

Ⅲ. 研究成果の刊行に関する一覧

研究成果の刊行に関する一覧表

書籍

著者氏名	論文タイトル名	書籍全体の 編集者名	書籍名	出版社名	出版地	出版年	ページ
広畑俊成	14.膠原病及び類縁疾患 ベーチェット病(内科).	山口徹 北原光夫 福井次矢 総編集	今日の治療指 針 2006(ポケッ ト版)	医学書院	東京	2006	608- 610
広畑俊成	6.抗リウマチ薬各論一本邦既 承認薬 6-2免疫抑制薬 4. タクロリム ス.	川合眞一 編集	抗リウマチ薬の 選び方と使い 方	南江堂	東京	2006	52-54
広畑俊成	7.抗リウマチ薬各論一本邦 未承認薬 7-2免疫抑制薬 1.アザチオ プリン、 2.シクロホスファミド、3.シクロ スポリン.	川合眞一 編集	抗リウマチ薬の 選び方と使い 方	南江堂	東京	2006	81-89
広畑俊成	III.骨・関節疾患 1.骨粗鬆症、 2.関節リウマチ、 3.変形性関節症.	石崎高志 鎌滝哲也 望月真弓 編集	薬物療法学	南江堂	東京	2006	205- 220
廣畑俊成	第4部疾患としてみた膠原病・ リウマチ“膠原病” H.Behçet 病.	住田孝之編集	「EXPERT膠原 病・リウマチ」改 定第2版	診断と治 療社	東京	2006	328- 337
田中 栄	骨の解剖(構造)と生理	菊地臣一 中村利孝 越智光夫	経験すべき外 傷・疾患97	Medical View	東京	2006	pp2-4
田中 栄	新しい治療薬③抗RANKL抗体	中村利孝 松本俊夫	骨粗鬆症診療 ハンドブック 改訂4版	医薬 ジャーナ ル社	東京	2006	pp285- 292
田中 栄	骨および標的臓器に対する作用	中村利孝 松本俊夫 加藤茂明	骨代謝と活性 型ビタミンD- 過渡と現在、そ して未来-	ライフサイ エンス出 版	東京	2006	pp51- 56

研究成果の刊行に関する一覧表

雑誌

発表者氏名	論文タイトル名	発表誌名	巻号	ページ	出版年
山田 真介 稲葉 雅章 後藤 仁志 田中 一匡 酒井 真礼 白川 久美 今西 康雄 西沢 良記	アレンドロネート投与後の骨量改善に伴う動脈硬化進展に及ぼす影響について	Osteoporosis Japan	14	39-41	2006
田中 一匡 稲葉 雅章 櫻井 真由美 山田 真介 酒井 真礼 後藤 仁志 西沢 良記	関節リウマチ患者では傍関節性骨粗鬆症の進展が末梢の動脈硬化進展に関与する	Osteoporosis Japan	14	291-293	2006
Yamada, S. Inaba, M. Goto, H. Ngata, M. Kumeda Y. Imanishi Y. Emoto, M. Ishimura E. Nishizawa, Y.	Associations between physical activity, peripheral atherosclerosis and one status in health Japanese women.	Atherosclerosis	188	196-202	2006
Tanaka, K. Inaba, M. Goto, H. Ngata-Sakurai, M. Sakai, S. Yamada, S. Ueda, M. Ishimura E. Nishizawa, Y.	Paraarticular trabecular bone loss at the ultradistal radius and increased arterial stiffening in postmenopausal patients with rheumatoid arthritis	J Rheumatol	33	652-658	2006
妻木 範行 村井 純子 岩井 貴男 岡本 美奈 吉川 秀樹	BMPシグナルと骨形成・骨吸収	The Bone	20	343-348	2006
名井 陽 玉井 宣行 荒木 信人 藤井 昌一 富田 哲也 古野 雅彦 吉川 秀樹	連通気孔構造を有するハイドロキシアパタイト人工骨の臨床応用, 物理学的特性・臨床的特徴・問題点	日整会誌	80	262-269	2006

坪井 秀規 吉川 秀樹	関節リウマチにみられる骨粗鬆症の臨床病態	Rheumatology	13	22-25	2006
向井 克容 細野 昇 坂浦 博伸 藤原 桂樹 富士 武史 吉川 秀樹	関節リウマチ中下位頸椎病変に対する椎弓形成術	別冊整形外科	50	108-112	2006
Ando,W. Hashimoto,J. Nampei,A. Tsuboi,H. Taakeshi,K. Ono,T. Nakamura,N. Ochi,T. Yoshikawa,H.	.Imatinib mesylate inhibits osteoclastogenesis and joint destruction in rats with collagen-induced arthritis	Journal of Bone Mineral Metabolism	24(CIA)	274-282	2006
Hirao,M. Tamai,N. Tsumaki,N. Yoshikawa,H. Myoui,A.	Oxygen tension regulates chondrocyte differentiation and function during endochondral ossification.,:	Journal of Biological Chemistry	281	3179-3192	2006
Hirohata,S. Miura,Y. Tomita,T. Yoshikawa,H. Ochi,T. Chiorazzi,N.	.Enhanced expression of mRNA for nuclear factor kB1(p50) in CD34+ cells of the bone marrow in rheumatoid arthritis	Arthritis Research & Therapy	8	R54	2006
Kaito,T. Mukai,Y. Nishikawa,M. Ando,W. Yoshikawa,H. Myoui,A.	Dual hydroxyapatite composite with porous and solid parts: Experimental study using canine lumbar interbody fusion model	J Biomed Mater Res B Appl Biomater	78	378-384	2006
Kawane,K. Ohtani,M. Miwa,K. Kizawa,T. Kanbara,Y. Yoshioka,Y Yoshikawa,H.	Chronic polyarthritis caused by mammalian DNA that escapes from degradation in macrophages	Nature	443	:998-1002	2006
Kunugiza,Y. Tomita,T. Tomita,N. Morishita,R. Yoshikawa,H.	Inhibitory effect of ribbon-type NFkB decoy oligodeoxynucleotides on osteoclast induction and activity in vitro and in vivo	Arthritis Research & Therapy	8	R103	2006
Matsubara,T. Myoui,A. Ikeba,F. Yoshikawa,H. Yoneda,T.	Critical role of cortactin in actin ring formation and osteoclastic bone resorption	Journal of Bone Miner Metabolism	24	368-372	2006
Nakase,T. Yoshikawa,H.	Potential roles of bone morphogenetic proteins (BMPs) in skeletal repair and regeneration	Journal of Bone and Mineral Metabolism	24	425-433	2006
Okamoto,M. Murai,J. Yoshikawa,H. Tsunami,N.	Bone morphogenetic proteins in bone stimulate osteoclasts and osteoblasts during bone development.	Journal of Bone Mineral Research	21	1022-1033	2006
Sakaura,H. Hosono,N. Mukai,Y. Fuji,R. Yoshikawa,H.	Paraparesis due to exacerbation of preexisting spinal pseudoarthrosis following infliximab therapy for advanced ankylosing spondylitis.	Spine Journal	6	325-329	2006

Shi,K. Hayashiba,K. Hashimoto,J. Sugamoto,K. Kawai,H. Yoshikawa,H.	Hydroxyapatite augmentation for bone atrophy in total ankle replacement in rheumatoid arthritis.	Journal of Foot and Ankle Surgery	45	316-321	2006
Tomita,T. Kunugiya,Y. Tomita,N. Takanno,H. Morishita,R. Kaneba,Y Yoshikawa,H.	E2F decoy oligodeoxynucleotide ameliorates cartilage invasion by infiltrating synovium derived from rheumatoid arthritis. International	Journal of Molecular Medicine	18	257-265	2006
Karassa FB. Afeltra A. Ambrozeic A. Chang D-M. Keyser FD. Doria A. Galeazzi M. Hirohata S. Hoffman IEA Inanc M. Massardo L. Mathieu A. Mok CC. Morozzi G. Sanna G. Sqindler AJ. Yzioufas AG. Yoshio T. Ioannidis JPA.	Accuracy of anti-ribosomal P protein antibody testing for the diagnosis of neuropsychiatric systemic lupus erythematosus. An international meta-analysis.	Arthritis Rheum	54	312-324	2006
Hirohata S	Role of bone marrow in the pathogenesis of rheumatoid arthritis.	Curr Rheum Rev	2	47-54	2006
Hirohata S. Miura Y. Tomita T. Yoshizawa H. Ochi T. Chiorazzi N.	Enhanced expression of mRNA for nuclear factor kB1 (p50) in CD34+ cells of the bone marrow in rheumatoid arthritis.	Arthritis Res Ther	8	R54	2006
Hirohata S	Is the long-term use of systemic corticosteroids beneficial in the management of Behcet's syndrome.	Nature Clin Practice Rheum	2	358-359	2006
広畑俊成	膠原病・セミナー/膠原病の特異病変における診療のポイント。中枢神経病変。	Medical Practice	23	624-638	2006
広畑俊成	特集 自己免疫疾患の新しい治療法—生物学的製剤を中心に— エファリズマブ(LFA抗体)とナタリズマブ(VLA-4抗体)。	最新医学	61	993-999	2006
広畑俊成	特集 内科疾患最新の治療—専門家からのアドバイス VI.膠原病・免疫・アレルギー Behcet病。	内科	97	1238-1239	2006

広畑俊成	関節リウマチ骨髄CD34+細胞からの線維芽細胞様細胞の分化.	Rheumatology Clinical Update	13	19-21	2006
広畑俊成	「関節リウマチ —積極的な治療へのパラダイム転換」Suggestion: 関節リウマチの病態形成における骨髄異常について.	治療学	40	732-733	2006
広畑俊成	特集 リウマチ・膠原病にみられる自己抗体—その対応抗原と臨床的意義 抗リボソームP抗体.	リウマチ科	36	58-64	2006
廣畑俊成	整形外科医が誤りやすい膠原病.	臨床整形外科	41	962-969	2006
Rosenthal AK. Gohr CM. Uzuki M. Masuda I.	Osteopontin promotes pathologic mineralization in articular cartilage	Matrix Biol	in press		2006
Yoshimura F. Kanno H. Uzuki M. Tajima K. Shimamura T. Sawai T.	Downregulation of inhibitor of apoptosis proteins in apoptotic human chondrocytes treated with tumor necrosis factor- α and actinomycin D	Osteoarthr Cartil	14	435-441	2006
Uzuki M. Otsuka K. Akiyama Y. Ohtsu T. Guy CD. Sawai T.	A case of vermiform appendix tumor	J Iwate Med Assoc	58	151-153	2006
Watanabe T. Sato A. Sawai T. Uzuki M. Goto H. Ymashita H. Akamatsu D. Sato H. Shimizu T. Miyama N. Nakano Y. Satomi S.	The elevated level of circulating matrix metalloproteinase-9 in patients with abdominal aortic aneurysms decreased to levels equal to those of healthy controls after an aortic repair	Ann Vasc Surg	20	317-321	2006
Matushita I. Uzuki M. Matsuno H. Sugiyama E. Kimura T.	Rheumatoid nodulosis during methotrexate therapy in a patient with rheumatoid arthritis	Mod Rheumatol	16	401-403	2006
宇月 美和 徳永 勢二 佐藤 克巳 澤井 高志	関節病変の病理	臨床リウマチ	18	103-113	2006
宇月 美和 徳永 勢二 貝山 潤 鎌滝 章央 澤井 高志	関節リウマチにおける軟骨破壊とヒアルロン酸代謝	リウマチ科	35	578-585	2006

貝山 潤 字月 美和	変形性膝関節症における関節液のヒアルロン酸とその性状の変化	岩手医誌	58	9-21	2006
Samba,K. Araki,K. Li,Z. Matsumoto,K. Suzuki,M. Nakagata,N. Takagi,K. Takeya,M. Yoshinobu,K. Araki,M. Imai,K. Abe,K. Yamamura,K.	A novel murine gene, Sickle tail (Skt), linked to the Danforth's short tail (Sd) locus, is required for normal development of the intervertebral disc.	Genetics	172	445-456	2006
Miura,K. Yoshinobu,K. Imaizumi,T. Hatuna,K. Miyamoto,Y. Yoneda,Y. Nakagata,N. Araki,M. Miyakawa,T. Yamamura,K. Araki,K.	Impaired expression of Importin/karyopherin b1 leads to post-implantation lethality.	Biochem Biophys Res Commun.	341	132-138	2006
Kishigami,S. Komatsu,Y. Takeda,H. Nomura- Kitabayashi,A. Yamauchi,Y. Abe,K. Yamamura,K. Mishina,Y.	An optimized beta-gal staining method for simultaneous detection of endogenous gene expression in early mouse embryos.	Genesis: J. Genet. Dev.	44	57-65	2006
Ohmuraya,M. Hirota,M. Araki,K. Baba,H. Yamamura,K.	Enhanced trypsin activity in pancreatic acinar cells deficient for serine protease inhibitor Kazal type 3	Pancreas	33	104-106	2006
Reifenberg,K. Hildt,E. Lecher,B. Wiese,E. Nusser,P. Ott,S. Yamamura,K. Rutter,G. Loehler,J.	IFN α expression inhibits LHBs storage disease and ground glass hepatocyte appearance, but exacerbates inflammation and apoptosis in HBV surface proteinaccumulating transgenic livers.	Liver International	26	986-993	2006
Kanakubo,S. Nomura,T. Yamamura,K. Miyazaki,J. Tamai,M. Osumi,N.	Abnormal migration and distribution of neural crest cells in Pax6 heterozygous mutant eye, a model for human eye diseases.	Genes to Cells	11	919-933	2006
Araki,K. Araki,M. Yamamura,K.	Negative Selection with the Diphtheria toxin A fragment Gene Improves Frequency of Cre-Mediated Cassette Exchange in ES Cells.	J. Biochem.	140	793-798	2006

Hikita A. Tanaka S. et al.	Negative Regulation of Osteoclastogenesis by Ectodomain Shedding of Receptor Activator of NF- κ B Ligand.	J Biol. Chem	281	36846-36855	2006
Sato K. Tanaka S. et al.	Th17 functions as an osteoclastogenic helper T cell subset that links T cell activation and bone destruction.	J Exp Med	203	2673-2682	2006
Sawada Y. Tanaka S. et al.	Force Sensing by Mechanical Extension of the Src Family Kinase Substrate p130Cas.	Cell	127	1015-1026	2006
Tanaka S, et al.	Molecular mechanism of the life and death of the osteoclast..	Ann N Y Acad Sci	1068	180-186	2006
Miyazaki T. Tanaka S. et al.	The role of c-Src kinase in the regulation of osteoclast function.	Mod Rheumatol	16	68-74	2006
大島 和也 下村 伊一郎 船橋 徹	肥満とカルシウム骨代謝	内分泌・糖尿病	23巻	108-116	2006
大島 和也, 下村 伊一郎	最新用語解説 基礎 アディポサイトカイン	骨粗鬆症治療	4巻	340-347	2005
Nakamura N. Shimaoka N. Tougan T. Onda H. Okuzaki D. Zhao H. Fujimori A. Yabuta N. Nagamori I. Tanaigawa A. Sato J. Hayashiba K. Suzuki R. Yukioka M. Ochi T.	Isolation and expression profiling of genes upregulated in bone marrow-derived mononuclear cells of rheumatoid arthritis patients.	DNA Res.	13(4)	169-183	2006
Fukunaga A. Nagai H. Yu X. Oniki S. Okazawa H. Motegi S. Suzuki R. Honma N. Matozaki T. Nishigori C. Horikawa T.	Src homology 2 domain-containing protein tyrosine phosphatase substrate 1 regulates the induction of Langerhans cell maturation.	Eur J Immunol	36(12)	3216-3226	2006
Fukui N. Ikeba Y. Ohnuki T. Hikita A. Tanaka S. Yamane S. Suzuki R. Sandell LJ. Ochi T.	Pro-inflammatory cytokine tumor necrosis factor- α induces bone morphogenetic protein-2 in chondrocytes via mRNA stabilization and transcriptional up-regulation.	J Biol Chem.	281(37)	273229-27241	2006
鈴木隆二	リウマチの破骨細胞	臨床整形外科	41(3)	260-263	2006
鈴木隆二 越智隆弘	関節リウマチに見られる特異な破骨細胞分化機序	Rheumatology Clinical Update :	13	15-18	2006

IV. 研究成果の刊行物

IV－I 総括研究刊行物

Review

Mesenchymal stromal cells

Nurse-like cells reside in the synovial tissue and bone marrow in rheumatoid arthritis

Takahiro Ochi¹, Hideki Yoshikawa², Tomoko Toyosaki-Maeda³ and Peter E Lipsky⁴

¹Sagamihara National Hospital, Sagamihara, Kanagawa, Japan

²Department of Orthopaedic Surgery, Osaka University Medical School, Suita, Osaka, Japan

³Department of Immunology, Shionogi Research Laboratories, Shionogi & Co. Ltd, Osaka, Japan

⁴National Institute of Arthritis and Musculoskeletal and Skin Diseases, Bethesda, MD 20892, USA

Corresponding author: Takahiro Ochi, t-ochi@sagamihara-hosp.gr.jp

Published: 8 February 2007

This article is online at <http://arthritis-research.com/content/9/1/201>

© 2007 BioMed Central Ltd

Arthritis Research & Therapy 2007, 9:201 (doi:10.1186/ar2105)

Abstract

A major question concerning the immunopathology of rheumatoid arthritis is why the disease is localized to particular joints. A possible explanation could be the presence within the synovium of cells that foster inflammation or easy accessibility of the synovium to migratory disease enhancing cells. Within both the bone marrow and the synovium, fibroblastic stromal cells play an important role in supporting the differentiation and survival of normal cells, and also contribute to the pathologic processes. Among fibroblastic stromal cells in synovial tissue and bone marrow, nurse-like cells are a unique population having the specific capacity to promote pseudoemperipolesis (adhesion and holding beneath) of lymphocytes, and also the ability to promote the growth and function of some populations of lymphocytes and monocytes. Nurse-like cells could therefore contribute to the immunopathogenesis of rheumatoid arthritis, and may contribute to the localization of inflammation within specific joints. The present review considers the evidence that supports these possibilities.

Introduction

Rheumatoid arthritis (RA) is a chronic autoimmune disease characterized by immunologically enhanced inflammation and damage to articular structures [1,2]. Rheumatoid synovium is a site of intense inflammation, with active involvement by various populations of infiltrating lymphocytes, myeloid cells, and resident synovial fibroblasts or synoviocytes [1]. One question that has not been addressed is why RA preferentially affects certain joints. Although the explanation for the localization of rheumatoid inflammation to particular joints is not clear, one possibility relates to the presence within the synovium of resident cells that can promote inflammation. In addition, cells that can be induced to migrate from adjacent bone marrow structures may contribute to the

local facilitation and propagation of inflammation and bone damage. The present review will focus on one such population, the nurse-like cells (NLCs) that populate the rheumatoid synovium and bone marrow.

Fibroblastic stromal cells in bone marrow and synovial tissue

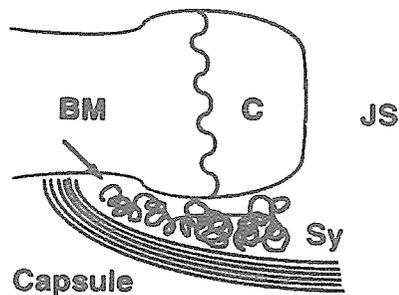
Initially, to examine the relationship between the epiphyseal bone marrow and synovial tissue, we employed the animal model of collagen-induced arthritis [3]. Fibroblastic stromal cells (FSCs) in the bone marrow of Lewis rats were labeled with a fluorescent probe or ³HTdr and were examined for their migration at the onset of arthritis [4]. Accompanying the induction of polyarthritis, a large number of labeled FSCs in bone marrow were found to migrate into the joint cavity through canals observed in the bare zone of the joint (Figure 1), and then to proliferate in the synovial tissue. This observation suggested the hypothesis that pathophysiological cells of RA could be produced in bone marrow, from which some of these cells could migrate into the joint space and potentially play roles in inflammation or tissue damage in and around articular structures. Based on these findings, we have studied FSCs of RA patients, comparing the characteristics of FSCs from bone marrow and FSCs from synovial tissue [5-7].

Nurse-like cells found in bone marrow and synovial tissue

Among the FSCs derived from the bone marrow and synovium of RA patients, a population of NLCs was identified by the capacity to carry out pseudoemperipolesis. The

BST-1 = bone marrow fibroblastic stromal cell antigen 1; FSC = fibroblastic stromal cell; GM-CSF = granulocyte/macrophage colony-stimulating factor; HLA = human major histocompatibility antigen; IFN = interferon; IL = interleukin; mAb = monoclonal antibody; NLC = nurse-like cell; RA = rheumatoid arthritis; RANKL = receptor activator of NF- κ B ligand; TNF = tumor necrosis factor; TRAP = tartrate-resistant acid phosphatase.

Figure 1



Migration of fibroblastic stromal cells from epiphyseal bone marrow (BM) into the joint space (JS) forming synovial (Sy) tissue in collagen-induced arthritis. C, cartilage.

function of the NLCs was reminiscent of thymic nurse cells [8,9], which have the capacity to interact with populations of thymic cells and gather them beneath their cell bodies in a process known as pseudoemperipolesis (adhesion and holding beneath). *In vivo*, such thymic nurse cells were thought to support the development and expansion of thymocytes and to also play a role in positive/negative selection of T cells in mouse and rat thymus. A very similar capacity to interact and support the maturation of some population of lymphocytes and monocytes was noted for FSCs of bone marrow [5,7] and for FSCs of synovial tissue [6,7] of RA patients, suggesting that the NLC function of FSCs could contribute to the pathophysiology of RA [7].

We established RA-NLC clones with the ability to promote pseudoemperipolesis from bone marrow [5] and synovial tissue [6] of RA patients. These RA-NLC clones were determined to be of mesenchymal origin, given that they expressed vimentin but not cytokeratin. They did not exhibit desmosomes or classical junctional complexes, both of which are characteristic features of epithelial cells. Elongated and branching mitochondria were present in the cytoplasm of the clones, and caveolae, which are unique to cells of mesenchymal origin, were present on the surface [5,6].

NLCs have a number of unique functional activities that could contribute to rheumatoid inflammation. Among these activities are their ability to promote antibody production by B cells, the capacity to protect lymphocytes from apoptosis, the ability to secrete large amounts of cytokines and chemokines that could promote the accumulation and activation of lymphocytes and monocytes, and their unique capacity to promote the differentiation of osteoclasts from myeloid precursors in a receptor activator of NF- κ B/receptor activator of NF- κ B ligand (RANKL)-independent manner [10].

Multipotent mesenchymal stem cells from bone marrow were also found to exist in the synovial membrane [11-14]. Those

cells were shown to have multipotency to develop into various cells such as cartilage, bone, fat, and muscle. Although it is currently unknown whether these cells can differentiate into NLCs, RA-NLCs are a more differentiated population. Multipotential mesenchymal stem cells from the synovial fluid and bone marrow of patients with inflammatory and degenerative arthritis were reported to be negative for CD45 and to be positive for D7-FIB, CD13, CD105, CD55, and CD10 [13]; these mesenchymal stem cells therefore have a very different phenotype from that of RA-NLCs mentioned in the following.

Surface phenotype of rheumatoid arthritis nurse-like cells

RA-NLC clones from bone marrow and synovial tissue [5-7] expressed CD29, CD44, CD49c, CD54, CD106, and HLA-A, HLA-B, and HLA-C (class I major histocompatibility complex), but did not express CD1a, CD18 (LFA-1), CD35, CD40, CD154, or CD56. RA-NLCs constitutively expressed CD106 after long-term culture in the absence of cytokine stimulation. Constitutive expression of CD106 appears to be a characteristic appearance of nurse cell lines, permitting them to be distinguished from fibroblasts [7]. Human dermal fibroblast also expressed CD29, CD49c, CD54, and class I major histocompatibility complex, whereas constitutive expression of CD106 was minimal. IFN γ (100 U/ml) stimulation of RA-NLCs induced expression of CD40 and HLA-DR (class II major histocompatibility complex), but not expression of CD35 or CD154. The surface phenotype of RA-NLCs was therefore similar to that of FSCs derived from synovial tissue and bone marrow cells from non-RA controls. Namely, the phenotype of NLCs derived from osteoarthritis patients and human skin nurse cells was similar to that of RA-NLCs. Enhanced expression of CD106 and CD157 by IFN γ (mentioned below) was the characteristic observation in RA-NLCs and was different from human dermal fibroblasts [7].

Expression of CD106 by RA-NLCs was modestly enhanced by culture with normal peripheral B cells, and was markedly enhanced by IFN γ . In contrast, expression of CD106 by human dermal fibroblasts was much less marked after stimulation with IFN γ or by culture with peripheral B cells. One of the features of NLCs is their capacity to promote the survival of B lymphocytes [5-7]. Such B-cell survival was reduced by a blocking anti-CD106 mAb to the same level as B cells cultured in medium alone.

One notable product of NLCs is human bone marrow fibroblastic stromal cell antigen 1 (BST-1). This product was originally cloned from a human bone marrow FSC cell line by surveying for any unknown factors [15], supporting the FSC-dependent growth of the murine pre-B-cell line DW34. A new growth factor was identified, having the ability to enhance DW34 cell growth, and it was designated BST-1 [16]. Human BST-1 is expressed in various tissues and cell

lines, such as umbilical vein endothelial cells, myeloid cells, as well as FSCs of bone marrow and also synovial cells in RA, but is not expressed in lymphoid cell lines. Notably, serum levels of BST-1 were higher (30-fold to 50-fold) in 7% of RA patients than in non-RA samples [17]. Human BST-1 was later designed as CD157, and the human *Bst-1* gene was assigned to chromosome 4q15, regulating humoral immune responses *in vivo* [18]. Expression of CD157 (BST-1) was detected on all RA-NLCs, as well as on human dermal fibroblasts. Expression of CD157 by RA-NLCs, but not by dermal fibroblasts, was enhanced by IFN γ . This enhancement was much more marked with bone marrow-derived RA-NLCs compared with synovium-derived RA-NLCs. It should be noted that expression of CD106 and CD157 mRNA was found in all RA-NLC clones. Soluble CD157 together with RA-NLCs further increased the survival of B cells, which was reduced by a blocking anti-CD157 polyclonal antibody [7].

Cytokine production by nurse-like cells of RA patients

RA-NLCs produced numerous cytokines [5-7]. RA-NLCs from both bone marrow and synovial tissue produced detectable levels of IL-6, IL-8, and granulocyte/macrophage colony-stimulating factor (GM-CSF), and the production of IL-6 and IL-8 was quite robust. RA-NLCs from bone marrow but not synovial tissue produced IL-7, whereas RA-NLCs from synovial tissue produced granulocyte colony-stimulating factor and a greater amount of IL-6. Regulation of the production of cytokines was examined by co-culture of RA-NLCs from synovial tissue in direct contact with B cells. Secretion of IL-6, IL-8, granulocyte colony-stimulating factor, and GM-CSF was markedly increased by co-culture with B cells. IL-1 β and TNF were only detected in the culture supernatants after co-culture with B cells. The effect of co-culture with B lymphocytes on the secretion of cytokines and immunoglobulin production by the B cells were examined under various culture conditions [5-7] (Table 1). After co-culture with B cells, the levels of IL-6, IL-8, granulocyte colony-stimulating factor, GM-CSF, and the levels of IgM were increased, and IL-1 β and TNF were detected. Direct contact with the B-cell clone was required for RA-NLCs to produce IL-1 β and TNF and higher levels of the other cytokines.

Inhibition of spontaneous apoptosis of lymphocytes and the effect of adhesion molecules

RA-NLCs were found to promote lymphocyte viability. Although peripheral blood B cells cultured in medium alone rapidly died, culture of B cells with RA-NLCs markedly increased the B-cell viability. The loss of viability of B cells cultured alone related to the induction of apoptosis, whereas co-culture of B cells with RA-NLCs substantially blocked their apoptosis. The mechanism of the prevention of apoptosis of B cells involved the contact-dependent upregulation of Bcl-x $_L$ by RA-NLCs [19].

The regulation of pseudoemperipolesis (adhesion and holding beneath) by RA-NLCs was examined using MC/car cells and a cloned RA-NLC line from synovial tissue [20]. Pretreatment with anti-CD29 (integrin β_1 chain) or anti-CD49d (integrin α_4 chain) reduced adhesion by MC/car cells by approximately 50%. This result indicated that integrin $\alpha_4\beta_1$ (very late antigen 4) on MC/car cells was involved, at least in part, in the cells' ability to participate in pseudoemperipolesis with RA-NLCs, although such interactions were not involved in IL-6 and IL-8 production by RA-NLCs. Pretreatment of MC/car cells with the Rho-specific inhibitor C3 transferase significantly inhibited the migration of MC/car cells underneath RA-NLCs in a concentration-dependent manner, whereas the same treatment did not inhibit the adhesion of the MC/car cells to RA-NLCs. In addition, RA-NLCs produced comparable levels of IL-6 and IL-8 when co-cultured with C3-treated transmigration-defective MC/car cells. The processes of pseudoemperipolesis, adhesion and holding beneath were therefore thought to be independent events [20]. Moreover, very late antigen 4 ($\alpha_4\beta_1$)-independent lymphocyte adhesion and not holding beneath induced the enhanced proinflammatory cytokine production by the RA-NLCs [20].

Regarding NLCs, another group reported that CD14(+) monocytes could differentiate into NLCs and support the viability of chronic lymphocytic leukemia B cells [21-23], and also support the viability of primary B cells in RA [24,25]. These effects were dependent on interactions between RA-NLC-expressed CD106 and B-cell-expressed very late antigen 4 [24], which were quite similar to the interactions between RA-NLCs and B cells we had previously reported [7]. Although the other group's NLCs were identified to be derived from CD14 myelomonocytic cells [22,23,25] we have not yet clarified the stem cell of our RA-NLCs, but it clearly appears to be of mesenchymal origin [5,6].

RANKL-independent differentiation of osteoclast-like cells supported by RA nurse-like cells

RA-NLCs also promoted a specific pathway of the differentiation of CD14(+) monocytes. After 3-4 weeks of co-culture, CD14(+) monocytes differentiated into tartrate-resistant acid phosphatase (TRAP)(+) mononuclear cells with abundant cytoplasm and an off-center nucleus without the involvement of RANKL. It was noted that RA-NLCs supported such differentiation of peripheral blood CD14(+) monocytes not only from RA patients, but also from normal control subjects [10]. The second step of differentiation from such TRAP(+) mononuclear cells into multinucleated bone-resorbing giant cells (osteoclast-like cells) could also be induced without RANKL in the presence of IL-3, IL-5, IL-7, or GM-CSF, and was inhibited by mAb to each cytokine [10]. Differentiation of these TRAP(+) mononuclear cells into multinucleated bone-resorbing giant cells could also be promoted by macrophage colony-stimulating factor and RANKL [26].

Table 1

Effects of co-culture on production of cytokines from rheumatoid arthritis nurse-like cells (RA-NLCs)

	Cytokines in cell culture supernatant (pg/ml) ^a								IgM (μg/ml) ^a		
	IL-1α	IL-1β	IL-6	IL-7	IL-8	G-CSF	GM-CSF	TNFα	TNFβ	Experiment 2	Experiment 3
Cytokine production from RA-NLCs derived from synovium and immunoglobulin from B cells ^b [6]											
RA-SNCs	<5.0	<10.0	2,200		4,300	460	40	<5.0	<5.0	<1.5	<1.5
B cells	<5.0	<10.0	<10.0		<10.0	<10.0	<2.5	<5.0	<5.0	1.8	2.7
B cells + RA-SNCs (separated) ^c	<5.0	<10.0	1,800		3,900	510	30	<5.0	<5.0	<1.5	<1.5
B cells + RA-SNCs	<5.0	153	15,900		34,500	2,400	740	690	<5.0	5.6	8.6
Cytokine production from RA-NLCs derived from bone marrow cells ^d [5]											
RA-BMNC-1 cell line	-	-	38,250	-	1,480	-	150	-			
+ MC/car cell line	-	320	89,015	-	33,510	755	915	275			
+ Molt-17 cell line	-	235	78,750	-	10,615	540	355	255			

RA-BMNCs, cytokine production from RA-NLCs derived from bone marrow cells; RA-SNCs, cytokine production from RA-NLCs derived from synovium; G-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte/macrophage colony-stimulating factor; -, not detectable.

^aThe amount of each cytokine and IgM in the culture supernatant was measured with an enzyme-linked immunosorbent assay kit. ^bB-cell clones (1×10^5) and RA-SNC3 (5×10^4) were cultured under the indicated conditions for 3 days in 24-well plates. ^cB-cell clones were cultured in a Millicell culture insert. ^dRA-BMNC cells (3×10^4 cells/well) were inoculated and cultured overnight, and 1×10^6 cells MC/car cells or Molt-17 cells were added to the culture. After 5 days of incubation, the culture supernatants were collected and the amount of each cytokine in the culture supernatant was measured with an enzyme-linked immunosorbent assay kit.

Expression of MMP-2, MMP-9, and MMP-12 was increased in both TRAP(+) mononuclear and multinucleated cells after differentiation by culture with RA-NLCs, and these cells could induce cartilage degeneration *in vitro* by a mechanism that was completely blocked by inhibitors of MMP-2 and MMP-9. Although MMP-2 expression was significantly increased in TRAP(+) mononuclear cells, expression of MMP-9 and MMP-12 was also higher in TRAP(+) multinucleated cells [27]. Of note, both TRAP(+) mononuclear and multinucleated cells differentiated by culture with RA-NLCs specifically expressed MMP-12 [27], whereas multinucleated cells expressing MMP-12 were clearly found near the bone erosions (S Yamane, M Maeda-Tanimura, Y Shimaoka, M Yukioka, T Toyosaki-Maeda, S Ishida, N Yamane, Y Tsuruta, T Itoh, N Fukui, *et al.*, unpublished observation). RA-NLCs were therefore found to promote the differentiation of CD14(+) monocytes in a characteristic two-step differentiation process into multinucleated osteoclast-like cells with the capacity to degrade bone and cartilage.

Although TNF [28], IL-1 [29], macrophage colony-stimulating factor, and RANKL [30] are very important factors for developing osteoclasts, the RANKL-independent two-step differentiation of CD14(+) monocyte supported by RA-NLCs [10,26] may be an alternative pathway to develop multinucleated osteoclast-like cells specifically in RA. Beside the destruction of bone tissue by osteoclasts or osteoclast-like cells, we could confirm that FSCs from RA patients inoculated *in vivo* showed aggressive behavior, invading

cartilage as reported previously [31-33], although we have not yet confirmed that pure RA-NLC lines have such function.

Comparison of the properties of RA nurse-like cells and fibroblast-like synoviocytes

A considerable amount of work has characterized another population of cells found in the rheumatoid synovium, namely fibroblast-like synoviocytes. The cells are thought to play a role in rheumatoid pathogenesis, especially because of their capacity to contribute to tissue damage [31-33]. RA-NLCs, however, have a number of specific attributes that suggest they may play a unique role in RA pathogenesis (Table 2).

Mechanisms of progressive proliferation of fibroblastic stromal cells specifically found in joint

To explain the remarkable proliferation of synovial tissue in the RA patient, various mechanisms have been reported such as the involvement of protooncogenes [34], inflammatory cytokines [35], and perturbations of Fas-mediated apoptosis [36]. As a mechanism specifically found in the synovial space but not in the bone marrow, we found that the interference with Fas-mediated apoptosis could upregulate specifically the growth of synovial FSCs [37,38]. In this regard, soluble Fas ligand was found to inhibit competitively the Fas-Fas ligand-mediated apoptosis [37] of FSCs bearing Fas. The levels of human soluble Fas ligand in synovial fluid from RA patients were found to be significantly higher than those from osteoarthritis patients.

Table 2**Comparison of the properties of rheumatoid arthritis nurse-like cells and fibroblast-like synoviocytes**

Property	Rheumatoid arthritis nurse-like cells	Fibroblast-like synoviocytes
Pseudoemperipolesis	+	-
Constitutive expression of CD106	+	-
Enhanced expression of CD106 and CD157 by IFN γ	+	-
Promote B-cell differentiation	+	-
Promote differentiation of osteoclast-like cells from CD14(+) monocytes	+	-
Inhibit lymphocyte apoptosis	+	-

In contrast, soluble Fas ligand was not detected in the peripheral blood, and also not in bone marrow blood in RA patients [38]. This mechanism, therefore, could at least partially upregulate the FSC growth in synovial tissue, but not in bone marrow.

Conclusion

A specific population of FSCs, RA-NLCs reside in both the bone marrow and synovium of RA patients and have the functional capacity to interact with lymphocyte and monocyte populations, inducing cellular differentiation and biologic activities that mimic pathophysiologic features of rheumatoid inflammation. These findings suggest that RA-NLCs may play an essential role in the development of local immune and inflammatory responses in the synovium and the bone marrow. RA-NLCs could therefore be central elements in the pathologic events in RA and might be appropriate targets for therapeutic intervention in RA.

Competing interests

The authors declare that they have no competing interests.

Acknowledgements

The work reported here has been supported in part by a grant-in-aid from the Health Science Research grant from the Ministry of Health and Welfare of Japan. The authors are grateful for the great collaboration and support of the people listed in each paper related to this review. Among them, we are especially grateful to Dr T Kishimoto, Dr T Hirano, Dr S Nagata, Dr T Suda, Dr M Miyasaka, Dr T Kaisho, and Dr K Ishihara of Osaka University Medical School, and to Dr R Suzuki and Miss T Uchida of the Research Center, Sagami National Hospital.

This review is part of a series on
Mesenchymal stromal cells
edited by Steffen Gay.

Other articles in this series can be found at
[http://arthritis-research.com/articles/
review-series.asp?series=ar_Mesenchymal](http://arthritis-research.com/articles/review-series.asp?series=ar_Mesenchymal)

References

- Eisenberg RA, Cohen PL: The role of immunologic mechanisms in the pathogenesis of rheumatic disease. In *Primer on the Rheumatic Diseases*. 10th edition, Edited by Schumacher HR, Klippel JH, Koopman WJ. Atlanta: Arthritis Foundation; 1993:27-35.
- Genant HK: Radiology of rheumatic diseases. In *Arthritis and Allied Conditions*. 9th edition, Edited by McCarty DJ. Philadelphia: Lea & Febiger; 1979:70-130.
- Trentham D, Townes A, Kang A: Autoimmunity to type II collagen: an experimental model of arthritis. *J Exp Med* 1977, **146**: 857-868.
- Nakagawa S, Toritsuka Y, Wakitani S, Denno K, Tomita T, Owaki H, Kimura T, Shino K, Ochi T: Bone marrow stromal cells contribute to synovial cell proliferation in rats with collagen induced arthritis. *J Rheumatol* 1996, **23**:2098-2103.
- Tomita T, Takeuchi E, Toyosaki-Maeda T, Oku H, Kaneko M, Takano H, Sugamoto K, Ohzono K, Suzuki R, Ochi T: Establishment of nurse-like stromal cells from bone marrow of patients with rheumatoid arthritis: indication of characteristic bone marrow microenvironment in patients with rheumatoid arthritis. *Rheumatology* 1999, **38**:854-963.
- Takeuchi E, Tomita T, Toyosaki-Maeda T, Kaneko M, Takano H, Hashimoto H, Sugamoto K, Suzuki R, Ochi T: Establishment and characterization of nurse cell-like stromal cell lines from synovial tissues of patients with rheumatoid arthritis. *Arthritis Rheum* 1999, **42**:221-228.
- Shimaoka Y, Attrep JF, Hirano T, Ishihara K, Suzuki R, Toyosaki T, Ochi T, Lipsky PE: Nurse-like cells from bone marrow and synovium of patients with rheumatoid arthritis promote survival and enhance function of human B cells. *J Clin Invest* 1998, **102**:606-618.
- Wekerle H, Ketelsen UP: Thymic nurse cells - Ia bearing epithelium involved in T-lymphocyte differentiation? *Nature* 1980, **283**:402-404.
- Wekerle H, Ketelsen UP, Ernst M: Thymic nurse cells. Lymphoepithelial cell complexes in murine thymuses: morphological and serological characterization. *J Exp Med* 1980, **161**: 925-944.
- Toyosaki-Maeda T, Takano H, Tomita T, Tsuruta Y, Maeda-Tanimura M, Shimaoka Y, Takahashi T, Iton T, Suzuki R, Ochi T: Differentiation of monocytes into multinucleated giant bone-resorbing cells: two-step differentiation induced by nurse-like cells and cytokines. *Arthritis Res* 2001, **3**:306-310.
- De Bari C, Dell'Accio F, Tylzanowski P, Luyten FP: Multipotent mesenchymal stem cells from adult human synovial membrane. *Arthritis Rheum* 2001, **44**:1928-1942.
- De Bari C, Dell'Accio F, Vanlauwe J, Eyckmans J, Khan IM, Archer CW, Jones EA, McGonagle D, Mitsiadis TA, Pitzalis C, Luyten FP: Mesenchymal multipotency of adult human periosteal cells demonstrated by single-cell lineage analysis. *Arthritis Rheum* 2006, **54**:209-1221.
- Jones EA, English A, Henshaw K, Kinsey SE, Markham AF, Emery P, McGonagle D: Enumeration and phenotypic characterization of synovial fluid multipotential mesenchymal progenitor

- cells in inflammatory and degenerative arthritis. *Arthritis Rheum* 2004, **50**:817-827.
14. Jones EA, English A, Kinsey SE, Straszynski L, Emery P, Ponchel F, McGonagle D: Optimization of a flow cytometry-based protocol for detection and phenotypic characterization of multipotent mesenchymal stromal cells from human bone marrow. *Cytom Part B (Clin Cytom)* 2006, **70B**:391-399.
 15. Kaisho T, Oritani K, Ishikawa J, Tanabe M, Muraoka O, Ochi T, Hirano T: Human bone marrow stromal cell lines from myeloma and rheumatoid arthritis that can support murine pre-B cell growth. *J Immunol* 1992, **149**:4088-4095.
 16. Kaisho T, Ishikawa J, Oritani K, Inazawa J, Tomizawa H, Muraoka O, Ochi T, Hirano T: BST-1, a surface molecule of bone marrow stromal cell lines that facilitates pre-B-cell growth. *Proc Natl Acad Sci USA* 1994, **91**:5325-5329.
 17. Lee BO, Ishikawa K, Denno K, Kobune Y, Itoh M, Muraoka O, Kaisho T, Sasaki T, Ochi T, Hirano T: Elevated levels of the soluble form of bone marrow stromal cell antigen 1 in the sera of patients with severe rheumatoid arthritis. *Arthritis Rheum* 1996, **39**:629-637.
 18. Ishihara K, Hirano T: BST-1/CD157 regulates the humoral immune responses in vivo. *Chem Immunol* 2000, **75**:235-255.
 19. Hayashida K, Shimaoka Y, Ochi T, Lipsky PE: Rheumatoid arthritis synovial stromal cells inhibit apoptosis and up-regulate Bcl-xL expression by B cells in a CD49/CD29-CD106-dependent mechanism. *J Immunol* 2000, **164**:1110-1116.
 20. Takeuchi E, Tanaka T, Umamoto E, Tomita T, Shi K, Takahi K, Suzuki R, Ochi T, Miyasaka M: VLA-4-dependent and -independent pathways in cell contact-induced proinflammatory cytokine production by synovial nurse-like cells from rheumatoid arthritis patients. *Arthritis Res* 2002, **4**:1-8.
 21. Burger JA, Tsukada N, Burger M, Zvaifler NJ, Dell'Aquila M, Kipps TJ: Blood-derived nurse-like cells protect chronic lymphocytic leukemia B cells from spontaneous apoptosis through stromal cell-derived factor-1. *Blood* 2000, **96**:2655-2663.
 22. Tsukada N, Burger JA, Zvaifler NJ, Kipps TJ: Distinctive features of 'nurselike' cells that differentiate in the context of chronic lymphocytic leukemia. *Blood* 2002, **99**:1030-1037.
 23. Nishio M, Endo T, Tsukada N, Ohata J, Kitada S, Reed JC, Zvaifler NJ, Kipps TJ: Nurselike cells express BAFF and APRIL, which can promote survival of chronic lymphocytic leukemia cells via a paracrine pathway distinct from that of SDF-1 α . *Blood* 2005, **106**:1012-1020.
 24. Burger JA, Zvaifler NJ, Tsukada N, Firestein GS, Kipps TJ: Fibroblast-like synoviocytes support B-cell pseudoemperipolesis via a stromal cell-derived factor-1 and CD106 (VCAM-1)-dependent mechanism. *J Clin Invest* 2001, **107**:305-315.
 25. Ohata J, Zvaifler NJ, Nishio M, Boyle DL, Kalled SL, Carson DA, Kipps TJ: Fibroblast-like synoviocytes of mesenchymal origin express functional B cell-activating factor of the TNF family in response to proinflammatory cytokines. *J Immunol* 2005, **174**:864-870.
 26. Tsuboi H, Udagawa N, Hashimoto J, Yoshikawa H, Takahashi N, Ochi T: Nurse-like cells from patients with rheumatoid arthritis support the survival of osteoclast precursors via macrophage colony-stimulating factor production. *Arthritis Rheum* 2005, **52**:3819-3828.
 27. Tsuboi H, Matsui Y, Hayashida K, Yamane S, Maeda-Tanimura M, Nampei A, Hashimoto J, Suzuki R, Yoshikawa H, Ochi T: Tartrate resistant acid phosphatase (TRAP) positive cells in rheumatoid synovium may induce the destruction of articular cartilage. *Ann Rheum Dis* 2003, **62**:196-203.
 28. Boyce BF, Li P, Yao Z, Zhang Q, Badell IR, Schwartz EM, O'Keefe RJ, and Xing L: TNF α and pathologic bone resorption. *Keio J Med* 2005, **54**:127-131.
 29. Wei S, Kitaura H, Zhou P, Ross P, Teitelbaum SL: IL-1 mediates TNF-induced osteoclastogenesis. *J Clin Invest* 2005, **115**:282-290.
 30. Saldenbergh-Kermanac'h N, Cohen-Solal M, Bessis N, De Vernejoul MC, Boissier MC: Role for osteoprotegerin in rheumatoid inflammation. *Joint Bone Spine* 2004, **71**:9-13.
 31. Gay S, Gay RE, Koopman WJ: Molecular and cellular mechanism of joint destruction in rheumatoid arthritis: two cellular mechanisms explain joint destruction? *Ann Rheum Dis* 1993, **52**:39-47.
 32. Firestein GS: Invasive fibroblast-like synoviocytes in rheumatoid arthritis. Passive responders or transformed aggressors? *Arthritis Rheum* 1996, **39**:1781-1790.
 33. Shigeyama Y, Pap T, Kunzler P, Rethage J, Simmen B, Gay RE, Gay S: Rheumatoid arthritis (RA) synovial fibroblasts express osteoclast differentiating factor (ODF) mRNA at sites of joint destruction [abstract]. *Arthritis Rheum* 1999, **42**:283.
 34. Gay S, Gay RE: Cellular basis and oncogene expression of rheumatoid joint destruction. *Rheumatol Int* 1989, **9**:105-113.
 35. Farahat MN, Yanni G, Poston R, Panayi GS: Cytokine expression in synovial membranes of patients with rheumatoid arthritis and osteoarthritis. *Ann Rheum Dis* 1993, **52**:870-875.
 36. Nagata S, Suda T: Fas and Fas ligand: lpr and gld mutations. *Immunol Today* 1995, **16**:39-43.
 37. Suda T, Hashimoto H, Tanaka M, Ochi T, Nagata S: Membrane Fas ligand kills human peripheral blood T lymphocytes, and soluble Fas ligand blocks the killing. *J Exp Med* 1997, **186**:2045-2050.
 38. Hashimoto H, Tanaka M, Suda T, Tomita T, Hayashida K, Takeuchi E, Kaneko M, Takano H, Nagata S, Ochi T: Soluble fas ligand in the joints of patients with rheumatoid arthritis and osteoarthritis. *Arthritis Rheum* 1998, **41**:657-662.

IV—II 分担研究刊行物

IV-II 分担研究刊行物

A) RA・骨粗鬆症の臨床的研究

- 1) RA重症化の早期診断マーカー開発研究
- 2) 薬物治療法確立研究
- 3) 人工骨開発適用研究

第7回日本骨粗鬆症学会奨励賞受賞演題

アレンドロネート投与後の骨量改善に伴う
動脈硬化進展に及ぼす影響について

山田真介¹⁾・稲葉雅章¹⁾・後藤仁志¹⁾・田中一匡¹⁾
酒井真礼²⁾・白川久美¹⁾・今西康雄¹⁾・西沢良記¹⁾

はじめに

以前より、骨粗鬆症の進展と動脈硬化進展の関連が示唆されている。たとえば、脳血管障害による片側不全麻痺患者では、身体活動性の低下した麻痺側において、健側と比較し有意な骨量の減少と動脈硬化の進展を認めており、両者の間には有意な負の相関を認める¹⁾。同様に、関節リウマチ患者(RA患者)では、健常人と比較して骨量が低下し²⁾、動脈壁肥厚度が進展している³⁾。つまりRA患者においても、骨量の減少と動脈硬化進展の間には何らかの関連が存在することが示唆される。また、一般女性と比較して女性RA患者では、心血管病変による死亡リスクが高いという報告がある⁴⁾。つまり、RA患者では一般の人々と比較し、動脈硬化の管理が非常に重要であることが示唆される。

1 RA合併症検査

当科ではRAに伴う内科合併症を検討する目的で、通院中のRA患者に対し動脈硬化の進展度と骨量を定期的に検査している。動脈硬化の評価については、動脈壁硬化度の指標となる上腕-足首間の脈波伝播速度(ba PWV)を測定し、骨量については体幹の骨量として腰椎骨密度(腰椎

BMD)を、四肢の骨量として大腿骨・踵骨の骨密度(大腿骨BMD)、踵骨OSIを測定している。また、骨代謝の状態を検討するために、骨吸収の指標として尿中NTX/Creを、骨形成の指標としてBAPを測定している。

2 健常人とRA患者の骨量と動脈壁硬化度の比較

現在、当科通院加療中の閉経後RA患者47名と、年齢・BMI・収縮期血圧をマッチさせた、一般の閉経後女性47名の骨量と動脈壁硬化について比較検討してみると、RA患者では有意に腰椎BMDおよび踵骨OSIが低下し、ba PWVが上昇していることがわかる(表1)。さらに、腰椎BMDも踵骨OSIもba PWVとの間に有意な負の相関を認めている(表2)。つまり、RA患者は身体活動性の低下などを危険因子とし早期から骨量減少をきたしており、その程度に伴い動脈硬化が進展していると示唆される。

3 目的と方法

RA患者の骨量を改善することにより、動脈壁硬化の進展を予防する可能性も示唆されることから、骨吸収抑制剤であるアレンドロネート

Influence on Atherosclerosis with the Improvement of Bone Mineral Density after Medication for Osteoporosis in Arthritis Rheumatism Patient

Shinsuke Yamada: Department of Metabolism, Endocrinology, and Molecular Medicine Osaka City University Graduate School of Medicine, et al.

Key words : Osteoporosis, Atherosclerosis, Alendronate, Arthritis rheumatism

¹⁾大阪市立大学大学院医学研究科代謝内分泌病態内科学, ²⁾阪和第二泉北病院

表1 健常人とRA患者の骨量と動脈硬化進展の比較

	コントロール	RA	p
人数(閉経後女性)	47	47	
年齢(歳)	60.0 ± 5.9	60.0 ± 9.7	0.9898
BMI(kg/m ²)	22.1 ± 3.3	21.3 ± 3.3	0.2628
収縮期血圧(mmHg)	125.0 ± 19.8	128.9 ± 20.3	0.3551
Total cholesterol(mg/dL)	224.9 ± 29.6	204.3 ± 32.9	0.0024
CRP(mg/dL)	-	1.3 ± 1.9	-
U-NTX/Cre(nmol BCE/mmol・Cr)	-	83.9 ± 37.0	-
腰椎BMD(g/cm ²)	0.874 ± 0.100	0.820 ± 0.154	0.0498
踵骨OSI(×10 ⁶)	2.495 ± 0.269	2.196 ± 0.266	<0.0001
ba PWV(cm/s)	1301.1 ± 214.6	1573.0 ± 381.9	<0.0001

表2 ba PWVと各パラメータの相関

	ba PWV(cm/s)			
	コントロール		RA	
	r	p	r	p
年齢(歳)	0.284	0.0532 ^s	0.627	<0.0001 [†]
BMI(kg/m ²)	-	0.5140	-	0.7466
収縮期血圧(mmHg)	0.595	<0.0001 [†]	0.625	<0.0001 [†]
Total Chol(mg/dL)	-	0.8802	-	0.2529
CRP(mg/dL)	-	-	-	0.5697
腰椎BMD(g/cm ²)	-	0.1461	-0.299	0.0460 [†]
踵骨OSI(×10 ⁶)	-	0.6007	-0.309	0.0437 [†]

[†]p<0.0001, [†]p<0.05, ^sp<0.1

(5mg/日)を投与し、動脈壁硬化の進展を予防しうるか否かにつき検討した。対象はRA患者16名、アレンドロネート非投与群、アレンドロネート投与群おのおの8名ずつ、すべて女性とした。アレンドロネート投与前および投与半年から1年後において、ba PWV、腰椎・大腿骨BMD、尿中NTX/Cre、BAPを測定した。対象の開始時の背景を表3に示す。

4 結 果

観察前後での各種パラメータの結果を表4、5に示す。血清CRP値は、非投与群・投与群ともにほとんど変化を認めなかった。骨代謝の状態については、非投与群において、NTX/CreおよびBAPとも有意な上昇を認め、骨代謝が亢進していることが示唆された。一方、投与群では、

NTX/Creは有意に減少しており、骨吸収が抑制されていることが示唆された。腰椎BMDについては、非投与群では変化を認めなかったが、投与群では有意な上昇を認めた。大腿骨BMDにおいても、非投与群では変化を認めなかったが、投与群では上昇傾向を認めた。つまり、投与群ではアレンドロネートが骨吸収を抑制することにより骨量が増加したと考えられた。ba PWVの変化については、統計学的に有意な結果とはならなかったが、その平均値は非投与群において約4%の増加を示し、投与群では約8%の減少を認めた。また、腰椎BMDとba PWVそれぞれの増加率は有意な負の相関(r=0.671, p=0.0337)を認めた。つまり、骨量が増加すればするほど動脈壁硬化が改善することが示唆された。