

Fig. 3. Expanded HOXA1 reduces PBX1-coupled transcriptional activity. Luciferase assay from the different cell lines. Cells were cotransfected with plasmids containing PBX1, HOXA1, and the enhancer b1-ARE fused to a luciferase reporter gene (Di Rocco et al., 2001). EGFP vector was used as an internal control. **A:** COS-7. **B:** Neuroblastoma cell line SK-N-SH. **C:** Embryonic carcinoma cell line P19 grown in serum-free medium. Bars represent the mean \pm SE of at least three independent experiments. * $P < 0.05$, ** $P < 0.01$.

Transcriptional Activities of Polyhistidine Variants of HOXA1 Coupled With PBX1

We next examined the physiological relevance of polyhistidine variants of HOXA1 protein in neuronal

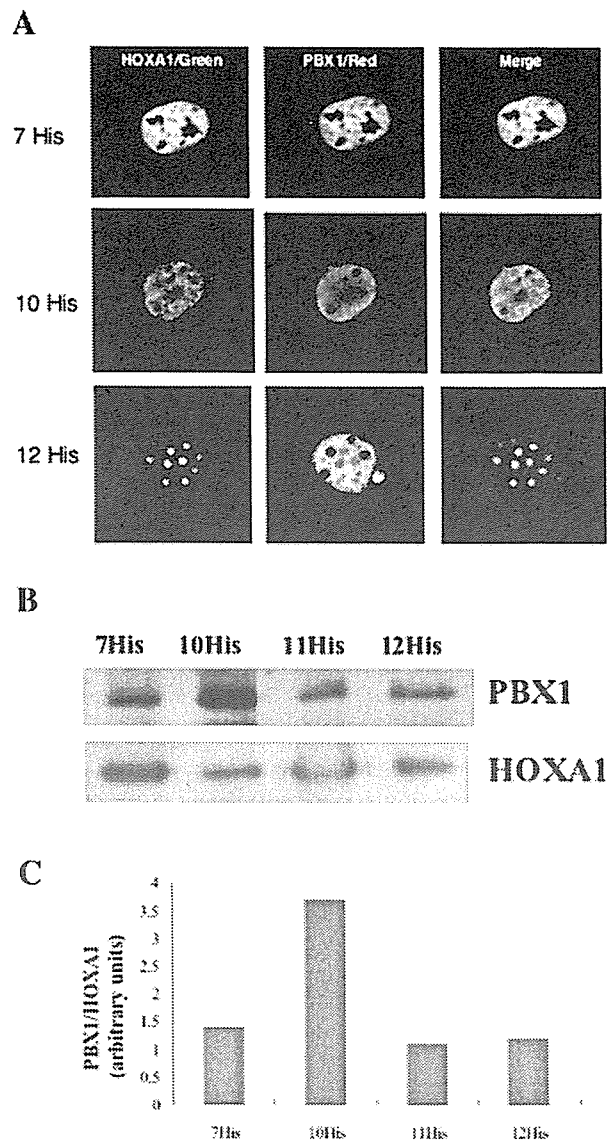


Fig. 4. HOXA1 and PBX1 interaction is impaired in expanded variant of HOXA1. **A:** Immunofluorescence images of COS-7 cotransfected with PBX1, an enhancer bi-ARE, and 7-His, 10-His, or 12-His repeat variants of HOXA1-EGFP. Fluorescence images were taken 24 hr posttransfection. Red signals: anti-PBX1. Note the absence of PBX1 signal in protein aggregates of the 12-His variant. **B:** Immunoprecipitation with anti-GFP and Western blotted with anti-PBX1 and anti-HOXA1. PBX1 protein shows decreased levels in both expanded and deleted variants of HOXA1-EGFP transfected to COS-7 cells. **C:** Quantification of bound PBX1 protein levels per HOXA1 protein from Western blot analysis. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

cells. PBX1 plays a major role in cooperative transcriptional activation with HOXA1 through an autoregulatory element, the b1-ARE (Di Rocco et al., 2001). To investigate the cooperative transcription of PBX1 with

the different variants of HOXA1 through b1-ARE, we simultaneously expressed HOXA1, PBX1, EGFP (an internal control), and b1-ARE luciferase reporter constructs in COS7, SK-N-SH, and P19 cells. Luciferase assay were performed 24 hr after transfection. The P19 embryonic cell line was grown under serum-free conditions. Growth in serum-free media itself committed EC cells to neural differentiation (Darmon et al., 1981) and addition of retinoic acid (RA) intensified this effect (Tanaka et al., 1992). However, nonneural cell types arise after treatment with RA under serum-containing conditions. Differentiation in serum-free media alone is accompanied by expression of only neuroectodermal/neural mRNAs, but treatment with RA invariably induces the cells to express both neuroectodermal/neural and endodermal mRNAs (Pachernik et al., 2005), so the serum-free medium condition was used in this experiment.

The transcriptional activities of expanded variants were significantly reduced in all cell lines, whereas the activities of deleted variant with 7-polyhistidine were enhanced in P19 cells compared with the activity of 10-polyhistidine variant (Fig. 3A–C). Immunofluorescence images revealed the inability of aggregated forms of expanded HOXA1 protein to bind efficiently with PBX1 protein (Fig. 4A). Inefficient interaction between expanded HOXA1 and PBX1 is clearly indicated by low levels of PBX1 protein in immunoprecipitation assay with anti-GFP (Fig. 4B,C).

Expanded and Deleted Polyhistidine Variants in HOXA1 Inhibit Neuronal Differentiation

Because HOXA1 is one of the neurodevelopmental genes, we examined whether polyhistidine variants had any effect on the process of neuronal differentiation. To accomplish this, RA-induced neuronal differentiation was performed in SK-N-SH cells. Phase microscopy shows the effect of RA treatment on the neurites of these cells (Fig. 5A). Immunocytochemistry with antibody against the neuron-specific protein MAP2 revealed neurite outgrowth in cells expressing the 10-histidine

HOXA1-GFP, 42 hr after transfection (Fig. 5B). In contrast, a greater number of cells expressing expanded and deleted forms of HOXA1-GFP failed to initiate neurite outgrowth. Total levels of MAP2 protein in cells overexpressing expanded and deleted variants were also significantly reduced (Fig. 5C,D). The appearance of

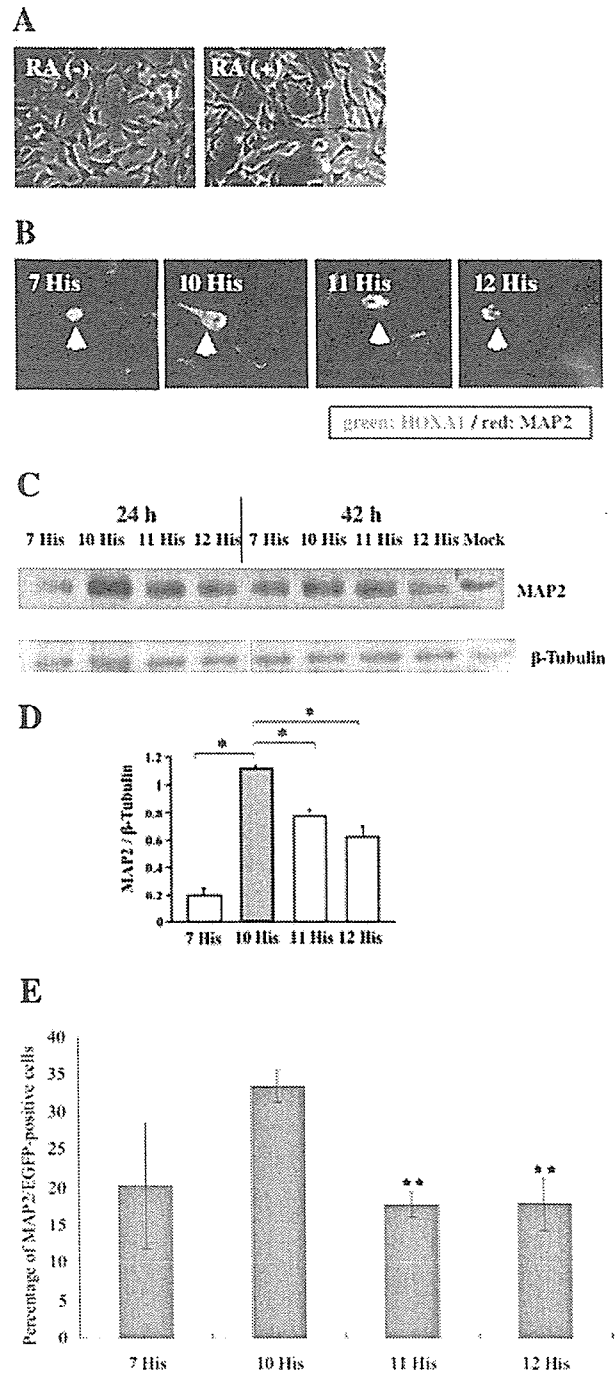


Fig. 5. Expanded and deleted polyhistidine repeats in HOXA1 inhibit neuronal differentiation. **A**: Retinoic acid (RA)-induced neuronal differentiation in SK-N-SH cells. Phase images of SK-N-SH cells treated with (right) or without (left) 10 μ M RA for 42 hr. **B**: HOXA1-EGFP-transfected SK-N-SH treated with 10 μ M RA for 42 hr. Immunofluorescence images were taken 42 hr after transfection. Expanded 11- and 12-His repeat variants as well as deleted 7-His repeat variants inhibit neuronal differentiation. Note the neurite outgrowth immunostained with MAP2 (red signal). **C**: Western blot analysis shows decreased MAP2 expression levels in the expanded and deleted forms. **D**: Quantification of MAP2 expression levels normalized with β -tubulin. **E**: Quantification of the percentage of EGFP-MAP2-double-positive cells. Error bars represent SEM; $n = 3$. * $P < 0.05$, ** $P < 0.01$. P values from a paired t -test in all experiments. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com.]

differentiated cells was also examined by scoring MAP2-positive transformants expressing the HOXA1-EGFP variants. EGFP-MAP2 double-positive cells quantification also confirmed that there is a significant decrease in the percentage of MAP2 levels particularly in the expanded variants.

DISCUSSION

In this current study, we have shown that polyhistidine repeat tract variants compromised HOXA1 function in transcription and neuronal differentiation. Moreover, we have presented evidence that cell death resulting from polyhistidine repeat expansion is mediated primarily by autophagy and not by a caspase-dependent mechanism. Autophagy is a type of cell death involving bulk degradation of cytoplasmic proteins or organelles in the lytic compartment. Inhibition of autophagy enhances protein aggregation and cell death (Ravikumar et al., 2002). Our data showed that 3-MA accelerates protein accumulations and enhances cell death, particularly in the expanded variants (Fig. 2E,F). Conversely, Rapamycin, an inducer of the autophagic process, decreases protein aggregates and reduces cell death (Fig. 2G). Taken together, our findings indicate that autophagy is involved in the degradation and clearance of aggregations of expanded HOXA1 variants.

HOXA1 splice variant 1 is reported to be active in E7–E8.5 and functions in the presumptive rhombomere 3 and 4 regions of the developing hindbrain (Zhang et al., 1994). However, the nonhomeodomain-containing variant is expressed in the endodermal derivative after E8.5 to the adult stage (Godwin et al., 1998). Therefore, we speculate that splice variant 1 may be active only in early stages of embryonic neurodevelopment. As one of the DNA-binding proteins and transcription factors in the HOX gene family, HOXA1 also relies on the activity of transcriptional cofactors aside from its DNA-binding properties (Pearson et al., 2005). A previous study has reported that HOXB1 and PBX1 cooperatively activate transcription under the control of b1-ARE (autoregulatory enhancer). Moreover, HOXA1 and HOXA2 are also able to activate transcription by b1-ARE in cooperation with PBX1 (Di Rocco et al., 2001). We examined the binding ability of our HOXA1 variants to PBX1 and b1-ARE by evaluating the cooperative transcriptional activation using a luciferase assay. Expanded variants transfected into COS-7 as well as SK-N-SH neuroblastoma and P19 embryonic carcinoma cells (EC) that were induced to undergo neuronal differentiation indeed showed significantly reduced activation compared with the wild type, whereas the deleted variant expressed in P19 cell line showed an enhanced activation (Fig. 3). However, the reason for increased transcriptional activity observed in the deleted variant in this cell line is as yet unknown. Immunofluorescence images and immunoprecipitation assay clearly confirmed the inability of aggregated forms of expanded HOXA1 protein to bind efficiently with PBX1 protein. The co-

operative interaction between Hox and Pbx is mediated by a conserved hexapeptide sequence located toward the N-terminal region from Hox homeodomain (Phelan and Fetherstone, 1997; Remacle et al., 2004). Our results suggest that polyhistidine variants in HOXA1 might also affect the binding efficiency of its homeodomain to certain cofactors and/or other target genes or proteins.

Human neuroblastoma SK-N-SH cells, from a malignant pediatric tumor derived from the neural crest, retains its ability to differentiate into the neuronal lineage when exposed to RA (Wainwright et al., 2001). HOXA1 is the first target gene activated by RA, followed by a sequential activation of other HOX genes (Simeone et al., 1990; Martinez-Ceballos et al., 2005). We showed an inhibition of neuronal differentiation by not only the extended but also the deleted variants (Fig. 5). Our study provides novel insights on the pathological implications of the polyhistidine tract in HOXA1 and leaves us with the intriguing possibility that polyhistidine repeat expansions and deletions may cause aberrations in neuronal morphogenesis or differentiation in general.

EC cells differentiate into various lineages depending on the presence of activators in the culture medium (Pachernik et al., 2005). Growth under serum-free conditions committed EC cells to neural differentiation. Differentiation of EC cells into endodermal-like cells is induced by serum. Interestingly, we found that, in EC cells committed to neural differentiation, transcriptional activity inversely coincided with polyhistidine repeat length. These results further strengthen our theory that polyhistidine length affects neurodevelopment. In agreement with our results, another study has proposed that HOXA1 may function as a stimulator of neuroectodermal and mesodermal differentiation and a repressor of embryonic endoderm formation (Martinez-Ceballos et al., 2005) and that aberration in HOXA1 could lead to increased expression of endodermal genes by RA and would lead to repression of neuroectodermal and mesodermal markers. Premature death of HOXA1-expressing cells may impair transcription and neuronal differentiation.

Recently, Tischfield and colleagues (2005) reported that patients with Bosley-Salih-Alohrainy syndrome were homozygous for HOXA1 truncating mutations, whereas heterozygotes had normal phenotypes. This mutation resulted in abnormal development of the central nervous system in the brainstem. Distinguishing phenotypes include horizontal gaze abnormalities, mental retardation, and autism spectrum disorder. Even low levels of Hoxa1 expression in Hoxa1^{+/-} cells are sufficient for normal activation of the Hoxa1 pathway and may explain why mice heterozygous for HOXA1 mutations appear normal (Pasqualetti et al., 2001; Martinez-Ceballos et al., 2005). In our previous report, we identified heterozygous polyhistidine repeat variants in HOXA1 gene from a Japanese population comprising normal and autistic individuals. No individuals homozygous for these mutations have been found. We speculate that there is a possibility

that phenotypic aberrations may exist in homozygous individuals. Future *in vivo* studies are essential to examine the physiological functions of the polyhistidine variants of HOXA1.

ACKNOWLEDGMENTS

We thank Prof. Vincenzo Zappavigna for providing the PBX1 and pAdMLARE reporter plasmids. R.C.P. is the recipient of a research scholarship from the Japanese Ministry of Education, Culture, Sports, Science and Technology.

REFERENCES

- Broustas CG, Gokhale PC, Rahman A, Dritschlo A, Ahmad I, Kasid U. 2004. BRCC2, a novel BH3-like domain-containing protein, induces apoptosis in a caspase-dependent manner. *J Biol Chem* 279:26780–26788.
- Carpenter EM, Goddard JM, Chisaka O, Manley NR, Capocchi MR. 1993. Loss of Hoxa1 (Hox-1.6) function results in the reorganization of the murine hindbrain. *Development* 118:1063–1075.
- Chisaka O, Musci TS, Capocchi MR. 1992. Developmental defects of the ear, cranial nerves and hindbrain resulting from targeted disruption of the mouse homeobox gene, Hox-1.6. *Nature* 355:516–520.
- Darmon M, Bottenstein J, Sato G. 1981. Neural differentiation following culture of embryonal carcinoma cells in a serum-free defined medium. *Dev Biol* 85:463–473.
- Di Rocco G, Gavalas A, Popperl H, Krumlauf R, Mavilio F, Zappavigna V. 2001. The recruitment of SOX/OCT complexes and the differential activity of HOXA1 and HOXB1 modulate the Hoxb1 auto-regulatory enhancer function. *J Biol Chem* 276:20506–20515.
- Gavalas A, Studer M, Lumsden A, Rijli FM, Krumlauf R, Chambon P. 1998. Hoxa1 and Hoxb1 synergize in patterning the hindbrain, cranial nerves and second pharyngeal arch. *Development* 125:1123–1136.
- Gehring WJ, Affolter M, Burglin T. 1994. Homeodomain proteins. *Annu Rev Biochem* 63:487–526.
- Godwin AR, Stadler HS, Nakamura K, Capocchi MR. 1998. Detection of targeted GFP-Hox gene fusions during mouse embryogenesis. *Proc Natl Acad Sci U S A* 95:13042–13047.
- Grier DG, Thompson A, Kwasniewska A, McGonigle GJ, Halliday HL, Lappin TR. 2005. The pathophysiology of HOX genes and their role in cancer. *J Pathol* 205:154–171.
- Huang H, Rastegar M, Bodner C, Goh SL, Rambaldi I, Featherstone M. 2005. MEIS C termini harbor transcriptional activation domains that respond to cell signaling. *J Biol Chem* 280:10119–10127.
- Lufkin T, Dierich A, LeMeur M, Chambon M. 1991. Disruption of the Hox-1.6 homeobox gene results in defects in a region corresponding to its rostral domain of expression. *Cell* 66:1105–1119.
- Martinez-Ceballos E, Chambon P, Gudas LJ. 2005. Differences in gene expression between wild type and Hoxa1 knockout embryonic stem cells after retinoic acid treatment or LIF removal. *J Biol Chem* 280:16484–16498.
- Pachernik J, Bryja V, Esner M, Kubala L, Dvorak P, Hampl A. 2005. Neural differentiation of pluripotent mouse embryonic carcinoma cells by retinoic acid: inhibitory effects of serum. *Physiol J* 54:115–122.
- Paraguison RC, Higaki K, Sakamoto Y, Hashimoto O, Miyake N, Matsumoto H, Yamamoto K, Sasaki T, Kato N, Nanba E. 2005. Polyhistidine tract expansions in HOXA1 result in intranuclear aggregation and increased cell death. *Biochem Biophys Res Commun* 336:1033–1039.
- Pasqualetti M, Neun R, Davenne M, Rijli FM. 2001. Retinoic acid rescues inner ear defects in Hoxa1 deficient mice. *Nat Genet* 1:34–39.
- Pearson JC, Lemons D, McGinnis W. 2005. Modulating hox gene functions during animal body patterning. *Nat Rev Neurosci* 6:893–904.
- Phelan ML, Featherstone MS. 1997. Distinct HOX N-terminal arm residues are responsible for specificity of DNA recognition by HOX monomers and HOX-PBX heteromers. *J Biol Chem* 272:8635–8643.
- Ravikumar B, Duden R, Rubinsztein DC. 2002. Aggregate-prone proteins with polyglutamine and polyalanine expansions are degraded by autophagy. *Hum Mol Genet* 11:1107–1117.
- Ravikumar B, Berger Z, Vacher C, O’Kane CJ, Rubinsztein DC. 2006. Rapamycin pre-treatment protects against apoptosis. *Hum Mol Genet* 15:1209–1216.
- Remacle S, Abbas L, De Backer O, Pacico N, Gavalas A, Gofflot F, Picard JJ, Rezsosahay R. 2004. Loss of function but no gain of function caused by amino acid substitutions in the hexapeptide of Hoxa1 *in vivo*. *Mol Cell Biol* 24:8567–8575.
- Shanmugam K, Featherstone MS, Saragovi HU. 1997. Residues flanking the HOX YPWM motif contribute to cooperative interactions with PBX. *J Biol Chem* 272:19081–19087.
- Simeone A, Acampora D, Arcioni L, Andrews PW, Boncinelli E, Mavilio F. 1990. Sequential activation of HOX 2 homeobox genes by retinoic acid in human embryonal carcinoma cells. *Nature* 346:763–766.
- Slupsky CM, Sykes DB, Gay GL, Sykes BD. 2001. The HoxB1 hexapeptide is a prefolded domain: implications for the Pbx1/Hox interaction. *Prot Sci* 10:1244–1253.
- Tanaka Y, Kawahata K, Nakata T, Hirokawa A. 1992. Chronological expression of microtubule-associated proteins (MAPs) in EC cell P19 after neural induction by retinoic acid. *Brain Res* 596:269–278.
- Tischfield MA, Bosley TM, Salih MAM, Alorainy IA, Sener EC, Nester MJ, Oystreck DT, Chan WM, Andrews C, Erickson RP, Engle EC. 2005. Homozygous HOXA1 mutations disrupt human brainstem, inner ear, cardiovascular and cognitive development. *Nat Genet* 37:1035–1037.
- Wainwright LJ, Lasorella A, Iavarone A. 2001. Distinct mechanisms of cell cycle arrest control the decision between differentiation and senescence in human neuroblastoma cells. *Proc Natl Acad Sci U S A* 98:9396–9400.
- Zhang M, Kim HJ, Marshall H, Gendron-Maguire M, Lucas DA, Baron A, Gudas T, Gridley LJ, Krumlauf R, Grippo JF. 1994. Ectopic Hoxa-1 induces rhombomere transformation in mouse hindbrain. *Development* 120:2431–2442.

REVIEW ARTICLE

Review of animal models for autism: implication of thyroid hormone

Miyuki Sadamatsu¹, Hirohiko Kanai¹, Xiaobin Xu², Ying Liu², and Nobumasa Kato²¹Department of Psychiatry, Shiga University of Medical Science, Otsu, and ²Department of Neuropsychiatry, Graduate School of Medicine, University of Tokyo, Tokyo, Japan

ABSTRACT Autism is a behaviorally defined disorder associated with characteristic impairments in social interactions and communication, as well as restricted and repetitive behaviors and interest. Its prevalence was once thought to be 2/10 000, but recently several large autism prevalence reviews revealed that the rate of occurrence was roughly 30/10 000. While it has been considered a developmental disorder, little is certain about its etiology. Neuroanatomical studies at the histological level in the brains of autistic patients provide many arguments in the etiology of autism. Results from postmortem and imaging studies have implicated many major structures of the brain including the limbic system, cerebellum, corpus callosum, basal ganglia and brainstem. There is no single biological or clinical marker for autism. While several promising candidate genes have been presented, the critical loci are yet unknown. Environmental influences such as rubella virus, valproic acid, and thalidomide exposure during pregnancy are also considered important, as concordance in monozygotic twins is less than 100% and the phenotypic expression of the disorder varies widely. It is thus hypothesized that non-genetic mechanisms contribute to the onset of autistic syndrome. In light of these ambiguities, hope is held that an animal model of autism may help elucidate matters. In this article, we overview most of the currently available animal models for autism, and propose the rat with mild and transient neonatal hypothyroidism as a novel model for autism.

Key Words: animal model, autism, hypothyroidism, thalidomide, rat

INTRODUCTION

Autism is a severe neurobiological disorder that develops in the first 3 years of life. It is characterized by impairments in social interactions and communication, as well as restricted and repetitive behaviors and interest. Its prevalence was once thought to be 2/10 000, but recently several large autism prevalence reviews revealed that the rate of occurrence is roughly 30/10 000, and its incidence is progressively increasing (Stokstad 2001; Muhle *et al.* 2004; Honda *et al.* 2005). The etiology of autism remains to be clarified. Since the first description by Kanner in 1943, autism has been attributed as the earliest manifestation of schizophrenia and then to a failure of parental nurturing. Currently its etiology is unanimously thought to derive from some developmental disorder of communication with a neurobiological basis.

The genetic component clearly plays an important role in the pathophysiology of the disorder, as there is a concordance rate of approximately 2–6% in dizygotic twins as opposed to the 60–95% concordance rate in monozygotic twins (Ritvo *et al.* 1985; Bailey *et al.* 1995). The prevalence rate of non-twin siblings of children with autism varies from 1–6% (Hallmayer *et al.* 2002). Nevertheless, the finding of the increasing prevalence rate of autism during the past 10 years may cast some doubt on whether genetics alone can explain the whole picture. A case-control study of a group of Swedish adults with Asperger syndrome (1999) has noted that the rate of autistic children with mild mental retardation remains relatively stable, while the rate is increased in children with severe mental retardation and with normal intelligence (Gillberg & Wing 1999). Some other epidemiological studies (Ehlers & Gillberg 1993; Kadesjo *et al.* 1999) indicate that the recent increase of autistic children is mainly attributable to the increase of so-called atypical autism characterized by a lesser degree of mental retardation or normal intelligence. This higher prevalence rate of high function autism or Asperger syndrome encouraged us to accept the concept of autism spectrum disorders (ASD).

Environmental factors can cause developmental disabilities. Case reports of autism associated with environmental factors, such as rubella virus, valproic acid, and thalidomide exposure during pregnancy, lead to the hypothesis that non-genetic mechanisms may also produce an autistic syndrome (Chess 1977). Although there clearly exists a genetic component in the pathophysiology of autism, ASD appears to be a syndrome of complex genetic traits, rather than the outcome of any single mutation. Furthermore, a varied burden of environmental factors may contribute to the broad spectrum disorders of autistic syndrome with a higher prevalence rate.

Since the 1970s, researchers have known that autism is a complex genetic disorder. Thus far a number of research groups including an international consortium have tried to identify the responsible gene(s) in autism. However, although several promising candidates have been presented, the critical loci are still not known. Therefore, the animal models for autism currently available are mainly derived from empiric data such as viral infection, thalidomide exposure and valproate exposure in human subjects. The rationale for some models arises from the similarities between clinical manifestations in autism and behavioral abnormalities exhibited by treated animals. In this review, we briefly introduce currently available animal models of autism, and then present our hypothesis, the pivotal role of mild neonatal hypothyroidism, as a putative animal model for studying the underlying mechanisms of autism and/or related neurodevelopmental disorders.

NEUROANATOMICAL AND NEUROIMAGING ASPECTS OF AUTISM

Neuroanatomical studies at the histological level in the brains of autistic patients provide many arguments in the etiology of autism.

Correspondence: Nobumasa Kato, MD, PhD, Department of Neuropsychiatry, Graduate School of Medicine, University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan. Email: katon-tky@umin.ac.jp

Received October 13, 2005; revised and accepted November 25, 2005.

Results from postmortem and imaging studies have implicated many major structures of the brain including the limbic system, cerebellum, corpus callosum, basal ganglia and brainstem.

Areas of the forebrain that have been found to be histologically abnormal include the hippocampus, subiculum, entorhinal cortex, amygdala, mammillary body, anterior cingulate gyrus and septum, structures which comprise a major portion of the limbic system. In comparison with controls, these areas showed reduced neuronal cell size and increased cell packing density (increased numbers of neurons per unit volume) bilaterally (Bauman & Kemper 1994). Golgi analysis of CA1 and CA4 pyramidal neurons has shown decreased complexity and extent of dendritic arbors in these cells (Raymond *et al.* 1996). In the amygdala, the most significant increase in cell packing density was noted in the most medially placed nuclei. With the exception of a single child of normal intelligence, the lateral nucleus has appeared to be uninvolved.

Outside of the limbic system, the most apparent and consistent abnormalities have been confined to the cerebellum and related inferior olive. Regardless of age, sex, or cognitive abilities, all the autistic brains reported to date have shown a significant decrease in the number of Purkinje cells (Bailey *et al.* 1998). With few exceptions, there has been an absence of glial hyperplasia (Bauman & Kemper 1996; Bailey *et al.* 1998) suggesting the cerebellar lesions are acquired early in development. A similar pattern of change in cell size has also been observed in the inferior olive of the brainstem but the number of neurons has been found to be preserved. Bailey *et al.* (Bailey *et al.* 1998) have noted neocortical malformations to be a prominent feature in their autopsy material. They found evidence of thickened cortices, areas of increased neuronal density, irregular laminar patterns, increased number of neurons in layer I, and abnormally oriented pyramidal cells.

The observation of postnatal brain enlargement is intriguing and a number of hypotheses have been posed to explain its origins. Clinically, the head circumference of the autistic child has been said to be either normal or slightly small at birth but later increases in size during early to mid-childhood (Courchesne *et al.* 2003; Lainhart 2003). More recently, imaging studies have indicated increased brain volume in autism (Aylward *et al.* 1999; Sparks *et al.* 2002; Herbert *et al.* 2003; Courchesne & Pierce 2005), most prominent between 2–4.5 years of age, and later appear to plateau during adolescence (Courchesne *et al.* 2001). Schumann *et al.* (Schumann *et al.* 2004) reported that children with autism (7.5–12.5 years of age) had larger right and left amygdala volumes as well as a right hippocampal volume larger than typically developing controls, even after controlling for total cerebral volume. Brambilla *et al.* (Brambilla *et al.* 2003) reviewed original MRI research papers published from 1966 to 2003 and concluded that increased total volumes of the brain, parieto-temporal lobe, and cerebellar hemisphere were the most replicated abnormalities in autism. Interestingly, recent papers suggest the size of amygdala, hippocampus, and corpus callosum may also be abnormal, although the results are controversial (Abell *et al.* 1999; Aylward *et al.* 1999; Sparks *et al.* 2002).

CURRENT ANIMAL MODELS FOR AUTISM

Neonatal Borna disease virus infected rat

Borna disease virus (BDV) is a negative strand, non-segmented RNA virus that is the prototypic member of Bornaviridae, a new class of virus in the Mononegavirales order, and is a human pathogen (De La Torre 1994). Host factors including the age at the time of inoculation, the genetic background and the immune status, as well as viral factors, influence the course of infection. In adult rats,

BDV usually causes an immune-mediated biphasic behavioral disease. After a varied incubation period, the onset of a hyperactive phase is observed, which can lead to rapid death in some animals. Excitability and hyperactivity, together with movement and posture disorders, are consistent clinical features in both natural and experimental infections. Some animals may have stereotyped behaviors. A chronic hypoactive phase with somnolence follows in conjunction with a decrease in the inflammatory reaction and high levels of virus in the Central Nervous System (CNS). During this chronic phase, symptoms resembling those of the initial phase may reemerge in the form of recurrent episodes (Taieb *et al.* 2001). Heightened viral gene expression in limbic system structures, together with astrogliosis and neuronal structural alterations within the hippocampal formation are the main histopathologic hallmarks of BDV infection in adult rats (De La Torre 2002).

When BDV is inoculated into a newborn Lewis rat, it causes a life-long persistent infection that is characterized by the lack of any significant inflammatory cell infiltration within the central nervous system (CNS) and the absence of clinical signs of BDV (Pletnikov *et al.* 1999a).

Intracranial injection of the BDV in a newborn rat pup within the first 24–48 h after birth is the most common way of inducing neonatal BDV infection in rats (Pletnikov *et al.* 2003). Infected rats have normal body shape and proportion but are overall smaller than uninfected control pups. Injury to the cerebellum is one of the most salient morphological features of neonatal infection. BDV infection induces a prominent loss of Purkinje cells (PC), with up to 75% of PCs dropping out by seven months. A loss of PCs and their dendrites in the molecular layer has been suggested to play a major role in markedly reducing cerebellum size (Hornig *et al.* 1999).

In addition to injury of the cerebellum, neonatal BDV infection affects the postnatal maturation of the hippocampus. BDV infection of dentate gyrus neurons is associated with their continuing loss and eventual complete disappearance by 45–55 postnatal days (PNDs) and replaced by reactive glial cells (Hornig *et al.* 1999; Gonzalez-Dunia *et al.* 2000).

Neonatal BDV infection also induces cortical shrinkage. It has been shown that up to 30% of cortical neurons are lost in BDV-infected rats by PND 45. Similar to the hippocampus, diminished immunoreactivity for GAP-43 and synaptophysin is observed in the neocortex of neonatally BDV-infected rats (Gonzalez-Dunia *et al.* 2000).

Neonatally BDV-infected rats have very robust astrogliosis. Astrocytes, oligodendrocytes, ependymal cells and Schwann cells in the peripheral nervous system all express BDV markers (Bautista *et al.* 1995; Pletnikov *et al.* 2002). In the late stages of neonatal infection, BDV spreads centrifugally by anterograde axonal transport and infects most inner organs innervated by peripheral or autonomic nerves.

BDV-induced neuroanatomical damage is likely to underlie the behavioral abnormalities observed in BDV-infected rats. BDV-associated behavioral deficits are as follows; (i) selectively deficient social behaviors; (ii) changes in emotional behavior; (iii) selectively reduced cognitive abilities in spatial memory and learning/contextual fear conditioning; and (iv) spontaneous locomotor hyperactivity, hyper-reactivity and stereotypy along with mild gait ataxia (Dittrich *et al.* 1989; Hornig *et al.* 1999; Bauer *et al.* 2002). Neonatally BDV-infected rats show no evidence of gross ataxia and have normal swimming speeds despite the significant cerebellar lesions (Bautista *et al.* 1995).

Pletnikov *et al.* (Pletnikov *et al.* 1999a) showed that BDV-infected adult Lewis rats exhibited locomotor hyperactivity and elevated defecation in a highly aversive and brightly lit open field,

whereas uninfected control rats showed slightly decreased ambulation when compared in a less aversive, dimly lit open field. Moreover, compared to controls, neonatally BDV-infected rats exhibited attenuated habituation of the acoustic startle at PND 23 and decreased startle responsiveness at PND 30. Prepulse inhibition of the acoustic startle reflex remained unaltered in BDV-infected rats (Pletnikov *et al.* 2001).

Another behavioral task requiring the integrity of the limbic system, particularly the hippocampus, is contextual fear conditioning. Freezing behavior and defecation response can be used in rats for assessing the amount of contextual fear conditioning. BDV-infected rats demonstrated attenuated conditional freezing in the context previously paired with either sudden loud noise or foot shock compared to control rats (Pletnikov *et al.* 1999a).

Both the hippocampus and cerebellum play a major role in the acquisition phase of the spatial navigation task and lesioning of these areas impairs acquisition of a hidden platform location in the Morris water maze (MWM). In the MWM, neonatally BDV-infected rats exhibited a performance deficit. At PND 72, BDV-infected rats had difficulties in learning the location of the platform over a series of swim trials (Rubin *et al.* 1999).

Neonatal BDV infection induced abnormal social interaction and communication in Lewis rats when tested as old as 30–35 days of age. Studies were conducted using the resident/intruder paradigm. A resident rat was isolated for one week in order to increase social motivation. An unfamiliar rat (intruder) was placed in the resident's cage. This scenario is conducive to social interactions between the rats, often resulting in play behavior (measured as number of pins, similar to a pin observed in a wrestling match) (Pletnikov *et al.* 1999b). As the result, control rat pairs exhibited significantly more pins than the pairs where either one or both rats were BDV-infected rats. Similarly, play soliciting behaviors (e.g. pounce, crawl over/under and darting) were reduced in BDV-infected rats. Nonsocial exploratory activity (e.g. ambulation and rearing) was similar in BDV-infected and non-infected residents. Duration of non-play social investigation (e.g. sniffing, approach, and follow) was higher in BDV-infected rats as compared to non-infected controls.

However, there is little serological evidence that suggests BDV infects humans (Chalmers *et al.* 2005), and its role in psychiatric disorders remains controversial.

Chemical teratogenic model of autism

Thalidomide exposure: rats

Thalidomide (THAL) was used worldwide at the end of the 1950s and beginning of the 1960s for the treatment of anxiety and insomnia. Lenz carried out analysis of hypoplastic malformations of the limbs and reported a correlation between the intake of THAL during pregnancy and the observed birth defects (Lenz *et al.* 1962). In addition to limb defects, THAL may give rise to a wide spectrum of malformations of various organ systems. Anomalies noted are heart defects, laryngeal and tracheal abnormalities, anotia, microtia, and hearing impairment, choanal atresia, microphthalmia, cloboma, intestinal atresia, aplastic or hypoplastic gallbladder, renal anomalies, criptorchism, vaginal and anal atresia, as well as dysfunction of cranial nerves, notably the 6th and 7th nerve (Miller & Stromland 1999).

Recent epidemiological studies have revealed that THAL exposure during the first trimester in humans causes higher incidence of autism in the offspring. Exposure between the 20th and 24th day of gestation led to an incidence of autism of 5 out of 15 cases (Stromland *et al.* 1994; Miller *et al.* 2005). This critical period for exposure corresponds to the time of early development of the Cen-

tral Nervous System (CNS), when the neural tube begins to form. On the basis of somite numbers in early embryos of rats and humans, E9-E11 in rats is considered to be from early somite stage corresponding to approximately E20-E24 in human embryos (Rice & Barone 2000). Models exposed to THAL showed a reduction of cell numbers in the cranial nerve motor nuclei, reductions in Purkinje cell number and cerebellar volume, and retarded migration of 5-HT neuron (Rodier *et al.* 1997; Narita *et al.* 2002). Narita *et al.* (Narita *et al.* 2002) reported that a significant increase of hippocampal serotonin concentration was observed in the group exposed to THAL on E9. E9 THAL exposure resulted in an increase of hippocampal serotonin and frontal cortex dopamine, as well as hyperserotonemia. These observations all parallel the reported human autistic pathologic findings (Rodier *et al.* 1997).

Although neurobehavioral investigations have been scarce, Vorhees *et al.* (Vorhees *et al.* 2001) have reported that male THAL exposed rat pups show significant increases in errors and latency in the multiple-T Cincinnati water maze. They also indicated that THAL exposure induced increased preweaning mortality and male specific, late onset reduction in growth in rat pups (Vorhees *et al.* 2001).

Valproic acid exposed rat model

While THAL may have a teratogenic effect in rodents that differs from that in primates (Schumacher *et al.* 1972), valproic acid (VPA) has a similar effect in rodents and humans (Kemper & Bauman 1993). The effect of VPA is observed if the rat brainstem is exposed to VPA *in utero* and the somatic effects are similar to those of THAL (Kemper & Bauman 1993). Offspring of female rats injected with VPA at the time of neural tube closure show brain abnormalities resembling those found in autistic patients (Christianson *et al.* 1994). There are several brainstem abnormalities found so far in rats exposed to VPA *in utero*: (i) diminished number of motor neurons in the oculomotor, trigeminal, abducens, and hypoglossus nuclei of cranial nerves; (ii) shortening of the region caudal to the facial nucleus and lengthening of the region rostral to the facial nucleus; (iii) smaller cerebellum with reduction of a number of Purkinje cells both in the hemispheres and vermis; and (iv) reduced cerebellar nucleus interpositus (Rodier *et al.* 1997; Ingram *et al.* 2000a)

Schneider *et al.* (Schneider *et al.* 2001) have suggested that rats exposed to VPA during gestation may resemble the abnormalities seen in autism both neurophysiologically and behaviorally. They have demonstrated that VPA exposed rat offspring exhibit (i) lower sensitivity to pain and higher sensitivity to non-painful stimuli; (ii) diminished acoustic prepulse inhibition; (iii) locomotor and repetitive/stereotypic-like hyperactivity combined with lower levels of exploratory activity; and (iv) decreased number of social behaviors and increased latency to social behaviors.

Neonatal amygdala lesioned rat

Results from neuroanatomical studies indicate that medial temporal lobe structures, especially amygdala, may be implicated in the pathogenesis of autism (Bachevalier 1996; Baron-Cohen *et al.* 2000). Some authors have noted similarities between autism and the Kluver–Bucy syndrome, a syndrome caused by bilateral lesions to the anterior temporal lobes in monkeys (Baron-Cohen *et al.* 2000). Monkeys with the Kluver–Bucy syndrome display features often seen in autistic subjects such as absence of social chattering, lack of facial expression and absence of emotional reactions. Other such similarities include repetitive abnormal movement patterns, increased aggression, and the tendency to examine objects by mouth or smell. Several post-mortem studies in autistic subjects

have demonstrated amygdala abnormalities with small neuronal size and increased cell-packing density (Bauman & Kemper 1985; Kemper & Bauman 1993; Bailey *et al.* 1998).

Experimental lesion studies in non-human primates provide further evidence for medial temporal lobe involvement in autism. Bilateral lesions to the medial temporal lobe in infant rhesus monkeys have resulted in long-term deficits in social behavior, an effect that is absent in monkeys receiving similar lesions in adulthood (Bachevalier 1996). Monkeys subjected to bilateral removal of the amygdala, hippocampus, and adjacent cortical areas were uninterested in and avoided social contacts. Those monkeys also developed autistic-like characteristics, such as unexpressive faces, very little eye contact, locomotor stereotypies, and self-directed activity (Prather *et al.* 2001; Bauman *et al.* 2004).

Neonatal ibotenic acid lesion of the amygdala in the rat has also been proposed as an animal model of autism. Excitotoxic lesions of the amygdala at PND 7, but not PND 21 in rat, produce multiple behavioral abnormalities persisting into adulthood, indicating neurodevelopmental deficits of structures connected to the amygdala (Daenen *et al.* 2002a). Lesioning the amygdala on PND 7 resulted in an adult animal with stereotypic-like increased ambulatory behaviors and decreased investigatory behaviors. Moreover, those animals exhibited increased locomotor reactivity to challenge with a low dose of apomorphine, reminiscent of supersensitivity of postsynaptic dopamine systems in the nucleus accumbens (Wolterink *et al.* 2001; Daenen *et al.* 2002a).

Other lesioned animals

There have been several reports suggesting that neonatally ventral hippocampus (VH) lesioned rats show many aspects of abnormalities in behavior and cellular formation reminiscent of schizophrenia. When tested as juveniles (PND 35), rats with the neonatal VH lesions are less social than controls (Sams-Dodd *et al.* 1997), but otherwise behave normally in motor tests involving exposure to stress and dopamine agonists. In adolescence and adulthood (PND 56 and older), lesioned animals display markedly changed behaviors such as motor hyperresponsiveness to stress and stimulants, and enhanced stereotypies. They also show deficits in PPI and latent inhibition, impaired social behaviors and working memory problems (Lipska & Weinberger 1993; Lipska & Weinberger 1994; Lipska *et al.* 1995).

However, other reports found that rats lesioned in the VH on PND 7 or PND 21, showed no differences in social behavior related or unrelated to social play behavior early in life or in adulthood (Wolterink *et al.* 2001; Daenen *et al.* 2002b). In monkeys, emotional behavior was not disturbed with damage in the hippocampal area only (Bauman *et al.* 2004). Wood *et al.* (Wood *et al.* 1997) suggested that the pattern of impairments associated with the excitotoxic VH lesion varies depending on the age at which lesioning occurs. Consequently, VH lesioned rats are still considered to be controversial as a model of autism.

Early prefrontocortical damage in humans has been shown to impair cooperative and reciprocal behavior, social interactions, and social cognition (Eslinger *et al.* 2004). It is suggested that dysfunctions and morphological abnormalities of the prefrontal cortex (PFC) are implicated in the pathophysiology of autism (Baron-Cohen *et al.* 1999). Neonatal PFC lesions have also been proposed as an adequate model to investigate early developmental aberrations (Schneider & Koch 2004). The total amount of self-grooming and social behaviors was reduced in PFC lesioned animals compared to controls. Neonatal PFC lesions reduced pinning in juvenile rats and lesioned rats showed an increase in the total number of so called 'partial rotations'. Partial rotation is an adult-like pattern of

defense, so investigators suggested that neonatal lesions of PFC lead to a behavioral shift of social play in juvenile rats to an adult-like pattern of defense (Schneider & Koch 2004).

There is growing evidence that the cerebellum is implicated in autism. Recently, many studies have demonstrated that the cerebellum is involved not only in the regulation of motor skills, but also in more complex integrated functions, such as classical conditioning, learning of motor skills, spatial learning, habituation of exploratory behavior and the acoustic startle response (McCormick & Thompson 1984; Leaton & Supple 1986; Leaton & Supple 1991; Dahhaoui *et al.* 1992a,b; Molinari *et al.* 1997). The cerebellum is further implicated in motivations and emotional behavior as well (Heath *et al.* 1980; Caston *et al.* 1998). Adult rats with midline lesions of the cerebellum performed at PND 10 exhibited the hyperactivity in the open field test as well as overt disinhibition tendencies in the anxiety and social discrimination tests (Bobee *et al.* 2000). These results indicate the involvement of the cerebellar vermis in the pathology of autism, considering a number of autistic subjects have a hypoplasia of cerebellar vermal lobules.

Genetic model

Recently, overwhelming evidence of genetic underpinnings of autism has generated much research. As this field is rapidly developing, many candidate loci for autism have been published in recent years. Spontaneous mutants or transgenic animal models can greatly help to delineate the role of these candidate genes.

The nonapeptide oxytocin (OT) is synthesized in the hypothalamus and released into the blood stream via axon terminals in the posterior pituitary or neurohypophysis. OT receptors are concentrated in several brain regions involved in social behavior in the mouse, including the olfactory bulbs, piriform cortex, amygdala and lateral septum. OT facilitates the formation of the mother-infant bond in sheep and stimulates nurturing behaviors in rodent females toward pups. In male rats, chronic OT treatment doubles the time spent in social contact. OT knockout mice (OTKO) fail to remember recently encountered individuals despite apparently normal olfactory and general cognitive abilities (Young 2001; Winslow & Insel 2002). Central injections of OT prior to the first encounter, but not after, completely rescue this very specific deficit and infusions of an OT antagonist inhibit social recognition in normal wild-type (WT) mice (Ferguson *et al.* 2000). Both WT and OTKO mice showed a similar neuronal activation in the initial encounter, as evidenced by the comparable c-Fos immunoreactivity in olfactory bulbs, piriform cortex, cortical amygdala, and the lateral septum. However, WT mice, but not OTKO mice, exhibited an induction of c-Fos in the medial amygdala, whereas OTKO, but not WT mice, showed dramatic increases in c-Fos in the somatosensory cortex and the hippocampus (Ferguson *et al.* 2000). These findings have an interesting parallel with recent neuroimaging studies in autistic human patients, suggesting that people with autism utilize alternative cortical areas to process social cues, areas that are typically activated by non-social cues in normal subjects (Schultz *et al.* 2000).

Recent genetic reports implicate a number of genes in the causation of autism and the Reelin gene (*RELN*) is one such gene (Fatemi *et al.* 2001). Persico *et al.* (Persico *et al.* 2001; Zhang *et al.* 2002) reported that individuals inheriting alleles of the Reelin gene that contain 11 CGG repeats in the 5'-UTR of the *RELN* mRNA have an increased risk of autism. Another group has reported that autistic patients and their first-degree relatives show significantly reduced plasma levels of full-length Reelin and its low molecular weight isoforms (Fatemi *et al.* 2001). The reeler mutation is a spontaneous recessive mutation in mice that leads, in the homozygous

state, to the absence of Reelin and to severe disorganization of cortical, hippocampal, and cerebellar development. In comparison to WT mice, heterozygous reeler mice (*rl/+*) displaying Reelin levels reduced by 50% do not show gross developmental abnormalities of the CNS, but do show a progressive loss of Purkinje cells in the cerebellum during the first postnatal weeks (Tueting *et al.* 1999). The loss of Purkinje cells is seen only in male *rl/+*, and not in female *rl/+* mice (Hadj-Sahraoui *et al.* 1996).

Another interesting example of genetically altered mouse models presenting autistic-like features is mice deficient for Dishevelled-1 (*Dvl1*) proteins. *Dvl1* is one of three mouse homologs of the *Drosophila* segment polarity gene *Dishevelled*. Mice deficient in *Dvl1* were reported to exhibit abnormal social interaction as well as deficits in sensorimotor gating, as measured by impaired prepulse inhibition (PPI) (Lijam *et al.* 1997). These mice have been noted as a potent model for autism or schizophrenia, but the deficits in social memory task and PPI were not replicated in *Dvl1*-null mice in a later study (Long *et al.* 2004).

NEONATAL HYPOTHYROIDISM RATS

Thyroid hormone is essential for brain development and maintenance of basal metabolic rates. The manipulation of thyroid hormone in laboratory animals typically increases activity levels and decreases performance during motivated learning tasks. It is well-known that hypothyroidism during the critical period of brain development induces irreversible dysfunction of the central nervous system. The timing of thyroid hormone manipulation plays a critical role in the degree to which developmental sequelae are expressed. The anatomical bases of behavioral and intellectual deficits may result from global reductions in brain size, premature termination of neuronal proliferation, non-migrated granule cells in the cerebellar cortex and caudate nucleus, decreased synaptic junctions in cerebellar cortex and malformed dendrites on Purkinje cells (Lewis *et al.* 1976). Humans with primary or secondary congenital hypothyroidism demonstrate deficits in academic skills as children, and as adults, decreased performance on neuropsychological tests and prolongation of latencies for visual- and auditory-evoked potentials (Murphy & Nagy 1976; Osterweil *et al.* 1992).

Lactating rats receiving 0.02% propylthiouracil (PTU) in their drinking water transfer the goitrogenic effect to the offspring through their milk. This treatment induces a temporary mild hypothyroid condition of the pups (Van Middlesworth & Norris 1980). We conducted experiments to investigate the effects of temporary neonatal PTU-induced hypothyroidism on the behavior of rats. Rat pups were treated with 0.02% PTU in drinking water to dams from day 0–19 post partum (Kato *et al.* 1982). The serum T4 level was depressed below the limit of detection at 2 weeks of age, but recovered to the normal level at 4 weeks of age (Akaike *et al.* 1991). The open field test was conducted at 3, 6, and 9 weeks of age. At 3 weeks of age, the number of ambulations did not differ between PTU rats and controls. At 6 and 9 weeks of age, the number of ambulations of the PTU rats was significantly greater than that of the control rats. Kato *et al.* reported extensive hyperactivity (Akaike *et al.* 1991; Akaike & Kato 1997) and attenuated habituation in the open field test in PTU rats after maturation, as shown in Figure 1 (Kato *et al.* 1992).

Spatial learning ability was further investigated in the PTU rats. Biel water maze tests at the age of 6 weeks showed an increase in errors with prolonged swimming time in the PTU rats. The radial arm maze test was performed to evaluate spatial maze learning. The test started at 13 weeks and revealed that the PTU animals required more trials until they showed the first well-performed trial. The

PTU rats showed more active moving from arm to arm compared to controls. However, while the number of total choices of PTU rats was increased the number of correct choices was smaller than the control values (Akaike *et al.* 1991; Akaike & Kato 1997). The performance of PTU rats was further assessed by the modified T-maze test and then the mirror image of the first trial (Fig. 2a). The performance of PTU rats was superior to that of the controls in the initial maze test, but it was clearly inferior to that of the controls in terms of a higher error frequency and a longer running time upon reversal of the route to the mirror image of the original (Fig. 2b). This was interpreted as inability to adapt to changes in the environment and a reference for the highly repetitive and routine response pattern initially acquired.

As stated earlier, the most apparent and consistent neuropathology in autistic patients lies in the cerebellum. In this regard, it might be of significance that PTU rats have retarded granular cell migration in the external granular layer (Sadamatu & Watanabe 2005). Furthermore, PTU rats exhibited a marked susceptibility to audiogenic seizures, starting from the age 7 weeks and persisting into adulthood (Yasuda *et al.* 2000).

These results suggest that mild hypothyroidism around the critical period causes permanent impairment of brain function, as manifested by hyperactivity, lack of habituation, spatial learning impairment and auditory hypersensitivity. It is thus expected that PTU-treated rats may serve a useful model for autism.

DISCUSSION

The extremely high concordance rate of autism in monozygotic twins, as compared in dizygotic twins, clearly indicates the important role of genetic factors. The published genome screens have found convergent evidence for linkage in several genomic regions, with regions on chromosome 2, 7, 15, 16 (IMGSAC 2001a). In particular a region on chromosome 7q showed increased allele sharing in all screens (Risch *et al.* 1999; IMGSAC 2001a,b; Bartlett *et al.* 2005). *RELN* (Persico *et al.* 2001) and *HOXA1* (Ingram *et al.* 2000b), both on chromosome 7q22, are the most prospective candidate genes. In human subjects, one report showed that blood levels of Reelin were reduced (Fatemi *et al.* 2002). Although the case-control and affected sib-pair findings fail to support a role for *RELN* in susceptibility to Autism Spectrum Disorder (ASD), the more powerful family-based association study demonstrates that *RELN* alleles with larger numbers of CCG repeats may play a role in the etiology of some cases of ASD, especially in children without delayed phrase speech (Zhang *et al.* 2002; Bonora *et al.* 2003). Recent studies have reported conflicting findings of an association between a variant of the *HOXA1* gene and autism (Ingram *et al.* 2000b; Conciatori *et al.* 2004; Gallagher *et al.* 2004). Thus, so far a single gene responsible for the pathogenesis of autism has not been found and, it seems unlikely that any single gene can explain the whole picture of autism.

Genes have two broad roles, the first being the template function and the second the transcriptional function. Although the template function is largely independent of outside forces, the transcriptional function is highly regulated and responsive to environmental factors.

The question of whether or not the actual number of autistic patients has increased is also a matter of debate. Honda *et al.* (Honda *et al.* 2005) first reported that childhood autism was more frequent in Japan than previously estimated. Cumulative incidence of childhood autism up to 5 years in the birth cohort in the Yokohama increased up to 27.2 per 10 000 in 1991 in the strictest sense, whereas it was 16.2 in 1988. If indeed, the prevalence of autism is

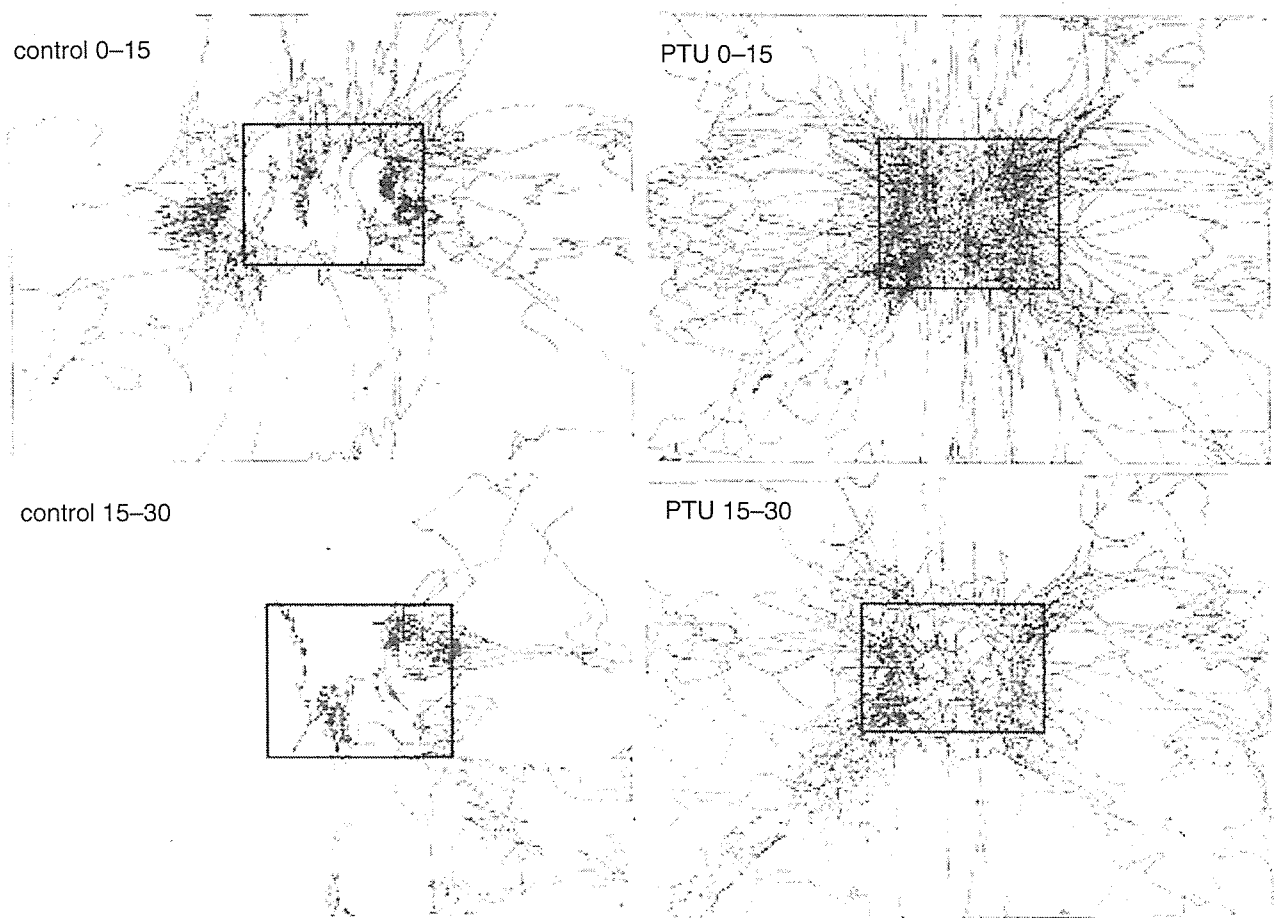


Fig. 1 Comparison of spontaneous movement in a PTU rat (right) and its littermate control (left) as detected by a multidimensional behavioral analyzer (Animex) at the age of 10 weeks. The device recorded the linear locomotion of the animal for two consecutive 15-min periods. The rectangle in the center of each figure indicates the base of the cage and traces outside the rectangle indicate rearing (Kato *et al.* 1992).

growing recently in some urban areas like Yokohama regardless of the rate, it seems plausible that environmental factors might contribute to the incidence of autism. In view of this, autism-like syndrome(s) due to environmental factors may not necessarily be the same as classical autism with mental retardation.

Results from postmortem and imaging studies have implicated many major structures of the brain including the limbic system, cerebellum, corpus callosum, basal ganglia and brainstem. However, is any single brain region able to explain such a broad spectrum dysfunction of ASD? Baron-Cohen *et al.* (Baron-Cohen *et al.* 2000) proposed the amygdala as an area responsible for the impairment of social behavior in autism, but recent data on the effects of amygdala lesions in macaque monkeys did not support their hypothesis (Amaral *et al.* 2003). Alternatively, a different hypothesis as to the brain region responsible for ASD may be derived from recent neuroimaging studies with human patients with the disorder. When viewing images of faces, autistic subjects, compared with unaffected subjects, exhibit a decreased activation of both the amygdala and cortical 'face' areas, and interestingly, also show an increase in other cortical regions typically activated while viewing non-social objects (Critchley *et al.* 2000). Autistic patients may have genetic and/or environmental impairments in some specific

brain areas, which, in turn, activate a different set of brain structures during social recognition (Ferguson *et al.* 2001).

Most of the neuroanatomical features highlighted by recent studies of autistic subjects indicate the aberration of very early fetal development, such as shortening and elongation of the brainstem, increased cell packing in the cerebral cortex and preceding enlargement of brain volume. This may imply the significance of a critical period when some genetic and/or environmental factors work in the fetus. The period determines the extent of organs involved, and each organ has its own period for maturation. Some environmental factors such as VPA or THAL disrupt specific points of cell proliferation and differentiation, and some factors such as thyroxine affect the maturation of some sets of organs in the CNS.

Neonatal mild hypothyroidism may provide a useful model for autism. The importance of thyroid hormone in brain development has been extensively documented. Recent studies further demonstrate that relatively subtle changes in circulating levels of thyroid hormone in pregnant women can affect the neurological outcome of their children (Morreale De Escobar *et al.* 2004; Pop & Vulmsa 2005). One candidate that affects thyroid function is endocrine disruptors. We currently focus on bisphenol-A, one of the endocrine disruptors known to alter thyroid function (Moriyama *et al.* 2002).

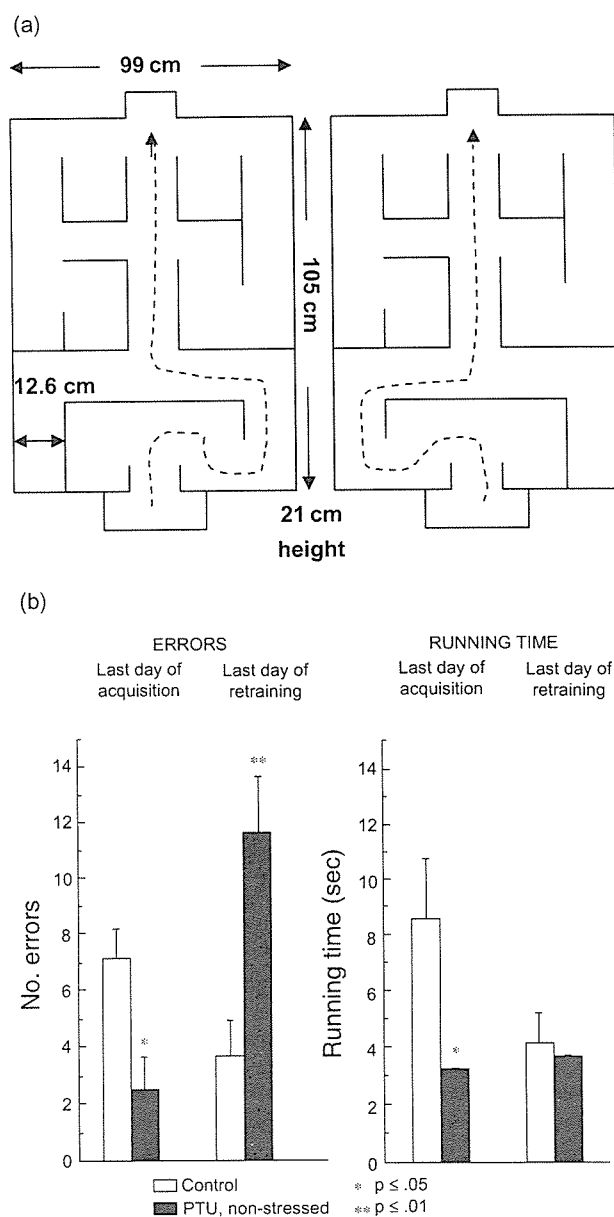


Fig. 2 (a) Diagram of T-maze for study of learning ability in rats. Rats were placed on a 23 h food deprivation schedule and trained to run the maze for food reinforcement once daily in 10 trial sessions. Route 'a' was employed for the learning phase of the study and route 'b' (the mirror image of 'a') for the relearning phase. (b) Maze-learning ability of PTU versus control rats. Average number of errors and running time were evaluated on the last day of task acquisition in the 'a' maze and the last day of retraining in the 'b' maze. Both error frequency and total running time were reduced in the PTU rats on the last day of task acquisition. In contrast, error frequency was significantly increased in the PTU rats on the last day of retraining. (Akaike *et al.* 1991; Akaike & Kato 1997)

Our preliminary data indicates that the administration of bisphenol-A at the environmental dose during the early postnatal period induces hyperactivity and learning impairment in male, but not female, rats after maturation (unpublished data).

Our understanding of the neuropathology of autism has advanced substantially over the past 20 years, but there are still so many questions that remain unsolved. Each of these models mentioned above seems to capture at least one of the pieces of the autism puzzle. It is hoped further studies will elucidate the whole picture of the neuropathology of autism.

ACKNOWLEDGMENTS

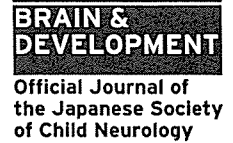
The work was supported by Research Grant on Mental Health, H17-004, from the Ministry of Health, Labor and Welfare, Tokyo, Japan.

REFERENCES

- Abell F, Krams M, Ashburner J *et al.* (1999) The neuroanatomy of autism: A voxel-based whole brain analysis of structural scans. *Neuroreport* **10**: 1647–1651.
- Akaike M, Kato N (1997) Abnormal behavior, spatial learning impairment and neuropeptides caused by temporary neonatal hypothyroidism. In: Hendrich CE (ed). *Recent Research Development in Neuroendocrinology – Thyroid Hormone and Brain Maturation*. Research Signpost, Trivandrum, pp. 39–48.
- Akaike M, Kato N, Ohno H, Kobayashi T (1991) Hyperactivity and spatial maze learning impairment of adult rats with temporary neonatal hypothyroidism. *Neurotoxicol Teratol* **13**: 317–322.
- Amaral D, Bauman MD, Schumann DM (2003) The amygdala and autism: Implications from non-human primate studies. *Genes Brain Behav* **2**: 295–302.
- Aylward E, Minshew NJ, Goldstein G *et al.* (1999) MRI volumes of amygdala and hippocampus in non-mentally retarded autistic adolescents and adults. *Neurology* **53**: 2145–2150.
- Bachevalier J (1996) Brief report: Medial temporal lobe and autism: A putative animal model in primates. *J Autism Dev Dis* **27**: 217–220.
- Bailey A, Le Couteur RA, Gottesman I *et al.* (1995) Autism as a strongly genetic disorder: Evidence from a British twin study. *Psychol Med* **25**: 63–77.
- Bailey A, Luthert P, Dean A *et al.* (1998) A clinicopathological study of autism. *Brain* **121**: 889–905.
- Baron-Cohen S, Ring HA, Wheelwright S *et al.* (1999) Social intelligence in the normal and autistic brain: An fMRI study. *Eur J Neurosci* **11**: 1891–1898.
- Baron-Cohen S, Ring HA, Bullmore ET, Wheelwright S, Ashwin C, Williams SCR (2000) The amygdala theory of autism. *Neurosci Biobehav Rev* **24**: 355–364.
- Bartlett C, Gharani N, Millonig JH, Brzustowicz LM (2005) Three autism candidate genes: A synthesis of human genetic analysis with other disciplines. *Int J Dev Neurosci* **23**: 221–234.
- Bauer M, Heinz A, Whybrow PC (2002) Thyroid hormones, serotonin and mood: Of synergy and significance in the adult brain. *Mol Psychiatry* **7**: 140–156.
- Bauman M, Kemper TL (1985) Histoanatomic observations of the brain in early infantile autism. *Neurology* **35**: 866–874.
- Bauman M, Kemper TL (1994) *Neuroanatomic Observations of the Brain in Autism*. Johns Hopkins University Press, Baltimore.
- Bauman M, Kemper TL (1996) Observations on the Purkinje cells in the cerebellar vermis in autism. *J Neuropathol Exp Neurol* **55**: 613.
- Bauman M, Lavenex P, Mason WA, Capitanio JP, Amaral DG (2004) The development of mother–infant interactions after neonatal amygdala lesions in rhesus monkeys. *J Neurosci* **24**: 711–721.
- Bautista J, Rubin SA, Moran TH, Schwartz GJ, Carbone KM (1995) Developmental injury to the cerebellum following perinatal Borna disease virus infection. *Dev Brain Res* **90**: 45–53.
- Bobee S, Mariette E, Tremblay-Leaveau H, Caston J (2000) Effects of early midline cerebellar lesion on cognitive and emotional functions in the rat. *Behav Brain Res* **112**: 107–117.

- Bonora E, Beyer KS, Lamb JA *et al.* (2003) Analysis of reelin as a candidate gene for autism. *Mol Psychiatry* **8**: 885–892.
- Brambilla P, Hardan A, Di Nemi SU, Perez J, Soares JC, Barale F (2003) Brain anatomy and development in autism: Review of structural MRI studies. *Brain Res Bull* **61**: 557–569.
- Caston J, Chianale C, Delhaye-Bouchaud N, Mariani J (1998) Role of the cerebellum in exploration behavior. *Brain Res* **808**: 232–237.
- Chalmers R, Thomas DR, Salmon RL (2005) Borna disease virus and the evidence for human pathogenicity: A systematic review. *QJM* **98**: 255–274.
- Chess S (1977) Follow-up report on autism in congenital rubella. *J Autism Child Schiz* **7**: 69–81.
- Christianson A, Chesler N, Kromberg JG (1994) Fetal valproate syndrome: Clinical and neuro-developmental features in two sibling pairs. *Dev Med Child Neurol* **36**: 361–369.
- Conciatori M, Stodgell CJ, Hyman SL *et al.* (2004) Association between the HOXA1 A218G polymorphism and increased head circumference in patients with autism. *Biol Psychiatry* **55**: 413–419.
- Courchesne E, Pierce K (2005) Brain overgrowth in autism during a critical time in development: Implications for frontal pyramidal neuron and interneuron development and connectivity. *Int J Dev Neurosci* **23**: 153–170.
- Courchesne E, Karns CM, Davis HR (2001) Unusual brain growth patterns in early life in patients with autistic disorder: An MRI study. *Neurology* **57**: 245–254.
- Courchesne E, Carper R, Akshoomoff N (2003) Evidence of brain overgrowth in the first year of life in autism. *JAMA* **290**: 337–344.
- Critchley H, Daly EM, Bullmore ET (2000) The functional neuroanatomy of social behaviour changes in cerebral blood flow when people with autistic disorder process facial expressions. *Brain* **123**: 2203–2212.
- Daenen E, Wolterink G, Gerrits MAFM, Van Ree JM (2002a) Amygdala or ventral hippocampal lesions at two early stages of life differentially affect open field behaviour later in life; an animal model of neurodevelopmental psychopathological disorders. *Behav Brain Res* **131**: 67–78.
- Daenen E, Wolterink G, Gerrits MAFM, Van Ree JM (2002b) The effects of neonatal lesions in the amygdala or ventral hippocampus on social behaviour later in life. *Behav Brain Res* **136**: 571–582.
- Dahhaoui M, Caston J, Lannou J, Avenel S (1992a) Role of the cerebellum in habituation exploration behavior in the rat. *Physiol Behav* **52**: 339–344.
- Dahhaoui M, Lannou J, Stelz T, Caston J, Guastavino JM (1992b) Role of the cerebellum in spatial orientation in the rat. *Behav Neural Biol* **58**: 180–189.
- De La Torre J (1994) Molecular biology of borna disease virus: Prototype of a new group of animal viruses. *J Virol* **68**: 7669–7675.
- De La Torre J (2002) Bornavirus and the brain. *J Infect Dis* **186**: S241–S247.
- Dittrich W, Bode L, Ludwig H, Kao M, Schneider K (1989) Learning deficiencies in Borna disease virus-infected but clinically healthy rats. *Biol Psychiatry* **26**: 818–828.
- Ehlers S, Gillberg C (1993) The epidemiology of Asperger syndrome. A total population study. *J Child Psychol Psychiatry* **34**: 1327–1350.
- Eslinger P, Flaherty-Craig CV, Benton AL (2004) Developmental outcomes after early prefrontal cortex damage. *Brain Cogn* **55**: 84–103.
- Fatemi S, Strydom JM, Halt AR, Realmuto GR (2001) Dysregulation of Reelin and Bcl-2 proteins in autistic cerebellum. *J Autism Dev Disord* **31**: 529–535.
- Fatemi S, Strydom JM, Egan EA (2002) Reduced blood levels of reelin as a vulnerability factor in pathophysiology of autistic disorder. *Cell Mol Neurobiol* **22**: 139–152.
- Ferguson J, Young LJ, Hearn EF, Matzuk MM, Insel TR, Winslow JT (2000) Social amnesia in mice lacking the oxytocin gene. *Nat Genet* **25**: 284–288.
- Ferguson J, Aldag JM, Insel TR, Young LJ (2001) Oxytocin in the medial amygdala is essential for social recognition in the mouse. *J Neurosci* **21**: 8278–8285.
- Gallagher L, Hawi Z, Kearney G, Fitzgerald M, Gill M (2004) No association between allelic variants of HOXA1/HOXB1 and autism. *Am J Med Genet* **124B**: 64–67.
- Gillberg C, Wing L (1999) Autism: Not an extremely rare disorder. *Acta Psychiatr Scand* **99**: 399–406.
- Gonzalez-Dunia D, Watanabe M, Syan S, Mallory M, Maslah E, De La Torre JC (2000) Synaptic pathology in Borna disease virus persistent infection. *J Virol* **74**: 3441–3448.
- Hadj-Sahraoui N, Frederic F, Delhaye-Bouchaud N, Mariani J (1996) Gender effect on Purkinje cell loss in the cerebellum of the heterozygous reeler mouse. *J Neurogenet* **11**: 45–58.
- Hallmayer J, Glasson EJ, Bower C *et al.* (2002) On the twin risk in autism. *Am J Hum Genet* **71**: 941–946.
- Heath R, Dempsey CW, Fontana CJ, Fitzjarrell AT (1980) Feedback loop between cerebellum and septal-hippocampal sites: Its role in emotion and epilepsy. *Biol Psychiatry* **15**: 541–556.
- Herbert M, Zeigler DA, Deutsch CK *et al.* (2003) Dissociations of cerebral cortex, subcortical and cerebral white matter volumes in autistic boys. *Brain* **126**: 1182–1192.
- Honda H, Shimizu Y, Imai M, Nitto Y (2005) Cumulative incidence of childhood autism: A total population study of better accuracy and precision. *Dev Med Child Neurol* **47**: 10–18.
- Hornig M, Weissenböck H, Horscroft N, Lipkin WL (1999) An infection-based model of neurodevelopmental damage. *PNAS* **96**: 12102–12107.
- Ingram J, Peckham SM, Tisdale B, Rodier PM (2000a) Perinatal exposure of rats to valproic acid reproduces the cerebellar anomalies associated with autism. *Neurotoxicol Teratol* **22**: 319–324.
- Ingram J, Stodgell CJ, Hyman SL, Figlewicz DA, Weitkamp LR, Rodier PM (2000b) Discovery of allelic variants of HOXA1 and HOXB1: Genetic susceptibility to autism spectrum disorders. *Teratology* **62**: 393–405.
- International Molecular Genetic Study of Autism Consortium (IMGSAC) (2001a) A genomewide screen for autism: Strong evidence for linkage to chromosomes 2q, 7q, and 16p. *Am J Hum Genet* **68**: 570–581.
- International Molecular Genetic Study of Autism Consortium (IMGSAC) (2001b) Further characterization of the autism susceptibility locus AUTS1 on chromosome 7q. *Hum Mol Genet* **10**: 973–982.
- Kadesjo B, Gillberg C, Hagberg B (1999) Brief report: Autism and Asperger syndrome in seven-year-old children: A total population study. *J Autism Dev Disord* **29**: 327–331.
- Kato N, Sundmark VC, Van Middlesworth L, Havlicek V, Friesen HG (1982) Immunoreactive somatostatin and beta-endorphin content in the brain of mature rats after neonatal exposure to propylthiouracil. *Endocrinology* **110**: 1851–1855.
- Kato N, Akaike M, Masui A, Naruse H (1992) *Brain Somatostatin in Possible Animal Models of Infantile Autism*. Elsevier Science Publishers, Amsterdam.
- Kemper T, Bauman ML (1993) The contribution of neuropathologic studies to the understanding of autism. *Neurol Clin* **11**: 175–187.
- Lainhart J (2003) Increased rate of head growth during infancy in autism. *JAMA* **290**: 393–394.
- Leaton R, Supple WF Jr (1986) Cerebellar vermis: Essential for long-term habituation of the acoustic startle response. *Science* **232**: 513–515.
- Leaton R, Supple WF Jr (1991) Medial cerebellum and long-term habituation of acoustic startle in rats. *Behav Neurosci* **105**: 804–816.
- Lenz W, Pfeiffer RA, Kosenow W, Hayman DJ (1962) Thalidomide and congenital abnormalities. *Lancet* **279**: 45–46.
- Lewis P, Patel AJ, Johnson AL, Balazs R (1976) Effect of thyroid deficiency on cell acquisition in the postnatal rat brain: A quantitative histological study. *Brain Res* **104**: 49–62.
- Lijam N, Paylor R, McDonald MP *et al.* (1997) Social interaction and sensorimotor gating abnormalities in mice lacking Dvl1. *Cell* **90**: 895–905.
- Lipska B, Weinberger DR (1993) Delayed effects of neonatal hippocampal damage on haloperidol-induced catalepsy and apomorphine-induced stereotypic behaviors in the rat. *Dev Brain Res* **75**: 213–222.
- Lipska B, Weinberger DR (1994) Gonadectomy does not prevent novelty or drug-induced motor hyperresponsiveness in rats with neonatal hippocampal damage. *Dev Brain Res* **78**: 253–258.
- Lipska B, Swerdlow NR, Geyer MA, Jaskiw GE, Braff DL, Weinberger DR (1995) Neonatal excitotoxic hippocampal damage in rats causes postpubertal changes in prepulse inhibition of startle and its disruption by apomorphine. *Psychopharmacology* **122**: 35–43.

- Long J, Laporte P, Paylor R, Wynshaw-Boris A (2004) Expanded characterization of the social interaction abnormalities in mice lacking Dvl1. *Genes Brain Behav* **3**: 51–62.
- McCormick D, Thompson RF (1984) Cerebellum: Essential involvement in the classically conditioned eyelid response. *Science* **223**: 296–299.
- Miller M, Stromland K (1999) Teratogen update: Thalidomide: A review, with a focus on ocular findings and new potential uses. *Teratology* **60**: 306–321.
- Miller M, Stromland K, Ventura L, Johansson M, Bandim JM, Gillberg C (2005) Autism associated with conditions characterized by developmental errors in early embryogenesis: A mini review. *Int J Dev Neurosci* **23**: 201–219.
- Molinari M, Leggio MG, Solida A *et al.* (1997) Cerebellum and procedural learning: Evidence from focal cerebellar lesions. *Brain* **120**: 1752–1762.
- Moriyama K, Tagam T, Akamizu T *et al.* (2002) Thyroid hormone action is disrupted by bisphenol A as an antagonist. *J Clin Endocrinol Metab* **87**: 5185–5190.
- Morreale De Escobar G, Obregon MJ, Escobar Del Rey F (2004) Role of thyroid hormone during early brain development. *Eur J Endocrinol* **151**: U25–U37.
- Muhle R, Trentacoste SV, Rapin I (2004) The genetics of autism. *Pediatrics* **113**: e472–e486.
- Murphy J, Nagy ZM (1976) Neonatal thyroxine stimulation accelerates the maturation of both locomotor and memory processes in mice. *J Comp Physiol Psychol* **90**: 1082–1091.
- Narita N, Kato M, Tazoe M, Miyazaki K, Narita M, Okado N (2002) Increased monoamine concentration in the brain and blood of fetal thalidomide- and valproic acid-exposed rat: Putative animal models for autism. *Pediatr Res* **52**: 576–579.
- Osterweil D, Syndulko K, Cohen SN *et al.* (1992) Cognitive function in non-demented older adults with hypothyroidism. *J Am Geriatr Soc* **40**: 325–335.
- Persico A, D'Agurra L, Maiorano N *et al.* (2001) Reelin gene alleles and haplotypes as a factor predisposing to autistic disorder. *Mol Psychiatry* **6**: 150–159.
- Pletnikov M, Rubin SA, Schwartz GJ, Moran TH, Sobotka TJ, Carbone KM (1999a) Persistent neonatal Borna disease virus (BDV) infection of the brain causes chronic emotional abnormalities in adult rats. *Physiol Behav* **66**: 823–831.
- Pletnikov M, Rubin SA, Vasudevan K, Moran TH, Carbone KM (1999b) Developmental brain injury associated with abnormal play behavior in neonatally Borna disease virus-infected Lewis rats: A model of autism. *Behav Brain Res* **100**: 43–50.
- Pletnikov M, Rubin SA, Carbone KM, Moran TH, Schwartz GJ (2001) Neonatal Borna disease virus infection (BDV)-induced damage to the cerebellum is associated with sensorimotor deficits in developing Lewis rats. *Dev Brain Res* **126**: 1–12.
- Pletnikov M, Moran TH, Carbone KM (2002) Borna disease virus infection of the neonatal rat: Developmental brain injury model of autism spectrum disorders. *Front Biosci* **7**: 593–607.
- Pletnikov M, Rubin SA, Moran TH, Carbone KM (2003) Exploring the cerebellum with a new tool: Neonatal Borna disease virus (BDV) infection of the rat's brain. *Cerebellum* **2**: 62–70.
- Pop V, Vulmsa T (2005) Maternal hypothyroxinaemia during (early) gestation. *Lancet* **365**: 1604–1606.
- Prather M, Lavenex P, Mauldin-Jourdain ML *et al.* (2001) Increased social fear and decreased fear of objects in monkeys with neonatal amygdala lesions. *Neuroscience* **106**: 653–658.
- Raymond G, Bauman ML, Kemper TL (1996) Hippocampus in autism: A Golgi analysis. *Acta Neuropathol* **91**: 117–119.
- Rice D, Barone S Jr (2000) Critical periods of vulnerability for the developing nervous system: Evidence from humans and animal models. *Environ Health Perspect* **108**: 511–533.
- Risch N, Spiker D, Lotspeich L *et al.* (1999) A genomic screen of autism: Evidence for a multilocus etiology. *Am J Hum Genet* **65**: 493–507.
- Ritvo E, Freeman BJ, Mason-Brothers A, Mo A, Ritvo AM (1985) Concordance for the syndrome of autism in 40 pairs of afflicted twins. *Am J Psychiatry* **142**: 74–77.
- Rodier P, Ingram JL, Tisdale B, Croog V (1997) Linking etiologies in humans and animal models: Studies of autism. *Reprod Toxicol* **11**: 417–422.
- Rubin S, Sylves P, Vogel M *et al.* (1999) Borna disease virus-induced hippocampal dentate gyrus damage is associated with spatial learning and memory deficits. *Brain Res Bull* **48**: 23–30.
- Sadamatu M, Watanabe K (2005) Is a neonatal hypothyroid rat useful as an animal model of autism? *Neurosci Res* **52** (Suppl. 1): 28.
- Sams-Dodd F, Lipska BK, Weinberger DR (1997) Neonatal lesions of the rat ventral hippocampus result in hyperlocomotion and deficits in social behaviour in adulthood. *Psychopharmacology (Berl)* **132**: 303–310.
- Schneider M, Koch M (2004) Deficient social and play behavior in juvenile and adult rats after neonatal cortical lesion: Effects of chronic pubertal cannabinoid treatment. *Neuropsychopharmacology* **30**: 1–14.
- Schneider T, Labuz D, Przewocki R (2001) Nociceptive changes in rats after prenatal exposure to valproic acid. *Pol J Pharmacol* **53**: 531–534.
- Schultz R, Gauthier I, Klin A *et al.* (2000) Abnormal ventral temporal cortical activity during face discrimination among individuals with autism and Asperger syndrome. *Arch Gen Psychiatry* **57**: 331–340.
- Schumacher H, Terapane J, Jordan RL, Wilson JG (1972) The teratogenic activity of a thalidomide analogue, EM 12 in rabbits, rats, and monkeys. *Teratology* **5**: 233–240.
- Schumann C, Hamstra J, Goodlin-Jones BL *et al.* (2004) The amygdala is enlarged in children but not adolescents with autism; the hippocampus is enlarged at all adults. *J Neurosci* **24**: 6392–6401.
- Sparks B, Friedman SD, Shaw DW *et al.* (2002) Brain structural abnormalities in young children with autism spectrum disorder. *Neurology* **59**: 184–192.
- Stokstad E (2001) New hints into the biological basis of autism. *Science* **294**: 34–37.
- Stromland K, Nordin V, Miller M, Akerstrom B, Gillberg C (1994) Autism in thalidomide embryopathy: A population study. *Dev Med Child Neurol* **36**: 351–356.
- Taieb O, Baleyte JM, Mazet P, Fillet AM (2001) Borna disease virus and psychiatry. *Eur Psychiatry* **16**: 3–10.
- Tueting P, Costa E, Dwivedi Y *et al.* (1999) The phenotypic characteristics of heterozygous reeler mouse. *Neuroreport* **10**: 1329–1334.
- Van Middlesworth L, Norris CH (1980) Audiogenic seizures and cochlear damage in rats after perinatal antithyroid treatment. *Endocrinology* **106**: 1686–1690.
- Vorhees C, Weisenburger WP, Minck DR (2001) Neurobehavioral teratogenic effects of thalidomide in rats. *Neurotoxicol Teratol* **23**: 255–264.
- Winslow J, Insel TR (2002) The social deficits of the oxytocin knockout mouse. *Neuropeptides* **36**: 221–229.
- Wolterink G, Daenen LEWPM, Dubbeldam S *et al.* (2001) Early amygdala damage in the rat as a model for neurodevelopmental psychopathological disorders. *Eur Neuropsychopharmacol* **11**: 51–59.
- Wood G, Lipska BK, Weinberger DR (1997) Behavioral changes in rats with early ventral hippocampal damage vary with age at damage. *Brain Res Dev Brain Res* **101**: 17–25.
- Yasuda S, Ishida N, Higashiyama A, Morinobu S, Kato N (2000) Characterization of audiogenic-like seizures in naive rats evoked by activation of AMPA and NMDA receptors in the inferior colliculus. *Exp Neurol* **164**: 396–406.
- Young L (2001) Oxytocin and vasopressin as candidate genes for psychiatric disorders: Lessons from animal models. *Am J Med Genet* **105**: 53–54.
- Zhang K, Liu X, Zhang C *et al.* (2002) Reelin gene alleles and susceptibility to autism spectrum disorders. *Mol Psychiatry* **7**: 1012–1017.



Tachykinin 1 (TAC1) gene SNPs and haplotypes with autism: A case-control study

Tetsuya Marui ^{a,*}, Ikuko Funatogawa ^b, Shinko Koishi ^c, Kenji Yamamoto ^d,
Hideo Matsumoto ^e, Ohiko Hashimoto ^f, Eiji Nanba ^g, Hisami Nishida ^h,
Toshiro Sugiyama ^c, Kiyoto Kasai ^a, Keiichiro Watanabe ⁱ, Yukiko Kano ⁱ,
Nobumasa Kato ^a, Tsukasa Sasaki ^{a,j}

^a Department of Neuropsychiatry, Graduate School of Medicine, University of Tokyo, Tokyo, Japan

^b Department of Hygiene and Public Health, Teikyo University School of Medicine, Tokyo, Japan

^c Aichi Children's Health and Medical Center, Obu, Japan

^d Department of Psychiatry, Kitasato University School of Medicine, Sagami-hara, Japan

^e Department of Psychiatry, Tokai University School of Medicine, Isehara, Japan

^f Department of Occupational Therapy, Faculty of Nursing and Rehabilitation, Aino University, Ibaraki, Japan

^g Gene Research Center, Tottori University, Yonago, Japan

^h Asunaro Hospital for Child and Adolescent Psychiatry, Tsu, Japan

ⁱ Department of Child Psychiatry, School of Medicine, University of Tokyo, Tokyo, Japan

^j Department of Psychiatry, Health Service Center, University of Tokyo, Tokyo, Japan

Received 18 July 2006; received in revised form 10 January 2007; accepted 24 January 2007

Abstract

Autism (MIM 209850) is a severe neurodevelopmental disorder characterized by disturbances in social interaction and communication, by repetitive body movements and restricted interests, and by atypical language development. Several twin and family studies have shown strong evidence for genetic factors in the etiology of autism. Glutamate is a major excitatory neurotransmitter in the human brain. Glutamate systems are involved in the pathophysiology of autism. There are many similarities between the symptoms evoked by glutamate antagonist treatment and symptoms of autism found in several human and animal studies. To elucidate the genetic background of autism, we analyzed the relationship between three single nucleotide polymorphisms (SNPs) of the Tachykinin 1 gene (TAC1) and autism, because TAC1 is located in the candidate region for autism and produces substance P and neurokinins. These products modulate glutamatergic excitatory synaptic transmission and are also involved in inflammation. Many different inflammation-related mechanisms could be involved in the autistic brain. Therefore, TAC1 may have some functions associated with the presumable pathophysiology of autism. We compared the allele and haplotype frequencies between autistic patients ($n = 170$) and normal controls ($n = 214$) in the Japanese population, but no significant difference was observed. Thus, the TAC1 locus is not likely to play a major role in the development of autism.

© 2007 Published by Elsevier B.V.

Keywords: Autistic disorder; Chromosome 7; TAC1; Genetic association; Glutamate system; Haplotype block

1. Introduction

Autism is a neurodevelopmental disorder characterized by early onset of the three cardinal symptoms: disturbance of social interaction, atypical language

* Corresponding author. Tel.: +81 3 5800 9263; fax: +81 42 379 4544.

E-mail address: PXX03135@nifty.ne.jp (T. Marui).

40 development, and restricted, repetitive, stereotyped pat-
 41 terns of behavior and interests [1]. The genetic involve-
 42 ment of autism has been supported by its higher
 43 concordance rates for monozygotic twins than for dizy-
 44 gotic [2,3] and the several chromosomal loci possibly
 45 linked to autism, including 7q22-q31 [1].

46 Glutamate is one of the neurochemicals speculated
 47 to contribute to the pathogenesis of autism, because
 48 (1) there are many similarities between the symptoms
 49 evoked by glutamate antagonist treatment and the
 50 symptoms of autism found in several human and
 51 animal studies [6]; (2) several neuropathological and
 52 brain-imaging studies of autistic patients have shown
 53 involvement of the cerebral regions where glutamater-
 54 gic neurons originate [6]; (3) the mRNA levels of the
 55 α -amino-3-hydroxy-5-methyl-4-isoxazole propionic
 56 acid (AMPA) receptor increases in autistic subjects
 57 [7]; (4) the aspartate/glutamate carrier SLC25A12
 58 gene and metabotropic glutamate receptor 8 gene
 59 have been reported to be associated with autism
 60 [8,9].

61 In this study, we focused on the Tachykinin 1 gene
 62 (TAC1; MIM162320) because it is located in 7q21-q22
 63 and encodes a precursor containing substance P and
 64 other neurokinins (Neurokinin A, Neurokinin K,
 65 and Neuropeptide γ) [10], some of which are impli-
 66 cated in the modulation of glutamate-driven neuro-
 67 transmission and excitotoxicity in the basal forebrain
 68 and other CNS regions [11]. Moreover, TAC1 mutant
 69 mice are less sensitive to nociceptive stimulation,
 70 which reminds us of some autistic patients ignorant
 71 of pain [6,13]. An increase in proinflammatory cyto-
 72 kines and the activation of microglia and astrocytes
 73 in the brain of autistic patients may be also associated
 74 with the inflammatory responses of TAC1 products
 75 [14].

76 Based on TAC1 biological information and its chro-
 77 mosomal location, we considered the gene worth analyz-
 78 ing as one of the autism candidate genes. In the present
 79 study, we tested for the presence of an association of
 80 three single nucleotide polymorphisms (SNPs) of
 81 TAC1, and haplotypes consisting of the SNPs, with aut-
 82 ism, using case-control design.

2. Subjects and methods

83

84 The patients comprised 170 unrelated Japanese with
 85 autism (147 males and 23 females, mean age = 20.8 years
 86 within a range of 3–41 years). The patients were recruited
 87 from the outpatient clinics of the departments of psychia-
 88 try, Tokyo University Hospital and Tokai University
 89 Hospital, and seven daycare facilities for subjects with
 90 developmental disorders. All the hospitals and facilities
 91 were located around Tokyo. All the subjects met the
 92 DSM-IV criteria for autistic disorder. The diagnoses were
 93 made by one or two experienced child psychiatrists
 94 through interviews and reviews of clinical records. Appar-
 95 ent physical anomalies were not observed in the subjects.
 96 The controls consisted of 214 unrelated Japanese healthy
 97 volunteers (145 males and 69 females, mean age = 34.6
 98 years within a range of 21–65 years). They were mainly
 99 recruited from the hospital and facility staff, and all of
 100 them resided in the same area (Kanto District or around
 101 Tokyo) as the patients. All the patients and controls were
 102 ethnically Japanese, with no parents or grandparents of
 103 ethnicity other than Japanese.

104 Confirmation of the diagnosis was conducted as fol-
 105 lows. Semi-structured behavior-observation of the
 106 patients and interviews with them and their parents were
 107 conducted for most of the cases by two experienced
 108 child psychiatrists independently. At the interview with
 109 the parent(s), the Child Behavior Questionnaire Revised
 110 (CBQ-R) [12] was used to assist evaluation of autism-
 111 specific behavior and symptoms. After the initial obser-
 112 vation and interview, we followed up by examining the
 113 patients' behavior and symptoms for several months
 114 (for at least 6 months in most of the cases) and those
 115 who were not considered to meet the DSM-IV criteria
 116 during the follow-up were excluded from the sample.

117 The present study was approved by the Ethical Com-
 118 mittee, the Faculty of Medicine of the University of
 119 Tokyo and Tokai University. The objective of the present
 120 study was clearly expressed and written informed consent
 121 was obtained from all subjects and healthy volunteers.

122 Peripheral blood was obtained and genomic DNA
 123 was extracted using the standard phenol-chloroform
 124 method. Single nucleotide polymorphisms (SNPs) were

Table 1
 Allele frequencies of 3 SNPs of the TAC1 gene in autism patients and controls

Locus	db SNP ID	Allele A/B ^a	Minor allele frequencies				χ^2	P-value	Odds ratio	95% confidence intervals		Location
			Patients		Controls					Lower	Upper	
			N	%	N	%						
SNP 1	rs2072100	C/T	107	31.8	136	32.2	0.01	0.91	1.02	0.75	1.38	int01
SNP 2	rs1229434	C/T	149	44.3	199	47.2	0.60	0.44	1.12	0.84	1.49	int06
SNP 3	rs1397202	A/G	68	20.1	82	19.2	0.09	0.76	0.95	0.66	1.35	int06

^a Alleles "B" are minor alleles.

Please cite this article in press as: Marui T et al., Tachykinin 1 (TAC1) gene SNPs and haplotypes with autism: ..., Brain Dev (2007), doi:10.1016/j.braindev.2007.01.010

125 analyzed using the ABI 7900HT sequence detection sys-
 126 tem (Applied Biosystems, Foster City, CA, USA). Three
 127 SNPs of the TAC1 gene were selected from the list of
 128 Assays-on-Demand™ Products for the ABI 7900HT to
 129 cover the full length of the gene, considering minor allele
 130 frequencies indicated in the products list. The db SNP
 131 IDs of the SNPs are shown in Table 1.

132 Statistical analyses were performed using the SAS/
 133 Genetics 9.1 software (SAS Institute Inc., Cary, North
 134 Carolina, USA). The frequencies of alleles and geno-
 135 types of each SNP were compared between patients
 136 and controls using the χ^2 test. To further analyze the
 137 haplotype structure in our sample, we computed the
 138 pair-wise D' values that indicated the linkage disequilib-
 139 rium (LD) between the SNPs. The frequencies of haplo-
 140 types consisting of SNPs, which were at a high linkage
 141 disequilibrium (haplotype block), were estimated. The
 142 exact P -values based on the likelihood ratio test with
 143 10,000 permutations were calculated for comparison of
 144 the haplotype frequencies between patients and controls.

145 **3. Results**

146 The allele frequencies of the SNPs of the TAC1 gene
 147 are summarized in Table 1. No significant difference was
 148 observed in genotypic distributions (not shown in the
 149 tables) or allele frequencies of the three markers of the
 150 TAC1 gene between patients and controls. The minor
 151 allele frequencies of the SNPs were higher than 19% in
 152 all three SNPs. For all assayed SNPs, none of the SNPs
 153 deviated from Hardy–Weinberg equilibrium.

154 The pair-wise D' and r^2 values of the SNPs within the
 155 TAC1 gene are summarized in Table 2. Considering r^2 ,
 156 these three SNPs within the TAC1 gene were not in
 157 complete linkage disequilibrium with each other. How-
 158 ever, all the pair-wise D' values of each SNP indicated
 159 1.00, forming a haplotype block. Regarding the pair-
 160 wise D' values, therefore, three marker haplotypes of
 161 SNP1-3 (rs2072100, rs1229434, rs1397202), were tested.

162 The estimated haplotype frequencies are shown in
 163 Table 3. No significant difference was observed in the

Table 2
 The strength of LD (denoted as D' and r^2) between pairs of SNPs of TAC1 in autism patients (the lower diagonal) and controls (the upper diagonal)

SNP	1	2	3
D'			
1		1.00	1.00
2	1.00		1.00
3	1.00	1.00	
r^2			
1		0.53	0.11
2	0.60		0.22
3	0.12	0.20	

Table 3
 Estimated frequencies and permutation P -values for association of major TAC1 haplotypes with rs2072100-rs1229434-rs1397202

Haplotype	Frequencies		χ^2	P -value
	Case	Control		
C-C-A	0.36	0.33	0.42	0.53
T-T-A	0.32	0.32	0.00	0.95
C-C-G	0.20	0.19	0.09	0.78
C-T-A	0.12	0.15	1.33	0.25

Global $P = 0.6958$

Haplotype frequencies were estimated >1%.

164 estimated haplotype distributions between patients and
 165 controls.

166 **4. Discussion**

167 We genotyped three SNPs within the TAC1 gene. No
 168 association was found in the allele and haplotype distri-
 169 butions between autistic patients and normal controls.

170 The 95% confidence intervals of the odds ratios were
 171 within 0.66 and 1.49 in all three SNPs of the gene. Con-
 172 sidering the sample size and minor allele frequencies in
 173 the present study, our results might have adequate sta-
 174 tistical power (>0.8) to contradict the effects of the gene
 175 with odds ratios of approximately 1.7 or more.

176 The TAC1 gene is located on 7q21.3 within the can-
 177 didate region for autism and has 7 exons spanning
 178 approximately 8.4 kb of genomic DNA [13]. Neurokinin
 179 A, neuropeptide K, and neuropeptide γ , as well as Sub-
 180 stance P, are produced from the TAC1 gene as a result
 181 of differential splicing and posttranslational processing
 182 [14] (Fig. 1). Substance P and other neurokinins are
 183 known as a mediators of pain and inflammation. They
 184 are produced in nociceptive primary sensory neurons
 185 and in many brain regions involved in pain signaling.
 186 Pain behavior evoked by thermal, mechanical, and
 187 chemical stimulation of somatic and visceral tissues were
 188 all reduced in the TAC1 mutant mice [8].

189 Besides pain and inflammation, the concentration of
 190 Substance P was elevated in the cerebrospinal fluid of
 191 depressed patients, and in the brain of a rat model of
 192 depression. Limbic structures contain a high density of
 193 Substance P terminals and Neurokinin 1 receptor sites.
 194 The tachykinin system therefore may play an important
 195 role in the regulation of emotional states and the devel-
 196 opment of anxiety disorders and depression [10].

197 In the Rett syndrome included in pervasive develop-
 198 mental disorders (PDD) with autistic symptoms,
 199 reduced expression of the TAC1 gene products was
 200 reported [15,16]. This fact may suggest that the TAC1
 201 gene might not be implicated in the whole spectrum of
 202 PDD.

203 Nonetheless, several facts, including decreased pain
 204 sensitivity, brain inflammation, and association of the

Please cite this article in press as: Marui T et al., Tachykinin 1 (TAC1) gene SNPs and haplotypes with autism: ..., Brain Dev (2007), doi:10.1016/j.braindev.2007.01.010

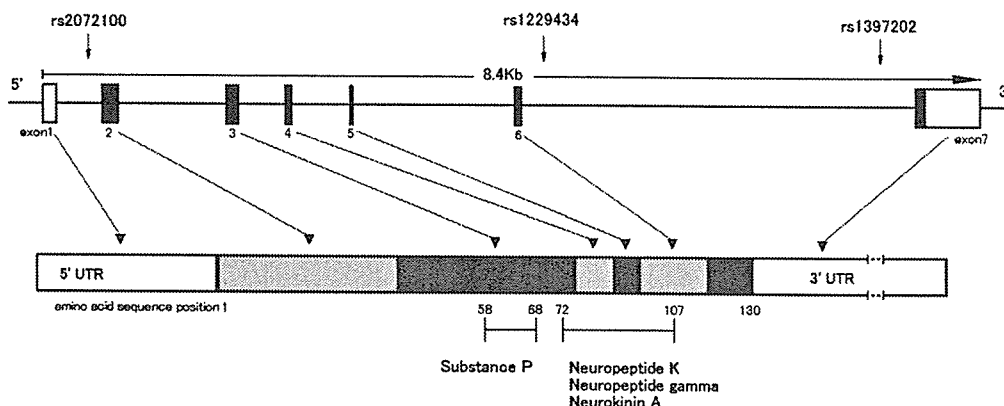


Fig. 1. This figure shows the TAC1 gene structure and positions of the SNPs (presented from 5' to 3' of the gene). All SNPs are denoted numerically with reference to Table 1. The vertical black bars indicate exons. Open boxes indicate the 5' and 3' untranslated regions. A horizontal line between the exons depicts introns. The TAC1 gene product is also indicated. Neurokinins are produced as a result of differential splicing and posttranslational processing of the product.

205 TAC1 gene with the glutamate system, suggested that
 206 Substance P or other neurokinins, which are products
 207 of the TAC1 gene, might contribute to the etiology of
 208 a part of autism. Although our data denied the associa-
 209 tion of the TAC1 gene with autism, these facts possibly
 210 support the necessity for this study.

211 The controls in this study were not age-matched to
 212 the patients. However, this is not likely to affect the
 213 result, considering the strong effect of genetic factors
 214 in autism compared with the rather small effect of envi-
 215 ronmental factors [2,17].

216 Another concern may be the population stratification
 217 of the sample, which could affect the results of studies in
 218 case-control design. This may not, however, significantly
 219 affect the present result, because the Japanese popula-
 220 tion is highly homogeneous, due to no major immigra-
 221 tion for more than a thousand years, compared with
 222 the European or North American population. No sub-
 223 jects in this study had parents or grandparents of ethnic-
 224 ity other than Japanese.

225 In conclusion, we analyzed three SNPs in the TAC1
 226 gene to determine whether there exists an association
 227 of TAC1 with autism using case-control design. To
 228 our knowledge, this is the first study that has examined
 229 the role of TAC1 polymorphisms in autism. No associ-
 230 ation was observed in this study.

231 **5. Uncited references**

232 [4,5].

233 **References**

234 [1] Folstein SE, Rosen-Sheidley B. Genetics of autism: complex
 235 aetiology for a heterogeneous disorder. *Nat Rev Genet*
 236 2001;2:943–55.

[2] Bailey A, Le Couteur A, Gottesman I, Bolton P, Simonoff E, Yuzda E, et al. Autism as a strongly genetic disorder: evidence from a British twin study. *Psychol Med* 1995;25:63–77. 237
 238
 239
 [3] Steffenburg S, Gillberg C, Hellgren L, Andersson L, Gillberg IC, Jakobsson G, et al. A twin study of autism in Denmark, Finland, Iceland, Norway and Sweden. *J Child Psychol Psychiatry* 1989;30:405–16. 240
 241
 242
 243
 [4] Carlsson ML. Hypothesis: is infantile autism a hypoglutamatergic disorder? Relevance of glutamate–serotonin interactions for pharmacotherapy. *J Neural Transm* 1998;105:525–35. 244
 245
 246
 [5] Purcell AE, Jeon OH, Zimmerman AW, Blue ME, Pevsner J. Postmortem brain abnormalities of the glutamate neurotransmitter system in autism. *Neurology* 2001;57:1618–28. 247
 248
 249
 [6] Ramoz N, Reichert JG, Smith CJ, Silverman JM, Beshalova IN, Davis KL, et al. Linkage and association of the mitochondrial aspartate/glutamate carrier SLC25A12 gene with autism. *Am J Psychiatry* 2004;161:662–9. 250
 251
 252
 253
 [7] Serajee FJ, Zhong H, Nabi R, Huq AH. The metabotropic glutamate receptor 8 gene at 7q31: partial duplication and possible association with autism. *J Med Genet* 2003;40:e42. 254
 255
 256
 [8] Cao YQ, Mantyh PW, Carlson EJ, Gillespie AM, Epstein CJ, Basbaum AI. Primary afferent tachykinins are required to experience moderate to intense pain. *Nature* 1998;392:390–4. 257
 258
 259
 [9] Zhang JP, Wei LC, Cao R, Chen LW. Differential co-expression of AMPA receptor subunits in substance P receptor-containing neurons of basal forebrain regions of C57/BL mice. *Neurochem Int* 2006;49:319–26. 260
 261
 262
 263
 [10] Bilkei-Gorzo A, Racz I, Michel K, Zimmer A. Diminished anxiety- and depression-related behaviors in mice with selective deletion of the Tac1 gene. *J Neurosci* 2002;22:10046–52. 264
 265
 266
 [11] Vargas DL, Nascimbene C, Krishnan C, Zimmerman AW, Pardo CA. Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann Neurol* 2005;57:67–81. 267
 268
 269
 [12] Izutsu T, Osada H, Tachimori H, Naganuma Y, Kato S, Kurita H. The usefulness of the child behavior questionnaire revised (CBQ-R) as a supplementary scale for diagnosis of pervasive developmental disorders. *Rinsyo-Seishin Igaku* 2001;30:525–32, in Japanese. 270
 271
 272
 273
 274
 [13] Ogden CA, Rich ME, Schork NJ, Paulus MP, Geyer MA, Lohr JB, et al. Candidate genes, pathways and mechanisms for bipolar (manic-depressive) and related disorders: an expanded convergent functional genomics approach. *Mol Psychiatry* 2004;9:1007–29. 275
 276
 277
 278
 [14] Krause JE, Chirgwin JM, Carter MS, Xu ZS, Hershey AD. Three rat preprotachykinin mRNAs encode the neuropeptides substance P and neurokinin A. *Proc Natl Acad Sci USA* 1987;84:881–5. 279
 280
 281

Please cite this article in press as: Marui T et al., Tachykinin 1 (TAC1) gene SNPs and haplotypes with autism: ..., *Brain Dev* (2007), doi:10.1016/j.braindev.2007.01.010

282	[15] Deguchi K, Antalffy BA, Twohill LJ, Chakraborty S, Glaze DG,	substance P in patients with Rett syndrome. <i>Ann Neurol</i>	287
283	Armstrong DD, et al. immunoreactivity in Rett syndrome.	1997;42:978–81.	288
284	<i>Pediatr Neurol</i> 2000;22:259–66.	[17] Szatmari P, Jones MB, Zwaigenbaum L, MacLean JE. Genetics	289
285	[16] Matsuishi T, Nagamitsu S, Yamashita Y, Murakami Y, Kimura	of autism: overview and new directions. <i>J Autism Dev Disord</i>	290
286	A, Sakai T, et al. Decreased cerebrospinal fluid levels of	1998;28:351–68.	291
			292

Please cite this article in press as: Marui T et al., Tachykinin 1 (TAC1) gene SNPs and haplotypes with autism: ..., *Brain Dev* (2007), doi:10.1016/j.braindev.2007.01.010

Revision of PNP-D-06-00190R2

Submitted to *Progress in Neuro-Psychopharmacology & Biological Psychiatry*
as an Original article.

This work has not been published and is not under review with another journal.

No association between the Neuronal Pentraxin II gene polymorphism and autism.

Tetsuya Marui,^{a*} Shinko Koishi,^b Ikuko Funatogawa,^c Kenji Yamamoto,^d
Hideo Matsumoto,^e Ohiko Hashimoto,^f Michiko Ishijima,^a Eiji Nanba,^g
Hisami Nishida,^h Toshiro Sugiyama,^b Kiyoto Kasai,^a Keiichiro Watanabe,^a
Yukiko Kano,^a Nobumasa Kato,^a Tsukasa Sasaki,^{a,i}

^aDepartment of Neuropsychiatry, Graduate School of Medicine, University of Tokyo,
Tokyo, Japan

^bAichi Children's Health and Medical Center, Obu, Japan

^cDepartment of Hygiene and Public Health, Teikyo University School of Medicine,
Tokyo, Japan

^dDepartment of Psychiatry, Kitasato University School of Medicine, Sagamihara, Japan

^eDepartment of Psychiatry, Tokai University School of Medicine, Isehara, Japan

^fDepartment of Occupational Therapy, Faculty of Nursing and Rehabilitation, Aino
University, Ibaraki, Japan

^gGene Research Center, Tottori University, Yonago, Japan

^hAsunaro Hospital for Child and Adolescent Psychiatry, Tsu, Japan

ⁱDepartment of Psychiatry, Health Service Center, University of Tokyo, Tokyo, Japan

Correspondence to:

Tetsuya Marui, MD,

Department of Neuropsychiatry, School of Medicine, University of Tokyo

7-3-1 Hongo, Bunkyo, Tokyo 113, Japan

Tel: +81-3-5800-9263

Fax: +81-42-379-4544

E-mail: PXX03135@nifty.ne.jp