significant insights into RA pathogenesis.[1,2] The first samples tested were synovial specimens,[3–8] and peripheral blood mononuclear cells (PBMC),[9] from RA and osteoarthritis (OA) patients, and cluster analysis of the resulting microarray gene-expression data revealed some candidate genes that may play a specific role in RA pathogenesis.

In other studies searching for key factors in RA pathogenesis, immunoscreening by using an antirheumatoid synovial cell antibody identified synoviolin/Hrd1 to be a highly expressed enzyme (E3 ubiquitin ligase) in the rheumatoid synovium.[10] Synoviolin appears to be a pathogenic factor for RA because mice overexpressing this enzyme developed spontaneous arthropathy, while heterozygous knockdown results in increased synovial cell apoptosis and resistance to collagen-induced arthritis.[11] It was proposed that the excess elimination of unfolded proteins due to synoviolin overexpression triggers synovial cell overgrowth.[12] Thus, synoviolin may play a pivotal role in the pathogenesis of arthropathy due to its functions in the quality control of proteins through the endoplasmic reticulum (ER)-associated degradation (ERAD) system; its elevated expression may therefore have an anti-apoptotic effect that causes synovial hyperplasia.

Bone marrow-derived mononuclear cells (BMMC) are another target for analyses aiming to identify the key genes that participate in RA pathogenesis because accumulating evidence suggests that BMMC cell abnormalities may contribute to the pathogenesis of RA and experimental arthritis models.[13–17] Moreover, RA patients suffer from defective central and peripheral B-cell tolerance checkpoints,[18] the first of which occurs in the bone marrow between the early immature and immature B-cell stages (the second counter selection step of autoantibody-expressing B cells takes place in the periphery, when the new emigrant becomes a mature naive B cell).[18,19] In addition, inflammatory changes similar to those found in RA synovium seem to occur in the subchondral bone marrow of the involved RA joint,[20] and synovial inflammatory tissue can reach the adjacent bone marrow by fully breaking the cortical barrier.[21] Thus, BMMC cells are an interesting subject for studies seeking to identify specific genes involved in RA pathogenesis.

To identify the genes whose expressions are dramatically induced or reduced in the pooled BMMC mRNAs of 50 RA patients as compared with 50 OA patients, we here subjected these pooled mRNAs to stepwise subtraction, which is a unique technique that we have developed previously.[22] This method permitted the comprehensive identification of those genes that are specifically up- or down-regulated during RA pathogenesis. In addition, we also used microarray analysis, since DNA microarray analyses on the BMMC of RA patients have not been described previously. As a control, we also subjected the BMMC RNA from OA patients to stepwise subtraction and microarray analysis to identify the genes that are specifically involved in OA pathogenesis. These analyses together resulted in the isolation of 103 RA-upregulated genes, of which amphiregulin (AREG) was revealed by quantitative real-time RT–PCR (QRT–PCR) to be the most conspicuously induced gene in RA patients. Interestingly, we also show here that AREG operates upstream of synoviolin in isolated synovial cells through an epidermal growth factor receptor (EFGR) signaling pathway. We discuss how AREG upregulation could contribute to RA pathogenesis.

2. Patients, Materials and Methods

2.1. Human subjects and ethical considerations

All RA patients satisfied the 1987 revised diagnostic criteria of the American College of Rheumatology (ACR: formerly the American Rheumatism Association).[23] All OA patients fulfilled the ACR criteria for hip or knee OA.[24] The RA and OA patient groups were largely matched in terms of their average age and sex (Supplementary Figure S1A and B). This study was reviewed and approved by the Internal Review Board of the Research Institute for Microbial Diseases, Osaka University. Accordingly, a written informed consent was obtained from each participant before obtaining human tissues.

2.2. Cell proliferation assay

The synovial cells from each patient were seeded onto uncoated 35 mm tissue culture plates at 1×10^5 cells/well and cultured in 5% FBS/DMEM. After 12 h, the cells were incubated in fresh 5% FBS/DMEM with (100 ng/ml) or without AREG (Sigma-Aldrich, A 7080). Four photos were taken from fixed areas in four quadrants near the central area of each plate at the 0, 1, 3 and 4 day time points. The cells at each time point were counted from these four photos and expressed as mean ± standard error (SE).

2.3. Statistical analysis

Significant differences were determined using the Spearman's rank correlation (Supplemenatry Figure S4) or the Mann–Whitney *U*-test (Figs 2, 4 and Supplementary Figure S3). The data are expressed as means ± SE. $P < 0.05$ or $P < 0.01$ was considered to be statistically significant.

3. RESULTS

3.1. Identification of RA- or OA-specific genes by stepwise subtraction and DNA microarray analysis

To isolate the putative RA-specific genes that are upregulated in BMMC of RA patients relative to those

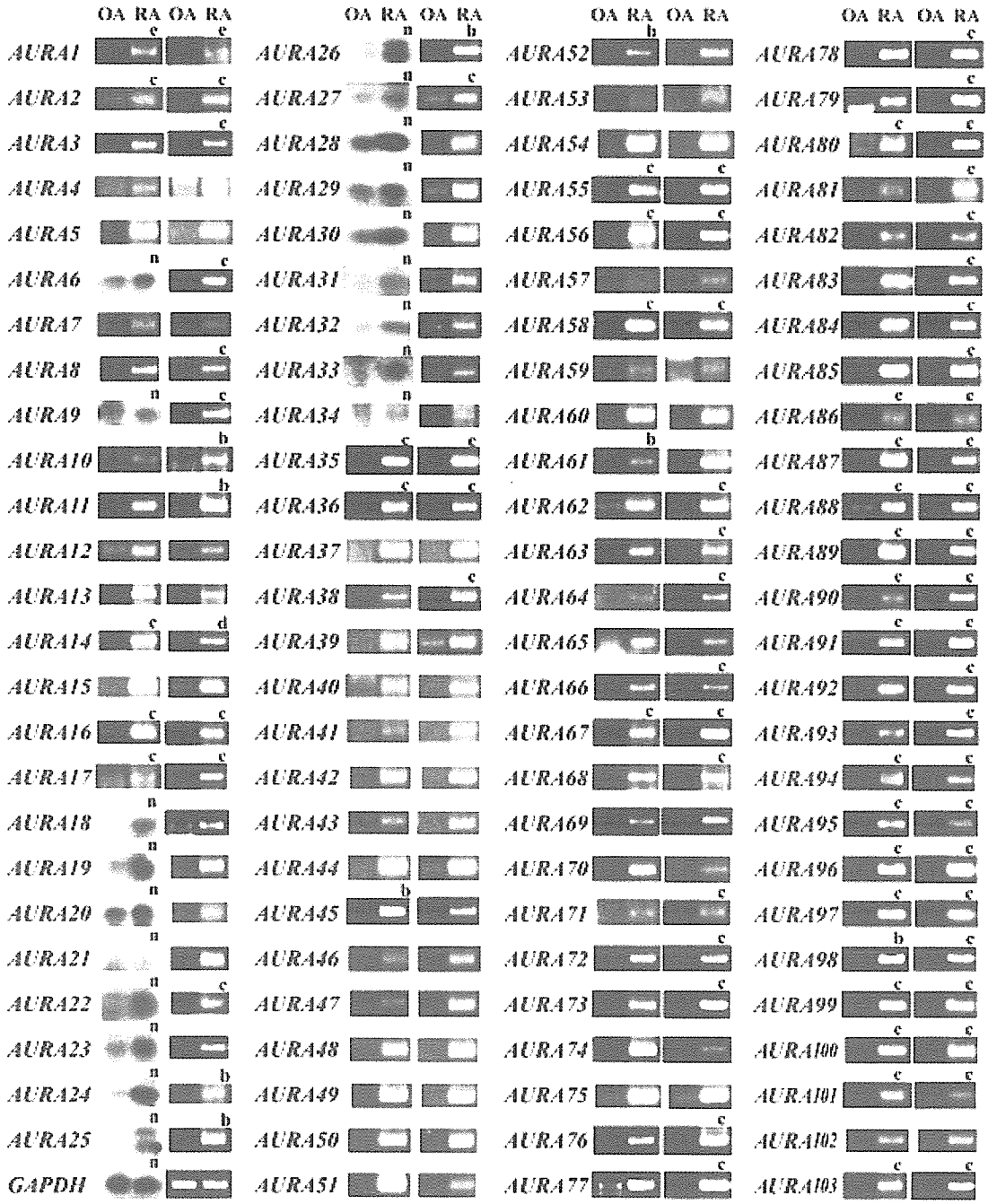


Figure 1. Northern blot or RT–PCR analysis of individual *AURA* cDNA clones to compare the expression levels of the genes in the BMMC of 50 RA patients and 50 OA patients (see Table 1 for their gene names). A northern blot or RT–PCR for *GAPDH* is also shown as a loading control. Left row: expression levels as detected by RT–PCR or northern blot analysis (denoted as n above each picture). Right row: confirmation of the expression level of each gene as determined by RT–PCR. The annealing temperature and amplification cycles for RT–PCR were always 50°C and 40 cycles, respectively, with the exception of the reactions denoted by a (50°C and 35 cycles, respectively), b (50°C and 30 cycles, respectively), c (55°C and 35 cycles, respectively), d (55°C and 40 cycles, respectively), and e (60°C and 40 cycles, respectively).

that are upregulated in OA patients, we first used our stepwise subtractive hybridization method. Briefly, we prepared a cDNA library from the pooled mRNA from the BMMC of 50 RA patients (Supplementary Figure S1A) by the linker-primer method using a pAP3neo vector.[25] Stepwise subtractive hybridization was then performed with the biotinylated pooled mRNA from the BMMC of 50 OA patients (Supplementary Figure S1A) to select candidate genes that may show upregulation in RA BMMC only as described previously.[22] To examine if the candidate genes are actually upregulated in RA but not OA BMMC, we performed northern blot analysis and/or RT–PCR using the pooled mRNA from the BMMC of 50 RA and 50 OA patients (Fig. 1). To reduce

the possibility of missing important RA-specific pathogenic genes by this method, we also performed a genome-wide complementary DNA microarray analysis using the Agilent Hu44K array with the same pooled RNA samples obtained from the BMMC of RA and OA patients that were described above. When we tested top 70 genes from the microarray list of RA-upregulated genes by northern blot analysis and/or RT–PCR as described above, we found that only 20 genes really displayed RA-upregulated expressions. Thus, we identified 103 RA-upregulated genes (Fig. 1) and named them *AURA* (*au*gmented in *RA*). As shown in Table 1, 15 *AURA* genes (*AURA1*~*AURA7* and *AURA10*~*AURA17*) are uncharacterized novel genes.

We also performed similar experiments to obtain candidate OA-upregulated genes by generating a cDNA library from the pooled mRNA from the BMMC of 50 OA patients (Supplementary Figure S1A) and then using biotinylated pooled mRNA from the BMMC of 50 RA patients for subtraction (Supplementary Figure S1A). DNA microarray analysis also yielded a number of candidate OA-specific genes, as described above. However, when we checked whether these candidate genes are truly specifically up-regulated in OA BMMCs by northern blot analysis and/or RT–PCR, we could confirm this for only two genes (Supplementary Figure S2). These two OA-upregulated genes encode nuclear receptor coactivator 1 and a hypothetical protein (FLJ20581). This result suggests that the gain of function due to the enhanced expression of the RA-upregulated candidate genes is important in the pathogenesis of RA. Thus, we subsequently concentrated our study on the RA-upregulated genes.

3.2. *Expression profiles of RA-upregulated genes in individual RA or OA patients*

To determine whether the upregulation of the 103 RA-specific candidate genes is widespread in many RA patients or occurs in only a few patients, we performed QRT–PCR using individually prepared RNA samples from the BMMC or PBMC of RA patients. Of the 103 candidate genes, 5 genes whose functions are unknown and 12 genes that may be related to growth regulation or immune response were analyzed by QRT–PCR. OA patients were also examined as negative controls. In every QRT–PCR, a standard RNA from the PBMC of a healthy volunteer (male, age 52) was used (denoted as normal with a relative intensity of 1.0). This allowed us to compare the expression profiles of the genes tested in this study. In addition, since we used this control, we could also compare the expression profiles of the genes in this study with those of other genes tested in our previous reports on other autoimmune diseases.[26]

Of the 17 tested *AURA* genes (denoted x in Table 1), AREG (*AURA9*) was the most conspicuously upregulated in the BMMC of many of the RA patients, while in contrast OA BMMCs invariably expressed this gene at very low levels (Fig. 2A). Similarly, the PBMC of many RA patients strongly expressed AREG, while only very low expression was detected in the PBMC of the OA patients (Fig. 2A). AREG is one of the EGF-like growth factors that stimulate cell growth by activating the EGF receptor (EGFR) signaling of the target cells in an autocrine/juxtacrine fashion.[27]

AURA1 was the next most conspicuously upregulated gene in the BMMC of many RA patients, while the BMMC of all OA patients showed only very low expression of this gene (Fig. 2B). However, unlike *AREG*, the PBMC of RA patients showed negligible enhancement in the expression of *AURA1*. *AURA1* encodes an uncharacterized protein containing a thioesterase domain (Fig. 2B inset) that may cleave thioester bonds of an unknown target.

The gene encoding FK506 (tacrolimus)-binding protein 5 (FKBP5 = *AURA45*) also showed enhanced expression in nearly half of the RA patient BMMC samples, while no such increase was observed in the OA patient BMMC samples or in the PBMC of the RA patients (Fig. 2C). FKBP5 is a cellular receptor for FK506 and has an immunosuppressive effect on activated T cells because it inhibits the protein phosphatase calcineurin.[28]

Nearly half of the RA patient BMMC samples showed 5- to 50-fold greater expression of *CLECSF9* (=*AURA24*), *TPST1* (=*AURA52*) and *AURA2* than the normal control PBMC sample (Fig. 2D–F). No such increase was observed in the BMMC of OA patients or in the PBMC of the RA patients. *CLECSF9* encodes a macrophage-inducible C-type lectin (Mincle) that harbors a calcium-dependent carbohydrate-recognition domain. *TPST1* is one of the two Golgi tyrosylprotein sulfotransferases (*TPST1* and *TPST2*) that mediate the post-translational modification tyrosine O-sulfation.

G0S2 (=*AURA8*), chemokine receptor 4 (CXCR4 = *AURA86*), nuclear factor-kappa B (NF-κB = *AURA25*) and *AURA17* showed augmented expression in both the BMMC and PBMC of some of the RA patients when compared to the expression in the BMMC and PBMC of the OA patients, although the differences between the RA and OA samples are not as significant as for the previously discussed genes (Supplementary Figure S3A–D). G0S2 is one of the G0/G1 switch (G0S) genes that are differentially expressed in lymphocytes during their lectin-induced switch from the G0 to the G1 phases of the cell cycle.[29] CXCR4, the receptor for a chemokine called stromal cell-derived factor-1 (SDF-1/CXCL12), is important in the migration, homing and survival of hematopoietic stem cells. SDF-1, which is secreted by ischemic myocardium, is involved in the homeostatic and inflammatory traffic of leukocytes, and is highly expressed in the synovial tissues of RA patients.[30] NF-κB

Table 1. List of *AURA* genes

AURA no.	Accession no.	Sequence description	SS/DM	QRT–PCR
AURA1	AK001968	Unknown cDNA (FLJ11106)	b	r
AURA2	BC022398	Unknown cDNA	b	r
AURA3	BC031341	Unknown cDNA (hypothetical protein MGC45871)		
AURA4	NM_052862.2	Unknown cDNA (hypothetical protein MGC21854)		
AURA5	AK097275.1	Unknown cDNA (FLJ39956) L-PLASTIN-like		
AURA6	BC019355	Unknown cDNA (ring finger protein 149: IMAGE:3956746)		
AURA7	AF078845.1	Unknown cDNA (16.7Kd protein)		
AURA8	M69199	Putative lymphocyte G0/G1 switch gene (G0S2)=Aile1	b	r
AURA9	AH002608	Amphiregulin	b	r
AURA10	AK026118	Unknown cDNA (Ch20-ORF43)		r
AURA11	AK094006	Unknown cDNA		
AURA12	AK095896.1	Unknown cDNA (FLJ38577)		
AURA13	BC014435	Unknown cDNA (IMAGE:4855747)		r
AURA14	ZF161365	Unknown cDNA (HSPC102)	m	
AURA15	FLJ23431	Unknown cDNA (FLJ23431) MHC class I -like		
AURA16	BC066334	Unknown cDNA (FLJ37760)		
AURA17	XM_058513	Unknown cDNA (DKFZp434H2111)	m	r
AURA18	BC016660	Heat shock 70 kDa protein 8		
AURA19	BC022347	Lactotransferrin		
AURA20	NM_001800.2	Cyclin-dependent kinase inhibitor 2D (p19) (CDKN2D)		
AURA21	X55668.1	Proteinase 3		
AURA22	BC013946	Kruppel-like factor 13		
AURA23	BC022463	Dual specificity phosphatase 1 (DUSP1)		r
AURA24	AY358499	C-type lectin, superfamily member 9 (CLECSF9)	b	r
AURA25	AY033600	NF-kB alpha	b	r
AURA26	AF194172	Androgen-regulated protein 6 (AIG6)	m	
AURA27	NM_021810	Cadherin-like 26 (CDH26)		
AURA28	X52053.1	HP-1 (corticostatin/defensin family)		r
AURA29	BC018857.2	Translation elongation factor 1 gamma		
AURA30	BC053585.1	Colony stimulating factor 3 receptor (granulocyte)		
AURA31	AY124010	Interleukin 1 receptor, type II (IL1R2)	m	
AURA32	BC020635	Ficolin 1 (FCN1: collagen/fibrinogen domain-containing)		
AURA33	BC106068	Microtubule-associated protein, RP/EB family, member 1		
AURA34	AF443591	Death effector domain-containing DNA binding protein2		
AURA35	BC032491	Ubiquitin-conjugating enzyme E2L 6 (UBE2L6)		
AURA36	BC004967	Ubiquitin associated domain containing 1 (UBADC1)		
AURA37	NM_006313.1	Ubiquitin specific protease 15 (USP15)		
AURA38	BC011358	ADP-ribosylation factor 1		
AURA39	AY366510.1	Pre-mRNA 3′end processing factor FIP1		
AURA40	NM_175039.1	Sialyltransferase 7D (SIAT7D). transcript variant 2		
AURA41	BC030230.2	Aminolevulinate, delta- synthase 2		
AURA42	NM_014390.1	Staphylococcal nuclease domain containing 1 (SND1)		
AURA43	NM_015999.2	Adiponectin receptor 1 (ADIPOR1)		
AURA44	BC033877.1	Finkel-Biskis-Reilly murine sarcoma virus (FBR-MuSV)		r
AURA45	NM_004117	FK506 binding protein 5 (FKBP5)	b	r
AURA46	NM_000211.1	Integrin beta 2 (antigen CD18 (p95)		
AURA47	BC015641.2	Enolase 1 (alpha)		

Table 1. continued.

AURA no.	Accession no.	Sequence description	SS/DM	QRT–PCR
AURA48	BC028299.1	Non-POU domain containing. octamer-binding.		
AURA49	BC000734.2	Eukaryotic translation initiation factor 3. subunit 648 kDa		
AURA50	NM_012198.2	Grancalcin. EF-hand calcium binding protein (GCA)		
AURA51	BC026690.2	CD97 antigen. transcript variant 2.		
AURA52	CR542060	Tyrosylprotein sulfotransferase 1 (TPST1)	m	r
AURA53	NM_005875.1	Translation factor sui1 homolog (GC20)		
AURA54	NM_004048.2	Beta-2-microglobulin (B2M)		
AURA55	BC017934	NudC domain containing 2 (NUDCD2)		
AURA56	NM_000569	Fc fragment of IgG, low affinity IIIa, receptor for (CD16)	b	
AURA57	BC018649.2	Polymerase (RNA) II (DNA directed)		
AURA58	BC013293	Synuclein, alpha (a molecular chaperone)		
AURA59	NM_033405.2	PRIC285		
AURA60	J02694.1	Myeloperoxidase		
AURA61	BC020219	Zinc finger protein 143 (clone pHZ-1)	m	
AURA62	BC071590	Nijmegen breakage syndrome 1 (nibrin)		
AURA63	BC003186	DNA replication complex GINS protein PSF2		r
AURA64	NM_006060	Zinc finger protein, subfamily 1A, 1 (ZNFN1A1)		
AURA65	BC015859	T-cell activation GTPase activating protein		
AURA66	Z50749	Sds22 (protein phosphatase regulatory subunit)-like		r
AURA67	AF411850	C-type lectin-like receptor CLEC-6	m	
AURA68	BC064831	HMT1 hnRNP methyltransferase-like 3		
AURA69	BC022797	Mof4 family associated protein 1		
AURA70	BC032437	Heterogeneous nuclear ribonucleoprotein A3		
AURA71	M87790	Anti-hepatitis A immunoglobulin lambda chain variable region		
AURA72	K01763	Haptoglobin alpha(1S)-beta precursor		
AURA73	BC016800	Aldolase A, fructose-bisphosphate, transcript variant		
AURA74	BC001391	Actin-like 6A, transcript variant 1		
AURA75	NM_003512.3	H2 histone, family 2AC (H2AC)		
AURA76	BC017558	H3 histone, family 3B (H3.3B)		
AURA77	BC032748	Myosin regulatory light chain MRCL3		
AURA78	S60099	APPH = amyloid precursor protein homolog		
AURA79	BC067100	Fas (TNFRSF6) associated factor 1		
AURA80	NM_000896	Cytochrome P450, family 4, subfamily F (CYP4F3)	b	
AURA81	BC010577	Granulin (an association partner of cyclin T1)		
AURA82	AF054186	p18		
AURA83	BC028626	Trinucleotide repeat containing 6B		
AURA84	L43631	Scaffold attachment factor B (SAF-B)		
AURA85	M11124	MHC HLA DQ alpha-chain mRNA from DRw9 cell line		
AURA86	AF025375	Chemokine (C-X-C motif) receptor 4 (CXCR4)	b	r
AURA87	BC000163	Vimentin (VIM)		
AURA88	BC071860	Lactate dehydrogenase B (LDHB)		
AURA89	BC100032	Ribosomal protein S13 (RPS13)		
AURA90	BC011852	Glutamine synthetase (GLUL)		
AURA91	NM_000045	Arginase, liver (ARG1)		
AURA92	BC006510	Cyclin B1		
AURA93	BC007063	Peroxiredoxin 1		
AURA94	NM_005746	Pre-B-cell colony enhancing factor 1 (PBEF1)	m	

Table 1. continued.

AURA no.	Accession no.	Sequence description	SS/DM	QRT–PCR
AURA95	BC018711	RNA-binding region (RNP1. RRM) containing 1		
AURA96	NM_001126	Adenylosuccinate synthase (ADSS)		
AURA97	BC008929	rab2 mRNA. YPT1-related and member of ras family		
AURA98	NM_004226	Serine/threonine kinase 17b (apoptosis-inducing) (STK17B)	m	
AURA99	BC096336	Insulin-degrading enzyme		
AURA100	AF501883	G protein Beta polypeptide 2 (GNB2)		
AURA101	BC007237	Myeloid/lymphoid or mixed-lineage leukemia		
AURA102	BC034149.1	Ribosomal protein S3		
AURA103	NM_020980	Aquaporin 9 (AQP9)	m	

Of 103 *AURA* genes, 83, 10 or 10 genes were identified by stepwise subtraction (SS) alone (no mark), by DNA microarray (DM) alone (denoted by m) or by both techniques (denoted by b), respectively. The *AURA* genes that were subjected to QRT–PCR analysis are denoted by r.
PRIC285: peroxisomal proliferator-activated receptor A interacting complex 285.

is a transcription factor that resides in the cytoplasm of every cell and translocates to the nucleus when activated by a wide variety of agents, including cytokines.[31] *AURA17* is an uncharacterized novel gene that encodes a large protein with 8 leucine rich repeats, Mitochondrial Rho (Miro) motif and protein tyrosine kinase domain (Supplementary Figure S3D inset).

We also tested seven other genes in RA and OA BMMC and PBMC samples by QRT–PCR, but none showed a widespread and conspicuous increase in expression in the RA BMMC samples (data not shown). Consequently, these genes appear to play a less significant role in RA pathogenesis. Since these experiments and those described above consumed almost all BMMC and PBMC samples from the RA and OA patients, the remaining *AURA* genes will have to be tested in the future with another RA patient set.

3.3. *Expression pattern of AURA genes in PBMC*

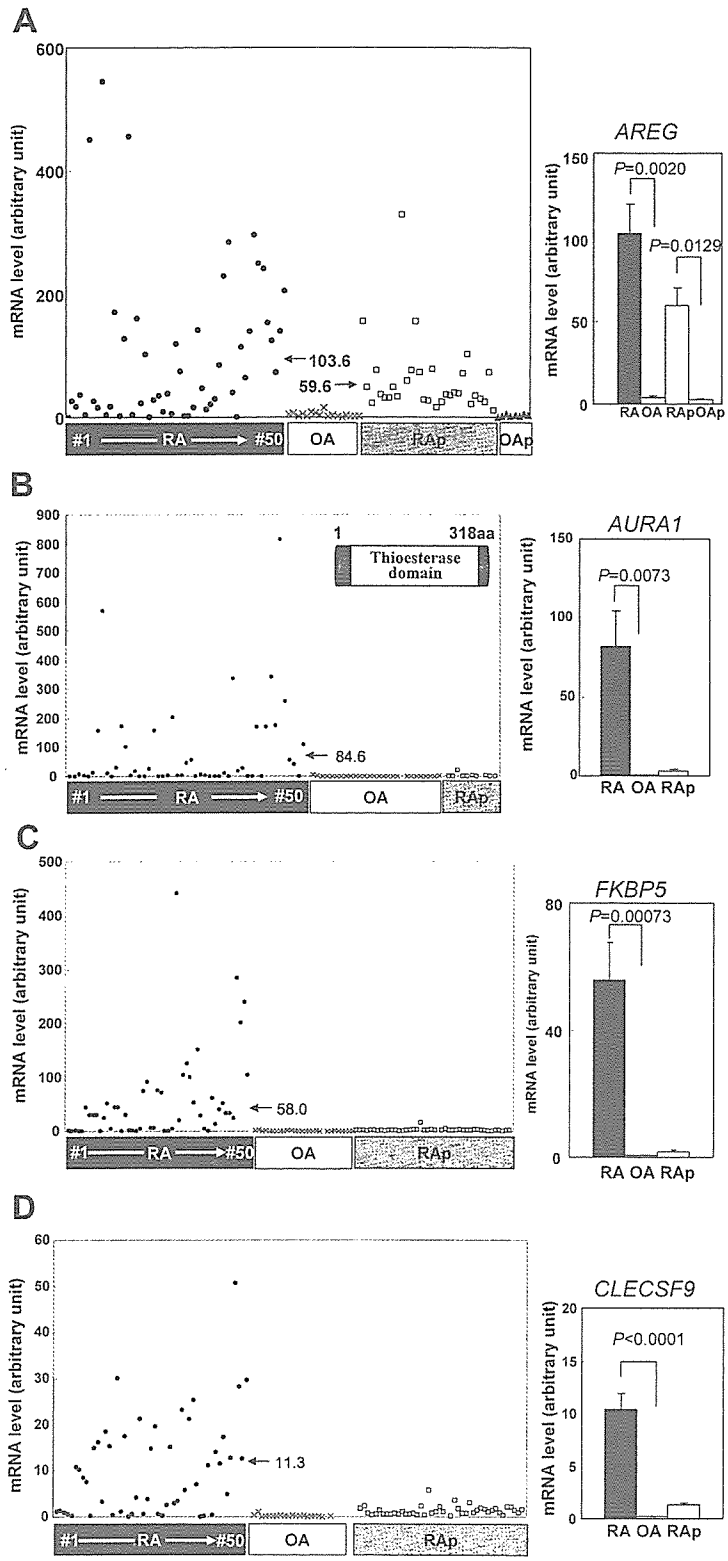
To determine whether the *AURA* genes are expressed in particular human blood cells, we performed RT–PCR on multiple tissue cDNA panels (MTC) from Clontech (Palo Alto, CA). As shown in Fig. 3, RT–PCR detected *AREG* mRNA in both monocytes (lane 4) and T and B cells (lanes 2–4), in particular in activated $CD4^+$ T cells (lane 8). *AURA1* is detected predominantly in resting $CD4^+$ (T helper/inducer; lane 3) and activated $CD4^+$ T (lane 8) cells. *CLECSF9* is expressed in most cell types except for activated $CD19^+$ T cells (lane 6), while *G0S2* is found primarily in monocytes (lanes a and 4). *FKBP5*, *TPST1*, *CXCR4*, *AURA2* and *NFκB* are ubiquitously expressed in most cell types. Thus, the analysis of the functions these *AURA* genes, apart from *AURA1* and *G0S2*, play in specific blood cells will not be easy because they are already expressed in normal blood. However, the function of *AURA1* can be studied by using $CD4^+$ T cells of RA and OA patients. In this study, however, we could not perform this analysis because of the low amounts of BMMC that we could obtain from the RA patients.

3.4. *AREG stimulates the growth of synovial cells*

Since AREG appears to be the most conspicuously unpregulated gene in many RA patients, we subjected it to further analysis. We first examined its ability to stimulate the growth of isolated synovial cells because AREG is one of the ligands of EGFR and is known to induce cell growth. Thus, we isolated synovial cells from synovial tissues that were obtained from five RA and three OA patients during joint reconstructive surgery. In the absence of AREG in the culture medium, the synovial cells from both the RA and OA patients grew at a similar rate (Fig. 4A and B). However, when AREG was present, the synovial cells from RA patients appeared to grow slightly faster than the synovial cells from OA patients, which is statistically significant ($P < 0.05$) (Fig. 4A).

To examine if this phenomenon is reflected in the signal transduction machinery of synovial cells, we investigated the activation of the EGFR signaling pathway in the AREG-treated and untreated RA synoviocytes. We first examined the phosphorylation of the extracellular signal-regulated kinases (ERK1/2) at Thr202 and Tyr204 by western blot analysis. ERK1/2 phosphorylation indicates the activation of the EGFR signaling pathway.[32] As shown in Fig. 5A, the phosphorylated ERK1/2 bands in the RA synoviocytes showed an increase in intensity when the cells had been treated with AREG; this effect peaked 8–12 h after AREG treatment but continued for 2–3 days. In contrast, the ERK1/2 protein levels remained largely unaffected by AREG treatment.

To compare the activation of EGFR signaling between RA and OA patients, we examined the activation of the EGFR signaling pathway in the synoviocytes from the five RA and three OA patients (Fig. 5B). We thus assessed the phosphorylated ERK1/2 expression levels by western blot analysis and expressed the results quantitatively by measuring the intensity of the lower phosphorylated band by densitometry and comparing it with the ERK1/2 band intensity (Fig. 5C). We found that the synoviocytes from the RA and OA patients expressed equivalent levels of EGFR and ERK1/2 proteins, regardless of AREG treatment. In contrast,



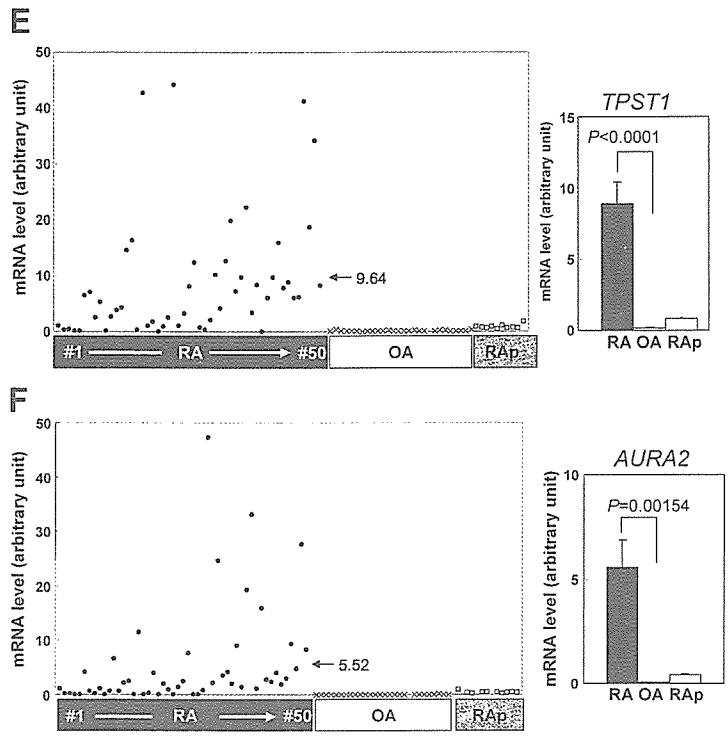


Figure 2. Expression levels of *AURA* genes in individual RA and OA patients. QRT–PCR analyses show that the mRNA levels of (A) *AREG*, (B) *AURA1*, (C) *FKBP5*, (D) *CLECSF9*, (E) *TPST1* and (F) *AURA2* are conspicuously upregulated in RA patient BMMC (and sometimes PBMC), while the BMMC and PBMC of OA patients show negligible upregulation. Expression levels in the BMMC for 50 RA patients (from #1 to 50) are arranged in the denoted order. The inset in (B) shows that the thioesterase domain occupies most of the Aura1 protein. The mean values of the samples analyzed in triplicate from each individual RA BMMC, RA PBMC, OA BMMC and OA PBMC are indicated by filled circles, open squares, x's, or filled triangles, respectively. The average values for the RA patient group are shown by the horizontal arrows. The bar graphs in the right panels show the average ± SE values of these measurements using the RA or OA BMMC or PBMC. All measurements are statistically significant when RA and OA are compared ($P < 0.01$).

AREG treatment upregulated the phosphorylated ERK1/2 expression levels much more strongly in the synoviocytes from RA2, RA3 and RA4 than in the synoviocytes of any of the OA patients. RA1 is an exception to this pattern as its limited phosphorylated ERK1/2 expression levels were similar to those in OA1–3. The AREG-induced upregulation of ERK1/2 phosphorylation was less apparent in the RA5 synovial cells because ERK1/2 was already activated in the absence of AREG.

Synoviolin plays a role in the synovial hyperplasia of RA by controlling the ERAD system.[10] To determine if the RA synovial cells have an abnormal ERAD system, we measured their levels of the ER stress proteins GRP78/BiP and GRP94, which protect cells from the stress-induced ER dysfunction that could lead to the accumulation of unfolded proteins.[33] We found that while the synovial cells of the RA and OA patients have similar levels of GRP78/BiP (Fig. 5B and D), the RA synoviocytes show enhanced levels of GRP94, irrespective of whether they have been stimulated with AREG. This suggests that at least part of the ER-stress responsive pathway, namely, that mediated by GRP94, is more activated in RA synoviocytes than in OA synoviocytes. Thus, the ERAD pathway does appear to be abnormally upregulated in RA synoviocytes. We confirmed by QRT–PCR that the BMMC and PBMC cells of RA patients RA1–5 show enhanced AREG mRNA levels, unlike the BMMC and PBMC of OA patients OA1–3 (Supplementary Figure S5A). Thus, chronic activation of AREG/EGFR signaling appears to be augmented in RA patients. Since AREG is expressed as transmembrane precursors that are cleaved in the extracellular domain to release soluble growth factor,[34] we speculated that the sera (PB) and bone marrow fluid (BM) of RA1–5 may show enhanced levels of cleaved AREG compared to the equivalent fluids of OA1–3. We tested this by enzyme-linked immunosorbent assay but found only one patient, RA2, showed levels of cleaved AREG that exceeded the detection level of the assay (Supplementary Figure S5B). Thus, it is not clear whether RA patients indeed secrete higher AREG levels than OA patients.

We also examined whether RA synoviocytes expressed higher synoviolin mRNA levels than OA synoviocytes in the presence or absence of AREG. However, we could not detect any significant differences between the RA and OA patients in this regard (Supplementary

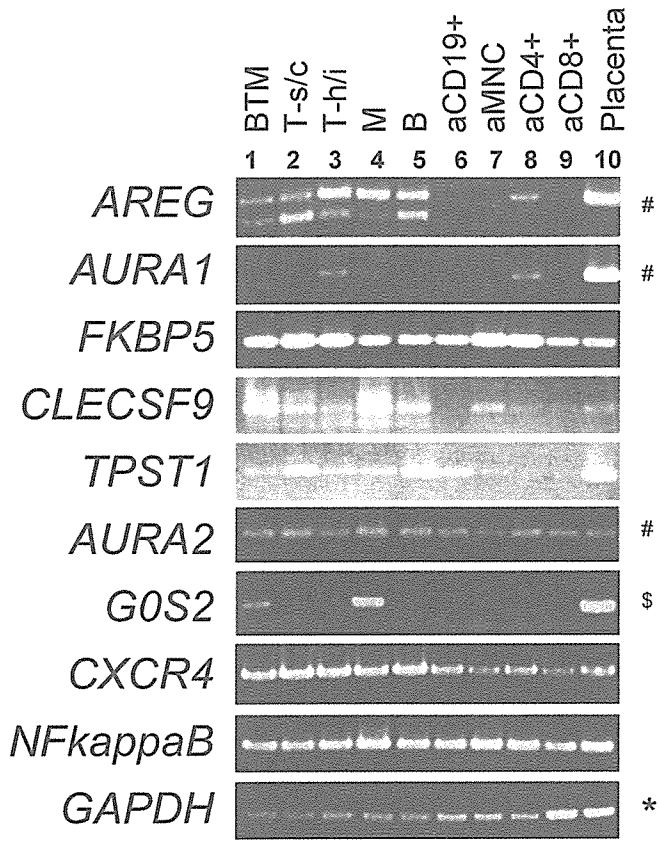


Figure 3. Determination by RT–PCR of the human blood cells that express *AREG*, *AURA1*, *FKBP5*, *CLECSF9*, *TPST1*, *AURA2*, *G0S2*, *CXCR4* and *NFκB*. RT–PCR was performed using the multiple tissue cDNA panel for human blood fractions (MTC, Clontech). *GAPDH* was also amplified as a loading control. PCR amplifications were conducted at 55°C and over 30 cycles except as indicated on the right of the panels: 55°C and 35 cycles (#), 55°C and 27 cycles (*) or 53°C and 25 cycles ($). Lane 1, mononuclear cells (B, T cells and monocytes). Lane 2, resting CD8+ cells (T-suppressor/cytotoxic cells). Lane 3, resting CD4+ cells (T-helper/inducer). Lane 4, resting CD14+ cells (monocytes). Lane 5, resting CD19+ cells (B cells). Lane 6, activated mononuclear cells. Lane 7, activated CD4+ cells. Lane 8, activated CD8+ cells. Lane 9, activated CD19+ cells. Lane 10, human placenta control cDNA served as a DNA size marker.

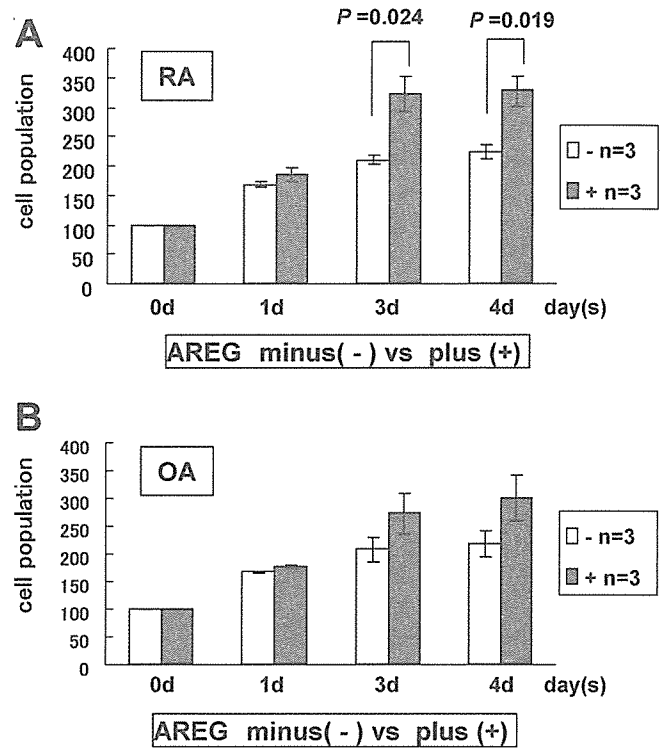


Figure 4. The effect of AREG on the proliferation of synoviocytes from RA and OA patients. The synovial cells from three RA patients (RA1, RA2 and RA3) (A) and three individual OA patients (OA1, OA2 and OA3) were counted on days 0, 1, 3 and 4 after incubation with or without AREG. The cell counts on days 1, 3 and 4 are expressed relative to 0 day. Statistically significant measurements are indicated ($P < 0.05$).

Figure S5C). It is not clear whether the synovial tissues of the patients would, like their cultured derivatives, show a similar lack of synoviolin upregulation.

4. Discussion

In this study, we report our comprehensive isolation of *AURA* genes that show augmented mRNA expression in the BMMC of RA patients as compared to their expression in OA patient BMMC (Fig. 1 and Table 1). Since RA patients suffer from defective central and peripheral B-cell tolerance checkpoints, and often display unusual immunoglobulin light chain repertoires that suggest impaired secondary recombination regulation,[18] we had expected that many immune response genes would be identified as *AURA* genes. Indeed, >10% of the *AURA* genes are directly related to immune responses; moreover, while the other *AURA* genes may seem at first glance to be unrelated to immune responses, many of these can also be linked to immune responses (Table 1). QRT–PCR analysis on individual patient samples revealed that the *AURA* genes discussed below are significantly increased in the BMMC of many of the 50 RA patients we tested (Fig. 2). Thus, the identification of these genes may help us to understand the pathogenesis of RA.

FKBP5, one of the cellular receptors for the immunosuppressant FK506, was expressed at higher mRNA levels in many RA patients than in the OA patients; this was true for the BMMC of the RA patients but not for their PBMC (Fig. 2C). FK506 has been suggested to be an effective drug for reducing the pain associated with RA.[35] This is because it can suppress inflammation by inhibiting the production by synovial cells of prostaglandin E2; it does so by suppressing the IL-1β production by leukocytes.[36] The enhanced FKBP5 expression in RA BMMC is not due to FK506 treatment since at the time of this study, treatment with FK506

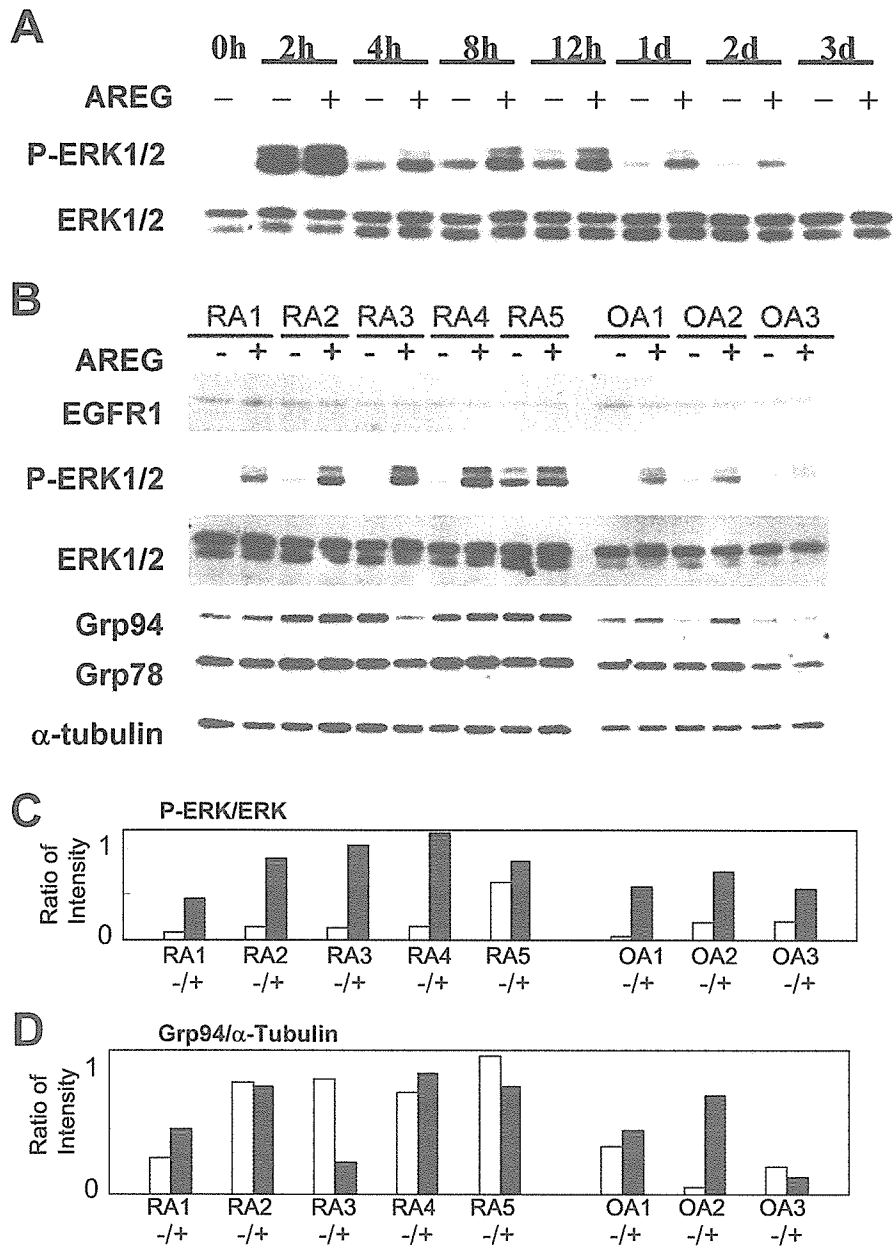


Figure 5. Western blot analysis of RA and OA synovial cells incubated in the presence or absence of AREG. (A) Expression levels of ERK1/2 and its phospho-form (P-ERK1/2) that is phosphorylated at Thr202 and Tyr204. Pooled synovial cells from five RA patients were incubated with (100 ng/ml) or without AREG for varying periods ranging from 0 h to 3 days. (B) Expression levels of EGFR1, ERK1/2, P-ERK1/2, Grp94, Grp78 and synoviolin in synovial cells from individual RA and OA patients that were incubated with or without AREG (100 ng/ml) for 8 h. Alpha-tubulin served as a loading control. (C) Relative optical densities of the western blot bands in (B) to determine P-ERK1/2 expression relative to ERK1/2 expression. (D) Relative optical densities of the western blot bands in (B) to determine Grp94 expression relative to alpha-tubulin expression.

was not permitted in Japan; consequently, none of the patients tested here have ever received FK506. In addition, the enhanced FKBP5 expression by RA BMMC does not correlate with therapeutic treatment using steroids. It remains possible, however, that the increased FKBP5 mRNA levels in the BMMC of RA patients may be due to treatment with other drugs. Alternatively, it may reflect genuine and spontaneous pathological events. Nevertheless, regardless of the cause of its elevated expression, the augmented FKBP expression may strongly inhibit the phosphatase activity of calcineurin, which could increase the dephosphorylation and thus inactivation of various substrates, including the NFAT family proteins and cytokines that are required for the expression of immunoregulatory molecules.

TPST1 mediates tyrosine sulfation within the trans-Golgi system, which affects 1% of all tyrosines in eukaryotic cells. It has been previously suggested that this post-translational modification may play an

important role in the pathogenesis of autoimmune diseases because it regulates mononuclear cell function at various stages of the immune response by enhancing interactions between ligands and receptors.[37] Notably, of the 62 identified target proteins of tyrosine sulfation, nine are cell adhesion molecules and chemokine receptors, which are both central players in leukocyte trafficking. Thus, the augmented expression of *TPST1* in RA patients may elevate the sulfation of crucial tyrosine residues in chemokine receptors that could constitutively increase their binding affinities with their ligands (e.g. the binding of CXCL12–CXCR4).

CLECSF9 belongs to the macrophage-inducible C-type lectin that serves multiple functions by recognizing carbohydrate chains; it plays important roles in macrophage function. Notably, a C-type lectin called DC-specific intercellular adhesion molecule 3-grabbing non-integrin is also highly expressed by macrophages in the synovium of RA patients.[38] However, the HH mRNA expression of macrophage-inducible C-type lectins is strongly induced in response to several inflammatory stimuli. Thus, the augmented expression of *CLECSF9* in the BMMC of RA patients may simply be due to the inflammation in the joint.

Unlike *FKBP5* and *TPST1* genes, the mRNA levels of G0S2, CXCR4 and NF-κB are increased in both the BMMC and PBMC of RA patients (Fig. 2 and Supplementary Figure S3). We previously showed that the PBMC of both systemic lupus erythematosus (SLE) patients and healthy young females express enhanced levels of *G0S2* mRNA.[26] Thus, G0S2 may not actually be involved in the pathogenesis of RA. With regard to the chemokine receptor CXCR4, it was also identified as a inflammation-related gene that is upregulated in synovial cells of patients with pigmented villonodular synovitis (PVNS), which is a joint problem that usually affects the hip or knee and involves the lining of the joint becoming swollen and growing.[8] The enhanced tyrosine sulfation of CXCR4 by augmented TPST1 activity, as described above, may also activate CXCR4, thereby elevating the ability of the CXCR4 ligand to induce the migration of bone marrow cells that could enhance the growth of synovial cells.[39] CXCR4 expression is also upregulated in the spinal cord of animals with experimental autoimmune encephalomyelitis, which is an animal model of autoimmune central nervous system inflammation.[40] With regard to NF-κB, this molecule along with the receptor activator of NF-κB (RANK) and its ligand RANKL have been found to play pivotal roles in the pathophysiological process of RA.[41] Thus, the increased mRNA levels of NF-κB in both the BMMC and PBMC of RA patients may contribute to the bone destruction mediated by activated NF-κB signaling pathway.[42]

AURA1 encodes a novel protein that is similar to thioesterase. Since the thioesterase homologs are widespread, functions of thioesterase vary in the human genome.[43] Thus, the physiological function of *AURA1* remains unknown. A possible role that it could play in RA pathogenesis is suggested by the following observations. First, the stable overexpression of acyl-CoA thioesterase III in human and murine T-cell lines increased both peroxisome numbers and lipid droplet formation, which suggests that it participates in the metabolic regulation of peroxisome proliferation in T cells.[44] Second, altered immune responsiveness is observed in mice deficient in palmitoyl protein thioesterase (PPT1) gene that is mutated in infantile neuronal ceroid lipofuscinosis.[45] Third, $CD4^+$ T cells are the prime mediators of RA in a mouse model SKG strain,[46] and *AURA1* expression is detected predominantly in resting and activated $CD4^+$ T cells (Fig. 3).

AREG is not directly related to immune responses but of all the genes examined, it showed the most conspicuously enhanced expression in both the BMMC and PBMC of many RA patients (Fig. 2A). We also found that the synovial cells of RA patients showed higher sensitivity to AREG, in terms of proliferation, than those of OA patients (Fig. 4). This is not due to augmented expression of EFGR (Fig. 5B, uppermost pane), but due to elevated activation of EGFR signaling pathway because the phosphorylation of ERK1/2 was more enhanced in AREG-treated RA patient synovial cells than that of AREG-treated OA patient synovial cells (Fig. 5). We here present a working hypothesis to explain how augmented AREG expression in BMMC and PBMC of RA patients and subsequent activation of EGFR signaling pathway lead to hyperproliferation of synovial cells in the joints of the RA patients (Fig. 6). Namely, this enhanced phosphorylation of ERK1/2 elevates the expression of many downstream target genes, which may also require the activation of the ERAD system. [12] Given that the Ets-binding site (EBS) of the proximal promoter of the synoviolin gene is responsible for its expression,[47] and that EBS-carrying genes are also activated by signaling events from the ERK pathway,[48] it is possible that the enhanced activation of EGFR signaling induced by AREG may directly activate the expression of synoviolin as well as that of other genes, thereby inducing the hyperproliferation of synovial cells. Thus, it is possible that the ERAD system in RA patients is hyperactivated by synoviolin because of augmented AREG expression in blood cells, possibly in the macrophages that occur in the vicinity of the synovial cells of RA patients, releasing augmented amount of AREG. This hypothesis should be tested more rigourously *in vivo* in the future because the experiments using the isolated synoviocyte cells in tissue culture medium may display distinct response to AREG. Likewise, examination of other EGF family proteins *in vivo* can also be interesting future subjects.

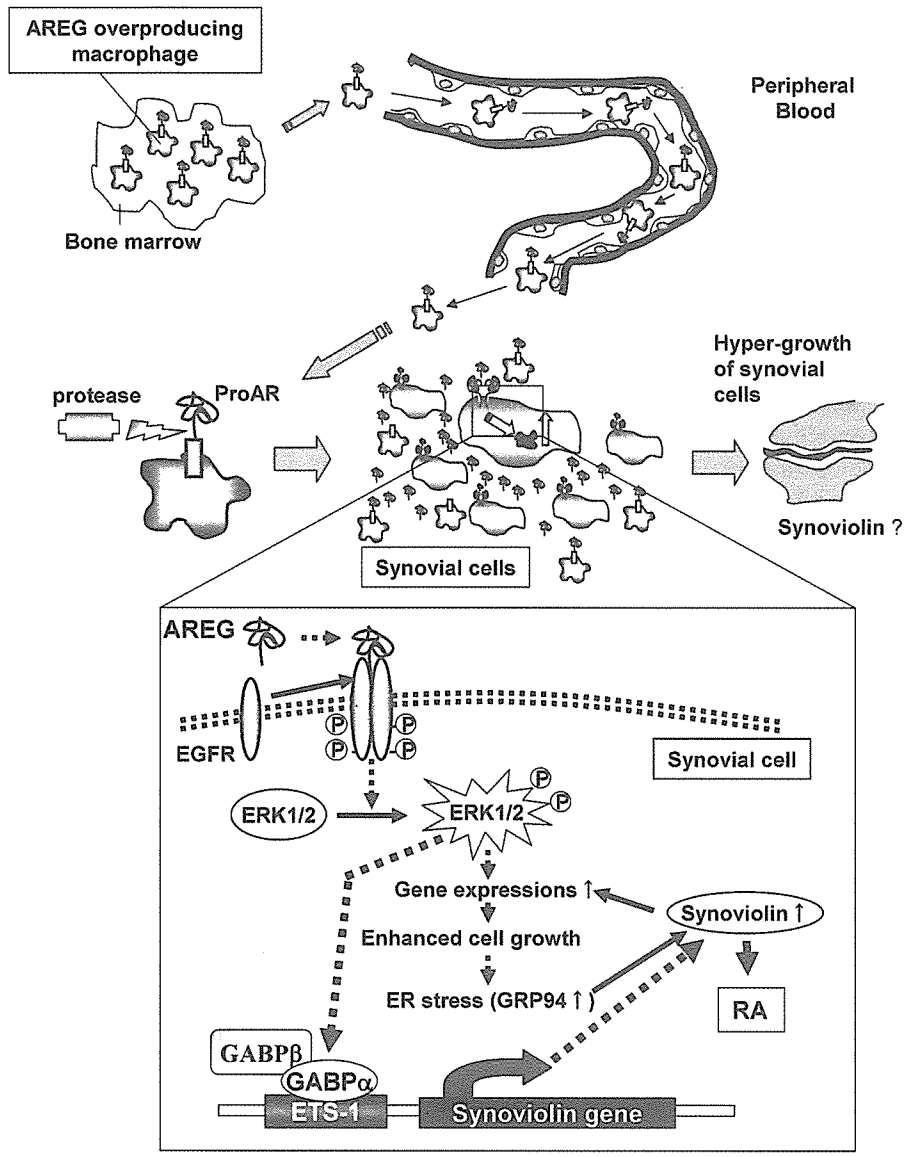


Figure 6. A working hypothesis to explain how augmented level of AREG in BMMC of RA patients may lead to hyperproliferation of synovial cells. Putative macrophages with enhanced expression of AREG precursor (ProAR) may approach to the synovial cells of the joint through blood flow, where they release AREG and activate the EGFR signaling pathway of synovial cells. Since Ets-binding site (ETS-1) of the proximal promoter of the synoviolin gene is one of the downstream targets of ERK pathway, the enhanced activation of EGFR signaling may directly activate the expression of synoviolin gene. The enhanced level of synoviolin activates the ERAD system, which may lead to hyperproliferation of synovial cells.

Overexpression of AREG has been linked to psoriasis in mice and humans.[49,50] Psoriasis is characterized by the hyperproliferation of keratinocytes and the loss of epidermal barrier function that leads to the infiltration of inflammatory cells into the epidermis and dermis.[51] AREG is also upregulated in a synoviocyte cell line derived from an RA patient in which the wild type and a dominant negative form of the orphan nuclear receptor Nurr1 were overexpressed.[52] Interestingly, AREG overexpression in the basal epidermis of transgenic mice induces a phenotype that is associated with synovial membrane inflammation.[49] Moreover, we showed previously that AREG expression is also enhanced in the PBMC of SLE and idiopathic thrombocytopenic purpura patients,[26] which suggests that AREG overexpression may also be associated with other autoimmune diseases. Notably, metalloprotease-mediated AREG shedding and the subsequent activation of EGFR appears to play a critical role in the secretion of IL-8 by the human airway epithelium-like NCI-H292 cells that is induced by tumor necrosis factor-α (TNF-α), a potent multifunctional cytokine that plays a central role in the pathogenesis of many inflammatory diseases like RA.[53] Since TNF-α-induced IL-8 secretion was completely inhibited by the neutralizing antibody against AREG,[53] this antibody could constitute a novel therapeutic tool for RA. Taken

together, we propose that enhanced expression of AREG in BMMC and PMBC may play a pivotal role in the pathogenesis of RA.

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Supplementary Data: Supplementary data are available online at www.dnaresearch.oxfordjournals.org.

References

1. Oertelt, S., Selmi, C., Invernizzi, P., Podda, M., and Gershwin, M. E. 2005, Genes and goals: an approach to microarray analysis in autoimmunity, *Autoimmune Rev.*, 4, 414–422.
2. Glocker, M. O., Guthke, R., Kekow, J., and Thiesen, H. J. 2006, Rheumatoid arthritis, a complex multifactorial disease: on the way toward individualized medicine, *Med. Res. Rev.*, **26**, 63–87.
3. van der Pouw Kraan, T. C., van Gaalen, F. A., Huizinga, T. W., Pieterman, E., Breedveld, F. C., and Verweij, C. L. 2003, Discovery of distinctive gene expression profiles in rheumatoid synovium using cDNA microarray technology: evidence for the existence of multiple pathways of tissue destruction and repair, *Genes Immun.*, 4, 187–196.
4. Ruschpler, P., Lorenz, P., Eichler, W., et al. 2003, High CXCR3 expression in synovial mast cells associated with CXCL9 and CXCL10 expression in inflammatory synovial tissues of patients with rheumatoid arthritis, *Arthritis Res. Ther.*, **5**, 241–252.
5. Devauchelle, V., Marion, S., Cagnard, N., et al. 2004, DNA microarray allows molecular profiling of rheumatoid arthritis and identification of pathophysiological targets, *Genes Immun.*, **5**, 597–608.
6. Cagnard, N., Letourneur, F., Essabbani, A., et al. 2005, Interleukin-32, CCL2, PF4F1 and GFD10 are the only cytokine/chemokine genes differentially expressed by in vitro cultured rheumatoid and osteoarthritis fibroblast-like synoviocytes, *Eur. Cytokine Netw.*, **16**, 289–292.
7. Lindberg, J., af Klint, E., Ulfgren, A. K., et al. 2006, Variability in synovial inflammation in rheumatoid arthritis investigated by microarray technology, *Arthritis Res. Ther.*, 8, R47.
8. Finis, K., Sultmann, H., Ruschhaupt, M., et al. 2006, Analysis of pigmented villonodular synovitis with genome-wide complementary DNA microarray and tissue array technology reveals insight into potential novel therapeutic approaches, *Arthritis Rheum.*, **54**, 1009–1019.
9. Bovin, L. F., Rieneck, K., Workman, C., et al. 2004, Blood cell gene expression profiling in rheumatoid arthritis: discriminative genes and effect of rheumatoid factor, *Immunol. Lett.*, **93**, 217–226.
10. Amano, T., Yamasaki, S., Yagishita, N., et al. 2003, Synoviolin/Hrd1, an E3 ubiquitin ligase, as a novel pathogenic factor for arthropathy, *Genes Dev.*, **17**, 2436–2449.
11. Yagishita, N., Ohneda, K., Amano, T., et al. 2005, Essential role of synoviolin in embryogenesis, *J. Biol. Chem.*, **280**, 7909–7916.
12. Yamasaki, S., Yagishita, N., Tsuchimochi, K., Nishioka, K., and Nakajima, T. 2005, Rheumatoid arthritis as a hyper-endoplasmic reticulum-associated degradation disease, *Athritis Res. Ther.*, **7**, 181–186.
13. Ochi, T., Hakomori, S., Adachi, M., et al. 1988, The presence of a myeloid cell population showing strong reactivity with monoclonal antibody directed to difucosyl type 2 chain in epiphyseal bone marrow adjacent to joints affected with rheumatoid arthritis (RA) and its absence in the corresponding normal and non-RA bone marrow, *J. Rheumatol.*, **15**, 1609–1615.
14. Hayashida, K., Ochi, T., Fujimoto, M., et al. 1992, Bone marrow changes in adjuvant-induced and collagen-induced arthritis, *Arthritis Rheum.*, **35**, 241–245.
15. Jongen-Lavrencic, M., Peeters, H. R., Wognum, A., Vreugdenhil, G., Breedveld, F. C., and Swaak, A. J. 1997, Elevated levels of inflammatory cytokines in bone marrow of patients with rheumatoid arthritis and anemia of chronic disease, *J. Rheumatol.*, **24**, 1504–1509.
16. Hirohata, S., Yanagida, T., Nampei, A., et al. 2004, Enhanced generation of endothelial cells from CD34+ cells of the bone marrow in rheumatoid arthritis: possible role in synovial neovascularization, *Arthritis Rheum.*, **50**, 3888–3896.
17. Nakamura-Kikuoka, S., Takahi, K., Tsuboi, H., et al. 2006, Limited VH gene usage in B-cell clones established with nurse-like cells from patients with rheumatoid arthritis, *Rheumatology*, **45**, 549–557.
18. Samuels, J., Ng, Y. S., Coupillaud, C., Paget, D., and Meffre, E. 2005, Human B cell tolerance and its failure in rheumatoid arthritis, *Ann. N. Y. Acad. Sci.*, **1062**, 116–126.
19. Wardemann, H., Yurasov, S., Schaefer, A., Young, J. W., Meffre, E., and Nussenzweig, M. C. 2003, Predominant autoantibody production by early human B cell precursors, *Science*, **301**, 1374–1377.
20. Bugatti, S., Caporali, R., Manzo, A., Vitolo, B., Pitzalis, C., and Montecucco, C. 2005, Involvement of subchondral bone marrow in rheumatoid arthritis: lymphoid neogenesis and in situ relationship to subchondral bone marrow osteoclast recruitment, *Arthritis Rheum.*, **52**, 3448–3459.
21. Jimenez-Boj, E., Redlich, K., Turk, B., et al. 2005, Interaction between synovial inflammatory tissue and bone marrow in rheumatoid arthritis, *J. Immunol.*, **175**, 2579–2588.
22. Fujii, T., Tamura, K., Masai, K., Tanaka, H., Nishimune, Y., and Nojima, H. 2002, Use of stepwise

subtraction to comprehensively isolate mouse genes whose transcription is up-regulated during spermiogenesis, *EMBO Rep.*, **3**, 367–372.
23. Arnett, F. C., Edworthy, S. M., Bloch, D. A., et al. 1988, The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis, *Arthritis Rheum.*, **31**, 315–324.
24. Altman, R. D. 1991, Criteria for classification of clinical osteoarthritis, *J. Rheumatol. Suppl.*, **27**, 10–12.
25. Kobori, M., Ikeda, Y., Nara, H., et al. 1998, Large scale isolation of osteoclast-specific genes by an improved method involving the preparation of a subtracted cDNA library, *Genes Cells*, **3**, 459–475.
26. Ishii, T., Onda, H., Tanigawa, A., et al. 2006, Isolation and expression profiling of genes upregulated in the peripheral blood cells of systemic lupus erythematosus patients, *DNA Res.*, **112**, 1–11.
27. Shoyab, M., McDonald, V. L., Bradley, J. G., and Todaro, G. J. 1988, Amphiregulin: a bifunctional growth-modulating glycoprotein produced by the phorbol 12-myristate 13-acetate-treated human breast adenocarcinoma cell line MCF-7, *Proc. Natl Acad. Sci. USA*, **85**, 6528–6532.
28. Fischer, G. and Aumuller, T. 2003, Regulation of peptide bond cis/trans isomerization by enzyme catalysis and its implication in physiological processes, *Rev. Physiol. Biochem. Pharmacol.*, **148**, 105–150.
29. Russell, L. and Forsdyke, D. R. 1991, A human putative lymphocyte G0/G1 switch gene containing a CpG-rich island encodes a small basic protein with the potential to be phosphorylated, *DNA Cell Biol.*, **10**, 581–591.
30. Santiago, B., Baleux, F., Palao, G., et al. 2006, CXCL12 is displayed by rheumatoid endothelial cells through its basic amino-terminal motif on heparan sulfate proteoglycans, *Arthritis Res. Ther.*, **8**, R43.
31. Ahn, K. S. and Aggarwal, B. B. 2005, Transcription factor NF-kappaB: a sensor for smoke and stress signals, *Ann. N. Y. Acad. Sci.*, **1056**, 218–233.
32. Shin, H. S., Lee, H. J., Nishida, M., et al. 2003, Betacellulin and amphiregulin induce upregulation of cyclin D1 and DNA synthesis activity through differential signaling pathways in vascular smooth muscle cells, *Circ. Res.*, **93**, 302–310.
33. Lee, A. S. 2001, The glucose-regulated proteins: stress induction and clinical applications, *Trends Biochem. Sci.*, **26**, 504–510.
34. Lee, D. C., Sunnarborg, S. W., Hinkle, C. L., et al. 2003, TACE/ADAM17 processing of EGFR ligands indicates a role as a physiological convertase, *Ann. N. Y. Acad. Sci.*, **995**, 22–38.
35. Curran, M. P. and Perry, C. M. 2005, Tacrolimus: in patients with rheumatoid arthritis, *Drugs.*, **65**, 993–1001.
36. Sasakawa, T., Sasakawa, Y., Ohkubo, Y., and Mutohm, S. 2005, FK506 inhibits prostaglandin E2 production from synovial cells by suppressing peripheral blood mononuclear cells, *Int. Immunopharmacol.*, **5**, 1291–1297.
37. Hsu, W., Rosenquist, G.L., Ansari, A. A., and Gershwin, M. E. 2005, Autoimmunity and tyrosine sulfation, *Autoimmune Rev.*, **4**, 429–435.
38. van Lent, P. L., Figdor, C. G., Barrera, P., et al. 2003, Expression of the dendritic cell-associated C-type lectin DC-SIGN by inflammatory matrix metalloproteinase-producing macrophages in rheumatoid arthritis synovium and interaction with intercellular adhesion molecule 3-positive T cells, *Arthritis Rheum.*, **48**, 360–369.
39. Crazzolara, R. and Bernhard, D. 2005, CXCR4 chemokine receptors, histone deacetylase inhibitors and acute lymphoblastic leukemia, *Leuk. Lymphoma.*, **46**, 1545–1551.
40. Glabinski, A. R., O'Bryant, S., Selmaj, K., and Ransohoff, R. M. 2000, CXC chemokine receptors expression during chronic relapsing experimental autoimmune encephalomyelitis, *Ann. N. Y. Acad. Sci.*, **917**, 135–144.
41. Wada, T., Nakashima, T., Hiroshi, N., and Penninger, J. M. 2006, RANKL-RANK signaling in osteoclastogenesis and bone disease, *Trends Mol. Med.*, **12**, 17–25.
42. Jimi, E. and Ghosh, S. 2005, Role of nuclear factor-kappaB in the immune system and bone, *Immunol. Rev.*, **208**, 80–87.
43. Akoh, C. C., Lee, G. C., Liaw, Y. C., Huang, T. H., and Shaw, J. F. 2004, GDSL family of serine esterases/lipases, *Prog. Lipid Res.*, **43**, 534–552.
44. Ishizuka, M., Toyama, Y., Watanabe, H., et al. 2004, Overexpression of human acyl-CoA thioesterase upregulates peroxisome biogenesis, *Exp. Cell Res.*, **297**, 127–141.
45. Jalanko, A., Vesa, J., Manninen, T., et al. 2005, Mice with Ppt1Deltaex4 mutation replicate the INCL phenotype and show an inflammation-associated loss of interneurons, *Neurobiol. Dis.*, **18**, 226–241.
46. Sakaguchi, N., Takahashi, T., Hata, H., et al. 2003, Altered thymic T-cell selection due to a mutation of the ZAP-70 gene causes autoimmune arthritis in mice, *Nature*, **426**, 454–460.
47. Tsuchimochi, K., Yagishita, N., Yamasaki, S., et al. 2005, Identification of a crucial site for synoviolin expression, *Mol. Cell. Biol.*, **25**, 7344–7356.
48. Hoffmeyer, A., Avots, A., Flory, E., Weber, C. K., Serfling, E., and Rapp, U. R. 1998, The GABP-responsive element of the interleukin-2 enhancer is regulated by JNK/SAPK-activating pathways in T lymphocytes, *J. Biol. Chem.*, **273**, 10112–10119.
49. Cook, P. W., Brown, J. R., Cornell, K. A., and Pittelkow, M. R. 2004, Suprabasal expression of human amphiregulin in the epidermis of transgenic mice induces a severe, early-onset, psoriasis-like skin pathology: expression of amphiregulin in the basal epidermis is also associated with synovitis, *Exp. Dermatol.*, **13**, 347–356.
50. Bhagavathula, N., Nerusu, K. C., Fisher, G. J., et al. 2005, Amphiregulin and epidermal hyperplasia: amphiregulin is required to maintain the psoriatic phenotype of human skin grafts on severe combined immunodeficient mice, *Am. J. Pathol.*, **166**, 1009–1016.
51. Ritchlin, C. T. 2005, Pathogenesis of psoriatic arthritis, *Curr. Opin. Rheumatol.*, **17**, 406–412.
52. Davies, M. R., Harding, C. J., Raines, S., et al. 2005, Nurr1 dependent regulation of pro-inflammatory mediators in immortalised synovial fibroblasts, *J. Inflamm. (Lond)*, **2**, 15.
53. Chokki, M., Mitsuhashi, H., and Kamimura, T. 2006, Metalloprotease-dependent amphiregulin release mediates tumor necrosis factor-alpha-induced IL-8 secretion in the human airway epithelial cell line NCI-H292, *Life Sci.*, **78**, 3051–3057.

リウマチの破骨因子

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リウマチの破骨因子

■新たなリウマチの破骨細胞分化誘導系

関節リウマチ(RA)の罹患者は日本国内で約80万人ともいわれ，これらの患者の機能障害をいかに軽減するかは医療福祉上・医療経済上重要な問題である．RAは四肢の関節破壊を主徴とするが，これに伴って加齢によるものよりはるかに高度な骨粗鬆症が生じ，これも患者の機能を大きく損ねる要因である．しかし高度な骨粗鬆の病態は解明されていない部分が多く，その予防・治療の方法はまったく確立されていないといっても過言ではない．

RAでは関節腔内からも骨破壊が進行するのが特徴的である．関節滑膜組織内に破骨細胞様の形態をとる多核巨細胞が存在することは以前より知られており，これが関節内からの骨吸収に関与する可能性も示唆されていた．われわれはRA関節腔内に前駆破骨細胞が存在することを予測し，RA患者関節液中に大量のCD14陽性単球様細胞の存在と，これらがサイトカインの刺激により骨吸収能を持つ成熟破骨細胞に分化することを見出し，前駆破骨細胞であるCD14陽性単球様細胞は，健常人末梢血単球とは異なる表面抗原プロファイルを有しており，分化段階の異なる細胞であることが示唆された[5]．

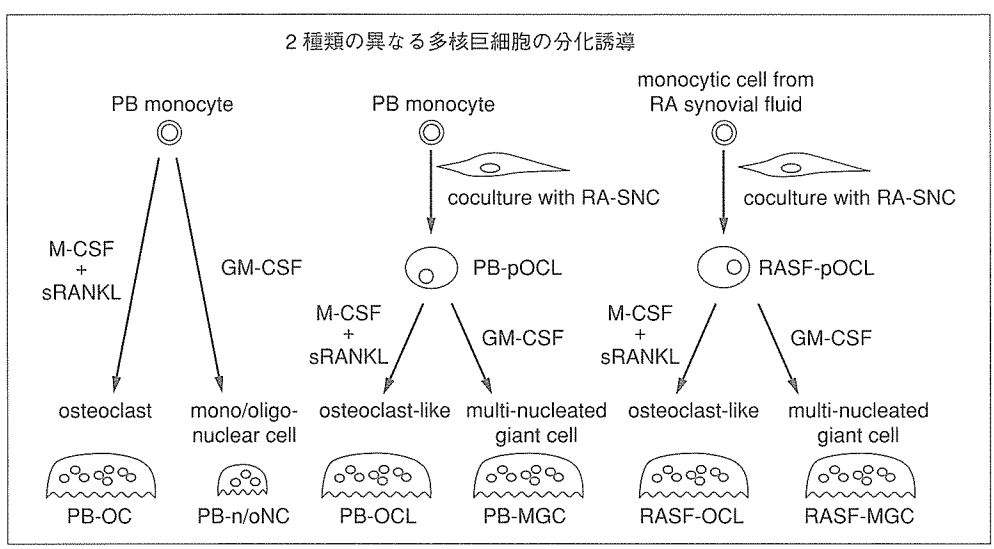
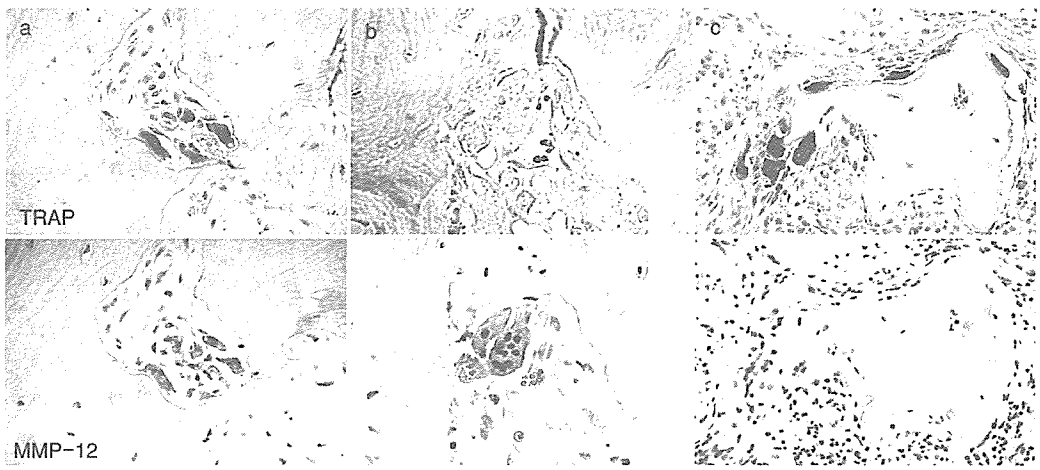


図1　M-CSF+RANKL依存性破骨細胞とM-CSF+RANKL非依存性破骨細胞

PB monocyte：末梢血単球　　M-CSF：マクロファージコロニー刺激因子
OCL　　：破骨細胞　　RASF：リウマチ滑膜線維芽細胞
RANKL　　：NF-κB活性化受容体リガンド

リウマチ骨吸収部位における MMP-12 の発現



健常な成人の骨・軟骨においては MMP-12 の発現は認められない.

図 2　RA 罹患部に存在する MMP-12 陽性の破骨細胞(文献 1)
a：$TRAP^{+}$ $MMP12^{+}$の破骨細胞
b：$TRAP^{-}$ $MMP12^{+}$の破骨細胞
c：$TRAP^{+}$ $MMP12^{-}$の破骨細胞

従来，破骨細胞はその分化および成熟のすべての過程において骨髄の間質細胞や骨芽細胞などの支持細胞が必要であるとされてきた．また近年，支持細胞の代わりに支持細胞の発現する分子(M-CSF，RANKL)を使用する破骨細胞誘導系も報告されている．一方，筆者らが RA 関節滑液から発見した RA 患者由来 CD14 陽性単球様細胞は，前駆破骨細胞としての性状を有しながらも，このような支持細胞や M-CSF＋RANKL を必要とせず，サイトカイン(IL-3, IL-5, IL-7 または GM-CSF)の刺激のみによって成熟破骨細胞へと分化する(図 1)．この分化に関わるこれらサイトカインは，RA 患者の関節液中に高頻度・高濃度に存在するものであることから，われわれが発見した CD14 陽性単球様細胞は関節腔内で直接的に破骨細胞に分化するものと推測される．

この仮説を実証することは，RA における骨吸収の新たなメカニズムを示すことになるが，いままでこれに類した報告は見当たらない．この方法は RA 患者から前駆破骨細胞を得る方法であり，培養前駆細胞は 98%以上の純度で繰り返し大量に実験に使用することが可能である．また，この方法では，末梢血単球から直接に破骨細胞を誘導してしまう M-CSF＋RANKL の実験系では得られない，前駆破骨細胞の性状解析と，前駆細胞から成熟破骨細胞への分化機序の解明が可能になると考えられる．

このような方向からの破骨細胞形成過程の解明は，われわれが RA 患者滑液から見出した方法以外のアプローチでは困難と考えられる．従来の実験系では解明できなかった，末梢血単球―前駆破骨細胞―成熟破骨細胞への分化過程と，各ステージに特異的な遺伝子産物の発現を明らかにすることができる．さらに得られた結果をもとに，骨代謝を各ステージ特異的に制御できる治療法を開発することが可能と考えている．

■RA 関節組織の免疫化学的検討

RA 患者の関節組織を免疫化学的に解析した結果，酒石酸耐性酸フォスファターゼ(tartrateresistant acid phosphatase；TRAP)陽性の単核球と多核巨細胞が滑膜組織と軟骨下骨付近に見出され，これらの関節滑膜組織中に存在していた TRAP 陽性単核球と多核巨細胞からは MMP(macrophage metalloelastase)-2 と MMP-9 が検出された．さらに TRAP 陽性多核巨細胞から MMP-12 と MMP-14 も検出された．RA 関節由来の TRAP 陽性単核球は，*in vitro* で軟骨組織中のプロテオグリカンを切断することも確認された．RA と変形性関節症(OA)の関節液中に見出される CD14 陽性前駆破骨細胞の性状と機能を比較するために，各疾患の患者の関節液の初代培養から得た CD14 陽性前駆破骨細胞を IL-3，IL-5，IL-7 または GM-CSF により刺激し，破骨細胞への分化誘導を行った．その結果，RA，OA それぞれの関節液由来前駆破骨細胞が破骨細胞へと分化したが，破骨細胞ごとの核数，単核細胞から多核細胞への融合率，骨吸収能のいずれにおいても RA 由来前駆破骨細胞によるもののほうが大きかった．このことより，RA 関節腔においては OA のそれより強く活性化された前駆破骨細胞が存在し，関節破壊に関与している可能性が示唆された[2]．

RA 患者において，組織マクロファージが関節滑膜組織中に集簇する機序を明らかにするため，われわれが以前からその存在を報告してきた RA 滑膜組織に特異的に存在する間質細胞(RA ナース細胞)[3,4]の培養上清による末梢血単球の遊走を測定したところ，RA ナース細胞培養上清は対照群である OA 由来間質細胞や皮膚由来線維芽細胞の培養上清と比較し，有意に単球の遊走を刺激する結果を得ている．さらに MCP-1(monocyte chemotactic protein-1)と IL-8 に対する中和抗体により単球遊走が阻害されたため，MCP-1 および IL-8 が RA ナース細胞における単球遊走に関与することが示唆された．また，TNFα 刺激により RA ナース細胞によるこれらの遊走因子産生が増強されることを見出している．

■今後の課題

RA 患者関節液の初代培養から CD14 陽性かつ破骨細胞特異的酵素である TRAP 陽性の単核球を見出し，その表面抗原プロファイルが健常人 CD14 陽性末梢血単球と異なることを発見した．また，これら CD14 陽性 TRAP 陽性単核球が IL-3，IL-5，IL-7 あるいは GM-CSF の存在下，支持細胞を必要とせずに強い骨吸収能を有する成熟破骨細胞へと分化することを確認している．前駆破骨細胞は高純度で長期にわたり維持され，最長 8 カ月まで繰り返し本誘導実験系に使用できることも明らかにした．さらに，健常人からの CD14 陽性単球を RA 滑膜ナース細胞と共培養することにより，正常末梢血由来単球は TRAP 陽性の前駆破骨細胞に分化し，IL-3，IL-5，IL-7 あるいは GM-CSF により関節液由来の前駆細胞同様，成熟破骨細胞へ分化することが判明した．また，RA に特異的に存在する RA ナース細胞が RA における破骨細胞の誘導に重要な関与をすることが示唆されてきた．

現在，RA 罹患部の免疫組織学検討から，M-CSF＋RANKL で誘導される破骨細胞が保有しない MMP-12 陽性の破骨細胞が RA 患者にのみ特徴的に存在することから(図 2)，RA における破骨細胞は，われわれが新たに見出した M-CSF＋RANKL 非依存性の分化経路による破骨細胞が，RA の骨破壊に関与している可能性を示唆したと考えている．

われわれの研究ではまず，RA 患者の関節における RA ナース細胞による前駆破骨細胞ならびに，これから分化誘導される破骨細胞に特異的に発現する遺伝子を現在探索中である．既に，この探索から見出された未解明の遺伝子に関しては，現在，これらがコードするタンパクの特異性の検討ならびに機能を解析中である．RA の骨破壊に

関与すると予想される同定されたタンパクのうち，破骨細胞の分化あるいは機能(骨吸収能)発現に関与するものをターゲットとして，その抑制による骨粗鬆の抑制を試みることが可能となってきている．

以上はRAの骨破壊に関わるわれわれ独自の発見であるが[5,6]，最近，米国・韓国・日本で複数のグループによっても，本実験系がRA病態を反映し，骨・関節破壊治療法開発に有用であると報告されたが，本実験系による破骨細胞分化解析はわれわれが先行している．先進各国における社会の高齢化を鑑みても，骨代謝の中心的役割を担う破骨細胞の分化・機能制御が医療福祉上極めて重要な課題であることは間違いないと考えている．

文　献

1) Kerkela E, Bohling T, Herva R, et al：Human macrophage metalloelastase(MMP-12)expression is induced in chondrocytes during fetal development and malignant transformation. Bone **29**：487-493, 2001
2) Takano H, Tomita T, Toyosaki-Maeda T, et al：Comparison of the activities of multinucleated bone-resorbing giant cells derived from CD14-positive cells in the synovial fluids of rheumatoid arthritis and osteoarthritis patients. Rheumatology(Oxford) **43**：435-441, 2004
3) Takeuchi E, Tomita T, Toyosaki-Maeda T, et al：Establishment and characterization of nurse cell-like stromal cell lines from synovial tissues of patients with rheumatoid arthritis. Arthritis Rheum **42**：221-228, 1999
4) Tomita T, Takeuchi E, Toyosaki-Maeda T, et al：Establishment of nurse-like stromal cells from bone marrow of patients with rheumatoid arthritis：indication of characteristic bone marrow microenvironment in patients with rheumatoid arthritis. Rheumatology(Oxford)**38**：854-863, 1999
5) Toyosaki-Maeda T, Takano H, Tomita T, et al：Differentiation of monocytes into multinucleated giant bone-resorbing cells：two-step differentiation induced by nurse-like cells and cytokines. Arthritis Res **3**：306-310, 2001
6) Tsuboi H, Matsui Y, Hayashida K, et al：Tartrate resistant acid phosphatase(TRAP)positive cells in rheumatoid synovium may induce the destruction of articular cartilage. Ann Rheum Dis **62**：196-203, 2003

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