_HUMAN	(Q14795)	Quercetin oxidase, cytoplasmic isoform 8 (QSOX8) (Quercetin oxidase)
QSOX9_HUMAN	(Q14796)	Quercetin oxidase, cytoplasmic isoform 9 (QSOX9) (Quercetin oxidase)
QSOX10_HUMAN	(Q14797)	Quercetin oxidase, cytoplasmic isoform 10 (QSOX10) (Quercetin oxidase)
QSOX11_HUMAN	(Q14798)	Quercetin oxidase, cytoplasmic isoform 11 (QSOX11) (Quercetin oxidase)
QSOX12_HUMAN	(Q14799)	Quercetin oxidase, cytoplasmic isoform 12 (QSOX12) (Quercetin oxidase)
QSOX13_HUMAN	(Q14800)	Quercetin oxidase, cytoplasmic isoform 13 (QSOX13) (Quercetin oxidase)
QSOX14_HUMAN	(Q14801)	Quercetin oxidase, cytoplasmic isoform 14 (QSOX14) (Quercetin oxidase)
QSOX15_HUMAN	(Q14802)	Quercetin oxidase, cytoplasmic isoform 15 (QSOX15) (Quercetin oxidase)
QSOX16_HUMAN	(Q14803)	Quercetin oxidase, cytoplasmic isoform 16 (QSOX16) (Quercetin oxidase)
QSOX17_HUMAN	(Q14804)	Quercetin oxidase, cytoplasmic isoform 17 (QSOX17) (Quercetin oxidase)
QSOX18_HUMAN	(Q14805)	Quercetin oxidase, cytoplasmic isoform 18 (QSOX18) (Quercetin oxidase)
QSOX19_HUMAN	(Q14806)	Quercetin oxidase, cytoplasmic isoform 19 (QSOX19) (Quercetin oxidase)
QSOX20_HUMAN	(Q14807)	Quercetin oxidase, cytoplasmic isoform 20 (QSOX20) (Quercetin oxidase)
QSOX21_HUMAN	(Q14808)	Quercetin oxidase, cytoplasmic isoform 21 (QSOX21) (Quercetin oxidase)
QSOX22_HUMAN	(Q14809)	Quercetin oxidase, cytoplasmic isoform 22 (QSOX22) (Quercetin oxidase)
QSOX23_HUMAN	(Q14810)	Quercetin oxidase, cytoplasmic isoform 23 (QSOX23) (Quercetin oxidase)
QSOX24_HUMAN	(Q14811)	Quercetin oxidase, cytoplasmic isoform 24 (QSOX24) (Quercetin oxidase)
QSOX25_HUMAN	(Q14812)	Quercetin oxidase, cytoplasmic isoform 25 (QSOX25) (Quercetin oxidase)
QSOX26_HUMAN	(Q14813)	Quercetin oxidase, cytoplasmic isoform 26 (QSOX26) (Quercetin oxidase)
QSOX27_HUMAN	(Q14814)	Quercetin oxidase, cytoplasmic isoform 27 (QSOX27) (Quercetin oxidase)
QSOX28_HUMAN	(Q14815)	Quercetin oxidase, cytoplasmic isoform 28 (QSOX28) (Quercetin oxidase)
QSOX29_HUMAN	(Q14816)	Quercetin oxidase, cytoplasmic isoform 29 (QSOX29) (Quercetin oxidase)
QSOX30_HUMAN	(Q14817)	Quercetin oxidase, cytoplasmic isoform 30 (QSOX30) (Quercetin oxidase)
QSOX31_HUMAN	(Q14818)	Quercetin oxidase, cytoplasmic isoform 31 (QSOX31) (Quercetin oxidase)
QSOX32_HUMAN	(Q14819)	Quercetin oxidase, cytoplasmic isoform 32 (QSOX32) (Quercetin oxidase)
QSOX33_HUMAN	(Q14820)	Quercetin oxidase, cytoplasmic isoform 33 (QSOX33) (Quercetin oxidase)
QSOX34_HUMAN	(Q14821)	Quercetin oxidase, cytoplasmic isoform 34 (QSOX34) (Quercetin oxidase)
QSOX35_HUMAN	(Q14822)	Quercetin oxidase, cytoplasmic isoform 35 (QSOX35) (Quercetin oxidase)
QSOX36_HUMAN	(Q14823)	Quercetin oxidase, cytoplasmic isoform 36 (QSOX36) (Quercetin oxidase)
QSOX37_HUMAN	(Q14824)	Quercetin oxidase, cytoplasmic isoform 37 (QSOX37) (Quercetin oxidase)
QSOX38_HUMAN	(Q14825)	Quercetin oxidase, cytoplasmic isoform 38 (QSOX38) (Quercetin oxidase)
QSOX39_HUMAN	(Q14826)	Quercetin oxidase, cytoplasmic isoform 39 (QSOX39) (Quercetin oxidase)
QSOX40_HUMAN	(Q14827)	Quercetin oxidase, cytoplasmic isoform 40 (QSOX40) (Quercetin oxidase)
QSOX41_HUMAN	(Q14828)	Quercetin oxidase, cytoplasmic isoform 41 (QSOX41) (Quercetin oxidase)
QSOX42_HUMAN	(Q14829)	Quercetin oxidase, cytoplasmic isoform 42 (QSOX42) (Quercetin oxidase)
QSOX43_HUMAN	(Q14830)	Quercetin oxidase, cytoplasmic isoform 43 (QSOX43) (Quercetin oxidase)
QSOX44_HUMAN	(Q14831)	Quercetin oxidase, cytoplasmic isoform 44 (QSOX44) (Quercetin oxidase)
QSOX45_HUMAN	(Q14832)	Quercetin oxidase, cytoplasmic isoform 45 (QSOX45) (Quercetin oxidase)
QSOX46_HUMAN	(Q14833)	Quercetin oxidase, cytoplasmic isoform 46 (QSOX46) (Quercetin oxidase)
QSOX47_HUMAN	(Q14834)	Quercetin oxidase, cytoplasmic isoform 47 (QSOX47) (Quercetin oxidase)
QSOX48_HUMAN	(Q14835)	Quercetin oxidase, cytoplasmic isoform 48 (QSOX48) (Quercetin oxidase)
QSOX49_HUMAN	(Q14836)	Quercetin oxidase, cytoplasmic isoform 49 (QSOX49) (Quercetin oxidase)
QSOX50_HUMAN	(Q14837)	Quercetin oxidase, cytoplasmic isoform 50 (QSOX50) (Quercetin oxidase)
QSOX51_HUMAN	(Q14838)	Quercetin oxidase, cytoplasmic isoform 51 (QSOX51) (Quercetin oxidase)
QSOX52_HUMAN	(Q14839)	Quercetin oxidase, cytoplasmic isoform 52 (QSOX52) (Quercetin oxidase)
QSOX53_HUMAN	(Q14840)	Quercetin oxidase, cytoplasmic isoform 53 (QSOX53) (Quercetin oxidase)
QSOX54_HUMAN	(Q14841)	Quercetin oxidase, cytoplasmic isoform 54 (QSOX54) (Quercetin oxidase)
QSOX55_HUMAN	(Q14842)	Quercetin oxidase, cytoplasmic isoform 55 (QSOX55) (Quercetin oxidase)
QSOX56_HUMAN	(Q14843)	Quercetin oxidase, cytoplasmic isoform 56 (QSOX56) (Quercetin oxidase)
QSOX57_HUMAN	(Q14844)	Quercetin oxidase, cytoplasmic isoform 57 (QSOX57) (Quercetin oxidase)
QSOX58_HUMAN	(Q14845)	Quercetin oxidase, cytoplasmic isoform 58 (QSOX58) (Quercetin oxidase)
QSOX59_HUMAN	(Q14846)	Quercetin oxidase, cytoplasmic isoform 59 (QSOX59) (Quercetin oxidase)
QSOX60_HUMAN	(Q14847)	Quercetin oxidase, cytoplasmic isoform 60 (QSOX60) (Quercetin oxidase)
QSOX61_HUMAN	(Q14848)	Quercetin oxidase, cytoplasmic isoform 61 (QSOX61) (Quercetin oxidase)
QSOX62_HUMAN	(Q14849)	Quercetin oxidase, cytoplasmic isoform 62 (QSOX62) (Quercetin oxidase)
QSOX63_HUMAN	(Q14850)	Quercetin oxidase, cytoplasmic isoform 63 (QSOX63) (Quercetin oxidase)
QSOX64_HUMAN	(Q14851)	Quercetin oxidase, cytoplasmic isoform 64 (QSOX64) (Quercetin oxidase)
QSOX65_HUMAN	(Q14852)	Quercetin oxidase, cytoplasmic isoform 65 (QSOX65) (Quercetin oxidase)
QSOX66_HUMAN	(Q14853)	Quercetin oxidase, cytoplasmic isoform 66 (QSOX66) (Quercetin oxidase)
QSOX67_HUMAN	(Q14854)	Quercetin oxidase, cytoplasmic isoform 67 (QSOX67) (Quercetin oxidase)
QSOX68_HUMAN	(Q14855)	Quercetin oxidase, cytoplasmic isoform 68 (QSOX68) (Quercetin oxidase)
QSOX69_HUMAN	(Q14856)	Quercetin oxidase, cytoplasmic isoform 69 (QSOX69) (Quercetin oxidase)
QSOX70_HUMAN	(Q14857)	Quercetin oxidase, cytoplasmic isoform 70 (QSOX70) (Quercetin oxidase)
QSOX71_HUMAN	(Q14858)	Quercetin oxidase, cytoplasmic isoform 71 (QSOX71) (Quercetin oxidase)
QSOX72_HUMAN	(Q14859)	Quercetin oxidase, cytoplasmic isoform 72 (QSOX72) (Quercetin oxidase)
QSOX73_HUMAN	(Q14860)	Quercetin oxidase, cytoplasmic isoform 73 (QSOX73) (Quercetin oxidase)
QSOX74_HUMAN	(Q14861)	Quercetin oxidase, cytoplasmic isoform 74 (QSOX74) (Quercetin oxidase)
QSOX75_HUMAN	(Q14862)	Quercetin oxidase, cytoplasmic isoform 75 (QSOX75) (Quercetin oxidase)
QSOX76_HUMAN	(Q14863)	Quercetin oxidase, cytoplasmic isoform 76 (QSOX76) (Quercetin oxidase)
QSOX77_HUMAN	(Q14864)	Quercetin oxidase, cytoplasmic isoform 77 (QSOX77) (Quercetin oxidase)
QSOX78_HUMAN	(Q14865)	Quercetin oxidase, cytoplasmic isoform 78 (QSOX78) (Quercetin oxidase)
QSOX79_HUMAN	(Q14866)	Quercetin oxidase, cytoplasmic isoform 79 (QSOX79) (Quercetin oxidase)
QSOX80_HUMAN	(Q14867)	Quercetin oxidase, cytoplasmic isoform 80 (QSOX80) (Quercetin oxidase)
QSOX81_HUMAN	(Q14868)	Quercetin oxidase, cytoplasmic isoform 81 (QSOX81) (Quercetin oxidase)
QSOX82_HUMAN	(Q14869)	Quercetin oxidase, cytoplasmic isoform 82 (QSOX82) (Quercetin oxidase)
QSOX83_HUMAN	(Q14870)	Quercetin oxidase, cytoplasmic isoform 83 (QSOX83) (Quercetin oxidase)
QSOX84_HUMAN	(Q14871)	Quercetin oxidase, cytoplasmic isoform 84 (QSOX84) (Quercetin oxidase)
QSOX85_HUMAN	(Q14872)	Quercetin oxidase, cytoplasmic isoform 85 (QSOX85) (Quercetin oxidase)
QSOX86_HUMAN	(Q14873)	Quercetin oxidase, cytoplasmic isoform 86 (QSOX86) (Quercetin oxidase)
QSOX87_HUMAN	(Q14874)	Quercetin oxidase, cytoplasmic isoform 87 (QSOX87) (Quercetin oxidase)
QSOX88_HUMAN	(Q14875)	Quercetin oxidase, cytoplasmic isoform 88 (QSOX88) (Quercetin oxidase)
QSOX89_HUMAN	(Q14876)	Quercetin oxidase, cytoplasmic isoform 89 (QSOX89) (Quercetin oxidase)
QSOX90_HUMAN	(Q14877)	Quercetin oxidase, cytoplasmic isoform 90 (QSOX90) (Quercetin oxidase)
QSOX91_HUMAN	(Q14878)	Quercetin oxidase, cytoplasmic isoform 91 (QSOX91) (Quercetin oxidase)
QSOX92_HUMAN	(Q14879)	Quercetin oxidase, cytoplasmic isoform 92 (QSOX92) (Quercetin oxidase)
QSOX93_HUMAN	(Q14880)	Quercetin oxidase, cytoplasmic isoform 93 (QSOX93) (Quercetin oxidase)
QSOX94_HUMAN	(Q14881)	Quercetin oxidase, cytoplasmic isoform 94 (QSOX94) (Quercetin oxidase)
QSOX95_HUMAN	(Q14882)	Quercetin oxidase, cytoplasmic isoform 95 (QSOX95) (Quercetin oxidase)
QSOX96_HUMAN	(Q14883)	Quercetin oxidase, cytoplasmic isoform 96 (QSOX96) (Quercetin oxidase)
QSOX97_HUMAN	(Q14884)	Quercetin oxidase, cytoplasmic isoform 97 (QSOX97) (Quercetin oxidase)
QSOX98_HUMAN	(Q14885)	Quercetin oxidase, cytoplasmic isoform 98 (QSOX98) (Quercetin oxidase)
QSOX99_HUMAN	(Q14886)	Quercetin oxidase, cytoplasmic isoform 99 (QSOX99) (Quercetin oxidase)
QSOX100_HUMAN	(Q14887)	Quercetin oxidase, cytoplasmic isoform 100 (QSOX100) (Quercetin oxidase)

表2 ラット尿中でMASCOT検索により同定されたタンパク質

NAME	ID	Description
ALBU_RAT	(P02770)	Serum albumin precursor
MUP_RAT	(P02781)	Major urinary protein precursor (MUP) (Alpha-2u-globulin) (Alpha(2)-uglobulin)
EGF_RAT	(P07622)	Pro-epidermal growth factor precursor (EGF) [Contains: Epidermal growth factor]
ZP1_RAT	(P22282)	Cystatin-related protein 1 precursor (CRP-1) [Prostatic: 22 kDa glycoprotein P22K16/P22K20]
UROG_RAT	(P27500)	Uromodulin precursor (Tamm-Horsfall urinary glycoprotein) (THUP)
CP3_RAT	(P05544)	Contraptin-like protease inhibitor 3 precursor (CPi-3) [Serine protease inhibitor 1] (SPI)
CP1_RAT	(P05545)	Contraptin-like protease inhibitor 1 precursor (CPi-1) [Kallikrein-binding protein] (KBP)
AMFN_RAT	(P15684)	Aminopetidase N (EC 3.4.11.2) (AMFN) (Aminyl aminopetidase) (Microsomal aminopetidase)
AMBP_RAT	(P04240)	AMBP protein precursor [Contains: Alpha-1-microglobulin; Inter-alpha-trypsin inhibitor lig]
KLK7_RAT	(P36373)	Glandular kallikrein-7, submandibular precursor (EC 3.4.21.35) [Tissue kallikrein]
TRFE_RAT	(P12346)	Sexostransferin precursor (Transferrin) (Siderophilin) (Beta-1-metal binding globulin)
A1AT_RAT	(P17475)	Alpha-1-antitrypsin precursor (Alpha-1-antitrypsin) (Alpha-1-proteinase inhibitor)
UPI1_RAT	(P81827)	Urinary protein 1 precursor (RUP-1) [Liver regeneration-related protein (LRG05)]
UPI2_RAT	(P83121)	Urinary protein 2 precursor (RUP-2)
ACTB_RAT	(P60711)	Actin, cytoplasmic 1 (Beta-actin)
UPI2_RAT	(P81828)	Urinary protein 1 precursor (RUP-1)
ACTC_RAT	(P86035)	Actin, alpha cardiac (Alpha-cardiac actin)
ACTB_RAT	(P60711)	Actin, alpha skeletal muscle (Alpha-actin-1)
AMYP_RAT	(P00880)	Pancreatic alpha-amylase precursor (EC 3.2.1.1) (PA) [(1,4-alpha-D-glucan glucanohydrolase)
PSC3_RAT	(P02780)	Prostatic stator-binding protein C3 chain precursor (Prostatein peptide C3)
KACA_RAT	(P01836)	Ig kappa chain C region, A allele
GGT1_RAT	(P07314)	Gamma-glutamyltransferase 1 precursor (EC 2.3.2.2) (Gamma-glutamyltransferase 1)
HEMO_RAT	(P20069)	Hemopexin precursor
SODC_RAT	(P07632)	Superoxide dismutase [Cu-Zn] (EC 1.15.1.1)
SLC31_RAT	(Q64319)	Neutral and basic amino acid transport protein (BAT) (B(0,+)-type amino acid transport prot
KLJ3_RAT	(P07647)	Submandibular glandular kallikrein-9 precursor (EC 3.4.21.35) [Tissue kallikrein] (S3 kall
HBG2_RAT	(P11517)	Hemoglobin beta-2 subunit (Hemoglobin beta-2 chain) [Beta-2-globin] (Hemoglobin beta chain
KLK1_RAT	(P00758)	Nerve growth factor gamma chain precursor (EC 3.4.21.35) (Gamma-NGF) [Tissue kallikrein]
B2MG_RAT	(P07151)	Beta-2-microglobulin precursor
MEP1A_RAT	(Q06420)	Meprin A alpha-subunit precursor (EC 3.4.24.16) (Entropetidase-2) (MEP-1) (Entropetidase-2)
NOTC2_RAT	(Q90W30)	Neurogenic locus notch homolog protein 2 precursor (Notch 2) [Contains: Notch 2 extracellu
PBAS_RAT	(P15369)	Probasin precursor (PB) (M-40)
SYCP1_RAT	(Q03410)	Synaptonemal complex protein 1 (SCP-1 protein)
SPASM_RAT	(Q03568)	Serine protease inhibitor A3M precursor (Serine protease inhibitor 2.4) (SPI-2.4)
HBA_RAT	(P01946)	Hemoglobin alpha-12 subunit (Hemoglobin alpha-12 chain) (Alpha-12-globin)
ATOX1_RAT	(Q9WHUC4)	Copper transport protein ATOX1 [Metal transport protein ATOX1]
PSC2_RAT	(P02781)	Prostatic stator-binding protein C2 chain precursor (Prostatein peptide C2)
MYH7_RAT	(P02781)	Myosin heavy chain, cardiac muscle beta isoform (MyHC-beta)
DNMT1_RAT	(Q02330)	DNA (cytosine-5)-methyltransferase 1 (EC 2.1.1.37) (Dnmt1) [DNA methyltransferase 1]
KLK12_RAT	(P38378)	Glandular kallikrein-12, submandibular precursor (EC 3.4.21.35) [Tissue kallikrein]
MYH10_RAT	(Q9JL70)	Myosin-10 (Myosin heavy chain, nonmuscle 1b) (Nonmuscle myosin heavy chain 1b)
PLEC1_RAT	(P30427)	Plectin 1 (PLTN) (PCN)
MYH6_RAT	(P02563)	Myosin heavy chain, cardiac muscle alpha isoform (MyHC-alpha)
ZP2_RAT	(P22283)	Cystatin-related protein 2 precursor (Prostatic: 22 kDa glycoprotein P22K15)
PCLO_RAT	(Q9JH56)	Plectin protein (Multidomain presynaptic cytomatrix protein)
MOES_RAT	(Q35763)	Moesin (Membrane-organizing extension spike protein)
GRK4_RAT	(P07507)	G protein-coupled receptor kinase 4 (EC 2.7.1.-) (G protein-coupled receptor kinase GRK4)
CAC1C_RAT	(Q02502)	Voltage-dependent L-type calcium channel alpha-1C subunit (Voltage-gated calcium channel alpha subunit Cav1.2)
CAC1H_RAT	(Q02503)	Voltage-dependent T-type calcium channel alpha-1H subunit
KLJ3_RAT	(P07647)	Glandular kallikrein-3, submandibular (EC 3.4.21.35) [Tissue kallikrein] (S1 kallikrein)
XAB2_RAT	(Q96P90)	XPA-binding protein 2 (Adaptor protein ATH-55)
PLMN_RAT	(Q01177)	Fibrinogen precursor (EC 3.4.21.7) [Contains: Fibrin heavy chain A; Activation peptide;
EZR1_RAT	(P31977)	Ezrin (p81) (Cytovillin) (Villin-2)
EZB2_RAT	(Q03180)	Translation initiation factor eIF-2B delta subunit (eIF-2B GDP-GTP exchange factor)
PGFRA_RAT	(P02786)	Alpha platelet-derived growth factor receptor precursor (EC 2.7.1.112) (PDGF-R-alpha)
FAS_RAT	(P12785)	Fatty acid synthase (EC 2.3.1.85) [Includes: Acyl-carrier-protein] S-acetyltransferase (E
THY2_RAT	(P06062)	Thyroglobulin precursor
CC4_RAT	(Q02502)	Complement C4 precursor [Contains: Complement C4 beta chain; Complement C4 alpha chain; C4
ARN1_RAT	(P41739)	Arn1 hydrocarbon receptor nuclear translocator (ARN1 protein) [Dioxin receptor, nuclear translocat
MAP2_RAT	(P15146)	Microtubule-associated protein 2 (MAP 2) (MAP-2)
MYH11_RAT	(Q03862)	Myosin-11 (Myosin heavy chain, smooth muscle isoform) (SMMHC) (Fragments)
HD_RAT	(P51111)	Huntingtin (Huntington's disease protein homolog) (HD protein)
CSAD_RAT	(Q04611)	Cysteine sulfonic acid decarboxylase (EC 4.1.1.29) (Sulfinoalanine decarboxylase)
FOLH1_RAT	(P07627)	Glutamate carboxypeptidase II (EC 3.4.17.21) (Membrane glutamate carboxypeptidase)
RFP3_RAT	(Q9ER28)	Rho-interacting protein 3 (p18RFP) (RIP3)
PLD2_RAT	(P70496)	Phospholipase D2 (EC 3.1.4.4) (PLD 2) (Choline phosphatase 2) (Phosphatidylcholine-hydroly
MAST1_RAT	(Q810W7)	Microtubule-associated serine/threonine-protein kinase 1 (EC 2.7.1.37) (Synaptrophin-associated serine/threonine-protein kinase)
DPD1_RAT	(Q9Q1Y9)	DNA polymerase gamma subunit 1 (EC 2.7.7.7) (Mitochondrial DNA polymerase catalytic subunit)
PASA_RAT	(Q83787)	Protein-activated serine/threonine kinase 3-kinase regulatory gamma subunit (PK3-kinase p65-alpha subunit) (Pak3
PAK3_RAT	(Q02529)	Serine/threonine-protein kinase PAK 3 (EC 2.7.1.37) (p21-activated kinase 3) (PAK-3) (Beta
CLH_RAT	(P11442)	Clathrin heavy chain
SH3_RAT	(Q9JLW4)	SH3 and multiple arylin repeat domains 3 (Shank3) (Proline-rich synapse associated protein 2) (ProSAP2) (SPANK-2)
PRIC1_RAT	(Q71QF9)	Protein-tyrosine phosphatase 1 (RETN/RSF-interacting LIM domain protein 1)
GBR2_RAT	(P18508)	Gamma-aminobutyric acid receptor gamma-2 subunit precursor (GABA(A) receptor)
B3AT2_RAT	(P23347)	Antion exchange protein 2 (Non-erythroid band 3-like protein) (AE2 anion exchanger) (Solute carrier family 4 member 2) (B3RP)
NOTC1_RAT	(Q07008)	Neurogenic locus notch homolog protein 1 precursor (Notch 1) [Contains: Notch 1 extracellu
VGFR1_RAT	(P03767)	Vascular endothelial growth factor receptor 1 precursor (EC 2.7.1.112) (VEGFR-1) (Tyrosine-protein kinase receptor FLT) (FLT-1)
YME1L1_RAT	(Q92558)	ATP-dependent metalloprotease YME1L1 (EC 3.4.24.-) (YME1-like protein 1) (ATP-dependent me
CYL20_RAT	(Q55156)	Cytoplasmic linker protein 2 (Cytoplasmic linker protein 115) (CLP-115)
PR37_RAT	(Q05347)	26S proteasome regulatory subunit 7 (MSS1 protein)
CAND1_RAT	(P91538)	Cullin-associated NEDD8-dissociated protein 1 (Cullin-associated and neddylation-dissociated protein 1) (p120 CAND1)
ABCC9_RAT	(Q05363)	Sulfonamide receptor 2
MYST3_RAT	(Q51709)	Histone acetyltransferase MYST3 (EC 2.3.1.48) (EC 2.3.1.-) (MYST protein 3)
SPTBN2_RAT	(Q90WNR)	Spectrin beta chain, brain 2 (Spectrin, non-erythroid beta chain 2) (Beta-II spectrin) (S)
ABL2_RAT	(Q06C51)	Actin-binding LIM protein 2 (Actin-binding LIM protein family member 2) (abLIM2)
CASP1_RAT	(P43527)	Caspase-1 precursor (EC 3.4.22.36) (CASP-1) (Interleukin-1 beta convertase) (IL-1BC)
IL6R_RAT	(P22273)	Interleukin-6 receptor alpha chain precursor (IL-6R-alpha) (IL-6R 1)
MINP1_RAT	(Q36217)	Multiple inositol polyphosphate phosphatase 1 precursor (EC 3.1.3.62) (Inositol (1,3,4,5)-tetrakisphosphate 3-phosphatase)
INP6B_RAT	(Q99WV1)	72 kDa inositol polyphosphate 5-phosphatase (EC 3.1.3.36) (Phosphatidylinositol-4,5-bispho
DVL1_RAT	(Q99WV9)	Segment polarity protein dishevelled homolog DVL-1 (Dishevelled-1) (DSH homolog 1)
PSSA_RAT	(Q03789)	Phosphatidylinositol 3-kinase regulatory gamma subunit (PK3-kinase p65-gamma subunit) (PtdIns-3-kinase p65-gamma) (p65PK)
KCM4_RAT	(Q02978)	Calcium-activated potassium channel alpha subunit 1 (Calcium-activated potassium channel,
TNFR1A_RAT	(P22934)	Tumor necrosis factor receptor superfamily member 1A precursor (p50) (TNF-R1)
MYO1E_RAT	(Q03356)	Myosin 1e (Myosin heavy chain myr 3)
PLCG2_RAT	(P24135)	1-phosphatidylinositol-4,5-bisphosphate phospholipase gamma 2 (EC 3.1.4.11)
PSDI_RAT	(Q08761)	26S proteasome non-ATPase regulatory subunit 1 (26S proteasome regulatory subunit RP2)
ALDOC_RAT	(P08117)	Fructose-bisphosphate aldolase C (EC 4.1.2.13) (Brain-type aldolase)
MYH6_RAT	(P12847)	Myosin heavy chain, fast skeletal muscle, embryonic
PCSK5_RAT	(P41413)	Proprotein convertase subtilisin/kexin type 5 precursor (EC 3.4.21.-)
KCNH1_RAT	(Q03472)	Potassium voltage-gated channel subfamily H member 1
SRP54_RAT	(Q064Y5)	Signal recognition particle 54 kDa protein (SRP54)
PTPRV_RAT	(Q04812)	Receptor-type tyrosine-protein phosphatase V precursor (EC 3.1.3.48)
CEL22_RAT	(Q02YF2)	Cedrelin EGF LAG seven-pass G-type receptor 2 (Multiple epidermal growth factor-like domains 3) (Fragment)
CAC1A_RAT	(P41283)	Voltage-dependent P/Q-type calcium channel alpha-1A subunit (Voltage-gated calcium channel
MTTP1_RAT	(Q99WZ3)	Membrane-bound transcription factor site-1 protease precursor (EC 3.4.21.-)
KCNQ1_RAT	(Q02207)	Potassium voltage-gated channel subfamily KQT member 1 (Voltage-gated potassium channel subunit Kv7.1)
RORB_RAT	(P45446)	Nuclear receptor ROR-beta (Nuclear receptor RZR-beta)
SULF1_RAT	(Q08V60)	Extracellular sulfatase Sufl-1 precursor (EC 3.1.6.-) (Sulfatase FP) (RSuFP1)
MBB1A_RAT	(Q36821)	Myb-binding protein 1A (PAR-interacting protein) (PIP)

表3 既知標準混合サンプルのICAT法による定量予測

BSA										
Time	m/z-Light	m/z-Heavy	価数	観イオン[M+H]	Area-Light	Area-Heavy	H : L	Ave.	理論比	実測/理論
22. 055-22. 297	684. 3770	687. 3873	3	2051. 1150	1892	1421	0. 751			
16. 134-16. 376	807. 4157	811. 9334	2	1613. 8234	1589	1486	0. 936	0. 843	1. 0	0. 843
LDH-M										
16. 617-17. 222	709. 8737	714. 3826	2	1418. 7394	13497	7673	0. 569		0. 667	0. 853
SOD										
20. 122-20. 363	586. 3432	590. 8665	2	1171. 6784	30469	20652	0. 678			
21. 451-22. 297	766. 1079	769. 1191	3	2296. 3077	3570	1691	0. 474	0. 576	0. 5	1. 152
RNase A										
14. 322-14. 926	543. 2754	547. 7853	2	1085. 5428	5303	6704	1. 264			
24. 109-24. 832	798. 7677	801. 7729	3	2394. 2871	12550	18428	1. 468			
14. 322-14. 926	837. 9311	842. 4474	2	1674. 8542	5699	8335	1. 463			
24. 349-24. 712	1197. 6519	1202. 1606	2	2394. 2958	19162	27464	1. 433	1. 407	1. 5	0. 938

表4 cICAT法による個別サイトソルタンパク量の比較 (プールドサイトソルに対し)

HG23 (40 / 41)		HK27 (49 / 54)		HG30 (53 / 56)		HG43 (55 / 56)		(比決定 / 同定)
Accession #	Ratio	Accession #	Ratio	Accession #	Ratio	Accession #	Avg HL	
ADHA_HUMAN (P03116)	1.3111	ADHA_HUMAN (P03116)	0.534	ADHA_HUMAN (P03116)	1.0718	ADHA_HUMAN (P03116)	0.5359	
ADHA_HUMAN (P07327)	1.3931	ADHA_HUMAN (P07327)	1.1474	ADHA_HUMAN (P07327)	1.2270	ADHA_HUMAN (P07327)	1.4888	
ADHB_HUMAN (P00325)	1.4522	ADHB_HUMAN (P00325)	1.1494	ADHB_HUMAN (P00325)	1.2739	ADHB_HUMAN (P00325)	1.5482	
ADHG_HUMAN (P00326)	1.4428	ADHG_HUMAN (P00326)	1.1885	ADHG_HUMAN (P00326)	1.2248	ADHG_HUMAN (P00326)	1.6434	
AKC1_HUMAN (Q04828)	1.1888	AKC1_HUMAN (Q04828)	1.4352	AKC1_HUMAN (Q04828)	1.5054	AKC1_HUMAN (Q04828)	1.4061	
AKC2_HUMAN (P52895)	1.1888	AKC2_HUMAN (P52895)	1.4352	AKC2_HUMAN (P52895)	1.5054	AKC2_HUMAN (P52895)	1.6081	
ALBU_HUMAN (P02788)	1.8222	ALBU_HUMAN (P02788)	2.8488	ALBU_HUMAN (P02788)	1.2220	ALBU_HUMAN (P02788)	2.4181	
ALFB_HUMAN (P05087)	1.303	ALFB_HUMAN (P05087)	1.6318	ALFB_HUMAN (P05087)	1.5541	ALFB_HUMAN (P05087)	1.054	
ARGL_HUMAN (P05088)	0.7382	ARGL_HUMAN (P05088)	0.9815	ARGL_HUMAN (P05088)	1.0188	ARGL_HUMAN (P05088)	0.9039	
ASSY_HUMAN (P00966)		ASSY_HUMAN (P00966)	1.1822	ASSY_HUMAN (P00966)	1.4727	ASSY_HUMAN (P00966)	1.1136	
BHMT_HUMAN (Q93028)	1.0285	BHMT_HUMAN (Q93028)	0.8033	BHMT_HUMAN (Q93028)	1.134	BHMT_HUMAN (Q93028)	0.9018	
BLP1_HUMAN (Q9LBR1)		BLP1_HUMAN (Q9LBR1)	1.2448					
CATA_HUMAN (P04040)		CATA_HUMAN (P04040)	1.5489	CATA_HUMAN (P04040)	0.8020	CATA_HUMAN (P04040)	1.2424	
CP5M_HUMAN (P11327)	1.1447	CP5M_HUMAN (P11327)	1.2454	CP5M_HUMAN (P11327)	1.2469	CP5M_HUMAN (P11327)	1.2690	
DEF1_HUMAN (P18865)		DEF1_HUMAN (P18865)	0.8348					
DHA1_HUMAN (P00352)	1.0551	DHA1_HUMAN (P00352)	1.2413	DHA1_HUMAN (P00352)	1.1567	DHA1_HUMAN (P00352)	1.1799	
DHAM_HUMAN (P05091)	1.1082	DHAM_HUMAN (P05091)	1.0569	DHAM_HUMAN (P05091)	1.5259	DHAM_HUMAN (P05091)	1.3544	
DHCA_HUMAN (P18152)	1.0251	DHCA_HUMAN (P18152)	1.1787	DHCA_HUMAN (P18152)	1.3268	DHCA_HUMAN (P18152)	1.2183	
DHE3_HUMAN (P00367)	1.5682			DHE3_HUMAN (P00367)		DHE3_HUMAN (P00367)	1.2133	
DHCO_HUMAN (Q00796)	1.5494							
DOPD_HUMAN (P20048)	0.5875	DOPD_HUMAN (P20048)	1.0614	DOPD_HUMAN (P20048)	0.7498	DHE4_HUMAN (P49448)	1.1804	
ENO A_HUMAN (P08733)	1.101			ENO A_HUMAN (P08733)	1.2876	ENO A_HUMAN (P08733)	1.4443	
ENO B_HUMAN (P13828)	1.0362			ENO B_HUMAN (P13828)	1.1582			
ENO O_HUMAN (P09104)	1.0362			ENO O_HUMAN (P09104)	1.1582			
FAAA_HUMAN (P16830)	0.9861	FAAA_HUMAN (P16830)	0.9871	F16P_HUMAN (P09487)	1.1654	F16P_HUMAN (P09487)	0.8798	
FAS_HUMAN (P49327)	1.3834	FAS_HUMAN (P49327)	2.3302	FAA_HUMAN (P16830)	1.199			
ETCD_HUMAN (Q93954)	1.3304	ETCD_HUMAN (Q93954)	1.4688	FAS_HUMAN (P49327)	2.7567	ETCD_HUMAN (Q93954)	0.8728	
GSP1_HUMAN (P03354)	1.1018	GSP1_HUMAN (P03354)	1.2817	GSP1_HUMAN (P03354)	1.5352	GSP1_HUMAN (P03354)	1.1307	
GSP2_HUMAN (P04406)	1.0108	GSP2_HUMAN (P04406)	1.1409	GSP2_HUMAN (P04406)	1.2589	GSP2_HUMAN (P04406)	1.011	
GHPR_HUMAN (Q9LBR7)	1.2783	GHPR_HUMAN (Q9LBR7)	1.4892	GHPR_HUMAN (Q9LBR7)	1.3408	GHPR_HUMAN (Q9LBR7)	1.5782	
GLYC_HUMAN (P34896)	0.9373	GLYC_HUMAN (P34896)	0.7853	GLYC_HUMAN (P34896)	1.0333	GLYC_HUMAN (P34896)	1.1558	
GTT1_HUMAN (P30711)		GTT1_HUMAN (P30711)		GTT1_HUMAN (P30711)		GTT1_HUMAN (P30711)	1.4208	
HAO1_HUMAN (Q9LBR8)		HAO1_HUMAN (Q9LBR8)	2.4438	HAO1_HUMAN (Q9LBR8)	1.8418	HAO1_HUMAN (Q9LBR8)	1.8087	
HEB_HUMAN (P02025)	2.1978	HEB_HUMAN (P02025)	2.0821	HEB_HUMAN (P02025)	2.0821	HEB_HUMAN (P02025)	1.811	
HEM2_HUMAN (P13710)		HEM2_HUMAN (P13710)	1.2531					
HMCM_HUMAN (P54888)		HMCM_HUMAN (P54888)	1.3887	HMCM_HUMAN (P54888)		HMCM_HUMAN (P54888)	1.0913	
HNT1_HUMAN (P49773)	0.7414			HNT1_HUMAN (P49773)	2.1378	HNT1_HUMAN (P49773)	1.5178	
IF41_HUMAN (P08842)	1.1581			IF41_HUMAN (P08842)	2.1378	IF41_HUMAN (P08842)	1.5178	
IF42_HUMAN (Q14240)	1.1581			IF42_HUMAN (Q14240)		IF42_HUMAN (Q14240)	1.5259	
LDHA_HUMAN (P00338)	0.9771	LDHA_HUMAN (P00338)	0.7848	LDHA_HUMAN (P00338)	1.2233	LDHA_HUMAN (P00338)	1.4545	
LDHB_HUMAN (P07185)	0.9771	LDHB_HUMAN (P07185)	0.8138	LDHB_HUMAN (P07185)	1.0186	LDHB_HUMAN (P07185)	1.0767	
LDHC_HUMAN (P07884)	0.9771	LDHC_HUMAN (P07884)	0.8138	LDHC_HUMAN (P07884)	1.0186	LDHC_HUMAN (P07884)	1.0767	
LGAL_HUMAN (Q04780)		LGAL_HUMAN (Q04780)	1.0446				0.993	
MDHM_HUMAN (P40928)		MDHM_HUMAN (P40928)	0.8712					
MPF_HUMAN (P14174)		MPF_HUMAN (P14174)	1.28	METL_HUMAN (Q00284)	1.7116	MPF_HUMAN (P14174)	0.8533	
				MPB1_HUMAN (P22712)	1.4313	MPB1_HUMAN (P22712)	1.5797	

いずれにおいてもH:L比が出ているタンパク質
 HG30以外においてH:L比が出ているタンパク質
 HG30以外で同定されているタンパク質

表5 マウス個体におけるシグナル変化の共通性の検索

Spot No	m/z	Charge	Olive oil						HEM						Quinolone					
			mouse-1	RT	mouse-2	RT	mouse-3	RT	mouse-1	RT	mouse-2	RT	mouse-3	RT	mouse-1	RT	mouse-2	RT	mouse-3	RT
1	888	2+	UP	26	UP	24	UP	28	DN	24	DN	24	AB	22	DN	27	DN	27	DN	26
2	854	2+	AB	24	AB	25	AB	29	UP	22	UP	24	UP	28	---	---	---	---	---	---
3	888	1+	UP	15	UP	16	PR	20	DN	15	DN	18	DN	20	DN	20	DN	22	DN	20
4	484	2+	UP	30	UP	30	UP	35	AB	27	AB	30	AB	34	---	---	---	---	---	---
5	488.7515	2+	UP	30	UP	32	UP	35	DN	30	DN	30	AB	27	DN	32	DN	34	DN	33
6	437	2+	UP	22	UP	25	UP	25	AB	18	AB	22	AB	27	DN	25	PR	26	DN	26
7	791	2+	AB	15	AB	18	AB	22	UP	15	UP	15	UP	15	---	---	---	---	---	---
8	789	1+	AB	22	AB	22	AB	22	UP	20	UP	24	UP	22	---	---	---	---	---	---
9	780	2+	UP	27	UP	27	UP	32	AB	25	AB	25	AB	30	AB	30	DN	25	AB	30
10	738	1+	AB	40	DN	45	AB	44	UP	44	UP	46	UP	48	---	---	---	---	---	---
11	888	2+	AB	22	AB	22	AB	22	UP	20	UP	25	UP	25	---	---	---	---	---	---
12	888	2+	DN	45	DN	46	DN	46	UP	44	UP	43	UP	48	---	---	---	---	---	---
13	881	2+	UP	25	UP	22	UP	30	AB	25	AB	25	AB	28	---	---	---	---	---	---
14	880	2+	PR	48	PR	38	DN	43	AB	36	AB	37	AB	40	---	---	---	---	---	---
15	813.7	2+	AB	70	AB	70	AB	70	UP	70	UP	70	UP	70	---	---	---	---	---	---
16	888	2+	UP	33	UP	36	DN	40	DN	33	DN	36	AB	40	---	---	---	---	---	---
17	842	2+	UP	55	UP	58	DN	58	AB	55	DN	53	AB	60	---	---	---	---	---	---
18	838	1+	AB	43	AB	40	AB	45	UP	44	UP	44	UP	48	---	---	---	---	---	---
19	823	2+	UP	50	UP	52	UP	57	DN	50	DN	51	DN	55	---	---	---	---	---	---
20	885	2+	UP	26	UP	27	UP	30	AB	25	AB	27	AB	29	DN	30	DN	30	AB	30
21	821	2+	PR	28	UP	28	UP	33	DN	25	DN	28	DN	28	---	---	---	---	---	---
22	818	2+	AB	25	AB	25	AB	25	PR	25	DN	27	DN	27	---	---	---	---	---	---
23	1081	2+	AB	20	AB	20	AB	20	UP	20	UP	24	UP	26	UP	24	UP	22	UP	20
24	1052	2+	UP	27	UP	27	UP	27	DN	25	DN	28	DN	25	---	---	---	---	---	---

DN: down regulated UP: upregulated PR: no change ---: not checked
RT: Retention time

表6 シグナル強度に差が見られた ICAT ペアとその普遍性

ID	m/z-L	m/z-H	Mass Diff	APAP 800 mg/kg												Control											
				11 Int	RT	14 Int	RT	12 Int	RT	13 Int	RT	2 Int	RT	3 Int	RT	5 Int	RT	1 Int	RT	4 Int	RT						
12-1	1253.71	1255.74	2.03	nc		LL w 35.7-38	HH w 35-38	nc		LL w 33.9-34.1	nps	LL w 33.9-34.1	nps	LL w 36.3-36.5	HH w 36.3-36.5	nc		HH w 37.4-38	HH w 37.4-38								
12-2	1186.89	1191.52	4.53	nc	w 47.4-47.7	LL w 44.5-45	HH w 44.3-44.8	nc		LL w 42.3-43.4	nps	LL w 42.3-43.4	nps	LL w 44.6-44.9	HH w 44.6-44.9	nc		HH w 46.3-46.7	HH w 46.3-46.7								
12-3	Ratio is not high																										
12-4	1075.42	1081.11	5.69																								
12-5	884.98	897.97	3.01	L	w 41.3-41.8	LL w 38.3-38.8	HH w 38-40.2	H	w 40.4-41.2	LL w 37.4-38.5	L	w 36.3-38.8	L	w 36.7-40.8	HH w 38.8-41	HH	w 41-42	HH w 41-42									
12-6	1003.00	1004.80	1.80	HH	w 37.9-38.9	LL w 35.6-36.4	HH w 35.3-38	HH	w 38.8-37.1	LL w 33.9-34.55	LL	w 35.3-38	LL	w 35.8-36.8	HH w 35.8-36.6	HH	w 37.2-38.1	HH w 37.2-38.1									
12-7	888.82	891.33	4.51	np		LL w 72.2-72.9	L	w 72.7-72.2	L	w 70.9-71.5	np	LL w 72.2-72.9	L	w 72.5-73.1	L	w 73-73.6	L	w 73-73.5									
12-8	841.39	845.91	4.52	LL	w 41.6-42.4	LL w 38.8-39.5	LL	w 38.9-41	LL	w 40-41.2	LL	w 37.3-38	LL	w 38.8-39.3	LL	w 39.6-40.15	LL	w 40.7-41.8									
12-9	843.98	847.90	3.02	nc		HH w 23.3-24.5	HH	w 26-23	HH	w 22.7-23.1	nc	HH w 23.3-24.5	HH	w 20.6-21.2	nc	m 21.9-22.4	nc	w 26.9-27.3									
12-10	733.89	738.98	3.01	L	w 51-51.5	LL w 48.8-49.5	HH w 48.8-49	HH	w 48.4-49.3	nc		LL w 48.8-49.5	HH	w 48.8-49.3	L	w 48.5-49.1	nc	w 48.7-49.2									
12-11	Not consistent																										
12-12	658.78	664.30	4.52	nc			HH		HH		L							HH									
12-13	530-531	Not consistent																									
12-14	497.53	500.54	3.01	LL	Not consistent	L		LL																			
12-15	491-492	Not consistent																									
12-16	Not consistent																										
12-17	448.70	453.22	4.52	Not consistent																							
12-18	Not real peptide peak																										
12-19	Not real peptide peak																										
12-20	588.78	593.36	4.52	np		L	w 44.8-45.5	HH	w 44.7-45.4	HH	w 41.5-42	H	w 43.3-44.2	H	w 45-46.3	nc	w 48.3-47.1	L	w 46.5-47								
13-1	383.85	388.36	3.01	L	w 34-35	LL w 31.9-32.8	HH	w 23-23.8	HH	w 23.3-24.2	H	w 18.9-20.5	H	w 21.1-22.3	nc	w 21.3-22.3	L	w 22.8-23.7	LL	w 27.3-28.4							
13-2	Not real pep																										
13-3	nc																										
13-4	Not consistent																										
13-5	456.70	461.22	4.52	Not consistent		LL					H		nps		nps		LL										
13-6	456.89	459.90	3.01	Not consistent		LL																					
13-7	Not consistent																										
13-8	Not consistent																										
13-9	Not consistent																										
13-10	506.85	512.91	3.01	L	w 38.5-39.8	LL w 36.9-38	HH	w 36.1-38.8	HH	w 48.3-41	H	w 37.5-39.2	L	w 38.8-39.5	nc	w 38.8-40.3	L	w 38.3-40.4	HH	w 41-41.8							
13-11	523.89	528.90	3.01	nc		LL		HH		HH		nc															
13-12	Not consistent																										

*Int peak intensity of higher one a 100+
m 80-100
w <50

Heavy peak is
HH Very high
H High
nc no change
L Low
LL Very Low

np Not observed
nps Not observed because of no spray
nA Not seen in Analyst

*Sample-label
Light
Heavy

表 7 候補ペプチドの同定結果

peptide ID	m/z-L	Identified	Possible candidates
12-5	684.96	Rat Serum Albumin	
12-10	725.99	<i>not identified</i>	
12-20	598.78	Major Urinary Protein Precursor	
13-1	393.85		<i>Retinoic acid receptor RXR-beta</i> <i>Receptor-interacting serine/threonine-protein kinase 5</i>
13-10	509.9		<i>E3 ubiquitin-protein ligase rifylin</i>

図表

VII 細胞レベルでのメタボロミクス技術の開発

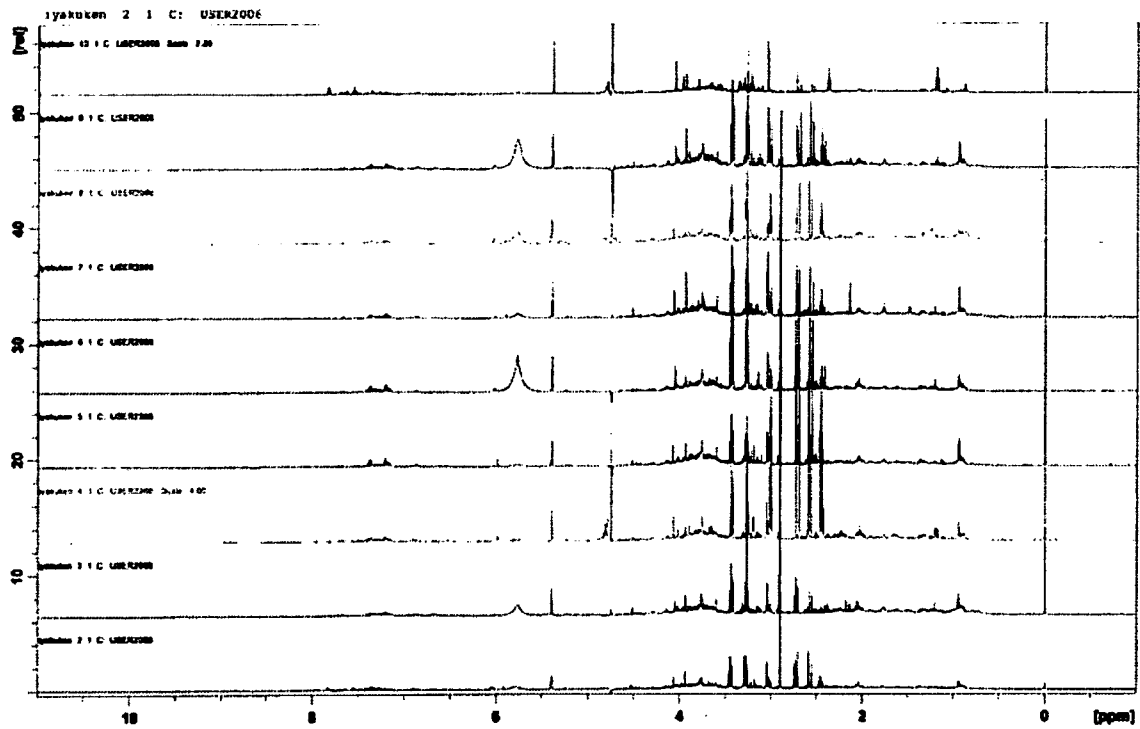


図1 病態モデルマウス尿試料のNMR

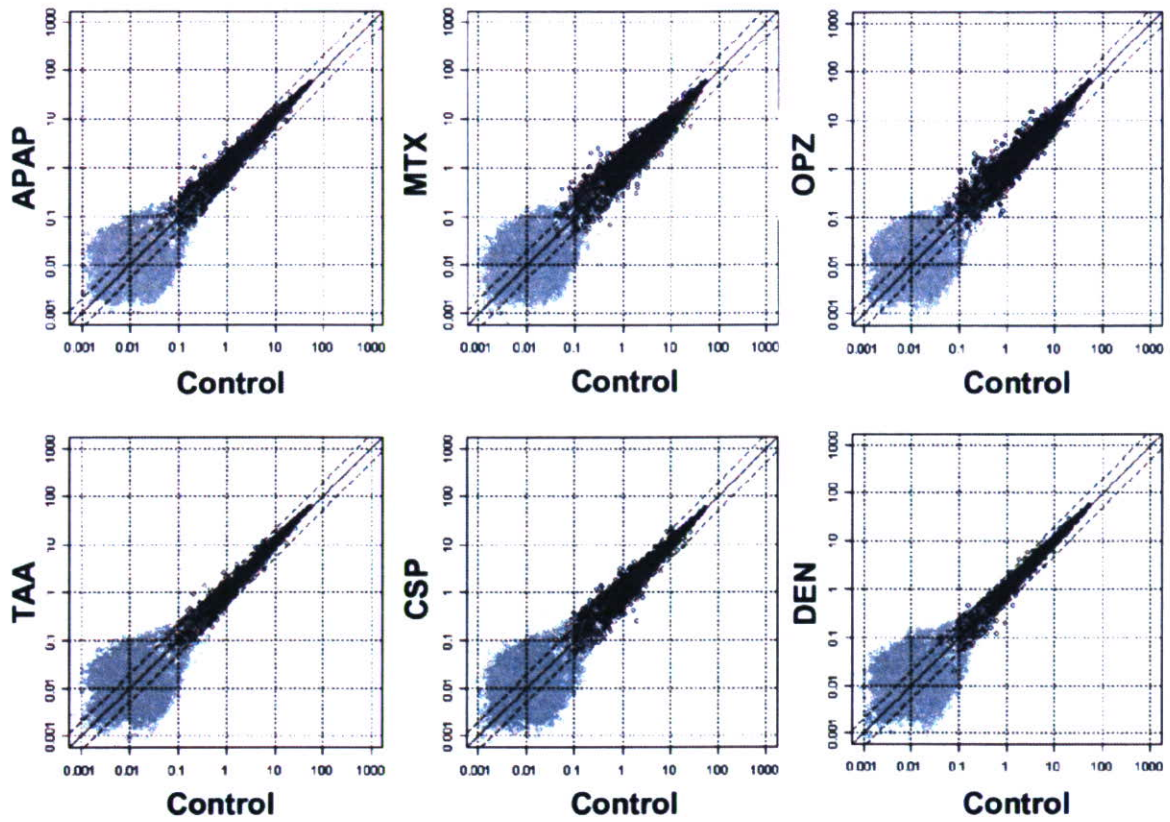


図2 遺伝子発現解析結果（薬剤処理とControlのスカッタープロット）

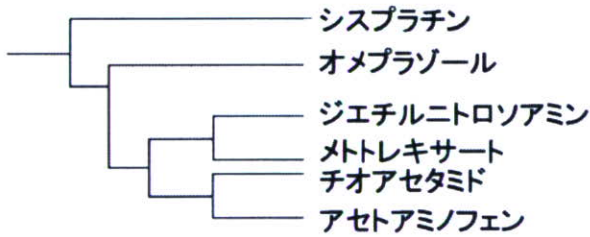


図3 薬剤処理による変動遺伝子を用いたクラスター解析

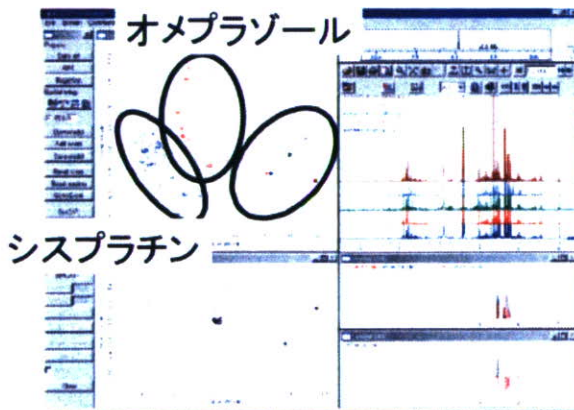


図4 薬剤処理細胞破碎液のPCA解析結果

研究成果の刊行に関する一覧表

雑誌

発表者氏名	論文タイトル名	発表誌名	巻号	ページ	出版年
I. Nakanishi, C. Nishizawa, K. Ohkubo, K. Takeshita, K. Suzuki, T. Ozawa, S. M. Hecht, M. Tanno, S. Sueyoshi, N. Miyata, <u>H. Okuda</u> , S. Fukuzumi, N. Ikota, K. Fukuhara	Hydroxyl Radical Generation via Photoreduction of a Simple Pyridine <i>N</i> -Oxide by an NADH Analogue	<i>Org. Biomol. Chem.</i>	3	3263 – 3265	2005
I. Nakanishi, T. Kawashima, K. Ohkubo, H. Kanazawa, K. Inami, M. Mochizuki, K. Fukuhara, <u>H. Okuda</u> , T. Ozawa, S. Itoh, S. Fukuzumi, and N. Ikota	Electron-Transfer Mechanism in Radical-Scavenging Reactions by a Vitamin E Model in a Protic Medium	<i>Org. Biomol. Chem.</i>	3	626 – 629	2005
Takayoshi Suzuki, Osamu Nagae, Yuka Kato, Hidehiko Nakagawa, Kiyoshi Fukuhara, <u>Naoki Miyata</u> ,	Photo-induced Nitric Oxide Release from Nitrobenzene Derivatives	<i>J. Am. Chem. Soc.</i>	127 (33)	11720-11726	2005
Takayoshi Suzuki, Azusa Matsuura, Akiyasu Kouketsu, Sinya Hisakawa, Hidehiko Nakagawa, <u>Naoki Miyata</u>	Design and synthesis of non-hydroxamate histone deacetylase inhibitors; identification of a selective histone acetylating agent	<i>Bioorganic & Medicinal Chemistry</i>	13	4332-4342	2005
Takayoshi Suzuki, Yuki Nagano, Akiyasu Kouketsu, Azusa Matsuura, Sakiko Maruyama, Mineko Kurotaki, Hidehiko Nakagawa, <u>Naoki Miyata</u>	Novel inhibitors of human histone deacetylases: design, synthesis, enzyme inhibition, and cancer cell growth inhibition of SAHA-based non-hydroxamates	<i>J. Med. Chem.</i>	48(4)	1019-1032	2005
Uhinya Usui, Takayoshi Suzuki, Yoshifumi Hattori, Kazuma Etoh, Hiroki Fujieda, Makoto Nishizuka, Masayoshi Imagawa, Hidehiko Nakagawa, Kohfuku Kohda, <u>Naoki Miyata</u>	Design, synthesis and biological activity of novel PPAR γ ligands based on rosiglitazone and 15d-PGJ2	<i>Bioorganic & Medicinal Chemistry Letters</i>	15	1547-1551	2005
Takayoshi Suzuki, Azusa Matsuura, Akiyoshi Kouketsu, Hidehiko Nakagawa, <u>Naoki Miyata</u> ,	Identification of a potent non-hydroxamate histone deacetylase inhibitor by mechanism- based drug design	<i>Bioorganic & Medicinal Chemistry Letters</i>	15	331-335	2005

Takayoshi Suzuki, Hidehiko Nakagawa, <u>Naoki Miyata</u>	Molecularly targeted approach to cancer therapy: design, synthesis and biological activity of non-hydroxamate histone deacetylase inhibitors	<i>J. of Syn. Org. Chem</i>	63(10)	1004-1015	2005
T. Suzuki, <u>N. Miyata</u>	Non-hydroxamate Histone Deacetylase Inhibitors	<i>Curr. Med. Chem.</i>	12	2867-2880	2005
Yamada K, <u>Suzuki T</u> , <u>Kohara A</u> , Kato TA, Hayashi M, Mizutani T, Saeki K.	Nitrogen-substitution effect on in vivo mutagenicity of chrysene	<i>Mutat Res.</i>	586	1-17	2005
Kanayasu-Toyoda T, Fujino T, Oshizawa T, <u>Suzuki T</u> , Nishimaki-Mogami T, Sato Y, Sawada J, Inoue K, Shudo K, Ohno Y, Yamaguchi T	HX531, a retinoid X receptor antagonist, inhibited the 9-cis retinoic acid-induced binding with steroid receptor coactivator-1 as detected by surface plasmon resonance	<i>J Steroid Biochem Mol Biol</i>	94	303-309	2005
Kawakami T, Hoshida Y, Karai F, Tanaka Y, Tateishi K, Ikenoue T, Obi S, Sato S, Teratani T, Shiina S, Kawabe T, <u>Suzuki T</u> , Hatano N, Taniguchi H, Omata M.	Proteomic analysis of sera from hepatocellular carcinoma patients after radiofrequency ablation treatment	<i>Proteomics</i>	16	4287-4295	2005
K. Fukuhara, M. Nagakawa, I. Nakanishi, K. Ohkubo, K. Imai, S. Urano, S. Fukuzumi, T. Ozawa, N. Ikota, M. Mochizuki, <u>N. Miyata</u> , <u>H. Okuda</u>	Structural Basis for DNA Cleaving-Activity of Resveratrol on the Presence of Cu(II)	<i>Bioorg. Med. Chem.</i>	14	1437-1443	2006
W. Hakamata, E. Yamamoto, M. Muroi, M. Mochizuki, M. Kurihara, <u>H. Okuda</u> , K. Fukuhara	Design and synthesis of α -glucosidase inhibitor having DNA cleaving activity	<i>J. Appl. Glycosci.</i>	53	255-260	2006
W. Hakamata, I. Nakanishi, Y. Masuda, T. Shimizu, H. Higuchi, Y. Nakamura, T. Oku, S. Saito, S. Urano, T. Ozawa, N. Ikota, <u>N. Miyata</u> , <u>H. Okuda</u> , K. Fukuhara	Planar catechin analogues with alkyl side chains, as a potent antioxidant and an α -glucosidase inhibitor	<i>J. Am. Chem. Soc.</i>	128	6524-6525	2006
W. Hakamata, M. Muroi, T. Nishio, T. Oku, A. Takatsuki, H. Osada, K. Fukuhara, <u>H. Okuda</u> , M. Kurihara	N-linked oligosaccharide processing enzymes as molecular targets for drug discovery	<i>J. Appl. Glycosci.</i>	53	149-154	2006

K. Fukuhara, M. Nagakawa, I. Nakanishi, K. Ohkubo, K. Imai, S. Urano, S. Fukuzumi, T. Ozawa, N. Ikota, M. Mochizuki, <u>N. Miyata</u> , <u>H. Okuda</u>	Structural Basis for DNA Cleaving-Activity of Resveratrol on the Presence of Cu(II)	<i>Bioorg. Med. Chem.</i>	14	1437 – 1443	2006.
<u>M. Miyata</u> , Y. Matsuda, H. Tsuchiya, H. Kitada, T. Akase, M. Shimada, K. Nagata, F. J. Gonzalez and Y. Yamazoe	Chenodeoxycholic acid-mediated activation of the farnesoid X receptor negatively regulates hydroxysteroid sulfotransferase	<i>Drug Metab. Pharmacokin</i>	21,	315-323	2006
<u>M. Miyata</u> , H. Watase, W. Hori, M. Shimada, K. Nagata, F. J. Gonzalez, and Y. Yamazoe	Role for enhanced fecal excretion of bile acid in hydroxysteroid sulfotransferase-mediated protection against lithocholic acid-induced liver toxicity	<i>Xenobiotica</i>	36	631-644	2006
Takayoshi Suzuki, <u>Naoki Miyata</u>	Epigenetic control using natural products and synthetic molecules	<i>Curr. Med. Chem.</i>	13	935-58	2006
Takayoshi Suzuki, <u>Naoki Miyata</u>	Rational design of non-hydroxamate histone deacetylase inhibitors	<i>Mini Rev. Med. Chem.</i>	6	515-526	2006
Shinya Usui, Hiroki Fujieda, Takayoshi Suzuki, Naoaki Yoshida, Hidehiko Nakagawa and <u>Naoki Miyata</u>	Identification of novel PPARalpha ligands by the structural modification of a PPARgamma ligand	<i>Bioorganic & Medicinal Chemistry Letters</i>	16	3249-3254	2006
Takayoshi Suzuki, Akiyasu Kouketsu, Yukihiro Itoh, Shinya Hisakawa, Satoko Maeda, Minoru Yoshida, Hidehiko Nakagawa, <u>Naoki Miyata</u>	Highly potent and selective histone deacetylase 6 inhibitors designed based on a small-molecular substrate	<i>J. Med. Chem.</i>	49	4809-4812	2006
Takayoshi Suzuki, Keiko Imai, Hidehiko Nakagawa, <u>Naoki Miyata</u>	2-Anilinobenzamides as SIRT Inhibitors	<i>Chem Med Chem.</i>	1	1059-1062	2006
Hidehiko Nakagawa, Ryo Ohyama, Ayako Kimata, Takayoshi Suzuki, <u>Naoki Miyata</u>	Hydroxyl radical scavenging by edaravone derivatives: Efficient scavenging by 3-methyl-1-(pyridin-2-yl)-5-pyrazolone with an intramolecular base	<i>Bioorg. Med. Chem. Lett.</i>	16	5939-5942	2006

Shizuka Ban, Hidehiko Nakagawa, Takayoshi Suzuki, <u>Naoki Miyata</u>	Novel membrane-localizing TEMPO derivatives for measurement of cellular oxidative stress at the cell membrane	<i>Bioorg. Med. Chem. Lett.</i>	17	1451-1454	2007
Takayoshi Suzuki, Shinya Hisakawa, Yukihiro Itoh, Sakiko Maruyama, Mineko Kurotaki, Hidehiko Nakagawa, <u>Naoki Miyata</u>	Identification of a potent and stable antiproliferative agent by the prodrug formation of a thiolate histone deacetylase inhibitor	<i>Bioorg. Med. Chem. Lett.</i>	17	1558-1561	2007
Shizuka Ban, Hidehiko Nakagawa, Takayoshi Suzuki, <u>Naoki Miyata</u>	Novel mitochondria-localizing TEMPO derivative for measurement of cellular oxidative stress in mitochondria	<i>Bioorg. Med. Chem. Lett.</i>	17	2055-2058	2007
Kanayasu-Toyoda, <u>T.</u> , <u>Suzuki</u> , T., Oshizawa, T., Uchida, E., Hayakawa, T., Yamaguchi, T.	Granulocyte colony-stimulating factor promotes the translocation of protein kinase Ci in neutrophilic differentiation cells	<i>J. Cell Physiol.</i>	211	189-196	2007
Zhan, L., Honma, M., Wang, L., Hayashi, M., Wu, D., Zhang, L., Rajaguru, P., <u>Suzuki</u> , T.	Microcystin-LR is not Mutagenic in vivo in the $\square/lacZ$ Transgenic Mouse (Muta TM Mouse)	<i>Genes and Environment.</i>	28	68-73	2006
Dertinger, S.D., Bishop, M.E., McNamee, J.P., Hayashi, M., <u>Suzuki</u> , T., Asano, N., Nakajima, M., Saito, J., Moore, M., Torous, D.K., Macgregor, J.T.	Flow cytometric analysis of micronuclei in peripheral blood reticulocytes: I. Intra- and interlaboratory comparison with microscopic scoring	<i>Toxicol. Sci.</i>	94	83-91	2006
<u>小原有弘</u> , <u>水澤博</u>	JCRB細胞バンクの事業の概要	分子細胞治療	5	152-156	2006
R. Shimazawa, N. Nagai, S. Toyoshima, <u>H. Okuda</u>	Present sate of new chiral drug development and review in Japan	<i>J. Health Sciences</i>	54	23-29	2008
K. Fukuhara, I. Nakanishi, A. Matsuoka, T. Matsumura, S. Honda, M. Hayashi, T. Ozawa, N. Miyata, S. Saito, N. Ikota, <u>H. Okuda</u>	Effect of methyl substitution on antioxidative property and genotoxicity of resveratrol	<i>Chem. Res. Toxicol.</i>	21	282-287	2008

S. Manda, I. Nakanishi, K. Ohkubo, Y. Uto, T. Kawashima, K. Fukuhara, <u>H. Okuda</u> , H. Hori, T. Ozawa, N. Ikota, S. Fukuzumi, and K. Anzai	Enhanced radical-scavenging activity of naturally-oriented Artepillin C Derivatives	<i>Chem. Commun.</i>		626-628	2008
I. Nakanishi, T. Shimada, K. Ohkubo, T. Shimizu, S. Urano, <u>H. Okuda</u> , <u>N. Miyata</u> , T. Ozawa, K. Anzai, S. Fukuzumi, N. Ikota, K. Fukuhara	Involvement of electron transfer in the radical-scavenging reaction of resveratrol	<i>Chem. Lett.</i>	36	1276-1277	2007
K. Fukuhara, S. Oikawa, N. Hakota, Y. Sakai, Y. Hiraku, T. Shoda, S. Saito, <u>N. Miyata</u> , S. Kawanishi, <u>H. Okuda</u>	9-Nitroanthracene derivative as a precursor of anthraquinone for photodynamic therapy	<i>Bioorg. Med. Chem.</i>	15	3869-3873	2007
M. Nomoto, M. <u>Miyata</u> , M. Shimada, K. Yoshinari, F. J. Gonzalez, S. Shibasaki, T. Kurosawa, Y. Shindo, Y. Yamazoe	ME3738 protects against lithocholic acid-induced hepatotoxicity, associated with enhancement in biliary bile acid and cholesterol output	<i>Eur. J. Pharmacol.</i>	574	192-200	2007
M. Ohbuchi, <u>M. Miyata</u> , D. Daichi, M. Shimada, K. Yoshinari and Y. Yamazoe	Involvement of N-hydroxylation of flutamide metabolites and hepatic GSH depletion in flutamide-induced hepatotoxicity	<i>Drug Metab. Rev.</i>	39, supplement 1	36	2007
矢本 敬、真鍋 淳	医薬品開発における毒性評価のバイオマーカー	ヒューマンサイエンス	19(2)	22-26	2008
Hiroki Tsumoto, Katsumasa Takahashi, Takayoshi Suzuki, Hidehiko Nakagawa, Kohfuku Kohda, <u>Naoki Miyata</u>	Preparation of C60-based active esters and coupling of C60 moiety to amines or alcohols	<i>Bioorg. Med. Chem. Lett.</i>	18	657-660	2008
Yukihiro Itoh, Takayoshi Suzuki, Akiyasu Kouketsu, Nobuaki Suzuki, Satoko Maeda, Minoru Yoshida, Hidehiko Nakagawa, and <u>Naoki Miyata</u>	Design, Synthesis, Structure-Selectivity Relationship, and Effect on Human Cancer Cells of a Novel Series of Histone Deacetylase 6-Selective Inhibitors	<i>J. Med. Chem.</i>	50	5425-5438	2007

Ayako Kimata, Hidehiko Nakagawa, Ryo Ohyama, Tomoko Fukuuchi, Shigeru Ohta, Takayoshi Suzuki, <u>Naoki Miyata</u>	New Series of Antiprion Compounds: Pyrazolone Derivatives Have the Potent Activity of Inhibiting Protease-Resistant Prion Protein Accumulation	<i>J. Med. Chem.</i>	50	5053-5056	2007
Shinya Usui, Hiroki Fujieda, Takayoshi Suzuki, N. Yoshida, Hidehiko Nakagawa, Michitaka Ogura, Makoto Makishima, <u>Naoki Miyata</u>	Synthesis and Evaluation of 2-Nonylaminopyridine Derivatives as PPAR Ligands	<i>Chem. Pharm. Bull.</i>	55	1053-1059	2007
Takayoshi Suzuki, Shinya Hisakawa, Yukihiko Itoh, Nobuaki Suzuki, Katsumasa Takahashi, Masatoshi Kawahata, Kentaro Yamaguchi, Hidehiko Nakagawa, <u>Naoki Miyata</u>	Design, synthesis, and biological activity of folate receptor-targeted prodrugs of thiolate histone deacetylase inhibitors	<i>Bioorg. Med. Chem. Lett.</i>	17	4208-4212	2007
Hiroki Fujieda, Shinya Usui, Takayoshi Suzuki, Hidehiko Nakagawa, Michitaka Ogura, Makoto Makishima, <u>Naoki Miyata</u> ,	Phenylpropanoic acid derivatives bearing a benzothiazole ring as PPARdelta-selective agonists	<i>Bioorg. Med. Chem. Lett.</i>	17	4351-4357	2007
Hidehiko Nakagawa, Nobuko Komai, Mitsuko Takusagawa, Yuri Miura, Tosifusa Toda, <u>Naoki Miyata</u> , Toshihiko Ozawa, Nobuo Ikota	Nitration of specific tyrosine residues of cytochrome C is associated with caspase-cascade inactivation	<i>Biol. Pharm. Bull.</i>	30	15-20	2007
Sanda T, Okamoto T, Uchida Y, Nakagawa H, Iida S, Kayukawa S, Suzuki T, Oshizawa T, Suzuki T, <u>Miyata N</u> , Ueda R	Proteome analyses of the growth inhibitory effects of NCH-51, a novel histone deacetylase inhibitor, on lymphoid malignant cells	<i>Leukemia</i>	21	2344-2453	2007
Hiroki Tsumoto, Chie Murata, <u>Naoki Miyata</u> , Kohfuku Kohda, Ryo Taguchi	Efficient identification and quantification of proteins using isotope-coded 1-(6-methylnicotinoyloxy)succinimides by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry	<i>Rapid Commun. Mass Spectrom.</i>	21	815-824	2007

Linxiang Li, Yoshihiro Abe, Kiyotada Kanagawa, Tomoko Shoji, Tadahiko Mashino, Masataka Mochizuki, Miho Tanaka, <u>Naoki Miyata</u>	Iron-chelating agents never suppress Fenton reaction but participate in quenching spin-trapped radicals	<i>Analytica Chimica Acta</i>	599	315-319	2007
Tomoko Shoji, Linxiang Li, Yoshihiro Abe, Masahiro Ogata, Yoshihisa Ishimoto, Ryoko Gonda, Tadahiko Mashino, Masataka Mochizuki, Michihisa Uemoto, <u>Naoki Miyata</u>	DMPO-OH radical formation from 5,5-dimethyl-1-pyrroline N-oxide (DMPO) in hot water	<i>Analytical Sciences</i>	23	219-221	2007
Watanabe T, Tobe K, Nakachi Y, Kondoh Y, Nakajima M, Hamada S, Namiki C, <u>Suzuki T</u> , Maeda S, Tadakuma A, Sakurai M, Arai Y, Hyogo A, Hoshino M, Tashiro T, Ito H, Inazumi H, Sakaki Y, Tashiro H, Furihata C.	Differential Gene Expression Induced by Two Genotoxic N-nitroso Carcinogens, Phenobarbital and Ethanol in Mouse Liver Examined with Oligonucleotide Microarray and Quantitative Real-time PCR.	<i>Genes and Environment</i>	29	115-127	2007
Luan Y, <u>Suzuki T</u> , Palanisamy R, Takashima Y, Sakamoto H, Sakuraba M, Koizumi T, Saito M, Matsufuji H, Yamagata K, Yamaguchi T, Hayashi M, Honma M.	Potassium bromate treatment predominantly causes large deletions, but not GC>TA transversion in human cells.	<i>Mutat Res.</i>	619	113-123	2007
Kanayasu-Toyoda T, Ishii-Watabe A, <u>Suzuki T</u> , Oshizawa T, Yamaguchi T.	A new role of thrombopoietin enhancing ex vivo expansion of endothelial precursor cells derived from AC133-positive cells.	<i>J Biol Chem.</i>	282	33507-33514	2007
Sanda T, Okamoto T, Uchida Y, Nakagawa H, Iida S, Kayukawa S, Suzuki T, Oshizawa T, <u>Suzuki T</u> , Miyata N, Ueda R.	Proteome analyses of the growth inhibitory effects of NCH-51, a novel histone deacetylase inhibitor, on lymphoid malignant cells	<i>Leukemia</i>	21	2344-2353	2007
Kanayasu-Toyoda, T., <u>Suzuki, T.</u> , Oshizawa, T., Uchida, E., Hayakawa, T., Yamaguchi, T.	Granulocyte colony-stimulating factor promotes the translocation of protein kinase Ci in neutrophilic differentiation cells	<i>J. Cell Physiol.</i>	211	189-196	2007
小原有弘, 水澤 博	JCRB 細胞バンク:厚生労働省	細胞工学	26(10)	1177-1178	2007
水澤 博, <u>小原有弘</u> , 増井徹、	我国におけるヒト研究資源の現状と将来	医学のあゆみ	222(2)	113	2007

Takeuchi M, Takeuchi K, Kohara A, Satoh M, Shioda S, Ozawa Y, et al.	Chromosomal instability in human mesenchymal stem cells immortalized with human papilloma virus E6, E7, and hTERT genes	<i>In Vitro Cell Dev. Biol. Anim.</i>	43	129-138	2007
Ono K., Satoh M., Yoshida T., Ozawa Y., Kohara A., Takeuchi M., Mizusawa H. Sawada H.	Species identification of animal cells by nested PCR targeted to mitochondrial DNA	<i>In Vitro Cell. Dev. Biol. Anim.</i>	43	168-175	2007
小原有弘、大谷梓、小澤裕、塩田節子、増井徹、水澤博.	培養細胞研究資源のマイコプラズマ汚染調査	<i>Tiss. Cult. Res. Commun.</i>	26	159-163	2007
水澤博、増井徹、竹内昌男、小原有弘	-190C気相式液体窒素保存システム、	<i>Tiss. Cult. Res. Commun.</i>	26	155-170	2007
水澤博、小澤裕、小原有弘、増井徹、佐藤元信、岩瀬秀、深海薫、西條薫、中村幸夫	培養細胞で頻発するクロソコントミネーションへの警戒	実験医学	26	561-567	2008

Hydroxyl radical generation *via* photoreduction of a simple pyridine *N*-oxide by an NADH analogue†

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Received 5th July 2005, Accepted 22nd July 2005

First published as an Advance Article on the web 2nd August 2005

Photoreduction of pyridine *N*-oxide, which has a key structure of antitumor agents for hypoxic solid tumors, by 1-benzyl-1,4-dihydropyridinamide in deaerated aprotic media resulted in generation of hydroxyl radical, leading to the oxidation of salicylic acid to 2,3- and 2,5-dihydroxybenzoic acids, and catechol.

Recently, considerable effort has been made to develop effective drugs against solid tumors, which exist under hypoxic (oxygen-poor) conditions in inefficient vascular systems.^{1,2} Tirapazamine (3-amino-1,2,4-benzotriazine 1,4-di-*N*-oxide), which has a heterocyclic *N*-oxide structure, is a clinically promising antitumor agent against hypoxic cells.³ The DNA damage induced by tirapazamine is proposed to result from the generation of hydroxyl radical ([•]OH) or the direct oxidation of the deoxyribose backbone of DNA after one-electron reduction of the *N*-oxide to form an activated intermediate.⁴⁻⁶ However, the actual DNA-damaging species has yet to be clarified. On the other hand, photosensitizers available for photodynamic therapy are advantageous to localize the toxicity to a selected site (tumor cells), thus avoiding toxicity to normal cells.⁷ However, since most photosensitizers require O₂ to produce reactive oxygen species, they are not effective toward anaerobic solid tumors. Thus, photoactivated compounds, which generate reactive oxygen species under anaerobic conditions, are certainly required for the development of drugs effective against solid tumors without affecting normal cells.

We report herein [•]OH generation from a simple unsubstituted pyridine *N*-oxide (PyO), which has a largely negative reduction potential, *via* one-electron reduction by photoexcited 1-benzyl-1,4-dihydropyridinamide (BNAH) used as a model compound

of dihydropyridinamide adenine dinucleotide (NADH) in DMF under anaerobic conditions. The effects of the substituent at the C-4 position of pyridine *N*-oxides on the mechanism of photoinduced electron transfer from BNAH to pyridine *N*-oxides as well as the reactivity of the corresponding radical anions are clarified based on the spectral and electrochemical data together with the calculated molecular structures by the density functional method, providing a valuable insight into the development of antitumor agents for hypoxic cells.

Salicylic acid (SA) was employed to detect [•]OH generated in the photoreaction of pyridine *N*-oxides with BNAH in deaerated DMF. SA reacts with [•]OH to form 2,3-dihydroxybenzoic acid (2,3-DHBA) and 2,5-dihydroxybenzoic acid (2,5-DHBA) as major products and catechol as a minor product.⁸⁻¹³ These oxidized products of SA are stable and are readily isolated and quantified by a reverse-phase HPLC equipped with an electrochemical detector (HPLC-ECD). SA does not react with O₂^{•-} at an appreciable rate as compared to [•]OH. Although SA also reacts with singlet oxygen (¹O₂), only 2,5-DHBA is formed exclusively, instead of the mixture of 2,3-DHBA, 2,5-DHBA, and catechol.¹⁴

Aprotic solvents, such as DMF and acetonitrile (MeCN) were used because of the poor solubility of pyridine *N*-oxides toward water, although the reactivity in aqueous media is important for an *in vivo* situation. When a deaerated DMF solution of PyO (5.0 × 10⁻³ mol dm⁻³) and BNAH (5.0 × 10⁻³ mol dm⁻³) was irradiated with UV light (λ > 290 nm) in the presence of SA (3.2 × 10⁻² mol dm⁻³), 2,3-DHBA, 2,5-DHBA, and catechol were detected by the HPLC-EC analysis as shown in Fig. 1a. The ratio of the yields of 2,5-DHBA, 2,3-DHBA, and catechol are 48:35:16. Irradiation of PyO or BNAH alone in the presence of SA resulted in no formation of oxidized products of SA (Fig. 1c and d). Under dark conditions, neither DHBA product nor catechol was formed even in the presence of all the components, *i.e.*, PyO, BNAH, and SA (Fig. 1e). These results demonstrate that [•]OH is generated in the photoreaction of PyO with BNAH in deaerated DMF as shown in Scheme 1. In fact, addition of

† Electronic supplementary information (ESI) available: Transient absorption spectra of the photoreaction between BNAH and PyO (S1), EPR spectrum of NO₂PyO^{•-} (S2), spectral change in the photoreaction between BNAH and NO₂PyO (S3), and DFT minimized structures (S4). See <http://dx.doi.org/10.1039/b509447j>

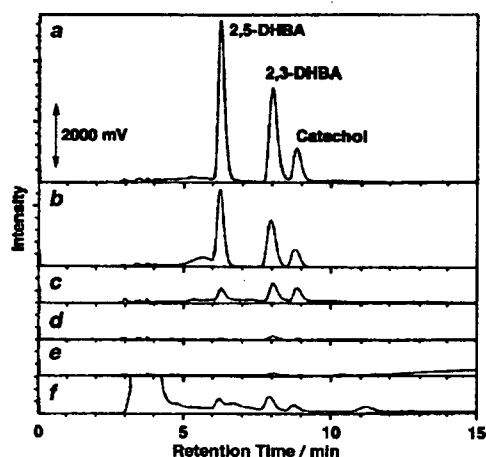
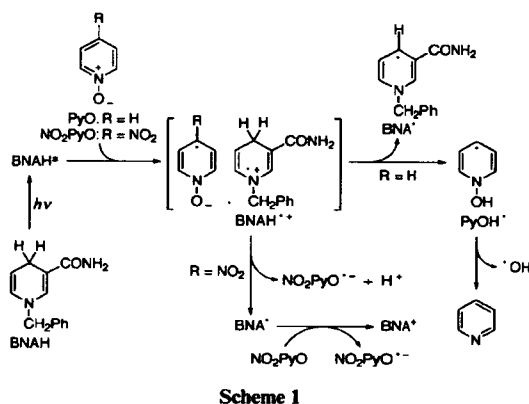


Fig. 1 HPLC-ECD chromatograms of products formed during irradiation (1 h) of (a) PyO (5.0×10^{-3} mol dm $^{-3}$), BNAH (5.0×10^{-3} mol dm $^{-3}$), and SA (3.2×10^{-2} mol dm $^{-3}$); (b) PyO (5.0×10^{-3} mol dm $^{-3}$), BNAH (5.0×10^{-3} mol dm $^{-3}$), SA (3.2×10^{-2} mol dm $^{-3}$), and ethanol (5.7×10^{-1} mol dm $^{-3}$); (c) PyO (5.0×10^{-3} mol dm $^{-3}$) and SA (3.2×10^{-2} mol dm $^{-3}$); (d) BNAH (5.0×10^{-3} mol dm $^{-3}$) and SA (3.2×10^{-2} mol dm $^{-3}$); (e) PyO (5.0×10^{-3} mol dm $^{-3}$), BNAH (5.0×10^{-3} mol dm $^{-3}$), and SA (3.2×10^{-2} mol dm $^{-3}$) without irradiation; (f) NO $_2$ PyO (5.0×10^{-3} mol dm $^{-3}$), BNAH (5.0×10^{-3} mol dm $^{-3}$), and SA (3.2×10^{-2} mol dm $^{-3}$) in deaerated DMF at 298 K.



ethanol, which is a $\cdot\text{OH}$ scavenger, resulted in a decrease in the yield of three oxidized SA products (Fig. 1b).

Photoirradiation of BNAH is known to give the singlet excited state, BNAH*.¹⁵ The fluorescence of BNAH* is quenched efficiently by the addition of PyO in deaerated MeCN. The quenching rate constant is determined as 1.4×10^{10} mol $^{-1}$ dm 3 s $^{-1}$, which is close to the diffusion limited value in MeCN. Nanosecond laser excitation of a deaerated MeCN solution of PyO and BNAH results in the formation of PyO \cdot^- ($\lambda_{\text{max}} = 540$ nm) and BNAH** ($\lambda_{\text{max}} = 380$ nm)¹⁶ (see Electronic Supplementary Information, Fig. S1†).

The reaction mechanism of electron-transfer reduction of PyO by BNAH is shown in Scheme 1. Photoinduced electron transfer from BNAH* to PyO takes place to produce a radical ion pair of BNAH** and PyO \cdot^- , since the oxidation potential of BNAH* ($E^{\circ}_{\text{ox}} = -2.65$ V vs. SCE) determined in DMF¹⁷ is more negative than the reported reduction potential of PyO ($E^{\circ}_{\text{red}} = -2.30$ V) in DMF.¹⁸ Proton transfer from BNAH**, thus produced, to PyO \cdot^- gives PyOH \cdot and BNA \cdot . The resulting PyOH \cdot then undergoes the N–O bond cleavage to produce $\cdot\text{OH}$ and pyridine. Judging from the more positive oxidation potential of BNA \cdot ($E^{\circ}_{\text{ox}} = -1.05$ V)¹⁷ in DMF than the reduction potential of PyO ($E^{\circ}_{\text{red}} = -2.30$ V),¹⁸ no electron transfer from BNA \cdot to PyO would occur and instead a coupling between two molecules of BNA \cdot may occur to give a dimer (BNA) $_2$.

When PyO is replaced by 4-nitropyridine *N*-oxide (NO $_2$ PyO), neither DHBA products nor catechol was produced after irradiation of a deaerated DMF solution containing NO $_2$ PyO, BNAH, and SA (Fig. 1f).

This result demonstrates that the radical anion of NO $_2$ PyO (NO $_2$ PyO \cdot^-) generated in photoinduced electron transfer from BNAH to NO $_2$ PyO is relatively stable and does not undergo the N–O bond cleavage. In fact, an EPR spectrum of NO $_2$ PyO \cdot^- , which has a *g* value of 2.0054, was observed after irradiation of a deaerated MeCN solution of NO $_2$ PyO and BNAH (Fig. S2a†). The hyperfine coupling constants (hfc) of the observed EPR spectrum of NO $_2$ PyO \cdot^- were determined by comparison of the observed spectrum with the computer-simulated spectrum (Fig. S2b†) and they were assigned based on the reported hfc values for NO $_2$ PyO \cdot^- in DMF (Fig. S2†).¹⁸

The UV-vis spectral titration (Fig. S3†) indicates that BNAH acts as a two-electron donor to reduce 2 equiv. of NO $_2$ PyO to NO $_2$ PyO \cdot^- . Since the E°_{ox} value of BNAH* (-2.65 V) is more negative than the E°_{red} value of NO $_2$ PyO (-0.77 V),¹⁸ photoinduced electron transfer from BNAH* to NO $_2$ PyO occurs to give a radical ion pair of NO $_2$ PyO \cdot^- and BNAH**. BNAH** undergoes deprotonation to give BNA \cdot . The delocalization of an electron on the NO $_2$ PyO \cdot^- molecule due to the electron-withdrawing NO $_2$ group may preclude the protonation of NO $_2$ PyO \cdot^- . In fact, no hyperfine structure due to the N–OH proton was observed in the EPR spectrum of NO $_2$ PyO \cdot^- . The subsequent electron transfer from BNA \cdot to NO $_2$ PyO may also occur rapidly, judging from the E°_{ox} value of BNA \cdot (-1.05 V), which is lower than the E°_{red} value of NO $_2$ PyO (-0.77 V).¹⁸ Thus, once photoinduced electron transfer from BNAH to NO $_2$ PyO occurs, 2 equiv. of NO $_2$ PyO \cdot^- are produced.

DFT calculations using B3LYP/6-31G* basis set for PyOH \cdot and NO $_2$ PyOH \cdot were carried out to investigate the difference in the reactivity of radical species of pyridine *N*-oxides depending on a substituent at the C-4 position (Fig. S4†). The calculated N–O bond length in PyOH \cdot (1.50 Å) is significantly longer than that in NO $_2$ PyOH \cdot (1.40 Å). This suggests that the N–O bond cleavage in PyOH \cdot may occur much easier than that in NO $_2$ PyOH \cdot . Furthermore, PyOH \cdot is significantly bent as indicated by the out-of-plane N–O bond bending angle (α) of 152°, whereas NO $_2$ PyOH \cdot is relatively flat ($\alpha = 169^\circ$). Similar results have been reported for the N–O bond fragmentation in *N*-methoxy-substituted aromatic compounds.¹⁹ The N–O bond cleavage requires mixing of π^* and σ^* orbitals, which is achieved by bending the N–O bond out of the plane of the aromatic ring.

In conclusion, photoreduction of a simple pyridine *N*-oxide by BNAH in deaerated aprotic medium resulted in generation of $\cdot\text{OH}$, which can oxidize SA to 2,3-DHBA, 2,5-DHBA, and catechol. The electron-withdrawing group such as NO $_2$ on the aromatic ring can significantly stabilize the radical anion of pyridine *N*-oxide, precluding the subsequent $\cdot\text{OH}$ release. We are currently investigating the detailed effects of various substituents on the photoreactivities of pyridine *N*-oxides in the presence of various reducing agents.

Acknowledgements

This work was partially supported by Grant-in-Aids for Scientific Research (A) (No. 16205020) and for Young Scientist (B) (No. 17790044) from the Ministry of Education, Culture, Sports, Science and Technology, Japan and by the Budget for Nuclear Research of the Ministry of Education, Culture, Sports, Science and Technology, Japan based on the screening and counselling Atomic Energy Commission.

Notes and references

- J. M. Brown, *Cancer Res.*, 1999, **59**, 5863.
- S. Kizaka-Kondoh, M. Inoue, H. Harada and M. Hiraoka, *Cancer Sci.*, 2003, **94**, 1021.